

the trial results indicated differences in the toxicity profiles of the two regimens, with relatively more frequent grade 3 or 4 mucositis, nausea and vomiting, and grade 2 alopecia with FOLFIRI chemotherapy and more frequent grade 3 or 4 neutropenia and neurotoxicity with FOLFOX chemotherapy.^{10,11}

In the last decade, the introduction of targeted biological therapy has transformed the paradigm for treatment of mCRC. Antiangiogenesis agents, such as bevacizumab,¹² and epidermal growth factor receptor antagonists, such as cetuximab,¹³ have been added to the standard combination chemotherapy (FOLFOX or FOLFIRI), with improved survival outcomes reported in patients with mCRC. Angiogenesis, the process by which new blood and lymphatic vessels are formed, is required to support growth in the embryo and young animals, as well as to allow tissue repair and remodeling in adults.¹⁴ For tumors, as with normal tissues, oxygen and nutrients are obtained and waste products are removed via the vasculature. Although tumors can grow in part by co-opting existing host vessels,¹⁵ most tumors induce new vessel formation (neovascularization), suggesting that angiogenesis is a hallmark of tumor growth and function throughout the tumor's life cycle. The predominant growth factor, or proangiogenic factor, of the angiogenesis pathway is vascular endothelial growth factor (VEGF).^{16,17} Its continuous expression by the tu-

mor makes VEGF a rational target for cancer therapy.¹⁷

This article focuses on the clinical review of ziv-aflibercept (ziv'' - a flib' er sept; Zaltrap, Regeneron Pharmaceuticals and sanofi-aventis), a novel fully humanized monoclonal antibody with antiangiogenic activity recently approved by the Food and Drug Administration (FDA) for use in combination with second-line FOLFIRI chemotherapy for patients with mCRC who have not responded to an oxaliplatin-containing first-line regimen.

Pharmacology

The mammalian VEGF family^{14,18} consists of six distinct glycoproteins, or growth factors: VEGF-A, VEGF-B, VEGF-C, VEGF-D, VEGF-E, and placental growth factor (PlGF). These growth factors are ligands that bind to VEGF receptors that mediate tumor angiogenesis. Among these, VEGF-A (or simply VEGF) is considered the principal driving force in tumor angiogenesis.¹⁸ Each ligand has specific affinity for different receptors.

There are three VEGF receptors: VEGFR-1, VEGFR-2, and VEGFR-3. All of these VEGF receptors are characterized by seven immunoglobulin (Ig)-like domains in the extracellular region, a single transmembrane domain, and an intracellular tyrosine kinase domain.¹⁹ These VEGF receptors are high-affinity (denoted by low dissociation constant) receptors, with tyrosine kinase activity expressed in normal and tumor vasculature (i.e.,

vascular endothelial cells).^{18,20} The major receptor for VEGF-A is VEGFR-2 (flk1/KDR). Although VEGF-A binds to VEGFR-1 with 10 times higher affinity than does VEGFR-2, the higher tyrosine kinase activity of VEGFR-2 makes it the most important effector in VEGF signaling.^{18,20} Of note, PlGF, which binds to VEGFR-1 but not VEGFR-2, also contributes to tumor angiogenesis.¹⁹

Ziv-aflibercept, a fully humanized soluble recombinant fusion protein, is created by fusing extracellular Ig domain 2 of VEGFR-1 and extracellular Ig domain 3 of VEGFR-2 to the Fc (constant) region of human IgG1, resulting in the formation of a more potent and high-affinity VEGF blocker than bevacizumab.¹⁹ This fusion protein has been demonstrated to have pharmacologically improved activity over bevacizumab (Table 1).²¹⁻²³ Since this fusion protein blocks the angiogenesis pathway by binding to all isoforms of VEGF-A, VEGF-B, and PlGF, it is also known as a "VEGF trap" or a composite decoy receptor (due to the fusion of both VEGFR-1 and VEGFR-2 to IgG1).²¹ By binding to these endogenous ligands, ziv-aflibercept inhibits the binding and activation of their cognate receptors, resulting in inhibition of downstream signaling events that mediate neovascularization and vascular permeability. This process results in maintenance of a more functional and normal vasculature,^{18,19} inhibition of tumor growth and tumor metastasis, and potentially improved drug delivery (due to "normalization" of tumor vasculature that enables tumor cells to become more sensitive to cytotoxic chemotherapy).²⁴

Pharmacokinetics and pharmacodynamics

Ziv-aflibercept binds 1:1 to endogenous VEGF to form stable inert complexes, which appear in the circulation at a maximum level within 24–48 hours of treatment.²⁵

Table 1.
Key Differences in Biological Properties of Ziv-aflibercept and Bevacizumab^{21-23,a}

Property	Ziv-aflibercept	Bevacizumab
Binding target(s)	VEGF-A, VEGF-B, PlGF	VEGF-A
Equilibrium dissociation constant, ^b pM	0.49	58
Elimination half-life, days	5–6	20

^aVEGF = vascular endothelial growth factor, PlGF = placental growth factor.
^bA measure of binding affinity, with smaller values indicating stronger binding.

These complexes are retained in the systemic circulation. Clearance occurs via a receptor- or pinocytome-mediated pathway that results in proteolysis.

Pharmacokinetic studies of ziv-aflibercept involved the measurement of plasma concentrations of free and VEGF-bound ziv-aflibercept.²⁶⁻²⁸ The level of VEGF-bound ziv-aflibercept indicates the amount of endogenous VEGF produced in normal and tumor tissues, whereas free ziv-aflibercept is available for binding with newly secreted VEGF. Free ziv-aflibercept concentrations exhibit linear kinetics at a dose range of 2–9 mg/kg, with steady-state concentrations of free ziv-aflibercept reached by the second dose. Maximum VEGF-bound ziv-aflibercept levels are reached at doses of 2 mg/kg i.v. or greater, indicating complete blockade of the ligand. In pharmacokinetic studies, the half-life of ziv-aflibercept increased (from 1.7 to 5.1 days) as doses were increased from 0.3 to 7.0 mg/kg i.v. Clearance was stable at doses in the range of 2–7 mg/kg. There was no accumulation of free ziv-aflibercept between treatment cycles.

Some studies also demonstrated that the biological effects of ziv-aflibercept correlated closely with free ziv-aflibercept levels in excess of VEGF-bound ziv-aflibercept levels.²⁶⁻²⁹ The mean maximum observed plasma concentration of free ziv-aflibercept, at doses ranging from 0.3 to 7.0 mg/kg (0.3 mg/kg is about a tenth of the biologically effective dose in humans), varied from 4 to 159 µg/mL.²⁹ Following the administration of ziv-aflibercept at 2- and 3-mg/kg doses, free drug concentrations were similar to VEGF-bound ziv-aflibercept concentrations. With doses of ≥4 mg/kg, free ziv-aflibercept concentrations were in excess of bound ziv-aflibercept concentrations throughout the dosing intervals in most patients. Thus, a 4-mg/kg i.v. dose provides a sufficient ziv-aflibercept concentration

to block endogenous production of VEGF.

Van Cutsem and colleagues²⁶ showed that clearance and concentrations of free and bound ziv-aflibercept at steady state in patients treated with irinotecan-based combination chemotherapy were comparable to those observed with ziv-aflibercept monotherapy. No clinically significant pharmacokinetic drug–drug interactions were found between ziv-aflibercept, irinotecan, and SN-38 the active metabolite of irinotecan).

Clinical trials

Phase I research. The safety and dose determination of single-agent ziv-aflibercept therapy were explored in a dose-escalation trial conducted by Lockhart and colleagues²⁹ that included 7 patients with mCRC among a total of 47 patients with refractory solid tumors. Seven dose levels were evaluated (0.3, 1.0, 2.0, 3.0, 4.0, 5.0, and 7.0 mg/kg); 3, 7, 6, 7, 7, 4, and 13 patients were treated at each respective dose level. The investigators also evaluated the safety and maximum tolerable dose (MTD) of ziv-aflibercept at doses ranging from 0.3 to 7.0 mg/kg i.v. every two weeks. The MTD was defined as the highest dose at which 2 of 3–6 treated patients experienced a dose-limiting toxicity (DLT): uncontrolled hypertension, grade 3 or 4 proteinuria, febrile neutropenia, grade 4 neutropenia, grade 3 or 4 thrombocytopenia, and any toxicity of any grade that led to drug discontinuation. DLTs were evaluated during each dose-escalation phase. Tumor response was evaluated after every two cycles (or four treatments per eight-week interval) according to the RECIST (Response Evaluation Criteria In Solid Tumors, version 1.0) scheme. Patients could elect to continue treatment until disease progression or intolerable toxicity or until they chose to withdraw consent.

Results showed that single-agent ziv-aflibercept was generally well tol-

erated, with a rate of partial response of 6% and a rate of stable disease for greater than one year of 4%. The types of treatment-related toxicities observed (e.g., fatigue, hypertension, proteinuria) were consistent with those reported in other trials evaluating antiangiogenic therapies.³⁰⁻³⁴ The median times to the onset of hypertension or proteinuria were 3.5 days (range, 1–21 days) and 15 days (range, 14–16 days), respectively. These adverse events were reversible on drug discontinuation or initiation of supportive care. Based on these observations and observed increases in the frequency and severity of adverse events at doses of ≥4 mg/kg, single-agent ziv-aflibercept 4 mg/kg i.v. every two weeks was determined to be the recommended dosage for use in Phase II trials.

In another study, investigators evaluated the safety, efficacy, DLT profile, and recommended dose of ziv-aflibercept in combination with irinotecan-based chemotherapy for second-line treatment of Japanese patients with mCRC, 10 (63%) of whom had received prior bevacizumab therapy.^{28,35} Ziv-aflibercept was administered at two dosage levels (2 and 4 mg/kg i.v. every two weeks, administered to 3 and 13 patients, respectively). The median numbers of cycles administered were 6 (range, 3–9) and 10 (range, 1–23), and the total numbers of cycles of the 2- and 4-mg/kg doses were 18 and 131, respectively. No DLTs were observed at either dosing level. The response rate and PFS for patients receiving the 4-mg/kg i.v. dose were 8% and 7.6 months, respectively. At both dosing levels, the most common adverse events (occurring in ≥40% of patients) included bone marrow suppression (e.g., neutropenia) and gastrointestinal symptoms (e.g., nausea, vomiting, diarrhea, stomatitis, decreased appetite); these toxicities are known to be associated with FOLFIRI regimens. In addition, toxicities known to be associated with

the antiangiogenesis therapy, such as hypertension and epistaxis, were also frequently observed (rates of $\geq 40\%$) with use of the 4-mg/kg dose. The most common grade 3 or 4 adverse events were neutropenia and hypertension. This study established the recommended dose and schedule of ziv-aflibercept (i.e., 4 mg/kg i.v. every two weeks) to be used in combination with irinotecan-based chemotherapy for mCRC.

Of note, ziv-aflibercept has been evaluated extensively for the treatment of patients with other advanced solid tumors.³⁰⁻³⁴ Most of the patients involved had received prior chemotherapy regimens, either as first- or higher-line therapies. Ziv-aflibercept was found to have very modest clinical activity as a monotherapy; however, when it was used as a component of combination therapies, partial-response rates ranged from 4% to 18%.

Phase II research. Tang and colleagues³⁶ conducted a multicenter open-label Phase II trial in 75 heavily treated mCRC patients with good performance status (i.e., Eastern Cooperative Oncology Group [ECOG] performance status of 2 or less). Patients had received a median of two prior regimens and were stratified as having received ($n = 51$) or not received ($n = 24$) prior bevacizumab therapy. The primary endpoints were response rate and PFS. After a median of 4 treatment cycles (range, 1–16 cycles) of single-agent ziv-aflibercept therapy (4 mg/kg i.v. every 2 weeks), median PFS was 2.4 months among patients who had received prior bevacizumab therapy and 2.0 months in the bevacizumab-naïve group. The efficacy of ziv-aflibercept did not seem to be affected by prior bevacizumab therapy (duration not specified). In the prior-bevacizumab cohort, 1 patient had a partial response and 6 patients (12%) had stable disease for greater than 16 weeks. In the bevacizumab-naïve cohort, the best response was stable disease

for greater than 16 weeks in 5 of 24 patients (21%). Overall, the most common adverse events of any grade were fatigue (68%), hypertension (51%), proteinuria (49%), headache (42%), voice alteration (31%), anorexia (24%), and joint pain (18%). Single-agent ziv-aflibercept appeared to be tolerated and to have modest antitumor activity in pretreated patients with mCRC, including those who had received prior bevacizumab therapy.

A notable limitation of the trial of Tang et al.³⁶ was that the activity of ziv-aflibercept against mCRC has never been directly compared with that of bevacizumab. The best conclusion that one can draw from this study is that ziv-aflibercept appears to be neither dramatically better nor worse than bevacizumab; this underlines the need for more studies with different drug combinations and direct comparisons with bevacizumab-based therapies.

Phase III research. Because of the acceptable clinical activity of ziv-aflibercept when used in combination therapies,^{28-31,35} a Phase III prospective, multinational, double-blind, parallel-arm trial (the VELOUR trial) was conducted in patients with mCRC who had experienced disease progression while receiving oxaliplatin-based chemotherapy (e.g., FOLFOLX).³⁷

The primary study endpoint was OS, as determined by the planned subgroup analysis. Secondary endpoints were PFS and response rate. A total of 1226 patients with mCRC and an ECOG score of 2 or less were randomly assigned in a 1:1 fashion to receive ziv-aflibercept ($n = 612$) or a placebo ($n = 614$) followed by FOLFIRI chemotherapy every two weeks. Patients were stratified based on prior bevacizumab treatment and performance status (an ECOG score of 0 or 1 versus a score of 2). The study had a power of 90% to detect a 20% improvement in OS in the ziv-aflibercept group. All pa-

tients had received prior oxaliplatin therapy.

Ziv-aflibercept was given as a 4-mg/kg i.v. infusion prior to FOLFIRI chemotherapy. The FOLFIRI regimen was administered according to the National Comprehensive Cancer Network (NCCN) colon cancer guideline (irinotecan hydrochloride 180 mg/m² i.v. on day 1 of each two-week cycle, to be administered concurrently with leucovorin 400 mg/m² i.v., followed by fluorouracil 400 mg/m² by i.v. bolus and then fluorouracil 2400 mg/m² by continuous i.v. infusion over 46 hours³⁸). The use of supportive care medications, including atropine (for prophylaxis of acute diarrhea due to irinotecan) and colony-stimulating factor (for prophylaxis of neutropenia) was permitted. Disease response was evaluated radiologically every six weeks. Treatment continued until disease progression or intolerable toxicity. The study groups were well balanced in terms of performance status and prior bevacizumab therapy. Approximately 30% of patients (186 of 612) in the ziv-aflibercept group had received bevacizumab, compared with 31% of patients (187 of 614) in the placebo group. The overall median age was 61 years; the majority of patients (87%) were white, 7% were Asians, and 3.5% were black.

At a median follow-up of 22.3 months, the two-year survival rates were 28.0% and 18.7% in the ziv-aflibercept and control groups, respectively. Relative to placebo users, patients treated with ziv-aflibercept had longer OS (13.5 months versus 12.1 months; hazard ratio, 0.817; 95% confidence interval, 0.71–0.94 months; $p = 0.0032$), a higher response rate (19.8% versus 11.1%, $p < 0.001$), and longer PFS (6.9 months versus 4.7 months, $p < 0.0001$). Patients receiving antiangiogenic therapy were given a median of seven cycles of ziv-aflibercept (the numbers of cycles of FOLFIRI and ziv-aflibercept varied among these patients); those in the control group received eight cycles

of placebo administration. Irinotecan and fluorouracil were administered for a median of 21 weeks in the ziv-aflibercept group and 18.1 weeks in the control group.

Cycle delays and dose modifications were more common among patients receiving ziv-aflibercept. Dose intensity (defined as the ratio of the delivered chemotherapy dose to the standard full dose, expressed as a percentage) was 83% in the ziv-aflibercept group versus 92% in the control group. No significant relationship was observed between treatment outcomes and prior bevacizumab exposure. In a subgroup analysis, a greater OS benefit related to ziv-aflibercept therapy was observed only in patients with liver metastasis as opposed to metastasis to other organs. The benefits in OS and PFS seen in the bevacizumab-pretreated group were consistent with those observed in the overall population.

Adverse events among VELOUR trial participants included diarrhea, asthenia, stomatitis, ulceration, nausea, and infections. These adverse events were primarily grade 1 or 2 and were more common in patients treated with ziv-aflibercept.

Overall, adverse events of all grades were similar in both study groups.

Grade 3 or 4 adverse events related to antiangiogenesis therapy that occurred more frequently in the ziv-aflibercept group relative to the control group included hypertension (19% versus 1.5%), hemorrhage (2.9% versus 1.7%), arterial thromboembolic events (1.8% versus 0.5%), venous thromboembolic events (8% versus 6%), and proteinuria (8% versus 1%). Surprisingly, a significant increase in chemotherapy-induced grade 3 or 4 toxicities—particularly diarrhea (19% versus 8%), asthenia (17% versus 11%), stomatitis (13% versus 5%), and infections (12% versus 7%)—was also observed with antiangiogenesis therapy (Table 2). This apparent increase in ziv-aflibercept-associated toxicities was not in line with Phase III study results on bevacizumab use in patients with mCRC.³⁸ However, prior bevacizumab treatment did not seem to impact the safety profile of ziv-aflibercept. The most common adverse events that led to treatment discontinuation included asthenia, infections, diarrhea, and hypertension.

Based on the results that demonstrated statistically significant improvement in OS, PFS, and response rate in patients with mCRC who received ziv-aflibercept as a second-

line therapy with or without prior bevacizumab therapy, FDA approved ziv-aflibercept for use in this patient population in 2012.²²

Safety

Overall, ziv-aflibercept appears to be tolerated, even in heavily pretreated mCRC populations. In the VELOUR trial, the most common adverse effects observed in patients receiving ziv-aflibercept in combination with the FOLFIRI regimen were leucopenia, neutropenia, thrombocytopenia, diarrhea, proteinuria, and hypertension.^{22,37} These adverse effects were mostly grade 1 or 2 and reversible. Of note, some of these common adverse effects (hypertension and proteinuria) were among the most common DLTs identified in Phase I and II trials²⁶⁻²⁹ and were consistent with the inhibition of the angiogenesis pathway. Other adverse effects occurring more commonly in ziv-aflibercept versus control groups (e.g., increases in liver function enzymes, stomatitis, fatigue, decreased appetite) may not be related to ziv-aflibercept's antiangiogenic effect.

Class-related adverse effects. Hypertension, proteinuria, and thromboembolism are recognized as the

Table 2.

Most Common Adverse Effects of Ziv-aflibercept in Phase III Trial^{37,a}

Adverse Effect	% Patients Affected			
	FOLFIRI + Ziv-aflibercept (n = 611)		FOLFIRI + Placebo (n = 605)	
	All Grades	Grade 3 or 4	All Grades	Grade 3 or 4
Leukopenia	78	16	72	12
Neutropenia	67	37	57	30
Thrombocytopenia	48	3	35	2
Hypertension	41	19	11	2
Diarrhea	69	19	57	8
Stomatitis	50	13	33	5
Proteinuria	62	8	41	1
Fatigue	48	13	39	8
AST elevation	62	3	54	2
ALT elevation	50	3	39	2

^aFOLFIRI = leucovorin, fluorouracil, and irinotecan; AST = aspartate transaminase; ALT = alanine transaminase.

hallmark class-related adverse effects associated with antiangiogenic therapy.³⁹⁻⁴² Hypertension associated with antiangiogenic therapy is often of low grade (1 or 2). Even grade 3 or 4 hypertension may still be reversible, and patients are managed with angiotensin-converting enzyme inhibitors, calcium channel blockers, or diuretics.⁴³

Proteinuria (all grades) was reported in 49% of patients in the VELOUR trial.³⁷ The frequency of grade 3 or 4 proteinuria was 7.8%; dose modification may be necessary for patients who develop this complication. The drug manufacturer recommends monitoring for proteinuria by urine dipstick (or urinalysis) and determination of the urinary protein-to-creatinine ratio (UPCR) prior to each dose of ziv-aflibercept.²² For patients with a UPCR greater than 1, analysis of a 24-hour urine collection is recommended.

In addition, inhibition of the VEGF signaling pathway leads to dysfunctional endothelial cells in the vascular endothelium, causing activation of tissue factor and an increased risk of thrombosis. Experience with bevacizumab demonstrated that bevacizumab-VEGF complexes induced platelet aggregation and led to the thromboembolic events observed in clinical trials.^{23,44} Currently, it remains unclear whether the biologically inert ziv-aflibercept-VEGF complex is associated with a lower frequency of thromboembolic events. In the VELOUR trial, grade 3 or 4 thromboembolic complications occurred in less than 1% of patients.³⁷ It is, however, important to note that cancer patients in general are at an increased risk for thrombosis due to their procoagulant disease state and that antiangiogenic therapy marginally increases this risk.⁴⁴

Black-box warning. Ziv-aflibercept carries an FDA-mandated black-box warning on treatment-related hemorrhage, gastrointestinal perforation, and compromised

wound healing.²² In clinical trials, the rate of bleeding and hemorrhage together was 37.8% in the ziv-aflibercept groups versus 19% in the control groups^{35,37}; the occurrence of grade 3 or 4 hemorrhage, including severe intracranial hemorrhage, was also slightly higher with ziv-aflibercept use (3% versus 1.7%).²² Patients receiving ziv-aflibercept should be monitored for signs and symptoms of bleeding. Therapy should be discontinued in patients who develop severe hemorrhage. Although the rates of gastrointestinal perforation were similar in both study groups in the VELOUR trial, patients receiving antiangiogenic therapy should be monitored and treatment should be discontinued if gastrointestinal perforation develops.

Pregnancy consideration. Ziv-aflibercept is classified as a pregnancy category D agent. It should be used in pregnancy only if the potential benefit justifies the potential risk to the fetus.²² Both female and male patients of reproductive potential must use effective contraception during treatment and for a minimum of three months after the last dose of ziv-aflibercept.

Immunogenicity and hypersensitivity. During clinical research, the frequency of development of antibodies to ziv-aflibercept was 3.1% in patients receiving ziv-aflibercept versus 1.7% in patients receiving a placebo²²; among patients who tested positive for anti-ziv-aflibercept antibody, neutralizing antibodies were detected in 17 of 48 patients administered ziv-aflibercept and in 2 of 40 placebo users. The impact of neutralizing antibodies on treatment efficacy and safety remains unclear. Since the antibody assay may be influenced by many factors, direct comparison of the frequency of anti-ziv-aflibercept antibody formation with the corresponding rate for other protein-based medications may be misleading. Currently, it is

not known if such antibody formation is persistent enough to cause infusion-related reactions or reduced ziv-aflibercept exposure. No data are available on the usefulness of routine screening for positive antibody.

In regard to the frequency of severe (grade 3 or 4) hypersensitivity reactions, no difference was found in ziv-aflibercept versus control patients in the VELOUR trial (0.3% versus 0.5%).³⁷

Other safety considerations. Ziv-aflibercept, known simply as aflibercept in European and other countries, is specifically named with the prefix “ziv” (derived from the brand name Zaltrap and the abbreviation i.v.) in the United States in accordance with the FDA requirement that drug manufacturers reduce the potential for confusion related to sound-alike and look-alike medication names. The World Health Organization’s international nonproprietary name for Zaltrap is aflibercept.

FDA required that the U.S. generic name ziv-aflibercept be used to clearly differentiate the formulation approved for use in managing mCRC from its ophthalmic counterpart, aflibercept, which is marketed under the proprietary name Eylea (Regeneron Pharmaceuticals) in a different strength.²² The scientific literature still uses aflibercept as the generic name for ziv-aflibercept. It is recommended that ziv-aflibercept be stored, segregated, and labeled with a specific warning to help distinguish the product from the ophthalmic product.

Dosage, preparation, and administration

The recommended dosage of ziv-aflibercept is 4 mg/kg i.v. administered every two weeks prior to FOLFIRI combination chemotherapy. Treatment is continued until disease progression or intolerable toxicity. Ziv-aflibercept is supplied as a ready-to-use single-dose vial

of either 200 or 100 mg (solution concentration, 25 mg/mL). After the withdrawal of the calculated dose from the vial, the drug is further diluted in a vehicle of 0.9% sodium chloride injection or 5% dextrose injection to yield a final ziv-aflibercept concentration of 0.6–8 mg/mL. Diluted solution must be refrigerated and used within four hours.²²

Ziv-aflibercept should be administered as an intermittent infusion over one hour through a 0.2- μ m polyethersulfone filter.²² An administration set made of one of the following materials may be used: polyvinyl chloride (PVC)-containing diethylhexyl phthalate (DEHP), DEHP-free PVC containing trioctyl trimellitate, polypropylene, polyethylene-lined PVC, and polyurethane. Ziv-aflibercept is administered alone prior to any component of FOLFIRI combination chemotherapy.

Ziv-aflibercept has not been shown to have clinically significant interactions with irinotecan, SN-38, and fluorouracil.^{26,37} The manufacturer has not conducted dedicated drug–drug interaction studies.²²

The dosage of ziv-aflibercept does not need to be adjusted for renal impairment.²² The VELOUR trial data indicated that no dosage adjustment is needed for mild hepatic impairment; however, no information is available on the potential need for dosage adjustments in severe hepatic impairment (defined in the VELOUR trial as a total bilirubin level greater than three times the upper limit of normal).³⁷

Ziv-aflibercept should be discontinued for any of the following toxicities: arterial thromboembolic events, fistula formation, gastrointestinal perforation, severe hemorrhage, hypertensive crisis, hypertensive encephalopathy, nephrotic syndrome, thrombotic microangiopathy, and reversible posterior leukoencephalopathy syndrome.^{22,44}

Dosage modifications should be made for patients with severe

or uncontrolled hypertension; ziv-aflibercept should be withheld until blood pressure is controlled, with the dose subsequently reduced to 2 mg/kg.²² For patient with substantial proteinuria (i.e., ≥ 2 g in 24 hours), ziv-aflibercept should be withheld and only administered when the protein level declines below that level. If proteinuria recurs, ziv-aflibercept must be discontinued until the urine protein level falls below 2 g in 24 hours, with therapy resumed at a reduced dose of 2 mg/kg.

Patients must be monitored for hematologic toxicity at baseline and prior to each cycle of ziv-aflibercept.²² Treatment with ziv-aflibercept and FOLFIRI chemotherapy must be delayed until the neutrophil count is $\geq 1.5 \times 10^9$ cells/L. Furthermore, ziv-aflibercept has been shown to delay and compromise wound healing. Treatment should be stopped at least four weeks prior to elective surgery and should not be resumed until at least four weeks after surgery or until the surgical wound is fully healed.

No dosage adjustment is necessary for the elderly. Based on a population pharmacokinetic analysis,²² race and sex did not have a clinically significant effect on exposure to free ziv-aflibercept. Currently, there is no recommendation for ziv-aflibercept dosing in obese patients. In one of the Phase I studies summarized above, patients weighing more than 100 kg had a 29% increase in systemic exposure to ziv-aflibercept compared with patients weighing 50–100 kg.³⁵ Furthermore, no recommendation for a maximum total dose of ziv-aflibercept has been made. In the VELOUR trial, a maximum patient weight was not specified.^{22,37}

The use of FOLFIRI combination chemotherapy requires the calculation of body surface area (BSA).²² In the VELOUR trial, if a patient's BSA was greater than 2 m², the amounts of irinotecan and fluorouracil used

were adjusted downward to the doses corresponding to a BSA of 2 m² for safety purposes.^{22,37}

Cost considerations

The average wholesale acquisition cost (WAC) of ziv-aflibercept is \$1,600 per 100-mg vial or \$3,200 per 200-mg vial.⁴⁵ Recently, the drug manufacturer discounted the cost by 50% for hospitals and clinics through its wholesalers and through group purchasing organizations. With a dose of 4 mg/kg and an average patient weight of 72 kg, an estimated dose for each treatment would be about 288 mg. Thus, each dose would cost approximately \$4,800. Based on the clinical trial data, the median treatment duration of ziv-aflibercept was 6.9 months (PFS).³⁷ The cost of ziv-aflibercept alone for this treatment course is about \$66,240. Since the drug was approved as an add-on to the standard second-line chemotherapy combination for mCRC (FOLFIRI), the total cost of the ziv-aflibercept–FOLFIRI regimen is approximately \$80,000 for a patient with an average BSA (1.73 m²). It is apparent that the use of ziv-aflibercept will result in a significant increase in overall mCRC treatment expenditure that may raise concerns among payers about the growing cost of oncology treatment.

On the other hand, the WAC of bevacizumab is \$596 for a 100-mg vial or \$2387 for a 400-mg vial.⁴⁵ Given the recommended dose of 5 mg/kg with standard chemotherapy, each dose of bevacizumab costs approximately \$2387—only half the cost of ziv-aflibercept. The significant difference in cost between bevacizumab and ziv-aflibercept has brought renewed attention to cost issues in cancer care.⁴⁶ Whether an increased cost is justified by additional clinical efficacy is a robust area of comparative effectiveness research that is likely to continue.

Place in therapy and future directions

The Phase III VELOUR trial demonstrated a statistically significant OS benefit, with improved PFS, in favor of ziv-aflibercept in combination with FOLFIRI chemotherapy relative to FOLFIRI chemotherapy alone in patients with mCRC that has progressed with an oxaliplatin-based regimen.³⁷ Common adverse effects are related to the mechanism-based effect of antiangiogenic therapy. Currently, there are no data to suggest that FOLFIRI plus ziv-aflibercept therapy has significant antitumor activity in patients whose disease has progressed during treatment with FOLFIRI plus bevacizumab. Ziv-aflibercept has only been shown to have antitumor activity when given in combination with FOLFIRI in FOLFIRI-naïve patients. Therefore, NCCN recommends ziv-aflibercept as a second-line option to be used in combination with FOLFIRI or irinotecan only in patients with disease progression while receiving therapy not containing irinotecan.³⁸

Considerable strides have been made regarding the use of bevacizumab beyond disease progression in the metastatic setting. In the setting of second-line chemotherapy that does not consist of FOLFIRI, current opinion seems to favor bevacizumab.⁴⁷⁻⁵⁰ Ziv-aflibercept has yet to show safety and efficacy with chemotherapy regimens other than FOLFIRI in the second-line setting. Recently, a large Phase II trial (AFFIRM) of FOLFOX with or without ziv-aflibercept therapy for first-line treatment demonstrated no improvement in efficacy outcomes with ziv-aflibercept use.⁵¹ In contrast, the activity of bevacizumab in the second-line treatment of patients with disease progression during first-line or even maintenance bevacizumab therapy is supported by some studies.⁴⁷⁻⁵⁰ Similar efficacy, however, cannot be assumed for ziv-aflibercept based on current data.^{22,37}

Ziv-aflibercept has the broad pharmacologic advantage of targeting endogenous ligands that are pro-angiogenic: VEGF-A, VEGF-B, and PlGF. How much of this advantage is translated to real clinical benefits is yet to be determined. On the other hand, the survival benefits conferred by bevacizumab have been clearly demonstrated in first-line, second-line, and maintenance treatment settings.⁴⁷⁻⁵⁰ Bevacizumab also seems to have a more favorable safety profile than ziv-aflibercept (i.e., a lower potential to induce proteinuria and minimal constitutional toxicities).²³

Ziv-aflibercept is appropriate for use as an add-on therapy in combination with irinotecan-based regimens in patients with mCRC who have not received irinotecan previously. At the time of writing, it cannot be recommended as first-line therapy, but an ongoing trial (NCT016522196) that is investigating the role of ziv-aflibercept in combination with FOLFOX chemotherapy in patients with previously untreated mCRC may shed light on the place of therapy for ziv-aflibercept.⁵²

Like bevacizumab, ziv-aflibercept may be complementary to other cytotoxic chemotherapy agents. It has been studied in several tumor types and may be useful in various combinations.^{26-31,40} Its distinctive mechanism of action may allow its use against other advanced solid tumors.⁵³ A Phase III trial of ziv-aflibercept plus docetaxel therapy for locally advanced or metastatic non-small-cell lung cancer demonstrated efficacy in terms of PFS and response rate but no major improvement in OS.⁵⁴ In another Phase III trial, combination therapy with ziv-aflibercept and gemcitabine was shown to produce no significant improvement in metastatic pancreatic cancer.⁵⁵ In metastatic prostate cancer refractory to hormonal therapy, recently published data from a Phase III study (the VENICE trial) showed that ziv-aflibercept plus docetaxel and

prednisone failed to improve survival outcomes.⁵⁶ Until more study results are available, due to the aforementioned cost and reimbursement issues, ziv-aflibercept is indicated for selected patients with mCRC who have good performance status and whose disease is refractory to first-line oxaliplatin-based chemotherapy.

Conclusion

Current clinical data are insufficient to directly compare ziv-aflibercept and bevacizumab when used with standard combination chemotherapy as first- or second-line regimens for mCRC. The role of ziv-aflibercept is currently limited to the second-line setting in combination with irinotecan-based regimens in mCRC patients who have not received irinotecan previously. The role of ziv-aflibercept for other tumor types is yet to be determined.

References

1. Parkin DM, Whelan SL, Ferlay J. Cancer incidence in five continents. Vol. VIII. Lyon, France: International Agency for Research on Cancer; 2002. IARC scientific publications no. 155.
2. American Cancer Society. Cancer facts and figures 2012. www.cancer.org/research/cancerfactsstatistics/cancerfactsfigures2012/index (accessed 2012 Dec 25).
3. National Cancer Institute. SEER (surveillance, epidemiology and end results) statistics. http://seer.cancer.gov (accessed 2013 Jan 17).
4. Cidon EU. The challenge of metastatic colorectal cancer. *Clin Med Insights Oncol.* 2010; 4:55-60.
5. Obrand DJ, Gordon PH. Incidence and patterns of recurrence following curative resection for colorectal carcinoma. *Dis Colon Rectum.* 1997; 40:15-24.
6. Kelly H, Goldberg RM. Systemic therapy for metastatic colorectal cancer: current opinions, current evidence. *J Clin Oncol.* 2005; 23:4553-60.
7. Saltz LB, Cox JV, Blanke C et al. Irinotecan plus fluorouracil and leucovorin for metastatic colorectal cancer. *N Engl J Med.* 2000; 343:905-14.
8. Goldberg RM, Sargent DJ, Morton RF et al. A randomized controlled trial of fluorouracil plus leucovorin, irinotecan, and oxaliplatin combinations in patients with previously untreated metastatic colorectal cancer. *J Clin Oncol.* 2004; 22:23-30.
9. Tournigand C, Andre T, Achille E et al. FOLFIRI followed by FOLFOX6 or the

- reverse sequence in advanced colorectal cancer: a randomized GERCOR study. *J Clin Oncol*. 2004; 22:229-37.
10. Song X, Zhao Z, Barber B et al. Characterizing medical care by disease phase in metastatic colorectal cancer. *J Oncol Pract*. 2011; 7:255-305.
 11. Ragnhainmar P, Hafstrom L, Nygren P et al. A systemic overview of chemotherapy effects in colorectal cancer. *Acta Oncol*. 2001; 40:282-308.
 12. Saltz LB, Clarke S, Diaz-Rubio E et al. Bevacizumab in combination with oxaliplatin-based chemotherapy as first-line therapy in metastatic colorectal cancer: a randomized phase III study. *J Clin Oncol*. 2008; 26:2013-9.
 13. Van Cutsem E, Kohne CH, Hitre E et al. Cetuximab and chemotherapy as initial treatment for metastatic colorectal cancer. *N Engl J Med*. 2009; 360:1408-17.
 14. Ferrara N. Vascular endothelial growth factor: basic science and clinical progress. *Endocr Rev*. 2004; 25:581-611.
 15. Holash J, Maisonpierre PC, Compton D et al. Vessel cooption, regression, and growth in tumors mediated by angiopoietins and VEGF. *Science*. 1999; 284:1994-8.
 16. Kerbel RS. Tumor angiogenesis. *N Engl J Med*. 2008; 358:2039-49.
 17. Folkman J, Klagsbrun M. Angiogenic factors. *Science*. 1987; 235:442-7.
 18. Ferrara N. Vascular endothelial growth factor as a target for anticancer therapy. *Oncologist*. 2004; 9(suppl 1):2-10.
 19. Chu QS. Aflibercept (AVE 0005): an alternative strategy for inhibiting tumor angiogenesis by vascular endothelial growth factors. *Expert Opin Biol Ther*. 2009; 9:263-71.
 20. Shibuya M. Tyrosine kinase receptor Flt/VEGFR family: its characterization related to antiangiogenesis and cancer. *Genes Cancer*. 2010; 1:1119-23.
 21. Holash J, Davis S, Papadopoulos N et al. VEGF-trap: a VEGF blocker with potent antitumor effects. *Proc Natl Acad Sci USA*. 2002; 99:11393-8.
 22. Zaltrap (ziv-aflibercept) prescribing information. Bridgewater, NJ: sanofi-aventis; 2012 Aug.
 23. Avastin (bevacizumab) prescribing information. South San Francisco, CA: Genentech; 2011 Feb.
 24. Jain RK. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science*. 2005; 307:58-62.
 25. Rudge JS, Holash J, Hylton D et al. VEGF trap complex formation measures production rates of VEGF, providing a biomarker for predicting efficacious angiogenesis blockade. *Proc Natl Acad Sci USA*. 2007; 104:18363-70.
 26. Van Cutsem E, Khayat D, Verslype C et al. Phase I dose-escalation study of intravenous aflibercept administered in combination with irinotecan, 5-fluorouracil and leucovorin in patients with advanced solid tumors. *Eur J Cancer*. 2013; 49:17-24.
 27. Khayat D, Tejpar S, Spano JP et al. Intravenous aflibercept administered in combination with irinotecan, 5-fluorouracil and leucovorin in patients with advanced solid tumours: results from the expansion cohort of a phase I study. *Eur J Cancer*. 2013; 49:790-7.
 28. Yamazaki K, Yoshino T, Yamaguchi K et al. Phase I dose escalation and pharmacokinetics study of intravenous aflibercept plus irinotecan, 5-fluorouracil, and folinic acid (FOLFIRI) in patients with metastatic colorectal cancer. *J Clin Oncol*. 2011; 29(suppl 4):538. Abstract.
 29. Lockhart AC, Rothenberg ML, Dupont J et al. Phase I study of intravenous vascular endothelial growth factor trap, aflibercept, in patients with advanced solid tumors. *J Clin Oncol*. 2010; 28:207-14.
 30. Verslype C, Spano J, van Cutsem E et al. Validation of the selected dose of aflibercept (VEGF-Trap) plus irinotecan, 5-fluorouracil, and leucovorin (I-IV5FU2) in a phase I clinical trial of patients with advanced solid tumors: preliminary results. *J Clin Oncol*. 2008; 26(suppl 15):14540. Abstract.
 31. Rixe O, Verslype C, Khayat D et al. A phase I dose-escalation (DE) and pharmacokinetics (PK) study of intravenous (iv) aflibercept (VEGF Trap) plus irinotecan, 5-fluorouracil, and leucovorin (I-IV5FU2) in patients with advanced solid tumors (STs). *J Clin Oncol*. 2008; 26(suppl 15):3557. Abstract.
 32. Fukasawa M, Kore M. Vascular endothelial growth factor-trap suppresses tumorigenicity of multiple pancreatic cancer cell lines. *Clin Cancer Res*. 2004; 10:327-32.
 33. Dupont J, Rothenberg MI, Spriggs DR et al. Safety and pharmacokinetics of intravenous VEGF trap in a phase I clinical trial of patients with advanced solid tumors. *J Clin Oncol*. 2005; 23(suppl 16):3029. Abstract.
 34. Isambert N, Freyer G, Zanetta S et al. Phase I dose-escalation study of intravenous aflibercept in combination with docetaxel in patients with advanced solid tumors. *Clin Cancer Res*. 2012; 18:1743-50.
 35. Yoshino T, Yamazaki K, Yamaguchi K. A phase I study of iv aflibercept with FOLFIRI in Japanese patients with previously treated metastatic colorectal cancer. *Invest New Drugs*. 2013; 31:910-7.
 36. Tang R, Cohen SJ, Kollmannsberger C et al. Phase II clinical and pharmacokinetic study of aflibercept in patients with previously treated metastatic colorectal cancer. *Clin Cancer Res*. 2012; 18:6023-31.
 37. Van Cutsem E, Tabernero J, Lakomy R et al. Addition of aflibercept to fluorouracil, leucovorin, and irinotecan improves survival in a phase III randomized trial in patients with metastatic colorectal cancer previously treated with an oxaliplatin-based regimen. *J Clin Oncol*. 2012; 30:3499-506.
 38. National Comprehensive Cancer Network. Clinical practice guidelines in oncology, colon cancer. V.3.2013. www.nccn.org (accessed 2013 Mar 6).
 39. Hurwitz HI, Fehrenbacher L, Hainsworth JD et al. Bevacizumab in combination with fluorouracil and leucovorin: an active regimen for first-line metastatic colorectal cancer. *J Clin Oncol*. 2005; 23:3502-8.
 40. Gaya A, Tse V. A preclinical and clinical review of aflibercept for the management of cancer. *Cancer Treat Rev*. 2012; 38:484-93.
 41. Roodhart JM, Langenberg MH, Witteveen E. The molecular basis of class side effects due to treatment with inhibitors of the VEGF/VEGFR pathway. *Curr Clin Pharmacol*. 2008; 3:132-43.
 42. Lankhorst S, Kappers MH, van Esch JH et al. Mechanism of hypertension and proteinuria during angiogenesis inhibition: evolving role of endothelin-1. *J Hypertension*. 2013; 31:444-54.
 43. Maitland ML, Bakris GL, Black HR et al. Initial assessment, surveillance, and management of blood pressure in patients receiving vascular endothelial growth factor signaling pathway inhibitors. *J Natl Cancer Inst*. 2010; 102:596-604.
 44. Eremina V, Quaggin SE. Biology of anti-angiogenic therapy-induced thrombotic microangiopathy. *Semin Nephrol*. 2010; 30:582-90.
 45. Drug topics. Ziv-aflibercept (Zaltrap). Red book online [Internet database]. Greenwood Village, CO: Thompson Micromedex. Updated periodically.
 46. Swain S. The high cost of a cancer drug: an oncologist's view. www.nytimes.com/2012/10/20/opinion/the-high-cost-of-a-cancer-drug-an-oncologists-view.html?_r=0 (accessed 2013 Mar 7).
 47. Arnold D, Andre T, Bennaoui J et al. Bevacizumab (BEV) plus chemotherapy (CT) continued beyond first progression in patients with metastatic colorectal cancer (mCRC) previously treated with BEV plus CT: results of a randomized phase III intergroup study (TML study). *J Clin Oncol*. 2012; 30(suppl 18):CRA3503. Abstract.
 48. Giantonio BJ, Catalano PJ, Neropol NJ et al. Bevacizumab in combination with oxaliplatin, fluorouracil, leucovorin (FOLFOX4) for previously treated metastatic colorectal cancer: results from the Eastern Cooperative Oncology Group Study E3200. *J Clin Oncol*. 2007; 25:1539-44.
 49. Masi G. A randomized phase III study evaluating the continuation of bevacizumab (BV) beyond progression in metastatic colorectal cancer (mCRC) patients (pts) who received BV as part of first-line treatment: results of the BEBYP trial by the Gruppo Oncologico Nord Ovest (GONO). Presentation at 37th European Society for Medical Oncology Congress, Vienna, Austria; 2012 Sep 28-Oct 2.
 50. Grothey A, Sugrue MM, Purdie DM et al. Bevacizumab beyond first progression is associated with prolonged overall survival

- al in metastatic colorectal cancer: results from a large observational cohort study (BRiTE). *J Clin Oncol*. 2008; 26:5326-34.
51. Pericay C, Folprechr G, Saunders M et al. Phase 2 randomized, noncomparative, open-label study of aflibercept and modified FOLFOX6 in the first-line treatment of metastatic colorectal cancer. (AFFIRM). *Ann Oncol*. 2012; 23(suppl 4):IV16.
 52. ClinicalTrials.gov. Aflibercept and FOLFOX6 treatment for previously untreated stage IV colorectal cancer. www.clinicaltrials.gov/show/NCT016522196 (accessed 2013 Mar 7).
 53. Teng LS, Jin KT, He KF et al. Clinical applications of VEGF-Trap (aflibercept) in cancer treatment. *J Chin Med Assoc*. 2010; 73:449-56.
 54. Novello S, Ramlay R, Gorbunova VA et al. Aflibercept in combination with docetaxel for second-line treatment of locally advanced or metastatic nonsmall-cell lung cancer (NSCLC): final results of a multinational placebo controlled phase III trial (EFC10261-VITAL). In: Proceedings of 14th World Conference on Lung Cancer. Amsterdam, Netherlands; 2011 Jul 3-7. Abstract O43.06
 55. Riess H, Manges R, Karasek P et al. Double-blind, placebo-controlled randomized phase III trial of aflibercept (A) plus gemcitabine (G) versus placebo (P) plus gemcitabine (G) in patients with metastatic pancreatic cancer: final results. In: Proceedings of 12th World Congress on Gastrointestinal Cancer. Barcelona, Spain; 2010 Jun 30–Jul 3. Abstract O-0006.
 56. Tannock I, Fizazi K, Ivanov S et al. Aflibercept versus placebo in combination with docetaxel/prednisone for first-line treatment of men with metastatic castration-resistant prostate cancer (mCRPC): results from the multinational phase III trial (VENICE). *J Clin Oncol*. 2013; 31(suppl 6):13. Abstract.

Increased Renal Expression of Vascular Endothelial Growth Factor (VEGF) and Its Receptor VEGFR-2 in Experimental Diabetes

Mark E. Cooper, Dimitria Vranes, Sherif Youssef, Steven A. Stacker, Alison J. Cox, Bishoy Rizkalla, David J. Casley, Leon A. Bach, Darren J. Kelly, and Richard E. Gilbert

It has been suggested that the cytokine vascular endothelial growth factor (VEGF) has an important role in the pathogenesis of diabetic retinopathy, but its role in nephropathy has not been clearly demonstrated. Assessment of VEGF, ^{125}I -VEGF binding, and vascular endothelial growth factor receptor-2 (VEGFR-2) in the kidney was performed after 3 and 32 weeks of streptozotocin-induced diabetes. Gene expression of both VEGF and VEGFR-2 was assessed by Northern blot analysis and the localization of the ligand and receptor was examined by *in situ* hybridization. VEGF and VEGFR-2 protein were also evaluated by immunohistochemistry. Binding of the radioligand ^{125}I -VEGF was evaluated by *in vitro* and *in vivo* autoradiography. Diabetes was associated with increased renal VEGF gene expression. VEGF mRNA and protein were localized to the visceral epithelial cells of the glomerulus and to distal tubules and collecting ducts in both diabetic and nondiabetic rats. Renal VEGFR-2 mRNA was increased after 3 weeks of diabetes but not in long-term diabetes. *In situ* hybridization and immunohistochemical studies revealed that glomerular endothelial cells were the major site of VEGFR-2 expression. In addition, VEGFR-2 gene expression was detected in cortical and renomedullary interstitial cells and on endothelial cells of peritubular capillaries. There was an increase in ^{125}I -VEGF binding sites after 3 but not 32 weeks of diabetes. The major VEGF binding sites were in the glomeruli. ^{125}I -VEGF binding was also observed in medullary rays and in the renal papillae. These studies indicate an early and persistent increase in renal VEGF gene expression in association with experimental diabetes. In addition, an early and transient increase in renal VEGF receptors was also observed in diabetic rats. These findings are consistent with a role for VEGF

in mediating some of the changes observed in the diabetic kidney. *Diabetes* 48:2229-2239, 1999

Vascular endothelial growth factor (VEGF), also known as vascular permeability factor, is a term used to describe a family of potent multifunctional cytokines (1). The VEGF-A gene produces five closely related species by alternative splicing. VEGF-A₁₆₅ is the most abundantly expressed species and is a 165-amino acid glycoprotein with 20% homology to the A and B chains of platelet-derived growth factor (1). At least two high-affinity receptor-binding sites have been reported for VEGF. These receptors have been cloned and named flt-1 (vascular endothelial growth factor receptor [VEGFR]-1) and flk-1/KDR/NYK (VEGFR-2) and were initially viewed to be endothelial-specific tyrosine kinases (2,3). More recently, it has been determined that these receptors are present on other cell types (4). Recent studies suggest that it is the biological interaction of VEGF with VEGFR-2 that is involved in inducing the spectrum of biological responses of VEGF (1,5). VEGF also has a selective action of promoting vascular permeability (6), being 50,000 times more potent than histamine on a molar basis (7). Radioligand binding studies with iodinated recombinant VEGF have confirmed a wide distribution of VEGF binding sites in the rat, including the kidney (8). More recently, VEGF binding has also been detected in the adult and fetal human kidney (9).

Recent studies have emphasized the potential role of VEGF in diabetic retinopathy (10,11), with several groups reporting an increase in VEGF and its receptors in the diabetic retina (12,13). It has been shown that there is a close relationship between proliferative diabetic retinopathy and nephropathy (14). Furthermore, several of the putative pathways implicated in the pathogenesis of diabetic retinopathy are also viewed to be involved in the development of diabetic nephropathy (15,16). However, the status of VEGF and its receptors in the kidney in experimental diabetes has not been previously characterized.

RESEARCH DESIGN AND METHODS

Experimental plan. Diabetes was induced by streptozotocin (Sigma, St. Louis, MO) injection (diabetic, 60 mg/kg i.v.) or sham injection with citrate buffer injection (control) after an overnight fast in Sprague-Dawley rats weighing 200–250 g. Before the rats were killed, blood was collected for determination of plasma glucose by the glucose oxidase technique (17) and systolic blood pressure was measured by tail cuff plethysmography (18). The animals were anesthetized with

From the Department of Medicine (M.E.C., D.V., S.Y., A.J.C., B.R., D.J.C., L.A.B., D.J.K., R.E.G.), University of Melbourne, Austin and Repatriation Medical Centre (Repatriation Campus), West Heidelberg, Victoria; and the Ludwig Institute for Cancer Research (S.A.S.), Royal Melbourne Hospital, Parkville, Victoria, Australia.

Address correspondence and reprint requests to Associate Professor Mark E. Cooper, Department of Medicine, Austin and Repatriation Medical Centre (Repatriation Campus), West Heidelberg, Victoria 3081, Australia. Email: cooper@austin.unimelb.edu.au.

Received for publication 11 February 1999 and accepted in revised form 20 July 1999.

AGE, advanced glycated end product; IC₅₀, concentration of unlabeled ligand required to displace 50% of the radioligand; NSB, nonspecific binding; PBS, phosphate-buffered saline; PKC, protein kinase C; ROD, relative optical density; SSC, sodium chloride-sodium citrate; TGF, transforming growth factor; VEGF, vascular endothelial growth factor; VEGFR-2, vascular endothelial growth factor receptor-2.

pentobarbital sodium (Nembutal 50 mg/kg body weight i.v.; Bomac Laboratories, Asquith, Australia) and killed after 3 or 32 weeks of diabetes. Both kidneys were then removed. One kidney was snap frozen in liquid nitrogen-cooled isopentane for use in extracting RNA for Northern hybridization and for *in vitro* autoradiographic binding studies. The other kidney was bisected and immediately fixed in freshly prepared 4% paraformaldehyde in preparation for *in situ* hybridization. A separate group of control and diabetic rats were prepared in a manner similar to that described above, except that both kidneys were removed after infusion of radioactive VEGF tracer for the *in vivo* autoradiographic binding studies.

Northern hybridization of VEGF and its receptors

Preparation of RNA and synthesis of probes. Total RNA was isolated from the kidney at the time when the animals were killed using the acid guanidium thiocyanate-phenol-chloroform extraction method (19). RNA purity and concentration were determined spectrophotometrically. Twenty-microgram samples were denatured and electrophoresed through 0.8% agarose formaldehyde gels. RNA integrity was verified by examination of the 28S and 18S ribosomal bands of ethidium bromide-stained material under ultraviolet light. RNA was then transferred by capillary action onto nylon filters (Hybond-N; Amersham, Bucks, U.K.) and fixed under ultraviolet light. Filters were then probed with the cDNA coding for either VEGF or its receptors, VEGFR-1 and VEGFR-2 (13). The probes were then labeled with [³²P]dCTP by randomly primed DNA synthesis (Boehringer Mannheim, Mannheim, Germany). Hybridization was performed at 42°C for 16 h in 50% formamide, 45 mmol/l Na₂HPO₄, 5× Denhardt's solution, 0.5% SDS, and sonicated salmon sperm DNA. After hybridization, filters were washed in solutions of decreasing ionic strength and increasing temperature, followed by exposure of filters to X-ray film for 5–14 days.

In situ hybridization of VEGF and VEGFR-2

Synthesis of riboprobes. Sense and antisense RNA probes for VEGF and VEGFR-2 were generated by *in vitro* transcription (Promega, Madison, WI), as previously described (13). In brief, linearized template (500 ng) was added to a reaction mixture of transcription buffer (final volume 20 µl) containing 6 mmol/l dithiothreitol, 333 µmol/l each of ATP, CTP, and GTP, 12 µmol/l UTP, 100 µCi [³²P]UTP (2,000 Ci/mmol, New-Dupont, Boston, MA), 20 U RNAsin (Boehringer-Mannheim), and 20 U RNA polymerase (Boehringer-Mannheim). The reaction mixture was incubated at 37°C for 90 min, after which the DNA template was digested with 1 U RNase-free DNase for 15 min. The riboprobe was precipitated by ammonium acetate and ethanol using yeast tRNA as a carrier and then reconstituted in 100 µl of water. Purified riboprobe length was adjusted to ~150 bases by alkaline hydrolysis.

***In situ* hybridization.** Four-micrometer-thick sections cut from paraformaldehyde-fixed paraffin-embedded kidney tissue were placed onto slides precoated with 3-aminopropyltriethoxysilane and left overnight at 37°C. Tissue sections were dewaxed in histolene, rehydrated in graded ethanols to deionized milliQ water, equilibrated in P buffer (50 mmol/l Tris-HCl, pH 7.5, 5 mmol/l EDTA), and incubated in 125 µg/ml Pronase E (Sigma) in P buffer for 10 min at 37°C. Sections were then washed in 0.1 mol/l sodium phosphate buffer (pH 7.2), briefly refixed in 4% paraformaldehyde for 10 min, rinsed in milliQ water, dehydrated in 70% ethanol, and air dried.

Hybridization buffer containing 2 × 10⁶ cpm/µl riboprobe in 300 mmol/l NaCl, 10 mmol/l Tris-HCl (pH 7.5), 10 mmol/l Na₂HPO₄, 5 mmol/l EDTA (pH 8.0), 1× Denhardt's solution, 50% formamide, 17 mg/ml yeast RNA, and 10% wt/vol dextran sulfate was heated to 85°C for 5 min. A 25-µl aliquot of this solution was then applied to each tissue section under coverslips. Hybridization of tissue to the riboprobe was performed overnight at 60°C in 50% formamide-humidified chambers. Sense probes for VEGF and its receptor, neuroepithelial tyrosine kinase (NYK), were used on a further set of tissue sections as controls for nonspecific binding.

After hybridization, slides were washed in 2× sodium chloride-sodium citrate (SSC) containing 50% formamide prewarmed to 50°C to remove coverslips. Sections were then washed in the above-mentioned solution for 1 h at 55°C, rinsed three times in RNase buffer (10 mmol/l Tris-HCl, pH 7.5, 1 mmol/l EDTA, pH 8.0, 0.5 mol/l NaCl), and then incubated with RNase A (150 µg/ml) for 1 h at 37°C. Sections were later washed in 2× SSC for 45 min at 55°C, dehydrated in graded ethanol, air dried, and exposed to Kodak X-Omat autoradiographic film (Kodak, Rochester, NY) for 1–3 days. Slides were then dipped in Ilford K5 nuclear emulsion (Ilford, Moberley, U.K.), stored in a light-free box with desiccant at room temperature for 2–3 weeks, and developed in Kodak D19, followed by fixation with Ilford Hypam. Sections were then stained with hematoxylin and eosin for examination under light microscopy.

Quantitation of Northern and *in situ* hybridization X-ray films. The relative intensity of autoradiograms of the Northern blots was determined by scanning densitometry (Ultrocan XL; LKB, Bromma, Sweden). All results were corrected for differences in RNA loading by rehybridization with an oligonucleotide probe for 18S rRNA. Densitometry of autoradiographic images from kidney sections, obtained by *in situ* hybridization, was performed as previously described (20), using a Micro Computer Imaging Device (MCID; Imaging Research, St.

Catharines, Ontario, Canada), according to the methods of Baskin and Stahl (21). In brief, *in situ* autoradiographic images were placed on a uniformly illuminating fluorescent light box (Northern Light Precision Luminator Model C60; Image Research) and captured using a video camera (Sony Video Camera Module CCD; Tokyo) connected to an IBM AT computer with a 512-by-512 pixel array imaging board with 256 gray levels. In view of the focal nature of glomerular [¹²⁵I]VEGF binding, the mean of 20–25 quantifications of these areas was assessed for each kidney section (22). After calibration, construction of a curve of optical versus radioactivity density was possible and quantitation of digitalized autoradiographic images is performed using MCID software (22). Results were then expressed as relative optical density (ROD) (23). All measurements were performed by an independent observer (B.R.) in a masked manner.

Immunohistochemistry. Sections 4 µm thick were placed onto slides, rehydrated, and treated with 1% H₂O₂/methanol followed by incubation in Protein Blocking Agent (Lipshaw-Immuno, Pittsburgh, PA) for 20 min at room temperature, as previously described (13). Sections were next incubated with a polyclonal rabbit antibody directed against the NH₂-terminal 21 amino acids of human VEGF (Santa Cruz Biotechnology, Santa Cruz, CA) overnight at 4°C, washed in phosphate-buffered saline (PBS), and incubated with biotinylated goat anti-rabbit IgG (DAKO, Carpinteria, CA). After washing with PBS, sections were incubated with avidin-biotin complex (Vector, Burlingame, CA). Peroxidase conjugates were subsequently localized by diaminobenzidine tetrahydrochloride as a chromogen. Sections were counterstained with hematoxylin. Antibody specificity had been previously confirmed by preabsorption of antibody with recombinant VEGF and by incubation without the primary antiserum (13). A similar procedure was used for assessment of VEGFR-2 protein using a polyclonal rabbit antibody directed against the COOH-terminal 20 amino acids of the mouse flk-1 protein (3).

VEGF binding in the kidney

Iodination of VEGF-165. Recombinant human VEGF (R + D Systems, Minneapolis, MN) (0.5 µg) was iodinated with ¹²⁵I using the chloramine T method (24). A 5-µl (0.5 mCi) aliquot of ¹²⁵I was added to the recombinant VEGF. Next, 50 µl of Chloramine T at a concentration of 5 mg/10 ml was added and the mixture allowed to react for 60 s. To halt the reaction, 50 µl of sodium metabisulphite (5 mg/ml) was added. After iodination, the labeled protein was separated on a Sep-pak C18 column (Millipore, Milford, MA).

***In vitro* autoradiography.** Frozen 20-µm sections were cut on a cryostat and placed onto gelatin-coated slides. Tissue sections were washed twice for 7.5 min in preincubation buffer containing 10 mmol/l HEPES, 100 mmol/l NaCl, 5 mmol/l KCl, 5 mmol/l MgCl₂, 1 mmol/l EGTA, 0.1% bovine serum albumin, and 0.5 mg/ml bacitracin. Tissue sections were then incubated for 3 h in incubation buffer, consisting of preincubation buffer and ¹²⁵I-VEGF tracer. The final amount of radioactivity in the incubation buffer was ~300 cpm/µl; a total of 2.4 × 10⁶ cpm was used with a final concentration of ~2 pmol/l ¹²⁵I-VEGF. Tissue sections were washed first in preincubation buffer for 4 min, followed by two 4-min washes in postincubation buffer consisting of 10 mmol/l HEPES, 100 mmol/l NaCl, 5 mmol/l KCl, 5 mmol/l MgCl₂, 1 mmol/l EGTA, and 0.5 mg/ml bacitracin. Once sections were dry, they were placed against X-ray film (Kodak X-o-Mat; Kodak, Rochester, NY) for 2 weeks. Competitive binding studies were performed with varying concentrations (10⁻¹² to 10⁻⁹ mol/l) of unlabeled VEGF. Nonspecific binding was determined by addition of 1 mol/l of unlabeled VEGF. Radioactive standards were made by using known dilutions of ¹²⁵I-VEGF onto standard disks of rat kidney slices, which were included in each slide cassette to allow calculation of autoradiographic density into units of radioactivity, as previously described (22). Quantitation of binding was performed using a computerized imaging system (MCID; Imaging Research), and one kidney from five rats per group was examined. By means of this imaging system, the density of radioactive VEGF that binds to each kidney section can be calculated. In view of the focal nature of ¹²⁵I-detected rat VEGF glomerular binding, the mean of 20–25 quantifications of these areas was assessed for each kidney section (22). At least five sections per rat were analyzed. The data were then analyzed by SIGMAPLOT (Jandel Scientific, San Rafael, CA) to yield a value for the concentration of unlabeled ligand required to displace 50% of the radioligand ¹²⁵I-VEGF (IC₅₀) (22).

***In vivo* autoradiography.** To further define cellular localization of ¹²⁵I-VEGF binding in the renal cortex, the tracer was infused *in vivo* in separate groups of rats (control, *n* = 6; diabetic, *n* = 5). This approach has been used when characterizing other binding sites, since it is associated with better preservation of renal morphology (25,26). Rats were anesthetized with pentobarbital sodium (Nembutal 50 mg/kg body weight) and the abdominal cavity was opened with a midline incision. For infusion of ¹²⁵I-VEGF, a 22-gauge catheter was inserted into the abdominal aorta below the renal arteries. For total binding studies, rats were infused with ~15 µCi ¹²⁵I-VEGF in 1 ml of 0.1 mol/l PBS at pH 7.4. After 10–15 min of circulation, the rats were perfused at systolic blood pressure (~140 mmHg) with PBS for 1 min followed by 2% glutaraldehyde for 2 min. After perfusion fixation, one kidney was removed and fixed in 10% neutral buffered formalin overnight before it was embedded in paraffin and sections pre-

TABLE 1
Metabolic parameters

	Short-term (3 weeks)		Long-term (32 weeks)	
	Control	Diabetic	Control	Diabetic
<i>n</i>	10	6	10	8
Glucose (mmol/l)	7.9 ± 0.4	17.7 ± 1.2*	8.2 ± 0.4	19.8 ± 1.5*
Blood pressure (mmHg)	129 ± 4	122 ± 8	135 ± 3	137 ± 2
Body weight (g)				
Week 0	232 ± 5	228 ± 8	230 ± 4	220 ± 6
End of study	316 ± 12	220 ± 6*	495 ± 16	344 ± 9*

Data are means ± SE. **P* < 0.01 vs. control.

pared for emulsion autoradiography. Nonspecific binding (NSB) was determined in separate animals by infusion of excess (10^{-9} mol/l) unlabeled VEGF proteins 3 min before infusion of 125 I-VEGF tracer. The procedure was then followed as described above. The other kidney was frozen in liquid nitrogen-cooled isopentane, and 20- μ m sections were cut on a cryostat and then dried and placed against X-ray film. Between 8 and 13 sections per animal were quantitated using the MCID imaging system as described above.

Statistical analysis. All data were compared by analysis of variance using the statistical package Statview SE+Graphics on an Apple Macintosh computer (Apple Computer, Cupertino, CA) and are shown as means ± SE. Differences between groups were assessed using Fisher's least significant differences test (27).

RESULTS

At the end of both short-term (3 weeks) and long-term (32 weeks) studies, all streptozotocin-induced diabetic animals had hyperglycemia (plasma glucose >15 mmol/l) and decreased body weight compared with the control groups

(Table 1). In the short-term diabetic group, no weight gain was observed over the 3-week period. No difference in blood pressure was observed between control and diabetic groups. **Northern analysis.** Representative Northern hybridization blots for both VEGF and VEGFR-2 in the short-term and long-term studies are shown in Fig. 1. By Northern analysis, short-term diabetes was associated with a threefold increase in VEGF mRNA when compared with control rats (Fig. 2A). There was also a twofold increase in VEGFR-2 gene expression with short-term diabetes (Fig. 2A). Long-term diabetes was associated with a twofold increase in VEGF mRNA (Fig. 2B). VEGFR-2 mRNA levels were not different between con-

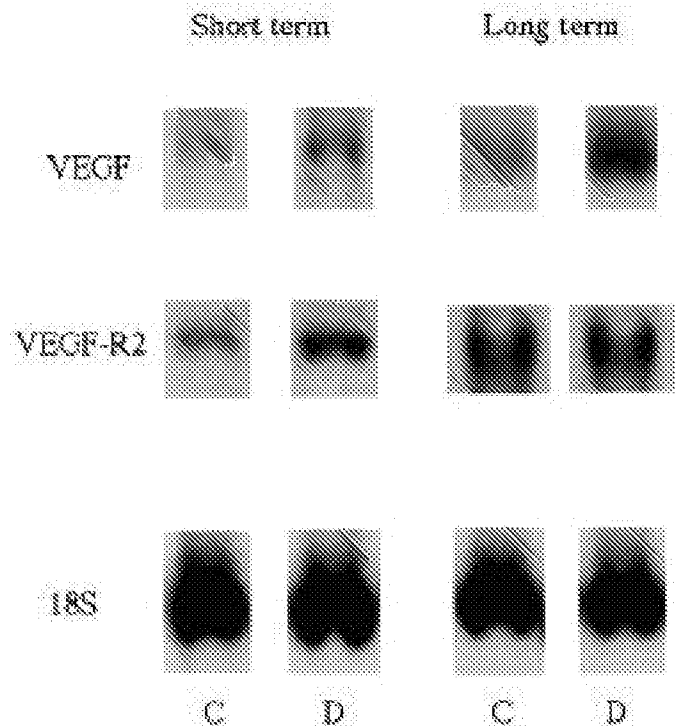


FIG. 1. Northern blot of VEGF, VEGFR-2, and 18S in control (C) and diabetic (D) rat kidneys. Increased gene expression of VEGF and VEGFR-2 is seen in kidneys of short-term diabetic rats. Increased gene expression of VEGF but not VEGFR-2 is seen in kidneys of long-term diabetic rats. 18S rRNA is similar in control and diabetic groups.

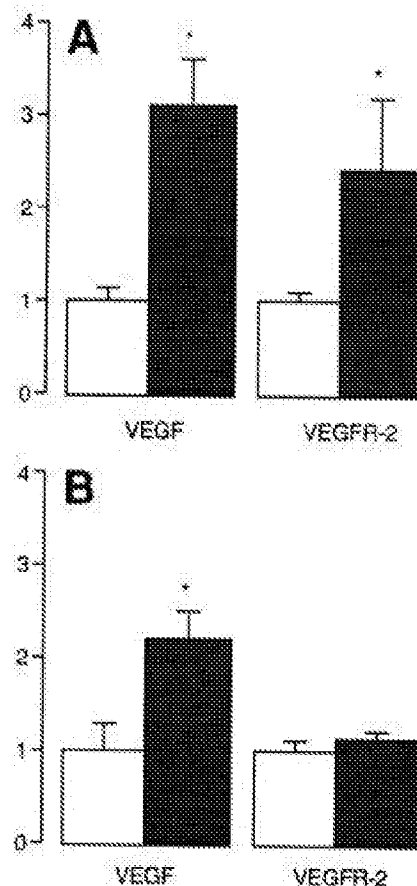


FIG. 2. Quantitation of kidney VEGF and VEGFR-2 mRNA in control (□) and diabetic (■) rats from short-term (A) and long-term (B) studies. Data are means ± SE of the ratio of optical density of VEGF and VEGFR-2 mRNA to that of 18S rRNA relative to control values (designated an arbitrary value of 1). **P* < 0.01; diabetic versus control.

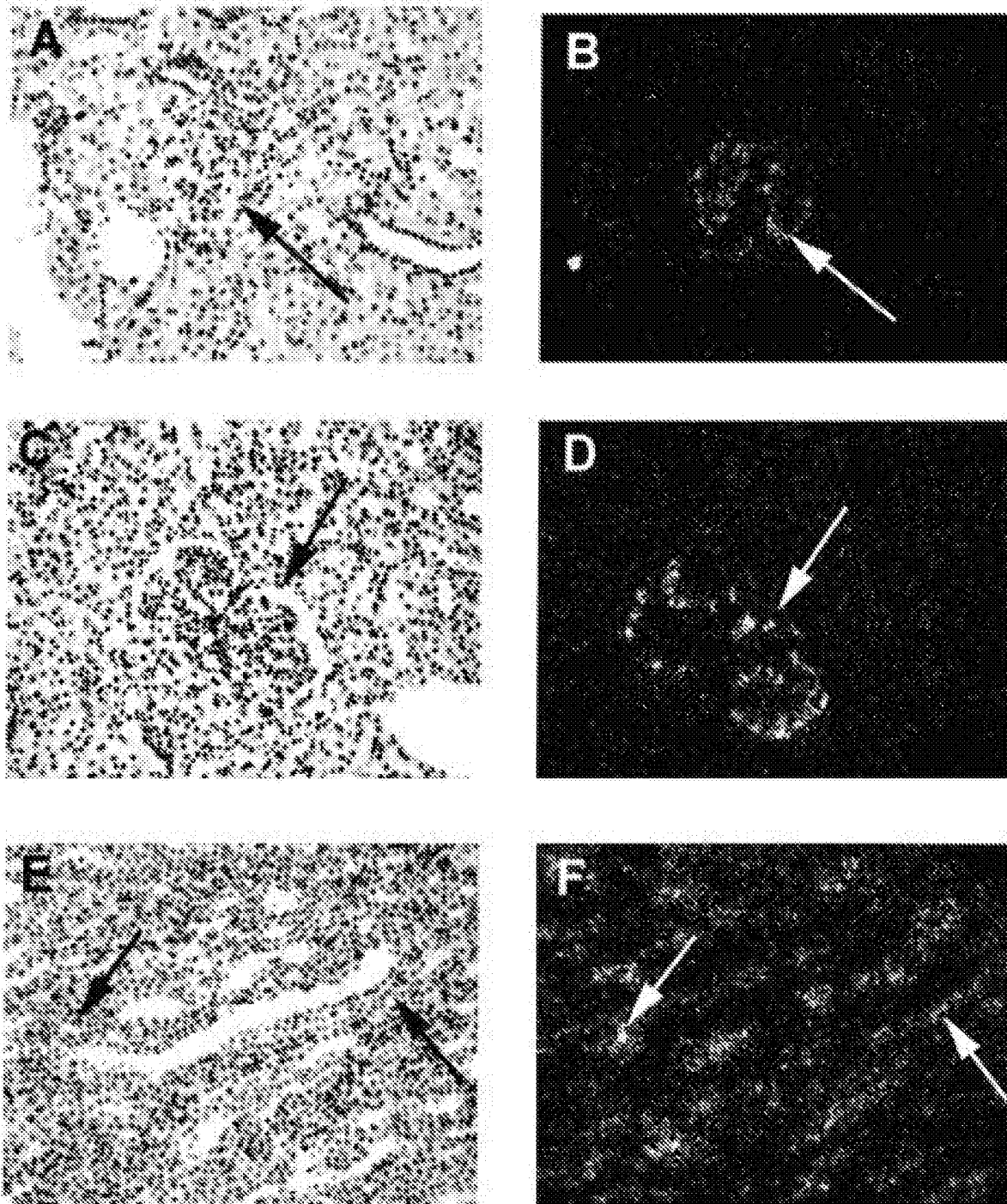


FIG. 3. In situ hybridization photomicrographs of VEGF mRNA. In the cortex, hybridization is present in the glomerular epithelial cells (arrow) in control (A: light field; B: dark field) and in greater abundance in diabetic (C: light field; D: dark field) rat kidneys. In both control and diabetic kidneys, collecting ducts expressed VEGF (control rats, E: light field; F: dark field). Original magnification 200 \times .

trol and diabetic rats (Fig. 2B). By contrast, there was no evidence of hybridization to a VEGFR-1 riboprobe in either control or diabetic rat kidney.

Localization of VEGF and VEGFR-2. Light microscopy following in situ hybridization of sections revealed abundant expression of VEGF mRNA in the visceral epithelial cells of glomeruli (Fig. 3A–D) and in distal tubules and collecting ducts (Fig. 3E and F). Gene expression for the VEGF recep-

tor, VEGFR-2, was detected in glomerular endothelial cells (Fig. 4A–D). In addition, VEGFR-2 mRNA was noted in cortical interstitial fibroblasts and endothelial cells of peritubular capillaries (Fig. 4E) and renomedullary interstitial cells (Fig. 4F). No difference in the pattern of distribution between control and diabetic rats was noted for either VEGF (Fig. 3A–D) or VEGFR-2 mRNA (Fig. 4A–D). No hybridization was observed with sense probes (Fig. 5).

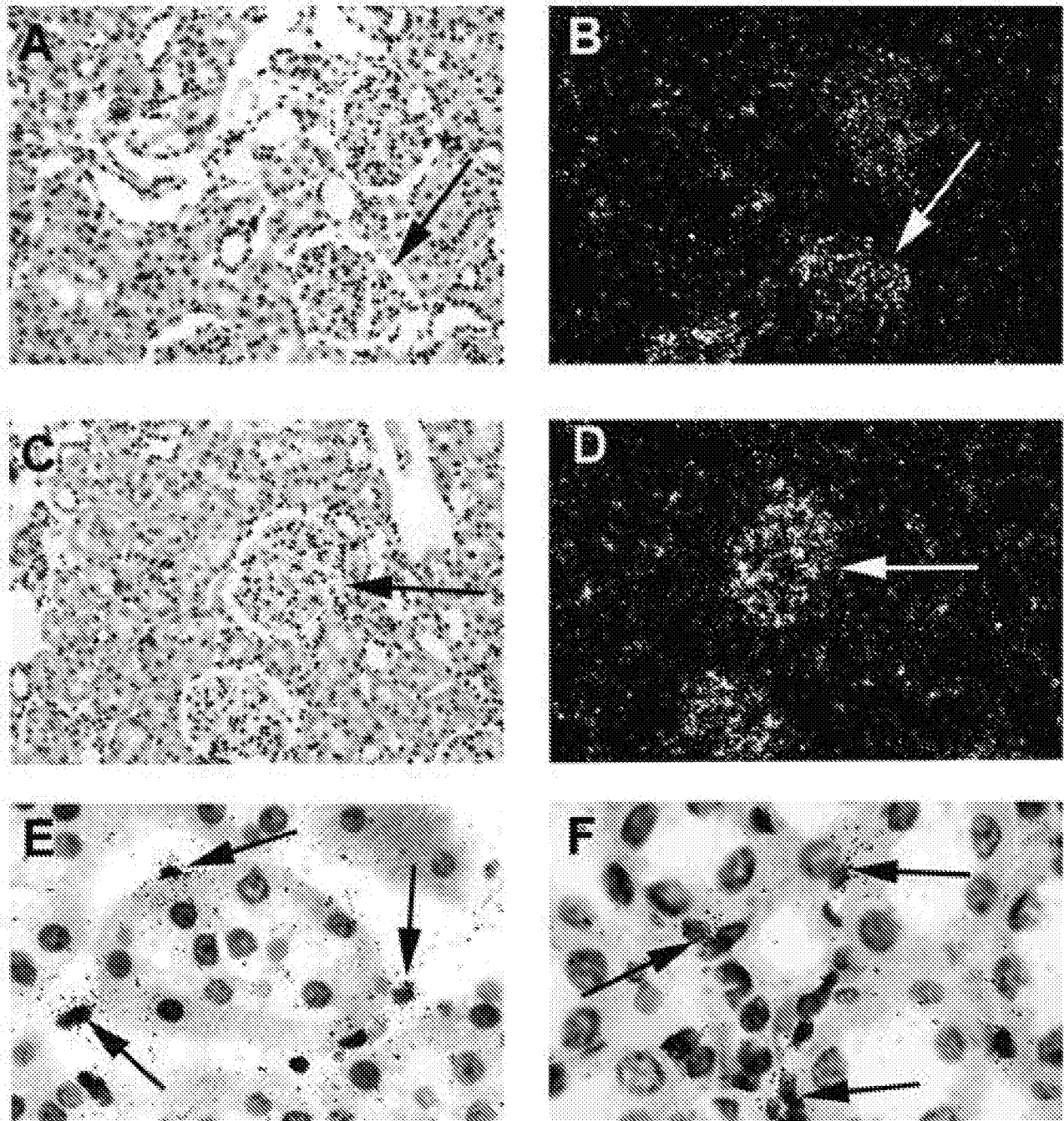


FIG. 4. In situ hybridization photomicrographs of VEGFR-2 mRNA. In the cortex, hybridization is present in the glomerular endothelial cells in control (A: light field; B: dark field) and in greater abundance in diabetic (C: light field; D: dark field) rat kidneys. Original magnification 200 \times . In both control and diabetic kidneys, VEGFR-2 was expressed in cortical fibroblasts, in endothelial cells of peritubular capillaries (E), and in renomedullary interstitial cells (F). Original magnification 1,000 \times .

Quantitation of in situ hybridization autoradiography.

Kidney sections labeled with the VEGF probe showed that hybridization within glomeruli was more intense in the short-term diabetic group compared with the age-matched nondiabetic group (control, $n = 10$; 0.213 ± 0.006 vs. diabetic, $n = 6$; 0.314 ± 0.006 , ROD units, mean \pm SE, $P = 0.002$).

Quantitation of glomerular VEGF mRNA expression in long-term diabetic rats showed similar results to that observed in the short-term diabetic animals. Glomerular VEGF mRNA expression in the diabetic group was increased

when compared with the nondiabetic group (control, $n = 5$; 0.247 ± 0.016 vs. diabetic, $n = 5$; 0.314 ± 0.020 ROD, $P = 0.03$).

Immunohistochemistry. VEGF protein was detected in glomerular epithelial cells, distal tubules, and collecting ducts (Fig. 6A). A similar distribution of VEGF protein was observed in diabetic rats, although the immunostaining appeared to be increased at these sites (Fig. 6B). There was only minimal VEGFR-2 protein detected in glomerular endothelial cells in control rats (Fig. 7A). Immunostaining of glomerular endothelial cells was increased in short-term diabetic rats (Fig. 7B).

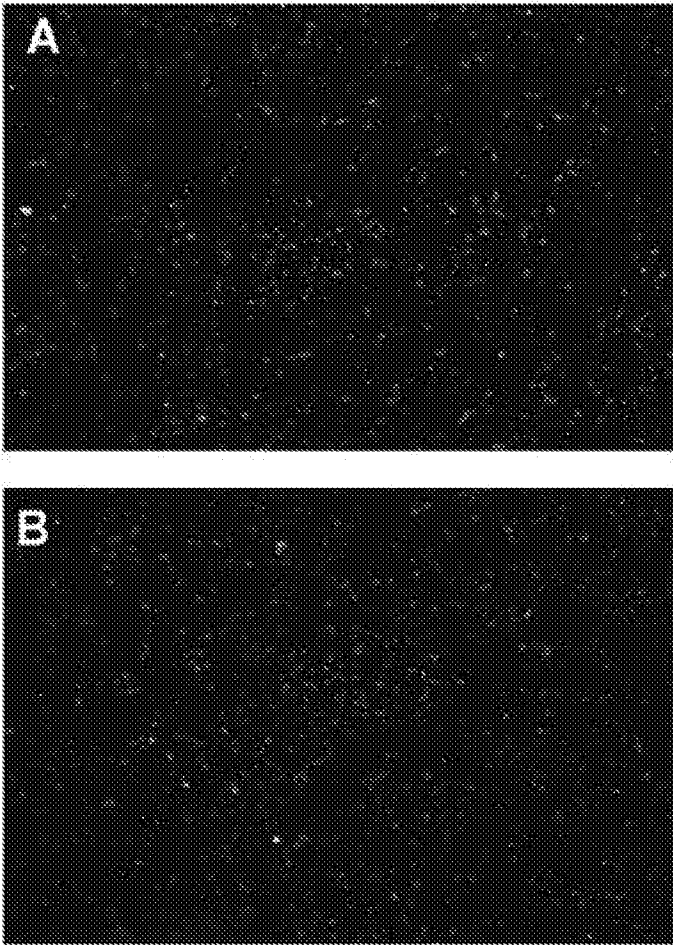


FIG. 5. In situ hybridization photomicrographs of sense VEGF (A) and VEGFR-2 (B) mRNA. No specific hybridization was detected in sections exposed to the VEGF and VEGFR-2 sense probes. Original magnification 300 \times .

¹²⁵I-VEGF binding in the diabetic kidney

In vitro studies. Maximal receptor binding of iodinated VEGF-165 was seen after 5 h at a concentration of ~ 2 pmol/l (specific activity of 2,000 Ci/nmol). Binding of ¹²⁵I-VEGF was visualized by X-ray film in both control and diabetic rat kidney sections. In both control and diabetic rats, ¹²⁵I-VEGF binding was observed in the renal cortex, medulla, and papilla (Fig. 8).

Total binding of VEGF in the diabetic rats was increased by $\sim 50\%$ after 3 weeks of diabetes. Competition studies between radiolabeled VEGF and purified unlabeled VEGF were performed. ¹²⁵I-VEGF binding was inhibited by 10^{-9} mol/l unlabeled VEGF (i.e., NSB). NSB was $<10\%$ in all experiments (Fig. 8).

Cortex: short-term diabetes. Both control and diabetic kidneys showed similar ¹²⁵I-VEGF binding characteristics (Fig. 9A) ($-\log IC_{50}$: control, 11.14 ± 0.14 mol/l vs. diabetic, 11.21 ± 0.03 mol/l, $n = 4$ /group). Renal cortical binding was increased in diabetes (control, 484 ± 11 dpm/mm²; diabetic, 687 ± 18 dpm/mm², $P = 0.001$, $n = 4$ /group).

Medulla. Medullary binding was also increased in the diabetic kidney (control, 368 ± 10 dpm/mm² vs. diabetic, 669 ± 18 dpm/mm², $P < 0.001$, $n = 4$ /group). The binding curve,

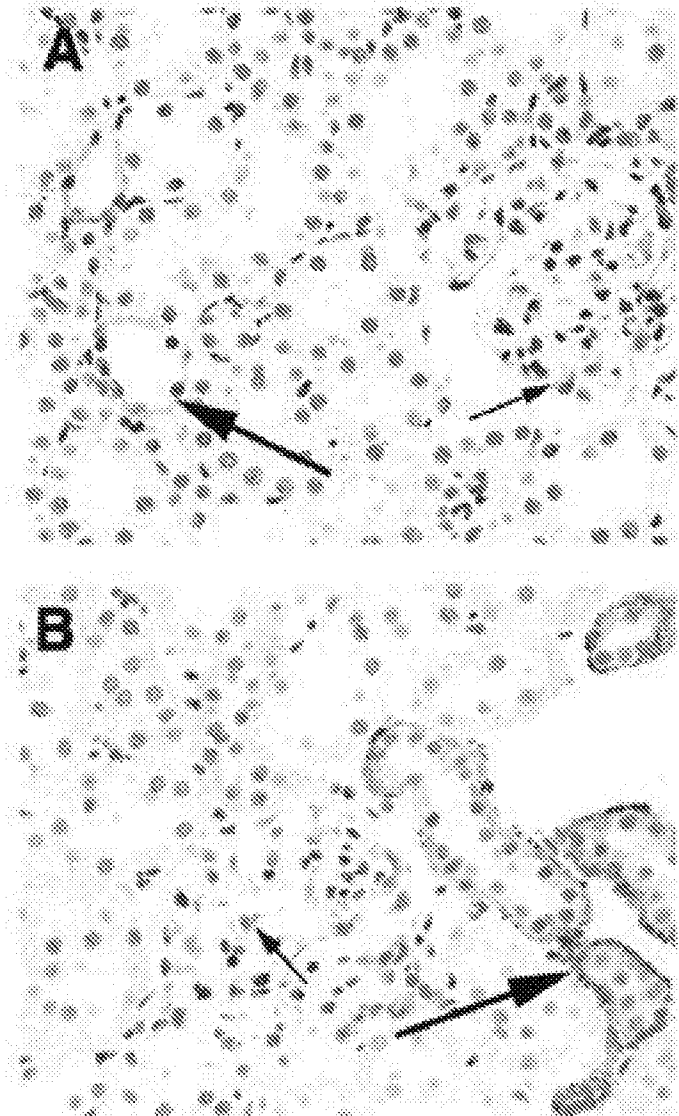


FIG. 6. Paraffin sections (4- μ m thick) of control (A) and diabetic (B) rat kidney were immunostained for VEGF and counterstained with hematoxylin. Original magnification 320 \times . The arrows identify immunolabeling to distal tubules and glomerular epithelial cells. There appeared to be increased immunostaining in the diabetic kidney.

shown in Fig. 9B, was analyzed, and it showed that binding characteristics were modestly, yet statistically, different between the control and diabetic kidneys (control, $-\log IC_{50}$: 10.85 ± 0.16 mol/l; diabetic, 11.16 ± 0.08 mol/l, $P < 0.001$, $n = 4$ /group).

In vivo binding studies. Localization of VEGF binding sites is shown in Fig. 10. There was increased glomerular binding in diabetic rats in comparison with control rats (control, $2,543 \pm 53$, $n = 6$ vs. $4,555 \pm 134$ dpm/mm², $n = 5$, $P < 0.001$).

DISCUSSION

The present study has documented increased expression of VEGF within the kidney after induction of diabetes. This increase persisted over the 8 months of diabetes. By means of in situ hybridization and immunohistochemical techniques, VEGF gene expression was localized to the epithelial cells of the glomerulus. This localization of VEGF is as pre-

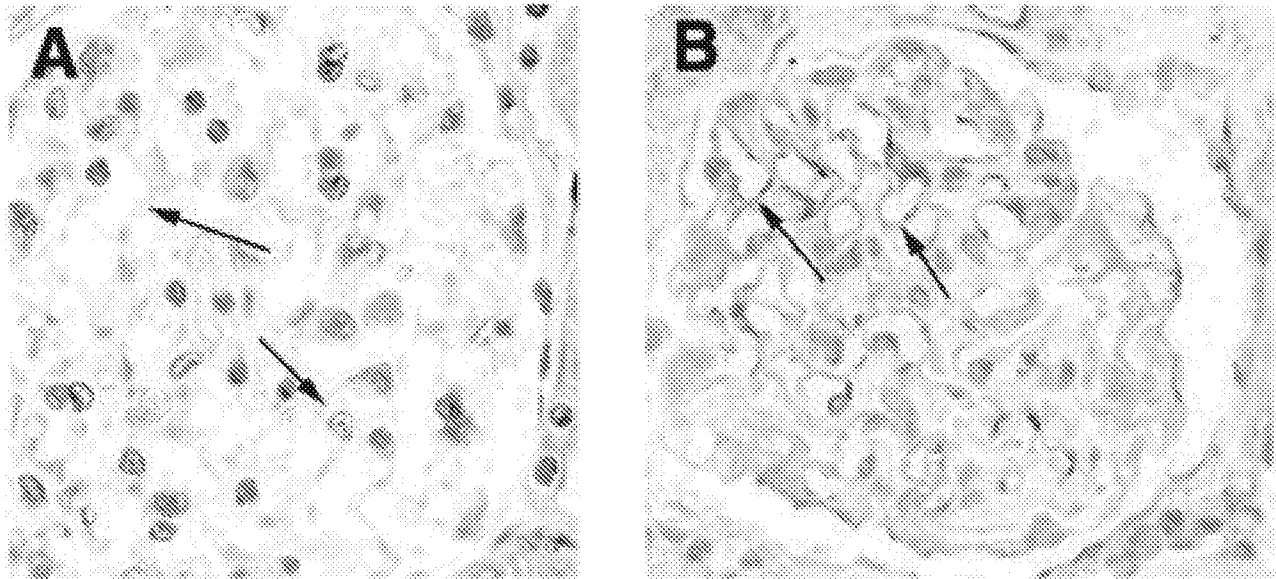


FIG. 7. Paraffin sections (4- μ m thick) of control (A) and diabetic (B) rat kidney were immunostained for VEGFR-2 and counterstained with hematoxylin. Original magnification 380 \times . The arrows identify immunolabeling to glomerular endothelial cells. There is increased immunostaining in the diabetic kidney.

viously described by other groups and is similar to that reported in human kidney (9). In addition, VEGF was detected in collecting ducts, as has been previously reported (28). Although diabetes was associated with increased VEGF gene expression, as assessed by both Northern analysis and by quantitative evaluation of the *in situ* hybridization studies, there was no difference in the spatial distribution of glomerular VEGF between control and diabetic rats.

There was also increased gene expression of VEGFR-2 in the kidneys from animals with short-term diabetes. Furthermore, in long-term diabetic rats, VEGFR-2 expression was also noted in the interstitium, including both fibroblasts and peritubular capillaries. This receptor has been shown to mediate the full biological spectrum of actions of VEGF (1). This increase in receptor gene expression was readily observed in short-term diabetes but was not evident in long-term diabetic rats. Although this difference in renal receptor expression between short- and long-term diabetic rats may relate to effects of duration of diabetes, one cannot exclude that other factors, such as the severity of diabetes or low-dose insulin treatment, may also influence VEGF receptor expression.

In situ hybridization studies revealed that VEGFR-2 was expressed in glomeruli, as has been previously reported (28). The endothelial distribution of this VEGF receptor is consistent with the specific mitogenic effect of VEGF on endothelial cells (29). Immunohistochemical studies confirmed that there was translation of the VEGFR-2 mRNA to protein with evidence of receptor protein expression in glomerular endothelial cells, as has been previously reported (9). In the present study, VEGFR-2 was also detected in both control and diabetic rats in cortical fibroblasts and renomedullary interstitial cells. The significance of this finding is as yet unknown, but these cells lie in close proximity to tubular capillaries, and the action of VEGF at these sites may involve effects on vascular integrity. These interstitial cells have been reported to have receptors for many growth

factors (30) and vasoactive hormones, such as angiotensin II and endothelin (31).

To complement the studies evaluating VEGFR-2 expression, both *in vivo* and *in vitro* autoradiographic techniques were used to assess 125 I-VEGF binding in control and diabetic rats. Specific VEGF binding sites were observed in the kidney, including the glomerulus, medullary rays, and papillae of the kidney, as has been previously described by Jakeman et al. (8). Further evaluation of 125 I-VEGF binding revealed an increase in specific binding in experimental diabetes consistent with translation of the VEGF receptor mRNA to receptor protein, thereby leading to increased binding of the ligand VEGF. The increase in VEGF binding suggested by *in vitro* autoradiography is consistent with the increased expression of VEGFR-2 detected in the diabetic kidney using both *in situ* hybridization and immunohistochemical techniques. These findings using three different methods support the notion that there is upregulation of VEGF receptors in the diabetic kidney.

In vivo autoradiography revealed a similar pattern to the *in vitro* studies with increased binding, primarily to glomeruli, in diabetes. Although the findings from the *in vivo* autoradiographic studies could be confounded by renal hemodynamic changes such as diabetes-related renal hyperperfusion-hyperfiltration, this is unlikely to explain the increased renal VEGF binding, since the *in vitro* autoradiographic studies, which are not confounded by systemic or renal hemodynamic factors, also showed increased binding of the radioligand 125 I-VEGF.

Although VEGFR-1 can also bind VEGF with high affinity, recent studies indicate that this protein does not initiate VEGF signal transduction pathways and is not primarily responsible for the biological actions of VEGF in the adult (5,32). This receptor has been previously reported to be present in the kidney (9). However, other groups have suggested that there is minimal or no expression of VEGFR-1 in the kidney (33–35). Indeed, in the present study, VEGFR-1 mRNA

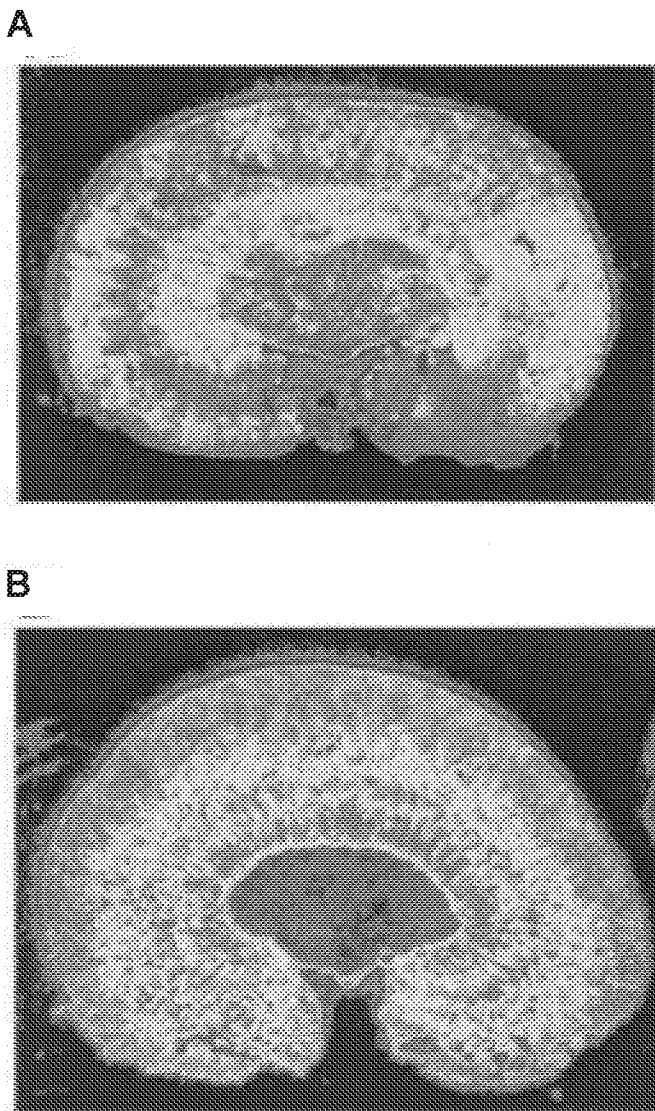


FIG. 8. Magnitude of ¹²⁵I-VEGF binding in the kidney from control (A) and diabetic (B) rats is indicated semiquantitatively in pseudocolored computer images (blue, nil; green, low; yellow, moderate; red, high).

could not be detected by Northern analysis in either control or diabetic rat kidney. By contrast, abundant VEGFR-1 mRNA expression was observed in rat lung and placenta (data not shown) and in retina, as has been previously reported by our group and others (12,13). It has been suggested that the dominant form of this receptor in the kidney is the soluble form that lacks an intracellular signaling domain (33). Nevertheless, one cannot totally exclude a contribution from this high-affinity receptor in explaining the increase in VEGF binding in the diabetic kidney.

The increase in VEGF in the diabetic kidney is consistent with *in vitro* studies suggesting that glucose can induce VEGF expression. Williams et al. (36) have reported that acute hyperglycemia induces VEGF gene expression in vascular smooth-muscle cells, via a protein kinase C (PKC)-dependent pathway. A link between PKC and VEGF is further suggested by studies with an orally active PKC β inhibitor (37). PKC β _{II} inhibition was associated not only with reduced VEGF-induced retinal permeability (37) but also with retar-

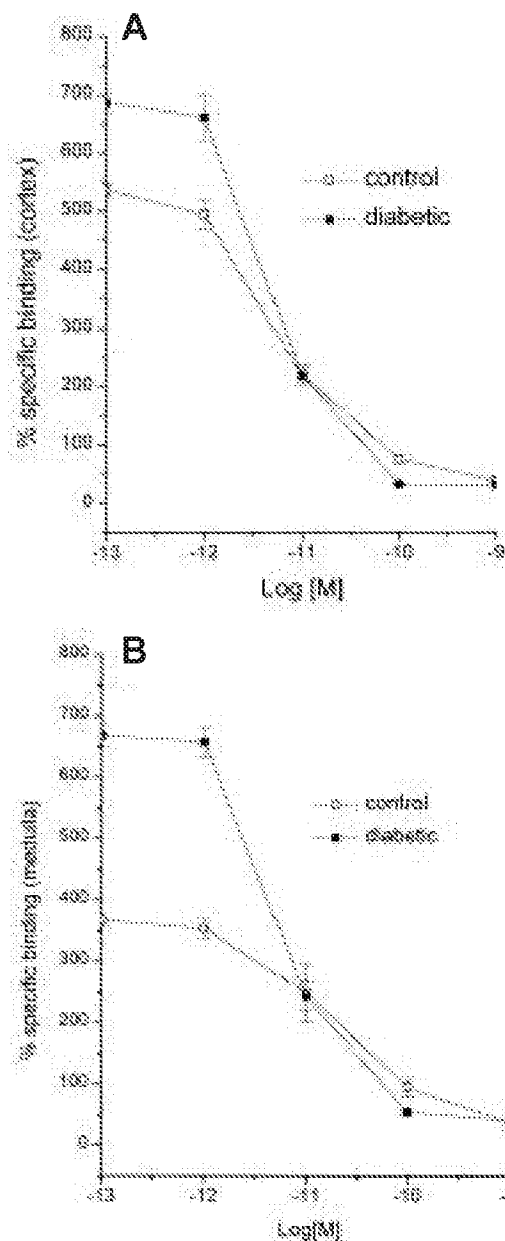


FIG. 9. Binding isotherms for VEGF generated by the incubation of tissue sections in [¹²⁵I]VEGF with increasing concentrations (10⁻¹² to 10⁻⁹ mol/l) of nonradioactive rat VEGF in control (□, n = 5) and diabetic (■, n = 5) rats in renal cortex (A) and medulla (B). The y-axis represents percent specific binding.

dation of development of albuminuria in streptozotocin-induced diabetic rats (38).

The persistent expression of VEGF in the diabetic kidney may relate not only to the effects of hyperglycemia per se but to the effects of advanced glycated end products (AGEs) that accumulate in diabetic tissues over weeks to months (39). These AGEs have been shown *in vitro* to activate VEGF expression in retinal Müller cells (40) and *in vivo* to upregulate VEGF expression in the retina, including cells within the ganglion and inner nuclear layers (41). Various other factors relevant to the pathogenesis of diabetic nephropathy have also been shown to promote VEGF expression, including stretch (42), angiotensin II (43), and a number of

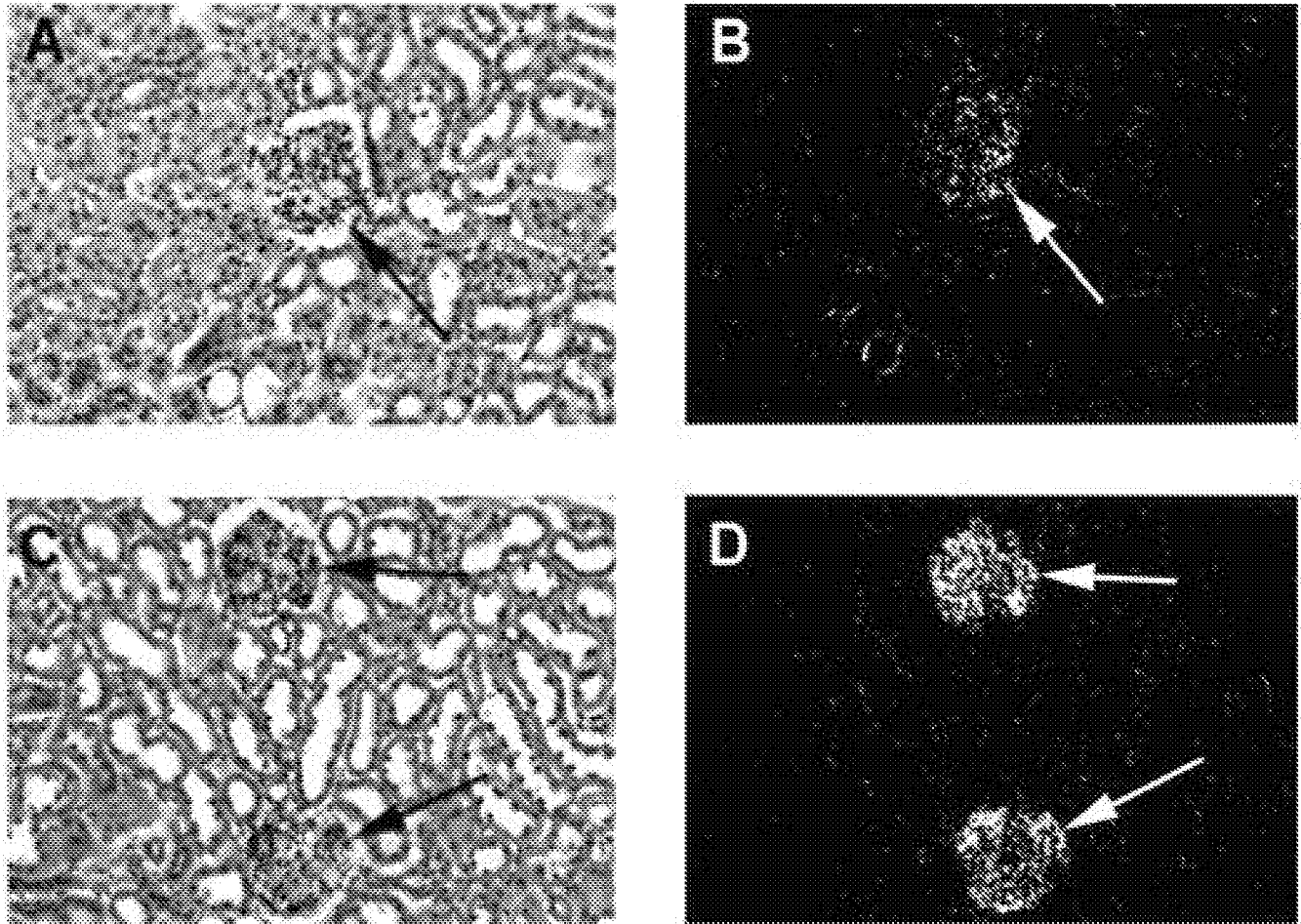


FIG. 10. In vivo binding of ^{125}I -VEGF in the kidney. Light microscopic autoradiographs of kidney cortex from rats administered ^{125}I -VEGF in vivo. Total binding for ^{125}I -VEGF in glomeruli in control (A: light field; B: dark field) and diabetic (C: light field; D: dark field) rats. Magnification 200 \times .

cytokines (44-46). Since it is likely that diabetic nephropathy occurs as a result of an interaction of metabolic factors and hemodynamic factors, it is of particular interest that VEGF expression may be particularly enhanced in the context of angiotensin II and hyperglycemia (47). Transforming growth factor (TGF)- β 1 has been reported to downregulate VEGFR-2 expression (48) but to increase gene and protein expression of its ligand, VEGF (49). Since TGF- β 1 has been reported to increase in the kidneys from long-term diabetic rats (50,51), it is possible that this cytokine was involved in reducing renal VEGFR-2 overexpression and stimulating VEGF expression in long-term diabetic rats, as was observed in the present study.

The increase in VEGF and VEGFR-2 has been previously reported in the retina in experimental diabetes by several groups (12,13). Hammes et al. (12) reported that there is minimal evidence of expression of VEGF or its receptors in the normal retina with clear evidence of significant synthesis of VEGF and its receptors in the diabetic retina. This was confirmed by Gilbert et al. (13), who also reported that in experimental diabetes there were increased levels of VEGF and VEGFR-2 in the retina. The present study in the kidney provides evidence that the changes observed in the retina are not restricted to one site but also occur in the kidney. From a clinical perspective, a close association has been noted between

vision-threatening retinopathy and diabetic renal disease (14,52). This association may be considered to be due to similar pathogenic mechanisms. VEGF must be viewed as a primary candidate for mediating both microvascular complications. The presence of increased ligand as well as increased receptor is suggestive of activation of the VEGF pathway in diabetes. Although it is likely that VEGF plays an important role in ocular neovascularization, its role in the kidney remains to be fully delineated (15). It is clear that VEGF and its receptors are vital for renal development, based on studies in VEGF and VEGF receptor knockout mice (53,54) and on studies involving postnatal administration of neutralizing antibodies to VEGF (55). The localization of VEGF to the glomerular epithelial cell (podocyte) in the current study is similar to the findings of others (56). It has been suggested that VEGF may contribute to the relaxing capacity of the renal vasculature so as to alleviate vascular injury and re-establish vascular integrity (56,57). The importance of this phenomenon in diabetes remains speculative, and, from the present study, it is not possible to determine if overexpression of VEGF in the kidney is deleterious or represents a compensatory mechanism to attenuate renal injury.

The major biological actions of VEGF appear to be the enhancement of vascular permeability and angiogenesis (1). In the retina, these two actions of VEGF are considered rel-

evant to the development of macular edema and proliferative retinopathy. By contrast, it remains uncertain whether these biological actions of VEGF are relevant to the changes observed in the diabetic kidney. Diabetes is associated with an early increase in albumin permeability, which ultimately leads to the development of albuminuria (58). VEGF is a very potent inducer of permeability (59), and it is possible that this action of VEGF is enhanced in diabetes due to increased synthesis of this peptide within the glomerulus. The importance of VEGF in mediating this effect in diabetes has been suggested by studies in which glucose-induced changes in albumin permeation and blood flow could be prevented by neutralizing antibodies to VEGF (60) or a nonpeptide VEGF antagonist (61).

With respect to VEGF's action as an endothelial mitogen-promoting angiogenesis, the status of the endothelial cell in the diabetic kidney and whether there is associated angiogenesis remains controversial. Østerby et al. (62) have suggested that angiogenesis may occur in human diabetes with duplication of the efferent arteriole. Several other groups have also reported increased blood vessel growth in the diabetic kidney (63–65), but the link of these phenomena to VEGF and VEGF receptor expression remains to be delineated. Indeed, a specific antagonist of VEGF will ultimately be required to determine if VEGF is involved in mediating many of the functional and possibly structural changes observed in diabetic nephropathy.

ACKNOWLEDGMENTS

This work was supported by grants from the Juvenile Diabetes Foundation International and the National Health and Medical Research Council of Australia. R.E.G. is the recipient of a Career Development Award from the Juvenile Diabetes Foundation International.

REFERENCES

- Ferrara N, Davissmyth T: The biology of vascular endothelial growth factor. *Endocrine Rev* 18:4–25, 1997
- Quinn TP, Peters KG, De Vries C, Ferrara N, Williams LT: Fetal liver kinase 1 is a receptor for vascular endothelial growth factor and is selectively expressed in vascular endothelium. *Proc Natl Acad Sci U S A* 90:7533–7537, 1993
- Oelrichs RB, Reid HH, Bernard O, Ziemiecki A, Wilks AF: NYK/FLK-1: a putative receptor protein tyrosine kinase isolated from E10 embryonic neuroepithelium is expressed in endothelial cells of the developing embryo. *Oncogene* 8:11–18, 1993
- Neufeld G, Cohen T, Gengrinovitch S, Poltorak Z: Vascular endothelial growth factor (VEGF) and its receptors. *FASEB J* 13:9–22, 1999
- Waltenberger J, Claesson-Welsh L, Sieghart A, Shibuya M, Heldin CH: Different signal transduction properties of KDR and Flt1, two receptors for vascular endothelial growth factor. *J Biol Chem* 269:26988–26995, 1994
- Monacci WT, Merrill MJ, Oldfield EH: Expression of vascular permeability factor/vascular endothelial growth factor in normal rat tissues. *Am J Physiol* 264:995–1002, 1993
- Brenchley PE: VEGF/VFP: a modulator of microvascular function with potential roles in glomerular pathophysiology. *J Nephrol* 9:10–17, 1996
- Jakeman LB, Winer J, Bennett GL, Altar CA, Ferrara N: Binding sites for vascular endothelial growth factor are localized on endothelial cells in adult rat tissues. *J Clin Invest* 89:244–253, 1992
- Simon M, Rockl W, Hornig C, Grone EF, Theis H, Weich HA, Fuchs E, Yayon A, Grone HJ: Receptors of vascular endothelial growth factor/vascular permeability factor (VEGF/VFP) in fetal and adult human kidney: localization and [1-125] VEGF binding sites. *J Am Soc Nephrol* 9:1032–1044, 1998
- Aiello LP, Avery RL, Arrigg PG, Keyt BA, Jampel HD, Shah ST, Pasquale LR, Thieme H, Iwanoto MA, Park JE, Nguyen HV, Aiello LM, Ferrara N, King GL: Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. *N Engl J Med* 331:1480–1487, 1994
- Adamis AP, Miller JW, Bernal MT, Damico DJ, Folkman J, Yeo TK, Yeo KT: Increased vascular endothelial growth factor levels in the vitreous of eyes with proliferative diabetic retinopathy. *Am J Ophthalmol* 118:445–450, 1994
- Hammes H-P, Lin J, Bretzel RG, Brownlee M, Breier G: Upregulation of the vascular endothelial growth factor/vascular endothelial growth factor receptor system in experimental background diabetic retinopathy of the rat. *Diabetes* 47:401–406, 1998
- Gilbert RE, Vranes D, Berka JL, Kelly DJ, Cox A, Wu LL, Stacker SA, Cooper ME: Vascular endothelial growth factor and its receptors in control and diabetic rat eyes. *Lab Invest* 78:1017–1027, 1998
- Gilbert RE, Tsalamandris C, Allen TJ, Colville D, Jerums G: Early nephropathy predicts vision-threatening retinal disease in patients with Type I diabetes mellitus. *J Am Soc Nephrol* 9:85–89, 1998
- Del Prete D, Angiani F, Ceol M, D'Angelo A, Forino M, Vianello D, Baggio B, Gambaro G: Molecular biology of diabetic glomerulosclerosis. *Nephrol Dial Transpl* 13 (Suppl. 8):20–25, 1998
- Cooper ME: Pathogenesis, prevention, and treatment of diabetic nephropathy. *Lancet* 352:213–219, 1998
- Schmidt FH: Enzymatic determination of glucose and fructose simultaneously. *Klin Woch* 39:1244–1247, 1961
- Bunag RD: Validation in awake rats of a tail-cuff method for measuring systolic pressure. *J Appl Physiol* 34:279–282, 1973
- Chomczynski P, Sacchi N: Single step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal Biochem* 162:156–159, 1987
- Gilbert RE, Cox A, Wu LL, Allen TJ, Hulthen L, Jerums G, Cooper ME: Expression of transforming growth factor- β 1 and type IV collagen in the renal tubulointerstitium in experimental diabetes: effects of ACE inhibition. *Diabetes* 47:414–422, 1998
- Baskin DG, Stahl WL: Fundamentals of quantitative autoradiography by computer densitometry for in situ hybridisation with emphasis on 33P. *J Histochem Cytochem* 41:1767–1776, 1993
- Wookey PJ, Tikellis C, Du HC, Qin HF, Sexton PM, Cooper ME: Amylin binding in rat renal cortex, stimulation of adenylyl cyclase, and activation of plasma renin. *Am J Physiol* 39:F289–F294, 1996
- Wu LL, Cox A, Roe CJ, Dziadek M, Cooper ME, Gilbert RE: Secreted protein acidic and rich in cysteine expression after subtotal nephrectomy and blockade of the renin-angiotensin system. *J Am Soc Nephrol* 8:1373–1382, 1997
- Greenwood PC, Hunter WM, Glover JS: The preparation of 125 I-labelled human growth hormone of high specific radioactivity. *Biochem J* 89:114–123, 1963
- Dean R, Zhuo JL, Alcorn D, Casley D, Mendelsohn FAO: Cellular localization of endothelin receptor subtypes in the rat kidney following in vitro labelling. *Clin Exp Pharmacol Physiol* 23:524–531, 1996
- Harris PJ, Cooper ME, Hiranyachaitada S, Berka JL, Kelly DJ, Nobes M, Wookey PJ: Anylin stimulates proximal tubular sodium transport and cell proliferation in the rat kidney. *Am J Physiol* 272:F13–F21, 1997
- Snedecor GW, Cochran WG: *Statistical Methods*. 7th ed. Ames, Iowa, Iowa State University Press, 1980, p. 228–236
- Simon M, Grone HJ, Jöhren O, Kullmer J, Plate KH, Risau W, Fuchs E: Expression of vascular endothelial growth factor and its receptors in human renal ontogenesis and in adult kidney. *Am J Physiol* 268:F240–F250, 1995
- Houck KA, Ferrara N, Winer J, Cachianes G, Li B, Leung DW: The vascular endothelial growth factor family: identification of a fourth molecular species and characterization of alternative splicing of RNA. *Mol Endocrinol* 5:1806–1814, 1991
- Seifert RA, Alpers CE, Bowen-Pope DF: Expression of platelet-derived growth factor and its receptors in the developing and adult mouse kidney. *Kidney Int* 54:731–746, 1998
- Zhuo JL, Dean R, Maric C, Aldred PG, Harris P, Alcorn D, Mendelsohn FAO: Localization and interactions of vasoactive peptide receptors in renomedullary interstitial cells of the kidney. *Kidney Int* 54:S22–S28, 1998
- Keyt BA, Nguyen HV, Berleau LT, Duarte CM, Park J, Chen H, Ferrara N: Identification of vascular endothelial growth factor determinants for binding kdr and flt-1 receptors: generation of receptor-selective VEGF variants by site-directed mutagenesis. *J Biol Chem* 271:5638–5646, 1996
- Williams B: A potential role for angiotensin II-induced vascular endothelial growth factor expression in the pathogenesis of diabetic nephropathy? *Miner Electrolyte Metab* 24:400–405, 1998
- Takahashi T, Shirasawa T, Miyake K, Maruyama N, Kasahara N, Kawamura T, Matsumura O, Mitarai T, Sakai O: Protein tyrosine kinases expressed in glomeruli and cultured glomerular cells: Flt-1 and VEGF expression in renal mesangial cells. *Biochem Biophys Res Commun* 209:218–226, 1995
- Kee N, McTavish AJ, Papillon J, Cybulsky AV: Receptor protein tyrosine kinases in perinatal developing rat kidney. *Kidney Int* 52:309–317, 1997

36. Williams B, Gallacher B, Patel H, Orme C: Glucose-induced protein kinase C activation regulates vascular permeability factor mRNA expression and peptide production by human vascular smooth muscle cells in vitro. *Diabetes* 46:1497-1503, 1997
37. Aiello LP, Bursell S-E, Clermont A, Duh E, Ishii H, Takagi C, Mori F, Ciulla TA, Wachs K, Jirousek M, Smith LEH, King GL: Vascular endothelial growth factor-induced retinal permeability is mediated by protein kinase C in vivo and suppressed by an orally effective β -isoform-selective inhibitor. *Diabetes* 46:1473-1480, 1997
38. Ishii H, Jirousek MR, Koya D, Takagi C, Xia P, Clermont A, Bursell S-E, Kern TS, Ballas LM, Heath WF, Stranum LE, Feener EP, King GL: Amelioration of vascular dysfunctions in diabetic rats by an oral PKC beta inhibitor. *Science* 272:728-731, 1996
39. Bucala R, Vlassara H: Advanced glycosylation end products in diabetic renal and vascular disease. *Am J Kidney Dis* 26:875-888, 1995
40. Hirata C, Nakano K, Nakamura N, Kitagawa Y, Shigeta H, Hasegawa G, Ogata M, Ikeda T, Sawa H, Nakamura K, Ienaga K, Obayashi H, Kondo M: Advanced glycation end products induce expression of vascular endothelial growth factor by retinal Muller cells. *Biochem Biophys Res Commun* 236:712-715, 1997
41. Lu M, Kuroki M, Armano S, Tolentino M, Keough K, Kim I, Bucala R, Adamis AP: Advanced glycation end products increase retinal vascular endothelial growth factor expression. *J Clin Invest* 101:1219-1224, 1998
42. Gruden G, Thomas S, Burt D, Lane S, Chusney G, Sacks S, Viberti GC: Mechanical stretch induces vascular permeability factor in human mesangial cells: mechanisms of signal transduction. *Proc Natl Acad Sci U S A* 94:12112-12116, 1997
43. Williams B, Baker AQ, Gallacher B, Lodwick D: Angiotensin II increases vascular permeability factor gene expression by human vascular smooth muscle cells. *Hypertension* 25:913-917, 1995
44. Williams B, Quimbaker A, Gallacher B: Serum and platelet-derived growth factor-induced expression of vascular permeability factor mRNA by human vascular smooth muscle cells in vitro. *Clin Sci* 88:141-147, 1995
45. Stavri GT, Zachary IC, Baskerville PA, Martin JF, Erusalimsky JD: Basic fibroblast growth factor upregulates the expression of vascular endothelial growth factor in vascular smooth muscle cells: synergistic interaction with hypoxia. *Circulation* 92:11-14, 1995
46. Stavri GT, Hong Y, Zachary IC, Breier G, Baskerville PA, Yia HS, Risau W, Martin JF, Erusalimsky JD: Hypoxia and platelet-derived growth factor-BB synergistically upregulate the expression of vascular endothelial growth factor in vascular smooth muscle cells. *FEBS Lett* 358:311-315, 1995
47. Natarajan R, Bai W, Lanting L, Gonzales N, Nadler J: Effects of high glucose on vascular endothelial growth factor expression in vascular smooth muscle cells. *Am J Physiol* 42:H2224-H2231, 1997
48. Mandriota SJ, Menoud PA, Pepper MS: Transforming growth factor β -1 down-regulates vascular endothelial growth factor receptor 2/flk-1 expression in vascular endothelial cells. *J Biol Chem* 271:11500-11505, 1996
49. Pertovaara I, Kaipainen A, Mustonen T, Orpana A, Ferrara N, Saksela O, Alitalo K: Vascular endothelial growth factor is induced in response to transforming growth factor- β in fibroblastic and epithelial cells. *J Biol Chem* 269:6271-6274, 1994
50. Gilbert RE, Wilkinson-Berka JL, Johnson DW, Cox A, Soulis T, Wu LL, Kelly DJ, Jerums G, Pollock CA, Cooper ME: Renal expression of transforming growth factor- β inducible gene-h3 (β -ig-h3) in normal and diabetic rats. *Kidney Int* 54:1052-1062, 1998
51. Yamamoto T, Nakamura T, Noble NA, Ruoslahti E, Border WA: Expression of transforming growth factor beta is elevated in human and experimental diabetic nephropathy. *Proc Natl Acad Sci U S A* 90:1814-1818, 1993
52. Vigstrup J, Mogensen CE: Proliferative diabetic retinopathy: at risk patients identified by early detection of microalbuminuria. *Acta Ophthalmol (Copenh)* 63:530-534, 1985
53. Shalaby F, Rossant J, Yamaguchi TP, Gertszenstein M, Wu XF, Breitman ML, Schuh AC: Failure of blood-island formation and vasculogenesis in flk-1-deficient mice. *Nature* 376:62-66, 1995
54. Ferrara N, Carver-Moore K, Chen H, Dowd M, Lu L, O'Shea KS, Powell-Braxton L, Hillan KJ, Moore MW: Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature* 380:439-442, 1996
55. Kitamoto Y, Tokunaga H, Tomita K: Vascular endothelial growth factor is an essential molecule for mouse kidney development: glomerulogenesis and nephrogenesis. *J Clin Invest* 99:2351-2357, 1997
56. Grone HJ, Simon M, Grone EF: Expression of vascular endothelial growth factor in renal vascular disease and renal allografts. *J Pathol* 177:259-267, 1995
57. Klanke B, Simon M, Rockl W, Weich HA, Stolte H, Grone HJ: Effects of vascular endothelial growth factor (VEGF) vascular permeability factor (VPF) on haemodynamics and permeability of the isolated perfused rat kidney. *Nephrol Dial Transplant* 13:875-885, 1998
58. Parving HH, Kasrup J, Smidt UM: Reduced transcapillary escape of albumin during acute blood pressure-lowering in type I (insulin-dependent) diabetic patients with nephropathy. *Diabetologia* 28:797-801, 1985
59. Roberts WG, Palade GE: Increased microvascular permeability and endothelial fenestration induced by vascular endothelial growth factor. *J Cell Sci* 108:2369-2379, 1995
60. Tilton RG, Kawamura T, Chang KC, Ido Y, Bjorcke RJ, Stephan CC, Brock TA, Williamson JR: Vascular dysfunction induced by elevated glucose levels in rats is mediated by vascular endothelial growth factor. *J Clin Invest* 99:2192-2202, 1997
61. Stephan CC, Chang KC, Lejeune W, Erichsen D, Bjorcke RJ, Rege A, Biediger RJ, Kogan TP, Brock TA, Williamson JR, Tilton RG: Role for heparin-binding growth factors in glucose-induced vascular dysfunction. *Diabetes* 47:1771-1778, 1998
62. Østerby R, Asphund J, Bangstad HJ, Nyberg G, Rudberg S, Viberti G, Walker JD: Neovascularization at the vascular pole region in diabetic glomerulopathy. *Nephrol Dial Transplant* 14:348-352, 1999
63. Nyengaard JR, Rasch R: The impact of experimental diabetes mellitus in rats on glomerular capillary number and sizes. *Diabetologia* 36:180-194, 1993
64. Min W, Yamanaka N: Three-dimensional analysis of increased vasculature around the glomerular vascular pole in diabetic nephropathy. *Virchows Arch A Pathol Anat Histopathol* 423:201-207, 1993
65. Vranes D, Dilley RJ, Cooper ME: Vascular changes in the diabetic kidney: effects of ACE inhibition. *J Diabetes Complications* 9:296-300, 1995

VEGF-mediated inflammation precedes angiogenesis in adult brain

Susan D. Croll,^{a,*} Richard M. Ransohoff,^b Ning Cai,^a Qing Zhang,^a Francis J. Martin,^a
Tao Wei,^b Lora J. Kasselmann,^a Jennifer Kintner,^a Andrew J. Murphy,^a
George D. Yancopoulos,^a and Stanley J. Wiegand^a

^aRegeneron Pharmaceuticals, Tarrytown, NY 10591, USA

^bDepartment of Neurosciences, Cleveland Clinic Foundation, Cleveland, OH 44195, USA

Received 24 November 2003; revised 4 February 2004; accepted 6 February 2004

Available online 28 March 2004

Abstract

Vascular endothelial growth factor (VEGF) has been shown to induce angiogenesis when infused continuously into adult rat brain tissue. In addition, VEGF has been shown to enhance permeability in brain vasculature. Adult rats were continuously infused with mouse VEGF into neocortex for up to 7 days. We studied the development of VEGF-induced vasculature in rat neocortex and evaluated the temporal expression of a wide variety of markers for inflammation and vascular leak in relation to the angiogenic response using immunohistochemistry and Western blot analysis. We report here that VEGF-mediated inflammation in brain is characterized by upregulation of ICAM-1 and the chemokine MIP-1 α , as well as a preferential extravasation of monocytes. VEGF causes a dramatic breakdown of the blood–brain barrier, which is characterized by decreased investment of the vasculature with astroglial endfeet. Perivascular cells, in contrast, increase around the newly formed cerebrovasculature. In addition, breakdown of the blood–brain barrier, leukocyte extravasation, and extracellular matrix deposition occur before vascular proliferation. Furthermore, administration of low doses of VEGF induces permeability and inflammation without appreciable vascular proliferation.

© 2004 Elsevier Inc. All rights reserved.

Keywords: Cytokine; Blood–brain barrier; Permeability; Vasculature; Chemokine; VEGF; Inflammation; MIP-1 α ; Monocytes

Introduction

Vascular endothelial growth factor (VEGF) has been shown to induce vascular proliferation in a variety of tissues (Bauters et al., 1994; Krum et al., 2002; Proescholdt et al., 1999; Rosenstein et al., 1998; Springer et al., 1998; Takeshita et al., 1994), and studies of VEGF null mutant mice have shown that VEGF is necessary for angiogenesis and vasculogenesis in the developing embryo (Ferrara et al., 1996). Consequently, VEGF has been evaluated in clinical trials as a pro-angiogenic factor to treat ischemic conditions (Isner, 1998; Isner et al., 1996; Losordo et al., 1998). Although VEGF's role as an angiogenic factor is well established, it was originally discovered and characterized based on its ability to increase vascular permeability (Senger et al., 1986). VEGF has been shown to increase vascular

leak of both proteins and particulates, and it has been proposed that this increase in permeability is a necessary prerequisite for the induction of angiogenesis (for reviews, see Dvorak et al., 1995, 1999).

Many human diseases are characterized by pathological angiogenesis, such as proliferative vascular retinopathies, psoriasis, rheumatoid arthritis, endometriosis, and tumors. In these conditions, VEGF levels are known to be increased before and/or during the angiogenic process (Bhushan et al., 1999; Brown et al., 1993, 1995; Detmar et al., 1994; Harada et al., 1998; Jackson et al., 1997; Kikuchi et al., 1998; Lashkari et al., 2000; Mahnke et al., 2000). In addition, VEGF levels are elevated in many human diseases, or animal models of human disease, which are characterized by inflammation and vascular leak, such as cerebral ischemia, tumor ascites, trauma, early diabetic retinopathy, pre-eclampsia, ovarian hyperstimulation syndrome, and status epilepticus (Baker et al., 1995; Boulton et al., 1998; Cobbs et al., 1998; Croll et al., 2002; Grad et al., 1998; Hayashi et al., 1997; Issa et al., 1999; Kovacs et al., 1996; Kraft et al.,

* Corresponding author. Regeneron Pharmaceuticals, 777 Old Saw Mill River Road, Tarrytown, NY 10591. Fax: +1-914-347-5045.

E-mail address: susan.croll@regeneron.com (S.D. Croll).

1999; Lee et al., 1999; Lemmyr et al., 1998; Levin et al., 1998; McClure et al., 1994; Neulen et al., 1995; Pichiule et al., 1999; Plate et al., 1999; Slevin et al., 2000; Spirin et al., 1999; Zebrowski et al., 1999). Therefore, a clear understanding of VEGF-mediated angiogenesis and vascular leak is critical if VEGF-related treatments for these pathological conditions are to be evaluated.

We have employed continuous infusion of exogenous VEGF protein into adult rat neocortex as a model for understanding more fully the nature and development of VEGF-induced changes in cerebral blood vessel structure and function. VEGF induces an angiogenic response when infused continuously into adult rat brain via osmotic minipump, or when delivered by way of an adenoviral vector (Proescholdt et al., 1999; Rosenstein et al., 1998). When applied to the brain surface or parenchyma, VEGF induces a rapid and dramatic increase in vascular permeability (Dobrogowska et al., 1998). Our findings show that VEGF acts either directly or indirectly as a potent pro-inflammatory cytokine in adult rat brain, and that increases in vascular permeability and indices of inflammation precede VEGF-induced angiogenesis of brain vasculature.

Materials and methods

Subjects

Adult male Sprague–Dawley rats (300–400 g) from Taconic laboratories (Germantown, NY) were housed two per cage within a temperature and humidity-stabilized animal facility with food and water available ad libitum. Animals were maintained on a 12:12 light–dark cycle (lights on 0600 h) and were allowed to acclimate for at least 1 week before any experimental manipulations. All experiments were conducted under the auspices of protocols approved by the Regeneron Animal Care and Use Committee.

Surgeries

Animals were anesthetized with chloral hydrate (170 mg/kg)–pentobarbital (35.2 mg/kg) ip. Cannulae (2 mm long; Plastics One, Roanoke, VA) were stereotaxically implanted in the right frontoparietal cortex (0.5 mm anterior, 3.2 mm lateral to bregma) as described previously (Croll et al., 1998). Polyvinyl catheters (Bolab, Lake Havasu City, AZ) were attached to the cannulae with cyanoacrylate glue, filled with sterile PBS, and heat-sealed (Loctite 495, Loctite Corp., Newington, CT). One week later (to permit surgical trauma to attenuate), animals were briefly anesthetized with either halothane (5% induction, 1.5% maintenance) in 1:2 oxygen–nitrous oxide or isoflurane (2.5% induction, 2% maintenance) in oxygen. An incision was made in the nape of the neck, the tip of the catheter was cut, and an osmotic minipump (model 2002, 0.5 μ l/h, Alza Corporation, Palo Alto, CA) was attached to the catheter using cyanoacrylate glue.

Minipumps containing VEGF were attached to pre-implanted PBS-filled catheters on day 0. Because the pumps had to first pump PBS from the catheters into the brains before VEGF protein could reach the brain, VEGF took an average of 21–23 h to reach the brain. Hence, the 1-day time point represents one to several hours of VEGF infusion, rather than a full day of VEGF infusion. Minipumps contained either recombinant mVEGF 164 in PBS, VEGF + Flt-Fc, or PBS (proteins produced at Regeneron). In some experiments, comparison proteins (hFc, hAngiopoietin 1, and hAngiopoietin 2) were infused instead of PBS at concentrations equaling or exceeding the VEGF dose to control for nonspecific protein effects. Angiopoietin 1 and 2 were selected for infusion to control for nonspecific effects of protein growth factors on brain tissue. The hFc protein was selected for infusion both as a general protein control and as a control for the hFc portion of the Flt-Fc. The responses seen in animals infused with control proteins were not different from those observed in animals infused with PBS, except for an occasional tendency toward increased microglial reaction at the infusion site. For dosing experiments, VEGF was given at 30, 60, 120, or 240 ng/day. Flt-Fc, which binds to VEGF with high affinity, was co-infused with either the 120 or 240 ng/day dose of VEGF at 4.8 or 9.6 μ g/day (an approximately 10-fold molar excess compared to VEGF). Because Flt-Fc binds to the exogenous VEGF, preventing it from binding to its endogenous receptors, any nonspecific effects of infusion of VEGF protein into brain tissue should be prevented by co-infusion with Flt-Fc. Indeed, nonspecific tissue irritation would be expected to worsen with the addition of increased protein load (i.e., VEGF + Flt-Fc as opposed to just VEGF). Studies of the chronological development of the VEGF-induced vascular responses and detailed analyses of the 7-day neovasculature used 240 ng/day of VEGF. All experiments were repeated at least once, with group sizes of three to six animals in each treatment group in each study, to ensure that the results obtained were consistent.

Immunohistochemistry

After an overdose of chloral hydrate–pentobarbital, animals were exsanguinated and perfused transcardially with heparinized saline (5 IU/l) followed by 4% buffered paraformaldehyde at 4°C. Some animals received a bolus intravenous injection of 0.026- μ m-diameter fluorescent microspheres (Duke Scientific, Palo Alto, CA) 30 min before exsanguination and perfusion. Brains were removed and placed in 30% buffered sucrose and later sectioned at 40 μ m in the coronal plane. Sections were stored in cryoprotectant (Watson et al., 1986) at –20°C until stained. Brains were immunostained with the antibodies listed in Table 1 using a biotinylated secondary antibody–avidin–peroxidase reaction (Vectastain Elite kit, Vector Laboratories, Burlingame, CA) as described previously (Morse et al., 1993).

Table 1
Antibodies used for immunostaining

Stains for	Antibody	Species	Dilution	Supplier	City
Vascular endothelial cells	RECA	mouse	1:250	Serotec	Raleigh, NC
Leukocytes	OX-1	mouse	1:10,000	Harlan Bioproducts	Cincinnati, OH
Microglia (macrophage)	OX-42	mouse	1:10,000	Serotec	Raleigh, NC
Astroglia	GFAP	rabbit	1:60,000	Dako	Carpinteris, CA
Immunoglobulin	Rat IgG	rabbit	1:1500	Vector Laboratories	Burlingame, CA
Nitric oxide synthase	eNOS	rabbit	1:250	Transduction Laboratories	Lexington, KY
T cells	R73	mouse	1:5000	Serotec	Raleigh, NC
Monocyte/macrophage	ED-1	mouse	1:15,000	Serotec	Raleigh, NC
ICAM-1	CD-54	mouse	1:10,000	Serotec	Raleigh, NC
Smooth muscle cells/pericytes	α -SMA	mouse	1:500	Dako	Carpinteris, CA
Fibrinogen	Fibrinogen	rabbit	1:20,000	Dako	Carpinteris, CA
VEGF	VEGF	goat	1 μ g/ml	R&D Systems	Minneapolis, MN
Proliferating cells	Ki67	mouse	1:100	PharMingen	San Diego, CA
Proliferating cells	BrdU	mouse	1:833	Becton-Dickinson	Franklin Lakes, NJ
Proliferating cells	Histone H3	rabbit	1 μ g/ml	Upstate Biotechnology	Lake Placid, NY
Proliferating cells	PCNA	mouse	1:100	Dako	Carpinteris, CA

Quantification

Changes in vascular quality were evaluated quantitatively using stereological techniques. Animals were perfused with 2% paraformaldehyde, the brains were removed, and then frozen in methylbutane on dry ice. Brains were stored at -80°C until sectioned on a cryostat. Within each group, a third of the brains each were pseudorandomly selected for sectioning at 10 μm in coronal, sagittal, and transverse planes. Stereology was performed on RECA-stained slides after acquisition of images by a DAGE MTI CCD725 video camera (Dage, Michigan City, IN) attached to a Nikon microscope (Morrell Instruments, Melville, NY). Images were analyzed using the NIH Image software (NIH, Bethesda, MD). All vasculature within the approximated field of VEGF diffusion was evaluated (a 3-mm square block of cortex centered on the cannula tip).

Percentage of tissue area containing vessels was determined by point-count stereology. An acetate containing points in a grid pattern were overlaid on the images taken from sections sampled every 200 μm throughout the approximately 1.5 mm extent of VEGF diffusion (for a total of approximately seven to eight sections analyzed per animal). All points (intersections) which laid over a vessel were counted and were represented as a proportion of total points. Points were counted if they either lay on a RECA-positive lumen or lay within a space completely encircled by a circumferential RECA-positive lumen. Spaces in the tissue produced by the cannula track were excluded from analysis. Percentage of vascular area for each brain was expressed as percentage of tissue area occupied by vessels averaged over all of the sections. Percentage of tissue area occupied by vasculature was compared for VEGF ($n = 7$) vs. PBS-infused ($n = 7$) animals using an independent-groups Student's t test with alpha set at 0.05.

Changes in the proportion of tissue area occupied by vasculature can be attributed either to changes in vascular diameter or changes in vessel length. Vessel diameters were

also measured using NIH Image on the same sections sampled for vessel density measurements. Vessels were sampled using a pseudorandomly oriented grid. Vessel diameter was taken by measuring the length of the line representing the smallest perpendicular distance across the vessel. Because more of the grid intersections in the VEGF animals than the PBS animals were overlying vasculature, more vessels were sampled for the VEGF than the PBS animals. However, the mean diameter was determined for each section and then for each animal, such that only one value was statistically analyzed per animal. Vascular diameter was compared for VEGF vs. PBS-infused animals using an independent groups Student's t test with alpha set at 0.05.

All stereological analyses and histological analyses were conducted by observers blind to the treatment condition of the animals from which the tissue was taken.

Western blot analysis

After 7 days of infusion with VEGF or PBS, animals were anesthetized with chloral hydrate-pentobarbital and were rapidly decapitated. The brains were removed and placed inside a 1.5-mm brain block (Zivic-Miller, Allentown, PA). The brain was cut coronally through the cannula track, and the 1.5-mm slab on either side of the track was taken and placed on a cold metal block. Using a fine scalpel blade, cuts were made 1.5 mm lateral and medial to the cannula track in the cortex. The tissue between the cuts in each slab was taken through the entire depth of cortex, placed in a vial, and immediately frozen on dry ice. Tissue was also taken from naive rats.

Tissue was homogenized in lysis buffer containing 1.0% NP40, 0.5% deoxycholic acid, 0.1% SDS, and protease inhibitors (1 Complete tablet, Boehringer Mannheim/Roche, Indianapolis, IN). Lysate (10–20 μl), diluted from 1:1 to 1:5 with sample buffer, was loaded onto each lane of a 12% Tri-Glycine pre-cast Western blot gel. A nonreducing gel was run at 150 V using a Novex Minicell

II gel electrophoresis unit (Novex/Invitrogen, Carlsbad, CA) for 2 h. Protein was transferred to a nitrocellulose membrane (Novex/Invitrogen) for 2–16 h. After transfer, membranes were washed in TBS-T and transferred into a solution containing primary antibody for all Westerns except for the rat IgG Western analysis (rabbit anti-fibrinogen antibody, 1:1000, Dako, Carpinteris, CA; mouse anti-ICAM-1 antibody, 1:1000, Serotec, Raleigh, NC). After an overnight incubation, membranes were washed in TBS-T and transferred to an HRP-conjugated secondary antibody (for rat IgG: goat anti-rat, 1:10,000, Boehringer Mannheim/Roche; for fibrinogen: goat anti-rabbit, 1:10,000, Chemicon, Temecula, CA; for ICAM-1: goat anti-mouse, 1:5000, Boehringer Mannheim/Roche) for 1 h. After washing, membranes were treated with ECL (Amersham Pharmacia Biotech, Piscataway, NJ) and transferred onto film (Kodak, Rochester, NY). Films were exposed from 0.5 s to 2 min. For all antibodies, secondary-only gels were run to control for nonspecific bands. In addition, positive protein controls were available and run for fibrinogen (Sigma, St. Louis, MO) and rat IgG (IgG1 κ , Serotec).

LightCycler PCR

Tissue was collected for RT-PCR analysis of chemokine levels as described for Western blots, except that equivalent tissue was also taken from the contralateral hemisphere of each animal. Tissues were taken after 2, 4, or 7 days of 240 ng/day VEGF. RNA levels were determined for MCP-1, RANTES, MIP-1 α , and GRO- α . Lightcycler PCR was performed as previously described (Lie and Petropoulos, 1998; Ransohoff et al., 2002; Schreiber et al., 2001). Data were analyzed using a 2 (treatment) \times 3 (time) \times 2 (hemisphere) mixed factorial ANOVA, with $\alpha = 0.05$. Tissue was collected from two separate cohorts of animals, and the data were generated by an experimenter blind to the treatment group of the animals. The control group for the first cohort was vehicle (PBS), and the control group for the second cohort was a protein, human Fc, infused at a dose equivalent to that of VEGF. Data from the two control groups were compared using an ANOVA and were combined for the final analysis after confirming statistically that the results from these groups did not differ. In addition, data from the two cohorts were not found to differ from each other overall by ANOVA.

In situ hybridization

Slides were taken for in situ hybridization from those prepared for the quantitative analysis. The MIP-1 α riboprobe was prepared as previously described (Glabinski et al., 1997). In situ hybridization was conducted as previously described (Davis et al., 1996), except that slides were exposed to emulsion for 4 weeks before developing. A control slide from each brain was hybridized to the sense probe, and no specific hybridization pattern was detected.

Results

VEGF dose response

PBS or VEGF (30, 60, 120, or 240 ng/day) were infused continuously into the neocortex of adult male Sprague–Dawley rats for 7 days. Tissue was collected and processed histologically for evaluation of vascular changes. Immunostaining for rat endothelial cell antigen (RECA) revealed that there was little obvious change in cerebral vascular density or morphology when VEGF was infused at a dose of 30 ng/day. At 60 ng/day, small increases in vascular density, tortuosity, and vessel diameter became apparent. These changes became progressively more marked when VEGF was infused at 120 and 240 ng/day (Fig. 1). To confirm the effect of VEGF infusion on vascular leak, tissue sections were also stained for rat IgG, which is excluded from the brain parenchyma of normal adult rats. In rats infused with PBS alone, only a very small amount of rat IgG staining was evident immediately adjacent to the cannula track. However, infusion of VEGF produced a marked extravasation of rat IgG within the brain parenchyma. The amount of rat IgG detected in brain increased in a dose-dependent manner (Fig. 1). Additional sections were immunostained with the pan-leukocyte marker OX-1 to assess infiltration of inflammatory cells into the brain. Few, if any, leukocytes were found in brains infused with PBS, but even the lowest dose of VEGF (30 ng/day), which produced no appreciable changes in vascular morphology, led to a marked extravasation of leukocytes (Fig. 1). The magnitude of the inflammatory response was again clearly dose-dependent. In contrast, infusion of comparison proteins such as human recombinant Angiopoietin-1 and human Fc produced no increase in the extravasation of rat IgG or leukocytes compared to PBS alone (Angiopoietin-1 data shown in Fig. 2C). Furthermore, co-infusion of a 10-fold molar excess of Fc-Fc blocked the leak, inflammation, and increased vasculature induced by VEGF administration (data not shown for leak and inflammation; vasculature shown in Fig. 2D).

Characterization of the VEGF-induced Neovasculature

Vascular morphology

An extensive characterization of the VEGF-induced neovasculature was undertaken following 7 days of infusion. Immunostaining for VEGF revealed that detectable levels of infused protein were apparent within a 1.5-mm radius from the cannula tip (Fig. 3A). RECA immunostaining further revealed that alterations in the normal morphology of brain vasculature were confined to the region of VEGF distribution (refer back to Fig. 2B). The central portion of the infused region was characterized by the presence of a tortuous, large-caliber vascular tangle. At the margin of the VEGF infusion, where VEGF immunostaining was lightest, cerebral vessel morphology was relatively normal,

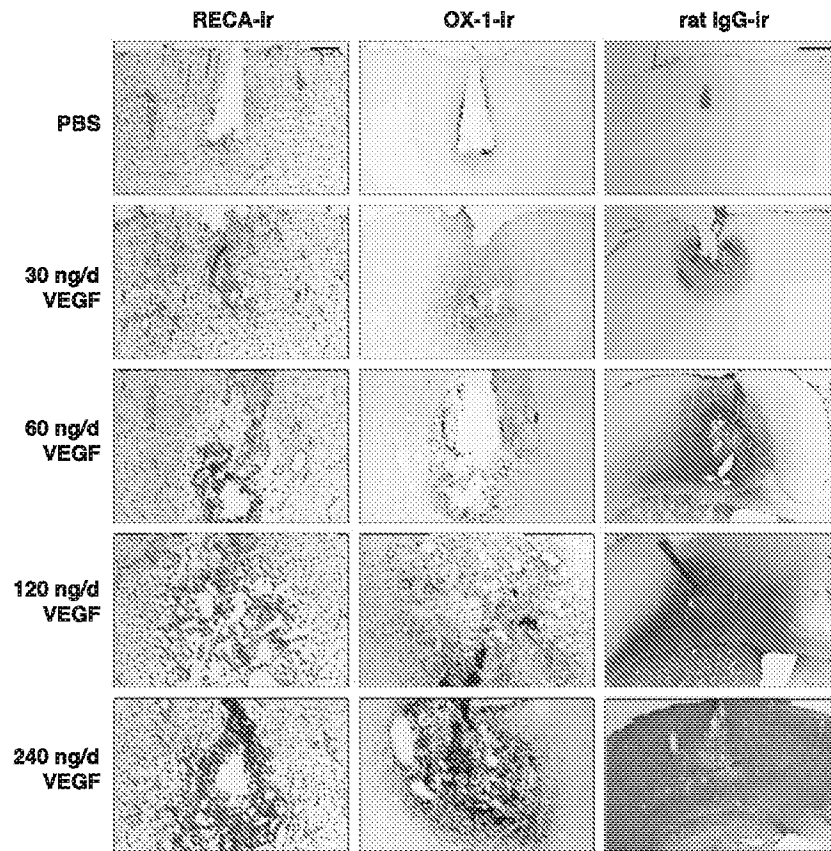


Fig. 1. Immunolocalization of rat endothelial cell antigen (RECA, for visualization of blood vessels), OX-1 (pan-leukocyte), and rat IgG (to visualize extravasated plasma proteins) in brains infused with either PBS or 30, 60, 120, or 240 ng/day of VEGF for 7 days. Angiogenesis, inflammation, and vascular leak were all dose-dependent effects of continuous VEGF infusion. The experiment was conducted twice ($n = 4-6$ per group each time) with comparable results. Scale bar = 300 μm for RECA and OX-1, and 625 μm for rat IgG.

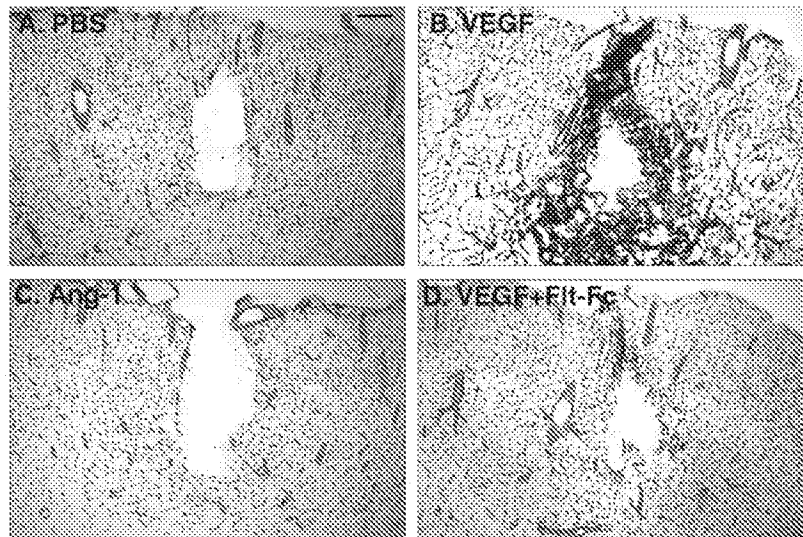


Fig. 2. Immunostaining for RECA after continuous infusion into cortex for 7 days. (A) Infusion of phosphate-buffered saline (PBS) results in minimal vascular change around the cannula track. (B) Infusion of 240 ng/day VEGF induces marked angiogenesis. (C) Infusion of Angiopoietin-1 cannot be differentiated from infusion of PBS. (D) Co-infusion of the receptor body Flt-Fc with VEGF completely prevents VEGF-induced angiogenesis. Each observation was made consistently (in a minimum of two experiments containing four animals per infusion group). Scale bar = 200 μm .

