

RAPID COMMUNICATION

Treatment of Autoimmune Disease by Intense Immunosuppressive Conditioning and Autologous Hematopoietic Stem Cell Transplantation

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Multiple sclerosis, systemic lupus erythematosus, and rheumatoid arthritis are immune-mediated diseases that are responsive to suppression or modulation of the immune system. For patients with severe disease, immunosuppression may be intensified to the point of myelosuppression or hematopoietic ablation. Hematopoiesis and immunity may then be rapidly reconstituted by reinfusion of CD34⁺ progenitor cells. In 10 patients with these autoimmune diseases, autologous hematopoietic stem cells were collected from bone marrow or mobilized from peripheral blood with either granulocyte colony-stimulating factor (G-CSF) or cyclophosphamide and G-CSF. Stem cells were enriched ex vivo using CD34⁺ selection and reinfused after either myelosuppressive conditioning with cyclophosphamide (200 mg/kg), methylprednisolone (4 g) and antithymocyte globulin (ATG; 90 mg/kg) or myeloablative conditioning with total body irradiation (1,200 cGy), methylprednisolone (4 g), and cyclophosphamide

(120 mg/kg). Six patients with multiple sclerosis, 2 with systemic lupus erythematosus, and 2 with rheumatoid arthritis have undergone hematopoietic stem cell transplantation. Mean time to engraftment of an absolute neutrophil count greater than 500/ μ L (0.5×10^9 /L) and a nontransfused platelet count greater than 20,000/ μ L (20×10^9 /L) occurred on day 10 and 14, respectively. Regimen-related nonhematopoietic toxicity was minimal. All patients improved and/or had stabilization of disease with a follow-up of 5 to 17 months (median, 11 months). We conclude that intense immunosuppressive conditioning and autologous T-cell-depleted hematopoietic transplantation was safely used to treat these 10 patients with severe autoimmune disease. Although durability of response is as yet unknown, all patients have demonstrated stabilization or improvement.

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INTENSIVE immunosuppression and hematopoietic stem cell transplantation has been proposed or initiated as a therapy for patients with severe autoimmune diseases (SADS) who have poor prognostic features.¹⁻¹¹ The rationale is to maximally suppress or ablate the immune system and then rescue the patient from prolonged cytopenias or hematopoietic failure by infusing either autologous or allogeneic hematopoietic progenitor cells (CD34⁺ cells). This approach is supported by hematopoietic stem cell transplantation in animal autoimmune disorders and in patients undergoing transplantation for a hematologic disease who also had a coincidental autoimmune disease. Animal autoimmune diseases may occur spontaneously or be induced either by immunization with self-peptides or adoptive transfer of disease-initiating lymphocytes. A spontaneous onset lupus-like illness occurs in Murthy Roth Lab lymphoproliferative (MRL/lpr) mice and New Zealand Black/New Zealand White (B/W) mice.^{12,13} In MRL/lpr and B/W mice, an allogeneic transplant from a nonsusceptible strain is required to cure disease.^{12,13} Alternatively, disease can be transferred from susceptible to nonsusceptible mice after bone marrow transplantation. In MRL/lpr mice, a single gene defect in Fas expression, a protein that signals for apoptosis, results in a lymphoproliferative response with lupus-like features.^{14,15} Therefore, it appears in these animal models that spontaneous-onset autoimmune disease may arise from a hematopoietic stem cell defect predisposing to immune dysregulation.

Induced autoimmune diseases require manipulation of a normal immune system (ie, immunization) to break self-tolerance. Why a potentially self-reactive repertoire exists is unknown, but environmental influences are necessary to break tolerance. Experimental autoimmune encephalomyelitis (EAE) is an induced animal autoimmune disease that mimics multiple sclerosis (MS). EAE may be cured by allogeneic, syngeneic, or autologous bone marrow transplantation,¹⁶⁻²⁰ although the relapse rate is higher after an autologous or syngeneic transplant.

In animal autoimmune disorders, genetically preordained diseases require an allogeneic transplant from a nonsusceptible strain for cure, whereas environmentally induced disease may be cured with either allogeneic or autologous transplantation.

