British Society for Haematology

DOI: 10.1002/jha2.252

REVIEW ARTICLE



The beginning of a new therapeutic era in acute myeloid leukemia

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Abstract

In the field of AML, the early 2000s were shaped by the advent of novel molecular biology technologies including high-throughput sequencing that improved prognostic classification, response evaluation through the quantification of minimal residual disease, and the launch of research on targeted therapies. Our knowledge of leukemogenesis, AML genetic diversity, gene-gene interactions, clonal evolution, and treatment response assessment has also greatly improved. New classifications based on chromosomal abnormalities and gene mutations are now integrated on a routine basis. These considerable efforts contributed to the discovery and development of promising drugs which specifically target gene mutations, apoptotic pathways and cell surface antigens as well as reformulate classical cytotoxic agents. In less than 2 years, nine novels drugs have been approved for the treatment of AML patients, and many others are being intensively investigated, in particular immune therapies. There are now numerous clinical research opportunities offered to clinicians, thanks to these new treatment options. We are only at the start of a new era which should see major disruptions in the way we understand, treat, and monitor patients with AML.

KEYWORDS

acute myeloid leukemia, CPX-351, enasidenib, FLT3 inhibitors, gemtuzumab ozogamycin, gilteritinib, glasdegib, IDH inhibitors, ivosidenib, midostaurin, monoclonal antibodies, oral azacitidine, TP53, venetoclax

1 | INTRODUCTION

It has long been written that there was no breakthrough for the treatment of acute myeloid leukemia (AML) as compared to other hematological malignancies including chronic myeloid leukemia, B-cell lymphoma (BCL), or multiple myeloma [1]. Going back more than 20 years ago, the situation was roughly similar for multiple myeloma. Younger patients were treated with high-dose melphalan and autologous stem-cell transplantation while older patients received low-dose melphalan and prednisone, the so-called MP regimen that has fallen

into oblivion over the years [2]. Now, there have been more than 10 novel drugs approved in myeloma targeting intracellular pathways, tumor microenvironment, and cell surface antigens the combinations of which have transformed a highly deadly disease into a chronic one [3, 4]. In AML, we have been relying for the past 40 years on the combination of cytarabine and an anthracycline, so-called ("7+3"), as induction chemotherapy in patients suitable for intensive treatments followed by high-dose cytarabine consolidation and eventually by allogeneic stem cell transplantation (cure being the goal), whereas older or unfit patients received low-dose cytarabine or more recently

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CELGENE 2110 APOTEX v. CELGENE IPR2023-00512

TABLE 1 Novel drugs recently approved in acute myeloid leukemia

Agent	Approval	Mechanism of action	Indication
Midostaurin	2017	FLT3 inhibition	Newly diagnosed AML with <i>FLT3</i> mutation in combination with standard induction, consolidation +/-maintenance
Enasidenib	2017	IDH2 mutant inhibition	Relapsed or refractory AML with IDH2 mutation
CPX-351	2017	Liposomal formulation including daunorubicin and cytarabine at a fixed 5-molar:1-molar ratio	Newly diagnosed, therapy-related AML (t-AML) or AML with myelodysplasia-related
changes (AML-MRC)			
Gemtuzumab ozogamicin	2017	CD33 monoclonal antibody linked to calicheamicin	Newly diagnosed, CD33-positive AML in combination with standard induction and relapsed or refractory CD33-positive AML
lvosidenib	2018	IDH1 mutant inhibition	Relapsed or refractory or newly diagnosed (unfit) AML with <i>IDH1</i> mutation
Glasdegib	2018	Hedgehog pathway inhibition	Unfit or older (≥75y) patients with newly diagnosed AML in combination with low-dose cytarabine
Venetoclax	2018	Bcl-2 inhibition	Unfit or older (≥75y) patients with newly diagnosed AML in combination with hypomethylating agents or low-dose cytarabine
Gilteritinib	2018	FLT3 inhibition	Relapsed or refractory AML with FLT3 mutation
Oral azacitidine	2020	Hypomethylating agent	Continued treatment of AML patients who achieved CR/CRi following intensive induction chemotherapy and are not able to complete intensive curative therapy (i.e., alloSCT)

hypomethylating agents, both inducing few complete responses (CRs) and little hope for cure (prolonged survival being the goal) [5]. Then, genomics changed the game. Firstly, targeted gene sequencing successively identified FLT3-ITD, CEBPA, and NPM1 mutations which helped improve prognostic classification, the response evaluation through quantification of molecular residual disease, and launched the research on targeted therapies leading to the registration of midostaurin as the first FLT3 inhibitor in AML [6, 7]. Soon after, thanks to the advance in high-throughput sequencing technologies, the first AML genome was reported in 2008 and subsequent studies identified several novel recurrent mutations with pathophysiological, prognostic, or therapeutic relevance such as IDH1, IDH2, or TP53 mutations [8–10]. Our knowledge of leukemogenesis, AML genetic diversity, gene-gene interactions, clonal evolution, and treatment response assessment has also greatly improved, and new classifications based on chromosomal abnormalities and gene mutations are now integrated in clinical environments [11-17]. These considerable efforts contributed to the discovery and development of promising drugs for AML specifically targeting gene mutations, apoptotic pathways, and cell surface antigens. Novel liposomal formulations of classical cytotoxic agents are also promising. Midostaurin, gemtuzumab ozogamycin, glasdegib, venetoclax, ivosidenib, enasidenib, gilteritinib, CPX-351, and oral azacitidine were approved by the Food and Drug Administration (FDA) of for AML patients in less than 3 years between 2017 and 2020 [18, 19] (Table 1).

Rather than just another review stacking up one novel drug after another, this article will try to outline the perspectives for the coming years in the field of AML [20, 21]. Indeed, there are now numerous clinical research opportunities offered to clinicians with these new molecules and with those under development. With this sudden over-abundance of choices, we feel like a lottery player who gets rich overnight and wonders what he's going to do with it all!

2 VENETOCLAX IN AML, THE MAGIC POTENTIATOR

The anti-apoptotic BCL 2 (BCL-2) protein is overexpressed in AML, especially in leukemic stem cells that are supposed to be responsible for chemoresistance and relapse. BCL-2 overexpression is a poor risk factor in AML and is associated with chemoresistance [22]. BCL-2 inhibition by small molecule inhibitors kills AML blasts, targets

oxidative phosphorylation, and selectively eradicates leukemic stem cells [23, 24]. In AML patients, contrary to chronic lymphocytic leukemia, venetoclax - an oral, selective, small-molecule inhibitor of BCL-2 - is not very active as a single agent with perhaps, the exception of NPM1 or IDH mutation subgroups [25, 26]. However, when combined to low-dose cytarabine or with hypomethylating agents, venetoclax dramatically increases the CR rates, thus demonstrating synergistic activity in patients. Venetoclax has been recently approved in combination with hypomethylating agents as first line therapy in AML patients who are ineligible to receive standard induction therapy on the basis of high response rates and promising response durations in a Phase 1b/2 trial [27, 28]. These results have been very recently confirmed in the randomized, placebo-controlled, VIALE-A Phase 3 trial which demonstrated the superiority of azacitidine plus venetoclax over azacitidine plus placebo in terms of CR rate (66.4% vs. 28.3%), duration of response (17.5 vs. 13.4 months), and OS (14.7 vs. 9.6 months) [29]. These differences are highly statistically and clinically significant. This major breakthrough is reminiscent of the one made more than 40 years ago when anthracyclines were added to cytarabine in patients eligible for intensive chemotherapy. Furthermore, the CR rate with azacitidine-venetoclax is >70% in the subgroups with IDH1 or IDH2 mutations [30]. These unprecedented results compared favorably with intensive chemotherapy in fit patients and generate hope for a cure in some patients. With the exception of the risk of tumor lysis syndrome and increased incidence of febrile neutropenia, no unexpected adverse event emerged, thus ensuring the widespread use of this combination in this difficult-to-treat population of unfit or elderly patients. Venetoclax combined with low-dose cytarabine was also superior to low-dose cytarabine in the VIALE-C Phase 3 trial [31].

Thus, doublet azacitidine-venetoclax is becoming the new standard of care in patients ineligible for intensive induction chemotherapy and the standard arm to overcome in clinical trials. Planned or on-going clinical trials are already comparing this doublet to triplets with small molecule inhibitors or monoclonal antibodies. For example, the first results of the azacitidine-venetoclax-ivosidenib triplet may reach a 100% rate of CR in treatment-naïve AML patients with *IDH1* mutations (EHA Library. DiNardo C. 06/12/20; 294963; S143).