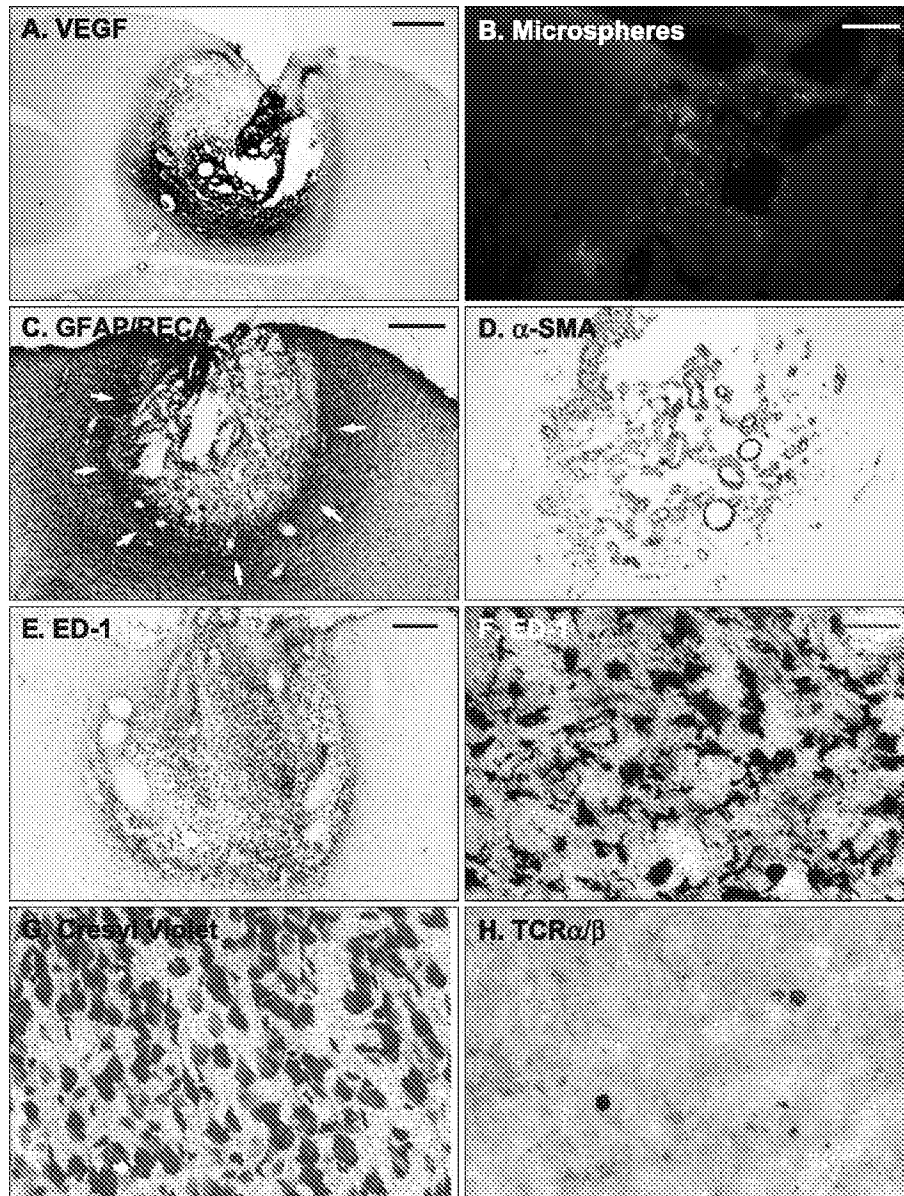


Fig. 3. Cortex infused with 240 ng/day VEGF for 7 days. For panels A–E, the area outside the region of VEGF diffusion displays normal vasculature and can be observed at the edges of the photomicrographs. PBS-infused brains have a similar appearance to the areas outside of the range of VEGF diffusion. (A) VEGF immunostaining shows diffusion of VEGF protein approximately 1.5 mm from the cannula track. (B) Fluorescent microspheres (0.026 μm in diameter) administered intravenously 30 min before sacrifice show that the vasculature within the infused site is acutely leaky at 7 days. (C) A GFAP (dark brown/black) stain shows that there is a dense halo of reactive astroglia located at the peripheral margin of the VEGF-infusion site (approximate boundary with unexposed tissue indicated by the white arrows). In contrast, the center of the VEGF-infusion site is relatively depleted of astrocytes, such that the neovasculature is not well invested with astroglial endfeet. (D) In contrast, α -smooth muscle actin (α SMA) immunostaining illustrates an increased investment of the neovasculature by α -SMA-positive perivascular cells. (E) ED-1, a macrophage/monocyte marker, reveals that most of the inflammatory infiltrate is of monocyte/macrophage origin. (F) A higher magnification view of ED-1-positive cell profiles. (G) Cresyl violet staining after 1 day of VEGF infusion shows the presence of occasional PMNs (example illustrated with an arrow). PMNs are also rarely observed at later time points after VEGF infusions. (H) T cell stain reveals that T cells are not major components of the VEGF-induced immune response. An arrow indicates a TCR-positive cell profile. All findings were confirmed in at least two independent experiments ($n = 4–6$ animals per group per experiment). Scale bars = 500 μm for A, 300 μm for B, 545 μm for C, 340 μm for D and E, and 30 μm for F, G, and H.

though the distance between vascular elements was clearly increased because of local edema. Despite the relative decrease in vascular density at the periphery of the VEGF infusion sites, quantification of vascular density by point-count stereology revealed a significant increase in mean

vascular density across the entire field of the infusion in VEGF vs. PBS-treated animals (Table 2). In addition, analysis of vessel diameters revealed a statistically significant increase in mean vessel caliber in the field of VEGF infusion (Table 2).

Table 2
Quantification of vascular parameters

Measure	Vehicle	VEGF	<i>t</i> test results
Vascular density (%)	15.3	36.9	<i>t</i> (12) = 4.429, <i>P</i> < 0.0008
Vascular diameter (μm)	7.1	17.1	<i>t</i> (12) = 4.469, <i>P</i> < 0.0008

Each value represents the mean value obtained after quantification of seven animals in each group after 7 days of infusion with 240 ng/day VEGF.

Vascular leak

VEGF-induced neovasculature was characterized by a markedly abnormal vascular permeability, as evidenced by the extravasation of both endogenous serum proteins to demonstrate chronic leak (rat IgG, Fig. 1) and fluorescent microspheres (0.026 μm in diameter), injected intravenously 30 min before sacrifice, to demonstrate acute leak (Fig. 3B). Both the endogenous serum proteins and the microspheres are normally excluded from the brain parenchyma, and there is no specific biological mechanism by which microspheres could be actively transported into the brain during pathological states. Therefore, leakage of microspheres not only demonstrates acute leak at time of sacrifice, but is also likely to reflect a generalized, nonspecific leak.

Astrocytes and perivascular cells

The leak of proteins and particulate matter into the brain suggested that the blood–brain barrier was compromised in the VEGF-affected neovasculature. One important contributor to the blood–brain barrier is astroglial endfeet. GFAP immunostaining revealed a marked decrease in GFAP immunoreactivity within the field of VEGF infusion and intensified staining of reactive glia along the boundary between the affected and unaffected tissue (Fig. 3C). Where VEGF concentrations were highest, the few remaining astroglial endfeet did not closely invest the blood vessels as they normally do in rat brain. Interestingly, immunostaining for α-smooth muscle actin (SMA) showed that the investment of vasculature with SMA-positive perivascular cells was not decreased but, rather, was dramatically increased by VEGF infusion (Fig. 3D).

Inflammation

As previously noted, VEGF infusion produced a marked local inflammatory response. Therefore, additional immunostaining studies were conducted to determine the identity of the leukocytes at the infusion site. ED-1 immunostaining revealed that the majority of the inflammatory cells located within the region of VEGF infusion were of monocyte/macrophage origin (Figs. 3E and F), although very few cells were polymorphonuclear cells (Fig. 3G) and almost none were lymphocytes (TCRα/β stain, Fig. 3H). Because the extravasation of inflammatory cells is typically accompanied by the upregulation of adhesion molecules, which initiate leukocyte rolling and sticking, we evaluated the expression of the adhesion

molecule ICAM-1. ICAM-1 was dramatically upregulated in the brains of the VEGF-infused animals (Fig. 4), coincident with both inflammation and the leak of rat IgG and fibrinogen into brain tissue (Fig. 4).

Development of VEGF-induced neovasculature

Vascular morphology

To evaluate the development of VEGF-induced vascular tangles, brains were taken after 1, 3, 5, or 7 days of VEGF infusion. After 1 day of infusion, there was no evidence of VEGF-induced vascular changes (Fig. 5). At 3 days of infusion, changes in vascular morphology were very subtle and variable (Fig. 5). Consistent changes in vascular morphology, including increased vessel diameter and tortuosity, were not clearly evident until 5 days after initiation of the infusion and were more pronounced at 7 days (Fig. 5).

Cellular proliferation

Immunostaining conducted with the proliferation marker Ki67 revealed that, consistent with the results observed with the RECA stain, there was no apparent proliferation of endothelial or perivascular cells after 1 day of infusion (data not shown). At 3 days, there were many Ki67-positive cells present within the field of VEGF diffusion, but few of these were endothelial cells (Fig. 6). Staining for proliferating cell nuclear antigen (PCNA), phosphorylated histone H3, and BrdU (injected at 50 mg ip daily in one experiment) all stained similar non-endothelial cell populations (data not shown). Most stained cells appeared

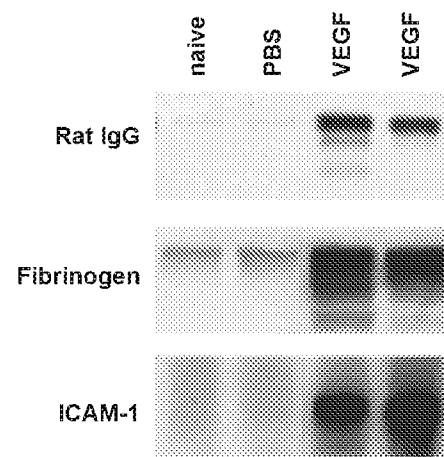


Fig. 4. Unreduced Western blot analyses of rat IgG, fibrinogen, and ICAM-1 found in tissue lysate from rat cortical tissue infused with 240 ng/day VEGF for 7 days. Results obtained from representative unoperated control animals (naive), PBS-infused animals, and two different VEGF-infused animals are illustrated. All Western blot analyses were conducted on two to three different sets of animals at two different protein concentrations (10 and 30 μg). Results were consistent for all proteins analyzed. Results of Westerns were also confirmed by immunostaining tissue sections in separate cohorts of animals (not shown for ICAM-1).

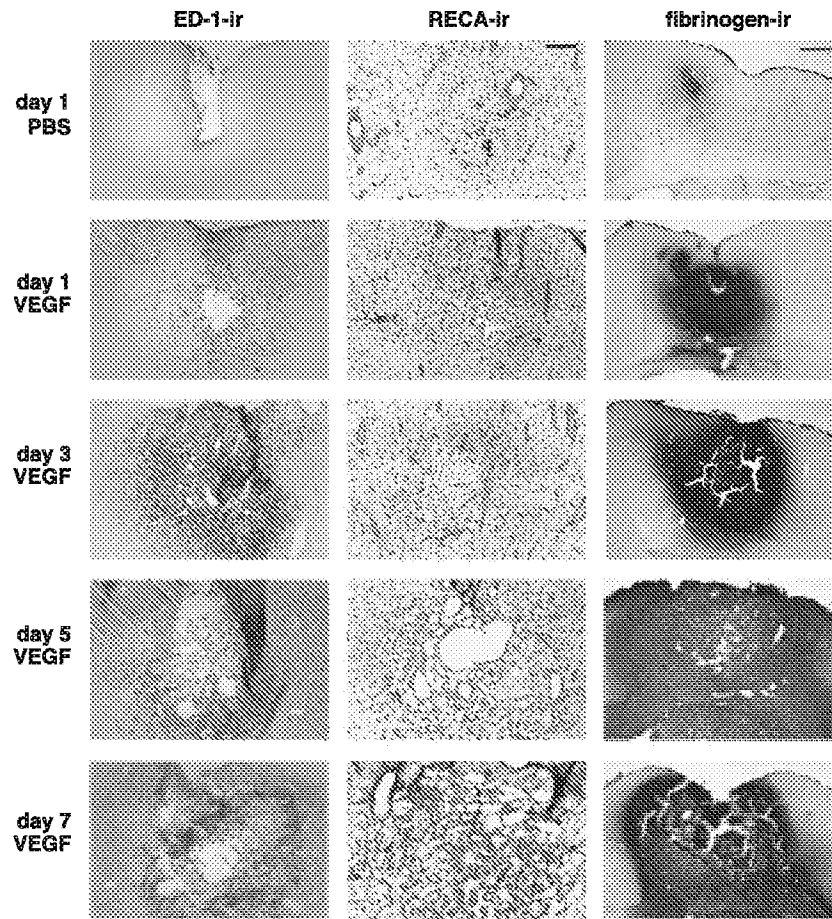


Fig. 5. Immunoreactivity for the monocyte/macrophage marker ED-1, RECA, and fibrinogen are shown for 1, 3, 5, and 7 days after the initiation of 240 ng/day VEGF infusion. Panels in the top row show results after 1 day of PBS infusion. Vascular leak and inflammation are markedly greater after 1 day of VEGF infusions than after 1 day of PBS infusions. Inflammation reaches a plateau by 3–5 days on infusion, and vascular leak plateaus by 5 days of infusions. The experiment was conducted three times ($n = 3–4$ per group per experiment), with each stain completed at least twice. Scale bar = 200 μm for ED-1, 250 μm for RECA, and 625 μm for fibrinogen.

to be of either glial or monocytic morphology (Fig. 6), including dramatic proliferation of both microglia and astroglia at day 7, in agreement with recent findings (astrocytes, Krum et al., 2002). More consistent increases in endothelial cell proliferation were observed at 5 and 7 days of infusion (day 7 shown in Fig. 6). In spite of the dramatic investment of vasculature by smooth muscle cells and pericytes during VEGF infusion, few cells double stained for both Ki67 and α -smooth muscle actin can be observed surrounding the vasculature, suggesting the possibility that these cells were recruited from another site (Fig. 6). Indeed, streams of cells positive for smooth muscle actin, some also positive for Ki67, can be observed in the supracallosal region during VEGF infusion (Fig. 6, 3d Ki67/ α -SMA).

Inflammatory cells

In contrast to the delayed effect of VEGF infusion on angiogenesis, OX-1 staining revealed that extravasation of leukocytes began to occur as early as 1 day after the initiation of VEGF infusions (Fig. 5). This inflammation

is likely to be a direct consequence of VEGF application because there is no more damage or tissue disruption in VEGF-infused animals than in vehicle or protein-infused animals at early time points (determined by Nissl staining, data not shown). Hematoxylin and eosin staining revealed that polymorphonuclear cells (PMNs) were present, but in very small numbers (refer back to Fig. 3G). T cells, as revealed by TCR α/β immunostaining, were very rarely observed. OX-1 immunostaining revealed, however, that the inflammatory response was marked at day 1, increased dramatically at day 3, and continued at high levels at days 5 and 7 of infusion (Fig. 5). The majority of the inflammatory cells at all time points evaluated were determined to be of monocyte/macrophage lineage based on ED1 immunostaining (compare the monocyte marker ED-1 in Fig. 5 to the pan-leukocyte marker OX-1 in Fig. 1).

Vascular leak

In agreement with the VEGF dose-response results, the temporal development of vascular leak paralleled the time

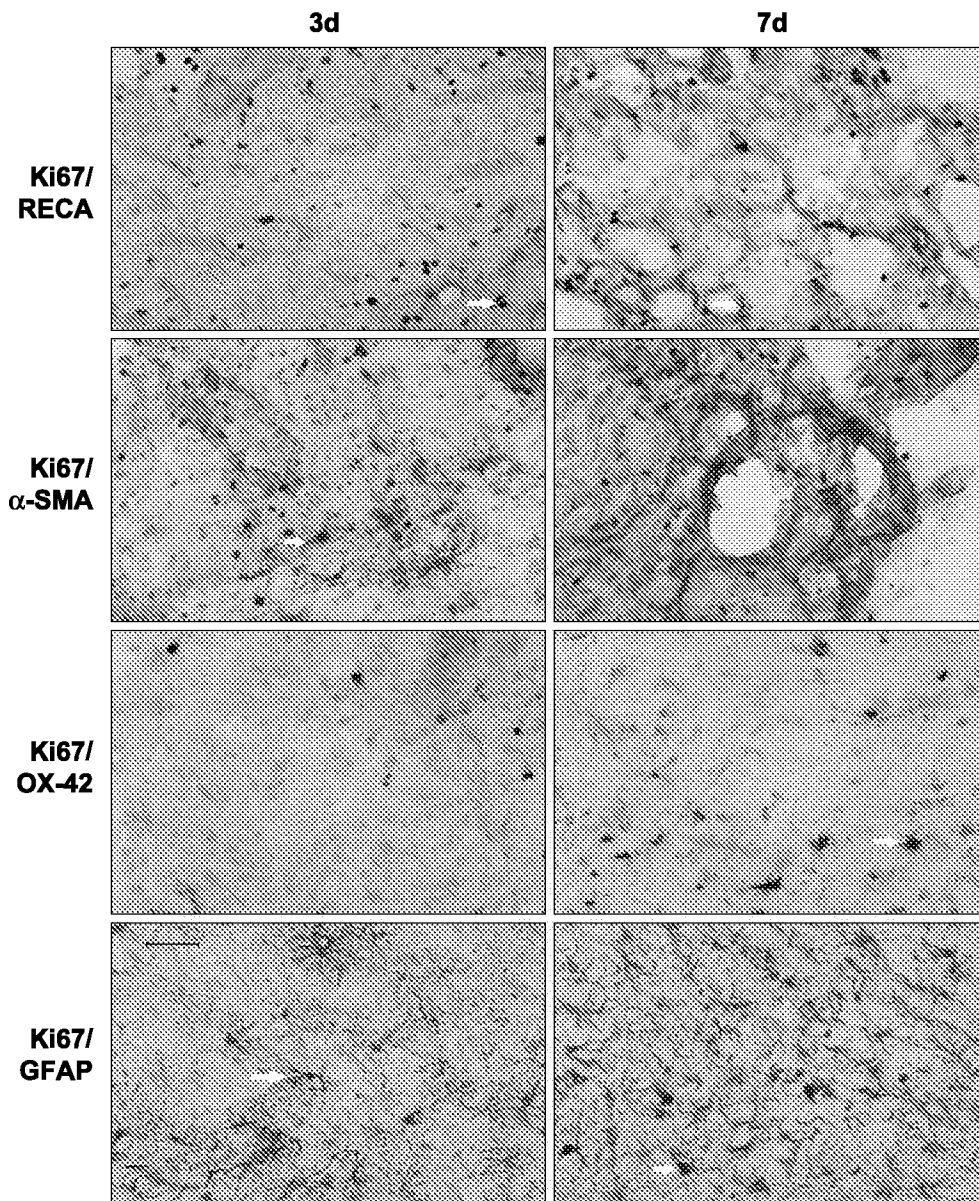


Fig. 6. Double immunoreactivity for Ki67 (a marker of proliferating cells), with RECA, α -SMA, OX-42, and GFAP show that VEGF results, either directly or indirectly, in the proliferation of multiple cell types. Data are shown at 3 and 7 days after the initiation of 240 ng/day VEGF infusion. Ki67 staining was not dramatic at 1 day of VEGF infusions (data not shown). RECA immunostaining reveals that although there are some candidate proliferating endothelial cells (examples shown by arrows), most proliferating cells, especially at 3 days of infusions, are not endothelial cells. In addition, smooth muscle cells did not account for the majority of proliferating cells, although an occasional proliferating smooth muscle actin-positive cell could be identified migrating toward the VEGF infusion site (see arrow). Most proliferating cells were OX-42-positive. Most of these double-positive cells appeared to be monocytic, but at 7 days of infusion, many proliferating cells with a microglial morphology could be identified (see arrow). Large numbers of GFAP-positive cells were double-stained for Ki67. These cells could be first observed at 3 days of VEGF infusions, but were even more numerous at 7 days of VEGF infusions (see arrows). Scale bar = 50 μ m.

course of leukocyte extravasation. That is, infusion of VEGF resulted in an influx of plasma proteins and leukocytes into brain parenchyma which preceded its effects on endothelial cell proliferation. A modest extravasation of plasma proteins and leukocytes was already apparent after 1 day of infusion, and dramatic leak and inflammation were consistently evident at 3, 5, and 7 days of infusion (Fig. 5, fibrinogen staining).

Chemokines

Chemokine mRNA levels were evaluated in brains infused for 2, 4, or 7 days with VEGF, PBS, or control protein (human Fc). LightCycler RT-PCR revealed dramatic increases in MCP-1 mRNA [$F(1,94) = 12.116$, $P < 0.0008$] and modest increases in RANTES mRNA [$F(1,94) = 9.049$, $P < 0.004$] in the infused cortices of both control and VEGF-infused brains, but upregulation of

these chemokines was not significantly associated with infusion of VEGF [$F(1,94) = 1.264$, $P > 0.26$ for MCP-1, $F(1,94) = 2.679$, $P > 0.10$ for RANTES, and additionally, $F(1,44) = 0.529$, $P > 0.47$ for GRO- α , which showed no upregulation in ipsilateral cortex]. In marked contrast, VEGF infusion produced a dramatic and selective upregulation of transcripts for MIP-1 α [$F(1,68) = 8.959$, $P < 0.004$; Fig. 7D].

In situ hybridization for MIP-1 α mRNA revealed that only a few cells expressing MIP-1 α mRNA were evident in PBS-infused brains, and these were located immediately adjacent to the cannula track (data not shown). In marked contrast, in VEGF-treated animals, MIP-1 α mRNA was expressed at high levels in many cells located preferentially near the periphery of the field of VEGF infusion (Fig. 7A). In contrast, most extravasated inflammatory cells are located at the core of the infusion (Fig. 7B). The cell bodies of the hybridized cells appeared dark and angular in the thionin counterstain, properties most typically found in microglia, but also sometimes observed in astroglia, pericytes, or other cell types (Fig. 7C).

Discussion

VEGF is a known mediator of angiogenesis and vascular permeability. Our results demonstrate that VEGF also acts, either directly or indirectly, as a potent pro-inflammatory cytokine. In addition, we show that vascular leak and inflammation precede the endothelial cell proliferation that occurs in response to VEGF.

Chronology and dose response of VEGF-induced vascular changes

Vascular changes with VEGF infusion into brain parenchyma occur with a significant delay upon exposure of normal brain tissues to even high levels of exogenous VEGF. This observation also appears to be the case when angiogenesis is initiated by endogenous VEGF. For instance, after cerebral ischemia, VEGF mRNA is increased within hours of the ischemic injury (Cobbs et al., 1998; Hayashi et al., 1997; Issa et al., 1999; Kovacs et al., 1996; Lee et al., 1999; Lemmyr et al., 1998; Pichiule et al., 1999; Plate et al., 1999), but the angiogenic response

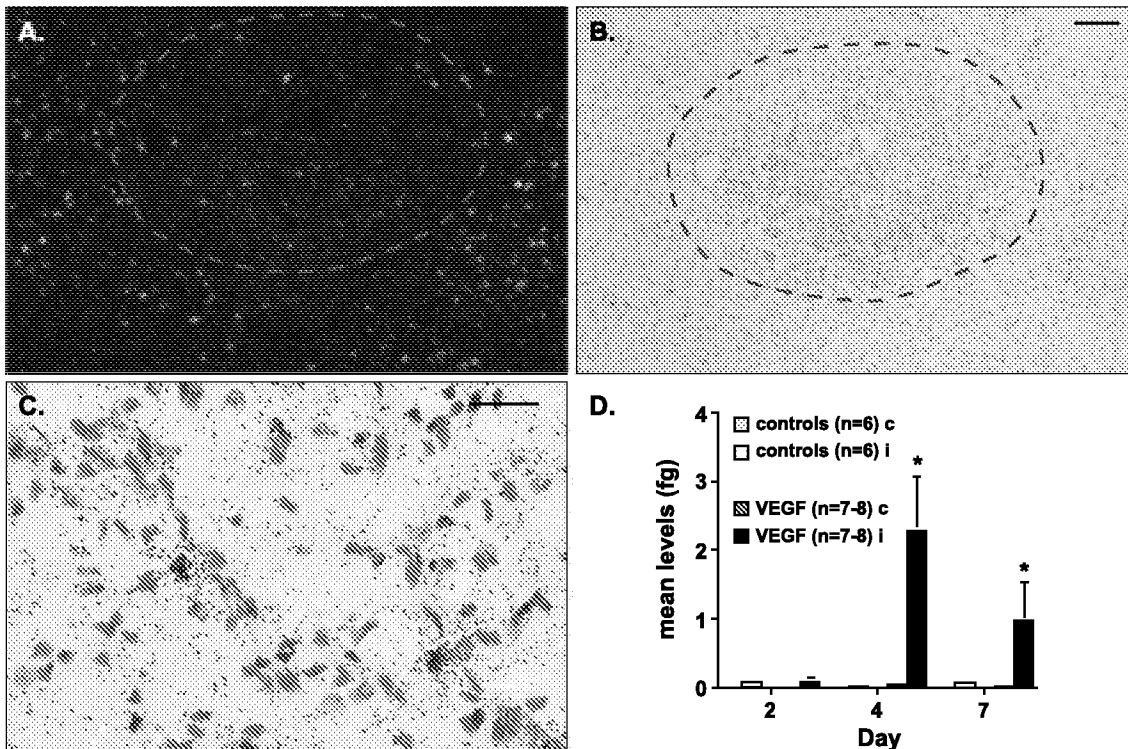


Fig. 7. MIP-1 α mRNAs after VEGF infusion. (A) Dark-field photomicrograph of MIP-1 α in situ hybridization after 7 days of VEGF infusion. The majority of MIP-1 α positive cells lie at the periphery of the infusion site, outlined by the dashed line in panels A and B. Only one or two positive cells were found adjacent to the cannula track in PBS-infused animals. (B) Bright-field photomicrograph from the same section as panel A showing that the majority of the extravasated inflammatory cells were in the core of the VEGF-infused region in which cells hybridized for MIP-1 α were sparse. Scale bar for A and B = 200 μ m. (C) Higher magnification view of area indicated in panel B showing that MIP-1 α positive cells (examples indicated by arrows) have an angular morphology. Scale bar = 50 μ m. (D) MIP-1 α mRNA levels determined by LightCycler PCR after 2, 4, or 7 days infusion of 240 ng/day VEGF. Results were obtained from two separate cohorts of animals, as described in Materials and methods. c = contralateral side, i = ipsilateral side, * $P < 0.05$, significantly different from control group.

is not evident until days later (Beck et al., 2000; Marti et al., 2000). Our observation of delayed angiogenesis was made based on data obtained with RECA immunostaining and proliferation markers. RECA will stain all mature vasculature and most vascular sprouts; however, we cannot rule out the possibility that earlier, RECA-negative basal lamina could have been present before the 3-day time point. Future studies might investigate very early events in the development of vascular sprouts in mature brain vasculature.

In contrast to the apparently delayed angiogenic response, blood–brain barrier compromise occurs very rapidly upon exposure to VEGF. We have observed vascular leak 1 day after VEGF minipump implantation, the earliest time point evaluated, but others have documented vascular permeability within 30 min of acute application (Dobrogowska et al., 1998). In addition, we showed that low doses of VEGF, which did not lead to appreciable angiogenesis within our 7-day infusions, rapidly and substantially increased vascular permeability. VEGF-mediated vascular leak was always accompanied by leukocyte extravasation. Hence, leukocyte extravasation, like vascular leak, occurred at earlier time points and lower doses than angiogenesis.

It has been proposed that VEGF-mediated increases in vascular permeability represent the first step in the angiogenic process (Dvorak et al., 1995, 1999). Vascular leak leads to the deposition of plasma proteins, such as fibrinogen, which can form an extracellular matrix on which endothelial cells can migrate. In this schema, vascular leak facilitates subsequent angiogenesis and may be a necessary pre-requisite for vascular proliferation.

VEGF and inflammation

Although VEGF's roles as a permeability and angiogenic factor are well established, less is understood about VEGF's role in the induction or augmentation of inflammation. We observed a reliable and dramatic induction of a leukocyte infiltrate at early time points of VEGF administration, before the development of obvious angiogenesis. In addition, we observed inflammation at low doses of VEGF insufficient to induce angiogenesis. Using immunostaining, we have shown that the majority of extravasated inflammatory cells are of monocyte/macrophage lineage. VEGF is a monocyte attractant *in vitro* (Heil et al., 2000; Waltenberger et al., 2000) and stimulates hematopoiesis *in vivo* (Hattori et al., 2001). The effect of VEGF on monocytes could be direct because monocytes express VEGFR1 (Flt-1) receptors (Sawano et al., 2001).

Consistent with the increased extravasation of leukocytes, we have detected an increase in the adhesion molecule ICAM-1 in the VEGF-infused cortex. VEGF has previously been shown to increase ICAM-1 both *in vitro* (Kim et al., 2001; Melder et al., 1996) and *in vivo*

(Miyamoto et al., 2000; Proescholdt et al., 1999). ICAM-1 is normally expressed at negligible levels in adult rat brain (refer to naive band in Fig. 4). The increases in this adhesion protein in VEGF-infused brains are consistent with an increased inflammatory response because adhesion molecules encourage leukocyte sticking and rolling in the brain vasculature (for review, see del Zoppo, 1997). VEGF also increases integrin expression on leukocytes (Byzova et al., 2000; Heil et al., 2000). The interaction of leukocyte integrins with ICAM-1 appears necessary for the transendothelial migration of monocytes in response to VEGF (Heil et al., 2000).

Once rolling, leukocytes extravasate into brain in the presence of a leukocyte chemoattractant protein. Among the most common leukocyte attractants are chemokines (for review, see Ransohoff and Tani, 1998). We have discovered that VEGF potently and selectively upregulates the beta chemokine MIP-1 α , which attracts or modulates the function of multiple types of inflammatory cells (Alam et al., 1992; Fahey et al., 1992; Schall et al., 1993). VEGF's effect on MIP-1 α does not generalize broadly to other chemokines, as VEGF did not significantly upregulate MCP-1, RANTES, or GRO- α . Trauma alone dramatically upregulated MCP-1, as previously reported (McTigue et al., 1998; Sun et al., 2000).

We have yet to determine whether VEGF's effects on the inflammatory chemokine MIP-1 α are direct or indirect, although an indirect effect seems likely given the delayed nature of the upregulation. Extravasated monocytes release many cytokines which could act on resident brain cells or newly extravasated immune cells to increase MIP-1 α . MIP-1 α levels increase in microglia and astrocytes upon exposure to cytokines or ischemia (McManus et al., 1998; Peterson et al., 1997; Takami et al., 1997). The increased expression of MIP-1 α in cells surrounding the VEGF infusion site could serve to further propagate the inflammatory response initiated by VEGF infusion.

MIP-1 α is increased in neurological disorders associated with vascular permeability and inflammation. For example, MIP-1 α is elevated both in human multiple sclerosis lesions (Baranzini et al., 2000; Boven et al., 2000) and in the lesions observed in the animal model, experimental allergic/autoimmune encephalitis (Fischer et al., 2000; Glabinski et al., 1998; Nygardas et al., 2000). In fact, the severity of symptoms in experimental allergic encephalitis is decreased in animals deficient in an MIP-1 α receptor, CCR1 (Rottman et al., 2000), although this decreased severity has not yet been observed in MIP-1 α deficient animals (Tran et al., 2000). MIP-1 α also increases within 4–6 h after cerebral ischemia (Gourmalia et al., 1999; Kim et al., 1995; Pang et al., 2002; Spleiss et al., 1998; Takami et al., 1997), concurrent with increased post-ischemic VEGF mRNA levels (Cobbs et al., 1998; Hayashi et al., 1997; Issa et al., 1999; Kovacs et al., 1996;

Lee et al., 1999; Lemmyr et al., 1998; Pichiule et al., 1999, Plate et al., 1999).

Examples of VEGF-induced inflammation *in vivo* have been reported previously, albeit sparsely, in the literature. Mice overexpressing endogenous VEGF exhibit marked inflammation in their skins (Detmar et al., 1998; Xia et al., 2003). VEGF delivered via adenovirus induces a monocyte/macrophage infiltrate in brain similar to the inflammatory response which we have observed (Proescholdt et al., 1999). This inflammatory response could not be induced using other adenoviruses, but could be induced using VEGF protein co-infused with viral particles. Here, we demonstrate that administration of VEGF alone is sufficient to induce an inflammatory reaction. Given the complete inhibition of VEGF's actions by Flt-Fc, the potential for this and similar reagents to inhibit not only angiogenesis, but also VEGF-induced inflammation, should be more fully explored.

Our understanding of the role of VEGF-mediated inflammation in the angiogenic process is still developing. However, increasing evidence suggests that, at least in the adult, VEGF-mediated inflammation is inseparable from, and might participate directly in, the initiation of the angiogenic process as has been proposed previously for vascular leak (Dvorak et al., 1995, 1999). Pathological retinal angiogenesis, which can be suppressed by blockade of VEGF-164, has been shown to be inhibited both by monocyte depletion (Ishida et al., 2003) and by deletion of the gene for ICAM-1 (Sakurai et al., 2003). In addition, placental growth factor (PlGF), a VEGF family member which acts at VEGFR1 but not VEGFR2, induces angiogenesis which is blocked after monocyte depletion in animals (Pipp et al., 2003), suggesting the possibility that angiogenesis mediated by VEGFR1 may be critically dependent on monocytes.

Monocytes secrete both degradative enzymes and a variety of trophic and regulatory factors which establish conditions conducive to endothelial cell proliferation and migration (Pakala et al., 2002). Proliferating vasculature expresses high levels of ICAM-1 coincident with increased numbers of adherent monocytes (Scholz et al., 2000). Monocytes, in particular, are frequently associated with proliferating blood vessels (Arras et al., 1998; Schaper and Buschmann, 1999; Scholz et al., 2000) and can open passages in existing vasculature to encourage the development of vascular sprouts (Molodovan et al., 2000). Furthermore, monocytes can differentiate into endothelial-like phenotypes *in vitro*, especially when treated with VEGF (Fernandez Pujol et al., 2000), raising the possibility that monocytes might contribute directly to angiogenesis as a source of new endothelial cells. Circulating blood cells have been shown previously to integrate into vascular endothelium (Shi et al., 1998). Thus, although the exact role(s) of inflammatory cells in pathological VEGF-mediated angiogenesis remains to be determined, a contributory role seems clear.

Acknowledgments

The authors are grateful to many colleagues at Regeneron for discussions and suggestions, especially Drs. Keith Anderson, John Rudge, Gavin Thurston, Jocelyn Holash, and Peter Maisonpierre. We are grateful to Drs. Nicholas Papadopoulos, Thomas Daly, and Kevin Bailey for production and characterization of protein reagents. Melissa Burrows, Tamar Katz, Dianna Barber, Robert Feeley, and Lisa Fox provided technical assistance, and Evan Burrows, Scott Staton, Vicki Lan, and Brian Ephraim produced the figures.

References

- Alam, R., Forsythe, P.A., Stafford, S., Lett-Brown, M.A., Grant, J.A., 1992. Macrophage inflammatory protein-1 α activates basophils and mast cells. *J. Exp. Med.* 176 (3), 781–786.
- Arras, M., Ito, W.D., Scholz, D., Winkler, B., Schaper, J., Schaper, W., 1998. Monocyte activation in angiogenesis and collateral growth in the rabbit hindlimb. *J. Clin. Invest.* 101 (1), 40–50.
- Baker, P.N., Krasnow, J., Roberts, J.M., Yeo, K.T., 1995. Elevated serum levels of vascular endothelial growth factor in patients with preeclampsia. *Obstet. Gynecol.* 86 (5), 815–821.
- Baranzini, S.E., Elfstrom, C., Chang, S.Y., Butunoi, C., Murray, R., Higuuchi, R., Oksenberg, J.R., 2000. Transcriptional analysis of multiple sclerosis brain lesions reveals a complex pattern of cytokine expression. *J. Immunol.* 165 (11), 6576–6582.
- Bauters, C., Asahara, T., Zheng, L.P., Takeshita, S., Bunting, S., Ferrara, N., Symes, J.F., Isner, J.M., 1994. Physiological assessment of augmented vascularity induced by VEGF in ischemic rabbit hindlimb. *Am. J. Physiol.* 267, H1263–H1271.
- Beck, H., Acker, T., Wiessner, C., Allegrini, P.R., Plate, K.H., 2000. Expression of angiopoietin-1, angiopoietin-2, and tie receptors after middle cerebral artery occlusion in the rat. *Am. J. Pathol.* 157 (5), 1473–1483.
- Bhushan, M., McLaughlin, B., Weiss, J.B., Griffiths, C.E., 1999. Levels of endothelial cell stimulating angiogenesis factor and vascular endothelial growth factor are elevated in psoriasis. *Br. J. Dermatol.* 141 (6), 1054–1060.
- Boulton, M., Foreman, D., Williams, G., McLeod, D., 1998. VEGF localisation in diabetic retinopathy. *Br. J. Ophthalmol.* 82 (5), 561–568.
- Boven, L.A., Montagne, L., Nottet, H.S., De Groot, C.J., 2000. Macrophage inflammatory protein-1 α (MIP-1 α), MIP-1 β , and RANTES mRNA semiquantification and protein expression in active demyelinating multiple sclerosis (MS) lesions. *Clin. Exp. Immunol.* 122 (2), 257–263.
- Brown, L.F., Berse, B., Jackman, R.W., Tognazzi, K., Manseau, E.J., Senger, D.R., Dvorak, H.F., 1993. Expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in adenocarcinomas of the gastrointestinal tract. *Cancer Res.* 53 (19), 4727–4735.
- Brown, L.F., Berse, B., Jackman, R.W., Tognazzi, K., Guidi, A.J., Dvorak, H.F., Senger, D.R., Connolly, J.L., Schnitt, S.J., 1995. Expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in breast cancer. *Hum. Pathol.* 26 (1), 86–91.
- Byzova, T.V., Goldman, C.K., Pampori, N., Thomas, K.A., Bett, A., Shattil, S.J., Plow, E.F., 2000. A mechanism for modulation of cellular responses to VEGF. Activation of the integrins. *Mol. Cell* 6 (4), 851–860.
- Cobbs, C.S., Chen, J., Greenberg, D.A., Graham, S.H., 1998. Vascular endothelial growth factor expression in transient focal cerebral ischemia in the rat. *Neurosci. Lett.* 249 (2–3), 79–82.

- Croll, S.D., Chesnutt, C.R., Rudge, J.S., Acheson, A., Ryan, T.E., Siuciak, J.A., DiStefano, P.S., Wiegand, S.J., Lindsay, R.M., 1998. Co-infusion with a TrkB-Fc receptor body carrier enhances BDNF distribution in the adult rat brain. *Exp. Neurol.* 152, 20–33.
- Croll, S.D., Goodman, J.H., Sollas, A.L., Shah, S.K., Scharfman, H.E., 2002. Upregulation of vascular endothelial growth factor (VEGF) in limbic neurons and glia following pilocarpine-induced seizures in rats. Program No. 598.2. 2002 Abstract Viewer/Itinerary Planner. Society for Neuroscience, Washington, DC. Online.
- Davis, S., Aldrich, T.H., Jones, P.F., Acheson, A., Compton, D.L., Jain, V., Ryan, T.E., Bruno, J., Radjiewski, C., Maisonpierre, P.C., Yancopoulos, G.D., 1996. Isolation of angiopoietin-1, a ligand for the Tie2 receptor, by secretion trap cloning. *Cell* 87, 1161–1169.
- del Zoppo, G.J., 1997. Microvascular responses to cerebral ischemia/inflammation. *Ann. N. Y. Acad. Sci.* 823, 132–147.
- Detmar, M., Brown, L.F., Claffey, K.P., Yeo, K.T., Koehler, O., Jackman, R.W., Berse, B., Dvorak, H.F., 1994. Overexpression of vascular permeability factor/vascular endothelial growth factor and its receptors in psoriasis. *J. Exp. Med.* 180 (3), 1141–1146.
- Detmar, M., Brown, L.F., Schon, M.P., Elicker, B.M., Velasco, P., Richard, L., Fukumura, D., Monsky, W., Claffey, K.P., Jain, R.K., 1998. Increased microvascular density and enhanced leukocyte rolling and adhesion in the skin of VEGF transgenic mice. *J. Invest. Dermatol.* 111 (1), 1–6.
- Dobrogowska, A., Lossinsky, A.S., Tamawski, M., Vorbrodt, A.W., 1998. Increased blood–brain barrier permeability and endothelial abnormalities induced by vascular endothelial growth factor. *J. Neurocytol.* 27, 163–173.
- Dvorak, H.F., Brown, L.F., Detmar, M., Dvorak, A.M., 1995. Vascular permeability factor/vascular endothelial growth factor, microvascular hyperpermeability, and angiogenesis. *Am. J. Pathol.* 146 (5), 1029–1039.
- Dvorak, H.F., Nagy, J.A., Feng, D., Brown, L.F., Dvorak, A.M., 1999. Vascular permeability factor/vascular endothelial growth factor and the significance of microvascular hyperpermeability in angiogenesis. *Curr. Top. Microbiol. Immunol.* 237, 97–132.
- Fahey, T.J., Tracey, K.J., Tekamp-Olson, P., Cousens, L.S., Jones, W.G., Shires, G.T., Cerami, A., Sherry, B., 1992. Macrophage inflammatory protein 1 modulates macrophage function. *J. Immunol.* 148 (9), 2764–2769.
- Fernandez Pujol, B., Lucibello, F.C., Gehling, U.M., Lindemann, K., Weidner, N., Zuzarte, M.L., Adamkiewicz, J., Elsasser, H.P., Muller, R., Havemann, K., 2000. Endothelial-like cells derived from human CD14 positive monocytes. *Differentiation* 65 (5), 287–300.
- Ferrara, N., Carver-Moore, K., Chen, H., Dowd, M., Lu, L., O’Shea, K.S., Powell-Braxton, L., Hillan, K.J., Moore, M.W., 1996. Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature* 380, 439–442.
- Fischer, F.R., Santambrogio, L., Luo, Y., Berman, M.A., Hancock, W.W., Dorf, M.E., 2000. Modulation of experimental autoimmune encephalomyelitis: effect of altered peptide ligand on chemokine and chemokine receptor expression. *J. Neuroimmunol.* 110 (1–2), 195–208.
- Glabinski, A., Tani, M., Strieter, R., Tuohy, V., Ransohoff, R., 1997. Synchronous synthesis of α - and β -chemokines by cells of diverse lineage in the central nervous system of mice with relapses of experimental autoimmune encephalomyelitis. *Am. J. Pathol.* 150, 617–630.
- Glabinski, A.R., Tuohy, V.K., Ransohoff, R.M., 1998. Expression of chemokines RANTES, MIP-1 α and GRO- α correlates with inflammation in acute experimental autoimmune encephalomyelitis. *Neuroimmunomodulation* 5 (3–4), 166–171.
- Gourmala, N.G., Limonta, S., Bochelen, D., Sauter, A., Boddeke, H.W., 1999. Localization of macrophage inflammatory protein: macrophage inflammatory protein-1 expression in rat brain after peripheral administration of lipopolysaccharide and focal cerebral ischemia. *Neuroscience* 88 (4), 1255–1266.
- Grad, S., Ertel, W., Keel, M., Infanger, M., Vonderschmitt, D.J., Maly, F.E., 1998. Strongly enhanced serum levels of vascular endothelial growth factor (VEGF) after polytrauma and burn. *Clin. Chem. Lab. Med.* 36 (6), 379–383.
- Harada, M., Mitsuyama, K., Yoshida, H., Sakisaka, S., Taniguchi, E., Kawaguchi, T., Ariyoshi, M., Saiki, T., Sakamoto, M., Nagata, K., Sata, M., Matsuo, K., Tanikawa, K., 1998. Vascular endothelial growth factor in patients with rheumatoid arthritis. *Scand. J. Rheumatol.* 27 (5), 377–380.
- Hattori, K., Dias, S., Heissig, B., Hackett, N.R., Lyden, D., Tateno, M., Hicklin, D.J., Zhu, Z., Witte, L., Crystal, R.G., Moore, M.A., Rafii, S., 2001. Vascular endothelial growth factor and angiopoietin-1 stimulate post-natal hematopoiesis by recruitment of vasculogenic and hematopoietic stem cells. *J. Exp. Med.* 193 (9), 1005–1014.
- Hayashi, T., Abe, K., Suzuki, H., Itoyama, Y., 1997. Rapid induction of vascular endothelial growth factor gene expression after transient middle cerebral artery occlusion in rats. *Stroke* 28, 2039–2044.
- Heil, M., Clauss, M., Suzuki, K., Buschmann, I.R., Willuweit, A., Fischer, S., Schaper, W., 2000. Vascular endothelial growth factor (VEGF) stimulates monocyte migration through endothelial monolayers via increased integrin expression. *Eur. J. Cell Biol.* 79 (11), 850–857.
- Ishida, S., Usui, T., Yamashiro, K., Kaji, Y., Amano, S., Ogura, Y., Hida, T., Oguchi, Y., Ambati, J., Miller, J.W., Gragoudas, E.S., Ng, Y.S., D’Amore, P.A., Shima, D.T., Adams, A.P., 2003. VEGF164-mediated inflammation is required for pathological, but not physiological, ischemia-induced retinal neovascularization. *J. Exp. Med.* 198 (3), 483–489.
- Isner, J.M., 1998. Arterial gene transfer of naked DNA for therapeutic angiogenesis: early clinical results. *Adv. Drug Delivery Rev.* 30 (1–3), 185–197.
- Isner, J.M., Picczek, A., Schainfeld, R., Blair, R., Haley, L., Asahara, T., Rosenfeld, K., Razvi, S., Walsh, K., Symes, J.F., 1996. Clinical evidence of angiogenesis after arterial gene transfer of phVEGF165 in patient with ischaemic limb. *Lancet* 348, 370–374.
- Issa, R., Krupinski, J., Bujny, T., Kumar, S., Kaluza, J., Kumar, P., 1999. Vascular endothelial growth factor and its receptor, KDR, in human brain tissue after ischemic stroke. *Lab. Invest.* 79, 417–425.
- Jackson, J.R., Minton, J.A., Ho, M.L., Wei, N., Winkler, J.D., 1997. Expression of vascular endothelial growth factor in synovial fibroblasts is induced by hypoxia and interleukin 1 β . *J. Rheumatol.* 24 (7), 1253–1259.
- Kikuchi, K., Kubo, M., Kadono, T., Yazawa, N., Iha, H., Tamaki, K., 1998. Serum concentrations of vascular endothelial growth factor in collagen diseases. *Br. J. Dermatol.* 139, 1049–1051.
- Kim, J.S., Gautam, S.C., Chopp, M., Zaloga, C., Jones, M.L., Ward, P.A., Welch, K.M., 1995. Expression of monocyte chemoattractant protein-1 and macrophage inflammatory protein-1 after focal cerebral ischemia in the rat. *J. Neuroimmunol.* 56 (2), 127–134.
- Kim, I., Moon, S.-O., Kim, S.H., Kim, H.J., Koh, Y.S., Koh, G.Y., 2001. VEGF stimulates expression of ICAM-1, VCAM-1 and e-selectin through nuclear factor- κ B activation in endothelial cells. *J. Biol. Chem.* 276 (10), 7614–7620.
- Kovacs, Z., Ikezaki, K., Samoto, K., Inamura, T., Fukui, M., 1996. VEGF andflt: expression time kinetics in rat brain infarct. *Stroke* 27, 1865–1873.
- Kraft, A., Weindel, K., Ochs, A., Marth, C., Zmija, J., Schumacher, P., Unger, C., Marne, D., Gastl, G., 1999. Vascular endothelial growth factor in the sera and effusions of patients with malignant and nonmalignant disease. *Cancer* 85 (1), 178–187.
- Krum, J.M., Mani, N., Rosenstein, J.M., 2002. Angiogenic and astroglial responses to vascular endothelial growth factor administration in adult rat brain. *Neuroscience* 110 (4), 589–604.
- Lashkari, K., Hirose, T., Yazdany, J., McMeel, J.W., Kazlauskas, A., Rahimi, N., 2000. Vascular endothelial growth factor and hepatocyte growth factor levels are differentially elevated in patients with advanced retinopathy of prematurity. *Am. J. Pathol.* 156 (4), 1337–1344.
- Lee, M.-Y., Ju, W.-K., Cha, J.-H., Son, B.C., Chun, M.-H., Kang, J.K., Park, C.K., 1999. Expression of vascular endothelial growth factor mRNA following transient forebrain ischemia in rats. *Neurosci. Lett.* 265, 107–110.

- Lennmyr, F., Ata, K.A., Funai, K., Olsson, Y., Terent, A., 1998. Expression of vascular endothelial growth factor (VEGF) and its receptors (Flt-1 and Flk-1) following permanent and transient occlusion of the middle cerebral artery in the rat. *J. Neuropathol. Exp. Neurol.* 57 (9), 874–882.
- Lievu, E.R., Rosen, G.F., Cassidenti, D.L., Yee, B., Meldrum, D., Wisot, A., Pedram, A., 1998. Role of vascular endothelial cell growth factor in Ovarian Hyperstimulation Syndrome. *Clin. Invest.* 102 (11), 1978–1985.
- Lie, Y.S., Petropoulos, C.J., 1998. Advances in quantitative PCR technology: 5' nuclease assays. *Curr. Opin. Biotechnol.* 9, 43–48.
- Losordo, D.W., Vale, P.R., Symes, J.F., Dunnington, C.H., Esakof, D.D., Maysky, M., Ashare, A.B., Lathi, K., Isner, J.M., 1998. Gene therapy for myocardial angiogenesis: initial clinical results with direct myocardial injection of rhVEGF165 as sole therapy for myocardial ischemia. *Circulation* 98 (25), 2800–2804.
- Mahnke, J.L., Dawood, M.Y., Huang, J.C., 2000. Vascular endothelial growth factor and interleukin-6 in peritoneal fluid of women with endometriosis. *Fertil. Steril.* 73 (1), 166–170.
- Marti, H.J., Bernaudin, M., Behlail, A., Schoch, H., Euler, M., Petit, E., Risau, W., 2000. Hypoxia-induced vascular endothelial growth factor expression precedes neovascularization after cerebral ischemia. *Am. J. Pathol.* 156 (3), 965–976.
- McClure, N., Healy, D.L., Rogers, P.A., Sullivan, J., Beaton, L., Haning Jr., R.V., Connolly, D.T., Robertson, D.M., 1994. Vascular endothelial growth factor as capillary permeability agent in ovarian hyperstimulation syndrome. *Lancet* 344, 235–236.
- McManus, C.M., Brosnan, C.F., Berman, J.W., 1998. Cytokine induction of MIP-1 α and MIP-1 β in human fetal microglia. *J. Immunol.* 160 (3), 1449–1455.
- McTigue, D., Tani, M., Kravacic, K., Chernosky, A., Kelner, G., Maciejewski, D., Maki, R., Ransohoff, R., Stokes, B., 1998. Selective chemokine mRNA accumulation in the rat spinal cord after contusion injury. *J. Neurosci. Res.* 53, 368–376.
- Melder, R.J., Koenig, G.C., Witwer, B.P., Safabakhsh, N., Munn, L.L., Jain, R.K., 1996. During angiogenesis, vascular endothelial growth factor and basic fibroblast growth factor regulate natural killer cell adhesion to tumor endothelium. *Nat. Med.* 2 (9), 992–997.
- Miyamoto, K., Khosrof, S., Bursell, S.E., Moromizato, Y., Aiello, L.P., Ogura, Y., Adamis, A.P., 2000. Vascular endothelial growth factor (VEGF)-induced retinal vascular permeability is mediated by intercellular adhesion molecule-1 (ICAM-1). *Am. J. Pathol.* 156 (5), 1733–1739.
- Molodovan, N.I., Goldschmidt-Clermont, P.J., Parker-Thornburg, J., Shapiro, S.D., Kolattukudy, P.E., 2000. Contribution of monocytes/macrophages to compensatory neovascularization: the drilling of metallo-elastase-positive tunnels in ischemic myocardium. *Circ. Res.* 87 (5), 378–384.
- Morse, J.K., Wiegand, S.J., Anderson, K.D., You, Y., Cai, N., Carnahan, J., Miller, J., DiStefano, P.S., Altar, C.A., Lindsay, R.M., Alderson, R.F., 1993. Brain-derived neurotrophic factor (BDNF) prevents the degeneration of medial septal cholinergic neurons following fimbria transection. *J. Neurosci.* 13, 4146–4156.
- Neulen, J., Yan, Z., Raczek, S., Weindel, K., Keck, C., Weich, H.A., Marne, D., Breckwoldt, M., 1995. Human chorionic gonadotrophin-dependent expression of vascular endothelial growth factor/vascular permeability factor in human granulosa cells: importance in ovarian hyperstimulation syndrome. *J. Clin. Endocrinol. Metab.* 80 (6), 1967–1971.
- Nygardas, P.T., Maatta, J.A., Hinkkanen, A.E., 2000. Chemokine expression by central nervous system resident cells and infiltrating neutrophils during experimental autoimmune encephalomyelitis in the BALB/c mouse. *Eur. J. Immunol.* 30 (7), 1911–1918.
- Pakala, R., Watanabe, T., Benedict, C.R., 2002. Induction of endothelial proliferation by angiogenic factors released by activated monocytes. *Cardiovasc. Radiat. Med.* 3 (2), 95–101.
- Pang, L., Ye, W., Che, X.M., Roessler, B.J., Betz, A.L., Yang, G.Y., 2002. Reduction of inflammatory response in the mouse brain with adenoviral-mediated transforming growth factor- β 1 expression. *Stroke* 32 (2), 544–552.
- Peterson, P.K., Hu, S., Slak-Johnson, J., Molitor, T.W., Chao, C.C., 1997. Differential production of and migratory response to beta chemokines by human microglia and astrocytes. *J. Infect. Dis.* 175 (2), 478–481.
- Pichiule, P., Chavez, J.C., Xu, K., LaManna, J.C., 1999. Vascular endothelial growth factor upregulation in transient global ischemia induced by cardiac arrest and resuscitation in rat brain. *Brain Res., Mol. Brain Res.* 74 (1–2), 83–90.
- Pipp, F., Heil, M., Issbrucker, K., Ziegelhoeffer, T., Martin, S., van den Heuvel, J., Weich, H., Fernandez, B., Golomb, G., Carmeliet, P., Schaper, W., Clauss, M., 2003. VEGFR-1-selective VEGF homologue PlGF is arteriogenic: evidence for a monocyte-mediated mechanism. *Circ. Res.* 92 (4), 378–385.
- Plate, K.H., Beck, H., Danne, S., Allegrini, P.R., Wiessner, C., 1999. Cell type specific upregulation of vascular endothelial growth factor in an MCA-occlusion model of cerebral infarct. *J. Neuropathol. Exp. Neurol.* 58 (6), 654–666.
- Proescholdt, M.A., Heiss, J.D., Walbridge, S., Mühlhauser, J., Capogrossi, M.C., Oldfield, E.H., Merrill, M.J., 1999. Vascular endothelial growth factor (VEGF) modulates vascular permeability and inflammation in rat brain. *J. Neuropathol. Exp. Neurol.* 58 (6), 613–627.
- Ransohoff, R., Tani, M., 1998. Do chemokines mediate leukocyte recruitment in post-traumatic CNS inflammation? *Trends Neurosci.* 21, 154–159.
- Ransohoff, R.M., Wei, T., Pavelko, K.D., Lee, J.C., Murray, P.D., Rodriguez, M., 2002. Chemokine expression in the central nervous system of mice with a viral disease resembling multiple sclerosis: roles of CD4+ and CD8+ T cells and viral persistence. *J. Virol.* 76 (5), 2217–2224.
- Rosenstein, J.M., Mani, N., Silverman, W.F., Krum, J.M., 1998. Patterns of brain angiogenesis after vascular endothelial growth factor administration in vivo and in vivo. *Proc. Natl. Acad. Sci. U. S. A.* 95, 7086–7091.
- Rottman, J.B., Slavin, A.J., Silva, R., Weiner, H.L., Gerard, C.G., Hancock, W.W., 2000. Leukocyte recruitment during onset of experimental allergic encephalomyelitis is CCR1 dependent. *Eur. J. Immunol.* 30 (8), 2372–2377.
- Sakurai, E., Taguchi, H., Anand, A., Ambati, B.K., Gragoudas, E.S., Miller, J.W., Adamis, A.P., Ambati, J., 2003. Targeted disruption of the CD18 or ICAM-1 gene inhibits choroidal neovascularization. *Invest. Ophthalmol. Visual Sci.* 44 (6), 2743–2749.
- Sawano, A., Iwai, S., Sakurai, Y., Ito, M., Shitara, K., Nakahata, T., Shibuya, M., 2001. Flt-1, vascular endothelial growth factor receptor 1, is a novel cell surface marker for the lineage of monocyte-macrophages in humans. *Blood* 97 (3), 785–791.
- Schall, T.J., Bacon, K., Camp, R.D., Kaspari, J.W., Goeddel, D.V., 1993. Human macrophage inflammatory protein α (MIP-1 α) and MIP-1 β chemokines attract distinct populations of lymphocytes. *J. Exp. Med.* 177 (6), 1821–1826.
- Schaper, W., Buschmann, I., 1999. Arteriogenesis, the good and bad of it. *Cardiovasc. Res.* 43 (4), 835–837.
- Scholz, D., Ito, W., Fleming, I., Deindl, E., Sauer, A., Wiesnet, M., Busse, R., Schaper, J., Schaper, W., 2000. Ultrastructure and molecular histology of rabbit hind-limb collateral artery growth (arteriogenesis). *Virchows Arch.* 436 (3), 257–270.
- Schreiber, R., Krivacic, K., Kirby, B., Vaccariello, S., Wei, T., Ransohoff, R.M., Zigmond, R., 2001. Monocyte chemoattractant protein (MCP)-1 is rapidly expressed by sympathetic ganglion neurons following axonal injury. *NeuroReport* 12, 601–606.
- Senger, D.R., Perruzzi, C.A., Feder, J., Dvorak, H.F., 1986. A highly conserved vascular permeability factor secreted by a variety of human and rodent tumor cell lines. *Cancer Res.* 46 (11), 5629–5632.
- Shi, Q., Rafii, S., Wu, M.H., Wijelath, E.S., Yu, C., Ishida, A., Fujita, Y., Kothari, S., Mohle, R., Sauvage, L.R., Moore, M.A., Storb, R.F., Hammond, W.P., 1998. Evidence for circulating bone marrow-derived endothelial cells. *Blood* 92 (2), 362–367.
- Slevin, M., Krapinski, J., Slowik, A., Kumar, P., Szczudlik, A., Gaffney, J.,

2000. Serial measurement of vascular endothelial growth factor and transforming growth factor- β 1 in serum of patients with acute ischemic stroke. *Stroke* 31 (8), 1863–1870.
- Spirin, K.S., Saghizadeh, M., Lewin, S.L., Zardi, L., Kenney, M.C., Ljubimov, A.V., 1999. Basement membrane and growth factor gene expression in normal and diabetic human retinas. *Curr. Eye Res.* 18 (6), 490–499.
- Spleiss, O., Gourmala, N., Boddeke, H.W., Sauter, A., Fiebich, B.L., Berger, M., Gebicke-Haerter, P.J., 1998. Cloning of rat HIV-1 chemokine coreceptor CKR5 from microglia and upregulation of its mRNA in ischemic and endotoxemic rat brain. *J. Neurosci. Res.* 53 (1), 16–28.
- Springer, M.L., Chen, A.S., Kraft, P.E., Bednarski, M., Blau, H.M., 1998. VEGF gene delivery to muscle: potential role for vasculogenesis in adults. *Mol. Cell* 2 (5), 549–558.
- Sun, D., Tani, M., Newman, T., Krivacic, K., Gill, P., Chernosky, A., Wei, T., Griswold, K., Ransohoff, R., Weller, R., 2000. Role of chemokines, neuronal projections and the blood–brain barrier in the enhancement of EAE following focal brain damage. *J. Neuropathol. Exp. Neurol.* 59, 1039–1043.
- Takami, S., Nishikawa, H., Minami, M., Nishiyori, A., Sato, M., Akaike, A., Satoh, M., 1997. Induction of macrophage inflammatory protein MIP-1 α mRNA on glial cells after focal cerebral ischemia in the rat. *Neurosci. Lett.* 227 (3), 173–176.
- Takeshita, S., Zheng, L.P., Brogi, E., Kearney, M., Pu, L.Q., Bunting, S., Ferrara, N., Symes, J.F., Isner, J.M., 1994. Therapeutic angiogenesis. A single intraarterial bolus of vascular endothelial growth factor augments revascularization in a rabbit ischemic hind limb model. *Clin. Invest.* 93 (2), 662–670.
- Tran, E.H., Kuziel, W.A., Owens, T., 2000. Induction of experimental autoimmune encephalomyelitis in C57BL/6 mice deficient in either the chemokine macrophage inflammatory protein-1 α or its CCR5 receptor. *Eur. J. Immunol.* 30 (5), 1410–1415.
- Waltenberger, J., Lange, J., Kranz, A., 2000. Vascular endothelial growth factor-A-induced chemotaxis of monocytes is attenuated in patients with diabetes mellitus: a potential predictor for the individual capacity to develop collaterals. *Circulation* 102 (2), 185–190.
- Watson, R.E., Wiegand, S.J., Clogh, R.W., Hoffman, G.E., 1986. Use of cryoprotectant to maintain long-term peptide immunoreactivity and tissue morphology. *Peptides* 7, 155–159.
- Xia, Y.P., Li, B., Hylton, D., Detmar, M., Yancopoulos, G.D., Rudge, J.S., 2003. Transgenic delivery of VEGF to mouse skin leads to an inflammatory condition resembling human psoriasis. *Blood* 102 (1), 161–168.
- Zebrowski, B.K., Liu, W., Ramirez, K., Akagi, Y., Mills, G.B., Ellis, L.M., 1999. Markedly elevated levels of vascular endothelial growth factor in malignant ascites. *Ann. Surg. Oncol.* 6 (4), 373–378.

Antibodies against Vascular Endothelial Growth Factor Improve Early Renal Dysfunction in Experimental Diabetes

AN S. DE VRIESE,* RONALD G. TILTON,[†] MARLIES ELGER,[‡]
 CLIFFORD C. STEPHAN,[†] WILHELM KRIZ,[‡] and NORBERT H. LAMEIRE*
 *Renal Unit, Ghent University, Belgium; [†]Departments of Cell Biology and Pharmacology, Texas
 Biotechnology Corporation, Houston, Texas; and the [‡]Institute of Anatomy and Cell Biology I, University of
 Heidelberg, Heidelberg, Germany.

Abstract. Vascular endothelial growth factor (VEGF) is a cytokine that potently stimulates angiogenesis, microvascular hyperpermeability, and endothelium-dependent vasodilation, effects that are largely mediated by endothelial nitric oxide synthase (eNOS). The expression of VEGF is pronounced in glomerular visceral epithelial cells, but its function in renal physiology and pathophysiology is unknown. VEGF expression is upregulated by high ambient glucose concentrations in several cell types *in vitro* and in glomeruli of diabetic rats. To assess the role of VEGF in the pathophysiology of early renal dysfunction in diabetes, monoclonal anti-VEGF antibodies (Ab) were administered to control and streptozotocin-induced diabetic rats for 6 wk after induction of diabetes. Based on *in*

vitro binding studies, an adequate serum VEGF inhibitory activity was achieved during the entire course of anti-VEGF Ab administration. Anti-VEGF Ab treatment but not administration of isotype-matched control Ab decreased hyperfiltration, albuminuria, and glomerular hypertrophy in diabetic rats. VEGF blockade also prevented the upregulation of eNOS expression in glomerular capillary endothelial cells of diabetic rats. Antagonism of VEGF had no effect on GFR and glomerular volume in control rats. These results identify VEGF as a pathogenetic link between hyperglycemia and early renal dysfunction in diabetes. Targeting VEGF may prove useful as a therapeutic strategy for the treatment of early diabetic nephropathy.

It is well established that hyperglycemia is a major risk factor for the development and progression of diabetic nephropathy. Hyperglycemia induces multiple cellular and molecular alterations that presage the development of renal vascular dysfunction. However, the exact sequence of events from exposure to high glucose concentrations to development of renal damage remains equivocal. Early alterations in the diabetic kidney include the development of glomerular hyperfiltration and glomerular hypertrophy, followed by thickening of the glomerular basement membrane, mesangial matrix accumulation, increased urinary albumin excretion rate (UAER), and ultimately progression to glomerular sclerosis. There is compelling evidence for the involvement of various growth factors, including insulin-like growth factor-I (IGF-I) and transforming growth factor- β (TGF- β), in the early renal changes in diabetes (1,2).

Vascular endothelial growth factor (VEGF) is a member of a family of heparin-binding growth factors that very potently stimulate endothelial cell proliferation and play a pivotal role in physiologic and pathologic angiogenesis (3). In addition, VEGF is one of the most potent vascular permeabilizing agents known and has the capacity to induce endothelium-dependent

vasodilation. Several lines of evidence indicate that endothelium-derived nitric oxide (NO) acts as a downstream mediator for VEGF (4,5). In cultured endothelial cells, VEGF stimulates endothelial NO synthase (eNOS) expression and activity, resulting in enhanced generation of bioactive NO (6). In the normal kidney, VEGF is strongly expressed in podocytes and its binding sites are localized mainly on glomerular endothelial cells (7–9). Because of this strategic anatomic location, it has been speculated that VEGF may play a role in the regulation of glomerular permeability and glomerular endothelial cell growth (10). Recent studies have demonstrated an upregulation of VEGF and its receptors in kidneys of diabetic rats (11,12). However, whether VEGF plays a causative role in the pathophysiology of diabetic nephropathy remains unknown. Nevertheless, in view of its known biologic properties, its high glomerular expression in physiologic circumstances, and its upregulation in diabetes, VEGF is an ideal candidate to provide a mechanistic link between hyperglycemia and glomerular hypertrophy, glomerular hyperfiltration, and increased glomerular permeability for macromolecules. We therefore investigated the possibility that inhibition of VEGF activity prevents the onset of early renal dysfunction in experimental diabetes. In addition, the effect of VEGF blockade on glomerular eNOS expression was studied.

Materials and Methods

Laboratory Animals

The studies were performed with female Wistar rats (Iffa Credo, Brussels, Belgium) that received care in accordance with the national

Received May 5, 2000. Accepted October 10, 2000.

Correspondence to Dr. An De Vriese, Renal Unit, University Hospital, OK12, De Pintelaan 185, B-9000 Ghent, Belgium. Phone: +32-9-2404524; Fax: +32-9-2404599; E-mail: an.devriese@rug.ac.be

1046-6673/1205-0993

Journal of the American Society of Nephrology

Copyright © 2001 by the American Society of Nephrology

guidelines for animal protection. Diabetes was induced by intravenous injection of streptozotocin (65 mg/kg; Pfanstiel Europe LTD, Davenham, UK). Slow-release insulin implants (Linsulin, Scarborough, Canada), with a release rate of 1 U/24 h, were used to maintain moderate hyperglycemia. Age-matched control rats received an intravenous injection of buffer solution and placebo implants composed of palmitic acid. Final experiments were carried out 6 wk after the induction of diabetes. At the beginning of each experiment, plasma samples were drawn for analysis of glucose, fructosamine, and total protein levels.

Anti-VEGF Antibody Treatment

Monoclonal anti-VEGF antibodies (Ab) and isotype-matched control Ab were prepared as described previously (13). Briefly, female 8-wk-old BALB/c mice (Harlan Sprague Dawley, Inc., Indianapolis, IN) were immunized, then received a booster three times, 21 d apart, by intraperitoneal and subcutaneous injections of 50 μ g of rh-VEGF₁₆₅, emulsified with an equal volume of Complete Freund's Adjuvant for the primary immunization and Incomplete Freund's Adjuvant for secondary immunizations. The mouse with the highest serum titer to rhVEGF₁₆₅ as measured by enzyme-linked immunosorbent assay received an intravenous injection of an additional 30 μ g of immunogen in phosphate-buffered saline (PBS), 21 d after the last immunization. Three d later, spleen cells were harvested for production of hybridomas to rhVEGF₁₆₅. Two hybridoma cell lines with highest Ab titer and neutralizing Ab activity were cloned three to four times by limiting dilution in 96-well microtiter plates. Ascites fluid was collected from pristane (Sigma Chemical Co., St. Louis, MO) primed BALB/c mice that received intraperitoneal injections of each of the cloned hybridomas (10^7 cells), and purified IgG was prepared by Protein A chromatography (Sigma). The isotype and light chain composition of the Ab and the characterization of neutralizing activity were performed as described previously (13).

Diabetic rats were either untreated ($n = 11$) or treated with anti-VEGF Ab ($n = 12$) or with isotype-matched control Ab ($n = 9$). One mg of the appropriate Ab was injected intraperitoneally three times per week, starting 2 d after the streptozotocin injection until the final experiments. Age-matched control rats were either untreated ($n = 12$) or treated with anti-VEGF Ab ($n = 7$) during 6 wk.

VEGF Binding Studies

To determine the efficiency of the anti-VEGF treatment, we obtained serum samples at baseline and after 2, 4, and 6 wk of treatment with anti-VEGF or control Ab. The samples were drawn from the tail vein just before the next Ab administration. Binding studies were performed using a fusion protein composed of the seven loop ectodomain of Flt-1 fused to the heavy chains of a mouse IgG_{2a} Ab. The fusion protein was captured onto Immulon 4 strip wells with IgG_{2a}-specific goat anti-mouse Ab (Sigma). Increasing concentrations of monoclonal anti-VEGF Ab (used as standards) and increasing dilutions of serum (1:50, 1:100, and 1:250) were incubated separately with 4 ng/ml ¹²⁵I-VEGF (Biomedical Technologies Inc., Stoughton, MA) for 30 min at 37°C before adding to the receptor. Nonspecific binding was defined as the binding measured in the presence of a 100-fold molar excess of unlabeled VEGF. Incubations were terminated after 60 min at 22°C by washing the wells once with 400 μ l of ice-cold PBS, then twice more with 200 μ l of ice-cold PBS. Bound VEGF was measured in a gamma spectrometer (Life Technologies, Inc., Schaumburg, IL).

Measurement of UAER and GFR

For determination of UAER, rats were housed in metabolic cages two times for 24 h and the urine was collected. Urine samples were stored at -20°C until analysis. Albumin was determined with an enzyme-linked immunosorbent assay kit specific for rat albumin (Nephlat, Exocell, Philadelphia, PA). To obtain an estimate of GFR, we measured inulin clearance. The rats were anesthetized with thiobutobarbital (Inactin, RBI, Natick, MA; 100 mg/kg intraperitoneally). The trachea was intubated, a jugular vein was cannulated for continuous infusion of isotonic saline at a rate matching diuresis, and a carotid artery was cannulated for drawing of blood samples. Inulin clearance was measured using the slope technique, as described previously (14). A single dose of FITC-inulin (Sigma, 80 mg/kg) was administered intravenously as a bolus, and plasma samples were obtained at $t = 3, 30, 120, 140, 160,$ and 180 min. FITC-inulin plasma levels were measured with a scanning fluorescence detector (Waters 474, Milford, MA). Inulin clearance was calculated as the ratio of administered dose and area under the curve of the inulin plasma levels.

Measurement of Kidney Weight and Estimation of Glomerular Volume

Kidneys of six animals from each experimental group were rinsed with PBS by retrograde aortic perfusion for 1 min. The right kidney was perfusion-fixed with 2% glutaraldehyde in PBS, stored in the same fixative for 24 h, and rinsed in PBS. Kidneys were embedded in paraffin and stained with Masson-Goldner's trichrome technique. Morphometric analysis was carried out with a semiautomatic image analysis system (VIDS IV, AiTectron, Düsseldorf, Germany) connected to a Zeiss photomicroscope. Cross-sectional glomerular tuft area (A_T , minimal convex polygon) was determined from the mean of 80 random glomerular profiles per animal. Mean glomerular tuft volume (V_T) was calculated as $V_T = \beta/k \times (A_T)(3/2)$, with $\beta = 1.38$, the shape coefficient for spheres and $k = 1.1$, a size distribution coefficient (15).

Immunocytochemistry for eNOS

The expression of eNOS in the glomeruli was investigated using indirect immunocytochemistry. After retrograde aortic perfusion with PBS for 1 min, the left kidney of six animals from each experimental group was removed and snap-frozen in melting isopentane cooled by liquid nitrogen. Five- μ m frozen sections were prepared with a 2800 Frigocut E cryostat microtome (Reichert-Jung GmbH, Nussloch, Germany) and fixed in acetone for 10 min. Nonspecific Ab staining was blocked by incubation with PBS containing 0.2% cold-water fish gelatin (Sigma), 2% bovine serum albumin (Sigma), and 2% fetal calf serum (Sigma) for 30 min at room temperature. Incubation with a mouse monoclonal Ab against human eNOS isotype IgG1 (Transduction Laboratories, Lexington, KY), diluted 1:1000 in PBS containing fish gelatin, was carried out overnight at 4°C. Ab binding was visualized with Cy3-conjugated goat anti-mouse IgG Ab (Jackson Laboratories, West Grove, PA). Sections were mounted and viewed with a Polyvar fluorescence microscope (Reichert-Jung). The specificity of the immunolabeling was confirmed by incubation without primary Ab and by incubation with nonspecific Ab.

The intensity of the eNOS Ab staining in the kidneys was evaluated by two independent investigators who were unaware of the status of the animals. Because the receptors for VEGF are localized mainly on glomerular capillary endothelial cells (7-9), eNOS staining was assessed semiquantitatively in these cells. For each kidney, 20 glomerular profiles, cut in equatorial section planes and successively appear-

ing in the visual field of the microscope when moving the section through the entire depth of the cortex, were evaluated. The number of capillary profiles stained by eNOS Ab was determined and normalized for glomerular surface area.

Immunocytochemical Localization of the Anti-VEGF and Control Ab

To exclude the possibility of precipitation of the Ab or Ab-antigen complexes in the glomeruli, we analyzed a separate group of kidneys (3 diabetes + anti-VEGF Ab, 1 control + anti-VEGF Ab, 2 diabetes + control Ab, 2 diabetes, 2 control). Kidneys were rinsed with PBS by retrograde aortic perfusion for 1 min, then perfusion-fixed with 2% paraformaldehyde and stored in the same fixative for 24 h before being washed with PBS and embedded in paraffin. To detect the murine anti-VEGF Ab, we deparaffinized 5- μ m kidney sections with xylene followed by 100% ethanol, rehydrated them with graded ethanol (100%, 95%, and 70%) followed by PBS, blocked them for 1 h at room temperature using 2% normal goat serum in PBS (Sigma), then incubated them for 1 h at room temperature with anti-mouse IgG, Cy3 conjugate (Sigma), diluted 1:500 in PBS containing 2% normal goat serum.

Statistical Analyses

The data are presented as mean \pm SEM. ANOVA and unpaired *t* tests were used as appropriate to test statistical significance. The significance level was set at *P* < 0.05.

Results

Characteristics of Laboratory Animals

Diabetic animals had significantly higher plasma glucose and fructosamine levels as compared with the age-matched control rats (Table 1). There were no differences in metabolic control between the Ab-treated and the untreated diabetic rats. Body weights were significantly lower in diabetic rats as compared with control rats. There were no significant differences in body weight among the diabetic groups. Kidney weight was significantly higher in all diabetic animals as compared with the control groups. The kidney weight in the diabetic animals that were treated with anti-VEGF Ab tended to be lower than in the other diabetic groups, but the difference was not significant (Table 1). Food consumption was approximately 70% higher in diabetic animals as compared with controls, without significant differences between the diabetic groups (Table 1).

VEGF Binding Studies

VEGF inhibitory activities were measured after dilution of the serum at 1:50, 1:100, and 1:250. The results of the 1:50 dilution are displayed in Figure 1. At baseline, VEGF inhibitory activity was low and not different between experimental groups. A significantly increased serum inhibitory activity was found at 2, 4, and 6 wk of treatment with anti-VEGF Ab in both diabetic and control rats. At 4 wk, inhibitory activity was somewhat lower in anti-VEGF-treated control rats as compared with anti-VEGF-treated diabetic rats. Because the inhibitory activity of undiluted serum of anti-VEGF-treated rats is almost 100%, the clinical relevance of this difference may be minimal. No changes in VEGF inhibitory activity were seen in diabetic rats during treatment with control Ab, except at 6 wk, when a lower inhibitory activity was noted. The inhibitory activities at 1:100 and 1:250 dilution of the serum were lower than those at 1:50 dilution but followed a similar pattern (data not shown).

Measurement of UAER and GFR

Untreated diabetic rats showed a marked elevation of the UAER after 6 wk of diabetes, which was partially reduced by anti-VEGF treatment but not by administration of control Ab (Figure 2). Inulin clearance was elevated in diabetic rats, as compared with control rats (Figure 3). The development of hyperfiltration in the diabetic rats was largely prevented by treatment with anti-VEGF Ab but not by injection of control Ab. The anti-VEGF Ab treatment did not significantly decrease GFR in control rats.

Glomerular Volume

Glomerular volume was significantly higher in diabetic rats as compared with age-matched control rats. Anti-VEGF Ab treatment decreased glomerular volume in diabetic rats but not in control animals (Figure 4). Administration of control Ab did not affect glomerular volume in diabetic rats.

Immunocytochemical Study of eNOS Expression

Positive eNOS staining was detectable in the endothelial cells of preglomerular vessels (arcuate arteries, interlobular arteries, afferent arterioles), of glomerular capillaries, and of postglomerular vessels (efferent arterioles, peritubular capillar-

Table 1. Gravimetric and biochemical characteristics of the experimental groups^a

	Control (n = 12)	Control + Anti- VEGF Ab (n = 7)	Diabetes (n = 11)	Diabetes + Anti- VEGF Ab (n = 12)	Diabetes + Control Ab (n = 9)
Glycemia (mg/dl)	144 \pm 12	117 \pm 8	430 \pm 36 ^b	438 \pm 63 ^b	459 \pm 31 ^b
Fructosamine (μ mol/g total protein)	1.78 \pm 0.04	1.79 \pm 0.06	4.00 \pm 0.23 ^b	4.17 \pm 0.31 ^b	3.92 \pm 0.28 ^b
Body weight (g)	285 \pm 4	273 \pm 5	253 \pm 6 ^b	259 \pm 8 ^b	254 \pm 7 ^b
Kidney weight (g)	0.91 \pm 0.03	0.88 \pm 0.02	1.20 \pm 0.05 ^b	1.12 \pm 0.03 ^b	1.20 \pm 0.05 ^b
Food consumption (g)	14.0 \pm 1.4	14.1 \pm 1.3	24.5 \pm 1.5 ^b	22.5 \pm 1.9 ^b	23.5 \pm 1.4 ^b

^a VEGF, vascular endothelial growth factor; Ab, antibody.

^b *P* < 0.001 versus controls.

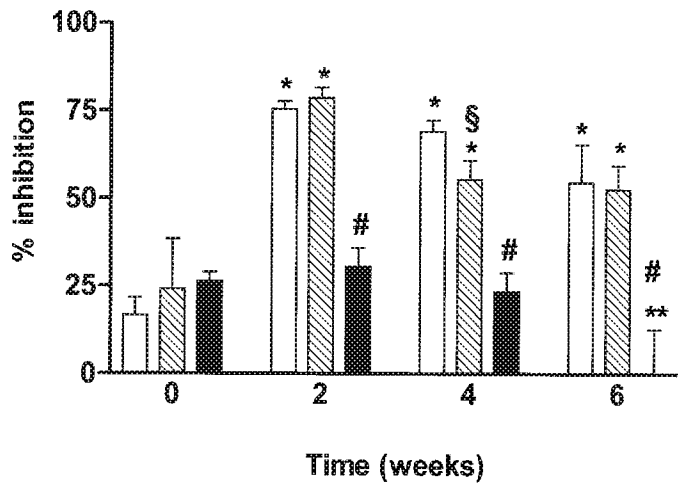


Figure 1. Serum inhibitory activities in diabetic rats + anti-vascular endothelial growth factor (VEGF) antibody (Ab) ($n = 12$, □), control rats + anti-VEGF Ab ($n = 7$, ▨) and diabetic rats + control Ab ($n = 9$, ■) before and 2, 4, and 6 wk after the start of treatment. Inhibitory activities are expressed as percentage of inhibition by 1:50 diluted serum of the binding between ^{125}I -VEGF and Flt-1 *in vitro*. Comparison among time points: *, $P < 0.001$ versus 0 wk; **, $P < 0.05$ versus 0, 2, and 4 wk. Comparison among groups: #, $P < 0.001$ versus diabetic + anti-VEGF Ab and control + anti-VEGF Ab; §, $P < 0.05$ versus diabetic + anti-VEGF Ab.

ies, vascular bundles in the outer medulla). In the glomeruli, the number of stained capillary profiles per glomerulus, normalized for glomerular surface area, was significantly higher in diabetic rats compared with control rats (12.6 ± 0.8 versus 9.1 ± 0.1 ; $P < 0.01$). The increased staining for eNOS in the glomerular capillary endothelial cells was also present in diabetic rats that were treated with control Ab (13.1 ± 1.33 ; $P < 0.05$ versus control) but not in anti-VEGF Ab-treated diabetic rats (8.1 ± 0.4 ; $P < 0.01$ versus diabetes and diabetes + control Ab; Figure 5).

Immunocytochemical Localization of the Anti-VEGF and Control Ab

No staining was detected in either glomeruli, tubules, or peritubular spaces, indicating that the anti-VEGF or control Ab did not accumulate in the kidney (data not shown).

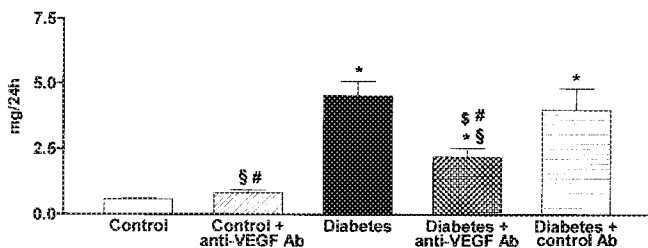


Figure 2. Albuminuria in untreated ($n = 12$) and anti-VEGF Ab-treated ($n = 7$) control rats and in untreated ($n = 11$), anti-VEGF Ab-treated ($n = 12$), and control Ab-treated ($n = 9$) diabetic rats. *, $P < 0.0001$ versus control; §, $P < 0.001$ versus diabetes; #, $P < 0.02$ versus diabetes + control Ab; §, $P < 0.01$ versus control + anti-VEGF Ab.

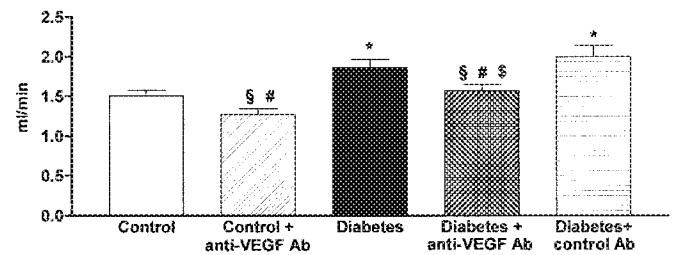


Figure 3. FITC-inulin clearance in untreated ($n = 12$) and anti-VEGF Ab-treated ($n = 7$) control rats and in untreated ($n = 11$), anti-VEGF Ab-treated ($n = 12$), and control Ab-treated ($n = 9$) diabetic rats. *, $P < 0.01$ versus control; §, $P < 0.05$ versus diabetes; #, $P < 0.01$ versus diabetes + control Ab; §, $P < 0.05$ versus control + anti-VEGF Ab.

Discussion

A potential role for VEGF in the pathophysiology of early diabetic nephropathy can be reasoned based on several lines of evidence. First, VEGF is constitutively expressed with a distinct glomerular localization, which suggests an important role in the regulation of glomerular permeability, blood flow, and endothelial cell growth (10). In the adult kidney, VEGF mRNA and protein expression is very pronounced in visceral epithelial cells (7-9). VEGF receptor mRNA and protein were detected in endothelial cells of glomerular capillaries (8,9,11) and pre- and postglomerular vessels (8). The finding that glomerular endothelial cells are a major binding site for exogenous ^{125}I -VEGF (9,11) is consistent with these observations. Although it remains unclear how VEGF crosses the glomerular basement membrane, it is reasonable to postulate the existence of a local regulatory mechanism, with generation of the peptide in podocytes, paracrine secretion, and binding on glomerular capillary endothelium.

Second, VEGF expression is increased in several cell types and tissues by high ambient glucose concentrations. High glucose levels in the culture medium upregulate VEGF expression in vascular smooth muscle cells (16), in retinal epithelial cells (17), and in glomerular endothelial cells (18). Upregulation of VEGF has been demonstrated in the retina of diabetic patients (19) and experimental animals (20). An increased renal expression of VEGF mRNA and protein was reported in

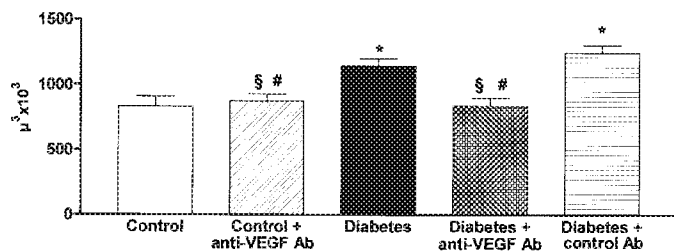
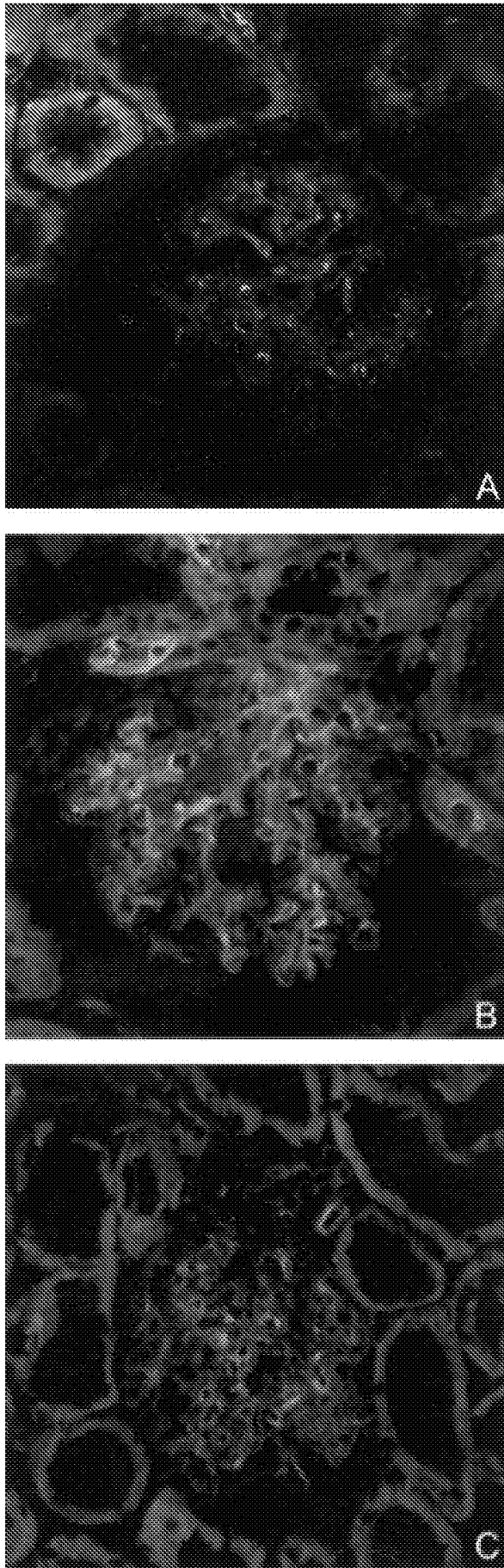


Figure 4. Glomerular volume in untreated ($n = 6$) and anti-VEGF Ab-treated ($n = 6$) control rats and in untreated ($n = 6$), anti-VEGF Ab-treated ($n = 6$), and control Ab-treated ($n = 6$) diabetic rats. *, $P < 0.01$ versus control; §, $P < 0.01$ versus diabetes; #, $P < 0.001$ versus diabetes + control Ab.



experimental rat models of diabetes type I (11) and type II (12). In addition, the expression of VEGF receptors was found to be upregulated in kidneys of diabetic rats (11).

Finally, interference with the VEGF-NO axis was shown to prevent microvascular dysfunction induced by high glucose levels in the dorsal skinfold chamber of the rat (13).

Although these studies suggest a causal role for VEGF in the pathophysiology of diabetic nephropathy, the evidence remains circumstantial. The present data are the first to support a causative role for VEGF in the early renal changes in diabetes. Streptozotocin-induced diabetic rats and age-matched control rats were treated chronically with murine monoclonal anti-VEGF Ab or with isotype-matched control Ab, the latter to exclude an effect of immunization. Based on *in vitro* binding studies, an adequate serum VEGF inhibitory activity was achieved during the entire course of anti-VEGF Ab administration. In diabetic rats, anti-VEGF treatment partially prevented early renal dysfunction. The decrease in GFR in the anti-VEGF Ab-treated diabetic rats could not merely be attributed to precipitation of antigen-Ab complexes in the glomerular basement membrane, because no extravascular murine Ab was detected at the end of the experiment by immunohistochemical analysis. The effects were specific for diabetes, because the anti-VEGF Ab did not affect filtration rate and glomerular volume in control rats. These findings are consistent with a previous report that a VEGF₁₆₅ aptamer did not affect glomerular morphology in normal rats (21). The present results indicate that the upregulation of VEGF and its receptors in the diabetic kidney, as reported by others (11,12), contributes to the pathophysiology of early renal dysfunction in diabetes. The beneficial effect of VEGF-blockade does not exclude the involvement of other growth factors in the pathogenesis of early diabetic renal dysfunction. Inhibition of TGF- β (22,23) and interference with the growth hormone/IGF-I system (24,25) were shown recently to prevent manifestations of early experimental diabetic nephropathy. Several diabetes-induced mediators interact in their adverse effects on the kidney; therefore, it is not surprising that correction of any of them results in amelioration of diabetic nephropathy. In addition, it should be noted that the deleterious effects of VEGF may be limited to early diabetes. When diabetic vascular disease is full-blown and results in tissue ischemia, VEGF may actually be essential for collateral vessel formation (26).

Several recent studies provided evidence that VEGF exerts its angiogenic, vascular permeability, and hemodynamic ef-

Figure 5. Immunocytochemical staining for endothelial nitric oxide synthase (eNOS) in control (A), control Ab-treated diabetic (B), and anti-VEGF Ab-treated (C) kidneys. Staining is detectable in glomerular capillaries and in afferent and efferent arterioles, visible at the vascular pole of the glomeruli. As compared with control rats, the glomeruli of control Ab-treated diabetic rats are characterized by increased number of eNOS-positive capillary profiles. Treatment of diabetic rats with neutralizing anti-VEGF Ab results in a lower number of stained capillaries, which is not different from controls. Magnification, $\times 330$.

ffects via upregulation of eNOS in endothelial cells (4–6). The action of NO as a downstream mediator of VEGF is commensurate with the upregulation of the NO system in early diabetes and its implication in the pathogenesis of early renal dysfunction. NOS blockade substantially reduces or even completely eliminates the hyperfiltration in experimental diabetes (27–29). Chronic administration of NG-nitro-L-arginine methyl ester to diabetic rats partially prevented the increase in glomerular volume in diabetic rats (30). In the present study, an increased expression of eNOS was documented in glomerular capillary endothelial cells of the diabetic rats, confirming earlier findings (27,30). VEGF blockade prevented the upregulation of eNOS in the diabetic glomeruli, thus supporting the contention that VEGF binds to its receptors on glomerular capillary endothelial cells and increases eNOS expression in these cells.

The mechanism(s) by which VEGF may affect glomerular permeability and filtration rate remains speculative. VEGF is known to induce fenestrations in endothelial cells *in vitro* (31), and it has been hypothesized that VEGF is involved in the induction and maintenance of the fenestrae in the glomerular capillary endothelial cells (9). It is, however, generally acknowledged that the capillary fenestrations do not represent the ultimate barrier for filtration but rather that the glomerular basement membrane and the podocyte foot processes with their interconnecting slit diaphragms restrict the passage of proteins. Alternatively, VEGF could affect the filtration barrier by increasing the production of NO in glomerular endothelial cells, which in turn diffuses to the podocytes and acts on the slit pores. In support of this hypothesis, local NOS blockade is known to reduce glomerular ultrafiltration coefficient (32). In addition, VEGF may increase glomerular filtration surface area by stimulating glomerular capillary endothelial cell growth. Nyengaard and Rasch (33) demonstrated that the increased glomerular filtration surface in diabetes results from a formation of new glomerular capillaries, in addition to a slight elongation of existing capillaries. This phenomenon is analogous to the capillary proliferation observed in diabetic retinas and in other vascular beds and thus may be understood as one expression of a generalized diabetic microangiopathy. Our finding that blockade of VEGF prevented the increase in glomerular tuft volume in diabetic rats supports the contention that VEGF is an essential growth factor for glomerular capillaries in pathophysiologic circumstances. In accordance, administration of a VEGF₁₆₅ aptamer was found to decrease glomerular endothelial cell regeneration in experimental glomerulonephritis (21).

The cause of the upregulation of VEGF in diabetes remains speculative, but multiple factors may be implicated. Several factors relevant to the pathogenesis of diabetic complications have been shown to promote VEGF expression in various cell types and tissues, including advanced glycation end products (34), angiotensin II (35), reductive stress (13), reactive oxygen species (36), TGF- β (37), and IGF-I (38). Importantly, protein kinase C (PKC), which is increasingly recognized as a central mediator of the damaging effects of hyperglycemia, has been shown to upregulate VEGF (16,39). Direct (40) or indirect (41) inhibition of PKC activation in diabetic rats produced similar

improvement of glomerular hyperfiltration and albuminuria as in our study, further supporting a link between PKC and VEGF. Taken together, many factors could act independently or in combination to increase VEGF expression in the diabetic kidney. However, the *in vivo* relevance of these pathways remains to be determined. In addition to increased renal expression of VEGF and its receptors (11,12), a disturbed feedback regulation of VEGF could contribute to the pathophysiologic effects of VEGF in the diabetic kidney. Because VEGF is a molecule with very high potency, the existence of powerful defense mechanisms may be assumed. One such mechanism may be the binding of VEGF to the heparin sulfate glycosaminoglycan side chains in the glomerular basement membrane (42). It is tempting to speculate that the reduced heparin content of the glomerular basement membrane in diabetes (43) may contribute to an increased access of VEGF to its receptors.

In conclusion, blockade of VEGF has renoprotective effects in early streptozotocin-induced diabetes in rats. In comparison with untreated diabetic rats, animals that were treated with monoclonal Ab against VEGF exhibited a smaller increase in GFR, glomerular volume, and UAER. The present study demonstrates a new mechanism by which hyperglycemia causes renal dysfunction. Targeting VEGF may prove useful as a therapeutic strategy for the treatment of early diabetic nephropathy.

Acknowledgments

The authors thank Rita De Smet, Julien Dupont, Bruni Haehnel, Inge Hartmann, Pascale Vogelee, and Marie-Anne Waterloos for their expert technical assistance. A.S.D.V. is supported by a grant from the Fund for Scientific Research-Flanders (N20/0).

References

1. Flyvbjerg A, Gronbaek H, Bak M, Nielsen B, Christiansen T, Hill C, Logan A, Orskov H: Diabetic kidney disease: The role of growth factors. *Nephrol Dial Transplant* 13: 1104–1107, 1998
2. Sharma K, Ziyadeh FN: Hyperglycemia and diabetic kidney disease. The case for transforming growth factor-beta as a key mediator. *Diabetes* 44: 1139–1146, 1995
3. Ferrara N: Role of vascular endothelial growth factor in the regulation of angiogenesis. *Kidney Int* 56: 794–814, 1999
4. Ziche M, Morbidelli L, Choudhuri R, Zhang HT, Donnini S, Granger HJ, Bicknell R: Nitric oxide synthase lies downstream from vascular endothelial growth factor-induced but not basic fibroblast growth factor-induced angiogenesis. *J Clin Invest* 99: 2625–2634, 1997
5. Tilton RG, Chang KC, Lejeune WS, Stephan CC, Brock TA, Williamson JR: Role for nitric oxide in the hyperpermeability and hemodynamic changes induced by intravenous VEGF. *Invest Ophthalmol Vis Sci* 40: 689–696, 1999
6. Hood JD, Meininger CJ, Ziche M, Granger HJ: VEGF upregulates eNOS message, protein, and NO production in human endothelial cells. *Am J Physiol* 274: H1054–H1058, 1998
7. Brown LF, Berse B, Tognazzi K, Manseau EJ, Van De Water L, Senger DR, Dvorak HF, Rosen S: Vascular permeability factor mRNA and protein expression in human kidney. *Kidney Int* 42: 1457–1461, 1992
8. Simon M, Gröne HJ, Jöhren O, Kullmer J, Plate KH, Risau W, Fuchs E: Expression of vascular endothelial growth factor and its

- receptors in human renal ontogenesis and in adult kidney. *Am J Physiol* 268: F240–F250, 1995
9. Simon M, Röckel W, Hornig C, Gröne EF, Theis H, Weich HA, Fuchs E, Yayon A, Gröne HJ: Receptors of vascular endothelial growth factor/vascular permeability factor (VEGF/VPF) in fetal and adult human kidney: Localisation and [¹²⁵I]VEGF binding sites. *J Am Soc Nephrol* 9: 1032–1044, 1998
 10. Brenchley PE: VEGF/VPF: A modulator of microvascular function with potential roles in glomerular pathophysiology. *J Nephrol* 9: 10–17, 1996
 11. Cooper ME, Vranes D, Youssef S, Stacker SA, Cox AJ, Rizkalla B, Casley DJ, Bach LA, Kelly DJ, Gilbert RE: Increased renal expression of vascular endothelial growth factor (VEGF) and its receptor VEGFR-2 in experimental diabetes. *Diabetes* 48: 2229–2239, 1999
 12. Tsuchida K, Makita Z, Yamagishi S, Atsumi T, Miyoshi H, Obara S, Ishida M, Ishikawa S, Yasumura K, Koike T: Suppression of transforming growth factor beta and vascular endothelial growth factor in diabetic nephropathy in rats by a novel advanced glycation end product inhibitor, OPB-9195. *Diabetologia* 42: 579–588, 1999
 13. Tilton RG, Kawamura T, Chang KC, Ido Y, Björck RJ, Stephan CC, Brock TA, Williamson JR: Vascular dysfunction induced by elevated glucose levels in rats is mediated by vascular endothelial growth factor. *J Clin Invest* 99: 2192–2202, 1997
 14. Kühnle HF, Linzmeier P, Doerge L: Determination of glomerular filtration rate in rats. In: *Experimental and Genetic Rat Models of Chronic Renal Failure*, edited by Gretz N, Strauch M, Basel, Karger, 1993, pp 331–336
 15. Weibel ER: *Stereological Methods: Practical Methods for Biological Morphometry*, London, Academic, 1979
 16. Williams B, Gallacher B, Patel H, Orme C: Glucose-induced protein kinase C activation regulates vascular permeability factor mRNA expression and peptide production by human vascular smooth muscle cells in vitro. *Diabetes* 46: 1497–1503, 1997
 17. Sone H, Kawakami Y, Okuda Y, Kondo S, Hanatani M, Suzuki H, Yamashita K: Vascular endothelial growth factor is induced by long-term high glucose concentration and up-regulated by acute glucose deprivation in cultured bovine retinal pigmented epithelial cells. *Biochem Biophys Res Commun* 221: 193–198, 1996
 18. Han DC, Chen S, Hong SW, Iglesias-de la Cruz MC, Ziyadeh FN: Increased expression of TGF- β 1, VEGF, and fibronectin in rat glomerular endothelial cells by high ambient glucose [Abstract]. *J Am Soc Nephrol* 10: 681A, 1999
 19. Mathews MK, Merges C, McLeod DS, Luttj GA: Vascular endothelial growth factor and vascular permeability changes in human diabetic retinopathy. *Invest Ophthalmol Vis Sci* 38: 2729–2741, 1997
 20. Gilbert RE, Vranes D, Berka JL, Kelly DJ, Cox A, Wu LL, Stacker SA, Cooper ME: Vascular endothelial growth factor and its receptors in control and diabetic rat eyes. *Lab Invest* 78: 1017–1027, 1998
 21. Ostendorf T, Kunter U, Eitner F, Loos A, Regele H, Kerjaschki G, Henniger DD, Janjic N, Floege J: VEGF(165) mediates glomerular endothelial repair. *J Clin Invest* 104: 913–923, 1999
 22. Ziyadeh FN, Hoffman BB, Han DC, Iglesias-De La Cruz MC, Hong SW, Isono M, Chen S, McGowan TA, Sharma K: Long-term prevention of renal insufficiency, excess matrix gene expression, and glomerular mesangial matrix expansion by treatment with monoclonal antitransforming growth factor-beta antibody in db/db diabetic mice. *Proc Natl Acad Sci USA* 97: 8015–8020, 2000
 23. Han DC, Hoffman BB, Hong SW, Gaa J, Ziyadeh FN: Therapy with antisense TGF-beta1 oligodeoxynucleotides reduces kidney weight and matrix mRNAs in diabetic mice. *Am J Physiol* 278: F628–F634, 2000
 24. Flyvbjerg A, Bennett WF, Rasch R, Kopchick JJ, Scarlett JA: Inhibitory effect of a growth hormone receptor antagonist (G120K-PEG) on renal enlargement, glomerular hypertrophy and urinary albumin excretion in experimental diabetes in mice. *Diabetes* 48: 377–382, 1999
 25. Segev Y, Landau D, Rasch R, Flyvbjerg A, Phillip M: Growth hormone receptor antagonism prevents early renal dysfunction in nonobese diabetic mice. *J Am Soc Nephrol* 10: 2374–2381, 1999
 26. Duh E, Aiello LP: Vascular endothelial growth factor and diabetes. The agonist versus antagonist paradox. *Diabetes* 48: 1899–1906, 1999
 27. Veelken R, Hilgers KF, Hartner A, Haas A, Böhrer KP, Sterzel RB: Nitric oxide synthase isoforms and glomerular hyperfiltration in early diabetic nephropathy. *J Am Soc Nephrol* 11: 71–79, 2000
 28. Komers R, Allen TJ, Cooper ME: Role of endothelium-derived nitric oxide in the pathogenesis of the renal hemodynamic changes of experimental diabetes. *Diabetes* 43: 1190–1197, 1994
 29. Mattar AL, Fujihara CK, Ribeiro MO, de Nucci G, Zatz R: Renal effects of acute and chronic nitric oxide inhibition in experimental diabetes. *Nephron* 74: 136–143, 1996
 30. Sugimoto H, Shikata K, Matsuda M, Kushiro M, Hayashi Y, Hiragushi K, Wada J, Makino H: Increased expression of endothelial cell nitric oxide synthase (eNOS) in afferent and glomerular endothelial cells is involved in glomerular hyperfiltration of diabetic nephropathy. *Diabetologia* 41: 1426–1434, 1998
 31. Esser S, Wolburg K, Wolburg H, Breier G, Kurzchalia T, Risau W: Vascular endothelial growth factor induces endothelial fenestrations in vitro. *J Cell Biol* 140: 947–959, 1998
 32. Gabbai FB, Blantz RC: Role of nitric oxide in renal hemodynamics. *Semin Nephrol* 19: 242–250, 1999
 33. Nyengaard JR, Rasch R: The impact of experimental diabetes mellitus in rats on glomerular capillary number and sizes. *Diabetologia* 36: 189–194, 1993
 34. Lu M, Kuroki M, Amano S, Tolentino M, Keough K, Kim I, Bucala R, Adamis AP: Advanced glycation end products increase retinal vascular endothelial growth factor expression. *J Clin Invest* 101: 1219–1224, 1998
 35. Gruden G, Thomas S, Burt D, Zhou W, Chusney G, Gnudi L, Viberti G: Interaction of angiotensin II and mechanical stretch on vascular endothelial growth factor production by human mesangial cells. *J Am Soc Nephrol* 10: 730–737, 1999
 36. Kuroki M, Voest EE, Amano S, Beerepoot LV, Takashima S, Tolentino M, Kim RY, Rohan RM, Colby KA, Yeo KT, Adamis AP: Reactive oxygen intermediates increase vascular endothelial growth factor expression in vitro and in vivo. *J Clin Invest* 98: 1667–1675, 1996
 37. Pertovaara L, Kaipainen A, Mustonen T, Orpana A, Ferrara N, Saksela O, Alitalo K: Vascular endothelial growth factor is induced in response to transforming growth factor- β in fibroblastic and epithelial cells. *J Biol Chem* 269: 6271–6274, 1994
 38. Punglia RS, Lu M, Hsu J, Kuroki M, Tolentino MJ, Keough K, Levy AP, Levy NS, Goldberg MA, D'Amato RJ, Adamis AP:

- Regulation of vascular endothelial growth factor expression by insulin-like growth factor I. *Diabetes* 1997; 1619-1626, 1997
39. Uchida K, Uchida S, Nitta K, Yumura W, Marumo F, Nihei H: Glomerular endothelial cells in culture express and secrete vascular endothelial growth factor. *Am J Physiol* 266: F81-F88, 1994
40. Ishii H, Jirousek MR, Koya D, Takagi C, Xia P, Clermont A, Bursell SE, Kern TS, Ballas LM, Heath WF, Stramm LE, Feener EP, King GL: Amelioration of vascular dysfunctions in diabetic rats by an oral PKC β inhibitor. *Science* 272: 728-731, 1996
41. Koya D, Lee IK, Ishii H, Kanoh H, King GL: Prevention of glomerular dysfunction in diabetic rats by treatment with d- α -tocopherol. *J Am Soc Nephrol* 8: 426-435, 1997
42. Houck KA, Leung DW, Rowland AM, Winer J, Ferrara N: Dual regulation of vascular endothelial growth factor bioavailability by genetic and proteolytic mechanisms. *J Biol Chem* 267: 26031-26037, 1992
43. Jensen T: Pathogenesis of diabetic vascular disease: Evidence for the role of reduced heparan sulphate proteoglycan. *Diabetes* 46[Suppl 2]: S98-S100, 1997

Glomerular-specific alterations of VEGF-A expression lead to distinct congenital and acquired renal diseases

See the related Commentary beginning on page 600.

Vera Eremina,¹ Manish Sood,¹ Jody Haigh,¹ András Nagy,¹ Ginette Lajoie,² Napoleone Ferrara,³ Hans-Peter Gerber,³ Yamato Kikkawa,⁴ Jeffrey H. Miner,⁴ and Susan E. Quaggin^{1,5}

¹The Samuel Lunenfeld Research Institute, Mount Sinai Hospital, Toronto, Ontario, Canada

²Princess Margaret Hospital, University Health Network, Department of Pathology, Toronto, Ontario, Canada

³Department of Molecular Oncology, Genentech Inc., South San Francisco, California, USA

⁴Department of Medicine, Washington University School of Medicine, Renal Division, St. Louis, Missouri, USA

⁵St. Michael's Hospital, Toronto, Ontario, Canada

Kidney disease affects over 20 million people in the United States alone. Although the causes of renal failure are diverse, the glomerular filtration barrier is often the target of injury. Dysregulation of VEGF expression within the glomerulus has been demonstrated in a wide range of primary and acquired renal diseases, although the significance of these changes is unknown. In the glomerulus, VEGF-A is highly expressed in podocytes that make up a major portion of the barrier between the blood and urinary spaces. In this paper, we show that glomerular-selective deletion or overexpression of VEGF-A leads to glomerular disease in mice. Podocyte-specific heterozygosity for VEGF-A resulted in renal disease by 2.5 weeks of age, characterized by proteinuria and endotheliosis, the renal lesion seen in preeclampsia. Homozygous deletion of VEGF-A in glomeruli resulted in perinatal lethality. Mutant kidneys failed to develop a filtration barrier due to defects in endothelial cell migration, differentiation, and survival. In contrast, podocyte-specific overexpression of the VEGF-164 isoform led to a striking collapsing glomerulopathy, the lesion seen in HIV-associated nephropathy. Our data demonstrate that tight regulation of VEGF-A signaling is critical for establishment and maintenance of the glomerular filtration barrier and strongly supports a pivotal role for VEGF-A in renal disease.

J. Clin. Invest. 111:707-716 (2003). doi:10.1172/JCI200317423.

Introduction

Glomeruli are highly specialized filtration barriers between the blood and urinary space. Each day, approximately 180 l of blood passes through these filters in the average adult human kidney. Although water and small solutes must pass freely through this barrier, critical blood proteins such as albumin and blood clotting factors must not. The filter has a number of unique characteristics that provide the essential properties for this renal filtration process and include highly specialized glomerular visceral epithelial cells (podocytes), a fenestrated glomerular capillary endothelial system, and intervening glomerular basement membrane (GBM) that is produced by both the podocytes and the

endothelial cells (1, 2) (Figure 1a). This filtration barrier is the target of injury and ultimate scarring in a wide variety of kidney diseases (3-8).

During glomerular development, the podocytes express numerous vascular growth factors such as VEGF-A, while the glomerular endothelial cells express the VEGF receptors fetal liver kinase 1 (Flk1) and fms-like tyrosine kinase 1 (Flt1) (9). In addition, the podocytes are geographically situated at the developing vascular cleft adjacent to incoming endothelial cells (2, 10) (Figure 1b). The location and gene expression profile of podocytes suggests that they are required to provide migratory cues to glomerular endothelial cells to establish the renal filtration barrier. Furthermore, similar to other fenestrated vascular beds in the body, podocytes continue to express VEGF-A in the mature glomerulus. This suggests that VEGF plays a role in maintaining the filtration barrier either through survival, proliferation, and/or differentiation cues to the adjacent specialized endothelia.

It is clear that VEGF is a critical mediator of vasculogenesis as heterozygous and homozygous null VEGF-A mice die with major vascular defects at 11.5 and 9.5 days after coitus, respectively (11, 12). However, its later role in specific vascular beds, such as the glomerulus, is less clear. Although dysregulation of VEGF-A expression

Received for publication November 18, 2002, and accepted in revised form January 15, 2003.

Address correspondence to: Susan E. Quaggin, The Samuel Lunenfeld Research Institute, Room 871Q, Mount Sinai Hospital, 600 University Avenue, Toronto, Ontario M5G 1X5, Canada. Phone: (416) 586-4800 ext. 2859; Fax: (416) 586-8588; E-mail: quaggin@mshri.on.ca.

Conflict of interest: The authors have declared that no conflict of interest exists.

Nonstandard abbreviations used: glomerular basement membrane (GBM); fetal liver kinase 1 (Flk1); fms-like tyrosine kinase 1 (Flt1); HIV-associated nephropathy (HIVAN); Wilms tumor suppressor gene (WT1); α -smooth muscle actin (VSMa).

has been demonstrated in a number of renal diseases, the significance of these changes is presently unknown.

To determine the role of VEGF-A in the glomerular filtration barrier, we generated mice with gain or loss of function of VEGF specifically in the podocyte, thus avoiding the embryo-lethal effects. The distinct glomerular-specific haploinsufficient and null phenotypes observed in this study demonstrate for the first time that the “dose” of VEGF is critical in the establishment and maintenance of later vascular beds, as it is in vascular formation during earlier stages of embryogenesis.

By 2.5 weeks of age, mice with podocyte-specific heterozygosity for VEGF developed endotheliosis and “bloodless glomeruli,” the renal lesion seen in preeclampsia, which progressed to nephrotic syndrome, a common glomerular syndrome seen in humans (13), and end-stage kidney failure by 9–12 weeks. The podocyte-specific homozygotes died at birth or within 18 hours of birth with hydrops (generalized swelling), kidney failure, and grossly abnormal glomeruli that lack mature endothelial cells. In contrast, overexpression of the 164 isoform of VEGF-A in podocytes also led to end-stage renal failure due to a collapsing glomerulopathy, which is the pathologic lesion seen in HIV-associated nephropathy (HIVAN) (14). This demonstrates that there is an ongoing requirement for tight regulation of VEGF signaling between the podocyte and glomerular endothelium; disruption in this regulation leads to dramatic and distinct renal phenotypes that are determined by the glomerular VEGF dose and suggests that VEGF is pivotal in a wide variety of renal diseases.

Methods

Cell-specific gene targeting. Three independent podocyte-specific Cre recombinase murine lines, A15, GG8, and V9, which all demonstrated 100% excision when crossed to the Z/EG reporter mouse strain (15), were bred to the floxed VEGF-A mouse (Figure 1c). The VEGF-A mouse has loxP sites inserted around the third exon (16). Site-specific recombination between the loxP sites of the VEGF gene results in a null VEGF allele (16).

To generate homozygous floxed VEGF-Cre recombinase mice, bitransgenic mice carrying both a nephrin-Cre transgene and one floxed VEGF-A allele were bred to homozygous floxed VEGF-A mice.

To generate transgenic founders that overexpress the 164 isoform of VEGF-A, a 645-bp fragment of the VEGF gene, including a Kozak consensus sequence (nucleotides 78–669 of GenBank accession no. NM009505) and initiating ATG, were subcloned into the XhoI and XbaI sites of a pNXRS vector between a 4.125-kb 5' fragment of the murine nephrin gene that is capable of podocyte-specific expression in vivo in the kidney (17) and a 0.97-kb poly(A) signal from the SV40 polyoma virus (Figure 1e).

Genotyping. Genomic DNA was isolated from mouse tails as described. The nephrin-Cre transgenic mice

were generated and genotyped as previously described (17). Floxed VEGF mice were received from Napoleone Ferrara (Genentech Inc.). Presence of the floxed VEGF gene was detected by PCR using the oligonucleotide primers muVEGF 419.F (5'-CC TGCCCTCAAGTACACCTT-3') and muVEGF 567.R (5'-TCCGTACGACGCATTCTAG-3') (both from Sigma-Genosys, The Woodlands, Texas, USA), which generate a 148-bp fragment of the VEGF allele in the presence of the loxP-1 site and a DNA fragment that is approximately 40 bp shorter than for the wild-type allele (16).

To identify founder mouse lines that carried the nephrin-VEGF-164 transgene, Southern blot analysis was performed. Briefly, the DNA was digested with EcoRI; the probe used was the 645-bp fragment encoding the VEGF-164 cDNA that recognized a 1.3-kb genomic fragment in the transgenic founders. To estimate transgene copy number, 1 µg, 2 µg, and 5 µg of genomic DNA from the transgenic founder or wild-type mice was blotted on Biodyne B membrane (P/N 60207, Pall Gelman Laboratory, Ann Arbor, Michigan, USA) and hybridized with the VEGF cDNA probe described above. The signal was quantified using the Quantity One quantitation software program (4.2.1 version) (Bio-Rad Laboratories, Hercules, California, USA) according to the manufacturer's instructions.

Phenotypic analysis. Urine was collected passively in an Eppendorf tube from 0-, 3-, 6-, and 9-week-old mice. A urine dipstick (Chemstrip 5L; Roche Diagnostics Corp., Indianapolis, Indiana, USA) was used to detect the presence or absence of protein and red blood cells in the urine. The standard colorimetric assay was performed according to the manufacturer's instructions. In addition, 2 µl of urine from transgenic or control mice was placed in 18 µl of Laemmli buffer (18), boiled, and loaded on a 12% SDS-PAGE gel. An SDS-PAGE low-range protein standard (Bio-Rad Laboratories Inc., Hercules, California, USA) was loaded in the first lane of the gel.

Blood samples were taken with a heparinized capillary tube by femoral vein stab after warming. A total of 120 µl of blood was collected and creatinine, urea, and blood chemistry measurements were recorded using a Stat Profile M7 (Nova Biomedical Corp., Waltham, Massachusetts, USA). The CBC (total blood count) was performed on a Coulter Counter (AcT diff; Beckman Coulter Canada, Ontario, Canada).

Histologic analysis. Embryonic tissues for histologic analysis were dissected, fixed in 10% formalin/PBS, and embedded in paraffin. Sections 4 µm thick were cut. Sections were stained with H&E, examined, and photographed with a DC200 Leica camera and Leica DMLB microscope (Leica Microsystems Inc., Deerfield, Illinois, USA). Tissue for electron microscopy was fixed in 1.5% glutaraldehyde, embedded in Spurr (Canemco Inc., Saint-Laurent, Quebec, Canada), and sectioned.

In situ hybridization and immunohistochemistry. Kidneys were dissected from mice on postnatal day 0 and at 1 week, 3 weeks, 6 weeks, or 9 weeks of age. Kidneys were

washed briefly in RNase-free PBS and fixed overnight in DEPC-treated 4% paraformaldehyde. These tissues were then placed in 30% sucrose for 12–24 hours, embedded in Tissue-Tek OCT 4583 compound (Sakura Finetek USA Inc., Torrance, California, USA) and snap frozen. Ten-micron tissue samples were cut on a Leica Jung cryostat (model CM3050; Leica Microsystems Inc.) and transferred to Superfrost microscope slides (Fisher Scientific Co., Pittsburgh, Pennsylvania, USA). The slides were stored at -20°C until needed. Digoxigenin-labeled probes were prepared according to the Roche Molecular Biochemicals protocol (Roche Molecular Biochemicals, Mannheim, Germany). Probes used for in situ analysis were nephrin (19), Wilms tumor suppressor gene (WT1; a kind gift of J. Kreidberg, Children's Hospital, Boston, Massachusetts, USA), podocin (a kind gift from C. Antignac, Institut National de la Santé et de la Recherche Médicale, Paris, France), α -smooth muscle actin (VSMA; a

kind gift from P. Igarashi, Southwestern University, Dallas, Texas, USA), VEGF-A (kind gift of A. Nagy, Samuel Lunenfeld Research Institute, Toronto, Canada). Details of the in situ analysis protocol may be obtained upon request.

Immunostaining was performed with antibodies to WT1 and PECAM as described (20).

Results

Mice that are heterozygous for VEGF-A in the podocyte develop endotheliosis and nephrotic syndrome. To determine whether there is any phenotype resulting from a reduction in VEGF-A gene dose within the podocyte, we used the Cre-loxP system. Nephrin-Cre recombinase mice were generated in our laboratory and are capable of site-specific recombination in 100% of podocytes at the capillary-loop stage in vivo (17). We generated mice that were heterozygous for the floxed VEGF-A allele and carried the nephrin-Cre transgene *VEGF-loxP^{+/+},Neph-Cre^{+/-}*

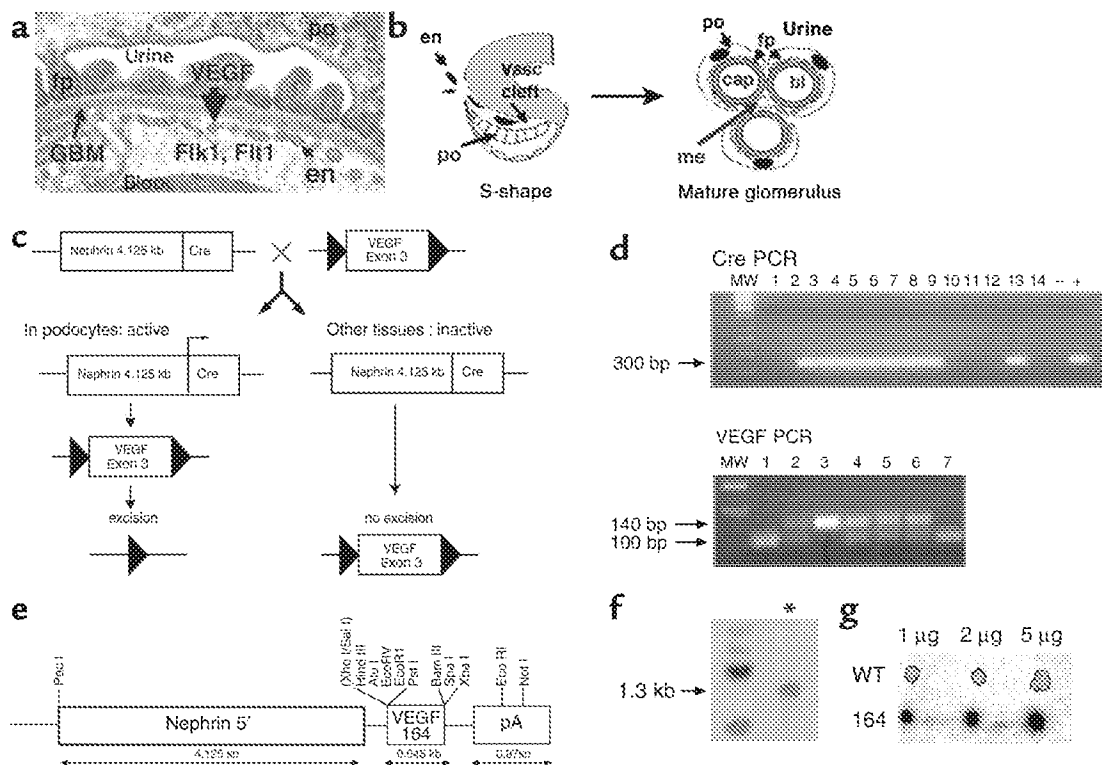


Figure 1

Expression and genomic targeting of VEGF-A within the glomerular filtration barrier. **(a)** Transmission electron micrograph of the glomerular filtration barrier that consists of podocytes (po) and their specialized foot processes (fp), fenestrated endothelium (en), and intervening GBM. VEGF-A is produced in the podocyte; the VEGF receptors Flk1 and Flt1 are expressed in the adjacent endothelial cells. **(b)** Development of the glomerular filtration barrier. In the S-shape stage, podocyte precursors (po) express VEGF-A. Endothelial cells (en) that express the VEGF receptors migrate into the vascular (Vasc) cleft and differentiate in direct apposition to podocytes. In the mature glomerulus, the fenestrated endothelial capillary loops (cap) remain in intimate contact with the VEGF-expressing podocytes (po). Mesangial cells (me) provide support to the capillary tuft. Urine is formed as blood (bl) is filtered from the capillaries, across the GBM, and through slit diaphragms that connect adjacent podocyte foot processes (fp). **(c)** Scheme to generate heterozygous and homozygous podocyte-specific VEGF knock-out mice. Triangles are 34 bp loxP sites. **(d)** The Cre recombinase transgene was identified as a 300 bp PCR product. The floxed VEGF allele measures 140 bp by PCR analysis, whereas the wild-type allele measures 100 bp. MW, molecular weight markers. **(e)** Transgenic construct used to overexpress the 164-isoform of VEGF. pA, poly(A). **(f)** Presence of the transgenic VEGF-164 gene was identified as a 1.3-kb band (*) by Southern blot analysis. **(g)** Dot blot analysis of transgene copy number. The transgenic founder mice (164) demonstrated a 30-fold increase in copy number compared with the wild type.

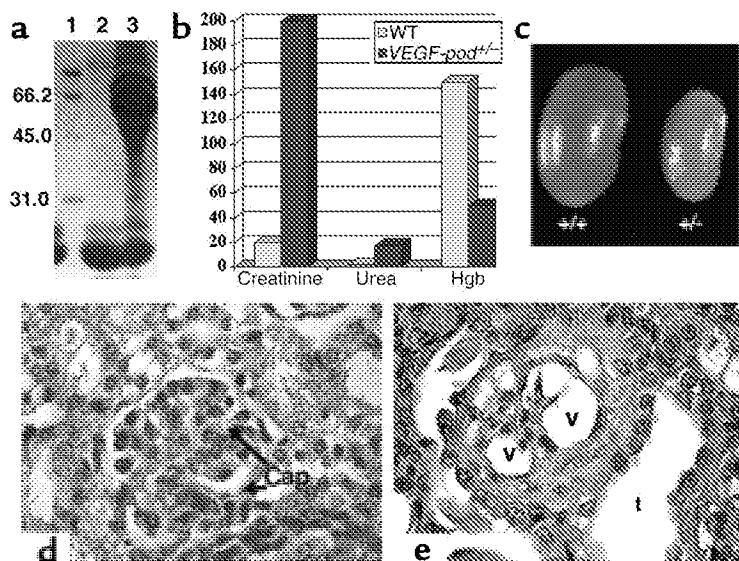


Figure 2 Heterozygous *VEGF-loxP^{+/-},Neph-Cre^{+/-}* mice develop nephrotic syndrome and end-stage renal failure by 9 weeks of age. (a) SDS-PAGE analysis was performed using 2 μ l of mouse urine. Lane 1 contains molecular weight markers, lane 2 shows urine from a *VEGF-loxP^{+/-},Neph-Cre^{+/-}* control aged 9 weeks, and lane 3 shows urine from a 9-week-old sick *VEGF-loxP^{+/-},Neph-Cre^{+/-}* animal. The presence of a large amount of albumin measuring 66.2 kDa is identified in the sick mouse and demonstrates damage to the kidney filter. In contrast, low-molecular-weight proteins, which are normally found in mouse urine, are not different. (b) Bar graph showing elevated creatinine (more than ten times higher than normal), elevated urea, and decreased hemoglobin (Hgb) in *VEGF-loxP^{+/-},Neph-Cre^{+/-}* mice (*VEGF-pod^{+/-}*) at 9 weeks of age compared with *VEGF-loxP^{+/-},Neph-Cre^{+/-}* and *VEGF-loxP^{+/-},Neph-Cre^{+/-}* mice (combined for analysis and considered as wild type). (c) Whole-mount image of a kidney from a sick 9-week-old *VEGF-loxP^{+/-},Neph-Cre^{+/-}* (+/-) mouse compared with that of a wild-type littermate (+/+). The affected kidney is pale and shrunken. Magnification: $\times 60$. (d) A wild-type glomerulus. Note the open capillary loops (Cap). $\times 350$. (e) A glomerulus from a heterozygous VEGF-A mouse. All the glomeruli are grossly distorted morphologically. Note the empty cytoplasmic vacuoles (v) that are present in podocytes. No patent capillary loops can be seen. Dilated tubules (t) can be seen and in most places are packed with proteinaceous material, consistent with nephrotic syndrome. Magnification: $\times 375$.

(Figure 1, c and d). They were born in the expected mendelian frequency but developed end-stage kidney failure by 9–12 weeks of age.

Physical examination of mice at this stage showed that 30/30 of the *VEGF-loxP^{+/-},Neph-Cre^{+/-}* mice had lethargy and decreased skin turgor. Urinalysis was performed and showed 3.0 g/l of protein (defined as “nephrotic range” proteinuria) and 250 red blood cells/ μ l of urine by dipstick analysis. SDS-PAGE analysis demonstrated massive albuminuria in all of these *VEGF-loxP^{+/-},Neph-Cre^{+/-}* mice (Figure 2a), which is pathognomonic for damage to the filtration barrier. Blood chemistry showed mice to have severely decreased renal function with an elevated serum creatinine that measured 200 μ M, more than ten times the normal value, markedly elevated urea, and a normochromic, normocytic anemia consistent with end-stage kidney failure (Figure 2b). Of note, fragmentation of red blood cells was not observed on the blood smear. The mice did

not demonstrate any gross signs of renal failure prior to 7–8 weeks of age.

At 9 weeks of age, the kidneys were pale and shrunken (Figure 2c). By light microscopy, the glomeruli looked histologically normal at birth and at 1, 3, and 6 weeks of age. However, by 9 weeks of age, the glomerular tufts were retracted with expansion of the mesangial matrix and were surrounded by podocytes containing large empty cytoplasmic vacuoles. The tubules were packed with protein (Figure 2e and data not shown).

Serial transmission EM studies demonstrated that the first detectable lesion occurred at 2.5 weeks of age with swelling of the endothelial cells (endotheliosis) and hyaline deposits (Figure 3a) that resemble the pathologic lesions seen in renal biopsies from patients with preeclampsia, a common disease of pregnancy (21). At this time, the podocytes and endothelial cells appeared ultrastructurally normal with well-formed foot processes and fenestrations, respectively. By 6.5 weeks of age, the GBM was expanded and endothelial fenestrations could no longer be identified (Figure 3b). By 9 weeks of age, the endothelial cells were necrotic and no podocyte foot processes could be identified (Figure 3b).

Molecular marker analysis confirmed the disappearance of differentiated podocytes with a complete absence of WT1, nephrin, and VEGF-A in the majority of glomeruli of terminally ill mice (Figure 4, g and h, and data not shown). On occasion, a single cell could be identified that stained positively for these markers (Figure 4h). These markers were all present at birth and at 3 and 6 weeks of age (Figure 4, d–f and data not

shown). The level of VEGF-A mRNA was consistently lower in the heterozygous VEGF glomeruli than in control glomeruli at the same developmental stage (Figure 4d). VSMA is not normally found in 9-week-old mesangial cells unless they are “activated” in glomerular injury; in the heterozygotes, occasional VSMA-positive cells were identified in the glomeruli (Figure 4i).

VEGF-A is required in the podocyte to establish the glomerular filtration barrier. In order to investigate the phenotype resulting from a complete absence of VEGF-A in the glomerulus, mice that were null for VEGF-A specifically in the podocyte (*VEGF-loxP^{+/-},Neph-Cre^{+/-}* mice) were generated ($n = 15$). These mice were born at the expected mendelian frequency but died at birth or within 18 hours of birth. Some of these mice were born with hydrops that can be seen in infants with congenital nephrotic syndrome (22).

Light microscopy demonstrated that all of the null VEGF-A glomeruli were small with no or few distin-

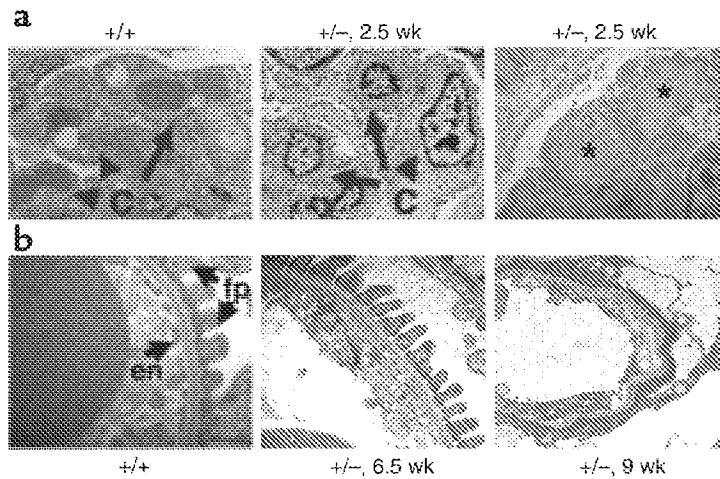


Figure 3

Heterozygous *VEGF-loxP^{+/-},Neph-Cre^{+/-}* mice demonstrate endotheliosis and loss of fenestrations. (a) At 2.5 weeks of age, wild-type glomerular capillary loops (c) are open and contain numerous red blood cells. In contrast, podocyte-specific VEGF-A heterozygotes (+/-) demonstrate bloodless glomeruli, and the capillary loops are filled with swollen endothelial cells, demonstrating endotheliosis, the classic renal lesion of preeclampsia. In addition, large subendothelial hyaline deposits (*) can be seen. (b) At 6.5 weeks of age, wild-type filtration barriers (+/+) are characterized by fenestrated endothelial cells (en) and well-formed podocyte foot processes (fp). In the podocyte-specific heterozygotes (+/-), the fenestrations are lost at 6.5 weeks of age, and by 9 weeks of age, the endothelial cells appear necrotic and no podocyte foot processes can be identified.

guishable glomerular capillary loops. Additionally, podocytes were present but tended to pile up in several layers and lacked well-formed slit diaphragms, the specialized intercellular junctions found between foot processes (Figure 5a and data not shown).

Immunohistochemical analysis with an antibody to PECAM that recognizes a cell surface receptor on endothelial cells was performed. Although endothelial cells were present in most immature glomeruli, they were markedly reduced in number and mature

glomeruli lacked endothelial cells altogether (Figure 5b). BrdU labeling was performed; labeled endothelial cells were easily identified in the vascular clefts of wild-type S-shape stage glomeruli but were never observed in podocyte-specific VEGF-A null glomeruli (data not shown).

EM studies demonstrated widespread but not complete effacement of podocyte foot processes (not shown). Small capillary loops with a GBM could be identified in some glomeruli. However, this GBM failed

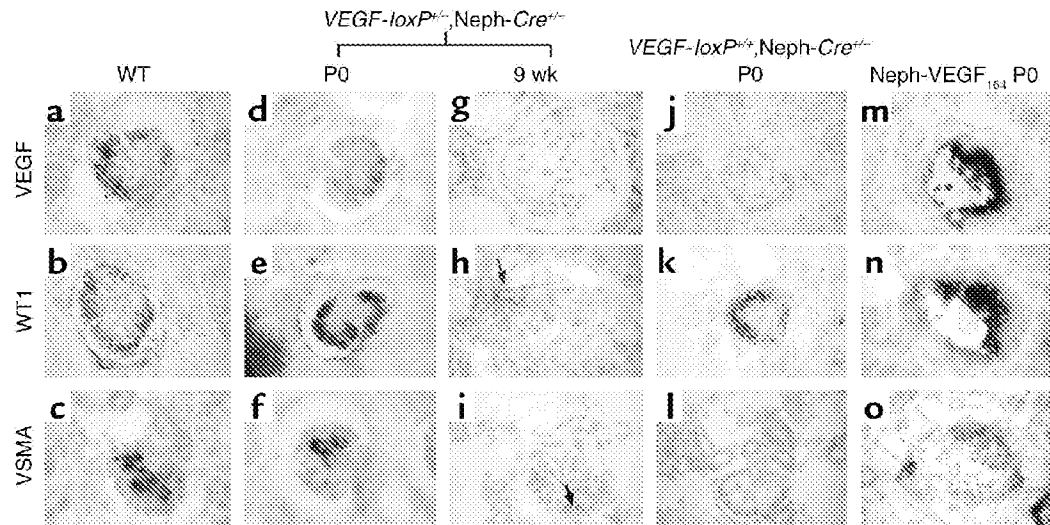


Figure 4

Digoxigenin-labeled in situ analysis of wild-type and mutant glomeruli. (a–c) Capillary loop-stage glomeruli from a newborn *VEGF-loxP^{+/-},Neph-Cre^{+/-}* control mouse demonstrate expression of VEGF-A and WT1 in podocytes, while VSMA is expressed in mesangial cells, which are found inside the glomerulus and are required to support the capillary structure. (d–f) At birth (P0), capillary loop-stage glomeruli from a heterozygous *VEGF-loxP^{+/-},Neph-Cre^{+/-}* mouse demonstrate normal levels of expression of WT1 and VSMA, while VEGF expression is consistently reduced at the mRNA level compared with the wild-type controls. (g–i) By 9 weeks of age, the heterozygous VEGF mice are clinically unwell. At this time, most glomeruli demonstrate a complete absence of markers of podocyte differentiation (i.e., no VEGF or WT1; both are absent). In h, a single WT1-positive cell can be identified (arrow). (i) VSMA is not usually present in glomeruli at 9 weeks; however, occasional VSMA-positive cells can also be identified and likely represent “activated” mesangial cells (arrow). (j–l) In the null *VEGF-loxP^{-/-},Neph-Cre^{+/-}* glomeruli at birth, no VEGF is seen in glomeruli as predicted due to podocyte-specific excision of both VEGF alleles. WT1 is present in differentiated podocytes. In contrast, VSMA is absent, demonstrating a defect in migration and/or differentiation of mesangial cells into the glomerulus. (m–o) In the nephrin-VEGF-164 mouse, both VEGF and WT1 are expressed in podocytes present within collapsed glomeruli. VEGF is markedly upregulated. In addition, VSMA and mesangial cells are present but appear to surround the collapsed glomerulus in a crescent shape. Magnification: $\times 350$.

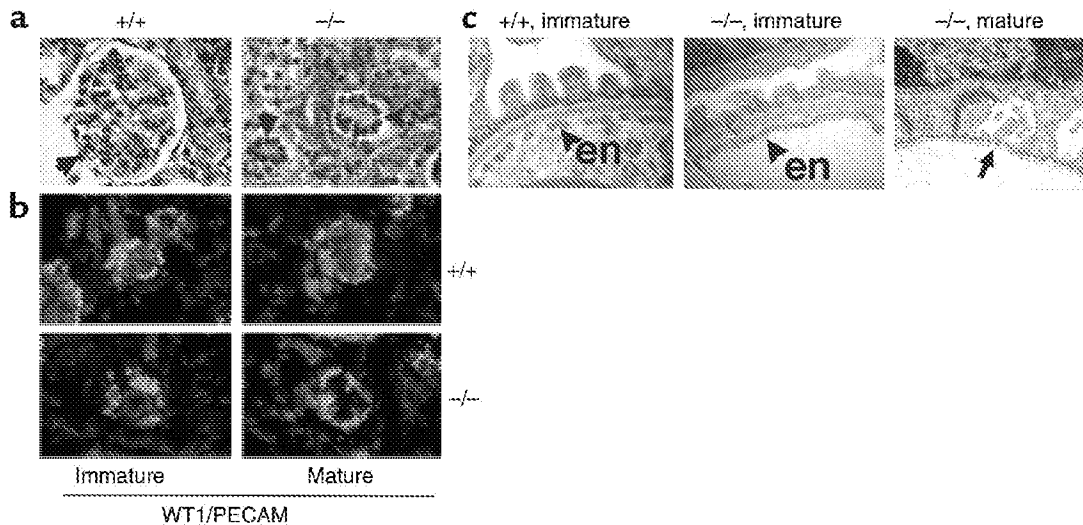


Figure 5

VEGF-null glomeruli do not form filtration barriers or fenestrations within endothelial cells. (a) The wild-type (+/+) glomerulus (arrow) has a lacy appearance due to open capillary loops. The VEGF-null glomeruli (-/-) fail to develop fully and lack visible capillary loops. Magnification: $\times 350$. (b) Immunohistochemical staining for WT1 (green), a marker for podocyte cells, and PECAM (red), a marker for endothelial cells, shows a reduced number of endothelial cells in immature (capillary loop-stage) VEGF-null glomeruli. In mature glomeruli, no endothelial cells remain. Magnification: $\times 300$. (c) Transmission EM of the filtration barrier in a wild-type (+/+) glomerulus clearly demonstrates fenestrated endothelium at the late capillary-loop stage, whereas no fenestrations are observed in endothelial cells (en) found in corresponding late capillary loop-stage VEGF-null glomeruli. In mature VEGF-null glomeruli, the basement membrane is seen (arrow), but the endothelial cells are missing. Magnification: $\times 20,000$.

to fuse (data not shown). Endothelial cells were seen only rarely in capillary loop glomeruli and always lacked fenestrations (Figure 5c). In contrast, fenestrations were easily observed in endothelial cells of capillary loop-stage wild-type glomeruli (Figure 5c). Capillary loops in fully differentiated (mature) glomeruli demonstrated an absence of endothelial cells (Figure 5c).

In situ analysis showed that the podocytes that were present expressed markers of differentiation appropriately, including WT1 (Figure 4k), nephrin, and podocin (data not shown), although VEGF-A was absent due to genomic excision of the VEGF gene (Figure 4j). Of note, VSMA, a marker for mesangial cells, was absent from mutant glomeruli, although some desmin staining could be identified (Figure 4l and data not shown).

Upregulation of VEGF-A in podocytes leads to a collapsing glomerulopathy and death at 5 days of age. Given the distinct phenotypes observed when the dosage of VEGF is reduced by excising one or both alleles from the podocyte, we next sought to determine the effect of increasing the level of VEGF within the podocyte and its effect on the adjacent endothelium. Transgenic founder lines that overexpressed the 164 isoform of VEGF-A (nephrin-VEGF-164) under regulation of a 4.125-kb podocyte-specific promoter from the murine nephrin gene (Figure 1e) were identified by Southern blot analysis (Figure 1f). Two independent founder mice were used for analysis. By dot blot analysis, each of these founder mice demonstrated a 30-fold increase in the VEGF copy number (Figure 1g).

The transgenic mice appeared normal at birth but became growth-retarded within 2 days. By 5 days of age,

the mice were clinically unwell and demonstrated albuminuria by dipstick analysis.

Grossly, the kidneys appeared normal in size, were hyperemic, and demonstrated cortical hemorrhages (Figure 6, a and b). Light microscopy showed global collapse of the glomerular tuft and dilation of proximal tubules that were packed with protein (Figure 6d and data not shown). Complete collapse of the capillary loops was illustrated by silver methenamine staining that recognizes the GBM (Figure 6f).

The few visible patent capillary loops were larger in diameter (Figure 6h) and multiple endothelial cell nuclei were visible within them (Figure 6, h and j) that were not seen in wild-type glomeruli (Figure 6, g and i). Although multiple endothelial cell nuclei could be identified within the few remaining patent glomerular capillary loops by EM, virtually all of the loops were collapsed and no endothelial cells could be identified. In addition, podocytes were abnormal and could be seen detaching from the GBM (data not shown).

In situ analysis confirmed that the majority of cells within the collapsed tufts were podocytes that continued to express WT1 (Figure 4n) and nephrin (not shown) and very high levels of VEGF-A that were upregulated five- to tenfold (Figure 4m and data not shown). Although mesangial cells were present as indicated by the presence of VSMA (Figure 4o), they were situated in a crescent shape at the periphery of the glomerulus.

Discussion

VEGF-A is a critical mediator of angiogenesis and vasculogenesis (11); both heterozygous and homozygous

knockout mice die during embryogenesis due to major vascular defects. This demonstrates a dosage sensitivity for VEGF during development in the whole embryo (11). Other studies have shown that VEGF-A is required for the establishment and maintenance of endothelial fenestrae in vitro (23, 24).

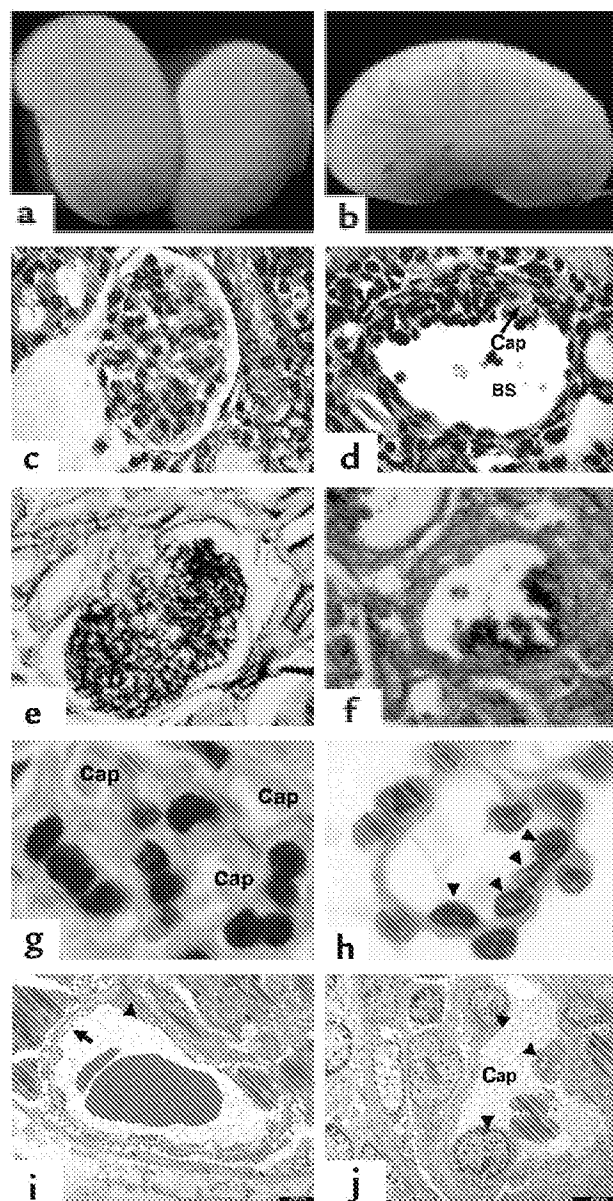
Given the expression pattern of VEGF-A in developing and mature podocytes, which are located in direct apposition to fenestrated endothelial cells, and the fact that VEGF-A expression is associated with a variety of renal diseases, we hypothesized that VEGF-A is required in developing podocytes to establish and maintain the filtration barrier. To test this hypothesis, we used the Cre-loxP system to manipulate levels of VEGF-A expression specifically within the podocyte. The mice developed distinct haploinsufficient, null, and overexpression phenotypes. Thus, similar to vascular development in the early embryo, tight regulation of VEGF signaling is essential in the establishment of later vascular beds such as the glomerulus. In addition, sequential reduction in VEGF-A levels led to a loss of fenestrations or failure of fenestrations to form, definitively demonstrating for the first time a role for VEGF-A in maintenance of endothelial fenestrations in vivo. Finally, the distinct and dramatic renal phenotypes observed with each alteration of VEGF level suggest that VEGF signaling is pivotal in glomerular health and establish its role in the pathogenesis of glomerular disease.