Patients with aplastic anemia, leukemia, or lymphoma and a coincidental autoimmune disease such as rheumatoid arthritis (RA), scleroderma, Crohn's disease, or MS have been treated by hematopoietic stem cell transplantation for their hematologic disorder.²¹⁻²⁹ In most reported cases this has also resulted in subsequent remission of their autoimmune disease. Although the number of anecdotal case reports is small, duration of remission appears better with an allogeneic graft compared with an unmanipulated autologous graft. Virtually no information is available on lymphocyte-depleted autologous transplantation in patients with autoimmune diseases. Because of the higher expected morbidity of allogeneic transplantation, consensus

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conferences have recommended initiating this approach with autologous stem cells.³

MATERIALS AND METHODS

Patient Selection

All three protocols (MS, systemic lupus erythematosus [SLE], and RA) are approved by the US FDA under IDE numbers BB-IDE 6440, BB-IDE 6559, and BB-IDE 6778, respectively. For all protocols, patients must be less than 60 years of age at the time of pretransplant evaluation and must meet the following criteria.

MS. Patients must have clinically definite MS using Poser criteria supported by characteristic magnetic resonance imaging (MRI) changes and absence of serologic or clinical signs of other autoimmune diseases.³⁰ In addition, the patients must fulfill both of the following criteria: (1) failure to stabilize active clinical progression with intravenous methylprednisolone administered for a minimum of 3 days at 1 g per day; and (2) a Kurtzke extended disability status scale (EDSS)³¹ of 5.0 to 8.0, with an increase in the EDSS by 1.5 points within the preceding 12 months in patients with an EDSS of 5.5 or less at the start of the evaluation period or an increase of 1 point in patients with an EDSS of 6 or greater at the start of the evaluation period. The increased EDSS must be sustained for at least 3 months before enrollment. Final eligibility is determined by a selection and safety monitoring committee consisting of Drs Jerry Wolinsky (University of Texas, Houston, TX) and Henry McFarland (National Institutes of Health, Bethesda, MD).

SLE. Patients may be enrolled if they fulfill any one of the following criteria: (1) biopsy-proven World Health Organization (WHO) class III or IV glomerulonephritis that has failed to respond to NIH short course cyclophosphamide therapy³² (500 to 1,000 mg/m² monthly for at least 6 months), with treatment failure defined as a failure of serum creatinine to return to normal or pre-exacerbation level; (2) vasculitis and/or immune complex deposition causing end organ signs or symptoms, eg, cerebritis, transverse myelitis, pulmonary hemorrhage, or cardiac failure, not controlled with corticosteroids and cyclophosphamide; (3) transfusion-dependent cytopenias that are immune-mediated and not controlled with danazol, prednisone, and an alkylating agent (cyclophosphamide or vincristine); or (4) catastrophic antiphospholipid syndrome, which is defined as an antiphospholipid titer greater than 5 standard deviations above the mean and two or more antiphospholipid related manifestations, including either cytopenias or vascular thrombosis that failed to respond to anticoagulant therapy.

RA. Patients must fulfill all of the following criteria: (1) an established clinical diagnosis of RA by the American College of Rheumatology criteria³³; (2) a positive rheumatoid factor; and (3) failure of at least two disease-modifying agents (methotrexate, gold, penicillamine, and hydroxychloroquine), where failure is defined as at least six swollen joints and either 30 or more involved (swelling, tenderness, deformity, pain on motion, and decreased motion) joints or answering less than 75% of the Activities of Daily Living (ADL) Health Assessments Questionnaire "without any difficulty."³⁴

Hematopoietic Stem Cell (HSC) Procurement

Hematopoietic stem cells were collected from bone marrow in the first 3 patients, but due to low CD34⁺ cell yield, the graft was supplemented with peripheral blood stem cells (PBSCs). All subsequent hematopoietic stem cells were collected using only peripheral blood. PBSCs were mobilized with either granulocyte colony-stimulating factor (G-CSF; Amgen, Thousand Oaks, CA) at 10 µg/kg subcutaneous daily with leukapheresis beginning on day 5 or cyclophosphamide (2.0 g/m²) and G-CSF (10 µg/kg/d) with leukapheresis initiated when the white blood cell count reached 1,000/µL (1.0 × 10⁹/L). Apheresis was continued daily until the number of harvested progenitor cells reached a

minimum of 2.0 × 10⁶ CD34⁺ cells/kg body weight after CD34 enrichment. The mobilized peripheral blood stem cells were lymphocyte-depleted via positive selection for CD34⁺ cells using the CEPRATE SC Stem Cell Concentrator (CellPro, Bothell, WA).

Conditioning Regimen

For MS, cyclophosphamide (120 mg/kg) in divided doses of 60 mg/kg/d was administered intravenously over 2 hours on days -6 and -5 and total body irradiation (TBI) was administered as 1,200 cGy divided 150 cGy twice a day on days -4, -3, -2, and -1 in the AP/PA position with 50% lung and 30% kidney and right lobe of the liver transmission blocks. One gram of methylprednisolone was administered intravenously on days -4, -3, -2, and -1.