There is an on-going debate on how to define patients unfit for intensive induction chemotherapy [32-34]. There are no absolute clear-cut criteria to unambiguously select the patients, and certain parameters used in the majority studies, such as performance status (a highly subjective criterion) or age, are questionable in this setting [34]. It has been suggesting that comorbidity, physical capacities, and nutritional status may be more relevant than performance status or age [35]. What about a 70-year-old patient with a normal karyotype, an ECOG performance status of 2 and no comorbidity? In routine, this patient is fit for chemotherapy especially when no valuable alternative is available [36]. However, patients of the VIALE-A study could be included in the trial on the sole basis of the ECOG criteria of 2; therefore it is likely that this Phase 3 trial has included a substantial number of patients that were otherwise fit for chemotherapy outside clinical trials. Therefore, many physicians will be tempted to replace intensive chemotherapy by azacitidine-venetoclax in older patients since CR rate and survival are comparable, and toxicity is likely to be lower with the doublet, although this important question should be the matter of prospective randomized trials.

Beyond the population of unfit patients, venetoclax is likely to become the drug of choice for combination therapy in virtually all AML patients. In relapse or refractory AML, the preliminary results of combinations with other small molecules inhibitors such as FLT3. IDH, or MDM2 inhibitors seem very promising [37, 38]. In patients fit for intensive chemotherapy, adding venetoclax to 7+3 is feasible and induces promising response rates, especially in patients with intermediate cytogenetic risk or with NPM1 or IDH mutations [25]. In these fit patients who have achieved CR after intensive induction chemotherapy, venetoclax in combination with intermediate dose cytarabine as consolidation therapy and/or with azacitidine as maintenance therapy so as to eradicate residual disease will be an important scope of investigation. To date, 91 clinical trials with venetoclax in AML are recorded on the clinicaltrials.gov Website, thus highlighting the tremendous interest for this drug in AML. This has been confirmed at the virtual 2020 American Society of Hematology meeting where no less than 10 oral communications dealt with venetoclax based-combinations. Venetoclax is becoming the drug of choice to combine with all kind of therapeutic strategies in AML including high or low intensity chemotherapy, novel agents, and tyrosine kinase inhibitors. In a phase 2 trial, a low-intensity backbone of cladribine/low dose cytarabine plus venetoclax alternating with azacitidine plus venetoclax for older patients with AML, yielded very high rates of durable MRD negative CR (96% CR/CRi and 80% MRD negative). With a median follow-up of 11+ months, the median OS was not reached, with 6- and 12-month OS rates of 86% and 70%, respectively [39]. Venetoclax has been also combined with intensive chemotherapy including CPX-351 or FLAG-Ida regimen [40, 41]. These combinations are feasible provided that the dose and duration of venetoclax exposure is reduced. Prolonged myelosuppression and infections remain a major problem with this type of combinations which will be reserved for suitable patients. In FLT3 mutated AML, combining venetoclax plus FLT3 inhibitors such as quizartinib or gilteritinib with or without hypomethylating agents also induced impressive response rates in R/R AML patients [42, 43]. However, AML patients with TP53 mutations did not appear to benefit from venetoclax basedtreatment. Indeed, the combination of venetoclax and decitabine was associated with inferior response rate, shorter response duration, higher MRD positivity, and a poor median OS of 5.2 months [44].

3 | IDH AND FLT3 INHIBITORS, THE SMART DRUGS WITH COMPANION BIOMARKERS

With the development and recent approval of drugs targeting specific mutations such as FLT3, IDH1, and IDH2, it has become critical that onco-hematology laboratories increase their turnaround in order to allow clinicians to prescribe these agents in a timely manner. It has long been postulated that AML represents an oncologic emergency and should be treated without delay, especially in younger patients treated by intensive chemotherapy [45]. However, recent studies have proven

that waiting a short period of time so as to characterise molecular alterations and design tailored treatments at diagnosis is safe [46, 47]. It is now crucial that clinicians be knowledgeable on a panel of gene mutations including at least *FLT3*, *IDH1*, *IDH2*, *NPM1*, *CEBPA*, and *TP53* at the time of diagnosis (48–72 h) in order to guide initial treatment. Furthermore, since AML clones and subclones are subject to clonal evolution under the therapeutic pressure of either chemotherapy or targeted agents, repeating molecular testing in each phase of the disease (refractory or relapse) should be mandatory. Recent advances in the treatment of AML with FLT3 or IDH inhibitors illustrate well this recent development.

3.1 | IDH1 inhibitors

Somatic mutations of isocitrate dehydrogenase 1 (*IDH1*^{R132}) genes are found in 6%–10% of AML patients [48]. *IDH1* mutations are most frequent in AML with normal karyotype and associated with *NPM1* and DNMT3A mutations at diagnosis [49]. Their prognostic impact mainly depends on the mutational context [50–52]. Furthermore, *IDH1* mutations, which have been described in clonal hematopoiesis, are considered as early event during leukemogenesis, stable at relapse and thus, have emerged as promising therapeutic targets. It is also noteworthy that the molecular landscape of AML with *IDH1* mutations observed at R/R disease under chemotherapy selection pressure differs from diagnosis with an increased frequency of *SRSF2*, *ASXL1*, *RUNX1*, *NRAS*, and *TP53* co-occurring mutations [49, 53].

Ivosidenib – an oral, targeted, small-molecule inhibitor of mutant IDH1 – has been evaluated as a single agent in a Phase 1 study in relapsed or refractory (R/R) AML with *IDH1* mutation [53]. The frequency of Grade 3 or higher treatment-related adverse events was low, mainly a prolongation of the QT interval, leukocytosis, and differentiation syndrome which are manageable [54]. CR or CR with partial hematologic recovery (CRh) was 30.4% with 21.8% CR, whereas CRi was 11.7%. It should be noted that clonal or subclonal m*IDH1* had similar CR/CRh rates. Furthermore, mutation clearance was observed in 21% of responding patients, thus demonstrating that deep response may be achieved in some patients. The median duration of response was 9.3 months in CR patients. The median OS was 8.8 months. Based on these promising results, ivosidenib was recently approved by the FDA.

The mechanisms of resistance to ivosidenib were studied in patients who failed to or relapsed after the response to this drug [55]. Receptor tyrosine kinase pathway mutations and mutations in *NRAS* and *PTPN11* were significantly associated with the lack of response to ivosidenib. Interestingly, emerging mutations in patients who relapsed or progressed under ivosidenib were *IDH* or non-*IDH*-related. Indeed, mutations of resistance in a second site of *IDH1* or the emergence of *IDH2*^{R140} clones were detected in ~25% of resistant patients, whereas potentially actionable mutations in genes such as *FLT3*, *NRAS*, or *KRAS* were also identified, thus indicating that molecular rescreening is important at each stage of the disease.

The preliminary results of ivosidenib combined with azacitidine when treating naive patients showed a CR rate of 70% and may reduce the emergence of mutant resistant clones [56].

3.2 | IDH2 inhibitors

Somatic mutations of the *IDH2* gene, either *IDH2*^{R140} or *IDH2*^{R172}, occur in 5%–15% and 1%–4% of AML, respectively [48]. Like IDH1, *IDH2* mutations are frequently found in normal karyotype AML [57, 58]. At diagnosis, *IDH2*^{R140} mutations are associated with *NPM1* and *DNMT3A* mutations whereas in the relapse/refractory setting, mutations of the *SRSF2*, *DNMT3A*, *RUNX1*, *ASXL1*, *NRAS*, and *BCOR* genes emerge as the most frequent co-mutations [11, 48, 59]. *IDH2*^{R172} mutations are associated with *DNMT3A* and *BCOR* mutations and mutually exclusive with *NPM1* and other class-defining mutations [49]. Therefore, AML with *IDH2*^{R172} has been recognized as a defined subgroup of the AML genomic classification [11].

Enasidenib - an oral, targeted, small-molecule inhibitor of mutant IDH2 - has been evaluated as a single agent in a Phase 1 study in mutant IDH2 R/R AML patients and subsequently approved by the FDA [60]. A low frequency of Grade 3 or higher treatment-related adverse events was reported, mainly leukocytosis and differentiation syndrome [59, 61]. The overall response rate was 40.3% including 19.3% CR and 6.8% CRi. The median OS was 9.3 months and reached 19.7 months in CR patients. CRs were observed in patients with subclonal IDH2 mutations, and a variant allele frequency of IDH2 mutant, which measures the mutational burden, was not associated with the response. Also, whereas in some CR patients, IDH2 mutation clearance was achieved, IDH2 mutational burden did not decrease in all responding patients during treatment, possibly due to the maturation of leukemic blasts into the functional neutrophils carrying the mutation. The mechanisms of resistance may involve the emergence of second-site IDH2 mutations, IDH2-mutant subclones with neomorphic mutations in IDH1, cooccurring mutations in NRAS, and other MAPK pathway effectors or complex clonal evolution [59, 62, 63].