Complete loss of VEGF-A in the glomerulus was 100% fatal in the perinatal period. The null *VEGF-loxP^{+/+},Neph-Cre^{+/-}* mice died within 18 hours of birth with generalized swelling (hydrops) and a failure of the glomerular filtration barrier to form. Although occasional endothelial cells were identified in most but not

all capillary loop-stage (immature) glomeruli, all of these endothelial cells lacked fenestrations. Upon glomerular maturation, no endothelial cells remained. The variability in this phenotype is likely due to the time of genomic excision of the VEGF-A floxed allele. VEGF starts to be expressed during the S-shape stage of glomerulogenesis, whereas nephrin-Cre-mediated excision takes place slightly later during the capillary loop stage (17). Thus it appears that transient expression of VEGF is sufficient to direct a reduced number of incoming endothelial cells but insufficient to maintain survival and proliferation of these cells. In addition, our results suggest that there is a threshold level required for VEGF to establish fenestrations that is not reached in VEGF-null glomeruli. VSMA, a marker of glomerular mesangial cells during glomerular development, was also absent from null glomeruli, demonstrating that mesangial cell differentiation and/or

Figure 6

Mice that overexpress the 164 isoform of VEGF-A in their podocytes develop collapsing glomerulopathy. (a and b) Whole-mount images of VEGF-overexpressing kidneys at 5 days. The kidneys demonstrate many surface hemorrhages. (c) A glomerulus stained with H&E from a wild-type littermate. (d) A glomerulus from a transgenic VEGF-overexpressing mouse demonstrates global collapse of the capillary tuft toward the vascular pole of the glomerulus. A single patent capillary loop that appears dilated is identified (Cap). In addition, Bowman's space (BS) is enlarged. (e) A 5-day-old wild-type glomerulus is stained with silver methenamine that recognizes basement membranes (black). Note the intricate pattern of GBM that lines the capillary loops between endothelial cells and podocytes. (f) In contrast, a transgenic glomerulus demonstrates complete collapse of the capillary network. (g) A high-power view of the capillary loops (Cap) in a wild-type glomerulus. (h) In contrast, the few patent capillary loops identified at 5 days of age in the transgenic mice demonstrate increased diameter and multiple endothelial cell nuclei (arrowheads). (i) A wild-type capillary loop at 5 days of age. Note the fenestrated endothelium (arrow). Although a portion of an endothelial cell body is identified (arrowhead), glomerular endothelial cell nuclei are difficult to find on EM sections. (j) In a transgenic patent capillary loop at 5 days of age, three endothelial cell nuclei are easily identified (arrowheads). Magnification in a and b: $\times 60$; in c-f: $\times 225$; in g and h: $\times 1,000$. In i, bar = 2,000 nm; in j, bar = 5,000 nm.



migration is dependent upon successful establishment of a glomerular capillary system.

In contrast, initial development of the glomerular filtration barrier was unaffected in the heterozygous *VEGF-loxP⁺,Neph-Cre^{-/-}* mice. However, by 2.5 weeks of age, marked swelling of the glomerular endothelial cells led to the appearance of bloodless glomeruli and endotheliosis, the pathognomonic lesion seen in preeclampsia. Although preeclampsia is a common and potentially fatal disease that affects 7–8% of all pregnancies, the pathogenesis of this disorder is poorly understood. Patients typically develop proteinuria, and renal biopsies performed early in the disease demonstrate endotheliosis that progresses to glomerulosclerosis in a subset of patients (21). By 9 weeks of age, all of the podocyte-specific VEGF heterozygotes developed end-stage kidney failure due to a severe form of glomerulosclerosis with loss of differentiated podocytes and endothelial cells. Although alterations in circulating levels of VEGF have been implicated in preeclampsia (25), the significance of these changes is unknown and has not been studied in tissues of affected organs such as the kidney. Our results suggest that downregulation of VEGF signaling within the glomerulus may be involved in the renal lesion of preeclampsia. Because the primary defect in preeclampsia is believed to lie in the placenta and/or trophoblast, it is interesting to speculate that some as-yet-unidentified factor that is generated by the placenta leads to downregulation of VEGF-A expression within the glomerulus and endotheliosis, and suggests future areas of potential investigation.

Although glomerular defects were not observed prior to 2.5 weeks of age, it is quite possible that earlier endothelial and/or podocyte defects exist that we were unable to detect. Furthermore, the dramatic loss of podocytes by 9 weeks of age suggests that upon stimulation with VEGF-A, endothelial cells “signal back” to the podocyte, and that endothelial cell damage disrupts these reciprocal signals, emphasizing the importance and dependence of reciprocal interactions between these two cell types.

In addition to its paracrine role in the glomerulus, it is possible that VEGF-A has an autocrine function that is required for podocyte survival. Presently, it is controversial whether Flk1 is expressed even at low levels in podocytes. However, we have crossed our nephrin-Cre recombinase mice with floxed Flk1 mice (a kind gift of J. Rossant’s lab at The Samuel Lunenfeld Research Institute). By 4 weeks of age, these mice have no phenotype that demonstrates an absence of Flk1-dependent autocrine signaling within the podocyte. VEGF-A is required for breast cancer cell survival *in vitro*; in this setting, VEGF-A appears to signal in an autocrine fashion through the VEGF coreceptor, neuropilin-1, in the absence of Flk1 (26). As neuropilin-1 is expressed in the podocyte, additional studies that target neuropilin-1 in the podocyte are required to definitively answer this question.

Previous studies that have globally reduced the expression of VEGF-A in the mouse by using neutralizing antibodies (16, 27) or expressing only the 120 isoform of VEGF-A (28) have reported glomerular defects that are different from those seen in our study. The intraperitoneal injection of neutralizing antibodies to human recombinant VEGF in postnatal day 1–3 mice led to mesangiolysis and an arrest in postnatal kidney development. Similarly, the postnatal administration of a soluble chimeric VEGF receptor (Flt1) led to hypocellular glomeruli with mesangial deposits and mesangial cell vacuolization (16). In addition, the authors observed a decrease in the number of glomerular capillaries and fewer endothelial cell fenestrations (16). In the developing kidney, VEGF-A is expressed both in podocytes and in tubular epithelial cells and adjacent metanephric mesenchymal cells (29). The differences seen between previous studies and the present one are most likely due to alteration of VEGF levels in multiple cell populations within the kidney and to a variable reduction of the VEGF dose, which may be more difficult to control with a circulating antibody or receptor. In addition, endogenous VEGF-A levels were upregulated in podocytes in a study by Kitamoto et al. (27). In our model, there is a complete absence of VEGF in podocytes. The glomerular phenotype was more severe in our null mice than in mice treated with blocking antibodies or the soluble Flt receptor, suggesting that the localized delivery of VEGF from the podocyte across the heparan sulfate-rich GBM to the VEGF receptors that face the GBM (30) is critical for its function *in vivo*.

More recently, Carmeliet and colleagues have reported that mice that express only the secreted 120 isoform of VEGF-A develop glomerulosclerosis by 6 weeks of age (28). Although endothelial cells are lost, the podocytes are reportedly normal. In our experiments, all isoforms of VEGF-A are lost from the kidney and the phenotype differs from the VEGF-120 mice. Together, these results clearly emphasize the importance of isoform-specific functions of VEGF-A within the glomerulus. Genomic targeting experiments that will address the role of the different VEGF isoforms within the podocyte are underway.

Given the exquisite sensitivity to VEGF dosage reduction in glomerular development and function, we also sought to determine the phenotype resulting from overexpression of the 164 isoform of VEGF-A specifically in the podocyte. The 164 isoform is secreted and bound by heparan sulfate in the GBM. These transgenic mice developed a dramatic glomerular phenotype and rapidly succumbed to end-stage renal failure. At the time of death, the majority of their glomeruli demonstrated global collapse of the tuft as seen in collapsing glomerulopathy and HIVAN (14). Why do the capillaries collapse? At birth and from 1–5 days of age, the glomeruli are present and filter urine. At this time, the patent glomerular capillary loops have greater diameters and a greater number of

endothelial cell nuclei than do wild-type capillaries. Other studies have shown that treatment of endothelial cells with increasing doses of VEGF-A leads to coalescence of endothelial cells and the formation of larger endothelial tubes, a process that has been termed “hyperfusion” (31, 32). In the absence of increased glomerular capillary flow, this would lead to a fall in intraluminal capillary pressure and collapse. Of clinical relevance, the tat protein from HIV has been shown to signal through Flk1 in endothelial cells in Kaposi sarcoma (33–35), and the podocyte has been identified as a reservoir for the HIV virus (36). Taken together, these results present a possible explanation for the similarity between the capillary collapse seen in the VEGF-A overexpression model and HIVAN.

It has been hypothesized that damage to the podocyte ultimately leads to the capillary collapse seen in HIVAN and other forms of collapsing glomerulopathies (37–40). However, our results demonstrate that capillary collapse can occur in the absence of dedifferentiation or dysregulation of podocytes. In fact, the capillaries also collapse in heterozygous *VEGF-loxP^{-/-},Neph-Cre^{+/-}* mice after the endothelial cells are lost, and in this case, the differentiated podocytes are lost. Thus, it is evident that a single mechanism or phenotype cannot explain all cases of capillary collapse in glomerular disease.

In summary, our results demonstrate an exquisite dosage sensitivity for VEGF-A in the developing glomerulus. Numerous clinical studies have documented that alterations in glomerular VEGF-A expression are associated with glomerular disease (41–44). Our results demonstrate that dysregulation of VEGF-A is not only associated with but also plays a pathogenic role in initiating glomerular injury. The Cre-loxP system and transgenic approach allowed us to engineer mice with three different doses of VEGF within the podocyte based on the allele copy number. Each VEGF level was associated with a distinct mechanism that led to one of three important glomerular phenotypes. These results provide insight into the molecular mechanisms that underlie a variety of common and clinically important human diseases, including preeclampsia and HIV, and suggest potential future avenues for therapeutic intervention. In addition, it is clear that interactions between podocytes and endothelial cells are critical during development of the glomerular filtration barrier and continue in the filtering glomerulus.

Finally, these results provide a note of caution for clinical trials aimed at altering VEGF levels. Although the podocyte has not been specifically targeted in these therapies, careful monitoring of renal function with a particular emphasis on the glomerular filtration barrier should be included in the clinical protocols.

Acknowledgments

We gratefully acknowledge the technical support of Lois Schwartz for generation of transgenic mice, Doug Holmyard for EM processing and images, the Toronto

Centre for Comparative Models of Human Disease for help with biochemical analysis of the mice, and Dragana Vukasovic for excellent secretarial assistance. We also thank Janet Rossant and Jordan Kreidberg for critically reviewing the manuscript and Wilhelm Kriz for invaluable assistance. S.E. Quaggin is the recipient of a Canada Research Chair. This work was funded by NIH grant 5 R21 DK-59148-02 and a Kidney Foundation of Canada Grant (to S.E. Quaggin). This work was inspired by the courage of Kelly Kalt (1983–2000).

- Mundel, P., and Reiser, J. 1997. New aspects of podocyte cell biology. *Kidney Blood Press. Res.* **20**:173–176.
- Abrahamson, D.R. 1991. Glomerulogenesis in the developing kidney. *Semin. Nephrol.* **11**:375–389.
- Kriz, W., and Lemley, K. 1999. The role of the podocyte in glomerulosclerosis. *Curr. Opin. Nephrol. Hypertens.* **8**:489–497.
- Kriz, W. 1997. Evolving role of the podocyte in chronic renal failure. *Kidney Blood Press. Res.* **20**:180–183.
- Shirato, I., et al. 1996. The development of focal segmental glomerulosclerosis in masugi nephritis is based on progressive podocyte damage. *Virchows Arch.* **429**:255–273.
- Coimbra, T.M., et al. 2000. Early events leading to renal injury in obese Zucker (fatty) rats with type II diabetes. *Kidney Int.* **57**:167–182.
- Floege, J., et al. 1997. Age-related glomerulosclerosis and interstitial fibrosis in Milan normotensive rats: a podocyte disease. *Kidney Int.* **51**:230–243.
- Pagralunan, M.E., et al. 1997. Podocyte loss and progressive glomerular injury in type II diabetes. *J. Clin. Invest.* **99**:342–348.
- Robert, B., Zhao, X., and Abrahamson, D.R. 2000. Coexpression of neuropilin-1, flk1, and VEGF(164) in developing and mature mouse kidney glomeruli. *Am. J. Physiol. Renal Physiol.* **279**:F275–F282.
- Saxen, L., and Sariola, H. 1987. Organogenesis of the kidney. *Pediatr. Nephrol.* **1**:385–392.
- Carmeliet, P., et al. 1996. Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. *Nature.* **380**:435–439.
- Ferrara, N., et al. 1996. Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature.* **380**:439–442.
- Somlo, S., and Mundel, P. 2000. Getting a foothold in nephrotic syndrome. *Nat. Genet.* **24**:333–335.
- Laurinavicius, A., Hurwitz, S., and Rennke, H.G. 1999. Collapsing glomerulopathy in HIV and non-HIV patients: a clinicopathological and follow-up study. *Kidney Int.* **56**:2203–2213.
- Novak, A., Guo, C., Yang, W., Nagy, A., and Lobe, C.G. 2000. Z/EG, a double reporter mouse line that expresses enhanced green fluorescent protein upon Cre-mediated excision. *Genesis.* **28**:147–155.
- Gerber, H.P., et al. 1999. VEGF is required for growth and survival in neonatal mice. *Development.* **126**:1149–1159.
- Eremina, V., Wong, M.A., Cui, S., Schwartz, L., and Quaggin, S.E. 2002. Glomerular-specific gene excision in vivo. *J. Am. Soc. Nephrol.* **13**:788–793.
- Laemmli, U.K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature.* **227**:680–685.
- Wong, M.A., Cui, S., and Quaggin, S.E. 2000. Identification and characterization of a glomerular-specific promoter from the human nephrin gene. *Am. J. Physiol. Renal Physiol.* **279**:F1027–F1032.
- Míner, J.H., and Li, C. 2000. Defective glomerulogenesis in the absence of laminin alpha5 demonstrates a developmental role for the kidney glomerular basement membrane. *Dev. Biol.* **217**:278–289.
- Kincaid-Smith, P. 1991. The renal lesion of preeclampsia revisited. *Am. J. Kidney Dis.* **17**:144–148.
- McDonald, R., Wiggelinkhuizen, J., and Kaschula, R.O. 1971. The nephrotic syndrome in very young infants. *Am. J. Dis. Child.* **122**:507–512.
- Esser, S., et al. 1998. Vascular endothelial growth factor induces endothelial fenestrations in vitro. *J. Cell Biol.* **140**:947–959.
- Roberts, W.G., and Palade, G.E. 1995. Increased microvascular permeability and endothelial fenestration induced by vascular endothelial growth factor. *J. Cell Sci.* **108**:2369–2379.
- Bielecki, D.A., Klonowska-Dziatkiewicz, E., Jarocki, S., and Urban, J. 2002. Growth factors in pregnancy complications with preeclampsia. *Ginekol Pol.* **73**:422–429.
- Bachelder, R.E., et al. 2001. Vascular endothelial growth factor is an autocrine survival factor for neuropilin-expressing breast carcinoma cells. *Cancer Res.* **61**:5736–5740.
- Kitamoto, Y., Tokunaga, H., and Tomita, K. 1997. Vascular endothelial growth factor is an essential molecule for mouse kidney development: glomerulogenesis and nephrogenesis. *J. Clin. Invest.* **99**:2351–2357.

28. Mattot, V., et al. 2002. Loss of the VEGF(164) and VEGF(188) isoforms impairs postnatal glomerular angiogenesis and renal arteriogenesis in mice. *J. Am. Soc. Nephrol.* **13**:1548-1560.
29. Miquerol, L., Gertsenstein, M., Harpal, K., Rossant, J., and Nagy, A. 1999. Multiple developmental roles of VEGF suggested by a LacZ-tagged allele. *Dev. Biol.* **212**:307-322.
30. Gengrinovitch, S., et al. 1999. Glypican-1 is a VEGF165 binding proteoglycan that acts as an extracellular chaperone for VEGF165. *J. Biol. Chem.* **274**:10816-10822.
31. Drake, C.J., and Little, C.D. 1995. Exogenous vascular endothelial growth factor induces malformed and hyperfused vessels during embryonic neovascularization. *Proc. Natl. Acad. Sci. U. S. A.* **92**:7657-7661.
32. Drake, C.J., and Little, C.D. 1999. VEGF and vascular fusion: implications for normal and pathological vessels. *J. Histochem. Cytochem.* **47**:1351-1356.
33. Morini, M., et al. 2000. Kaposi's sarcoma cells of different etiologic origins respond to HIV-Tat through the Flk-1/KDR (VEGFR-2): relevance in AIDS-KS pathology. *Biochem. Biophys. Res. Commun.* **273**:267-271.
34. Ganju, R.K., et al. 1998. Human immunodeficiency virus tat modulates the Flk-1/KDR receptor, mitogen-activated protein kinases, and components of focal adhesion in Kaposi's sarcoma cells. *J. Virol.* **72**:6131-6137.
35. Vene, R., Benelli, R., Noonan, D.M., and Albini, A. 2000. HIV-Tat dependent chemotaxis and invasion, key aspects of tat mediated pathogenesis. *Clin. Exp. Metastasis.* **18**:533-538.
36. Marras, D., et al. 2002. Replication and compartmentalization of HIV-1 in kidney epithelium of patients with HIV-associated nephropathy. *Nat. Med.* **8**:522-526.
37. Barisoni, L., Kriz, W., Mundel, P., and D'Agati, V. 1999. The dysregulated podocyte phenotype: a novel concept in the pathogenesis of collapsing idiopathic focal segmental glomerulosclerosis and HIV-associated nephropathy. *J. Am. Soc. Nephrol.* **10**:51-61.
38. Bruggeman, L.A., et al. 1997. Nephropathy in human immunodeficiency virus-1 transgenic mice is due to renal transgene expression. *J. Clin. Invest.* **100**:84-92.
39. Conaldi, P.G., et al. 2002. Human immunodeficiency virus-1 tat induces hyperproliferation and dysregulation of renal glomerular epithelial cells. *Am. J. Pathol.* **161**:53-61.
40. Husain, M., et al. 2002. HIV-1 Nef induces proliferation and anchorage-independent growth in podocytes. *J. Am. Soc. Nephrol.* **13**:1806-1815.
41. Shahbazi, M., et al. 2002. Vascular endothelial growth factor gene polymorphisms are associated with acute renal allograft rejection. *J. Am. Soc. Nephrol.* **13**:260-264.
42. Kim, Y.G., et al. 2000. Vascular endothelial growth factor accelerates renal recovery in experimental thrombotic microangiopathy. *Kidney Int.* **58**:2390-2399.
43. Cha, D.R., et al. 2000. Role of vascular endothelial growth factor in diabetic nephropathy. *Kidney Int. Suppl.* **77**:S104-S112.
44. Ostendorf, T., et al. 1999. VEGF(165) mediates glomerular endothelial repair. *J. Clin. Invest.* **104**:913-923.

Structure, Expression and Receptor-Binding Properties of Novel Vascular Endothelial Growth Factors

U. ERIKSSON¹ and K. ALITALO²

1 Introduction	41
2 Identification and Properties of VEGF-B/VRF.	42
3 Identification and Properties of VEGF-C/VRP.	45
4 Identification and Properties of VEGF-D/FIGF	51
5 Primary Structures of VEGF-Related Growth Factors.	52
6 Perspectives	54
References.	55

1 Introduction

Vascular endothelial growth factor (VEGF), an important regulator of endothelial cell physiology, was identified some 10 years ago and has, since then, been recognised as the major growth factor relatively specific for endothelial cells (reviewed in FERRARA and DAVIS-SMYTH 1997). VEGF is a dimeric glycoprotein, closely related to placenta growth factor (PlGF). Both VEGF and PlGF are distantly related in structure to the platelet-derived growth factors A and B (PDGF A and PDGF B) (HELDIN et al. 1993). Three novel growth factors belonging to the family of VEGF, PlGF and the two PDGFs were recently discovered. These growth factors, termed vascular endothelial growth factor B/VEGF-related factor (VEGF-B/VRF) (GRIMMOND et al. 1996; OLOFSSON et al. 1996a), vascular endothelial growth factor C/VEGF-related protein (VEGF-C/VRP) (JOUKOV et al. 1996; LEE et al. 1996) and *c-fos*-induced growth factor (FIGF) (ORLANDINI et al. 1996) share structural features typical of the VEGF/PDGF growth factor family. The prominent structural similarities between VEGF-related growth factors, several of which target endothelial cells, and FIGF suggest the possibility that FIGF also targets endothelial cells, despite its identification as a fibroblast growth factor. Based on these criteria,

¹Ludwig Institute for Cancer Research, Stockholm Branch, Box 240, S-171 77 Stockholm, Sweden

²Molecular/Cancer Biology Laboratory, Haartman Institute, PL21 (Haartmaninkatu 3), FIN-00014 University of Helsinki, Finland

we propose that the name FIGF should be changed to VEGF-D to indicate its structural and functional relatedness to the other VEGFs.

The rapidly expanding list of growth factors belonging to the VEGF-family is surprising, but underscores the complexity of regulation of endothelial cell functions and the heterogeneity among different subpopulations of endothelial cells. In this review, we will summarise known structural and functional properties of the novel VEGFs, i.e. VEGF-B, VEGF-C and VEGF-D.

2 Identification and Properties of VEGF-B/VRF

A serendipitously found partial, mouse complementary deoxyribonucleic acid (cDNA) clone, encoding a VEGF-related peptide, was used to isolate full-length mouse and human cDNA clones from an adult mouse-heart cDNA library and from a human tumour cell cDNA library, respectively (OLOFSSON et al. 1996a). The full-length cDNAs encoded a homologue of VEGF and, in analogy with the nomenclature of the PDGFs, the new protein was denoted VEGF-B. Independently, another group of researchers found the same gene when attempting to identify the gene for multiple endocrine neoplasia type 1 (MEN1). The protein encoded by this gene was designated VEGF-related factor (VRF, GRIMMOND et al. 1996).

The mouse and human genes for VEGF-B are almost identical, and both span about 4 kb of DNA. The genes are composed of seven exons and their exon-intron organisation resembles that of the VEGF and PlGF genes (Fig. 1) (GRIMMOND et al. 1996; OLOFSSON et al. 1996b; TOWNSON et al. 1996). Presently, two isoforms of VEGF-B, generated by alternative splicing of mRNA, have been recognised (GRIMMOND et al. 1996; OLOFSSON et al. 1996b; TOWNSON et al. 1996). These two secreted forms of VEGF-B have 167 (VEGF-B₁₆₇) and 186 (VEGF-B₁₈₆) amino acid residues, respectively. The isoforms have an identical *N*-terminal domain of 115 amino acid residues, excluding the signal sequence, while the *C*-terminal domains differ. The common *N*-terminal domain is encoded by exons 1–5. Differential use of the remaining three exons gives rise to the two splice isoforms. By the use of an alternative splice-acceptor site in exon 6, an insertion of 101 bp introduces a frame shift and a stop of the coding region of VEGF-B₁₆₇ cDNA (see Fig. 1). Thus, the two VEGF-B isoforms will have different *C*-terminal domains which are unrelated to each other. In VEGF and PlGF, several isoforms are encoded by the use of alternative splice-acceptor sites and different combinations of exons in the genes, but the corresponding transcripts are translated using the same reading frame. The use of partially overlapping, but different reading frames is fairly uncommon among higher eukaryotes.

The different *C*-terminal domains of the two splice isoforms of VEGF-B affect their biochemical and cell biological properties. The *C*-terminal domain of VEGF-B₁₆₇ is structurally related to the corresponding region in VEGF, with several conserved cysteine residues and stretches of basic amino acid residues (see Sect. 5).

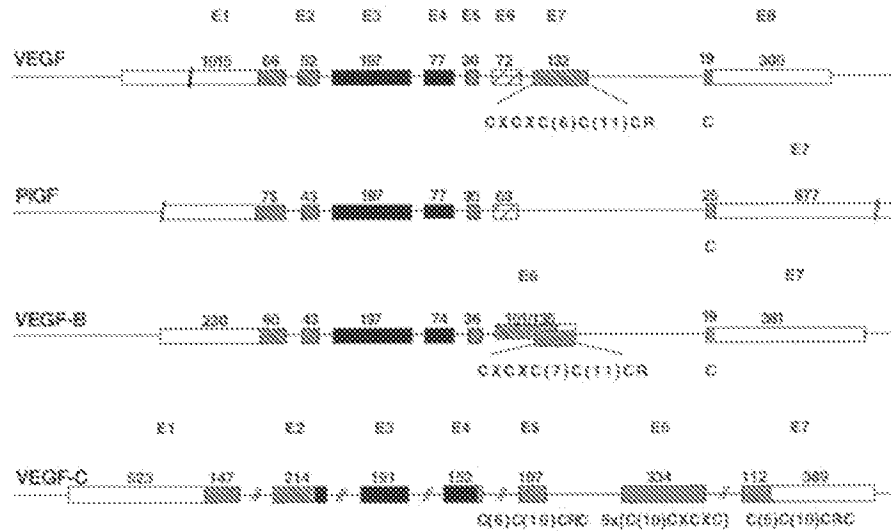


Fig. 1. Exon-intron organization of genes of the vascular endothelial growth factor (VEGF) family. The exons are shown as boxes and their lengths (bp) are indicated; the non-coding portions are white; grey boxes denote sequences encoding C- and N-terminal peptides; black boxes denote sequences encoding the VEGF homology domain. The striped box indicates the exon encoding part of the heparin-binding region. Certain cysteine motifs encoded by the different exons are shown. The structures of the genes are from TISCHER et al. 1991, OLOFSSON et al. 1996b, CHILOV et al. 1997 and DiPALMA et al. 1997. The figure was modified from CHILOV et al. 1997

Thus, this domain is highly hydrophilic and basic and, accordingly, VEGF-B₁₆₇ will remain cell-associated on secretion, unless the producing cells are treated with heparin or high salt concentrations. The cell-associated molecules binding VEGF-B₁₆₇ are likely to be cell surface or pericellular heparan sulphate proteoglycans. It is likely that the cell-association of this isoform occurs via its unique basic C-terminal region, as noted for the highly basic splice variants of VEGF. This suggestion is further supported by the observation that a fusion protein of glutathione-S-transferase and the unique C-terminal domain of VEGF-B₁₆₇ binds tightly to a heparin-Sepharose column (B. Olofsson and the authors, unpublished observation).

The C-terminal domain of the second splice isoform, VEGF-B₁₈₆, has no significant similarity with known amino acid sequences in the databases. The hydrophobic character of this domain, with several conserved alanine, proline, serine and threonine amino acid residues contrasts with the properties of the hydrophilic and basic C-terminal domain in VEGF-B₁₆₇. This is supported by the observation that VEGF-B₁₈₆ does not remain cell-associated on its secretion. Recent evidence suggests that this isoform is proteolytically processed, which regulates the biological properties of the protein (OLOFSSON et al. 1998 and unpublished data).

Isoforms of both human and mouse VEGF-B lack the consensus sequence for N-linked glycosylation (NXT/S), unlike the other growth factors of the PDGF/VEGF-family. However, VEGF-B₁₈₆ is O-glycosylated, presumably in the unique C-terminal domain rich in serine and threonine residues (OLOFSSON et al. 1996b).

The VEGF-B isoforms are produced as disulphide-linked homodimers and, under reducing conditions, the apparent molecular masses of secreted VEGF-B₁₆₇ and VEGF-B₁₈₆ isoforms are 21 kDa and 32 kDa, respectively (OLOFSSON et al. 1996a,b). The secreted 32 kDa form of VEGF-B₁₈₆ is *O*-glycosylated, while the unmodified intracellular form of VEGF-B₁₈₆ has an apparent molecular mass of 26kDa.

It is well documented that VEGF can form naturally occurring heterodimers with PlGF (DiSALVO et al. 1995) and such heterodimers might display functional properties distinct from those of both VEGF and PlGF homodimers. Analysis of both isoforms of VEGF-B showed that disulphide-linked heterodimers with VEGF are generated when both are co-expressed in recipient cells (OLOFSSON et al. 1996a,b), but it has not been established whether naturally occurring VEGF-VEGF-B heterodimers exist. Homodimers of VEGF₁₆₅ are secreted from cells in a soluble form, while heterodimers of VEGF-B₁₆₇-VEGF remain cell-associated. In contrast, heterodimers of VEGF-B₁₈₆ and VEGF are freely secreted into the cell-culture medium. Thus, VEGF-B₁₆₇ appears to determine the release of the heterodimers from cells, and heterodimerization of VEGF with either of the two isoforms of VEGF-B might, therefore, control the release and bioavailability of VEGF-VEGF-B heterodimers (OLOFSSON et al. 1996a,b). It is presently unknown whether the VEGF-B polypeptides perform their *in vivo* function as homodimers, as heterodimers with VEGF, or as both.

The ability of VEGF-B isoforms to affect the release of VEGF-VEGF-B heterodimers from the producing cells is intriguing, since the two growth factors are co-expressed in many tissues, most prominently in the heart (OLOFSSON et al. 1996a). The almost identical patterns of expression of the two VEGF-B isoforms, predominantly in embryonic and adult muscle tissues (myocardium and skeletal muscle), makes it unlikely that differential expression of VEGF-B isoforms would contribute to a genetically controlled mechanism involved in the release of VEGF-VEGF-B heterodimers.

Conditioned medium from 293 cells transfected with an expression vector generating VEGF-B₁₆₇ stimulated thymidine incorporation into DNA in human umbilical vein endothelial cells (HUVECs) and bovine capillary endothelial (BCE) cells. This suggested that VEGF-B is an endothelial cell mitogen and that it may be angiogenic *in vivo* (OLOFSSON et al. 1996a). However, the possibility remains that at least part of the mitogenic activity is contributed by VEGF-VEGF-B heterodimers, as recombinant VEGF-B₁₈₆ homodimers have no detectable mitogenic activity on endothelial cells (OLOFSSON et al. 1998).

Despite their close structural similarities, the receptor-binding properties of VEGF-B differ from those of VEGF. Using soluble VEGF receptor (VEGFR) extracellular-domain fusion proteins, we have established that VEGF-B binds to VEGFR-1 with high affinity, but not to VEGFR-2 or -3 (OLOFSSON et al. 1998). The affinity for VEGFR-1 and not VEGFR-2 is not surprising, considering that certain receptor-specific epitopes defined for VEGF (KEYT et al. 1996) predict this reactivity. Thus, the acidic residues in loop 2 of VEGF, important for binding to VEGFR-1, are almost identical in VEGF-B. However, analysis of several VEGF-B

mutants in which these acidic residues have been replaced by alanine residues show that the corresponding residues of VEGF-B has some effect for VEGFR-1 binding (OLOFSSON et al. 1998). Conversely, the basic residues in loop 3 of VEGF, important for VEGFR-2 binding, are not present in VEGF-B. The selective binding of VEGF-B and PlGF to VEGFR-1 suggests that the two growth factors may be differentially expressed functional homologues.

The expression of VEGF-B during most of murine development suggests that VEGF-B has a role during the establishment of the vascular system. Results from the knockout studies of VEGF have shown that VEGF-B is unable to compensate for the loss of even a single allele of VEGF (CARMELIET et al. 1996; FERRARA et al. 1996). Given that VEGF-B does not bind VEGFR-2, this is not surprising, and the functions of VEGF and VEGF-B are, thus, clearly distinct. Furthermore, the role of VEGF-B may extend beyond the vascular system as it is expressed early during the development of the central nervous system. VEGF-B expression was detected in 8-day-old embryos in structures most likely corresponding to parts of the neural tube (LAGERCANTZ et al. 1996). On day 11.5–12.5 p.c., VEGF-B was strongly expressed in the developing heart (OLOFSSON et al. 1996a and unpublished observations). Later, on day 14 p.c., VEGF-B is expressed in most tissues of the embryo, although most prominently in heart, spinal cord and cerebral cortex. On day 17, most of the *in situ* hybridization signal is concentrated in the heart, brown fat and spinal cord (LAGERCANTZ et al. 1996).

One of the unique features of VEGF expression is its upregulation under hypoxic conditions (GOLDBERG and SCHNEIDER 1994; STEIN et al. 1995) and by a variety of other stimuli, including several growth factors and cytokines (FINKENZELLER et al. 1992; GARRIDO et al. 1993; PERTOVAARA et al. 1994; FRANK et al. 1995; COHEN et al. 1996). The regulation of VEGF-B mRNA is apparently very different, as neither hypoxia nor several growth factors alter the level of expression of this gene (ENHOLM et al. 1997).

The VEGF-B gene was localised to chromosome 11q13, proximal to the cyclin D1 gene, which is amplified in a number of human carcinomas (PAAVONEN et al. 1996). The amplification of cyclin D1, however, was not accompanied by amplification of VEGF-B in several mammary carcinoma cell lines studied (PAAVONEN et al. 1996).

3 Identification and Properties of VEGF-C/VRP

A factor stimulating tyrosine phosphorylation of Flt4 (subsequently referred to as VEGFR-3), a receptor tyrosine kinase closely related to VEGFR-1 and VEGFR-2, was identified in conditioned medium from PC-3 prostatic adenocarcinoma cells. Receptor-affinity chromatography using the VEGFR-3 extracellular domain led to the purification of the stimulating factor. The partial amino acid sequence was obtained from the purified factor and a 5' fragment of the cDNA encoding it was

amplified by serial polymerase chain reactions (PCR) using degenerate primers. A full-length cDNA was then cloned from a library prepared from PC-3 cells, using the labelled PCR-amplified 5' fragment as a probe (JOUKOV et al. 1996). The full-length cDNA encoded a novel homologue of VEGF and was subsequently denoted VEGF-C. Independently, an expressed sequence tag (EST) was identified in the database as being homologous with VEGF. Using the partial EST clone as the probe, a full-length VEGF-C cDNA clone was isolated. The protein encoded by this cDNA was designated VEGF-related protein (VRP) (LEE et al. 1996).

The human VEGF-C cDNA encodes a protein of 419 amino acid residues, with a predicted molecular mass of 46.9 kDa. However, the newly synthesised VEGF-C product is a pre-pro-protein, consisting of an *N*-terminal signal sequence followed by an *N*-terminal peptide, the VEGF-homology domain, and a *C*-terminal pro-peptide (JOUKOV et al. 1996). VEGF-C is secreted as a disulphide bonded homodimer, and most of it is proteolytically processed from the precursor polypeptide, which contains three putative *N*-glycosylation sites; two of these remain in mature, fully processed VEGF homology domain. Based on our results, we propose the VEGF-C proteolytic processing model schematically presented in Fig. 2. This model resembles the model for the proteolytic processing of PDGF, especially of PDGF-B (ÖSTMAN et al. 1988, 1992) in that: (1) the proteolytic cleavages occur after the formation of disulphide-bonded precursor dimers, (2) both *N*- and *C*-terminal peptides may be subject to cleavage, and (3) a variety of processed forms are secreted. However, there are several important differences between PDGF-B and VEGF-C, concerning both their processing and the structures of the mature growth factors.

The homologous part of VEGF-C is about 30% identical with VEGF₁₆₅, 27% with VEGF-B₁₆₇, 25% with PIGF-1 and 22–24% with PDGF-A and PDGF-B. Fully processed VEGF-C binds to and activates both VEGFR-3 and VEGFR-2. A single class of high-affinity sites was observed in porcine aortic endothelial (PAE)/VEGFR-3 cells ($K_d = 135$ pM) and PAE/VEGFR-2 cells ($K_d = 410$ pM). These values are of similar magnitude to the affinities reported for the VEGF-VEGFR-2 interaction (TERMAN et al. 1992; WALTENBERGER et al. 1994). VEGF-C and VEGF displace each other from VEGFR-2, indicating that the same region of this receptor is involved in the binding of both ligands. Surprisingly, none of the three basic residues reported to be critical for VEGFR-2 binding by VEGF (KEYT et al. 1996) are conserved in VEGF-C. VEGF-C also dose-dependently stimulated autophosphorylation of VEGFR-3 and VEGFR-2, but in agreement with previous reports (LEE et al. 1996), we could not detect binding to VEGFR-1 (JOUKOV et al. 1996, 1997).

The human and mouse VEGF-C genes both comprise over 40 kb of genomic DNA and consist of seven exons, all containing coding sequences (Fig. 1). The VEGF-C gene was localised to human chromosome 4q34, close to the human aspartylglucosaminidase gene (PAAVONEN et al. 1996). The VEGF homology domain of VEGF-C is encoded by exons 3 and 4. Exons 5 and 7 encode cysteine-rich motifs of the type C(6)C(10)CRC, and exon 6 encodes C(10)CXCXC motifs typical of a silk protein (CHILOV et al. 1997). The upstream promoter sequences contain

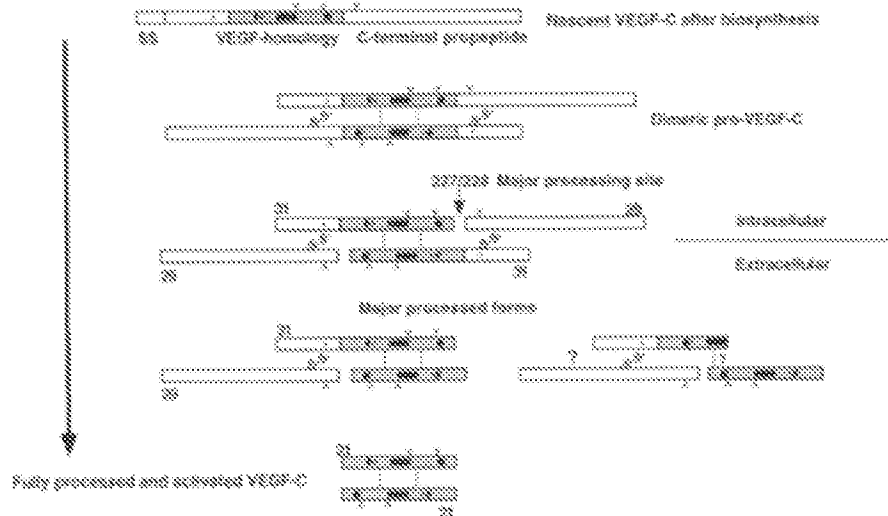


Fig. 2. Schematic model of the proteolytic processing of VEGF-C. The regions of VEGF-C polypeptide are marked as follows: *SS* signal sequence; *grey box* VEGF-homology domain; *open boxes* *N*-terminal and *C*-terminal peptides. Cysteine residues are shown as *black ovals*, the cysteine residues in the *C*-terminal pro-peptide are not marked for clarity. Putative sites of *N*-linked glycosylation are indicated by *Y*. The major proteolytic cleavage site between residues 227/228 is indicated by an *arrow*. This cleavage occurs as early as during secretion of the protein from the cellular compartment and it creates the major processed forms consisting of disulphide-linked 29/31-kDa polypeptides. Disulphide bonds are marked as *-S-S-*; the *dashed lines* indicate the detection of both covalent or non-covalent interactions. The proposed structure of the alternatively processed VEGF-C is indicated with a *question mark*. Several intermediate forms are omitted to simplify the scheme. The figure was adopted from Joukov et al. (1997)

conserved putative binding sites for Sp-1, AP-2 and nuclear factor κ B (NF- κ B) transcription factors, but no TATA box, and show serum-stimulated promoter activity when transfected into cells. The VEGF-C gene structure is, thus, assembled from exons encoding pro-peptides and distinct cysteine-rich domains in addition to the VEGF homology domain, showing both similarities and distinct differences compared with other members of the VEGF/PDGF gene family.

VEGF-C mRNA was detected in Northern-blot analyses of many embryonal and adult tissues. In adult humans, the VEGF-C mRNA is expressed most prominently in heart, placenta, ovary, small intestine and the thyroid gland. Tumour cells express, almost exclusively, a 2.4-kb mRNA form, suggesting that it corresponds to the described VEGF-C cDNA clone obtained from the PC-3 tumour cell line (Joukov et al. 1996). The identity of another 2.0-kb mRNA, hybridizing with the VEGF-C probe in analysis of many tissues remains to be determined. Two VEGF-C cDNA clones were obtained that contained 152-bp and 557-bp deletions, corresponding to exon 2 or exons 2-4, respectively (LEE et al. 1996; CHILOV et al. 1997) Due to the shift of the reading frame, which occurs 15 amino acid residues downstream of these deletions, the predicted proteins encoded by the two deleted cDNAs contain either no or only part of the core cysteine knot region similar to that in VEGF.

Also the mouse VEGF-C cDNA was cloned and shown to encode a protein of 415 amino acid residues, which is 85% identical with human VEGF-C and similarly processed (KUKK et al. 1996). In *in situ* hybridization, mouse VEGF-C mRNA was detected in 8.5-day-old embryos in the cephalic mesenchyme, along the somites, in the tail region and extraembryonally in the allantois. In embryos 12.5 days p.c., VEGF-C mRNA was particularly prominent in regions where the lymphatic vessels are generated from embryonic veins, such as perimetanephric, axillary and jugular areas. The signal was also detected between the developing vertebrae, in the lung mesenchyme, in the neck region and in the developing forehead. The developing mesenterium, which is rich in lymphatic vessels, also showed strong VEGF-C expression (KUKK et al. 1996). The distribution of the VEGFR-3 mRNA follows a somewhat similar temporal and spatial pattern (KAIPAINEN et al. 1995; KUKK et al. 1996). This suggests a paracrine mode of ligand-receptor interaction, with VEGF-C expressed in mesenchymal cells adjacent to the VEGFR-3 positive endothelia. — The juxtaposed VEGFR-3 and VEGF-C expression patterns suggest that VEGF-C functions in the formation of the venous and lymphatic vascular systems during embryogenesis. Constitutive expression of VEGF-C in adult tissues further suggests that this growth factor is also involved in the maintenance of functions of, for example, differentiated lymphatic endothelium, where VEGFR-3 is expressed (KAIPAINEN et al. 1993, 1995; KUKK et al. 1996).

VEGF-C expression was detected in embryos as early as day 7 p.c. (KUKK et al. 1996). This was striking, considering the appearance of VEGFR-3 mRNA first on day 8.5 of gestation (KAIPAINEN et al. 1995). This suggests a possible role of VEGF-C during earlier stages of embryonic development. Such a function might be exercised through the ability of VEGF-C to function as a ligand for VEGFR-2, which is expressed in presumptive progenitors of yolk-sac blood islands as early as day 7 p.c. Interestingly, VEGFR-2 is essential for the development of both haematopoietic and endothelial cell lineages (CARMELIET et al. 1996; FERRARA et al. 1996). We, therefore, investigated the effect of VEGF-C on VEGFR-2 positive cells isolated from the primitive streak of gastrulating quail embryos. VEGF binding triggers endothelial differentiation of these cells, whereas haemopoietic differentiation appears to be mediated by binding of a so-far unidentified VEGFR-2 ligand. We could show that, like VEGF (EICHMANN et al. 1997), VEGF-C also triggers endothelial differentiation of these cells, presumably via VEGFR-2 (EICHMANN et al. 1998). These results indicate that VEGF and VEGF-C can act in a redundant manner via VEGFR-2.

Our results demonstrate that proteolytic processing allows VEGF-C to bind to and activate VEGFR-2 and increases its affinity and activity towards VEGFR-3 (JOUKOV et al. 1997). The biosynthesis of VEGF-C as a precursor may prevent unwanted angiogenic effects via VEGFR-2 and allow VEGF-C to signal preferentially via VEGFR-3. In certain circumstances, proteolytic processing would release mature VEGF-C, which is able to signal via both VEGFR-3 and VEGFR-2. It is also possible that activation of both VEGFR-3 and VEGFR-2, either as homo- or as heterodimers, is necessary to elicit a complete biological response to VEGF-C. Similarly, heterodimers of VEGFR-1 and VEGFR-2 could be important for the

biological activities of VEGF. In the case of VEGF-C, proteolytic processing might provide a regulatory mechanism that provides the possibility for fine tuning of the biological functions of this growth factor.

The major secreted VEGF-C form contains the C-terminal pro-peptide, which has an unusual structure with tandemly repeated cysteine-rich motifs and is linked via disulphide bonds with the N-terminal peptide. The possible function of this, apparently in itself an inactive C-terminal half of VEGF-C, is unknown. It has domains of striking similarity to a secretory silk protein and contains short motifs homologous with the epidermal growth factor (EGF)-like domains of other secreted proteins, such as fibrillin, laminin and tenascin. All of these proteins are known to participate in protein-protein or protein-cell surface interactions. One can, thus, speculate that partially processed VEGF-C may stay associated with the extracellular matrix via its C-terminal pro-peptide (Fig. 2). Cleavage of the N-terminal pro-peptide in certain conditions by, as yet, unknown proteases would then release the active VEGF-C.

Like VEGF, VEGF-C also stimulates the migration of endothelial cells and increases vascular permeability, albeit at concentrations higher than required for VEGF (JOUKOV et al. 1996; LEE et al. 1996). About 50-fold higher concentrations of VEGF-C were required to induce the proliferation of blood vascular endothelial cells. These activities are probably mediated through VEGFR-2 activation (PARK et al. 1994; WALTENBERGER et al. 1994). The lower specific activity of VEGF-C in these assays may depend on its lower affinity for VEGFR-2 and on its inability to bind VEGFR-1, precluding the formation of VEGFR-1-VEGFR-2 heterodimers, which may be required for maximal biological responses to VEGF (WALTENBERGER et al. 1994; DiSALVO et al. 1995; CAO et al. 1996; CLAUSS et al. 1996).

In order to better understand the function of VEGF-C, *in vivo*, its cDNA was expressed via a human keratin promoter in the basal cells of stratified squamous epithelia (JELTSCH et al. 1997). Histological examination of the transgenic mice showed that the dermis was atrophic and its connective tissue was replaced by large lymphatic vessels. In ultrastructural analysis, these vessels were shown to have overlapping endothelial junctions, anchoring filaments in the vessel wall, and a discontinuous or even partially absent basement membrane. The endothelium was also characterised by positive staining with monoclonal antibodies to desmoplakins I and II, expressed in lymphatic, but not in vascular, endothelial cells (SCHMELZ et al. 1994). VEGFR-3 and VEGFR-2, and the Tie-1 endothelial-receptor tyrosine kinase mRNAs were detected in endothelial cells lining the abnormal vessels. The VEGF-C-receptor interaction in transgenic mice apparently transduced a mitogenic signal because, in contrast to littermate controls, the lymphatic endothelium of the skin from young transgenic mice showed increased DNA synthesis. In fluorescent microlymphography, a typical honeycomb-like network with similar mesh sizes was detected in both control and transgenic mice, but the diameter of the vessels was approximately twice as large in the transgenic mice. Thus, the endothelial proliferation induced by VEGF-C led to hyperplasia of the superficial lymphatic network, but did not induce the sprouting of new vessels. Also, a relatively specific lymphangiogenic response was obtained when recombinant VEGF-

C was applied to the differentiated chick chorioallantoic membrane (OH et al. 1997).

These effects of VEGF-C overexpression were unexpectedly specific, particularly as VEGF-C is also capable of binding to and activating VEGFR-2 of blood vessel endothelial cells. In vivo, the specific effects of VEGF-C on lymphatic endothelial cells may reflect a requirement for the formation of VEGFR-3-VEGFR-2 heterodimers for endothelial cell proliferation. Such possible heterodimers may help to explain how three homologous VEGFs exert partially redundant, yet strikingly specific, biological effects. Thus, VEGF-C induces specific lymphatic endothelial proliferation and hyperplasia of the lymphatic vasculature in vivo. Further studies should establish the role of VEGF-C in lymphangiomas and in tumour metastasis via the lymphatic vasculature as well as in various other disorders involving the lymphatic system and their treatment.

Both VEGF and VEGF-C are potent vascular-permeability factors. Surprisingly, we have found that the recombinant mature VEGF-C, in which Cys156 was replaced by a Ser residue, is a selective agonist of VEGFR-3 (Joukov et al. 1998). This mutant, designated Δ NAC156S, binds and activates VEGFR-3, but neither binds VEGFR-2 nor activates its autophosphorylation and downstream signalling to the ERK/MAPK pathway. Unlike VEGF-C, Δ NAC156S neither induces vascular permeability in vivo nor stimulates migration of bovine capillary endothelial cells in culture. These data point out the critical role of VEGFR-2-mediated signal transduction for the vascular permeability activity of VEGF-C, and strongly suggest that the redundancy of biological effects of VEGF and VEGF-C is caused by their ability to bind to and activate VEGFR-2. However, the possibility exists that there are additional receptors for VEGF and VEGF-C, that are responsible for vascular permeability. The Δ NAC156S mutant may provide a valuable tool for the analysis of VEGF-C effects mediated selectively via VEGFR-3. The ability of Δ NAC156 S to form homodimers also emphasises differences in the structural requirements for VEGF and VEGF-C dimerization.

Serum and its component growth factors, PDGF, EGF and transforming growth factor- β (TGF- β), and tumor promoters were found to stimulate VEGF-C, but not VEGF-B, mRNA expression (ENHOLM et al. 1997). Serum induction of VEGF-C mRNA occurred independently of protein synthesis; with a slight increase of the mRNA half-life, whereas VEGF-B mRNA was very stable. However, hypoxia, *Ras* oncoprotein and mutant p53 tumour suppressor, which are potent inducers of VEGF mRNA did not increase VEGF-B or VEGF-C mRNA levels. We have also studied the regulation of VEGF-C by angiogenic pro-inflammatory cytokines. Interleukin (IL)-1 induced a concentration- and a time-dependent increase in VEGF-C, but not in VEGF-B, mRNA steady-state levels in human lung fibroblasts, mainly due to increased transcription (RISTIMAKI et al. 1998). Tumour necrosis factor alpha (TNF α) and IL-1 also elevated VEGF-C mRNA steady-state levels, whereas the IL-1 receptor antagonist and dexamethasone inhibited the effect of IL-1. Hypoxia, which is an important inducer of VEGF expression, had no effect on VEGF-B or VEGF-C mRNA levels (ENHOLM et al. 1997). IL-1 and TNF α also stimulated the production of VEGF-C protein by the fibroblasts (RISTIMAKI et al.

1998). Our data suggest that in addition to VEGF, VEGF-C may also serve as a lymphangiogenic or angiogenic stimulus at sites of cytokine activation. In particular, these results raise the possibility that certain pro-inflammatory cytokines regulate the lymphatic vessels indirectly via VEGF-C.

4 Identification and Properties of VEGF-D/FIGF

A partial cDNA for FIGF was first isolated from a differential-display screening of murine fibroblast mRNAs from cells with or without a targeted inactivation of the *c-fos* locus (ORLANDINI et al. 1996). The full-length murine cDNA clone was found to encode a protein of 358 amino acid residues, including a hydrophobic putative signal sequence, with significant similarities to the PDGF/VEGF family of growth factors (see Sect. 5). FIGF was shown to stimulate mitosis of fibroblasts in a dose-dependent manner. However, based on the strong structural similarities to the PDGF/VEGF family of growth factors, we propose that FIGF should be renamed to VEGF-D to highlight this relationship and to get a rational nomenclature of the novel VEGFs. VEGF-D can be viewed as having a VEGF homology domain and long *N*- and *C*-terminal extensions. The fact that VEGF-D is most closely related to VEGF-C is apparent for two reasons (see Sect. 5): first, the VEGF homology domain of VEGF-D is much more closely related to that found in VEGF-C than to those of the other family members; second, of the other factors in the VEGF family, only VEGF-C has long *N*- and *C*-terminal extensions similar to those in VEGF-D. The presence of these extensions in VEGF-C and VEGF-D, thus, defines a new subfamily of the VEGFs.

The similarity between VEGF-D and VEGF-C exists also at the functional level, as receptor-binding studies demonstrated that VEGF-D and VEGF-C exhibit similar receptor specificities (ACHEN et al. 1998). This protein is likely to be processed in a similar fashion to VEGF-C (data not shown). A region of VEGF-D corresponding to the fully processed, mature VEGF-C can bind to the extracellular domain of VEGFR-2 and induce tyrosine phosphorylation of both VEGFR-2 and VEGFR-3. When expressed in insect cells, the full-length VEGF-D was not proteolytically processed; this protein was unable to activate VEGFR-3 and activation of VEGFR-2 was, at best, marginal. Given that VEGF-D can also activate VEGFR-3, it is possible that VEGF-D could be involved in the regulation of the growth and/or differentiation of lymphatic endothelium just like VEGF-C.

The notion that VEGF-D and VEGF-C may have similar biological functions is further supported by their similar expression patterns. For example, both genes are strongly expressed in heart, muscle and small intestine, whereas expression was undetectable in peripheral blood leucocytes, brain and liver (JOUKOV et al. 1996; LEE et al. 1996; ACHEN et al. 1998). Nevertheless, the expression patterns are not identical. A second VEGF-D transcript was detected only in skeletal muscle.

Like VEGF-C, human VEGF-D was also mitogenic for bovine aortic endothelial cells. This response is likely to involve VEGFR-2. Mouse VEGF-D has

previously been reported to induce proliferation and morphological alterations of cultured fibroblasts, but the receptors responsible for mediating these effects have not been identified (ORLANDINI et al. 1996). It would be of interest to determine whether or not the cultured fibroblasts used for such studies expressed VEGFR-2 or VEGFR-3. A summary of the receptor-binding properties of known members of the VEGF family of growth factors is illustrated in Fig. 3.

5 Primary Structures of VEGF-Related Growth Factors

Seven polypeptides with significant similarities to VEGF and two PDGFs have been discovered so far. A multiple amino acid sequence alignment of human VEGF, PlGF, VEGF-B, VEGF-C, VEGF-D and the two PDGFs show that a central core of the proteins is well conserved during evolution (Fig. 4A). A major part of this core region is located between the eight invariant cysteine residues, shown to be involved in inter- and intramolecular disulphide bonding of VEGF and the two PDGFs. This region is encoded by the two well-conserved exons, E3 and E4, in the corresponding genes of the VEGFs (see Fig. 1). The overall amino acid sequence identity in this region varies between 20% and 56% in pairwise comparisons of available amino acid residues. Outside this central core, the overall amino acid identities are much weaker, although individual pairs of these proteins display higher sequence similarities. The receptor binding epitopes in these group of growth factors, at least for VEGF (KEYT et al. 1996) and the two PDGFs (HELDIN et al. 1993), are confined within the central core defined by the eight invariant cysteine residues. Thus, this region defines a structural and functional minimal domain. Determination of the three dimensional structures of VEGF (MULLER

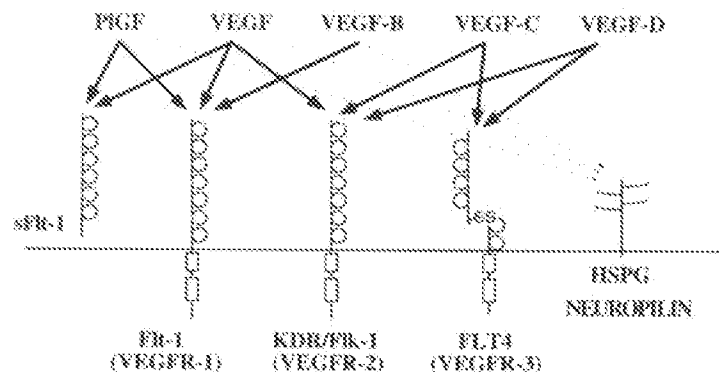


Fig. 3. Interactions of VEGF-related growth factor with their receptors. A schematic illustration of the receptor-binding characteristics of VEGF-related growth factors with soluble Flt-1 (sFlt-1), Flt-1 (*VEGFR-1*), KDR/Flk-1 (*VEGFR-2*), Flt-4 (*VEGFR-3*) and with heparin sulphate proteoglycans (*HSPG*) and neuropilin (SOKER et al. 1998)

```

VEGF-C      M H I L G F F S V A C S L L A A A L L F G P P R E A P A A A A A F E S G L D L S D 40
VEGF-D      - - - - - M Y G E W G M G N I L M M F H V Y L V Q G F R S E H G P V 29
PDGF-A      - - - - - - - - - - - - - - - - - - - - - - - - - - - M R T L A C 6
PDGF-B      - - - - - - - - - - - - - - - - - - - - - - - - - - - M N R C W A L F L S 10

VEGF-C      A E P D A G E A T A Y A S K D L E E Q L R S V S S V D E L M T V L Y F E Y W K M 80
VEGF-D      K D F S F E R S S R S M L E R S E Q Q I R A A S S L E E L L - - - Q I A H S E 65
PDGF-A      L L L L G C G Y L A H V L A E E A E I P R E V I E R L A R S Q I H S I R D L Q R 46
PDGF-B      L C C Y L R L V S A E G D P I P E E L Y E M L S D H S I R S F D D L Q R L L H G 50

VEGF 165    M N F L L S W V H W S L A L L L V L H H A K W S Q A A P M A E G G G Q N H H E V 40
PIGF-2      M P V M R L F P C F L Q L L A G L A L P A V P E Q W A L S A G N G S S E V E V 40
VEGF-B167   - - - - - M S P L L R R L L L A A L L Q L A P A Q A P V S Q P D L A P G H Q R R V 35
Pox Orf VEGF - - - - - - - - - - - - - - - M K L L V G I L V A V C L H Q Y L L N A D S N T 24
VEGF-C      Y K C Q L R E G G W Q H N R E Q A N L N S R T E E T I K F A A A H Y N T E I - L 119
VEGF-D      D W K L W R C R L K L K S L A S M D S R S A S H R S T R F A A T F Y D T E T - L 104
PDGF-A      L L E I D S V G S E D S L D T S L F A R G V H - A T K H V P E K R P L P I R 84
PDGF-B      D P - - - - G E E D G A E L D L N M T R S H S G G E L E S L A R I G R S L G S 85

VEGF 165    V K F M D V Y Q R S Y C H P I E T L V D I F O E Y P D E I E Y I F K - - P S C V 78
PIGF-2      V P F Q E V W G R S Y C R A L E R L V D V V S E Y P S E V E H M E S - - P S C V 78
VEGF-B167   V S W I D V Y T R A T C Q P R E L V V P L T V E L M G T L V A K Q L V - - P S C V 73
Pox Orf VEGF K G W S E V L K G S E C K P R P J V V P V S S T H P E L T S Q R F N - - P P C V 62
VEGF-C      K S I D N E W R K T Q C M P R E V C I D V G K E E G V A T N T F F K - - P P C V 157
VEGF-D      K V Y D E E W Q R T Q C S P R E T C V E V A S E L G K T T N T F F K - - P P C V 142
PDGF-A      K R S I E E A V P A V C K T R T V I Y E I P R S Q V D E T S A N F L I W P P C V 124
PDGF-B      L T I A E P A M I A E C K T R T E V F E I S R R L I D R T N A N E L V W P P C V 125

VEGF 165    P L M R C G G C C N D E G L R C V P T E R S N I T M Q I M R I K - P H Q G Q H F 117
PIGF-2      S L L R C T G C C G D E D L H C V P V E T A N V T M Q L L K I R - S G D I R P S Y 117
VEGF-B167   T V Q R C G G C C P D D G L E C V P T G Q H Q V R M Q I L M I R Y E - - S I Q L 111
Pox Orf VEGF T L M R C G G C C N D E S L E C V R T R E V N V S M E L L G A S G S G S N G M Q 102
VEGF-C      S V Y R C G G C C N S E G L Q C M N T S T S Y L S K T L F E I T V P L S Q G P K 197
VEGF-D      N V F R C G G C C N E E G V M C M N T S T S Y I S K Q L F E I S V E L T S V P E 182
PDGF-A      E V K R C T G C C N T S S V K C Q P S R V H H R S V K V A K V E Y V R K K F K L 164
PDGF-B      E V Q R C S G C C N N R N V Q C R R T Q V Q L R P V Q V R K I E I V R K K P I F 165

VEGF 165    G E M S P L Q H N K - - C E C R P K K - - - - - - - - - D R A R Q E N P C G F 145
PIGF-2      V E L T P S Q H V R - - C E C R P L R E - - - - - K M K F E R R P K G R G K R R 151
VEGF-B167   G E M S L E E H S Q - - C E C R P K K - - - - - D S A V K P D S P R P L C P R 144
Pox Orf VEGF R L S F V E H K K - - - C D C R P P F T T T P P T T T R P P R R R R 133
VEGF-C      P V T I S F A N H T S - C R C M S K L D - - - V Y R Q V H S I I R R S L P A T - 232
VEGF-D      L V P V K I A N H T G - C K C L L P T G P - - - - R H P Y S I I R R S I Q T P E 216
PDGF-A      K E L V Q V R L E H L E C A C A T T S L N P D Y R E E D T G R P L R E S G K K R K 204
PDGF-B      K K A T V T L E D H L A C K C E T V A A A R P V T R S F G G S Q E Q R A K K T P Q 205

VEGF 165    C S S E R R K K H L F V Q D P O T C K C S C K N T D S - L R C K A R Q L E L N E R T 184
PIGF-2      R E N Q R F T D C H L C G D A V P R R R 170
VEGF-B167   C T Q H H Q R P D P R T - - - - - C R C R C R R R S F L R C Q G R G L E L N P D T 180
VEGF-C      L P Q C Q A A N K T C P T N Y M W N N H I C R C L A Q E D F M F S S L A G D D S 272
VEGF-D      E D E C P H S K K L C P I D M L W D N T K C K C V L Q D E - T P L P G T E D H S 255
PDGF-A      R K R L K P T 211
PDGF-B      T R V T I R T V R V R R P E K G K H R K F K H T H D K T A L X E T L G A 241

VEGF 165    C R C D K P R R 192
VEGF-B167   C R C R K L R R 188
VEGF-C      T D G F H D I C G P N K E L D E B T C Q C V C R A G L R P A S C G P H K E L D R 312
VEGF-D      Y L Q E P T L C G P H M T F D E D R - - - - - - - - - - - - - - - - - - - - 273

VEGF-C      N S C Q C V C K N K I F P S Q C G A N R E F D E N T C Q C V C K R T C P R N Q P 352
VEGF-D      - - C E C V C K A P C P G D L I Q H P E N - - - - - C S C F E C K E S I E S C C 306

VEGF-C      L N P G K C A C E C T E S P Q K C L L K G K K F H H Q T C S C Y R R P C T N R Q 392
VEGF-D      Q K H K I - - - - - - - - - - - - - - - - - - - - - - - - - - - P H P D T C S C E D R - C P F H T 327

VEGF-C      K A C E P G F S Y S E E V C R C V P S Y W K R P Q M S 419
VEGF-D      R T C A S R K P A C G K H W R F P K E T R A Q G L Y S Q E N F 358

```

A

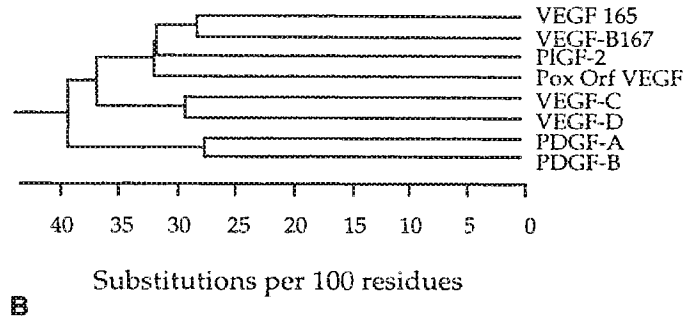


Fig. 4A, B. Amino acid sequence alignment of VEGF-related growth factors. **A.** Multiple amino acid sequence alignment of three novel VEGFs, e.g. VEGF-B, VEGF-C and VEGF-D and a comparison with VEGF, placenta growth factor (PlGF) and the two platelet-derived growth factors (PDGFs) as well as with the interesting viral homologue of the poxvirus *orf* virus (LYTTLE et al. 1994). The amino acid sequences were aligned using the Crystal algorithm, and the alignment was refined manually. The boxed residues are within two distance units using the PAM 250 matrix. **B.** An unrooted phylogenetic tree of the VEGF-related growth factors based on the amino acid sequence alignment in **A**.

et al. 1997) and PDGF-B (OEFNER et al. 1992) by X-ray crystallography have also shown that this region forms distinct domains with remarkable structural similarities.

A phylogenetic analysis of the amino acid sequences of these growth factors shows that they can be grouped into three separate subfamilies, consisting of VEGF, PlGF, VEGF-B, and VEGF-C and D, and the two PDGFs, respectively (Fig. 4B).

6 Perspectives

The discovery of three novel members of VEGF family increases our understanding of the complexity of the regulatory signals for endothelial cells and promotes new areas of research in vascular biology. Many of the already-established experimental models and approaches used in VEGF studies might obviously be applied to studies of the novel VEGFs. However, not only endothelial functions should be taken into consideration here, as recent results show that VEGF might induce certain biological effects via the targeting of non-endothelial cells (MIDY and PLOUET 1994; GABRILOVICH et al. 1996).

The main questions regarding the biological roles of the novel VEGFs are not answered yet. In this regard, different molecular genetic and transgenic approaches, including gene targeting, are of great importance. Studies on VEGF-C have shown that it acts as a specific growth factor for endothelial cells of lymphatic vessels (JELTSCH et al. 1997). Studies on VEGF-B and VEGF-D are likely to provide additional information on the role of these growth factors in endothelial cell function and physiology.

Important issues also concern the analysis of tissue-specific regulation of VEGF-B, VEGF-C and VEGF-D expression by hypoxia, various growth factors and other agents or conditions known to regulate VEGF expression. Similarly, the function of the different splicing forms of VEGF-B, VEGF-C and VEGF-D should be explored. Such alternatively spliced isoforms of these growth factors might possess different functions *in vivo*, e.g. due to the differences in their receptor specificity/affinity, bioavailability, stability and proteolytic processing, and their ability to form heterodimers with other VEGF family members. The latter property might be of particular importance, as heterodimers of the various growth factors might express biological properties distinct from those of the corresponding homodimers. Finally, the discovery of VEGF-B, VEGF-C and VEGF-D highlights the structural similarities of VEGF family polypeptides and may simplify the search for novel homologous molecules.

Acknowledgements. We kindly thank all of our coworkers and collaborators for outstanding contributions to the basic findings summarized in the review and the funding agencies for continued support.

References

- Achen MG, Jeltsch M, Kukk E, Makinen T, Vitali A, Wilks AF, Alitalo K, Stacker SA (1998) Vascular endothelial growth factor-D (Vegf-D) is a ligand for the tyrosine kinases Vegf receptor 2 (Flk1) and Vegf receptor 3 (Flt4). *Proc Natl Acad Sci USA* 95:548–553
- Cao Y, Chen H, Zhou L, Chiang MK, Anand-Apte B, Weatherbee JA, Wang Y, Fang F, Flanagan JG, Tsang ML (1996) Heterodimers of placenta growth factor/vascular endothelial growth factor. Endothelial activity, tumor cell expression, and high affinity binding to Flk-1/KDR. *J Biol Chem* 271:3154–3162
- Carmeliet P, Ferreira V, Breier G, Pollefeyt S, Kieckens L, Gertsenstein M, Fahrig M, Vandenhoeck A, Harpal K, Eberhardt C, Declercq C, Pawling J, Moons L, Collen D, Risau W, Nagy A (1996) Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. *Nature* 380:435–439
- Chilov D, Kukk E, Taira S, Jeltsch M, Kaukonen J, Palotie A, Joukov V, Alitalo K (1997) Genomic organization of human and mouse genes for vascular endothelial growth factor C. *J Biol Chem* 272:25176–25183
- Clauss M, Weich H, Breier G, Knies U, Rockl W, Waltenberger J, Risau W (1996) The vascular endothelial growth factor receptor Flt-1 mediates biological activities – implications for a functional role of placenta growth factor in monocyte activation and chemotaxis. *J Biol Chem* 271:17629–17634
- Cohen T, Nahari D, Cerem LW, Neufeld G, Levi BZ (1996) Interleukin 6 induces the expression of vascular endothelial growth factor. *J Biol Chem* 271:736–741
- DiPalma T, Tucci M, Russo G, Maglione D, Lago C, Romano A, Saccone S, DellaValle G, De Gregorio L, Dragani T, Viglietto G, Persico M (1997) The placenta growth factor gene of the mouse. *Mamm Genome* 7:6–12
- DiSalvo J, Bayne ML, Conn G, Kwok PW, Trivedi PG, Soderman DD, Palisi TM, Sullivan KA, Thomas KA (1995) Purification and characterization of a natural occurring vascular endothelial growth factor-placenta growth factor heterodimer. *J Biol Chem* 270:7717–7723
- Eichmann A, Corbel C, Nataf V, Vaigot P, Breant C, Le Douarin NM (1997) Ligand-dependent development of the endothelial and hemopoietic lineages from embryonic mesodermal cells expressing vascular endothelial growth factor receptor 2. *Proc Natl Acad Sci USA* 94:5141–5146
- Eichmann A, Corbel C, Jaffredo T, Bréant C, Joukov V, Kumar V, Alitalo K, le Douarin N (1998) Avian VEGF-C: cloning, embryonic expression pattern and stimulation of the differentiation of VEGFR2 expressing endothelial cell precursors. *Dev Biol* (in press)

- Enholm B, Paavonen K, Ristimäki A, Kumar V, Gunji Y, Kiefstrom J, Kivinen L, Laiho M, Olofsson B, Joukov V, Eriksson U, Alitalo K (1997) Comparison of VEGF, VEGF-B, VEGF-C and Ang-1 mRNA regulation by serum, growth factors, oncoproteins and hypoxia. *Oncogene* 14:2475–2483
- Ferrara N, Davis-Smyth T (1997) The biology of vascular endothelial growth factor. *Endocr Rev* 18:4–25
- Ferrara N, Carver-Moore K, Chen H, Dowd M, Lu L, O'Shea KS, Powell-Braxton L, Hillan KJ, Moore MW (1996) Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature* 380:439–442
- Finkenzeller G, Marme D, Weich HA, Hug H (1992) Platelet-derived growth factor-induced transcription of the vascular endothelial growth factor gene is mediated by protein kinase C. *Cancer Res* 52:4821–4823
- Frank S, Hubner G, Breier G, Longaker MT, Greenhalgh DG, Werner S (1995) Regulation of vascular endothelial growth factor expression in cultured keratinocytes. Implications for normal and impaired wound healing. *J Biol Chem* 270:12607–12613
- Gabrilovich DI, Chen HL, Girgis KR, Cunningham HT, Meny GM, Nadaf S, Kavanaugh D, Carbone DP (1996) Production of vascular endothelial growth factor by human tumors inhibits the functional maturation of dendritic cells. *Nat Med* 2:1096–1103
- Garrido C, Saule S, Gospodarowicz D (1993) Transcriptional regulation of vascular endothelial growth factor gene expression in ovarian bovine granulosa cells. *Growth Factors* 8:109–117
- Goldberg MA, Schneider TJ (1994) Similarities between the oxygen-sensing mechanisms regulating the expression of vascular endothelial growth factor and erythropoietin. *J Biol Chem* 269:4355–4359
- Grimmond S, Lagercrantz J, Drinkwater C, Silins G, Townson S, Pollock P, Gotley D, Carson E, Rakar S, Nordenskjöld M, Ward L, Hayward N, Weber G (1996) Cloning and characterization of a novel human gene related to vascular endothelial growth factor. *Genome Res* 6:124–131
- Heldin C, Östman A, Westermark B (1993) Structure of platelet-derived growth factor: Implications for functional properties. *Growth Factors* 8:245–252
- Jeltsch M, Kaipainen A, Joukov V, Meng XJ, Lakso M, Rauvala H, Swartz M, Fukumura D, Jain RK, Alitalo K (1997) Hyperplasia of lymphatic vessels in VEGF-C transgenic mice. *Science* 276:1423–1425
- Joukov V, Pajusola K, Kaipainen A, Chilov D, Lahtinen I, Kukk E, Saksela O, Kalkkinen N, Alitalo K (1996) A novel endothelial growth factor, VEGF-C, is a ligand for the Flt4 (VEGFR-3) and KDR (VEGFR-2) receptor tyrosine kinases. *EMBO J* 15:290–298
- Joukov V, Sorsa T, Kumar V, Jeltsch M, Claesson-Weish L, Cao Y, Saksela O, Kalkkinen N, Alitalo K (1997) Proteolytic processing regulates receptor specificity and activity of VEGF-C. *EMBO J* 16:3898–3911
- Joukov V, Kumar V, Sorsa T, Arighi E, Weich H, Saksela O, Alitalo K (1998) A recombinant mutant vascular endothelial growth factor C that has lost vascular endothelial growth factor receptor-2 binding, activation and vascular permeability activities. *J Biol Chem* 273:6599–6602
- Kaipainen A, Korhonen J, Pajusola K, Aprelikova O, Persico M, Terman B, Alitalo K (1993) The related FLT4, FLT1 and KDR receptor tyrosine kinases show distinct expression patterns in human fetal endothelial cells. *J Exp Med* 178:2077–2088
- Kaipainen A, Korhonen J, Mustonen T, van Hinsbergh VW, Fang GH, Dumont D, Breitman M, Alitalo K (1995) Expression of the *fms*-like tyrosine kinase 4 gene becomes restricted to lymphatic endothelium during development. *Proc Natl Acad Sci USA* 92:3566–3570
- Keyt BA, Nguyen HV, Berleau LT, Duarte CM, Park J, Chen H, Ferrara N (1996) Identification of vascular endothelial growth factor determinants for binding KDR and FLT-1 receptors. Generation of receptor-selective VEGF variants by site-directed mutagenesis. *J Biol Chem* 271:5638–5646
- Kukk E, Lymboussaki A, Taira S, Kaipainen A, Jeltsch M, Joukov V, Alitalo K (1996) VEGF-C receptor binding and pattern of expression with VEGFR-3 suggests a role in lymphatic vascular development. *Development* 122:3829–3837
- Lagercrantz J, Larsson C, Grimmond S, Fredriksson M, Weber G, Piehl F (1996) Expression of the VEGF-related factor gene in pre- and postnatal mouse. *Biochem Biophys Res Commun* 220:147–152
- Lee J, Gray A, Yuan J, Luoh S-M, Avraham H, Wood WI (1996) Vascular endothelial growth factor-related protein: a ligand and specific activator of the tyrosine kinase receptor Flt4. *Proc Natl Acad Sci USA* 93:1988–1992
- Lytle DJ, Fraser KM, Fleming SB, Mercer AA, Robinson AJ (1994) Homologs of vascular endothelial growth factor are encoded by the poxvirus orf virus. *J Virol* 68:84–92
- Midy V, Plouet J (1994) Vasculotropin/vascular endothelial growth factor induces differentiation in cultured osteoblasts. *Biochem Biophys Res Commun* 199:380–386
- Muller YA, Li B, Christinger HW, Wells JA, Cunningham BC, De Vos AM (1997) Vascular endothelial growth factor – crystal structure and functional mapping of the kinase domain receptor binding site. *Proc Natl Acad Sci USA* 94:7192–7197

- Oefner C, D'Arcy A, Winkler FK, Eggimann B, Hosang M (1992) Crystal structure of human platelet-derived growth factor BB. *EMBO J* 11:3921-3926
- Oh SJ, Jeltsch MM, Birkenhäger R, McCarthy JEG, Weich HA, Christ B, Alitalo K, Wiltling J (1997) VEGF and VEGF-C: Specific induction of angiogenesis and lymphangiogenesis in the differentiated avian chorioallantoic membrane. *Dev Biol* 188:96-109
- Olofsson B, Pajusola K, Kaipainen A, von Euler G, Joukov V, Saksela O, Orpana O, Pettersson R, Alitalo K, Eriksson U (1996a) Vascular endothelial growth factor B, a novel growth factor for endothelial cells. *Proc Natl Acad Sci USA* 93:2576-2581
- Olofsson B, Pajusola K, Voneuler G, Chilov D, Alitalo K, Eriksson U (1996b) Genomic organization of the mouse and human genes for vascular endothelial growth factor B (VEGF-B) and characterization of a second splice isoform. *J Biol Chem* 271:19310-19317
- Olofsson B, Korpeleinen E, Mandriota S, Pepper MS, Aase K, Jeltsch MM, Shibuya M, Alitalo K, Eriksson U (1998) VEGF-B binds VEGFR-1 and regulates plasminogen activator activity in endothelial cells. *Proc Natl Acad Sci USA* (submitted)
- Orlandini M, Marconcini L, Ferruzzi R, Oliviero S (1996) Identification of a C-fos-induced gene that is related to the platelet-derived growth factor/vascular endothelial growth factor family. *Proc Natl Acad Sci USA* 93:11675-11680
- Östman A, Rall L, Hammacher A, Wormstead MA, Coit D, Valenzuela P, Betsholtz C, Westermark B, Heldin CH (1988) Synthesis and assembly of a functionally active recombinant platelet-derived growth factor AB heterodimer. *J Biol Chem* 263:16202-16208
- Östman A, Thyberg J, Westermark B, Heldin CH (1992) PDGF-AA and PDGF-BB biosynthesis: pro-protein processing in the Golgi complex and lysosomal degradation of PDGF-BB retained intracellularly. *J Cell Biol* 118:509-519
- Paavonen K, Horelli-Kuitunen N, Chilov D, Kukk E, Pennanen S, Kallioniemi OP, Pajusola K, Olofsson B, Eriksson U, Joukov V, Palotie A, Alitalo K (1996) Novel human vascular endothelial growth factor genes VEGF-B and VEGF-C localize to chromosomes 11q13 and 4q34, respectively. *Circulation* 93:1079-1082
- Park J, Chen H, Winer J, Houck K, Ferrara N (1994) Placenta growth factor. *J Biol Chem* 269:25646-25654
- Pertovaara L, Kaipainen A, Mustonen T, Orpana A, Ferrara N, Saksela O, Alitalo K (1994) Vascular endothelial growth factor is induced in response to transforming growth factor-beta in fibroblastic and epithelial cells. *J Biol Chem* 269:6271-6274
- Ristimäki A, Narko K, Enholm B, Joukov V, Alitalo K (1998) Proinflammatory cytokines regulate expression of the lymphatic endothelial mitogen vascular endothelial growth factor-C. *J Biol Chem* 273:8413-8418
- Schmelz M, Moll R, Kuhn C, Franke WW (1994) Complexus adhaerentes, a new group of desmoplakin-containing junctions in endothelial cells: II. Different types of lymphatic vessels. *Differentiation* 57:97-117
- Soker S, Takashima S, Quan Miao H, Neufeld G, Klagsbrun M (1998) Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor. *Cell* 92:735-745
- Stein I, Neeman M, Shweiki D, Itin A, Keshet E (1995) Stabilization of vascular endothelial growth factor mRNA by hypoxia and hypoglycemia and coregulation with other ischemia-induced genes. *Mol Cell Biol* 15:5363-5368
- Terman B, Dougher-Vermazen M, Carrison M, Dimitrov D, Armellino D, Gospodarowicz D, Böhlen P (1992) Identification of the KDR tyrosine kinase as a receptor for vascular endothelial growth factor. *Biochem Biophys Res Commun* 187:1579-1586
- Tischer E, Mitchell R, Hartman T, Silva M, Gospodarowicz D, Fiddes J, Abraham J (1991) The human gene for vascular endothelial growth factor: multiple protein forms are encoded through alternative exon splicing. *J Biol Chem* 266:11947-11954
- Townson S, Lagercrantz J, Grimmond S, Silins G, Nordenskjöld M, Weber G, Hayward N (1996) Characterization of the murine VEGF-related factor gene. *Biochem Biophys Res Commun* 220:922-928
- Waltenberger J, Claesson-Welsh L, Siegbahn A, Shibuya M, Heldin CH (1994) Different signal transduction properties of KDR and Flt1, two receptors for vascular endothelial growth factor. *J Biol Chem* 269:26988-26995

Vascular Endothelial Growth Factor: Molecular and Biological Aspects

N. FERRARA

1	Introduction	1
2	Biological Activities of Vascular Endothelial Growth Factor	2
3	Organization of the VEGF Gene and Characteristics of the VEGF Proteins	4
4	Regulation of VEGF Gene Expression	6
4.1	Oxygen Tension	6
4.2	Cytokines	6
4.3	Differentiation and Transformation	7
5	The VEGF Receptors	8
6	The VEGFR-1 and VEGFR-2 Tyrosine Kinases	8
6.1	Binding Characteristics	8
6.2	Signal Transduction	9
6.3	Regulation	11
7	Role of VEGF and its Receptors in Physiological Angiogenesis	11
7.1	Distribution of VEGFR-1 and VEGFR-2 mRNA	11
7.2	The VEGFR-1, VEGFR-2 and VEGF Gene Knockouts in Mice	12
8	Role of VEGF in Corpus Luteum Angiogenesis	13
9	Role of VEGF in Pathological Angiogenesis	14
9.1	Tumor Angiogenesis	14
9.2	Angiogenesis Associated with Other Pathological Conditions	16
10	VEGF and Therapeutic Angiogenesis	18
11	Conclusions	20
	References	21

1 Introduction

The development of a vascular supply is a fundamental requirement for organ development and differentiation during embryogenesis as well as for wound healing and reproductive functions in the adult (FOLKMAN 1995). Angiogenesis is also implicated in the pathogenesis of a variety of disorders: proliferative retinopathies,

Department of Cardiovascular Research, Genentech, Inc., 460 Point San Bruno Boulevard, South San Francisco, CA 94080, USA

age-related macular degeneration, tumors, rheumatoid arthritis and psoriasis (FOLKMAN 1995; GARNER 1994).

The search for positive regulators of angiogenesis has yielded several candidates, including fibroblast growth factors a and b (aFGF, bFGF), transforming growth factors alpha and beta (TGF- α , TGF- β), hepatocyte growth factor (HGF), tumor necrosis factor alpha (TNF- α), angiogenin, interleukin-8 (IL-8), etc. (FOLKMAN and SHING 1992; RISAU 1997). However, in spite of extensive research, there is still significant debate as to their role as endogenous mediators of angiogenesis. The negative regulators identified so far include thrombospondin (GOOD et al. 1990; DiPIETRO 1997), the 16-kilodalton *N*-terminal fragment of prolactin (FERRARA et al. 1991), angiostatin (O'REILLY et al. 1994) and endostatin (O'REILLY et al. 1997).

This chapter discusses the molecular and biological properties of the vascular endothelial growth factor (VEGF) proteins. Over the last few years, several additional members of the VEGF gene family have been identified, including VEGF-B, VEGF-C, Placenta growth factor (PlGF) and VEGF-D. This chapter focuses primarily on VEGF, also referred to as "VEGF-A". For a description of the other members of the family, the reader is referred to the appropriate chapters in this book. Work done by several laboratories over the last few years has elucidated the pivotal role of VEGF and its receptors in the regulation of normal and abnormal angiogenesis (FERRARA and DAVIS-SMYTH 1997). The finding that the loss of even a single VEGF allele results in embryonic lethality points to an irreplaceable role played by this factor in the development and differentiation of the vascular system (FERRARA et al. 1996; CARMELIET et al. 1996). Furthermore, VEGF-induced angiogenesis has been shown to result in a therapeutic effect in animal models of coronary or limb ischemia and, most recently, in a human patient affected by critical leg ischemia (FERRARA and DAVIS-SMYTH 1997).

2 Biological Activities of Vascular Endothelial Growth Factor

Vascular endothelial growth factor (VEGF) is a mitogen for vascular endothelial cells derived from arteries, veins and lymphatics, but is devoid of consistent and appreciable mitogenic activity for other cell types (FERRARA and DAVIS-SMYTH 1997). VEGF promotes angiogenesis in tri-dimensional *in vitro* models, inducing confluent microvascular endothelial cells to invade collagen gels and form capillary-like structures (PEPPER et al. 1992). Also, VEGF induces sprouting from rat aortic rings embedded in a collagen gel (Nicosia et al. 1994). VEGF also elicits a pronounced angiogenic response in a variety of *in vivo* models, including the chick chorioallantoic membrane (LEUNG et al. 1989), the primate iris (TOLENTINO et al. 1996) etc.

VEGF induces expression of the serine proteases urokinase-type and tissue-type plasminogen activators (PA), and also PA inhibitor 1 (PAI-1) in cultured

bovine microvascular endothelial cells (PEPPER et al. 1991). Moreover, VEGF increases expression of the metalloproteinase interstitial collagenase in human umbilical-vein endothelial cells (HUVEC), but not in dermal fibroblasts (UNEMORI et al. 1992). Other studies have shown that VEGF promotes expression of the urokinase receptor (uPAR) in vascular endothelial cells (MANDRIOTA et al. 1995). Additionally, VEGF stimulates hexose transport in cultured vascular endothelial cells (PEKALA et al. 1990).

VEGF is known also as vascular permeability factor (VPF), based on its ability to induce vascular leakage in the guinea-pig skin (DVORAK et al. 1995). DVORAK and colleagues proposed that an increase in microvascular permeability is a crucial step in angiogenesis associated with tumors and wounds (DVORAK 1986). According to this hypothesis, a major function of VPF/VEGF in the angiogenic process is the induction of plasma-protein leakage. This effect would result in the formation of an extravascular fibrin gel, a substrate for endothelial and tumor cell growth (DVORAK et al. 1987). Recent studies have also suggested that VEGF may induce fenestrations in endothelial cells (ROBERTS and PALADE 1995, 1997). Topical administration of VEGF acutely resulted in the development of fenestrations in the endothelium of small venules and capillaries, even in regions where endothelial cells are not normally fenestrated, and was associated with increased vascular permeability (ROBERTS and PALADE 1995, 1997).

MELDER et al. (1996) have shown that VEGF promotes expression of VCAM-1 and ICAM-1 in endothelial cells. This induction results in the adhesion of activated natural killer (NK) cells to endothelial cells, mediated by specific interaction of endothelial VCAM-1 and ICAM-1 with CD18 and VLA-4 on the surface of NK cells.

VEGF has been reported to have certain regulatory effects on blood cells. CLAUSS et al. (1990) reported that VEGF may promote monocyte chemotaxis, while BROXMEYER et al. (1995) have shown that VEGF induces colony formation by mature subsets of granulocyte-macrophage progenitor cells. These findings may be explained by the common origin of endothelial and hematopoietic cells and the presence of VEGF receptors in progenitor cells as early as hemangioblasts in blood islands in the yolk sac. Furthermore, GABRILOVICH et al. (1996) have reported that VEGF may have an inhibitory effect on the maturation of host professional antigen-presenting cells, such as dendritic cells. VEGF was found to inhibit immature dendritic cells, without having a significant effect on the function of mature cells. These findings led to the suggestion that VEGF may also facilitate tumor growth by allowing the tumor to avoid the induction of an immune response (GABRILOVICH et al. 1996).

VEGF induces vasodilatation *in vitro* in a dose-dependent fashion (KU et al. 1993; YANG et al. 1996) and produces transient tachycardia, hypotension and a decrease in cardiac output when injected intravenously in conscious, instrumented rats (YANG et al. 1996). Such effects appear to be caused by a decrease in venous return, mediated primarily by endothelial cell-derived nitric oxide (NO), as assessed by the requirement for an intact endothelium and the prevention of the effects by *N*-methyl-arginine (YANG et al. 1996). Accordingly, VEGF has no direct effect on contractility or rate in the isolated rat heart *in vitro* (YANG et al. 1996). These

hemodynamic effects, however, are not unique to VEGF: other angiogenic factors, such as aFGF and bFGF, also have the ability to induce NO-mediated vasodilatation and hypotension (CUEVAS et al. 1991, 1996).

3 Organization of the VEGF Gene and Characteristics of the VEGF Proteins

The human VEGF gene is organized in eight exons, separated by seven introns. The coding region spans approximately 14 kb (HOUCK et al. 1991; TISCHER et al. 1991). The human VEGF gene has been assigned to chromosome 6p21.3 (VINCENTI et al. 1996). It is now well established that alternative exon splicing of a single VEGF gene results in the generation of four different molecular species, having respectively 121, 165, 189 and 206 amino acids following signal sequence cleavage (VEGF₁₂₁, VEGF₁₆₅, VEGF₁₈₉, VEGF₂₀₆). VEGF₁₆₅ lacks the residues encoded by exon 6, while VEGF₁₂₁ lacks the residues encoded by exons 6 and 7. Compared with VEGF₁₆₅, VEGF₁₂₁ lacks 44 amino acids; VEGF₁₈₉ has an insertion of 24 amino acids, highly enriched in basic residues; and VEGF₂₀₆ has an additional insertion of 17 amino acids (HOUCK et al. 1991). Analysis of the VEGF gene promoter region reveals a single major transcription start which lies near a cluster of potential Sp1 factor binding sites.

VEGF₁₆₅ is the predominant molecular species produced by a variety of normal and transformed cells. Transcripts encoding VEGF₁₂₁ and VEGF₁₈₉ are detected in the majority of cells and tissues expressing the VEGF gene (HOUCK et al. 1991). In contrast, VEGF₂₀₆ is a very rare form, so far identified only in a human fetal liver complementary deoxyribonucleic acid (cDNA) library (HOUCK et al. 1991). The genomic organization of the murine VEGF gene has been also described (SHIMA et al. 1996). Similarly to the human gene, the coding region of the murine VEGF gene encompasses approximately 14kb and is comprised of eight exons interrupted by seven introns. Analysis of exons suggests the generation of three isoforms: VEGF₁₂₀, VEGF₁₆₄ and VEGF₁₈₈. Therefore, murine VEGFs are shorter than human VEGF by one amino acid. However, a fourth isoform comparable with VEGF₂₀₆ is not predicted, since an in-frame stop codon is present in the region corresponding to the human VEGF₂₀₆ open reading frame. Analysis of the 3' untranslated region of the rat VEGF messenger ribonucleic acid (mRNA) has revealed the presence of four potential polyadenylation sites (LEVY et al. 1996). A frequently used site is about 1.9kb further downstream from the previously reported transcription termination codon (CONN et al. 1990). The sequence within this 3' untranslated region reveals a number of sequence motifs that are known to be involved in the regulation of mRNA stability (LEVY et al. 1996).

Native VEGF is a basic, heparin-binding, homodimeric glycoprotein of 45,000Da (FERRARA and HENZEL 1989). These properties correspond to those of VEGF₁₆₅, the major isoform (HOUCK et al. 1992). VEGF₁₂₁ is a weakly acidic

polypeptide that fails to bind to heparin (HOUCK et al. 1992). VEGF₁₈₉ and VEGF₂₀₆ are more basic and bind to heparin with a greater affinity than VEGF₁₆₅ (HOUCK et al. 1992). Such differences in the isoelectric point and the affinity for heparin may affect the bioavailability of VEGF profoundly.

VEGF₁₂₁ is a freely diffusible protein; VEGF₁₆₅ is also secreted, although a significant fraction remains bound to the cell surface and the extracellular matrix (ECM). In contrast, VEGF₁₈₉ and VEGF₂₀₆ are almost completely sequestered in the ECM (PARK et al. 1993). However, these isoforms may be released in a soluble form by heparin or heparinase, suggesting that their binding site is represented by proteoglycans containing heparin-like moieties. The long forms may also be released by plasmin following cleavage at the carboxy (COOH) terminus. This action generates a bioactive proteolytic fragment with a molecular weight of ~34,000Da (HOUCK et al. 1992).

Plasminogen activation and generation of plasmin have been shown to play an important role in the angiogenesis cascade. Thus, proteolysis of VEGF is likely also to occur *in vivo*. KEYT et al. (1996a) have shown that the bioactive product of plasmin action is comprised of the first 110 amino (NH₂)-terminal amino acids of VEGF. These findings suggest that the VEGF proteins may become available to endothelial cells by at least two different mechanisms: as freely diffusible proteins (VEGF₁₂₁, VEGF₁₆₅) or following protease activation and cleavage of the longer isoforms. However, loss of heparin binding, whether it is due to alternative splicing of RNA or plasmin cleavage, results in a substantial loss of mitogenic activity for vascular endothelial cells: compared with VEGF₁₆₅, VEGF₁₂₁ or VEGF₁₁₀ which demonstrate a 50- to 100-fold reduced potency when tested in endothelial cell growth assay (KEYT et al. 1996a).

It has been suggested that the stability of VEGF-heparan-sulfate-receptor complexes contributes to effective signal transduction and stimulation of endothelial cell proliferation (KEYT et al. 1996a). Thus, VEGF has the potential to express structural and functional heterogeneity to yield a graded and controlled biological response. Very recently, POLTORAK et al. (1997) provided evidence for the existence of an additional, alternatively spliced molecular species of VEGF. A VEGF isoform containing exons 1-6 and 8 of the VEGF gene was found to be expressed as a major VEGF mRNA form in several cell lines derived from carcinomas of the female reproductive system. This mRNA is predicted to encode a VEGF form of 145 amino acids (VEGF₁₄₅). Recombinant VEGF₁₄₅ induced the proliferation of vascular endothelial cells, albeit at much lower potency than VEGF₁₆₅. VEGF₁₄₅ binds to the kinase domain region (KDR) receptor, also denoted VEGF receptor-2 (VEGFR-2) on the surface of endothelial cells. It also binds to heparin with an affinity similar to that of VEGF₁₆₅.

Recently, MULLER et al. (1997) determined the crystal structure of VEGF at a resolution of 2.5Å. Overall, the VEGF monomer resembles that of platelet-derived growth factor (PDGF), but its *N*-terminal segment is helical rather than extended. The dimerization mode of VEGF is similar to that of PDGF and very different from that of TGF-β. Functional analysis of the binding epitopes for two receptor-

blocking antibodies reveal different binding determinants near each of the VEGFR-2 binding hot spots.

4 Regulation of VEGF Gene Expression

4.1 Oxygen Tension

Among the mechanisms that have been proposed to participate in the regulation of VEGF gene expression, oxygen tension plays a major role, both *in vitro* and *in vivo*. VEGF mRNA expression is rapidly and reversibly induced by exposure to low pO₂ in a variety of normal and transformed cultured-cell types (MINCHENKO et al. 1994; SHIMA et al. 1995). Also, ischemia caused by occlusion of the left anterior descending coronary artery results in a dramatic increase in VEGF RNA levels in the pig and rat myocardium, suggesting that VEGF may mediate the spontaneous revascularization that follows myocardial ischemia (BANAI et al. 1994b; HASHIMOTO et al. 1994). Furthermore, hypoxic upregulation of VEGF mRNA in neuroglial cells, secondary to the onset of neuronal activity, has been proposed to play an important physiological role in the development of the retinal vasculature (STONE et al. 1995).

Similarities exist between the mechanisms leading to hypoxic regulation of VEGF and erythropoietin (Epo) (GOLDBERG and SCHNEIDER 1994). Hypoxia-inducibility is conferred on both genes by homologous sequences. By deletion and mutation analysis, a 28-base sequence has been identified in the 5' promoter of the rat and human VEGF gene, which mediates hypoxia-induced transcription (LEVY et al. 1995; LIU et al. 1995). Such a sequence reveals a high degree of homology and similar protein-binding characteristics as the hypoxia-inducible factor 1 (HIF-1) binding site, within the Epo gene (MADAN and CURTIN 1993). HIF-1 has been identified as a mediator of transcriptional responses to hypoxia and is a basic, heterodimeric, helix-loop-helix protein (WANG and SEMENZA 1995). When reporter constructs containing the VEGF sequences that mediate hypoxia-inducibility were co-transfected with expression vectors encoding HIF-1 subunits, reporter gene transcription was much greater than that observed in cells transfected with the reporter alone, both in hypoxic and normoxic conditions (FORSYTHE et al. 1996).

However, transcriptional activation is not the only mechanism leading to VEGF upregulation in response to hypoxia (IKEDA et al. 1995; LEVY et al. 1996). Increased mRNA stability has been identified as a significant post-transcriptional component. Sequences that mediate increased stability were identified in the 3' untranslated region of the VEGF mRNA.

4.2 Cytokines

Various cytokines or growth factors may upregulate VEGF mRNA expression. Epidermal growth factor (EGF), TGF- β or keratinocyte growth factor (KGF)

result in a marked induction of VEGF mRNA expression (FRANK et al. 1995) EGF also stimulates VEGF release by cultured glioblastoma cells (GOLDMAN et al. 1993). In addition, treatment of quiescent cultures of epithelial and fibroblastic cell lines with TGF- β resulted in induction of VEGF mRNA and release of VEGF protein in the medium (PERTOVAARA et al. 1994). Based on these findings, it has been proposed that VEGF may function as a paracrine mediator for indirectly acting angiogenic agents, such as TGF- β (PERTOVAARA et al. 1994). Furthermore, IL-1- β induces VEGF expression in aortic smooth muscle cells (LI et al. 1995). Both IL-1- α and prostaglandin E₂(PGE₂) have been shown to induce expression of VEGF in cultured synovial fibroblasts, suggesting the participation of such inductive mechanisms in inflammatory angiogenesis (BEN-AV et al. 1995). IL-6 has also been shown to significantly induce VEGF expression in several cell lines (COHEN et al. 1996). IGF-1, a mitogen implicated in the growth of several malignancies, has also been shown to induce VEGF mRNA and protein in cultured colorectal carcinoma cells (WARREN et al. 1996). Such induction was mediated by a combined increase in the transcriptional rate of the VEGF gene and in the stability of the mRNA.

4.3 Differentiation and Transformation

Cell differentiation has been shown to play an important role in the regulation of VEGF gene expression (CLAFFEY et al. 1992). The VEGF mRNA is upregulated during the conversion of 3T3 pre-adipocytes into adipocytes or during the myogenic differentiation of C2C12 cells. Conversely, VEGF gene expression is repressed during the differentiation of the pheochromocytoma cell line PC12 into non-malignant, neuron-like cells.

Specific transforming events also result in induction of VEGF gene expression. A mutated form of the murine p53 tumor-suppressor gene has been shown to result in induction of VEGF mRNA expression in NIH 3T3 cells in transient transfection assays (KIESER et al. 1994). Likewise, oncogenic mutations or amplification of *ras* lead to VEGF upregulation (RAK et al. 1995; GRUGEL et al. 1995). Interestingly, expression of oncogenic *ras*, either constitutive or transient, potentiated the induction of VEGF by hypoxia (MAZURE et al. 1996). Moreover, the von Hippel-Lindau (VHL) tumor-suppressor gene has been recently implicated in the regulation of VEGF gene expression (SIEMEISTER et al. 1996; ILIOPOULOS et al. 1996; GNARRA et al. 1996).

The VHL tumor-suppressor gene is inactivated in patients with VHL disease and in most sporadic clear cell renal carcinomas. Although the function of the VHL protein remains to be fully elucidated, it is known that such a protein interacts with the elongin BC subunits in vivo, and regulates RNA polymerase-II elongation activity in vitro by inhibiting formation of the elongin ABC complex. Human renal cell carcinoma cells, either lacking endogenous wild-type VHL gene or expressing an inactive mutant, demonstrated altered regulation of VEGF gene expression, which was corrected by introduction of the wild-type VHL gene. Most of the endothelial cells' mitogenic activity released by tumor cells expressing the mutant

VHL gene was neutralized by anti-VEGF antibodies (SIEMEISTER et al. 1996). These findings suggest that VEGF is a key mediator of the abnormal vascular proliferations and solid tumors characteristic of VHL syndrome.

ILIOPOULOS et al. (1996) have shown that one function of the VHL protein is to provide a negative regulation of a series of hypoxia-inducible genes, including the VEGF, PDGF-B chain and the glucose transporter GLUT1 genes. In the presence of a mutant VHL, mRNAs for such genes were produced under both normoxic and hypoxic conditions. Reintroduction of wild-type VHL resulted in inhibition of mRNA production under normoxic conditions and restored the characteristic hypoxia-inducibility of those genes (ILIOPOULOS et al. 1996). In addition, GNARRA et al. (1996) have shown that VHL regulates VEGF expression at a post-transcriptional level and that VHL inactivation in target cells causes a loss of VEGF suppression, leading to formation of a vascular stroma. Interestingly, despite fivefold differences in VEGF mRNA levels, VHL overexpression did not affect VEGF transcription initiation.

5 The VEGF Receptors

Two classes of high-affinity VEGF binding sites on the surface of bovine endothelial cells were described initially, with K_d values of 10 pM and 100 pM (VAISMAN et al. 1990; PLOUET and MOUKADIRI 1990). Lower affinity binding sites on mononuclear phagocytes were subsequently described (SHEN et al. 1993). It has been suggested that such binding sites are involved in mediating the chemotactic effects of VEGF for monocytes (CLAUSS et al. 1990).

Ligand autoradiography studies on fetal and adult rat tissue sections demonstrated that high-affinity VEGF binding sites are localized to the vascular endothelium of large or small vessels in situ (JAKEMAN et al. 1992, 1993). VEGF binding was apparent not only on proliferating, but also on quiescent endothelial cells (JAKEMAN et al. 1992, 1993). Also, the earliest developmental identification of high-affinity VEGF binding was in hemangioblasts in the blood islands in the yolk sac (JAKEMAN et al. 1993).

6 The VEGFR-1 and VEGFR-2 Tyrosine Kinases

6.1 Binding Characteristics

Two VEGF receptor tyrosine kinases (RTKs) have been identified. The VEGFR-1 (fms-like-tyrosine kinase) (DE VRIES et al. 1992) and VEGFR-2 (KDR; TERMAN et al., 1992) receptors bind VEGF with high affinity. The murine homologue of

VEGFR-2 (also denoted fetal liver kinase-1; Flk-1), shares 85% sequence identity with human KDR (MATTHEWS et al. 1991). Both VEGFR-1 and VEGFR-2 have seven immunoglobulin (Ig)-like domains in the extracellular domain (ECD), a single transmembrane region and a consensus tyrosine kinase sequence, which is interrupted by a kinase-insert domain (SHIBUYA et al. 1990; TERMAN et al. 1991; MATTHEWS et al. 1991). VEGFR-1 has the highest affinity for rhVEGF₁₆₅, with a K_d of approximately 10–20pM (DE VRIES et al. 1992). VEGFR-2 has a somewhat lower affinity for VEGF: the K_d has been estimated to be approximately 75–125pM (TERMAN et al. 1992).

A cDNA coding an alternatively spliced soluble form of VEGFR-1 (sVEGFR-1), lacking the seventh Ig-like domain, the transmembrane sequence and the cytoplasmic domain, has been identified in HUVEC (KENDALL et al. 1996). This sVEGFR-1 receptor binds VEGF with high affinity (K_d 10–20pM) and is able to inhibit VEGF-induced mitogenesis and may be a physiological negative regulator of VEGF's action (KENDALL et al. 1996).

An additional member of the family of RTKs with seven Ig-like domains in the ECD is VEGFR-3 (also denoted Flt-4; PAJUSOLA et al. 1992; GALLAND et al. 1992; FINNERTY et al. 1993) which, however, is not a receptor for VEGF, but rather binds a newly identified ligand called VEGF-C or VEGF-related peptide (VRP) (see Chapt. 3).

Recent studies have mapped the binding site for VEGF to the second immunoglobulin-like domain of VEGFR-1 and VEGFR-2. Deletion of the second domain of VEGFR-1 completely abolished the binding of VEGF. Introduction of the second domain of VEGFR-2 into an VEGFR-1 mutant lacking the homologous domain restored VEGF binding. However, the ligand specificity was characteristic of the VEGFR-2. To further test this hypothesis, chimeric receptors, where the first three or just the second Ig-like domains of VEGFR-1 replaced the corresponding domains in VEGFR-3, were created. Both swaps conferred upon VEGFR-3 the ability to bind VEGF with an affinity nearly identical to that of wild-type VEGFR-1. Furthermore, transfected cells expressing these chimeric VEGFR-3 receptors exhibited increased DNA synthesis in response to VEGF or PlGF (DAVIS-SMYTH et al. 1996).

An application of these structure–function studies is the generation of inhibitors of VEGF activity. The first three Ig-like domains of VEGFR-1 fused to a heavy chain Fc potently inhibits VEGF bioactivity across species. The Fc may confer sufficient half-life and stability when injected systemically (CHAMOW and ASHKENAZI 1996). Therefore, this agent may be a useful tool in determining the role of endogenous VEGF in several *in vivo* models.

6.2 Signal Transduction

VEGF has been shown to induce the phosphorylation of at least 11 proteins in bovine aortic endothelial cells (GUO et al. 1995). Phospholipase C (PLC)- γ , and two proteins that associate with PLC- γ were phosphorylated in response to VEGF.

Furthermore, immunoblot analysis to search for mediators of signal transduction that contain SH2 domains demonstrated that VEGF induces phosphorylation of phosphatidyl inositol 3-kinase, *ras* GTPase activating protein (GAP) and several other proteins. These findings suggest that VEGF promotes the formation of multimeric aggregates of VEGF receptors with proteins that contain SH2 domains. These studies, however, did not identify which VEGF receptor(s) are involved in these events. Recently, it was suggested that NO mediates, at least in part, the mitogenic effect of VEGF on cultured microvascular endothelium isolated from coronary venules (MORBIDELLI et al. 1996). The proliferative effect of VEGF was reduced by pre-treatment of the cells with NO-synthase inhibitors. Exposure of the cells to VEGF induced a significant increment in cyclic guanosine monophosphate (cGMP) levels. These findings suggest that VEGF stimulates proliferation of post-capillary endothelial cells through the production of NO and cGMP accumulation.

Several studies have indicated that VEGFR-1 and VEGFR-2 have different signal transduction properties (WALTENBERGER et al. 1994; SEETHARAM et al. 1995). Porcine aortic endothelial cells lacking endogenous VEGF receptors display chemotaxis and mitogenesis in response to VEGF when transfected with a plasmid coding for VEGFR-2 (WALTENBERGER et al. 1994). In contrast, transfected cells expressing VEGFR-1 lack such responses (WALTENBERGER et al. 1994; SEETHARAM et al. 1995). VEGFR-2 undergoes strong ligand-dependent tyrosine phosphorylation in intact cells, while VEGFR-1 reveals a weak or undetectable response (WALTENBERGER et al. 1994; SEETHARAM et al. 1995). In addition, VEGF stimulation results in weak tyrosine phosphorylation that does not generate any mitogenic signal in transfected NIH 3T3 cells expressing VEGFR-1 (SEETHARAM et al. 1995). These findings agree with other studies showing that PlGF, which binds with high affinity to VEGFR-1, but not to VEGFR-2, lacks direct mitogenic or permeability-enhancing properties or the ability to effectively stimulate tyrosine phosphorylation in endothelial cells (PARK et al. 1994).

It seems, then, that interaction with VEGFR-2 is a critical requirement to induce the full spectrum of VEGF biological responses. In further support of this conclusion, VEGF mutants that bind selectively to VEGFR-2 are fully active endothelial-cell mitogens (KEYT et al. 1996b). These findings led to cast doubt on the role of VEGFR-1 as a truly signaling receptor. However, more recent evidence indicates that VEGFR-1 indeed signals, although our understanding of these events is fragmentary. CUNNINGHAM et al. (1995) have demonstrated an interaction between VEGFR-1 and the p85 subunit of phosphatidyl inositol 3-kinase (CUNNINGHAM et al. 1995), suggesting that p85 couples VEGFR-1 to intracellular signal transduction systems and implicate elevated levels of phosphatidyl inositol (3,4,5) P3 levels in this process (CUNNINGHAM et al. 1995). Also, members of the *Src* family, such as *Fyn* and *Yes*, show an increased level of phosphorylation following VEGF stimulation in transfected cells expressing VEGFR-1, but not VEGFR-2 (WALTENBERGER et al. 1994). Furthermore, it has been shown that a specific biological response, the migration of monocytes in response to VEGF (or PlGF), is mediated by VEGFR-1 (BARLEON et al. 1996).

6.3 Regulation

The expression of VEGFR-1 and -2 genes is largely restricted to the vascular endothelium. The promoter region of VEGFR-1 has been cloned and characterized and a 1-kb fragment of the 5' flanking region, essential for endothelial-specific expression, was identified (MORISHITA et al. 1995). Likewise, a 4-kb 5' flanking sequence has been identified in the promoter of the human VEGFR-2 that confers endothelial cell-specific activation (PATTERSON et al. 1995).

Similarly to VEGF, hypoxia has been proposed to play an important role in the regulation of VEGF-receptor gene expression. Exposure of rats to acute or chronic hypoxia led to pronounced upregulation of both VEGFR-1 and VEGFR-2 genes in the lung vasculature (TUDER et al. 1995). Also, VEGFR-1 and -2 mRNAs were substantially upregulated throughout the heart, following myocardial infarction in the rat (LI et al. 1996). However, in vitro studies have yielded unexpected results. Hypoxia increases VEGF receptor number by 50% in cultured bovine retinal capillary endothelial cells, but the expression of VEGFR-2 is not induced although, paradoxically, shows an initial downregulation (TAKAGI et al. 1996). BROGI et al. (1996) have proposed that the hypoxic upregulation of VEGFR-2 observed in vivo is not direct, but requires the release of an unidentified paracrine mediator from ischemic tissues.

Recent studies have provided evidence of a differential transcriptional regulation of the VEGFR-1 and VEGFR-2 genes by hypoxia (GERBER et al. 1997). When HUVEC were exposed to hypoxic conditions, in vitro, increased levels of VEGFR-1 expression were observed. In contrast, VEGFR-2 mRNA levels were unchanged or slightly repressed. Promoter deletion analysis demonstrated that a 430-bp region of the VEGFR-1 promoter was required for transcriptional activation in response to hypoxia. This region includes a heptamer sequence matching the HIF-1 consensus binding site previously found in other hypoxia inducible genes. The element mediating the hypoxia response was further defined as a 40-bp sequence, including the putative HIF-1 binding site, but was not found in the VEGFR-2 promoter. These findings indicate that, unlike the VEGFR-2 gene, the VEGFR-1 receptor gene is directly upregulated by hypoxia via a hypoxia-inducible enhancer element located at position -976 to -937 of the VEGFR-1 promoter (GERBER et al. 1997). Also, recent studies have shown that both TNF- α (PATTERSON et al. 1996) and TGF- β (MANDRIOTA et al. 1996) have the ability to inhibit the expression of the VEGFR-2 gene in cultured endothelial cells.

7 Role of VEGF and its Receptors in Physiological Angiogenesis

7.1 Distribution of VEGF, VEGFR-1 and VEGFR-2

The proliferation of blood vessels is crucial for a wide variety of physiological processes, such as embryonic development, normal growth and differentiation, wound healing and reproductive functions.

During embryonic development, VEGF expression is first detected within the first few days following implantation in the giant cells of the trophoblast (BREIER et al. 1992; JAKEMAN et al. 1993). At later developmental stages in the mouse or rat embryos, the VEGF mRNA is expressed in several organs, including heart, vertebral column, kidney and along the surface of the spinal cord and brain. In the developing mouse brain, the highest levels of mRNA expression are associated with the choroid plexus and the ventricular epithelium (BREIER et al. 1992). In the human fetus (16–22 weeks), VEGF mRNA expression is detectable in virtually all tissues and is most abundant in lung, kidney and spleen (SHIFREN et al. 1994).

In situ hybridization studies have shown that the VEGFR-2 mRNA is expressed in the yolk sac and intraembryonic mesoderm and later on in angioblasts, endocardium and small and large vessel endothelium (QUINN et al. 1993; MILLAUER et al. 1993). These findings strongly suggested a role for VEGFR-2 in the regulation of vasculogenesis and angiogenesis. Other studies have demonstrated that expression of VEGFR-2 mRNA is first detected in the proximal-lateral embryonic mesoderm, which gives rise to the heart (YAMAGUCHI et al. 1993). VEGFR-2 is then detectable in endocardial cells of the heart primordia and, subsequently, in the major embryonic and extraembryonic vessels (YAMAGUCHI et al. 1993). These studies have indicated that VEGFR-2 may be the earliest marker of endothelial-cell precursors. The VEGFR-1 mRNA is selectively expressed in vascular endothelial cells, in both fetal and adult mouse tissues (PETERS et al. 1993). Similarly to the high-affinity VEGF binding, the VEGFR-1 mRNA is expressed in both proliferating and quiescent endothelial cells, suggesting a role for VEGFR-1 in the maintenance of endothelial cells (PETERS et al. 1993).

VEGF expression is also detectable around microvessels in areas where endothelial cells are normally quiescent, such as kidney glomerulus, pituitary, heart, lung and brain (FERRARA et al. 1992; MONACCI et al. 1993). These findings raised the possibility that VEGF may be required not only to induce active vascular proliferation but, at least in some circumstances, also for the maintenance of the differentiated state of blood vessels (FERRARA et al. 1992). In agreement with this hypothesis, ALON et al. (1995) have shown that VEGF acts as a survival factor, at least for the developing retinal vessels. They propose that hyperoxia-induced vascular regression in the retina of neonatal animals is a consequence of inhibition of VEGF production by glial cells. Accordingly, intraocular administration of VEGF to newborn rats at the onset of hyperoxia was able to prevent cell apoptosis and regression of the retinal vasculature (ALON et al. 1995).

7.2 The VEGFR-1, VEGFR-2 and VEGF Gene Knockouts in Mice

Recent studies have demonstrated that both VEGFR-1 and VEGFR-2 are essential for normal development of embryonic vasculature. However, their respective roles in endothelial-cell proliferation and differentiation appear to be distinct (FONG et al. 1995; SHALABY et al. 1995). Mouse embryos homozygous for a targeted mutation in the VEGFR-1 locus died in utero between day 8.5 and day 9.5 (FONG et al.

1995). Endothelial cells developed in both embryonic and extraembryonic sites, but failed to organize in normal vascular channels. Mice in which the VEGFR-2 gene had been inactivated lacked vasculogenesis and also failed to develop blood islands. Hematopoietic precursors were severely disrupted and organized blood vessels failed to develop throughout the embryo or the yolk sac, resulting in death in utero between day 8.5 and day 9.5 (SHALABY et al. 1995).

However, these findings do not necessarily imply VEGF as being equally essential, since other ligands might potentially activate the VEGFR-1 and -2 and, thus, substitute VEGF's action. Very recent studies (CARMELIET et al. 1996; FERRARA et al. 1996) have generated direct evidence for the role played by VEGF in embryonic vasculogenesis and angiogenesis. Inactivation of the VEGF gene in mice resulted in embryonic lethality in heterozygous embryos, between day 11 and day 12. The VEGF^{+/−} embryos were growth retarded and also exhibited a number of developmental anomalies. The forebrain region appeared significantly underdeveloped. In the heart region, the outflow region was grossly malformed; the dorsal aortae were rudimentary, and the thickness of the ventricular wall was markedly decreased. The yolk sac revealed a markedly reduced number of nucleated red blood cells within the blood islands. Also, the vitelline veins failed to fuse with the vascular plexus of the yolk sac. Significant defects in the vasculature of other tissues and organs, including placenta and nervous system, were observed. In situ hybridization confirmed expression of VEGF mRNA in heterozygous embryos. Thus, the VEGF^{+/−} phenotype appears to be due to gene dosage and not to maternal imprinting.

While several heterozygous phenotypes have been described (BRANDON et al. 1995), this may be the first example of embryonic lethality following the loss of a single allele of a gene that is not maternally imprinted. Therefore, VEGF and its receptors are essential for blood island formation and angiogenesis, such that even reduced concentrations of VEGF are inadequate to support a normal pattern of development. However, inactivation of the PlGF gene does not result in embryonic lethality, even in the homozygous state (CARMELIET and COLLEN 1997). PlGF^{−/−} mice are viable and fertile, although they may have some impairment of wound healing.

8 Role of VEGF in Corpus Luteum Angiogenesis

The development and endocrine function of the ovarian corpus luteum (CL) are dependent on the growth of new capillary vessels. Although several molecules have been implicated as mediators of CL angiogenesis, at present, there is no direct evidence for the involvement of any. The VEGF mRNA is temporally and spatially related to the proliferation of blood vessels in the rat, mouse and primate ovary and in the rat uterus, suggesting that VEGF is a mediator of the cyclical growth of blood vessels that occurs in the female reproductive tract (PHILLIPS et al. 1990; RAVINDRANATH et al. 1992; SHWEIKI et al. 1993; CULLINAN-BOVE and KOOS 1993).

Very recently, the hypothesis that VEGF is a mediator of CL angiogenesis has been examined in a rat model of hormonally induced ovulation (FERRARA et al. 1998). Treatment with Flt (1-3)-IgG resulted in virtually complete suppression of CL angiogenesis. This effect was associated with inhibition of CL development and progesterone release. Failure of maturation of the endometrium was also observed. Areas of ischemic necrosis were demonstrated in the CL of treated animals; however, no effect on the pre-existing ovarian vasculature was observed. These findings demonstrate that, in spite of the redundancy of potential mediators, VEGF is essential for CL angiogenesis. Furthermore, they have implications in the control of fertility and the treatment of ovarian disorders characterized by hypervascularity and hyperplasia, such as polycystic ovary syndrome.

9 Role of VEGF in Pathological Angiogenesis

9.1 Tumor Angiogenesis

In 1945, ALGIRE and CHALKLEY, on the basis of microscopic observations of the vascular development of tumor xenografts in transparent chambers in mice, proposed that the growth of solid tumors is dependent on the development of a new vascular supply derived from the host (ALGIRE and CHALKLEY 1945). In 1971, FOLKMAN proposed inhibition of angiogenesis as a novel strategy to treat cancer (FOLKMAN 1971). Since then, extensive research has been devoted to the identification of tumor angiogenesis factor(s).

Many tumor cell lines secrete VEGF *in vitro* (FERRARA et al. 1992). *In situ* hybridization studies have demonstrated that the VEGF mRNA is markedly up-regulated in the vast majority of human tumors examined so far. These include: lung (VOLM et al. 1997a,b), breast (BROWN et al. 1995a; YOSHII et al. 1996), gastrointestinal tract (BROWN et al. 1993b; SUZUKI et al. 1996), kidney (BROWN et al. 1993a), bladder (BROWN et al. 1993a), ovary (OLSON et al. 1994), endometrium (GUIDI et al. 1996) and uterine cervix (GUIDI et al. 1995) carcinomas, angiosarcoma (HASHIMOTO et al. 1995), germ cell tumors (VIGLIETTO et al. 1996) and several intracranial tumors, including glioblastoma multiforme (SHWEIKI et al. 1992; PLATE et al. 1992; PHILLIPS et al. 1993) and sporadic, as well as VHL syndrome-associated capillary hemangioblastoma (BERKMAN et al. 1993; WIZIGMANN VOOS et al. 1995). In glioblastoma multiforme and other tumors with significant necrosis, the expression of VEGF mRNA is highest in hypoxic tumor cells adjacent to necrotic areas (SHWEIKI et al. 1992; PLATE et al. 1992; PHILLIPS et al. 1993). A correlation exists between the degree of vascularization of the malignancy and VEGF mRNA expression (BERKMAN et al. 1993; WIZIGMANN VOOS et al. 1995; GUIDI et al. 1995). In virtually all specimens examined, the VEGF mRNA was expressed in tumor cells, but not in endothelial cells. In contrast, the mRNAs for VEGFR-1 and -2 were upregulated in the endothelial cells associated with the tumor (BROWN et al.

1993b; PLATE et al. 1993). These findings are consistent with the hypothesis that VEGF is primarily a paracrine mediator (FERRARA et al. 1993).

Immunohistochemical studies have localized the VEGF protein not only to the tumor cells, but also to the vasculature (PLATE et al. 1992; BROWN et al. 1993b). This localization indicates that tumor-secreted VEGF accumulates in the target cells (QU et al. 1995). Interestingly, recent studies have suggested that the angiogenesis mediated by the human immunodeficiency virus (HIV-1) Tat protein (ALBINI et al. 1996a) requires activation of VEGFR-2 (ALBINI et al. 1996b). Tat induces growth of Kaposi's sarcoma (KS) spindle cells and has been implicated in the vasculature of the KS lesions (ALBINI et al. 1996b).

Elevations in VEGF levels have been detected in the serum of some cancer patients (KONDO et al. 1994). Also, a correlation has been noted between VEGF expression and microvessel density in primary breast cancer sections (TOI et al. 1996). A post-operative survey indicated that the relapse-free survival rate of patients with VEGF-positive tumors was significantly worse than that of VEGF-negative, suggesting that expression of VEGF is associated with stimulation of angiogenesis and with early relapse in primary breast cancer (GASPARINI et al. 1997). A similar correlation has been described in gastric-carcinoma patients (MAEDA et al. 1996). VEGF-positivity in tumor sections was correlated with vessel involvement, lymph node metastasis and liver metastasis. Furthermore, patients with VEGF-positive tumors had a worse prognosis than those with VEGF-negative tumors (MAEDA et al. 1996).

The availability of specific monoclonal antibodies capable of inhibiting VEGF-induced angiogenesis in vivo and in vitro (KIM et al. 1992) made it possible to generate direct evidence for a role of VEGF in tumorigenesis. In a study published by KIM et al. (1993), such antibodies were found to exert a potent inhibitory effect on the growth of three human tumor cell lines injected subcutaneously in nude mice, the SK-LMS-1 leiomyosarcoma, the G55 glioblastoma multiforme and the A673 rhabdomyosarcoma. The growth inhibition ranged between 70% and more than 95%. Subsequently, other tumor cell lines were found to be inhibited in vivo by this treatment (WARREN et al. 1995; MELNYK et al. 1996; ASANO et al. 1995; BORGSTROM et al. 1998a).

In agreement with the hypothesis that inhibition of neovascularization is the mechanism of tumor suppression, the density of blood vessels was significantly lower in sections of tumors from antibody-treated animals than in controls. Furthermore, neither the antibodies nor VEGF had any effect on the in vitro growth of the tumor cells (KIM et al. 1993). Intravital videomicroscopy techniques have allowed a more direct verification of the hypothesis that anti-VEGF antibodies indeed block tumor angiogenesis (BORGSTROM et al. 1996). Non-invasive imaging of the vasculature revealed a nearly complete suppression of tumor angiogenesis in anti-VEGF treated animals compared with controls, at all time points examined (BORGSTROM et al. 1996).

VEGF is a mediator of the in vivo growth of human colon carcinoma HM7 cells in a nude mouse model of liver metastasis (WARREN et al. 1995). Treatment with anti-VEGF monoclonal antibodies resulted in a dramatic decrease in the

number and size of metastases. Similarly, administration of anti-VEGF neutralizing antibodies inhibited primary tumor growth and metastasis of A431 human epidermoid carcinoma cells in severe combined immune deficient (SCID) mice (MELNYK et al. 1996) or HT-1080 fibrosarcoma cells implanted in BALB/c nude mice (ASANO et al. 1995).

Recently, BORGSTROM et al. (1998b) have shown that a combination treatment that includes anti-VEGF monoclonal antibody and doxorubicin results in a significant enhancement of the efficacy of either agent alone and led, in some cases, to complete regression of tumors derived from MCF-7 breast carcinoma cells in nude mice.

Intravital fluorescence microscopy and video imaging analysis have also been applied to address the important issue regarding the effects of VEGF on permeability and other properties of tumor vessels (YUAN et al. 1996). Treatment with anti-VEGF monoclonal antibodies was initiated after tumor xenografts had already been established and vascularized, and resulted in time-dependent reductions in vascular permeability (YUAN et al. 1996). These effects were accompanied by striking changes in the morphology of vessels, with dramatic reduction in diameter and tortuosity. This reduction in diameter is expected to block the passage of blood elements and eventually stop the flow in the tumor vascular network. A regression of blood vessels was observed after repeated administrations of anti-VEGF antibody. These findings suggest that tumor vessels require constant stimulation with VEGF in order to maintain not only their proliferative properties, but also some key morphological features (YUAN et al. 1996).

An independent verification of the hypothesis that the VEGF action is required for tumor angiogenesis has been provided by the finding that retrovirus-mediated expression of a dominant negative VEGFR-2 mutant, which inhibits signal transduction through wild-type VEGFR-2, suppresses the growth of glioblastoma multiforme and other tumor cell lines in vivo (MILLAUER et al. 1994).

9.2 Angiogenesis Associated with Other Pathological Conditions

Diabetes mellitus, occlusion of central retinal vein or prematurity with subsequent exposure to oxygen can all be associated with intraocular neovascularization (GARNER 1994). The new blood vessels may lead to vitreous hemorrhage, retinal detachment, neovascular glaucoma and eventual blindness (GARNER 1994). Diabetic retinopathy is the leading cause of blindness in the working population (OLK and LEE 1993). All of these conditions are known to be associated with retinal ischemia (PATZ 1980). In 1948, MICHAELSON proposed that a key event in the pathogenesis of these conditions was the release by the ischemic retina of diffusible angiogenic factor(s) ("factor X") responsible for retinal and iris neovascularization into the vitreous (MICHAELSON 1948). VEGF, by virtue of its diffusible nature and hypoxia-inducibility, was an attractive candidate as a mediator of intraocular neovascularization. Accordingly, elevations of VEGF levels in the aqueous and

vitreous of eyes with proliferative retinopathy have been described (AIELLO et al. 1994; ADAMIS et al. 1994; MALECAZE et al. 1994).

In a large series, a strong correlation was found between levels of immunoreactive VEGF in the aqueous and vitreous humors and active proliferative retinopathy. VEGF levels were undetectable or very low (<0.5ng/ml) in the eyes of patients affected by non-neovascular disorders or diabetes without proliferative retinopathy (AIELLO et al. 1994). In contrast, the VEGF levels were in the range 3–10ng/ml in the presence of active proliferative retinopathy associated with diabetes, occlusion of central retinal vein or prematurity. In agreement with these findings, *in situ* hybridization studies have demonstrated upregulation of VEGF mRNA in the retina of patients with proliferative retinopathies secondary to diabetes, central retinal vein occlusion, retinal detachment or intraocular tumors (PE'ER et al. 1996).

More direct evidence for a role of VEGF as a mediator of intraocular neovascularization has been generated in a primate model of iris neovascularization and in a murine model of retinopathy of prematurity (MILLER et al. 1994; PIERCE et al. 1995). In the former, intraocular administration of anti-VEGF antibodies dramatically inhibits the neovascularization that follows occlusion of central retinal veins (ADAMIS et al. 1996). Likewise, soluble VEGFR-1 or VEGFR-2 fused to an IgG suppresses retinal angiogenesis in the mouse model (AIELLO et al. 1995).

Neovascularization is also a major cause of visual loss in age-related macular degeneration (AMD), the overall leading cause of blindness (GARNER 1994). Most AMD patients demonstrate atrophy of the retinal pigment epithelia and characteristic formations called "drusen". A significant percentage of AMD patients (~20%) manifest the neovascular (exudative) form of the disease. In this condition, the new vessels stem from the extraretinal choriocapillary (GARNER 1994). Leakage and bleeding from these vessels may lead to damage to the macula and, ultimately, to loss of central vision. Because of the proximity of the lesions to the macula, laser photocoagulation or surgical therapy are of very limited value. Very recent studies have documented the immunohistochemical localization of VEGF in surgically resected choroidal neovascular membranes from AMD patients (LOPEZ et al. 1996; KVANTA et al. 1996). These findings suggests a role for VEGF in the progression of AMD-related choroidal neovascularization, raising the possibility that a pharmacological treatment with monoclonal antibodies or other VEGF inhibitors may constitute a therapy for this condition.

Two independent studies have suggested that VEGF is involved in the pathogenesis of rheumatoid arthritis (RA), an inflammatory disease in which angiogenesis plays a significant role (KOCH et al. 1994; FAVA et al. 1994). The RA synovium is characterized by the formation of pannus, an extensively vascularized tissue, which invades and destroys the articular cartilage (FASSBENDER and SIMLING-ANNENFELD 1983). Levels of immunoreactive VEGF were found to be high in the synovial fluid of RA patients, while they were very low or undetectable in the synovial fluid of patients affected by other forms of arthritis or by degenerative joint disease (KOCH et al. 1994; FAVA et al. 1994). Furthermore, anti-VEGF antibodies significantly reduced the endothelial cell chemotactic activity of the RA synovial fluid (KOCH et al. 1994).

It has been shown that VEGF expression is increased in psoriatic skin (DETMAR et al. 1994). Increased vascularity and permeability are characteristic of psoriasis. Also, VEGF mRNA expression has been examined in three bullous disorders with subepidermal blister formation, bullous pemphigoid, erythema multiforme and dermatitis herpetiformis (BROWN et al. 1995b).

Angiogenesis is also important in the pathogenesis of endometriosis, a condition characterized by ectopic endometrium implants in the peritoneal cavity. Recently, elevation of VEGF in the peritoneal fluid of patients with endometriosis has been reported (MCLAREN et al. 1996; SHIFREN et al. 1996). Immunohistochemistry indicated that activated peritoneal fluid macrophages, as well as tissue macrophages within the ectopic endometrium, are the main source of VEGF in this condition. (MCLAREN et al. 1996; SHIFREN et al. 1996). VEGF upregulation has also been implicated in the hypervascularity of the ovarian stroma that characterizes Stein-Leventhal syndrome (KAMAT et al. 1995). Moreover, SATO et al. (1995) proposed that VEGF may be responsible for the characteristic hypervascularity of Graves' disease. Thyroid-stimulating hormone (TSH), insulin phorbol ester, dibutyl cAMP and Graves' IgG were found to stimulate VEGF mRNA expression in cultured human thyroid follicles (SATO et al. 1995).

10 VEGF and Therapeutic Angiogenesis

The availability of agents able to promote the growth of new collateral vessels would be, potentially, of major therapeutic value for disorders characterized by inadequate tissue perfusion, and might constitute an alternative to surgical reconstruction procedures. For example, chronic limb ischemia, most frequently caused by obstructive atherosclerosis affecting the superficial femoral artery, is associated with a high rate of morbidity and mortality, and treatment is currently limited to surgical revascularization or endovascular interventional therapy (GRAOR and GRAY 1991); no pharmacological therapy has been shown to be effective for this condition.

It has been shown that intraarterial or intramuscular administration of recombinant human (rh)VEGF₁₆₅ may significantly augment perfusion and development of collateral vessels in a rabbit model, in which chronic hindlimb ischemia was created by surgical removal of the femoral artery (TAKESHITA et al. 1994). These studies provided angiographic evidence of neovascularization in the ischemic limbs. Arterial gene transfer with cDNA encoding VEGF also led to revascularization in the same rabbit model to an extent comparable with that achieved with the recombinant protein (TAKESHITA et al. 1996a,b). In addition, the hypothesis that the angiogenesis initiated by the administration of VEGF improved muscle function in ischemic limbs was tested by WALDER et al. (1996). A single intraarterial injection of rhVEGF₁₆₅ augmented muscle function in this rabbit model of peripheral limb ischemia. This exercise-induced hyperemia was signifi-

cantly improved in ischemic limbs treated with rhVEGF₁₆₅ (WALDER et al. 1996). Such improvement in perfusion was, however, not seen in other non-ischemic tissues, including the contralateral limb. Similarly, BAUTERS et al. (1994) have shown that both maximal flow velocity and maximal blood flow are significantly increased in ischemic limbs following VEGF administration.

Other studies have shown that VEGF administration also leads to a recovery of normal endothelial reactivity in dysfunctional endothelium. Following obstruction of a large artery and development of collateral vessels, the increase in blood flow that normally follows acetylcholine infusion is severely blunted; serotonin paradoxically leads to a decrease in blood flow (BAUTERS et al. 1995). Thirty days after a single intraarterial bolus of VEGF₁₆₅, restoration of the normal increase in blood flow was demonstrated in the ischemic rabbit hindlimb, following acetylcholine or serotonin infusion (BAUTERS et al. 1995).

BANAI et al. (1994a) have shown that VEGF administration results in increased coronary blood flow in a dog model of coronary insufficiency. Following occlusion of the left circumflex coronary artery, daily intraluminal injections of rhVEGF distal to the occlusion resulted in a significant enhancement of collateral blood flow over a 4-week period. In addition, HARADA et al. (1996) demonstrated that extraluminal administration of as little as 2µg of rhVEGF by an osmotic pump results in a significant increase in coronary blood flow in a pig model of chronic myocardial ischemia created by ameroid occlusion of the left proximal circumflex artery. Also, magnetic resonance imaging provided a non-invasive assessment of the benefits secondary to VEGF administration in the porcine model (PEARLMAN et al. 1995). Image series converted to a space-time map demonstrated a reduction in the size of the ischemic zone and a decreased delay in contrast arrival after VEGF treatment. These findings demonstrated improvement in cardiac global and regional function and reduced infarct size, resulting from enhanced collateral blood supply (PEARLMAN et al. 1995; WARE and SIMONS 1997).

A further potential therapeutic application of VEGF is the prevention of restenosis following percutaneous transluminal angioplasty (PTA). Between 15% and 75% of patients undergoing PTA for occlusive coronary or peripheral arterial disease develop restenosis within 6 months (GRAOR and GRAY 1991). It has been proposed that damage to the endothelium is a crucial event, triggering fibrocellular intimal proliferation (ESSED et al. 1983). Therefore, the induction of rapid re-endothelialization may be an effective strategy to prevent the cascade of events leading to neointima formation and ultimately to restenosis in patients. Recent evidence shows that VEGF accelerates re-endothelialization and also attenuates intimal hyperplasia in balloon-injured rat carotid artery or rabbit aorta (ASAHARA et al. 1995; CALLOW et al. 1994).

Recently, the hypothesis that VEGF may result in therapeutically significant angiogenesis in humans has been tested by ISNER et al. (1996b) in a gene-therapy trial in patients with severe limb ischemia. A case report of an interim analysis of this trial has been published (ISNER et al. 1996a). Arterial gene transfer of 2000µg naked plasmid DNA encoding VEGF₁₆₅, applied to the hydrogel polymer coating of an angioplasty balloon, resulted in angiographic and histologic evidence of

angiogenesis in the knee mid-tibial and ankle levels 4 weeks after transfer. Such effects persisted at a 12-week view (ISNER et al. 1996a).

11 Conclusions

The recent findings that heterozygous mutations inactivating the VEGF gene result in profound deficits in vasculogenesis and blood island formation, leading to early intrauterine death, emphasize the pivotal role played by this molecule in the development of the vascular system. Future studies, using inducible gene knockout technology (KUHN et al. 1995) should help determine the timing, when the embryo is most vulnerable to VEGF deficiency.

The elucidation of the signal transduction properties of the VEGF receptors holds the promise to dissect the pathways leading to such fundamental biological events as endothelial cell differentiation, morphogenesis and angiogenesis. Furthermore, a more complete understanding of the signaling events involving other endothelial cell-specific tyrosine kinases as well as cell-adhesion molecules and their interrelation with the VEGF/VEGF receptor system should provide a more integrated view of the biology of the endothelial cell, both in normal and abnormal circumstances. In this context, recent studies have shown that VEGF-mediated angiogenesis requires a specific vascular integrin pathway, mediated by $\alpha v\beta 5$ (FRIEDLANDER et al. 1995). Furthermore, a ligand selective for the endothelial cell-specific tyrosine kinase Tie-2 has been recently identified and named angiopoietin (Ang)-1 (DAVIS et al. 1996). Gene knockout studies have shown that Ang-1 is required for the correct assembly of the vessel wall (SURI et al. 1996). Ang-1 seems to play a crucial role in mediating reciprocal interactions between the endothelium and surrounding matrix and mesenchyme, and plays a later role in angiogenesis than VEGF. Also, unlike VEGF, Ang-1 does not directly stimulate endothelial cell growth. Interestingly, very recent studies provide evidence for the existence of Ang-2, a natural antagonist for the Tie-2 receptor. (MAISONPIERRE et al. 1997). Transgenic expression of Ang-2 disrupted blood vessel formation. The interrelation between the VEGF and Ang systems is likely to be an area of intense investigation in vascular biology.

An attractive possibility is that recombinant VEGF or gene therapy with the VEGF gene may be used to promote endothelial cell growth and collateral vessel formation. This would represent a novel therapeutic modality for conditions that frequently are refractory to conservative measures and unresponsive to pharmacological therapy. rhVEGF₁₆₅ is already in clinical trials for the treatment of myocardial ischemia associated with coronary artery disease.

The high expression of VEGF mRNA in human tumors, the presence of the VEGF protein in ocular fluids of individuals with proliferative retinopathies and in the synovial fluid of RA patients, as well as the localization of VEGF in AMD lesions, strongly supports the hypothesis that VEGF is a key mediator of angio-

genesis associated with various disorders. Therefore, anti-VEGF antibodies or other inhibitors of VEGF, used alone or in combination with other agents, may be of therapeutic value for a variety of malignancies and other disorders. Very recently, a humanized version of a high-affinity anti-VEGF monoclonal antibody, which retains the same affinity and efficacy as the original murine antibody, has been generated (PRESTA et al. 1997) and is being tested in humans as a treatment for solid tumors, alone or in combination with chemotherapy.

In conclusion, in spite of the plurality of factors potentially involved in angiogenesis, one specific factor, VEGF, appears to play an irreplaceable role in a variety of physiological and pathological circumstances.