For lupus and RA, cyclophosphamide (200 mg/kg) was administered in divided doses of 50 mg/kg/d intravenously over 1 to 2 hours on days -7, -6, -5, and -4. Antithymocyte globulin (ATG; 90 mg/kg) was administered in doses of 30 mg/kg/d on days -6, -5, and -4 and infused over 10 to 12 hours beginning 8 to 10 hours after the infusion of cyclophosphamide. Methylprednisolone (1 g) was administered intravenously 30 minutes before each dose of ATG.

Definition of Disease Status

Outcome was based on assessments before transplant and at 1, 2, 3, and 6 months and yearly after transplantation.

MS. Improvement was defined as a decrease in the Kurtzke EDSS³¹ by at least 1 point or increase in the Scripps NRS³⁵ by at least 10 points. Deterioration was defined as an increase in the Kurtzke EDSS by at least 1 point or decrease in the Scripps NRS by at least 10 points. Stabilization of active disease was defined as absence of any new or progressive neurologic deficits and no significant change in the EDSS or NRS scores. MRI was performed at approximately the same intervals to monitor occurrence of new lesions and activity of lesions as determined by gadolinium enhancement.

SLE. Outcome was based on serology (C3, C4, anti-Ds-DNA, ANA, Sm, anti-SSA, anti-SS-B, and lupus anticoagulant), lupus disease activity index (SLEDAI),³⁶ and response of pretransplant abnormalities in involved organ systems (eg, serum creatinine; 24-hour urine protein and creatinine clearance in nephritis; left ventricular ejection fraction in myocarditis; and chest radiograph and pulmonary function tests in pneumonitis). Improvement was defined as a 50% improvement in any baseline parameter with no deterioration in any objective parameter.

RA. Assessment parameters were tender joint count, swollen joint count, patient's assessment of pain, patient's global assessment of disease, physician's global assessment, Health Assessment Questionnaire Activities of Daily Living (ADL), and acute-phase reactant value. Definition of improvement was greater than 20% improvement in both tender and swollen joint count and 20% improvement in at least three of the other five assessment parameters.³⁷ Criteria for complete remission requires that five or more of the following be fulfilled for at least 2 consecutive months: (1) duration of morning stiffness not exceeding 15 minutes, (2) no fatigue, (3) no joint pain (by history), (4) no joint tenderness or pain on motion, (5) no soft tissue swelling in joints or tendon sheaths, and (6) erythrocyte sedimentation rate less than 30 mm/h for a female or 20 mm/h for a male.³⁸

Immunologic Assays

Pretransplant and posttransplant, two- and three-color immunophenotyping was performed on EDTA anticoagulated whole blood. The infused stem cell products were assessed by three-color immunophenotyping. The panel of fluorescein isothiocyanate (FITC), phycoerythrin (PE), PE-cyanin 5 (PE-Cy5), or PerCp fluorochromes included antibodies to CD45 and CD34 (Becton Dickinson, Mountain View, CA), and

Table 1. Hematopoietic Stem Cell Collection in Patients With Autoimmune Diseases

Patient	Method of PBSC Collection	No. of Apheresis	CD34 ⁺ Cells/kg Infused ×10 ⁶	CD3 ⁺ Cells/kg Infused ×10 ⁶ (log depletion)	CD19 ⁺ Cells/kg Infused ×10 ⁶ (log depletion)
MS #1	G-CSF	2 plus 1 bone marrow harvest	2.12	0.50 (2.5)	0.12 (2.51)
MS #2	G-CSF	2 plus 1 bone marrow harvest	3.91	1.4 (1.93)	1.0 (1.9)
MS #3	G-CSF	2 plus 1 bone marrow harvest	2.42	1.06 (2.2)	0.5 (1.9)
MS #4	G-CSF	1	2.57	0.23 (2.6)	1.28 (1.6)
MS #5	G-CSF	1	2.02	1.04 (2.5)	0.23 (2.3)
MS #6	G-CSF	1	2.76	0.26 (2.86)	0.72 (1.92)
SLE #1	Cy + G-CSF	3	2.46	1.5 (1.4)	0.0004 (3.74)
SLE #2	Cy + G-CSF	5	2.0	1.0 (2.0)	0.06 (2.51)
RA #1	Cy + G-CSF	2	2.16	0.86 (2.7)	0.12 (2.18)
RA #2	Cy + G-CSF	1	6.32	0.60 (2.5)	0.10 (2.43)

Abbreviation: Cy, cyclophosphamide (2.0 g/m²).