A recent randomized Phase 2 trial of azacitidine versus azacitidine plus enasidenib in newly diagnosed AML patients unfit for intensive chemotherapy showed a significantly higher CR rate with the combination compared to azacitidine alone (53% vs. 12%) and a median duration of response of 24.1 months in the combination arm (EHA Library. DiNardo C. 06/12/20; 294959; S139).

Whereas IDH inhibitors in combination with azacitidine yielded some very interesting response rates compared to azacitidine alone, one open issue will be to determine which induction regimen to choose between azacitidine-venetoclax and azacitidine-IDH inhibitors. Will it be more appropriate to start with the standard azacitidine-venetoclax and reserve the IDH inhibitors for relapse? Or will triplets eradicate the disease at first line? Which inhibitor will we use in patients with other concomitant targetable mutations, such as FLT3 or TP53? In fit patients, the addition of ivosidenib or enasidenib to intensive chemotherapy is under evaluation in an international Phase 3 trial involving several cooperative AML study groups (NCT03839771). Several others IDH1 or IDH2 inhibitors including drugs that target both mutations are under investigation [64].

3.3 | FLT3 inhibitors

Mutations in the *FLT3* gene are among the most common mutations in AML, occurring in up to 30% of patients [65]. There are two distinct activating *FLT3* mutations: internal tandem duplications (ITD) in the juxta-membrane domain and point mutations in the tyrosine kinase domain 2 (TKD). *FLT3* mutations are associated with an aggressive disease course, especially *FLT3*-ITD which generally predicts an early relapse and poor prognosis. Through clonal selection under chemotherapy, a higher mutant allelic burden is frequently observed at relapse, thus indicating that AML cells have become more addicted to FLT3 signalling. This is an important point because at least in preclinical settings, FLT3-mutant allelic burden and clinical status (i.e., diagnosis versus relapse samples) are predictive of the response to FLT3 inhibitors in AML [66].

Midostaurin - a staurosporine analog with a multikinase inhibitory activity against KIT, PDGFR, PKC, VEGF, and FLT3 (amongst others) - was the first FLT3 inhibitor to be approved for frontline therapy in AML patients with FLT3 mutations and fit for intensive chemotherapy [67]. In the randomized Phase 3 trial RATIFY, midostaurin and placebo were added to standard "7+3" induction chemotherapy and high-dose cytarabine consolidation followed by a 12-months single agent maintenance in younger patients (18-60 years) with FLT3-ITD or FLT3-TKD mutations. The CR rates were similar in both groups. However, midostaurin significantly improved the 4-year overall OS from 44.3% to 51.4%, compared with placebo [7]. The benefit of midostaurin was observed in FLT3-ITD patients, whatever the allelic ratio, and in FLT3-TKD patients. A subsequent exploratory analysis based on the FLT3-ITD patient of the RATIFY trial showed that the impact of midostaurin was significant in the three prognostic subgroups of the ELN2017 classification which includes NPM1, RUNX1, ASXL1, and TP53 as well as cytogenetic risk [68]. Furthermore, the benefit of midostaurin was independent from the allogeneic stem cell transplantation. A Phase 2 study suggested that older patients aged 60-70 years may also benefit from midostaurin [69]. Midostaurin was approved in Europe for induction, consolidation, and maintenance, whereas it was only approved for induction and consolidation in the US indicating that the need for midostaurin as maintenance is uncertain. However, this postulate could also apply to the consolidation phase since RATIFY was not designed to demonstrate that midostaurin treatment beyond induction chemotherapy was essential. Anyhow, since this approval and because FLT3 mutated patients present with a high tumor burden at diagnosis, the results of FLT3 mutational screen must been found rapidly.

Two randomized Phase 3 trials with second generation FLT3 inhibitors were recently conducted in R/R AML patients with *FLT3* mutations (gilteritinib) or *FLT3*-ITD only mutations (quizartinib) [70, 71]. In both studies, the FLT3 inhibitor as a single agent was superior to the standard of care with high or low intensity chemotherapy in terms of response and OS. However, while gilteritinib has been broadly

approved in North America, Europe and Japan, quizartinib was only registered in Japan.

Gilteritinib is an oral, small molecule inhibitor, highly selective of FLT3 acting against both FLT3-ITD and FLT3-TKD mutations and only acting marginally against cKIT [72, 73] which distinguishes it from quizartinib and likely explains the weak myelosuppression observed in patients. Gilteritinib also targets AXL, another tyrosine kinase implicated in the resistance to chemotherapy and FLT3 inhibitors [74, 75]. In the pivotal Phase 3 ADMIRAL study, AML patients with R/R FLT3mutated AML were randomized between 120-mg/day gilteritinib and a standard of care with high or low intensity regimen defined by physicians prior to randomization [70]. It is important to keep in mind that few patients of the ADMIRAL trial had previously been exposed to midostaurin, which is no longer the case. Gilteritinib induced higher CR/CRh and CR rates (34.0% vs. 15.3% and 21.1% vs. 10.5%, respectively) and significantly improved OS (median OS, 9.3 vs. 5.6 months). Adverse events were more frequent in the standard chemotherapy arm, with the exception of liver transaminase elevations. QTc prolongation, differentiation syndrome, and lipase elevation are very rare events in the context of gilteritinib treatment (<5%), whereas posterior reversible encephalopathy syndromes have been exceptionally reported [54, 76]. Off-target activating mutations in genes of the RAS/MAPK pathway have been identified as a key mechanism of the resistance to gilteritinib and confirmed in patients of the ADMIRAL trial who relapsed on gilteritinib treatment in whom in-target FLT3-F691L mutations were also detected [77, 78].

Phase 3 randomized trials comparing first line intensive chemotherapy plus midostaurin or second generation inhibitors are ongoing, and many combinations with hypomethylating agents or targeted agents are also being investigated [79]. Furthermore, other novel FLT3 inhibitors are under development, and it is foreseeable that clinicians may have a handful of FLT3 inhibitors in order to deal with clonal evolution, drug-drug interactions, adverse events, or co-morbid conditions just like BCR-ABL inhibitors for Philadelphia-chromosome-positive leukemia [80–83].

Patients with FLT3-ITD mutations are generally candidates to allogeneic stem cell transplantation, even though the post-transplant relapse rate remains a problem [84, 85]. Interestingly, sorafenib - a multikinase inhibitor with potent activity against FLT3 - has demonstrated clinical activity in FLT3-ITD patients having relapsed after transplantation [86]. A subsequent comprehensive preclinical study elegantly demonstrated that sorafenib, like other FLT3 inhibitors, increased the IL-15 production by FLT3-ITD leukemic cells leading to the potentiation of the allogeneic CD8+ T cell response as well as disease eradication in preclinical models [87]. As a clinical translation of this finding, the randomized Phase 3 SORMAIN trial demonstrated that sorafenib maintenance therapy reduces the risk of relapse and death after transplantation in AML patients with FLT3-ITD mutations [88]. Reducing the risk of relapse after allogeneic stem cell transplantation by maintenance therapy with non-cytotoxic drugs is an active field of research going beyond FLT3 inhibitors, and virtually all the small molecules inhibitors having shown efficacy in AML will be assessed in this context [89, 90].

4 | TP53: WANTED, DEAD, OR ALIVE

Although the TP53 tumor suppressor gene is the most frequently mutated gene in human cancer, its incidence in AML is relatively low (5%-20%) and increases with age or in therapy-related AML [48]. Patients with TP53 mutations have one of the worst prognosis in AML because their disease is both chemo and immune resistant, as shown by the poor response rate to standard treatments including intensive chemotherapy and hypomethylating agents and the high rate of relapse after allogeneic stem cell transplantation [91-94]. Treatment of this AML subgroup is an urgent unmet medical need, and therefore, huge efforts have been made to drug the undruggable. The first glimmer of hope came from eprenetapopt (APR-246), which is the first-in-class small molecule that selectively targets TP53 mutated cancer cells [95, 96]. Furthermore, eprenetapopt acts in synergy with azacitidine [97]. In two recent phase 2 trials designed for TP53 mutated myelodysplastic syndrome or AML, eprenetapopt combined with azacitidine induced an overall response rate of 62%-71% (44%-47% CR) with a median duration of response at 8-10.4 months. [98, 99]. Neurologic toxicity emerged as the main adverse effect with this drug. Furthermore, it has been recently shown that AML with TP53 mutations are associated with an infiltration of cytotoxic lymphocytes in the tumor microenvironment, indicating that immune intervention could be of value in this subgroup of patients [100, 101]. Altogether and even though these data are immature, there is reasonable hope to improve the outcome of patients with TP53 mutated-AML in the near future.