References

- Adamis AP, Miller JW, Bernal MT, D'Amico DJ, Folkman J, Yeo TK, Yeo KT (1994) Increased vascular endothelial growth factor levels in the vitreous of eyes with proliferative diabetic retinopathy. *Am J Ophthalmol* 118:445-450
- Adamis AP, Shima DT, Tolentino MJ, Gragoudas ES, Ferrara N, Folkman J, D'Amore PA, Miller JW (1996) Inhibition of vascular endothelial growth factor prevents retinal ischemia-associated iris neovascularization in a nonhuman primate. *Arch Ophthalmol* 114:66-71
- Aiello LP, Avery RL, Arrigg PG, Keyt BA, Jampel HD, Shah ST, Pasquale LR, Thieme H, Iwamoto MA, Park JE, Nguyen H, Aiello LM, Ferrara N, King GL (1994) Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. *N Engl J Med* 331:1480-1487
- Aiello LP, Pierce EA, Foley ED, Takagi H, Chen H, Riddle L, Ferrara N, King GL, Smith LE (1995) Suppression of retinal neovascularization in vivo by inhibition of vascular endothelial growth factor (VEGF) using soluble VEGF-receptor chimeric proteins. *Proc Natl Acad Sci USA* 92:10457-10461
- Albini A, Benelli R, Presta M, Rusnati M, Ziche M, Rubartelli A, Paglialunga G, Bussolino F, Noonan D (1996a) HIV-tat protein is a heparin-binding angiogenic growth factor. *Oncogene* 12:289-297
- Albini A, Soldi R, Giunciuglio D, Giraudo E, Benelli R, Primo L, Noonan D, Salio M, Camussi G, Rockl W, Bussolino F (1996b) The angiogenesis induced by HIV-1 tat protein is mediated by the Flk-1/KDR receptor on vascular endothelial cells. *Nat Med* 2:1371-1375
- Algire GH, Chalkley HW (1945) Vascular reactions of normal and malignant tissues in vivo. I. Vascular reactions of mice to wounds and to normal and neoplastic transplants. *J Natl Cancer Inst* 6:73-85
- Alon T, Hemo I, Itin A, Pe'er J, Stone J, Keshet E (1995) Vascular endothelial growth factor acts as a survival factor for newly formed retinal vessels and has implications for retinopathy of prematurity. *Nat Med* 1:1024-1028
- Asahara T, Bauters C, Pastore C, Kearney M, Rossow S, Bunting S, Ferrara N, Symes JF, Isner JM (1995) Local delivery of vascular endothelial growth factor accelerates reendothelialization and attenuates intimal hyperplasia in balloon-injured rat carotid artery (see comments). *Circulation* 91:2793-2801
- Asano M, Yukita A, Matsumoto T, Kondo S, Suzuki H (1995) Inhibition of tumor growth and metastasis by an immunoneutralizing monoclonal antibody to human vascular endothelial growth factor/vascular permeability factor121. *Cancer Res* 55:5296-5301
- Banai S, Jaktlish MT, Shou M, Lazarous D, Scheinowitz M, Biro S, Epstein S, Unger E (1994a) Angiogenic-induced enhancement of collateral blood flow to ischemic myocardium by vascular endothelial growth factor. *Circulation* 89:2183-2189
- Banai S, Shweiki D, Pinson A, Chandra M, Lazarovich G, Keshet E (1994b) Upregulation of vascular endothelial growth factor expression induced by myocardial ischemia: implications for coronary angiogenesis. *Cardiovasc Res* 28:1176-1179
- Barleon B, Sozzani S, Zhou D, Weich HA, Mantovani A, Marme D (1996) Migration of human monocytes in response to vascular endothelial growth factor (VEGF) is mediated via the VEGF receptor flt-1. *Blood* 87:3336-3343

- Bauters C, Asahara T, Zheng LP, Takeshita S, Bunting S, Ferrara N, Symes JF, Isner JM (1994) Physiological assessment of augmented vascularity induced by VEGF in ischemic rabbit hindlimb. *Am J Physiol* 267:H1263-H1271
- Bauters C, Asahara T, Zheng LP, Takeshita S, Bunting S, Ferrara N, Symes JF, Isner JM (1995) Recovery of disturbed endothelium-dependent flow in the collateral-perfused rabbit ischemic hindlimb after administration of vascular endothelial growth factor. *Circulation* 91:2802-2809
- Ben-Av P, Crofford LJ, Wilder RL, Hia T (1995) Induction of vascular endothelial growth factor expression in synovial fibroblasts. *FEBS Lett* 372:83-87
- Berkman RA, Merrill MJ, Reinhold WC, Monacci WT, Saxena A, Clark WC, Robertson JT, Ali IU, Oldfield EH (1993) Expression of the vascular permeability factor/vascular endothelial growth factor gene in central nervous system neoplasms. *J Clin Invest* 91:153-159
- Borgstrom P, Hillan KJ, Sriramarao P, Ferrara N (1996) Complete inhibition of angiogenesis and growth of microtumors by anti-vascular endothelial growth factor neutralizing antibody: novel concepts of angiostatic therapy from intravital videomicroscopy. *Cancer Res* 56:4032-4039
- Borgstrom P, Hillan K, Sriramarao P, Ferrara N (1998a) Anti-Vascular endothelial growth factor antibody inhibits angiogenesis and growth of human prostate cancer in vivo. *Prostate* (in press)
- Borgstrom P, Hillan KJ, Sriramarao P, Ferrara N (1998b) Combination treatment with anti-vascular endothelial growth factor antibody and doxorubicin suppresses growth of breast carcinoma cells (submitted)
- Brandon EP, Idzerda RL, McKnight GS (1995) Targeting the mouse genome: a compendium of knockouts (part III). *Curr Biol* 5:873-881
- Breier G, Albrecht U, Sterrer S, Risau W (1992) Expression of vascular endothelial growth factor during embryonic angiogenesis and endothelial cell differentiation. *Development* 114:521-532
- Brogi E, Schatteman G, Wu T, Kim EA, Varticovski L, Keyt B, Isner JM (1996) Hypoxia-induced paracrine regulation of vascular endothelial growth factor receptor expression. *J Clin Invest* 97:469-476
- Brown LF, Berse B, Jackman RW, Tognazzi K, Manseau EJ, Dvorak HF, Senger DR (1993a) Increased expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in kidney and bladder carcinomas. *Am J Pathol* 143:1255-1262
- Brown LF, Berse B, Jackman RW, Tognazzi K, Manseau EJ, Senger DR, Dvorak HF (1993b) Expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in adenocarcinomas of the gastrointestinal tract. *Cancer Res* 53:4727-4735
- Brown LF, Berse B, Jackman RW, Tognazzi K, Guidi AJ, Dvorak HF, Senger DR, Connolly JL, Schnitt SJ (1995a) Expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in breast cancer. *Hum Pathol* 26:86-91
- Brown LF, Harrist TJ, Yeo KT, Stahle-Backdahl M, Jackman RW, Berse B, Tognazzi K, Dvorak HF, Detmar M (1995b) Increased expression of vascular permeability factor (vascular endothelial growth factor) in bullous pemphigoid, dermatitis herpetiformis, and erythema multiforme. *J Invest Dermatol* 104:744-749
- Broxmeyer HE, Cooper S, Li ZH, Lu L, Song HY, Kwon BS, Warren RE, Donner DB (1995) Myeloid progenitor cell regulatory effects of vascular endothelial cell growth factor. *Int J Hematol* 62:203-215
- Callow AD, Choi ET, Trachtenberg JD, Stevens SL, Connolly DT, Rodi C, Ryan US (1994) Vascular permeability factor accelerates endothelial regrowth following balloon angioplasty. *Growth Factors* 10:223-228
- Carmeliet P, Collen D (1997) Genetic analysis of blood vessel formation: role of endothelial versus smooth muscle cells. *Trends Cardiovasc Med* 8:271-281
- Carmeliet P, Ferreira V, Breier G, Pollefeyt S, Kieckens L, Gertsenstein M, Fahrig M, Vandenhoeck A, Harpal K, Eberhardt C, Declercq C, Pawling J, Moons L, Collen D, Risau W, Nagy A (1996) Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. *Nature* 380:435-439
- Chamow SM, Ashkenazi A (1996) Immunoadhesins: principles and applications. *Trends Biotechnol* 14:52-60
- Claffey KP, Wilkison WO, Spiegelman BM (1992) Vascular endothelial growth factor. Regulation by cell differentiation and activated second messenger pathways. *J Biol Chem* 267:16317-16322
- Clauss M, Gerlach M, Gerlach H, Brett J, Wang F, Familletti PC, Pan YC, Olander JV, Connolly DT, Stern D (1990) Vascular permeability factor: a tumor-derived polypeptide that induces endothelial cell and monocyte procoagulant activity, and promotes monocyte migration. *J Exp Med* 172:1535-1545
- Cohen T, Nahari D, Cerem LW, Neufeld G, Levi BZ (1996) Interleukin 6 induces the expression of vascular endothelial growth factor. *J Biol Chem* 271:736-741

- Conn G, Bayne ML, Soderman DD, Kwok PW, Sullivan KA, Palisi TM, Hope DA, Thomas KA (1990) Amino acid and cDNA sequences of a vascular endothelial cell mitogen that is homologous to platelet-derived growth factor. *Proc Natl Acad Sci USA* 87:2628–2632
- Cuevas P, Carceller F, Ortega S, Zazo M, Nieto I, Gimenez-Gallego G (1991) Hypotensive activity of fibroblast growth factor. *Science* 254:1208–1210
- Cuevas P, Garcia-Calvo M, Carceller F, Reimers D, Zazo M, Cuevas B, Munoz-Willery I, Martinez-Coso V, Lamas S, Gimenez-Gallego G (1996) Correction of hypertension by normalization of endothelial levels of fibroblast growth factor and nitric oxide synthase in spontaneously hypertensive rats. *Proc Natl Acad Sci USA* 93:11996–12001
- Cullinan-Bove K, Koos RD (1993) Vascular endothelial growth factor/vascular permeability factor expression in the rat uterus: rapid stimulation by estrogen correlates with estrogen-induced increases in uterine capillary permeability and growth. *Endocrinology* 133:829–837
- Cunningham SA, Waxham MN, Arrate PM, Brock TA (1995) Interaction of the Flt-1 tyrosine kinase receptor with the p85 subunit of phosphatidylinositol 3-kinase. Mapping of a novel site involved in binding. *J Biol Chem* 270:20254–20257
- Davis S, Aldrich TH, Jones PF, Acheson A, Compton DL, Jain V, Ryan TE, Bruno J, Radziejewski C, Maisonpierre PC, Yancopoulos GD (1996) Isolation of angiopoietin-1, a ligand for the TIE2 receptor, by secretion-trap expression cloning. *Cell* 87:1161–1169
- Davis-Smyth T, Chen H, Park J, Presta LG, Ferrara N (1996) The second immunoglobulin-like domain of the VEGF tyrosine kinase receptor Flt-1 determines ligand binding and may initiate a signal transduction cascade. *EMBO J* 15:4919–4927
- de Vries C, Escobedo JA, Ueno H, Houck K, Ferrara N, Williams LT (1992) The fms-like tyrosine kinase, a receptor for vascular endothelial growth factor. *Science* 255:989–991
- Detmar M, Brown LF, Claffey KP, Yeo KT, Kocher O, Jackman RW, Berse B, Dvorak HF (1994) Overexpression of vascular permeability factor/vascular endothelial growth factor and its receptors in psoriasis. *J Exp Med* 180:1141–1146
- DiPietro LA (1997) Thrombospondin as a regulator of angiogenesis. In: Rosen E, Goldberh I (eds) *Regulation of angiogenesis*. Springer, Berlin Heidelberg New York, pp 295–314
- Dvorak HF (1986) Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *N Engl J Med* 315:1650–1659
- Dvorak HF, Brown LF, Detmar M, Dvorak AM (1995) Vascular permeability factor/vascular endothelial growth factor, microvascular hyperpermeability, and angiogenesis. *Am J Pathol* 146:1029–1039
- Dvorak HF, Harvey VS, Estrella P, Brown LF, McDonagh J, Dvorak AM (1987) Fibrin containing gels induce angiogenesis. Implications for tumor stroma generation and wound healing. *Lab Invest* 57:673–686
- Essed CD, Brand MVD, Becker AE (1983) Transluminal coronary angioplasty and early restenosis. *Br Heart J* 49:393–402
- Fassbender HJ, Simling-Annenfeld M (1983) The potential aggressiveness of synovial tissue in rheumatoid arthritis. *J Pathol* 10:845–851
- Fava RA, Olsen NJ, Spencer-Green G, Yeo KT, Yeo TK, Berse B, Jackman RW, Senger DR, Dvorak HF, Brown LF (1994) Vascular permeability factor/endothelial growth factor (VPF/VEGF): accumulation and expression in human synovial fluids and rheumatoid synovial tissue. *J Exp Med* 180:341–346
- Ferrara N, Davis-Smyth T (1997) The biology of vascular endothelial growth factor. *Endocr Rev* 18:4–25
- Ferrara N, Henzel WJ (1989) Pituitary follicular cells secrete a novel heparin-binding growth factor specific for vascular endothelial cells. *Biochem Biophys Res Commun* 161:851–858
- Ferrara N, Clapp C, Weiner R (1991) The 16 K fragment of prolactin specifically inhibits basal or fibroblast growth factor stimulated growth of capillary endothelial cells. *Endocrinology* 129:896–900
- Ferrara N, Houck K, Jakeman L, Leung DW (1992) Molecular and biological properties of the vascular endothelial growth family of proteins. *Endocr Rev* 13:18–32
- Ferrara N, Winer J, Burton T, Rowland A, Siegel M, Phillips HS, Terrell T, Keller GA, Levinson AD (1993) Expression of vascular endothelial growth factor does not promote transformation but confers a growth advantage in vivo to Chinese hamster ovary cells. *J Clin Invest* 91:160–170
- Ferrara N, Carver Moore K, Chen H, Dowd M, Lu L, O'Shea KS, Powell Braxton L, Hillan KJ, Moore MW (1996) Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature* 380:439–442
- Ferrara N, Chen H, Davis-Smyth T, Gerber H-P, Nguyen T-N, Peers D, Chisholm V, Hillan KJ, Schwall RH (1998) Vascular endothelial growth factor is essential for corpus luteum angiogenesis (submitted)

- Finnerty H, Kelleher K, Morris GE, Bean K, Merberg DM, Kriz R, Morris JC, Sookdeo H, Turner KJ, Wood CR (1993) Molecular cloning of murine FLT and FLT4. *Oncogene* 8:2293–2298
- Folkman J (1971) Tumor angiogenesis: therapeutic implications. *N Engl J Med* 285:1182–1186
- Folkman J (1995) Angiogenesis in cancer, vascular, rheumatoid and other disease. *Nat Med* 1:27–31
- Folkman J, Shing Y (1992) Angiogenesis. *J Biol Chem* 267:10931–10934
- Fong GH, Rossant J, Gertsenstein M, Breitman ML (1995) Role of the Flt-1 receptor tyrosine kinase in regulating the assembly of vascular endothelium. *Nature* 376:66–70
- Forsythe JA, Jiang BH, Iyer NV, Agani F, Leung SW, Koos RD, Semenza GL (1996) Activation of vascular endothelial growth factor gene transcription by hypoxia-inducible factor 1. *Mol Cell Biol* 16:4604–4613
- Frank S, Hubner G, Breier G, Longaker MT, Greenhaig DG, Werner S (1995) Regulation of VEGF expression in cultured keratinocytes. Implications for normal and impaired wound healing. *J Biol Chem* 270:12607–12613
- Friedlander M, Brooks PC, Shaffer RW, Kincaid CM, Varner JA, Cheresch DA (1995) Definition of two angiogenic pathways by distinct alpha v integrins. *Science* 270:1500–1502
- Gabrilovich DI, Chen HL, Girgis KR, Cunningham HT, Meny GM, Nadaf S, Kavanaugh D, Carbone DP (1996) Production of vascular endothelial growth factor by human tumors inhibits the functional maturation of dendritic cells. *Nat Med* 2:1096–1103
- Galland F, Karamysheva A, Mattei MG, Rosnet O, Marchetto S, Birnbaum D (1992) Chromosomal localization of FLT4, a novel receptor-type tyrosine kinase gene. *Genomics* 13:475–478
- Garner A (1994) Vascular diseases. Pathobiology of ocular disease. Dekker, New York
- Gasparini G, Toi M, Gion M, Verderio P, Dittadi R, Hanatani M, Matsubara I, Vinante O, Bonoldi E, Boracchi P, Gatti C, Suzuki H, Tominaga T (1997) Prognostic significance of vascular endothelial growth factor protein in node-negative breast carcinoma. *J Natl Cancer Inst* 89:139–147
- Gerber HP, Condorelli F, Park J, Ferrara N (1997) Differential transcriptional regulation of the two VEGF receptor genes. Flt-1, but not Flk-1/KDR, is up-regulated by hypoxia. *J Biol Chem* 272:23659–23667
- Gnarra JR, Zhou S, Merrill MJ, Wagner JR, Krumm A, Papavassiliou E, Oldfield EH, Klausner RD, Linehan WM (1996) Post-transcriptional regulation of vascular endothelial growth factor mRNA by the product of the VHL tumor suppressor gene. *Proc Natl Acad Sci USA* 93:10589–10594
- Goldberg MA, Schneider TJ (1994) Similarities between the oxygen-sensing mechanisms regulating the expression of vascular endothelial growth factor and erythropoietin. *J Biol Chem* 269:4355–4361
- Goldman C, Kim J, Wenf W-L, King V, Brock T, Gillespie Y (1993) Epidermal growth factor stimulates vascular endothelial growth factor production by malignant glioma cells. A model of glioblastoma multiforme pathophysiology. *Mol Biol Cell* 4:121–133
- Good D, Polverini P, Rastinejad F, Beau M, Lemons R, Frazier W, Bouck N (1990) A tumor suppressor-dependent inhibitor of angiogenesis is immunologically and functionally indistinguishable from a fragment of thrombospondin. *Proc Natl Acad Sci USA* 87:6624–6628
- Graor RA, Gray BH (1991) Interventional treatment of peripheral vascular disease. *Peripheral vascular diseases*. Mosby, St Louis
- Grugel S, Finkenzeller G, Weindel K, Barleon B, Marme D (1995) Both v-Ha-Ras and v-Raf stimulate expression of the vascular endothelial growth factor in NIH 3T3 cells. *J Biol Chem* 270:25915–25919
- Guidi AJ, Abu-Jawdeh G, Berse B, Jackman RW, Tognazzi K, Dvorak HF, Brown LF (1995) Vascular permeability factor (vascular endothelial growth factor) expression and angiogenesis in cervical neoplasia. *J Natl Cancer Inst* 87:1237–1245
- Guidi AJ, Abu-Jawdeh G, Tognazzi K, Dvorak HF, Brown LF (1996) Expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in endometrial carcinoma. *Cancer* 78:454–460
- Guo D, Jia Q, Song HY, Warren RS, Donner DB (1995) Vascular endothelial cell growth factor promotes tyrosine phosphorylation of mediators of signal transduction that contain SH2 domains. Association with endothelial cell proliferation. *J Biol Chem* 270:6729–6733
- Harada K, Friedman M, Lopez JJ, Wang SY, Li J, Prasad PV, Pearlman JD, Edelman ER, Sellke FW, Simons M (1996) Vascular endothelial growth factor administration in chronic myocardial ischemia. *Am J Physiol* 270:H1791–H1802
- Hashimoto E, Ogita T, Nakaoka T, Matsuoka R, Takao A, Kira Y (1994) Rapid induction of vascular endothelial growth factor expression by transient ischemia in rat heart. *Am J Physiol* 267:H1948–H1954
- Hashimoto M, Ohsawa M, Ohnishi A, Naka N, Hirota S, Kitamura Y, Aozasa K (1995) Expression of vascular endothelial growth factor and its receptor mRNA in angiosarcoma. *Lab Invest* 73:859–863

- Houck KA, Ferrara N, Winer J, Cachianes G, Li B, Leung DW (1991) The vascular endothelial growth factor family: identification of a fourth molecular species and characterization of alternative splicing of RNA. *Mol Endocrinol* 5:1806-1814
- Houck KA, Leung DW, Rowland AM, Winer J, Ferrara N (1992) Dual regulation of vascular endothelial growth factor bioavailability by genetic and proteolytic mechanisms. *J Biol Chem* 267:26031-26037
- Ikeda E, Achen MG, Breier G, Risau W (1995) Hypoxia-induced transcriptional activation and increased mRNA stability of vascular endothelial growth factor in C6 glioma cells. *J Biol Chem* 270:19761-19766
- Iliopoulos O, Levy AP, Jiang C, Kaelin WG Jr, Goldberg MA (1996) Negative regulation of hypoxia-inducible genes by the von Hippel-Lindau protein. *Proc Natl Acad Sci USA* 93:10595-10599
- Isner JM, Pieczek A, Schainfeld R, Blair R, Haley L, Asahara T, Rosenfield K, Razvi S, Walsh K, Symes JF (1996a) Clinical evidence of angiogenesis after arterial gene transfer of phVEGF165 in patient with ischaemic limb (see comments). *Lancet* 348:370-374
- Isner JM, Walsh K, Symes J, Pieczek A, Takeshita S, Lowry J, Rosenfield K, Weir L, Brogi E, Juraj D (1996b) Arterial gene transfer for therapeutic angiogenesis in patients with peripheral artery disease. *Hum Gene Ther* 7:959-988
- Jakeman LB, Winer J, Bennett GL, Altar CA, Ferrara N (1992) Binding sites for vascular endothelial growth factor are localized on endothelial cells in adult rat tissues. *J Clin Invest* 89:244-253
- Jakeman LB, Armanini M, Phillips HS, Ferrara N (1993) Developmental expression of binding sites and mRNA for vascular endothelial growth factor suggests a role for this protein in vasculogenesis and angiogenesis. *Endocrinology* 133:848-859
- Kamat BR, Brown LF, Mansueti EJ, Senger DR, Dvorak HF (1995) Expression of vascular permeability factor/vascular endothelial growth factor by human granulosa and theca lutein cells. Role in corpus luteum development. *Am J Pathol* 146:157-165
- Kendall RL, Wang G, Thomas KA (1996) Identification of a natural soluble form of the vascular endothelial growth factor receptor, FLT-1, and its heterodimerization with KDR. *Biochem Biophys Res Commun* 226:324-328
- Keyt BA, Berleau LT, Nguyen HV, Chen H, Heinsohn H, Vandlen R, Ferrara N (1996a) The carboxyl-terminal domain (111-165) of vascular endothelial growth factor is critical for its mitogenic potency. *J Biol Chem* 271:7788-7795
- Keyt BA, Nguyen HV, Berleau LT, Duarte CM, Park J, Chen H, Ferrara N (1996b) Identification of vascular endothelial growth factor determinants for binding KDR and FLT-1 receptors. Generation of receptor-selective VEGF variants by site-directed mutagenesis. *J Biol Chem* 271:5638-5646
- Kieser A, Weich H, Brandner G, Marme D, Kolch W (1994) Mutant p53 potentiates protein kinase C induction of vascular endothelial growth factor expression. *Oncogene* 9:963-969
- Kim KJ, Li B, Houck K, Winer J, Ferrara N (1992) The vascular endothelial growth factor proteins: identification of biologically relevant regions by neutralizing monoclonal antibodies. *Growth Factors* 7:53-64
- Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N (1993) Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumor growth in vivo. *Nature* 362:841-844
- Koch AE, Harlow L, Haines GK, Amento EP, Unemori EN, Wong W-L, Pope RM, Ferrara N (1994) Vascular endothelial growth factor: a cytokine modulating endothelial function in rheumatoid arthritis. *J Immunol* 152:4149-4156
- Kondo S, Asano M, Matsuo K, Ohmori I, Suzuki H (1994) Vascular endothelial growth factor/vascular permeability factor is detectable in the sera of tumor-bearing mice and cancer patients. *Biochim Biophys Acta* 1221:211-214
- Ku DD, Zaleski JK, Liu S, Brock TA (1993) Vascular endothelial growth factor induces EDRF-dependent relaxation in coronary arteries. *Am J Physiol* 265:H586-H592
- Kuhn R, Schwenk F, Aguett M, Rajewsky K (1995) Inducible gene targeting in mice. *Science* 269:1427-1429
- Kvanta A, Algvere PV, Berglin L, Seregard S (1996) Subfoveal fibrovascular membranes in age-related macular degeneration express vascular endothelial growth factor. *Invest Ophthalmol Vis Sci* 37:1929-1934
- Leung DW, Cachianes G, Kuang WJ, Goeddel DV, Ferrara N (1989) Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science* 246:1306-1309
- Levy AP, Levy NS, Wegner S, Goldberg MA (1995) Transcriptional regulation of the rat vascular endothelial growth factor gene by hypoxia. *J Biol Chem* 270:13333-13340

- Levy AP, Levy NS, Goldberg MA (1996) Post-transcriptional regulation of vascular endothelial growth factor by hypoxia. *J Biol Chem* 271:2746-2753
- Li J, Perrella MA, Tsai JC, Yet SF, Hsieh CM, Yoshizumi M, Patterson C, Endego WO, Zhou F, Lee M (1995) Induction of vascular endothelial growth factor gene expression by interleukin-1 beta in rat aortic smooth muscle cells. *J Biol Chem* 270:308-312
- Li J, Brown LF, Hibberd MG, Grossman JD, Morgan JP, Simons M (1996) VEGF, flk-1, and flt-1 expression in a rat myocardial infarction model of angiogenesis. *Am J Physiol* 270:H1803-H1811
- Liu Y, Cox SR, Morita T, Kourembanas S (1995) Hypoxia regulates vascular endothelial growth factor gene expression in endothelial cells. Identification of a 5' enhancer. *Circ Res* 77:638-643
- Lopez PF, Sippy BD, Lambert HM, Thach AB, Hinton DR (1996) Transdifferentiated retinal pigment epithelial cells are immunoreactive for vascular endothelial growth factor in surgically excised age-related macular degeneration-related choroidal neovascular membranes. *Invest Ophthalmol Vis Sci* 37:855-868
- Madan A, Curtin PT (1993) A 24-base-pair sequence 3' to the human erythropoietin gene contains a hypoxia-responsive transcriptional enhancer. *Proc Natl Acad Sci USA* 90:3928-3932
- Maeda K, Chung YS, Ogawa Y, Takatsuka S, Kang SM, Ogawa M, Sawada T, Sowa M (1996) Prognostic value of vascular endothelial growth factor expression in gastric carcinoma. *Cancer* 77:858-863
- Maisonpierre PC, Suri C, Jones PF, Bartunkova S, Wiegand SJ, Radziejewski C, Compton D, McClain J, Aldrich TH, Papadopoulos N, Daly TJ, Davis S, Sato TN, Yancopoulos GD (1997) Angiopoietin-2, a natural antagonist for Tie-2 that disrupts in vivo angiogenesis. *Science* 277:55-60
- Malecaze F, Clemens S, Simorer-Pinotel V, Mathis A, Chollet P, Favard P, Bayard F, Plouet J (1994) Detection of vascular endothelial growth factor mRNA and vascular endothelial growth factor-like activity in proliferative diabetic retinopathy. *Arch Ophthalmol* 112:1476-1482
- Mandriota SJ, Seghezzi G, Vassalli JD, Ferrara N, Wasi S, Mazzieri R, Mignatti P, Pepper MS (1995) Vascular endothelial growth factor increases urokinase receptor expression in vascular endothelial cells. *J Biol Chem* 270:9709-9716
- Mandriota SJ, Menoud PA, Pepper MS (1996) Transforming growth factor beta 1 down-regulates vascular endothelial growth factor receptor 2/flk-1 expression in vascular endothelial cells. *J Biol Chem* 271:11500-11505
- Matthews W, Jordan CT, Gavin M, Jenkins NA, Copeland NG, Lemischka IR (1991) A receptor tyrosine kinase cDNA isolated from a population of enriched primitive hematopoietic cells and exhibiting close genetic linkage to c-kit. *Proc Natl Acad Sci USA* 88:9026-9030
- Mazure NM, Chen EY, Yeh P, Laderoute KR, Giaccia AJ (1996) Oncogenic transformation and hypoxia synergistically act to modulate vascular endothelial growth factor expression. *Cancer Res* 56:3436-3440
- McLaren J, Prentice A, Charnock-Jones DS, Smith SK (1996) Vascular endothelial growth factor (VEGF) concentrations are elevated in peritoneal fluid of women with endometriosis. *Hum Reprod* 11:220-223
- Melder RJ, Koenig GC, Witwer BP, Safabakhsh N, Munn LL, Jain RK (1996) During angiogenesis, vascular endothelial growth factor and basic fibroblast growth factor regulate natural killer cell adhesion to tumor endothelium (see comments). *Nat Med* 2:992-997
- Melnyk O, Shuman MA, Kim KJ (1996) Vascular endothelial growth factor promotes tumor dissemination by a mechanism distinct from its effect on primary tumor growth. *Cancer Res* 56:921-924
- Michaelson IC (1948) The mode of development of the vascular system of the retina with some observations on its significance for certain retinal disorders. *Trans Ophthalmol Soc UK* 68:137-180
- Millauer B, Witzmann Voos S, Schnurch H, Martinez R, Moller NP, Risau W, Ullrich A (1993) High affinity VEGF binding and developmental expression suggest Flk-1 as a major regulator of vasculogenesis and angiogenesis. *Cell* 72:835-846
- Millauer B, Shawver LK, Plate KH, Risau W, Ullrich A (1994) Glioblastoma growth inhibited in vivo by a dominant-negative Flk-1 mutant. *Nature* 367:576-579
- Miller JW, Adamis AP, Shima DT, D'Amore PA, Moulton RS, O'Reilly MS, Folkman J, Dvorak HF, Brown LF, Berse B et al (1994) Vascular endothelial growth factor/vascular permeability factor is temporally and spatially correlated with ocular angiogenesis in a primate model. *Am J Pathol* 145:574-584
- Minchenko A, Bauer T, Salceda S, Caro J (1994) Hypoxic stimulation of vascular endothelial growth factor expression in vivo and in vitro. *Lab Invest* 71:374-379
- Monacci WT, Merrill MJ, Oldfield EH (1993) Expression of vascular permeability factor/vascular endothelial growth factor in normal rat tissues. *Am J Physiol* 264:C995-C1002

- Morbidegli L, Chang CH, Douglas JG, Granger HJ, Ledda F, Ziche M (1996) Nitric oxide mediates mitogenic effect of VEGF on coronary venular endothelium. *Am J Physiol* 270:H411–H415
- Morishita K, Johnson DE, Williams LT (1995) A novel promoter for vascular endothelial growth factor receptor (flt-1) that confers endothelial-specific gene expression. *J Biol Chem* 270:27948–27953
- Muller YA, Li B, Christinger HW, Wells JA, Cunningham BC, de Vos AM (1997) Vascular endothelial growth factor: crystal structure and functional mapping of the kinase domain receptor binding site. *Proc Natl Acad Sci USA* 94:7192–7197
- Nicosia R, Nicosia SV, Smith M (1994) Vascular endothelial growth factor, platelet-derived growth factor and indulin-like growth factor stimulate angiogenesis in vitro. *Am J Pathol* 145:1023–1029
- O'Reilly MS, Holmgren L, Shing Y, Chen C, Rosenthal RA, Moses M, Lane WS, Cao Y, Sage EH, Folkman J (1994) Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma (see comments). *Cell* 79:315–328
- O'Reilly MS, Boehm T, Shing Y, Vasios G, Lane WS, Flynn E, Birkhead JR, Olsen BR, Folkman J (1997) Endostatin: an endogenous inhibitor of angiogenesis and tumor growth. *Cell* 88:277–285
- Oik RJ, Lee CM (1993) Diabetic retinopathy: practical management. Lippincott, Philadelphia
- Olson TA, Mohanraj D, Carson LF, Ramakrishnan S (1994) Vascular permeability factor gene expression in normal and neoplastic human ovaries. *Cancer Res* 54:276–280
- Pajusola K, Aprelikova O, Korhonen J, Kaipainen A, Pertovaara L, Alitalo R, Alitalo K (1992) FLT4 receptor tyrosine kinase contains seven immunoglobulin-like loops and is expressed in multiple human tissues and cell lines. *Cancer Res* 52:5738–5743
- Park JE, Keller H-A, Ferrara N (1993) The vascular endothelial growth factor isoforms (VEGF): differential deposition into the subepithelial extracellular matrix and bioactivity of extracellular matrix-bound VEGF. *Mol Biol Cell* 4:1317–1326
- Park JE, Chen HH, Winer J, Houck KA, Ferrara N (1994) Placenta growth factor. Potentiation of vascular endothelial growth factor bioactivity, in vitro and in vivo, and high affinity binding to Flt-1 but not to Flk-1/KDR. *J Biol Chem* 269:25646–25654
- Patterson C, Perrella MA, Hsieh CM, Yoshizumi M, Lee ME, Haber E (1995) Cloning and functional analysis of the promoter for KDR/flk-1, a receptor for vascular endothelial growth factor. *J Biol Chem* 270:23111–23118
- Patterson C, Perrella MA, Endege WO, Yoshizumi M, Lee ME, Haber E (1996) Downregulation of vascular endothelial growth factor receptors by tumor necrosis factor-alpha in cultured human vascular endothelial cells. *J Clin Invest* 98:490–496
- Patz A (1980) Studies on retinal neovascularization. *Invest Ophthalmol Vis Sci* 19:1133–1138
- Pearlman JD, Hibberd MG, Chuang ML, Harada K, Lopez JJ, Gladstone SR, Friedman M, Selke FW, Simons M (1995) Magnetic resonance mapping demonstrates benefits of VEGF-induced myocardial angiogenesis. *Nat Med* 1:1085–1089
- Pe'er J, Folberg R, Itin A, Gnessin H, Hemo I, Keshet E (1996) Upregulated expression of vascular endothelial growth factor in proliferative diabetic retinopathy. *Br J Ophthalmol* 80:241–245
- Pekala P, Marlow M, Heuvelman D, Connolly D (1990) Regulation of hexose transport in aortic endothelial cells by vascular permeability factor and tumor necrosis factor-alpha, but not by insulin. *J Biol Chem* 265:18051–18054
- Pepper MS, Ferrara N, Orci L, Montesano R (1991) Vascular endothelial growth factor (VEGF) induces plasminogen activators and plasminogen activator inhibitor-1 in microvascular endothelial cells. *Biochem Biophys Res Commun* 181:902–906
- Pepper MS, Ferrara N, Orci L, Montesano R (1992) Potent synergism between vascular endothelial growth factor and basic fibroblast growth factor in the induction of angiogenesis in vitro. *Biochem Biophys Res Commun* 189:824–831
- Pertovaara L, Kaipainen A, Mustonen T, Orpana A, Ferrara N, Saksela O, Alitalo K (1994) Vascular endothelial growth factor is induced in response to transforming growth factor-beta in fibroblastic and epithelial cells. *J Biol Chem* 269:6271–6274
- Peters KG, De Vries C, Williams LT (1993) Vascular endothelial growth factor receptor expression during embryogenesis and tissue repair suggests a role in endothelial differentiation and blood vessel growth. *Proc Natl Acad Sci USA* 90:8915–8919
- Phillips HS, Hains J, Leung DW, Ferrara N (1990) Vascular endothelial growth factor is expressed in rat corpus luteum. *Endocrinology* 127:965–967
- Phillips HS, Armanini M, Stavrou D, Ferrara N, Westphal M (1993) Intense focal expression of vascular endothelial growth factor mRNA in human intracranial neoplasms: Association with regions of necrosis. *Int J Oncol* 2:913–919

- Pierce EA, Avery RL, Foley ED, Aiello LP, Smith LE (1995) Vascular endothelial growth factor/vascular permeability factor expression in a mouse model of retinal neovascularization. *Proc Natl Acad Sci USA* 92:905-909
- Plate KH, Breier G, Weich HA, Risau W (1992) Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo. *Nature* 359:845-848
- Plate KH, Breier G, Millauer B, Ullrich A, Risau W (1993) Up-regulation of vascular endothelial growth factor and its cognate receptors in a rat glioma model of tumor angiogenesis. *Cancer Res* 53:5822-5827
- Plouet J, Moukadir HJ (1990) Characterization of the receptors for vasculotropin on bovine adrenal cortex-derived capillary endothelial cells. *J Biol Chem* 265:22071-22075
- Poltorak Z, Cohen T, Sivan R, Kandelis Y, Spira G, Vlodaysky I, Keshet E, Neufeld G (1997) VEGF145, a secreted vascular endothelial growth factor isoform that binds to extracellular matrix. *J Biol Chem* 272:7151-7158
- Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Humanization of an anti-VEGF monoclonal antibody for the therapy of solid tumors and other disorders. *Cancer Res* 57:4593-4599
- Qu H, Nagy JA, Senger DR, Dvorak HF, Dvorak AM (1995) Ultrastructural localization of vascular permeability factor/vascular endothelial growth factor (VPF/VEGF) to the albuminal plasma membrane and vesiculovacuolar organelles of tumor microvascular endothelium. *J Histochem Cytochem* 43:381-389
- Quinn TP, Peters KG, De Vries C, Ferrara N, Williams LT (1993) Fetal liver kinase 1 is a receptor for vascular endothelial growth factor and is selectively expressed in vascular endothelium. *Proc Natl Acad Sci USA* 90:7533-7537
- Rak J, Mitsuhashi Y, Bayko L, Filmus J, Shirasawa S, Sasazuki T, Kerbel RS (1995) Mutant ras oncogenes upregulate VEGF/VPF expression: implications for induction and inhibition of tumor angiogenesis. *Cancer Res* 55:4575-4580
- Ravindranath N, Little-Ihrig L, Phillips HS, Ferrara N, Zeleznik AJ (1992) Vascular endothelial growth factor messenger ribonucleic acid expression in the primate ovary. *Endocrinology* 131:254-260
- Risau W (1997) Mechanisms of angiogenesis. *Nature* 386:671-674
- Roberts WG, Palade GE (1995) Increased microvascular permeability and endothelial fenestration induced by vascular endothelial growth factor. *J Cell Sci* 108:2369-2379
- Roberts WG, Palade GE (1997) Neovasculature induced by vascular endothelial growth factor is fenestrated. *Cancer Res* 57:765-772
- Sato K, Yamazaki K, Shizume K, Kanaji Y, Obara T, Ohsumi K, Demura H, Yamaguchi S, Shibuya M (1995) Stimulation by thyroid-stimulating hormone and Grave's immunoglobulin G of vascular endothelial growth factor mRNA expression in human thyroid follicles in vitro and ft mRNA expression in the rat thyroid in vivo. *J Clin Invest* 96:1295-1302
- Seetharam L, Gotoh N, Maru Y, Neufeld G, Yamaguchi S, Shibuya M (1995) A unique signal transduction from FLT tyrosine kinase, a receptor for vascular endothelial growth factor VEGF. *Oncogene* 10:135-147
- Shalaby F, Rossant J, Yamaguchi TP, Gertsenstein M, Wu XF, Breitman ML, Schuh AC (1995) Failure of blood-island formation and vasculogenesis in Flk-1-deficient mice. *Nature* 376:62-66
- Shen H, Clauss M, Ryan J, Schmidt AM, Tjiburg P, Borden L, Connolly D, Stern D, Kao J (1993) Characterization of vascular permeability factor/vascular endothelial growth factor receptors on mononuclear phagocytes. *Blood* 81:2767-2773
- Shibuya M, Yamaguchi S, Yamane A, Ikeda T, Tojo A, Matsushime H, Sato M (1990) Nucleotide sequence and expression of a novel human receptor-type tyrosine kinase (flt) closely related to the fms family. *Oncogene* 5:519-527
- Shifren JL, Doldi N, Ferrara N, Mesiano S, Jaffe RB (1994) In the human fetus, vascular endothelial growth factor is expressed in epithelial cells and myocytes, but not vascular endothelium: implications for mode of action. *J Clin Endocrinol Metab* 79:316-322
- Shifren JL, Tseng JF, Zaloudek CJ, Ryan IP, Meng YG, Ferrara N, Jaffe RB, Taylor RN (1996) Ovarian steroid regulation of vascular endothelial growth factor in the human endometrium: implications for angiogenesis during the menstrual cycle and in the pathogenesis of endometriosis. *J Clin Endocrinol Metab* 81:3112-3118
- Shima DT, Adamis AP, Ferrara N, Yeo KT, Yeo TK, Allende R, Folkman J, D'Amore PA (1995) Hypoxic induction of endothelial cell growth factors in retinal cells: identification and characterization of vascular endothelial growth factor (VEGF) as the mitogen. *Mol Med* 1:182-193
- Shima DT, Kuroki M, Deutsch U, Ng YS, Adamis AP, D'Amore PA (1996) The mouse gene for vascular endothelial growth factor. Genomic structure, definition of the transcriptional unit, and

- characterization of transcriptional and post-transcriptional regulatory sequences. *J Biol Chem* 271:3877-3883
- Shweiki D, Itin A, Soffer D, Keshet E (1992) Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. *Nature* 359:843-845
- Shweiki D, Itin A, Neufeld G, Gitay-Goren H, Keshet E (1993) Patterns of expression of vascular endothelial growth factor (VEGF) and VEGF receptors in mice suggest a role in hormonally-mediated angiogenesis. *J Clin Invest* 91:2235-2243
- Siemeister G, Weindel K, Mohrs K, Barleon B, Martiny-Baron G, Marme D (1996) Reversion of deregulated expression of vascular endothelial growth factor in human renal carcinoma cells by von Hippel-Lindau tumor suppressor protein. *Cancer Res* 56:2299-2301
- Stone J, Itin A, Alon T, Pe'er J, Gnessin H, Chan Ling T, Keshet E (1995) Development of retinal vasculature is mediated by hypoxia-induced vascular endothelial growth factor (VEGF) expression by neuroglia. *J Neurosci* 15:4738-4747
- Suri C, Jones PF, Patan S, Bartunkova S, Maisonpierre PC, Davis S, Sato TN, Yancopoulos GD (1996) Requisite role of angiopoietin-1, a ligand for the TIE2 receptor, during embryonic angiogenesis. *Cell* 87:1171-1180
- Suzuki K, Hayashi N, Miyamoto Y, Yamamoto M, Ohkawa K, Ito Y, Sasaki Y, Yamaguchi Y, Nakase H, Noda K, Enomoto N, Arai K, Yamada Y, Yoshihara H, Tujimura T, Kawano K, Yoshikawa K, Kamada T (1996) Expression of vascular permeability factor/vascular endothelial growth factor in human hepatocellular carcinoma. *Cancer Res* 56:3004-3009
- Takagi H, King GL, Ferrara N, Aiello LP (1996) Hypoxia regulates vascular endothelial growth factor receptor KDR/Flk gene expression through adenosine A2 receptors in retinal capillary endothelial cells. *Invest Ophthalmol Vis Sci* 37:1311-1321
- Takeshita S, Zhung L, Brogi E, Kearney M, Pu L-Q, Bunting S, Ferrara N, Symes JF, Isner JM (1994) Therapeutic angiogenesis: a single intra-arterial bolus of vascular endothelial growth factor augments collateral vessel formation in a rabbit ischemic hind-limb model. *J Clin Invest* 93:662-670
- Takeshita S, Tsurumi Y, Couffinhal T, Asahara T, Bauters C, Symes J, Ferrara N, Isner JM (1996a) Gene transfer of naked DNA encoding for three isoforms of vascular endothelial growth factor stimulates collateral development in vivo. *Lab Invest* 75:487-501
- Takeshita S, Weir L, Chen D, Zheng LP, Riessen R, Bauters C, Symes JF, Ferrara N, Isner JM (1996b) Therapeutic angiogenesis following arterial gene transfer of vascular endothelial growth factor in a rabbit model of hindlimb ischemia. *Biochem Biophys Res Commun* 227:628-635
- Terman BI, Carrion ME, Kovacs E, Rasmussen BA, Eddy RL, Shows TB (1991) Identification of a new endothelial cell growth factor receptor tyrosine kinase. *Oncogene* 6:1677-1683
- Terman BI, Dougher Vermazen M, Carrion ME, Dimitrov D, Armellino DC, Gospodarowicz D, Bohlen P (1992) Identification of the KDR tyrosine kinase as a receptor for vascular endothelial cell growth factor. *Biochem Biophys Res Commun* 187:1579-1586
- Tischer E, Mitchell R, Hartman T, Silva M, Gospodarowicz D, Fiddes JC, Abraham JA (1991) The human gene for vascular endothelial growth factor. Multiple protein forms are encoded through alternative exon splicing. *J Biol Chem* 266:11947-11954
- Toi M, Kondo S, Suzuki H, Yamamoto Y, Inada K, Imazawa T, Taniguchi T, Tominaga T (1996) Quantitative analysis of vascular endothelial growth factor in primary breast cancer. *Cancer* 77:1101-1106
- Tolentino MJ, Miller JW, Gragoudas ES, Chatzistefanou K, Ferrara N, Adamis AP (1996) Vascular endothelial growth factor is sufficient to produce iris neovascularization and neovascular glaucoma in a nonhuman primate. *Arch Ophthalmol* 114:964-970
- Tuder RM, Flook BE, Voelkel NF (1995) Increased gene expression for VEGF and the VEGF receptors KDR/Flk and Flt in lungs exposed to acute or to chronic hypoxia. Modulation of gene expression by nitric oxide. *J Clin Invest* 95:1798-1807
- Unemori EN, Ferrara N, Bauer EA, Amento EP (1992) Vascular endothelial growth factor induces interstitial collagenase expression in human endothelial cells. *J Cell Physiol* 153:557-562
- Vaisman N, Gospodarowicz D, Neufeld G (1990) Characterization of the receptors for vascular endothelial growth factor. *J Biol Chem* 265:19461-19466
- Viglietto G, Romano A, Maglione D, Rambaldi M, Paoletti I, Lago CT, Califano D, Monaco C, Mineo A, Santelli G, Manzo G, Botti G, Chiappetta G, Persico MG (1996) Neovascularization in human germ cell tumors correlates with a marked increase in the expression of the vascular endothelial growth factor but not the placenta-derived growth factor. *Oncogene* 13:577-587
- Vincenti V, Cassano C, Recchi M, Persico G (1996) Assignment of the vascular endothelial growth factor gene to human chromosome 6p21.3. *Circulation* 93:1493-1495

- Voim M, Koomagi R, Mattern J (1997a) Prognostic value of vascular endothelial growth factor and its receptor Flt-1 in squamous cell lung cancer. *Int J Cancer* 74:64-68
- Voim M, Koomagi R, Mattern J, Stammler G (1997b) Angiogenic growth factors and their receptors in non-small cell lung carcinomas and their relationships to drug response in vitro. *Anticancer Res* 17:99-103
- Walder CE, Errett CJ, Bunting S, Lindquist P, Ogez JR, Heinsohn HG, Ferrara N, Thomas GR (1996) Vascular endothelial growth factor augments muscle blood flow and function in a rabbit model of chronic hindlimb ischemia. *J Cardiovasc Pharmacol* 27:91-98
- Waltenberger J, Claesson Welsh L, Siegbahn A, Shibuya M, Heldin CH (1994) Different signal transduction properties of KDR and Flt1, two receptors for vascular endothelial growth factor. *J Biol Chem* 269:26988-26995
- Wang GL, Semenza GL (1995) Purification and characterization of hypoxia-inducible factor 1. *J Biol Chem* 270:1230-1237
- Ware JA, Simons M (1997) Angiogenesis in ischemic heart disease. *Nat Med* 3:158-164
- Warren RS, Yuan H, Matli MR, Gillett NA, Ferrara N (1995) Regulation by vascular endothelial growth factor of human colon cancer tumorigenesis in a mouse model of experimental liver metastasis. *J Clin Invest* 95:1789-1797
- Warren RS, Yuan H, Matli MR, Ferrara N, Donner DB (1996) Induction of vascular endothelial growth factor by insulin-like growth factor 1 in colorectal carcinoma. *J Biol Chem* 271:29483-29488
- Wizigmann Voos S, Breier G, Risau W, Plate KH (1995) Up-regulation of vascular endothelial growth factor and its receptors in von Hippel-Lindau disease-associated and sporadic hemangioblastomas. *Cancer Res* 55:1358-1364
- Yamaguchi TP, Dumont DJ, Conlon RA, Breitman ML, Rossant J (1993) flk-1, an flt-related receptor tyrosine kinase is an early marker for endothelial cell precursors. *Development* 118:489-498
- Yang R, Thomas GR, Bunting S, Ko A, Ferrara N, Keyt B, Ross J, Jin H (1996) Effects of vascular endothelial growth factor on hemodynamics and cardiac performance. *J Cardiovasc Pharmacol* 27:838-844
- Yoshiji H, Gomez DE, Shibuya M, Thorgeirsson UP (1996) Expression of vascular endothelial growth factor, its receptor, and other angiogenic factors in human breast cancer. *Cancer Res* 56:2013-2016
- Yuan F, Chen Y, Dellian M, Safabakhsh N, Ferrara N, Jain RK (1996) Time-dependent vascular regression and permeability changes in established human tumor xenografts induced by an anti-vascular endothelial growth factor/vascular permeability factor antibody. *Proc Natl Acad Sci USA* 93:14765-14770

Promoting the formation of new collateral vessels in ischemic tissues using angiogenic growth factors (therapeutic angiogenesis) is an exciting frontier of cardiovascular medicine. Conversely, inhibition of the action of key regulators of angiogenesis, such as VEGF, constitutes a promising approach for the treatment of solid tumors and intraocular neovascular syndromes. These concepts are being tested now in clinical trials.

Clinical applications of angiogenic growth factors and their inhibitors

NAPOLIONE FERRARA¹ &
KARI ALITALO²

In embryos, blood vessels form through two distinct processes, vasculogenesis and angiogenesis. Vasculogenesis involves the de novo differentiation of endothelial cells from mesodermal precursors, whereas in angiogenesis new vessels are generated from pre-existing ones¹. Vasculogenesis takes place only during embryonic development and leads to the formation of a primary vascular plexus. Later these rather uniformly sized endothelial channels are remodeled into a mature system consisting of a tree-like hierarchy of large and small vessels. New capillaries then form through angiogenesis, either by sprouting or by splitting (intussusception) from their vessels of origin. In adults, angiogenesis is essential for the female reproductive cycle, and for repair, remodeling and regeneration of tissues, for example during wound healing². Neovascularization is also important in pathological processes such as tumor growth and metastasis².

The known endothelial cell specific growth factors and their receptors can be classified into vascular endothelial growth factor (VEGF) and angiopoietin (Ang) families³ (Fig. 1). Among the various angiogenic factors, VEGF is probably the most essential for the development and differentiation of the vascular system⁴. Loss of a single VEGF allele results in embryonic lethality^{5,6} (Fig. 2). Even selective inactivation of the heparin-binding isoforms of VEGF, leaving one functional isoform (VEGF₁₂₀), is insufficient for the proper development of the cardiovascular system and results in myocardial ischemia and perinatal or early postnatal lethality⁷. Also, other angiogenic factors, such as FGFs may work more indirectly, some of them through the VEGFs and their receptors⁸, so that a thorough knowledge of the signal transduction pathways of VEGFs and angiopoietins is essential for their use in therapeutic settings.

Therapeutic angiogenesis and inhibition of arterial restenosis

An exciting frontier of cardiovascular medicine is therapeutic angiogenesis. Promoting the formation of new collateral vessels on the ischemic myocardium, leg muscles and other tissues would have an important effect on the treatment of disorders for which pharmacological intervention has been ineffective in controlled trials and for which therapy is now limited to surgical revascularization or endovascular interventional therapy⁹.

Several angiogenic molecules have been tested in animal models, including bFGF, aFGF, FGF-5, VEGF isoforms, VEGF-C, HGF/SF and Ang-1/Ang-2. The factors tested most extensively are VEGF and bFGF. In some cases, the recombinant protein was tested. In others, gene transfer using naked DNA or adenoviral vectors was used. A single intra-arterial administration of 500–1000 µg of rhVEGF₁₆₅ augmented perfusion and development of collateral vessels in a rabbit model of hindlimb ischemia in which the femoral artery was surgically removed¹⁰. Similar results were obtained in the same model

with intramuscular or intra-arterial administration of aFGF, bFGF, HGF/SF and VEGF-C (refs. 11–14). VEGF administration after removal of the femoral artery

not only resulted in increased vascularization but also led to recovery of the normal endothelial reactivity to various mediators¹⁵. Arterial gene transfer with cDNA encoding VEGF isoforms also led to revascularization to an extent comparable to that achieved with the recombinant protein¹⁶. Moreover, administration of a VEGF₁₆₅ adenovirus vector shortly after common iliac artery ligation in the rat was capable of stimulating an angiogenic response that protects against subsequent occlusion of the femoral artery, indicating that gene transfer of VEGF might be useful in the prophylaxis of advancing arterial occlusive disease¹⁷. As little as 2 µg rhVEGF delivered over 4 weeks periaortally, distal to the occlusion, resulted in a significant increase in coronary blood flow and functional improvement in a pig model of chronic myocardial ischemia¹⁸. Very similar results were obtained using bFGF (ref. 19). Unexpectedly, even a single intracoronary administration of VEGF (or bFGF) was efficacious in this model to an extent comparable to that of 4-week infusion, despite the fact that only a small fraction of protein localizes to the ischemic area²⁰. Given such results, it is conceivable that young and otherwise healthy animals are very responsive to exogenous growth factors in the context of ischemia. At least some of this responsiveness may be due to the upregulation of VEGF receptors in the endothelia of ischemic tissues²¹. Adenovirus-mediated gene transfer of VEGF₁₂₁ (ref. 22) or FGF-5 (ref. 23) also resulted in collateral vessel growth and functional improvement in porcine models of myocardial ischemia.

These encouraging animal studies led to clinical trials using recombinant VEGF₁₆₅, aFGF, bFGF or gene therapy with plasmid or with adenoviral vectors. There is considerable debate whether gene therapy or administration of recombinant protein would be preferable. Delivery of angiogenic proteins by gene therapy might not only minimize their systemic side effects, such as hypotension (VEGF) or nephrotoxicity (bFGF), but also provide a slow release of the encoded factor for 1–2 weeks, leading to a more lasting angiogenic response. However, slow release of the recombinant protein, using microspheres or heparin-alginate formulations, might achieve the same results, without the potential risks associated with the use of viral vectors.

Arterial gene transfer of naked plasmid DNA encoding VEGF₁₆₅ in a patient with severe limb ischemia produced angiographic and histologic evidence of angiogenesis in the knee, mid-tibial and ankle levels 4 weeks after the transfer²⁴. In a subsequent study, the VEGF₁₆₅ plasmid cDNA was injected intramuscularly²⁵. Gene transfer was done in ten limbs of nine patients with nonhealing ischemic ulcers and/or rest pain due

to peripheral arterial disease. Improvement in the ankle-brachial index and distal flow in eight limbs were reported²⁵. Additional small trials by the same group have also shown that local injection of the VEGF₁₆₅ plasmid DNA resulted in clinical improvement in patients affected by myocardial ischemia²⁶ or Burger's disease (thromboangiitis obliterans)²⁷. However, none of these studies were placebo-controlled. Clinical trials using VEGF-C naked DNA or adenovirus mediated gene transfer of VEGF₁₂₁ in myocardial ischemia patients are now in phase I. Femoral angiograms from a patient with limb ischemia, before and 3 months after transfection of a VEGF₁₆₅ plasmid/liposome expression vector, show increased vascular density after the treatment (Fig. 3). However, the trial is ongoing and some caution should be used in interpreting such data, until more patients and the effect of placebo are more extensively evaluated.

Clinical trials using recombinant VEGF₁₆₅ and bFGF are also ongoing. In a phase I study in patients with coronary ischemia in which rhVEGF₁₆₅ was administered by intracoronary infusion, the molecule was safely tolerated at all doses tested²⁸. There was evidence of improvement in perfusion in seven of fifteen subjects and improved collateralization in five of seven who underwent follow-up coronary angiography. However, a subsequent placebo-controlled phase II study, in which rhVEGF was delivered as a single intracoronary infusion, followed by three intravenous infusions, has not demonstrated clinical benefit²⁹. The treatment was not better than placebo in treadmill time and pain relief, at least at 60 days²⁹. Brief exposures to rhVEGF₁₆₅, such as those achieved in this trial, may be insufficient to trigger and maintain a therapeutically meaningful angiogenic response, especially in the context of extensive atherosclerotic disease. Also, systemic administration of rhVEGF₁₆₅ or other factor may fail to generate an appropriate angiogenic concentration gradient from ischemic to non-ischemic areas, a requisite aspect of angiogenesis in a variety of physiological and pathological circumstances¹. Moreover, the placebo effect is probably greater than initially suspected, and even patients with very compromised myocardial function may show a substantial improvement with placebo. A phase II study with bFGF for coronary ischemia is now ongoing.

Local gene transfer into the vascular wall offers a promising alternative for the treatment of the complication of restenosis after percutaneous transcatheter angioplasty (PTCA) and coronary stenting. Restenosis occurs in many treated patients in 6 months, leading to obstruction in 20–35% of the patients³⁰. The pathogenesis of restenosis depends on endothelial damage, which also predisposes arteries to other pathological conditions, such as spasms or thrombosis. Prophylaxis of restenosis could therefore be based on strategies for endothelial protection or enhancement of endothelial repair and endothelial growth factors or vascular gene transfer could be used for this³¹. Re-endothelization in balloon-injured rat carotid artery was accelerated by a single dose of recombinant VEGF injected into the bloodstream or locally^{32,33}. Vessel status was also improved by injection of VEGF plasmid into adventitial surface of rabbit carotid arteries³⁴. Intravascular gene transfer in the arterial

wall was not very efficient³⁵, but secreted proteins such as VEGF could be used for therapeutic gene transfer trials using infusion-perfusion catheters³⁶ or histamine-induced increase of endothelial permeability³⁷. Because VEGF and VEGF-C share one receptor (VEGFR-2) but differ in the other receptor, VEGF-C and VEGF₁₆₅ might have overlapping but distinct effects in the vessel wall. However, VEGF-C gene transfer inhibits intimal thickening early, and the protective effect is at least equal to that seen with VEGF₁₆₅ gene transfer³⁸.

Therapeutic inhibition of vascular endothelial growth factor

Tumors

The growth of tumor xenografts in transparent chambers in mice is preceded by an increase in vascular density, indicating that the rapid growth of tumors depends on the development of a neovascular supply³⁹. In 1971, inhibition of angiogenesis was proposed as a valid strategy for the treatment of solid tumors and the search for the mediator(s) of tumor angiogenesis was begun⁴⁰.

Although inhibition of bFGF (ref. 41) or angiopoietin/Tie2 (refs. 42,43) may inhibit tumor growth, so far VEGF and its receptors constitute the most extensively investigated system in tumor angiogenesis and are now a main target of anti-cancer strategies. VEGF mRNA is substantially upregulated in most human tumors¹. Although tumor cells represent the main source of VEGF, tumor-associated stroma is also an important site of VEGF production⁴⁴. There is a correlation between VEGF expression and microvessel density in primary breast cancer sections⁴⁵. A similar correlation has been described in several other malignancies, including gastric carcinoma⁴⁶. Furthermore, there are increases in plasma levels of VEGF in tumor patients compared with tumor-free individuals, and high VEGF levels before chemotherapy are associated with a poor outcome⁴⁷.

Direct evidence for involvement of VEGF in tumorigenesis was first demonstrated using monoclonal antibodies against

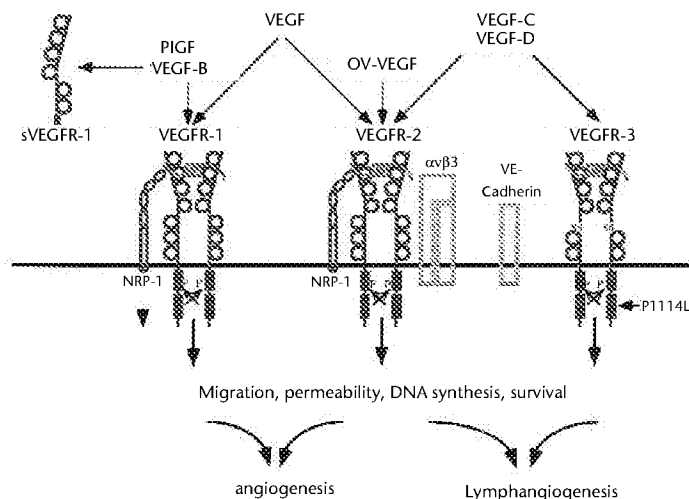


Fig. 1 VEGFs, their receptors and some of their endothelial effects in cells and tissues. Ligand binding induces receptor dimerization and subsequent auto/transphosphorylation, activates various signal transduction pathways and leads to differential cellular responses. sVEGFR-1, soluble VEGFR-1; HSPG, heparan sulphate proteoglycan; NP-1, neuropilin-1; α₃, integrin α₃, reported to make a molecular complex with activated VEGFR-2; ref. 95). VE-cadherin is also able to form a complex with VEGFR-2, a requirement for VEGF-dependent anti-apoptotic signals involving the PI3-kinase/Akt pathway⁹⁶. P1114L, point mutation of VEGFR-3 affecting patients in a family with lymphoedema⁹⁷.

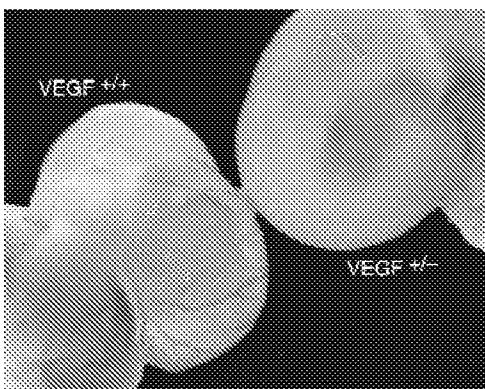


Fig. 2 Yolk sac of E10.5 VEGF^{+/+} and VEGF^{+/-} mouse embryos⁵. There is an apparent absence of vasculature in the yolk sac of the heterozygous, which die around E11. This is probably the only example among vertebrates of lethality after inactivation of a single allele of a gene that is not maternally imprinted.

VEGF in human xenografts in nude mice⁴⁸. These initial studies showed that several tumor cell lines can be substantially growth-inhibited by this treatment⁴⁸. These findings were extended to a broad variety of tumor cell lines, including carcinomas, sarcomas and gliomas⁴. Intravital videomicroscopy techniques have augmented our understanding of VEGF in tumorigenesis^{49,50}. Non-invasive imaging of the vasculature demonstrated a nearly complete suppression of tumor-associated angiogenesis in animals treated with monoclonal antibodies against VEGF compared with controls, providing a direct verification that inhibition of angiogenesis is the mechanism of tumor suppression after anti-VEGF treatment⁴⁹. Intravital microscopy techniques have also been used to investigate the effects of VEGF on the permeability and other properties of tumor vessels⁵⁰. Treatment with antibodies against VEGF resulted in time-dependent reductions in vascular permeability, in the diameter and tortuosity and eventually to a regression of tumor blood vessels; thus, VEGF is also an essential survival factor for tumor endothelial cells⁵⁰. Further evidence that VEGF action is required for tumor angiogenesis has been provided by the finding that retrovirus-mediated expression of a dominant negative VEGFR-2 mutant, which inhibits signal transduction through wild-type VEGFR-2 receptor, suppresses the growth of glioblastoma multiforme as well as other tumor cell lines *in vivo*⁵¹. Furthermore, high local expression of the soluble extracellular domain of VEGFR-1 or VEGFR-2, achieved by administration of the recombinant proteins, adenoviral-mediated gene transfer or by stable transfection of tumor cells, may significantly inhibit tumor growth, metastasis and mortality rate in nude mice^{52,53}.

Several strategies have been used to generate VEGF inhibitors suitable for clinical trials. One approach involves the 'humanization' of mouse monoclonal antibodies. A chief advantage of 'humanized' antibodies is a high degree of specificity, combined with a long half-life and little or no immunogenicity. A 'humanized' high-affinity monoclonal antibody against VEGF (rhuMab VEGF) with the same affinity and biological properties as the original murine antibody has been described⁵⁴. Toxicological studies in primates have shown that the effects of rhuMab VEGF are limited to inhibition of angiogenesis in the female reproductive tract and in the epiphyseal growth plate in

sexually immature animals that have not completed statural growth⁵⁵. rhuMab VEGF is now in phase II clinical trials for the treatment of non-small cell lung carcinoma and colorectal carcinoma in conjunction with standard chemotherapy and for breast and renal cell carcinoma as a single agent. In addition, small molecules that inhibit VEGFR-2 signal transduction are undergoing phase II clinical trials in cancer patients⁵⁶. Furthermore, monoclonal antibodies against VEGFR-2 are entering clinical trials.

Retinal ischemia and other conditions

Diabetes mellitus, occlusion of the central retinal vein or prematurity with subsequent exposure to oxygen can all be associated with intraocular neovascularization⁵⁷. A common denominator among these conditions is retinal ischemia⁵⁷. The new blood vessels may lead to vitreous hemorrhage, retinal detachment, neovascular glaucoma, and eventual blindness. Diabetic retinopathy is the leading cause of blindness in the working population. The hypothesis that ischemia-induced VEGF may be pathogenic in these conditions was initially tested by measuring VEGF levels in the eye fluids of patients. In a large series with 165 patients, a strong correlation was found between concentrations of VEGF in both aqueous and vitreous and active proliferative retinopathy associated with diabetes, occlusion of central retinal vein or prematurity⁵⁸. Direct evidence for the role of VEGF as a mediator of intraocular neovascularization has been generated in several animal models, including a primate model of iris neovascularization and a mouse model of retinopathy of prematurity. In the former, intraocular administration of monoclonal antibodies against VEGF substantially inhibits the neovascularization that follows the occlusion of central retinal veins⁵⁹. Likewise, soluble VEGFR-1 or VEGFR-2 extracellular domains fused to the immunoglobulin γ Fc domain suppress retinal angiogenesis in the mouse model⁶⁰. There is also evidence that growth hormone/insulin-like growth factor-1 is involved in ischemia-induced retinal neovascularization⁶¹.

Neovascularization is a principal cause of visual loss also in the wet form of age-related macular degeneration (AMD), the overall leading cause of blindness⁶². Several studies have documented the immunohistochemical localization of VEGF in surgically resected choroidal neovascular membranes from AMD patients⁶³. These findings suggest involvement of VEGF in the progression of AMD-related choroidal neovascularization. Anti-VEGF strategies for AMD are now being explored in clinical trials. One approach consists in the intravitreal administration of a recombinant humanized anti-VEGF Fab antibody fragment. Another strategy involves the injection of 2'-fluoropyrimidine RNA oligonucleotide ligands (aptamers)⁶⁴.

VEGF inhibition may also have therapeutic value for the treatment of ischemic-reperfusion related brain edema and injury. VEGF antagonism has shown beneficial effects in a mouse model of cortical ischemia⁶⁵; reducing acutely the volume of edematous tissue and resulting in a significant sparing of cortical tissue.

VEGF is important in angiogenesis in the female reproductive tract. VEGF inhibition results in suppression of corpus luteum angiogenesis in rodents⁶⁶ and primates⁵⁵. VEGF inhibitors might be used to treat conditions characterized by ovarian hyperplasia and hypervascularity, such as the polycystic ovary syndrome⁶⁶. VEGF-dependent angiogenesis may also be important pathogenically in endometriosis. Furthermore, VEGF is a

mediator of the ovarian growth and increased vascular permeability of ovarian hyperstimulation syndrome, a potentially fatal condition characterized by massive ovarian enlargement that may follow medical induction of ovulation with gonadotropins⁶⁷.

Perspectives

VEGF₁₆₅ binds to neuropilin-1, which functions as a ligand binding subunit of putative transmembrane receptors mediating specific signals for different semaphorins, the molecules mediating the collapse of axonal growth cones⁶⁸. Neuropilin is expressed in endothelial cells and enhances the mitogenic effects of VEGFR-2 upon VEGF₁₆₅ stimulation. Thus, there may be an as-yet ill-defined cross-regulation of cellular signals between these two families of factors. These findings lead to the intriguing conclusion that the processes of axon guidance and development of a network of capillary tubes share at least some common molecular mechanisms. In addition, the angiopoietin receptor/Tie and ephrin families of endothelial tyrosine kinases have important functions in the formation and maintenance of the vascular system⁶⁹⁻⁷¹. Endothelial cell-specific members of the TGF- β receptor and Notch families have also been described^{72,73}. Given this complexity of vascular endothelial signaling, therapies using VEGF alone or any other single angiogenic factor may produce incompletely functioning or unstable endothelial channels with defective arteriovenous and pericellular differentiation, characteristic of many tumors⁷⁴. Combinations of growth factors may be preferable in future therapies directed to neovascularization of tissues, with an adequate investment of the formed vessels with periendothelial matrix and pericyte/smooth muscle cells. In fact, a more heterogeneous set of genes coordinating angiogenic functions may be provided by active ongoing research of hypoxia-regulated gene expression in mammalian cells⁷⁵. Also, some virus-encoded proteins, such as the VEGFR-2 activating HIV Tat protein⁷⁶, Kaposi sarcoma herpesvirus-associated G-protein-coupled receptor⁷⁷ or Orf virus encoded VEGF-E⁷⁸⁻⁸⁰ may offer new insights into the mechanism of regulation of angiogenesis.

Although recent research has focused on the combination of VEGF and Ang-1 as being especially promising, it is not known now which growth factor combinations will prove to be the most effective therapeutically. VEGF and bFGF have a very synergistic effect in the induction of angiogenesis, both *in vitro* and *in vivo*⁴. The interaction between VEGF and HGF/SF is also being actively investigated. Although transgenic expression of Ang-1 in the skin epidermis under the keratin (K)14 promoter has been associated with neovascularization⁸¹, other studies, using defined amounts of the recombinant protein in a model of adult neovascularization, have failed to demonstrate strong angiogenic responses to Ang-1, unless it is used in combination with VEGF (refs. 71,82). This discrepancy may be explained by the fact that the expression of the K14 promoter is initiated already at midgestation, and thus the results may reflect persistence of the fetal neovascularization. It is possible, however, that Ang-1 may provide a co-factor for combination therapies. A further unresolved issue is the correct dosage of growth factor(s). This seems particularly important for a molecule like VEGF, which has several isoforms and such a tight dose-response effect that a 50% reduction in expression results in lethality during embryonic life^{5,6}. Conversely, continuous local overexpression of VEGF may result in a hemangioma-like vasculature and thus can be deleterious⁸³.

Also, it is unknown whether an angiogenic treatment may be sufficient to induce functional blood vessels for prolonged periods or will need to be re-administered periodically in order to maintain such vessels.

A K14-driven VEGF-C transgene induced lymphangiogenesis but no angiogenesis in mouse skin⁸⁴, and recombinant VEGF-C also stimulated lymphatic vessel hyperplasia in mature chick chorioallantoic membrane⁸⁵. Thus, besides angiogenesis, it may also become possible to direct therapeutic lymphangiogenesis in patients, such as after evacuation of axillary lymph nodes in breast carcinoma surgery.

Despite the potential redundancy of tumor angiogenesis factors, inhibition of VEGF alone seems sufficient to achieve considerable tumor growth suppression in a wide variety of models. However, it remains to be established whether tumors are able to activate, after prolonged therapy, alternative angiogenic pathways that might confer resistance to the treatment. These issues should be addressed in the current clinical trials with various VEGF inhibitors. A challenge now in anti-VEGF (and anti-angiogenic) therapy is devising appropriate and reliable markers to monitor tumor progression. There is considerable debate whether blood vessel count in biopsy specimens^{45,46} may provide a reliable indicator of response to the treatment. There are also efforts to identify surrogate endpoints, applying non-invasive approaches, such as magnetic resonance imaging⁸⁶.

VEGF is not only a mitogen but also a potential survival factor for endothelial cells⁴. Such a 'maintenance' function seems to be developmentally regulated, as it is very dependent on the age of the animal⁸⁷. VEGF inactivation during early postnatal life, achieved by *Cre-loxP*-mediated inducible gene targeting of by administration of a soluble VEGFR-1 chimeric protein, results in regression of the vasculature, kidney failure and lethality⁸⁷. However, in adult animals a similar treatment has no effects on the existing vasculature. Therefore, a process of maturation occurs in endothelial cells such that VEGF eventually is not essential for survival. This switch seems to take place in the mouse around the fourth postnatal week. Absence of pericyte

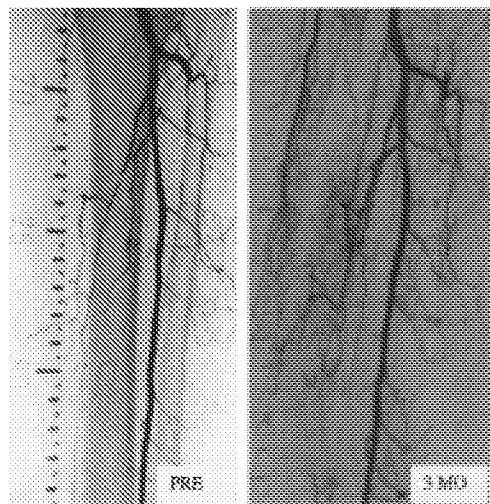


Fig. 3 Angiography of the lower extremity of a patient with limb ischemia before (PRE) and 3 months after (3 MO) the transfection of a VEGF₁₆₅ plasmid/liposome expression vector, showing strongly increased vascular density after the treatment. Courtesy H. Manninen, P. Matsi, K. Mäkinen, M. Hilpeläinen, M. Laitinen, E. Alhava and S. Ylä-Herttua, A. I. Virtanen Institute and Kuopio University Hospital (Kuopio, Finland).

coverage in immature vessels may be a factor determining their dependence on VEGF (ref. 88). However, other evidence suggests that the molecular/intracellular nature of this switch may be more complex and mostly still to be determined⁸⁷. In juvenile animals, VEGF is essential for endochondral bone formation and longitudinal growth^{89,55}. In the fully developed animal, VEGF may be required mainly for active angiogenic processes such as corpus luteum development or wound healing. Nevertheless, VEGF may be important for endothelial homeostasis in the adult in certain circumstances; for example, during disease states. Indeed, prolonged VEGF inhibition failed to induce glomerular damage in normal primates⁵⁵ or rodents^{87,90}, despite the strong constitutive expression of the VEGF mRNA in podocytes and other cell types in the adult kidney⁴. However, administration of VEGF inhibitors to rats with mesangioproliferative nephritis results in impaired glomerular endothelial regeneration and increased endothelial cell death⁹⁰.

Some CD34⁺ hematopoietic progenitor cells mobilized by GM-CSF from human peripheral blood, bone marrow, fetal liver or umbilical cord blood were shown to express VEGFR-2 on their surface⁹¹, and VEGFR-2 is expressed on human hematopoietic stem cells⁹². Endothelial progenitor cells expand and differentiate into endothelial cells after addition of bFGF and VEGF to the cultures, and they can thus be considered to provide endothelial progenitor cells⁹¹⁻⁹³. The endothelial progenitor cells from bone marrow may be mobilized using the stromal-derived factor 1 chemokine, the GM-CSF cytokine or tissue hypoxia⁹⁴. As these cells may be capable of participating in active angiogenesis after entry into the circulatory system⁹⁴, they provide an interesting possibility for the delivery of cellular or gene therapy to sites of neovascularization.

Finally, the first placebo-controlled clinical study with rhVEGF may have brought a more realistic assessment of the potential of therapeutic angiogenesis and raised a number of questions. For example, how can one explain the discrepancy between the considerable efficacy observed even with very small amounts of growth factors in animal models of coronary or limb ischemia and the rather disappointing clinical results? An essential difference may lie in the fact that young and otherwise healthy animals are able to mount an effective endogenous angiogenic response that can be maximized by an additional stimulus provided by a recombinant protein or gene therapy. In contrast, patients with extensive atherosclerotic disease may have poor responses. It is possible, however, that a more persistent exposure to an individual growth factor or to a combination of growth factors may be effective. Clinical trials now ongoing should answer at least some of these questions over the next 2-3 years.

1. Risau, W. Mechanisms of angiogenesis. *Nature* **386**, 671-674 (1997).
2. Folkman, J. Angiogenesis in cancer, vascular, rheumatoid and other diseases. *Nature Med.* **1**, 27-31 (1995).
3. Korpelainen, E.I. & Alitalo, K. Signaling angiogenesis and lymphangiogenesis. *Curr. Opin. Cell Biol.* **10**, 159-164 (1998).
4. Ferrara, N. & Davis-Smyth, T. The biology of vascular endothelial growth factor. *Endocr. Rev.* **18**, 4-25 (1997).
5. Ferrara, N. *et al.* Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature* **380**, 439-442 (1996).
6. Carmeliet, P. *et al.* Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. *Nature* **380**, 435-439 (1996).
7. Carmeliet, P. *et al.* Impaired myocardial angiogenesis and ischemic cardiomyopathy in mice lacking the vascular endothelial growth factor isoforms VEGF164 and VEGF188. *Nature Med.* **5**, 495-502 (1999).
8. Seghezzi, G. *et al.* Fibroblast growth factor-2 (FGF-2) induces vascular endothelial growth factor (VEGF) expression in the endothelial cells of forming capillaries: an autocrine mechanism contributing to angiogenesis. *J. Cell Biol.* **171**, 1659-1673 (1998).

9. Rivard, A. & Isner, J.M. Angiogenesis and vasculogenesis in treatment of cardiovascular disease. *Mol. Med.* **4**, 429-440 (1998).
10. Takeshita, S. *et al.* Therapeutic angiogenesis. A single intra-arterial bolus of vascular endothelial growth factor augments revascularization in a rabbit ischemic hind limb model. *J. Clin. Invest.* **93**, 662-670 (1994).
11. Pu, L.Q. *et al.* Enhanced revascularization of the ischemic limb by angiogenic therapy. *Circulation* **88**, 208-215 (1993).
12. Asahara, T. *et al.* Synergistic effect of vascular endothelial growth factor and basic fibroblast growth factor on angiogenesis *in vivo*. *Circulation* **92**, I1365-I1371 (1995).
13. Van Belle, E. *et al.* Potentiated angiogenic effect of scatter factor/hepatocyte growth factor via induction of vascular endothelial growth factor: the case for paracrine amplification of angiogenesis. *Circulation* **97**, 381-390 (1998).
14. Witzensbichler, B. *et al.* Vascular endothelial growth factor-C (VEGF-C/VEGF-2) promotes angiogenesis in the setting of tissue ischemia. *Am. J. Pathol.* **153**, 381-394 (1998).
15. Bauters, C. *et al.* Physiological assessment of augmented vascularity induced by VEGF in ischemic rabbit hindlimb. *Am. J. Physiol.* **267**, H1263-H1271 (1994).
16. Takeshita, S. *et al.* Gene transfer of naked DNA encoding for three isoforms of vascular endothelial growth factor stimulates collateral development *in vivo*. *Lab. Invest.* **75**, 487-501 (1996).
17. Mack, C.A. *et al.* Salvage angiogenesis induced by adenovirus-mediated gene transfer of vascular endothelial growth factor protects against ischemic vascular occlusion. *J. Vasc. Surg.* **27**, 699-709 (1998).
18. Pearlman, J.D. *et al.* Magnetic resonance mapping demonstrates benefits of VEGF-induced myocardial angiogenesis. *Nature Med.* **1**, 1085-1089 (1995).
19. Harada, K. *et al.* Basic fibroblast growth factor improves myocardial function in chronically ischemic porcine hearts. *J. Clin. Invest.* **94**, 623-630 (1994).
20. Lopez, J.J. *et al.* VEGF administration in chronic myocardial ischemia in pigs. *Cardiovasc. Res.* **40**, 272-281 (1998).
21. Li, J. *et al.* VEGF, flk-1, and flt-1 expression in a rat myocardial infarction model of angiogenesis. *Am. J. Physiol.* **270**, H1803-H1811 (1996).
22. Mack, C.A. *et al.* Biologic bypass with the use of adenovirus-mediated gene transfer of the complementary deoxyribonucleic acid for vascular endothelial growth factor 121 improves myocardial perfusion and function in the ischemic porcine heart. *J. Thor. Cardiovasc. Surg.* **115**, 168-176 (1998).
23. Giordano, F. *et al.* Intracoronary gene transfer of fibroblast growth factor-5 increases blood flow and contractile function in an ischemic region of the heart. *Nat. Med.* **2**, 534-539 (1996).
24. Isner, J.M. *et al.* Clinical evidence of angiogenesis after arterial gene transfer of phVEGF165 in patient with ischaemic limb. *Lancet* **348**, 370-374 (1996).
25. Baumgartner, I. *et al.* Constitutive expression of phVEGF165 after intramuscular gene transfer promotes collateral vessel development in patients with critical limb ischemia. *Circulation* **97**, 1114-1123 (1998).
26. Losordo, D.W. *et al.* Gene therapy for myocardial angiogenesis: initial clinical results with direct myocardial injection of phVEGF165 as sole therapy for myocardial ischemia. *Circulation* **98**, 2800-2804 (1998).
27. Isner, J.M. *et al.* Treatment of thromboangiitis obliterans (Buerger's disease) by intramuscular gene transfer of vascular endothelial growth factor: preliminary clinical results. *J. Vasc. Surg.* **28**, 964-975 (1998).
28. Henry, T.D. *et al.* Results of intracoronary recombinant human vascular endothelial growth factor (rhVEGF) administration trial. *J. Am. Coll. Cardiol.* **31**, 65A (810-811) (1998).
29. Henry, T.D. *et al.* Double blind, placebo controlled, trial of recombinant human vascular endothelial growth factor: the VIVA trial. *J. Am. Coll. Cardiol.* **33**, 384A 874 (1999).
30. Bittl, J.A. Advances in coronary angioplasty. *N. Engl. J. Med.* **335**, 1290-1302 (1996).
31. Yla-Herttuala, S. Vascular gene transfer. *Curr. Opin. Lipidol.* **8**, 72-76 (1997).
32. Asahara, T. *et al.* Local delivery of vascular endothelial growth factor accelerates re-endothelialization and attenuates intimal hyperplasia in balloon-injured rat carotid artery. *Circulation* **91**, 2793-2801 (1995).
33. Burke, P.A., Lehmann-Bruinsma, K. & Powell, J.S. Vascular endothelial growth factor causes endothelial proliferation after vascular injury. *Biochem. Biophys. Res. Comm.* **207**, 348-354 (1995).
34. Laitinen, M. *et al.* VEGF gene transfer reduces intimal thickening via increased production of nitric oxide in carotid arteries. *Hum. Gene Ther.* **8**, 1737-1744 (1997).
35. Laitinen, M. *et al.* Adenovirus-mediated gene transfer to lower limb artery of patients with chronic critical leg ischemia. *Hum. Gene Ther.* **9**, 1481-1486 (1998).
36. Camenzind, E. *et al.* Intracoronary heparin delivery in humans. Acute feasibility and long-term results. *Circulation* **92**, 2463-2472 (1995).
37. Greelish, J.P. *et al.* Stable restoration of the sarcoglycan complex in dystrophic muscle perfused with histamine and a recombinant adeno-associated viral vector. *Nature Med.* **5**, 439-443 (1999).
38. Hiltunen, M.O. *et al.* Intravascular Adenovirus-mediated VEGF-C gene transfer inhibits neointima formation in valvulotomy-denuded rabbit aorta. *Circulation* (in press).
39. Algire, G.H. & Chalkley, H.W. Vascular reactions of normal and malignant tissues *in vivo*. I. Vascular reactions of mice to wounds and to normal and neoplastic transplants. *J. Natl. Cancer Inst.* **6**, 73-85 (1945).
40. Folkman, J. Tumor angiogenesis: therapeutic implications. *N. Engl. J. Med.* **285**, 1182-1186 (1971).
41. Enzoli, B. *et al.* Block of AIDS-Kaposi's sarcoma (KS) cell growth, angiogenesis, and lesion formation in nude mice by antisense oligonucleotide targeting basic

- fibroblast growth factor. A novel strategy for the therapy of KS. *J. Clin. Invest.* **94**, 1736–1746 (1994).
42. Lin, P. *et al.* Inhibition of tumor angiogenesis using a soluble receptor establishes a role for Tie2 in pathologic vascular growth. *J. Clin. Invest.* **100**, 2072–2078 (1997).
 43. Lin, P. *et al.* Antiangiogenic gene therapy targeting the endothelium-specific receptor tyrosine kinase Tie2. *Proc. Natl. Acad. Sci. USA* **95**, 8829–8834 (1998).
 44. Fukumura, D. *et al.* Tumor induction of VEGF promoter activity in stromal cells. *Cell* **94**, 715–725 (1998).
 45. Gasparini, G. *et al.* Prognostic significance of vascular endothelial growth factor protein in node-negative breast carcinoma. *J. Natl. Cancer Inst.* **89**, 139–147 (1997).
 46. Maeda, K. *et al.* Prognostic value of vascular endothelial growth factor expression in gastric carcinoma. *Cancer* **77**, 853–863 (1996).
 47. Salven, P., Ruotsalainen, T., Mattson, K. & Joensuu, H. High pre-treatment serum level of vascular endothelial growth factor (VEGF) is associated with poor outcome in small-cell lung cancer. *Int. J. Cancer* **79**, 144–146 (1998).
 48. Kim, K.J. *et al.* Inhibition of vascular endothelial growth factor induced angiogenesis suppresses tumour growth in vivo. *Nature* **362**, 841–844 (1993).
 49. Borgström, P., Hillan, K.J., Sriramarao, P. & Ferrara, N. Complete inhibition of angiogenesis and growth of microtumors by anti-vascular endothelial growth factor neutralizing antibody: novel concepts of angiostatic therapy from intravitreal videomicroscopy. *Cancer Res.* **56**, 4032–4039 (1996).
 50. Yuan, F. *et al.* Time-dependent vascular regression and permeability changes in established human tumor xenografts induced by an anti-vascular endothelial growth factor/vascular permeability factor antibody. *Proc. Natl. Acad. Sci. USA* **93**, 14765–14770 (1996).
 51. Millauer, B. *et al.* Dominant-negative inhibition of Flk-1 suppresses the growth of many tumor types *in vivo*. *Cancer Res.* **56**, 1615–1620 (1996).
 52. Kong, H.L. *et al.* Regional suppression of tumor growth by *in vivo* transfer of a cDNA encoding a secreted form of the extracellular domain of the flt-1 vascular endothelial growth factor receptor. *Hum. Gene Ther.* **9**, 823–833 (1998).
 53. Goldman, C.K. *et al.* Paracrine expression of a native soluble vascular endothelial growth factor receptor inhibits tumor growth, metastasis, and mortality rate. *Proc. Natl. Acad. Sci. USA* **95**, 8795–8800 (1998).
 54. Presta, L.G. *et al.* Humanization of an anti-vascular endothelial growth factor monoclonal antibody for the therapy of solid tumors and other disorders. *Cancer Res.* **57**, 4593–4599 (1997).
 55. Ryan, A.M. *et al.* Preclinical safety evaluation of rhuMABVEGF, an antiangiogenic humanized monoclonal antibody. *Toxicol Pathol.* **27**, 78–86, 1999.
 56. Strawn, L.M. *et al.* Flk-1 as a target for tumor growth inhibition. *Cancer Res.* **56**, 3540–3545 (1996).
 57. Patz, A. Studies on retinal neovascularization. *Invest. Ophthalmol. Vis. Sci.* **19**, 1133–1138 (1980).
 58. Aiello, L.P. *et al.* Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. *N. Engl. J. Med.* **331**, 1480–1487 (1994).
 59. Adamis, A.P. *et al.* Inhibition of vascular endothelial growth factor prevents retinal ischemia-associated iris neovascularization in a nonhuman primate. *Arch. Ophthalmol.* **114**, 66–71 (1996).
 60. Aiello, L.P. *et al.* Suppression of retinal neovascularization *in vivo* by inhibition of vascular endothelial growth factor (VEGF) using soluble VEGF-receptor chimeric proteins. *Proc. Natl. Acad. Sci. USA* **92**, 10457–10461 (1995).
 61. Smith, L.E. *et al.* Essential role of growth hormone in ischemia-induced retinal neovascularization. *Science* **276**, 5319–5321 (1997).
 62. Garner, A. in *Pathobiology of Ocular Diseases* 2nd edn. (eds. Garner, A. & Klintworth, G.K.) 1625–1710 (Marcel Dekker, New York, 1994).
 63. Lopez, P.F., Sippy, B.D., Lambert, H.M., Thach, A.B. & Hinton, D.R. Transdifferentiated retinal pigment epithelial cells are immunoreactive for vascular endothelial growth factor in surgically excised age-related macular degeneration-related choroidal neovascular membranes. *Invest. Ophthalmol. Vis. Sci.* **37**, 855–868 (1996).
 64. Ruckman, J. *et al.* 2'-Fluoropyrimidine RNA-based aptamers to the 165-amino acid form of vascular endothelial growth factor (VEGF165). Inhibition of receptor binding and VEGF-induced vascular permeability through interactions requiring the exon 7-encoded domain. *J. Biol. Chem.* **273**, 20556–20567 (1998).
 65. van Bruggen, N. *et al.* VEGF antagonism reduces cerebral edema formation and tissue damage following ischemic-reperfusion injury in the mouse brain. *J. Clin. Inv.* (in the press).
 66. Ferrara, N. *et al.* Vascular endothelial growth factor is essential for corpus luteum angiogenesis. *Nature Med.* **4**, 336–340 (1998).
 67. McClure, N. *et al.* Vascular endothelial growth factor as a capillary permeability agent in ovarian hyperstimulation syndrome. *Lancet.* **344**, 235–269, 1994.
 68. Soker, S., Takashima, S., Miao, H.Q., Neufeld, G. & Klagsbrun, M. Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor. *Cell* **92**, 735–745 (1998).
 69. Adams, R.H. *et al.* Roles of ephrinB ligands and Eph receptors in cardiovascular development: demarcation of arterial/venous domains, vascular morphogenesis, and sprouting angiogenesis. *Genes Dev.* **13**, 295–306 (1999).
 70. Wang, H.U., Chen, Z.-F. & Anderson, D.J. Molecular distinction and angiogenic interaction between embryonic arteries and veins revealed by ephrin-B2 and its receptor Eph-B4. *Cell* **93**, 741–753 (1998).
 71. Shyu, K.-G., Manor, O., Magner, M., Yancopoulos, G.D. & Isner, J.M. Direct intramuscular injection of plasmid DNA encoding angiopoietin-1 but not angiopoietin-2 augments revascularization in the rabbit ischemic hindlimb. *Circulation* **98**, 2081–2087 (1998).
 72. Roelen, B.A. & van Rooijen, M.A. Mummery CL. Expression of ALK-1, a type 1 serine/threonine kinase receptor, coincides with sites of vasculogenesis and angiogenesis in early mouse development. *Dev. Dyn.* **209**, 418–430 (1997).
 73. Uyttendaele, H. *et al.* Notch4/int-3, a mammary proto-oncogene, is an endothelial cell-specific mammalian Notch gene. *Development* **122**, 2251–2259 (1996).
 74. Jain, R.K. Delivery of molecular and cellular medicine to solid tumors. *J. Control. Release* **53**, 49–67 (1998).
 75. Carmeliet, P. *et al.* Role of HIF-1alpha in hypoxia-mediated apoptosis, cell proliferation and tumour angiogenesis. *Nature* **394**, 485–490 (1998).
 76. Albini, A. *et al.* The angiogenesis induced by HIV-1 Tat protein is mediated by the Flk-1/KDR receptor on vascular endothelial cells. *Nat. Med.* **2**, 1371–1375 (1996).
 77. Bais, C. *et al.* G-protein-coupled receptor of Kaposi's sarcoma-associated herpesvirus is a viral oncogene and angiogenesis activator. *Nature* **391**, 86–89, 1998.
 78. Ogawa, S. *et al.* A novel type of vascular endothelial growth factor: VEGF-E (NZ-7 VEGF) preferentially utilizes KDR/Flk-1 receptor and carries a potent mitotic activity without heparin-binding domain. *J. Biol. Chem.* **273**, 31273–31282 (1998).
 79. Meyer, M. *et al.* A novel vascular endothelial growth factor encoded by Orf virus, VEGF-E, mediates angiogenesis via signaling through VEGFR-2 (KDR) but not VEGFR-1 (Flt-1) receptor tyrosine kinases. *EMBO J.* **18**, 363–374 (1999).
 80. Wise, L.M. *et al.* Vascular endothelial growth factor (VEGF)-like protein from orf virus NZ2 binds to VEGFR2 and Neuropilin-1. *Proc. Natl. Acad. Sci.* **96**, 3071–3076 (1999).
 81. Suri, C. *et al.* Increased vascularization in mice overexpressing angiopoietin-1. *Science* **282**, 468–471 (1998).
 82. Asahara, T. *et al.* Tie2 receptor ligands, angiopoietin-1 and angiopoietin-2, modulate VEGF-induced postnatal neovascularization. *Circ. Res.* **83**, 233–240 (1998).
 83. Springer, M.L., Chen, A.S., Kraft, P.E., Bednarski, M. & Blau, H.M. VEGF gene delivery to muscle: potential role for vasculogenesis in adults. *Mol. Cell* **2**, 549–558 (1998).
 84. Jeltsch, M. *et al.* Hyperplasia of lymphatic vessels in VEGF-C transgenic mice. *Science* **276**, 1423–1425 (1997).
 85. Oh, S.J. *et al.* VEGF and VEGF-C: specific induction of angiogenesis and lymphangiogenesis in the differentiated avian chorioallantoic membrane. *Dev. Biol.* **188**, 96–109 (1997).
 86. Pham, C.D. *et al.* Magnetic resonance imaging detects suppression of tumor vascular permeability after administration of antibody to vascular endothelial growth factor. *Cancer Invest.* **16**, 225–230 (1998).
 87. Gerber, H.P. *et al.* VEGF is required for growth and survival in neonatal mice. *Development* **126**, 1149–1159 (1999).
 88. Benjamin, L., Hemo, I. & Keshet, E. A plasticity window for blood vessel remodeling is defined by pericyte coverage of the preformed endothelial network and is regulated by PDGF-B and VEGF. *Development* **125**, 1591–1598 (1998).
 89. Gerber, H. P. *et al.* VEGF couples hypertrophic cartilage remodeling, ossification and angiogenesis during endochondral bone formation. *Nat. Med.*, **5**, 623–628 (1999).
 90. Ostendorf, T. *et al.* VEGF₁₆₅ mediates glomerular endothelial repair. *J. Clin. Invest.* **104**, 913–923 (1999).
 91. Asahara, T. *et al.* Isolation of putative progenitor endothelial cells for angiogenesis. *Science* **275**, 964–967 (1997).
 92. Shi, Q. *et al.* Evidence for circulating bone marrow-derived endothelial cells. *Blood* **92**, 362–367 (1998).
 93. Ziegler, B.L. *et al.* KDR receptor: a key marker defining hematopoietic stem cells. *Science* **285**, 1553–1558 (1999).
 94. Takahashi, T. *et al.* Ischemia- and cytokine-induced mobilization of bone marrow-derived endothelial progenitor cells for neovascularization. *Nature Med.* **5**, 434–438 (1999).
 95. Soldi, R. *et al.* Role of $\alpha_v\beta_3$ in the activation of vascular endothelial growth factor receptor-2. *EMBO J.* **18**, 882–892 (1999).
 96. Carmeliet, P. *et al.* Targeted deficiency or cytosolic truncation of the VE-cadherin gene in mice impairs VEGF-mediated endothelial survival and angiogenesis. *Cell* **98**, 147–157 (1999).
 97. Ferrell, R.E. *et al.* Hereditary lymphedema: evidence for linkage and genetic heterogeneity. *Hum. Mol. Genet.* **7**, 2073–2078 (1998).

¹Department of Molecular Oncology

Genentech

DNA Way

South San Francisco, California 94080, USA

²Molecular/Cancer Biology Laboratory

Haartman Institute University of Helsinki

PI 21 (Haartmaninkatu 3)

00014 Helsinki, Finland

Correspondence should be addressed to N.F.; email: nf@gene.com, or K.A.; email: Kari.Aitalo@Helsinki.FI

Amelioration of Long-Term Renal Changes in Obese Type 2 Diabetic Mice by a Neutralizing Vascular Endothelial Growth Factor Antibody

Allan Flyvbjerg,¹ Frederik Dagnæs-Hansen,² An S. De Vriese,³ Bieke F. Schrijvers,^{1,3} Ronald G. Tilton,⁴ and Ruth Rasch⁵

Diabetic nephropathy in type 2 diabetic patients is a frequent complication associated with increased morbidity and mortality. Various growth factors and cytokines have been implicated in the pathogenesis of diabetic kidney disease, including vascular endothelial growth factor (VEGF). To explore a role for VEGF in renal changes in type 2 diabetes, we examined the renal effects of a neutralizing murine VEGF antibody in the diabetic *db/db* mouse, a model of obese type 2 diabetes. One group of *db/db* mice was treated for 2 months with a VEGF antibody, while another *db/db* group was treated for the same period with an isotype-matched irrelevant IgG. A third group consisting of nondiabetic *db/+* mice was treated with the same isotype-matched IgG for 2 months. Placebo-treated *db/db* mice showed a pronounced increase in kidney weight, glomerular volume, basement membrane thickness (BMT), total mesangial volume, urinary albumin excretion (UAE), and creatinine clearance (CrCl) when compared with nondiabetic controls. In VEGF antibody-treated *db/db* mice, increases in kidney weight, glomerular volume, BMT, and UAE were attenuated, whereas the increase in CrCl was abolished. VEGF antibody administration tended to reduce expansion in total mesangial volume. These effects in diabetic animals were seen without impact on body weight, blood glucose, insulin levels, or food consumption. In conclusion, chronic inhibition of VEGF in *db/db* mice ameliorates the diabetic renal changes seen in type 2 diabetes. *Diabetes* 51:3090–3094, 2002

The incidence of type 2 diabetes is increasing worldwide. The development of diabetic nephropathy is seen in 30–40% of type 2 diabetic individuals, with an associated increased morbidity and mortality. Accordingly, diabetic nephropathy is the most common cause of end-stage renal failure in the Western world. Mechanisms underlying the development of diabetic kidney disease in type 2 diabetes are complex. Among the many potential pathogenic mechanisms responsible for the development of diabetic kidney disease, growth factors have been suggested to be important players. Accordingly, growth hormone/IGFs and transforming growth factor (TGF)- β have been shown to have measurable effects on the development of diabetic kidney changes in animal models of type 1 diabetes (1). Recently, the vascular endothelial growth factor (VEGF) system has been proposed to play a role in the development of diabetic renal changes in animal models of type 1 diabetes (1–4); the potential role of the VEGF system in renal complications of type 2 diabetes remains unknown.

The aim of the present study was to explore the role of VEGF in the development of renal changes in type 2 diabetes. Accordingly, a specific neutralizing murine VEGF antibody was administered for 2 months in *db/db* mice, a genetic model of type 2 diabetes characterized by obesity, sustained hyperglycemia, hyperinsulinemia, lack of ketonuria, and progressive renal kidney disease (5–8).

RESEARCH DESIGN AND METHODS

Animals. Adult female *db/db* mice (C57BLKS/J-*lepr^{db}/lepr^{db}*) and their age-matched nondiabetic *db/+* littermates (C57BLKS/J-*lepr^{db/+}*) (M&B, Ry, Denmark) were used. Nondiabetic *db/+* mice had a body weight of 19–20 g, and the *db/db* mice had an initial weight of 30–41 g. Intervention with VEGF antibody administration was initiated at 8 weeks of age because 100% of the *db/db* mice become frankly hyperglycemic from week 7–8 (8). The *db/db* mice were included in the study 1–2 weeks after development of diabetes, at the age of 8 weeks. The mice were housed six to eight per cage in a room with a 12:12 h artificial light cycle (7:00 A.M. to 7:00 P.M.), a temperature of 21 \pm 1°C, and a humidity of 55 \pm 5%. The animals had free access to standard chow (Altromin no. 1324; Altromin, Lage, Germany) and tap water throughout the experiment. The study complied with Danish regulations for care and use of laboratory animals.

Study design. The *db/db* mice were randomized into two groups of 12 per group. One group of *db/db* mice was treated with intraperitoneal injections of a neutralizing VEGF antibody, and the other group was treated with an isotype-matched irrelevant IgG, as were the nondiabetic *db/+* mice ($n = 6$). The VEGF antibody and irrelevant IgG were administered intraperitoneally in an initial bolus dose of 300 μ g, followed by doses of 100 μ g three times weekly. The VEGF antibody and irrelevant IgG were dissolved in 0.154 mol/l

From the ¹Medical Department M and Medical Research Laboratories, Institute of Experimental Clinical Research, Aarhus University Hospital, Aarhus, Denmark; the ²Department of Medical Microbiology and Immunology, University of Aarhus, Aarhus, Denmark; the ³Renal Unit, Department of Internal Medicine, Ghent University Hospital, Ghent, Belgium; the ⁴Department of Pharmacology, Texas Biotechnology Corporation, Houston, Texas; and the ⁵Department of Cell Biology, Institute of Anatomy, Aarhus University, Aarhus, Denmark.

Address correspondence and reprint requests to Dr. Allan Flyvbjerg, MD, DMSc, Medical Department M and Medical Research Laboratories, Institute of Experimental Clinical Research, Aarhus University Hospital, Aarhus Kommunehospital, DK-8000 Aarhus C, Denmark. E-mail: allan.flyvbjerg@dadlnet.dk.

Received for publication 4 April 2002 and accepted in revised form 12 July 2002.

BMT, basement membrane thickness; CrCl, creatinine clearance; LM, light microscopy; STZ, streptozotocin; TGF, transforming growth factor; UAE, urinary albumin excretion; VEGF, vascular endothelial growth factor; VEGFR, VEGF receptor.

TABLE 1

Mean body weight, blood glucose, and food consumption at day 0 and 60 in placebo-treated controls, placebo-treated diabetic *db/db* mice, and VEGF antibody-treated diabetic *db/db* mice

	Day 0			Day 60		
	Body weight (g)	Blood glucose (mmol/l)	Food consumption (g/24 h)	Body weight (g)	Blood glucose (mmol/l)	Food consumption (g/24 h)
Control, placebo	19.6 ± 0.4	5.4 ± 0.3	5.5 ± 0.6	21.3 ± 0.2	5.5 ± 0.3	5.9 ± 0.8
Diabetic, placebo	41.4 ± 0.7*	18.6 ± 1.0*	8.5 ± 0.7*	47.3 ± 1.0*	19.8 ± 1.4*	9.1 ± 1.1*
Diabetic, VEGF antibody	40.6 ± 0.8*	18.4 ± 1.0*	8.8 ± 0.8*	46.2 ± 0.9*	18.8 ± 2.0*	8.9 ± 0.9*

Data are means ± SE ($n = 6-12$ in each group). * $P < 0.01$ vs. nondiabetic controls.

NaCl and injected in a volume of 0.5 ml. A full characterization of the VEGF antibody used has been described elsewhere (2,4). Briefly, 8-week-old female Balb/C mice were immunized by repeated intraperitoneal and subcutaneous injections of 50 µg rhVEGF₁₆₅, which was emulsified with complete Freund's adjuvant for the primary immunization and incomplete Freund's adjuvant for the subsequent immunizations. Mice with the highest serum titer to VEGF₁₆₅ received an additional injection of 30 µg VEGF₁₆₅ in PBS, and 3 days later, spleen cells were harvested for production of hybridomas to rhVEGF₁₆₅. Two hybridoma cell lines with the highest antibody titer and neutralizing activity were cloned three to four times in microplates and injected intraperitoneally (10⁷ cells). Ascites fluid was collected, and purified IgG was prepared by protein A chromatography, with a further characterization of the neutralizing activity as described previously (2).

Body weight, food consumption, and blood glucose were determined at initiation of the experiment and every 2 weeks. Blood glucose was measured in tail-vein blood as described below. After 8 weeks, mice were placed in metabolic cages to collect 24-h urine samples for urinary albumin excretion (UAE) and urinary creatinine determinations. At sacrifice, mice were anesthetized with pentobarbital (50 mg/kg i.p.) and nonfasting blood samples were drawn from the retro-orbital venous plexus using heparinized capillary tubes. Serum samples were stored at -80°C until analysis was performed. In all animals, the right and left kidneys were removed and weighed. The middle piece of the right kidney (including the papilla) was fixed in 4% paraformaldehyde for determination of glomerular volume by light microscopy (LM) (see below). The middle piece of the left kidney (including the papilla) was fixed in 0.1 mol/l cacodylate buffer with 1% glutaraldehyde and 2% paraformaldehyde for later determination of basement membrane thickness (BMT) and mesangial fraction by electron microscopy (see below). In addition, liver and heart were removed, weighed, and snap frozen in liquid nitrogen.

Determination of blood glucose and serum insulin. Blood glucose was measured at day 0 and every 2 weeks in tail-vein blood by Precision Xtra Plus (Abbott Laboratories, MediSense Products, Bedford, MA), and urine was tested for glucose and ketone bodies by Combur⁵ Test D (Roche Diagnostics, Mannheim, Germany). Serum insulin was measured by an ultrasensitive rat insulin enzyme-linked immunosorbent assay (DRG Diagnostics, Marburg, Germany). Semilog linearity of mouse serum and rat insulin was found at multiple dilutions, indicating antigen similarity between mouse and rat insulin. The intra- and interassay coefficients of variation were <5% and <10% for the insulin assays.

Determination of UAE and creatinine clearance. The urinary albumin concentration was determined by radioimmunoassay as previously described (9) using rat albumin antibody and rat albumin standard. Semilog linearity of mouse urine and rat albumin (in the standard) was found at multiple dilutions, indicating antigen similarity between mouse and rat albumin. Urine samples were stored at -20°C until assay was performed. Serum and urinary creatinine concentrations were measured by an automated technique adapted from the method of Jaffé and corrected for the prevailing glucose content due to interference in the Jaffé reaction. The creatinine clearance (CrCl) was expressed in milliliters per hour. The intra- and interassay coefficients of variation were <5% and <10% for both assays.

Estimation of glomerular volume. The middle part of the right kidney (containing the papilla) was embedded in paraffin for LM examination. Two micron-thick sections were cut on a rotation microtome and stained with *p*-aminosalicylic acid and hematoxylin. The mean glomerular tuft volume (V_G) was determined from the mean glomerular cross-sectional area (A_G) at a magnification of 400×, as previously described (10-12). The areas were determined with a two-dimensional version of the nucleator (CAST; Olympus, Copenhagen, Denmark) (12) by LM as the average area of a total of 40-50 glomerular profiles (tuft omitting the proximal tubular tissue within the Bowman capsule). V_G was calculated as $V_G = \beta/k \times (A_G)^{3/2}$, where $\beta = 1.38$,

which is the shape coefficient for spheres (the idealized shape of glomeruli), and $k = 1.1$, which is a size distribution coefficient (10-12).

Estimation of mesangial fraction, total mesangial volume, and BMT.

The middle part of the left kidney (containing the papilla) was embedded in Epon 825 for electron microscopy examination. Thin sections were cut on a Reichert Ultracut (Leica, Vienna, Austria) and stained with uranyl acetate and lead citrate. From an electron microscope (Tecnai 12; Phillips, Eindhoven, Holland), images covering the whole glomerular profile were recorded with a MegaView video camera (Soft Imaging System, Münster, Germany) onto a monitor. Measurements of mesangial regions were performed at a final magnification of 3,200×. Four to six glomeruli were measured from two blocks. Mesangial fractions were determined by point counting of mesangial regions as fraction of the tuft. The total mesangial volume was calculated by multiplying the mesangial fraction by the total glomerular volume. For measurements of BMT, randomized fields were recorded at a magnification of 30,000× from the same sections described above. BMT was measured, applying the orthogonal intercept method as previously described (13). About 60 measurements were performed per glomerulus, and BMT is given as a harmonic mean.

Statistical analysis. For repeated measurements, ANCOVA was used to evaluate differences with Student's *t* test for unpaired comparisons. A *P* value <0.05 was considered statistically significant. For data not following a normal distribution, the Mann-Whitney rank-sum test was used. All data are expressed as means ± SE, with *n* indicating the number of mice studied. Statistical analysis was performed using SPSS for Windows.

RESULTS

Body weight, blood glucose, food consumption, and serum insulin. The *db/db* mice had a greater body weight than the nondiabetic *db/+* mice, as was also the case for food consumption (Table 1). Mean blood glucose levels were 18-19 mmol/l in *db/db* mice throughout the study, and 5-6 mmol/l in *db/+* animals (Table 1). The *db/db* mice had severe hyperinsulinemia (Table 2). VEGF antibody administration did not affect any of the above parameters in *db/db* mice throughout the study duration (Tables 1 and 2).

Kidney weight, glomerular volume, BMT, and mesangial volume. Placebo-treated *db/db* mice showed an in-

TABLE 2

Mean serum insulin, liver weight, and heart weight at day 60 in placebo-treated controls, placebo-treated diabetic *db/db* mice, and VEGF antibody-treated diabetic *db/db* mice

	Day 60		
	Serum insulin (µg/l)	Liver weight (mg)	Heart weight (mg)
Control, placebo	2.86 ± 0.29	1,136 ± 36	103 ± 4
Diabetic, placebo	18.82 ± 1.60*	2,272 ± 88*	109 ± 3
Diabetic, VEGF antibody	16.85 ± 1.67*	1,891 ± 65†	103 ± 3

Data are means ± SE ($n = 6-12$ in each group). * $P < 0.01$ vs. nondiabetic controls; † $P < 0.05$ vs. the two other groups.

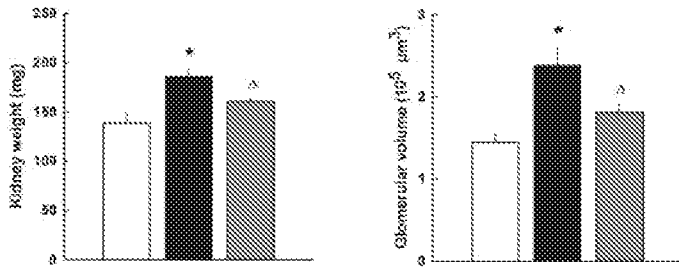


FIG. 1. Mean right kidney weight and glomerular volume on day 60 in nondiabetic controls (□), placebo-treated diabetic *db/db* mice (■), and VEGF antibody-treated diabetic *db/db* mice (▨). Values are means + SE ($n = 6-12$ in each group). * $P < 0.01$ vs. nondiabetic controls; $\Delta P < 0.05$ vs. nondiabetic controls and placebo-treated *db/db* mice.

crease in kidney weight of 34% at day 60 (187 ± 7 vs. 139 ± 11 mg, $P < 0.01$) when compared with nondiabetic *db/+* controls (Fig. 1). In VEGF antibody-treated *db/db* mice, a significantly smaller increase in kidney weight was observed versus placebo-treated *db/db* mice (161 ± 4 , $P < 0.05$), although the kidney weight was higher than that seen in nondiabetic controls ($P < 0.01$). The same pattern of changes was seen in glomerular volume (Fig. 1). Total glomerular volume increased by 65% in placebo-treated *db/db* mice compared with nondiabetic controls (2.38 ± 0.22 vs. $1.44 \pm 0.11 \times 10^5 \mu\text{m}^3$, $P < 0.01$). VEGF antibody treatment in *db/db* mice partially prevented the increase in glomerular volume versus placebo-treated *db/db* mice ($1.81 \pm 0.11 \times 10^5 \mu\text{m}^3$, $P < 0.01$). The glomerular volume was, however, still elevated above that of nondiabetic controls ($P < 0.05$). BMT increased by 18% in placebo-treated *db/db* mice when compared with nondiabetic controls (176 ± 6 vs. 149 ± 4 nm, $P < 0.05$), while an insignificant increase was seen in VEGF antibody-treated *db/db* mice (160 ± 6 nm, NS), with a value significantly lower than that of placebo-treated *db/db* mice ($P < 0.05$) (Fig. 2). Both diabetic groups had a significant increase in mesangial fraction ($P < 0.05$, data not shown), and total glomerular mesangial volume tended ($0.05 < P < 0.10$) to be lower in the VEGF antibody-treated *db/db* group (Fig. 2).

UAE and CrCl. A pronounced increase in UAE was observed in placebo-treated *db/db* mice at day 60 versus nondiabetic *db/+* controls (4.57 ± 0.75 vs. 1.15 ± 0.16 $\mu\text{g}/24$ h, $P < 0.01$), with a considerably lower level in *db/db* mice treated with the VEGF antibody (1.85 ± 0.34 $\mu\text{g}/24$ h, $P < 0.01$ vs. placebo-treated *db/db* mice) (Fig. 3). Placebo-treated *db/db* mice showed a pronounced increase in CrCl

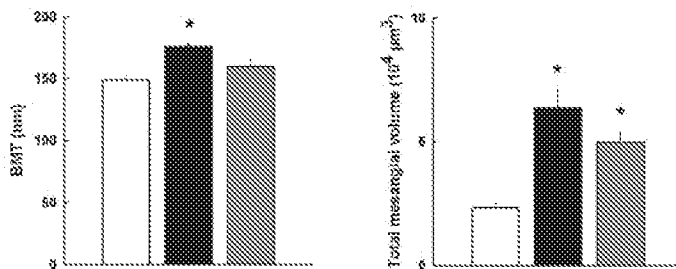


FIG. 2. BMT and total mesangial volume on day 60 in nondiabetic controls (□), placebo-treated diabetic *db/db* mice (■), and VEGF antibody-treated diabetic *db/db* mice (▨). Values are means + SE ($n = 6-12$ in each group). * $P < 0.05$ vs. nondiabetic controls and VEGF antibody-treated diabetic *db/db* mice.

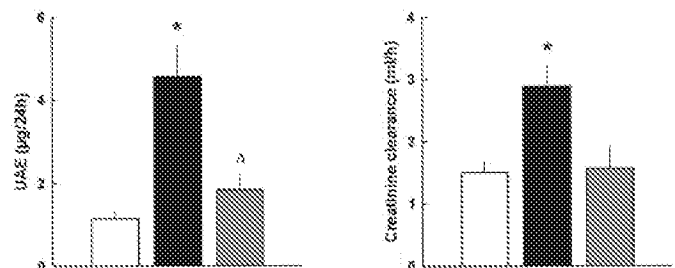


FIG. 3. Mean 24-h UAE and CrCl on day 60 in nondiabetic controls (□), placebo-treated diabetic *db/db* mice (■), and VEGF antibody-treated diabetic *db/db* mice (▨). Values are means + SE ($n = 6-12$ in each group). * $P < 0.01$ vs. nondiabetic controls; $\Delta P < 0.05$ vs. nondiabetic controls and placebo-treated *db/db* mice.

when compared with nondiabetic controls (1.51 ± 0.18 vs. 2.89 ± 0.35 ml/h, $P < 0.05$), with normalization in the VEGF antibody-treated *db/db* mice (1.57 ± 0.36 ml/h) (Fig. 3).

Liver and heart weight. The placebo-treated *db/db* mice had greater liver weights than the *db/+* mice, whereas VEGF antibody-treated *db/db* mice had less liver weight gain (Table 2). There were no significant differences in heart weight among the three groups.

DISCUSSION

The *db/db* mouse, which expresses a leptin receptor defect in the hypothalamus, is a genetic model of type 2 diabetes characterized by obesity, sustained hyperglycemia, hyperinsulinemia, and lack of ketonuria. Previously, this model has been shown to present with robust diabetic renal changes characterized by increased renal/glomerular volume, BMT, UAE, and mesangial volume within 2 months of diabetes (5-8).

The major new finding of the present study is an amelioration of diabetes-induced renal changes in *db/db* mice by VEGF antibody administration. Accordingly, antibody administration attenuated the increase in renal/glomerular volume, BMT, and UAE and abolished the increase in CrCl. These effects were seen without affecting metabolic control, insulin levels, body weight, or food consumption, indicating that VEGF plays a causal role in the development of late renal changes in a model of type 2 diabetes.

The VEGF system consists of different isoforms of homodimeric glycoproteins (14-21). Furthermore, at least two high-affinity VEGF receptors (VEGFR-1 and -2) have been described (17). VEGF has pronounced angiogenic actions (15,18-20) and causes vasodilation and increased vascular permeability (14,18). The expression of VEGF was initially described to be markedly increased in highly vascularized rapidly growing tumors (22), and VEGF has been shown to be a potent mitogenic factor for endothelial cells (20,21). The two VEGFRs (VEGFR-1 and -2), also known as the *fms*-like tyrosine kinase and fetal liver kinase 1, are high-affinity transmembrane tyrosine kinase receptors (17). Both VEGF and the two VEGFRs are expressed in the kidney (3,23-27). VEGF expression and specific VEGF binding have been described in rat (23) and human kidney (24-26). VEGF has been localized to epithelial glomerular cells (i.e., podocytes) (3,26,27), distal tubules, and renal collecting ducts (3,25). Furthermore, VEGFR-2 has been localized mainly to glomerular endo-

thelial cells and cortical interstitial fibroblasts (3). Mesangial cells, glomerular endothelial cells, vascular smooth muscle cells, and proximal and distal tubular cells are capable of producing VEGF *in vitro* (27–29). High glucose has been shown to stimulate VEGF expression in vascular smooth muscle cells (30). Also, in a recent study in OLETF rats (an experimental rat model of type 2 diabetes), renal VEGF mRNA and glomerular VEGF immunoreactivity were reported to be elevated over a diabetes duration of 9–68 weeks (31). In another study, changes in renal VEGF levels were described in streptozotocin (STZ)-induced diabetic rats (a rat model of type 1 diabetes) with a diabetes duration of 3 and 32 weeks (3). VEGF mRNA and protein were mainly localized to the glomerular epithelial cells and VEGFR-2 mRNA mainly to glomerular endothelial cells (3). VEGF mRNA and peptide were increased in diabetic animals at both time points examined, whereas the expression of VEGFR-2 and VEGFR binding were increased only at 3 weeks (3).

Although the area of identifying and developing specific antagonists of a pathophysiologically enhanced VEGF system in oncology and different eye diseases has attracted increasing interest (32), no studies have appeared on the effect of VEGF antagonists in diabetic kidney disease of type 2 diabetes. Direct evidence for a role of VEGF in the early renal changes observed in a model of type 1 diabetes (i.e., STZ-induced diabetic rats) has been published using the same VEGF antibody (4). Six weeks treatment with the VEGF antibody abolished the diabetes-associated hyperfiltration and partially blocked the increase in UAE (4). VEGF antibody administration in nondiabetic control rats had no impact on any renal parameters, indicating a diabetes-specific effect of VEGF antibody administration in diabetes (4). In the present study, using a mouse model of type 2 diabetes, administration of the VEGF antibody was shown to ameliorate both the classical early features of diabetic kidney disease, i.e., renal/glomerular hypertrophy and hyperfiltration (measured as CrCl), and more importantly, late renal changes (i.e., BMT), with a tendency to reduce total mesangial volume. The *db/db* mouse has previously been reported to develop decreased CrCl within 2 months after the onset of diabetes, suggesting a progressive diabetic kidney disease with loss of kidney function (8). In the present study, however, several lines of evidence indicated that placebo-treated *db/db* mice presented with renal hyperfunction, which was partially or fully normalized by VEGF antibody treatment, i.e., partial effect on kidney weight, glomerular volume, UAE, and normalization of elevated CrCl. The reason for this discrepancy is unknown, but may be explained by a variable susceptibility to diabetes in subbreedings of the *db/db* mouse strain.

The observation that VEGF antibody treatment abolished the increase in BMT and renal hyperfiltration and partially blocked the increase in UAE is interesting in view of the well-known actions of VEGF on vascular permeability (14,18) and the anatomical localization of the VEGF system in the glomerulus (i.e., podocytes and glomerular endothelial cells) (3,25–27). These results indicate that administration of a specific, neutralizing VEGF antibody in *db/db* mice fully or partly restores the abnormally increased albumin permeability in the diabetic kidney,

which is believed to be caused by abnormalities in the filtration barrier due to increased membrane pore size and reduced anion charge. Although VEGF expression has been described in glomerular epithelial cells (3,26), VEGF antibody administration only tended to reduce total mesangial volume in the present study. These results suggest that the primary role of VEGF in the diabetic renal changes in type 2 diabetes is linked to the diabetes-associated permeability changes, while the role of VEGF in mesangial expansion, if any, seems to be secondary. In this context, it is interesting that administration of a neutralizing TGF- β antibody in *db/db* mice has been shown to ameliorate diabetes-associated glomerular matrix expansion without affecting either elevated UAE or renal VEGF expression (33).

Although currently unproven, several potential pathways involved in diabetes-induced vascular changes (1) may involve VEGF as a downstream cytokine. *In vitro*, VEGF has been shown to be stimulated by IGF-I (34), and furthermore, IGF-I receptor blockade in an ischemia-induced retinopathy model has been shown to reduce the intracellular VEGF-mediated mitogen-activated protein kinases along with ameliorating retinal neovascularization (35). Blockade of protein kinase C β activity with a specific inhibitor (LY333531) suppresses the VEGF-induced alterations in retinal leakage, retinal blood flow, and ischemia-induced retinal neovascularization (36). In addition, ACE inhibition in diabetic rats has been shown to reduce diabetes-associated retinal changes in VEGF expression and vascular permeability (37). Also, in the study described above in a rat model of type 2 diabetes (31), it was shown that long-term administration of an advanced glycation end product inhibitor (OPB-9195) abolished the enhanced renal VEGF mRNA and glomerular VEGF immunoreactivity along with renoprotection, in terms of normalization of diabetes-induced renal collagen IV accumulation and a reduction of the rise in UAE (31).

In conclusion, the present data strongly support the hypothesis that VEGF is an important pathogenetic factor in the development of long-term renal changes in type 2 diabetes. Further studies are warranted to fully elucidate the role of VEGF as a downstream mediator for some of the well-known pathways leading to diabetic renal damage.

ACKNOWLEDGMENTS

This study was supported by the Danish Medical Research Council (Grant 9700592), the Eva and Henry Fränkels Memorial Foundation, the Danish Kidney Foundation, the Ruth König Petersen Foundation, the Danish Diabetes Association, the Novo Foundation, the Nordic Insulin Foundation, the Johanne and Aage Louis Petersen Memorial Foundation, the Institute of Experimental Clinical Research, the University of Aarhus, and the Aarhus University-Novo Nordisk Center for Research in Growth and Regeneration (Danish Medical Research Council Grant 9600822). B.F.S. is supported by a grant from the Institute for the Promotion of Innovation by Science and Technology in Flanders. A.S.D. is supported by a grant from the Fund for Scientific Research Flanders (N20/0).

The excellent technical assistance by Karen Mathiassen, Birgitte Gran, and Kirsten Nyborg is highly appreciated.

REFERENCES

1. Flyvbjerg A: Putative pathophysiological role of growth factors and cytokines in experimental diabetic kidney disease. *Diabetologia* 43:1205–1223, 2000
2. Tilton RG, Kawamura T, Chang KC, Ido Y, Bjerrcke RJ, Stephan CC, Brock TA, Williamson JR: Vascular dysfunction induced by elevated glucose levels in rats is mediated by vascular endothelial growth factor. *J Clin Invest* 99:2192–2202, 1997
3. Cooper ME, Vranes D, Youssef S, Stacker SA, Cox AJ, Rizkalla B, Casley DJ, Bach LA, Kelly DJ, Gilbert RE: Increased renal expression of vascular endothelial growth factor (VEGF) and its receptor VEGFR-2 in experimental diabetes. *Diabetes* 48:2229–2239, 1999
4. De Vriese AS, Tilton RG, Elger M, Stephan CC, Kriz W, Lameire NH: Antibodies against vascular endothelial growth factor improve early renal dysfunction in experimental diabetes. *J Am Soc Nephrol* 12:993–1000, 2001
5. Koenig RJ, Cerami A: Synthesis of hemoglobin A1c in normal and diabetic mice: potential model of basement membrane thickening. *Proc Natl Acad Sci U S A* 72:3687–3691, 1975
6. Gartner K: Glomerular hyperfiltration during onset of diabetes mellitus in two strains of diabetic mice (C57b1/6J *db/db* and C57b1/ksj *db/db*). *Diabetologia* 15:59–63, 1978
7. Bower G, Brown DM, Steffes MW, Vernier RL, Mauer SM: Studies of the glomerular mesangium and the juxtaglomerular apparatus in the genetically diabetic mouse. *Lab Invest* 43:333–341, 1980
8. Cohen MP, Clements RS, Cohen JA, Shearman CW: Prevention of decline in renal function in the diabetic *db/db* mouse. *Diabetologia* 39:270–274, 1996
9. Flyvbjerg A, Bennett WF, Rasch R, Kopchick JJ, Scarlett JA: Inhibitory effect of a growth hormone receptor antagonist (G120K-PEG) on renal enlargement, glomerular hypertrophy, and urinary albumin excretion in experimental diabetes in mice. *Diabetes* 48:377–382, 1999
10. Gundersen HJ, Bagger P, Bendtsen TF, Evans SM, Korbo L, Marcussen N, Møller A, Nielsen K, Nyengaard JR, Pakkenberg B: The new stereological tools: disector, fractionator, nucleator and point sampled intercepts and their use in pathological research and diagnosis. *APMIS* 96:857–881, 1988
11. Pagtakman ME, Kasch R, Remke HG, Meyer TW: Morphometric analysis of effects of angiotensin II on glomerular structure in rats. *Am J Physiol* 268:F82–F88, 1995
12. Weibel ER: *Stereologic Methods: Practical Methods for Biological Morphometry*. London, Academic Publishers, 1979, p. 51–57
13. Jensen EB, Gundersen HJ, Østerby R: Determination of membrane thickness distribution from orthogonal intercepts. *J Microsc* 115:19–33, 1979
14. Senger DR, Galli SJ, Dvorak AM, Perruzzi CA, Harvey VS, Dvorak HF: Tumor cells secrete a vascular permeability factor that promotes accumulation of ascites fluid. *Science* 219:983–985, 1983
15. Keck PJ, Hauser SD, Krivi G, Sanzo K, Warren T, Feder J, Connolly DT: Vascular permeability factor, an endothelial cell mitogen related to PDGF. *Science* 246:1309–1312, 1989
16. Tisher E, Mitchell R, Hartman T, Silva M, Gospodarowicz D, Fiddes JC, Abraham JA: The human gene for vascular endothelial growth factor: multiple protein forms are encoded through alternative exon splicing. *J Biol Chem* 266:11947–11954, 1991
17. Neufeld G, Cohen T, Gengrinovitch S, Poltorak Z: Vascular endothelial growth factor (VEGF) and its receptors. *FASEB J* 13:9–22, 1999
18. Senger DR, Connolly DT, van de Water L, Feder J, Dvorak HF: Purification and NH₂-terminal amino acid sequence of guinea pig tumor-secreted vascular permeability factor. *Cancer Res* 50:1774–1778, 1990
19. Leung DW, Cachianes G, Kuang WJ, Goeddel DV, Ferrara N: Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science* 246:1306–1309, 1989
20. Plate KH, Breier G, Weich HA, Risau W: Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo. *Nature* 359:845–848, 1992
21. Ferrara N, Houck KA, Jakeman LB, Winer J, Leung DW: The vascular endothelial growth factor family of polypeptides. *J Cell Biochem* 47:211–218, 1991
22. Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N: Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumour growth in vivo. *Nature* 362:841–844, 1993
23. Jakeman LB, Winer J, Bennett GL, Altar CA, Ferrara N: Binding sites for vascular endothelial growth factor are localized on endothelial cells in adult rat tissues. *J Clin Invest* 89:244–253, 1992
24. Brown LF, Berse B, Tognazzi K, Manseau EJ, van de Water L, Senger DR, Dvorak HF, Rosen S: Vascular permeability factor mRNA and protein expression in human kidney. *Kidney Int* 42:1457–1461, 1992
25. Simon M, Grone HJ, Jöhren O, Küllmer J, Plate KH, Risau W, Fuchs E: Expression of vascular endothelial growth factor and its receptors in human renal ontogenesis and in adult kidney. *Am J Physiol* 268:F240–F250, 1995
26. Simon M, Rockl W, Hornig C, Grone EF, Theis H, Weich HA, Fuchs E, Yayon A, Grone HJ: Receptors of vascular endothelial growth factor/vascular permeability factor (VEGF/VPF) in fetal and adult human kidney: localization and [¹²⁵I]VEGF binding sites. *J Am Soc Nephrol* 9:1044–1044, 1998
27. Williams B: A potential role for angiotensin II-induced vascular endothelial growth factor expression in the pathogenesis of diabetic nephropathy. *Miner Electrolyte Metab* 24:400–405, 1998
28. Pupilli C, Lasagni L, Romagnani P, Bellini F, Mammelli M, Misciglia N, Mavilia C, Vellei U, Villari D, Serio M: Angiotensin II stimulates the synthesis and secretion of vascular permeability factor/vascular endothelial growth factor in human mesangial cells. *J Am Soc Nephrol* 10:245–255, 1999
29. Gruden G, Thomas S, Burt D, Zhou W, Chusney G, Gnudi L, Viberti G: Interaction of angiotensin II and mechanical stretch on vascular endothelial growth factor production by mesangial cells. *J Am Soc Nephrol* 10:730–737, 1999
30. Natarajan R, Bai W, Lanting L, Gonzales N, Nadler J: Effects of high glucose on vascular endothelial growth factor expression in vascular smooth muscle cells. *Am J Physiol* 42:H2224–H2231, 1997
31. Tsuchida K, Makita Z, Yamagishi S, Atsumi T, Miyoshi H, Obara S, Ishida M, Ishikawa S, Yasumura K, Koike T: Suppression of transforming growth factor and vascular endothelial growth factor in diabetic nephropathy in rats by a novel advanced glycation end product inhibitor, OPB-9195. *Diabetologia* 42:579–588, 1999
32. Duh E, Aiello LP: Vascular endothelial growth factor and diabetes: the agonist versus antagonist paradox. *Diabetes* 48:1899–1906, 1999
33. Ziyadeh FN, Hoffman BB, Han DC, Iglesias-de la Cruz MC, Hong SW, Isono M, Chen S, McGowan TA, Kumar S: Long-term prevention of renal insufficiency, excess matrix gene expression, and glomerular mesangial matrix expansion by treatment with monoclonal antitransforming growth factor- β antibody in *db/db* mice. *Proc Natl Acad Sci U S A* 97:8015–8020, 2000
34. Rubin J, Wang H, Tashjian AH Jr, Patterson C: Enhanced expression of vascular endothelial growth factor in human SaOS-2 osteoblast-like cells and murine osteoblasts induced by insulin-like growth factor I. *Endocrinology* 137:2262–2268, 1996
35. Smith LE, Shen W, Perruzzi C, Soker S, Kinose F, Xu X, Robinson G, Driver S, Bischoff J, Zhang B, Schaeffer JM, Senger DR: Regulation of vascular endothelial growth factor-dependent retinal neovascularization by insulin-like growth factor I receptor. *Nat Med* 5:1390–1395, 1999
36. Aiello LP, Bursell SE, Clermont A, Duh E, Ishii H, Takagi C, Mori F, Ciulla TA, Wasy K, Jirousek M, Smith LE, King GL: Vascular endothelial growth factor-induced retinal permeability is mediated by protein kinase C in vivo and suppressed by an orally effective β -isoform-selective inhibitor. *Diabetes* 46:1473–1480, 1997
37. Gilbert RE, Kelly DJ, Cox AJ, Wilkinson-Berka JL, Rumble JR, Osicka T, Panagiotopoulos S, Lee V, Hendrich EC, Jerums G, Cooper ME: Angiotensin converting enzyme inhibition reduces retinal overexpression of vascular endothelial growth factor and hyperpermeability in experimental diabetes. *Diabetologia* 43:1360–1367, 2000

12. A. Nicholson, S. Frieda, A. Pearce, R. Silverstein, *Arterioscler. Thromb. Vasc. Biol.* **15**, 269 (1995).

13. G. Endemann *et al.*, *J. Biol. Chem.* **268**, 11811 (1993).

14. S. Nozaki *et al.*, *J. Clin. Invest.* **96**, 1859 (1995).

15. J. Savill, I. Dransfield, N. Hogg, C. Haslett, *Nature* **343**, 170 (1990).

16. J. Savill, N. Hogg, Y. Ren, C. Haslett, *J. Clin. Invest.* **90**, 1513 (1992).

17. Y. Ren, R. Silverstein, J. Allen, J. Savill, *J. Exp. Med.* **181**, 1857 (1995).

18. S. W. Ryeom, J. R. Sparrow, R. L. Silverstein, *J. Cell Sci.* **109**, 387 (1996).

19. K. Schneitz, P. Spielmann, M. Nöll, *Genes Dev.* **7**, 114 (1993).

20. P. Heitzler, unpublished observations.

21. M. Boedigheimer and A. Laughon, *Development* **118**, 1291 (1993).

22. L. Frank and C. Rushlow, *ibid.* **122**, 1343 (1996).

23. Berkeley Drosophila Genome Project, unpublished data.

24. Single embryos were squished in 10 μ l of 10 mM tris-HCl (pH8.0), 1 mM EDTA, and 25 mM NaCl containing Proteinase K (200 μ g/ml; Boehringer Mannheim) and incubated at 37°C for 30 min, followed by 2 min at 95°C. The PCR was performed with 2.5 U of Taq polymerase and 100 ng of each primer. Two pairs of primers were designed to amplify 634 base pairs (bp) of the *crq* genomic sequence and 200 bp of the genomic region of the *doom* gene as an internal control [A. J. Harvey, A. P. Bidwadi, L. K. Miller, *Mol. Cell. Biol.* **17**, 2835 (1997)]. *Crq*-specific primers were 5'-TGCCACCGATGCTGCAGAT-3' and 5'-AGCCGAATATGAT TCCGTAAGT-3'. *Doom*-specific primers were 5'-AGGGTAAACGCCACAGAATGT-3' and 5'-GATATCGTGTAGTGGCCCG-3'. The PCR cycles were 94°C for 1 min, 65°C for 1 min, and 72°C for 1 min for 30 cycles. In embryos from the W88 stock, 20 of 79 were missing the *crq*-specific band.

25. CRQ immunostaining was used to genotype each embryo. Peroxidase immunostaining detected all hemocytes [R. E. Nelson *et al.*, *EMBO J.* **13**, 3438 (1994)]. The nuclear dye 7-AAD labeled all DNA and allowed for the identification of apoptotic corpses. Unless otherwise specified, stage 11 to 16 embryos were fixed with standard procedures (44). Fixed devitelinized embryos were incubated in phosphate-buffered saline (PBS), 0.0125% saponin, 1% bovine serum albumin, and 4% normal goat serum (PSN) for 1 hour at room temperature and then incubated with the primary antibodies at a 1:1000 dilution in PSN overnight at 4°C. After several washes in PBS, the embryos were incubated for 1 hour at room temperature with the following secondary antibodies: fluorescein isothiocyanate-conjugated goat antibody to mouse and Cy5-conjugated goat antibody to rabbit (Jackson ImmunoResearch) used at a 1:1000 dilution in PSN. Finally, embryos were washed three times in PBS for 20 min and subsequently incubated with 7-AAD (5 μ g/ml) in PBS for 30 min. Embryos were quickly washed twice in PBS, mounted in Vectashield (Vector), and viewed by confocal microscopy (Leica TCS NT 4D).

26. The efficiency of engulfment was quantified by counting the number of engulfed corpses per macrophage in at least five fields of four embryos of each genotype. A P.I., that is, the mean number of engulfed corpses per macrophage, was calculated for each embryo, and the mean P.I. was derived for each genotype.

27. C. Phelps and A. Brand, *Methods* **14**, 367 (1998).

28. N. Franc and K. White, data not shown.

29. Stage 12 to 14 w. *UAS-crq;hs-Gal4/+* and control w; *hs-Gal4* embryos were heat-shocked for 1 hour at 39°C, aged for 2 hours at 25°C, and fixed and embedded in Spurr's resin (34). Serial 1- μ m sections of two embryos of each genotype were stained with a solution of methylene, toluidine blue, and borax (44) and viewed by standard microscopy.

30. J. Pugin *et al.*, *Immunity* **1**, 509 (1994).

31. A. Devitt *et al.*, *Nature* **392**, 505 (1998).

32. Stage 11 embryos were microinjected with a solution of PBS and 2 mM Na₂S₂O₈ containing about 6 \times 10⁹ tetramethyl rhodamine isothiocyanate (TRITC)-labeled fluorescent *E. coli* (K-12 strain) or *S. aureus* (Wood strain) bioparticles (heat-killed bacteria; Molecular Probes) with standard microinjection proce-

dures (44). After injection, embryos were kept in the dark at 18°C for 14 to 16 hours, incubated for 1 hour at 4°C, mounted, and viewed under Nomarski and fluorescence with a confocal microscope.

33. J. Abrams, A. Lux, H. Steller, M. Krieger, *Proc. Natl. Acad. Sci. U.S.A.* **89**, 10375 (1992).

34. K. White *et al.*, *Science* **264**, 677 (1994).

35. M. E. Grether, J. M. Abrams, J. Agapite, K. White, H. Steller, *Genes Dev.* **9**, 1694 (1995).

36. P. Chen, W. Nordstrom, B. Gish, J. M. Abrams, *ibid.* **10**, 1773 (1996).

37. After CRQ immunostaining, the amount of fluorescence seen in five isolated macrophages of each genotype was quantified with a confocal microscope. For each macrophage, the fluorescence of serial sections of 0.5 μ m was quantified from top to bottom of the cell. After subtracting the background fluorescence, the total amount of fluorescence within each macrophage was calculated.

38. M. Hortsch *et al.*, *Int. J. Dev. Biol.* **42**, 33 (1998).

39. J.-L. Dimarcq *et al.*, *Insect Biochem. Mol. Biol.* **27**, 877 (1997).

40. E. Gateff *et al.*, in *Invertebrate Systems In Vitro*, E. Kurstak, K. Maramorosch, A. Dubendorfer, Eds. (North-Holland, Elsevier, Amsterdam, 1980), pp. 517-533.

41. L. Nagy, P. Tontonoz, J. Alvarez, H. Chen, R. Evans, *Cell* **93**, 229 (1998).

42. A. Pearson, A. Lux, M. Krieger, *Proc. Natl. Acad. Sci. U.S.A.* **92**, 4056 (1995).

43. V. Rodrigues, P. Cheah, K. Ray, W. Chia, *EMBO J.* **14**, 3007 (1995).

44. M. Ashburner, *Drosophila: A Laboratory Manual* (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

45. We thank J. Fessler and M. Hortsch for providing antibodies; E. Nöll, N. Perrimon, the Bloomington Stock Center, and I. Dobens for fly stocks; M. Krieger for the Dii AcLDLs; Y. Ge and W. Fowle for assistance with confocal microscopy and histology; the lab of T. Orr-Weaver for advice on nuclear dyes; I. Ando for suggesting the bacteria assay experiment; and J. Settleman and the members of the Ezekowitz and White laboratories for helpful comments on this work. This work was supported by grants from the Shiseido Company of Japan to Massachusetts General Hospital/Harvard Medical School (N.C.F. and K.W.), from NIH (K.W. and A.E.), and from the Human Frontiers in Science Program (A.E. and N.C.F.).

5 February 1999; accepted 12 May 1999

Vessel Cooption, Regression, and Growth in Tumors Mediated by Angiopoietins and VEGF

J. Holash,¹ P. C. Maisonpierre,¹ D. Compton,¹ P. Boland,¹ C. R. Alexander,¹ D. Zagzag,² G. D. Yancopoulos,^{1*} S. J. Wiegand^{1*}

In contrast with the prevailing view that most tumors and metastases begin as avascular masses, evidence is presented here that a subset of tumors instead initially grows by coopting existing host vessels. This coopted host vasculature does not immediately undergo angiogenesis to support the tumor but instead regresses, leading to a secondarily avascular tumor and massive tumor cell loss. Ultimately, however, the remaining tumor is rescued by robust angiogenesis at the tumor margin. The expression patterns of the angiogenic antagonist angiopoietin-2 and of pro-angiogenic vascular endothelial growth factor (VEGF) suggest that these proteins may be critical regulators of this balance between vascular regression and growth.

It is widely accepted that most tumors and metastases originate as small avascular masses that belatedly induce the development of new blood vessels once they grow to a few millimeters in size (1-3). Initial avascular growth would be predicted for tumors that arise in epithelial structures that are separated from the underlying vasculature by a basement membrane and for experimental tumors that are implanted into avascular settings (such as the cornea pocket) or into a virtual

space (such as the subcutaneum) (2, 3). However, there is also evidence to suggest that tumors in more natural settings do not always originate avascularly, particularly when they arise within or metastasize to vascularized tissue (4). In such settings, tumor cells may coopt existing blood vessels (4). The interplay between this coopting of existing vessels and subsequent tumor-induced angiogenesis has not been extensively examined nor has the role of angiogenic factors in this process.

The pro-angiogenic vascular endothelial growth factors (VEGFs) and the angiopoietins are the only known growth factor families that are specific for the vascular endothelium because expression of their receptors is restricted to these cells (5, 6). The angiopoietins include both receptor activators [angiopoietin-1 (Ang-1)] and receptor antagonists [angiopoietin-2

¹Regeneron Pharmaceuticals, 777 Old Saw Mill River Road, Tarrytown, NY 10591, USA. ²Microvascular and Molecular Neuro-Oncology Laboratory, Department of Pathology, Kaplan Cancer Center, New York University Medical Center, New York, NY 10016, USA.

*To whom correspondence should be addressed. E-mail: gdy@regpha.com (G.D.Y.); stan.wiegand@regpha.com (S.J.W.)

REPORTS

(Ang-2)] (7–10). The VEGFs and the angiopoietins seem to play complementary and coordinated roles in vascular development (9, 11). During development, VEGF acts via the Flk1/KDR receptor to promote endothelial cell differentiation, proliferation, and primitive vessel formation (12). Ang-1 subsequently acts via the Tie2 receptor to remodel these primitive vessels and is then thought to help maintain and stabilize the mature vessels by promoting interactions between endothelial cells and surrounding support cells (6–9, 11, 13, 14). In adults, Ang-2 is expressed primarily at sites of vascular remodeling (9, 11), where it is thought to block the constitutive stabilizing action of Ang-1. It has been proposed that destabilization by Ang-2 in the absence of VEGF leads to frank vessel regression, whereas such destabilization in the presence of high VEGF levels facilitates the angiogenic response (9, 11). In tumors, hypoxia-induced VEGF (15) apparently recapitulates its developmental actions by contributing to the onset of tumor-associated angiogenesis, and antagonists of VEGF have been shown to inhibit the growth of many tumors (16).

To explore the possibility that VEGFs and angiopoietins collaborate during tumor angiogenesis, we studied early angiogenic events using the rat C6 glioma model (17). Remarkably, even the smallest C6 gliomas at just 1 week after implantation (<1 mm in diameter) were found to be well vascularized (Fig. 1, A and A'). As previously noted (17), this is attributable to the coopting of existing brain blood vessels by the implanted tumor cells. The vessels within these early tumors were similar to normal brain vessels in caliber and heterogeneity. There was no evidence of angiogenesis, as judged by the lack of vascular sprouts, non-canalized endothelial cell chains, and hyperplastic vessels. By 2 weeks after implantation, the tumors had grown to ≥ 2 mm in diameter but still showed no obvious angiogenic response. Rather, they exhibited a dramatic decrease in vessel density, presumably due to tumor growth in the absence of compensatory angiogenesis (Fig. 1, B and B'). The vessels within the tumors were distinctly larger and more homogeneous in caliber than the microvasculature of the normal brain. By 4 weeks after implantation, the tumors measured several millimeters in diameter and showed marked changes in comparison with tumors at earlier stages of development (Fig. 1, C and C'). Blood vessels within the core of the tumor had undergone dramatic regression, with no evidence of a local, compensatory angiogenic response. The centers of the tumors were largely bereft of vessels, leading to massive tumor cell death (Fig. 1, C and C'). The remaining cells in the tumor interior were organized in cuffs of pseudopalisading cells around the few surviving internal vessels (Fig. 1, C and C'). In contrast to the tumor interior, the tumor periphery displayed robust angiogenesis (Fig. 1, C and C').

Regression of coopted blood vessels was a very early event that preceded tumor cell death. Apoptotic cells were predominantly found in blood vessels in early-stage tumors, whereas at later stages there was widespread apoptosis of tumor cells (Fig. 2, A through C). Staining with markers for both endothelial cells and supporting pericytes or smooth muscle cells revealed that vessel regression

was associated with progressive disengagement of endothelial cells from surrounding support cells (Fig. 1, D through G).

The apparent association of tumor vessel regression, apoptosis, and disruption of endothelial cell interactions with support cells raised the possibility that blockade of the stabilizing action of Ang-1 might be contributing to tumor vessel regression. Consistent with this possibil-

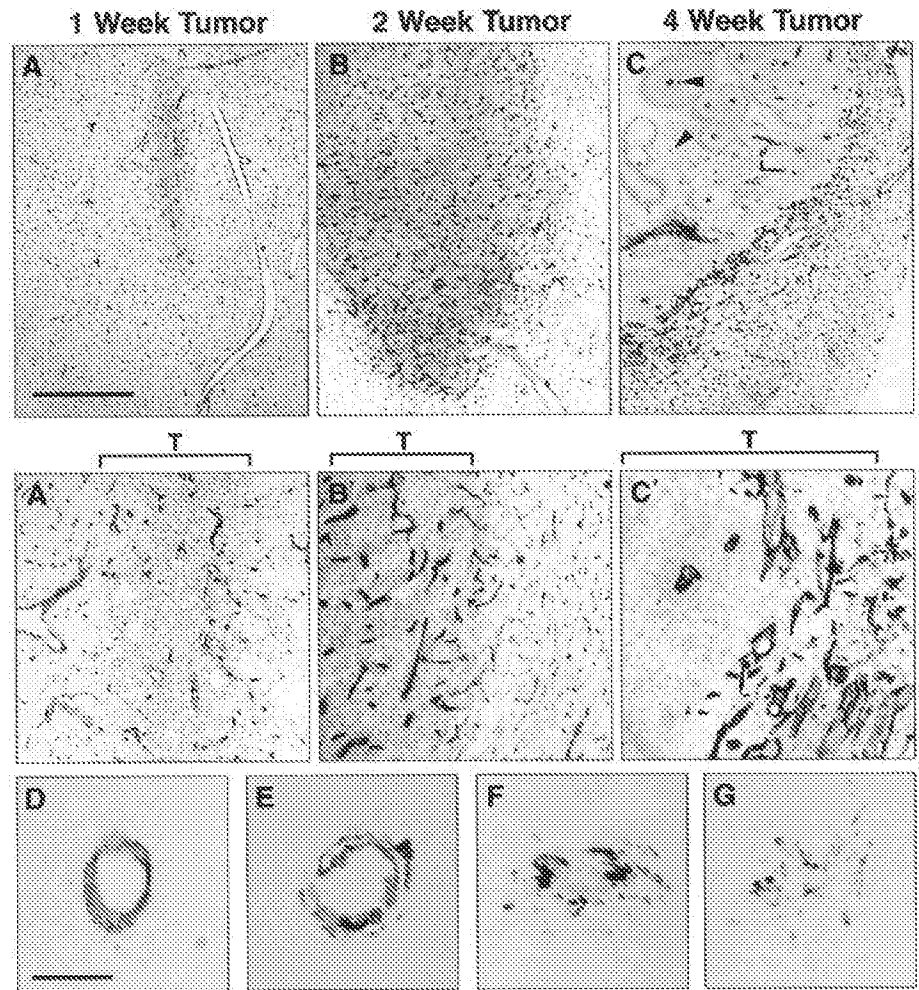


Fig. 1. Sections from rat C6 gliomas (28) showing progressive vessel regression, accompanied by dissociation of endothelial and smooth muscle cells. (A and A') Small 1-week tumors that measure a fraction of a millimeter in width are well vascularized as determined by RECA immunostaining (29), apparently because they coopt and grow around existing vessels. [T, tumor; scale bar in (A), 1 mm for (A) through (C) and 200 μ m for (A') through (C')] The vessels in early tumors resemble vessels in surrounding brain tissue in both density and morphology. (B and B') Two-week tumors continue to have extensive internal vasculature, although the vessel density is less than that in surrounding brain tissue, presumably because of the growth of the tumor in the absence of compensatory angiogenesis from existing internal vessels; the caliber of the internal vessels within these tumors does become dilated and relatively uniform compared to normal brain vessels. (C and C') Within large 4-week tumors, internal vessels regress with accompanying loss of surrounding tumor (necrotic tumor areas are unstained). Surviving internal vessels are sparse and uniform, are centrally located with respect to surrounding cuffs of well-stained viable tumor cells, and exhibit no evidence of compensatory angiogenesis; although robust angiogenesis is apparent at the margin of the tumor, where increased density of ectatic vessels is noted. Arrowheads in (C) depict a patent (top) and a regressed (bottom) vessel, each surrounded by either a surviving or regressed cuff of tumor. (D through G) Immunostaining with antibodies to SMA (black) and RECA (brown) (29) shows that pericytes and smooth muscle cells detach from the vessel wall in tumors. (D) shows a vessel wall in normal brain tissue in which RECA and SMA staining are essentially superimposed, whereas (E) through (G) depict vessels within tumors with progressive detachment of SMA-positive cells and vessel regression. Scale bar in (D) indicates 50 μ m for (D) through (G).

Fig. 2. Detection of apoptosis in rat C6 gliomas. Vessel-specific apoptosis (25, 29) is evident in early tumors (A and B), and this is followed by widespread apoptosis of tumor cells at later stages (C); arrowheads denote vessel-specific apoptotic figures (stained black) in panels. Scale bar in (A), 10 μ m. Flow cytometry experiments (E through H) indicate that Ang-1 can be as effective as VEGF in preventing apoptosis of serum-starved endothelial cells, as judged by a decreased percentage of endothelial cells with hypodiploid DNA content (see percentages over the sub-G₀/G₁ peak delineated by brackets). Cell number is shown on the y axis. PI, propidium iodide. However, in contrast to VEGF, Ang-1 cannot promote DNA synthesis in these cells (D) (30). Similar data have just been reported (37).

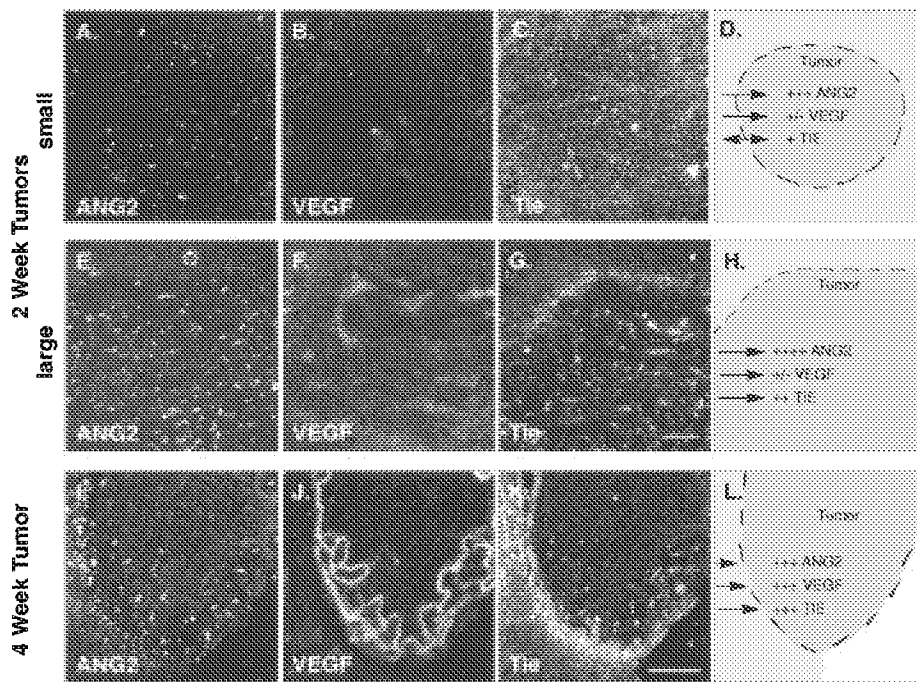
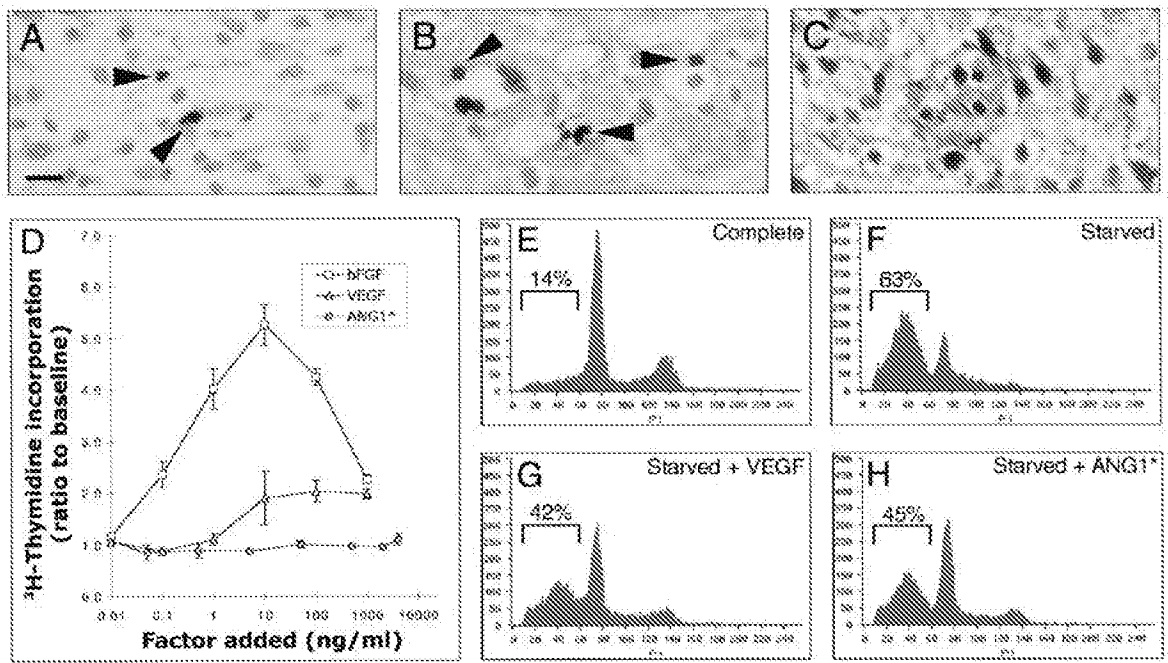


Fig. 3. In situ hybridization analysis of Ang-2, VEGF, and Tie mRNA in two different 2-week rat gliomas (small and large) and a large 4-week rat glioma (29, 32). At 2 weeks, the vessels within both a small tumor (A through D) and a larger tumor (E through H) consistently express high levels of Ang-2 mRNA (A and E). In contrast, up-regulation of Tie mRNA (C and K) is restricted to the larger tumor. Induction of VEGF is minimal in small tumors (B) and is still modest and patchy in larger tumors (F). In very large 4-week tumors, the tumor is secondarily avascular because of massive vessel regression and thus has few internal vessels, but has a hypervascular plexus at the tumor border. The few internal and the many rim vessels are now marked by both Ang-2 and Tie (I and K), although expression of Ang-2 is more punctate than that of Tie. The remaining live tumor cuffs around vessels show dramatically up-regulated VEGF expression (J). This VEGF expression is highest in palisading, presumably hypoxic, tumor cells that are furthest from vessels; large areas within the tumor, between palisading cells, are necrotic. (D), (H), and (L) outline the boundaries of the tumor within the brain and indicate the relative levels of expression of Ang-2, VEGF, and Tie. Scale bar in (G) indicates 500 μ m for (A) through (H); scale bar in (K) indicates 1 mm for (I) through (L).

ity, Ang-1 was found to be anti-apoptotic for cultured endothelial cells (Fig. 2, E through H), and expression of its antagonist, Ang-2, was found to be induced in the endothelium of coopted tumor vessels before their regression (Fig. 3A). In contrast, marked induction of VEGF expression occurred much later in tumor progression, in the hypoxic periphery of tumor cells surrounding the few remaining internal vessels, as well as adjacent to the robust plexus of vessels at the tumor margin (Fig. 3, B, F, and J). Expression of Ang-2 continued to mark not only the few surviving internal vessels but also the angiogenic vessels at the tumor margin (Fig. 3I), which suggests that the destabilizing action of Ang-2 facilitates the angiogenic action of VEGF at the tumor rim. Ang-1 expression did not change significantly throughout tumor development. Consistent with its expression in C6 glioma cells in culture (18), in relatively small tumors Ang-1 mRNA was expressed in a diffuse pattern by the tumor cells themselves (19) at levels just above that in the normal brain. Unlike VEGF, Ang-1 was not expressed at elevated levels in hypoxic regions of large tumors (20–22).