CD3, CD4, CD8, CD29, CD45RA, CD19, CD16, and CD56 (Coulter Cytometry; Coulter, Hialeah, FL).

Supportive Care

Patients were treated on a hepa-filtered hematology/oncology floor. A low microbial diet, fluconazole (400 mg/d oral or intravenous), and valacyclovir (500 mg TID oral or intravenous) were started upon admission and discontinued when the absolute neutrophil count (ANC) rebounded to 500/μL. Oral ciprofloxacin (750 mg orally BID) was started upon admission and switched to intravenous piperacillin/tazobactam or cefipime when the ANC fell below 500/μL. Subcutaneous G-CSF (5 μg/kg) was started the day of hematopoietic stem cell infusion and continued until the ANC was greater than 1,000/μL for 3 consecutive days. For the first 6 months after transplantation, patients were treated with either daily oral fluconazole or itraconazole and either Bactrim DS once orally three times a week or, for patients with lupus, aerosolized pentamidine (300 mg) monthly.

RESULTS

Hematopoietic Stem Cell Collection

In general, one to five daily 10 to 20 L apheresis were required to obtain greater than 2.0×10^6 CD34⁺ cells/kg after lymphocyte depletion. In 3 patients with MS, apheresis was used to supplement the bone marrow harvests (Table 1). Positive selection for CD34⁺ stem cells resulted in a median 2.3 log depletion of T (CD3⁺) and B (CD19⁺) cells.

Toxicity

Nonhematologic toxicity was limited to grade 0-1 for the gastrointestinal system (nausea, vomiting, and diarrhea) according to NCI common toxicity criteria (Table 2). Median time to an absolute neutrophil count greater than 500/μL ($0.5 \times 10^9/L$) and platelet count greater than 20,000/μL ($20 \times 10^9/L$) occurred on day 10 and 14, respectively. The median time to hospital discharge was day 14 after transplant. Five patients had positive cultures from either stool (candida and clostridium), blood (staphylococcus), or percutaneous intravenous central catheter (streptococcus) during the period of neutropenia. After engraftment, no posttransplant opportunistic infections have occurred, with the exception of a single case of dermatomal varicella zoster occurring 6 months after transplantation.

MS. Pretransplant G-CSF was well tolerated without exacerbation of neurologic symptoms. Transient elevation of hepatic transaminases ($5 \times$ normal) occurred in 2 patients while receiving G-CSF for stem cell mobilization. These values normalized within 7 days without intervention. Patients with MS tolerated chemotherapy, TBI, and G-CSF without neurologic deterioration or exacerbation.

SLE. The first patient with lupus started transplantation while in acute renal failure with a creatinine level of 5.0 mg/dL. In this patient, dialysis was initiated before starting the condi-

Table 2. Toxicity

Patient	ANC >500/μL (day)	Platelet >20,000/μL	No. of Days With Fever (>100.5)	Positive Cultures	Maximum NCI Toxicity, Renal	Maximum NCI Toxicity, Pulmonary	Maximum NCI Toxicity, Hepatic	Maximum NCI Toxicity, CNS
MS #1	11	14	3	Stool-candida	1-trace hematuria	0	1 elevated transaminase	0
MS #2	11	14	9	Blood-staphylococcus	1-trace hematuria	0	3 elevated transaminase	0
MS #3	12	14	0	0	0	0	0	0
MS #4	11	12	0	0	0	0	0	0
MS #5	11	13	0	0	0	0	0	0
MS #6	10	12	0	Stool-clostridium difficile	0	0	1 elevated transaminase	0
SLE #1	10	14	2	Blood-staphylococcus epidermidis	3-creatinine 6.8 mg/dL	0	2 elevated transaminase	0
SLE #2	8	14	2	0	2	4	3 elevated transaminase	0
RA #1	8	12	1	0	1-trace hematuria	0	1 elevated transaminase	0
RA #2	9	12	3	PICC line streptococcus sangoisi	1-trace hematuria	0	2 elevated transaminase	0

Abbreviation: PICC, percutaneous intravenous central catheter.

tioning regimen. High-dose chemotherapy was associated with a cell-lysis effect, acid base and electrolyte disorders, and volume disturbances. The second patient underwent transplant for recurrent alveolar hemorrhage refractory to corticosteroids and cyclophosphamide. Before starting transplantation, she required supplemental face mask oxygen. With the exception of volume and electrolyte disturbances, organ function generally stabilized or began to improve during conditioning or shortly after onset of neutropenia.