5 | CHEMOTHERAPY'S NOT DEAD (YET)

A sizable proportion of intermediate and good-risk AML patients are cured by intensive chemotherapy in one shot (i.e., without the need for allogeneic stem cell transplantation and without relapse). For these patients who generally receive one 7 + 3 induction cycle then three cycles of intermediate-to-high dose cytarabine, treatment is definitely completed in 6 months, and thereafter, the patients are treatmentfree for the rest of their life. In these patients, the added value of new drugs will be challenging to demonstrate both in terms of efficacy and duration of treatment because most new drugs have been developed so as to be administered on a long-term basis. However, well-known adverse events including profound myelosuppression, gastro-intestinal toxicity, severe mucositis, and infections as well as the strong impact of chemotherapy on quality of life remain of concern. CPX-351, a dualdrug liposomal combination of daunorubicin and cytarabine with a synergistic drug ratio, was approved for the treatment of adults with newly diagnosed therapy-related AML or AML with myelodysplasia-related changes [102]. Long-term results of the pivotal phase 3 trial shown at ASH 2020 confirmed the superiority of CPX-351 over standard "3+7" chemotherapy and the particular good outcome of patients who were allografted in first response after CPX-351 [103]. Interestingly, CPX-351 accumulates in the bone marrow where it has been shown to be taken up to a greater extent by AML cells than normal bone marrow cells and sparing normal tissues [104]. In a clinical context, this translates into improved efficacy (a higher response rate) but also into increased tolerability to induction chemotherapy. Nurses were probably among the first to notice this curious effect of CPX-351 compared to free daunorubicin and cytarabine: much less mucositis, gastrointestinal (GI) toxicity, and no hair loss. It is generally recognized by caregivers that CPX-351 is better tolerated than the standard 7 + 3, even though the Phase 3 trial did not clearly demonstrate this point. Furthermore, when ranking adverse events, it is noteworthy that hair-loss is the most common side effect reported as severe by the patients, while caregivers are more prone to declare infections, suggesting that quality of life with CPX-351 should be better than with 7+3 [105]. Therefore, it is tempting to foresee that CPX-351 indications could extend beyond the label to be broadly applied in de novo AML. In this regard, the results of the ongoing Phase 3 trial of the German AMLSG study group, currently assessing CPX-351 versus standard "7+3" in newly diagnosed AML and intermediate- or adverse genetics (\geq 18 y) (NCT03897127), will be of great importance for AML patients fit for chemotherapy.

6 | REVISITING THE CONCEPT OF REMISSION MAINTENANCE WITH ORAL THERAPY IN AML

Preventing relapse after induction and consolidation therapy in AML has long been a major challenge especially in older patients ineligible for allogeneic stem cell transplantation [106]. While there have been some interesting data with drugs such as the histamine dihydrochloride-interleukin 2 combination, which was associated with prolonged disease-free survival or more recently with norethandrolone which improved OS, there have been no approved drug in this indication [107, 108]. CC-486, an oral formulation of azacitidine has been recently developed for maintenance therapy in AML. Oral dosing of this drug that is not bioequivalent to standard injectable azacitidine, allows for extended drug exposure and prolonged pharmacodynamic effects. CC-486 was recently approved for maintenance of first CR after intensive chemotherapy in adult patients with AML not able to proceed to allogeneic stem cell transplantation. Indeed, in the pivotal randomized phase 3 trial (QUAZAR AML-001) that randomized 472 patients who were 55 years or older, in first CR after intensive chemotherapy and not candidates for transplantation, oral azacitidine improved OS compared to placebo (median OS, 24.7 vs. 14.8 months) [109]. Several communications at 2020 American Society of Hematology provided a better understanding of the QUAZAR AML-001 results. Indeed, the comparison of patients who received no consolidation (20% of the cohort), 1 consolidation (45%), or \geq 2 consolidations (35%), showed that CC-486 was associated with consistent survival benefits vs. placebo regardless of number of prior consolidation cycles ([110]). Furthermore, this study included long-term longitudinal assessment of MRD. CC-486 improved OS compared to placebo in patients who were either MRD+ (median 14.6 vs. 10.4 months) or MRD- (median 30.1 vs. 24.3 months) at study entry and induced in a higher rate of MRD+ to MRD- conversion (37% vs. 19%) [111]. GI events were the most common treatment-emergent adverse events reported in patients who received CC-486. These GI events were low-grade and decreased in frequency after initial treatment cycles [112]. Beyond the maintenance phase, there is much to be anticipated with this oral hypomethylating agent that could replace the injectable form in other phases of treatment alone or in combination with new agents.

7 | MONOCLONAL ANTIBODIES: WILL THE SECOND WAVE HAPPEN?

B-cell non-Hodgkin lymphoma has rituximab; multiple myeloma has daratumumab and acute lymphoblastic leukemia blinatumomab. What about AML? After small molecules inhibitors, the second wave of new game-changing drugs in AML may be represented by monoclonal antibodies. There is indeed a great deal of excitement in AML, thanks to various immune therapies, including naked antibodies targeting surface antigens expressed by leukemic stem cells, bispecific T-cell engager antibodies, or antibodies targeting immune checkpoint receptors [113, 114]. Naked monoclonal antibodies targeting CD47, CD70, or TIM3 that are expressed on leukemic stem cells display a very good safety profile as single agent and are excellent candidates for combination with hypomethylating agents. For example, anti-CD47 magrolimab combined with azacitidine induced a CR/CRi rate of 56% as first line treatment in unfit AML patients [115]. Anti-CD70 cusatuzumab has been shown to eradicate leukemic stem cells in xenotransplantation experiments and to also reduce the frequency of leukemic stem cells in AML patients in combination with azacitidine [116]. Bispecific T-cell engager such as flotetuzumab (CD3XCD123) or AMG-330 (CD3XCD33) are more challenging to manage because of the cytokine release syndrome, but may be powerful as single agents especially when the tumor microenvironment is immune-infiltrated. It has been shown that an immune interferon- γ signature associated with chemoresistance was predictive of response to flotetuzumab in R/R AML patients, thus suggesting that some immune therapy indications could be guided by a companion biomarker [101]. The use of bispecific antibodies in consolidation or maintenance could also be a valuable option for the eradication of the residual disease, while avoiding the cytokine release syndrome which is correlated with the tumor burden. Many other very promising antibodies are under development in AML [117]. It will be a huge challenge to integrate these new immunotherapies into the extraordinarily changing landscape of AML and adapt them to the disease' biological heterogeneity.

8 | NEXT-GENERATION SEQUENCING TO FOLLOW MINIMAL RESIDUAL DISEASE: WILL WE HAVE OUR CRYSTAL BALL?

Complete remission has been defined morphologically by a threshold of <5% bone marrow blasts together with the recovery of peripheral blood counts and no evidence of extramedullary disease. However, more than 50% of the patients who have reached this stage will ultimately relapse because of a high burden of residual disease that is now better measured, thanks to multiparameter flow cytometry, real-time

quantitative polymerase chain reaction, and more recently by nextgeneration sequencing (NGS) [118, 119]. Many studies have shown that the higher the measurable residual disease (MRD), the higher the risk of relapse; and recent guidelines have included MRD in the response criteria [16, 120–124]. Achieving CR with negative MRD is a major goal after first line treatment in order to guide subsequent treatments during consolidation, including allogeneic stem cell transplantation [124]. Furthermore, MRD may become a valuable early surrogate marker of survival endpoints for clinical trials. Indeed, with the introduction of novel drugs in the different phases of post-remission therapy – such as maintenance and in the treatment of relapse as well as the strong impact of allogeneic stem cell transplantation – it will become very challenging to build successful clinical trials based on OS or eventfree survival endpoints.