We also examined human glioblastomas (20, 21). Ang-2 was not detectable in the normal human brain, but its expression was dramatically induced in coopted tumor vessels, preceding vessel regression. As in the rat C6 model, this occurred in association with a disruption of interactions between endothelial and smooth muscle cells and with endothelial cell apoptosis. Diffuse Ang-1 expression in the human tumors also resembled that seen in the rat model (20–22).

REPORTS

To examine whether these findings are generalizable to other tumor types, we implanted rat RBA mammary adenocarcinoma cells into rat brains. Rather than growing avascularly, the implanted RBA cells rapidly associated with and migrated along cerebral blood vessels in a manner even more striking than that observed with the glioma cells (Fig. 4, A and D). Consistent with the well-vascularized state of these early tumors, there was minimal up-regulation of VEGF (22). However, the coopted vessels displayed striking and specific up-regulation of Ang-2, which was not detectable in the vessels of adjacent brain tissue (Fig. 4B). Preliminary analysis of RBA tumors at a later stage indicated that Ang-2 expression was associated with a pattern of vascular regression (in the absence of VEGF) and angiogenesis (in the presence of VEGF), as was the case with gliomas (22). Ang-1 was not expressed in cultured RBA cells or the tumors themselves (22).

Examination of a model of tumor metastasis, in which the mouse lung is colonized by intravenously injected Lewis lung carcinoma cells, yielded similar results. Tiny tumor metastases (arrowheads, Fig. 4, E and F) as well as moderately sized tumor nodules (arrows, Fig. 4, E and F) were closely associated with pulmonary vessels, and these vessels showed dramatic induction of Ang-2 expression (Fig. 4F). Progressively larger tumor nodules appeared to be characterized by vessel regression as well as neo-angiogenesis, again correlating with Ang-2 and VEGF expression (22).

In summary, our analyses of several different tumor models suggest a modification of the prevailing view that most malignancies and metastases originate as avascular masses that only belatedly induce angiogenic support. Our findings indicate that a subset of tumors rapidly coopts existing host vessels to form an initially well-vascularized tumor mass. Perhaps as part of a host defense mechanism, there is widespread regression of these initially coopted vessels, leading to a secondarily avascular tumor and massive tumor cell loss; however, the remaining tumor is ultimately rescued by robust angiogenesis at the tumor margin.

The expression patterns of VEGF and the natural Tie2 receptor antagonist Ang-2 strongly implicate them in these processes. There is a striking induction of Ang-2 expression in coopted vessels before induction of VEGF expression in the adjacent tumor cells, providing perhaps the earliest marker of tumor vasculature. The intense autocrine expression of Ang-2 by endothelial cells in tumor-associated vessels may counter a paracrine stabilization or survival signal provided by low-level constitutive expression of Ang-1 in normal tissues. We hypothesize that Ang-2 "marks" the coopted vessels for regression by an apoptotic mechanism that may involve disrupted interactions between

endothelial cells and the surrounding extracellular matrix and supporting cells. Subsequently, VEGF up-regulation coincident with Ang-2 expression at the tumor periphery is associated with robust angiogenesis. This late expression of tumor-derived VEGF may nullify the regression signal provided by Ang-2, which is consistent with the observation that VEGF is required for tumor vessel survival (23).

The angiogenic properties of tumor-derived VEGF may actually be facilitated when vessels are destabilized by Ang-2. Newly formed tumor vessels are often tenuous, poorly differentiated, and undergo regressive changes even as blood vessel proliferation continues. The failure of many solid tumors to form a well-differentiated and stable vasculature may be attributable to the fact that newly formed tumor vessels continue to overexpress Ang-2. In fact, hyper-vascular hepatomas with aberrant vasculatures

show high levels of Ang-2 expression in their endothelium (24). Thus, a persistent blockade of Tie2 signaling, which is otherwise constitutively activated in many normal adult tissues (14), may prevent tumor vessel differentiation and maturation and contribute to their generally tenuous and leaky nature.

In tumors, Ang-2 and VEGF apparently reprise the roles they play during vascular remodeling in normal tissues, acting to regulate the previously underappreciated balance between vascular regression and growth. Our findings bolster the case for anti-VEGF therapies in cancer, not only to prevent further angiogenesis but also perhaps to promote the regression of fragile new tumor vessels. Ang-2 appears to be the earliest marker of blood vessels that have been perturbed by invading tumor cells. As such, Ang-2 may prove to be useful in the imaging of very

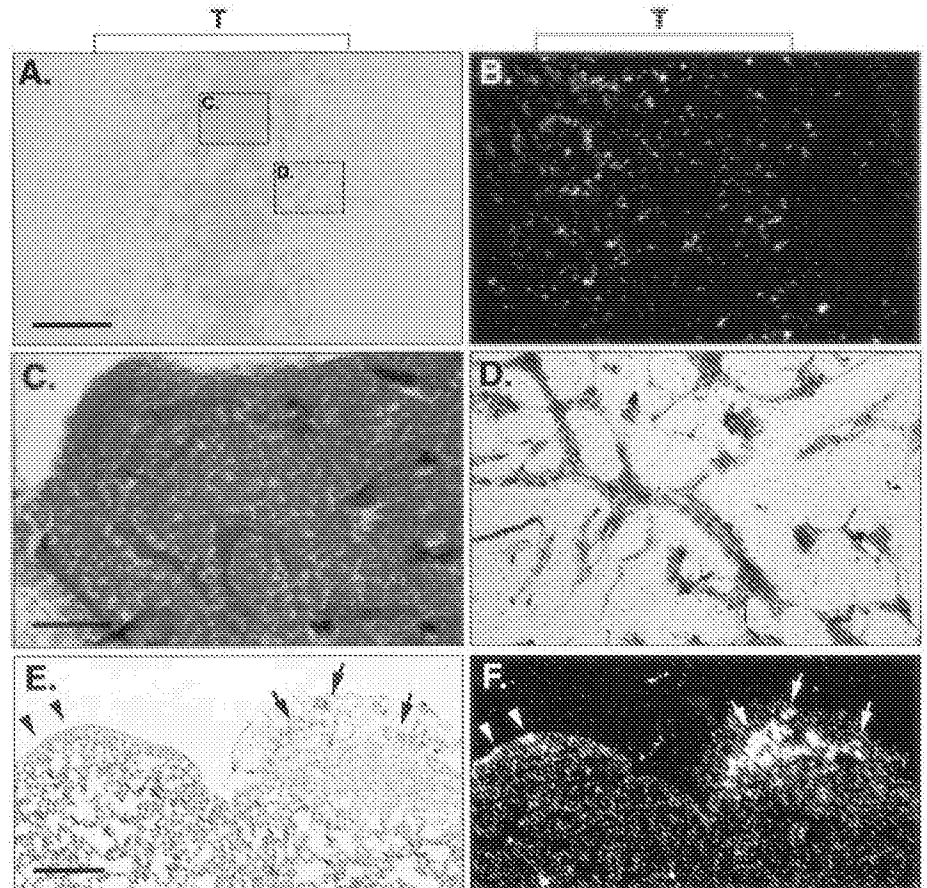


Fig. 4. In situ hybridization analysis (32) of rat RBA mammary carcinomas (28) and mouse Lewis lung carcinomas (28), showing up-regulation of Ang-2 mRNA in coopted tumor vessels. (A) A section through a mammary carcinoma stained with cresyl violet demonstrates the invasiveness of the tumor cells in the brain. The boxes within (A) delineate regions of the tumor core and periphery. Similar regions in specimens stained with an antibody to RECA (29) are shown in (C) and (D). The dramatic homing of tumor cells to blood vessels is especially apparent in (D). (B) Whereas VEGF is typically weak or undetectable at this tumor stage (22), Ang-2 is highly expressed in a punctate manner by blood vessels. (E) A section through a Lewis lung carcinoma stained with Papanicolaou demonstrates a metastasis only a few cells thick (left, arrowheads) and a slightly larger metastasis (right, arrows). Vessels, stained black with antibodies to PECAM (29), lie within these small tumors. (F) The vessels coopted by the small Lewis lung metastases exhibit dramatic induction of Ang-2. Scale bar in (A) indicates 500 μ m for (A) and (B); scale bar in (C) indicates 25 μ m for (C) and 50 μ m for (D); scale bar in (E) indicates 500 μ m for (E) and (F).

small tumors and metastases and possibly in schemes designed to specifically target chemotoxic therapy to tumor vasculature.

References and Notes

1. J. Folkman, *N. Engl. J. Med.* **285**, 1182 (1971).
2. _____, *J. Natl. Cancer Inst.* **82**, 4 (1990).
3. D. Hanahan and J. Folkman, *Cell* **86**, 353 (1996).
4. P. Wesseling, J. A. van der Laak, H. de Leeuw, D. J. Ruiter, P. C. Burger, *J. Neurosurg.* **81**, 902 (1994); L. Holmgren, M. S. O'Reilly, J. Folkman, *Nature Med.* **1**, 149 (1995); F. Pezzella, et al., *Am. J. Pathol.* **151**, 1417 (1997).
5. D. J. Dumont, T. P. Yamaguchi, R. A. Conlon, J. Rossant, M. L. Breitman, *Oncogene* **7**, 1471 (1992); J. Partanen et al., *Mol. Cell. Biol.* **12**, 1698 (1992); A. Iwama et al., *Biochem. Biophys. Res. Commun.* **195**, 301 (1993); P. C. Maisonnier, M. Goldfarb, G. D. Yancopoulos, G. Gao, *Oncogene* **8**, 1631 (1993); T. N. Sato, Y. Qin, C. A. Kozak, K. L. Audus, *Proc. Natl. Acad. Sci. U.S.A.* **90**, 9355 (1993); H. Schnurch and W. Risau, *Development* **119**, 957 (1993); S. F. Ziegler, T. A. Bird, S. A. Schneringer, K. A. Schooley, P. R. Baum, *Oncogene* **8**, 663 (1993); N. Ferrara, *Curr. Top. Microbiol. Immunol.* **237**, 1 (1999).
6. D. J. Dumont et al., *Genes Dev.* **8**, 1897 (1994); T. N. Sato et al., *Nature* **376**, 70 (1995).
7. S. Davis et al., *Cell* **87**, 1161 (1996).
8. C. Suri et al., *ibid.*, p. 1171.
9. P. C. Maisonnier et al., *Science* **277**, 55 (1997).
10. D. Valenzuela et al., *Proc. Natl. Acad. Sci. U.S.A.* **96**, 1904 (1999).
11. A diagram depicting the complementary and coordinated roles of VEGF and the angiopoietins during vascular growth and development can be found at Science Online (Web Fig. 1 at www.sciencemag.org/feature/data/1039904.shl).
12. F. Shalaby et al., *Nature* **376**, 62 (1995); P. Carmeliet et al., *ibid.* **380**, 435 (1996); N. Ferrara et al., *ibid.*, p. 439.
13. C. Suri et al., *Science* **282**, 468 (1998).
14. A. Wong et al., *Circ. Res.* **81**, 567 (1997).
15. D. Shweiki, M. Neeman, A. Itin, E. Keshet, *Proc. Natl. Acad. Sci. U.S.A.* **92**, 768 (1995).
16. K. J. Kim et al., *Nature* **362**, 841 (1993); B. Millauer, L. K. Shawver, K. H. Plate, W. Risau, A. Ullrich, *ibid.* **367**, 576 (1994); R. S. Warren, H. Yuan, M. R. Matli, N. A. Gilllett, N. Ferrara, *J. Clin. Invest.* **95**, 1789 (1995); B. Millauer et al., *Cancer Res.* **56**, 1615 (1996); C. K. Goldman et al., *Proc. Natl. Acad. Sci. U.S.A.* **95**, 8795 (1998).
17. N. Nagano, H. Sasaki, M. Aoyagi, K. Hirakawa, *Acta Neuropathol.* **86**, 117 (1993).
18. B. Erholm et al., *Oncogene* **14**, 2475 (1997).
19. A figure showing the expression patterns of Tie1, Tie2, Ang-1, Ang-2, and VEGF in early rat C6 gliomas can be found at Science Online (Web Fig. 2 at www.sciencemag.org/feature/data/1039904.shl).
20. A. Stratmann, W. Risau, K. H. Plate, *Am. J. Pathol.* **153**, 1459 (1998).
21. A complete description of angiopoietin expression in human glioblastomas can be found at Science Online (Web Fig. 3 at www.sciencemag.org/feature/data/1039904.shl); D. Zagzag et al., *Exp. Neurol.*, in press.
22. J. Holash et al., unpublished observations; D. Zagzag et al., unpublished observations.
23. L. Benjamin, D. Golijanin, A. Itin, D. Podes, E. Keshet, *J. Clin. Invest.* **103**, 159 (1999).
24. S. Tanaka et al., *ibid.*, p. 341.
25. S. D. Morgenbesser et al., *EMBO J.* **14**, 743 (1995).
26. J. K. Morse et al., *J. Neurosci.* **13**, 4146 (1993).
27. D. M. Valenzuela et al., *Neuron* **10**, 963 (1993).
28. Tumor cells were obtained from the American Type Culture Collection and grown in culture. About 1.0×10^5 C6 or RBA cells were suspended in $\sim 2 \mu\text{l}$ of phosphate-buffered saline (PBS) and injected stereotactically over a period of 5 to 10 min into the right striatum (AP + 0.5; ML ~ -3.0 ; DV ~ -6.0 relative to Bregma) of adult male Sprague-Dawley rats. About 5.0×10^5 Lewis lung carcinoma cells were suspended in $50 \mu\text{l}$ of serum-free media and injected into the jugular vein of adult male C57BL mice.
29. Animals were anesthetized and either decapitated or

perfused with 4% paraformaldehyde. Brains for thick sliding microtome sections ($40 \mu\text{m}$) were post-fixed in 4% paraformaldehyde overnight and then equilibrated in 35% sucrose. Fixed brains for thin sections ($10 \mu\text{m}$) were equilibrated in 17% sucrose. Fixed and fresh brains were frozen in methylbutane, chilled in dry ice, and sectioned on a cryostat. For TUNEL labeling (25), brains were immersion-fixed in 10% neutral buffered formalin and embedded in paraffin. Fixed sections were immunostained with a monoclonal antibody to rat endothelial cell antigen (RECA 1; 1:250; Serotec) and a biotinylated horse anti-mouse secondary antibody (1:1500; Vector) or with a monoclonal antibody to PECAM (CD31; 1:100; Pharmingen) and a biotinylated rabbit anti-rat secondary antibody (1:150; Vector) as previously described (26). A similar protocol was used for double labeling. Sections were initially labeled with a monoclonal antibody to alpha smooth muscle actin (SMA; 1:500; DAKO) and a biotinylated goat anti-mouse immunoglobulin G IIa secondary antibody (1:1250; Amersham). SMA staining was visualized with a Vectastain Elite kit (Vector), and a black reaction product was generated by nickel sulfate enhancement. After SMA labeling, sections were then reblocked and labeled with antibody to RECA (1:100). A brown reaction product was used.

30. Human umbilical vein endothelial cells (HUVECs) (Clonetics, San Diego, CA) were maintained in recommended medium on gelatin-coated plastic. For DNA synthesis assays, 1×10^4 cells were plated in 96-well microtiter wells and grown for 24 hours in basal medium plus 0.5% fetal bovine serum. Cells were re-fed with the same medium plus purified factors and grown for 20 hours, with 1 mCi tritiated thymidine (80 Ci/mmol; Amersham) being present for the last 3 hours of incubation. Cells were rinsed and fixed with trichloroacetic acid, and thymidine incorporation was measured

by standard liquid scintillation techniques. Ang-1* (ANG1*) was a modified form of human Ang-1, described previously (9); VEGF was murine VEGF-164, produced and purified from baculovirus-infected insect cells; bFGF was human basic fibroblast growth factor (R&D Systems). For assessing resistance to apoptosis, plates of $\sim 80\%$ confluent HUVECs were rinsed twice with basal medium and grown for 18 to 20 hours in basal medium and bovine serum albumin (0.5 mg/ml), plus or minus purified factors. Both adherent and non-adherent cells were harvested, pooled, and fixed in 70% ethanol at -20°C overnight. Cells were washed in PBS, incubated for 30 min with ribonuclease A (5 kunitz units/ml; Sigma) and propidium iodide (50 $\mu\text{g}/\text{ml}$; Sigma). Cellular DNA content, as judged by propidium iodide fluorescence, was measured by flow cytometry (MoFlo, Cytomation, Fort Collins, CO).

31. A. Papapetropoulos et al., *Lab. Invest.* **79**, 213 (1999).
32. Fresh frozen or fixed sections were probed with ^{35}S -labeled cRNAs (27). Probes for VEGF and Ang-1 and Ang-2 have been described (9). For Tie1, a 1.3-kb fragment of rat Tie1 spanning the last 309 codons and 375 base pairs of the 3' untranslated sequence was used, and for Tie2 a 460-base pair fragment spanning codons 771 through 924 within the kinase domain was used. This probe does not cross-hybridize to Tie1 mRNA in Northern blots.
33. We thank B. Luan, J. Zheng, P. Burfeind, S. Zabski, and F. Martin for excellent technical assistance; E. Burrows and C. Murphy for graphics work; A. Hooper and D. Friedlander for data on human gliomas; and M. Grumet for intellectual discussions. Supported in part by a grant from the Children's Brain Tumor Foundation to D.Z. and by Procter & Gamble Pharmaceuticals, Inc. All animal studies were done in accordance with institutional guidelines.

9 March 1999; accepted 10 May 1999

Initiation of Mammalian Liver Development from Endoderm by Fibroblast Growth Factors

Joonil Jung,¹ Minghua Zheng,¹ Mitchell Goldfarb,² Kenneth S. Zaret^{1*}

The signaling molecules that elicit embryonic induction of the liver from the mammalian gut endoderm or induction of other gut-derived organs are unknown. Close proximity of cardiac mesoderm, which expresses fibroblast growth factors (FGFs) 1, 2, and 8, causes the foregut endoderm to develop into the liver. Treatment of isolated foregut endoderm from mouse embryos with FGF1 or FGF2, but not FGF8, was sufficient to replace cardiac mesoderm as an inducer of the liver gene expression program, the latter being the first step of hepatogenesis. The hepatogenic response was restricted to endoderm tissue, which selectively coexpresses FGF receptors 1 and 4. Further studies with FGFs and their specific inhibitors showed that FGF8 contributes to the morphogenetic outgrowth of the hepatic endoderm. Thus, different FGF signals appear to initiate distinct phases of liver development during mammalian organogenesis.

Identifying the molecular signals that initiate organogenesis from the gut is important for understanding the fundamental mechanisms

of developmental regulation, hereditary digestive disorders, and tissue regeneration. Different segments of the mammalian gut endoderm give rise to the liver, lung, pancreas, thyroid, and gastrointestinal tract. Typically, a portion of the endoderm will begin to express genes specific to one of these tissues, and then the newly specified cells will proliferate out of the endoderm layer to form a tissue bud, initiating morphogenesis (1, 2). In *Drosophila*, the initial specification of tissues

¹Department of Molecular Biology, Cell Biology, and Biochemistry, Brown University, Box G-J363, Providence, RI 02912, USA. ²Brookdale Center for Molecular Biology, Mount Sinai School of Medicine, New York, NY 10029, USA.

*To whom correspondence should be addressed: E-mail: zaret@brown.edu

Vessel Cooption, Regression, and Growth in Tumors Mediated by Angiopoietins and VEGF

J. Holash, P. C. Maisonpierre, D. Compton, P. Boland, C. R. Alexander, D. Zagzag, G. D. Yancopoulos and S. J. Wiegand

Science **284** (5422), 1994-1998.
DOI: 10.1126/science.284.5422.1994

ARTICLE TOOLS	http://science.sciencemag.org/content/284/5422/1994
REFERENCES	This article cites 35 articles, 10 of which you can access for free http://science.sciencemag.org/content/284/5422/1994#BIBL
PERMISSIONS	http://www.sciencemag.org/help/reprints-and-permissions

Use of this article is subject to the Terms of Service

Science (print ISSN 0036-8075; online ISSN 1095-9203) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. 2017 © The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works. The title *Science* is a registered trademark of AAAS.

Intravitreal Aflibercept Injection for Macular Edema Resulting from Central Retinal Vein Occlusion

One-Year Results of the Phase 3 GALILEO Study

Jean-François Korobelnik, MD,^{1,2,3} Frank G. Holz, MD,⁴ Johann Roeder, MD,⁵ Yuichiro Ogura, MD,⁶ Christian Simader, MD,⁷ Ursula Schmidt-Erfurth, MD,⁷ Katrin Lorenz, MD,⁸ Miki Honda, MD,⁹ Robert Vitti, MD,¹⁰ Alyson J. Berliner, MD, PhD,¹⁰ Florian Hiemeyer, MS,¹¹ Brigitte Stemper, MD,^{11,12} Oliver Zeitz, MD,^{11,13} Rupert Sandbrink, MD,^{11,14} for the GALILEO Study Group*

Purpose: To evaluate the efficacy and safety of intravitreal aflibercept injections for treatment of macular edema secondary to central retinal vein occlusion (CRVO).

Design: A randomized, multicenter, double-masked phase 3 study.

Participants: A total of 177 treatment-naïve patients with macular edema secondary to CRVO were randomized in a 3:2 ratio.

Methods: Patients received either 2-mg intravitreal aflibercept or sham injections every 4 weeks for 20 weeks. From week 24 to 48, the aflibercept group received aflibercept as needed (pro re nata [PRN]), and the sham group continued receiving sham injections.

Main Outcome Measures: The primary efficacy end point was the proportion of patients who gained 15 letters or more in best-corrected visual acuity (BCVA) at week 24. This study reports week 52 results including the proportion of patients who gained 15 letters or more in BCVA and the mean change from baseline BCVA and central retinal thickness. Efficacy end points at week 52 were all exploratory.

Results: At week 52, the mean percentage of patients gaining 15 letters or more was 60.2% in the aflibercept group and 32.4% in the sham group ($P = 0.0004$). Aflibercept patients, compared with sham patients, had a significantly higher mean improvement in BCVA (+16.9 letters vs. +3.8 letters, respectively) and reduction in central retinal thickness ($-423.5 \mu\text{m}$ vs. $-219.3 \mu\text{m}$, respectively) at week 52 ($P < 0.0001$ for both). Aflibercept patients received a mean of 2.5 injections (standard deviation, 1.7 injections) during PRN dosing. The most common ocular adverse events in the aflibercept group were related to the injection procedure or the underlying disease, and included macular edema (33.7%), increased intraocular pressure (17.3%), and eye pain (14.4%).

Conclusions: Treatment with intravitreal aflibercept provided significant functional and anatomic benefits after 52 weeks as compared with sham. The improvements achieved after 6 monthly doses at week 24 largely were maintained until week 52 with as-needed dosing. Intravitreal aflibercept generally was well tolerated. *Ophthalmology* 2014;121:202-208 © 2014 by the American Academy of Ophthalmology.



*Group members listed online in Appendix 1 (<http://aaajournal.org>).

The most common cause of vision loss in patients with central retinal vein occlusion (CRVO) is macular edema, which resolves spontaneously in only 30% of nonischemic cases and may not resolve in ischemic cases.^{1,2} Several lines of evidence indicate that vascular endothelial growth factor (VEGF) may play a key role in the pathophysiology of macular edema secondary to CRVO. Vascular endothelial growth factor is released in response to retinal hypoxia, which occurs in CRVO as a result of impaired capillary blood flow.³ Vascular endothelial growth factor stimulates angiogenesis and may result in neovascularization of the retina, the anterior segment, or

both, as well as vascular leakage resulting in macular edema.³ In CRVO patients, the vitreous level of VEGF correlates with the severity of macular edema.⁴ Furthermore, intravitreal injections of the anti-VEGF agents ranibizumab or aflibercept significantly improve visual and anatomic outcomes in patients with macular edema secondary to CRVO.⁵⁻⁹

Intravitreal aflibercept (historically known in the scientific literature as VEGF Trap-Eye; Regeneron Pharmaceuticals, Inc, Tarrytown, NY, and Bayer Healthcare Pharmaceuticals, Berlin, Germany) is a fusion protein of key domains from human VEGF receptors 1 and 2 with the

constant region (Fc) of human immunoglobulin G that binds to multiple VEGF-A isoforms with a higher affinity than ranibizumab and bevacizumab.¹⁰ Studies of intravitreal aflibercept injections in patients with neovascular age-related macular degeneration (AMD) demonstrate that aflibercept given monthly for 3 initial administrations and then once every 2 months improves visual and anatomic outcomes as effectively and safely as monthly ranibizumab over a 1-year period.¹¹ The efficacy and safety of intravitreal aflibercept for the treatment of macular edema secondary to CRVO was investigated in 2 parallel trials performed in Europe and in the Asia Pacific region (GALILEO) and in the United States (COPERNICUS).^{5,7,9} The primary efficacy end point of the GALILEO study was at week 24 and was published previously.⁹ Herein, we report the 52-week results of the GALILEO study.

Methods

Study Design

The GALILEO study is an 18-month, randomized, double-masked, phase 3 study comparing the efficacy and safety of intravitreal aflibercept with sham for the treatment of macular edema secondary to CRVO. The study protocol was approved by the institutional review board or ethics committee at each site. All patients signed a written consent form before initiation of the study-specific procedures. The study was registered with ClinicalTrials.gov (identifier no. NCT01012973) and was conducted across 63 sites in Europe and the Asia Pacific region in compliance with ethical guidelines from the Declaration of Helsinki and International Conference on Harmonization. Data for this 52-week report were collected between October 2009 and July 2011.

The design and eligibility criteria for the GALILEO study have been described previously.⁹ Only 1 eye from each patient was included in the study. Patients were randomized in a 3:2 ratio to receive 2 mg intravitreal aflibercept (IVT-AFL 2Q4) or sham injections in the study eye once every 4 weeks for 20 weeks, for a total of 6 doses (Fig 1). From weeks 24 to 52, patients in the aflibercept group were evaluated monthly and received aflibercept as needed (pro re nata [PRN]; IVT-AFL 2Q4 + PRN) if they had more than a 50- μ m increase in central retinal thickness (CRT) compared with the lowest previous measurement, new or persistent cystic changes within the neurosensory retina or sub-retinal fluid, persistent diffuse edema of 250 μ m or more in the central subfield, loss of 5 letters or more from the best prior measurement in conjunction with any increase in CRT, or an increase of 5 letters or more in best-corrected visual acuity (BCVA) from the most recent visit, potentially suggesting further improvements on a subsequent injection. If none of the retreatment criteria were met, patients received a sham injection to maintain masking. Patients in the sham group continued to receive sham injections at all visits through week 52. All patients were eligible to receive panretinal laser photocoagulation at any time during the study if they progressed to neovascularization of the anterior segment, optic disc, or elsewhere in fundus. Given that there was no approved treatment for CRVO when the GALILEO study was designed, no other rescue treatment was prespecified. The GALILEO study design included a full year of treatment with sham based on the request from health authorities. However, considering this long duration of sham treatment, the visual acuity and other ocular findings were monitored carefully by a team of masked medical reviewers. If, at any time, this review team had the impression that a patient may not benefit from further study

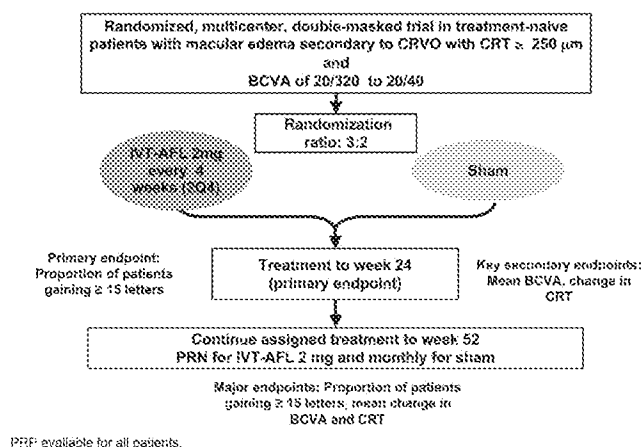


Figure 1. Diagram showing the GALILEO study design. BCVA = best-corrected visual acuity; CRT = central retinal thickness; CRVO = central retinal vein occlusion; IVT-AFL = intravitreal aflibercept; PRN = pro re nata (as needed); PRP = panretinal photocoagulation; 2Q4 = every 4 weeks.

participation or would be treated more adequately outside the study, the investigator was queried and asked to provide a reassessment of the patient. Investigators then used their medical judgment ultimately to determine whether it would still be beneficial for the patient to continue the study.

Outcome Measures

The primary efficacy end point of the GALILEO study was the proportion of patients achieving a gain of 15 letters or more in BCVA from baseline to week 24, which was published previously.⁹ Herein, we report the 52-week results of the GALILEO study. Efficacy end points at week 52 all were exploratory and included the proportion of patients who gained 15 letters or more in BCVA; mean change from baseline BCVA and CRT; proportion of patients progressing to neovascularization of the anterior segment, optic disc, or elsewhere in the fundus; and change from baseline in the mean 25-item National Eye Institute Visual Function Questionnaire total and subscale (distance activities, near activities, and vision dependency) scores.

The efficacy and safety end points were assessed as described previously.⁹ The BCVA and CRT were assessed at baseline and every 4 weeks afterward to week 52. Fundus photography and fluorescein angiography were performed at screening (days -21 to -1) and weeks 12, 24, 36, and 52. Retinal perfusion status was determined by fluorescein angiography. Perfused and nonperfused retinas were defined as those with less than 10 disc areas and 10 disc areas or more, respectively, of capillary nonperfusion on fluorescein angiography. Vision-related quality of life was assessed at baseline and weeks 24 and 52 using the 25-item National Eye Institute Visual Function Questionnaire, which was administered by masked site personnel before intravitreal injections.

Statistical Analyses

The efficacy end points were analyzed in the full analysis set (FAS), which included all randomized patients who received any study treatment and had a baseline and at least 1 postbaseline BCVA assessment. In a prespecified analysis of proportions of patients who gained 15 letters or more at week 24 (the primary efficacy end point), patients who discontinued before week 24 were

considered to be nonresponders. In a prespecified analysis of proportions of patients who gained 15 letters or more at week 52, the missing values were imputed by the last-observation-carried-forward method. Between-group differences in the proportion of patients who gained 15 letters or more were evaluated with a 2-sided Cochran-Mantel-Haenszel test.

Continuous variables were analyzed with an analysis of covariance, except for BCVA, which was assessed using an analysis of variance. The last-observation-carried-forward approach was used to impute missing values. For sensitivity, additional analyses were performed using observed values at week 52. The proportion of patients with neovascularization by week 52 was analyzed using a Cochran-Mantel-Haenszel test. Safety from baseline to week 24 was analyzed in the safety analysis set, which included all randomized patients who received any study treatment. Safety from weeks 24 to 52 was analyzed in week 24 completers within the safety analysis set.

Results

Of 240 patients screened, 106 patients were randomized to the IVT-AFL 2Q4 + PRN group, and 71 patients were randomized to the sham group. A total of 104 (98.1%) patients in the IVT-AFL 2Q4 + PRN group and 68 (95.8%) patients in the sham group were treated in the study and were included in the safety analysis set. One patient did not have any postbaseline BCVA value, and therefore was excluded from the FAS. Thus, the FAS included 103 patients in the IVT-AFL 2Q4 + PRN group and 68 patients in the sham group. Overall, 15 (14.2%) patients in the IVT-AFL 2Q4 + PRN group and 19 (26.8%) patients in the sham group discontinued the study before week 52. Major reasons for discontinuation in the IVT-AFL 2Q4 + PRN group were protocol violation (5 patients [4.7%]), withdrawal of consent (4 patients [3.8%]), and adverse events (4 patients [3.8%]). Major reasons for discontinuation in the sham group were lack of efficacy (6 patients [8.5%]), withdrawal of consent (6 patients [8.5%]), and adverse events (4 patients [5.6%]). No patient in the IVT-AFL 2Q4 + PRN group discontinued the study treatment because of a lack of efficacy.

Demographics and baseline disease characteristics of patients were similar in both treatment groups.⁹ Approximately half of patients had CRVO for less than 2 months (53.4% in the IVT-AFL 2Q4 + PRN group and 51.5% in the sham group, FAS). Most patients had a perfused retina (86.4% in the IVT-AFL 2Q4 + PRN group and 79.4% in the sham group) and a baseline BCVA of 35 letters or better (>20/200; 83.5% in the IVT-AFL 2Q4 + PRN group and 82.4% in the sham group).⁹

Visual Outcomes

At week 24, the proportion of patients who gained 15 letters or more in BCVA was 60.2% and 22.1% in the IVT-AFL 2Q4 and sham groups, respectively (patients who discontinued before week 24 were considered to be nonresponders; $P < 0.0001$).⁹ At week 52, the proportion of patients who gained 15 letters or more in BCVA was 60.2% in the IVT-AFL 2Q4 + PRN group versus 32.4% in the sham group (last observation carried forward; Fig 2A). More patients in the sham group had 15 letters or more of improvement in BCVA at week 52 compared with week 24 (32.4% vs. 22.1%, respectively). At week 52, patients treated with IVT-AFL 2Q4 + PRN maintained the improvements in BCVA achieved at week 24.

The proportion of patients who gained 10 or more letters and 30 or more letters or those who lost more than 0, more than 10, and more than 15 letters at week 52 are shown in Table 1. Overall, higher proportions of sham patients lost more than 0, more than

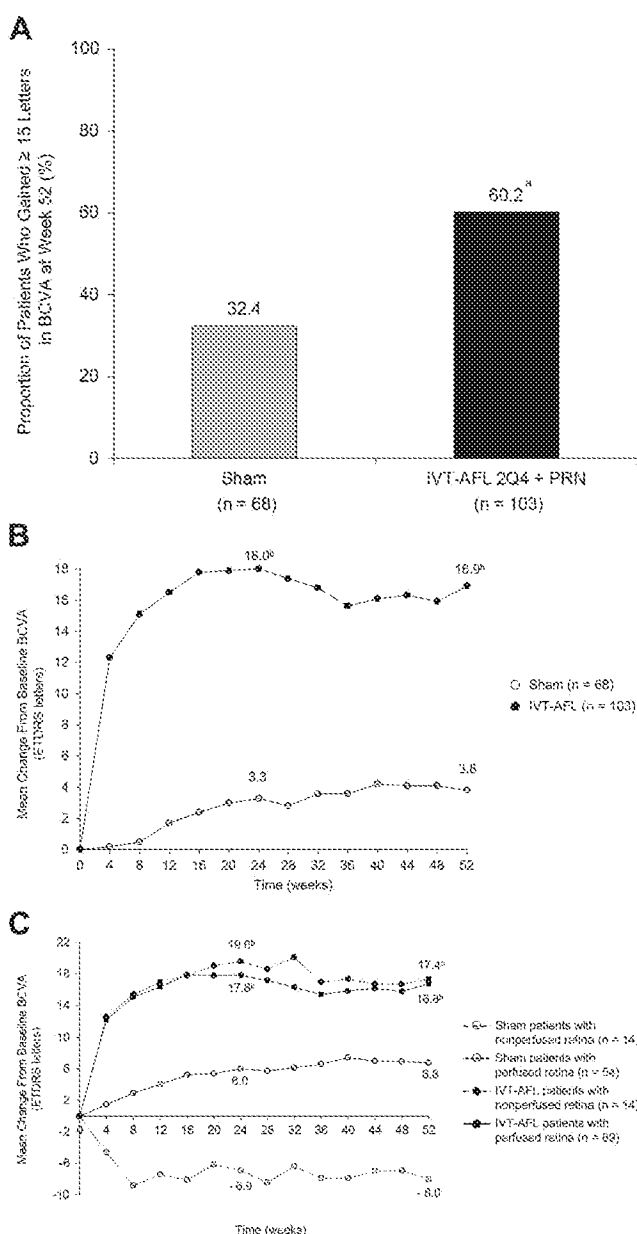


Figure 2. Graphs showing visual outcomes during the 52 weeks of the study: (A) percentage of patients who gained 15 letters or more at week 52, (B) mean change from baseline best-corrected visual acuity (BCVA), and (C) mean change from baseline BCVA by the status of retinal perfusion at baseline. Treatment frequency with intravitreal aflibercept (IVT-AFL) was every 4 weeks (2Q4) and pro re nata (PRN; as needed), respectively, before and after week 24. * $P = 0.0004$ vs. sham; ^b $P < 0.0001$ vs. sham; ^c $P < 0.001$ vs. sham. ETDRS = Early Treatment Diabetic Retinopathy Study.

10, and more than 15 letters compared with patients treated with IVT-AFL 2Q4 + PRN at week 52 (Table 1).

The mean change from baseline BCVA in the IVT-AFL 2Q4 + PRN and sham groups was 18.0 versus 3.3 letters at week 24 and 16.9 versus 3.8 letters at week 52 ($P < 0.0001$ for both; Fig 2B). When stratified by the baseline retinal perfusion status, patients treated with IVT-AFL 2Q4 + PRN had a similar mean ± standard deviation (SD) change from baseline BCVA in the perfused and nonperfused subgroups (+16.8±14.7 letters vs. +17.4±16.1

Table 1. Patients with Vision Gain and Loss at Week 52

	Week 52	
	Sham (n = 68)	Intravitreal Aflibercept Injection Monthly from Baseline to Week 24 plus Pro Re Nata Treatment from Weeks 24 to 52 (n = 103)
Vision gain, n (%)		
≥30 letters	5 (7.4)	15 (14.6)
≥15 letters	22 (32.4)	62 (60.2)
≥10 letters	26 (38.2)	74 (71.8)
Vision loss, n (%)		
>0 letters	30 (44.1)	11 (10.7)
>10 letters	16 (23.5)	1 (1.0)
>15 letters	10 (14.7)	1 (1.0)

letters, respectively; Fig 2C). In contrast, eyes with a perfused retina in the sham group gained a mean ± SD of 6.8±17.5 letters, whereas those with a nonperfused retina lost a mean of 8.0±15.8 letters at 52 weeks (Fig 2C). Regardless of the treatment group, patients with a baseline BCVA of 20/200 or worse had a greater BCVA gain than those with a baseline BCVA of better than 20/200 (9.4 vs. 2.5 letters for sham and 21.1 vs. 16.0 letters for IVT-AFL 2Q4 + PRN, respectively). Patients who had the disease for less than 2 months in the sham and IVT-AFL 2Q4 + PRN groups gained a mean of 2.1 letters and 19.5 letters from baseline, respectively, whereas those having the disease for 2 months or more gained a mean of 5.5 letters and 13.7 letters from baseline, respectively.

Anatomic Outcomes

At week 24, the mean CRT reduction from baseline was 448.6 μm and 169.3 μm in the IVT-AFL 2Q4 and sham groups, respectively (P < 0.0001). With the start of PRN dosing at week 24, CRT slightly increased in the IVT-AFL 2Q4 + PRN group, but then remained stable through week 52 (Fig 3). At week 52, the mean CRT reduction from baseline was significantly greater in the IVT-AFL 2Q4 + PRN group than in the sham group (423.5 μm vs. 219.3 μm, respectively; P < 0.0001). Regardless of the retinal perfusion status, patients treated with IVT-AFL 2Q4 + PRN had a greater CRT reduction (±SD) than those treated with sham (412.4±238.1 μm vs. 201.2±226.4 μm for the perfused subgroup and 494.6±318.4 μm vs. 294.3±258.6 μm for the nonperfused subgroup, respectively). During the 52-week study, 6 (5.8%) patients in the IVT-AFL 2Q4 + PRN group and 6 (8.8%) patients in the sham group developed neovascularization. In each group, 3 patients had a nonperfused

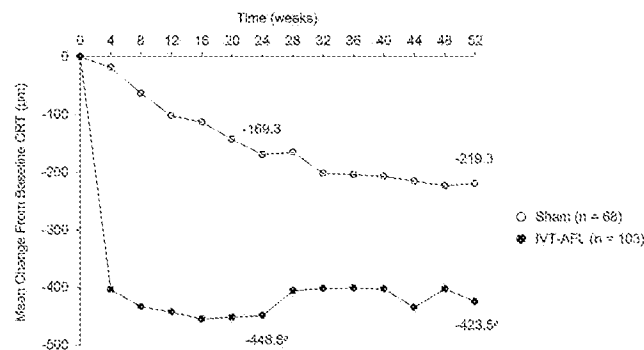


Figure 3. Graph showing the mean change from baseline central retinal thickness (CRT) during the 52 weeks of the study. Treatment frequency with intravitreal aflibercept (IVT-AFL) was every 4 weeks and pro re nata (as needed), respectively, before and after week 24. *P<0.0001 vs. sham.

retina at baseline, and 5 had disease duration of less than 2 months at baseline. In the IVT-AFL 2Q4 + PRN group, 4 patients demonstrated anterior segment neovascularization, 1 patient demonstrated neovascularization elsewhere in the fundus, and 1 patient demonstrated neovascularization both in anterior segment and elsewhere in the fundus. In the sham group, 4 patients demonstrated neovascularization of elsewhere in the fundus, 1 patient demonstrated anterior segment neovascularization, and 1 patient demonstrated neovascularization of optic disc. Panretinal photocoagulation was performed for 3 (4.4%) of the sham patients and 2 (1.9%) of the IVT-AFL 2Q4 + PRN patients.

Patient-Reported Outcomes

A clinically relevant improvement in the mean 25-item National Eye Institute Visual Function Questionnaire total score (≥4-point increase) was observed in both IVT-AFL 2Q4 + PRN group (7.8 points) and sham group (4.5 points) at week 52 (Table 2). The mean change from baseline to week 52 in near activities subscore was the highest among subscales, with IVT-AFL 2Q4 + PRN patients reporting a mean change of 12.2 points versus sham patients reporting a mean change of 5.0 points. No difference was noted between the 2 groups in the dependency subscale.

Study Drug Injections

During the 52 weeks of treatment, the mean (±SD) number of injections was 11.8±2.8 in the IVT-AFL 2Q4 + PRN group and 10.5±4.2 in the sham group. Most IVT-AFL 2Q4 + PRN patients (64 of 91 patients completing week 52 [70.3%]) received 3 or fewer IVT-AFL injections during weeks 24 to 52, with a mean ± SD of 2.5±1.7 injections during the PRN phase of study (Table 3). Patients who received 3 PRN injections or fewer had relatively higher BCVA gains than those who received 4 to 6 injections (Table 3). The median time to the first PRN intravitreal aflibercept injection was 83 days (95% confidence interval, 62–88 days).

Safety

The percentage of patients experiencing at least 1 ocular treatment-emergent adverse event (TEAE) in the sham and intravitreal aflibercept groups was 64.7% and 54.8% from baseline to week 24 and 50.9% and 69.1% from week 24 to week 52, respectively. The most common ocular TEAEs reported for the study eye in the intravitreal aflibercept group as compared with the sham group were eye pain (11.5% vs. 4.4%, respectively), increased intraocular pressure (8.7% vs. 5.9%, respectively), and conjunctival hemorrhage (8.7% vs. 4.4%, respectively) from baseline to week 24 and worsening of macular edema (35.1% vs. 10.5%, respectively), increased intraocular pressure (13.4% vs. 3.5%, respectively), and reduced visual acuity (11.3% vs. 1.8%, respectively) from weeks 24 to 52. All adverse events of intraocular pressure elevation were mild, except for 1 severe event that occurred in a sham patient before week 24. Ocular treatment-emergent serious adverse events (SAEs) are shown in Table 4. Most ocular SAEs were related to the disease state or injection procedure, and there were no clinically relevant differences between the treatment groups in terms of frequency or pattern of SAEs.

The incidence of nonocular TEAEs was similar in the sham and intravitreal aflibercept groups from baseline to week 24 (54.4% and 45.2%, respectively) and from weeks 24 to 52 (50.9% vs. 51.5%, respectively). Nasopharyngitis was the most commonly reported nonocular TEAE in both the sham and intravitreal aflibercept groups from baseline to week 24 (8.8% vs. 7.7%, respectively) and from weeks 24 to 52 (19.3% vs. 9.3%, respectively). Nonocular SAEs occurred in a small group of patients with a similar frequency in both the sham and intravitreal aflibercept groups from

Table 2. Change from Baseline to Weeks 24 and 52 in the National Eye Institute 25-Item Visual Function Questionnaire Score

	Baseline to Week 24*				Baseline to Week 52†			
	Mean Change		Difference in Least Square Mean Change (95% Confidence Interval)	P Value	Mean Change		Difference in Least Square Mean Change (95% Confidence Interval)	P Value
	Intravitreal Aflibercept Injection Monthly from Baseline to Week 24	Sham			Intravitreal Aflibercept Injection Monthly from Baseline to Week 24 Plus PRN Treatment from Week 24 to 52	Sham		
Total score	3.5	7.5	4.2 (1.7–6.8)	0.0013	4.5	7.8	3.6 (1.1–6.0)	0.0049
Distance activities subscore	2.4	6.3	3.5 (–0.3 to 7.2)	0.0689	3.9	8.4	4.2 (0.4–7.9)	0.0283
Near activities subscore	1.6	10.4	8.6 (4.0–13.2)	0.0003	5.0	12.2	6.9 (3.1–10.8)	0.0005
Dependency subscore	2.4	3.7	2.1 (–1.6 to 5.8)	0.2552	3.1	3.8	1.6 (–1.7 to 4.8)	0.3423

PRN = pro re nata (as needed).

*n = 65 for sham and n = 96 for intravitreal aflibercept injection monthly from baseline to week 24.

†n = 67 for sham and n = 97 for intravitreal aflibercept injection monthly from baseline to week 24 plus PRN treatment from week 24 to 52 (except for the total score, which was n = 98).

baseline to week 24 (7.4% and 5.8%, respectively) and from weeks 24 to 52 (8.8% and 6.2%, respectively). None of the nonocular SAEs were reported for more than 1 patient from baseline to week 24. During weeks 24 to 52, nonocular SAEs reported for more than 1 patient were pneumonia (1 patient in each treatment group) and syncope (2 patients in the sham group and 1 patient in the aflibercept group). No adverse event was adjudicated as an Anti-Platelet Trialists' Collaboration-defined arterial thromboembolic event during the course of study. There were no deaths during the 52 weeks of this study.

Discussion

The findings of the current study demonstrate that the improvements in BCVA and CRT achieved with monthly intravitreal aflibercept injections in the first 24 weeks of treatment largely were maintained during the PRN (as-needed) phase of study, with monthly monitoring and a mean of 2.5 injections from weeks 24 to 52. Of note, there was also a marked improvement in BCVA with aflibercept in a subgroup of patients with nonperfused retinas at

baseline, in contrast to a particularly poor response in the sham group. The visual improvements with aflibercept enhanced vision-related quality of life, particularly in near visual activities. In this study, aflibercept generally was well tolerated, and the most common adverse events were those typically associated with intravitreal injections or the underlying disease. The increase in macular edema seen in aflibercept patients during the PRN dosing phase suggests that some patients would have benefited from more regular dosing, rather than being treated in response to the recurrence of disease.

The sister study of GALILEO, the COPERNICUS study, demonstrated comparable improvements in BCVA and CRT with intravitreal aflibercept injections.^{5,7} However, the sham groups in the 2 studies were not comparable during weeks 24 to 52 because, in the COPERNICUS study, sham patients received aflibercept PRN starting from week 24, whereas in the GALILEO study, sham patients continued to receive sham treatments through week 52. In the COPERNICUS study, patients receiving sham plus IVT-AFL PRN

Table 3. Distribution of Pro Re Nata Injections during Weeks 24 through 52 and Best-Corrected Visual Acuity Gains at Week 52 in Patients Treated with Intravitreal Aflibercept Injection Every 4 Weeks from Baseline to Week 24 and Pro Re Nata from Weeks 24 to 52

No. of Pro Re Nata Injections	Intravitreal Aflibercept Patients, n (%; n = 91*)	Change (Standard Deviation) from Baseline in Best-Corrected Visual Acuity at Week 52, † No. of Letters
0	13 (14.3)	19.8 (11.4)‡
1	12 (13.2)	
2	18 (19.8)	21.1 (12.8)§
3	21 (23.1)	
4	17 (18.7)	13.1 (13.5)
5	3 (3.3)	
6	7 (7.7)	

BCVA = best-corrected visual acuity; SD = standard deviation.

*Patients completing week 52.

†Because of the small number of patients in each injection category, BCVA gains at week 52 were shown for patients who received 0 to 1, 2 to 3, and 4 to 6 injections. The mean BCVA ± SD at baseline was 58.2±15.5 letters, 49.4±15.9 letters, and 55.4±15.0 letters for patients who received 0 to 1, 2 to 3, and 4 to 6 injections, respectively.

‡For both 0 and 1 injection categories.

§For both 2 and 3 injection categories.

||For 4 to 6 injection categories.

Table 4. Ocular Treatment-Emergent Serious Adverse Events in the Study Eye Occurring from Baseline to Week 24 and Weeks 24 to 52

Serious Adverse Event	Baseline to Week 24*		Week 24 to Week 52 [†]	
	Sham (n = 68)	Intravitreal Aflibercept Injection Monthly from Baseline to Week 24 (n = 104)	Sham (n = 57)	Intravitreal Aflibercept Injection Monthly from Baseline to Week 24 and Pro Re Nata Treatment from Weeks 24 to 52 (n = 97)
Total no. of patients with ≥1 SAE, n (%)	5 (7.4)	2 (1.9)	2 (3.5)	8 (8.2)
Glaucoma	1 (1.5)	0 (0)	1 (1.8)	0 (0)
Iris neovascularization	0 (0)	1 (1.0)	0 (0)	0 (0)
Macular edema	2 (2.9)	0 (0)	0 (0)	4 (4.1)
Reduced visual acuity	1 (1.5)	0 (0)	0 (0)	1 (1.0)
Vitreous detachment	0 (0)	1 (1.0)	0 (0)	0 (0)
Vitreous hemorrhage	1 (1.5)	0 (0)	1 (1.8)	1 (1.0)
Macular fibrosis	0 (0)	0 (0)	0 (0)	1 (1.0)
Macular ischemia	0 (0)	0 (0)	0 (0)	1 (1.0)
Retinal detachment	0 (0)	0 (0)	0 (0)	1 (1.0)
Retinal vein occlusion	0 (0)	0 (0)	0 (0)	1 (1.0)

SAE = treatment-emergent serious adverse event.

*Safety analysis set.

[†]Week 24 completers within safety analysis set.

did not achieve visual and anatomic improvements as robustly as those receiving aflibercept from the inclusion at week 52, suggesting that patients with macular edema secondary to CRVO may benefit from initiating treatment early with aflibercept.⁷

Treatment of CRVO with monthly intravitreal injections for 6 months followed by monthly monitoring and PRN injections for an additional 6 months has been studied for the Ranibizumab for the Treatment of Macular Edema after Central Retinal Vein Occlusion Study: Evaluation of Efficacy and Safety (CRUISE) trial.⁸ Visual and anatomic outcomes reported from the CRUISE study are comparable with those from the COPERNICUS and GALILEO studies, with gains achieved during the fixed monthly dosing phase largely maintained under PRN dosing with monthly monitoring.^{8,12} However, it is suggestive that the steeper decline in visual acuity between months 6 and 7 with 0.5 mg ranibizumab in the CRUISE study, compared with the smaller decline seen with aflibercept during the same time period in the GALILEO study, is reflective of a longer duration of effect with aflibercept.

The GALILEO results at week 52 corroborate the robust effect on visual and anatomic measures seen at week 24 in patients with macular edema secondary to CRVO, after 24 weeks of fixed monthly dosing with aflibercept. Originally, the PRN dosing regimen was introduced to investigate the feasibility of extending the treatment interval after the initial monthly aflibercept dosing phase. It has been demonstrated that the PRN dosing regimen largely maintained the improvements seen at week 24 with monthly monitoring. During the PRN dosing phase, an average of 2.5 injections was given in the IVT-AFL 2Q4 + PRN group, which approximates the 3 injections that would have been administered using a bimonthly dosing regimen, as has been established for aflibercept in wet AMD patients.¹¹ From a practical perspective, the advantage of PRN dosing therefore is questionable: Although PRN dosing may lead to fewer injections than a fixed monthly regimen, it comes

with the requirement of monthly visits. Therefore, a good alternative option would be flexibly adjusting the treatment interval using a treat-and-extend algorithm. This may help to preserve visual and anatomic gains better over PRN dosing as well as to reduce the challenges and cost of monthly monitoring.

Acknowledgments. The authors thank Hadi Moini, PhD, Regeneron Pharmaceuticals, Inc, for editorial assistance.

References

- McIntosh RL, Rogers SL, Lim L, et al. Natural history of central retinal vein occlusion: an evidence-based systematic review. *Ophthalmology* 2010;117:1113–23.
- Wong TY, Scott IU. Clinical practice. Retinal-vein occlusion. *N Engl J Med* 2010;363:2135–44.
- Pournaras CJ, Rungger-Brandle E, Riva CE, et al. Regulation of retinal blood flow in health and disease. *Prog Retin Eye Res* 2008;27:284–330.
- Noma H, Funatsu H, Mimura T, et al. Role of soluble vascular endothelial growth factor receptor-2 in macular oedema with central retinal vein occlusion. *Br J Ophthalmol* 2011;95:788–92.
- Boyer D, Heier J, Brown DM, et al. Vascular endothelial growth factor Trap-Eye for macular edema secondary to central retinal vein occlusion: six-month results of the phase 3 COPERNICUS study. *Ophthalmology* 2012;119:1024–32.
- Brown DM, Campochiaro PA, Singh RP, et al; CRUISE Investigators. Ranibizumab for macular edema following central retinal vein occlusion: six-month primary end point results of a phase III study. *Ophthalmology* 2010;117:1124–33.
- Brown DM, Heier JS, Clark WL, et al. Intravitreal aflibercept injection for macular edema secondary to central retinal vein occlusion: 1-year results from the phase 3 COPERNICUS study. *Am J Ophthalmol* 2013;155:429–37.
- Campochiaro PA, Brown DM, Awh CC, et al. Sustained benefits from ranibizumab for macular edema following central retinal vein occlusion: twelve-month outcomes of a phase III study. *Ophthalmology* 2011;118:2041–9.

9. Holz FG, Roider J, Ogura Y, et al. VEGF Trap-Eye for macular oedema secondary to central retinal vein occlusion: 6-month results of the phase III GALILEO study. *Br J Ophthalmol* 2013;97:278–84.
10. Papadopoulos N, Martin J, Ruan Q, et al. Binding and neutralization of vascular endothelial growth factor (VEGF) and related ligands by VEGF Trap, ranibizumab and bevacizumab. *Angiogenesis* 2012;15:171–85.
11. Heier JS, Brown DM, Chong V, et al; VIEW 1 and VIEW 2 Study Groups. Intravitreal aflibercept (VEGF Trap-Eye) in wet age-related macular degeneration. *Ophthalmology* 2012;119:2537–48.
12. Heier JS, Campochiaro PA, Yau L, et al. Ranibizumab for macular edema due to retinal vein occlusions: long-term follow-up in the HORIZON trial. *Ophthalmology* 2012;119:802–9.

Footnotes and Financial Disclosures

Originally received: May 17, 2013.

Final revision: July 20, 2013.

Accepted: August 8, 2013.

Available online: September 30, 2013. Manuscript no. 2013-793.

¹ Service d'ophtalmologie, Hôpital Pellegrin, Centre Hospitalier Universitaire de Bordeaux, Bordeaux, France.

² Université Bordeaux Segalen, Bordeaux, France.

³ INSERM, L'Institut de Santé Publique, d'Épidémiologie et de Développement (ISPED), Centre INSERM U897-Epidémiologie-Biostatistique, Bordeaux, France.

⁴ Department of Ophthalmology, University of Bonn, Bonn, Germany.

⁵ Department of Ophthalmology, University of Kiel, Kiel, Germany.

⁶ Department of Ophthalmology and Visual Science, Nagoya City University Graduate School of Medical Science, Nagoya, Japan.

⁷ Department of Ophthalmology, Medical University of Vienna, Vienna, Austria.

⁸ Department of Ophthalmology, University Medical Center, Johannes Gutenberg-Universität Mainz, Mainz, Germany.

⁹ Department of Ophthalmology, Juntendo University Urayasu Hospital, Chiba, Japan.

¹⁰ Regeneron Pharmaceuticals, Inc, Tarrytown, New York.

¹¹ Bayer HealthCare AG, Berlin, Germany.

¹² Department of Neurology, University of Erlangen-Nürnberg, Germany.

¹³ Klinik und Poliklinik für Augenheilkunde, Universitätsklinikum Hamburg-Eppendorf, Hamburg, Germany.

¹⁴ Department of Neurology, Heinrich-Heine-Universität, Düsseldorf, Germany.

*The investigators from the GALILEO study are listed in Appendix 1, available at <http://aaojournal.org>.

Presented at: American Academy of Ophthalmology Annual Meeting, November 2012, Chicago, Illinois.

The GALILEO study was funded by Regeneron Pharmaceuticals, Inc, Tarrytown, New York, and Bayer HealthCare, Berlin, Germany. The sponsors participated in the design and conduct of the study, analysis of the data, and preparation of the manuscript.

Financial Disclosure(s): The author(s) have made the following disclosure(s):

Jean-François Korobelnik: Consultant—Alcon, Allergan, Bayer HealthCare, Carl Zeiss Meditec, Novartis, Thea

Frank G. Holz: Consultant—Acucela, Alcon, Allergan, Bayer HealthCare, Genentech, Heidelberg Engineering, Novartis, Pfizer; Financial support—Alcon, Allergan, Bayer HealthCare, Carl Zeiss Meditec, GlaxoSmithKline, Heidelberg Engineering, Novartis, Optos; Lecturer—Alcon, Bayer HealthCare, Heidelberg Engineering, Novartis, Pfizer

Johann Roider: Consultant and Financial support—Bayer HealthCare

Yuichiro Ogura: Consultant—Alcon, Bayer HealthCare, Santen, Wakamoto; Financial support - Bayer HealthCare

Christian Simader: the author's institution, the Medical University of Vienna, has received payments from Bayer HealthCare for data monitoring/reviewing, statistical analysis, and travel

Ursula Schmidt-Erfurth: Consultant—Bayer HealthCare, Alcon, Allergan, Boehringer, Novartis; Financial support—Bayer HealthCare; Advisory board—Alcon, Allergan, Boehringer, Novartis; Lecturer—Alcon, Allergan, Boehringer, Novartis

Katrin Lorenz: Consultant—Bayer HealthCare; Advisory board—GeneSignal SAS, Sensimed AG; Financial support—FP7 European Union, Bayer HealthCare, Ivantis, Inc; Lecturer—Bayer HealthCare, MSD Pharmaceuticals

Robert Vitti: Employee—Regeneron Pharmaceuticals, Inc.

Alyson J. Berliner: Employee—Regeneron Pharmaceuticals, Inc.

Florian Hiemeyer: Employee—Bayer HealthCare

Brigitte Stemper: Employee—Bayer HealthCare

Oliver Zeitz: Employee—Bayer HealthCare

Rupert Sandbrink: Employee—Bayer HealthCare

Correspondence:

Jean-François Korobelnik, MD, Service d'Ophtalmologie, Hôpital Pellegrin Place Amélie Raba Léon, 33000 Bordeaux, France. E-mail: jean-francois.korobelnik@chu-bordeaux.fr.

Targeted Therapy for Metastatic Colorectal Cancer: Role of Afibercept

Edith P. Mitchell

Abstract

Worldwide, colorectal cancer (CRC) is the third most commonly diagnosed cancer in male individuals and the second most commonly diagnosed cancer in female individuals. Survival outcomes are less than optimal for patients with metastatic disease, with a 5-year survival in the 5% to 8% range. The development of new chemotherapeutic agents and effective combination regimens for metastatic colorectal cancer (mCRC) has increased median overall survival (OS) to the 24- to 28-month range. Because of the recognition that vascular endothelial growth factors (VEGFs) and their receptors are primary regulators of physiologic and pathologic angiogenesis and lymphangiogenesis, leading to neovascularization and tumor growth, the targeting of the angiogenic pathway has become a focus of key therapeutic strategies in mCRC. Therapeutic regimens that include bevacizumab, an inhibitor of VEGF-A, in combination with cytotoxic chemotherapy, have resulted in improved response rate (RR) and survival in mCRC. However, the effects of VEGF-A inhibition are often temporary, with resistance and disease progression developing in most patients. Proposed models include intrinsic and adaptive resistance, mediated by factors other than VEGF-A. Afibercept (known as ziv-aflibercept in the United States; Zaltrap®, Regeneron Pharmaceuticals; sanofi-aventis), a novel recombinant fusion protein, is an angiogenic factor trap that blocks the binding of VEGF-A, VEGF-B, and placental growth factor. Phase I/II clinical trials have demonstrated effective activity in mCRC, with acceptable safety and tolerability. A recent phase III randomized double-blind trial in patients previously treated with oxaliplatin reported significant improvement in OS, progression-free survival (PFS), and RR with afibercept compared with placebo when administered in combination with irinotecan and fluorouracil. Adverse events were consistent with anti-VEGF therapy. Thus afibercept represents a potential new treatment option for patients with mCRC.

Clinical Colorectal Cancer, Vol. 12, No. 2, 73-85 © 2013 Elsevier Inc. All rights reserved.

Keywords: Antiangiogenesis, Metastatic colorectal cancer (mCRC), Placental growth factor, VEGF-A, VEGF-B, afibercept

Introduction

Colorectal cancer (CRC) is the third most commonly diagnosed cancer in male individuals and the second most commonly diagnosed cancer in female individuals, with more than 1.2 million new cases worldwide; in 2008 it was the cause of 608,700 deaths.¹ Although CRC incidence and death rates have shown decreases in the United States, it is anticipated that 143,460 new cases of CRC will be diagnosed and approximately 51,690 Americans will die of the disease, accounting for approximately 9% of cancer deaths.² The lifetime incidence of CRC in patients at average risk is approximately 5%, with 90% of cases occurring after age 50 years.³ In the United States, CRC incidence declined roughly 2% to 3% per year between 1992 and 2008.⁴ Information from the Surveillance Epidemiology and

End Results (SEER) database suggests that incidence rates are increasing in the 40- to 44-year age group.⁵ Furthermore, CRC incidence in the United States is 25% higher in men than in women and is almost 20% higher in African Americans than in whites.² For most patients with metastatic disease, treatment remains palliative, and survival outcomes are less than optimal. Despite advances in management, metastatic disease is associated with poor 5-year survival, with a rate of approximately 10%.⁶ It is estimated that 20% of patients with CRC have metastatic disease at the time of diagnosis, whereas many others will experience metastases during the course of the disease.⁷ SEER data suggest that 64% of all patients, including all stages of disease, who are treated for CRC survive 5 years.³ A recent clinical trial achieved an overall survival (OS) rate of 72.9% at 6 years in patients with stage III disease who were treated with oxaliplatin-based postoperative adjuvant chemotherapy.⁸

In the early 1970s, Folkman et al reported the importance of novel growth and remodeling of blood vessels in the growth and proliferation of tumors.⁹ This work led to the development of biological

Submitted: Apr 30, 2012; Revised: Aug 1, 2012; Accepted: Aug 6, 2012; Epub: Oct 24, 2012

Thomas Jefferson University, 233 South 10th Street, BLSB 502, Philadelphia, PA.
E-mail contact: edith.mitchell@jefferson.edu

agents that target angiogenesis. The elucidation of the role of vascular endothelial growth factors (VEGFs) in tumorigenesis contributed to important advances for the treatment of patients with a variety of solid tumors, including metastatic CRC (mCRC). During the past 2 decades, the addition of bevacizumab combined with cytotoxic chemotherapy has resulted in unprecedented advances in the treatment of mCRC, with improved response, progression-free survival (PFS), and OS.^{10,11} This review will discuss the role of angiogenesis in mCRC, the principles of resistance to antiangiogenic therapy, and the development of aflibercept (known as ziv-aflibercept in the United States; Zaltrap®, Regeneron Pharmaceuticals; sanofi-aventis) as a novel antiangiogenic agent for mCRC.

Angiogenesis

Angiogenesis is a pivotal process for growth, invasion, and metastasis in many solid tumors.^{9,12} Although physiologic angiogenesis is a highly regulated process,^{13,14} pathologic angiogenesis manifests as an abnormal increase in proliferating endothelial cells and as structural and functional abnormalities of tumor vasculature.^{15,16} These abnormalities include loss of normal vascular hierarchy, development of an irregular and leaky endothelial layer, compression of vessels, abnormal blood flow, loss of functional lymphatic vessels, increased interstitial pressure, and development of hypoxia and acidosis in the tumor microenvironment. Other abnormalities involve the pericytes and basement membrane of the tumor vasculature, and endothelial cells in tumor vessels show altered gene expression.^{16,17}

Angiogenesis is controlled by a complex signaling network that involves multiple interacting proangiogenic and antiangiogenic signals, including VEGF, angiopoietins, Notch, and integrins.^{18,19} The variety of pathways involved in angiogenesis offers numerous possible therapeutic targets. The VEGF family has been widely implicated as a key regulator of tumor angiogenesis^{20,21} and is composed of 5 growth factors: VEGF-A, VEGF-B, VEGF-C, VEGF-D, and placental growth factor (PlGF).^{15,21,22} These growth factors differentially bind and activate 3 cell surface tyrosine kinase receptors, VEGFR-1, VEGFR-2, and VEGFR-3, the activities of which can be enhanced by coreceptors neuropilin receptor (NRP)-1 and NRP-2.²³⁻²⁵

Mechanism of Angiogenesis

VEGF-A was the first member of the VEGF family to be identified and is recognized as the most potent inducer and positive regulator of the normal and pathologic angiogenic cascade.^{26,27} It regulates blood vessel proliferation and vascular permeability, and its expression is associated with poor prognosis in a variety of human cancers.²⁸ The biological effects of VEGF-A include endothelial cell proliferation, survival, migration, invasion, chemotaxis of bone marrow progenitors, vascular permeability, and vasodilation, which are mediated by its binding and activation of receptor tyrosine kinases VEGFR-1 and VEGFR-2.¹⁵ Although VEGF-A binds VEGFR-1 with approximately 10 times higher affinity than VEGFR-2, the higher kinase activity of VEGFR-2 makes it the most important effector of VEGF-A signaling.^{19,29,26,29-32} Although VEGF-A is the best-characterized member of the VEGF family, experimental and clinical evidence indicates that other VEGFs, such as VEGF-B and PlGF, play an important role in tumor biological processes and pathologic angiogenesis.^{22,33}

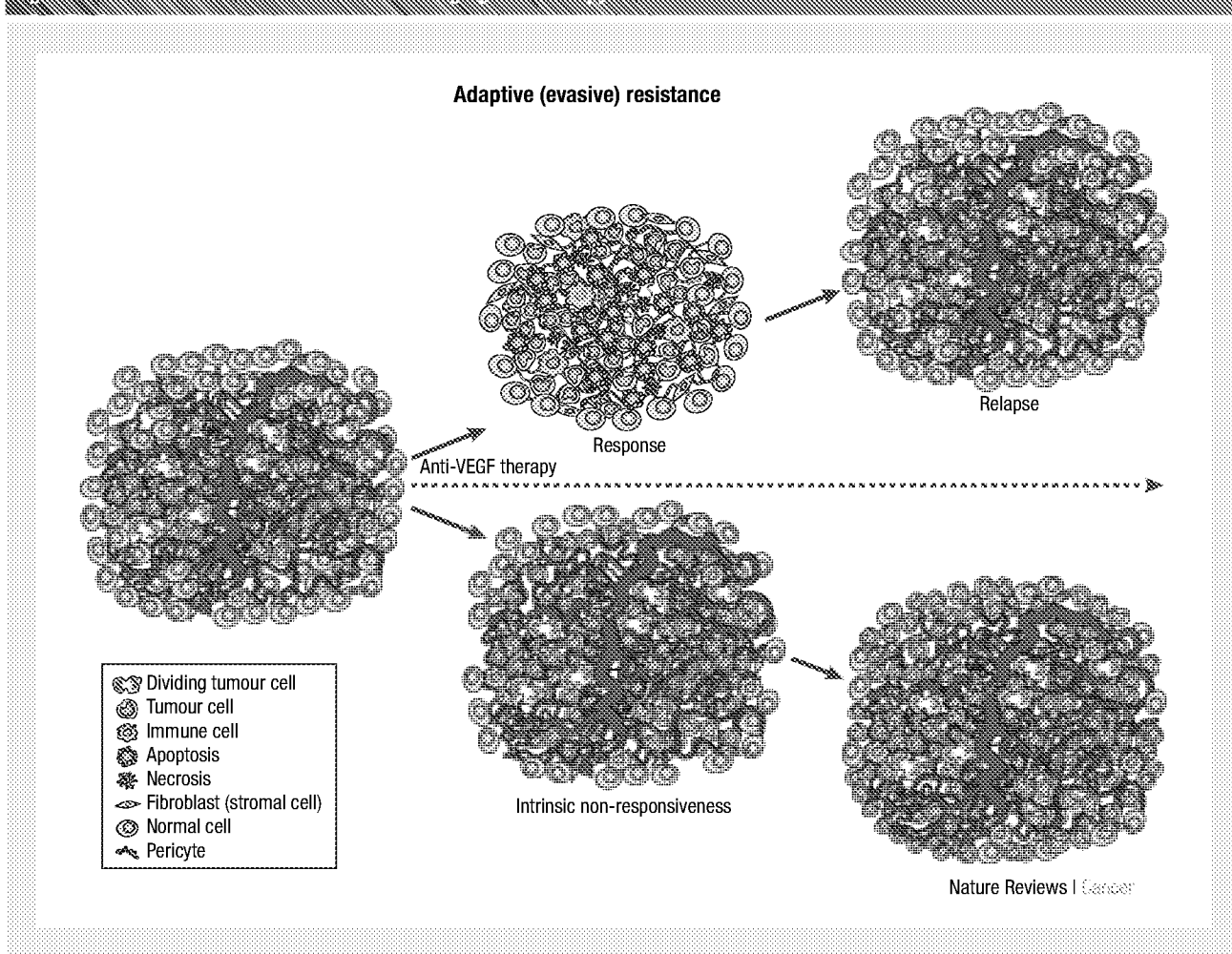
VEGF-B shares close structural homology with VEGF-A; although these 2 factors are coexpressed in many tissues, VEGF-B is more broadly expressed in skeletal muscle and the pancreas.³⁴ VEGF-B binds VEGFR-1 and NRP1 and although still under investigation, it could play a role in tumorigenesis and angiogenesis.³⁵ This is postulated because both VEGF-B and VEGFR-1 are upregulated in a number of different tumor types—in some cases, correlating with poor prognosis, metastasis, and relapse.²² VEGF-B has been shown to have pleiotropic effects on vascular cell adhesion,^{33,36} and although it seems to be dispensable for the growth of blood vessels, VEGF-B may play a role in the survival of preexisting blood vessels under pathologic conditions.³⁶

PlGF exists as at least 4 different isoforms originated by alternative splicing: PlGF-1, PlGF-2, PlGF-3, and PlGF-4.³⁷ Similar to VEGF-B, PlGF is also homologous to VEGF-A and binds VEGFR-1.^{22,38} Levels of both PlGF-1 and PlGF-2 have been shown to be elevated in human colorectal tumors.³⁹ There is also evidence that PlGF potentiates the response to VEGF-A by signaling through VEGFR-1,⁴⁰ and this signaling stimulates the recruitment of bone marrow-derived macrophages to the tumor site, where they release angiogenic factors.⁴¹

Because of the central role of the VEGF family in angiogenesis, and the increased VEGF expression in many tumor types, this family of growth factors has become an important therapeutic target. Anti-VEGF therapies currently available for the treatment of mCRC include the monoclonal antibody bevacizumab,³² which binds VEGF-A, as well as the monoclonal antibodies cetuximab³³ and panitumumab,⁴⁴ which indirectly inhibit angiogenesis by targeting the EGF receptor. Nonetheless, there has been a lack of patient response to anti-VEGF therapies, and the clinical benefits observed are short-lived. Tumor regrowth and disease progression often occur after an initial response, indicating that cancer cells are able to functionally evade therapeutic inhibition of angiogenesis.⁴⁵⁻⁴⁷

Mechanisms of Resistance To Antiangiogenic Therapy

Despite initial success, resistance to antiangiogenic therapy eventually develops, thus limiting survival benefits. Because multiple pathways contribute to angiogenesis, it is possible that 1 or more may contribute to the development of resistance. Possible specific mechanisms of resistance include upregulation of VEGF receptors, the fibroblast growth factor signaling pathway, interleukin-12, hepatocyte growth factor, increased pericyte coverage of tumor blood vessels to support vasculature, and recruitment of alternative angiogenic factors and pathways such as VEGF-C or VEGF-D.^{48,49} Two broad mechanisms, adaptive and intrinsic resistance, have been proposed to explain the lack of effectiveness of anti-VEGF therapy (Figure 1). Adaptive resistance develops after an initial positive antiproliferative response, whereas intrinsic resistance exists in tumors before treatment.⁵⁰ The key mechanisms include (1) revascularization after upregulation of alternative proangiogenic pathways, (2) protection of the tumor vasculature either by the recruitment of vascular progenitor cells and proangiogenic monocytes from the bone marrow to the peritumoral area or by increasing protective pericyte coverage, (3) enhanced capability of tumor cells to migrate and invade local tissue to coopt normal vasculature, and (4) increased metastatic spread and tumor cell growth in lymph nodes and distant organs. Intrinsic re-

Figure 1. Mechanisms of Resistance to Antiangiogenic Therapy⁵⁴

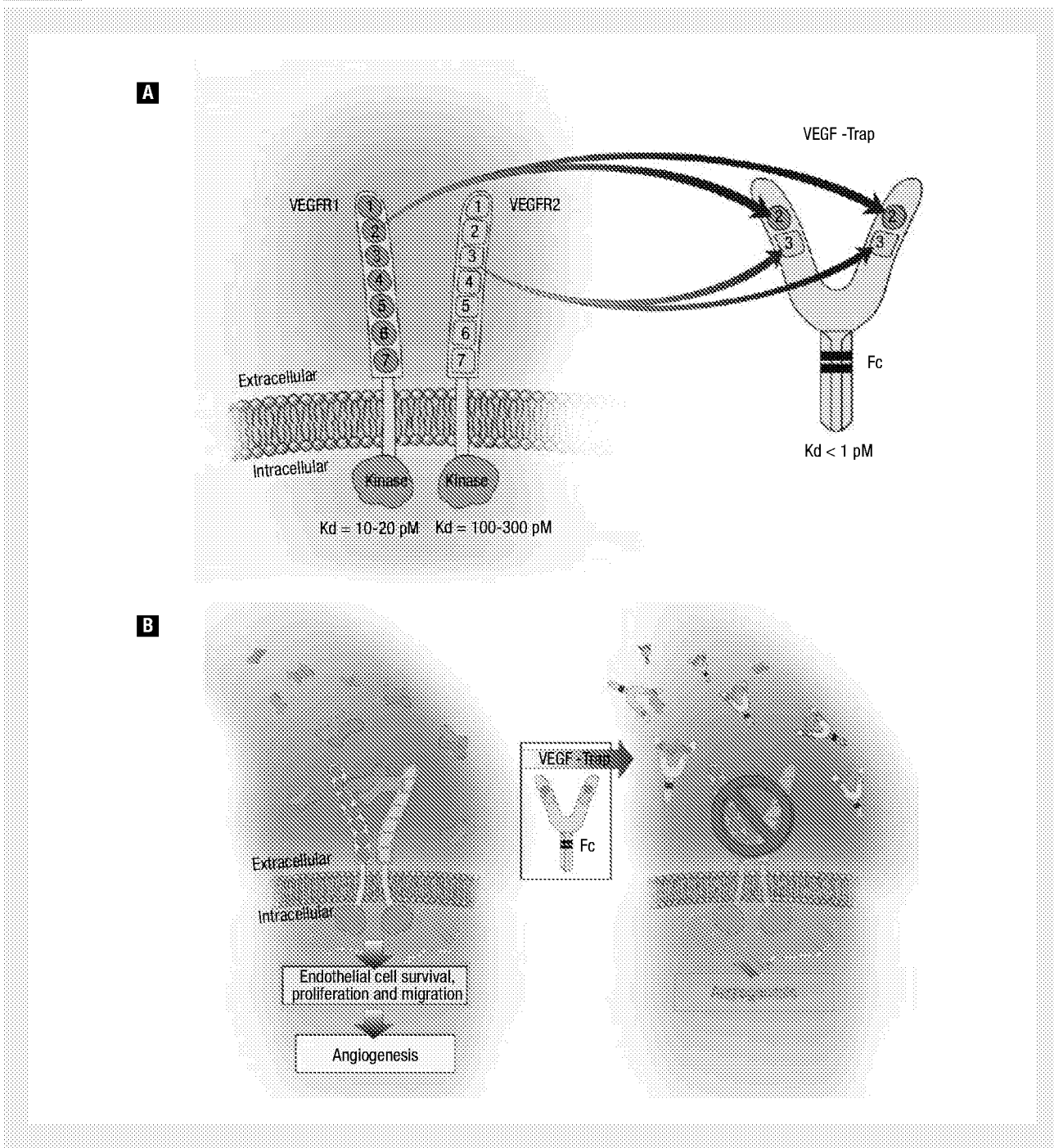
Reprinted by permission from Macmillan Publishers Ltd: [NAT REV CANCER] Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. 2008;8(8):592-603, Copyright (2008).

sistance is thought to involve many of these same mechanisms. In addition to rapid adaptation, intrinsic resistance may result from (1) multiplicity of proangiogenic signals, (2) vascular protection mediated by inflammatory cell or hypovascular tumor microenvironment characterized by indifference toward angiogenesis inhibitors, or (3) invasive cooption of normal vessels in the absence of neoangiogenesis (Figure 1).⁵⁰ It has been suggested that future cancer therapeutic strategies will integrate inhibitors of angiogenesis with drugs targeting resistance mechanisms to provide more enduring efficacy.^{49,50} Two recent studies in mCRC have shown changes in circulating angiogenic biomarkers after the inhibition of VEGF-A with bevacizumab. The addition of bevacizumab to 5-fluorouracil (5-FU), leucovorin, irinotecan, and oxaliplatin (FOLFOXIRI) resulted in a prolonged and significant reduction in levels of VEGF-A through the time of disease progression. However, the soluble VEGFR-2 and PlGF levels increased from baseline during treatment, potentially contributing to disease progression.⁵¹ This is consistent with observations from other studies that demonstrate that PlGF promotes tumor angiogenesis and tumor growth and may also contribute to tumor “escape” by providing sufficient angiogenic signals when VEGF-A is blocked.^{22,52,53} In a second study, patients with mCRC

(n = 43) with no previous chemotherapy for metastatic disease were treated with 5-FU, leucovorin, and irinotecan (FOLFIRI) in combination with bevacizumab.⁵⁴ There was no association between baseline levels of VEGF or VEGFR-2 and differences in PFS or OS.⁵⁴ However, compared with baseline levels, initial treatment resulted in an increase in PlGF ($P = .01$), soluble VEGFR-2 ($P = .03$), and eotaxin ($P = .01$), whereas hepatocyte growth factor ($P = .046$), basic fibroblast growth factor ($P = .047$), PlGF ($P < .001$), stromal derived factor-1 ($P = .038$), and macrophage chemoattractant protein-3 ($P < .001$) were elevated before disease progression.⁵⁴ At the time of progression, the PlGF level declined from its peak but remained greater than baseline ($P < .01$), whereas the VEGFR-2 ($P < .001$) and soluble VEGFR-2 ($P = .005$) levels declined to less than baseline.⁵⁴ It was suggested that increases in the levels of uninhibited angiogenic factors, such as PlGF, are compensatory mechanisms to stimulate new vessel growth in preparation for disease progression.⁵⁴

The development of resistance by some tumor types to long-term antiangiogenic therapy has contributed to the search and development of treatment options intended to address the proposed mechanisms of resistance to antiangiogenic therapy. A review of many of these strategies,

Figure 2. Structure and Mechanism of Action of Aflibercept.⁵⁵ (A) Aflibercept consists of a Domain 2 of VEGFR-1 and Domain 3 of VEGFR-2 fused to the Fc portion of IgG1. (B) Aflibercept binds and blocks its substrate and prevents it from activating the native receptors.



Used with permission from: *Inflammation and Allergy Drug Targets*, Vol. 10, Stewart MW. Aflibercept (VEGF-TRAP): the next anti-VEGF drug, pages 497-508, Copyright Elsevier (2011).

such as regorafenib, cediranib, AMG 386, and RO5323441 (TB 403) can be found in a recent review by Tejpar et al.⁴⁸

Aflibercept

Aflibercept, a soluble recombinant fusion protein, is a multiple angiogenic factor trap rationally designed to block the angiogenesis

network by not only binding VEGF-A but also uniquely targeting VEGF-B and PlGF.⁵⁵⁻⁵⁷ Aflibercept was developed by fusing sections of the second immunoglobulin (Ig) domain of VEGFR-1 and the third Ig domain of VEGFR-2 to the F_c portion of human IgG1 (Figure 2).^{55,58} Aflibercept has now been approved by the US Food and Drug Administration, with the US name of ziv-aflibercept, for

Table 1. Pharmacokinetics in Free Aflibercept⁶³

Parameter	Dose Level (mg/kg)						
	0.3	1.0	2.0	3.0	4.0	5.0	7.0
C_{max} ($\mu\text{g/mL}$)	4.00 \pm 9%	17.9 \pm 31%	34.5 \pm 11%	46.7 \pm 30%	97.4 \pm 43%	86.8 \pm 34%	159 \pm 21%
C_{last} ($\mu\text{g/mL}$)	0.147 \pm 38%	0.659 \pm 116%	2.36 \pm 71%	4.06 \pm 63%	11.0 \pm 51%	9.63 \pm 28%	14.4 \pm 55%
AUC (day- $\mu\text{g/mL}$)	9.34 \pm 15%	50.9 \pm 54%	125 \pm 35%	226 \pm 34%	293 \pm 15%	428 \pm 64%	605 \pm 46%
$t_{1/2}$ (day)	1.70 \pm 21%	2.58 \pm 50%	3.76 \pm 42%	6.18 \pm 38%	5.51 \pm 18%	7.43 \pm 38%	5.14 \pm 37%
V_{ss} (L)	4.51 \pm 29%	5.88 \pm 22%	5.58 \pm 21%	7.74 \pm 33%	7.88 \pm 38%	9.89 \pm 31%	6.12 \pm 29%
Cl (L/d)	1.95 \pm 42%	1.87 \pm 51%	1.13 \pm 31%	1.14 \pm 48%	1.10 \pm 38%	1.27 \pm 65%	0.915 \pm 39%

Abbreviations: AUC = area under the curve extrapolated to infinity; Cl = total body clearance; C_{last} = last measurable (non-zero) plasma concentration; C_{max} = maximum observed plasma concentration; $t_{1/2}$ = apparent terminal half-life; V_{ss} = volume of distribution at steady state.

Values are \pm coefficient of variation (%).

Reprinted with permission. ©2009 American Society of Clinical Oncology. All rights reserved. Lockhart, AC et al. *J Clin Oncol* 2010; 28:207-14.

the use in combination with FOLFIRI in the treatment of mCRC that is resistant to or has progressed following an oxaliplatin-containing regimen.⁵⁹

Pharmacokinetics and Pharmacodynamics

In animals, aflibercept has been found to have negligible binding to extracellular tissues.^{55,60} Aflibercept forms a stable and inert 1:1 complex with VEGF, which appears in the circulation at maximal levels within 24 to 48 hours of treatment.⁶¹ Clearance occurs via F_c receptor- or pinocytosis-mediated pathways that result in proteolysis.⁶¹ With a K_d of 0.49 pM, aflibercept binds VEGF-A with affinities that are 19- and 181-fold higher than the native VEGF receptors fused to F_c (VEGFR-1 and VEGFR-2, respectively).⁶² Aflibercept also exhibits strong affinity for PlGF-2, with a K_d of approximately 45 pM.⁵⁵

The pharmacokinetics of subcutaneous aflibercept were characterized in a phase I trial by Tew et al. Patients ($n = 38$) received aflibercept subcutaneously at doses of 25, 50, 100, 200, 400, or 800 $\mu\text{g/kg}$ weekly or 800 $\mu\text{g/kg}$ twice weekly for 10 weeks, with continuation until disease progression.⁵⁶ Dose-dependent increases of free aflibercept were observed peaking within 3 days after the dose. Clearance of free aflibercept was relatively rapid, with an elimination half-life ($t_{1/2}$) up to 3 days compared with approximately 18 days for bound aflibercept. Formation of the VEGF/aflibercept complex appeared to saturate at approximately 800 $\mu\text{g/kg}$ weekly, suggesting that much of the VEGF produced by the patient is being captured at this dose.⁵⁶

When administered intravenously in doses ranging from 0.3 to 7.0 mg/kg every 2 weeks ($n = 47$), the mean maximum plasma concentration of free aflibercept increased with doses ranging from 4 to 159 $\mu\text{g/mL}$, whereas the exposure (area under the curve [AUC]) increased more than the dose proportionally in the 0.3- to 2.0-mg/kg dose range (Table 1).⁶³ The apparent $t_{1/2}$ also increased with doses ranging from 1.7 to 5.1 days. Clearance decreased at low doses but was stable at 2.0 to 7.0 mg/kg, and the concentration of free aflibercept remained greater than bound aflibercept throughout the dosing intervals at doses of ≥ 2.0 mg/kg, indicating binding saturation of endogenous VEGF. Bound aflibercept concentrations increased after cycle 1 and up to 3 weeks after the first dose, which indicates that steady state was not achieved during this time.⁶³

Aflibercept Activity In Animal and Human Models

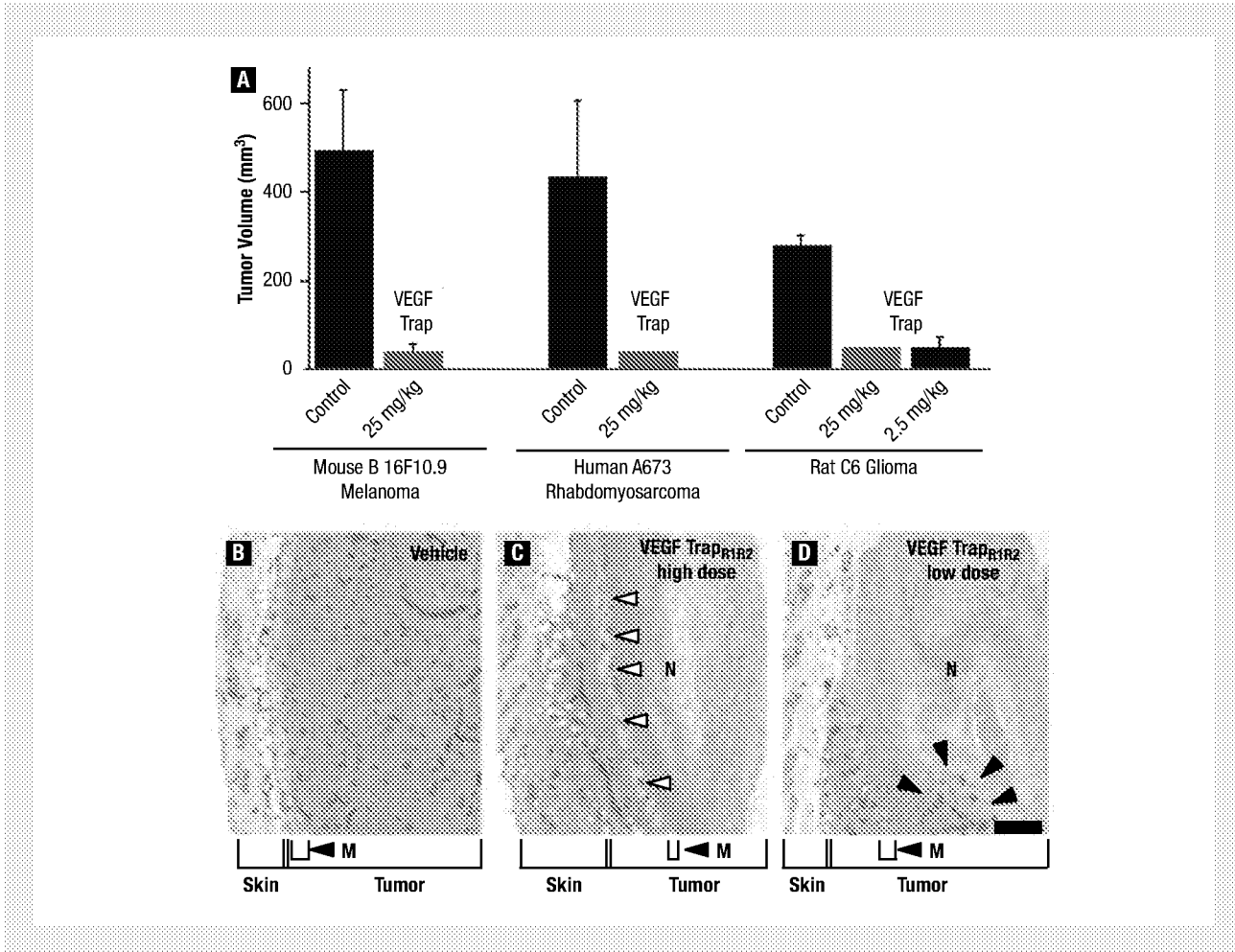
Subcutaneous single-agent aflibercept was effective at inhibiting tumor growth in mice models of mouse B16F10.9 melanoma, human A673 rhabdomyosarcoma, and rat C6 glioma, and almost completely blocked tumor-associated angiogenesis at the highest dose of 25 mg/kg (Figure 3A).⁵⁵ Aflibercept 2.5 mg/kg, comparable to the dose that inhibited tumor growth, was less effective at completely blocking tumor-associated angiogenesis, with small pockets of tumor-associated vessels observed (Figure 3B).⁵⁵ Similar dose-dependent effects have been observed in a xenograft model of neuroblastoma, whereas high doses of aflibercept led to greater regression of coopted vascular structures, which occurs during the initial phase of tumor growth.⁶⁴ Single-agent aflibercept also diminished tumor vasculature and volume in lung tumors in mice.⁶⁵ Furthermore, pre-existing lung micrometastases markedly decreased in both size and cell number compared with controls, with evidence of apoptosis after 1 dose of aflibercept.⁶⁵

The effect of aflibercept on activated signaling pathways in endothelial cells has been investigated in SCID mice bearing K1735 tumors or COLO 205 human colon cancer tumors.⁶⁶ Aflibercept significantly decreased tumor endothelial p-ERK, p-STAT3, and p-AKT expression ($P \leq .05$), with signs of antiangiogenesis.⁶⁶

Aflibercept has been investigated alone and in combination with cytotoxic chemotherapy in several 3-arm investigations in mice. In a mammary adenocarcinoma study, aflibercept 40 mg/kg/dose twice weekly was as active as the highest non-toxic dose of 5-FU (90 mg/kg/dose), with the combination showing synergistic activity at all doses tested. Single-agent aflibercept and irinotecan were equally active in mice with advanced-stage human colon cancer, with the combination demonstrating synergy (Figure 4).⁶⁷ In BALB/c nude female mice with early-stage B16 melanoma, as well as gemcitabine in SCID mice with advanced human colon cancer, single-agent activity with aflibercept was comparable to docetaxel in additional studies. In both studies, the combination was more active than either agent alone.⁶⁸ Furthermore, these 4 studies presented little to no overlap in host toxicity with combination therapy.

The combination of aflibercept and docetaxel has shown activity in HT1080 tumors with vasculature that exhibits resistance to VEGF blockade. Although single-agent therapy exhibited only modest ef-

Figure 3 Effect of Fully-Neutralizing VEGF Trap on Tumor Cell Growth and Vascularity.⁶⁹ Aflibercept dramatically inhibits the subcutaneous growth and vascularity of implanted tumors from diverse tissues and species. (A) Aflibercept substantially blocked the growth of the indicated subcutaneously implanted tumors at the indicated doses twice weekly for 2 weeks (OS and B16F10.9) or 1.6 weeks (A673). Error bars represent standard error of mean, n = five mice/treatment group. The differences between control tumor volumes and aflibercept treated tumor volumes were analyzed by using Student's *t* tests and were found to be significant at the following levels: B16F10, *P* = .01; A673, *P* = .06; OS, *P* < .001. (B-D) Histologic analysis reveals that aflibercept not effectively block blood vessel growth in these implanted tumors. Sections of C6 tumors stained with antibodies to platelet endothelial cell adhesion molecule reveal that vehicle treated animals had large tumors that were highly vascularized (B), whereas animals treated with aflibercept 2.5 mg/kg (C) had tumors that were largely avascular with large areas of necrosis (N). Viable tumors appeared to be vascularized because of cooption of preexisting host vessels (white arrowheads) associated with hypodermal musculature (M) and dermis. Treatment with aflibercept 2.5 mg/kg greatly shrank tumor growth (C) and resulted in large necrotic regions (N). Although small patches of vessels were occasionally apparent (black arrows) (Bar = 100 μ m).



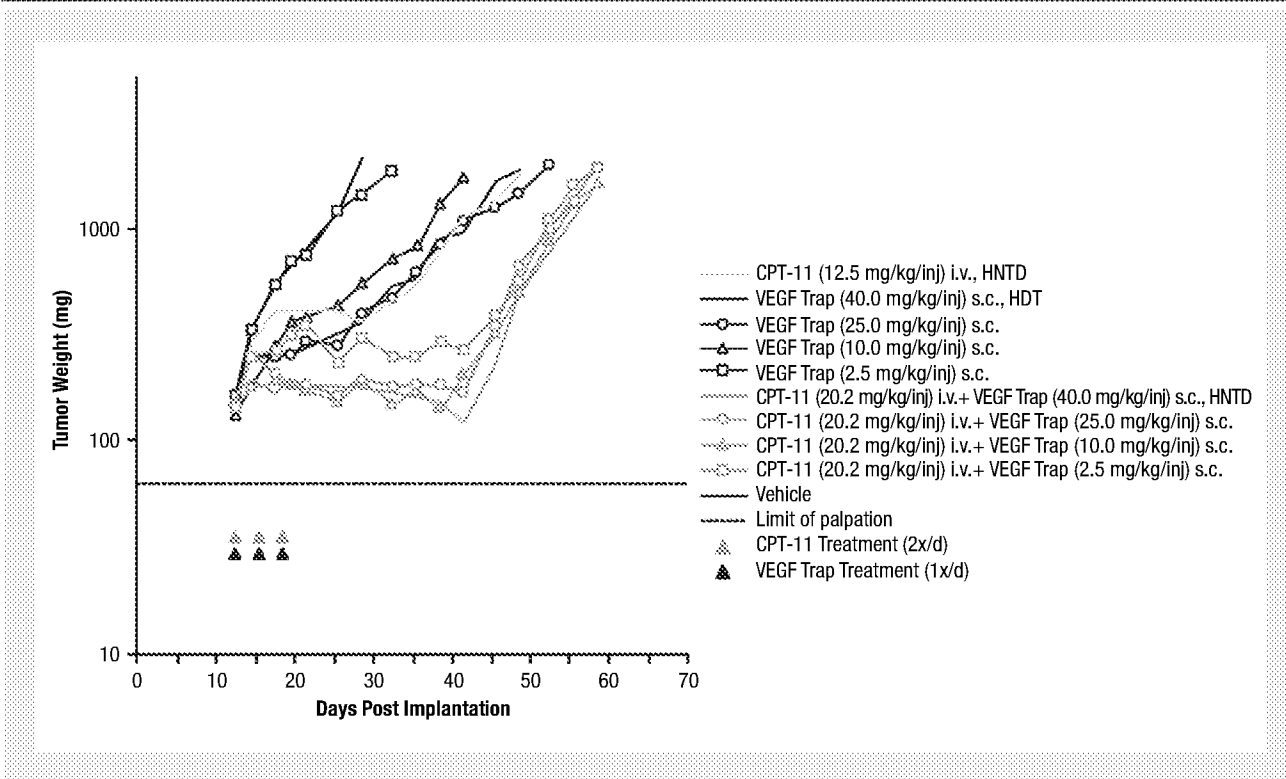
Reprinted from *Proceedings of the National Academy of the Sciences of the United States of America*, Vol. 99, Holash J et al., VEGF-Trap: A VEGF blocker with potent antitumor effects, 11303-11308, Copyright (2002) National Academy of Sciences, USA.

ffects on tumor vessels, the combination of aflibercept and docetaxel resulted in pruned vessels with less branching, some vessels with punctuate endothelial cell staining, and an increase in terminal dUTP nick end labeling-positive endothelial cells.⁶⁹

Compared with bevacizumab and doxorubicin, enhanced antitumor effects with aflibercept have been shown in vivo in human VEGF-expressing acute myeloid leukemia models, in which aflibercept treatment induced increased tumor ablation and areas of central necrosis surrounded by rims of proliferating leukemia cells. This

suggests both direct and indirect effects with aflibercept in combination with doxorubicin, not simply the decreased tumor blood flow that was also observed.⁷⁰ Aflibercept treatment followed by doxorubicin showed progressive anthracycline accumulation in extramedullary acute myeloid leukemia sites and marrow, resulting in up to 2-fold higher doxorubicin concentrations after 24 hours. In contrast, mice not pretreated with aflibercept generally showed progressive drug clearance over time. Although the aflibercept mechanisms leading to variable doxorubicin levels in different

Figure 4 Synergy Between Aflibercept (VEGF Trap) and Irinotecan in Colon Cancer Xenografts (colon HCT-116 in HCT nude mice)



Reprinted by permission from sanofi-aventis U.S. LLC, Inc.; Chiron M et al. Synergistic activity of aflibercept (VEGF Trap) in combination with 5-fluorouracil or irinotecan in preclinical tumor models. AACR Meeting Abstracts. October 22-26, 2007: A13.

tissues are uncertain, there is the suggestion that inefficient drug delivery by leukemia-associated vasculature, which may mediate chemoresistance, may be positively influenced by aflibercept.⁷⁰

In summary, preclinical studies in various mouse and human models have shown antiangiogenic activity with aflibercept alone or in combination with cytotoxic chemotherapy, causing vascular remodeling and tumor growth inhibition and regression. Furthermore, pretreatment with aflibercept increases tumor cell kill compared with single-agent cytotoxic chemotherapy by increasing cytotoxic drug exposure and possibly by other mechanisms.

Clinical Trials

The efficacy and safety of aflibercept alone or in combination with various chemotherapy regimens has been explored in several phase I^{56,63,71-76} and phase II⁷⁷⁻⁸⁷ trials in patients with advanced solid tumors or non-Hodgkin lymphoma. Solid tumors included breast, colorectal, endometrial, gastric, glioblastoma, lung, melanoma, ovarian, pancreas, sarcoma, thyroid, urothelial, and others. Aflibercept has also been investigated in a phase III trial involving patients with mCRC.

Phase I Trials

Aflibercept has been administered subcutaneously or intravenously in phase I clinical trials.^{56,63,71-76} Three phase I clinical trials have investigated the safety, pharmacokinetics, and pharmacodynamics of aflibercept as a single agent every 2 weeks in patients with

advanced solid tumors (Table 2). Lockhart et al enrolled 47 patients with refractory solid tumors at a starting dose of 0.3 mg/kg, which was 10-fold less than the dose from primate studies that produced no observed effect.⁶³ Patients who tolerated the first 2 doses of aflibercept were eligible to receive treatment at 7 dose levels on the long-term tolerability study. Three patients had an objective partial response (PR). The following dose-limiting toxicities occurred: grade 3 elevation in alanine aminotransferase levels at 1.0 mg/kg, grade 3 dyspnea and arthralgia at 2.0 mg/kg, and grade 3 hypertension at 4.0 mg/kg. The study was subsequently modified to allow for cohort expansion, provide more intensive blood pressure control before defining a dose-limiting toxicity, and allow for higher levels of proteinuria. The subsequent dose-limiting toxicities occurred at the 7.0-mg/kg dose level with 1 episode each of proteinuria and rectal ulceration. The most common antiangiogenic adverse events (AEs) were dysphonia, hypertension, and proteinuria. Other common, generally grade 1, AEs were fatigue, nausea, and vomiting. Maximal VEGF/aflibercept complex levels were reached at doses of 2 mg/kg and greater.

A second trial involved 38 patients with advanced solid tumors who had received a median of 5 previous chemotherapy regimens.⁵⁶ The initial cohort of 3 patients received subcutaneously administered aflibercept at 25 μ g/kg to investigate the pharmacokinetics of aflibercept. Five additional patients were enrolled at either the maximum administered dose or the maximum tolerated dose (dose level at less

Aflibercept in Metastatic Colorectal Cancer

Table 2. Results of Phase I, II, and III Studies of Aflibercept in Combination With Standard-of-Care Therapies in Metastatic Colorectal Cancer

Patient Population	Treatment	Results	Major Adverse Events (All Grades)	Major Findings	Reference
Phase I					
Refractory solid tumors (n = 47) including CRC (n = 7)	Aflibercept (0.3-7 mg/kg)	PR 6%; SD (> 1 y) 4% (CRC group not specified) DLT: rectal ulceration and proteinuria (7 mg/kg) Maximum VEGF blockade at doses \geq 2 mg/kg	Fatigue 64%, dysphonia 47%, HTN 38%, nausea 36%, constipation 32%, headache 32%, vomiting 28%, arthralgia 26%	Aflibercept was safely administered at doses that demonstrated antitumor activity	Lockhart et al ⁶³
Refractory solid tumors (n = 38) including CRC (n = 6)	Aflibercept (0.025-0.8 mg/kg)	SD (\geq 10 wk) 47% (CRC group not specified) Maximum VEGF blockade at 0.8 mg/kg	Proteinuria 37%, fatigue 32%, injection-site reaction 18%, nausea 17%, musculoskeletal discomfort 16%, anorexia 16%, HTN 13%, hoarseness 11%	Aflibercept was well tolerated and had manageable side effects	Tew et al ⁶⁴
Heavily pretreated advanced solid tumors (n = 54) including CRC (n = 9)	Aflibercept (2-9 mg/kg) + docetaxel	DLT at cycle 1 included neutropenic infection (2 mg/kg), grade 2 HTN and grade 3 dysphonia (7 mg/kg), grade 2 HTN (9 mg/kg) PR 9%, SD 69% (CRC group not specified)	Neutropenia 98%, fatigue 89%, hemorrhage 81%, anemia 81%, stomatitis 72%, dysphonia 64%, HTN 53%, musculoskeletal disorders 53%, alopecia 51%	Aflibercept 6 mg/kg with docetaxel 75 mg/m ² is recommended for further investigation	Isambert et al ⁶⁵
Pretreated mCRC (n = 16)	Aflibercept (2 or 4 mg/kg) + irinotecan + 5-FU + leucovorin (fixed doses)	Response rate 8%, PFS 7.6 mo No DLT	Grade 3/4: neutropenia (75%), hypertension (25%)	Recommended dose of aflibercept is 4 mg/kg in combination with FOLFIRI	Yamazaki et al ⁶²
Heavily pretreated advanced solid tumors (n = 16) including CRC (n = 2)	Aflibercept (4-6 mg/kg) + docetaxel + cisplatin	DLT without G-CSF prophylaxis: 17% (4 mg/kg) and 14% (5 mg/kg) DLT with G-CSF: 0% CR 7%, PR 7%, SD 50% (CRC group not specified)	Grade 2: epistaxis 19%, proteinuria 17%, dysphonia 13% Grade 3/4: HTN 13%	Recommended dose of aflibercept is 6 mg/kg in combination with standard docetaxel + cisplatin	Freyer et al ⁷⁴
Heavily pretreated advanced solid tumors (n = 32) including CRC (n = 1)	Aflibercept (2-5 mg/kg) + oxaliplatin + leucovorin + 5-FU (FOLFOX)	No DLT during dose escalation Free/bound aflibercept ratio (free:bound concentration): 0.74-4.5 PR 16%, SD 31% in CRC; PR 100%	Fatigue 81%, nausea 72%, skin/subcutaneous tissue disorder 56%, anemia 53%, diarrhea 53%, vomiting 53%, abdominal pain 50%, headache 50%, HTN 50%, musculoskeletal/connective tissue disorder 50%	Aflibercept 4 mg/kg with FOLFOX is recommended for further investigation	Limentani et al ⁷¹
Advanced solid tumors (n = 38) including CRC (n = 23)	Aflibercept (2-6 mg/kg) + irinotecan + 5-FU + leucovorin (LV5-FU2)	PR 18%, SD 50% in CRC: PR 17%, SD 61% (> 12 mo 26%) DLTs were grade 3 proteinuria (4 mg/kg), grade 3 stomatitis and esophageal reflux (5 mg/kg), and febrile neutropenia and grade 3 stomatitis (6 mg/kg)	Diarrhea 92%, fatigue 92%, nausea 84%, stomatitis 82%, anemia 82%, neutropenia 76%, HTN 74%, dysphonia 74%, infections 71%, musculoskeletal disorder 68%, hemorrhage 68%, constipation 66%, vomiting 61%, anorexia 61%, headache 61%, dyspnea 55%, alopecia 50%, abdominal pain 42%, thrombocytopenia 42%	Aflibercept 4 mg/kg with LV5-FU2 is recommended for further investigation	Rixe et al ⁷²
Advanced solid tumors (n = 27) including CRC (n = 19)	Aflibercept (4 mg/kg) + irinotecan + 5-FU + leucovorin (I-LV5-FU2)	PR 4%, SD 78% in CRC: PR 5%	Grade 2: HTN 26%, dysphonia 11%, epistaxis 4%, proteinuria 4% Grade 3/4: HTN 15%	Aflibercept 4 mg/kg with I-LV5-FU2 is recommended for further investigation	Verslype et al ⁶⁹
Phase II					
Previously treated mCRC (n = 51)	Aflibercept (4 mg/kg)	Bevacizumab-naïve: RR 29%, 4 mo PFS 29% Previous bevacizumab: RR 30%, 4 mo PFS 26% (CRC group not specified)	Fatigue 78%, HTN 56%, proteinuria 49%, headache 43%, voice alteration 31%, anorexia 24%, joint pain 18%	Single agent aflibercept is active and well tolerated in previously treated mCRC	Tang et al ⁶⁸
Phase III					
mCRC previously treated with oxaliplatin-based therapy (n = 1226)	Aflibercept (4 mg/kg) + irinotecan + 5-FU (FOLFIRI) vs. placebo + irinotecan + 5-FU	Median OS: 13.5 vs. 12.06 mo (P = .0032) Median PFS: 6.9 vs. 4.67 mo (P = .0007) ORR: 19.8% vs. 11.1% (P = .0001)	Proteinuria (62% vs. 41%), HTN (41% vs. 11%), hemorrhage (38% vs. 19%), dysphonia (25% vs. 3%), headache (22% vs. 9%)	Addition of aflibercept to FOLFIRI resulted in significant improvement in OS and PFS and was unaffected by previous treatment with bevacizumab	Van Cutsem ⁷³

Abbreviations: CR = complete response; DLT = dose-limiting toxicity; 5-FU = 5-fluorouracil; FOLFOX = oxaliplatin + leucovorin + 5-FU; G-CSF = granulocyte colony-stimulating factor; HTN = hypertension; mCRC = metastatic colorectal cancer; ORR = overall response rate; OS = overall survival; PFS = progression-free survival; PR = partial response; RR = response rate; SD = stable disease; VEGF = vascular endothelial growth factor.

than which 2 of 6 patients exhibited a dose-limiting toxicity), which ever was lower. A duration of \geq 10 weeks with stable disease was achieved in 18 patients (47%). The maximum administered dose was

800 μ g/kg twice weekly, whereas the maximum tolerated dose was not reached. Dose-limiting toxicity was observed in 4 patients and resolved in 3 patients and included confusion and decreased oral

Table 3 Adverse Events Reported in a 2-Stage Phase I Trial of Afibercept in Patients with Metastatic Colorectal Cancer⁷³

Adverse Event	Most Frequent Adverse Events— Afibercept (n = 51)	
	All Grades (%)	Grade 3/4 (%)
Fatigue	78.4	5.9
Decreased appetite	23.5	—
Joint pain	17.6	—
Anti-VEGF associated events		
Proteinuria	49.0	7.8
Hypertension	54.9	7.8
Dysphonia	31.4	—
Headache	43.1	5.9

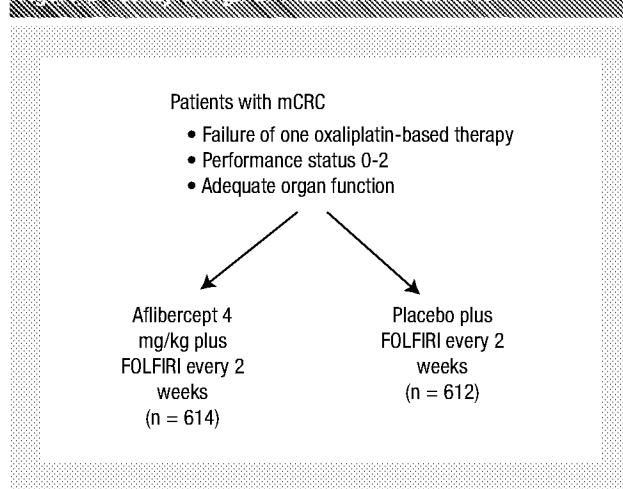
Abbreviation: VEGF = vascular endothelial growth factor.

intake, grade 3 proteinuria, and grade 3 leukopenia. Pulmonary embolism was diagnosed in the fourth patient after initiation of afibercept, but was subsequently identified on baseline computed tomographic scan. Grade 3/4 AEs included hypertension, proteinuria, nausea, leukopenia, pulmonary embolism, and cerebral ischemia. Levels of the VEGF/afibercept complex seen after the first dose did not increase appreciably between the 800 $\mu\text{g}/\text{kg}$ once weekly and twice weekly dose levels.⁵⁶

The aims of a third study were to document the dose-limiting toxicities occurring during the first treatment cycle and to establish the recommended dosage in patients with advanced solid tumors.⁷³ Patients with metastatic or unresectable disease or those for whom no standard conventional therapy existed received afibercept 2 mg/kg with docetaxel 75 mg/m² on day 1 every 3 weeks until disease progression or unacceptable toxicity. The dose of afibercept was escalated to 4, 5, 6, 7, and 9 mg/kg if no dose-limiting toxicity was observed in a cohort of 3 patients. If dose-limiting toxicity was observed in 1 patient, the cohort was expanded to 6 patients. The recommended dose was defined as the highest dose at which 2 of 3 to 6 patients experienced dose-limiting toxicity. The safety and activity of afibercept were studied further in an expanded group after the recommended dose had been quantified. Seven patients experienced PR, and 18 patients experienced stable disease for more than 3 months. Dose-limiting toxicity during cycle 1 related to afibercept was observed at the 7 and 9 mg/kg dose levels; consequently, 6 mg/kg was determined to be the recommended dose for afibercept. All patients experienced 1 or more AEs. Antiangiogenic AEs included epistaxis, proteinuria, dysphonia, and hypertension.

The efficacy, safety, dose-limiting toxicities, recommended dose, and pharmacokinetics of afibercept in combination with fixed doses of FOLFIRI as second-line therapy have been investigated in 16 patients with mCRC.⁷⁶ Afibercept was administered at dosages of 2 or 4 mg/kg every 2 weeks, with 3 to 6 patients to be recruited at each dose. An additional 10 patients were to be treated at the recommended dose, defined as the highest dose of afibercept at which < 33% of all evaluable patients experienced dose-limiting toxicity dur-

Figure 5 Study Design of Phase II Study⁷⁷



ing the first 2 cycles. A total of 16 patients (3 receiving doses of 2 mg/kg and 13 receiving doses of 4 mg/kg) received a total of 131 cycles with no dose-limiting toxicity observed. The most common grade 3/4 AEs was neutropenia, including 1 case of febrile neutropenia and hypertension.

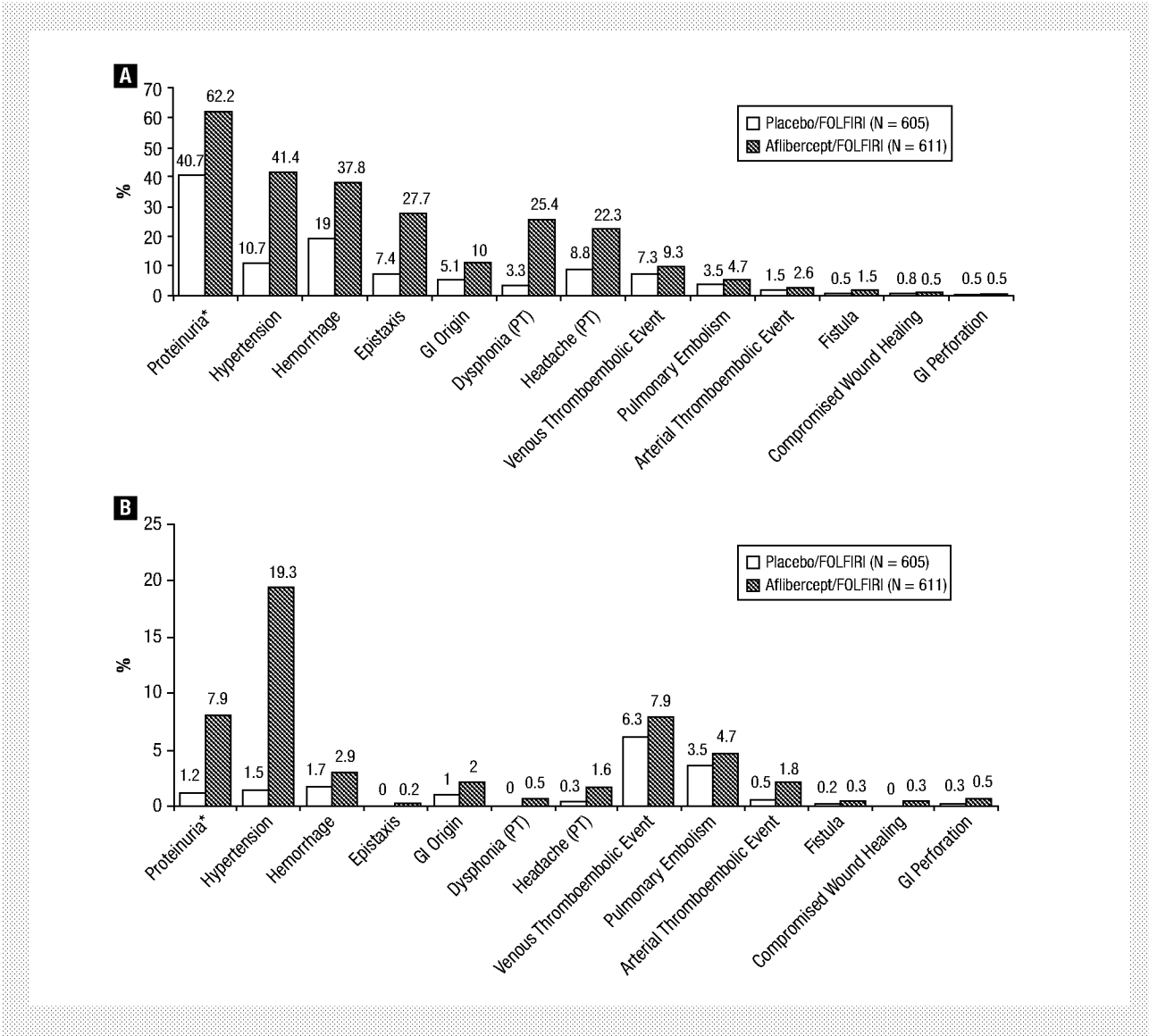
As single-agent therapy or in combination with chemotherapy, these trials suggested that afibercept 4 mg/kg every 2 weeks or 6 mg/kg every 3 weeks were optimal doses based on antitumor efficacy and acceptable safety. In addition, the fraction of free afibercept was found to be greater than bound afibercept at these doses, with no apparent impact from concomitant chemotherapy on afibercept pharmacokinetics. As expected, hypertension and proteinuria were the most common afibercept-related AEs.

Phase II Trials

Phase II trials of afibercept have investigated the efficacy and safety of single-agent afibercept 4 mg/kg every 2 weeks given intravenously. Patients with gynecologic cancer,^{77,80} lung cancer,^{78,87} malignant ascites,^{79,81} glioblastoma,⁸² mCRC,⁸³ metastatic gynecologic soft tissue sarcomas,⁸⁸ melanoma,⁸⁵ or urothelial cancer⁸⁶ were generally heavily pretreated. Modest to moderate improvement in PFS was generally observed. For example, in patients with lung cancer (n = 98), the median PFS was 2.7 months and OS was 6.2 months, with 29% surviving at 12 months.⁸⁷ In patients with metastatic or locally advanced urothelial cancer previously treated with 1 platinum-containing regimen (n = 22), PFS was 2.79 months.⁸⁶ Disease progression was the most frequent reason for study discontinuation in these phase II trials. The most common afibercept-related grade 3/4 AEs were those expected with antiangiogenic therapy, eg, hypertension, proteinuria, and fatigue, as well as headache and abdominal pain/perforation.

In heavily treated patients with mCRC, including those previously treated with bevacizumab, results from an open-label multicenter 2-stage phase II trial of patients with good performance status (PS) (Eastern Cooperative Oncology Group PS \leq 2) (n = 51) demonstrated that afibercept was well tolerated with modest activity (Table 3).⁸³ Twenty-seven patients had received previous bevacizumab therapy. After an average of 5.6 cycles of afibercept 4 mg/kg intra-

Figure 6. Adverse Events Reported in Phase III VELOUR Trial of Aflibercept in Patients With Metastatic Colorectal Cancer (mCRC). (A) Percentages of All Grade Adverse Events. (B) Percentages of Grade 3/4 Adverse Events.



Abbreviations: GI = gastrointestinal, PT = preferred term.

venously every 2 weeks, the median PFS was 3.4 months in the group previously treated with bevacizumab and 2.0 months in bevacizumab-naïve patients. The most common aflibercept-related AEs were fatigue, hypertension, proteinuria, headache, voice alteration, anorexia, and joint pain (Table 2). Grade 3/4 treatment-related AEs occurring in more than 1 patient were hypertension (8%), proteinuria (8%), fatigue (6%), and headache (6%).

Aflibercept 6 mg/kg has also been investigated in combination with docetaxel 75 mg/m² given every 3 weeks in patients with recurrent epithelial ovarian, primary peritoneal, or fallopian tube carcinoma (n = 46).⁸⁰ After a median of 6 treatment cycles and a median follow-up of 24 months, an objective response (according to Response Evaluation Criteria in Solid Tumors) was achieved in 25/46 (54%) patients, 11 (24%) of whom achieved a complete response.

The median PFS and OS were 6.4 and 26.6 months, respectively. AEs associated with aflibercept were grade 1/2 hypertension in 5 patients (11%) and grade 2 hypotension in 1 patient (2%).

Phase III Trial

Aflibercept in combination with FOLFIRI has also been investigated in a phase III trial (VELOUR; <http://ClinicalTrials.gov> ID NCT00561470) in patients with previously treated mCRC (n = 1226) who had adequate organ function and a PS of 0 to 2 and in whom a previous oxaliplatin-based regimen had failed.⁵⁷ Patients were a median of 61 years, 58.6% were men, 97.9% had a PS of 0 to 1, 56.4% had metastases in more than 1 organ, and 30.4% had received previous treatment with bevacizumab. Aflibercept 4 mg/kg on day 1 every 2 weeks (n = 614) or placebo (n = 612) was added to

Table 4. Results in Second-Line Treatment of Metastatic Colorectal Cancer With Afibercept vs Bevacizumab

Study	Patient Population	Treatment	Results	Grade \geq 3 AEs
VELOUR (Van Cutsem et al ⁵⁷)	mCRC previously treated with oxaliplatin-based therapy (n = 1226)	Afibercept (4 mg/kg) + irinotecan + 5-FU (FOLFIRI) vs. placebo + irinotecan + 5-FU	Median OS: 13.5 vs. 12.06 mo (P = .0032) Median PFS: 6.9 vs. 4.67 mo (P = .0007) ORR: 19.8% vs. 11.1% (P = .0001)	Diarrhea (19.3% vs. 7.8%), asthenia (16.8% vs. 10.6%), stomatitis (13.8% vs. 5.0%), infections/infections (12.3% vs. 6.9%), HTN (19.3% vs. 1.5%), GI/abdominal pains (5.4% vs. 3.3%), headache (1.6% vs. 0.3%), hand-foot syndrome (2.8% vs. 0.5%), thromboembolic event (9.6% vs. 6.7%), neutropenia (36.7% vs. 29.5%), thrombocytopenia (3.4% vs. 1.6%), proteinuria (7.8% vs. 1.2%)
E3200 (Giantonio et al ⁵³)	mCRC previously treated with fluoropyrimidine and irinotecan (n = 829)	FOLFOX4 + bevacizumab 10 mg/kg vs. FOLFOX4 vs. bevacizumab 10 mg/kg	Median OS: 12.9 vs. 10.8 vs. 10.2 mo (P = .0011) PFS: 7.3 vs. 4.7 vs. 2.7 mo (P < .0001) ORR: 22.7% vs. 8.6% vs. 3.3% (P < .0001) (P values are for FOLFOX4 + bevacizumab vs. FOLFOX4)	HTN (6.2% vs. 1.8% vs. 7.3%), bleeding (3.4% vs. 0.4% vs. 2.1%), vomiting (10.1% vs. 3.2% vs. 4.7%), proteinuria (0.7% vs. 0% vs. 0%), neuropathy (16.3% vs. 9.2% vs. 0.8%), thromboembolism (3.4% vs. 2.5% vs. 0.4%), cardiac/cerebrovascular ischemia (0.9% vs. 0.4% vs. 0.4%)
TML (Arnold et al ⁵¹)	mCRC after first progression with oxaliplatin- or irinotecan-based chemotherapy plus bevacizumab (n = 820)	Irinotecan- or oxaliplatin-based chemotherapy + bevacizumab 2.5 mg/kg vs. irinotecan- or oxaliplatin-based chemotherapy	Median OS: 11.2 vs. 9.8 mo (P = .0062) Median PFS: 5.7 vs. 4.1 mo (P < .0001) ORR: 5.4% vs. 3.9% (P = .3113)	Neutropenia (16% vs. 13%), diarrhea (10% vs. 8%), leukopenia (4% vs. 3%), vomiting (4% vs. 3%), abdominal pain (4% vs. 3%), asthenia (6% vs. 4%), stomatitis (3% vs. 1%), HTN (92% vs. 1%), venous thromboembolism (5% vs. 3%)

Abbreviations: AEs = adverse events; 5-FU = 5-fluorouracil; FOLFOX4 = oxaliplatin, 5-FU, and leucovorin; GI = gastrointestinal; HTN = hypertension; mCRC = metastatic colorectal cancer; ORR = overall response rate; OS = overall survival; PFS = progression-free survival.

FOLFIRI (Figure 5). At data cutoff, median follow-up was 22.28 months, and 863 patients had died. In the afibercept arm, patients experienced significant improvement in median OS, PFS, and overall response rate (ORR) vs. placebo.⁵⁷ Prespecified subgroup analyses demonstrated that the overall results were irrespective of previous exposure to bevacizumab, age, sex, race, previous hypertension, number of metastatic sites, or location of primary tumor.⁸⁹ However, a greater treatment effect on OS with afibercept was observed in patients with liver metastases only (hazard ratio, 0.649; 95.34% CI, 0.492-0.855; $P = .0899$).⁸⁹ Discontinuation because of AEs was 26.6% in the afibercept arm and 12.1% in the placebo arm.⁵⁷ The most common reasons for discontinuation in the afibercept arm were asthenia/fatigue, infections, diarrhea, hypertension, and venous thromboembolic events. Grade 3/4 AEs occurring with at least 2% greater incidence in patients treated with afibercept vs. placebo were diarrhea, asthenia/fatigue, stomatitis/ulceration, infections, hypertension, gastrointestinal/abdominal pains, neutropenia/neutropenic complications, and proteinuria (Figure 6A and B).⁵⁷ In patients in the afibercept arm with and without previous bevacizumab treatment, respectively, the incidence (grade 3/4) of hypertension (16.6% vs. 20.5%), hemorrhage (3.2% vs. 2.8%), and venous and arterial thromboembolic events (8.0% vs. 7.8% and 2.1% vs. 1.7%) were similar.⁸⁹

Possible Role for Afibercept in Therapy

As new therapies for second-line treatment of mCRC are introduced, it is important to develop an understanding of their potential role in treatment. Since bevacizumab is a standard treatment for second-line management of patients with mCRC, comparison of afibercept with bevacizumab may provide some insight regarding the potential role for afibercept. Although there is no prospective

comparison of afibercept with bevacizumab, examination of relevant clinical trials such as the phase III VELOUR trial described earlier for afibercept and the E3200 and TML (ML18147) trials for bevacizumab may be helpful. The E3200 trial randomized patients previously treated with a fluoropyrimidine and irinotecan to the combination of oxaliplatin, 5-FU, and leucovorin (FOLFOX4) with (n = 286) or without (n = 291) bevacizumab or bevacizumab alone (n = 243).⁵⁰ The bevacizumab alone arm was closed early after an interim analysis showed inferior survival compared with the other 2 arms. In addition, 36% of patients experienced 1 or more grade \geq 3 toxicity. The TML study investigated the efficacy and safety of bevacizumab with standard second-line chemotherapy in patients whose disease progressed after bevacizumab plus standard first-line chemotherapy.⁵¹ The choice of oxaliplatin- or irinotecan-based second-line chemotherapy was dependent on first-line therapy.

In the VELOUR study, the addition of afibercept to FOLFIRI resulted in significant improvement in OS and PFS and was unaffected by previous treatment with bevacizumab (Table 4). Selected side effects commonly observed with FOLFIRI (diarrhea, stomatitis, infection, neutropenia) were more common in patients treated with afibercept, as were anti-VEGF AEs—ie, hypertension, mucosal bleeding, and proteinuria. Although infrequent, gastrointestinal perforation, hemorrhage, and arterial thromboembolism were also more common with afibercept. Grade \geq 3 AEs were more common with the addition of afibercept compared with placebo (83.4% vs. 62.5%). In the E3200 study, bevacizumab monotherapy provided little benefit, but the addition of bevacizumab significantly improved OS and PFS with a significant increase in grade \geq 3 hypertension, bleeding, vomiting, and neuropathy compared with FOLFOX4. In the TML study, the combination of bevacizumab with standard chemo-

therapy significantly prolonged OS and PFS with no increase in AEs beyond disease progression.

Conclusion

Aflibercept is a multiple angiogenic factor trap that binds VEGF-A, VEGF-B, and PlGF. Clinical trials demonstrate effective antitumor activity with an acceptable safety and tolerability profile. In VELOUR and a phase II trial of patients with mCRC, no unexpected AEs were reported, and AEs observed were those typically associated with anti-VEGF agents, namely hypertension and proteinuria. The phase III VELOUR trial showed that patients with mCRC receiving aflibercept in combination with FOLFIRI experienced statistically significant improvements in OS, PFS, and ORR when compared with those receiving placebo after failure with an oxaliplatin-containing regimen. Furthermore, results of the phase III VELOUR trial demonstrated a 24% decrease in risk of disease progression, an improvement in response rate from 11% to 19.8% and in survival at 2 years from 19% to 28%. These benefits were consistent regardless of previous bevacizumab therapy. In conclusion, aflibercept represents a potential new option in combination with FOLFIRI in the treatment of mCRC.

Acknowledgments

Medical editorial writing assistance was provided by Samantha Taylor, PhD, and Alfredo Toschi, PhD, of Phase Five Communications Inc., supported by sanofi-aventis, U.S. LLC, in collaboration with Regeneron Pharmaceuticals. The author retained full editorial control over the content of the manuscript and received no compensation from any party for her work. E.P.M. discloses that she acts as a consultant to sanofi-aventis, U.S. LLC and Regeneron Pharmaceuticals; her institution also receives research support from Genentech Inc.

Disclosure

The author retained full editorial control over the content of the manuscript and received no compensation from any party for her work. E.P.M. discloses that she acts as a consultant to sanofi-aventis, U.S. LLC and Regeneron Pharmaceuticals; her institution also receives research support from Genentech Inc.

References

- Jemal A, Bray F, Center MM, et al. Global cancer statistics. *CA Cancer J Clin* 2011; 61:69-90.
- American Cancer Society. Cancer facts and figures 2012. Atlanta: American Cancer Society 2012. Available at: <http://www.cancer.org/Research/CancerFactsFigures/index>. Accessed: January 1, 2012.
- Hovlader N, Noone AM, Krapcho M, et al. SEER cancer statistics review, 1975-2008. Bethesda, MD: National Cancer Institute. Available at: http://seer.cancer.gov/csr/1975_2008/. Accessed: January 1, 2012.
- Siegel RL, Ward EM, Jemal A. Trends in colorectal cancer incidence rates in the United States by tumor location and stage, 1992-2008. *Cancer Epidemiol Biomarkers Prev* 2012; 21:411-6.
- Davis DM, Marcet JE, Frattini JC, et al. Is it time to lower the recommended screening age for colorectal cancer? *J Am Coll Surg* 2011; 213:352-61.
- Cidón EU. The challenge of metastatic colorectal cancer. *Clin Med Insights Oncol* 2010; 4:55-60.
- Song X, Zhao Z, Barber B, et al. Characterizing medical care by disease phase in metastatic colorectal cancer. *J Oncol Pract* 2011; 7:255-305.
- André T, Boni C, Navarro M, et al. Improved overall survival with oxaliplatin, fluorouracil, and leucovorin as adjuvant treatment in stage II or III colon cancer in the MOSAIC trial. *J Clin Oncol* 2009; 27:3109-16.
- Folkman J, Merler E, Abernathy C, et al. Isolation of a tumor factor responsible for angiogenesis. *J Exp Med* 1971; 133:275-88.

- Hurwitz H, Fehrenbacher L, Novotny W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N Engl J Med* 2004; 350:2335-42.
- Fuchs CS, Marshall J, Mitchell E, et al. Randomized, controlled trial of irinotecan plus infusional, bolus, or oral fluoropyrimidines in first-line treatment of metastatic colorectal cancer: results from the BICC-C study. *J Clin Oncol* 2007; 25:4779-86.
- Folkman J. Role of angiogenesis in tumor growth and metastasis. *Semin Oncol* 2002; 29(suppl 16):15-8.
- Folkman J. Fundamental concepts of the angiogenic process. *Curr Mol Med* 2003; 3:643-51.
- Chung AS, Lee J, Ferrara N. Targeting the tumour vasculature: insights from physiological angiogenesis. *Nat Rev Cancer* 2010; 10:505-14.
- Ellis LM, Hicklin DJ. VEGF targeted therapy: mechanisms of anti-tumour therapy. *Nat Rev* 2008; 8:579-91.
- Baluk P, Hashizume H, McDonald DM. Cellular abnormalities of blood vessels as targets in cancer. *Curr Opin Genet Dev* 2005; 15:102-11.
- Fukumura D, Duda DG, Munn LL, et al. Tumor microvasculature and microenvironment: novel insights through intravital imaging in pre-clinical models. *Microcirculation* 2010; 17:206-25.
- Carmeliet P. Angiogenesis in life, disease and medicine. *Nature* 2005; 438:932-6.
- Ferrara N. Vascular endothelial growth factor as a target for anticancer therapy. *Oncologist* 2004; 9 suppl 1:2-10.
- Ferrara N, Kerbel RS. Angiogenesis as a therapeutic target. *Nature* 2005; 438:967-74.
- Grothey A, Galanis E. Targeting angiogenesis: progress with anti-VEGF treatment with large molecules. *Nat Rev Clin Oncol* 2009; 6:507-18.
- Fischer C, Mazzone M, Jonckx B, et al. FLT1 and its ligands VEGFB and PlGF: drug targets for anti-angiogenic therapy? *Nat Rev Cancer* 2008; 8:942-56.
- Veikkola T, Alitalo K. VEGFs, receptors and angiogenesis. *Semin Cancer Biol* 1999; 9:211-20.
- Chu QS. Aflibercept (AVE0005): an alternative strategy for inhibiting tumour angiogenesis by vascular endothelial growth factors. *Expert Opin Biol Ther* 2009; 9:263-71.
- Ellis LM, Hicklin DJ. Pathways mediating resistance to vascular endothelial growth factor-targeted therapy. *Clin Cancer Res* 2008; 14:6371-5.
- Ferrara N. Vascular endothelial growth factor: basic science and clinical progress. *Endocr Rev* 2004; 25:581-611.
- Neufeld G, Cohen T, Gengrinovitch S, et al. Vascular endothelial growth factor (VEGF) and its receptors. *FASEB J* 1999; 13:9-22.
- Shweiki D, Neeman M, Itin A, et al. Induction of vascular endothelial growth factor expression by hypoxia and by glucose deficiency in multicell spheroids: implications for tumor angiogenesis. *Proc Natl Acad Sci U S A* 1995; 92:768-72.
- Shibuya M. Tyrosine kinase receptor Flt/VEGFR family: its characterization related to angiogenesis and cancer. *Genes Cancer* 2010; 1:1119-23.
- Waltenberger J, Claesson-Welsh L, Sieghahn A, et al. Different signal transduction properties of KDR and Flr1, two receptors for vascular endothelial growth factor. *J Biol Chem* 1994; 269:26988-95.
- Terman BI, Dougher-Vermazen M, Carrion ME, et al. Identification of the KDR tyrosine kinase as a receptor for vascular endothelial cell growth factor. *Biochem Biophys Res Commun* 1992; 187:1579-86.
- Shalaby F, Rossant J, Yamaguchi TP, et al. Failure of blood-island formation and vasculogenesis in Flk-1-deficient mice. *Nature* 1995; 376:62-6.
- Li X, Lee C, Tang Z, et al. VEGF-B: a survival, or an angiogenic factor? *Cell Adh Migr* 2009; 3:322-7.
- Olofsson B, Pajusola K, Kaipainen A, et al. Vascular endothelial growth factor B, a novel growth factor for endothelial cells. *Proc Natl Acad Sci U S A* 1996; 93:2576-81.
- Olofsson B, Korpelainen E, Pepper MS, et al. Vascular endothelial growth factor B (VEGF-B) binds to VEGF receptor-1 and regulates plasminogen activator activity in endothelial cells. *Proc Natl Acad Sci U S A* 1998; 95:11709-14.
- Zhang F, Tang Z, Hou X, et al. VEGF-B is dispensable for blood vessel growth but critical for their survival, and VEGF-B targeting inhibits pathological angiogenesis. *Proc Natl Acad Sci U S A* 2009; 106:6152-7.
- Yang W, Ahn H, Hinrichs M, et al. Evidence of a novel isoform of placenta growth factor (PlGF-4) expressed in human trophoblast and endothelial cells. *J Reprod Immunol* 2003; 60:53-60.
- Sawano A, Takahashi T, Yamaguchi S, et al. Flt-1 but not KDR/Flk-1 tyrosine kinase is a receptor for placenta growth factor, which is related to vascular endothelial growth factor. *Cell Growth Differ* 1996; 7:213-21.
- Escudero-Esparza A, Martin TA, Davies ML, et al. PGF isoforms, PLGF-1 and PGF-2, in colorectal cancer and the prognostic significance. *Cancer Genomics Proteomics* 2009; 6:239-46.
- Carmeliet P, Moons L, Luttun A, et al. Synergism between vascular endothelial growth factor and placental growth factor contributes to angiogenesis and plasma extravasation in pathological conditions. *Nat Med* 2001; 7:575-83.
- De Groot JF, Lamborn KR, Chang SM, et al. Phase II study of aflibercept in recurrent malignant glioma: a North American Brain Tumor Consortium study. *J Clin Oncol* 2011; 29:2689-95.
- Avastin [package insert]. South San Francisco, CA: Genentech Inc; 2009.
- Erbitinix [package insert]. Branchburg, NJ: Imclone LLC; 2012.
- Vectibix [package insert]. Thousand Oaks, CA: Amgen Inc; 2011.
- Saitz LB, Lenz HJ, Kindler HL, et al. Randomized phase II trial of cetuximab, bevacizumab, and irinotecan compared with cetuximab and bevacizumab alone in irinotecan-refractory colorectal cancer: the bond-2 study. *J Clin Oncol* 2007; 25:4557-61.

46. Kindler HL, Niedzwiecki D, Hollis D, et al. Gemcitabine plus bevacizumab compared with gemcitabine plus placebo in patients with advanced pancreatic cancer: phase III trial of the cancer and Leukemia Group B (CALGB 80303). *J Clin Oncol* 2010; 28:3617-22.
47. Barrios CH, Liu MC, Lee SC, et al. Phase III randomized trial of sunitinib versus capecitabine in patients with previously treated HER2-negative advanced breast cancer. *Breast Cancer Res Treat* 2010; 121:121-31.
48. Tejpar S, Prehen H, Mazzone M. Overcoming resistance to antiangiogenic therapies. *Oncologist* 2012; 17:1039-50. Epub 2012 Jul 6.
49. Moreno Garcia V, Basu B, Mollifé LR, et al. Combining antiangiogenics to overcome resistance: rationale and clinical experience. *Clin Cancer Res* 2012; 18:3750-61.
50. Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. *Nat Rev Cancer* 2008; 8:592-603.
51. Loupakis F, Cremolini C, Fioravanti A, et al. Pharmacodynamic and pharmacogenetic angiogenesis-related markers of first-line FOLFIRI plus bevacizumab schedule in metastatic colorectal cancer. *Br J Cancer* 2011; 104:1262-9.
52. Yao J, Wu X, Zhuang G, et al. Expression of a functional VEGFR-1 in tumor cells is a major determinant of anti-PlGF antibodies efficacy. *Proc Natl Acad Sci U S A* 2011; 108:11590-5.
53. Fischer C, Jonckx B, Mazzone M, et al. Anti-PlGF inhibits growth of VEGF(R)-inhibitor-resistant tumors without affecting healthy vessels. *Cell* 2007; 131:463-75.
54. Koperz S, Hoff PM, Morris JS, et al. Phase I trial of infusional fluorouracil, irinotecan, and bevacizumab for metastatic colorectal cancer: efficacy and circulating angiogenic biomarkers associated with therapeutic resistance. *J Clin Oncol* 2010; 28:453-9.
55. Holash J, Davis S, Papadopoulos N, et al. VEGF-Trap: a VEGF blocker with potent antitumor effects. *Proc Natl Acad Sci U S A* 2002; 99:11393-8.
56. Tew WP, Gordon M, Murren J, et al. Phase I study of aflibercept administered subcutaneously to patients with advanced solid tumors. *Clin Cancer Res* 2010; 16:358-66.
57. Van Cutsem E, Tabernero J, Lakomy R, et al. Intravenous (iv) aflibercept versus placebo in combination with irinotecan/5-FU (FOLFIRI) for second-line treatment of metastatic colorectal cancer (mCRC): results of a multinational phase III trial (EFC10262-VELOUR). Paper presented at: Congress on Gastrointestinal Cancer. European Society for Medical Oncology (ESMO). June 25, 2011; Barcelona, Spain.
58. Jin K, Shen Y, He K, et al. Aflibercept (VEGF Trap): one more double-edged sword of anti-VEGF therapy for cancer? *Clin Transl Oncol* 2010; 12:526-32.
59. Zaltrap [prescribing information]. Bridgewater, NJ: Regeneron Pharmaceuticals, Inc./sanofi-aventis, U.S. LLC; 2012.
60. Rudge JS, Holash J, Hylton D, et al. VEGF Trap complex formation measures production rates of VEGF, providing a biomarker for predicting efficacious angiogenic blockade. *Proc Natl Acad Sci U S A* 2007; 104:18363-70.
61. Dixon JA, Oliver SC, Olson JL, et al. VEGF Trap-eye for the treatment of neovascular age-related macular degeneration. *Expert Opin Investig Drugs* 2009; 18:1573-80.
62. Papadopoulos N, Martin J, Ruan Q, et al. Binding and neutralization of vascular endothelial growth factor (VEGF) and related ligands by VEGF Trap, ranibizumab and bevacizumab. *Angiogenesis* 2012; 15:171-85.
63. Lockhart AC, Rothenberg ML, Dupont J, et al. Phase I study of intravenous vascular endothelial growth factor trap, aflibercept, in patients with advanced solid tumors. *J Clin Oncol* 2010; 28:207-14.
64. Kim ES, Serur A, Huang J, et al. Potent VEGF blockade causes regression of coopted vessels in a model of neuroblastoma. *Proc Natl Acad Sci U S A* 2002; 99:11399-404.
65. Huang J, Frischer JS, Serur A, et al. Regression of established tumors and metastases by potent vascular endothelial growth factor blockade. *Proc Natl Acad Sci U S A* 2003; 100:7785-90.
66. Lassoued W, Murphy D, Tsai J, et al. Effect of VEGF and VEGF Trap on vascular endothelial cell signaling in tumors. *Cancer Biol Ther* 2011; 10:1326-33.
67. Chiron M, Vignaud P, Lejeune P, et al. Synergistic activity of aflibercept (VEGF Trap) in combination with 5-fluorouracil and irinotecan in preclinical tumor models. *AACR Meeting Abstracts* 2007; San Francisco, CA, October 2007:A13.
68. Lejeune P, Chiron M, Moigne RL, et al. Combination of the antiangiogenic agent aflibercept (VEGF Trap) with docetaxel and gemcitabine results in greater antitumor activity in tumor-bearing mice. Presented at: the American Association for Cancer Research (AACR) Annual Meeting, April 12-16, 2008; San Diego, CA. Abstract 1107.
69. Abraham C, Li B, Parveen A, et al. Combination of aflibercept (VEGF Trap) and docetaxel produces increased antitumor effects associated with enhanced changes to tumor vasculature. Presented at: the American Association for Cancer Research (AACR). Annual Meeting, April 17-21, 2010; Washington, DC. Abstract 5427.
70. Lal D, Park JA, Demock K, et al. Aflibercept exerts antivascular effects and enhances levels of anthracycline chemotherapy in vivo in human acute myeloid leukemia models. *Mol Cancer Ther* 2010; 9:2737-51.
71. Limentani SA, Just R, Purdham A, et al. A phase I dose escalation and pharmacokinetic (PK) study of intravenous (iv) aflibercept (VEGF Trap) plus FOLFIRI in patients (pts) with advanced solid tumors: preliminary results. *J Clin Oncol* 2008; 26(suppl): abstract:3556. Available at: www.asco.org. Accessed: September 15, 2012.
72. Rixe O, Verslype C, Khayat D, et al. A phase I dose escalation (DE) and pharmacokinetics (PK) study of intravenous (iv) aflibercept (VEGF Trap) plus irinotecan, 5-fluorouracil, and leucovorin (I-LV5FU2) in patients with advanced solid tumors (STs). *J Clin Oncol* 2008; 26(suppl): abstract:3557. Available at: www.asco.org. Accessed: September 15, 2012.
73. Isambert N, Freyer G, Zanetta S, et al. Phase I dose-escalation study of intravenous aflibercept in combination with docetaxel in patients with advanced solid tumors. *Clin Cancer Res* 2012; 18:1743-50.
74. Freyer G, Fumoleau P, You B, et al. A phase I dose escalation and pharmacokinetic (PK) study of intravenous (iv) aflibercept (VEGF Trap) plus docetaxel (D) and cisplatin (C) in patients (pts) with advanced solid tumors: preliminary results. *J Clin Oncol* 2008; 26(suppl):abstract:14539. Available at: www.asco.org. Accessed: September 15, 2012.
75. Patnaik A, Pipas M, Rosen L, et al. A phase I dose escalation and pharmacokinetic (PK) study of intravenous (iv) aflibercept (VEGF Trap) plus weekly gemcitabine (gem) in patients (pts) with advanced solid tumors: preliminary results. *J Clin Oncol* 2008; 26(suppl):abstract:3558. Available at: www.asco.org. Accessed: September 15, 2012.
76. Yamazaki K, Yoshino T, Yamaguchi K, et al. Phase I dose escalation and pharmacokinetics study of intravenous aflibercept plus irinotecan, 5-fluorouracil, and folic acid (FOLFIRI) in patients with metastatic colorectal cancer. *J Clin Oncol* 2011; 29(suppl): abstract:538. Available at: www.asco.org. Accessed: September 15, 2012.
77. Tew WP, Colombo N, Ray-Coquard I, et al. VEGF-Trap for patients (pts) with recurrent platinum-resistant epithelial ovarian cancer (EOC): preliminary results of a randomized, multicenter phase II study. *J Clin Oncol* 2007; 25(suppl): abstract: 5508. Available at: www.asco.org. Accessed: September 15, 2012.
78. Mareselli E, Miller VA, Leigh NB, et al. Phase II study of the efficacy and safety of intravenous (iv) AVE0005 (VEGF Trap) given every 2 weeks in patients (Pts) with platinum- and erlotinib-resistant adenocarcinoma of the lung (NSCLA). *J Clin Oncol* 2007; 25(suppl): abstract:7627. Available at: www.asco.org. Accessed: September 15, 2012.
79. Colombo N, Mangili G, Maramoliri S, et al. A phase II study of aflibercept in patients with advanced epithelial ovarian cancer and symptomatic malignant ascites. *Gynecol Oncol* 2012; 125:42-7.
80. Coleman RL, Duska LR, Ramirez PT, et al. Phase 1-2 study of docetaxel plus aflibercept in patients with recurrent ovarian, primary peritoneal, or fallopian tube cancer. *Lancet Oncol* 2011; 12:1109-17.
81. Goriieb WH, Amant F, Advani S, et al. Intravenous aflibercept for treatment of recurrent symptomatic malignant ascites in patients with advanced ovarian cancer: a phase 2, randomised, double-blind, placebo-controlled study. *Lancet Oncol* 2012; 13:154-62.
82. De Groot JF, Wen PY, Lamborn K, et al. Phase II single arm trial of aflibercept in patients with recurrent temozolomide-resistant glioblastoma: NABTC 0601. *J Clin Oncol* 2008; 26(suppl): abstract:2020. Available at: www.asco.org. Accessed: September 15, 2012.
83. Tang P, Cohen SJ, Bjarnason GA, et al. Phase II trial of aflibercept (VEGF Trap) in previously treated patients with metastatic colorectal cancer (MCR): a PMH phase II consortium trial. *J Clin Oncol* 2008; 26(suppl): abstract:4027. Available at: www.asco.org. Accessed: September 15, 2012.
84. Townsley C, Hirre H, Hoskins P, et al. A phase II study of aflibercept (VEGF Trap) in recurrent or metastatic gynecologic soft-tissue sarcomas: a study of the Princess Margaret Hospital Phase II Consortium. *J Clin Oncol* 2009; 27(suppl):abstract 5591. Available at: www.asco.org. Accessed: September 15, 2012.
85. Tarhini AA, Christensen S, Frankel P, et al. Phase II study of aflibercept (VEGF Trap) in recurrent inoperable stage III or stage IV melanoma of cutaneous or ocular origin. *J Clin Oncol* 2009; 27(suppl):abstract:9028. Available at: www.asco.org. Accessed: September 15, 2012.
86. Twardowski P, Sradler WM, Frankel P, et al. Phase II study of aflibercept (VEGF-Trap) in patients with recurrent or metastatic urothelial cancer, a California Cancer Consortium trial. *Urology* 2010; 76:923-6.
87. Leigh NB, Raez LE, Besse B, et al. A multicenter, phase 2 study of vascular endothelial growth factor trap (Aflibercept) in platinum- and erlotinib-resistant adenocarcinoma of the lung. *J Thorac Oncol* 2010; 5:1054-9.
88. Mackay HJ, Buckanovich RJ, Hirre H, et al. A phase II study single agent of aflibercept (VEGF Trap) in patients with recurrent or metastatic gynecologic carcinomas and uterine leiomyosarcoma. A trial of the Princess Margaret Hospital, Chicago and California cancer phase II consortia. *Gynecol Oncol* 2012; 125:136-40.
89. Tabernero J, Van Cutsem E, Lakomy R, et al. Results from VELOUR, a phase 3 study of aflibercept versus placebo in combination with FOLFIRI for the treatment of patients with previously treated metastatic colorectal cancer. Paper presented at: 2011 European Multidisciplinary Congress; September 23-27, 2011; Stockholm, Sweden. Abstract 6LBA.
90. Giantonio BJ, Catalano PJ, Neropoli NJ, et al. Bevacizumab in combination with oxaliplatin, fluorouracil, and leucovorin (FOLFIRI) for previously treated metastatic colorectal cancer: results from the Eastern Cooperative Oncology Group Study E3200. *J Clin Oncol* 2007; 25:1539-44.
91. Arnold D, Andre T, Bennouna J, et al. Bevacizumab (BEV) plus chemotherapy (CT) continued beyond first progression in patients with metastatic colorectal cancer (mCRC) previously treated with BEV plus CT: results of a randomized phase III intergroup study (TML study). *J Clin Oncol* 2012; 30(suppl):abstract CRA3503. Available at: www.asco.org. Accessed: September 15, 2012.
92. Stewart MW. Aflibercept (VEGF-TRAP): the next anti-VEGF drug. *Inflamm Allergy Drug Targets* 2011; 10:497-508.
93. Verslype C, Spano J, Van Cutsem E, et al. Validation of the selected dose of aflibercept (VEGF Trap) plus irinotecan, 5-fluorouracil, and leucovorin (I-LV5FU2) in a phase I clinical trial of patients (pts) with advanced solid tumors (STs): preliminary results. *J Clin Oncol* 2008; 26(suppl): abstract:14540. Available at: www.asco.org. Accessed: September 15, 2012.