RA. In both patients, conditioning was well tolerated without any pulmonary, renal, cardiac, or hepatic dysfunction. However, 1 patient had an immediate wheal and flare response to the subcutaneous test dose of ATG. After desensitization, full-dose ATG was administered by continuous intravenous infusion for 24 hours and then discontinued without administration over the last 48 hours due to urticaria.

Clinical Outcome

MS. All 6 patients had rapidly progressive disease despite maximal immunosuppressive therapy during the year before transplant. Since transplantation, disease has not progressed despite stopping all immunosuppressive and immune modulating medications. Posttransplant follow-up ranges from 5 to 17 months, with a median of 11 months, and 3 patients are greater than 1 year from transplant. All patients have experienced subjective and objective neurologic improvements (Table 3), but the Kurtzke EDSS remains unchanged due to its dependence in the upper range on lower extremity motor function for scoring. In contrast, the Scripps NRS, which includes more emphasis on upper extremity function, incontinence, and cognitive ability, has demonstrated a greater than 10 point improvement in 3 patients. Improvement in the NRS did not begin until several months after transplantation. The 3 patients whose NRS

improved by more than 10 points had gadolinium enhancement on their pretransplant MRI and had lower EDSS scores (6.0 to 7.5). In contrast, 2 of the 3 patients who had stabilization of disease but no significant improvement did not have pretransplant gadolinium enhancement and higher pretransplant EDSS scores (8.0 to 8.5). In all 6 patients, MRI showed no new or enhancing lesions after transplantation.

SLE. Both patients have had no evidence of active disease since transplantation (Table 4). The first patient, a 24-year-old white woman, was diagnosed with SLE at 11 years of age. Her C3, C4, ANA, and anti-ds DNA have never been normal, even during clinical remissions. She was treated at various times with plasmapheresis, corticosteroids, pulse cyclophosphamide, hydroxychloroquine, methotrexate, and azathioprine, none of which allowed steroid tapering to less than 20 mg/d. At transplantation, she exhibited active lupus manifested by a malar rash, arthralgias, hematuria, diffuse abdominal pain, ascites, and a large pericardial effusion. Renal function was rapidly decreasing, with a serum creatinine level of 5.0 mg/dL (398 $\mu\text{mol/L}$), red blood cell (RBC) casts in the urine, a 24-hour urine protein level of 3.8 g, and biopsy-proven WHO class IV glomerulonephritis. Pancytopenia was present and serology and complement were abnormal (Table 4). Over the 12 months since transplant, the malar rash, arthralgias, pleural and pericardial effusions, and cytopenias have resolved. Renal function has stabilized at a creatinine level of 1.9 mg/dL and 24-hour urine protein level of 0.2 g. For the first time since disease onset 13 years ago, complement components (C3, C4) and antinuclear antibodies are normal with the patient off all immunosuppressive medications, including corticosteroids.

The second patient presented at 15 years of age with flu-like symptoms followed by respiratory arrest. She was transferred to a tertiary care facility in which pulmonary biopsy showed

Table 3. Clinical Outcome After Hematopoietic Stem Cell Transplantation for MS

Patient	Time Since Transplant (mo)	Stable or Improved Disease (mo)	No. of Relapses	EDSS Pretransplant and Most Recent Posttransplant	NRS Pretransplant and Most Recent Posttransplant	MRI	Neurological Changes
MS #1 secondary progressive	17	17	0	8.5/8.5	15/19	No new lesions	Improved cognition, decreased incontinence
MS #2 secondary progressive	16	16	0	8.0/8.0	30/29	No new lesions	Decreased incontinence, decreased tremor
MS #3 primary progressive	15	15	0	8.0/8.0	34/37	No new lesions	Improved stamina and strength, now able to swim
MS #4 secondary progressive	6	6	0	6.5/6.0	49/63	No new lesions	Less dysarthria, correction of gaze deviation
MS #5 primary progressive	5	5	0	7.0/6.5	29/58	No new lesions	Improved strength
MS #6 secondary progressive	5	5	0	6.0/6.0	51/61	No new lesions	Improved strength and cerebellar function

The EDSS is divided into 20 half points from 0 (normal) to 10 (death). Increasing points indicate worse neurologic deficits. In the higher range (5.0 to 8.0), the EDSS is weighted by ambulation. For example, ambulation less than 200 meters without assist is 5.0; wheelchair bound is 8.0. In the NRS, normal neurologic function is 100 points, whereas lower numbers indicate progressive neurologic disability.