The advent of the NGS is critical to the evaluation of MRD and will be more and more used with the advance in NGS technologies which will improve NGS sensitivity and identify very low MRD levels [125]. In addition to the quantitative aspect, this technique will make it possible to detect which mutations persist in remission and those that will drive relapse, leaving room for targeted pre-emptive therapeutic interventions before morphological relapse, although pre-leukemic clonal hematopoiesis may interfere with the understanding of the results in some cases. Preliminary results have also shown that single-cell sequencing could improve the understanding of disease heterogeneity and the dynamics of clonal architecture during morphological remission [126]. Thus, it is likely that maintenance treatment will be soon guided by sequential NGS during the remission phase. In addition, two very important studies have described the pre-AML mutational landscape that is present in peripheral blood of health individuals several years prior to the diagnosis of overt disease, suggesting that early detection of AML, monitoring and interventional treatment may become a reality in future. [127, 128]

9 | FUTURE DIRECTIONS

Using new drugs as a single agent or in wise combinations during induction, consolidation and/or maintenance or at relapse, is the major challenge we will have to face in the 10 years to come. In younger patients, the first objective will be to increase the percentage of patients definitely cured by first line treatment. One important point will be to determine if the depth of the response reached by combining intensive chemotherapy and targeted agents eventually followed by novel strategies of immunotherapy will challenge allogeneic stem cell transplantation. This may become a hot topic in the field. In older patients, increasing response rates, duration of response and ultimately OS while maintaining a good quality of life will be major breakthroughs. Like in multiple myeloma, AML in older patients may become a chronic disease with successive lines of treatments, including maintenance, which could be eventually guided by a modern MRD follow-up. These are only two probable scenarios but others are obviously possible. With this multitude of therapeutic choices and methods to better assess the response and progression of the disease, it is clear that we

are coming out of the prehistoric era of AML treatment and that all physicians and biologists involved in AML will be writing an exciting new story.

CONFLICT OF INTEREST

Research grants from Abbvie, Amgen, Novartis, Celgene, Jazz Pharmaceuticals, Agios, Chugai, MaatPharma, Astellas, Roche, and Daiichi-Sankyo; an advisory role for Abbvie, Janssen, Jazz Pharmaceuticals, Novartis, Celgene, Otsuka, Astellas, Daiichi-Sankyo, Macrogenics, Roche, Takeda, and Pfizer.

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REFERENCES

- Dohner H, Weisdorf DJ, Bloomfield CD. Acute myeloid leukemia. N Engl J Med. 2015;373:1136–52.
- Barlogie B, Shaughnessy J, Tricot G, Jacobson J, Zangari M, Anaissie E, et al. Treatment of multiple myeloma. Blood. 2004;103:20– 32.
- Palumbo A, Anderson K. Multiple myeloma. N Engl J Med. 2011;364:1046-60.
- Rajkumar SV. Multiple myeloma: 2020 update on diagnosis, riskstratification and management. Am J Hematol. 2020;95:548–67.
- 5. Dombret H, Gardin C. An update of current treatments for adult acute myeloid leukemia. Blood. 2016;127:53–61.
- Grimwade D, Ivey A, Huntly BJ. Molecular landscape of acute myeloid leukemia in younger adults and its clinical relevance. Blood. 2016;127:29–41.
- Stone RM, Mandrekar SJ, Sanford BL, Laumann K, Geyer S, Bloomfield CD, et al. Midostaurin plus chemotherapy for acute myeloid leukemia with a FLT3 mutation. N Engl J Med. 2017;377:454–64.
- Ley TJ, Ding L, Walter MJ, Mclellan MD, Lamprecht T, Larson DE, et al. DNMT3A mutations in acute myeloid leukemia. N Engl J Med. 2010;363:2424–33.
- Mardis ER, Ding L, Dooling DJ, Larson DE, Mclellan MD, Chen K, et al. Recurring mutations found by sequencing an acute myeloid leukemia genome. N Engl J Med. 2009;361:1058–66.
- Ley TJ, Mardis ER, Ding L, Fulton B, Mclellan MD, Chen K, et al. DNA sequencing of a cytogenetically normal acute myeloid leukaemia genome. Nature. 2008;456:66–72.
- Papaemmanuil E, Gerstung M, Bullinger L, Gaidzik VI, Paschka P, Roberts ND, et al. Genomic classification and prognosis in acute myeloid leukemia. N Engl J Med. 2016;374:2209–21.
- Dohner H, Estey E, Grimwade D, Amadori S, Appelbaum FR, Buchner T, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. Blood. 2017;129:424-47.
- CANCER GENOME ATLAS RESEARCH Network, Ley TJ, Miller C, Ding L, Raphael BJ, Mungall AJ. Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia. N Engl J Med. 2013;368:2059–74.
- Welch JS, Ley TJ, Link DC, Miller CA, Larson DE, Koboldt DC, et al. The origin and evolution of mutations in acute myeloid leukemia. Cell. 2012;150:264–78.
- Ding L, Ley TJ, Larson DE, Miller CA, Koboldt DC, Welch JS, et al. Clonal evolution in relapsed acute myeloid leukaemia revealed by whole-genome sequencing. Nature. 2012;481:506–10.
- Jongen-Lavrencic M, Grob T, Hanekamp D, Kavelaars FG, Al Hinai A, Zeilemaker A, et al. Molecular minimal residual disease in acute myeloid leukemia. N Engl J Med. 2018;378:1189–99.

- Yamashita M, Dellorusso PV, Olson OC, Passegue E. Dysregulated haematopoietic stem cell behaviour in myeloid leukaemogenesis. Nat Rev Cancer. 2020;20:365–82.
- Miyamoto K, Minami Y. Cutting edge molecular therapy for acute myeloid leukemia. Int J Mol Sci. 2020;21:5114.
- Wei AH, Tiong IS. Midostaurin, enasidenib, CPX-351, gemtuzumab ozogamicin, and venetoclax bring new hope to AML. Blood. 2017;130:2469–74.
- Short NJ, Konopleva M, Kadia TM, Borthakur G, Ravandi F, Dinardo CD, et al. Advances in the treatment of acute myeloid leukemia: new drugs and new challenges. Cancer Discov. 2020;10:506–25.
- 21. Saygin C, Carraway HE. Emerging therapies for acute myeloid leukemia. J Hematol Oncol. 2017;10:93.
- "type="Periodical">Campos L, Rouault JP, Sabido O, Oriol P, Roubi N, Vasselon C, et al. High expression of bcl-2 protein in acute myeloid leukemia cells is associated with poor response to chemotherapy. Blood. 1993;81:3091–6.
- Lagadinou ED, Sach A, Callahan K, Rossi RM, Neering SJ, Minhajuddin M, et al. BCL-2 inhibition targets oxidative phosphorylation and selectively eradicates quiescent human leukemia stem cells. Cell Stem Cell. 2013;12:329–41.
- Pan R, Hogdal LJ, Benito JM, Bucci D, Han L, Borthakur G, et al. Selective BCL-2 inhibition by ABT-199 causes on-target cell death in acute myeloid leukemia. Cancer Discov. 2014;4:362–75.
- Chua CC, Roberts AW, Reynolds J, Fong CY, ting SB, Salmon JM, et al. Chemotherapy and venetoclax in elderly acute myeloid leukemia trial (CAVEAT): a phase Ib dose-escalation study of venetoclax combined with modified intensive chemotherapy. J Clin Oncol. 2020;38:3506– 17.
- Konopleva M, Pollyea DA, Potluri J, Chyla B, Hogdal L, Busman T, et al. Efficacy and biological correlates of response in a phase II study of venetoclax monotherapy in patients with acute myelogenous leukemia. Cancer Discov. 2016;6:1106–17.
- Dinardo CD, Pratz K, Pullarkat V, Jonas BA, Arellano M, Becker PS, et al. Venetoclax combined with decitabine or azacitidine in treatment-naive, elderly patients with acute myeloid leukemia. Blood. 2019;133:7–17.
- Dinardo CD, Pratz KW, Letai A, Jonas BA, Wei AH, Thirman M, et al. Safety and preliminary efficacy of venetoclax with decitabine or azacitidine in elderly patients with previously untreated acute myeloid leukaemia: a non-randomised, open-label, phase 1b study. Lancet Oncol. 2018a;19:216–28.
- Dinardo CD, Jonas BA, Pullarkat V, Thirman MJ, Garcia JS, Wei AH, et al. Azacitidine and venetoclax in previously untreated acute myeloid leukemia. N Engl J Med. 2020;383:617–29.
- Pollyea DA, Dinardo CD, Arellano ML, Pigneux A, Fiedler W, Konopleva M, et al. Results of venetoclax and azacitidine combination in chemotherapy ineligible untreated patients with acute myeloid leukemia with IDH 1/2 mutations. Blood. 2020;136:5–7.
- Wei AH, Montesinos P, Ivanov V, Dinardo CD, Novak J, Laribi K, et al. Venetoclax plus LDAC for newly diagnosed AML ineligible for intensive chemotherapy: a phase 3 randomized placebo-controlled trial. Blood. 2020c;135:2137–45.
- 32. Klepin HD. Definition of unfit for standard acute myeloid leukemia therapy. Curr Hematol Malig Rep. 2016;11:537–44.
- Podoltsev NA, Stahl M, Zeidan AM, Gore SD. Selecting initial treatment of acute myeloid leukaemia in older adults. Blood Rev. 2017;31:43-62.
- Rao AV. Fitness in the elderly: how to make decisions regarding acute myeloid leukemia induction. Hematol Am Soc Hematol Educ Program. 2016;2016:339–47.
- Hamaker ME, Prins MC, Stauder R. The relevance of a geriatric assessment for elderly patients with a haematological malignancy–a systematic review. Leuk Res. 2014;38:275–83.