ARTICLES

Blockade of Dll4 inhibits tumour growth by promoting non-productive angiogenesis

Irene Noguera-Troise¹, Christopher Daly¹, Nicholas J. Papadopoulos¹, Sandra Coetzee¹, Pat Boland¹, Nicholas W. Gale¹, Hsin Chieh Lin¹, George D. Yancopoulos¹ & Gavin Thurston¹

Tumour growth requires accompanying expansion of the host vasculature, with tumour progression often correlated with vascular density. Vascular endothelial growth factor (VEGF) is the best-characterized inducer of tumour angiogenesis. We report that VEGF dynamically regulates tumour endothelial expression of Delta-like ligand 4 (Dll4), which was previously shown to be absolutely required for normal embryonic vascular development. To define Dll4 function in tumour angiogenesis, we manipulated this pathway in murine tumour models using several approaches. Here we show that blockade resulted in markedly increased tumour vascularity, associated with enhanced angiogenic sprouting and branching. Paradoxically, this increased vascularity was non-productive—as shown by poor perfusion and increased hypoxia, and most importantly, by decreased tumour growth—even for tumours resistant to anti-VEGF therapy. Thus, VEGF-induced Dll4 acts as a negative regulator of tumour angiogenesis; its blockade results in a striking uncoupling of tumour growth from vessel density, presenting a novel therapeutic approach even for tumours resistant to anti-VEGF therapies.

Tumour growth depends on expansion of the host vasculature into the tumour, through the process of tumour angiogenesis¹. The connection between tumour growth and angiogenesis prompted the development of several approaches to limit tumour angiogenesis and thus control tumour growth. The best validated of these approaches involves blockade of the VEGF pathway. Blockade of VEGF controls tumour growth in numerous preclinical models^{2,3}, and recent results show that potent blockers of VEGF can completely prevent tumour angiogenesis in some models, thereby severely inhibiting tumour growth⁴. The promise of VEGF-blocking approaches has recently been realized in the clinic, as a VEGF-blocking antibody has been shown to have important effects on tumour progression and overall survival in cancer patients^{5,6}. However, despite the critical role for VEGF in tumour angiogenesis, it is also clear that in some cases tumour growth and angiogenesis can proceed even in the face of potent VEGF blockade^{7–9}. Thus, additional angiogenesis-targeted therapies are necessary for tumours resistant to VEGF blockade.

Mouse genetic studies have demonstrated that, in addition to the VEGF pathway, other signalling pathways are also required for normal embryonic vascular development (for reviews, see refs 10–12), raising the possibility that these pathways may also be important during tumour angiogenesis. One signalling pathway implicated in vascular development by gene deletion studies is the Notch pathway¹³. On binding a transmembrane ligand from the Delta/Jagged families, Notch transmembrane receptors generally provide signals to guide cell fate decisions^{14,15}. In particular, Delta-like ligand 4 (Dll4) is absolutely required for normal vascular development^{16–18} and is strongly expressed in tumour vessels^{17,19,20}.

To determine whether the Dll4/Notch pathway has a role during tumour angiogenesis, we manipulated this pathway in experimental tumour models in mice using a variety of genetic and pharmacologic approaches. We report that the Dll4/Notch pathway is a critical negative regulator of tumour angiogenesis, acting to restrain excessive VEGF-induced vascular sprouting and angiogenesis. Increased Dll4/Notch activity resulted in decreased tumour vascular density, whereas blockade of activity resulted in markedly increased vessel

density. Paradoxically, this increased vascularity seemed to be non-productive and resulted in decreased tumour growth, even for tumours that are resistant to anti-VEGF therapy. Our findings provide a striking example of an uncoupling of tumour growth from tumour vascular density, and support the model that the Dll4/Notch pathway normally acts as a negative regulator of angiogenic sprouting induced by VEGF or other pathways.

RESULTS

VEGF induces high expression of Dll4 in tumour vessels. To confirm and extend earlier reports that Dll4 expression was markedly and specifically induced in blood vessels during tumour angiogenesis^{17,19,20}, we used a combination of Dll4 detection approaches in two different tumour models. First, we exploited 'Dll4 reporter mice' in which a β -galactosidase reporter gene was driven by the *Dll4* promoter¹⁷. Lewis lung carcinomas implanted into these mice showed strong reporter-based staining of tumour vessels, apparently at higher levels than of vessels in surrounding normal tissue (Fig. 1a, b). At higher resolution, immunostaining for the β -galactosidase reporter protein and comparison with adjacent sections in which all vessels were immunostained with CD-31/PECAM antibodies revealed strong *Dll4* reporter expression in tumour blood vessels and relatively weak staining in adjacent subcutaneous and dermal blood vessels¹⁷ (Supplementary Fig. 1). To confirm that the β -galactosidase/*Dll4* reporter (*Dll4-LacZ*) construct marked sites of Dll4 protein, we generated polyclonal antibodies to murine Dll4; these antibodies also immunostained tumour blood vessels selectively (Fig. 1c, d). These findings were further confirmed by *in situ* hybridization for *Dll4* messenger RNA, which revealed prominent expression in the vessels of C6 glioma tumours¹⁹ (Supplementary Fig. 1). Thus, Dll4 is indeed specifically expressed in the tumour vasculature, particularly in the smaller vessels. Moreover, expression of Dll4 was dependent on continuous VEGF signalling, because blockade of VEGF with VEGF Trap, a recombinant soluble receptor that potently blocks VEGF-A and placental growth factor (PlGF)⁴, caused a rapid and marked decrease in the expression of Dll4 by tumour vessels (Fig. 1e).

¹Regeneron Research Laboratories, 777 Old Saw Mill River Road, Tarrytown, New York 10591, USA.

Activating and blocking the Dll4/Notch pathway in tumours. To manipulate the Dll4/Notch pathway in tumours, we first exploited a retroviral approach to overexpress forms of Dll4, which we reasoned would serve as blockers or activators, in tumour cells. On the basis of previous studies that used soluble versions of Dll to inhibit Notch signalling²¹, we generated retroviral vectors encoding a soluble dimerized version of Dll4 in which the extracellular region of Dll4 was fused to the human IgG1 Fc constant region (termed Dll4-Fc) as a presumed blocker, as well as full-length membrane-bound Dll4 that presumably would act as an activator; these constructs were transduced into rat C6 tumour cells to produce C6 Dll4-Fc and C6 Dll4 cells. Importantly, C6 tumour cells that overexpressed Dll4-Fc or Dll4 did not have different growth characteristics *in vitro* than control tumour cells (data not shown).

The expected changes on Notch signalling in the host (mouse) stroma of subcutaneously implanted C6 rat tumour cells were confirmed by quantitative messenger RNA analyses using probes specific for the mouse versions (to detect expression in the host stromal cells and not in the rat tumour cells) of three genes that are characteristic targets of Notch signalling (*HES1*, *HEY2* and *NRARP*)^{22–27}. That is, in C6 Dll4-Fc tumours, host Notch signalling was consistently reduced as reflected by decreased levels of these target genes, whereas in

C6 Dll4 tumours the Notch pathway was activated (Supplementary Fig. 3). Similarly, in co-culture studies in which the transduced rat tumour cells were mixed with human umbilical vein endothelial cells, expression levels of the *HES1*, *HEY2* and *NRARP* genes in the cultured endothelial cells (assayed using human specific probes so as to specifically detect the endothelial versions of these transcripts) were reduced by co-culture with C6 Dll4-Fc cells and induced by C6 Dll4 cells (Supplementary Fig. 4). Finally, treatment of cultured human umbilical vein endothelial cells with purified Dll4-Fc protein rapidly and consistently repressed Notch signalling, as reflected by decreased expression of these target genes (Supplementary Fig. 5).

Blockade of Dll4/Notch results in decreased tumour growth. To explore the effects of Dll4/Notch pathway manipulation in tumours, we next examined tumours derived from C6 Dll4-Fc and C6 Dll4 cells for their vascular morphologies and tumour growth rates. Strikingly, Dll4-Fc and full-length Dll4 seemed to promote reciprocal changes in the tumour vasculature: the vasculature in C6 Dll4-Fc tumours was much more highly branched and had more fine interconnections than that of control tumours (Fig. 2a–f). The leading front of the vessels was replete with sprouts and filopodia (Fig. 2e, h). In contrast, the vasculature in C6 Dll4 tumours was notably

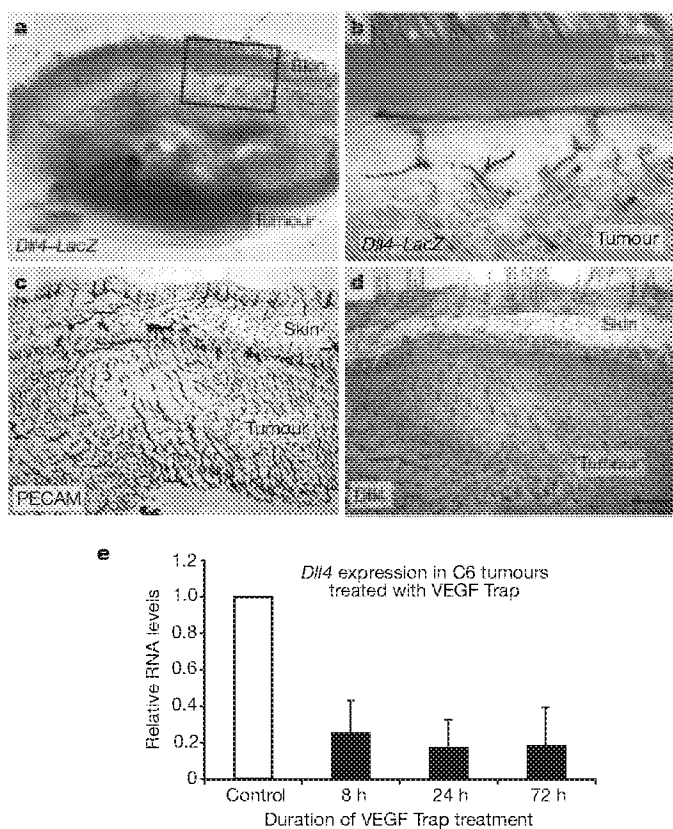


Figure 1 | Dll4 is expressed in tumour vessels, and its expression is dependent on VEGF signalling. a, b, Lewis lung tumours were grown subcutaneously in heterozygous *Dll4*-targeted mice in which the *Dll4* gene was replaced with the gene encoding β -galactosidase. a, *Dll4* expression, revealed by staining for LacZ (blue), was strong in tumour tissue, but weak in normal skin tissue. b, Higher magnification image showing *Dll4* expression in tumour blood vessels. c, Section of tumour showing immunoreactivity for CD31/PECAM is equally strong in tumour vessels and adjacent skin vessels. d, In contrast, immunoreactivity for *Dll4* is strong in tumour vessels and weaker in vessels of the adjacent skin. Scale bar: 400 μ m (c, d). e, Expression of *Dll4* in tumour vessels is decreased by VEGF blockade. Rat C6 tumours grown subcutaneously were treated for the indicated times with VEGF Trap. Tumours were analysed for expression of murine *Dll4* by microarray analysis. Data show mean \pm s.d. of *Dll4* expression compared with control C6 tumours from three tumours per time point ($n = 3$).

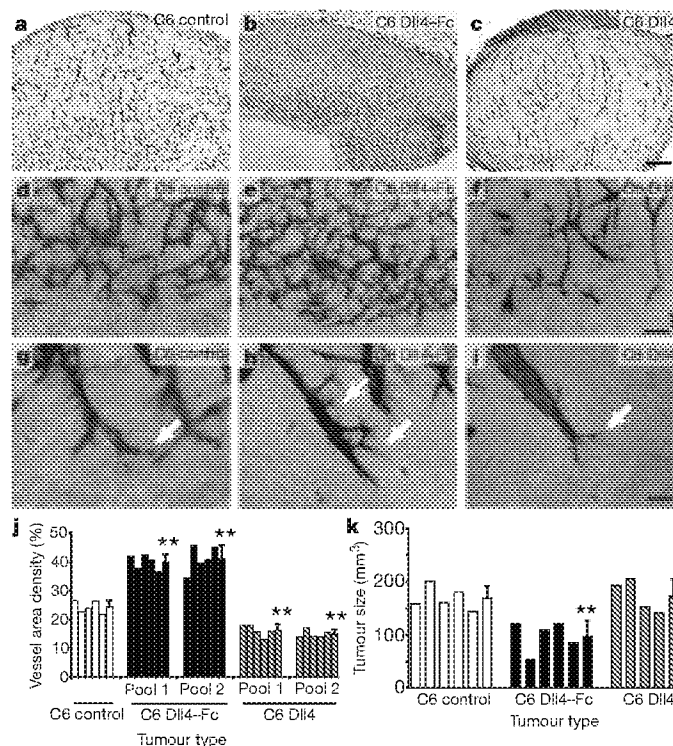


Figure 2 | Blockade of Dll4/Notch signalling results in smaller C6 tumours with increased vessel density. a–i, Micrographs showing tumours stained for CD31 (black) in control C6 tumours (a, d, g), in C6 tumours overexpressing Dll4-Fc (b, e, h), and in C6 tumours overexpressing full-length Dll4 (c, f, i). Micrographs show tumour sections at low, medium and high magnification. C6 Dll4-Fc tumours contain a dense network of vessel structures, with numerous cellular processes, particularly at the leading front (b, e). Reciprocally, C6 Dll4 tumours contain relatively sparse and unbranched vessels (c, f). The leading cells in the actively growing vascular front of Dll4-Fc tumours have more cellular processes than those in control tumours, and reciprocally, such cells in Dll4 tumours have fewer processes (arrows, g–i). Scale bars, 400 μ m (a–c); 100 μ m (d–f); 20 μ m (g–i). j, Quantification of vessel area density by morphometry shows increased vessel density in C6 Dll4-Fc tumours and decreased vessel density in C6 Dll4 tumours. k, C6 Dll4-Fc tumours are smaller than C6 GFP control tumours, whereas C6 Dll4 tumours are the same size. Quantification was done on two independently isolated pools of C6 Dll4-Fc and C6 Dll4 cells (results from a single pool are shown). Data from individual tumours are shown, as well as mean \pm s.d. (j, k) for each group ($n = 4–5$); ** $P < 0.01$.

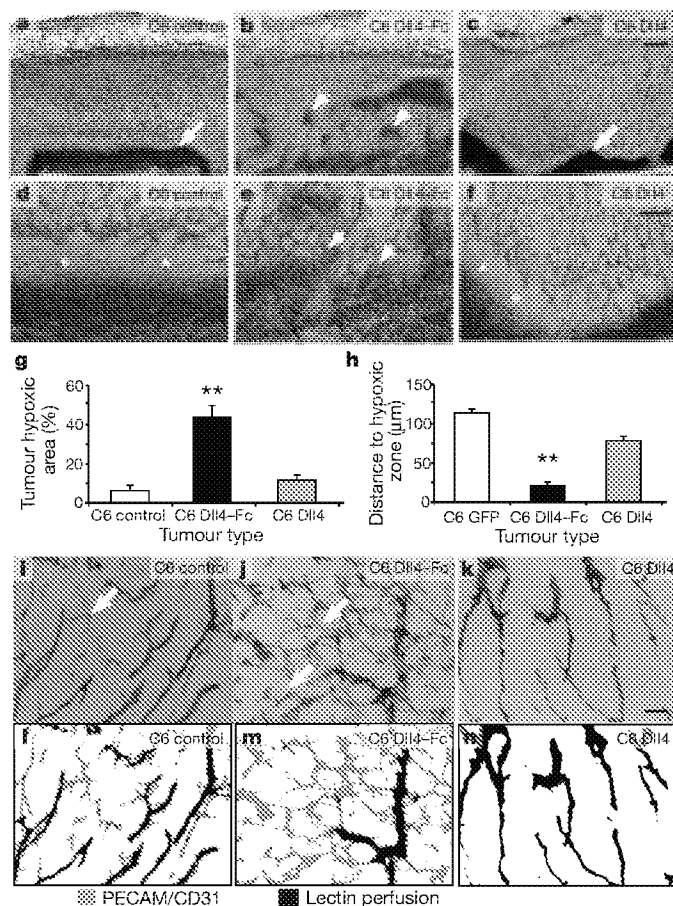


Figure 3 | Despite an increase in blood vessel density, tumours overexpressing Dll4-Fc have increased hypoxia and poor vascular perfusion. **a–f**, C6 tumours were stained for CD31/PECAM (brown) and for hypoxia (HypoxyProbe, black). Control C6 tumours of this size had no evident necrosis, and had very little hypoxia in the region of vascularized tumour, but did have a prominent rim of hypoxia at the base of the tumour (a, black staining, arrow) that was separated from the leading front of tumour vessels (asterisks in d). In comparison, C6 Dll4-Fc tumours contained increased areas of hypoxia in the region of vascularized tumour (arrowheads, b). In addition, hypoxia was found immediately adjacent to, and intermingled with, the leading front of tumour vessels (arrowheads, e). C6 Dll4 tumours had few areas of hypoxia in the region of vascularized tumour, and the hypoxic region (arrow, c) was typically separated from the leading front of vessels (asterisks, f), although more variability was seen than in control tumours. **g**, Quantification of hypoxic region within the vascularized tumour. C6 Dll4-Fc tumours contained significantly more hypoxia. **h**, Quantification of the distance between the leading front of vessels and the hypoxic zone, as marked with white asterisks in panels d, e, f. The distance to the hypoxic region was significantly less in C6 Dll4-Fc tumours than in control tumours. Values are mean \pm s.d. (**g**, **h**) for each group ($n = 4–5$); $**P < 0.01$. **i–n**, Comparison of vessel perfusion and overall CD31/PECAM immunoreactivity reveals decreased vessel function in C6 Dll4-Fc tumours. Tumour vessels were stained by *in vivo* intravascular injection of biotinylated lectin (stained in black) to mark perfused vessels, and by immersion in CD31/PECAM antibodies (stained in brown) to mark all vascular structures. **i–l**, Computer-generated colour images represent thresholds to show perfused vessels (black) and non-perfused vascular structures (green). Vascular structures in control C6 tumours (i, l) had a mixture of only brown staining, showing lack of perfusion (particularly in the fine processes, arrow) as well as a combination of black and brown staining showing perfused vessels. In C6 Dll4-Fc tumours (j, m), many vascular structures, particularly the numerous fine processes, were not perfused by intravascular tracer and thus stained only for CD31 (arrows). Reciprocally, in C6 Dll4 tumours (k, n), the unbranched vascular structures, which lack fine processes, were stained by both perfused lectin (black) and CD31 (brown), showing that vascular structures are functional. Scale bars, 100 μ m (**a–c**); 40 μ m (**d–f**); 20 μ m (**i–n**).

1034

straighter and relatively unbranched (Fig. 2c, f), and relatively devoid of sprouts and filopodia (Fig. 2f, i).

These obvious morphologic changes were reflected by quantitation of vascular area densities in these tumours, with the C6 Dll4-Fc tumours exhibiting increased vascular density as compared with control tumours, whereas the C6 Dll4 tumours exhibited slightly decreased vascular density (Fig. 2j). Paradoxically, the effects of Dll4-Fc and full-length Dll4 on vascular density were opposite to those that might be expected with respect to their effects on tumour growth. Despite increased vascular density, C6 Dll4-Fc tumours were consistently smaller than control tumours, whereas C6 Dll4 tumours were not substantially different in size from control tumours for small tumours (Fig. 2k) or when C6 Dll4 tumours were harvested at larger sizes (Supplementary Fig. 6). These results have two important implications: first, that the Dll4/Notch pathway normally serves as a negative regulator of sprouting and branching activity during tumour angiogenesis, so that blockade of this pathway results in increased tumour angiogenesis; and second, that the increased angiogenesis resulting from blockade of this pathway is in some sense 'non-productive', such that it does not support more robust tumour growth and instead seems to be associated with reduced tumour growth.

Blockade of Dll4/Notch increases tumour hypoxia. To account for the apparent paradox of increased tumour vessel density and decreased tumour growth in C6 Dll4-Fc tumours, we examined the possibility that the increased network of vessels might not be

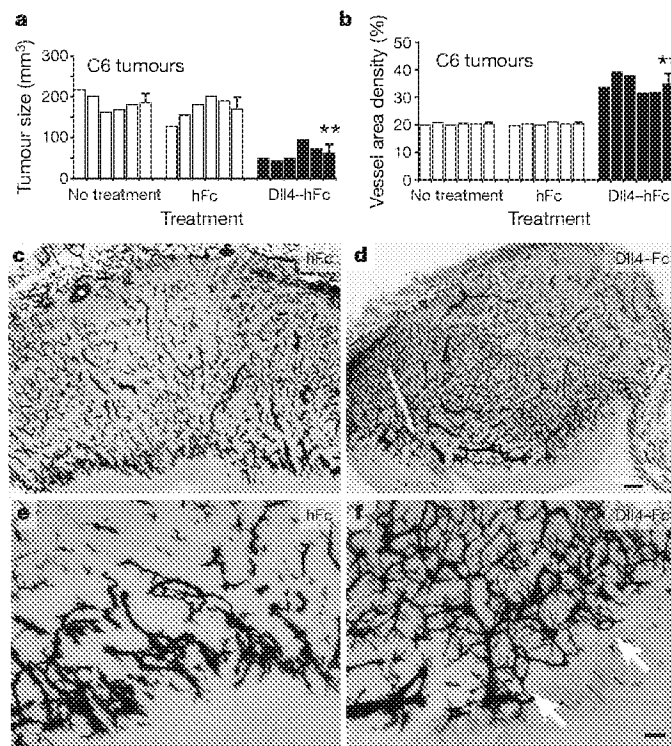


Figure 4 | Systemic delivery of Dll4-Fc using adenovirus results in smaller C6 tumours and increased vessel density, similar to effects of local tumour overexpression. **a**, C6 tumours were smaller in mice treated with systemically delivered Dll4-Fc, but not by control hFc. **b**, The vessel area density is increased in C6 tumours treated by systemic Dll4-Fc compared with untreated tumours or those treated with systemic hFc. The graphs in **a** and **b** show data from individual tumours, as well as mean \pm s.d. for each group ($n = 5$). $**$ denotes significantly different than the control tumour group ($P < 0.01$). **c–f**, Micrographs showing tumour vessel morphology in control tumours (hFc; **c**, **e**) and in tumours treated systemically with Dll4-Fc (Dll4-Fc; **d**, **f**). The vessel density is increased in C6 Dll4-Fc tumours, particularly at the leading front (**f**, arrows). Tumour sections stained for CD31 (PECAM, black). Scale bars, 200 μ m (**c**, **d**); 50 μ m (**e**, **f**).

optimally functional. In agreement with this possibility, histologic assessment showed more extensive tumour hypoxia in C6 Dll4-Fc tumours than in control tumours (Fig. 3a, b; hypoxic regions stained in black). Moreover, whereas the hypoxic region in control tumours was separated from the growing front of tumour vessels by an avascular zone that was itself not hypoxic (corresponding to the oxygen diffusion distance; white asterisks, Fig. 3d), areas of hypoxia were interspersed with the tumour vasculature in C6 Dll4-Fc tumours (arrowheads, Fig. 3b, e), indicating that this vasculature was not efficiently delivering oxygen to the surrounding tumour. Quantitative analysis showed that Dll4-Fc tumours contained sevenfold more total hypoxic area within the vascularized tumour (Fig. 3g), and that the hypoxic rim was less separated from the leading front of tumour vessels (Fig. 3h). Corresponding values in the C6 Dll4 tumours were not substantially different from controls

(Fig. 3c), although there was more variability in the distance from the vascular front to the hypoxic rim (white asterisks, Fig. 3f).

The increase in tumour hypoxia in the C6 Dll4-Fc tumours suggested that the dense network of vessels was not fully perfused. We compared the distribution of vessel perfusion (marked by intravascular lectin as a tracer) with the immunohistochemical staining of endothelial cells (stained with CD31/PECAM-1 antibodies) as a marker of total vasculature. Most of the larger vessels of control tumours were perfused, although—as expected—these vessels were associated with some non-perfused sprouts and smaller vessels emanating from the larger vessels (Fig. 3i, brown reveals total vasculature, black reveals perfused vessels; Fig. 3l, computer-generated colour depiction, with green showing total vasculature and black showing perfused vessels). In contrast, many of the vascular processes in the C6 Dll4-Fc tumours were not perfused (Fig. 3j, m), suggesting that the increased vessel density seen in these tumours is not part of a functional vascular network. Reciprocally, the relatively straight and unbranched vessels seen in the C6 Dll4 tumours were almost completely perfused (Fig. 3k, n).

Together, the tumour hypoxia and perfusion analyses support the notion that the increased vascular network that results from inhibition of the Dll4/Notch pathway (by Dll4-Fc) is not optimally functional and is instead 'non-productive'.

Systemic Dll4-Fc decreases tumour growth. To confirm and extend the finding that locally produced Dll4-Fc promotes excessive angiogenesis that paradoxically blunts tumour growth, we used an adenoviral delivery approach to determine whether systemic Dll4-Fc could also produce this effect. Adenoviruses expressing Dll4-Fc, or human Fc (hFc) as control, were injected intravenously into mice at the time of implanting subcutaneous C6 tumours. The intravenously injected adenovirus infects hepatocytes, and in the case of Dll4-Fc or hFc, produced high serum levels of the encoded protein of $74 \pm 20 \mu\text{g ml}^{-1}$ (range of $50\text{--}100 \mu\text{g ml}^{-1}$, $n = 5$ mice). Circulating Dll4-Fc resulted in an approximately 70% reduction in the size of subcutaneous C6 tumours (Fig. 4a), whereas circulating control hFc had no effect. When examined by histology, circulating Dll4-Fc also caused an increase in the density of the tumour vessels (Fig. 4b). The overall increased vessel density produced by circulating Dll4-Fc was associated with a dense mesh of highly branched and sprouted vessels, particularly at the leading front (Fig. 4c–f), similar to that produced by Dll4-Fc overexpression in the tumour cells. Gene expression analysis confirmed that systemic Dll4-Fc suppressed the Notch pathway in the tumour vessel, as indicated by decreased expression of *HES1* and *NRARP* (data not shown). Thus, inhibition of Dll4/Notch signalling by either local or systemic Dll4-Fc results in smaller C6 tumours and excessive but apparently non-productive tumour angiogenesis. Importantly, the systemic treatment did not seem to have untoward effects on the host animals; in addition, preliminary analysis of normal tissues did not reveal obvious changes in tissue vascularity (Supplementary Information).

Dll4-Fc or Dll4-blocking antibody act in multiple tumour models. To further extend the above findings, we used systemic injection of purified recombinant Dll4-Fc protein as the treatment, and tried additional tumour models. The above studies were carried out using C6 gliomas, which are relatively sensitive to the effects of VEGF blockade⁴, so we assessed the response to Dll4/Notch blockade in other tumours that are more resistant to VEGF blockade. In previous experiments using both bevacizumab (Avastin) and VEGF Trap, we had developed a model of HT1080 tumours (HT1080-(resistance model)RM), which is relatively resistant to both Avastin and VEGF Trap (G.T., I.N.-T. and J. Rudge, unpublished results). In contrast to Avastin and VEGF Trap, Dll4-Fc protein was quite effective in reducing the growth of HT1080-RM tumours (Fig. 5a). In a separate experiment (Fig. 5b), we assessed the growth curves of HT1080-RM tumours treated with Dll4-Fc, VEGF Trap or control protein (all at 25 mg kg^{-1} , three times per week; treatment began when tumours were approximately 100 mm^3 in size, arrow). Dll4-Fc

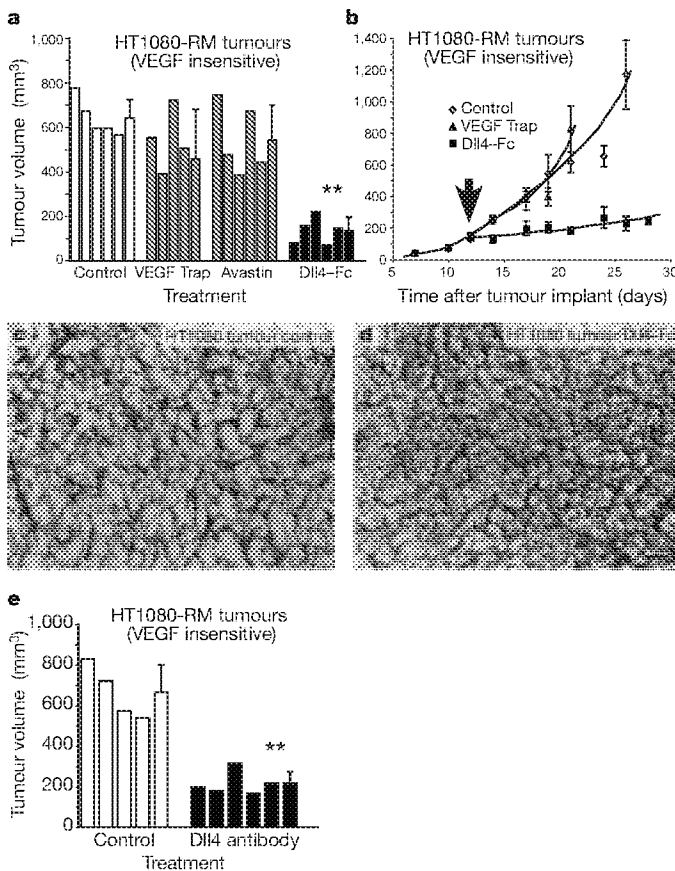


Figure 5 | Systemic delivery of Dll4-Fc or blocking Dll4 antibodies to mice bearing tumours that are resistant to blockade of VEGF results in decreased tumour growth and dramatic changes in tumour vessels. **a**, Size of HT1080-RM tumours treated with Dll4-Fc protein or VEGF Trap or bevacizumab (Avastin, all at 25 mg kg^{-1}). Tumours were treated with subcutaneous injections starting once tumours had reached approximately 100 mm^3 . Dll4-Fc treated tumours were smaller than controls, whereas those treated with blockers of VEGF were almost the same size as controls. **b**, Tumour growth curves from a separate experiment in which HT1080-RM tumours were treated from 100 mm^3 (arrow) with VEGF Trap, Dll4-Fc, or hFc (25 mg kg^{-1} , three times per week). Error bars are s.d. Whereas tumours treated with VEGF Trap grew as rapidly as control tumours, those treated with Dll4-Fc were restrained for at least 2 weeks of treatment. **c**, **d**, Dll4-Fc caused an increase in tumour vessel density and dramatic changes in vessel morphology in HT1080-RM tumours. Tumour sections were stained for CD31. Scale bar, $50 \mu\text{m}$ (**c**, **d**). **e**, Antibodies to Dll4 strongly suppressed tumour growth, similar to the effect of Dll4-Fc. HT1080-RM tumours were treated from a size of 100 mm^3 with polyclonal antibodies to Dll4 (10 mg kg^{-1} , three times per week) or with rabbit IgG as a control. The bar graphs show data from individual tumours, as well as the mean \pm s.d. for each group ($n = 4\text{--}5$).

treatment resulted in a prolonged suppression of tumour growth, whereas VEGF Trap had almost no impact on tumour growth in this resistant tumour model. Dll4-Fc treatment of HT1080-RM tumours also had a marked effect on tumour vessels. The already-dense vasculature of control HT1080 tumours (Fig. 5c) was further increased by treatment with Dll4-Fc (Fig. 5d), inducing an apparent increase in vascular sprouting and branching, and an apparent disorganization to the network.

To verify the specificity of blocking with Dll4-Fc, we also generated polyclonal antibodies to the extracellular portion of Dll4, which inhibited the binding of Dll4 to Notch1 in *in vitro* assays (Supplementary Fig. 7). Systemic treatment of mice bearing resistant HT1080-RM tumours with blocking Dll4 antibodies also reduced tumour growth (Fig. 5e) and caused marked changes in tumour vessels (data not shown).

As for HT1080 tumours, growth of mouse mammary tumours, which are also resistant to VEGF blockade (not shown), was strongly suppressed by systemic treatment with Dll4-Fc (Supplementary Fig. 8). Again, the vascularity of mouse mammary tumours was further increased by treatment with Dll4-Fc (Supplementary Fig. 8). Thus, as with C6 tumours, treatment of other tumours with systemic Dll4-Fc resulted in decreased tumour growth, accompanied by a denser and more highly branched tumour vasculature.

Discussion

To determine whether the Dll4/Notch pathway has a role during tumour angiogenesis, we manipulated this pathway in tumours using a variety of approaches. Our findings suggest that tumour-derived VEGF induces Dll4 expression in angiogenic endothelial cells as a critical negative regulator of vascular growth, acting to restrain excessive vascular sprouting and branching, and allowing angiogenesis to proceed at a productive rate (Supplementary Fig. 9). Thus, increasing Dll4/Notch activity resulted in decreased vascular density associated with less sprouting and branching of the vascular network. In contrast, Dll4/Notch blockade was associated with enhanced angiogenic sprouting and branching, resulting in a marked increase in tumour vessel density but a decrease in vessel function (Supplementary Fig. 9). Previously, angiogenesis-based treatment of tumours has focused on trying to block angiogenesis; however, our results using Dll4 blockade suggest an alternative approach based on promoting 'non-functionality' in the growing tumour vasculature. Although our studies suggest that VEGF blockade may be equally or more effective than Dll4 blockade in many tumour models, certain models that are resistant to VEGF blockade can still be sensitive to Dll4 blockers.

We, and others, have shown that Dll4 is specifically expressed in remodelling vessels and is the major Notch ligand in the vasculature. Thus, rather than a general blockade of the Notch pathway, specific blockade of Dll4 may lead to more specific disruption of tumour growth without significant impairment of Notch function in normal host tissues, and thus might be well tolerated in long-term treatments. It seems likely that biological therapeutic agents, which can be specific to a particular ligand or receptor in this complex pathway, may prove more potent and specific than more general pathway blockers, such as the γ -secretase inhibitors that not only block all Notch signalling but also other important γ -secretase-mediated signalling as well.

Our findings provide a striking example of an uncoupling of tumour growth from tumour vascular density. Although a large literature supports the notion that tumour growth rate may correlate with tumour vascular density, other studies argue that tumour angiogenesis must be regulated to be productive. For example, a recent study suggests that tumours may have higher vascular densities than is necessary to support their growth, and thus tumour angiogenesis may often exceed an optimally productive rate²⁸. Consistent with the concept of excessive tumour vessel density, some recent studies suggest that pruning of the vasculature might actually improve tumour

perfusion and oxygenation^{29–31}. The present studies with blockers of Dll4/Notch seem to provide the other side of the argument; in particular, that Dll4 blockade may further compromise tumour vasculature function by causing excessive non-productive angiogenesis, which can in turn inhibit tumour growth. The overall message seems to be that even tumour vascular networks require a regulated balance of growth factors to generate a hierarchy of well-organized and well-functioning vessels. VEGF clearly has a key angiogenic role in a wide variety of tumours, but Dll4 blockade may present a new therapeutic opportunity in cancer, and one that might be beneficial for patients with tumours that are resistant to anti-VEGF therapies.

METHODS

Engineered retroviruses. Retroviruses engineered to express green fluorescent protein (GFP) (control), GFP plus Dll4-hFc or GFP plus Dll4 were used to transduce C6 rat glioma tumour cells. Tumour cell pools, sorted by flow cytometry, were implanted subcutaneously in severe combined immunodeficient (SCID) mice (8–10 weeks old). Tumours were harvested and processed for histology and/or gene expression analysis.

Engineered adenoviruses. Adenoviruses engineered to express hFc or Dll4-Fc were injected into the jugular vein of mice bearing subcutaneous C6 glioma, mouse mammary or HT1080 tumours. Adenoviruses provided systemic production of the engineered proteins.

Antibodies. Anti-Dll4 polyclonal antibodies were generated by immunizing rabbits against murine Dll4-hFc protein. Serum was depleted for antibodies with reactivity to human Fc and then used to stain tumour sections or treat tumour-bearing mice.

Reporter mice. Dll4 Lac/Z reporter mice¹⁷ generated using Velocigene technology³² were implanted with Lewis lung tumour cells. Tumours were stained with antibodies to CD31/Pecam-1 and/or β -galactosidase, or reacted with 5-bromo-4-chloro-3-indolyl- β -D-galactoside (X-gal), and counterstained with pyronin-Y.

In vitro assays. *In vitro* assays to determine the effect of Dll4-Fc and full length Dll4 on Notch signalling used confluent human umbilical vein endothelial cells (VEC Technologies) treated with Dll4-Fc protein (10 μ g ml⁻¹) for 2 to 8 h. RNA was extracted and analysed by Taqman, using probes and primers specific for human HES1, HEY2 and NRARP. In other experiments, human umbilical vein endothelial cells (50% confluent) were co-cultured with C6 glioma cells and assayed at 24 h.

Assaying hypoxia and vessel perfusion. To measure hypoxia and vessel perfusion, HypoxyProbe-1 (Chemicon; 60 mg kg⁻¹) was injected intraperitoneally one hour before sacrifice. Tumours were processed for histological analysis, and tumour sections were stained using anti-Hypoxyprobe antibody. To mark vessel perfusion, mice were injected through the jugular vein with biotinylated *Lycopersicon esculentum* lectin (100 μ g, Vector Laboratories). Lectin circulated for 3 min, and then tumours were subsequently stained for lectin bound to the endothelial cell surface³³.

Received 17 June; accepted 16 October 2006.

- Folkman, J. The role of angiogenesis in tumor growth. *Semin. Cancer Biol.* **3**, 65–71 (1992).
- Ferrara, N. Vascular endothelial growth factor as a target for anticancer therapy. *Oncologist* **9** (Suppl. 1), 2–10 (2004).
- Rudge, J. S. *et al.* VEGF trap as a novel antiangiogenic treatment currently in clinical trials for cancer and eye diseases, and VelociGene-based discovery of the next generation of angiogenesis targets. *Cold Spring Harb. Symp. Quant. Biol.* **70**, 411–418 (2005).
- Holash, J. *et al.* VEGF-Trap: a VEGF blocker with potent antitumor effects. *Proc. Natl Acad. Sci. USA* **99**, 11393–11398 (2002).
- Hurwitz, H. *et al.* Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N. Engl. J. Med.* **350**, 2335–2342 (2004).
- Laskin, J. J. & Sandler, A. B. First-line treatment for advanced non-small-cell lung cancer. *Oncology* **19**, 1671–6; discussion 1678–80 (2005).
- Casanovas, O., Hicklin, D. J., Bergers, G. & Hanahan, D. Drug resistance by evasion of antiangiogenic targeting of VEGF signalling in late-stage pancreatic islet tumors. *Cancer Cell* **8**, 299–309 (2005).
- Jain, R. K., Duda, D. G., Clark, J. W. & Loeffler, J. S. Lessons from phase III clinical trials on anti-VEGF therapy for cancer. *Nature Clin. Pract. Oncol.* **3**, 24–40 (2006).
- Kerbel, R. S. *et al.* Possible mechanisms of acquired resistance to anti-angiogenic drugs: implications for the use of combination therapy approaches. *Cancer Metastasis Rev.* **20**, 79–86 (2001).
- Yancopoulos, G. D. *et al.* Vascular-specific growth factors and blood vessel formation. *Nature* **407**, 242–248 (2000).
- Jain, R. K. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science* **307**, 58–62 (2005).

12. Carmeliet, P. Angiogenesis in life, disease and medicine. *Nature* **438**, 932–936 (2005).
13. Shawber, C. J. & Kitajewski, J. Notch function in the vasculature: insights from zebrafish, mouse and man. *Bioessays* **26**, 225–234 (2004).
14. Artavanis-Tsakonas, S., Rand, M. D. & Lake, R. J. Notch signaling: cell fate control and signal integration in development. *Science* **284**, 770–776 (1999).
15. Gridley, T. Notch signaling during vascular development. *Proc. Natl Acad. Sci. USA* **98**, 5377–5378 (2001).
16. Duarte, A. *et al.* Dosage-sensitive requirement for mouse Dll4 in artery development. *Genes Dev.* **18**, 2474–2478 (2004).
17. Gaie, N. W. *et al.* Haploinsufficiency of delta-like 4 ligand results in embryonic lethality due to major defects in arterial and vascular development. *Proc. Natl Acad. Sci. USA* **101**, 15949–15954 (2004).
18. Krebs, L. T. *et al.* Haploinsufficient lethality and formation of arteriovenous malformations in Notch pathway mutants. *Genes Dev.* **18**, 2469–2473 (2004).
19. Mailhos, C. *et al.* Delta4, an endothelial specific notch ligand expressed at sites of physiological and tumor angiogenesis. *Differentiation* **69**, 135–144 (2001).
20. Patel, N. S. *et al.* Up-regulation of delta-like 4 ligand in human tumor vasculature and the role of basal expression in endothelial cell function. *Cancer Res.* **65**, 8690–8697 (2005).
21. Hicks, C. *et al.* A secreted Delta1–Fc fusion protein functions both as an activator and inhibitor of Notch1 signaling. *J. Neurosci. Res.* **68**, 655–667 (2002).
22. Taylor, K. L., Henderson, A. M. & Hughes, C. C. Notch activation during endothelial cell network formation *in vitro* targets the basic HLH transcription factor HESR-1 and downregulates VEGFR-2/KDR expression. *Microvasc. Res.* **64**, 372–383 (2002).
23. Iso, T., Kedes, L. & Hamamori, Y. HES and HRP families: multiple effectors of the Notch signaling pathway. *J. Cell. Physiol.* **194**, 237–255 (2003).
24. Shawber, C. J., Das, I., Francisco, E. & Kitajewski, J. Notch signaling in primary endothelial cells. *Ann. NY Acad. Sci.* **995**, 162–170 (2003).
25. Karsan, A. The role of notch in modeling and maintaining the vasculature. *Can. J. Physiol. Pharmacol.* **83**, 14–23 (2005).
26. Lassar, E. *et al.* Nrarp is a novel intracellular component of the Notch signaling pathway. *Genes Dev.* **15**, 1885–1899 (2001).
27. Krebs, L. T., Deftos, M. L., Bevan, M. J. & Gridley, T. The *Nrarp* gene encodes an ankyrin-repeat protein that is transcriptionally regulated by the Notch signaling pathway. *Dev. Biol.* **238**, 110–119 (2001).
28. Krneta, J. *et al.* Dissociation of angiogenesis and tumorigenesis in follistatin- and activin-expressing tumors. *Cancer Res.* **66**, 5686–5695 (2006).
29. Lee, C. G. *et al.* Anti-vascular endothelial growth factor treatment augments tumor radiation response under normoxic or hypoxic conditions. *Cancer Res.* **60**, 5565–5570 (2000).
30. Jain, R. K. Tumor angiogenesis and accessibility: role of vascular endothelial growth factor. *Semin. Oncol.* **29**, 3–9 (2002).
31. Jain, R. K. Antiangiogenic therapy for cancer: current and emerging concepts. *Oncology* **19**, 7–16 (2005).
32. Valenzuela, D. M. *et al.* High-throughput engineering of the mouse genome coupled with high-resolution expression analysis. *Nature Biotechnol.* **21**, 652–659 (2003).
33. Thurston, G., Baluk, P., Hirata, A. & McDonald, D. M. Permeability-related changes revealed at endothelial cell borders in inflamed venules by lectin binding. *Am. J. Physiol.* **271**, H2547–H2562 (1996).
34. Liu, Z. J. *et al.* Inhibition of endothelial cell proliferation by Notch1 signaling is mediated by repressing MAPK and PI3K/Akt pathways and requires MAML1. *FASEB J.* **20**, 1009–1011 (2006).

Supplementary Information is linked to the online version of the paper at www.nature.com/nature.

Acknowledgements We acknowledge the following Regeneron colleagues: Y. Wei for gene expression analysis, A. Adier, A. Rafique, B. Li, H. Huang, E. Pasnikowski, J. McClain, E. Burova, D. Hylton, P. Burfeind and J. Griffiths for technical assistance, S. Staton for assistance with graphics, and S. Wiegand, I. Lobov, T. Daly, S. Davis, E. Ioffe, J. Holash and J. Rudge for scientific input.

Author Contributions I. N.-T. directed and helped perform tumour experiments, generation of tumour lines, immunohistochemical staining, and data analysis. C.D. directed, helped perform, and analysed *in vitro* experiments. N.J.P. helped develop protein reagents and biochemical assays. S.C. performed and helped analyse tumour experiments and construction of tumour cell lines. P.B. performed and helped analyse immunohistochemical studies. N.W.G. helped perform and analyse experiments with gene-targeted mice. H.C.L. helped perform and analyse gene expression studies. G.D.Y. helped analyse and interpret results. G.T. helped design experiments, analyse data and interpret results.

Author Information Reprints and permissions information is available at www.nature.com/reprints. The authors declare competing financial interests: details accompany the paper on www.nature.com/nature. Correspondence and requests for materials should be addressed to G.T. (Gavin.Thurston@Regeneron.com).

VEGF Trap complex formation measures production rates of VEGF, providing a biomarker for predicting efficacious angiogenic blockade

John S. Rudge*, Jocelyn Holash[†], Donna Hylton, Michelle Russell, Shelly Jiang, Raymond Leidich, Nicholas Papadopoulos, Erica A. Pyles, Al Torri, Stanley J. Wiegand, Gavin Thurston, Neil Stahl, and George D. Yancopoulos*

Regeneron Pharmaceuticals, Inc., 777 Old Saw Mill River Road, Tarrytown, NY 10591

This contribution is part of the special series of Inaugural Articles by members of the National Academy of Sciences elected on April 20, 2004.

Contributed by George D. Yancopoulos, September 21, 2007 (sent for review July 16, 2007)

VEGF is the best characterized mediator of tumor angiogenesis. Anti-VEGF agents have recently demonstrated impressive efficacy in human cancer trials, but the optimal dosing of such agents must still be determined empirically, because biomarkers to guide dosing have yet to be established. The widely accepted (but unverified) assumption that VEGF production is quite low in normal adults led to the notion that increased systemic VEGF levels might quantitatively reflect tumor mass and angiogenic activity. We describe an approach to determine host and tumor production of VEGF, using a high-affinity and long-lived VEGF antagonist now in clinical trials, the VEGF Trap. Unlike antibody complexes that are usually rapidly cleared, the VEGF Trap forms inert complexes with tissue- and tumor-derived VEGF that remain stably in the systemic circulation, where they are readily assayable, providing unprecedented capability to accurately measure VEGF production. We report that VEGF production is surprisingly high in non-tumor-bearing rodents and humans, challenging the notion that systemic VEGF levels can serve as a sensitive surrogate for tumor load; tumor VEGF contribution becomes significant only with very large tumor loads. These findings have the important corollary that anti-VEGF therapies must be sufficiently dosed to avoid diversion by host-derived VEGF. We further show that our assay can indicate when VEGF is optimally blocked; such biomarkers to guide dosing do not exist for other anti-VEGF agents. Based on this assay, VEGF Trap doses currently being assessed in clinical trials are in the efficacious range.

affibercept | angiogenesis | tumor | endothelial cell

VEGF is critical in many settings of physiological and pathological angiogenesis (1). In particular, high VEGF expression is characteristic of many types of cancers (1), suggesting that it might be an attractive target for therapeutic intervention aimed at preventing tumors from recruiting the blood supply that they need to survive (2). The first attempts at validating this particular approach were taken by Ferrara and colleagues (3), who demonstrated that a murine anti-human VEGF antibody suppressed the growth of human tumor cell lines implanted in nude mice. This led to the generation of a humanized monoclonal antibody, bevacizumab (Avastin; Genentech, South San Francisco, CA), which yielded impressive results in a controlled clinical trial in patients with metastatic renal cell cancer (4, 5). At doses of 3 and 10 mg/kg, bevacizumab treatment resulted in a significant prolongation in time to tumor progression compared with placebo, although the increased efficacy of the higher dose in this study suggested that the maximally efficacious dose may not yet have been attained (4, 5). Bevacizumab was subsequently granted FDA approval based on the demonstration that it significantly improved the progression-free and overall survival in patients with metastatic colorectal cancer when given in combination with irinotecan 5-FU/LV chemotherapy (6). Sev-

eral other drugs designed to block VEGF signaling have since been developed and recently approved [BAY 43-9006 (sorafenib) and SU11248 (sunitinib)] or are proceeding through clinical trials [PTK787 (vatalanib), ZD6474 (zactima), ZD6126, SU5416 (semaxanib), and AG-013736] (7-9).

As new anti-VEGF agents proceed through the clinic, it would be very useful to have biomarkers that could either identify patients whose tumors depend most on VEGF or that could guide dosing by indicating when optimal VEGF blockade has been achieved. Unfortunately, accepted biomarkers do not currently exist for VEGF blockade and are few and far between for other targeted agents, such as epidermal growth factor receptor for colon cancer, Kit for gastrointestinal stromal tumor, and HER2/NEU for breast cancer (10). VEGF itself has been suggested as a candidate biomarker for guiding the application of anti-VEGF therapies. It is widely assumed that VEGF production is quite low in healthy adults in the absence of active angiogenesis. Were that the case, blood levels of VEGF in cancer patients might provide a useful index of tumor VEGF production (11, 12). However, because VEGF is rapidly cleared from the systemic circulation (having a half-life of only minutes), the sensitivity of assays measuring VEGF in the peripheral blood leads to a wide variability for blood levels of VEGF in published reports. Furthermore, VEGF is present at substantial levels within platelets and released upon their lysis such that preparation of peripheral blood samples that avoid contamination from platelet-derived VEGF becomes difficult. These limitations are reflected in the disparate values reported for circulating VEGF levels in cancer patients, which range from 0.04 to 1 ng/ml, calling into question the utility of plasma VEGF levels as a useful biomarker for guiding anti-angiogenic therapy (11, 13-19).

VEGF Trap is a fully human soluble decoy receptor protein that consists of a fusion of the second Ig domain of human VEGF receptor (VEGFR) 1 and the third Ig domain of human VEGFR2 with the constant region (Fc) of human Ig IgG1 (20).

Author contributions: J.S.R., J.H., and G.D.Y. designed research; J.S.R., S.J.W., G.T., and N.S. analyzed data; J.H., D.H., M.R., S.J., R.L., N.P., E.A.P., A.T., and G.T. performed research; N.P. contributed new reagents/analytic tools; and J.S.R. and G.D.Y. wrote the paper.

The authors declare no conflict of interest.

Freely available online through the PNAS open access option.

Abbreviations: AMD, age-related macular degeneration; MALLS, multiangled laser light scattering; SEC, size exclusion chromatography; VEGFR, VEGF receptor.

*To whom correspondence may be addressed. E-mail: john.rudge@regeneron.com or george@regeneron.com.

[†]Present address: Novartis, 1400 53rd Street, Emeryville, CA 94608.

This article contains supporting information online at www.pnas.org/cgi/content/full/0708865104/DC1.

© 2007 by The National Academy of Sciences of the USA

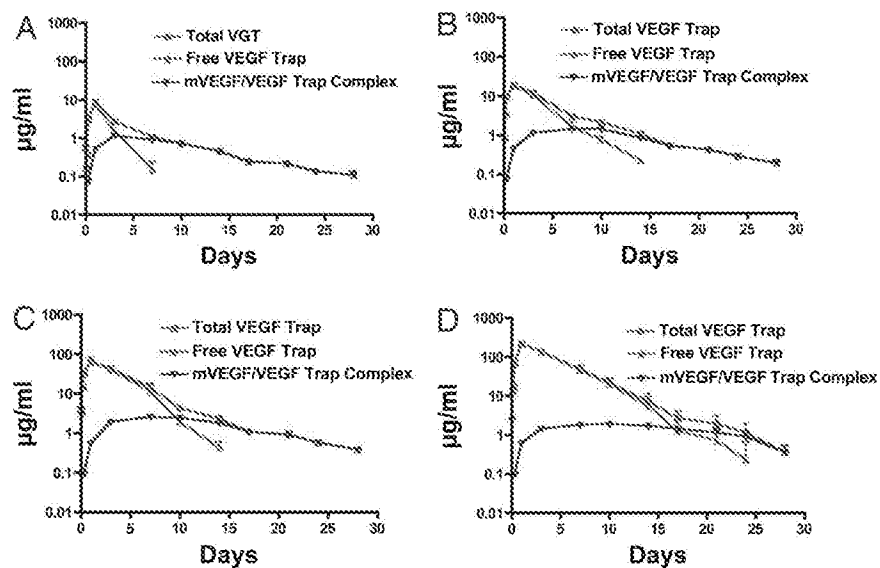


Fig. 1. s.c. injection of VEGF Trap into SCID mice at different doses reveals different levels of circulating free VEGF Trap but similar levels of circulating mouse VEGF–VEGF Trap complex. At all doses ranging from 1 mg/kg (A) to 25 mg/kg (D), a steady-state level of VEGF–VEGF Trap complex is achieved, which plateaus at ≈ 1 $\mu\text{g}/\text{ml}$. Dose-dependent levels of free VEGF Trap are observed as follows: 1 mg/kg to 10 $\mu\text{g}/\text{ml}$ C_{max} falling below complex levels at 4 days (A); 2.5 mg/kg to 20 $\mu\text{g}/\text{ml}$ C_{max} falling below complex levels at 7 days (B); 10 mg/kg to 80 $\mu\text{g}/\text{ml}$ C_{max} falling below complex levels at 9 days (C); and 25 mg/kg to 200 $\mu\text{g}/\text{ml}$ C_{max} falling below complex levels at 17 days (D). The half-life of VEGF Trap is ≈ 2 days at doses > 2.5 mg/kg. ($n = 6$ for each dose.)

The VEGF Trap was engineered to have optimized pharmacokinetic properties and a very high affinity for all isoforms of VEGF-A (< 1 pM), as well as placental growth factor, a closely related angiogenic factor (20). VEGF Trap has shown robust antitumor effects in numerous mouse models of cancer and is now in clinical trials (21, \ddagger , \S , \parallel). Here, we show that—unlike VEGF antibodies that tend to form multimeric immune complexes that are rapidly cleared from the circulation and can form immune complex deposits in tissues—the VEGF Trap forms a stable and inert 1:1 complex with VEGF. This VEGF–VEGF Trap complex has a long plasma half-life and can readily be measured in the systemic circulation, thus affording a reliable way to measure the rates of VEGF production in both tumor-bearing and non-tumor-bearing adult animals and humans. This unique ability to capture and thus precisely measure total VEGF levels, regardless of whether the VEGF comes from tumor or normal host tissues, allows for the unprecedented opportunity to accurately determine tumor and host VEGF production rates. Surprisingly, we find that total body VEGF production rates are quite high in normal adult rodents and humans, with the fractional contribution made by tumors being comparatively small. This finding has the important implication that therapies directed toward neutralizing VEGF produced by tumors must be provided in sufficient amounts so as to avoid being largely consumed by the significant levels of VEGF produced by the rest of the body. Toward this end, measurement of VEGF Trap complex allows the identification of VEGF Trap doses required to completely capture and block tumor-derived VEGF, providing a useful guide for optimizing angiogenic blockade; such assays do not exist for other anti-VEGF agents. Based on this

assay, we report that VEGF Trap doses currently being assessed in clinical trials appear to be in the efficacious range.

Results

VEGF Trap Forms an Inert Complex with VEGF That Remains Stably in the Circulation. Initial studies to determine the clearance rate of VEGF Trap revealed that it could form stable detectable complexes with endogenous VEGF in normal adult mice. After single injections of increasing amounts of VEGF Trap, we measured total VEGF Trap, uncomplexed/unbound or “free” VEGF Trap, and VEGF Trap–mouse VEGF “complex” at various times after injection (Fig. 1 A–D represent increasing amounts of injected VEGF Trap). Because no exogenous VEGF was provided, complexes represent the association of VEGF Trap with endogenous murine VEGF. As expected, total VEGF Trap levels increased proportional to dose (determined by combining free VEGF Trap levels with complex levels) (Fig. 1, see green curves). Somewhat unexpectedly, substantial levels of VEGF Trap complexed with mouse VEGF accumulated rapidly (Fig. 1, see blue curves). At all doses of VEGF Trap tested, maximal levels of complex (≈ 1 – 2 $\mu\text{g}/\text{ml}$) were attained within 24–48 h of injection and sustained at this level for at least several days. Consistent with conversion of free VEGF Trap into complexed VEGF Trap, most of the injected VEGF Trap is initially found in the free, unbound form, but after reaching peak levels (≈ 24 h after injection) free VEGF Trap in the circulation declines progressively (Fig. 1, note that red curves, corresponding to free VEGF Trap, initially overlap at early time points with green curves, representing total VEGF Trap, but then drop, as is most obvious at the lowest dose). Levels of free VEGF Trap decline because of a “consumption” (binding VEGF, thus being converted to complex) and clearance, which occurs at an identical rate for free and bound Trap. Thus, as long as free VEGF Trap remains in excess of bound, maximal steady-state levels of complex are maintained in the circulation. VEGF Trap is also able to bind placental growth factor with high affinity and is capable of forming stable circulating placental growth factor–VEGF Trap complexes *in vivo* with the same profile as VEGF–

[†]Rixe, G., Verslype, C., Méric, J. B., Tejpar, S., Bloch, J., Crabbe, M., Khayat, D., Furfine, E. S., Assadourian, S., Van Cutsem, E. (2006) *J. Clin. Oncol.* 24:13161 (abstr.).

[§]Mulay, M., Limentani, S. A., Carroll, M., Furfine, E. S., Cohen, D. P., Rosen, L. S. (2006) *J. Clin. Oncol.* 24:13061 (abstr.).

[¶]Tew, W. P., Colombo, N., Ray-Coquard, I., Oza, A., del Campo, J., Scambia, G., Spriggs, D. (2007) *J. Clin. Oncol.* 25:5508 (abstr.).

^{||}Massarelli, E., Miller, V. A., Leigh, N., Rosen, P., Albain, K., Hart, L., Melnyk, O., Sternas, L., Akerman, J., Herbst, R. S. (2007) *J. Clin. Oncol.* 25:7627 (abstr.).

VEGF Trap complex, albeit at ≈ 10 -fold lower levels (data not shown).

In separate experiments, the bioavailability of VEGF Trap and the efficiency of VEGF capture were determined by injecting s.c. [supporting information (SI) Fig. 7*A*] or i.v. (SI Fig. 7*B*) preformed complexes of the Trap and its VEGF target, or both agents separately. The results show that the bioavailability of s.c. (SQ) injected complex was essentially identical to that of i.v. injected complex, indicating that negligible complex was depositing within tissues. Moreover, whether the VEGF Trap was injected as a preformed complex with VEGF (single bolus) or the Trap and its target were injected separately, similar levels of complex were rapidly noted in the circulation, indicating that the Trap efficiently captures its target and brings it into the systemic circulation. In addition, VEGF Trap is also capable of sequestering VEGF already bound in target tissues as shown by injecting VEGF before VEGF Trap (SI Fig. 7). Thus, VEGF Trap efficiently captures and forms inert complexes with VEGF that enter and remain stably in the circulation, readily accessible for measurement.

Although VEGF Trap Forms a 1:1 Complex with VEGF, VEGF Antibodies Form Heterogeneous, Multimeric Immune Complexes with VEGF. The above findings suggested that VEGF Trap might behave very differently than VEGF antibodies, because antibodies commonly form multimeric immune complexes that rapidly deposit in tissues and thus are rapidly cleared from the circulation. Because immune complexes rapidly disappear, the amount of captured ligand cannot be determined from levels of bound or unbound antibodies remaining in the circulation. To demonstrate directly that the VEGF Trap behaves in a fundamentally different way than antibodies, we compared VEGF Trap complex formation and clearance with that of a well characterized VEGF antibody, bevacizumab (Avastin). As predicted, size exclusion chromatography (SEC) of a preformed VEGF Trap-VEGF₁₆₅ complex revealed a single major homogenous peak, with an approximate molecular mass (as judged by comparison to molecular mass standards, data not shown) of ≈ 150 kDa corresponding to that expected of a 1:1 complex between VEGF Trap (≈ 110 kDa) and VEGF₁₆₅ (≈ 40 kDa) (Fig. 2*A*, solid red line); a minor peak of free excess VEGF₁₆₅ was also seen, as was a small shoulder of higher molecular mass. The molecular masses of the peaks were confirmed by using coupled multiangled laser light scattering (MALLS) (dashed red lines in Fig. 2*A*). In contrast, SEC of preformed bevacizumab-VEGF₁₆₅ complexes revealed a heterogeneous mixture corresponding to very high molecular masses (Fig. 2*A*, solid blue line) in addition to the small peak of free excess VEGF₁₆₅. The purity of free VEGF Trap, bevacizumab, and VEGF was $>97\%$, as determined by SEC (data not shown). Coupled MALLS analysis revealed molecular masses of the heterogeneous mixture ranging from 370 kDa (corresponding to a multimer consisting of two bevacizumab molecules, each with a molecular mass of ≈ 145 kDa, and two VEGF₁₆₅ molecules, each with a molecular mass of ≈ 40 kDa) to $>2,000$ kDa (corresponding to much larger multimers) (Fig. 2*A*, dashed blue line). Consistent with the apparent tendency of bevacizumab to form multimeric immune complexes with VEGF, preformed bevacizumab-VEGF₁₆₅ complexes rapidly disappeared from the circulation when injection intravenously, as would be expected for multimeric immune complexes (SI Fig. 8; note that the levels of Bevacizumab when complexed with VEGF rapidly drop compared with the levels of free Bevacizumab that remain much higher), and in contrast to what was described above with VEGF Trap complexes that remain stably in the circulation. Because immune complexes can often be cleared by depositing in the renal glomeruli, we further explored apparent differences in the clearance of bevacizumab-VEGF and VEGF Trap-VEGF complexes by performing im-

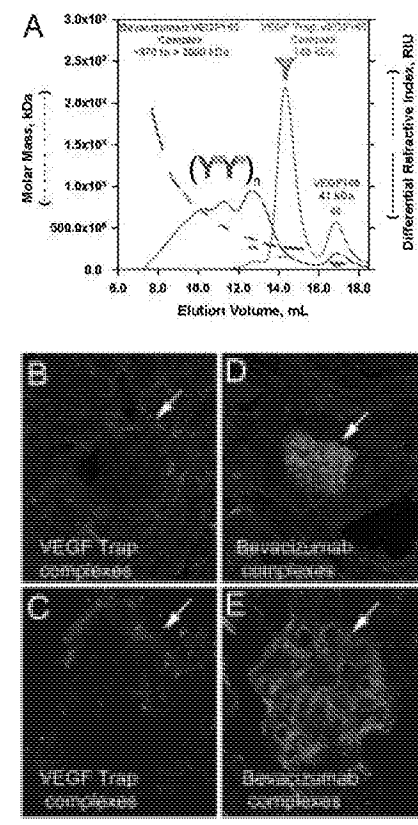


Fig. 2. The molar masses of VEGF Trap-VEGF and bevacizumab-VEGF complexes were determined by MALLS coupled to SEC. (*A*) Using a 1:2 molar ratio of VEGF Trap to VEGF₁₆₅, discrete peaks were observed at ≈ 17 ml for VEGF (41 kDa) and ≈ 14.5 ml for VEGF Trap-VEGF complex (148 kDa) with SEC (red line) and MALLS (dashed red line). In contrast, a 1:2 molar ratio of bevacizumab to VEGF₁₆₅ revealed a heterogeneous multimeric complex that ranged in molar mass from ≈ 370 kDa to $>2,000$ kDa (SEC, solid blue line; MALLS, dashed blue line). (*B–E*) One milligram of a preformed complex of VEGF Trap and VEGF₁₆₅ (*B* and *C*) or bevacizumab and VEGF₁₆₅ (*D* and *E*) were injected into the left ventricle of 2- to 3-month-old C57bl/6 mice. After 10 min, mice were killed, and their kidneys were processed for immunocytochemistry, using an anti human Fc reporter antibody to the human Fc moiety present on both VEGF Trap and bevacizumab. Significant staining was observed in the glomeruli of bevacizumab/VEGF treated mice but not in the glomeruli of VEGF Trap/VEGF treated mice (white arrows).

munostaining in the kidney. After i.v. administration, renal glomeruli stained strongly for bevacizumab-VEGF complexes (Fig. 2*D* and *E*) but not for VEGF Trap-VEGF complexes (Fig. 2*B* and *C*). Current evidence indicates that, as a class, pharmacological agents that block VEGF signaling may produce mechanism-based effects on kidney function. Deposition of immune complexes as noted for bevacizumab/VEGF in the renal glomeruli could further accentuate renal toxicity in a nonspecific and non-class-dependent manner.

VEGF Trap Complex Formation Reveals Unexpectedly High Production of Endogenous VEGF in Normal Adult Mice. As shown above, VEGF antibodies form immune complexes that rapidly deposit in tissues and thus do not allow for easy ascertainment of the amount of complex formed. In contrast, VEGF Trap forms inert complexes with VEGF that remain stably in the circulation and are thus readily accessible for measurement. In fact, the above findings demonstrate that, if VEGF Trap is present at sufficient levels so as to be in excess of Trap bound in complexes, the steady-state levels of VEGF Trap complex in the circulation reflect the total amount of VEGF produced. Daily production

rates of VEGF can be calculated by assuming that steady-state levels of VEGF Trap–VEGF complex reflect a balance between production of VEGF leading to formation of complex, and clearance of the resulting complex. Based on experimentally determined values for the steady-state levels of complex and its clearance (see *Materials and Methods*), we estimate that mice produce $\approx 0.065 \mu\text{g}$ of VEGF per day per ml of the volume of distribution, or $\approx 0.006 \mu\text{g}$ per gram of tissue per day. Because VEGF is active at picomolar levels, this at first seems to be a surprisingly high level of production for a normal adult animal (see below for comparison to tumor production rates). However, it should be noted that in the absence of VEGF Trap, any VEGF that enters the systemic circulation is rapidly cleared. For this reason, among others noted above, it has not proven possible to consistently and reliably measure systemic VEGF levels, preventing accurate estimation of VEGF production rates in normal adult animals.

Tumor-Derived VEGF Represents a Minority of Total Body VEGF Under Conditions of Minimal Tumor Burden. Next, we compared the total body production rate of VEGF, as determined above, with tumor production rates of VEGF. Toward this end, we implanted mice with tumors, allowed these tumors to grow to 0.5–3% of total body weight (average mouse weight, ≈ 25 g) and measured levels of VEGF Trap complex in these mice to compare them to complex levels found in healthy, non-tumor-bearing mice. Surprisingly, in mice bearing four different types of rodent tumors, the total levels of complex were not markedly different from those seen in non-tumor-bearing mice ($1\text{--}2 \mu\text{g/ml}$; see Fig. 3*A* and compare with Fig. 1). This finding implies that tumor-derived VEGF represented only a small proportion of total body VEGF or circulating bioavailable VEGF in these mice.

To further validate this unanticipated finding, we analyzed VEGF Trap complex levels in mice bearing human tumors, where it is possible to distinguish complexes formed with endogenous mouse VEGF with those formed with human VEGF derived from the implanted tumors by analyzing human VEGF–VEGF Trap complex levels in mouse serum. The levels of mouse-derived complexes (Fig. 3*B*) in these animals were equivalent to those of non-tumor-bearing mice (Fig. 2, above) and mice bearing rodent-derived tumors (Fig. 3*A*). In contrast, the levels of VEGF Trap complexed with tumor-derived human VEGF were an order of magnitude lower ($0.08\text{--}0.2 \mu\text{g/ml}$) (Fig. 3*B*). This result was seen in mice bearing tumors of three different human cell lines (SK-NEP, A673, and HT1080). Together, these studies demonstrate that normal total body production of VEGF eclipses the production from tumors that may weigh as much as 3% of body weight (mouse weight ranges from 23 to 29 g). Thus, it is unlikely that total levels of free VEGF in the systemic circulation would provide a sensitive index of tumor burden, even if accurate measurement of unbound VEGF in blood samples were readily achievable. Moreover, the above findings suggest that therapeutic compounds designed to bind and inactivate tumor-derived VEGF would have to be provided at sufficient levels to avoid being diverted by significant levels of VEGF normally produced by the rest of the body.

VEGF Trap Complex Levels Provide Guidance on When Efficacious VEGF Blockade Is Achieved. Based on the results above, it is evident that drugs that bind and neutralize VEGF must engage significant levels of VEGF derived from normal tissues, in addition to that originating from tumors. Therefore, we reasoned that measurements of VEGF Trap complex might provide a useful guide to when the dose of VEGF Trap sufficient to substantially neutralize both host and tumor-derived VEGF had been achieved. Indeed, for three different tumors [B16F1 mouse melanoma (Fig. 4*A*); A673 human rhabdomyosarcoma (Fig. 4*B*); and MMT mouse mammary carcinoma (Fig. 4*C*)], increasing the VEGF

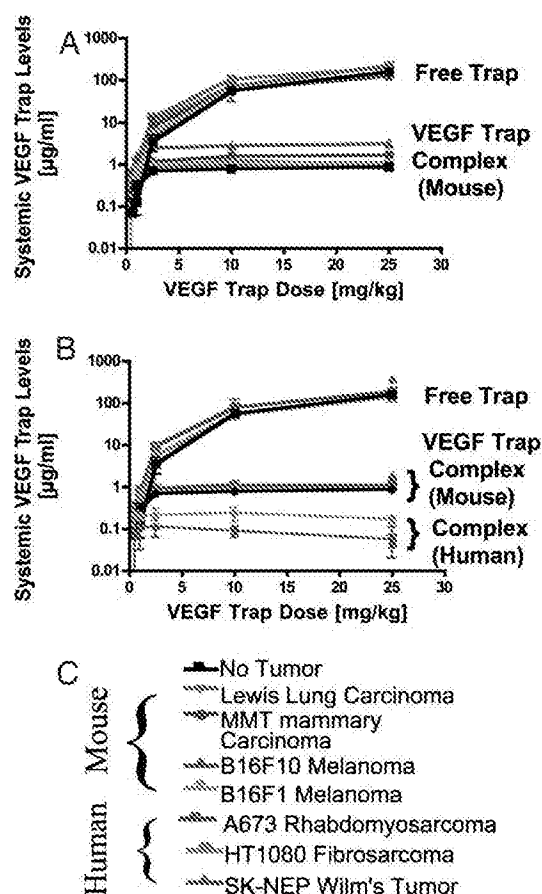


Fig. 3. In mice bearing tumors <3% body weight, the tumor pool of VEGF production is modest compared with endogenous mouse tissue VEGF production. (*A* and *B*) Mouse (*A*) or human (*B*) tumors were allowed to grow to $\approx 100 \text{ mm}^3$, and then VEGF Trap was administered twice per week for 1–2 weeks at 0.5, 1, 2.5, 10, and 25 mg/kg. At the termination of the experiment, free VEGF Trap, mouse, and human complex levels were measured in serum. In all cases, regardless of terminal tumor volume, levels of circulating mouse complex were $\approx 1 \mu\text{g/ml}$, whereas human complex levels in the mice bearing human tumors were $\approx 0.1 \mu\text{g/ml}$. Free Trap levels increased incrementally, with the dose levels rising above complex levels at the 2.5 mg/kg dose and reaching $\approx 100 \mu\text{g/ml}$ at the 25 mg/kg dose. (*n* = 6 for each dose). (*C*) Legend of mouse and human tumor types used.

Trap dose resulted in progressive, marked improvements in anti-tumor efficacy until a dose at which free VEGF Trap substantially exceeded maximal steady-state levels of complex was reached (Fig. 4). For all three tumor types, this was achieved at a dose of 2.5 mg/kg VEGF Trap given twice weekly: at this dose, free VEGF Trap (blue curve) is severalfold the level of complex (green curve), and past this point further dose escalation yields only modest incremental increases in complex levels (green curve) and in anti-tumor efficacy (red curve). In other tumor types, such as U87 glioblastoma, higher levels of VEGF Trap are required to achieve maximal efficacy (22).

Human VEGF/VEGF Trap Complex Levels Are Directly Related to Tumor Size. The finding that conventionally sized s.c. tumors in mice produced <10% the amount of total body VEGF prompted us to determine whether there is a consistent relationship between tumor size and VEGF production levels. Human tumors (A673 rhabdomyosarcoma) were implanted into mice and allowed to grow to various sizes before injecting VEGF Trap. In this case, we could define a clear linear relationship between tumor size (Fig. 5*A*) and complex levels (Fig. 5*B*), note that the assay reflects

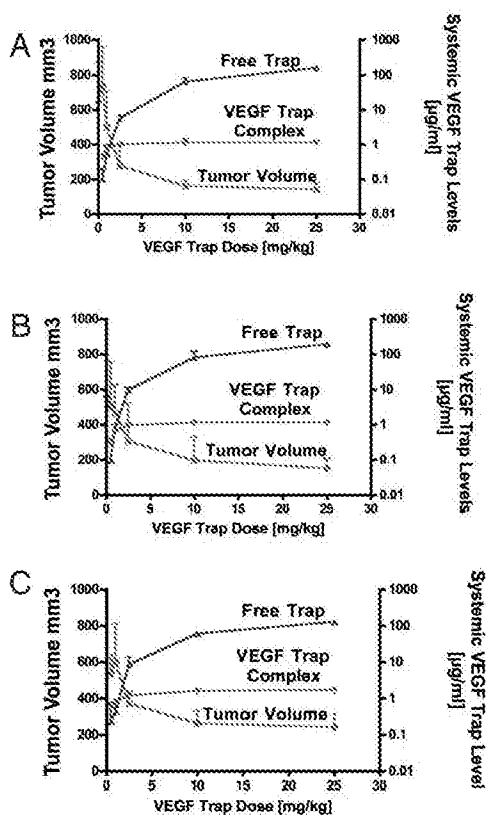


Fig. 4. VEGF Trap Complex provides guidance on when optimal VEGF blockade is achieved for antitumor purposes. In mice bearing B16F1 mouse melanoma tumors (A), A673 human rhabdomyosarcoma (B), and MMT mouse mammary carcinoma tumors (C) grown to ≈ 100 mm³ before treatment, increasing the dose of VEGF Trap from 0.5 mg/kg twice per week to 25 mg/kg twice per week results in a steady-state of mouse complex at ≈ 1 μ g/ml at 1–2.5 mg/kg and free circulating VEGF Trap levels of ≈ 10 μ g/ml at the 2.5 mg/kg dose, rising to ≈ 100 μ g/ml at the 25 mg/kg dose. Tumors remain quite large at the 0.5 and 1 mg/kg doses but begin to show a significant lack of growth at the 2.5 mg/kg dose, where free Trap levels rise above steady-state complex levels ($n = 6$ for each dose). Tumors were treated with VEGF Trap from 6–13 (B16F1), 4–13 (MMT), and 12–18 (A673) days after implantation.

levels of complexes containing only human VEGF to specifically detect only tumor-derived complex). The amount of complex per unit weight of tumor was similar across different-sized tumors (Fig. 5C), indicating that tumors maintained their rates of VEGF production as they grew. Linear regression analysis confirmed that there was a very strong correlation between A673 tumor volume and circulating human VEGF complex (Fig. 5D).

At these larger tumor sizes, the amount of complex (ranging from ≈ 0.8 to 5 μ g/ml) contributed by the tumor matched or even exceeded that contributed by the rest of the body, confirming that tumors do indeed make substantially more VEGF per cell than does the average cell in the normal adult host. For example, in the largest tumors (weighing $\approx 10\%$ of the total mass of the mouse, Fig. 5A), the tumor-derived human VEGF-VEGF Trap complex levels (≈ 5 μ g/ml, Fig. 5B) were ≈ 3 -fold above the levels of murine VEGF-VEGF Trap complex, indicating that the tumors made ≈ 30 times the amount of VEGF per unit of weight compared with normal, adult tissues.

VEGF Trap Complex Formation in Human Subjects With and Without Cancer. Very large tumors that substantially contribute to VEGF Trap complex formation in mice are generally not seen in the human patient. This in turn suggests that it is unlikely that most tumors in human patients become large enough to make a

readily detectable contribution to total body VEGF production. To determine whether or not this was indeed the case, we studied VEGF Trap complex formation in non-cancer patients [patients suffering from age-related macular degeneration (AMD)] and then compared these results with complex formation in cancer patients. In the AMD patients, the lowest dose of VEGF Trap tested (0.3 mg/kg, i.v.) was insufficient to neutralize all VEGF, as evidenced by the levels of free Trap quickly falling below those of bound VEGF Trap, and bound VEGF Trap did not approach the maximal steady-state levels seen with higher doses (Fig. 6A and B). However, doses of 1.0 and 3.0 mg/kg (i.v.) maintained substantial free Trap levels throughout the dosing period (Fig. 6A), and maximal complex levels were attained, as evidenced by equivalent levels of complex being generated at the two higher doses (≈ 1 –2 μ g/ml, see Fig. 6B). In cancer patients with advanced solid tumors or non-Hodgkin's lymphoma, remarkably similar results were obtained. That is, similar doses of VEGF Trap were required to saturate VEGF binding and complex formation (Fig. 6C–E). In addition, the maximal steady-state levels of VEGF-VEGF Trap complex were similar to those seen in non-cancer patients (Fig. 6B, D, and E). These findings indicate that, consistent with our findings in mice, endogenous VEGF production in adult human subjects is quite high, whether or not the individuals harbor tumors (Fig. 6E).

Using the same approach as was used for the mouse (see *Materials and Methods*), human production rates of VEGF in humans were found to be ≈ 0.0025 μ g per gram of tissue per day, which is remarkably similar to that calculated for mice (see above). If our findings in animal models continue to be predictive, these VEGF Trap levels achieved in ongoing clinical studies should be in the efficacious range.

Discussion

At present, there are a number of anti-angiogenic agents targeting the VEGF pathway that are proceeding through clinical trials or already approved for the treatment of cancer (9). One major challenge is the lack of objective measures to guide dosing to determine when sufficient blockade has been achieved or to inform pharmacological response to these drugs. VEGF itself has been suggested as a potential biomarker for the above purposes, based on the assumption that VEGF in the peripheral circulation was primarily derived from the tumor and therefore accurately reflected tumor burden (19). However, to date it has proven difficult to accurately measure systemic levels of VEGF, correlate these levels with tumor burden, or use them as a guide to dosing (11, 12). Here, we describe the use of the VEGF Trap, a potent VEGF antagonist that forms a stable, inert complex with VEGF, as an index that allows for the accurate assessment of VEGF production rates. In addition, this unique property of the VEGF Trap allows accurate assessment of the amounts of VEGF made by a resident tumor compared with the rest of the body. Furthermore, in animals, this approach has been shown to provide a useful guide to selecting dosing regimens that substantially block available VEGF. This has not been possible with anti-VEGF antibodies, as VEGF-antibody complexes are rapidly cleared.

We find unexpectedly high levels of VEGF production in the normal adult setting, where it has long been assumed that, in the absence of ongoing angiogenesis, VEGF production rates would be quite low (11, 12). However, the unexpectedly high rates of VEGF production in non-tumor-bearing adult mice and humans is consistent with the recent realization that VEGF likely plays an ongoing role in the “quiescent” vasculature of normal adults (23). For example, treating normal adult mice and monkeys with VEGF antagonists can increase hematocrit (a measure of the proportion of the blood volume occupied by red blood cells) (24). Similarly, VEGF antagonists can also increase blood pressure (25), indicating that VEGF is involved in regulating

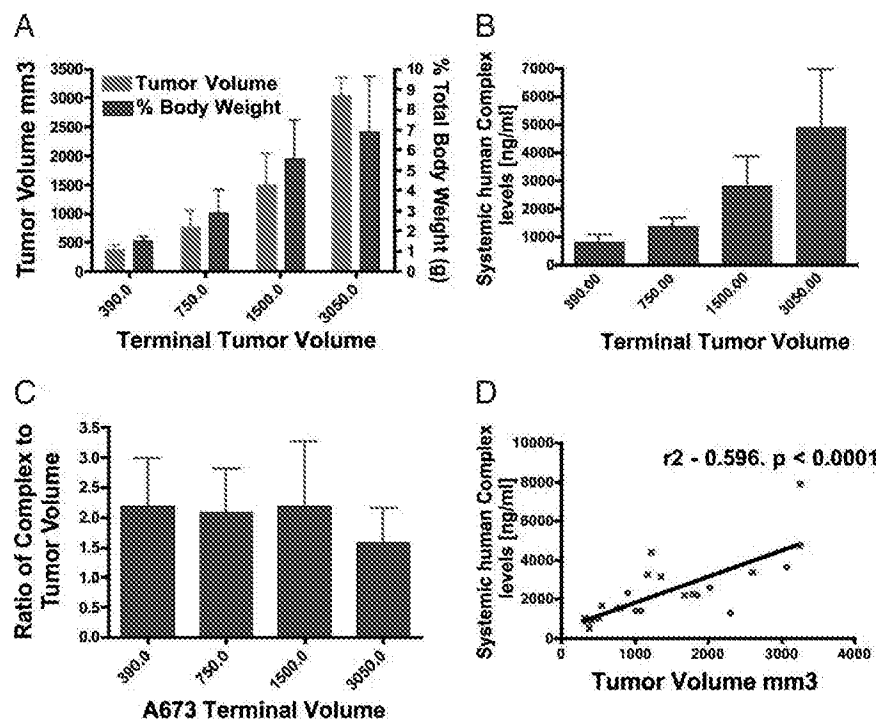


Fig. 5. Human VEGF–VEGF Trap complex levels are directly related to tumor size. Human A673 rhabdomyosarcoma tumors were grown in mice to ≈ 100 , ≈ 300 , ≈ 500 , and ≈ 750 mm³, at which point they were treated with a single bolus of 25 mg/kg. VEGF Trap, tumor volume, and human complex levels were measured after 2 weeks ($n = 6$). (A) Increasing tumor volume equates with an increase in tumor burden. (B) Increasing human tumor burden is reflected in an increase in circulating human VEGF–VEGF Trap complex. (C) The ratio of human VEGF–VEGF Trap complex to tumor volume remains steady at ≈ 2 -fold. (D) Linear regression analysis comparing systemic levels of human VEGF–VEGF Trap to tumor volume reveals that increasing tumor volume directly correlates with increasing complex levels. ($P < 0.0001$.)

vascular tone in the adult (26). We make the unexpected observation that constitutive VEGF production by normal adult tissues is sufficient to mask the lower levels made by most tumors, making it difficult to use peripheral levels of VEGF as a reliable indicator of tumor burden. However, in mice, VEGF production by tumors is clearly related to tumor size, and, when tumors become quite large, the VEGF Trap complex assays readily detect the tumors' VEGF contribution.

Our observations are consistent with recent studies by Bocci *et al.* (27), which reported that plasma VEGF levels are normally very low or undetectable, but are rapidly increased upon treatment with blocking VEGFR2 antibodies. In these experiments, the observed acute increase in circulating VEGF was not associated with increased VEGF expression in normal tissues, or the tumors, but reflected displacement of VEGF from VEGF receptors. It was also noted that maximal VEGF release occurred at antibody doses that produced near optimal anti-tumor effects, suggesting that maximal VEGF receptor blockade was attained. By extension, the induced increases in plasma VEGF could be used to guide dosing of anti-VEGFR antibodies.

The findings reported by Bocci *et al.* also support the notion that, in normal adult tissues, there is substantial basal production of VEGF, which is locally sequestered and thus not readily measured in the periphery unless it is dislodged. However, measurement of VEGF in the circulation after its displacement by anti-VEGFR antibodies cannot account for VEGF sequestered by binding to sites other than VEGFR1 or VEGFR2 (e.g., neuropilins or heparin) and thus cannot be used to calculate total VEGF production rates in host or tumors. Studies with the VEGF Trap, which also displaces tissue bound VEGF, extend these findings by precisely determining and comparing host and tumor production rates of VEGF. We also show that the observations made in mice seem to also apply to humans and that

the levels of VEGF Trap complexed to VEGF can serve as a sensitive guide for the effective dosing of this particular therapeutic candidate. By extension, determination of the dose required to achieve maximum levels of circulating complexes involving a blocker and its target could serve as a useful guide for the dosing of any therapeutic agent that forms long-lived inert circulating complexes with its target.

The sustained circulating levels of VEGF complex observed after VEGF Trap administration is not seen with VEGF-blocking. Unlike the VEGF Trap, which forms an inert 1:1 complex with VEGF that retains the same circulating half-life as unbound VEGF Trap, antibodies to VEGF form heterogeneous multimeric complexes with their antigens, which are cleared much more rapidly than the unbound antibodies. Thus, such immune complexes are not accessible for assays in the systemic circulation, and it is not possible to use systemic levels of such complexes as a guide to VEGF production or to having achieved efficacious antibody levels. Moreover, the formation of such immune complexes could produce undesirable off-mechanism effects. For example, Meyer *et al.* report that bevacizumab forms immune complexes with VEGF that can induce platelet aggregation, which they suggest "might be a possible cause for unexpected arterial thromboembolic events in clinical trials."** In addition, immune complexes can deposit in tissues, including the kidney, potentially contributing to renal damage; consistent with this, we show that VEGF antibodies complexed to VEGF have a much higher propensity to deposit in kidney glomeruli compared with VEGF Trap complexes. Further consistent with this, Gerber *et al.* have reported "anti-VEGF (antibody) depo-

**Meyer, T., Robson, T., Amirhosravi, A., Langer, F., Desai, H., Amaya, M., Elias, P., Francis, J. L. (2007) *Am. Soc. Hematol.* 108:1091 (abstr.).

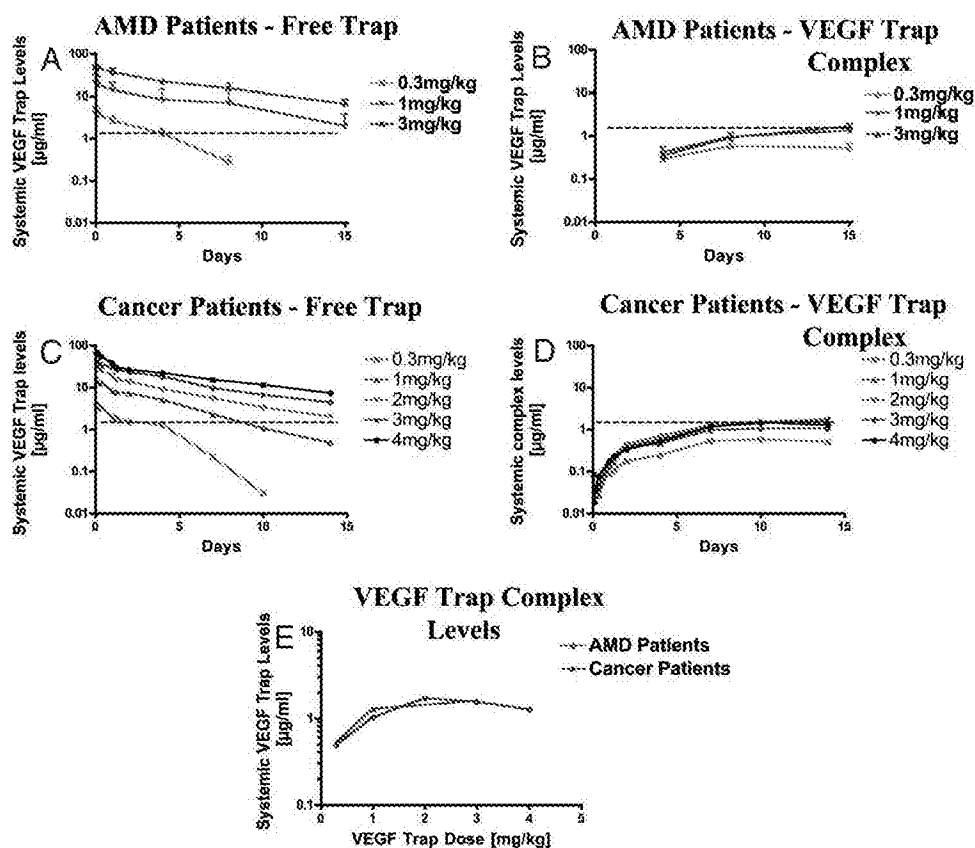


Fig. 6. Circulating free VEGF Trap and human VEGF-VEGF Trap complex levels are very similar in the plasmas of AMD and cancer patients. (A and B) Patients with AMD received a single i.v. bolus of VEGF Trap at 0.3, 1, or 3 mg/kg, and free VEGF Trap and complex levels were measured at 2 and 4 h and 1, 4, 8, and 15 days ($n = 7$, 0.3 mg/kg; $n = 7$, 1 mg/kg; $n = 5$, 3 mg/kg). (C and D) Patients with cancer received a single i.v. bolus of VEGF Trap at 0.3, 1, 2, 3, or 4 mg/kg, and free VEGF Trap and complex levels were measured at 1, 2, 4, and 8 h and 1, 2, 4, 7, 10, and 14 days ($n = 3$, 0.3 mg/kg; $n = 7$, 1 mg/kg; $n = 6$, 2 mg/kg; $n = 5$, 3 mg/kg; $n = 7$, 4 mg/kg). (E) Complex levels in AMD patients at 15 days and cancer patients at 14 days were plotted against the different doses revealing an almost exact overlap. Dotted lines denote the steady-state circulating levels of VEGF-VEGF Trap complex in AMD and cancer patients.

sition in glomeruli" with complement C3 staining and glomerulosclerosis, "which was generally more severe in animals treated with high-affinity mAbs," to VEGF (28). Thus, VEGF Trap, which does not form multimeric immune complexes but instead forms inert 1:1 complexes with VEGF, may not share the same adverse effect profile as anti-VEGF antibodies that can form immune complexes.

In summary, our studies show that assays of free and bound VEGF Trap can serve as useful indicators for the proportion of bioavailable VEGF that is bound and neutralized at a given dose of VEGF Trap. In mice, the majority of endogenous VEGF is captured at doses that result in maximal, steady-state levels of VEGF Trap-VEGF complex, at which point near-optimal efficacy is typically attained. Use of this assay in cancer patients might similarly allow for rapid determination of dosing regimens that are likely to be efficacious. Importantly, application of these assays in early stage clinical trials in patients indicates that the doses currently being evaluated in ongoing clinical studies are in the efficacious range (25, 29–32, ¶).

Materials and Methods

ELISAs. Free VEGF Trap and Complex Measurement. Levels of free VEGF Trap were measured by using a functional ELISA, which uses VEGF₁₆₅ as the capture and an antibody to the IgG2 domain of VEGFR1 as the report. Mouse VEGF-VEGF Trap complex is measured by using an antibody to mouse VEGF as the capture and the same antibody as above as the report. Human VEGF-

VEGF Trap complex is measured by using an antibody to human VEGF as the capture and an antibody to human Fc as the report. **MALLS Coupled to SEC.** A multiangle laser light scattering instrument was coupled to a size exclusion column to measure the molar mass and aggregation of VEGF₁₆₅ bound to VEGF Trap or bevacizumab.

Immunocytochemistry of VEGF Trap/VEGF and Bevacizumab-VEGF Complex Deposition in Kidney Glomeruli. Preformed VEGF₁₆₅-VEGF Trap or VEGF₁₆₅-bevacizumab complexes were injected into the left ventricles of C57bl6 mice, and deposition in the kidney was determined immunocytochemically.

Calculation of VEGF Production Rates Based on Steady-State VEGF-VEGF Trap Complex Levels in Mouse and Man. Endogenous VEGF production rates were determined by using the following equation: Complex production rate [$\mu\text{g/day per ml of volume of distribution (ml-d)}$] = $0.5 \times C_{ss} \mu\text{g/ml per } t_{1/2} \text{ days} = 0.5 \times C_{ss}/t_{1/2} \mu\text{g/ml-d}$. Because VEGF accounts for 1/4 of the mass of the complex, the VEGF production rate ($\mu\text{g per day per ml of volume of distribution}$) = $0.25 \times (0.5 \times C_{ss} \mu\text{g/ml per } t_{1/2} \text{ days}) = 0.125 \times C_{ss}/t_{1/2} \mu\text{g/ml-d}$.

Tumor Implantation. Tumor cell lines were implanted s.c. into the right flank of 7- to 9-week-old male SCID/CB17 mice, and serum samples were taken at termination of the experiment. To assess the relationship between tumor volume and human complex levels, A673 tumors were grown in mice to different sizes, at

which point the mice were treated with a single bolus of 25 mg/kg VEGF Trap, and tumor volume and mouse and human complex levels measured over a 2-week period.

Human Clinical Trials. Clinical trial design for the studies presented herein are available in refs. 25, 29, and 31–34.

Data Analysis. Linear regression analysis comparing circulating human VEGF–VEGF Trap complex levels with tumor volume was done by using the data analysis package in GraphPad Prism. Pharmacokinetic analyses were done by using the WinNonlin PK/PD modeling and analysis package (Pharsight, Mountain

View, CA). Molar masses of proteins and their complexes were determined by using ASTRA software (Wyatt Technology, Santa Barbara, CA) as described in ref. 35.

Additional Details. For a more detailed description of the methods, see *SI Materials and Methods*.

We thank the following Regeneron colleagues: Alain Thibaut, Robert Terifay, Jesse M. Cedarbaum, Chris Daly, Ella Ioffe, Thomas Daly, Douglas McDonald, Nicholas Gale, Samuel Davis, and Len Schleifer and our Sanofi-Aventis colleagues Paul Juniewicz, Marie-Christine Bissery, Friedhelm Blatt, and Marielle Chiron for valuable scientific and editorial input.

1. Ferrara N (2002) *Nat Rev Cancer* 2:795–803.
2. Folkman J (1971) *N Engl J Med* 285:1182–1186.
3. Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N (1993) *Nature* 362:841–844.
4. Yang JC, Haworth L, Sherry RM, Hwu P, Schwartzentruber DJ, Topalian SL, Steinberg SM, Chen HX, Rosenberg SA (2003) *N Engl J Med* 349:427–434.
5. Yang JC (2004) *Clin Cancer Res* 10:6367S–6370S.
6. Hurwitz H, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, Berlin J, Baron A, Griffing S, Holmgren E, et al. (2004) *N Engl J Med* 350:2335–2342.
7. Mass RD, Sarkar S, Holden SN, Hurwitz H (2005) *J Clin Oncol* 23:249S–249S.
8. Jubb AM, Oates AJ, Holden S, Koepfen H (2006) *Nat Rev Cancer* 6:626–635.
9. Quesada AR, Munoz-Chapuli R, Medina MA (2006) *Med Res Rev* 26:483–530.
10. Baker M (2005) *Nat Biotechnol* 23:297–304.
11. Hyodo I, Doi T, Endo H, Hosokawa Y, Nishikawa Y, Tanimizu M, Jinno K, Kotani Y (1998) *Eur J Cancer* 34:2041–2045.
12. Sugimoto H, Hamano Y, Charytan D, Cosgrove D, Kieran M, Sudhakar A, Kalluri R (2003) *J Biol Chem* 278:12605–12608.
13. El-Houseini ME, Abdel-Aziz SA, El-Desouky GI, Abdel-Hady S, El-Hamad MF, Kamel AM (2004) *J Egypt Natl Canc Inst* 16:57–61.
14. Ascierto PA, Leonardi E, Ottaviano A, Napolitano M, Scala S, Castello G (2004) *Anticancer Res* 24:4255–4258.
15. Shimanuki Y, Takahashi K, Cui R, Hori S, Takahashi F, Miyamoto H, Fukuruchi Y (2005) *Lung* 183:29–42.
16. Adams J, Carder PJ, Downey S, Forbes MA, MacLennan K, Allgar V, Kaufman S, Hallam S, Bicknell R, Walker JJ, et al. (2000) *Cancer Res* 60:2898–2905.
17. Jimeno A, Daw NC, Amador ML, Cusatis G, Kulesza P, Krailo M, Ingle AM, Blaney SM, Adamson P, Hidalgo M (2007) *Pediatr Blood Cancer* 49:352–357.
18. Shaked Y, Bocci G, Munoz R, Man S, Ebos JM, Hicklin DJ, Bertolini F, D'Amato R, Kerbel RS (2005) *Curr Cancer Drug Targets* 5:551–559.
19. Bremnes RM, Camps C, Sirena R (2006) *Lung Cancer* 51:143–158.
20. Holash J, Davis S, Papadopoulos N, Croll SD, Ho L, Russell M, Boland P, Leidich R, Hylton D, Burova E, et al. (2002) *Proc Natl Acad Sci USA* 99:11393–11398.
21. Rudge JS, Thurston G, Davis S, Papadopoulos N, Gale N, Wiegand SJ, Yancopoulos GD (2005) *Cold Spring Harbor Symp Quant Biol* 70:411–418.
22. Wachsberger PR, Burd R, Cardi C, Thakur M, Daskalakis C, Holash J, Yancopoulos GD, Dicker AP (2007) *Int J Radiat Oncol Biol Phys* 67:1526–1537.
23. Kamba T, Tam BY, Hashizume H, Haskell A, Sennino B, Mancuso MR, Norberg SM, O'Brien SM, Davis RB, Gowen LC, et al. (2006) *Am J Physiol Heart Circ Physiol* 290:H560–H576.
24. Tam BY, Wei K, Rudge JS, Hoffman J, Holash J, Park SK, Yuan J, Hefner C, Chartier C, Lee JS, et al. (2006) *Nat Med* 12:793–800.
25. Nguyen QD, Shah SM, Hafiz G, Quinlan E, Sung J, Chu K, Cedarbaum JM, Campochiaro PA (2006) *Ophthalmology* 113:1522.e1–1522.e14.
26. Inai T, Mancuso M, Hashizume H, Baffert F, Haskell A, Baluk P, Hu-Lowe DD, Shalinsky DR, Thurston G, Yancopoulos GD, et al. (2004) *Am J Pathol* 165:35–52.
27. Bocci G, Man S, Green SK, Francia G, Ebos JM, du Manoir JM, Weinerman A, Emmenegger U, Ma L, Thorpe P, et al. (2004) *Cancer Res* 64:6616–6625.
28. Gerber H-P, Wu X, Yu L, Wiesmann C, Liang XH, Lee CV, Fuh G, Olsson C, Damico L, Xie D, et al. (2007) *Proc Natl Acad Sci USA* 104:3478–3483.
29. Konner J, Dupont J (2004) *Clin Colorectal Cancer* 4(Suppl 2):S81–S85.
30. Shah SM, Tatlipinar S, Quinlan E, Sung JU, Tabandeh H, Nguyen QD, Fahmy AS, Zimmer-Galler I, Symons RC, Cedarbaum JM, et al. (2006) *Invest Ophthalmol Vis Sci* 47:5460–5468.
31. Dupont J, Schwartz J, Koutcher J, Spriggs D, Gordon M, Mendelson D, Murren J, Lucarelli A, Cedarbaum J (2004) *Proc Am Soc Clin Oncol* 22:3009.
32. Dupont J, Rothenberg ML, Spriggs DR, Cedarbaum JM, Furfine ES, Cohen DP, Dancy I, Lee H, Cooper W, Lockhart AC (2005) *Proc Am Soc Clin Oncol* 23:3029.
33. Tew WP, Colombo N, Ray-Coquard I, Oza A, del Campo J, Scambia G, Spriggs D (2007) *Am Soc Clin Oncol* 25:5508.
34. Massarelli E, Miller VA, Leigh N, Rosen P, Albain K, Hart L, Melnyk O, Sternas L, Akerman J, Herbst RS (2007) in *Am Soc Clin Oncol* 25:7627.
35. Wen J, Arakawa T, Philo JS (1996) *Anal Biochem* 240:155–166.

Intravitreal Aflibercept Injection for Neovascular Age-related Macular Degeneration

Ninety-Six-Week Results of the VIEW Studies

Ursula Schmidt-Erfurth, MD,¹ Peter K. Kaiser, MD,² Jean-François Korobelnik, MD,³ David M. Brown, MD,⁴ Victor Chong, MD,⁵ Quan Dong Nguyen, MD,⁶ Allen C. Ho, MD,⁷ Yuichiro Ogura, MD,⁸ Christian Simader, MD,¹ Glenn J. Jaffe, MD,⁹ Jason S. Slakter, MD,¹⁰ George D. Yancopoulos, MD, PhD,¹¹ Neil Stahl, PhD,¹¹ Robert Vitti, MD,¹¹ Alyson J. Berliner, MD, PhD,¹¹ Yuhwen Soo, PhD,¹¹ Majid Anderesi, MD,¹² Olaf Sowade, MD,¹² Oliver Zeitz, MD,^{12,13} Christiane Norenberg, MS,¹² Rupert Sandbrink, MD, PhD,^{12,14} Jeffrey S. Heier, MD¹⁵

Purpose: To determine efficacy and safety of intravitreal aflibercept in patients with neovascular age-related macular degeneration (AMD) during a second year of variable dosing after a first-year fixed-dosing period.

Design: Two randomized, double-masked, active-controlled, phase 3 trials.

Participants: Two thousand four hundred fifty-seven patients with neovascular AMD.

Methods: From baseline to week 52, patients received 0.5 mg intravitreal ranibizumab every 4 weeks (Rq4), 2 mg aflibercept every 4 weeks (2q4), 0.5 mg aflibercept every 4 weeks (0.5q4), or 2 mg aflibercept every 8 weeks (2q8) after 3 monthly injections. During weeks 52 through 96, patients received their original dosing assignment using an as-needed regimen with defined retreatment criteria and mandatory dosing at least every 12 weeks.

Main Outcome Measures: Proportion of eyes at week 96 that maintained best-corrected visual acuity (BCVA; lost <15 letters from baseline); change from baseline in BCVA.

Results: Proportions of eyes maintaining BCVA across treatments were 94.4% to 96.1% at week 52 and 91.5% to 92.4% at week 96. Mean BCVA gains were 8.3 to 9.3 letters at week 52 and 6.6 to 7.9 letters at week 96. Proportions of eyes without retinal fluid decreased from week 52 (60.3% to 72.4%) to week 96 (44.6% to 54.4%), and more 2q4 eyes were without fluid at weeks 52 and 96 than Rq4 eyes (difference of 10.4% [95% confidence interval {CI}, 4.9–15.9] and 9.0% [95% CI, 3.0–15.1]). Patients received on average 16.5, 16.0, 16.2, and 11.2 injections over 96 weeks and 4.7, 4.1, 4.6, and 4.2 injections during weeks 52 through 96 in the Rq4, 2q4, 0.5q4, and 2q8 groups, respectively. The number of injections during weeks 52 through 96 was lower in the 2q4 and 2q8 groups versus the Rq4 group (differences of –0.64 [95% CI, –0.89 to –0.40] and –0.55 [95% CI, –0.79 to –0.30]; $P < 0.0001$, post hoc analysis). Incidences of Antiplatelet Trialists' Collaboration–defined arterial thromboembolic events were similar across groups (2.4% to 3.8%) from baseline to week 96.

Conclusions: All aflibercept and ranibizumab groups were equally effective in improving BCVA and preventing BCVA loss at 96 weeks. The 2q8 aflibercept group was similar to ranibizumab in visual acuity outcomes during 96 weeks, but with an average of 5 fewer injections. Small losses at 96 weeks in the visual and anatomic gains seen at 52 weeks in all arms were in the range of losses commonly observed with variable dosing. *Ophthalmology* 2014;121:193-201 © 2014 by the American Academy of Ophthalmology.



The introduction of antiangiogenic therapy to treat neovascular age-related macular degeneration (AMD) has vastly changed common paradigms in this important entity usually referred to as “the leading cause of legal blindness in the developed world.”¹ The prospective, masked, randomized, pivotal trials for ranibizumab, called the Anti-Vascular Endothelial Growth Factor (VEGF) Antibody for the Treatment of Predominantly Classic Choroidal

Neovascularization in AMD (ANCHOR) and the Minimally Classic/Occult Trial of the Anti-VEGF Antibody Ranibizumab in the Treatment of Neovascular AMD (MARINA), showed clear superiority of monthly intravitreal ranibizumab administration compared with sham or with the previous gold standard, photodynamic therapy.^{2,3} After approval in 2006 to treat neovascular AMD, intravitreal ranibizumab was embraced quickly by the ophthalmic

community. Odds for visual acuity loss resulting from neovascular AMD markedly decreased with fixed monthly ranibizumab therapy.⁴ Thus, this monthly treatment regimen was included in the ranibizumab Food and Drug Administration label.

Although the visual results in the clinical trials were excellent with the monthly dosing regimens, in clinical practice, the repetitive office visits and injections represent an overwhelming management challenge for patients and their families. Evaluations of actual treatment patterns revealed that most patients were examined and treated far less frequently than recommended by the results of the studies, leading to inferior outcomes.^{5,6} Undertreatment prevents patients from optimally benefitting from one of the major therapeutic breakthroughs in ophthalmology.

To reduce the treatment burden and still conform to a structured regimen, treatment intervals were expanded in studies such as the Phase IIIB, Multicenter, Randomized, Double-Masked, Sham Injection-Controlled Study of the Efficacy and Safety of Ranibizumab in Patients with AMD-Related Subfoveal Choroidal Neovascularization (CNV), with or without Classic CNV (PIER), and the Efficacy and Safety of Monthly versus Quarterly Ranibizumab Treatment in Neovascular Age-Related Macular Degeneration (EXCITE).^{7,8} However, the visual acuity benefit of ranibizumab therapy was reduced markedly when treatment intervals were increased up to 3 months. It was recognized that treatment of recurrence had to take place in a timely manner to prevent functional loss.

Pro re nata (PRN) treatment, or treatment as needed, was evaluated first in the Prospective Optical Coherence Tomography Imaging of Patients with Neovascular AMD Treated with Intra-Ocular Ranibizumab (PRONTO) study, a small, single-center, carefully monitored investigator-driven trial.⁹ Physicians used an optical coherence tomography (OCT)-guided variable-dosing regimen with intravitreal ranibizumab and achieved outcomes comparable with those observed in the phase 3 clinical studies, which used a fixed monthly monitoring and dosing regimen. In contrast, the Open-Label Extension Trial of Ranibizumab for Choroidal Neovascularization Secondary to Age-Related Macular Degeneration (HORIZON) trial, a PRN extension trial after monthly ranibizumab for 2 years, reported that the initial benefit achieved by 2 years of monthly retreatment was lost progressively when switching to a PRN treatment paradigm.¹⁰

The Comparison of Age-Related Macular Degeneration Treatments Trials (CATT) group subsequently designed a large, multicenter, prospective, randomized trial that compared a fixed monthly regimen with a flexible as-needed regimen using the 2 most commonly used anti-VEGF therapies, ranibizumab and bevacizumab.¹¹ Unlike the PRONTO study,⁹ the indication for retreatment in the CATT study was focused strictly on the presence of fluid on OCT, rather than on overall retinal thickness changes; injection was indicated whenever intraretinal, subretinal, or sub-retinal pigment epithelium fluid was identified during monthly OCT monitoring. Although the primary outcome showed noninferiority between ranibizumab and bevacizumab when administered according to similar

regimens, the visual acuity gains and morphologic improvement were greater for the monthly groups as compared with the as-needed groups, especially in year 2.¹¹ To achieve these results, the total number of injections in the as-needed ranibizumab and bevacizumab arms was high: 6.9 and 7.7 injections over the first year and 12.6 and 14.1 injections over 2 years, respectively, with monthly monitoring visits. It is also important to note that the bevacizumab as-needed group did not meet the noninferiority criteria with an as-needed dosing schedule.¹²

Intravitreal aflibercept, a fusion protein of key domains from human VEGF receptors 1 and 2 with the constant region (Fc) of human immunoglobulin G, recently was approved for the treatment of neovascular AMD.¹³ As a designed molecule featuring optimal pharmacologic characteristics to inhibit intraocular VEGF, intravitreal aflibercept injection offers improved binding affinity and superior pharmacokinetics in an iso-osmotic formulation.^{14,15}

The VEGF Trap-Eye: Investigation of Efficacy and Safety in Wet AMD (VIEW 1 and 2) studies were the largest controlled trials of anti-VEGF agents in AMD ever performed, recruiting more than 2400 patients with treatment-naïve neovascular AMD from more than 360 centers worldwide.¹⁶ The focus of the trials was to compare the standard of care (ranibizumab 0.5 mg at monthly intervals) with 2 doses (2 and 0.5 mg) of intravitreal aflibercept and 2 regimens (monthly and every 2 months after 3 initial monthly doses). All intravitreal aflibercept groups were clinically equivalent to monthly ranibizumab in maintaining visual acuity at week 52.¹⁶ This result also was true when drug was administered every 2 months, which allowed a substantially reduced monitoring and treatment frequency, and thus introduced a novel treatment strategy to manage neovascular AMD.¹⁶

After the 52-week primary end point, a follow-up phase of the VIEW trials, up to 96 weeks, was based on a protocol that required a switch of all regimens from the fixed monthly or every 2 months regimen to a variable regimen requiring at least quarterly dosing (capped PRN); interim injections were allowed based on an assessment of anatomic and visual parameters. The aim of the current study was to investigate the safety and efficacy of an extended treatment interval after 1 year of rigorously scheduled fixed treatments. The 96-week data for the integrated VIEW studies describing characteristics and outcomes of a variable dosing regimen are presented and discussed in this article.

Methods

Design

The VIEW 1 and 2 studies were 2 similarly designed randomized, double-masked, active-controlled, parallel-group, multicenter, 96-week phase 3 trials comparing the efficacy and safety of intravitreal aflibercept and ranibizumab in patients with neovascular AMD.¹⁶ The VIEW 1 study was carried out from July 2007 through July 2011 in the United States and Canada, and the VIEW 2 study was carried out from April 2008 through August 2011 in Europe, the Middle East, the Asia-Pacific region, and

Latin America. Patients were screened and/or randomized at 362 sites in the VIEW studies. Each institutional review board or ethics committee approved the study protocols. Both trials were registered with ClinicalTrials.gov (identifier nos. NCT00509795 and NCT00637377), and all patients signed a written consent form before initiation of the study-specific procedures. The VIEW 1 and 2 studies were conducted in compliance with regulations of the Health Insurance Portability and Accountability Act and the tenets of the Declaration of Helsinki.

The design of VIEW studies has been described previously.¹⁶ In brief, patients 50 years of age and older with active, subfoveal, CNV lesions (or juxtafoveal lesions with leakage affecting the fovea) secondary to neovascular AMD were eligible for enrollment if CNV made up at least 50% of total lesion size and BCVA was between 25 and 73 Early Treatment Diabetic Retinopathy Study (ETDRS) letters (20/320–20/40 Snellen equivalent). Only 1 eye from each patient was included in the study. Patients were randomized in a 1:1:1:1 ratio to receive 1 of the following 4 regimens in the study eye for the first 52 weeks: (1) 0.5 mg intravitreal ranibizumab every 4 weeks (Rq4), (2) 2 mg intravitreal aflibercept every 4 weeks (2q4), (3) 0.5 mg intravitreal aflibercept every 4 weeks (0.5q4), and (4) 2 mg intravitreal aflibercept every 8 weeks (2q8) after 3 initial monthly injections. During the follow-up period from weeks 52 to 96, patients continued to receive the same dose of study drugs as in the first 52 weeks, but received injections at least every 12 weeks, with monthly evaluations for interim injections based on prespecified retreatment criteria (mandatory quarterly dosing with examination-guided interim injections or capped-PRN). Criteria for retreatment were new or persistent fluid on OCT, an increase in central retinal thickness of 100 μ m or more compared with the lowest previous value, loss of 5 ETDRS letters or more from the best previous score in conjunction with recurrent fluid on OCT, new-onset classic neovascularization, new or persistent leak on fluorescein angiography, new macular hemorrhage, or a time lapse of 12 weeks since the previous injection.

Outcome Measures

The primary efficacy end point of the VIEW 1 and VIEW 2 studies was noninferiority of the intravitreal aflibercept regimens to ranibizumab in the proportion of patients maintaining visual acuity (losing <15 ETDRS letters) at week 52.¹⁶ Prespecified secondary efficacy end points compared the change among treatment groups in visual acuity and anatomic outcomes from baseline to week 52.¹⁶ Prespecified primary and secondary efficacy outcomes of the VIEW 1 and VIEW 2 studies at week 52 have been reported previously.¹⁶ Efficacy end points evaluated after week 52 all were exploratory and included the proportion of patients maintaining visual acuity (losing <15 ETDRS letters), the mean change in BCVA from baseline, the proportion of patients gaining 15 letters or more, mean change from baseline CNV size, and the proportion of patients without retinal fluid at week 96. The mean change in central retinal thickness also was determined from baseline through week 96. Additional end points during the exploratory follow-up phase were the number of study drug injections and the proportion of patients receiving fewer than 6 injections and 6 injections or more between weeks 52 and 96.

Patients were evaluated for BCVA at screening, at the day of treatment initiation, and every 4 weeks thereafter through week 96, as well as 1 week after the first treatment for safety reasons. In the VIEW 1 study, OCT was performed at screening, at the day of treatment initiation, and at weeks 4, 12, 24, 36, and 52 and every 4 weeks thereafter through week 96. In the VIEW 2 study, OCT was performed at every visit. The OCT images were obtained with

a time-domain Stratus instrument (Carl Zeiss Meditec, Dublin, CA) and was evaluated by an independent central reading center (VIEW 1, Duke Reading Center, Durham, NC; VIEW 2, Vienna Reading Center, Vienna, Austria). Fundus photography and fluorescein angiography were performed at screening and at weeks 24, 52, 72, and 96, and the results were evaluated by an independent central reading center (Digital Angiography Reading Center, New York, NY). Areas of visible active CNV (classic, occult, or both) were identified when angiographic analyses showed evidence of visible neovascular tissue accompanied by late leakage or pooling of dye.

Statistical Analysis

Data from the VIEW 1 and VIEW 2 studies were pooled for the purpose of presentation in this report. The proportion of patients maintaining visual acuity (losing <15 ETDRS letters) at week 52 was analyzed in the per-protocol set as defined previously.¹⁶ The proportion of patients maintaining visual acuity (losing <15 ETDRS letters) at week 96 was analyzed in the full analysis set, which included all randomized patients who received any study medication and had a baseline BCVA measurement and at least 1 BCVA assessment after baseline. All other visual and anatomic end points were analyzed in the full analysis set. The last-observation-carried-forward approach was used to impute missing data. Safety end points at weeks 52 and 96 were analyzed in the safety analysis set, which included all patients who received any study medication. Treatment experience over the 2 years of study was analyzed in the safety analysis set. Treatment experience in the second year was analyzed in patients who completed study treatments. Between-group differences in the number of injections from weeks 52 to 96 were analyzed with an analysis of variance in a post hoc analysis.

Results

Patient Disposition and Baseline Characteristics

The VIEW 1 and 2 studies randomized a total of 2457 patients; 2419 (98.5%) patients received at least 1 dose of study medication, and 2245 (91.4%) patients completed 52 weeks of study. A total of 2235 (91.0%) patients entered the second year, and 2063 (84.0%) patients completed 96 weeks of study. The percentage of patients completing the study was similar among treatment groups at both weeks 52 and 96 (Table 1, available at <http://aaajournal.org>). Reasons for discontinuation before week 96 included consent withdrawal occurring in 5.0% to 6.5% of patients and adverse events occurring in 2.6% to 4.9% of patients across treatment groups (Table 1, available at <http://aaajournal.org>). Baseline demographics and disease characteristics were evenly balanced among all treatment groups (Table 2).

Efficacy

The proportion of patients maintaining visual acuity ranged from 94.4% to 96.1% at week 52 (Fig 1A). Both monthly and every 2 months intravitreal aflibercept regimens were statistically noninferior (with a margin within 5%) to monthly ranibizumab at week 52 (mean of Rq4 minus intravitreal aflibercept, $-0.9%$ [95% confidence interval (CI), -3.5 to 1.7] for 2q4; $-1.7%$ [95% CI, -4.2 to 0.9] for 0.5q4; and $-0.9%$ [95% CI, -3.5 to 1.7] for 2q8). Largely similar proportions of patients (91.5% to 92.4%) maintained visual acuity across all treatment groups at week 96 (Fig 1A). The mean increase in BCVA from baseline was largely similar among treatment groups throughout the 96 weeks of the study (Fig 1B). At week 96, the mean BCVA gains were 7.9 letters, 7.6 letters, 6.6 letters, and 7.6 letters in the Rq4,

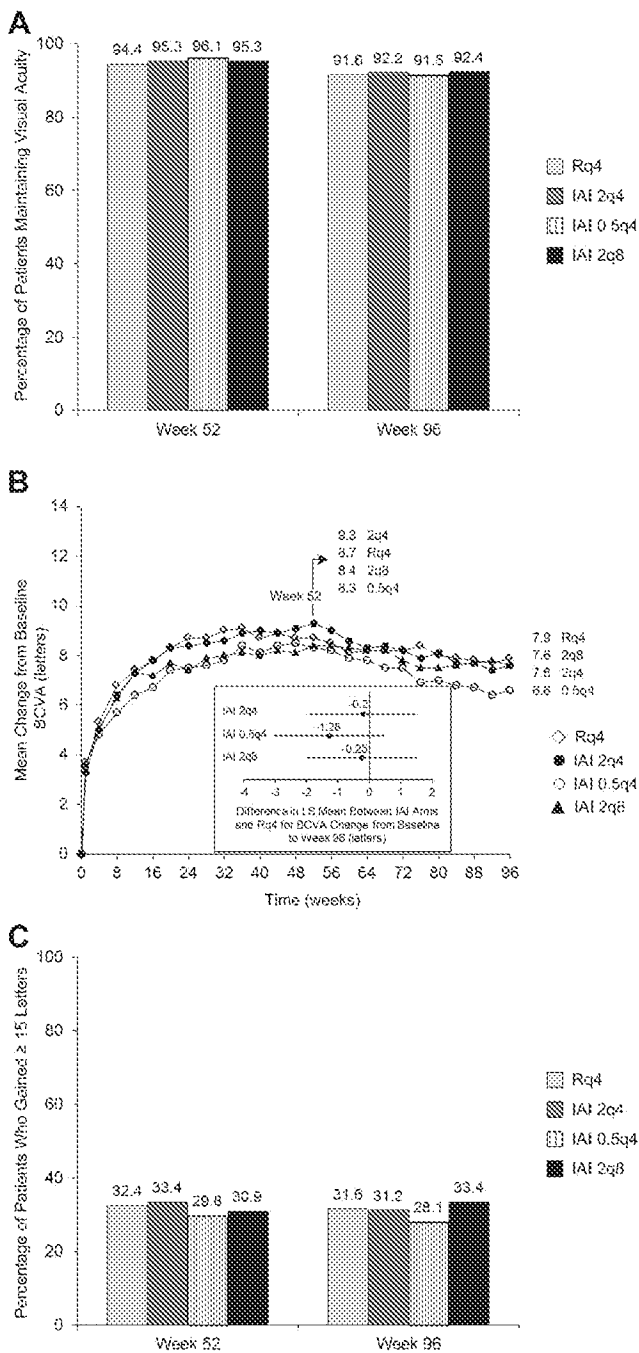


Figure 1. Graphs showing visual acuity outcomes in the total study cohort. **A**, Proportion of patients maintaining visual acuity (losing <15 Early Treatment Diabetic Retinopathy Study letters). Per-protocol and full analysis sets were used for weeks 52 and 96, respectively. At week 52, n = 538, n = 559, n = 538, and n = 535 for Rq4, 2q4, 0.5q4, and 2q8, respectively. At week 96, n = 595, n = 613, n = 597, and n = 607 for Rq4, 2q4, 0.5q4, and 2q8, respectively. **B**, Mean change from baseline best-corrected visual acuity (BCVA). The inset shows the difference in least square (LS) mean (with 95% confidence interval) between intravitreal aflibercept arms and ranibizumab (aflibercept minus ranibizumab) for BCVA change from baseline to week 96, full analysis set. **C**, Proportion of patients who gained 15 letters or more, full analysis set. At weeks 52 and 96, n = 595, n = 613, n = 597, and n = 607 for Rq4, 2q4, 0.5q4, and 2q8, respectively. Missing values were imputed using the

2q4, 0.5q4, and 2q8 groups, respectively; these gains represented a 1- to 2-letter loss in all groups during the capped PRN (modified quarterly dosing) phase, compared with the gains observed at week 52 (8.7, 9.3, 8.3, and 8.4 letters, respectively). Overall, 29.8% to 33.4% of patients in all treatment groups gained 15 letters or more from baseline to week 52 (Fig 1C). The proportions of patients who gained 15 letters or more from baseline to week 96 were similar and ranged from 28.1% to 33.4% (Fig 1C). Across treatment groups, largely similar proportions of patients had a BCVA of 20/40 or better or had an improvement from baseline BCVA of 0 letters or more, 10 letters or more, and 30 letters or more at weeks 52 and 96 (Table 3, available at <http://aaojournal.org>).

During the capped PRN phase (requiring at least quarterly dosing), there was a minor loss in the anatomic improvements that had been seen at week 52. At week 96, patients had an average increase in central retinal thickness of 10 μm, 10 μm, 10 μm, and 6 μm from week 52 in the Rq4, 2q4, 0.5q4, and 2q8 groups, respectively (Fig 2A). The proportion of patients with no retinal fluid on time-domain OCT (observed cases) ranged from 60.3% to 72.4% at week 52, with higher percentages of 2q4 and 2q8 patients having no retinal fluid compared with Rq4 patients (mean of aflibercept minus Rq4, 10.4% [95% CI, 4.9–15.9] for 2q4 and 5.7% [95% CI, 0–11.4] for 2q8). The percentage of patients with no retinal fluid decreased from week 52 to week 96 in all treatment groups. Nevertheless, a higher percentage of 2q4 patients had no retinal fluid at week 96 compared with Rq4 patients (mean of 2q4 minus Rq4, 9.0% [95% CI, 3.0–15.1]; Fig 2B). In contrast, the mean decreases in CNV area were maintained from week 52 (range, 3.9–5.3 mm²) to week 96 (range, 3.7–5.1 mm²) in all treatment groups. A lower CNV area was observed at week 52 for 2q4 in comparison with Rq4 (least squares mean of 2q4 minus Rq4, -0.74 mm² [95% CI, -1.27 to -0.21]), but was not maintained at week 96.

Number of Injections

The mean number of injections from week 0 to week 96 was 16.5 (standard deviation [SD], 3.7), 16.0 (SD, 3.2), 16.2 (SD, 4.0), and 11.2 (SD, 2.9) in the Rq4, 2q4, 0.5q4, and 2q8 groups, respectively. The mean number of injections from week 52 to week 96 was 4.7 (SD, 2.2), 4.1 (SD, 1.8), 4.6 (SD, 2.2), and 4.2 (SD, 1.7) in the Rq4, 2q4, 0.5q4, and 2q8 groups, respectively. In a post hoc analysis, this number of injections from week 52 to week 96 was lower in the 2q4 and 2q8 groups versus the Rq4 group: mean of aflibercept minus Rq4, -0.64 (95% CI, -0.89 to -0.40) for 2q4 and -0.55 (95% CI, -0.79 to -0.30) for 2q8 (P < 0.0001 for both). The proportion of patients who received fewer than 6 injections and 6 injections or more during weeks 52 to 96 are shown in Figure 3A. Overall, higher percentages of 2q4 and 2q8 patients received fewer than 6 injections compared with Rq4 patients, whereas a higher percentage of Rq4 patients received 6 injections or more compared with 2q4 and 2q8 patients (Fig 3A, B).

Safety

Safety profiles of both intravitreal aflibercept and ranibizumab were favorable. Ocular adverse events occurring in 10% or more of

last-observation-carried-forward method in (A), (B), and (C). The outcomes for the aflibercept and ranibizumab groups were similar in (A), (B), and (C) at both weeks 52 and 96. IAI = intravitreal aflibercept injection; Rq4 = 0.5-mg intravitreal ranibizumab every 4 weeks; 2q4 = 2 mg every 4 weeks; 0.5q4 = 0.5 mg every 4 weeks; 2q8 = 2 mg every 8 weeks after 3 initial monthly injections.

Table 2. Patient Demographics and Baseline Characteristics, Full Analysis Set

	0.5 mg Intravitreal Ranibizumab Every 4 Weeks (n = 595)	Intravitreal Aflibercept Injection 2 mg Every 4 Weeks (n = 613)	Intravitreal Aflibercept Injection 0.5 mg Every 4 Weeks (n = 597)	Intravitreal Aflibercept Injection 2 mg Every 8 Weeks after 3 Initial Monthly Injections (n = 607)
Female, n (%)	341 (57.3)	370 (60.4)	314 (52.6)	353 (58.2)
Race, n (%)				
White	509 (85.5)	521 (85.0)	510 (85.4)	504 (83.0)
Asian	60 (10.1)	70 (11.4)	66 (11.1)	73 (12.0)
Other*	26 (4.4)	22 (3.6)	21 (3.5)	30 (5.0)
Age (SD), yrs	75.6 (8.7)	75.9 (8.4)	76.5 (8.5)	75.8 (8.8)
BCVA (SD), letters	53.9 (13.4)	54.0 (13.6)	53.6 (13.8)	53.6 (13.5)
Central retinal thickness (SD), μm	296 (123) [†]	299 (126) [†]	296 (132) [†]	306 (134) [§]
Area of CNV (SD), mm^2	7.1 (5.3)	7.4 (5.5) [¶]	7.1 (4.9) [*]	7.2 (5.4) ^{**}
Type of CNV, n (%)				
Minimally classic	205 (34.5)	217 (35.4)	200 (33.5)	216 (35.6)
Occult	231 (38.8)	233 (38.0)	234 (39.2)	228 (37.6)
Predominantly classic	152 (25.5)	159 (25.9)	161 (27.0)	159 (26.2)
Missing	7 (1.2)	4 (0.7)	2 (0.3)	4 (0.7)
Total lesion size (SD), mm^2	7.5 (5.6)	7.9 (5.8) [¶]	7.5 (5.2) [*]	7.6 (5.6) ^{**}
Total NEI VFQ score (SD)	72.4 (18.1) ^{††}	70.3 (18.1) ^{††}	72.6 (18.0) ^{††}	70.4 (18.0) ^{§§}

BCVA = best-corrected visual acuity; CNV = choroidal neovascularization; NEI VFQ = National Eye Institute Visual Function Questionnaire; SD = standard deviation.

*Included American Indian or Alaska Native, Black or African American, Native Hawaiian or other Pacific Islander, multiracial patients, and those who did not report their race.

[†]n = 594.

[‡]n = 611.

[§]n = 603.

^{||}n = 589.

[¶]n = 610.

^{*}n = 596.

^{**}n = 605.

^{††}n = 609.

^{††}n = 592.

^{§§}n = 599.

patients across treatment groups were conjunctival hemorrhage (range, 21.7%–28.1%) and eye pain (range, 7.0%–10.8%) from baseline to week 52, and conjunctival hemorrhage (range, 23.7%–29.9%), retinal hemorrhage (range, 13.6%–16.2%), reduced visual acuity (range, 11.3%–13.0%), eye pain (range, 8.9%–12.1%), vitreous detachment (range, 7.7%–10.0%), and increased intraocular pressure (range, 6.2%–10.8%) from baseline to week 96. Any intraocular inflammatory response (predefined adverse event of interest) was reported in 0.8%, 0.7%, 0.3%, and 0.2% of patients from baseline to week 52 and in 1.5%, 1.1%, 0.8%, and 0.5% of patients from baseline to week 96 in the Rq4, 2q4, 0.5q4, and 2q8 groups, respectively. Serious ocular adverse events were infrequent and occurred with a similar rate across all treatment groups (Table 4). Major serious systemic adverse events were fall and pneumonia from baseline to week 52, and fall, pneumonia, atrial fibrillation, and myocardial infarction from baseline to week 96 (Table 5, available at <http://aaojournal.org>). In general, serious systemic adverse events were typical of those reported in this population of elderly patients who receive intravitreal treatment for neovascular AMD. The incidence of arterial thromboembolic events as defined by the Antiplatelet Trialists' Collaboration criteria was similar among treatment groups from both baseline to week 52 and from baseline to week 96 (Table 6). The percentage of deaths was 1.2%, 0.7%, 0.5%, and 1.5% from baseline to week 52 and 2.7%, 2.1%, 3.2%, and 3.3% from baseline to week 96 in the Rq4, 2q4, 0.5q4, and 2q8 groups,

respectively. The incidences and patterns of deaths were not different among treatment groups.

Discussion

The results from the follow-up regimen of mandatory quarterly dosing with intervening as-needed injections (capped PRN) in the second year of the VIEW studies confirm the sustained improvements in visual acuity, central retinal thickness, and CNV size achieved by fixed dosing regimens of intravitreal aflibercept and ranibizumab during the first year.¹⁶ All intravitreal aflibercept regimens were as effective as ranibizumab in increasing visual acuity and reducing retinal thickness and CNV size over 2 years of the VIEW studies. Small decreases in visual and anatomic improvements from week 52 to 96 were observed in all treatment groups, similar to declines seen in other randomized clinical trials when switching to treatment regimens with a variable component.¹¹ Of note was a decrease in the proportion of patients with no retinal fluid from week 52 to 96 after switching to a more variable dosing regimen in all treatment groups. Nevertheless, more patients in the 2q4 group had no

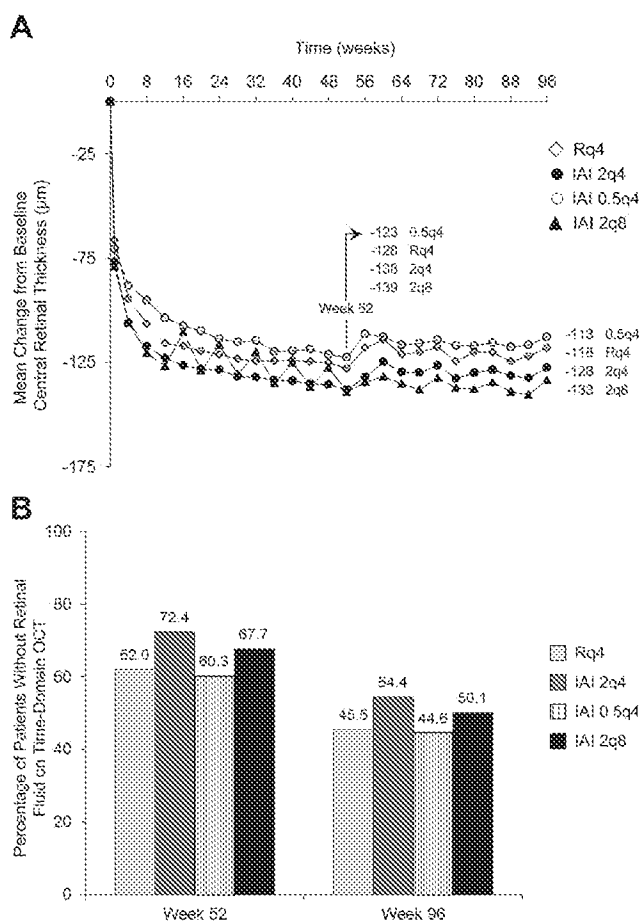


Figure 2. Graphs showing anatomic outcomes in total study cohort. **A**, Mean change from baseline central retinal thickness, full analysis set. Missing values were imputed using the last-observation-carried-forward method. The outcomes for the aflibercept and ranibizumab groups were similar at both weeks 52 and 96. **B**, Proportion of patients without fluid on time-domain optical coherence tomography (OCT) images. Observed values in full analysis set. Number of patients included in the Rq4, 2q4, 0.5q4, and 2q8 groups were 537, 558, 527, and 539 at week 52, and 508, 522, 493, and 505 at week 96, respectively. IAi = intravitreal aflibercept injection; Rq4 = 0.5-mg intravitreal ranibizumab every 4 weeks; 2q4 = 2 mg every 4 weeks; 0.5q4 = 0.5 mg every 4 weeks; 2q8 = 2 mg every 8 weeks after 3 initial monthly injections.

retinal fluid at week 96, as did both the 2q4 and 2q8 groups at week 52, compared with the Rq4 group. Subtle decreases in the visual and anatomic improvements from week 52 to 96 are likely the result of the variable dosing regimen used. A fixed dosing regimen may provide predictable visual and anatomic outcomes and may mitigate loss of visual and anatomic improvements.

Patients in the 2q8 group achieved visual and anatomic improvements similar to those in the Rq4 and 2q4 groups, but with a mean of 5 fewer injections over 2 years. The significantly fewer average number of injections (post hoc analysis) in the follow-up phase in both 2q4 and 2q8 groups compared with the Rq4 group was driven by more patients in the Rq4 arm receiving the most intense therapy (≥ 6 injections; 14.0% and 15.9% vs. 26.5%, respectively). These findings suggest that patients with greater disease

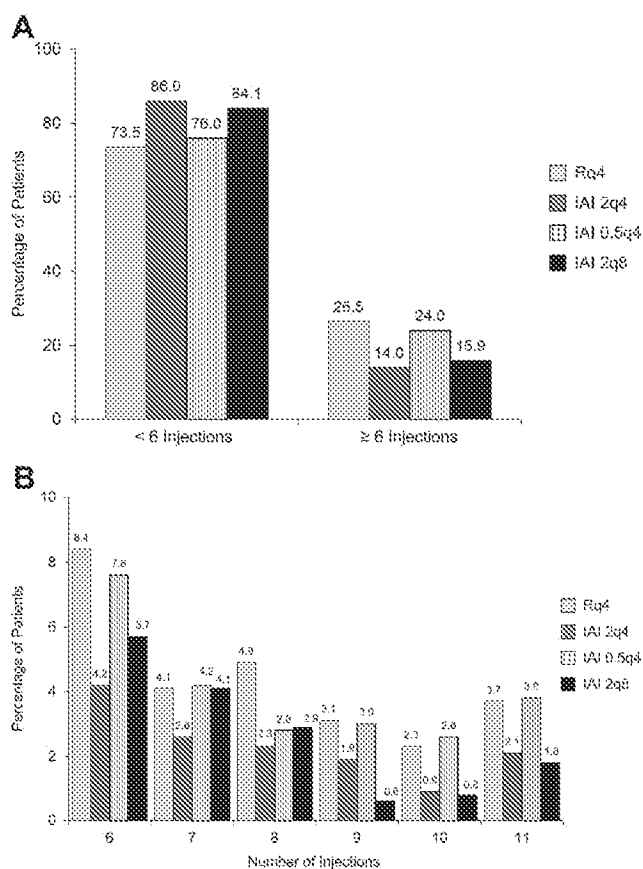


Figure 3. Graphs showing proportion of patients by the number of injections during weeks 52 to 96. **A**, Percentages of patients who received fewer than 6 injections and 6 or more injections. **B**, Percentage of patients who received 6 to 11 injections. The maximum number of injections was 11 in all treatment groups. Patients who completed the second year follow-up phase medications were included in the analyses shown in (A) and (B). Number of patients included in the Rq4, 2q4, 0.5q4, and 2q8 groups were 513, 529, 499, and 511, respectively, in (A) and (B). IAi = intravitreal aflibercept injection; Rq4 = 0.5-mg intravitreal ranibizumab every 4 weeks; 2q4 = 2 mg every 4 weeks; 0.5q4 = 0.5 mg every 4 weeks; 2q8 = 2 mg every 8 weeks after 3 initial monthly injections.

activity may require fewer injections using intravitreal aflibercept.

Over the 2 years of treatment, a generally favorable safety profile was observed for both intravitreal aflibercept and ranibizumab. No unexpected safety signals were observed with intravitreal aflibercept. The incidence of ocular treatment-emergent adverse events was balanced across all treatment groups, with the most frequent events associated with the injection procedure, the underlying disease, the aging process, or a combination thereof. The incidences of arterial thromboembolic events and death were similar across all treatment groups.

At the time the VIEW studies were designed, the efficacy of variable regimens of anti-VEGF agents was being evaluated as a recommended standard of care in several studies. Current clinical evidence shows that variable regimens, which are unpredictable and require monthly monitoring, are less effective to maintain visual and anatomic improvements gained by fixed dosing regimens. Debate

Table 4. Serious Ocular Adverse Events in the Study Eye Occurring in More Than 1 Patient in Any Treatment Group, Safety Analysis Set

Serious Adverse Event	Baseline to Week 52				Baseline to Week 96			
	0.5 mg Intravitreal Ranibizumab Every 4 Weeks (n = 595)	2 mg Intravitreal Aflibercept Injection Every 4 Weeks (n = 613)	0.5 mg Intravitreal Aflibercept Injection Every 4 Weeks (n = 601)	2 mg Intravitreal Aflibercept Injection Every 8 Weeks after 3 Initial Monthly Injections (n = 610)	0.5 mg Intravitreal Ranibizumab Every 4 Weeks (n = 595)	2 mg Intravitreal Aflibercept Injection Every 4 Weeks (n = 613)	0.5 mg Intravitreal Aflibercept Injection Every 4 Weeks (n = 601)	2 mg Intravitreal Aflibercept Injection Every 8 Weeks after 3 Initial Monthly Injections (n = 610)
Total patients with at least 1 ocular SAE, n (%)	19 (3.2)	13 (2.1)	11 (1.8)	12 (2.0)	26 (4.4)	22 (3.6)	19 (3.2)	24 (3.9)
Macular hole	0	0	2 (0.3)	0	0	0	2 (0.3)	0
Posterior capsule opacification	2 (0.3)	0	0	0	2 (0.3)	0	0	0
Retinal detachment	1 (0.2)	0	2 (0.3)	0	3 (0.5)	1 (0.2)	2 (0.3)	0
Retinal hemorrhage	3 (0.5)	2 (0.3)	1 (0.2)	3 (0.5)	4 (0.7)	3 (0.5)	5 (0.8)	5 (0.8)
Retinal pigment epithelial tear	1 (0.2)	0	1 (0.2)	2 (0.3)	1 (0.2)	0	1 (0.2)	3 (0.5)
Reduced visual acuity	3 (0.5)	2 (0.3)	3 (0.5)	5 (0.8)	5 (0.8)	4 (0.7)	3 (0.5)	7 (1.1)
Endophthalmitis	3 (0.5)	3 (0.5)	0	0	5 (0.8)	4 (0.7)	1 (0.2)	0
Cataract	1 (0.2)	1 (0.2)	1 (0.2)	1 (0.2)	1 (0.2)	4 (0.7)	3 (0.5)	4 (0.7)
Macular degeneration	0	0	0	1 (0.2)	0	0	0	2 (0.3)
Increased intraocular pressure	1 (0.2)	0	1 (0.2)	1 (0.2)	1 (0.2)	0	1 (0.2)	2 (0.3)

SAE = serious adverse event.

continues as to whether the losses in visual acuity are offset by the reductions in treatment and monitoring burden, especially if monitoring is not maintained at the monthly frequency mandated in our study and in the CATT study. Proponents of the treat-and-extend regimen, a treatment

strategy involving gradual extension of the treatment and monitoring intervals after initially treating monthly until the macula is dry, suggest that this regimen may result in visual acuity outcomes similar to those seen with monthly therapy, but this has not been demonstrated in a large, randomized

Table 6. Antiplatelet Trialists' Collaboration-Defined Arterial Thromboembolic Events, Safety Analysis Set

Any APTC event, n (%)	Baseline to Week 52					Baseline to Week 96				
	0.5 mg Intravitreal Ranibizumab Every 4 Weeks (n = 595)	2 mg Intravitreal Aflibercept Injection Every 4 Weeks (n = 613)	0.5 mg Intravitreal Aflibercept Injection Every 4 Weeks (n = 601)	2 mg Intravitreal Aflibercept Injection Every 8 Weeks after 3 Initial Monthly Injections (n = 610)	All Intravitreal Aflibercept Injections (n = 1824)	0.5 mg Intravitreal Ranibizumab Every 4 Weeks (n = 595)	2 mg Intravitreal Aflibercept Injection Every 4 Weeks (n = 613)	0.5 mg Intravitreal Aflibercept Injection Every 4 Weeks (n = 601)	2 mg Intravitreal Aflibercept Injection Every 8 Weeks after 3 Initial Monthly Injections (n = 610)	All Intravitreal Aflibercept Injections (n = 1824)
Any APTC event, n (%)	9 (1.5)	6 (1.0)	12 (2.0)	14 (2.3)	32 (1.8)	19 (3.2)	15 (2.4)	23 (3.8)	22 (3.6)	60 (3.3)
Nonfatal MI	6 (1.0)	3 (0.5)	6 (1.0)	6 (1.0)	15 (0.8)	12 (2.0)	6 (1.0)	12 (2.0)	7 (1.1)	25 (1.4)
Nonfatal stroke	1 (0.2)	2 (0.3)	3 (0.5)	3 (0.5)	8 (0.4)	5 (0.8)	5 (0.8)	3 (0.5)	5 (0.8)	13 (0.7)
Vascular death	2 (0.3)	1 (0.2)	3 (0.5)	5 (0.8)	9 (0.5)	3 (0.5)	5 (0.8)	8 (1.3)	11 (1.8)	24 (1.3)

APTC = Antiplatelet Trialists' Collaboration; MI = myocardial infarction.

clinical trial.^{17,18} The 1-year outcomes of the VIEW studies demonstrate that the average patient can obtain results clinically equivalent to monthly ranibizumab with 2 mg intravitreal aflibercept administered every 8 weeks after 3 initial monthly injections.¹⁶ It is conceivable that a continuation of the every-2-months fixed-dosing regimen using intravitreal aflibercept into the second year would have maintained more effectively the visual and anatomic improvements achieved during the first year. Such a fixed-dosing regimen thus would allow for better outcomes with a substantially lower number of monitoring visits. In addition, a fixed, every-2-months dosing regimen with aflibercept (requiring 5 injections) would approximate the 4.2 injections given with the capped PRN (modified quarterly dosing) regimen in the second year of the VIEW studies. Future studies may shed additional light on the benefit of continuing with an every-2-months fixed-dosing regimen instead of using variable dosing regimens.

Acknowledgments. The authors thank Hadi Moini, PhD, and S. Balachandra Dass, PhD, of Regeneron Pharmaceuticals, Inc, for editorial and administrative assistance to the authors who wrote the manuscript.