Abbreviations: EDSS, Kurtzke extended status disability scale²¹; MRI, magnetic resonance spectroscopy; NRS, Scripps Neurologic Rating Scale.²⁵

Table 4. Clinical Outcome After Hematopoietic Stem Cell Transplantation for SLE

	Patient SLE #1			
	Pretransplant	2 mo Posttransplant	6 mo Posttransplant	12 mo Posttransplant
SLEDAI score	37	0	0	0
Hemoglobin (mg/dL)	6.5	10.0	12.3	12.4
White blood cells/ μ L	2,000	6,000	6,100	6,100
Platelets (no./ μ L)	100,000	240,000	215,000	235,000
24-hour urine protien (g)	3.6	0.6	0.87	0.2
Serum creatinine (mg/dL)	5.0	2.6	2.7	1.9
C3 (68-164 mg/dL)	52 low	100 normal	114 normal	113 normal
C4 (7-47 mg/dL)	6 low	40 normal	35 normal	35 normal
ANA	1:320 high	1:40 normal	1:40 normal	1:80 increased
Anti-double-stranded DNA	1:160 high	1:60 borderline	1:60 borderline	1:60 borderline

	Patient SLE #2*		
	Pretransplant	2 mo Posttransplant	6 mo Posttransplant
SLEDAI score	32	0	0
Hemoglobin (mg/dL)	8.0	10.1	11.8
Platelets (no./ μ L)	135,000	236,000	242,000
Chest roentograph	Bilateral Infiltrates	Normal	Normal
	Alveolar Hemorrhage		
	Vasculitis		
Pulmonary function test, DLCO	50% predicted	50% predicted	50% predicted
Serum creatinine (mg/dL)	1.3	1.1	1.1
C3	62 low	190 high	167 normal
C4	10 normal	45 normal	29 normal
ANA	1:1280 high	1:40 normal	1:160 increased
Anti-double-stranded DNA	1:320 high	Negative	Negative

The SLEDAI is a measure of activity or inflammation in nine organ systems. It has a theoretical maximum of 105 points. A normal person has a SLEDAI of zero.

Abbreviation: SLEDAI, SLE disease activity index.²⁶

*White blood cell count and 24-hour urine protein on SLE patient no. 2 was always normal and is not included in the table.

alveolar hemorrhage and vasculitis and renal biopsy showed WHO class IV glomerulonephritis. The ANA was 1:1,280. Subsequently, her disease manifested predominately as alveolar hemorrhage requiring intubation on a second occasion despite high-dose corticosteroids, plasmapheresis, and pulse cyclophosphamide. Within 3 months of disease onset she was referred for transplantation. At the time of transplant, the patient had active pneumonitis manifested as hemoptysis, pulmonary infiltrates, and hypoxia. Since transplantation, hemoptysis and pulmonary

infiltrates have resolved. Pulmonary diffusion capacity has remained unchanged at 50% of normal. Corticosteroids are being gradually tapered and have been decreased from 80 to 25 mg/d.

RA. The first patient has met the 50% response criteria for improvement (Table 5). She was a 46-year-old woman diagnosed with RA 7 years before transplantation. She was treated with nonsteroidal anti-inflammatory drugs, hydroxychloroquine, cyclosporine, gold, methotrexate, dapsone, sulfasalazine,

Table 5. Clinical Outcome After Hematopoietic Stem Cell Transplantation for RA

	Patient No. 1					Patient No. 2	
	Pretransplant	1 mo After Transplant	3 mo After Transplant	6 mo After Transplant	12 mo After Transplant	Pretransplant	3 mo After Transplant
Swollen joint count	27	0	4	3	2	18	4
Tender joint count	41	3	7	3	4	21	22
Patient's global assessment of disease	65% poor	35% fair-well	25% well	15% very well	10% very well	75 poor	50 poor
Physician global assessment of disease	75% poor	5% very well	5% very well	10% very well	10% very well	67 fair	35 fair
ADL % without problems	0%	60%	80%	85%	80%	0%	25%
Sed rate	48 high	—	15 normal	—	15 normal	75 high	65 high

The ADL address ability to perform activities of daily living in eight categories: dressing and grooming, arising, eating, walking, hygiene, reach, grip, and other. A normal person's ADL is 100%.

Abbreviations: ADL, activities of daily living questionnaire²⁴; sed rate, sedimentation rate.

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