- 36. Othus M, Kantarjian H, Petersdorf S, Ravandi F, Godwin J, Cortes J, et al. Declining rates of treatment-related mortality in patients with newly diagnosed AML given 'intense' induction regimens: a report from SWOG and MD Anderson. Leukemia. 2014;28:289–92.
- Perl AE, Daver NG, Pratz KW, Maly J, Hong W-J, bahceci E, et al. Venetoclax in combination with gilteritinib in patients with relapsed/refractory acute myeloid leukemia: a phase 1b study. Blood. 2019;134:3910–0.
- Daver NG, Garcia JS, Jonas BA, Kelly KR, Assouline S, Brandwein JM, et al. Updated results from the venetoclax (Ven) in combination with idasanutlin (Idasa) arm of a phase 1b trial in elderly patients (Pts) with relapsed or refractory (R/R) AML ineligible for cytotoxic chemotherapy. Blood. 2019;134:229–9.
- 39. kadia TM, Borthakur G, Pemmaraju N, Daver N, Dinardo CD, Sasaki K, et al. Phase II study of venetoclax added to cladribine + low dose AraC (LDAC) alternating with 5-azacytidine demonstrates high rates of minimal residual disease (MRD) negative complete remissions (CR) and excellent tolerability in older patients with newly diagnosed acute myeloid leukemia (AML). Blood. 2020;136: 17–9.
- 40. Lachowiez C, Konopleva M, Kadia TM, Daver N, Loghavi S, Wang SA, et al. Interim analysis of the phase 1b/2 study of the BCL-2 inhibitor venetoclax in combination with standard intensive AML induction/consolidation therapy with FLAG-IDA in patients with newly diagnosed or relapsed/refractory AML. Blood. 2020;136:18–20.
- Kadia TM, Borthakur G, Takahashi K, Dinardo CD, Daver N, Pemmaraju N, et al. Phase II study of CPX-351 plus venetoclax in patients with acute myeloid leukemia (AML). Blood. 2020;136:20–22.
- 42. Daver N, Altman JK, Maly J, Levis M, Ritchie E, Litzow M, et al. Efficacy and safety of venetoclax in combination with gilteritinib for relapsed/refractory FLT3-mutated acute myeloid leukemia in the expansion cohort of a phase 1b study. Blood. 2020;136:20–22.
- 43. Yilmaz M, Kantarjian HM, Muftuoglu M, Kadia TM, Konopleva M, Borthakur G, et al. Quizartinib with decitabine +/- venetoclax is highly active in patients (Pts) with FLT3-ITD mutated (mut) acute myeloid leukemia (AML): clinical report and signaling Cytof profiling from a phase IB/II trial. Blood. 2020;136:19–20.
- 44. Kim K, Maiti A, Kadia TM, Ravandi F, Daver N, Pemmaraju N, et al. Outcomes of TP53-mutant acute myeloid leukemia with venetoclax and decitabine. Blood. 2020;136:33–6.
- 45. Sekeres MA, Elson P, Kalaycio ME, Advani AS, Copelan EA, Faderl S, et al. Time from diagnosis to treatment initiation predicts survival in younger, but not older, acute myeloid leukemia patients. Blood. 2009;113:28–36.
- 46. Bertoli S, Berard E, Huguet F, Huynh A, Tavitian S, Vergez F, et al. Time from diagnosis to intensive chemotherapy initiation does not adversely impact the outcome of patients with acute myeloid leukemia. Blood. 2013;121:2618–26.
- 47. Röllig C, Kramer M, Schliemann C, Mikesch J-H, Steffen B, Krämer A, et al. Time from diagnosis to treatment does not affect outcome in intensively treated patients with newly diagnosed acute myeloid leukemia. Blood. 2019;134:13.
- Bullinger L, Dohner K, Dohner H. Genomics of Acute Myeloid Leukemia Diagnosis and Pathways. J Clin Oncol. 2017;35:934-46.
- Duchmann M, Micol J-B, Duployez N, Raffoux E, Thomas X, Marolleau J-P, et al. Prognostic significance of concurrent gene mutations in intensively treated patients with IDH1/2 mutated AML. Blood. 2019;134:1416–6.
- Patel JP, Gonen M, Figueroa ME, Fernandez H, Sun Z, Racevskis J, et al. Prognostic relevance of integrated genetic profiling in acute myeloid leukemia. N Engl J Med. 2012;366:1079–89.
- Paschka P, Schlenk RF, Gaidzik VI, Habdank M, Kronke J, Bullinger L, et al. IDH1 and IDH2 mutations are frequent genetic alterations in acute myeloid leukemia and confer adverse prognosis in cytoge-

netically normal acute myeloid leukemia with NPM1 mutation without FLT3 internal tandem duplication. J Clin Oncol. 2010;28:3636– 43.

- 52. Abbas S, Lugthart S, Kavelaars FG, Schelen A, Koenders JE, Zeilemaker A, et al. Acquired mutations in the genes encoding IDH1 and IDH2 both are recurrent aberrations in acute myeloid leukemia: prevalence and prognostic value. Blood. 2010;116:2122–6.
- Dinardo CD, Stein EM, De Botton S, Roboz GJ, Altman JK, Mims AS, et al. Durable remissions with ivosidenib in IDH1-mutated relapsed or refractory AML. N Engl J Med. 2018;378:2386–98.
- 54. Dinardo CD, Wei AH. How I treat acute myeloid leukemia in the era of new drugs. Blood. 2020;135:85–96.
- 55. Choe S, Wang H, Dinardo CD, Stein EM, De Botton S, Fathi AT, et al. Molecular mechanisms mediating relapse following ivosidenib monotherapy in patients with IDH1-mutant relapsed or refractory acute myeloid leukemia. Blood. 2019;134:545–5.
- Dinardo CD, Stein AS, Stein EM, Fathi AT, Frankfurt O, Schuh AC, et al. Mutant IDH1 inhibitor ivosidenib (IVO; AG-120) in combination with azacitidine (AZA) for newly diagnosed acute myeloid leukemia (ND AML). J Clin Oncol. 2019b;37:7011–11.
- Ward PS, Patel J, Wise DR, Abdel-Wahab O, Bennett BD, Coller HA, et al. The common feature of leukemia-associated IDH1 and IDH2 mutations is a neomorphic enzyme activity converting alphaketoglutarate to 2-hydroxyglutarate. Cancer Cell. 2010;17:225– 34.
- Figueroa ME, Abdel-Wahab O, Lu C, Ward PS, Patel J, Shih A, et al. Leukemic IDH1 and IDH2 mutations result in a hypermethylation phenotype, disrupt TET2 function, and impair hematopoietic differentiation. Cancer Cell. 2010;18:553–67.
- Amatangelo MD, Quek L, Shih A, Stein EM, Roshal M, David MD, et al. Enasidenib induces acute myeloid leukemia cell differentiation to promote clinical response. Blood. 2017;130:732–41.
- Stein EM Enasidenib, a targeted inhibitor of mutant IDH2 proteins for treatment of relapsed or refractory acute myeloid leukemia. Future Oncol. 2018;14:23–40.
- 61. Fathi AT, Dinardo CD, Kline I, Kenvin L, Gupta I, attar EC, et al. Differentiation syndrome associated with enasidenib, a selective inhibitor of mutant isocitrate dehydrogenase 2: analysis of a phase 1/2 study. JAMA Oncol. 2018;4:1106–10.
- Intlekofer AM, Shih AH, Wang B, Nazir A, Rustenburg AS, Albanese SK, et al. Acquired resistance to IDH inhibition through trans or cis dimer-interface mutations. Nature. 2018;559:125–9.
- Quek L, David MD, Kennedy A, Metzner M, Amatangelo M, Shih A, et al. Clonal heterogeneity of acute myeloid leukemia treated with the IDH2 inhibitor enasidenib. Nat Med. 2018;24:1167–77.
- Golub D, Iyengar N, Dogra S, Wong T, Bready D, Tang K, et al. Mutant isocitrate dehydrogenase inhibitors as targeted cancer therapeutics. Front Oncol. 2019;9:417.
- 65. Papaemmanuil E, Dohner H, Campbell PJ. Genomic classification in acute myeloid leukemia. N Engl J Med. 2016;375:900–1.
- Pratz KW, Sato T, Murphy KM, Stine A, Rajkhowa T, Levis M FLT3mutant allelic burden and clinical status are predictive of response to FLT3 inhibitors in AML. Blood. 2010;115:1425–32.
- Weisberg E, Boulton C, Kelly LM, Manley P, Fabbro D, Meyer T, et al. Inhibition of mutant FLT3 receptors in leukemia cells by the small molecule tyrosine kinase inhibitor PKC412. Cancer Cell. 2002;1:433–43.
- Döhner K, Thiede C, Jahn N, Panina E, Gambietz A, Larson RA, et al. Impact of NPM1/FLT3-ITD genotypes defined by the 2017 European LeukemiaNet in patients with acute myeloid leukemia. Blood. 2020;135:371–80.
- Schlenk RF, Weber D, Fiedler W, Salih HR, Wulf G, Salwender H, et al. Midostaurin added to chemotherapy and continued singleagent maintenance therapy in acute myeloid leukemia with FLT3-ITD. Blood. 2019;133:840–51.