References

1. Eye Diseases Prevalence Research Group. Causes and prevalence of visual impairment among adults in the United States. *Arch Ophthalmol* 2004;122:477–85.
2. Brown DM, Kaiser PK, Michels M, et al; ANCHOR Study Group. Ranibizumab versus verteporfin for neovascular age-related macular degeneration. *N Engl J Med* 2006;355:1432–44.
3. Rosenfeld PJ, Brown DM, Heier JS, et al; MARINA Study Group. Ranibizumab for neovascular age-related macular degeneration. *N Engl J Med* 2006;355:1419–31.
4. Bressler NM, Doan QV, Varma R, et al. Estimated cases of legal blindness and visual impairment avoided using ranibizumab for choroidal neovascularization: non-Hispanic white population in the United States with age-related macular degeneration. *Arch Ophthalmol* 2011;129:709–17.
5. Cohen SY, Dubois L, Tadayoni R, et al. Results of one-year's treatment with ranibizumab for exudative age-related macular degeneration in a clinical setting. *Am J Ophthalmol* 2009;148:409–13.
6. Dadgostar H, Ventura AA, Chung JY, et al. Evaluation of injection frequency and visual acuity outcomes for ranibizumab monotherapy in exudative age-related macular degeneration. *Ophthalmology* 2009;116:1740–7.
7. Regillo CD, Brown DM, Abraham P, et al; PIER Study Group. Randomized, double-masked, sham-controlled trial of ranibizumab for neovascular age-related macular degeneration: PIER Study year 1. *Am J Ophthalmol* 2008;145:239–48.
8. Schmidt-Erfurth U, Eldem B, Guymer R, et al; EXCITE Study Group. Efficacy and safety of monthly versus quarterly ranibizumab treatment in neovascular age-related macular degeneration: the EXCITE study. *Ophthalmology* 2011;118:831–9.
9. Lalwani GA, Rosenfeld PJ, Fung AE, et al. A variable-dosing regimen with intravitreal ranibizumab for neovascular age-related macular degeneration: year 2 of the PrONTO study. *Am J Ophthalmol* 2009;148:43–58.
10. Singer MA, Awh CC, Sadda S, et al. HORIZON: an open-label extension trial of ranibizumab for choroidal neovascularization secondary to age-related macular degeneration. *Ophthalmology* 2012;119:1175–83.
11. Comparison of Age-related Macular Degeneration Treatments Trials (CATT) Research Group, Martin DF, Maguire MG, Fine SL, et al. Ranibizumab and bevacizumab for treatment of neovascular age-related macular degeneration: two-year results. *Ophthalmology* 2012;119:1388–98.
12. CATT Research Group. Ranibizumab and bevacizumab for neovascular age-related macular degeneration. *N Engl J Med* 2011;364:1897–908.
13. Papadopoulos N, Martin J, Ruan Q, et al. Binding and neutralization of vascular endothelial growth factor (VEGF) and related ligands by VEGF Trap, ranibizumab and bevacizumab. *Angiogenesis* 2012;15:171–85.
14. Holash J, Davis S, Papadopoulos N, et al. VEGF-Trap: a VEGF blocker with potent antitumor effects. *Proc Natl Acad Sci U S A* 2002;99:11393–8.
15. Stewart MW, Rosenfeld PJ, Penha FM, et al. Pharmacokinetic rationale for dosing every 2 weeks versus 4 weeks with intravitreal ranibizumab, bevacizumab, and aflibercept (vascular endothelial growth factor Trap-eye). *Retina* 2012;32:434–57.
16. Heier JS, Brown DM, Chong V, et al; VIEW 1 and VIEW 2 Study Groups. Intravitreal aflibercept (VEGF Trap-Eye) in wet age-related macular degeneration. *Ophthalmology* 2012;119:2537–48.
17. Engelbert M, Zweifel SA, Freund KB. Long-term follow-up for type 1 (subretinal pigment epithelium) neovascularization using a modified “treat and extend” dosing regimen of intravitreal antivascular endothelial growth factor therapy. *Retina* 2010;30:1368–75.
18. Gupta OP, Shienbaum G, Patel AH, et al. A treat and extend regimen using ranibizumab for neovascular age-related macular degeneration clinical and economic impact. *Ophthalmology* 2010;117:2134–40.

Footnotes and Financial Disclosures

Originally received: May 15, 2013.

Final revision: July 18, 2013.

Accepted: August 8, 2013.

Available online: September 30, 2013. Manuscript no. 2013-782.

¹ Department of Ophthalmology, Medical University of Vienna, Vienna, Austria.

² Department of Ophthalmology, Cole Eye Institute, Cleveland, Ohio.

³ Department of Ophthalmology, Centre Hospitalier Universitaire de Bordeaux, Université Bordeaux 2, Bordeaux, France.

⁴ Retina Consultants of Houston, Houston, Texas.

⁵ Oxford Eye Hospital, University of Oxford, Oxford, United Kingdom.

⁶ Wilmer Eye Institute, Johns Hopkins University, Baltimore, Maryland.

⁷ Wills Eye Hospital and Mid Atlantic Retina, Philadelphia, Pennsylvania.

⁸ Department of Ophthalmology, Nagoya City University, Nagoya, Japan.

⁹ Department of Ophthalmology, Duke University, Durham, North Carolina.

¹⁰ Vitreous-Retina-Macula Consultants of New York, New York, New York.

¹¹ Regeneron Pharmaceuticals, Inc, Tarrytown, New York.

¹² Bayer HealthCare, Berlin, Germany.

¹³ Universitätsklinikum Hamburg-Eppendorf, Klinik und Poliklinik für Augenheilkunde, Hamburg, Germany.

¹⁴ Department of Neurology, Heinrich-Heine-Universität Düsseldorf, Düsseldorf, Germany.

¹⁵ Ophthalmic Consultants of Boston and Tufts University School of Medicine, Boston, Massachusetts.

Presented at: American Academy of Ophthalmology Annual Meeting, November 2012.

Financial Disclosure(s):

The author(s) have made the following disclosure(s): Ursula Schmidt-Erfurth: Consultant – Bayer HealthCare, Alcon, Allergan, Boehringer, Novartis; Financial support – Bayer HealthCare; Advisory board – Alcon, Allergan, Boehringer, Novartis; Lecturer – Alcon, Allergan, Boehringer, Novartis

Peter K. Kaiser: Consultant – Alcon, Bayer HealthCare, Genentech, Novartis, Regeneron Pharmaceuticals

Jean-Francois Korobelnik: Consultant – Bayer HealthCare, Carl Zeiss Meditec, Novartis, Roche, Thea; Advisory board – Alcon, Allergan; Financial support – Regeneron Pharmaceuticals

David M. Brown: Consultant and Financial support – Alcon, Allergan, Bayer HealthCare, Genentech/Roche, Novartis, Regeneron Pharmaceuticals, Thrombogenics

Victor Chong: Consultant – Allergan, Bayer HealthCare, Novartis, Quantel; Financial support – Allergan, Novartis, Bayer HealthCare; Lecturer – Bayer HealthCare, Heidelberg, Novartis; Travel – Bayer Healthcare.

Quan Dong Nguyen: Consultant – Bausch & Lomb, Santen; Financial support – Genentech, Pfizer, Regeneron Pharmaceuticals

Allen C. Ho: Consultant – Regeneron Pharmaceuticals; Financial support – Regeneron Pharmaceuticals, Alcon, Allergan, Genentech, Neovista, Ophthalmotech, Oraya, P.R.N., Q.L.T., Regeneron Pharmaceuticals, Second Sight; Lecturer – Alcon, Allergan, Genentech, Neovista, Ophthalmotech, Oraya, P.R.N., Q.L.T., Regeneron Pharmaceuticals, Second Sight

Yuichiro Ogura: Consultant – Alcon, Bayer HealthCare, Santen; Lecturer – Alcon, Santen, Novartis; Financial support – Bayer HealthCare

Christian Simader: the author's institution, the Medical University of Vienna, has received funding from Bayer Healthcare for data monitoring/reviewing, statistical analysis, and travel

Glenn J. Jaffe: the author's institution, Duke University, has received research funding from Regeneron Pharmaceuticals to serve as a masked reading center
Jason S. Slakter: Consultant – Lpath, Ohr, Oraya, Regeneron Pharmaceuticals; Financial support and Lecturer – Regeneron Pharmaceuticals; the author's institution, Vitreous-Retina-Macula Consultants of New York, has received research funding from Bayer HealthCare, Centor, Genentech, Genzyme, GlaxoSmithKline, Kanghong Biotech, Lpath, NeoVista, Ohr, Oraya, Regeneron Pharmaceuticals, and Santen to serve as an Angiography Reading Center

George D. Yancopoulos: Employee – Regeneron Pharmaceuticals

Neil Stahl: Employee – Regeneron Pharmaceuticals

Robert Vitti: Employee – Regeneron Pharmaceuticals

Alyson J. Berliner: Employee – Regeneron Pharmaceuticals

Yuhwen Soo: Employee – Regeneron Pharmaceuticals

Majid Anderesi: Employee – Bayer HealthCare

Olaf Sowade: Employee – Bayer HealthCare

Oliver Zeitz: Employee – Bayer HealthCare

Christiane Norenberg: Employee – Bayer HealthCare

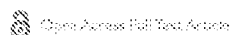
Rupert Sandbrink: Employee – Bayer HealthCare

Jeffrey S. Heier: Consultant – Acucela, Aerpio, Alimera, Allergan, Bausch & Lomb, Bayer HealthCare, Dutch Ophthalmic, Endo Optiks, Forsight, Genzyme, Heidelberg Engineering, Kala Pharmaceuticals, Kanghong, LPath, Nicox, Notal Vision, Ohr Pharmaceutical, Ophthalmotech, Oraya, QLT, Regeneron Pharmaceuticals, Roche, Sequenom, Thrombogenics, Vertex, Xcovery; Financial support – Acucela, Aerpio, Alcon, Alimera, Allergan, Bayer HealthCare, Fovea, Genentech, Genzyme, GlaxoSmithKline, LPath, Neovista, Notal Vision, Novartis, Ohr Pharmaceutical, Ophthalmotech, Paloma, Regeneron Pharmaceuticals

The VEGF Trap-Eye: Investigation of Efficacy and Safety in Wet AMD (VIEW) studies were funded by Regeneron Pharmaceuticals, Inc, Tarrytown, New York, and Bayer HealthCare, Berlin, Germany. The sponsor-authors participated in the design and conduct of the study, analysis of the data, and preparation of the manuscript.

Correspondence:

Jeffrey S. Heier, MD, Ophthalmic Consultants of Boston, 50 Staniford Street, Suite 600, Boston, MA 02114. E-mail: jsheier@eyeboston.com.



Aflibercept in wet AMD: specific role and optimal use

F Semeraro¹
F Morescalchi¹
S Duse¹
F Parmeggiani²
E Gambicorti¹
C Costagliola³

¹Department of Medical and Surgical Specialties, Radiological Specialties and Public Health, Ophthalmology Clinic, University of Brescia, Brescia, Italy; ²Department of Ophthalmology, University of Ferrara, Ferrara, Italy; ³Department of Health Science, Ophthalmology Clinic, University of Molise, Campobasso, Italy

Background: Vascular endothelial growth factor (VEGF) is a naturally occurring glycoprotein in the body that acts as a growth factor for endothelial cells. It regulates angiogenesis, enhances vascular permeability, and plays a major role in wet age-related macular degeneration. The consistent association between choroidal neovascularization and increased VEGF expression provides a strong reason for exploring the therapeutic potential of anti-VEGF agents in the treatment of this disorder. Blockade of VEGF activity is currently the most effective strategy for arresting choroidal angiogenesis and reducing vascular permeability, which is frequently the main cause of visual acuity deterioration. In recent years, a number of other molecules have been developed to increase the efficacy and to prolong the durability of the anti-VEGF effect. Aflibercept (EYLEA[®]; Regeneron Pharmaceutical Inc and Bayer), also named VEGF Trap-eye, is the most recent member of the anti-VEGF armamentarium that was approved by the US Food and Drug Administration in November 2011. Because of its high binding affinity and long duration of action, this drug is considered to be a promising clinically proven anti-VEGF agent for the treatment of wet maculopathy.

Objective: This article reviews the current literature and clinical trial data regarding the efficacy and the pharmacological properties of VEGF-Trap eye and describes the possible advantages of its use over the currently used “older” anti-VEGF drugs.

Methods: For this review, a search of PubMed from January 1989 to May 2013 was performed using the following terms (or combination of terms): vascular endothelial growth factors, VEGF, age-related macular degeneration, VEGF-Trap eye in wet AMD, VEGF-Trap eye in diabetic retinopathy, VEGF-Trap eye in retinal vein occlusions, aflibercept. Studies were limited to those published in English.

Results and conclusion: Two Phase III clinical trials, VEGF Trap-eye Investigation of Efficacy and Safety in Wet AMD (VIEW) 1 and 2, comparing VEGF Trap-eye to ranibizumab demonstrated the noninferiority of this novel compound. The clinical equivalence of this compound against ranibizumab is maintained even when the injections are administered at 8-week intervals, which indicates the potential to reduce the risk of monthly intravitreal injections and the burden of monthly monitoring.

Keywords: aflibercept, AMD, neovascularization, VEGF, VEGF inhibition, VEGF-Trap eye

Correspondence: Francesco Semeraro
Ophthalmology Clinic, Spedali Civili di
Brescia, Piazzale Spedali Civili I,
25123 Brescia, Italy
Tel +39 030 399 5308
Fax +39 030 338 8191
Email semeraro@med.unibs.it

Introduction

The neovascular form of age-related macular degeneration (AMD), also known as wet AMD, is characterized by the formation of subretinal choroidal neovascularization (CNV) and is the cause of most cases of blindness in the elderly. Wet AMD is the major cause of severe vision loss in developed nations and is estimated to affect >2.5 million people worldwide.^{1,2} The patients affected by exudative AMD often experience rapid



loss of fine resolution central vision over several months, and early visual stabilization is a key issue in preserving visual acuity.³

Vascular endothelial growth factor (VEGF) is a naturally occurring glycoprotein in the body that acts as a growth factor selective for endothelial cells. It regulates angiogenesis, enhances vascular permeability, and plays a leading role in wet AMD. The consistent association between CNV and increased VEGF expression provides a strong reason for exploring the therapeutic potential of anti-VEGF agents for the treatment of this disorder.⁴ Blockade of VEGF actions is currently the most effective strategy in arresting choroidal angiogenesis and reducing vascular permeability, which is frequently the main cause of visual acuity deterioration.⁵

Although pegaptanib (Macugen®; Eyetech Pharmaceuticals Inc, FL, USA and Pfizer Inc, New York, NY, USA) was the first VEGF inhibitor approved by the US Food and Drug Administration (FDA). Important advances in the on-label treatment of CNV in AMD have been achieved with the introduction of ranibizumab (Lucentis; Genentech USA, Inc, San Francisco, CA, USA) in 2006. The off-label use of bevacizumab (Avastin; Genentech USA, Inc) has also shown efficacy for treating wet AMD and other exudative retinal diseases and despite the lack of clinical trials to support its safety or efficacy, anecdotal evidence led to its widespread popularity prior to the approval of ranibizumab.

Aflibercept (EYLEA®; Regeneron Pharmaceutical Inc, Tarrytown, NY, USA and Bayer, Basel, Switzerland), also named VEGF Trap-eye, is the most recent member of the anti-VEGF family. This drug has been recently developed to afford a more potent and prolonged anti-VEGF effect and was approved by the FDA in November 2011.⁶ This article reviews the efficacy and summarizes the pharmacological properties of VEGF Trap-eye and describes the possible advantages of its use over the currently used “older” anti-VEGF drugs.

Overview of VEGF and its pathological effects in neovascular AMD

VEGF-A (usually simply referred to as VEGF) is a growth factor encoded by a gene family that also includes placental growth factor (PlGF), VEGF-B, VEGF-C, VEGF-D, and the orf virus encoded VEGF-E.⁷ Differences in exon splicing result in the generation of four main VEGF isoforms: VEGF₁₂₁, VEGF₁₆₅, VEGF₁₈₉, and VEGF₂₀₆, which have 121, 165, 189, and 206 amino acids after cleavage of the signal sequence, respectively.⁸

VEGF stimulates the growth of vascular endothelial cells derived from arteries, veins, and the lymphatic system.⁹ It also induces the formation of thin-walled endothelium-lined structures (ie, angiogenesis) in a variety of in vivo models,¹⁰ and induces rapid elevations in microvascular permeability.¹¹ VEGF acts also as a survival factor for endothelial cells, both in vitro and in vivo.^{12,13} Although endothelial cells represent the primary target of VEGF, several studies have demonstrated that VEGF has mitogenic effects on nonendothelial cell types¹⁴ and promotion effects on monocyte migration.¹⁵ VEGF protects neurons from insults such as hypoxia and glutamate toxicity¹⁶ and it stimulates neurogenesis in vitro and in vivo.¹⁷

VEGF contributes mainly at the initiation stage of CNV by promoting both angiogenesis and vasculogenesis. It acts as an endothelial cell specific mitogen as part of the angiogenesis pathway, and also as a chemoattractant for endothelial cell precursors, inducing their mobilization and differentiation in the vasculogenesis pathway.¹⁶ In addition to these activities, VEGF affects vascular permeability by inducing formation of pores in vascular endothelial cells^{17,18} and by disrupting the intercellular junction between these cells.¹⁹ In turn, this leads to extravasation of fluid, proteins, and circulating cells which disrupts the retinal anatomy and separates the retina from underlying structures, potentially causing severe vision loss.

Although other growth factors can induce the development of blood vessels (ie, transforming growth factor- β , interleukins, insulin-like growth factor-1, and epidermal growth factor), only VEGF appears to be both sufficient and essential for physiologic and pathologic angiogenesis. For this reason, the biochemical pathways involving VEGF are the most studied targets for new potential drugs against neovascular pathologies. Anti-VEGF therapy can arrest choroidal angiogenesis and reduce vascular permeability, which is frequently the main cause of visual acuity deterioration. Pegaptanib and ranibizumab have been approved by the FDA for the treatment of wet AMD, and the off-label use of a third agent, bevacizumab, has shown efficacy for treating wet AMD and other exudative retinal diseases. Pegaptanib was the first anti-VEGF drug FDA approved in December 2004.^{20–22} However, because it was proven to be less efficacious than other anti-VEGF drugs, possibly owing to its selective binding of VEGF₁₆₅, it is no longer widely used in most countries. Ranibizumab and bevacizumab, which are nonselective anti-VEGF drugs, are currently the most extensively used drugs worldwide for wet AMD as well as for many other ocular diseases in which VEGF is overexpressed.²³

The development of new agents for wet AMD has focused on both improving efficacy and extending the duration of action in comparison with the commonly used anti-VEGF drugs ranibizumab and bevacizumab, which are considered the standard drugs. Ranibizumab is a monoclonal humanized antibody fragment and bevacizumab is a whole monoclonal antibody, and both show a high binding affinity for all isoforms of VEGF. These agents appear to have similar efficacy profiles and mechanisms of action, ie, they block the extracellular availability of VEGF which can arrest choroidal angiogenesis and reduce vascular permeability for a limited period of time.²⁴⁻²⁷

Bevacizumab has a lower binding affinity for VEGF than ranibizumab.²⁸ However, bevacizumab is approximately three times larger than ranibizumab (149 kDa versus 48 kDa), and its substantially higher molecular weight results in an intravitreal half-life that is 36% higher than that of ranibizumab. Accumulating clinical evidence has demonstrated that the effects of a single intravitreal dose of either bevacizumab or ranibizumab effectively reduces the effect of VEGF on CNV for 4–6 weeks in most eyes.^{29,30}

Ranibizumab, which is the only widely used drug that is currently approved by the FDA for the treatment of neovascular AMD, is most extensively studied. Several ranibizumab Phase III clinical trials that have studied different treatment schedules, doses, and populations have obtained good results with monthly injections, ie, a mean number of 25 intravitreal injections over 2 years.^{31,32}

Despite the off-label status of bevacizumab, however, it is preferred over ranibizumab by nearly 60% of physicians³³ because of its significantly lower price (ranibizumab, US \$1,950 versus bevacizumab, US \$50) and similar efficacy. The FDA originally approved bevacizumab in 2004 for the treatment of metastatic colorectal cancer.³⁴ To deliver an intravitreal injection, the physician or pharmacist makes numerous unit doses from a vial of bevacizumab, dramatically lowering the cost of the drug. Moreover, many reports and a 2-year multicenter, randomized clinical trial (the Comparisons of Age-Related Macular Degeneration Treatment Trial [CATT]) demonstrated its near equivalency to ranibizumab with monthly dosing (+7.8 letters versus +8.8 letters) and insignificant poorer outcomes with as-needed dosing (+5.0 versus +6.7 letters).^{24,25} Moreover, while the systemic half-life of the unbound product of bevacizumab (20 days) was longer than that of ranibizumab (6 hours), severe systemic adverse events occurred at similar frequencies in patients receiving bevacizumab and ranibizumab in the CATT trial.^{26,35,36}

The main problem with the current anti-VEGF therapy is that monthly intravitreal injections are required for

maintaining vision. This necessitates an excessive time commitment from patients and institutions, and increases the physical and psychological discomfort and financial burdens for the patients. On the other hand, evidence from the SAILOR (Safety Assessment of Intravitreal Lucentis fOR AMD),³⁷ PIER (A Phase IIIb, Multicenter, Randomized, Double-Masked, Sham Injection-Controlled Study of the Efficacy and Safety of Ranibizumab in Subjects with Subfoveal Choroidal Neovascularization [CNV] with or without Classic CNV Secondary to Age-Related Macular Degeneration),^{38,39} and EXCITE (Efficacy and Safety of Ranibizumab in Patients With Subfoveal Choroidal Neovascularization [CNV] Secondary to Age-Related Macular Degeneration)⁴⁰ studies indicates that the efficacy decreases if treatment frequency is reduced. After the loading dose of monthly injections for 3 months of ranibizumab, vision decreases or returns to baseline in most patients if the frequency is reduced to one injection every 2, 3, or 4 months.

Although monthly injections of anti-VEGF represent the best way to preserve vision, most retina surgeons use individualized treatment protocols with monthly assessments after the first three intravitreal injections of anti-VEGF, and further injections are given only if signs of disease activity persist as observed on optical coherence tomography (OCT). This strategy is also abbreviated as “PRN dosing” from the Latin phrase *Pro Re Nata*, which means “as circumstances arise.” The PrONTO (Prospective Optical Coherence Tomography [OCT] Imaging of Patients With Neovascular AMD Treated With Intra-Ocular Ranibizumab) study used this strategy and obtained visual outcomes similar to those achieved with monthly injections while reducing the number of injections from 25 to 10 over 2 years.⁴¹ However, even with this dosing regimen, patients are still required to make monthly visits to the office and undergo frequent and expensive testing because of the constant risk of CNV recurrence.

A treatment approach that aims to reduce the number of injections and the number of visits is the “treat and extend” method. It consists of 3 monthly injections and a follow-up examination after 6 weeks. If the follow-up examination shows evidence of exudation, the patient is treated and told to undergo a follow-up examination in 4 weeks, otherwise the patient is still treated but the follow-up period is extended to 8 weeks. A similar evaluation is performed at the next follow-up visit. However, there is not much evidence in favor of this treatment method. Thus, research on new compounds is focused on inhibiting the VEGF signaling pathway for a more prolonged period.¹

Aflibercept (EYLEA[®]; Regeneron Pharmaceutical Inc and Bayer), or VEGF Trap-eye, is a novel compound derived from the native VEGF receptor (VEGFR) that binds to all VEGF and VEGF-B isoforms as well as to PlGF.⁴² VEGF Trap-eye promises to decrease the injection frequency in conjunction with the “treat and extend” or “PRN” strategies and appears to serve as an effective alternative drug for patients who are less responsive to the previously approved anti-VEGF drugs.

Structure and mechanism of action

The FDA approved VEGF Trap-eye (EYLEA[®], Regeneron Pharmaceutical Inc, and Bayer) for the treatment of subfoveal CNV caused by wet AMD on November 18, 2011.⁴³ VEGF Trap-eye is an intraocular formulation of aflibercept, a product used in oncology (Zaltrap; Regeneron Pharmaceutical Inc), that has been specifically purified and buffered to minimize the risk of eye toxicity when injected intravitreally.⁴⁴ It is a fully human, recombinant fusion protein that has the property to “trap,” that is to catch, hold, and block certain molecules. Aflibercept was constructed from portions of the human VEGFR fused to the FC portion of a human IgG1.⁴⁵

Circulating VEGF initiates a biochemical cascade by activating three membrane spanning tyrosine kinase receptors: VEGFR-1, VEGFR-2, and VEGFR-3.^{46,47} VEGFR-1 (fms-like tyrosine kinase-1, Flt-1) was the first VEGF receptor identified more than a decade ago.⁴⁸ VEGFR-1 releases tissue specific growth factors, recruits endothelial progenitors, and induces matrix metalloproteinases. It is thought to modulate VEGFR-2 signaling and to act as a dummy/decoy receptor by sequestering VEGF and preventing it from binding to VEGFR-2.⁷ VEGFR-2 (kinase insert domain-containing receptor or KDR) is considered the major mediator of the mitogenic, angiogenic, permeability enhancing, and anti-apoptotic effects of VEGF.⁷

Both VEGFR-1 and VEGFR-2 have seven Ig-like binding sequences for VEGF (two of which are incorporated in VEGF Trap-eye) in the extracellular region, a single transmembrane region, and a consensus tyrosine kinase sequence that is interrupted by a kinase insert domain.^{49–51} The third member of the same family of receptor tyrosine kinases is VEGFR-3.⁵² This protein is not a receptor for VEGF, but binds VEGF-C and VEGF-D.⁵³ Because VEGFR-1 possesses a higher affinity for VEGF than VEGFR-2, drug developers have used its binding sequences for VEGF Trap-eye.

Structurally, aflibercept is a soluble decoy receptor of 115 kDa that is made by the second binding domain of VEGFR-1 and the third binding domain of VEGFR-2, which then are fused to the FC region of a human IgG1 (Figure 1).

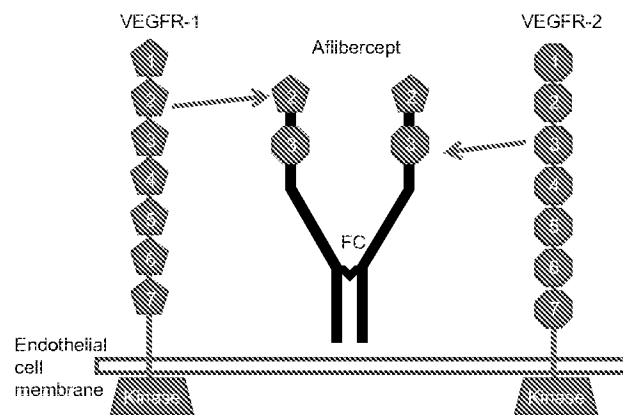


Figure 1 Diagram showing the structure of the vascular endothelial growth factor receptor-1 and -2 and the structure of aflibercept (VEGF Trap-eye).

Notes: Aflibercept (VEGF Trap-eye) is generated by a fusion that includes the second binding domain of vascular endothelial growth factor receptor (VEGFR)-1 and the third binding domain of VEGFR-2 attached to a FC fragment of a human IgG.

Abbreviation: FC, fragment crystallizable region.

The intermediate size of aflibercept (115 kDa compared to 48 kDa for ranibizumab and 148 kDa for bevacizumab) results in an estimated intravitreal half-life of 7.1 days and a duration of clinical action possibly as long as 2.5 months, which exceeds the 1-month intravitreal binding activity of ranibizumab.^{54,55} The molecular configuration of aflibercept allows it to bind to all of the VEGF isoforms more tightly than their native receptors (the dissociation constant [K_d] of aflibercept for VEGF₁₆₅ = 0.49 pmol/L).⁴² Thus, this compound effectively prevents VEGF from binding and activating its cognate receptors (the K_d of VEGFR-1 and VEGFR-2 for VEGF₁₆₅ are 9.33 and 88.8 pmol/L,

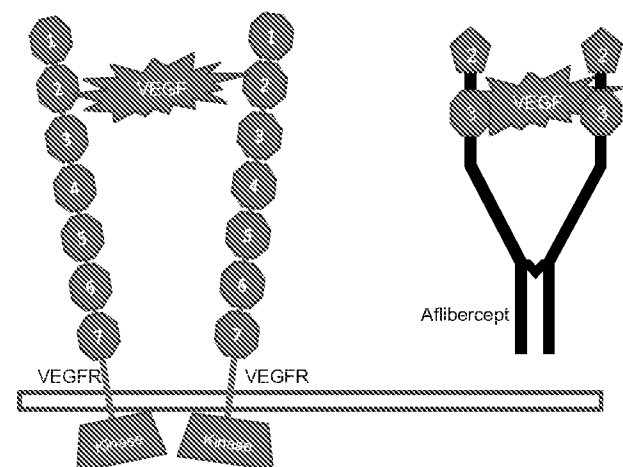


Figure 2 Vascular endothelial growth factor binds to two vascular endothelial growth factor receptors which induces the angiogenic response by activating the tyrosine kinase.

Notes: Vascular endothelial growth factor receptor (VEGFR)-2 is shown. Aflibercept (VEGF Trap-eye) binds all vascular endothelial growth factor (VEGF) isoforms more tightly than their native receptors, thus preventing binding of VEGF to its cognate receptors.

respectively) (Figure 2).⁵⁶ Moreover, the binding affinity of aflibercept ($K_d = 0.49$ pmol/L) is almost 100 times higher than that of ranibizumab ($K_d = 46$ pmol/L) and bevacizumab ($K_d = 58$ pmol/L).^{54,55} This was primarily attributed to the association rate constant for aflibercept binding to human VEGF₁₆₅, which is almost 80 times faster than the corresponding association rate constant values for ranibizumab and bevacizumab.

Because of these characteristics, the ability of aflibercept to block VEGF induced activation of VEGFR-1 and -2 in vitro is much stronger than that of ranibizumab and bevacizumab. Additionally, it blocks both PIGF-1 and PIGF-2 mediated activation of VEGFR-1, whereas ranibizumab and bevacizumab do not show such activity. A presumably important functional difference between aflibercept and the other anti-VEGF drugs currently in use is that it can bind and inhibit VEGF as well as PIGF-1 and -2 and VEGF-B, which have also been implicated in pathological vascular remodeling. Experimental evidence shows that targeting VEGF-B and PIGF inhibits CNV and suggests that PIGF synergizes with VEGF in promoting vascular pathology in wet AMD.⁵⁷

Pharmacodynamics, pharmacokinetics, and metabolism

Aflibercept forms a stable, inert 1:1 complex with either VEGF, VEGF-B, or the PIGF ligand preventing the activation of their receptors, VEGFR-1 and -2.⁵⁶ The highest intravitreal dose used in pivotal trials for aflibercept is 2 mg, which is 100-fold lower than the dose allowed in oncology (4–6 mg/kg).^{44,60} Following intravitreal injection of 2 mg of aflibercept, the drug can be detected in plasma as a free drug (a minor quantity) or in a complex bound with VEGF. The drug is rapidly cleared from circulation via pinocytotic proteolysis and glomerular filtration after forming a complex with VEGF via the same pathways that metabolize antibodies.

Following intravitreal injection of 2 mg of aflibercept, the mean maximal plasma concentration of unbound VEGF Trap-eye is attained in 1–3 days, and was estimated to be 200-fold lower than the concentration required for maximal systemic VEGF binding. The systemic half-life of unbound aflibercept is 1.5 days, which is inferior to that of bevacizumab (20 days) and closer to the systemic half-life of ranibizumab (6 hours).⁵⁹ Free aflibercept has never been detected in plasma at 2 weeks after intravitreal injection and cannot accumulate in plasma in the loading phase.⁴⁴ Thus, an intravitreal aflibercept dose of 2 mg would be predicted to cause negligible systemic

activity and have a systemic safety profile similar to that of ranibizumab.

Therapeutic efficacy

The first surveys regarding the use of aflibercept in treatment of wet AMD emerged from a preclinical study conducted on animal models. This study, published in 2003, showed the first evidence that VEGF Trap-eye is capable of suppressing CNV and VEGF mediated breakdown of the blood–retinal barrier in transgenic mice with laser induced CNV, which was treated with subcutaneous or intravitreal administration.⁵⁸ The initial use of aflibercept for wet AMD consisted of intravenous injections with doses between 0.3 mg/kg and 3 mg/kg (the usual oncologic dose is 4 mg/kg) and administered every 2 weeks to 25 patients.⁶⁰ Macular thickness decreased by an average of 66% and vision improved in many patients. Patients receiving the higher dose (3 mg/kg) experienced more systemic hypertension and proteinuria than those treated with the lower dose (1 mg/kg). However, the promising effects obtained intravenously encouraged researchers to transition the trial to intravitreal injections.

The Phase I Clinical Evaluation of Anti-angiogenesis in the Retina Intravitreal Trial (CLEAR-IT 1)⁶⁰ investigation was a small trial (21 patients) designed to determine the maximum tolerated dose, the bioactivity, and the safety and tolerability of intravitreally administered aflibercept in patients with wet AMD. This study confirmed that aflibercept doses between 0.05 mg and 4 mg were well tolerated. At 6 weeks after a single injection, most patients experienced an improvement in visual acuity (mean visual gain, 4.4 letters) and showed a decrease in macular thickness (–105 μ m). Almost 50% of the patients followed for 12 weeks did not show retinal leakage and maintained vision gain.^{61,62} On the basis of the results of CLEAR-IT 1, the developers hoped to show that an intravitreal formulation of aflibercept could be administered less frequently than once a month.

In a Phase II dose and interval ranging trial, 159 patients with wet AMD (CLEAR-IT 2) were randomized into five treatment groups: the first two groups received 3 monthly aflibercept injections of 0.5 mg or 2 mg and the other three groups received only one aflibercept injection of 0.5 mg, 2 mg, or 4 mg.⁶⁴ Final global evaluations were performed at 12 weeks. Although visual improvement at week 8 was similar in patients receiving a single dose or two doses (5.7 letters), the average vision in all groups improved more in patients treated monthly (mean gain of ≥ 8 letters) at 12 weeks. After 12 weeks, the reduction in macular thickness experienced by the patients receiving three monthly injections

exceeded that of patients treated only once.⁶³ For this reason, a second part of the CLEAR-IT 2 study was designed in which aflibercept treatment was provided as needed (PRN) from week 12 to 52 and monthly OCT and fluorescein angiography (FAG) examinations were performed, starting with a reinjection of all patients at week 12.⁶⁴ A decision to perform reinjection was made if any of the following conditions were observed: central retinal thickness increase of ≥ 100 μm , loss of at least five lines in the visual acuity chart approved by the Early Treatment Diabetic Retinopathy Study (ETDRS),⁶⁵ persistent fluid on OCT, new onset of classic neovascularization, persistent leakage on FAG, or the presence of a new hemorrhage on clinical examination. An average of two injections was required, with a mean time to the first injection of 129 days. After 1 year (week 52), the average improvement in vision was +5.3 letters. Patients initially treated with 2 mg every 4 weeks had the best visual improvement (mean gain of 9 letters).

The CLEAR-IT 2 study provided the first indication that aflibercept may be dosed as needed with excellent gains in vision.⁶⁶ Additionally, patients receiving a monthly “loading” dose for 3 months achieved superior visual results than those receiving single injections. Many patients required only two injections after the loading phase and at the last visit after 1 year. Thus, three different dosing regimens were identified for the Phase III studies:⁶⁷ 0.5 mg monthly, 2 mg monthly, or 2 mg every 2 months after the loading phase of three initial monthly doses.

In Phase III, two equivalent pivotal clinical trials of VEGF Trap-eye, VEGF Trap-eye Investigation of Efficacy and Safety in Wet AMD (VIEW) 1 and 2, were conducted to determine if VEGF Trap-eye was noninferior and clinically equivalent to ranibizumab, the drug considered to be the standard against which all subsequent drugs should be compared.^{66,67} The VIEW 1 study enrolled 1,217 patients in the US and Canada, and the VIEW 2 study enrolled 1,240 patients in Europe, Asia, Japan, and Latin America. Each trial randomized patients among three treatment regimens: 0.5 mg of aflibercept given monthly, 2 mg given monthly, and 2 mg given every two months after 3 monthly loading doses for 3 months. Both studies evaluated the noninferiority efficacy in comparison with a fourth arm of the study in which patients received 0.5 mg of ranibizumab monthly. The first noninferiority endpoint was the percentage of patients who maintained their visual acuity (decrease in vision less than -15 letters); the second noninferiority endpoint was the percentage of patients who gained vision.

After the first year, both the VIEW 1 and 2 studies were continued for a second year (52–96 weeks) in which a modified PRN strategy was adopted. Patients were assessed monthly and were treated only if necessary (with the same drug and dose as in the first year), but the injection was repeated at least every three months in all cases. At week 52, the proportion of patients who maintained their vision (lost < 15 ETDRS letters) was approximately 95% when using 2 mg of aflibercept (either monthly or every 2 months after the loading phase). The same results were obtained with 0.5 mg of ranibizumab given monthly. The gains in vision were comparable among the drugs administered monthly: a mean gain of +10.9 letters and +7.6 letters in the aflibercept group and a mean gain of +8.1 letters and +9.4 letters in those receiving ranibizumab, in VIEW 1 and VIEW 2,⁶⁷ respectively.

In VIEW 1, patients receiving 2 mg of aflibercept every 4 weeks gained more vision than those receiving ranibizumab (+10.9 letters versus +8.1 letters; $P = 0.0054$).⁶⁷ Improvements in macular thickness were not statistically different among any of the treatment groups. VIEW 2 patients receiving 2 mg of aflibercept every 8 weeks showed bimonthly fluctuations in macular thickness without corresponding fluctuations in visual acuity.⁶⁷ The safety of aflibercept was excellent and was comparable with that of ranibizumab in both the VIEW 1 and VIEW 2 studies. Severe extraocular adverse events such as stroke and myocardial infarction occurred with similar frequencies in patients receiving aflibercept (0.7% and 2.6%, respectively) and in patients receiving ranibizumab (1.6% and 2.6%, respectively) in both VIEW trials.

In VIEW 1, the mean vision gain from the baseline (best corrected visual acuity) BCVA at week 52 was greater in the 2 mg aflibercept every month group when compared with the ranibizumab group (mean gain of +10.9 versus +8.1 ETDRS letters).⁶⁷ Conversely, a statistically significant difference was not found in vision gain in comparison to ranibizumab (mean gain of +7.6 letters versus +9.4 letters) in VIEW 2.⁶⁷ The reason for this difference in vision results is unknown. However, it is likely that racial and ethnic differences existed between the two trials. Several reports have suggested that the incidence of polypoidal choroidal vasculopathy, which has been suggested to be a variant of neovascular AMD, is markedly high in African-American people, relatively high in the Asian population, and low in white people with AMD.^{68,69} Polypoidal CNV does not respond well to anti-VEGF therapy alone and should be treated with a combination of photodynamic therapy and anti-VEGF therapy for better results. Thus, a limitation of the two trials was the inclusion of all CNV types by using FAG but not indocyanine green angiography.

A comparative subanalysis of the data will be required to address this difference.

However, both VIEW studies showed that 2 mg injections of VEGF Trap-eye every two months delivered a comparable gain in visual acuity to monthly ranibizumab (+7.9 versus +8.1 letters in VIEW 1; +8.9 versus +9.4 letters in VIEW 2).⁶⁷ Additional efficacy was not demonstrated when VEGF Trap-eye was administered every 4 weeks compared with every 8 weeks, thus suggesting that patients would not require monthly examinations. In the two trials, approximately one third of patients receiving 2 mg of aflibercept every second month experienced a clinical improvement in visual acuity (ranging from +7 to +10 letters). Based on the 1-year efficacy (maintenance of vision) and safety results of the VIEW trials, the FDA approved a regimen of 2 mg of VEGF Trap-eye every 8 weeks for the treatment of wet AMD.⁷⁰ The recommended treatment regimen includes three loading injections at 4-week intervals, followed by injections every 8 weeks. During the second year (52–96 weeks), patients were assessed monthly and, if necessary, were treated via a modified PRN protocol with a new injection performed not less frequently than once every three months. Between weeks 52 and 96, patients initially receiving 2 mg of aflibercept every 8 weeks and those initially receiving ranibizumab every 4 weeks maintained previous gains in vision.

In an integrated analysis of the VIEW 1 and VIEW 2 studies,⁷⁰ the visual acuity gain from baseline in the aflibercept group that received 2 mg every 8 weeks was +7.6 letters at week 96 compared to +8.4 letters at week 52, with an average of 11.2 injections over 2 years and 4.2 injections during the second year. The visual acuity gain from baseline in the monthly ranibizumab group was +7.9 letters at week 96 compared to +8.7 letters at week 52, with an average of 16.5 injections over 2 years and 4.7 injections during the second year.⁷⁰

Only 16% of the patients received six or more injections during the second year.⁷⁰ In comparison, patients receiving ranibizumab monthly during the first year and PRN the second year received an average of 16.5 injections: 12 during the first year and an average of 4.7 injections over the second year. Approximately 26.5% of the patients required six or more injections during the second year. During year 2 of the VIEW trials,⁷⁰ 48% of the patients receiving 2 mg of aflibercept and 40% of the patients receiving ranibizumab received the minimum number (three) of injections.

In both studies,⁶⁷ the ocular adverse events experienced across the four treatment groups were those commonly associated with intravitreal injections:^{35,36} conjunctival hem-

orrhages, eye pain, and vitreous floaters. Systemic adverse events, such as falls, pneumonia, cancer, and cardiovascular disease were also balanced across the groups and were those commonly found in elderly AMD patients. No evidence of an increased risk of thromboembolic events such as stroke or myocardial infarction was found.⁷¹

VEGF Trap-eye: other clinical uses in retinal disease

The VEGF cytokine also plays an important role in the pathogenesis of vascular retinal diseases like diabetic retinopathy, central retinal vein occlusion (CRVO), and branch retinal vein occlusion. It causes an increase in retinal capillary permeability and leakage of fluid into the retina and macula, leading to significant loss of central vision.⁷² VEGF expression, which is upregulated by hypoxia, was found to be elevated in the ocular fluids of patients with diabetic macular edema (DME) and CRVO.⁷³ Anti-VEGF compounds have been successfully used as the first line of treatment for diabetic retinopathy⁷⁴ and macular edema due to CRVO, and have replaced laser photocoagulation in some cases.⁷⁵

Several anti-VEGF agents have been evaluated in numerous clinical trials from 2008 to the present day. Most notably, these include prospective clinical trials regarding intravitreal ranibizumab for the treatment of DME in RD (READ2 [Ranibizumab for Edema of the mAcUla in Diabetes], RESOLVE [Safety and Efficacy of Ranibizumab in Diabetic Macular Edema With Center Involvement], RESTORE [A 12 Month Core Study to Assess the Efficacy and Safety of Ranibizumab (Intravitreal Injections) in Patients With Visual Impairment Due to Diabetic Macular Edema and a 24 Month Open-label Extension Study], RISE [A Study of Ranibizumab Injection in Subjects With Clinically Significant Macular Edema (ME) With Center Involvement Secondary to Diabetes Mellitus (RISE)], RIDE [A Study of Ranibizumab Injection in Subjects With Clinically Significant Macular Edema (ME) With Center Involvement Secondary to Diabetes Mellitus (RIDE)]),⁷⁶ which demonstrated the superiority of this anti-VEGF compound over both sham injection and focal grid laser.^{77–82} Aflibercept was evaluated in a double-masked, prospective, randomized, multicenter Phase II trial, entitled DME And VEGF Trap-eye: INvestigation of Clinical Impact (DA VINCI),^{83,84} in which 221 patients with clinically significant DME with central macular involvement were randomized and 219 patients were treated with a balanced distribution over five groups. These groups included monthly doses of 0.5 or 2 mg of VEGF Trap-eye, monthly doses of 2 mg of VEGF Trap-eye for 3 months and then

every 8 weeks, monthly doses of 2 mg of VEGF Trap-eye for 3 months and then PRN, and macular laser therapy.^{83,84} The mean improvements in BVCA at 52 weeks in the VEGF Trap-eye groups were +11.0, +13.1, +9.7, and +12.0 letters, respectively, versus -1.3 letters in the laser group. It is interesting to note these similar results with longer dosing intervals of treatment.

The DA VINCI study^{83,84} showed that in addition to the benefits related to the reduction of central macular edema, aflibercept provides secondary benefits related to the nonprogression of retinopathy with the prevents the development of vascular neoproliferation.⁸⁴ Aflibercept has turned out to be a promising option in DME therapy because of its high binding affinity and extended duration of action. The latter quality is very important in view of the fact that diabetic retinopathy is a chronic disease and that a large percentage of affected patients are of working age.

Presently, no published randomized clinical trials have directly compared any of the anti-VEGF drugs for the treatment of diabetic retinopathy. However, two Phase III clinical studies, the VIVID (VEGF Trap-Eye In Vision Impairment Due to DME)⁸⁵ and the VISTA (Study of Intravitreal Administration of VEGF Trap-Eye in Patients With Diabetic Macular Edema)⁷⁵ studies, have been initiated and are evaluating the efficacy and safety of VEGF Trap-eye in comparison with laser treatment over a period of 1 and 2 years, respectively. Finally, a three arm study comparing ranibizumab versus bevacizumab versus aflibercept – the DRCR protocol T – is now in the enrollment phase.⁷⁵

In September 2012, the FDA approved aflibercept injection for the treatment of macular edema following CRVO.⁸⁶ This approval was based on data from the Phase III COPERNICUS (Controlled Phase 3 Evaluation of Repeated intravitreal administration of VEGF Trap-Eye In Central retinal vein occlusion: Utility and Safety)^{87,88} and GALILEO (General Assessment Limiting Infiltration of Exudates in central retinal vein Occlusion with EYLEA) studies.⁸⁹ In both studies, the results regarding the quality of vision and anatomical outcomes were superior in the aflibercept treated group than in the sham control group. The initial 6-month phase was similar among these studies, during which patients were randomized to receive either an intravitreal injection of 2 mg of aflibercept or a sham injection every month, but the second 6-month phase was different between the two studies. In the GALILEO study,⁸⁹ patients in the treatment group were treated on a PRN basis with aflibercept, while patients in the placebo group continued to receive treatment with sham injections;

in the COPERNICUS study,⁸⁸ all patients were treated with aflibercept on a PRN basis.

In both the COPERNICUS and GALILEO studies, aflibercept injection resulted in an improvement in visual acuity of >15 letters in 56.1% and 60.2% of patients, respectively, at week 24 compared with those receiving sham injections (12.3% and 22.1%, respectively).^{87,89} At week 52 in the COPERNICUS study,⁸⁸ the improvement in visual acuity was 55.3% in the aflibercept/aflibercept PRN patients compared with 30.1% in the sham/aflibercept PRN patients. In the GALILEO study, in which control patients did not receive any aflibercept injections, the improvement was 60.2% and 32.4%, respectively.^{88,90} The results of these studies showed that it is possible to maintain an excellent visual outcome and to extend the range of administration while using the PRN strategy. These data indicate that aflibercept provides benefits to patients with CRVO and using this drug as needed may become a first line approach that will reduce the burden of monthly injections.

Conclusion

In conclusion, aflibercept, or VEGF Trap-eye, may be considered an attractive alternative to other anti-VEGF agents because it appears to offer visual outcomes similar to ranibizumab and bevacizumab with a longer duration of action. For the first time, an anti-VEGF drug can be given at 2-month intervals with results comparable to ranibizumab given every 4 weeks.⁹¹

Aflibercept was shown to be generally well tolerated in the VIEW I and II studies, and the ocular adverse events and adverse events were similar to those of ranibizumab. Patients receiving 2 mg of aflibercept every 8 weeks achieved visual acuity gains similar to those receiving ranibizumab with five fewer injections, on average, over 2 years. Patients who required the most intense therapy received, on average, 1.4 fewer injections in the group receiving 2 mg of aflibercept every 8 weeks when compared to the ranibizumab group in the second year.

Although the future direction of the development of therapeutic management techniques should be driven by improving results, reducing the burden and the cost of treatment should also be considered. In particular, the cost of AMD treatments with the approved anti-VEGF agents is much higher by any metric compared to any previous AMD and retinal treatment. Economic consideration is an important influencing factor in the selection of drugs for individual patients, and the comparable safety and reduced injection burden of aflibercept in comparison with ranibizumab enhances its cost effectiveness. For those clinicians using ranibizumab, the transition to

aflibercept (which costs \$100 less than ranibizumab) will be easy because the total cost of aflibercept treatment will be even lower than the presumed per vial cost after accounting for the fact that the cost will be lowered further by the greater time interval between injections. However, the transition to aflibercept from off-label bevacizumab (which costs \$1,800 less than aflibercept) will be slower for cost conscious physicians. In this case, the relative merits of the more expensive, but less frequently dosed, aflibercept compared to the more frequently dosed, lower cost alternative of off-label bevacizumab must also be considered.

Moreover, aflibercept can be used in shifting patients treated with bevacizumab to aflibercept, as this monthly injection was the only regimen shown to be equivalent to ranibizumab in the comparison of AMD treatment trials.²⁵ Yet another strategy woven into combination therapy stems from the observation that most visual improvements with anti-VEGF agents occur in the first three months, raising the possibility of an initial (albeit high cost) loading treatment with a subsequent (lower cost) maintenance treatment. The addition of new drugs to these combination strategies may diminish both maintenance and loading therapies, achieving better results.

Furthermore, a major concern with chronic therapies is the reduction of the biological effect, which can limit long-term efficacy. This phenomenon has been called tachyphylaxis and has been described as a progressive decrease in the therapeutic response after repetitive administration of anti-VEGF drugs.⁹² A retrospective review from the National Eye Institute found that between five and ten injections of bevacizumab were required before tachyphylaxis occurred.⁹³ Nonresponder patients, or patients who experience tachyphylaxis, will need alternative treatment strategies to break the cycle of monthly injections with the same stagnant results. A possible solution would be to combine drugs with different mechanisms of action or different pharmacokinetics, for example, switching the treatment to different VEGF blockers. Several reports have shown that administration of aflibercept to eyes that had persistent fluid despite prolonged bevacizumab or ranibizumab therapy resulted in rapid resolution of the subretinal fluid and the flattening of pigment epithelial detachments.⁶ This indicates that aflibercept can be used with success in patients who show resistance to conventional anti-VEGF drugs and suggest that aflibercept works remarkably well as a “salvage” therapy.

In light of the above analysis based on the literature, the personal opinion of the authors on the therapy for maculopathy is that the best approach for wet AMD is an “attack

on several fronts.” In this sense, the first line drugs are anti-VEGF agents that can be used in combination with drugs that inhibit the actions of molecules involved in angiogenesis, including integrins, complements, and PIGF, and with compounds that are able to maintain and preserve the integrity of the retinal photoreceptors and of the choriocapillaris. However, because an effective combination therapy is still several years away, aflibercept promises to become the leading medication in the treatment of wet AMD in the coming years because of its ability to inhibit angiogenesis.

Disclosure

The authors report no conflicts of interest in this work. This review received no specific grant from any funding agency in the public, commercial, or not for profit sector.

References

1. Chappelov AV, Kaiser PK. Neovascular age-related macular degeneration: potential therapies. *Drugs*. 2008;68(8):1029–1036.
2. La Cour M, Kiilgaard JF, Nissen MH. Age-related macular degeneration: epidemiology and optimal treatment. *Drugs Aging*. 2002;19(2):101–133.
3. Bloch SB, Larsen M, Munch IC. Incidence of legal blindness from age-related macular degeneration in Denmark: year 2000–2010. *Am J Ophthalmol*. 2012;153(2):209–213.
4. Campa C, Costagliola C, Incorvaia C, et al. Inflammatory mediators and angiogenic factors in choroidal neovascularization: pathogenetic interactions and therapeutic implications. *Mediators Inflamm*. 2010;2010.
5. Stewart MW. The expanding role of vascular endothelial growth factor inhibitors in ophthalmology. *Mayo Clin Proc*. 2012;87(1):77–88.
6. Stewart MW. Clinical and differential utility of VEGF inhibitors in wet age-related macular degeneration: focus on aflibercept. *Clin Ophthalmol*. 2012;6:1175–1186.
7. Ferrara N. Vascular endothelial growth factor: basic science and clinical progress. *Endocr Rev*. Aug 2004;25(4):581–611.
8. Alitalo K, Tammela T, Petrova TV. Lymphangiogenesis in development and human disease. *Nature*. 2005;438:946–953.
9. Lee S, Jilani SM, Nikolova GV, Carpizo D, Iruela-Arispe ML. Processing of VEGF-A by matrix metalloproteinases regulates bioavailability and vascular patterning in tumors. *J Cell Biol*. 2005;169:681–691.
10. Leung DW, Cachianes G, Kuang WJ, Goeddel DV, Ferrara N. Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science*. 1989;246:1306–1309.
11. Alon T, Hemo I, Itin A, Stone J, Keshet E. Vascular endothelial growth factor acts as a survival factor for newly formed retinal vessels and has implications for retinopathy of prematurity. *Nat Med*. 1995;1:1024–1028.
12. Gerber HP, Dixit V, Ferrara N. Vascular endothelial growth factor induces expression of the antiapoptotic proteins Bcl-2 and A1 in vascular endothelial cells. *J Biol Chem*. 1998;273:13313–13316.
13. Carmeliet P. Angiogenesis in life, disease and medicine. *Nature*. 2005;438:932–936.
14. Rosenstein JM, Krum JM. New roles for VEGF in nervous tissue – beyond blood vessels. *Exp Neurol*. 2004;187:246–253.
15. Jin K, Zhu Y, Sun Y, Mao XO, Xie L, Greenberg DA. Vascular endothelial growth factor (VEGF) stimulates neurogenesis in vitro and in vivo. *Proc Natl Acad Sci U S A*. 2002;99:11946–11950.
16. Asahara T, Takahashi T, Masuda H, et al. VEGF contributes to postnatal neovascularization by mobilizing bone marrow-derived endothelial progenitor cells. *EMBO J*. 1999;18:3964–3972.

17. Roberts WG, Palade GE. Neovascularization induced by vascular endothelial growth factor is fenestrated. *Cancer Res.* 1997;57:765–772.
18. Roberts WG, Palade GE. Increased microvascular permeability and endothelial fenestration induced by vascular endothelial growth factor. *J Cell Sci.* 1995;108(6):2369–2379.
19. Monsky WL, Fukumura D, Gohongi T, et al. Augmentation of transvascular transport of macromolecules and nanoparticles in tumors using vascular endothelial growth factor. *Cancer Res.* 1999;59:4129–4135.
20. Gragoudas ES, Adamis AP, Cunningham ET Jr, Feinsod M, Guyer DR; VEGF Inhibition Study in Ocular Neovascularization Clinical Trial Group. Pegaptanib for neovascular age-related macular degeneration. *N Engl J Med.* 2004;351:2805–2816.
21. VEGF Inhibition Study in Ocular Neovascularization (V.I.S.I.O.N.) Clinical Trial Group, Chakravarthy U, Adamis AP, et al. Year 2 efficacy results of 2 randomised controlled clinical trials of pegaptanib for neovascular age-related macular degeneration. *Ophthalmology.* 2006;113:1508.e1–e25.
22. Singerman LJ, Masonson H, Patel M, et al. Pegaptanib sodium for neovascular age-related macular degeneration: third-year safety results of the VEGF Inhibition Study in Ocular Neovascularisation (VISION) trial. *Br J Ophthalmol.* 2008;92:1606–1611.
23. Frampton JE. Ranibizumab: a review of its use in the treatment of neovascular age-related macular degeneration. *Drugs Aging.* 2013;30(5):331–358.
24. Martin DF, Maguire MG, Ying GS, Grunwald JE, Fine SL, Jaffe GJ, CATT Research Group. Ranibizumab and bevacizumab for neovascular age-related macular degeneration. *N Engl J Med.* 2011;364(20):1897–1908.
25. Comparison of Age-related Macular Degeneration Treatments Trials (CATT) Research Group, Martin DF, Maguire MG, et al. Ranibizumab and bevacizumab for treatment of neovascular age-related macular degeneration: two-year results. *Ophthalmology.* 2012;119(7):1388–1398.
26. Ahfat FG, Zaidi FH. Bevacizumab vs ranibizumab—an appraisal of the evidence from CATT and IVAN. *Eye (Lond).* Mar 2013;27(3):289–290.
27. Costagliola C, Romano M, Corte MD, et al. Intravitreal bevacizumab for treatment-naïve patients with subfoveal occult choroidal neovascularization secondary to age-related macular degeneration: a 12-month follow-up study. *Retina.* Oct 2009;29(9):1227–1234.
28. Steinbrook R. The price of sight—ranibizumab, bevacizumab, and the treatment of macular degeneration. *N Engl J Med.* 2006;355:1409–1412.
29. Meyer CH, Krohne TU, Holz FG. Intracocular pharmacokinetics after a single intravitreal injection of 1.5 mg versus 3.0 mg of bevacizumab in humans. *Retina.* 2011;31(9):1877–1884.
30. Conrad PW, Zacks DN, Johnson MW. Intravitreal bevacizumab has initial clinical benefit lasting eight weeks in eyes with neovascular age-related macular degeneration. *Clin Ophthalmol.* 2008;2(4):727–733.
31. Rosenfeld PJ, Brown DM, Heier JS, et al; MARINA Study Group. Ranibizumab for neovascular age-related macular degeneration. *N Engl J Med.* 2006;355:1419–1431.
32. Brown DM, Kaiser PK, Michels M, et al; ANCHOR Study Group. Ranibizumab versus verteporfin for neovascular age-related macular degeneration. *N Engl J Med.* 2006;355:1432–1444.
33. Brechner RJ, Rosenfeld PJ, Babish JD, Caplan S. Pharmacotherapy for neovascular age-related macular degeneration: an analysis of the 100% 2008 medicare fee-for-service part B claims file. *Am J Ophthalmol.* 2011;151(5):887–895.e1.
34. Bevacizumab prescribing information. Available from: http://www.accessdata.fda.gov/drugsatfda_docs/label/2009/125085s01681bl.pdf. Accessed on July 24, 2013.
35. Semeraro F, Morescalchi F, Parmeggiani F, Arcidiacono B, Costagliola C. Systemic adverse drug reactions secondary to anti-VEGF intravitreal injection in patients with neovascular age-related macular degeneration. *Curr Vasc Pharmacol.* 2011;9(5):629–646.
36. Costagliola C, Agnifili L, Arcidiacono B, et al. Systemic thromboembolic adverse events in patients treated with intravitreal anti-VEGF drugs for neovascular age-related macular degeneration. *Expert Opin Biol Ther.* 2012;12(10):1299–1313.
37. Boyer DS, Heier JS, Brown DM, et al. A phase IIIb study to evaluate the safety of ranibizumab in subjects with neovascular age-related macular degeneration. *Ophthalmology.* 2009;116(9):1731–1739.
38. Regillo CD, Brown DM, Abraham P, et al. Randomised, double-masked, sham-controlled trial of ranibizumab for neovascular age-related macular degeneration: PIER Study year 1. *Am J Ophthalmol.* 2008;145:239–248.
39. Michels M, Francom S, Wilson L. Systemic safety and risk factors associated with intravitreal ranibizumab in patients with choroidal neovascularization (CNV) secondary to age-related macular degeneration (AMD). In: 26th Annual Meeting of the American Society of Retina Specialists; October 11–15, 2008; Maui, HI, USA.
40. Schmidt-Erfurth U, Eldem B, Guymer R, et al. EXCITE Study Group. Efficacy and safety of monthly versus quarterly ranibizumab treatment in neovascular age-related macular degeneration: the EXCITE study. *Ophthalmology.* 2011;118(5):831–839.
41. Lalwani GA, Rosenfeld PJ, Fung AE, et al. A variable-dosing regimen with intravitreal ranibizumab for neovascular age-related macular degeneration: year 2 of the PrONTO Study. *Am J Ophthalmol.* Jul 2009;148(1):43–58.
42. Papadopoulos N, Martin J, Ruan Q, et al. Binding and neutralization of vascular endothelial growth factor (VEGF) and related ligands by VEGF Trap, ranibizumab and bevacizumab. *Angiogenesis.* 2012;15(2):171–185.
43. US Food and Drug Administration. FDA approves Eylea for eye disorder in older people. November 18, 2011. Available from: <http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm280601.htm>. Accessed December 4, 2011.
44. Dixon JA, Oliver SC, Olson JL, Mandava N. VEGF Trap—Eye for the treatment of neovascular age-related macular degeneration. *Expert Opin Investig Drugs.* 2009;18(10):1573–1580.
45. Aflibercept: AVE 0005, AVE 005, AVE0005, VEGF Trap—Regeneron, VEGF Trap (R1R2), VEGF Trap—Eye. *Drugs R D.* 2008;9(4):261–269.
46. Vaisman N, Gospodarowicz D, Neufeld G. Characterization of the receptors for vascular endothelial growth factor. *J Biol Chem.* 1990;265:19461–19466.
47. Jakeman LB, Armanini M, Philips HS, Ferrara N. Developmental expression of binding sites and mRNA for vascular endothelial growth factor suggests a role for this protein in vasculogenesis and angiogenesis. *Endocrinology.* 1993;133:848–859.
48. De Vries C, Escobedo JA, Ueno H, Houck K, Ferrara N, Williams LT. The fms-like tyrosine kinase, a receptor for vascular endothelial growth factor. *Science.* 1992;255:989–991.
49. Shibuya M, Yamaguchi S, Yamane A, et al. Nucleotide sequence and expression of a novel human receptor-type tyrosine kinase (flt) closely related to the fms family. *Oncogene.* 1990;5(4):519–524.
50. Matthews W, Jordan CT, Gavin M, Jenkins NA, Copeland NG, Lemischka IR. A receptor tyrosine kinase cDNA isolated from a population of enriched primitive hematopoietic cells and exhibiting close genetic linkage to c-kit. *Proc Natl Acad Sci U S A.* 1991;88:9026–9030.
51. Terman BI, Carrion ME, Kovacs E, Rasmussen BA, Eddy RL, Shows TB. Identification of a new endothelial cell growth factor receptor tyrosine kinase. *Oncogene.* 1991;6:1677–1683.
52. Pajusola K, Aprelikova O, Korhonen J, et al. FLT4 receptor tyrosine kinase contains seven immunoglobulin-like loops and is expressed in multiple human tissues and cell lines. *Cancer Res.* 1992;52:5738–5743.
53. Karkkainen MJ, Makinen T, Alitalo K. Lymphatic endothelium: a new frontier of metastasis research. *Nat Cell Biol.* 2002;4:E2–E5.
54. Stewart MW, Rosenfeld PJ. Predicted biological activity of intravitreal VEGF Trap. *Br J Ophthalmol.* 2008;92(5):667–668.

55. Stewart MW. What are the half-lives of ranibizumab and aflibercept (VEGF Trap-eye) in human eyes? Calculations with a mathematical model. *Eye Reports*. 2011;1:e5.
56. Holash J, Davis S, Papadopoulos N, et al. VEGF-Trap: a VEGF blocker with potent antitumor effects. *PNAS*. 2002;99(17):11392–11398.
57. Rakic JM, Lambert V, Munaut C, et al. Mice without uPA, tPA, or plasminogen genes are resistant to experimental choroidal neovascularization. *Invest Ophthalmol Vis Sci*. 2003;44(4):1732–1739.
58. Saishin Y, Saishin Y, Takahashi K, et al. VEGF-TRAP(R1R2) suppresses choroidal neovascularization and VEGF-induced breakdown of the blood-retinal barrier. *J Cell Physiol*. May 2003;195(2):241–248.
59. EYLEA™ (aflibercept) injection: US prescribing information. Tarrytown, NY, USA; Regeneron Pharmaceuticals, Inc. Available from: <http://www.regeneron.com/Eylea/eylea-fpi.pdf>. Accessed April 2, 2012.
60. Nguyen QD, Shah SM, Hafiz G, et al; CLEAR-AMD 1 Study Group. A phase I trial of an IV-administered vascular endothelial growth factor trap for treatment in patients with choroidal neovascularization due to age-related macular degeneration. *Ophthalmology*. 2006;113:1522.e1–1532.e14.
61. Do DV, Nguyen QD, Browning DJ, et al. Results of a phase I study of intravitreal VEGF Trap in subjects with diabetic macular edema: the CLEAR-IT DME Study. *IOVS*. 2007;48:ARVO E-abstract 1430/B486.
62. Nguyen QD, Shah SM, Browning DJ, et al. A phase I study of intravitreal vascular endothelial growth factor trap-eye in patients with neovascular age-related macular degeneration. *Ophthalmology*. 2009;116:2141–2148.
63. Brown DM, Heier JS, Challa T, et al; CLEAR-IT 2 Investigators. Primary endpoint results of a phase II study of vascular endothelial growth factor trap-eye in wet age-related macular degeneration. *Ophthalmology*. 2011;118(6):1089–1097.
64. Heier JS, Boyer D, Nguyen QD, et al; CLEAR-IT 2 Investigators. The 1-year results of CLEAR-IT 2, a phase 2 study of vascular endothelial growth factor trap-eye dosed as-needed after 12-week fixed dosing. *Ophthalmology*. 2011;118:1098–1106.
65. Cantrill HL. The diabetic retinopathy study and the early treatment diabetic retinopathy study. *International Ophthalmology Clinics*. 1984;24(4):13–29.
66. Stewart MW, Rosenfeld PJ, Penha FM, et al. Pharmacokinetic rationale for dosing every 2 weeks versus 4 weeks with intravitreal ranibizumab, bevacizumab, and aflibercept (vascular endothelial growth factor trap-eye). *Retina*. 2012;32(3):434–457.
67. Heier JS, Brown DM, Chong V, et al; VIEW 1 and VIEW 2 Study Groups. Intravitreal aflibercept (VEGF trap-eye) in wet age-related macular degeneration. *Ophthalmology*. 2012;119(12):2537–2548.
68. Sho K, Takahashi K, Yamada H, et al. Polypoidal choroidal vasculopathy: incidence, demographic features, and clinical characteristics. *Arch Ophthalmol*. 2003;121(10):1392–1396.
69. Squirrel DM, Bacon JF, Brand CS. To investigate the prevalence of polypoidal choroidal vasculopathy in presumed age-related peripapillary subretinal neovascular membranes. *Clin Experiment Ophthalmol*. 2009;37(4):368–372.
70. Regeneron Pharmaceuticals, Inc. Two Year Results of Phase 3 Studies with EYLEA™ (aflibercept) injection in wet AMD show sustained improvement in visual acuity [press release]. Tarrytown, NY: Regeneron Pharmaceuticals, Inc; December 5, 2011. Available from: <http://investor.regeneron.com/releasedetail.cfm?releaseid=629800>. Accessed June 27, 2013.
71. Regeneron Pharmaceuticals, Inc. Bayer and Regeneron report positive top-line results of two phase 3 studies with VEGF Trap-Eye in wet age-related macular degeneration [press release]. Tarrytown, NY: Regeneron Pharmaceuticals, Inc; November 22, 2010. Available from: <http://newsroom.regeneron.com/releasedetail.cfm?releaseid=532099>. Accessed June 27, 2013.
72. Antonetti DA, Barber AJ, Hollinger LA, Wolpert EB, Gardner TW. Vascular endothelial growth factor induces rapid phosphorylation of tight junction proteins occludin and zonula occluden 1. A potential mechanism for vascular permeability in diabetic retinopathy and tumors. *J Biol Chem*. 1999;274:23463–23476.
73. Pierce EA, Avery RL, Foley ED, Aiello LP, Smith LE. Vascular endothelial growth factor/vascular permeability factor expression in a mouse model of retinal neovascularization. *Proc Natl Acad Sci U S A*. 1995;92:905–909.
74. Rinaldi M, Chiosi F, Dell’Omo R, et al. Intravitreal pegaptanib sodium (Macugen®) for treatment of diabetic macular oedema: a morphologic and functional study. *Br J Clin Pharmacol*. 2012;74(6):940–946.
75. Bethke W. For DME, one size does not fit all [webpage on the Internet]. Review of Ophthalmology; August 8, 2011. Available from: <http://www.revophth.com/content/i/1599/c/29625/>. Accessed April 05, 2013.
76. ClinicalTrials. Available from: <http://www.clinicaltrials.gov/>. Accessed July 24, 2013.
77. Nguyen QD, Shah SM, Heier JS, et al; READ-2 Study Group. Primary endpoint (six-months) results of the Ranibizumab for Edema of the macula in Diabetes (READ-2) study. *Ophthalmology*. 2009;116:2175–2181.
78. Nguyen QD, Shah SM, Khwaja AA, et al; READ-2 Study Group. Two-year outcomes of the Ranibizumab for Edema of the macula in Diabetes (READ-2) study. *Ophthalmology*. 2010;117:2146–2151.
79. Massin P, Bandello F, Garweg JG, et al. Safety and efficacy of ranibizumab in diabetic macular edema (RESOLVE study): a 12-month, randomized, controlled, double-masked, multicenter phase II study. *Diabetes Care*. 2010;33:2399–2405.
80. Mitchell P, Bandello F, Schmidt-Erfurth U, et al; RESTORE Study Group. The RESTORE study: ranibizumab monotherapy or combined with laser versus laser monotherapy for diabetic macular edema. *Ophthalmology*. 2011;118:615–625.
81. Nguyen QD, Brown DM, Marcus DM, et al; RISE and RIDE Research Group. Ranibizumab for diabetic macular edema: results from 2 phase III randomized trials: RISE and RIDE. *Ophthalmology*. 2012;119:789–801.
82. Boyer DS, Rundle AC, Zhang J, Hopkins JJ, Ehrlich JS. Long-term efficacy and safety of ranibizumab in diabetic macular edema (DME): 36-month results from RISE and RIDE, two phase III clinical trials. In: 45th Annual Scientific Meeting of the Retina Society; October 5, 2012; Washington DC, USA.
83. Do DV, Schmidt-Erfurth U, Gonzalez VH, et al. The DAVINCI study: phase 2 primary results of VEGF Trap-Eye in patients with diabetic macular edema. *Ophthalmology*. 2011;118:1819–1826.
84. Do DV, Nguyen QD, Boyer D, et al; DAVINCI Study Group. One-year outcomes of the DAVINCI study of VEGF Trap-Eye in eyes with diabetic macular edema. *Ophthalmology*. 2012;119:1658–1665.
85. Regeneron Pharmaceuticals Inc. Regeneron and Bayer Initiate Phase 3 Trial of EYLEA® (aflibercept) Injection for the Treatment of Diabetic Macular Edema in Asia and Russia. Available from: <http://investor.regeneron.com/releasedetail.cfm?ReleaseID=741127>. Accessed July 24, 2013.
86. Regeneron Announces FDA Approval of EYLEA® (aflibercept) Injection For Macular Edema Following Central Retinal Vein Occlusion. Available from <http://investor.regeneron.com/releasedetail.cfm?releaseid=708835>. Accessed July 24, 2013.
87. Boyer D, Heier J, Brown DM, et al. Vascular endothelial growth factor Trap-Eye for macular edema secondary to central retinal vein occlusion: six-month results of the phase 3 COPERNICUS study. *Ophthalmology*. 2012;119:1024–1032.
88. Brown DM, Heier JS, Clark LW, et al. Intravitreal aflibercept injection for macular edema secondary to central vein occlusion: 1-year results from the phase 3 COPERNICUS study. *Am J Ophthalmol*. 2013;155:429–437.

89. Holz FG, Roider J, Ogura Y, et al. VEGF Trap-Eye for macular oedema secondary to central retinal vein occlusion: 6-month results of the phase III GALILEO study. *Br J Ophthalmol*. 2013;97:278–284.
90. Holz FG, Ogura Y, Roider J, et al. Intravitreal aflibercept injection for macular edema in central retinal vein occlusion: 1-year results of the phase 3 GALILEO Study [Poster 6929; online] Available from: <https://www.abstractsonline.com>.
91. US Food and Drug Administration. *Eylea*. US Food and Drug Administration. Available from: http://www.accessdata.fda.gov/drugsatfda_docs/label/2012/125387s004lbl.pdf. Accessed June 27, 2013.
92. Schaal S, Kaplan HJ, Tezel TH. Is there tachyphylaxis to intravitreal anti-vascular endothelial growth factor pharmacotherapy in age-related macular degeneration? *Ophthalmology*. 2008;115(12):2199–2205.
93. Forooghian F, Cukras C, Meyerle CB, Chew EY, Wong WT. Tachyphylaxis after intravitreal bevacizumab for exudative age-related macular degeneration. *Retina*. 2009;29(6):723–731.

Drug Design, Development and Therapy

Publish your work in this journal

Drug Design, Development and Therapy is an international, peer-reviewed open-access journal that spans the spectrum of drug design and development through to clinical applications. Clinical outcomes, patient safety, and programs for the development and effective, safe, and sustained use of medicines are a feature of the journal, which

Submit your manuscript here: <http://www.dovepress.com/drug-design-development-and-therapy-journal>

Dovepress

has also been accepted for indexing on PubMed Central. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.



Aflibercept versus placebo in combination with docetaxel and prednisone for treatment of men with metastatic castration-resistant prostate cancer (VENICE): a phase 3, double-blind randomised trial

Ian F Tannock, Karim Fizazi, Sergey Ivanov, Camilla Thellenberg Karlsson, Aude Fléchon, Iwona Skoneczna, Francisco Orlandi, Gwenaëlle Gravis, Vsevolod Matveev, Sevil Bavbek, Thierry Gil, Luciano Viana, Osvaldo Arén, Oleg Karyakin, Tony Elliott, Alison Birtle, Emmanuelle Magherini, Laurence Hattéville, Daniel Petrylak, Bertrand Tombal, Mark Rosenthal, on behalf of the VENICE investigators

Summary

Lancet Oncol 2013; 14: 760–68
Published Online
June 4, 2013
[http://dx.doi.org/10.1016/S1470-2045\(13\)70184-0](http://dx.doi.org/10.1016/S1470-2045(13)70184-0)
See Comment page 681

Background Docetaxel plus prednisone is standard first-line chemotherapy for men with metastatic castrate-resistant prostate cancer. Aflibercept is a recombinant human fusion protein that binds A and B isoforms of VEGF and placental growth factor, thereby inhibiting angiogenesis. We assessed whether the addition of aflibercept to docetaxel and prednisone would improve overall survival in men with metastatic castrate-resistant prostate cancer compared with the addition of placebo to docetaxel and prednisone.

Princess Margaret Cancer Centre, Toronto, Canada (Prof I F Tannock DSc); Institut Gustave Roussy, University of Paris Sud, Villejuif, France (Prof K Fizazi MD); Russian Science Centre of X-ray, Moscow, Russia (S Ivanov MD); Onkologiska Kliniken Norrlands Universitetssjukhus, Umeå, Sweden (C T Karlsson MD); CRLC Léon Bérard, Lyon, France (A Fléchon MD); Centrum Onkologii-Instytut, Warszawa, Poland (I Skoneczna MD); Oncomed, Providencia, Santiago, Chile (F Orlandi MD); Institut Paoli Calmettes, Marseille, France (G Gravis MD); NN Blokhin Russian Cancer Research Centre, Moscow, Russia (V Matveev MD); Istanbul Universitesi Onkoloji Enstitüsü Çapa, Istanbul, Turkey (Prof S Bavbek MD); Institut Jules Bordet, Université Libre de Bruxelles, Bruxelles, Belgium (T Gil MD); Fundacao Pio XII - Hospital de Cancer de Barretos, Barretos, Brazil (L Viana MD); Instituto Nacional del Cancer, Santiago, Chile (O Arén MD); Medical Radiological Research Center, Obninsk, Russia (Prof O Karyakin MD); Christie Hospital, Manchester, UK (T Elliott PhD); Rosemere Cancer Centre, Lancs Teaching Hospitals NHS Foundation Trust, Preston, UK (A Birtle MD); Sanofi R&D, Vitry sur Seine, France (E Magherini MD, L Hattéville MSc); Yale Cancer Center, New Haven, CT, USA (Prof D Petrylak MD); Cliniques

Methods VENICE was a phase 3, multicentre, randomised double-blind placebo-controlled parallel group study done in 31 countries (187 sites). Men with metastatic castrate-resistant prostate cancer, adequate organ function, and no prior chemotherapy were treated with docetaxel (75 mg/m² intravenously every 3 weeks) and oral prednisone (5 mg twice daily) and randomly allocated (1:1) to receive aflibercept (6 mg/kg) or placebo, intravenously, every 3 weeks. Treatment allocation was done centrally via an interactive voice response system, using a computer-generated sequence with a permuted-block size of four and stratified according Eastern Co-operative Group performance status (0–1 vs 2). Patients, investigators, and other individuals responsible for study conduct and data analysis were masked to treatment assignment. Aflibercept or placebo vials were supplied in identical boxes. The primary endpoint was overall survival using intention-to-treat analysis. This is the primary analysis of the completed trial. The study is registered with ClinicalTrials.gov, number NCT00519285

Findings Between Aug 17, 2007, and Feb 11, 2010, 1224 men were randomly allocated to treatment: 612 to each group. At final analysis, median follow-up was 35 months (IQR 29–41) and 873 men had died. Median overall survival was 22.1 months (95.6% CI 20.3–24.1) in the aflibercept group and 21.2 months (19.6–23.8) in the placebo group (stratified hazard ratio 0.94, 95.6% CI 0.82–1.08; p=0.38). We recorded a higher incidence of grade 3–4 gastrointestinal disorders (182 [30%] vs 48 [8.0%]), haemorrhagic events (32 [5.2%] vs ten [1.7%]), hypertension (81 [13%] vs 20 [3.3%]), fatigue (97 [16%] vs 46 [7.7%]), infections (123 [20%] vs 60 [10%]) and treatment-related fatal adverse events (21 [3.4%] vs nine [1.5%]) in the aflibercept group than in the placebo group.

Interpretation Aflibercept in combination with docetaxel and prednisone given as first-line chemotherapy for men with metastatic castrate-resistant prostate cancer resulted in no improvement in overall survival and added toxicity compared with placebo. Docetaxel plus prednisone remains the standard treatment for such men who need first-line chemotherapy.

Funding Sanofi and Regeneron Pharmaceuticals Inc.

Introduction

Most men with metastatic prostate cancer respond to androgen-deprivation therapy with orchiectomy or a gonadotropin-releasing hormone agonist, but the disease progresses to a castration-resistant state. Secondary responses to hormonal agents such as androgen-receptor inhibitors can occur after progression on primary androgen-deprivation therapy, and secondary responses to androgen-deprivation therapy might become more frequent as new inhibitors of androgen synthesis (eg, abiraterone acetate¹) or androgen-receptor signalling (eg, enzalutamide²) are used.

Men with metastatic castration-resistant prostate cancer can benefit from chemotherapy. Mitoxantrone was the first chemotherapy approved for such men, based on findings that showed improved pain control in trials that compared treatment with mitoxantrone and corticosteroid with corticosteroid alone.^{3,4} Since 2004, standard first-line chemotherapy has been docetaxel (75 mg/m² every 3 weeks) and oral prednisone or prednisolone (5 mg twice daily), after demonstration of improved survival compared with mitoxantrone and prednisone.^{5,6} Findings from this trial also showed better pain control, prostate-specific antigen (PSA), and

health-related quality-of-life response in men randomised to receive docetaxel and prednisone,⁵⁻⁷ and were lent support by findings from the SWOG-9916 trial, which also compared docetaxel-based chemotherapy with mitoxantrone and prednisone.⁸

Many phase 3 trials⁹⁻¹⁵ have assessed the addition of targeted agents to docetaxel plus prednisone in attempts to improve overall survival in men with metastatic castrate-resistant prostate cancer. Agents assessed include DN101 (high-dose calcitriol), G-VAX vaccine (Cell Genesys, South San Francisco, CA, USA), atrasentan, zibotentan (inhibitors of endothelin-1 receptor-A), lenalidomide, bevacizumab (an inhibitor of VEGF), and dasatinib (an inhibitor of the Src protein and other tyrosine kinases), but none of these agents improved survival, and all of them added toxicity. Another trial assessing docetaxel and prednisone with custirsen (OGX-011; an inhibitor of clusterin synthesis) is ongoing (NCT01188187).

Aflibercept (also known as VEGF-trap and ziv-aflibercept; Sanofi, Paris France; and Regeneron, Tarrytown, NY, USA) is a recombinant fusion protein consisting of extracellular domains of the human VEGF receptor (VEGFR) fused to the Fc portion of human immunoglobulin G1.^{16,17} It contains sequences encoding the immunoglobulin domain 2 from VEGFR1 fused to the immunoglobulin domain 3 from VEGFR2, which in turn is fused to the hinge region of the human immunoglobulin G1 Fc domain. Aflibercept has high binding affinity to the isoform VEGF-A, and also binds VEGF-B and platelet-derived growth factors PlGF1 and PlGF2, thereby inhibiting angiogenesis.¹⁶⁻¹⁸ Aflibercept has been assessed alone and with chemotherapy, including docetaxel, in preclinical models and showed activity against the DU 145 prostatic carcinoma in immune-deprived mice.¹⁸ Aflibercept has been assessed in phase 1 and 2 clinical trials with docetaxel,^{19,20} although no phase 2 trial of this combination has been done for men with metastatic castrate-resistant prostate cancer. We examined the combination of aflibercept with docetaxel and prednisone in the first-line treatment of such men. We aimed to show or exclude an improvement in overall survival compared with placebo with docetaxel and prednisone.

Methods

Trial design and participants

VENICE was a phase 3, multicentre randomised double-blind placebo-controlled parallel group study done in 31 countries (187 sites). Eligible participants had histologically or cytologically confirmed prostate cancer and evidence of metastatic disease that had progressed on hormonal therapy or after surgical castration. Criteria of progression before study entry were as defined by the Prostate Cancer Clinical Trials Working Group 2 (PCWG2):²¹ the requirements were an increase in measurable disease, new lesions (more than two lesions

if on bone scan), or two successive rises in serum PSA concentrations and an absolute value of 2 ng/mL or greater. Patients' serum testosterone needed to be 0.50 ng/mL or lower, and treatment with an agonist of luteinising-hormone-releasing hormone was continued in participants who were receiving it. If patients had initial complete androgen blockade, or had reduction in PSA for 3 months or more after addition of an anti-androgen, previous anti-androgen therapy was stopped 4-6 weeks before randomisation, depending on the agent used. Participants were required to have an Eastern Cooperative Group (ECOG) performance status of 0-2. Adequate organ and bone marrow function was determined by values of serum creatinine, aspartate, and alanine aminotransferases less than 1.5 × upper limit of normal, bilirubin within the normal range, haemoglobin concentrations of 100 g/L or greater, an absolute neutrophil count of 1.5×10^9 /L or greater, and a platelet count of 100×10^9 /L or greater.

Men were ineligible if they had received previous cytotoxic chemotherapy for prostate cancer, except estramustine or adjuvant or neoadjuvant treatment completed 3 years or more before enrolment. They could not have received inhibitors of VEGF or its receptors. Previous treatment with radiotherapy, surgery, or estramustine must have been completed 28 days or more before randomisation, but patients receiving bisphosphonates were eligible. Participants were ineligible if they had received previous isotope therapy (eg, strontium-89 or samarium-153), whole-pelvic irradiation, or previous radiotherapy to more than 30% of the bone marrow. Patients who had no history of brain metastases, uncontrolled spinal-cord compression, or carcinomatous meningitis, and no active prior malignancy (except basal or squamous-cell skin cancer) within the previous 5 years were eligible. Other exclusion criteria included myocardial infarction, severe or unstable angina pectoris, coronary or peripheral artery bypass graft, congestive heart failure, cerebrovascular accident, or transient ischaemic attack within the previous 6 months; treatment-resistant peptic ulcer disease, erosive oesophagitis or gastritis, infectious or inflammatory bowel disease, diverticulitis, pulmonary embolism, or uncontrolled hypertension within 3 months or deep-vein thrombosis within 4 weeks of randomisation. Patients were also ineligible if they had active bleeding or grade 3 neuropathy, but those receiving warfarin with good control of anticoagulation were eligible. Participants were excluded if they had AIDS or known HIV disease needing antiretroviral therapy or any severe acute or chronic medical disorder. Men with reproductive potential, or their partners, were required to use contraception.

The institutional review boards in all participating centres approved the study, and men were required to sign the institutional review board-approved informed consent form before enrolment. Trial data were

Universitaires Saint Luc, Bruxelles, Belgium (Prof B Tombal MD); and Royal Melbourne Hospital, Melbourne, Australia (Prof M Rosenthal PhD)

Correspondence to: Prof Ian F Tannock, Division of Medical Oncology and Hematology, Princess Margaret Cancer Centre, 610 University Avenue, Toronto, ON M5G 2M9, Canada
ian.tannock@uhn.ca

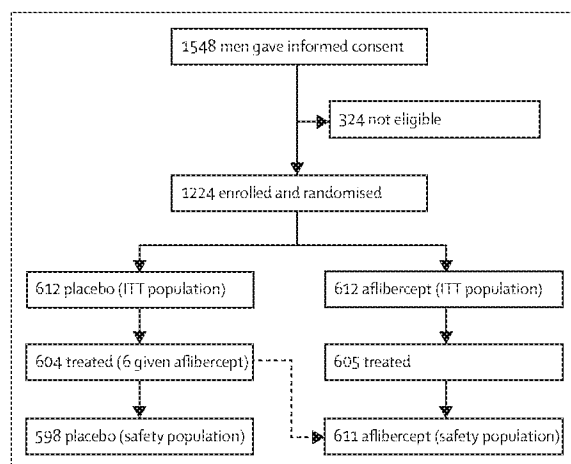


Figure 1: Trial profile

ITT=intention to treat.

monitored by an independent data monitoring committee, which met at 6-month intervals and provided recommendations to the study chair and trial committee.

Randomisation and masking

Generation of the patient randomisation list and management of treatment assignment was assigned to a third-party contractor: an interactive voice response system provider (S-CLINICA, Brussels, Belgium). Treatment was assigned centrally via the interactive voice response system, with patients randomly allocated in a one-to-one ratio to either the control or the experimental group, using a permuted-block size of four and stratified according to ECOG performance status (0–1 vs 2). There was strict adherence to the randomisation list and no corrections allowed during the course of the study. The treatment code was released for study analysis only after the database was locked on Feb 7, 2012. Aflibercept or placebo vials were supplied in identical boxes corresponding to patient kits: each kit was labelled with a unique kit number. Patients, investigators, and all other people responsible for study conduct and data analyses were masked to treatment assignment. During the course of the study, an external statistician (independent from the sponsor) did unmasked safety and efficacy analyses (interim analyses) for the purpose of the independent data monitoring committee reviews. Access to these data and analyses was restricted to the committee members. A site was able to break the code for safety reasons only in exceptional circumstances (when knowledge of the investigational product was essential for treating the patient) by calling the interactive voice response system. If treatment allocation was revealed, the treating physician was to document the date and reason for code breaking in the case report form, and the patient discontinued the investigational product.

Procedures

Docetaxel (75 mg/m²) and aflibercept (6 mg/kg) or matched placebo, were administered intravenously every 3 weeks. Prednisone or prednisolone (5 mg) were administered orally twice daily, from day 1 continuously. Dexamethasone (or another corticosteroid) was given before and after docetaxel, and standard anti-emetics were provided. Use of granulocyte-colony stimulating factor was at the discretion of the treating clinician. Aflibercept was administered in 5 mmol/L phosphate, 5 mmol/L citrate, 100 mmol/L sodium chloride, 20% (weight to volume) sucrose, and 0.1% (weight to volume) polysorbate 20, pH 6.0, supplied in sealed, sterile, single-use 5 mL vials; contents of the vial were diluted before infusion. Placebo consisted of sterile aqueous buffered vehicle pH 6.0, which was otherwise identical to aflibercept. Aflibercept or matching placebo were given over 1 h on day 1 every 3 weeks, followed immediately by docetaxel over 1 h. For patients with a body surface area greater than 2.2 m², the dose of docetaxel was adjusted to a maximum body surface area of 2.2 m².

Dose adjustments or cycle delays were planned in case of toxicity. Aflibercept or placebo could be reduced to 3 mg/kg in the case of grade 3 hypertension not controlled by added medication after 2 weeks; it was discontinued if there was on going grade 3 hypertension or grade 3–4 thromboembolic events, haemorrhage, or gastrointestinal perforation. Docetaxel could be reduced to 60 mg/m² and further to 45 mg/m² in the event of febrile neutropenia, or grade 3–4 thrombocytopenia, stomatitis, or diarrhoea; one dose reduction was allowed for grade 2 peripheral neuropathy, cutaneous reactions, or increased liver enzymes, but treatment with the drug was stopped if these persisted. Once a dose had been decreased, a patient's dose could not be increased back to the previous level. No more than one treatment omission, and no more than 2 weeks of delay in treatment, were allowed; drugs were discontinued in patients needing greater dose reductions or delays due to toxicity. No decision to discontinue treatment was to be made for increase in PSA or pain alone within the first 12 weeks. Patients were to be treated for at least 12 weeks in the absence of clinical evidence of disease progression until progressive disease, unacceptable toxicity, or patient refusal of further study treatment. All patients were followed-up until death or the study cutoff date, whichever came first.

The primary outcome was overall survival. Key secondary endpoints were PSA response, time to first skeletal-related event, and progression-free survival. PSA response was defined (for patients with baseline PSA ≥ 10 ng/mL) as a 50% or greater decrease of serum PSA concentrations from baseline, confirmed at least 3 weeks later; increases in PSA during the first 12 weeks were ignored in the assessment of PSA response, as per recommendations of PCWG2.²¹ Skeletal-related events included pathological fractures, spinal-cord compression,

need for bone irradiation (including radioisotopes or bone surgery), and change of treatment (eg, introduction or change in route of administration of bisphosphonates to treat bone pain). Progression-free survival was defined as a composite endpoint including tumour progression, PSA progression, occurrence of a skeletal-related event, pain progression, and radiotherapy for cancer-related symptoms or death, whichever came first. Other secondary endpoints included tumour response (assessed by Response Evaluation Criteria In Solid Tumors criteria²³), PSA progression-free survival (assessed by PCWG2 criteria²¹), and pain response and pain progression-free survival (assessed by the Present Pain Intensity Scale from the McGill-Melzack questionnaire and an analgesic consumption diary⁸). Data for health-related quality of life were obtained using the Functional Assessment of Cancer Therapy-Prostate²³ questionnaire and a Trial Outcome Index and will be reported separately.

Safety was assessed on the basis of treatment received by the frequency, severity, seriousness, and relation to treatment of treatment-emergent adverse events, which were assessed by clinical examination, including body weight, ECOG performance status and blood pressure, laboratory data (complete blood count, biochemistry, urinalysis, and other tests as indicated clinically), and concomitant drugs. We used the National Cancer Institute Common Terminology Criteria for Adverse Events (version 3.0) to grade severity of adverse events.

Statistical analysis

The primary analysis of overall survival was comparison between the two treatment groups using a log-rank test stratified at randomisation by ECOG performance status. The study was designed to detect a hazard ratio (HR) of 0.80 with 90% power, which required 873 deaths; the planned sample size was 1200. Estimates of the HR and its $(1-\alpha)\%$ CI (α being the two-sided nominal significance level of 0.044 at final analysis) were calculated with a Cox proportional hazard model stratified as described above; differences in overall survival were assessed by the stratified Kaplan-Meier method.

Exploratory analyses were done with Cox proportional hazard modelling to assess consistency of the treatment effect across subgroups and to examine the effect of covariates on overall survival. Covariates included age, ethnic origin, ECOG performance status, previous hypertension, on-going zoledronic acid treatment at study entry, time from initiation of hormonal therapy to progressive disease, and geographical region.

Two interim analyses of overall survival were planned for futility when 437 (50%) deaths had occurred and for early assessment of efficacy when 655 (75%) deaths had occurred. Type I and type II errors were protected by a group sequential approach with an O'Brien Fleming α -spending function and a non-binding gamma (-5) β -spending function.²⁴ The overall two-sided α nominal significance level for overall survival was 0.05.

	Aflibercept (N=612)	Placebo (N=612)
Age (years)	68 (43-88)	68 (40-87)
<65	195 (32%)	225 (37%)
65-74	183 (46%)	259 (42%)
≥75	134 (22%)	128 (21%)
White	560 (92%)	552 (90%)
Eastern Co-operative Group performance status		
0	283 (46%)	285 (47%)
1	303 (50%)	299 (49%)
2	26 (4.3%)	28 (4.6%)
Region		
Western Europe	227 (37%)	219 (36%)
Eastern Europe	132 (22%)	131 (21%)
North America	95 (16%)	81 (13%)
South America	71 (12%)	88 (14%)
Other	87 (14%)	93 (15%)
Previous prostatectomy	211 (35%)	214 (35%)
Previous radiotherapy		
Prostate	193 (32%)	202 (33%)
Bone	131 (21%)	134 (22%)
Stage at diagnosis		
Stage I-II	86 (14%)	82 (13%)
Stage III	81 (13%)	72 (12%)
Stage IV	321 (52%)	322 (53%)
Unknown	124 (20%)	136 (22%)
Gleason score		
2-4	22 (3.6%)	27 (4.4%)
5-7	263 (43%)	281 (46%)
8-10	295 (48%)	278 (45%)
Unknown	32 (5.2%)	26 (4.2%)
Years from diagnosis to randomisation	4.1 (0.4-18.6)	3.8 (0.3-21.5)
Previous hormonal therapy		
Number of regimens		
1	59 (10%)	73 (12%)
2	184 (30%)	167 (27%)
≥3	361 (59%)	361 (59%)
Unknown	8 (1.3%)	11 (1.8%)
Surgical castration	131 (21%)	147 (24%)
Gonadotropin-releasing hormone	526 (86%)	518 (85%)
Anti-androgen	567 (93%)	564 (92%)
Ketoconazole	37 (6.0%)	40 (6.5%)
Estramustine	32 (5.2%)	40 (6.5%)
Ongoing zoledronic acid	157 (26%)	165 (27%)
Criteria for progression		
Tumour progression	380 (62%)	372 (61%)
Increasing prostate-specific antigen concentration only	232 (38%)	239 (39%)
Sites of disease		
Bone	544 (89%)	541 (88%)
Lymph nodes	334 (55%)	330 (54%)
Visceral involvement	171 (28%)	176 (29%)
Prostate-specific antigen (ng/ml)	82.6 (0-6138)	92.9 (0-3821)
Data are median (range) or n (%).		

Table 1: Baseline characteristics

	Aflibercept (N=611)	Placebo (N=598)
Number of cycles administered	8 (1-50)	9 (1-36)
Number of cycles per patient		
1-3	137 (22%)	59 (10%)
4-6	131 (21%)	129 (22%)
7-9	131 (21%)	116 (19%)
10	62 (10%)	74 (12%)
11-15	98 (16%)	130 (22%)
>16	52 (8.5%)	90 (15%)
Median duration of treatment in weeks	24.0 (3-150)	28.9 (3-108)
Median relative dose intensity of aflibercept or placebo	0.97 (0.1-1.1)	0.99 (0.1-1.1)
Median relative dose intensity of docetaxel	0.93 (0.2-1.1)	0.97 (0.2-1.1)
Patients with at least one cycle delayed	302 (49%)	214 (36%)
Patients with at least one dose modification of aflibercept or placebo	65 (11%)	14 (2.3%)
Patients with at least one dose modification of docetaxel	189 (31%)	97 (16%)
Patients receiving granulocyte colony stimulating factor	150 (25%)	89 (15%)
Reason for treatment discontinuation		
Adverse event	266 (44%)	127 (21%)
Disease progression	186 (30%)	334 (56%)
Investigator decision	47 (7.7%)	75 (13%)
Other reason	112 (18%)	62 (10%)

Data are median (range) or n (%).

Table 2. Treatment received (safety population)

We used a closed-test hierarchical procedure to control the type I error rate when analysing key secondary endpoints; we did formal statistical testing only if the difference in prior endpoint was statistically significant. The procedure was done in the following order: overall survival (primary endpoint), PSA response, time to skeletal-related event, then progression-free survival. We used stratified Kaplan-Meier methods to assess differences in time-to-event endpoints and stratified Cochran-Mantel-Haenszel to test for comparison of response rates. We used SAS (version 9.2) for all statistical analyses.

This study is registered with ClinicalTrials.gov, number NCT00519285

Role of the funding source

The study was designed through collaboration between employees of the sponsor (including EM) and the study chair (IFT), and coauthors (MR, BT, and DP). During the study, the primary data were collected and managed by the sponsor (including EM and LH); at study conclusion, data were analysed by statisticians employed by the sponsor (including LH), and the detailed clinical study report was primarily drafted by the sponsor (including EM) and made available to the study chair (IFT). At the conclusion of the trial, authors employed by the sponsor (EM and LH) had full access to the raw data. The article was drafted by the study chair and corresponding author (IFT), and was amended after review by all authors. The

corresponding author had final responsibility to submit for publication.

Results

Between Aug 17, 2007, and Feb 11, 2010, 1224 patients with metastatic castrate-resistant prostate cancer were recruited from 187 centres in 31 countries: 612 patients were randomly allocated to each group (figure 1). Seven patients (1%) randomised to aflibercept and eight patients (1%) randomised to placebo were not treated: six patients in the placebo group received at least one dose of aflibercept in error (figure 1). The independent data monitoring committee recommended that the study continued without modification. The trial was analysed at median follow-up of 35 months (IQR 29-41) when 873 participants had died, as per the statistical plan.

Baseline demographic and clinical characteristics were much the same between the two groups (table 1). Most men had ECOG performance status 0-1 and most had received two or more lines of hormonal therapy. Most men had bone metastases, about half had lymph node metastases, and about a third had visceral disease (table 1).

More patients in the aflibercept group than in the placebo group stopped treatment because of an adverse event, resulting in a larger number of patients in the aflibercept than in the placebo group receiving three or fewer treatment cycles (table 2). Patients in the aflibercept group also had more delays and dose adjustments for both aflibercept and docetaxel, although the median dose intensity for aflibercept with placebo and docetaxel, calculated as a percentage of the ideal dose between first and last treatments, exceeded 93%. Further anticancer treatments administered after discontinuation of study treatment were much the same between treatment groups (appendix).

We recorded no between-group difference in median overall survival (HR for overall survival 0.94 [95.6% CI 0.82-1.08]; table 3 and figure 2). Exploratory analysis also showed survival to be similar for subgroups defined by ECOG performance status, age, geographical region, and other patient characteristics (data not shown).

More patients in the aflibercept group had a tumour response than did those in the placebo group, but we detected no between-group difference in any other of the secondary outcomes (tables 3 and 4), including time to first skeletal-related event and progression-free survival (table 3).

We recorded greater toxicity in the aflibercept group: grade 3-4 toxicities that were most common included gastrointestinal disorders such as diarrhoea, nausea and vomiting, stomatitis and ulceration, perforation, and fistula (table 5). Haemorrhage (mostly epistaxis), hypertension, fatigue, infection, neutropenia and its complications, dysphonia, and proteinuria were also more common in the aflibercept group than in the placebo group. The incidence of neuropathy and thromboembolic events was much the same between the

	Aflibercept		Placebo		p value
	Number assessable	Median (95% CI)	Number assessable	Median (95% CI)	
Median overall survival (months)	612 patients (428 deaths)	22.1 (95.6% CI 20.3–24.1)	612 patients (445 deaths)	21.2 (95.6% CI 19.6–23.8)	0.38
Time to first skeletal-related event (months)	612 patients (497 events)	15.3 (14.1–16.7)	612 patients (516 events)	15.0 (13.7–16.4)	0.31
PFS (months)	612 patients (592 events)	6.9 (6.2–7.4)	612 patients (592 events)	6.2 (5.6–6.9)	0.31
PSA-PFS (months)	608 patients (567 events)	8.3 (7.8–8.8)	606 patients (571 events)	8.1 (7.6–8.6)	0.42
Pain-PFS (months)	287 patients (244 events)	9.2 (8.2–10.4)	201 patients (263 events)	9.7 (8.5–11.5)	0.87

Unless otherwise indicated figures in parentheses represent 95% confidence intervals. Except for the primary endpoint of overall survival, p values are descriptive and exploratory; they are not corrected for multiplicity. PFS=progression-free survival. PSA=prostate-specific antigen.

Table 3: Time to event efficacy outcomes, by treatment group

two groups. 34 (5.6%) patients in the aflibercept group and 20 (3.3%) died in the absence of disease progression, with 21 (3.4%) patients in the aflibercept group and nine (1.5%) in the placebo group judged to be related to treatment. The most common cause of fatal events in the absence of disease progression (and of the difference between the group) was infection (15 [2.5%] with aflibercept and four [0.7%] with placebo); other causes of fatal adverse events, including cardiac events, respiratory problems, and gastrointestinal events (including haemorrhage), were evenly distributed between the two groups. The median age of men with a fatal adverse event was 74 years (IQR 70–77) (compared with 68 years [62–74] for all participants).

Discussion

Findings from this large, international phase 3 trial showed no improvement in overall survival compared with placebo when aflibercept was added to docetaxel and prednisone to treat men with metastatic castrate-resistant prostate cancer. The median survival in both groups (21–22 months) was longer than anticipated for docetaxel and prednisone at the time the trial was designed (19 months, based on the results of the TAX327 trial⁵). This better-than-expected survival of men with metastatic castrate-resistant prostate cancer given chemotherapy probably reflects that men included in recent clinical trials are offered chemotherapy earlier in the course of their disease; it was probably not due to availability of more effective treatments after progression of disease on docetaxel, because few patients received new agents such as abiraterone, enzalutamide, or cabazitaxel that have been shown to improve survival (appendix).^{1,2,25}

A limitation of our trial, and indeed of most clinical trials, is that entry criteria excluded patients with various types of comorbidity, which are common in elderly men with prostate cancer. Also, targeted agents generally add toxicity, and such toxicity is often underestimated from data in the original clinical trials that led to licensing of these agents.²⁶ Entry criteria for this trial were broadly similar to those for the TAX327 trial, which established docetaxel plus prednisone as standard first-line

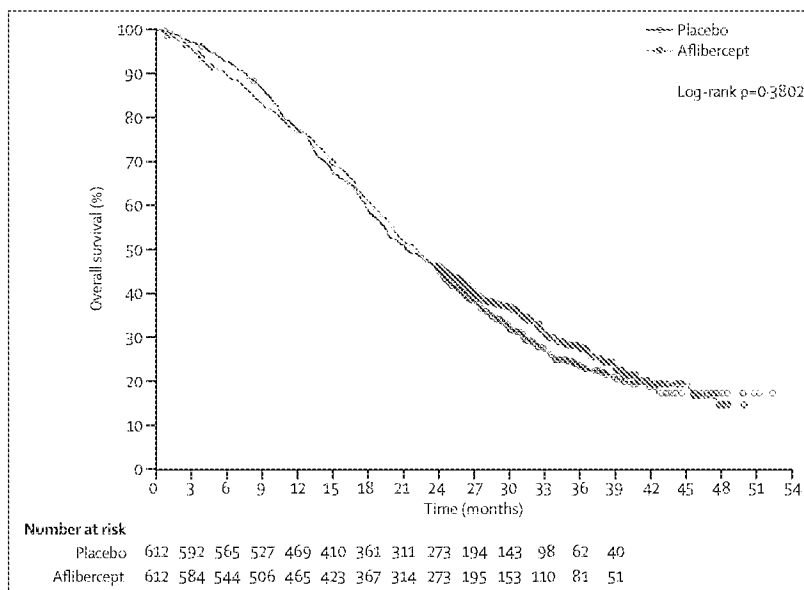


Figure 2: Kaplan-Meier curves for overall survival

	Aflibercept		Placebo		p value
	Number of events/number assessable	% (95% CI)	Number of events/number assessable	% (95% CI)	
PSA response	384/560	68.6 (64.7–72.4)	355/559	63.5 (59.5–67.5)	0.075
tumour response	124/323	38.4 (33.4–44.0)	90/320	28.1 (23.2–33.1)	0.0043
Pain response	24/67	35.8 (24.3–47.3)	31/67	46.3 (34.3–58.2)	0.20

p values are descriptive and exploratory, they are not corrected for multiplicity. PSA=prostate-specific antigen.

Table 4: Other secondary outcomes

chemotherapy for men with metastatic castrate-resistant prostate cancer (panel). In a comparison of men with such cancer receiving first-line docetaxel plus prednisone within and outside of clinical trials at the Princess

	Aflibercept (N=611)		Placebo (N=598)	
	All grades	Grades 3-4	All grades	Grades 3-4
Any	607 (99%)	470 (77%)	585 (98%)	290 (49%)
Gastrointestinal disorders	513 (84%)	182 (30%)	409 (68%)	48 (8%)
Diarrhoea	276 (45%)	35 (6%)	215 (36%)	20 (3%)
Nausea and vomiting	198 (32%)	17 (3%)	193 (32%)	4 (<1%)
Stomatitis and ulceration	345 (57%)	89 (15%)	125 (21%)	7 (1%)
Perforation	16 (3%)	16 (3%)	1 (<1%)	1 (<1%)
Fistula	14 (2%)	5 (<1%)	1 (<1%)	0
Haemorrhagic events	270 (44%)	22 (5%)	142 (24%)	30 (2%)
Epistaxis	208 (34%)	14 (2%)	56 (9%)	0
Gastrointestinal haemorrhage	62 (10%)	12 (2%)	34 (6%)	6 (1%)
Vascular disorders	264 (43%)	97 (16%)	142 (24%)	43 (7%)
Hypertension	218 (36%)	81 (13%)	69 (12%)	20 (3%)
Venous thrombotic events	23 (4%)	21 (3%)	36 (6%)	35 (6%)
Arterial thrombotic events	12 (2%)	7 (1%)	18 (3%)	15 (3%)
Asthenia or fatigue	376 (62%)	97 (16%)	349 (58%)	46 (8%)
Infection	338 (55%)	123 (20%)	251 (42%)	60 (10%)
Upper respiratory tract infections	97 (16%)	4 (<1%)	84 (14%)	3 (<1%)
Fever	68 (11%)	5 (<1%)	58 (10%)	1 (<1%)
Neutropenic complications	92 (15%)	87 (14%)	42 (7%)	41 (7%)
Oedema	69 (11%)	3 (<1%)	170 (28%)	5 (<1%)
Alopecia	224 (37%)	0	269 (45%)	0
Nail changes	156 (26%)	14 (2%)	153 (26%)	2 (<1%)
Nervous system	320 (52%)	58 (10%)	325 (54%)	56 (9%)
Headache	97 (16%)	4 (<1%)	45 (8%)	0
Peripheral neuropathy	111 (18%)	14 (2%)	156 (26%)	16 (3%)
Respiratory	409 (67%)	65 (11%)	236 (40%)	24 (4%)
Dysphoria	230 (38%)	3 (<1%)	36 (6%)	0
Breathing abnormalities	103 (17%)	14 (2%)	76 (13%)	4 (<1%)
Cough	116 (19%)	4 (<1%)	85 (14%)	0
Musculoskeletal	229 (38%)	27 (4%)	286 (48%)	37 (6%)
Appetite disorders	192 (31%)	12 (2%)	111 (19%)	5 (<1%)
Eye problems	163 (27%)	7 (1%)	108 (18%)	3 (<1%)
Proteinuria	275 (45%)	38 (7%)	214 (36%)	7 (1%)

Data are number of patients (%). We included all side-effects for which the frequency of grade 3-4 events was greater than 5% in either group, and those expected with inhibitors of angiogenesis.

Table 5. Treatment-emergent adverse events

Margaret Cancer Centre (Toronto, Canada), median survival was longer for men treated as part of a clinical trial,²⁷ suggesting that the median survival of men in the VENICE trial is probably longer than would be achieved in general oncological practice.

There were no differences in secondary time-to-event endpoints, including time to first skeletal-related event and progression-free survival, between the study groups. However, in patients treated with aflibercept, we detected more tumour responses than in those given placebo, and although not statistically significant, PSA response was greater with aflibercept than it was with placebo. Aflibercept added toxicity and led to more study discontinuations than placebo. The data have not been analysed to compare the proportion of men who

Panel: Research in context

Systematic review

We did a systematic review of PubMed using the search terms "docetaxel", "metastatic", "prostate cancer", and "randomized trial". We identified two phase 3 trials: TAX327 and SWOG 99-16. Findings from these trials showed improved survival in men with metastatic castration-resistant prostate cancer who received chemotherapy with docetaxel and prednisone (TAX327) or docetaxel and estramustine (SWOG 99-16), compared with the previous approved standard of mitoxantrone and prednisone; docetaxel given intravenously every 3 weeks with daily oral prednisone or prednisolone (approved as standard first-line chemotherapy for metastatic castrate-resistant prostate cancer). A systematic review in February, 2013, of PubMed, Google, and meeting reports of ASCO and ESMO using the same search terms identified seven large phase 3 trials that attempted, unsuccessfully, to improve overall survival in men with castrate-resistant prostate cancer by combining docetaxel with a targeted agent. One of these negative trials assessed bevacizumab, an anti-angiogenic agent that inhibits VEGF.

Interpretation

Our findings showed that aflibercept, an inhibitor of angiogenesis that inhibits a broader spectrum of angiogenic growth factors than bevacizumab, did not increase overall survival in men with metastatic castrate-resistant prostate cancer when combined with docetaxel and prednisone, but increased toxicity. The standard first-line chemotherapy treatment for such men remains docetaxel with prednisone or prednisolone.

had predefined levels of improvement in health-related quality of life in the two groups of the study, and this will be reported subsequently. However, that the addition of aflibercept to docetaxel and prednisone would improve health-related quality of life seems unlikely in view of the greater toxicity with aflibercept.

Both the primary tumour and metastases from prostate (and other) cancers require angiogenesis to provide nutrients and thereby allow cancer cells to survive and proliferate. Aflibercept inhibits a broader spectrum of angiogenic growth factors (VEGF-A, VEGF-B, PlGF1, and PlGF2)^{16,17} than does bevacizumab, and we expected that this broader spectrum would lead to more effective inhibition of angiogenesis and improved efficacy compared with placebo. The greater number of tumour responses and non-statistically significantly higher PSA response in the aflibercept group suggest that aflibercept has biological activity, and appropriate biomarkers could identify subpopulations that would benefit from treatment. Potential relations between putative biomarkers and efficacy or safety will be assessed by the investigators; however, concentrations of VEGF have not been

associated with efficacy in trials assessing bevacizumab.²⁸ Many growth factors stimulate angiogenesis, and activation of alternate angiogenic pathways is a potential cause of resistance to VEGF inhibitors. In attempts to inhibit multiple points of the angiogenic pathways simultaneously, receptor tyrosine kinase inhibitors in combination with bevacizumab have been studied, but have resulted in additional toxicity without therapeutic benefit.²⁹

The additional toxicity seen when aflibercept was added to docetaxel led to a higher proportion of patients completing fewer than four treatment cycles (22% vs 10%) and to more frequent discontinuation of study treatment (44% vs 21%) than with placebo. The shorter treatment duration might have prevented differences in potential anti-tumour activity from translating into improvements in overall survival and other time-to-event endpoints. The proportion of fatal adverse events that were probably due to toxicity of treatment was 3.4% in the aflibercept group and 1.5% in the placebo group. The number of toxic deaths in the control group was slightly higher than in previous phase 3 studies, in which the incidence of fatal adverse events varied between 0% and 1.2% for patients receiving docetaxel and prednisone,^{3,9,13} and was mainly due to infection. The occurrence of fatal toxic events in the aflibercept group was similar to the 4% noted for men receiving docetaxel and bevacizumab in the only other study that assessed an inhibitor of angiogenesis, CALGB 90401,¹³ and the types of grade 3–4 toxicity in the aflibercept group were similar to those reported for bevacizumab. The age distribution of patients treated in these studies is similar, although in the present study men with a fatal adverse event were older and more likely to have a previous cardiovascular risk factor than were those who did not have a fatal adverse event.

Aflibercept has been assessed with standard chemotherapy in other large placebo-controlled phase 3 clinical trials for metastatic colorectal, pancreatic, and lung cancer.^{30–32} Only the trial in second-line metastatic colorectal cancer, in which aflibercept was used in combination with FOLFIRI (the VELOUR study³⁰) resulted improved overall survival (a median of about 6 weeks), which led to approval of aflibercept by the US Food and Drug Administration for this indication.

Nine large phase 3 trials (including this trial) have now assessed docetaxel and prednisone with or without a targeted agent for men with metastatic castrate-resistant prostate cancer, and findings from the eight reported trials (findings from the SYNERGY trial are awaited; NCT01188187) have shown no difference in survival and increased toxicity in most of the experimental groups (table 6).^{9,15} Together, these eight trials recruited 7800 men and probably cost close to US\$1 billion. The VENICE trial was based on minimal preclinical and early clinical data, and the decision to proceed rapidly to phase 3 was based

	Number of participants	Partner drug	Result
ASCENT II ⁹	353	DN103 (Calcitriol)	Poorer survival in experimental group versus control Increased toxicity
VITAL II ¹⁰	408	GVAX vaccine	Poorer survival in experimental group versus control
SWOG S0421 ¹¹	991	Atrasentan	No difference in PFS or survival
ENTHUSE ¹²	594	Zibotentan	No between-group difference in survival Increased toxicity
CALGB 90401 ¹³	1050	Bevacizumab	No between-group difference in survival (better PFS) Increased toxicity
MAINSAIL ¹⁴	1059	Lenalidomide	No between-group difference in survival Increased toxicity
READY ¹⁵	1522	Dasatinib	No between-group difference in survival or other endpoints Increased toxicity
VENICE (present study)	1224	Aflibercept (VEGF-Trap)	No between-group difference in survival Increased toxicity
SYNERGY ¹⁶		Custirsen (OGX-011)	Study not completed (no data available)

PFS=progression-free survival. *NCT01188187

Table 6: Trials comparing docetaxel plus prednisone alone or with a targeted agent in men with metastatic castrate-resistant prostate cancer

on the expectation that anti-angiogenic agents would be effective and that docetaxel, prednisone, and bevacizumab would become the new standard of care. Several of the other trials of docetaxel and prednisone were based on minimal preclinical or early clinical data.^{10–15} Prostate cancer investigators and sponsors should learn from this experience: future large trials should proceed only after rigorous preclinical and phase 2 data show substantial preliminary evidence of benefit, and they should have tighter criteria for early stopping if substantial benefit can be excluded at interim analyses.

Contributors

IFT (study chair) and MR, BT, and DP (cochairs), along with Sanofi employees (including EM and LH) contributed to the study design, study conduct, and study follow-up to the final analysis. Sanofi employees (including EM and LH) did the interim and final analyses and provided interpretation of the results. The detailed clinical study report was drafted primarily by the sponsor (including EM) and made available to IFT, who approved it. IFT drafted the paper, and contributed to subsequent drafts with all authors. IFT, KF, SI, CTK, AF, IS, FO, GG, VM, SB, TG, LV, OA, AK, TE, AB, BT, and MR were investigators in the study and contributed to the recruitment, treatment of the patients, and collection of the data.

Conflicts of interest

Sanofi funded this trial. KF, AF, IS, FO, GG, SB, TE, AB, DP, BT, and MR have acted as paid consultants to Sanofi. IFT has received research funding from Sanofi. EM and LH are employees of Sanofi and hold stock in the company. SI, CTK, VM, TG, LV, OA, and OK declare that they have no conflicts of interest.

Acknowledgments

This study was funded by Sanofi and Regeneron Pharmaceuticals Inc.

References

- de Bono JS, Logothetis CJ, Molina A, et al. Abiraterone and increased survival in metastatic prostate cancer *N Engl J Med* 2011; **364**: 1995–2005.
- Scher HI, Fizazi K, Saad F, et al. Increased survival with enzalutamide in prostate cancer after chemotherapy. *N Engl J Med* 2012; **367**: 1187–97.

- 3 Tannock IF, Osoba D, Stockler MR, et al. Chemotherapy with mitoxantrone plus prednisone or prednisone alone for symptomatic hormone-resistant prostate cancer: a Canadian randomized trial with palliative end points. *J Clin Oncol* 1996; **14**: 1756–64.
- 4 Kantoff PW, Halabi S, Conaway M, et al. Hydrocortisone with or without mitoxantrone in men with hormone-refractory prostate cancer: results of the cancer and leukemia group B 9182 study. *J Clin Oncol* 1999; **17**: 2506–13.
- 5 Tannock IF, de Wit R, Berry WR, et al. Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer. *N Engl J Med* 2004; **351**: 1502–12.
- 6 Berthold DR, Pond GR, Soban F, et al. Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer: updated survival in the TAX327 study. *J Clin Oncol* 2008; **26**: 242–45.
- 7 Berthold DR, Pond GR, Roessner M, et al. Treatment of hormone-refractory prostate cancer with docetaxel or mitoxantrone: relationships between prostate-specific antigen, pain, and quality of life response and survival in the TAX-327 study. *Clin Cancer Res* 2008; **14**: 2763–67.
- 8 Petrylak DP, Tangen CM, Hussain MH, et al. Docetaxel and estramustine compared with mitoxantrone and prednisone for advanced refractory prostate cancer. *N Engl J Med* 2004; **351**: 1513–20.
- 9 Scher HI, Jia X, Chi K, et al. Randomized, open label phase III trial of docetaxel plus high-dose calcitriol versus docetaxel plus prednisone for patients with castration-resistant prostate cancer. *J Clin Oncol* 2011; **29**: 2191–98.
- 10 Small E, Demkow T, Gerritsen WR, et al. A phase III trial of GVAX immunotherapy for prostate cancer in combination with docetaxel versus docetaxel plus prednisone in symptomatic, castration-resistant prostate cancer. ASCO Genitourinary Cancers Symposium; abstr 7. <http://meetinglibrary.asco.org/content/20295-64> (accessed May 11, 2013).
- 11 Quinn DI, Tangen CM, Hussain M, et al. SWOG S0421: Phase III study of docetaxel and abiraterone versus docetaxel and placebo for men with advanced castrate resistant prostate cancer. *Proc Am Soc Clin Oncol* 2012; **30** (suppl): abstr 4511.
- 12 Fizazi K, Higano C, Nelson J, et al. Phase III, randomized, placebo-controlled study of docetaxel in combination with zibotentan (ZD4054) in patients with metastatic castration-resistant prostate cancer. *J Clin Oncol* (in press).
- 13 Kelly WK, Halabi S, Carducci M, et al. Randomized, double-blind, placebo-controlled phase III trial comparing docetaxel and prednisone with or without bevacizumab in men with metastatic castration-resistant prostate cancer: CALGB 90401. *J Clin Oncol* 2012; **30**: 1534–40.
- 14 Petrylak DP, Fizazi K, Sternberg C, et al. LBA24—a phase 3 study to evaluate the efficacy and safety of docetaxel and prednisone with or without lenalidomide in patients with castrate-resistant prostate cancer: the MAINSAIL trial. *Annals Oncol* 2012; **23**(suppl 9): LBA24.
- 15 Araujo JC, Trudel GC, Saad F, et al. Overall survival (OS) and safety of dasatinib/docetaxel versus docetaxel in patients with metastatic castration-resistant prostate cancer (mCRPC): Results from the randomized phase III READY trial. *Proc Am Soc Clin Oncol* 2013; **31** (suppl 6): abstr LBAS.
- 16 Holash J, Davis S, Pispadopoulos N, et al. VEGFTrap: a VEGF blocker with potent antitumor effects. *Proc Natl Acad Sci USA* 2002; **99**: 11393–98.
- 17 Lassoued W, Murphy D, Tsai J, Queslari R, Thurston G, Lee WM. Effect of VEGF and VEGF Trap on vascular endothelial cell signaling in tumors. *Cancer Biol Ther* 2011; **10**: 1326–33.
- 18 Chiron M, Lejeune P, Pascale B, Bladt F, Vrignaud P, Bissery M-C. Broad spectrum of antitumor activity of aflibercept (VEGF Trap) in tumor-bearing mice. American Association for Cancer Research Annual Meeting; San Diego, CA, USA; April 12–16, 2008; abstract 380.
- 19 Coleman RL, Duska LR, Ramirez PT, et al. Phase 1–2 study of docetaxel plus aflibercept in patients with recurrent ovarian, primary peritoneal, or fallopian tube cancer. *Lancet Oncol* 2011; **12**: 1109–17.
- 20 Isambert N, Freyer G, Zanetta S, et al. Phase I dose-escalation study of intravenous aflibercept in combination with docetaxel in patients with advanced solid tumors. *Clin Cancer Res* 2012; **18**: 1743–50.
- 21 Scher HI, Halabi S, Tannock I, et al. Design and end points of clinical trials for patients with progressive prostate cancer and castrate levels of testosterone: recommendations of the Prostate Cancer Clinical Trials Working Group. *J Clin Oncol* 2008; **26**: 1148–59.
- 22 Therasse P, Arbuck SG, Eisenhauer EA, et al. New guidelines to evaluate the response to treatment in solid tumors. European Organization for Research and Treatment of Cancer, National Cancer Institute of the United States, National Cancer Institute of Canada. *J Natl Cancer Inst* 2000; **92**: 205–16.
- 23 Esper P, Mo F, Chodak G, Sinner M, Cella D, Pienta KJ. Measuring quality of life in men with prostate cancer using the functional assessment of cancer therapy-prostate instrument. *Urology* 1997; **50**: 920–28.
- 24 Fleming TR, Harrington DP, O'Brien PC. Designs for group sequential tests. *Control Clin Trials* 1984; **5**: 348–61.
- 25 De Bono JS, Oudard S, Ozguroglu M, et al. Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: a randomised open-label trial. *Lancet* 2010; **376**: 1147–54.
- 26 Seruga B, Sterling L, Wang L, Tannock IF. Reporting of serious adverse drug reactions of targeted anticancer agents in pivotal phase III clinical trials. *J Clin Oncol* 2011; **29**: 174–85.
- 27 Templeton A, Wang L, Vera-Badillo F et al. From trial to practice: The Princess Margaret Hospital experience with docetaxel and prednisone for men with metastatic castration resistant prostate cancer. *Proc Am Soc Clin Oncol* 2013; **31** (suppl 6): abstr 125.
- 28 Jubb AM, Harris AL. Biomarkers to predict the clinical efficacy of bevacizumab in cancer. *Lancet Oncol* 2010; **11**: 1172–83.
- 29 Sosman J, Puzanov I. Combination targeted therapy in advanced renal cell carcinoma. *Cancer* 2009; **115**: 2368–75.
- 30 Van Cutsem E, Tabernero J, Lakomy R, et al. Addition of aflibercept to fluorouracil, leucovorin, and irinotecan improves survival in a phase III randomized trial in patients with metastatic colorectal cancer previously treated with an oxaliplatin-based regimen. *J Clin Oncol* 2012; **30**: 3499–506.
- 31 Rougier P, Riess H, Manges R, et al. Randomised, placebo-controlled, double-blind, parallel-group phase III study evaluating aflibercept in patients receiving first-line treatment with gemcitabine for metastatic pancreatic cancer. *Eur J Cancer* 2013; published online April 30. DOI: 10.1016/j.ejca.2013.04.002.
- 32 Ramlau R, Gorbunova V, Ciuleanu TE, et al. Aflibercept and docetaxel versus docetaxel alone after platinum failure in patients with advanced or metastatic non-small-cell lung cancer: a randomized, controlled phase III trial. *J Clin Oncol* 2012; **30**: 3640–47.

REVIEW

Complementary actions of VEGF and Angiopoietin-1 on blood vessel growth and leakage*

Gavin Thurston

Regeneron Pharmaceuticals Inc, 777 Old Saw Mill River Road, Tarrytown, NY 10591, USA

Abstract

Vascular endothelial growth factor (VEGF) and Angiopoietins are families of vascular-specific growth factors that regulate blood vessel growth, maturation and function. To learn more about the effects of these factors *in vivo*, we have overexpressed VEGF-A or Angiopoietin-1 (Ang1) in two systems in mice, and examined the effects on blood vessel growth and function. In one set of studies, VEGF, Ang1, or both factors, were transgenically overexpressed in the skin under the keratin-14 (K14) promoter. The skin of mice overexpressing VEGF (K14-VEGF) had numerous tortuous, capillary-sized vessels which were leaky to the plasma tracer Evans blue under baseline conditions. In contrast, the skin of mice overexpressing Ang1 (K14-Ang1) had enlarged dermal vessels without a significant increase in vessel number. These enlarged vessels were less leaky than those of wild-type mice in response to inflammatory stimuli. In double transgenic mice overexpressing VEGF and Ang1, the size and number of skin vessels were both increased; however, the vessels were not leaky. In a second set of studies, VEGF or Ang1 was systemically delivered using an adenoviral approach. Intravenous injection of adenovirus encoding VEGF (Adeno-VEGF) resulted in widespread tissue oedema within 1–2 days after administration, whereas injection of Adeno-Ang1 resulted in the skin vessels becoming less leaky in response to topical inflammatory stimuli or local injection of VEGF. The decreased leakage was not accompanied by morphological changes. Thus, overexpressing VEGF appears to promote growth of new vessels accompanied by plasma leakage, whereas overexpressing Ang1 promotes the enlargement of existing vessels and a resistance to leakage. Further understanding of the interrelationship of these factors during normal development could lead to their application in the treatment of ischaemic diseases.

Key words adenovirus; angiogenesis; inflammation; Tie-2 receptor; transgenic mice; vascular endothelial growth factor; vascular leakage.

Introduction

Two families of endothelial-specific growth factors, vascular endothelial growth factors and Angiopoietins, are necessary for the formation of blood vessels. These factors seem to act in co-ordinated and complementary ways to produce mature blood vessels. Vascular

endothelial growth factor A (VEGF-A, here called VEGF), the initial member of the VEGF family to be identified (Ferrara et al. 1991; Dvorak et al. 1992), is essential for early blood vessel formation and angiogenesis. Mice deficient in even one allele for VEGF die in embryogenesis due to a decrease in endothelial cell number and severe defects in blood vessel formation (Carmeliet et al. 1996; Ferrara et al. 1996). Angiopoietin-1, the first member of a second family of endothelial-specific factors (Davis et al. 1996; Maisonpierre et al. 1997; Valenzuela et al. 1999), is essential for a later stage of blood vessel formation. Mice deficient for Ang1 die by embryonic day 12.5 (E12.5) due to defects in vessel remodelling and maturation (Suri et al. 1996). VEGF and Ang1 act via distinct endothelial-cell-specific

Correspondence

Dr Gavin Thurston, Regeneron Pharmaceuticals Inc, 777 Old Saw Mill River Road, Tarrytown, NY 10591, USA. Tel. +1 914 345 7575; fax: +1 914 347 5045; e-mail: Gavin.Thurston@Regeneron.com

*From a paper presented at an Anatomical Society of Great Britain and Ireland symposium on the modulation of endothelial cell permeability, Royal Holloway College, UK, January 2002

Accepted for publication 24 April 2002

tyrosine kinase receptors, which are also essential for embryogenesis and blood vessel formation. Deletion of the VEGF receptor VEGF-R2 (Fong et al. 1995; Shalaby et al. 1995) or the Angiotensin receptor (Tie-2) (Dumont et al. 1994; Sato et al. 1995) is also lethal to the embryo, with phenotypes broadly similar to those in the corresponding ligand-deficient mice.

Thus a model has evolved to describe the role of these factors in developmental angiogenesis. In particular, VEGF-A and its endothelial cell receptor VEGF-R2 are believed to play a role in vasculogenesis and early angiogenesis, whereas Ang1 and its receptor are believed to be involved in blood vessel remodelling and maturation (Hanahan, 1997; Yancopoulos et al. 2000). However, because of the early lethality of the gene-targeted embryos, it has been difficult to fully define the role of these factors in adult and pathological angiogenesis.

To gain further insight into the function of VEGF and Ang1 *in vivo*, we have used two approaches to overexpress these factors in mice. In particular, we have used tissue-specific transgenic mice and adenoviral vectors. In the transgenic approach, VEGF or Ang1 was

overexpressed continuously in the mouse skin from mid-embryonic stages into adulthood. The factors, and their effects, were localized to the skin (Fig. 1A). In the adenoviral approach, VEGF or Ang1 was expressed systemically in otherwise normal adult mice, and the factors acted for a defined duration on blood vessels throughout the mouse (Fig. 1B). Comparing and contrasting these different approaches and the different factors has helped reveal the distinct actions of VEGF and Ang1 on blood vessel growth and leakage.

Transgenic overexpression – K14-VEGF and K14-Ang1 mice

Several groups have generated mice which overexpress VEGF-A in the skin, under either K14 or K5 promoters (Detmar et al. 1998; Larcher et al. 1998; Thurston et al. 1999). We generated K14-VEGF₁₆₄ transgenic mice (Thurston et al. 1999), which appeared normal but had some redness in the skin of the ears and snout. The epidermis of the K14-VEGF mice was thickened, and the dermis contained infiltrating leucocytes. Lesions appeared in the ear skin of older mice (Thurston et al.

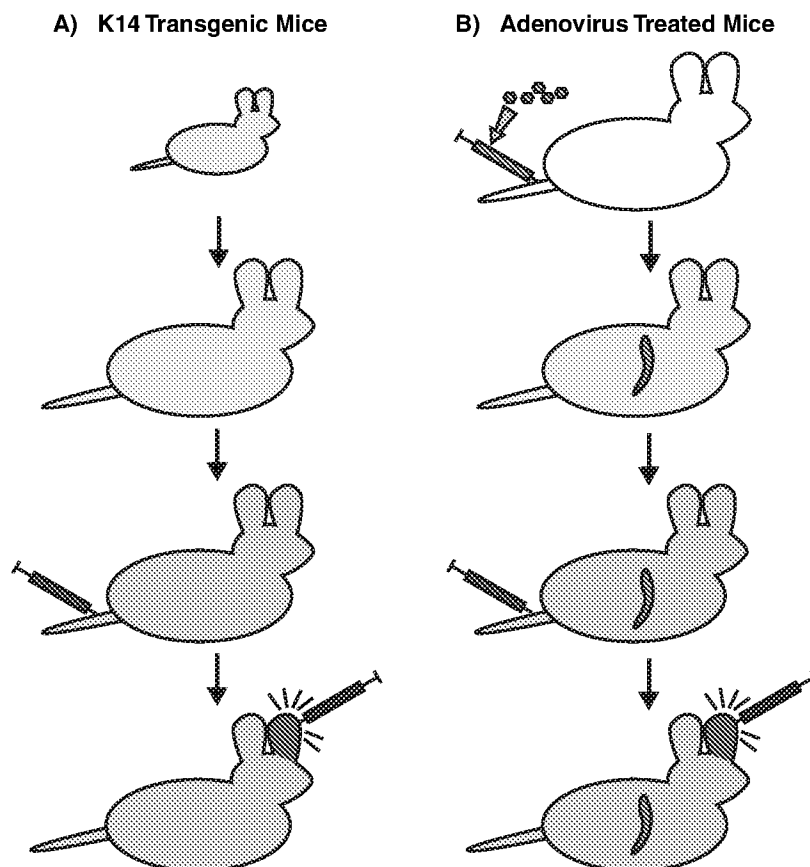


Fig. 1 Outline of Evans blue leakage experiments in (A) K14 transgenic mice and (B) adenovirus treated mice. (A) In transgenic mice, secreted factor (VEGF or Ang1 – green) is overexpressed in embryonic skin and throughout the lifetime of the mouse. Evans blue dye (blue) was injected into adult mice, and then inflammatory stimuli (red) were applied topically to ear skin on one side (other ear served as control). (B) In adenovirus experiments, adenovirus encoding factor (VEGF or Ang1 – green hexagons) was injected intravenously into normal adult mice (white). Factor was overexpressed in liver and secreted into circulation. Inflammatory stimuli were applied topically to the distal site: the ear skin on one side.

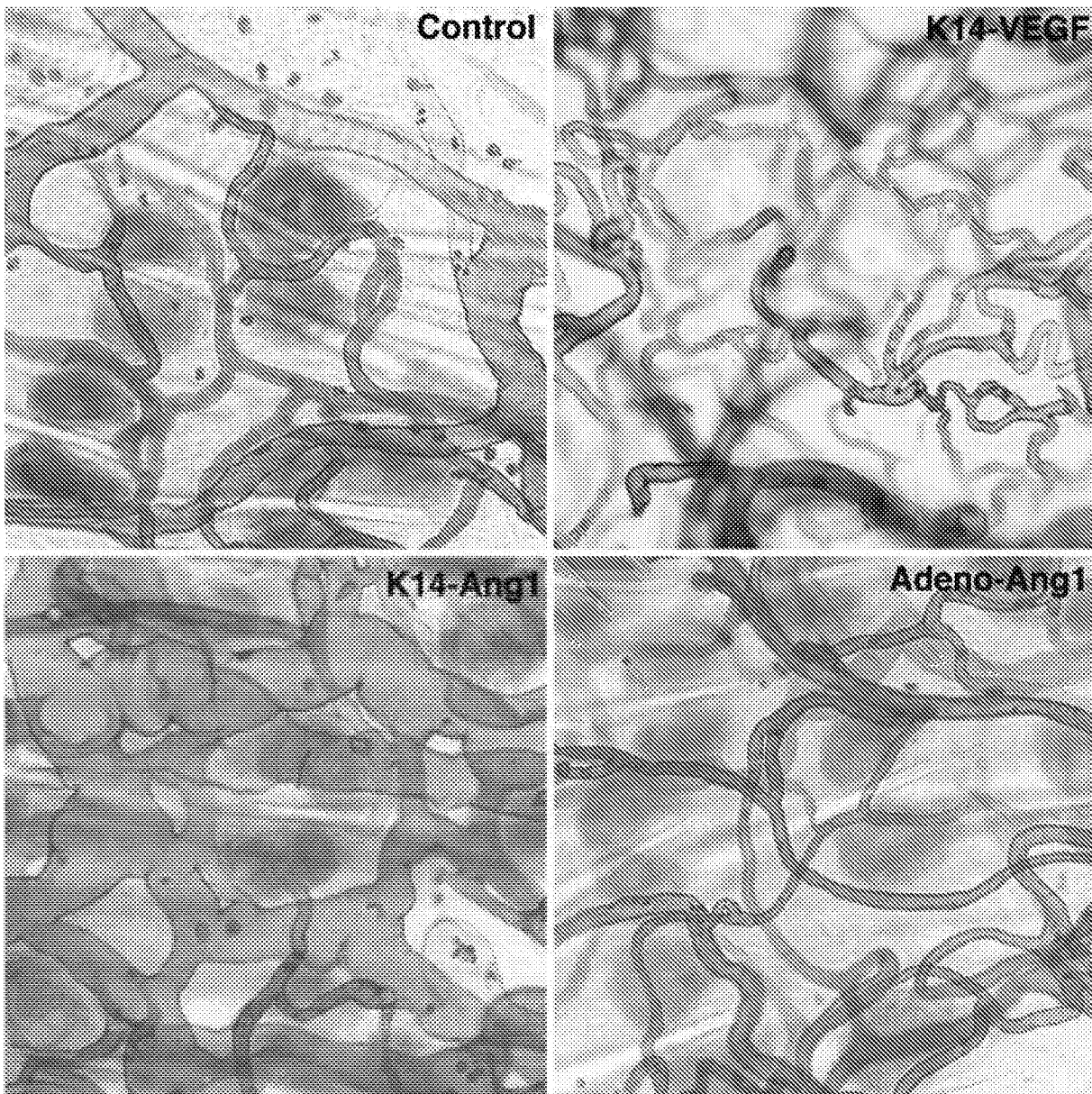


Fig. 2 Morphology of blood vessels in ear skin of control, K14-VEGF, K14-Ang1, and Adeno-Ang1 mice. Vessels were stained by intravascular perfusion of biotinylated *Lycopersicon esculentum* lectin and DAB-peroxidase reaction, and examined in whole mounts (Thurston et al. 1999, 2000). Lectin binds to the luminal surface of endothelium and reveals blood vessels.

1999). Upon examination of tissue sections stained for endothelial cells with antibodies to platelet endothelial cell adhesion molecule (PECAM, CD31) or whole mounts stained with intravascular lectin, the skin of K14-VEGF mice showed increased blood vessel density characterized by increased numbers of small capillary-sized vessels near the epidermis and surrounding the hair follicles (Fig. 2). The tortuous skin vessels of K14-VEGF mice showed leakage of the plasma tracer Evans blue under baseline conditions (Thurston et al. 1999). In addition, the basement membrane of these

vessels could be labelled by intravascular perfusion of Ricin lectin, indicating defects in the endothelial barrier. Application of inflammatory stimuli to the ear skin resulted in even larger amounts of plasma leakage. The VEGF transgenic mice highlight the role of VEGF as a potent angiogenic factor, but also emphasize that the resultant vessels can be leaky and inflamed.

Mice overexpressing Ang1 under the K14 promoter were also produced (Suri et al. 1998). The skin of K14-Ang1 mice was notably reddened, but the epidermis was normal in thickness and the dermis did not contain

infiltrating cells. The dermal vessels were dramatically increased in diameter compared to control mice (Fig. 2), but only moderately increased in number. The enlarged vessels were in the position of capillaries subjacent to the epidermis and surrounding the hair follicles. The enlarged vessels had an increased number of endothelial cells, indicating that Ang1 increased endothelial cell proliferation or survival (Thurston et al. 1999). Unlike K14-VEGF mice, the vessels in K14-Ang1 mice were not leaky under baseline conditions and, remarkably, seemed to be resistant to plasma leakage induced by inflammatory mediators such as histamine, serotonin and mustard oil.

K14-Ang1 mice were bred to K14-VEGF mice (Thurston et al. 1999). The skin of the resultant double transgenic K14-Ang1/VEGF mice was dramatically reddened, and the vascularity of the skin was higher than either K14-VEGF or K14-Ang1 mice. The morphology of the dermal vessels appeared to be a combination of the Ang1 and VEGF effects. In particular, numerous small vessels and enlarged vessels were both present (Thurston et al. 1999). The dermis of K14-Ang1/VEGF mice was normal in thickness and did not contain infiltrating leucocytes. Furthermore, the skin vessels in K14-Ang1/VEGF mice were not leaky to Evans blue or Ricin lectin under baseline conditions (Thurston et al. 1999). Thus, Ang1 seems to inhibit some of the inflammatory actions of VEGF, but Ang1 and VEGF appear to act on distinct signalling pathways for vessel growth.

Adenoviral overexpression – Adeno-VEGF and Adeno-Ang1

Adenoviruses encoding VEGF₁₆₄, Ang1, or green fluorescent protein (GFP) as a control, driven by the cytomegalous viral (CMV) promoter were injected intravenously into mice (age 8–10 weeks) (Thurston et al. 2000). GFP was localized in the liver, confirming that most (> 95%) of the adenoviral gene expression is in hepatocytes (Yao et al. 1996; Michou et al. 1997). High levels of VEGF or Ang1 (10 µg mL⁻¹) were detected in the serum within 1 day after injection of adenovirus and, depending upon the strain of mouse, the levels remained above 500 ng mL⁻¹ for 10 days or longer.

Following injection of adeno-VEGF (1 × 10⁸ pfu or more), mice became lethargic and died within 2–3 days. Histological examination of various organs in the mice given adeno-VEGF revealed evidence of widespread oedema (Thurston et al. 2000). In contrast,

mice injected with adeno-Ang1 (1 × 10⁹ pfu) appeared healthy and remained active. Similar to the K14-Ang1 mice, the blood vessels in the skin of mice given adeno-Ang1 became resistant to the plasma leakage normally induced by local injection of VEGF or topical application of mustard oil (Thurston et al. 2000). The resistance to leakage was found at 1 day after intravenous injection of adenovirus. However, unlike the K14-Ang1 mice, adult mice given adeno-Ang1 did not appear reddened, and the morphology of the skin blood vessels was normal for at least 7 days after adenoviral injection (Fig. 2) (Thurston et al. 2000). Subsequent experiments have shown that the antileakage action can be duplicated by intraperitoneal injection of Ang1 proteins (E. Joffe et al. unpublished results); thus this effect is not due to the adenoviral production of Ang1.

Discussion

Our experiments, using two approaches to overexpress VEGF and Ang1, demonstrate that these factors act separately and distinctly on blood vessels. In both overexpression systems, Ang1 resulted in vessel enlargement and resistance to leak, whereas VEGF resulted in leakiness and, if expressed locally, in vessel sprouting. The actions on blood vessel growth appear to be able to take place independently because, when given together, VEGF caused vessel sprouting and Ang1 caused vessel enlargement. However, at least in the situation where the two factors are overexpressed transgenically, the antileakage action of Ang1 appears to predominate.

In contrast to the transgenic mice in which Ang1 was overexpressed throughout development, Ang1 given to adults did not cause vessel enlargement in the skin, even when given for 50 days. Why did adenoviral expression of Ang1 not cause vessel enlargement? One possibility is that the vasculature in adults is less plastic than in embryonic and neonatal mice, and thus does not enlarge in response to Ang1. Another possibility is that Ang1 acts differently depending on whether it is delivered via the circulation or locally in the interstitium. Studies in which neonatal mice are treated with Ang1, and those in which Ang1 is inducibly overexpressed in adult mice using inducible transgenic approaches, may help shed light on this question.

Our findings using overexpression systems support and extend the studies of gene-targeted mice. Both sets of studies show clearly that VEGF and Ang1 play

distinct and complementary roles in blood vessel development. Our overexpression studies help to pinpoint the distinctions, in particular highlighting the vessel enlargement and maturation actions of Ang1, and the vessel sprouting and leakiness actions of VEGF. However, it is not known whether the actions of these factors that were highlighted in our experimental overexpression systems are indeed analogous to their roles in normal development. For example, does VEGF induce leakage during normal blood vessel development? Does Ang1 have an antileakage action in development? Does coexpression of Ang1 normally prevent VEGF-induced leakage? Does Ang1 help vessels to enlarge in development? It must be borne in mind that the readouts from the assays used in experimental studies may not have a direct correlate in normal blood vessel growth. Nevertheless, our studies help define the range of actions that can be induced by endothelial-specific factors.

One clinical application of Ang1 may be to use it in combination with VEGF to grow normal, non-leaky vessels. Therapeutic application of VEGF has been tested in a number of situations to induce the growth of new blood vessels (therapeutic angiogenesis – Isner & Losordo, 1999). However, a potential side-effect of large amounts of VEGF may be the growth of leaky vessels. Indeed, some reports have suggested this may be the case (Baumgartner et al. 1998; Springer et al. 1998; Lee et al. 2000). Co-administration of Ang1 may help reduce the leak-inducing actions of VEGF without suppressing the vessel growth actions. Further studies comparing the actions of VEGF and Ang1 will undoubtedly help define the discrete steps of normal blood vessel development.

Acknowledgments

I would like to thank Donald M. McDonald (UCSF), John Rudge, Ella Ioffe and George Yancopoulos (Regeneron Pharmaceuticals) for helpful discussions.

References

- Baumgartner I, Pieczek A, Manor O, Blair R, Kearney M, Walsh K, et al. (1998) Constitutive expression of phVEGF165 after intramuscular gene transfer promotes collateral vessel development in patients with critical limb ischemia. *Circulation* **97**, 1114–1123.
- Carmeliet P, Ferreira V, Breier G, Pollefeyt S, Kieckens L, Gertsenstein M, et al. (1996) Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. *Nature* **380**, 435–439.
- Davis S, Aldrich TH, Jones PF, Acheson A, Compton DL, Jain V, et al. (1996) Isolation of angiopoietin-1, a ligand for the TIE2 receptor, by secretion-trap expression cloning. *Cell* **87**, 1161–1169.
- Detmar M, Brown LF, Schon MP, Elicker BM, Velasco P, Richard L, et al. (1998) Increased microvascular density and enhanced leukocyte rolling and adhesion in the skin of VEGF transgenic mice. *J. Invest. Dermatol.* **111**, 1–6.
- Dumont DJ, Gradwohl G, Fong GH, Puri MC, Gertsenstein M, Auerbach A, et al. (1994) Dominant-negative and targeted null mutations in the endothelial receptor tyrosine kinase, tek, reveal a critical role in vasculogenesis of the embryo. *Genes Dev.* **8**, 1897–1909.
- Dvorak HF, Nagy JA, Berse B, Brown LF, Yeo KT, Yeo TK, et al. (1992) Vascular permeability factor, fibrin, and the pathogenesis of tumor stroma formation. *Ann. NY Acad. Sci.* **667**, 101–111.
- Ferrara N, Carver-Moore K, Chen H, Dowd M, Lu L, O'Shea KS, et al. (1996) Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene. *Nature* **380**, 439–442.
- Ferrara N, Leung DW, Cachianes G, Winer J, Henzel WJ (1991) Purification and cloning of vascular endothelial growth factor secreted by pituitary folliculostellate cells. *Meth. Enzymol.* **198**, 391–405.
- Fong GH, Rossant J, Gertsenstein M, Breitman ML (1995) Role of the Flt-1 receptor tyrosine kinase in regulating the assembly of vascular endothelium. *Nature* **376**, 66–70.
- Hanahan D (1997) Signaling vascular morphogenesis and maintenance. *Science* **277**, 48–50.
- Isner JM, Losordo DW (1999) Therapeutic angiogenesis for heart failure. *Nat. Med.* **5**, 491–492.
- Larcher F, Murillas R, Bolontrade M, Conti CJ, Jorcano JL (1998) VEGF/VPF overexpression in skin of transgenic mice induces angiogenesis, vascular hyperpermeability and accelerated tumor development. *Oncogene* **17**, 303–311.
- Lee RJ, Springer ML, Blanco-Bose WE, Shaw R, Ursell PC, Blau HM (2000) VEGF gene delivery to myocardium: deleterious effects of unregulated expression. *Circulation* **102**, 898–901.
- Maisonpierre PC, Suri C, Jones PF, Bartunkova S, Wiegand SJ, Radziejewski C, et al. (1997) Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis. *Science* **277**, 55–60.
- Michou AI, Santoro L, Christ M, Julliard V, Pavirani A, Mehtali M (1997) Adenovirus-mediated gene transfer: influence of transgene, mouse strain and type of immune response on persistence of transgene expression. *Gene Ther.* **4**, 473–482.
- Sato TN, Tozawa Y, Deutsch U, Wolburg-Buchholz K, Fujiwara Y, Gendron-Maguire M, et al. (1995) Distinct roles of the receptor tyrosine kinases Tie-1 and Tie-2 in blood vessel formation. *Nature* **376**, 70–74.
- Shalaby F, Rossant J, Yamaguchi TP, Gertsenstein M, Wu XF, Breitman ML, et al. (1995) Failure of blood-island formation and vasculogenesis in Flk-1-deficient mice. *Nature* **376**, 62–66.
- Springer ML, Chen AS, Kraft PE, Bednarski M, Blau HM (1998)

- VEGF gene delivery to muscle: potential role for vasculogenesis in adults. *Mol Cell* **2**, 549–558.
- Suri C, Jones PF, Patan S, Bartunkova S, Maisonpierre PC, Davis S, et al.** (1996) Requisite role of angiopoietin-1, a ligand for the TIE2 receptor, during embryonic angiogenesis. *Cell* **87**, 1171–1180.
- Suri C, McClain J, Thurston G, McDonald DM, Zhou H, Oldmixon EH, et al.** (1998) Increased vascularization in mice overexpressing angiopoietin-1. *Science* **282**, 468–471.
- Thurston G, Suri C, Smith K, McClain J, Sato TN, Yancopoulos GD, et al.** (1999) Leakage-resistant blood vessels in mice transgenically overexpressing angiopoietin-1. *Science* **286**, 2511–2514.
- Thurston G, Rudge JS, Ioffe E, Zhou H, Ross L, Croll SD, et al.** (2000) Angiopoietin-1 protects the adult vasculature against plasma leakage. *Nat. Med.* **6**, 460–463.
- Valenzuela DM, Griffiths JA, Rojas J, Aldrich TH, Jones PF, Zhou H, et al.** (1999) Angiopoietins 3 and 4: diverging gene counterparts in mice and humans. *Proc. Natl Acad. Sci. USA* **96**, 1904–1909.
- Yancopoulos GD, Davis S, Gale NW, Rudge JS, Wiegand SJ, Holash J** (2000) Vascular-specific growth factors and blood vessel formation. *Nature* **407**, 242–248.
- Yao SN, Farjo A, Roessler BJ, Davidson BL, Kurachi K** (1996) Adenovirus-mediated transfer of human factor IX gene in immunodeficient and normal mice: evidence for prolonged stability and activity of the transgene in liver. *Viral Immunol.* **9**, 141–153.