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- Perl AE, MARTINELLI G, Cortes JE, Neubauer A, Berman E, Paolini S, et al. Gilteritinib or chemotherapy for relapsed or refractory FLT3mutated AML. N Engl J Med. 2019;381:1728–40.
- Cortes JE, Khaled S, Martinelli G, Perl AE, Ganguly S, Russell N, et al. Quizartinib versus salvage chemotherapy in relapsed or refractory FLT3-ITD acute myeloid leukaemia (QuANTUM-R): a multicentre, randomised, controlled, open-label, phase 3 trial. Lancet Oncol. 2019;20:984-97.
- Lee LY, Hernandez D, Rajkhowa T, Smith SC, Raman JR, Nguyen B, et al. Preclinical studies of gilteritinib, a next-generation FLT3 inhibitor. Blood. 2017;129:257–60.
- Mori M, Kaneko N, Ueno Y, Yamada M, Tanaka R, Saito R, et al. Gilteritinib, a FLT3/AXL inhibitor, shows antileukemic activity in mouse models of FLT3 mutated acute myeloid leukemia. Invest New Drugs. 2017;35:556-65.
- 74. Ben-Batalla I, Schultze A, Wroblewski M, Erdmann R, Heuser M, Waizenegger JS, et al. Axl, a prognostic and therapeutic target in acute myeloid leukemia mediates paracrine crosstalk of leukemia cells with bone marrow stroma. Blood. 2013;122:2443–52.
- Dumas PY, Naudin C, Martin-Lanneree S, Izac B, Casetti L, Mansier O, et al. Hematopoietic niche drives FLT3-ITD acute myeloid leukemia resistance to quizartinib via STAT5-and hypoxia-dependent upregulation of AXL. Haematologica. 2019;104:2017–27.
- Mcmahon CM, Canaani J, Rea B, Sargent RL, Qualtieri JN, Watt CD, et al. Gilteritinib induces differentiation in relapsed and refractory FLT3-mutated acute myeloid leukemia. Blood Adv. 2019a;3:1581– 5.
- 77. Smith CC, Levis MJ, Perl AE, Martinelli G, Neubauer A, Berman E, et al. Emerging mutations at relapse in patients with FLT3-mutated relapsed/refractory acute myeloid leukemia who received gilteritinib therapy in the phase 3 admiral trial. Blood. 2019;134:14–14.
- Mcmahon CM, Ferng T, Canaani J, Wang ES, Morrissette JJD, Eastburn DJ, et al. Clonal selection with RAS pathway activation mediates secondary clinical resistance to selective FLT3 inhibition in acute myeloid leukemia. Cancer Discov. 2019b;9:1050–63.
- Ambinder AJ, Levis M. Potential targeting of FLT3 AML. Haematologica. 2021;106:671–81.
- Megías-Vericat JE, Solana-Altabella A, Ballesta-López O, Martínez-Cuadrón D, Montesinos P. Drug-drug interactions of newly approved small molecule inhibitors for acute myeloid leukemia. Ann Hematol. 2020;99:1989–2007.
- Antar AI, Otrock ZK, Jabbour E, Mohty M, Bazarbachi A. FLT3 inhibitors in acute myeloid leukemia: ten frequently asked questions. Leukemia. 2020;34:682–96.
- Wang ES. Beyond midostaurin: which are the most promising FLT3 inhibitors in AML? Best Pract Res Clin Haematol. 2019;32:101103.
- Daver N, Schlenk RF, Russell NH, Levis MJ. Targeting FLT3 mutations in AML: review of current knowledge and evidence. Leukemia. 2019a;33:299–312.
- Oran B, Cortes J, Beitinjaneh A, Chen HC, De Lima M, Patel K, et al. Allogeneic transplantation in first remission improves outcomes irrespective of FLT3-ITD allelic ratio in FLT3-ITD-positive acute myelogenous leukemia. Biol Blood Marrow Transplant. 2016;22:1218–26.
- Gaballa S, Saliba R, Oran B, Brammer JE, Chen J, Rondon G, et al. Relapse risk and survival in patients with FLT3 mutated acute myeloid leukemia undergoing stem cell transplantation. Am J Hematol. 2017;92:331–7.
- Metzelder SK, schroeder T, Finck A, Scholl S, Fey M, Gotze K, et al. High activity of sorafenib in FLT3-ITD-positive acute myeloid leukemia synergizes with allo-immune effects to induce sustained responses. Leukemia. 2012;26:2353–9.
- Mathew NR, Baumgartner F, Braun L, O'sullivan D, Thomas S, Waterhouse M, et al. Sorafenib promotes graft-versus-leukemia activity in mice and humans through IL-15 production in FLT3-ITD-mutant leukemia cells. Nat Med. 2018;24:282–91.

- Burchert A, Bug G, Fritz LV, Finke J, Stelljes M, Röllig C, et al. Sorafenib maintenance after allogeneic hematopoietic stem cell transplantation for acute myeloid leukemia with FLT3-internal tandem duplication mutation (SORMAIN). J Clin Oncol. 2020;38:2993– 3002.
- Appelbaum FR. Maintenance therapy after allogeneic hematopoietic cell transplantation for acute myeloid leukemia. Best Pract Res Clin Haematol. 2019;32:101109.
- Sterling C, Webster J. Harnessing the immune system after allogeneic stem cell transplant in acute myeloid leukemia. Am J Hematol. 2020;95:529–47.
- Lindsley RC, Saber W, Mar BG, Redd R, Wang T, Haagenson MD, et al. Prognostic mutations in myelodysplastic syndrome after stem-cell transplantation. N Engl J Med. 2017;376:536–47.
- Wong TN, Ramsingh G, Young AL, Miller CA, Touma W, Welch JS, et al. Role of TP53 mutations in the origin and evolution of therapyrelated acute myeloid leukaemia. Nature. 2015;518:552–5.
- Bally C, Adès L, Renneville A, Sebert M, Eclache V, Preudhomme C, et al. Prognostic value of TP53 gene mutations in myelodysplastic syndromes and acute myeloid leukemia treated with azacitidine. Leuk Res. 2014;38:751–5.
- Middeke JM, Herold S, Rücker-Braun E, Berdel WE, Stelljes M, Kaufmann M, et al. TP53 mutation in patients with high-risk acute myeloid leukaemia treated with allogeneic haematopoietic stem cell transplantation. Br J Haematol. 2016;172:914–22.
- 95. Perdrix A, Najem A, Saussez S, Awada A, Journe F, Ghanem G, et al. PRIMA-1 and PRIMA-1(Met) (APR-246): from mutant/wild type p53 reactivation to unexpected mechanisms underlying their potent antitumor effect in combinatorial therapies. Cancers (Basel). 2017;9:172.
- Sallman DA. To target the untargetable: elucidation of synergy of APR-246 and azacitidine in TP53mutant myelodysplastic syndromes and acute myeloid leukemia. Haematologica. 2020;105:1470-2.
- Maslah N, Salomao N, Drevon L, Verger E, Partouche N, Ly P, et al. Synergistic effects of PRIMA-1^{Met}(APR-246) and 5-azacitidine in TP53-mutated myelodysplastic syndromes and acute myeloid leukemia. Haematologica. 2020;105:1539–51.
- Cluzeau T, Sebert M, Rahmé R, Cuzzubbo S, Lehmann-Che J, Madelaine I, et al. Eprenetapopt plus azacitidine in TP53-mutated myelodysplastic syndromes and acute myeloid leukemia: a phase II study by the groupe francophone des myélodysplasies (GFM). J Clin Oncol. 2021;39:1575–83.
- Sallman DA, Dezern AE, Garcia-Manero G, Steensma DP, Roboz GJ, Sekeres MA, et al. Eprenetapopt (APR-246) and azacitidine in TP53mutant myelodysplastic syndromes. J Clin Oncol. 2021;39:1584–94.
- Dufva O, Polonen P, Bruck O, Keranen MAI, Klievink J, Mehtonen J, et al. Immunogenomic landscape of hematological malignancies. Cancer Cell. 2020;38:380–99.e13.
- Vadakekolathu J, Minden MD, Hood T, Church SE, Reeder S, Altmann H, et al. Immune landscapes predict chemotherapy resistance and immunotherapy response in acute myeloid leukemia. Sci Transl Med. 2020;12:eaaz0463.
- 102. Lancet JE, Uy GL, Cortes JE, Newell LF, Lin TL, Ritchie EK, et al. CPX-351 (cytarabine and daunorubicin) liposome for injection versus conventional cytarabine plus daunorubicin in older patients with newly diagnosed secondary acute myeloid leukemia. J Clin Oncol. 2018;36:2684-92.
- 103. Uy GL, Newell LF, Lin T, Goldberg SL, Wieduwilt MJ, Ryan RJ, et al. Long-term outcomes of allogeneic hematopoietic cell transplantation in patients enrolled in CPX-351-301, a randomized phase 3 study of CPX-351 versus 7+3 in older adults with newly diagnosed, high-risk and/or secondary AML. Blood. 2020;136:44–45.
- Tolcher AW, Mayer LD. Improving combination cancer therapy: the CombiPlex(R) development platform. Future Oncol. 2018;14:1317– 32.

- 105. Crossnohere NL, Richardson DR, Reinhart C, O'donoghue B, Love SM, Smith BD, et al. Side effects from acute myeloid leukemia treatment: results from a national survey. Curr Med Res Opin. 2019;35:1965– 70.
- Rashidi A, Walter RB, Tallman MS, Appelbaum FR, Dipersio JF. Maintenance therapy in acute myeloid leukemia: an evidence-based review of randomized trials. Blood. 2016;128:763–73.
- 107. Yang LP, Perry CM. Histamine dihydrochloride: in the management of acute myeloid leukaemia. Drugs. 2011;71:109–22.
- Pigneux A, Béné MC, Guardiola P, Recher C, Hamel JF, Sauvezie M, et al. Addition of androgens improves survival in elderly patients with acute myeloid leukemia: a GOELAMS study. J Clin Oncol. 2017;35:387–93.
- 109. Wei AH, Döhner H, Pocock C, Montesinos P, Afanasyev B, Dombret H, et al. Oral azacitidine maintenance therapy for acute myeloid leukemia in first remission. N Engl J Med. 2020;383:2526–37.
- 110. Wei A, Roboz GJ, Dombret H, Döhner H, Schuh AC, Montesinos P, et al. CC-486 improves overall survival (OS) and relapse-free survival (RFS) for patients with acute myeloid leukemia (AML) in first remission after intensive chemotherapy (IC), regardless of amount of consolidation received: results from the phase III QUAZAR AML-001 maintenance trial. Blood. 2020;136:38–40.
- 111. Roboz GJ, Ravandi F, Wei AH, Dombret H, Döhner H, Thol F, et al. CC-486 prolongs survival for patients with acute myeloid leukemia (AML) in remission after intensive chemotherapy (IC) independent of the presence of measurable residual disease (MRD) at study entry: results from the QUAZAR AML-001 maintenance trial. Blood. 2020;136:32–33.
- 112. Ravandi F, Pocock C, Selleslag D, Montesinos P, Sayar H, Musso M, et al. Gastrointestinal events and management strategies for patients with acute myeloid leukemia (AML) in first remission receiving CC-486 in the randomized, placebo-controlled, phase III QUAZAR AML-001 maintenance trial. Blood. 2020;136:22–23.
- 113. Assi R, Kantarjian H, Ravandi F, Daver N. Immune therapies in acute myeloid leukemia: a focus on monoclonal antibodies and immune checkpoint inhibitors. Curr Opin Hematol. 2018;25:136–45.
- 114. Guy DG, Uy GL. Bispecific antibodies for the treatment of acute myeloid leukemia. Curr Hematol Malig Rep. 2018;13:417–25.
- 115. Sallman DA, Malki MA, Asch AS, Lee DJ, Kambhampati S, Donnellan WB, et al. Tolerability and efficacy of the first-in-class anti-CD47 antibody magrolimab combined with azacitidine in MDS and AML patients: Phase Ib results. J Clin Oncol. 2020;38:7507–7.
- 116. Riether C, Pabst T, Höpner S, Bacher U, Hinterbrandner M, Banz Y, et al. Targeting CD70 with cusatuzumab eliminates acute myeloid leukemia stem cells in patients treated with hypomethylating agents. Nat Med. 2020;26:1459–67.
- 117. Vago L, Gojo I. Immune escape and immunotherapy of acute myeloid leukemia. J Clin Invest. 2020;130:1552–64.

- 118. Short NJ, Ravandi F. How close are we to incorporating measurable residual disease into clinical practice for acute myeloid leukemia? Haematologica. 2019;104:1532–41.
- 119. "type="Periodical">Schuurhuis GJ, Heuser M, Freeman S, Bene MC, Buccisano F, Cloos J, et al. Minimal/measurable residual disease in AML: a consensus document from the European LeukemiaNet MRD Working Party. Blood. 2018;131:1275–91.
- Ivey A, Hills RK, Simpson MA, Jovanovic JV, Gilkes A, Grech A, et al. Assessment of minimal residual disease in standard-risk AML. N Engl J Med. 2016;374:422–33.
- 121. Balsat M, Renneville A, Thomas X, De Botton S, Caillot D, Marceau A, et al. Postinduction minimal residual disease predicts outcome and benefit from allogeneic stem cell transplantation in acute myeloid leukemia with NPM1 mutation: a study by the acute leukemia French Association Group. J Clin Oncol. 2017;35:185–93.
- 122. Freeman SD, Virgo P, Couzens S, Grimwade D, Russell N, Hills RK, et al. Prognostic relevance of treatment response measured by flow cytometric residual disease detection in older patients with acute myeloid leukemia. J Clin Oncol. 2013;31:4123–31.
- 123. Dohner H, Estey E, Grimwade D, Amadori S, Appelbaum FR, Buchner T, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. Blood. 2017;129:424-47.
- 124. Venditti A, Piciocchi A, Candoni A, Melillo L, Calafiore V, Cairoli R, et al. GIMEMA AML1310 trial of risk-adapted, MRD-directed therapy for young adults with newly diagnosed acute myeloid leukemia. Blood. 2019;134:935–45.
- 125. Salk JJ, Schmitt MW, Loeb LA. Enhancing the accuracy of nextgeneration sequencing for detecting rare and subclonal mutations. Nat Rev Genet. 2018;19:269–85.
- 126. Ediriwickrema A, Aleshin A, Reiter JG, Corces MR, Kohnke T, Stafford M, et al. Single-cell mutational profiling enhances the clinical evaluation of AML MRD. Blood Adv. 2020;4:943–52.
- 127. Abelson S, Collord G, Ng SWK, Weissbrod O, Mendelson Cohen N, Niemeyer E, et al. Prediction of acute myeloid leukaemia risk in healthy individuals. Nature. 2018;559:400–4.
- 128. Desai P, Mencia-Trinchant N, Savenkov O, Simon MS, Cheang G, Lee S, et al. Somatic mutations precede acute myeloid leukemia years before diagnosis. Nat Med. 2018;24:1015–23.

How to cite this article: Récher C. The beginning of a new therapeutic era in acute myeloid leukemia. eJHaem. 2021;2:823–833. https://doi.org/10.1002/jha2.252