Christopher Kent, Senior Editor

PUBLISHED 5 AUGUST 2019

Anti-VEGF 2019: The State of the Art

Surgeons share the latest thinking about the benefits and risks of anti-VEGF therapy, and what lies ahead.

There's no question that anti-VEGF drugs have caused a sea-change in the treatment of several retinal diseases. Conditions that were basically untreatable have now become manageable, preventing what was once almost inevitable blindness. Yet there's still plenty of room for improvement; results vary, and not all patients avoid vision loss.

"When I started my career, neovascular age-related macular degeneration was a horrible disease," recalls K. Bailey Freund, MD, a retina specialist and a clinical professor of ophthalmology at the New York University School of Medicine. (Dr. Freund has been a principal investigator in several pivotal trials of novel treatments for retinal diseases.) "Nothing we could do would really help our patients. Today, certain patients do extremely well—particularly the ones that have Type 1 (subretinal pigment epithelium) neovascularization under the fovea. Others do well initially, but eventually the non-neovascular aspect of the disease kicks in and they ultimately end up losing vision, more from macular atrophy than exudative complications."

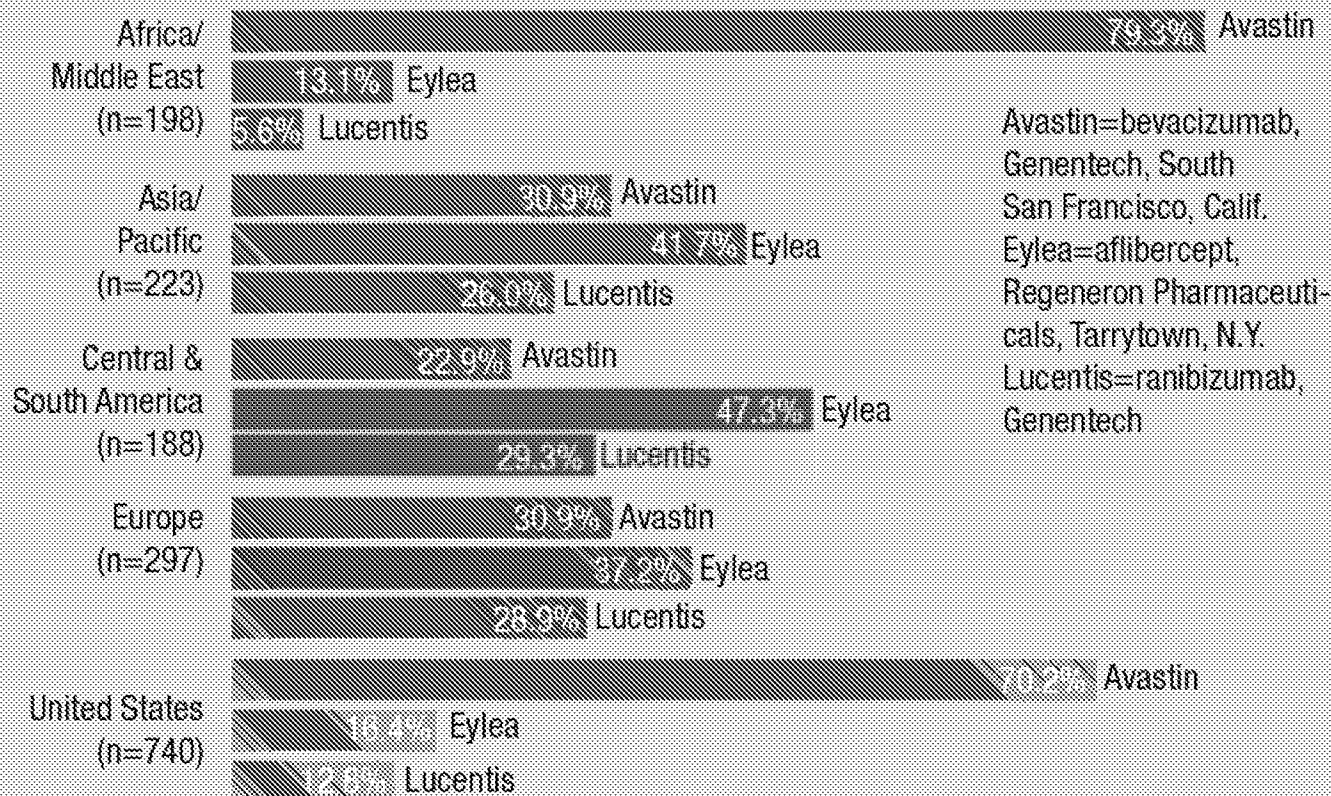
Here, retina specialists answer key questions about the current state of anti-VEGF drug treatments for macular degeneration and other retinal diseases; share the latest insights, which suggest that neovascularization and subretinal fluid are not always a bad thing; and discuss what the upcoming years may hold.

Anti-VEGF Drug Choice

"Each anti-VEGF option has an advantage that certain people prefer," notes Philip Rosenfeld, MD, PhD, professor of ophthalmology at the Bascom Palmer Eye Institute at the University of Miami Miller School of Medicine. "For example, bevacizumab is cheap. In certain countries surgeons can get a rebate for using ranibizumab, so some doctors use it because it's financially rewarding for the practice. Aflibercept is the preferred drug for certain neovascular types of disease; for example, I think it works best in fibrovascular retinal pigment epithelial detachments with a significant serous component.

"Personally, I treat with bevacizumab and aflibercept," he says. "In my non-hospital-based setting, I always start with bevacizumab and try to get a durability that exceeds six, eight or 10 weeks. If I can't get beyond that six-to-eight week interval, then I switch to aflibercept. Some surgeons prefer ranibizumab, and others prefer to use aflibercept exclusively. However, many times when patients are in Medicare Advantage plans, the choice of drug is decided by the provider."

What is your first-line anti-VEGF agent for wet AMD?



Data from the Global Trends in Retina Survey, conducted by the American Society of Retina Specialists, in conjunction with the 20th Annual ASRS Preferences and Trends (PAT) Survey.³ Members of 42 retina societies around the world participated in the 2018 Global Trends in Retina Survey. (Reprinted with permission.)

Dr. Rosenfeld notes that The American Society of Retina Specialists runs an annual Patterns and Trends survey, in which they ask about anti-VEGF drug choices every year. “We don’t have the results for 2019 as we speak,” he says, “but the replies for 2018 indicated that in the U.S., bevacizumab is the first-line choice for treating wet AMD for about 70 percent of retina specialists. Ranibizumab is first-line for 12 percent, and aflibercept is first-line for 16 percent. Outside the U.S.—except in Africa and the Middle East—it looks like aflibercept wins. However, treating physicians are litigating for access to bevacizumab in the U.K. In France, it’s still very hard to get.” (He notes that at a recent retina meeting in Italy, most treating physicians said that they’ve used all three drug options—ranibizumab, bevacizumab and aflibercept.)

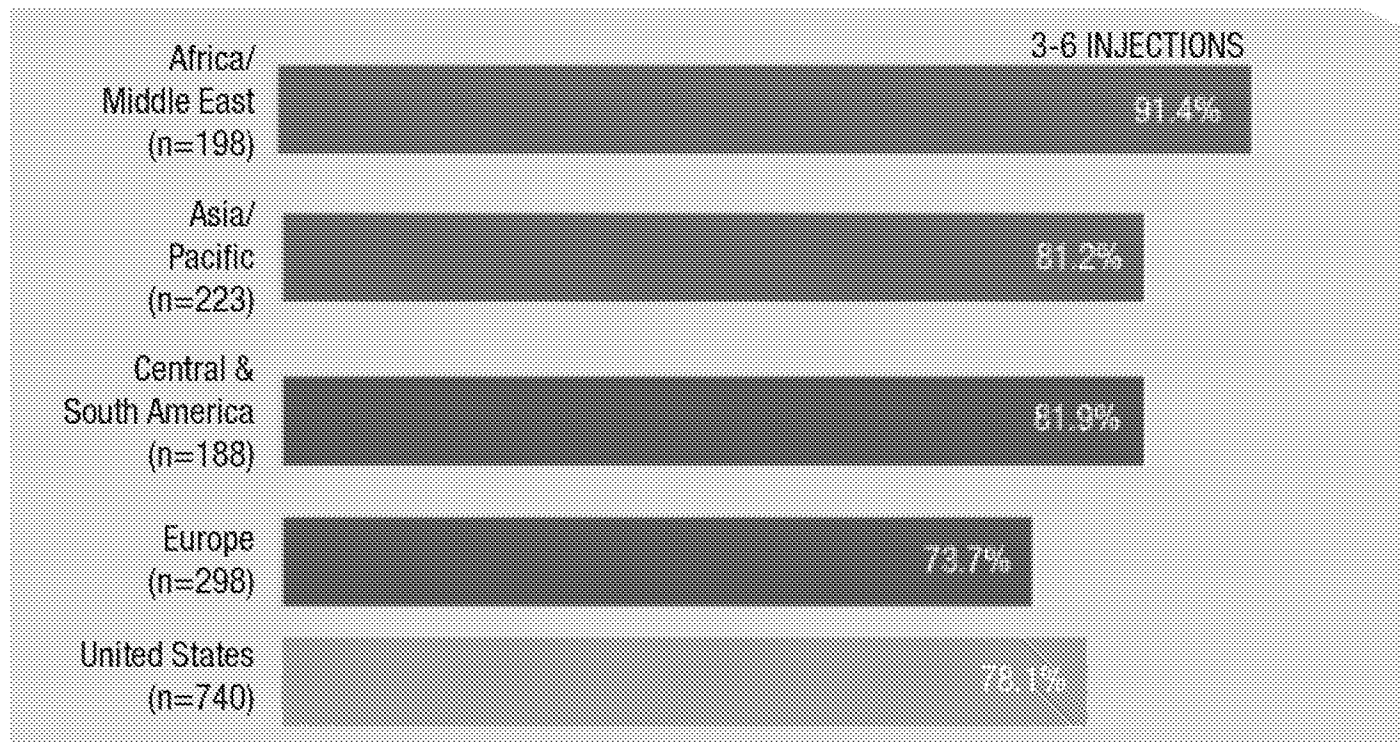
Dr. Freund says he finds the current anti-VEGF agents to be fairly similar in terms of efficacy and safety, so he chooses his drug regimen based on the individual patient’s presentation. “I might choose one drug over another when a patient is somewhat refractory to treatment,” he says. “This is particularly true for eyes that have what I currently refer to as aneurysmal Type 1 neovascularization, more commonly known as polypoidal choroidal vasculopathy. I see many patients with this lesion growth pattern because it was first described in our practice, and I’ve published on it extensively.

“Eyes with this lesion growth pattern may have a more robust response to aflibercept,” he continues. “Clinical trial evidence from the PLANET study showed that many of these eyes respond well to aflibercept monotherapy, while data from the EVEREST II trial indicates that eyes with aneurysmal Type 1 neovascularization receiving

ranibizumab may require the addition of photodynamic therapy in order to optimize visual outcomes.

“There’s another form of neovascular AMD that I call pachychoroid neovascuopathy,” he notes. “These eyes have choroidal findings that are similar to those seen in eyes with central serous chorioretinopathy. Also, they lack some of the characteristic clinical findings we associate with typical macular degeneration in elderly Caucasian patients, such as soft drusen. Those eyes may benefit from aflibercept, since aflibercept has been shown to decrease choroidal thickness more than the other agents. I find that eyes which are somewhat refractory to anti-VEGF therapy may not respond as well to bevacizumab, compared to the other agents.

Physicians who consider switching anti-VEGF agents due to inadequate response after three to six injections



More data from the 2018 Global Trends in Retina Survey, conducted by the American Society of Retina Specialists.³ (For more information, see the caption for the chart on page 27.)

“In the final analysis, a typical patient responds well to all of these agents,” he says. “Numerous studies show that bevacizumab is noninferior to the alternative FDA-approved options. However, for patients managed on a treat-and-extend regimen, I’m less comfortable extending my dosing interval beyond eight weeks with bevacizumab. So, for patients with no out-of-pocket drug expense, we may choose a different agent that could require fewer injections, particularly when frequent office visits would be very difficult.”

Of course, some patients don’t respond to the first drug the treating physician tries, but Dr. Freund points out that switching refractory patients to an alternative anti-VEGF is not a cure-all. “When aflibercept was approved, many practices did what we then called ‘switcher’ studies,” notes Dr. Freund. “We took patients who were poorly controlled with monthly bevacizumab or ranibizumab and switched them to aflibercept, expecting that because most eyes in the aflibercept trials could be maintained on injections every eight weeks, the same might happen with these refractory cases. But the eyes we were switching were very different from the type of newly-diagnosed cases

enrolled in clinical trials. All we typically found was that aflibercept might get rid of more fluid for a little longer than the prior agents. We did not find that eyes showing persistent fluid with monthly ranibizumab were fluid-free for eight weeks with aflibercept.”

Regarding the treatment protocol, Dr. Rosenfeld says he believes that most clinicians treat their patients using a treat-and-extend or modified treat-and-extend regimen. “This usually means performing monthly dosing until the macular fluid is resolved, and then extending the interval until macular fluid recurs,” he explains. “At that point the interval is shortened, and eventually a treatment interval is defined for that particular patient. When I attended the FLORetina meeting in Florence, Italy recently, this protocol seemed to be the general consensus among the attendees.”

Is Neovascularization Bad?

Clearly, neovascularization can be part of the problem in retinal diseases such as macular degeneration. However, many researchers note that anti-VEGF drugs aren't really treating the neovascularization—and they're beginning to suspect that neovascularization may actually be a good thing in some patients.

“There are three types of macular neovascularization,” Dr. Rosenfeld explains. “The major type is what's called Type 1, which occurs under the retinal pigment epithelium. Today, with OCTA, particularly swept-source OCTA, we can identify Type 1 neovascularization long before exudation occurs. For that reason there's been some debate about whether we should treat Type 1 lesions with anti-VEGF therapy before exudation starts.

“I think the consensus is that we should not,” he says. “Instead, we should watch the lesions and treat when symptomatic exudation develops. The reason is that those lesions don't go away because of treatment. What we're doing is training them not to leak. We calm the lesion down, but we don't make it go away.

Do Anti-VEGF Drugs Cause Atrophy?

Philip Rosenfeld, MD, PhD, professor of ophthalmology at the Bascom Palmer Eye Institute at the University of Miami Miller School of Medicine, believes the concern about anti-VEGF therapy possibly promoting the formation of macular atrophy is still present, but receding. “There's still some debate about whether the drugs themselves are influencing the formation of macular atrophy,” he notes, “or whether it's just normal disease progression, or whether it's a characteristic of certain lesion types, such as Type 3 macular neovascularization, also known as retinal angiomatous proliferation, which tends to form atrophy.”

K. Bailey Freund, MD, a retina specialist and a clinical professor of ophthalmology at the New York University School of Medicine, admits that a connection between anti-VEGF treatment and atrophy is biologically plausible. “Continuous VEGF suppression could do that, but we're not actually providing continuous suppression,” he points out. “We give a large bolus of the drug, and over time the concentration drops to a very low level. Then we give another bolus. This may be one reason that the studies suggesting a possible link between anti-VEGF therapy and atrophy have really only shown this association when patients were treated continuously every month. Even these studies haven't really proven that the

anti-VEGF treatment is causing the atrophy.”

Dr. Freund advises clinicians to use their judgment. “Look at the data and use common sense,” he says. “It might be reasonable to reduce injection frequency for eyes with Type 3 lesions, but not in eyes with large vascularized pigment epithelial detachments, particularly those with aneurysms—more commonly known as polyps—as these eyes appear resistant to macular atrophy, but are at risk for catastrophic hemorrhages if undertreated. So, look closely at the type of eye you're treating and adjust your protocol accordingly.

“The data indicate that more patients in the United States and around the world lose vision to undertreatment than to overtreatment,” he adds. “So, while I think retina specialists should be concerned about causing atrophy, they should also be concerned about the disease itself, and make sure it's kept under control.”

Dr. Rosenfeld agrees. “When patients come to us with exudative disease, not treating them isn't an option,” he says. “So they get the injections, and we try to balance the need for reinjection and eliminating macular fluid against our concern about the possibility that overtreatment may exacerbate atrophy.”

—CK

“In fact, we’re starting to believe that when the Type 1 lesions go away, you’re left with macular atrophy,” he says. “Many of my colleagues believe that you need to manage the neovascularization, but you don’t want to overtreat it. You want the neovascularization to stay there and just stop leaking. To put it another way, the field has evolved to believe that neovascularization is not the enemy; exudation and hemorrhage are the enemy. The neovascularization may actually be serving a very important function—providing nutritional support for the retinal pigment epithelium. Its appearance may be a normal physiologic response to a pathological condition in which you’re losing the choriocapillaris and there’s low-grade inflammation in the back of the eye. You need this neovascular complex. You just don’t want the exudation.”

“We know that patients that look clinically dry can harbor ‘non-exudative’ Type 1 neovascular tissue, which proliferates beneath the RPE with little clinical or optical coherence tomography evidence of its presence,” says Dr. Freund. “There’s no fluid and no hemorrhage. Today, however, we have tools like OCTA that let us detect these vessels. The most notable fact is that some of these patients never develop pathologic exudation. The challenge is that we don’t know exactly what percentage of those patients will eventually develop vision-threatening fluid and hemorrhage.”

Dr. Freund frequently collaborates with Christine A. Curcio, PhD, a professor of ophthalmology at the University of Alabama at Birmingham, who is an expert in the histology of macular degeneration. “We’ve been performing clinicopathologic correlations for AMD for close to a decade,” he says. “If patients from my practice consent, we enroll them in our donor program, and the eye bank recovers the whole globe following their death. Our mission is to correlate the histology with the retinal imaging done in the clinic.

“We’re currently studying the eyes of a patient followed for 10 years,” he says. “When she first presented, she was in her late 70s, and she’d already lost central vision in one eye from scarring due to neovascular AMD. Surprisingly, even though fluorescein angiography and OCT documented Type 1 neovascularization in her other eye, this eye never developed sight-threatening exudation, and retained excellent visual acuity for more than a decade. This unusual disease course was documented with multimodal retinal imaging, which we’re now correlating with high-resolution histology. Unexpectedly, it turns out that more than half of her macula was sitting on top of neovascular tissue, and the overlying photoreceptors maintained a healthy structure. The neovascular tissue appears to have maintained nutritional support to the fovea as the patient’s native choriocapillaris failed due to aging. So, by the time she died, the health of her outer retina appears to have been maintained by the very same tissue that we all fear is going to cause patients to lose vision—tissue that we often do everything in our power to suppress with our anti-VEGF agents.”

Dr. Freund notes that the idea that Type 1 neovascularization could be beneficial isn’t new. “This idea was first proposed by Hans E. Grossniklaus, MD and Richard Green, MD,” says Dr. Freund. “They hypothesized that Type 1 neovascularization had the potential to recapitulate the normal choriocapillaris, and this concept appears to be supported by the findings in our current donor. In some patients, these non-exudative vessels may actually be helping to preserve the overlying retina. So, the goal may not be to destroy or completely inhibit their growth but to carefully monitor eyes with these vessels so that pathologic exudation can be caught and treated early.

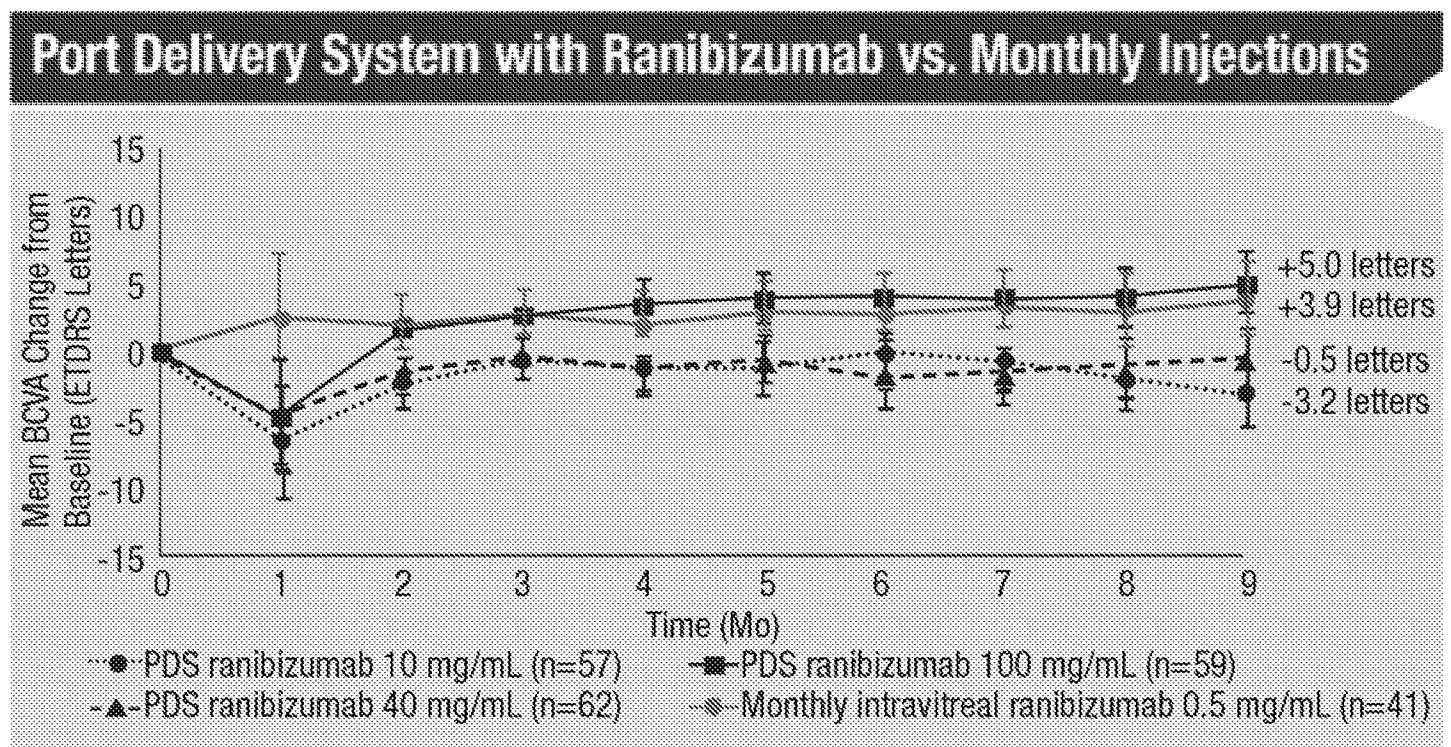
“That means we need to have the right balance,” he concludes. “We shouldn’t be so aggressive that we destroy a potentially protective mechanism that could benefit our patients in future, but we also shouldn’t let it get out of control and damage vision quickly with catastrophic bleeding.”

Is Fluid Always a Bad Thing?

“Studies have shown that eyes treated with intravitreal anti-VEGF therapy which continue to manifest subretinal fluid—eyes that don’t go completely dry—may do as well, or even better, than eyes which dry up completely,” notes Dr. Freund. “So, a little bit of fluid may not be such a bad thing.”

Dr. Freund refers to the three main types of neovascularization occurring in exudative AMD, and says evidence is mounting that they may warrant different treatment regimens. "I believe retinal specialists should look at the eye's presenting imaging characteristics, which define a lesion's subtype," he says. "Different subtypes seem to respond differently to treatment and call for different levels of monitoring. For example, it's my strong belief that eyes presenting with Type 1 neovascularization beneath the fovea that continue to show a small amount of subretinal fluid, despite frequent dosing, are more resistant to geographic atrophy than eyes with other neovascular lesion patterns. In these eyes, a little bit of fluid is probably not such a bad thing. The main threat to vision in eyes like this appears to be the occurrence of subretinal hemorrhage if treatment is discontinued.

"In contrast, atrophy is a greater concern in eyes that have intraretinal neovascularization known as a Type 3 neovascularization or RAP (retinal angiomatous proliferation) pattern," he continues. "Greater susceptibility to macular atrophy in eyes with Type 3 lesions has been observed in several large clinical trials. Patients presenting with Type 3 neovascularization are often older individuals with thin choroids and focal areas of macular hyperpigmentation, both of which increase susceptibility to macular atrophy. For eyes presenting with Type 3 neovascularization, I'll often first try PRN dosing, for three reasons: lesion quiescence may be long-lasting; recurrences predictably occur at the initial site of activity; and recurrent exudation with Type 3 lesions is rarely accompanied by large hemorrhages."



Data from the Phase II LADDER clinical trial. All patients were previously treated with and were responsive to anti-VEGF therapy. (Vertical bars=95% confidence intervals.)⁴

Dr. Freund notes that Type 1 and Type 3 are the two most common variants of neovascular AMD. "Type 2 and mixed lesions with a component of Type 2 are less common, but the vessels in these eyes have broken through the RPE to proliferate in the subretinal space," he says. "You have to be careful with these cases, since recurrent exudation from Type 2 lesions is in direct contact with the vulnerable photoreceptors. In that situation irreversible vision loss can occur quickly."

Dr. Freund points out that the implication is that different types of lesions should be treated differently. "Don't simply assume that all neovascular AMD is the same and should be treated with the same regimen," he says. "Over the short term, these slight variations in regimen may not seem to make a difference, but we often treat patients for the

rest of their lives. Little differences in how a patient does on one regimen versus another end up making a big difference over five, 10 or 15 years of treatment.”

Dr. Freund says that OCT may provide clues to the best treatment protocol. “If you see a shallow, irregular PED with heterogeneous internal contents in a patient with other findings of AMD, there’s a high probability that this finding represents neovascular tissue,” he says. “If there’s no exudation, eyes with this finding should be monitored closely, and the patient should be instructed to use an Amsler grid to monitor for conversion to active wet disease.

“When this type of vascularized pigment epithelial detachment, or Type 1 pattern, is subfoveal and associated with exudation, I use a treat-and-extend regimen of anti-VEGF therapy to control exudation,” he continues. “However, my goal is not to flatten the PED itself. I believe that intense treatment has the potential to convert a vascularized PED into avascular fibrotic tissue—tissue that will no longer be able to support the overlying retina. If there’s no hemorrhage, I’m not concerned about a little bit of persistent subretinal fluid, as long as it’s not increasing over time.”

Dr. Freund says that many of his patients have demonstrated the wisdom of this approach. “I have many patients I’ve been treating with anti-VEGF therapy for 10 or more years, some of whom have had as many as 100 injections,” he says. “Many eyes still have close to 20/20 vision.”

How Important is OCTA?

“Right now we treat based on exudation—the fluid that’s visible on structural OCT,” notes Dr. Rosenfeld. “However, people are working hard to see if the information we pick up with OCTA may be able to influence how we treat. Many papers have been published about using OCTA to look at different types of neovascular structures that exist in these eyes with wet AMD, hoping that some of what they find would be predictive of how often we need to treat, and/or the outcome of treatment. The results have been underwhelming, which is probably because the anti-VEGF drugs treat the exudation, not the neovascularization. Thus, changes in the vasculature revealed by OCTA may be less important than the presence of exudation on structural OCT.

“However, OCTA does have a role,” he says. “It’s a very powerful way to identify eyes with dry macular degeneration that are at higher risk of exudation, so we can follow them differently. With OCTA—in particular SS-OCTA—you can see the neovascularization many months before patients actually develop exudation. It’s growing silently. This has been known since the 1970s, when autopsy eyes with dry macular degeneration were shown to harbor neovascularization; it just wasn’t leaking or bleeding. Using ICG angiography in the 1990s, retina specialists also showed that this neovascularization existed in their living patients. However, we couldn’t routinely screen patients with ICG angiography, so this discovery was largely forgotten until OCTA came along.

“Knowing that this neovascularization is present, via OCTA, is valuable information,” he says. “We recently conducted a two-year study involving 227 patients, which is currently in press. We compared eyes with and without this nonexudative neovascularization and found that there’s a 14-fold increased risk of exudation over a period of two years when neovascularization is present. So OCTA serves a purpose—just not for monitoring lesions once you start treating. Instead, it can help to identify these lesions before exudation develops.

“For this reason, I use OCTA to check all of my dry AMD patients to see if they have these lesions,” he says. “It changes how I manage my patients. If they have the lesions, I usually follow them every two months. If they don’t have the lesions, I usually follow them every six months. It also helps me educate patients so they understand what’s going on and become partners in their own management.”

Dr. Freund says he uses OCTA frequently, but doesn’t believe it’s essential for diagnosing or treating AMD. “Often, OCT alone is sufficient to diagnose neovascular AMD or to arouse high suspicion that there’s a problem,” he notes. “In some cases, when there’s a characteristic OCT pattern of specific neovascular features, dye angiography may

not be needed, especially if there's a hemorrhage. I think OCTA is helpful if, like myself, you're interested in identifying the neovascular lesion type to help individualize your treatment algorithm.

"Does individualizing the treatment regimen necessarily lead to better outcomes?" he adds. "I think so, but this remains to be proven. For now, I'd say that OCTA is a valuable research tool. In the future, it may become essential for a clinician managing macular degeneration."

Dr. Rosenfeld believes clinicians will benefit from using OCTA. "I think everyone who practices retinal care needs access to OCTA, whether it's for diabetes or AMD or vein occlusions," he says. "It's a remarkable tool that allows us to see the full extent of the underlying disease, and in certain situations helps us manage it better. OCTA is now our number one tool for diagnosing proliferative diabetic retinopathy. It helps us determine the extent of vascular nonperfusion in diabetics and vein occlusion, and it's particularly useful for identifying the subclinical, nonexudative lesions in dry AMD. Furthermore, patients love it compared with dye-based angiograms. Meanwhile, of course, structural OCT is very useful for managing all of these conditions as well. But with OCTA scans, you get both the structural OCT and the angiographic OCT."

Dr. Rosenfeld points out one big problem with OCTA, however, at least in the United States. "We don't have a unique billing code for OCTA," he explains. "The scan time is about the same as routine OCT, but the time it takes for the physician to interact with the instrument, look at the scans and extract the information by manipulating the segmentation boundaries to get the best possible image isn't currently reimbursed. So, while the equipment is more expensive and it takes more time to evaluate the scans, the reimbursement is the same as for the more typical OCT. Outside the U.S., OCTA has a higher reimbursement, so the technology has become very popular. But in the U.S., it's a difficult product to sell, because doctors want the revenue from dye-based angiography.

"Despite that concern, I believe it's worth using OCTA," he concludes. "With it, you achieve increased patient satisfaction and patient wait time is decreased. You can see more patients, because dye-based angiography takes a long time to perform—and it's risky. So from an economic standpoint I think OCTA wins, even without the unique billing code."

The Syringe Factor

K. Bailey Freund, MD, a retina specialist and a clinical professor of ophthalmology at the New York University School of Medicine, notes that the type of syringe used to deliver the anti-VEGF medication has become a point of consideration. "Unless a syringe is silicone-free, droplets of silicone can end up in the medication," he explains. "Then, if you use syringes that don't have reservoir at the tip, which can trap the silicone as it's pushed down by the plunger, you're more likely to inject some of that silicone into the eye. Compounding pharmacies often use fixed-needle insulin syringes which lack a reservoir at the tip. As a result, some silicone oil often gets into the vitreous.

"Lucentis now offers a pre-filled syringe that's been designed, in part, to minimize this problem," he continues. "Also, with aflibercept, or ranibizumab dispensed in a vial, most retinal specialists use syringes with either a Luer-Lock or Luer-Slip design. These syringes have a dead space at the tip, so when the plunger is pushed all the way down, not all of the fluid in the syringe is in-

jected into the vitreous. However, that type of syringe is frequently not used for bevacizumab, in part because the presence of a dead space requires that more drug be added, which increases the cost.

"The SCORE trial investigators reported this issue with fixed-needle syringes when they were looking at a syringe pre-filled with an Allergan steroid," he says. "Furthermore, a study published in 2017 compared the amount of silicone expressed from syringes of different types.¹ It showed quite convincingly that when you use syringes with fixed needles, you're more likely to see silicone droplets expressed from the needle tip. I'm also a co-author of a recently published editorial in *Retina* which discusses this topic.²"

Dr. Freund admits that the number of incidents related to this problem and compounded bevacizumab is very small. "Nevertheless, there are potential issues with a compounded drug," he says. "All of those factors weigh into which drug you choose, even if you think there's not much of a difference between the drugs."

—CK

What's Next?

As effective as the current options are, new drugs and technology in the pipeline have surgeons hoping for even better outcomes—and a reduced injection burden.

• **Brolucizumab.** This is an investigational anti-VEGF drug from Novartis. “We hope it will provide a greater duration of action,” Dr. Rosenfeld explains.

Dr. Freund notes that brolucizumab was compared head to head with aflibercept in a trial. “The trial compared the labeled dosing of Eylea—every four weeks times three, extended to once every eight weeks—to brolucizumab, which is given every four weeks for three injections and then extended to every 12 weeks,” he explains. “With brolucizumab, a little more than half of the eyes could be maintained on the 12-week dosing to the final 48-week endpoint. Unfortunately, the trial design makes it hard to do a direct comparison between the drugs because patients treated with aflibercept were never extended beyond eight weeks. However, during the first three months, all the eyes were dosed every four weeks. During that period, there was more improvement in OCT thickness with brolucizumab than with aflibercept. “Part of the reason for this finding is that brolucizumab is a smaller molecule in a more concentrated formulation,” he points out. “When you inject the same volume of both drugs, the molar concentration of brolucizumab is approximately 10 times that of aflibercept. That means you’re getting a lot more drug in the eye with each injection of brolucizumab.”

One reason surgeons look forward to new options is the possibility that they’ll help to treat refractory patients. However, Dr. Freund points out that it could be problematic to use brolucizumab for this purpose, at least at the outset, because of the trial design. “Let’s say you have a patient that doesn’t dry up as much as you hope using one of the other agents,” he says. “In the trials of the other drugs there were arms in which patients were treated monthly to the endpoint. Brolucizumab didn’t have that, so it seems unlikely that the initial approval will include long-term monthly dosing. The reality is, if a refractory patient can’t go for four or five weeks on the other agents, they’ll probably have trouble going for eight or 12 weeks with brolucizumab. That may limit how much you can use that drug for these difficult, refractory patients.

“I believe the company is doing other studies that may eventually get a label allowing monthly treatment,” he adds, “but at the launch that could be a bit of an issue.”

• **Abicipar pegol.** Another drug being tested is Allergan’s abicipar pegol. “The trial design was basically 12-month dosing head-to-head between abicipar pegol and monthly ranibizumab,” Dr. Freund explains. “The new drug met its noninferiority endpoint, which is fairly impressive. Patients went to 12 weeks and seemed to do just as well as patients getting ranibizumab every month.

“The issue is that there’s been some inflammation with abicipar pegol,” he continues. “Initially about 15 percent of patients had inflammation. However, the company subsequently released data from a trial called MAPLE, where a change in the manufacturing of the drug reduced the inflammation rate to 8 or 9 percent, and so far, there’s no evidence that the inflammation caused in any of these patients was of much concern. So, you could decide to try abicipar pegol on your patient once. If the patient develops inflammation, you could stop. The risk/benefit ratio would be pretty good: There may be only a 9-percent chance that that the patient will have inflammation, but there’s close to a 90-percent chance that the patient will be able to be dosed once every 12 weeks. With brolucizumab you’d only have a 50-50 chance of reaching every 12 weeks. That’s one way to look at the data.”

• **Sustained delivery.** A key way to reduce the injection burden, of course, is via sustained delivery. “No one likes injections,” notes Dr. Freund. “But now Genentech has an implantable port that’s a refillable reservoir. Theoretically, you’d only have to do two refills a year.”

Dr. Rosenfeld says that data from the LADDER study, involving Genentech’s Port Delivery System with ranibizumab, was presented at the FLORetina meeting. “It appears to provide some extended benefit for patients,” he says. “Less-frequent dosing was needed. In addition, there’s a sustained-release tyrosine kinase inhibitor from Graybug Vision (Redwood City, California) that may or may not prove to be a long-term solution for this problem.”

• **Home monitoring.** “The key to monitoring those lesions is going to be some kind of home monitoring that’s reliable,” notes Dr. Rosenfeld. “Our dream device would be an OCT that a patient could use every day, or every other day. That would allow us to identify exudation as soon as it develops. Home OCT monitoring would be useful for following patients who have non-exudative, neovascular lesions that we can identify with OCT angiography, who are at high risk for exudation, as well as patients on a treat-and-extend regimen. If they were testing themselves with home OCT monitoring, we’d know as soon as the fluid develops or recurs.

“Notal Vision has a prototype that’s being tested that uses an AI algorithm capable of picking up exudation,” he continues. “There are other strategies being developed as well, but the Notal Vision device may become available within a year or two. Patients wouldn’t buy the instrument; it would be more of a lease situation. Medicare currently covers the cost of home monitoring, so for a nominal fee, perhaps in addition to Medicare coverage, you’d be able to have your patients monitored at home.”

• **Gene therapy.** “Two companies have gene therapies that involve injecting a viral vector into the eye,” says Dr. Freund. “Adverum Biotechnologies is in a Phase I trial with an intravitreal injection, and Regenxbio will soon be conducting a Phase II trial involving subretinal delivery of gene therapy. The viral vector inserts DNA into cells in the eye, so they start to produce either ranibizumab or aflibercept on their own. Theoretically, you could inject it once and you’d be done. To me that’s very exciting.” REVIEW

Dr. Freund is a consultant for Allergan, Novartis, Zeiss, OptoVue and Heidelberg Engineering, and receives research support from Genentech Roche. Dr. Rosenfeld has received research funding from and is a consultant for Carl Zeiss Meditec.

1. Emerson GG. Silicone oil droplets are more common in fluid from BD insulin syringes as compared to other syringes. *Journal Vitreoretinal Diseases* 2017;1:6:401-406.

2. Sharma A, Kumar N, Bandello F, Loewenstein A, Freund KB. Understanding intravitreal silicone oil droplets due to intravitreal injections. *Retina* 2019;39:7:1233-1235.

3. Singh RP, Stone TW, eds. 2018 Global Trends in Retina Survey: Chicago, IL. American Society of Retina Specialists; 2018.

4. Campochiaro PA, Marcus DM, Awh CC, et al. The Port Delivery System with ranibizumab for neovascular age-related macular degeneration: Results from the randomized Phase 2 Ladder clinical trial. *Ophthalmology* 2019 Apr 1. pii: S0161-6420(18)33328-1. doi: 10.1016/j.ophtha.2019.03.036. [Epub ahead of print]

Copyright © 2020 Jobson Medical Information LLC unless otherwise noted.

All rights reserved. Reproduction in whole or in part without permission is prohibited.

Transgenic delivery of VEGF to mouse skin leads to an inflammatory condition resembling human psoriasis

Yu-Ping Xia, Baosheng Li, Donna Hylton, Michael Detmar, George D. Yancopoulos, and John S. Rudge

Gene therapy approaches involving vascular endothelial growth factor (VEGF) to promote therapeutic angiogenesis are under consideration for conditions ranging from ischemic heart disease to nonhealing skin ulcers. Here we make the surprising observation that the transgenic delivery of VEGF to the skin results in a profound inflammatory skin condition with many of the cellular and molecular features of psoriasis, including the characteristic vascular changes, epidermal al-

terations, and inflammatory infiltrates. Even longstanding psoriatic disease remains dependent on the transgenic VEGF in this model because it can be effectively reversed by the addition of VEGF Trap, a potent VEGF antagonist. Previous attempts to faithfully replicate the psoriatic phenotype through the transgenic delivery of epidermal keratinocyte growth factors or inflammatory mediators generated phenotypes with only partial resemblance to human psoriasis, leaving

unanswered questions about the etiology of this disease. The ability of transgenic VEGF to induce a psoriasiform phenotype suggests a new etiology and treatment approach for this disease and further substantiates emerging concerns about possible proinflammatory adverse effects that might be associated with therapeutic attempts to deliver VEGF. (Blood. 2003;102:161-168)

© 2003 by The American Society of Hematology

Introduction

Vascular endothelial growth factor (VEGF) is a potent mediator of angiogenesis, prompting recent efforts to therapeutically exploit this factor in conditions involving pathologically decreased blood flow, such as ischemic heart disease and nonhealing skin ulcers. Some recent studies,¹⁻⁶ however, have raised concerns about whether the delivery of VEGF could also have deleterious consequences. Here we make the surprising observation that chronic transgenic delivery of VEGF to the skin can result in a profound inflammatory condition with many of the cellular and molecular hallmarks of human psoriasis, such as hyperplastic and inflamed dermal blood vessels,⁷ epidermal thickening (termed acanthosis) with aberrant keratinocyte differentiation,⁸ and characteristic inflammatory infiltrates.^{9,10}

It has long been known that the reddened appearance of psoriatic skin is caused by hyperplastic dermal blood vessels, that vascular changes occur early in this disease, and that levels of VEGF are elevated in psoriatic skin.^{7,11-13} In addition, the dermal vessels in psoriasis appear to be highly abnormal in that they are hyperpermeable, contributing to the edema that characterizes psoriatic skin, and in that they express markers of an inflamed vasculature, such as E-selectin, vascular cell adhesion molecule-1 (VCAM-1), and intracellular adhesion molecule-1 (ICAM-1).¹⁴⁻¹⁶ In addition, the levels of soluble adhesion molecules in the sera of psoriasis patients, particularly E-selectin, provide a valuable surrogate marker of disease severity and therapeutic efficacy of various treatments.¹⁷⁻²⁰ Despite this, most efforts at understanding the etiology of this disease have focused on the epidermal and inflammatory aberrations. In the abnormally thickened epidermis, the top layer (termed stratum corneum), usually consisting of

cornified keratinocytes lacking nuclei, instead contains cells with nuclei (termed parakeratosis). Furthermore, this keratinized upper layer is excessively thickened (termed hyperkeratosis). Most striking, the epidermis produces highly abnormal and characteristic fingerlike projections into the underlying dermis, termed rete ridges. The typical inflammatory cell infiltrate seen in psoriasis is composed of epidermal microabscesses, increased numbers of mast cells, neutrophils and macrophages in the dermis, and activated T cells in the dermis and epidermis.^{21,22} T-cell subsets achieve a unique distribution as psoriasis evolves, with CD4⁺ T cells congregating primarily in the dermis while CD8⁺ T cells migrate from their normal dermal position to the epidermis, which is usually free of leukocytes.^{23,24}

It is clear that psoriatic skin is a hotbed of epidermal growth factors and inflammatory mediators.^{23,25-30} Supportive evidence of a key role for such mediators comes from patients who respond to immunosuppressive, anti-inflammatory, and antiproliferative therapies such as cyclosporine, methotrexate, tacrolimus, corticosteroids, and ultraviolet-light-activated psoralen. However, extensive efforts aimed at transgenically delivering inflammatory mediators or keratinocyte growth factors to the skin have not completely reproduced the psoriatic phenotype^{28,31-36} (Figure 1), which has thus far only been faithfully modeled in animals by transplanting human psoriatic skin onto mice with severe combined immunodeficiency disease (SCID)^{10,23} (Figure 1). Factors such as keratinocyte growth factor, transforming growth factor- α (TGF- α), and interleukin-20 (IL-20) promote some degree of epidermal hyperplasia, in certain cases with associated inflammation, but fail to produce many of the hallmarks of human psoriasis.^{28,34,37,38} Transgenic

From Regeneron Pharmaceuticals, Tarrytown, NY; and Cutaneous Biology Research Center, Department of Dermatology, Massachusetts General Hospital, Harvard Medical School, Charlestown.

Submitted December 16, 2002; accepted March 3, 2003. Prepublished online as *Blood* First Edition Paper, March 20, 2003; DOI 10.1182/blood-2002-12-3793.

Supported in part by National Institutes of Health/National Cancer Institute grants CA69184 and CA86410 (M.D.).

Reprints: Y.-P. Xia, Regeneron Pharmaceuticals, Inc, 777 Old Saw Mill River Rd, Tarrytown, NY 10591; e-mail: yuping.xia@regeneron.com.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 U.S.C. section 1734.

© 2003 by The American Society of Hematology

Model Phenotype	Epidermal Changes				Vascular Changes	Inflammatory Changes	Injury Response	Percentage of Psoriasisiform Phenotype	
	Thickening	Altered Differentiation	rete Ridges	Papillomatosis	Dilation of Papillary Loops	Epidermal T Cell Infiltrate	Intra-epidermal Micro-abscesses		"Koebner" Phenomena
Natural Disease	Human Psoriasis								100%
Xeno-transplant	Human Psoriasis (SCID)								100%
Transgenic Mice	K14-VEGF								100%
	K14-Angiostatin (Gock et al)								100%
	VEGF ₁₆₄ Inducible (Garcia et al)								36-52%
	K10-DME6 (Bessing et al)								Is weak and patchy expression
	Transgenic P121 (Garcia et al)								5-30%
	K14-EGF (Gao et al)								0%
	K14-IGF1 (Vassar et al)								0%
	K14-IL10 (Grove et al)								Part
	K14-IL12 (Garcia et al)								0%
K14-IL20 (Kucharski et al)								0%	
Knockout Mouse	CD18 (Omland et al)								In PLN absent
Spontaneous Mutation	Fuzzy Skin								100%

Figure 1. Summary of various mouse models and their resemblance to human psoriasis. The top line indicates whether characteristic changes are seen in human psoriasis (Y for yes, N for no). Other models are compared with the human standard and are blocked in green if they match human psoriasis or in red if they do not match. Question marks and the yellow blocks indicate that the feature in question was not examined. Note that only 2 models precisely match human psoriasis in all the features indicated here—a xenotransplantation model in which human psoriatic skin was transplanted onto a SCID mouse (second line) and the K14-VEGF transgenic mouse discussed (third line).

delivery of amphiregulin has resulted in the most promising transgenic model, but this model still lacks the characteristic rete ridge projections seen in human psoriasis, and it is also prone to papillomatosis, which is not typical in the human disease²⁹ (Figure 1). In addition, a chronic inflammatory skin condition developed in mice in which CD18 was knocked out, but only when the CD18-deficient 129/Sv mice were backcrossed onto the PL/J strain.³⁹ Again, these mice lacked the rete ridge structures that are highly characteristic of human psoriasis (Figure 1).

The problems with the above transgenic models of psoriasis raise the possibility that there is an upstream predisposition for psoriasis that can somehow be triggered so as to lead to the extremely diverse cytokine and growth factor abnormalities that drive psoriasis and that none of these individual downstream mediators is sufficient to induce the full spectrum of psoriatic disease. Consistent with the notion that psoriasis involves an underlying predisposition, the wounding of asymptomatic skin in psoriatic patients can trigger a complete psoriatic response adjacent to the wound, in a classic reaction termed the Koebner phenomenon.⁴⁰

Our studies suggest that excess VEGF may provide just such a predisposition by inducing a vascular inflammatory response that then predisposes to more widespread tissue inflammation closely resembling the psoriatic state. We report that young mice transgenically overexpressing VEGF in the skin initially lack overt disease but have a predisposition such that wounding can elicit the psoriatic phenotype, analogous to the Koebner phenomenon in humans. In older transgenic mice, the condition progresses until a profound inflammatory skin condition spontaneously develops with many of the cellular and molecular hallmarks of psoriasis, from characteristic epidermal alterations including dramatic rete ridge formation to the inflammatory infiltrates typical of psoriasis. Even late-stage disease remains dependent on transgenic VEGF because it can be effectively reversed by the addition of a potent VEGF antagonist. The ability of transgenic VEGF to induce a psoriasisiform phenotype suggests a new etiology and treatment approach for this disease and further substantiates emerging concerns⁶ about possible proinflammatory adverse effects that might be associated with therapeutic attempts to deliver VEGF.

Materials and methods

K14-VEGF transgenic mice

A keratin-14 (K14)-based expression vector and a mouse cDNA encoding VEGF₁₆₄ were used to generate K14-VEGF transgenic mice on the FVB

genetic background, as previously described⁴¹; mice homozygous for this transgene were used throughout the studies described here. All animals in the facility are cared for in accordance with the "Guide for the Care and Use of Laboratory Animals" (National Research Council, 1996).

Tissue processing and immunostaining

Tissue from the K14-VEGF transgenic and wild-type littermate mice used in these studies was matched according to sex, age, and wound site. Fixed sections were immunostained with antimouse platelet-endothelial cell adhesion molecule-1 (PECAM-1) (CD31; BD PharMingen, San Diego, CA), antimouse CD4 (BD PharMingen), antimouse CD8 (BD PharMingen), antimouse F4/80 (Serotec, Oxford, England), or antimouse VEGF (R&D Systems, Minneapolis, MN) following the manufacturer's instructions. Stainings for keratinocyte proliferation and differentiation markers or leukocyte adhesion molecules were performed as previously described⁴¹ with rabbit polyclonal antibody against mouse keratin 6 (K6) (Babco, Richmond, CA) and rat monoclonal antibodies against mouse E-selectin (CD62E), ICAM-1 (CD54), and VCAM-1 (CD106; BD PharMingen) using the Vectastain ABC kit (Vector Laboratories, Burlingame, CA).

Histology

Hematoxylin and eosin (H&E) staining and trichrome staining were performed according to protocols previously described.⁴²

Injection of VEGF Trap

VEGF Trap is a fusion of the immunoglobulin 2 domain of human VEGFR1, the immunoglobulin 3 domain of human VEGFR2, and the Fc domain of human immunoglobulin G1 (IgG1), creating a forced homodimer that binds VEGF with high affinity (dissociation equilibrium constant, 1-5 pM) and prolonged in vivo half-life (1-2 days in mice).⁴³ The 6-month-old K14-VEGF homozygous transgenic mice were treated by the systemic administration of VEGF Trap by subcutaneous injection at a site distant from the psoriatic skin. Mice were treated with either 25 mg/kg VEGF Trap or 12.5 mg/kg human Fc, corresponding to an equal molar concentration as a control, using an injection schedule of every 3 days for 12 days that resulted in a total of 4 injections per animal. Mouse ear tissue was harvested on day 14 for subsequent histologic analyses.

Results

Young K14-VEGF transgenic mice display a mild pre-psoriatic phenotype

As previously reported, K14-VEGF transgenic mice overexpressing VEGF in the epidermis are fertile and overtly healthy.^{2,4}

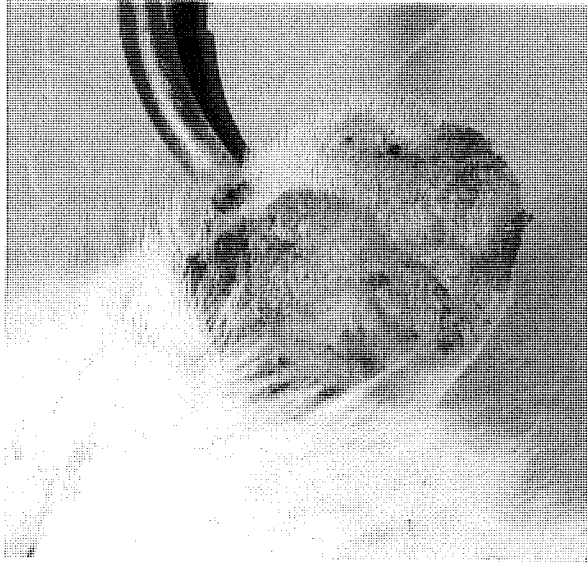


Figure 2. Psoriasisform phenotype. Erythematous, scaly, and thickened skin lesions with associated edema develop in homozygote K14-VEGF transgenic mice older than 5 months.

However, the ear skin of mice homozygous for this transgene is visibly redder than that of their wild-type *FVB* littermates. By 3 months, occasional focal skin lesions begin to develop on the ear and, to a lesser extent, on the dorsal and lateral skin. This condition worsens with age such that pronounced skin lesions are observed on the ears, neck, and snout by 5 months of age, with lesions

characterized by erythematous and scaly skin (Figure 2). These lesions coincided with sites of highest expression of the VEGF transgene (data not shown).

An initial histologic screen of the ear skin from young K14-VEGF transgenic mice, at 3 months of age, revealed a mild and potentially pre-psoriatic phenotype in these young mice using standard H&E staining—that is, the epidermis of these mice exhibited moderate acanthosis (epidermal hyperplasia) (Figure 3B, left inset), focal parakeratosis (keratinocytes in the stratum corneum retain nuclei) (Figure 3B, left inset), and mild rete ridge formation on the ventral ear surface (Figure 3B, arrows), compared with age-matched control littermates (Figure 3A). In the dermal compartment, edema contributing to an approximately 2- to 3-fold increase in tissue thickness was observed in the K14-VEGF mice, as was inflammatory cell infiltration in the subepidermal dermis (Figure 3; compare panels A and B).

Consistent with high-level transgenic overexpression of VEGF in the epidermis of these mice, VEGF protein was observed in the epidermis (where it is produced) and on dermal microvessels (where it presumably accumulates after diffusion into the dermis) in patterns (Figure 3A-B; compare right insets), remarkably reminiscent of those seen in human psoriasis.¹²

Young K14-VEGF transgenic mice exhibit a dramatic Koebner-like psoriatic response to injury

In contrast to the mild changes seen under basal conditions, creation of an excisional wound in the dorsal ear skin of 3-month-old K14-VEGF transgenic mice resulted in dramatic invaginations of the epidermis on the ventral side of the ear apposing the wound.

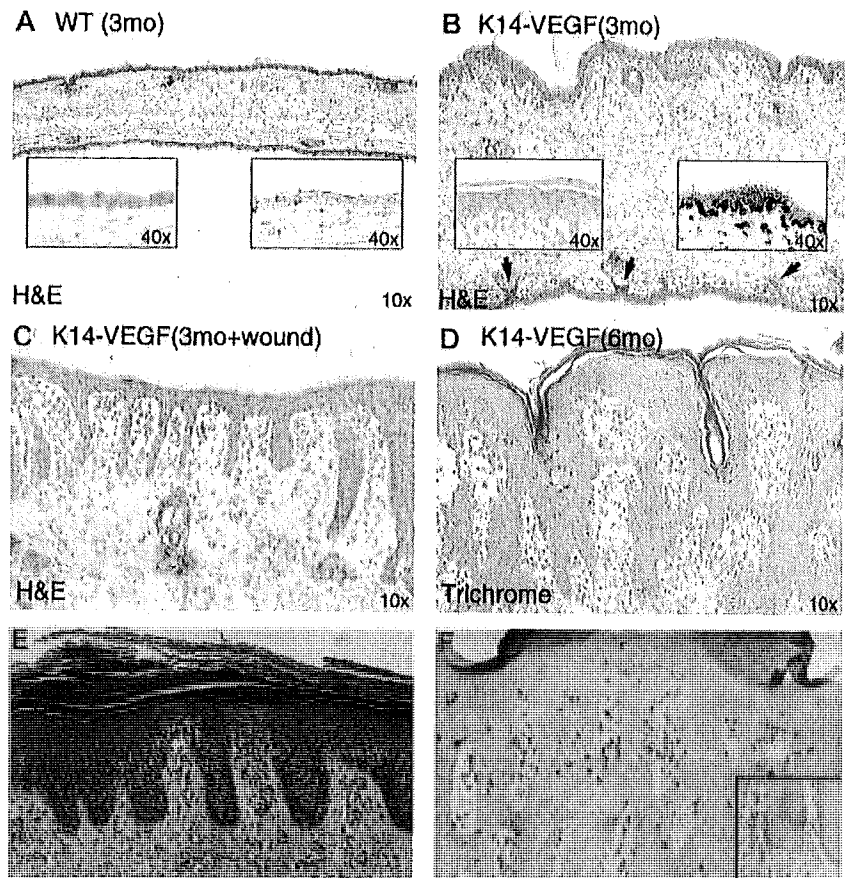


Figure 3. Histologic examination of ear skin from K14-VEGF transgenic mice, using H&E-stained tissue sections. (A) Control, wild-type littermate. Left inset shows epidermis at higher magnification; while right inset shows epidermis is negative for VEGF immunostaining. (B) Transgenic mouse (3 months of age) with edema, mild rete ridge formation on the ventral ear surface (arrows), epidermal acanthosis (left inset), and VEGF immunostaining in epidermis and in dermal microvessels (right inset). (C) Wound-induced rete ridge formation in a 3-month-old VEGF transgenic mouse. (D) Six-month-old VEGF transgenic mice showed spontaneous extensive rete ridge formation and anastomosis. (E) Early-stage human psoriasis shown for comparison. (F) Fully developed human psoriasis shown for comparison. Panels E-F reproduced with permission from Elder et al⁴⁰ and Nickoloff and Wrone-Smith,²³ respectively. Original magnifications: × 10 (A-D); and × 40 (insets).

These invaginations resembled the prominent rete ridge structures present in early human psoriasis (Figure 3E) and were observed in all 10 mouse ears used in the wounding study. Five ears were quantified for rete ridge counts (Figure 3C; Table 1). The induction of rete ridge formation in human pre-psoriatic skin has also been documented after skin injury and is termed the Koebner phenomenon.⁴⁰ Accompanying these rete ridge structures were increased dermal cellularity (inflammatory infiltrate), hyperkeratosis, and focal parakeratosis. These data suggest that to induce a severe psoriatic phenotype in the 3-month-old K14-VEGF transgenic animals, it would be necessary to introduce another stimulus such as wounding. This is consistent with the clinical evolution from pre-psoriatic skin to psoriatic lesion in humans (Koebner phenomenon).

Dramatic psoriasiform epidermal lesions spontaneously develop in older K14-VEGF mice

As the K14-VEGF transgenic mice aged, they spontaneously began to develop dramatic lesions resembling full-fledged psoriasis. K14-VEGF transgenic animals older than 5 months of age developed pronounced epidermal rete ridges and marked cutaneous inflammation (Figure 3D). In these lesions, psoriasiform hyperplasia with elongated rete ridges and anastomosis of neighboring rete ridges was found (Figure 3D), revealing a striking resemblance to fully developed psoriasis in humans (Figure 3F). It is important to note that rete ridge formation is one of the most characteristic and longest-recognized histologic features of human psoriasis and that no other transgenic mouse model results in rete ridge formation (Figure 1).

Hyperplastic and inflamed cutaneous blood vessels in K14-VEGF transgenic mice are similar to those observed in human psoriasis

To understand the nature of the visible skin redness in the K14-VEGF transgenic mice, we immunocytochemically stained ear sections with an antibody to an endothelial-cell-specific antigen, PECAM-1. When compared with microvessels in wild-type skin (Figure 4A) those in 3-month-old K14-VEGF mice with wound-induced psoriasis were obviously dilated and tortuous. The superficial vascular plexus showed that the most prominent angiogenic alterations consisted of vertically oriented vascular tufts in dermal papillae, similar to human psoriasis (Figure 4B). In the unwounded skin of older K14-VEGF mice (6 months of age), these enlarged vessels became more prominent within dermal papillae that were surrounded by a hyperproliferative epidermis undergoing rete ridge anastomosis (Figure 4C). Elongated and enlarged vessels found in the dermal papillae of K14-VEGF mice have a remarkable resemblance to the long, ectatic vessel loops seen in the dermal papillae of human psoriatic skin.

Because the K14-VEGF mice exhibited enlarged and tortuous vessels in dermal papillae analogous to those seen in human

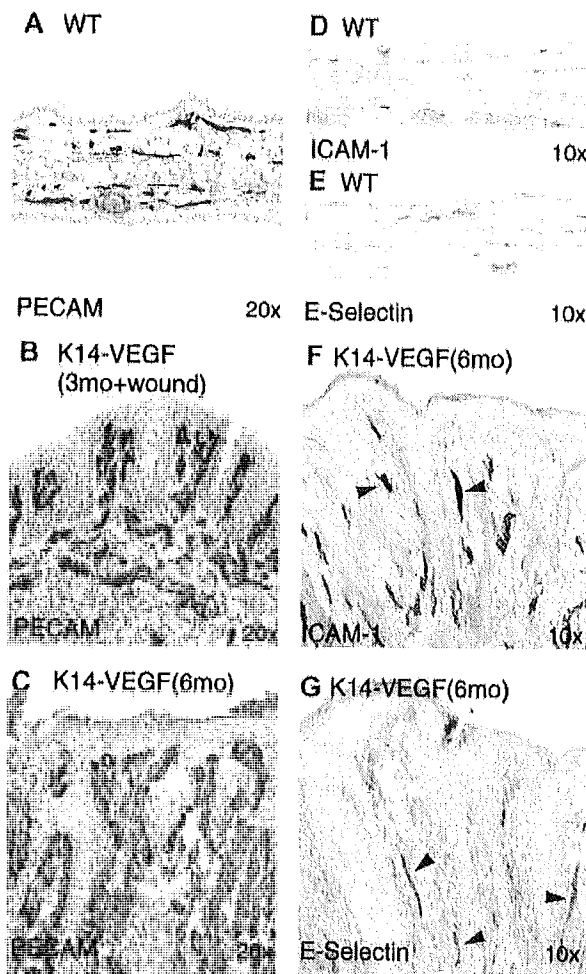


Figure 4. Hyperplastic and inflamed cutaneous blood vessels in K14-VEGF transgenic mice. Immunostaining was performed on cryosections of ear skin from wild-type littermate controls (A, D-E) and transgenic mice (B-C, F-G). PECAM staining showed increased vascular density mostly in the papillary dermis in wound-induced psoriasis in the 3-month-old transgenic mice (B). Enlarged vessels in 6-month-old transgenic mice showed vessels enclosed by anastomosing epidermal rete ridges (C). Immunostaining of E-selectin (G) and ICAM-1 (F) showed positive signals on dermal microvessels in transgenic mice (arrowheads). Original magnifications: $\times 20$ (A-C); and $\times 10$ (D-G).

psoriatic skin, we next explored whether these hyperplastic vessels also exhibited features of vascular inflammation seen in patients with psoriasis. In particular, the induction of specific endothelial cell adhesion molecules is a hallmark of the hyperplastic and inflamed vessels seen in human psoriatic skin lesions, including the induction of E-selectin (CD62E),¹⁵ VCAM-1 (CD106),¹⁶ and ICAM-1 (CD54).¹⁴ Similar to findings in human psoriasis, the expression of these cell adhesion molecules were prominent in hyperplastic vessels in the psoriasiform skin from K14-VEGF mice (Figure 4F-G).

Abnormal epidermal proliferation and differentiation in K14-VEGF mice resembling that seen in human psoriasis

Epidermal analysis of the psoriasiform lesions in K14-VEGF transgenic mice revealed hyperkeratosis (increased thickness of the stratum corneum) and parakeratosis (retention of nuclei in the cornified keratinocytes) (Figure 5A). Human psoriasis is characterized by similar hyperkeratosis and parakeratosis⁴⁴ and by altered epidermal hyperproliferation, as reflected by thickening of the

Table 1. Koebner phenomenon in young K14-VEGF transgenic mice

	Wild-type littermates, wounded	K14-VEGF mice, unwounded	K14-VEGF mice, wounded
Rete ridge counts/unit length of epithelium			
\pm SD, mm	0	2.0 \pm 1.3	11.3 \pm 5.2

Young is defined as approximately 3 months old. Each category of littermates consisted of 5 mice.

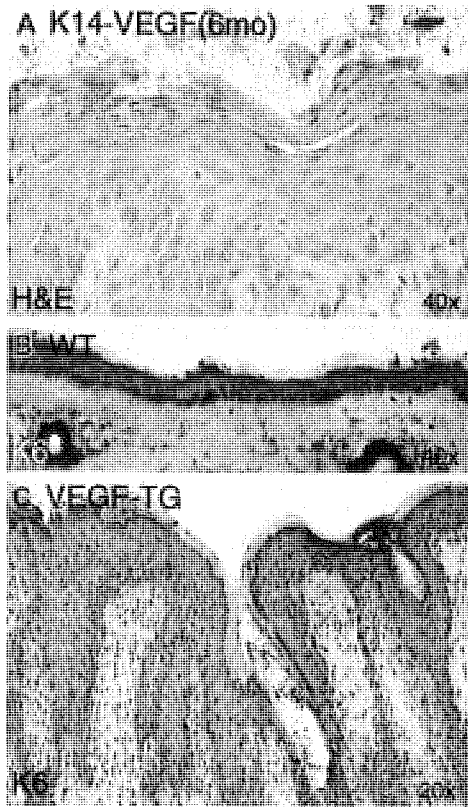


Figure 5. Abnormal epidermal proliferation and differentiation in K14-VEGF transgenic mice. Parakeratosis and hyperkeratosis were observed in H&E-stained skin sections from 6-month-old transgenic mice (A). Immunostaining of keratin K6 showed strong up-regulation throughout the epidermis (compare panels B and C) in 6-month-old transgenic mice.

epidermis and aberrant expression of hyperproliferation-associated keratins K6 and K16 throughout the epidermis, which are normally restricted to sporadic basal keratinocytes and hair follicles.⁴⁵ Another similarity we found between the psoriasiform lesions of K14-VEGF mice and human psoriasis was the strong expression of K6 throughout the hyperplastic epidermis of K14-VEGF mice (Figure 5C); normal mouse skin, with the exception of hair follicles and occasional basal keratinocytes, did not express K6 (Figure 5B).

K14-VEGF mice exhibit epidermal microabscesses and inflammatory infiltrates characteristic of human psoriasis

Neutrophil-filled lesions resembling the epidermal microabscesses found in advanced human psoriasis were observed in the epidermis of 6-month-old K14-VEGF transgenic mice (Table 2). One type of microabscess, mimicking the location of Munro microabscesses

Table 2. Quantitation of psoriatic phenotype in older K14-VEGF transgenic mice

	Wild-type littermates	K14-VEGF mice
Mice with lesions, %*	0	98
Intraepidermal CD8 ⁺ T-cell counts/unit length of epithelium ± SD, mm†	0	5.55 ± 2.10
Intraepidermal microabscesses per specimen‡	0	4.2

Older is defined as older than 5 months.
*n > 120 mice; †n = 4 mice; ‡n = 5 mice.

described in human psoriasis, was localized within the stratum corneum (Figure 6A),^{46,47} and a second type of microabscess, resembling Kogoj microabscesses seen in human psoriasis, was localized immediately beneath the stratum corneum (Figure 6B).^{46,47} The presence of microabscesses in human psoriatic skin is a key feature used in the clinical diagnosis of human psoriasis.^{46,47}

Analysis of the inflammatory cell infiltrate in 3-month-old K14-VEGF mice revealed a significant increase in both mast cells (as determined by staining for toluidine blue, which stains mast cell granules; Figure 6D) and macrophages (as determined with an antibody to the murine macrophage marker F4/80 antigen (Figure 6G) when compared with wild-type littermate controls (Figure 6C-F). Further increases in mast cells and macrophages were evident as the animals aged to 6 months (Figure 6E-H).

To assess the number and distribution of CD4⁺ and CD8⁺ T-lymphocytes, we immunostained for these cells in the 3- and 6-month-old transgenic animals. Results revealed massive infiltration of CD4⁺ T-lymphocytes localized primarily to the dermis of 3-month- (data not shown) and 6-month-old transgenic mice (Figure 6I). The overall level of CD8⁺ T-lymphocytes that infiltrated into the transgenic skin was significantly lower than that of

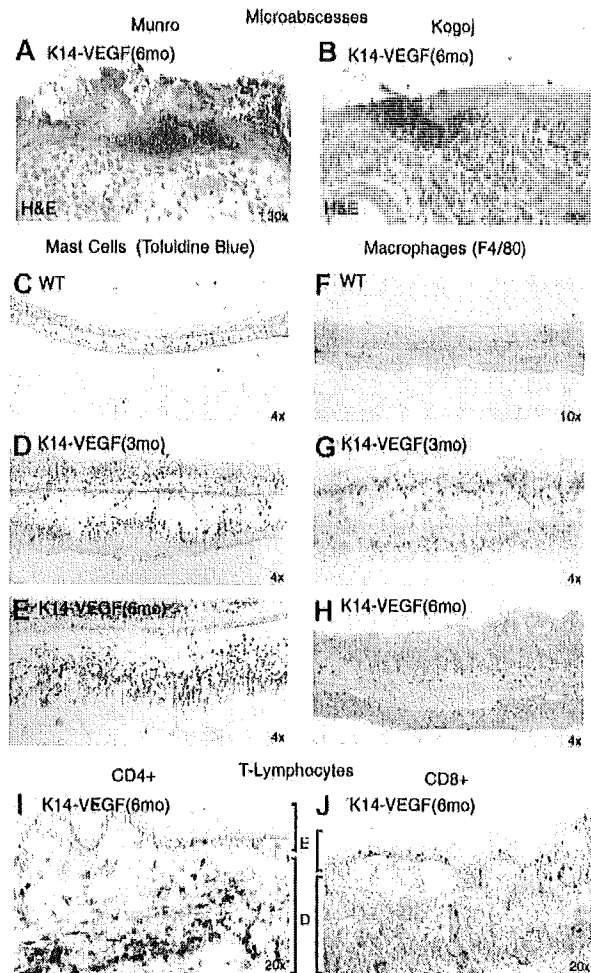


Figure 6. K14-VEGF mice exhibit epidermal microabscesses and inflammatory infiltrates characteristic of human psoriasis. (A) Munro-like microabscesses. (B) Kogoj-like microabscesses. Progressive increase of mast cell (C-E) and macrophage (F-H) density with 3-month-old (D, G) and 6-month-old (E, H) transgenic mice compared with wild-type littermates (C, F). CD4⁺ T-lymphocytes were detected primarily in the dermis (I) and CD8⁺ T-lymphocytes in the epidermis of 6-month-old transgenic mice (J). E indicates epidermis; D, dermis.

the CD4⁺ T-lymphocytes. In young K14-VEGF mice, these CD8⁺ T-lymphocytes were detected in the dermis and the epidermis (data not shown), whereas the CD8⁺ lymphocytes translocated and became localized to the epidermis in the 6-month-old transgenic mice (Figure 6J; Table 2). This complementary localization of CD4⁺ versus CD8⁺ lymphocytes in dermis and epidermis is also characteristic of human psoriatic skin.

Treatment with VEGF Trap normalizes the psoriatic phenotype in K14-VEGF transgenic mice

Our data demonstrate that many of the histologic and immunologic hallmarks of human psoriasis appear when VEGF is chronically overexpressed in mouse epidermis. To confirm the role of VEGF in the initiation and maintenance of this psoriatic phenotype and to attempt to ameliorate it, we used a potent VEGF inhibitor, VEGF Trap.⁴³ Six K14-VEGF transgenic mice at 6 months of age with obvious psoriatic lesions were systemically treated with VEGF Trap at a dose of 25 mg/kg every 3 days for 12 days. Although similar lesions in K14-VEGF mice do not regress spontaneously or after control treatments, in 4 of the K14-VEGF mice treated with VEGF Trap, pronounced visual improvement was observed in lesions on gross inspection. The other 2 K14-VEGF mice treated with VEGF Trap displayed moderate improvement. Using enzyme-linked immunosorbent assay (ELISA) to detect antibodies raised against VEGF Trap in these animals, we detected an obvious immune response only in the latter 2 moderate responders, suggesting partial immunoneutralization of VEGF Trap in these mice (data not shown). Histologic evaluation of all 6 K14-VEGF mice treated with VEGF Trap revealed near-complete resolution of the rete ridge elongations (Figure 7A-B), normalization of epidermal architecture and diminution of parakeratosis (Figure 7C-D),

and reduction in vascular hyperplasia (Figure 7E-F). In addition, the K6 marker of aberrant epidermal differentiation was normalized by VEGF Trap treatment (Figure 7G-H), as were markers of vascular inflammation E-selectin (Figure 7I-J), ICAM-1 in basal keratinocytes and vasculature (data not shown), and CD8⁺ T-lymphocyte distribution (Figure 7K-L).

Numerous reports have correlated the level of disease activity in psoriasis with the levels of soluble vascular adhesion molecules in the sera of patients with psoriasis, in particular soluble E-selectin that is presumably cleaved from the surface of inflamed dermal vessels.¹⁷⁻²⁰ Further correlating our animal model with the human condition, we find that serum levels of E-selectin are much higher in K14-VEGF mice (249.42 ± 38.64 ng/mL) than in control mice (37.34 ± 7.06 ng/mL), and, just as important, these increased levels are reduced with VEGF Trap treatment (to 96.74 ± 16.25 ng/mL). Altogether, our data indicate that VEGF is continuously required to maintain the psoriasiform lesions in older K14-VEGF mice and that even longstanding disease can be dramatically reversed with VEGF blockade.

Discussion

The underlying pathogenic mechanism and the key molecule(s) that are causative for psoriasis have not yet been identified. Recent studies for causative agents have focused on molecular mediators of inflammation or keratinocyte growth. However, attempts to mimic human psoriasis by transgenically overexpressing such mediators in mice do not completely recapitulate the human disease in all its pathologic aspects (Figure 1). In addition, the most

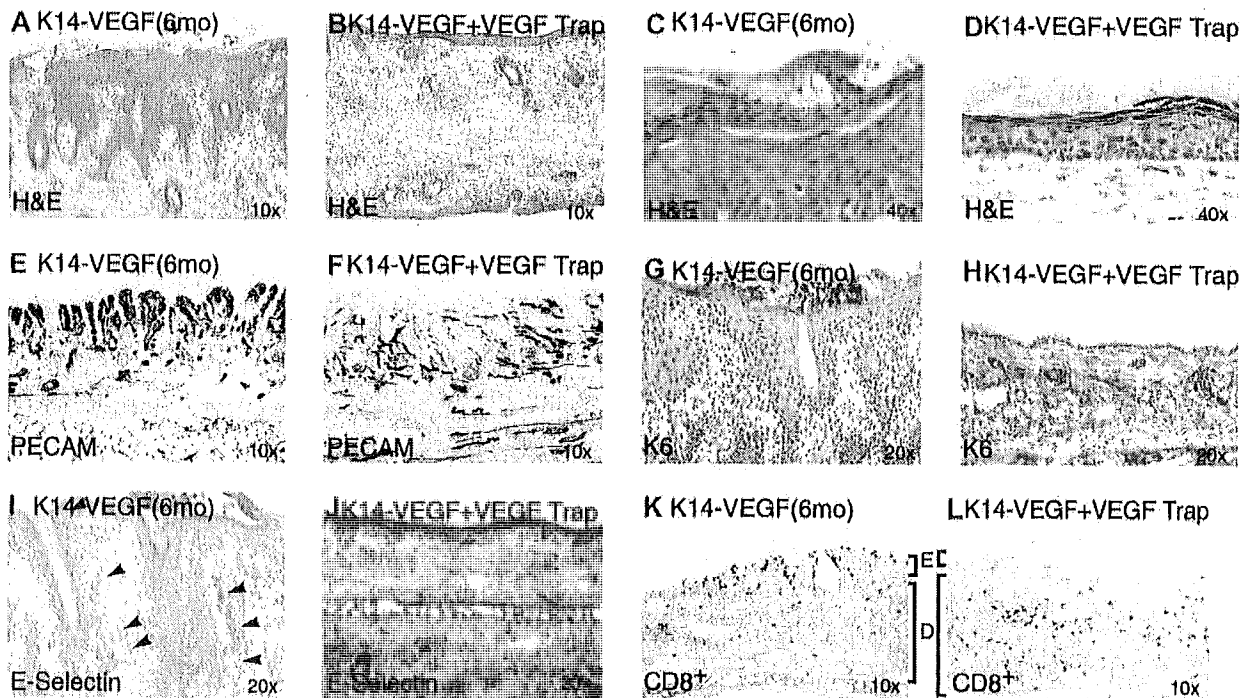


Figure 7. VEGF Trap normalizes the psoriatic phenotype in K14VEGF transgenic mice. Transgenic mice with severe skin lesions were injected with VEGF Trap (25 mg/kg) on days 0, 3, 7, and 12. Tissue was harvested on day 14 for histologic analysis. H&E staining of mouse ear skin treated with VEGF Trap showed clear resolution of rete ridges (compare panels A and B) and decreased parakeratosis/hyperkeratosis (compare panels C and D). Immunostaining with PECAM showed a drop-off of microvessels in the papillary dermis (compare panels E and F). Immunostaining with keratin K6 and E-selectin each showed remarkable down-regulation of signals in the epidermis (compare panels G and H), dermal capillaries (compare panels I and J), respectively. Arrowheads in panel I denote positive staining for E selectin. CD8⁺ T-lymphocytes shifted localization from the epidermis to the dermis in treated animals (compare panels K and L). E indicates epidermis; D, dermis.

faithful animal model of psoriasis to date requires the transplantation of human psoriatic skin to SCID mice²³ (Figure 1). Our findings lend credence to earlier suggestions that vascular changes might be among the earliest markers of the human psoriatic state.^{7,11} In particular, our studies suggest that VEGF, which has previously been shown to be dramatically elevated in human psoriatic skin,¹² might play a causative role in the vascular changes seen in this disease and also in epidermal and inflammatory alterations. Along these lines, we demonstrate that excess VEGF in the skin is sufficient to create a predisposition to a psoriatic phenotype and that such overexpression eventually leads to the spontaneous development of a psoriasiform condition in mice that recapitulates human psoriasis in many of its features—not only the hyperplastic and inflammatory vascular changes but also the characteristic epidermal alterations and tissue inflammatory cell infiltrates (Figure 1). Additional emerging evidence for a role of VEGF in the etiology of psoriasis comes from recent genetic analyses showing an association between VEGF promoter polymorphisms and the development of psoriatic symptoms (M.D., personal oral communication, November 2002). The induction of psoriasis by VEGF seems to be specific in that skin-specific transgenic delivery of another angiogenic factor, Ang1, does not result in a similar phenotype.¹⁴

In our model, it is clear that excess VEGF does not immediately cause full-blown disease. It takes up to 5 to 6 months for the development of obvious spontaneous disease. Overexpression of VEGF in the adult animal skin by viral gene transfer does not induce psoriasis because short-term VEGF expression is not sufficient to induce the psoriatic phenotype. In fact, it has been shown that the injection of a nonreplicating adenoviral vector, engineered to express VEGF₁₆₄, into the ears of athymic mice only temporarily induced the formation of dilated and leaky angiogenic vessels.⁴⁸ Thus, it seems likely that acute overexpression of VEGF creates a dilated, leaky, and inflamed cutaneous vasculature but that chronic overexpression is necessary to yield a more widespread inflammatory condition in the skin with profound epidermal changes resembling psoriasis. Although it is unclear how VEGF results in such widespread changes, it seems likely that the inflamed vasculature, which exhibits elevations in vascular adhesion molecules such as E-selectin, ICAM-1, and VCAM-1, plays a primary role by promoting the extravasation of inflammatory cells to the skin that then lend their own cytokine and chemokine mediators to the process. This inflammatory infiltrate and the tissue edema promoted by the leaky vessels may well compromise the normal barrier function of the skin, allowing the entry of exogenous antigens and further exacerbating the immune state. The creation of a diverse inflammatory milieu may then secondarily lead to epidermal alterations that seem to occur after the initial vascular and inflammatory changes in our model. Regardless of the mechanism by which chronically elevated VEGF in our model results in a psoriasiform phenotype, the maintenance of this abnormal state remains dependent on VEGF—we show that VEGF blockade late in this process can effectively reverse almost all the

observed abnormalities. It should perhaps not be surprising, based on previous studies indicating that VEGF can act as a potent and pleiotropic inflammatory agent, that transgenic delivery of VEGF to the skin can lead to a profound inflammatory skin condition. For example, a recent *in vitro* study using human umbilical vein endothelial cells (HUVECs) showed that VEGF stimulated the expression of ICAM-1, VCAM-1, and E-selectin through nuclear factor- κ B activation.⁴⁹ VEGF has also been shown to induce monocyte activation and chemotaxis through VEGF receptor-1 (Flt-1), which is expressed on monocytes.^{50,51} In addition, VEGF has been shown to induce expression of the chemokine IL-8, which is a potent modulator of the transendothelial migration of neutrophils.⁵²

Although we do not yet precisely understand how transgenic overexpression of VEGF eventually leads to a psoriasiform condition in mice, it seems impossible to ignore the possibility that VEGF may play a key causative role in human psoriasis, and it seems important to follow up on this possibility and on the implications for understanding and treating the human disease. Conventional psoriasis treatments that attempt to control the inflammatory response and subsequent epidermal hyperproliferation rely on immunosuppressants and antiproliferative therapy, involving considerable toxicity often without complete resolution. The use of a specific VEGF antagonist, such as VEGF Trap, to eliminate the hyperplastic vascular phenotype, suppress the associated inflammatory state, and reduce the levels of surrogate markers (such as E-selectin) of the disease in human psoriasis may provide a novel therapeutic strategy with minimal adverse side effects. It is also important to note that some existing or emerging therapies for psoriasis may act in part by blocking the VEGF pathway. For example, calcineurin inhibition by cyclosporine or FK-506 may block VEGF production or action,^{53,54} and tumor necrosis factor- α (TNF- α) and IL-1 seem to be potent inducers of VEGF. This induction may be important for some of their pathologic actions.^{55,56}

Our findings demonstrate that prolonged VEGF overexpression has powerful proinflammatory capabilities *in vivo* and leads to a skin phenotype resembling human psoriasis. VEGF is likely a key factor in the link between inflammation and angiogenesis. Therefore, its role should be explored in a variety of other inflammatory conditions. Furthermore, our findings substantiate emerging concerns⁶ about potential adverse effects that might be associated with therapeutic attempts to chronically deliver VEGF for proangiogenic purposes, particularly with regard to its profound proinflammatory capabilities.

Acknowledgments

We thank Drs Nick Gale and Virginia Hughes for maintaining transgenic mouse lines, Dr Dan Ragland for help with mast cell staining, and Drs Thomas Hawighorst and Jennifer Silva for contributing to the immunostaining. We also thank Scott Staton and Vicki Lan for image processing.

References

1. Suri C, McClain J, Thurston G, et al. Increased vascularization in mice overexpressing angiopoietin-1. *Science*. 1998;282:468-471.
2. Delmar M, Brown LF, Schon MP, et al. Increased microvascular density and enhanced leukocyte rolling and adhesion in the skin of VEGF transgenic mice. *J Invest Dermatol*. 1998;111:1-6.
3. Larcher F, Murillas R, Bolontrade M, Conti CJ, Jorcano JL. VEGF/VPF overexpression in skin of transgenic mice induces angiogenesis, vascular hyperpermeability and accelerated tumor development. *Oncogene*. 1998;17:303-311.
4. Thurston G, Suri C, Smith K, et al. Leakage-resistant blood vessels in mice transgenically overexpressing angiopoietin-1. *Science*. 1999;286:2511-2514.
5. Celletti FL, Waugh JM, Amabile PG, Brendolan A, Hilfiker PR, Dake MD. Vascular endothelial growth factor enhances atherosclerotic plaque progression. *Nat Med*. 2001;7:425-429.
6. Epstein SE, Kornowski R, Fuchs S, Dvorak HF. Angiogenesis therapy: amidst the hype, the neglected potential for serious side effects. *Circulation*. 2001;104:115-119.

7. Braverman IM, Sibley J. Role of the microcirculation in the treatment and pathogenesis of psoriasis. *J Invest Dermatol.* 1982;78:12-17.
8. Van de Kerkhof PC, Van Erp PE. The role of epidermal proliferation in the pathogenesis of psoriasis. *Skin Pharmacol.* 1996;9:343-354.
9. Nickoloff BJ. The immunologic and genetic basis of psoriasis. *Arch Dermatol.* 1999;135:1104-1110.
10. Nickoloff BJ. Characterization of lymphocyte-dependent angiogenesis using a SCID mouse: human skin model of psoriasis. *J Invest Dermatol Symp Proc.* 2000;5:67-73.
11. Schubert C, Christophers E. Mast cells and macrophages in early relapsing psoriasis. *Arch Dermatol Res.* 1985;277:352-358.
12. Detmar M, Brown LF, Claffey KP, et al. Overexpression of vascular permeability factor/vascular endothelial growth factor and its receptors in psoriasis. *J Exp Med.* 1994;180:1141-1146.
13. Bhushan M, McLaughlin B, Weiss JB, Griffiths CE. Levels of endothelial cell stimulating angiogenesis factor and vascular endothelial growth factor are elevated in psoriasis. *Br J Dermatol.* 1999;141:1054-1060.
14. Griffiths CE, Voorhees JJ, Nickoloff BJ. Characterization of intercellular adhesion molecule-1 and HLA-DR expression in normal and inflamed skin: modulation by recombinant gamma interferon and tumor necrosis factor. *J Am Acad Dermatol.* 1989;20:617-629.
15. Groves RW, Allen MH, Barker JN, Haskard DO, MacDonald DM. Endothelial leucocyte adhesion molecule-1 (ELAM-1) expression in cutaneous inflammation. *Br J Dermatol.* 1991;124:117-123.
16. Groves RW, Ross EL, Barker JN, MacDonald DM. Vascular cell adhesion molecule-1: expression in normal and diseased skin and regulation in vivo by interferon gamma. *J Am Acad Dermatol.* 1993;29:67-72.
17. Kowalick L, Neuber K, Weichenthal M, Kohler I, Ring J. Elevated serum-soluble ELAM-1 levels in patients with severe plaque-type psoriasis. *Arch Dermatol Res.* 1994;286:414-416.
18. Czech W, Schopf E, Kapp A. Soluble E-selectin in sera of patients with atopic dermatitis and psoriasis—correlation with disease activity. *Br J Dermatol.* 1996;134:17-21.
19. Krasowska D, Pietrzak A, Lecwicz-Torun B. Serum level of sELAM-1 in psoriatic patients correlates with disease activity. *J Eur Acad Dermatol Venereol.* 1999;12:140-142.
20. Szepletowski J, Wasik F, Bielicka E, Nockowski P, Noworolska A. Soluble E-selectin serum levels correlate with disease activity in psoriatic patients. *Clin Exp Dermatol.* 1999;24:33-36.
21. Bos JD, De Rie MA. The pathogenesis of psoriasis: immunological facts and speculations. *Immunol Today.* 1999;20:40-46.
22. Nickoloff BJ. Skin innate immune system in psoriasis: friend or foe? *J Clin Invest.* 1999;104:1161-1164.
23. Nickoloff BJ, Wrone-Smith T. Injection of psoriatic skin with CD4+ T cells induces psoriasis. *Am J Pathol.* 1999;155:145-158.
24. Weinstein GD, Krueger JG. Overview of psoriasis. In: Weinstein GD, Gottlieb AB, eds. *Therapy of Moderate to Severe Psoriasis.* Portland, OR: National Psoriasis Foundation; 1994:1-22.
25. Barker JN, Karabin GD, Stooft TJ, Sarma VJ, Dixit VM, Nickoloff BJ. Detection of interferon-gamma mRNA in psoriatic epidermis by polymerase chain reaction. *J Dermatol Sci.* 1991;2:106-111.
26. Kupper TS. The activated keratinocyte: a model for inducible cytokine production by non-bone marrow-derived cells in cutaneous inflammatory and immune responses. *J Invest Dermatol.* 1990;94(suppl):146S-150S.
27. Grossman RM, Krueger J, Yourish D, et al. Interleukin 6 is expressed in high levels in psoriatic skin and stimulates proliferation of cultured human keratinocytes. *Proc Natl Acad Sci U S A.* 1989;86:6367-6371.
28. Blumberg H, Conklin D, Xu WF, et al. Interleukin 20: discovery, receptor identification, and role in epidermal function. *Cell.* 2001;104:9-19.
29. Cook PW, Piepkorn M, Clegg CH, et al. Transgenic expression of the human amphiregulin gene induces a psoriasis-like phenotype. *J Clin Invest.* 1997;100:2286-2294.
30. Cooper KD, Hammerberg C, Baadsgaard O, et al. Interleukin-1 in human skin: dysregulation in psoriasis. *J Invest Dermatol.* 1990;95(suppl):24S-26S.
31. Cheng J, Turksen K, Yu QC, Schreiber H, Teng M, Fuchs E. Cachexia and graft-vs.-host-disease-type skin changes in keratin promoter-driven TNF alpha transgenic mice. *Genes Dev.* 1992;6:1444-1456.
32. Groves RW, Mizutani H, Kieffer JD, Kupper TS. Inflammatory skin disease in transgenic mice that express high levels of interleukin 1 alpha in basal epidermis. *Proc Natl Acad Sci U S A.* 1995;92:11874-11878.
33. Carroll JM, Crompton T, Seery JP, Watt FM. Transgenic mice expressing IFN-gamma in the epidermis have eczema, hair hypopigmentation, and hair loss. *J Invest Dermatol.* 1997;108:412-422.
34. Guo L, Yu QC, Fuchs E. Targeting expression of keratinocyte growth factor to keratinocytes elicits striking changes in epithelial differentiation in transgenic mice. *EMBO J.* 1993;12:973-986.
35. Blessing M, Schirmacher P, Kaiser S. Overexpression of bone morphogenetic protein-6 (BMP-6) in the epidermis of transgenic mice: inhibition or stimulation of proliferation depending on the pattern of transgene expression and formation of psoriatic lesions. *J Cell Biol.* 1996;135:227-239.
36. Carroll JM, Romero MR, Watt FM. Suprabasal integrin expression in the epidermis of transgenic mice results in developmental defects and a phenotype resembling psoriasis. *Cell.* 1995;83:957-968.
37. Vassar R, Fuchs E. Transgenic mice provide new insights into the role of TGF-alpha during epidermal development and differentiation. *Genes Dev.* 1991;5:714-727.
38. Schon MP. Animal models of psoriasis: what can we learn from them? *J Invest Dermatol.* 1999;112:405-410.
39. Bullard DC, Scharfetter-Kochanek K, McArthur MJ, et al. A polygenic mouse model of psoriasisiform skin disease in CD18-deficient mice. *Proc Natl Acad Sci U S A.* 1996;93:2116-2121.
40. Miller RA. The Koebner phenomenon. *Int J Dermatol.* 1982;21:192-197.
41. Streit M, Riccardi L, Velasco P, et al. Thrombospondin-2: a potent endogenous inhibitor of tumor growth and angiogenesis. *Proc Natl Acad Sci U S A.* 1999;96:14888-14893.
42. Xia YP, Zhao Y, Marcus J, et al. Effects of keratinocyte growth factor-2 (KGF-2) on wound healing in an ischaemia-impaired rabbit ear model and on scar formation. *J Pathol.* 1999;188:431-438.
43. Holash J, Davis S, Papadopoulos N, et al. VEGF-Trap: a VEGF blocker with potent antitumor effects. *Proc Natl Acad Sci U S A.* 2002;99:11393-11398.
44. Weinstein GD, Frost P. Abnormal cell proliferation in psoriasis. *J Invest Dermatol.* 1968;50:254-259.
45. Stoler A, Duvic M, Fuchs E. Unusual patterns of keratin expression in the overlying epidermis of patients with dermatofibromas: biochemical alterations in the epidermis as a consequence of dermal tumors. *J Invest Dermatol.* 1989;93:728-738.
46. Eider D, Elenitsas R, Jaworsky C, Johnson B. Algorithmic classification of skin disease for differential diagnosis. In: *Histopathology of the skin.* New York, NY: Lippincott-Raven; 1997:156-163.
47. Altman E, Kamino H. Diagnosis: psoriasis or not? what are the clues? *Semin Cutan Med Surg.* 1999;18:25-35.
48. Sundberg C, Nagy JA, Brown LF, et al. Glomeruloid microvascular proliferation follows adenoviral vascular permeability factor/vascular endothelial growth factor-164 gene delivery. *Am J Pathol.* 2001;158:1145-1160.
49. Kim I, Moon SO, Kim SH, Kim HJ, Koh YS, Koh GY. Vascular endothelial growth factor expression of intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion molecule 1 (VCAM-1), and E-selectin through nuclear factor-kappa B activation in endothelial cells. *J Biol Chem.* 2001;276:7614-7620.
50. Barleon B, Sozzani S, Zhou D, Weich HA, Mantovani A, Marme D. Migration of human monocytes in response to vascular endothelial growth factor (VEGF) is mediated via the VEGF receptor flt-1. *Blood.* 1996;87:3336-3343.
51. Clauss M, Weich H, Breier G, et al. The vascular endothelial growth factor receptor Flt-1 mediates biological activities: implications for a functional role of placenta growth factor in monocyte activation and chemotaxis. *J Biol Chem.* 1996;271:17629-17634.
52. Lee TH, Avraham H, Lee SH, Avraham S. Vascular endothelial growth factor modulates neutrophil transendothelial migration via upregulation of interleukin-8 in human brain microvascular endothelial cells. *J Biol Chem.* 2002;277:10445-10451.
53. Hernandez GL, Volpert OV, Iniguez MA, et al. Selective inhibition of vascular endothelial growth factor-mediated angiogenesis by cyclosporin A: roles of the nuclear factor of activated T cells and cyclooxygenase 2. *J Exp Med.* 2001;193:607-620.
54. Cho ML, Cho CS, Min SY, et al. Cyclosporine inhibition of vascular endothelial growth factor production in rheumatoid synovial fibroblasts. *Arthritis Rheum.* 2002;46:1202-1209.
55. Paleolog EM, Young S, Stark AC, McCloskey RV, Feldmann M, Maini RN. Modulation of angiogenic vascular endothelial growth factor by tumor necrosis factor alpha and interleukin-1 in rheumatoid arthritis. *Arthritis Rheum.* 1998;41:1258-1265.
56. Paleolog E. Target effector role of vascular endothelium in the inflammatory response: insights from the clinical trial of anti-TNF alpha antibody in rheumatoid arthritis. *Mol Pathol.* 1997;50:225-233.

Electronic Patent Application Fee Transmittal

Application Number:	16055847
Filing Date:	06-Aug-2018
Title of Invention:	USE OF A VEGF ANTAGONIST TO TREAT ANGIOGENIC EYE DISORDERS
First Named Inventor/Applicant Name:	George D. Yancopoulos
Filer:	Karl Bozicevic/Kimberly Zuehlke
Attorney Docket Number:	REGN-008CIPCON3

Filed as Large Entity

Filing Fees for Utility under 35 USC 111(a)

Description	Fee Code	Quantity	Amount	Sub-Total in USD(\$)
Basic Filing:				
Pages:				
Claims:				
Miscellaneous-Filing:				
Petition:				
Patent-Appeals-and-Interference:				
Post-Allowance-and-Post-Issuance:				
Extension-of-Time:				

Description	Fee Code	Quantity	Amount	Sub-Total in USD(\$)
Miscellaneous:				
SUBMISSION- INFORMATION DISCLOSURE STMT	1806	1	240	240
Total in USD (\$)				240

Electronic Acknowledgement Receipt

EFS ID:	39027245
Application Number:	16055847
International Application Number:	
Confirmation Number:	3451
Title of Invention:	USE OF A VEGF ANTAGONIST TO TREAT ANGIOGENIC EYE DISORDERS
First Named Inventor/Applicant Name:	George D. Yancopoulos
Customer Number:	96387
Filer:	Karl Bozicevic/Kimberly Zuehlke
Filer Authorized By:	Karl Bozicevic
Attorney Docket Number:	REGN-008CIPCON3
Receipt Date:	31-MAR-2020
Filing Date:	06-AUG-2018
Time Stamp:	20:00:19
Application Type:	Utility under 35 USC 111(a)

Payment information:

Submitted with Payment	yes
Payment Type	CARD
Payment was successfully received in RAM	\$240
RAM confirmation Number	E20203UK00473429
Deposit Account	
Authorized User	

The Director of the USPTO is hereby authorized to charge indicated fees and credit any overpayment as follows:

File Listing:					
Document Number	Document Description	File Name	File Size(Bytes)/ Message Digest	Multi Part /.zip	Pages (if appl.)
1	Transmittal Letter	0725US04_2020-03-31_Supp_IDS_trans_REGN-008CIPCON3.pdf	50733 775b25233e250ae2b14c905685e69d70eb10966a	no	2
Warnings:					
Information:					
2	Information Disclosure Statement (IDS) Form (SB08)	0725US04__2020-03-31_Supp_IDS_SB08A_REGN-008CIPCON3.pdf	36495 867caa9c3cb01ea1b0add9eef61cf513e53b5fd2	no	2
Warnings:					
Information:					
This is not an USPTO supplied IDS fillable form					
3	Non Patent Literature	Chatziralli_2018.pdf	1099422 fa8ce24228ee66646858acc7a38e7903fd5b157f	no	7
Warnings:					
Information:					
4	Non Patent Literature	Chung_2013.pdf	218908 041564f7a01739052957d3e9faf294746cc30266	no	10
Warnings:					
Information:					
5	Non Patent Literature	Cooper_1999.pdf	1217098 bcf4c7b7a55190f3168a8ea18183e7372994f2d8	no	11
Warnings:					
Information:					
6	Non Patent Literature	Croll_2004.pdf	2647465 622951c3c1068cf513951f9878c58568c890ab4d	no	15
Warnings:					

Information:					
7	Non Patent Literature	DeVriese_2001.pdf	472800	no	8
			315d7ee33b1ea94e70b3c2865ed739fbffb6c83		
Warnings:					
Information:					
8	Non Patent Literature	Eremina_2001.pdf	4088162	no	10
			24fe6cad852d68ab47d929eac59854d2373a3ed		
Warnings:					
Information:					
9	Non Patent Literature	Eriksson_and_Alitalo.pdf	2081053	no	17
			0a8d888a86fb2a6feed4a4cde01201eaff906a68		
Warnings:					
Information:					
10	Non Patent Literature	Ferrara.pdf	3622466	no	30
			ee47a2652aac7189cb40aa871921d0fb894747e		
Warnings:					
Information:					
11	Non Patent Literature	Ferrara_1999.pdf	437001	no	6
			1062f3fbd144b35ad7169cb169448a7aac2ab9e3		
Warnings:					
Information:					
12	Non Patent Literature	Flyvbjerg_2002.pdf	137211	no	5
			f16c80ee6a4d6123ed86ac9a82a2dcc1fb09e27f		
Warnings:					
Information:					
13	Non Patent Literature	Holash_1999.pdf	1253369	no	6
			3b1a7a106cecfcd3e27755b634ac865eba231		
Warnings:					
Information:					

14	Non Patent Literature	Korobelnik_2014.pdf	475993	no	7
			a018a34e0e817b45dc88156395903a71df424106		
Warnings:					
Information:					
15	Non Patent Literature	Mitchell_2013.pdf	2555569	no	13
			aa12fff21717962d60a0f87ccaee3586ef3f5c62		
Warnings:					
Information:					
16	Non Patent Literature	Noguera-Troise_2006.pdf	4146971	no	6
			41fa22c0bc3a6e37d0b1d0b3f90f2c1a5413daa9		
Warnings:					
Information:					
17	Non Patent Literature	Rudge_2007.pdf	2813823	no	8
			c03cb1fcf59ebe11e95d7e2d7094407b49be5260		
Warnings:					
Information:					
18	Non Patent Literature	Schmidt-Erfurth_2014.pdf	647240	no	9
			3dc7c75d15c920d816a7c2c519e2d6ff72485267		
Warnings:					
Information:					
19	Non Patent Literature	Semeraro_2013.pdf	239051	no	12
			9c73e3927d9432e267858cf261f42789ddf8ca9a		
Warnings:					
Information:					
20	Non Patent Literature	Tannock_2013.pdf	264317	no	9
			7699deca5f6f643b791e1b7f84a8b8705e664be1		
Warnings:					
Information:					

21	Non Patent Literature	Thurston_2002.pdf	430961 cd5fea4cd11d72c0b3f1cd4c029ea57ddd818398	no	6
Warnings:					
Information:					
22	Non Patent Literature	Anti-VEGF_2019.pdf	3877709 38be305da8c8aa0c9220de7e694de59f835d769c	no	10
Warnings:					
Information:					
23	Non Patent Literature	Xia_Blood_2013.pdf	2098399 07486287a13cc7c26c9adf9542d1998a9a491e29	no	8
Warnings:					
Information:					
24	Fee Worksheet (SB06)	fee-info.pdf	30914 1179b6d1be16e9e9e7ddc5263c98163a9cc8d2d	no	2
Warnings:					
Information:					
Total Files Size (in bytes):				34943130	
<p>This Acknowledgement Receipt evidences receipt on the noted date by the USPTO of the indicated documents, characterized by the applicant, and including page counts, where applicable. It serves as evidence of receipt similar to a Post Card, as described in MPEP 503.</p> <p><u>New Applications Under 35 U.S.C. 111</u> If a new application is being filed and the application includes the necessary components for a filing date (see 37 CFR 1.53(b)-(d) and MPEP 506), a Filing Receipt (37 CFR 1.54) will be issued in due course and the date shown on this Acknowledgement Receipt will establish the filing date of the application.</p> <p><u>National Stage of an International Application under 35 U.S.C. 371</u> If a timely submission to enter the national stage of an international application is compliant with the conditions of 35 U.S.C. 371 and other applicable requirements a Form PCT/DO/EO/903 indicating acceptance of the application as a national stage submission under 35 U.S.C. 371 will be issued in addition to the Filing Receipt, in due course.</p> <p><u>New International Application Filed with the USPTO as a Receiving Office</u> If a new international application is being filed and the international application includes the necessary components for an international filing date (see PCT Article 11 and MPEP 1810), a Notification of the International Application Number and of the International Filing Date (Form PCT/RO/105) will be issued in due course, subject to prescriptions concerning national security, and the date shown on this Acknowledgement Receipt will establish the international filing date of the application.</p>					

Electronically Filed

INFORMATION DISCLOSURE STATEMENT	Attorney Docket No.	REGN-008CIPCON3
	Confirmation No.	3451
	First Named Inventor	George D. Yancopoulos
	Application Number	16/055,847
	Filing Date	August 6, 2018
	Group Art Unit	1647
	Examiner Name	Jon McClelland Lockard
	Address to: Commissioner for Patents P.O. Box 1450 Alexandria, VA 22313-1450	Title: <i>“Use of a VEGF Antagonist to Treat Angiogenic Eye Disorders”</i>

Sir:

Applicants submit herewith documents which may be material to the examination of this application and in respect of which there may be a duty to disclose in accordance with 37 C.F.R. § 1.56. This submission is not intended to constitute an admission that any document referred to therein is "prior art" for this invention unless specifically designated as such. A listing of the documents is shown on enclosed Form PTO/SB/08A and copies of the foreign patents and non-patent literature are also enclosed.

The Examiner is requested to make the documents listed on the enclosed PTO/SB/08A of record in this application. Applicants would appreciate the Examiner initialing and returning the initialed copy of form PTO/SB/08A, indicating the documents cited therein have been considered and made of record herein.

Statements

No statement

.....
 PTA Statement under 37 CFR § 1.704(d)(1): Each item of information contained in the information disclosure statement filed herewith:

(i) Was first cited in any communication from a patent office in a counterpart foreign or international application or from the Office, and this communication was not received by any individual designated in § 1.56(c) more than thirty days prior to the filing of the information disclosure statement; or

(ii) Is a communication that was issued by a patent office in a counterpart foreign or international application or by the Office, and this communication was not received by any individual designated in § 1.56(c) more than thirty days prior to the filing of the information disclosure statement.

- IDS Statement under 37 CFR § 1.97(e)(1):** Each item of information contained in the information disclosure statement was first cited in any communication from a foreign patent office in a counterpart foreign application not more than three months prior to the filing of the information disclosure statement; or
- IDS Statement under 37 CFR § 1.97(e)(2):** No item of information contained in the information disclosure statement was cited in a communication from a foreign patent office in a counterpart foreign application, and, to the knowledge of the person signing the certification after making reasonable inquiry, no item of information contained in the information disclosure statement was known to any individual designated in § 1.56(c) more than three months prior to the filing of the information disclosure statement.

Fees

- No fee is believed to be due.
- The appropriate fee set forth in 37 C.F.R. §1.17(p) accompanies this information disclosure statement.

The Commissioner is hereby authorized to charge any underpayment of fees up to a strict limit of \$3,000.00 beyond that authorized on the credit card, but not more than \$3,000.00 in additional fees due with any communication for the above referenced patent application, including but not limited to any necessary fees for extensions of time, or credit any overpayment of any amount to Deposit Account No. 50-0815, order number REGN-008CIPCON3.

Respectfully submitted,
BOZICEVIC, FIELD & FRANCIS LLP

Date: 31 March 2020

By: /Karl Bozicevic, Reg. No. 28,807/
Karl Bozicevic
Reg. No. 28,807

BOZICEVIC, FIELD & FRANCIS LLP
201 Redwood Shores Parkway, Suite 200
Redwood City, CA 94065
Telephone: (650) 327-3400
Facsimile: (650) 327-3231

To: docket@bozpat.com,,
From: PAIR_eOfficeAction@uspto.gov
Cc: PAIR_eOfficeAction@uspto.gov
Subject: Private PAIR Correspondence Notification for Customer Number 96387

Apr 01, 2020 04:13:03 AM

Dear PAIR Customer:

Regeneron - Bozicevic, Field & Francis
201 REDWOOD SHORES PARKWAY
SUITE 200
REDWOOD CITY, CA 94065
UNITED STATES

The following USPTO patent application(s) associated with your Customer Number, 96387 , have new outgoing correspondence. This correspondence is now available for viewing in Private PAIR.

The official date of notification of the outgoing correspondence will be indicated on the form PTOL-90 accompanying the correspondence.

Disclaimer:

The list of documents shown below is provided as a courtesy and is not part of the official file wrapper. The content of the images shown in PAIR is the official record.

Application	Document	Mailroom Date	Attorney Docket No.
16055847	NOA	04/01/2020	REGN-008CIPCON3
	INTV.SUM.EX	04/01/2020	REGN-008CIPCON3
	1449	04/01/2020	REGN-008CIPCON3
	1449	04/01/2020	REGN-008CIPCON3

To view your correspondence online or update your email addresses, please visit us anytime at <https://sportal.uspto.gov/secure/myportal/privatepair>.

If you have any questions, please email the Electronic Business Center (EBC) at EBC@uspto.gov with 'e-Office Action' on the subject line or call 1-866-217-9197 during the following hours:

Monday - Friday 6:00 a.m. to 12:00 a.m.

Thank you for prompt attention to this notice,

UNITED STATES PATENT AND TRADEMARK OFFICE
PATENT APPLICATION INFORMATION RETRIEVAL SYSTEM



UNITED STATES PATENT AND TRADEMARK OFFICE

UNITED STATES DEPARTMENT OF COMMERCE
United States Patent and Trademark Office
Address: COMMISSIONER FOR PATENTS
P.O. Box 1450
Alexandria, Virginia 22313-1450
www.uspto.gov

Table with 5 columns: APPLICATION NO., FILING DATE, FIRST NAMED INVENTOR, ATTORNEY DOCKET NO., CONFIRMATION NO. Includes application details for George D. Yancopoulos and examiner information for Lockard, Jon McClelland.

Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Notice of the Office communication was sent electronically on above-indicated "Notification Date" to the following e-mail address(es):

docket@bozpat.com

CORRECTED
Notice of Allowability

Application No.
16/055,847

Applicant(s)
Yancopoulos, George D.

Examiner
JON M LOCKARD

Art Unit
1647

AIA (FITF) Status
No

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address--

All claims being allowable, PROSECUTION ON THE MERITS IS (OR REMAINS) CLOSED in this application. If not included herewith (or previously mailed), a Notice of Allowance (PTOL-85) or other appropriate communication will be mailed in due course. **THIS NOTICE OF ALLOWABILITY IS NOT A GRANT OF PATENT RIGHTS.** This application is subject to withdrawal from issue at the initiative of the Office or upon petition by the applicant. See 37 CFR 1.313 and MPEP 1308.

1. This communication is responsive to the IDS filed 31 March 2020.
 A declaration(s)/affidavit(s) under **37 CFR 1.130(b)** was/were filed on _____.
2. An election was made by the applicant in response to a restriction requirement set forth during the interview on _____; the restriction requirement and election have been incorporated into this action.
3. The allowed claim(s) is/are 21-23 (renumbered as claims 1-3, respectively). As a result of the allowed claim(s), you may be eligible to benefit from the **Patent Prosecution Highway** program at a participating intellectual property office for the corresponding application. For more information, please see http://www.uspto.gov/patents/init_events/pph/index.jsp or send an inquiry to PPHfeedback@uspto.gov.
4. Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).

Certified copies:

- a) All b) Some *c) None of the:
- Certified copies of the priority documents have been received.
 - Certified copies of the priority documents have been received in Application No. _____.
 - Copies of the certified copies of the priority documents have been received in this national stage application from the International Bureau (PCT Rule 17.2(a)).

* Certified copies not received: _____.

Applicant has THREE MONTHS FROM THE "MAILING DATE" of this communication to file a reply complying with the requirements noted below. Failure to timely comply will result in ABANDONMENT of this application.

THIS THREE-MONTH PERIOD IS NOT EXTENDABLE.

5. CORRECTED DRAWINGS (as "replacement sheets") must be submitted.
 including changes required by the attached Examiner's Amendment / Comment or in the Office action of Paper No./Mail Date _____.
- Identifying indicia such as the application number (see 37 CFR 1.84(c)) should be written on the drawings in the front (not the back) of each sheet. Replacement sheet(s) should be labeled as such in the header according to 37 CFR 1.121(d).**
6. DEPOSIT OF and/or INFORMATION about the deposit of BIOLOGICAL MATERIAL must be submitted. Note the attached Examiner's comment regarding REQUIREMENT FOR THE DEPOSIT OF BIOLOGICAL MATERIAL.

Attachment(s)

- | | |
|---|---|
| 1. <input type="checkbox"/> Notice of References Cited (PTO-892) | 5. <input checked="" type="checkbox"/> Examiner's Amendment/Comment |
| 2. <input checked="" type="checkbox"/> Information Disclosure Statements (PTO/SB/08),
Paper No./Mail Date _____. | 6. <input type="checkbox"/> Examiner's Statement of Reasons for Allowance |
| 3. <input type="checkbox"/> Examiner's Comment Regarding Requirement for Deposit
of Biological Material _____. | 7. <input type="checkbox"/> Other _____. |
| 4. <input type="checkbox"/> Interview Summary (PTO-413),
Paper No./Mail Date _____. | |

/CHRISTINE J SAOUD/
Primary Examiner, Art Unit 1647

Notice of Pre-AIA or AIA Status

1. The present application is being examined under the pre-AIA first to invent provisions.

Information Disclosure Statement

2. The information disclosure statement (IDS) submitted on 31 March 2020 was filed after the mailing date of the Non-Final rejection on 01 October 2019. The submission is in compliance with the provisions of 37 CFR 1.97. Accordingly, the information disclosure statement is being considered by the examiner. The Examiner would also like to note that he is aware of co-pending U.S. Patent Application No. 16/397,267, filed 29 April 2019.

EXAMINER'S COMMENT

3. The information disclosure statement (IDS) filed 31 March 2020 has been considered by the Examiner. After careful consideration, the Examiner has determined that none of the information contained therein raises new issues of patentability.

Advisory Information

Any inquiry concerning this communication or earlier communications from the examiner should be directed to **Jon M. Lockard** whose telephone number is **(571) 272-2717**. The examiner can normally be reached on Monday through Friday, 8:00 AM to 4:30 PM.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, **Joanne Hama**, can be reached on **(571) 272-2911**. The fax number for the organization where this application or proceeding is assigned is **571-273-8300**.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at **866-217-9197** (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

/Christine J Saoud/
Primary Examiner, Art Unit 1647

/J.L/
Examiner, Art Unit 1647
June 24, 2020

INFORMATION DISCLOSURE STATEMENT BY APPLICANT			Application Number	16/055,847	
			Filing Date	August 6, 2018	
			First Named Inventor	George D. Yancopoulos	
			Art Unit	1647	
			Examiner Name	Jon M. Lockard	
Sheet	1	of	2	Attorney Docket Number	REGN-008CIPCON3

U.S. PATENT DOCUMENTS						
Examiner Initial*	Cite No.	Patent Number		Issue Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear
		Number-Kind Code (if known)				
	1	7070959		2006-07-04	Papadopoulos	
	2	8092803		2012-01-10	Furfine et al.	
	3	10406226		2019-09-10	Dix et al.	
	4	10464992		2019-11-05	Furfine et al.	

U.S. PATENT APPLICATION PUBLICATIONS						
Examiner Initial*	Cite No.	Publication Number		Publication Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear
		Number-Kind Code (if known)				
	1	2019/0388539		2019-12-26	Dix et al.	
	2	2020/0017572		2020-01-16	Furfine et al.	

FOREIGN PATENT DOCUMENTS							
Examiner Initial*	Cite No.	Foreign Document Number		Publication Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear	T
		Country Code-Number-Kind Code (if known)					
	1						
	2						

NON PATENT LITERATURE DOCUMENTS			
Examiner Initials*	Cite No.	Include name of the author (in CAPITAL LETTERS), title of the article (when appropriate), title of the item (book, magazine, journal, serial, symposium, catalog, etc.), date, page(s), volume-issue number(s), publisher, city and/or country where published.	T
	1	ANONYMOUS "Anti-VEGF 2019: The State of the Art" Review of Ophthalmology (published August 5, 2019)	
	2	CHATZIRALLI et al. "Intravitreal aflibercept for neovascular age-related macular degeneration in patients aged 90 years or older: 2-year visual acuity outcomes" Eye (2018) 32:1523-1529	
	3	CHUNG et al. "Ziv-aflibercept: A novel angiogenesis inhibitor for the treatment of metastatic colorectal cancer" Am J Heath-Syst Pharm (November 1, 2013) 70:1887-1896	
	4	COOPER et al., "Increased Renal Expression of Vascular Endothelial Growth Factor (VEGF) and Its Receptor VEGFR-2 in Experimental Diabetes" Diabetes (1999) 48:2229-2239	
	5	CROLL et al., "VEGF-mediated inflammation precedes angiogenesis in adult brain" Experimental Neurology (2004) 187:388-402	
	6	DeVRIESE et al., "Antibodies against Vascular Endothelial Growth Factor Improve Early Renal Dysfunction in Experimental Diabetes" J. Am. Soc. Nephrol (2001) 12:993-1000	
	7	EREMINA et al., "Glomerular-specific alterations of VEGF-A expression lead to distinct congenital and acquired renal diseases" Journal of Clinical Investigation (March 2003) 111(5):707-716	
	8	ERIKSSON et al., "Structure, Expression and Receptor-Binding Properties of Novel Vascular Endothelial Growth Factors" Vascular Growth Factors and Angiogenesis, Springer (1999) pp. 41-57	

Examiner Signature		Date Considered	
--------------------	--	-----------------	--

*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

INFORMATION DISCLOSURE STATEMENT BY APPLICANT			Application Number	16/055,847	
			Filing Date	August 6, 2018	
			First Named Inventor	George D. Yancopoulos	
			Art Unit	1647	
			Examiner Name	Jon M. Lockard	
Sheet	2	of	2	Attorney Docket Number	REGN-008CIPCON3

NON PATENT LITERATURE DOCUMENTS			
Examiner Initials*	Cite No.	Include name of the author (in CAPITAL LETTERS), title of the article (when appropriate), title of the item (book, magazine, journal, serial, symposium, catalog, etc.), date, page(s), volume-issue number(s), publisher, city and/or country where published.	T
	9	FERRARA, N. "Vascular Endothelial Growth Factor: Molecular and Biological Aspects" <i>Advances in Organ Biology</i> (1999) pp. 1-30	
	10	FERRARA et al., "Clinical applications of angiogenic growth factors and their inhibitors" <i>Nature Medicine</i> (December 1999) 5(12):1359-1364	
	11	FLYVBJERG et al., "Amelioration of Long-Term Renal Changes in Obese Type 2 Diabetic Mice by a Neutralizing Vascular Endothelial Growth Factor Antibody" <i>Diabetes</i> (October 2002) 51:3090-3094	
	12	HOLASH et al., "Vessel Cooption, Regression, and Growth in Tumors Mediated by Angiopoietins and VEGF" <i>Science</i> (June 18, 1999) 284(5422):1994-1998	
	13	KOROBELNIK et al., "Intravitreal Aflibercept Injection for Macular Edema Resulting from Central Retinal Vein Occlusion" <i>American Academy of Ophthalmology</i> (2014) 121(1):202-208	
	14	MITCHELL, Edith P. "Targeted Therapy for Metastatic Colorectal Cancer: Role of Aflibercept" <i>Clinical Colorectal Cancer</i> (2013) 12(2):73-85	
	15	NOGUERA-TROISE et al., "Blockade of D114 inhibits tumour growth by promoting non-productive angiogenesis" <i>Nature</i> (December 2006) 444:1032-1037	
	16	RUDGE et al., "VEGF Trap complex formation measures production rates of VEGF, providing a biomarker for predicting efficacious angiogenic blockade" <i>PNAS</i> (November 20, 2007) 104(47):18363-18370	
	17	SCHMIDT-ERFURTH et al., "Intravitreal Aflibercept Injection for Neovascular Age-related Macular Degeneration" <i>Ophthalmology</i> (2014) 121:193-201	
	18	SEMERARO et al., "Aflibercept in wet AMD: specific role and optimal use" <i>Drug Design, Development and Therapy</i> (August 2, 2013) 7:711-722	
	19	TANNOCK et al., "Aflibercept versus placebo in combination with docetaxel and prednisone for treatment of men with metastatic castration-resistant prostate cancer (VENICE): a phase 3, double-blind randomized trial" <i>Lancet Oncol</i> (2013) 14:760-768	
	20	THURSTON, Gavin "Complementary actions of VEGF and Angiopoietin-1 on blood vessel growth and leakage" <i>J. Anat.</i> (2002) 200:575-580	
	21	XIA et al., "Transgenic delivery of VEGF to mouse skin leads to an inflammatory condition resembling human psoriasis" <i>Blood</i> (July 1, 2003) 102(1):161-168	

Examiner Signature	/JON M LOCKARD/	Date Considered	06/24/2020
--------------------	-----------------	-----------------	------------

*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

REQUEST FOR CONTINUED EXAMINATION(RCE)TRANSMITTAL (Submitted Only via EFS-Web)

Application Number	16055847	Filing Date	2018-08-06	Docket Number (if applicable)	REGN-008CIPCON3	Art Unit	1647
First Named Inventor	George D. Yancopoulos			Examiner Name	Jon Lockard		

This is a Request for Continued Examination (RCE) under 37 CFR 1.114 of the above-identified application.
 Request for Continued Examination (RCE) practice under 37 CFR 1.114 does not apply to any utility or plant application filed prior to June 8, 1995, or to any design application. The Instruction Sheet for this form is located at WWW.USPTO.GOV

SUBMISSION REQUIRED UNDER 37 CFR 1.114

Note: If the RCE is proper, any previously filed unentered amendments and amendments enclosed with the RCE will be entered in the order in which they were filed unless applicant instructs otherwise. If applicant does not wish to have any previously filed unentered amendment(s) entered, applicant must request non-entry of such amendment(s).

Previously submitted. If a final Office action is outstanding, any amendments filed after the final Office action may be considered as a submission even if this box is not checked.

Consider the arguments in the Appeal Brief or Reply Brief previously filed on _____

Other _____

Enclosed

Amendment/Reply

Information Disclosure Statement (IDS)

Affidavit(s)/ Declaration(s)

Other _____

MISCELLANEOUS

Suspension of action on the above-identified application is requested under 37 CFR 1.103(c) for a period of months _____
 (Period of suspension shall not exceed 3 months; Fee under 37 CFR 1.17(i) required)

Other _____

FEES

The RCE fee under 37 CFR 1.17(e) is required by 37 CFR 1.114 when the RCE is filed.

The Director is hereby authorized to charge any underpayment of fees, or credit any overpayments, to Deposit Account No 500815

SIGNATURE OF APPLICANT, ATTORNEY, OR AGENT REQUIRED

<input checked="" type="checkbox"/>	Patent Practitioner Signature
	Applicant Signature

Signature of Registered U.S. Patent Practitioner			
Signature	Karl Bozicevic, Reg. No. 28,807/	Date (YYYY-MM-DD)	2020-06-30
Name	Karl Bozicevic	Registration Number	28807

This collection of information is required by 37 CFR 1.114. The information is required to obtain or retain a benefit by the public which is to file (and by the USPTO to process) an application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.11 and 1.14. This collection is estimated to take 12 minutes to complete, including gathering, preparing, and submitting the completed application form to the USPTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden, should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, P.O. Box 1450, Alexandria, VA 22313-1450.

If you need assistance in completing the form, call 1-800-PTO-9199 and select option 2.

Privacy Act Statement

The Privacy Act of 1974 (P.L. 93-579) requires that you be given certain information in connection with your submission of the attached form related to a patent application or patent. Accordingly, pursuant to the requirements of the Act, please be advised that: (1) the general authority for the collection of this information is 35 U.S.C. 2(b)(2); (2) furnishing of the information solicited is voluntary; and (3) the principal purpose for which the information is used by the U.S. Patent and Trademark Office is to process and/or examine your submission related to a patent application or patent. If you do not furnish the requested information, the U.S. Patent and Trademark Office may not be able to process and/or examine your submission, which may result in termination of proceedings or abandonment of the application or expiration of the patent.

The information provided by you in this form will be subject to the following routine uses:

1. The information on this form will be treated confidentially to the extent allowed under the Freedom of Information Act (5 U.S.C. 552) and the Privacy Act (5 U.S.C. 552a). Records from this system of records may be disclosed to the Department of Justice to determine whether the Freedom of Information Act requires disclosure of these records.
2. A record from this system of records may be disclosed, as a routine use, in the course of presenting evidence to a court, magistrate, or administrative tribunal, including disclosures to opposing counsel in the course of settlement negotiations.
3. A record in this system of records may be disclosed, as a routine use, to a Member of Congress submitting a request involving an individual, to whom the record pertains, when the individual has requested assistance from the Member with respect to the subject matter of the record.
4. A record in this system of records may be disclosed, as a routine use, to a contractor of the Agency having need for the information in order to perform a contract. Recipients of information shall be required to comply with the requirements of the Privacy Act of 1974, as amended, pursuant to 5 U.S.C. 552a(m).
5. A record related to an International Application filed under the Patent Cooperation Treaty in this system of records may be disclosed, as a routine use, to the International Bureau of the World Intellectual Property Organization, pursuant to the Patent Cooperation Treaty.
6. A record in this system of records may be disclosed, as a routine use, to another federal agency for purposes of National Security review (35 U.S.C. 181) and for review pursuant to the Atomic Energy Act (42 U.S.C. 218(c)).
7. A record from this system of records may be disclosed, as a routine use, to the Administrator, General Services, or his/her designee, during an inspection of records conducted by GSA as part of that agency's responsibility to recommend improvements in records management practices and programs, under authority of 44 U.S.C. 2904 and 2906. Such disclosure shall be made in accordance with the GSA regulations governing inspection of records for this purpose, and any other relevant (i.e., GSA or Commerce) directive. Such disclosure shall not be used to make determinations about individuals.
8. A record from this system of records may be disclosed, as a routine use, to the public after either publication of the application pursuant to 35 U.S.C. 122(b) or issuance of a patent pursuant to 35 U.S.C. 151. Further, a record may be disclosed, subject to the limitations of 37 CFR 1.14, as a routine use, to the public if the record was filed in an application which became abandoned or in which the proceedings were terminated and which application is referenced by either a published application, an application open to public inspections or an issued patent.
9. A record from this system of records may be disclosed, as a routine use, to a Federal, State, or local law enforcement agency, if the USPTO becomes aware of a violation or potential violation of law or regulation.

Electronically filed		
PRELIMINARY AMENDMENT UNDER 37 C.F.R. §1.115	Attorney Docket No.	REGN-008CIPCON3
	Confirmation No.	3451
	First Named Inventor	George D. Yancopoulos
	Application Number	16/055,847
	Filing Date	August 6, 2018
	Group Art Unit	1647
	Examiner Name	Jon McClelland Lockard
	Address to: Commissioner for Patents P.O. Box 1450 Alexandria, VA 22313-1450	

Sir:

This Preliminary Amendment is being submitted concurrently with a Request for Continued Examination (RCE). In view of the remarks put forth below, reconsideration and allowance are respectfully requested.

Amendments to the Claims are reflected in the listing of claims which begins on page 2 of this paper.

Remarks/Arguments begin on page 3 of this paper.

AMENDMENTS TO THE CLAIMS

1. - 20. (Canceled)

21. **(Previously Presented)** A method for treating macular edema following retinal vein occlusion in a human subject comprising administering 2 mg aflibercept to the subject by intravitreal injection once every 4 weeks.

22. **(Previously Presented)** The method of claim 21 wherein the aflibercept is administered in a volume of 0.05 ml.

23. **(Previously Presented)** The method of claim 22 wherein the aflibercept is in a pharmaceutical formulation comprising a pharmaceutically acceptable carrier.

REMARKS

FORMAL MATTERS

Claims 21-23 are pending in this application

Claims 1-20 were previously cancelled.

No claims are amended.

No new matter is added.

ALLOWED CLAIMS

The claims that are pending here and shown above are identical to the claims that were allowed in the Notice of Allowance dated April 1, 2020.

This request for continued examination is filed for the purpose of citing additional publications in an IDS and thereby fully complying with Applicant's duty of disclosure.

STATEMENT UNDER 37 C.F.R. §§1.56 AND 1.2*

Applicant hereby advises the Examiner of the status of a co-pending application in compliance with the Applicant's duty to disclose under 37 C.F.R. §§1.56 and 1.2 (see also MPEP §2001.06(b)) as discussed in *McKesson Info. Soln. Inc., v. Bridge Medical Inc.*, 487 F.3d 897; 82 USPQ2d 1865 (Fed. Cir. 2007).

The Applicant wishes to bring to the Examiner's attention U.S. Patent Application No. 13/940,370, filed July 12, 2013 which issued on February 9, 2016 as U.S. Patent 9,254,338.

The Applicant wishes to bring to the Examiner's attention U.S. Patent Application No. 14/972,560, filed December 17, 2015 which issued on June 6, 2017 as U.S. Patent No. 9,669,069.

The Applicant wishes to bring to the Examiner's attention U.S. Patent Application No. 15/471,506, filed March 28, 2017 which issued on November 20, 2018 as U.S. Patent No. 10,130,681.

The Applicant wishes to bring to the Examiner's attention co-pending U.S. Patent Application No. 16/159,282, filed October 12, 2018 for which a Request for Continued Examination was filed on June 30, 2020.

The Applicant wishes to bring to the Examiner's attention co-pending U.S. Patent Application No. 16/397,267, filed April 29, 2019 for which an Office Action was mailed on May 12, 2020.

These documents are available on PAIR, and thus are not provided with this communication. Please inform the undersigned if there is any difficulty in obtaining the documents

from PAIR.

*This Statement is not an admission that any of the listed patents/applications are relevant to the instant claims.

CONCLUSION

Applicant submits that all the claims are in condition for allowance, which action is requested. If the Examiner finds that a telephone conference would expedite the prosecution of this application, the Examiner is invited to telephone the undersigned at the number provided.

The Commissioner is hereby authorized to charge any underpayment of fees up to a strict limit of \$3,000.00 beyond that authorized on the credit card, but not more than \$3,000.00 in additional fees due with any communication for the above referenced patent application, including but not limited to any necessary fees for extensions of time, or credit any overpayment of any amount to Deposit Account No. 50-0815, order number REGN-008CIPCON3.

Respectfully submitted,
BOZICEVIC, FIELD & FRANCIS LLP

Date: 30 June 2020

By: /Karl Bozicevic, Reg. No. 28,807/
Karl Bozicevic, Reg. No. 28,807

BOZICEVIC, FIELD & FRANCIS LLP
201 Redwood Shores Parkway, Suite 200
Redwood City, CA 94065
Telephone: (650) 327-3400
Direct: (650) 833-7735
Facsimile: (650) 327-3231

Electronically Filed

INFORMATION DISCLOSURE STATEMENT	Attorney Docket No.	REGN-008CIPCON3
	Confirmation No.	3451
	First Named Inventor	George D. Yancopoulos
	Application Number	16/055,847
	Filing Date	August 6, 2018
	Group Art Unit	1647
	Examiner Name	Jon McClelland Lockard
	Title: <i>“Use of a VEGF Antagonist to Treat Angiogenic Eye Disorders”</i>	

Address to:
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

Sir:

Applicants submit herewith documents which may be material to the examination of this application and in respect of which there may be a duty to disclose in accordance with 37 C.F.R. § 1.56. This submission is not intended to constitute an admission that any document referred to therein is "prior art" for this invention unless specifically designated as such. A listing of the documents is shown on enclosed Form PTO/SB/08A and copies of the foreign patents and non-patent literature are also enclosed.

The Examiner is requested to make the documents listed on the enclosed PTO/SB/08A of record in this application. Applicants would appreciate the Examiner initialing and returning the initialed copy of form PTO/SB/08A, indicating the documents cited therein have been considered and made of record herein.

Statements

No statement

PTA Statement under 37 CFR § 1.704(d)(1): Each item of information contained in the information disclosure statement filed herewith:

(i) Was first cited in any communication from a patent office in a counterpart foreign or international application or from the Office, and this communication was not received by any individual designated in § 1.56(c) more than thirty days prior to the filing of the information disclosure statement; or

(ii) Is a communication that was issued by a patent office in a counterpart foreign or international application or by the Office, and this communication was not received by any individual designated in § 1.56(c) more than thirty days prior to the filing of the information disclosure statement.

IDS Statement under 37 CFR § 1.97(e)(1): Each item of information contained in the information disclosure statement was first cited in any communication from a foreign

patent office in a counterpart foreign application not more than three months prior to the filing of the information disclosure statement; or

- IDS Statement under 37 CFR § 1.97(e)(2):** No item of information contained in the information disclosure statement was cited in a communication from a foreign patent office in a counterpart foreign application, and, to the knowledge of the person signing the certification after making reasonable inquiry, no item of information contained in the information disclosure statement was known to any individual designated in § 1.56(c) more than three months prior to the filing of the information disclosure statement.

Fees

- No fee is believed to be due.
- The appropriate fee set forth in 37 C.F.R. §1.17(p) accompanies this information disclosure statement.

The Commissioner is hereby authorized to charge any underpayment of fees up to a strict limit of \$3,000.00 beyond that authorized on the credit card, but not more than \$3,000.00 in additional fees due with any communication for the above referenced patent application, including but not limited to any necessary fees for extensions of time, or credit any overpayment of any amount to Deposit Account No. 50-0815, order number REGN-008CIPCON3.

Respectfully submitted,
BOZICEVIC, FIELD & FRANCIS LLP

Date: 30 June 2020

By: /Karl Bozicevic, Reg. No. 28,807/
Karl Bozicevic
Reg. No. 28,807

BOZICEVIC, FIELD & FRANCIS LLP
201 Redwood Shores Parkway, Suite 200
Redwood City, CA 94065
Telephone: (650) 327-3400
Facsimile: (650) 327-3231

INFORMATION DISCLOSURE STATEMENT BY APPLICANT				Application Number	16/055,847
				Filing Date	August 6, 2018
				First Named Inventor	George D. Yancopoulos
				Art Unit	1647
				Examiner Name	Jon McClelland Lockard
Sheet	1	of	2	Attorney Docket Number	REGN-008CIPCON3

U.S. PATENT DOCUMENTS						
Examiner Initial*	Cite No.	Patent Number		Issue Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear
		Number-Kind Code (if known)				
	1					
	2					

U.S. PATENT APPLICATION PUBLICATIONS						
Examiner Initial*	Cite No.	Publication Number		Publication Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear
		Number-Kind Code (if known)				
	1					
	2					

FOREIGN PATENT DOCUMENTS							
Examiner Initial*	Cite No.	Foreign Document Number		Publication Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear	T
		Country Code-Number-Kind Code (if known)					
	1						
	2						

NON PATENT LITERATURE DOCUMENTS						
Examiner Initials*	Cite No.	Include name of the author (in CAPITAL LETTERS), title of the article (when appropriate), title of the item (book, magazine, journal, serial, symposium, catalog, etc.), date, page(s), volume-issue number(s), publisher, city and/or country where published.				T
	1	Bayer Investor News, "VEGF Trap-Eye: New Data Confirm Successes in the Treatment of Age-related Macular Degeneration" (September 28, 2008)				
	2	Regeneron Press Release "Positive Interim Phase 2 Data Reported For VEGF Trap-Eye In Age-Related Macular Degeneration" (March 27, 2007)				
	3	Regeneron Press Release "VEGF TRAP-Eye Phase 2 Wet AMD Results Reported At Arvo Annual Meeting" (May 9, 2007)				
	4	Regeneron Press Release "Regeneron Reports Second Quarter Financial And Operating Results" (August 1, 2007)				
	5	Regeneron Pharmaceuticals, Inc., "Regeneron and Bayer Healthcare Initiate Phase 3 Global Development Program for VEGF Trap-Eye In Wet Age-Related Macular Degeneration (AMD)" (August 2, 2007)				
	6	Regeneron Press Release "Regeneron Announces Positive Primary Endpoint Results From A Phase 2 Study Of VEGF Trap-Eye In Age-Related Macular Degeneration" (October 1, 2007)				
	7	Regeneron Press Release "Regeneron Reports Fourth Quarter And Full Year 2007 Financial And Operating Results" (February 27, 2008)				
	8	Regeneron Pharmaceuticals, Inc., "Regeneron and Bayer HealthCare Announce Encouraging 32-Week Follow-up Results from a Phase 2 Study of VEGF Trap-Eye in Age-Related Macular Degeneration" (April 28, 2008)				
	9	Regeneron Pharmaceuticals, Inc., "Regeneron and Bayer HealthCare Announce VEGF Trap-Eye Achieved Durable Improvement in Vision over 52 Weeks in a Phase 2 Study in Patients with Age-related Macular Degeneration" (August 19, 2008)				

Examiner Signature		Date Considered	
--------------------	--	-----------------	--

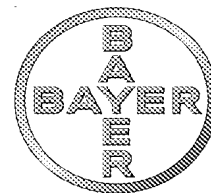
*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

INFORMATION DISCLOSURE STATEMENT BY APPLICANT				Application Number	16/055,847
				Filing Date	August 6, 2018
				First Named Inventor	George D. Yancopoulos
				Art Unit	1647
				Examiner Name	Jon McClelland Lockard
Sheet	2	of	2	Attorney Docket Number	REGN-008CIPCON3

NON PATENT LITERATURE DOCUMENTS			
Examiner Initials*	Cite No.	Include name of the author (in CAPITAL LETTERS), title of the article (when appropriate), title of the item (book, magazine, journal, serial, symposium, catalog, etc.), date, page(s), volume-issue number(s), publisher, city and/or country where published.	T
	10	Regeneron Pharmaceuticals, Inc. "Regeneron Reports Full Year and Fourth Quarter 2008 Financial and Operating Results" (February 26, 2009)	
	11	Regeneron Pharmaceuticals, Inc. "Bayer and Regeneron Extend Development Program for VEGF Trap-Eye to Include Central Retinal Vein Occlusion" (April 30, 2009)	
	12	Regeneron Press Release "First Patient Enrolled In Regeneron And Bayer Healthcare VEGF Trap-Eye Phase 3 Program In Central Retinal Vein Occlusion" (July 23, 2009)	
	13	Regeneron Press Release "Regeneron Schedules November 22, 2010 Teleconference And Webcast To Discuss Results Of Two Phase 3 Studies With VEGF Trap-Eye In Wet Age-Related Macular Degeneration" (November 19, 2010)	
	14	Regeneron Press Release "Regeneron And Bayer Start Phase 3 Trial To Extend Ophthalmology Research & Development Program For VEGF Trap-Eye In Asia" (January 18, 2011)	
	15	Regeneron Press Release "Regeneron To Webcast Investor Briefing On VEGF Trap-Eye Clinical Program On Sunday, February 13th At 9 Am Et" (February 9, 2011)	
	16	Regeneron Press Release "Regeneron Submits Biologics License Application To FDA For VEGF Trap-Eye For Treatment Of Wet Age-Related Macular Degeneration" (February 22, 2011)	
	17	Regeneron Press Release "Regeneron And Bayer Announce Start Of Phase 3 Clinical Program In Diabetic Macular Edema" (April 8, 2011)	
	18	Regeneron Pharmaceuticals, Inc., "FDA Grants Priority Review for VEGF Trap-Eye for the Treatment of Wet Age-Related Macular Degeneration" (April 18, 2011)	
	19	Regeneron Press Release "VEGF Trap-Eye Submitted for EU Marketing Authorization for Treatment of Wet Age-Related Macular Degeneration (June 7, 2011)"	
	20	Regeneron Pharmaceuticals, Inc., "Regeneron Announces EYLEA™ (aflibercept ophthalmic solution) Receives Unanimous Recommendation for Approval for Treatment of Wet AMD from FDA Advisory Committee" (June 17, 2011)	
	21	Regeneron Press Release "Regeneron Announces Clinical Presentations at ASRS 2011 Annual Meeting" (August 17, 2011)	
	22	Regeneron Pharmaceuticals, Inc., "Regeneron Announces FDA Approval of EYLEA™ (aflibercept) Injection for the Treatment of Wet Age-Related Macular Degeneration: CORRECTED (November 18, 2011)	
	23	Regeneron Pharmaceuticals, Inc., "Regeneron and Bayer Initiate Phase 3 Clinical Program for the Treatment of Wet Age-Related Macular Degeneration in China" (November 28, 2011)	
	24	Regeneron Pharmaceuticals, Inc., "Two Year Results of Phase 3 Studies with EYLEA™ (aflibercept) Injection in wet AMD Show Sustained Improvement in Visual Acuity" (December 5, 2011)	

Examiner Signature		Date Considered	
--------------------	--	-----------------	--

*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.



Investor News

Final Phase 2 Results Presented at Retina Society Meeting

VEGF Trap-Eye: New Data Confirm Successes in the Treatment of Age-related Macular Degeneration

Statistically significant reduction in the size of the affected area of the retina demonstrated

Leverkusen, Germany, September 28, 2008 – VEGF Trap-Eye can achieve durable improvements in visual acuity and in biologic measurement parameters in the formation of new blood vessels in the treatment of age-related macular degeneration (AMD). This was shown in the final evaluation of a Phase 2 study presented at the annual meeting of the Retina Society in Scottsdale, Arizona. These parameters include retinal thickness and active choroidal neovascularization lesion size (the damaged part of the retina). Bayer HealthCare and Regeneron Pharmaceuticals, Inc (Nasdaq:REGN) are developing VEGF Trap-Eye together. The treatment successes continued for up to a year.

The study showed that VEGF Trap-Eye was also associated with a reduction in the size of the choroidal neovascular membrane (CNV), the active lesion that is the underlying cause of vision loss in patients with wet AMD. Patients receiving monthly doses of VEGF Trap-Eye of either 2.0 or 0.5 milligrams (mg) for 12 weeks followed by PRN dosing achieved mean improvements in visual acuity versus baseline of 9.0 letters ($p < 0.0001$ versus baseline) and 5.4 letters ($p < 0.085$ versus baseline), respectively. Patients in the 2.0 mg monthly cohort also achieved a statistically significant 1.75 mm² reduction in total lesion size. A reduction in total lesion size was not seen in the cohort initially dosed with 0.5 mg monthly.

"Progression of the active CNV lesion and resulting vision impairment are an inevitable consequence of untreated wet AMD. The reduction in total active CNV lesion size achieved with VEGF Trap-Eye treatment in this Phase 2 clinical study could potentially translate into clinically meaningful outcomes in the larger, controlled Phase 3 studies that

are underway," stated Jason Slakter, M.D., head of the independent reading center for the study and a Clinical Professor of Ophthalmology, New York University School of Medicine, New York.

In this double-masked Phase 2 trial, participants were initially treated with either monthly or quarterly fixed dosing for 12 weeks and then continued to receive treatment for another 40 weeks on a PRN (as needed) dosing schedule. Patients receiving fixed monthly doses of VEGF Trap-Eye of either 2.0 or 0.5 milligrams (mg) for 12 weeks (i.e. 4 fixed doses) followed by PRN dosing achieved mean improvements in visual acuity versus baseline of 9.0 letters ($p < 0.0001$ versus baseline) and 5.4 letters ($p < 0.085$ versus baseline), respectively, at the end of one year. The proportion of patients with vision of 20/40 or better (part of the legal minimum medical requirement for an unrestricted driver's license in the U.S.) increased from 23 percent at baseline to 45 percent at week 52 in patients initially treated with 2.0 mg monthly and from 16 percent at baseline to 47 percent at week 52 in patients initially treated with 0.5 mg monthly. During the week 12 to week 52 PRN dosing period, patients initially dosed on a 2.0 mg monthly schedule received, on average, only 1.6 additional injections and those initially dosed on a 0.5 mg monthly schedule received, on average, 2.5 injections.

Patients receiving monthly doses of VEGF Trap-Eye of either 2.0 or 0.5 mg for 12 weeks followed by PRN dosing also achieved mean decreases in retinal thickness versus baseline of 143 microns ($p < 0.0001$ versus baseline) and 125 microns ($p < 0.0001$ versus baseline) at week 52, respectively.

While PRN dosing following a fixed quarterly dosing regimen (with dosing at baseline and week 12) also yielded improvements in visual acuity and retinal thickness versus baseline at week 52, the results generally were not as robust as those obtained with initial monthly treatment.

"Anti-VEGF therapy has dramatically changed the treatment paradigm for wet AMD, and improvement in visual acuity is now feasible in most patients. The biggest challenge we have is that with our current drugs, the majority of patients need frequent injections into their eye to maintain their visual acuity gains," stated David M. Brown, M.D., a study investigator and a retinal specialist at The Methodist Hospital in Houston. "These study results reinforce our interest in further exploring whether continued administration of

VEGF Trap-Eye on an as-needed basis after an initial period of fixed dosing can maintain a durability of effect over time in controlled Phase 3 clinical studies.”

VEGF Trap-Eye was generally well tolerated and there were no drug-related serious adverse events. There was one reported case of eye inflammation (culture-negative endophthalmitis/uveitis) in the study eye, which was deemed not to be drug-related. The most common adverse events were those typically associated with intravitreal injections.

About the Phase 3 Program in Wet AMD

Regeneron and Bayer HealthCare initiated a Phase 3 global development program for VEGF Trap-Eye in wet AMD in August 2007. In two Phase 3 trials, VIEW 1 and VIEW 2 (VEGF Trap-Eye: Investigation of Efficacy and Safety in Wet age related macular degeneration), the companies are evaluating VEGF Trap-Eye dosed 0.5 mg every 4 weeks, 2 mg every 4 weeks, or 2 mg every 8 weeks (following three monthly doses) in direct comparison with ranibizumab (Lucentis[®], a registered trademark of Genentech, Inc.) administered 0.5 mg every four weeks according to its U.S. label during the first year of the studies. PRN dosing will be evaluated during the second year of each study. The VIEW 1 study (http://www.regeneron.com/vegfrtrap_eye.html) is currently enrolling patients in the United States and Canada and the VIEW 2 study (www.view2study.com) is currently enrolling patients in Europe, Asia Pacific, Japan and Latin America. The companies are collaborating on the global development of VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About VEGF Trap-Eye

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye, which lead to the development of wet AMD. The VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related placental growth factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. Blockade of VEGF, which can prevent abnormal blood vessel formation and vascular leak, has proven beneficial in the treatment of wet AMD.

About Wet AMD

Age-related Macular Degeneration (AMD) is a leading cause of acquired blindness. Macular degeneration is diagnosed as either dry (nonexudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating blind spots in central vision, and it can account for blindness in wet AMD patients. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe.

About Bayer HealthCare

The Bayer Group is a global enterprise with core competencies in the fields of health care, nutrition and high-tech materials. Bayer HealthCare, a subsidiary of Bayer AG, is one of the world's leading, innovative companies in the healthcare and medical products industry and is based in Leverkusen, Germany. The company combines the global activities of the Animal Health, Consumer Care, Diabetes Care and Pharmaceuticals divisions. The pharmaceuticals business operates under the name Bayer Schering Pharma. Bayer HealthCare's aim is to discover and manufacture products that will improve human and animal health worldwide. Find more information at www.bayerhealthcare.com.

Bayer Schering Pharma is a worldwide leading specialty pharmaceutical company. Its research and business activities are focused on the following areas: Diagnostic Imaging, General Medicine, Specialty Medicine and Women's Healthcare. With innovative products, Bayer Schering Pharma aims for leading positions in specialized markets worldwide. Using new ideas, Bayer Schering Pharma aims to make a contribution to medical progress and strives to improve the quality of life. Find more information at www.bayerscheringpharma.de.

Bayer AG, Investor Relations contacts:

Dr. Alexander Rosar (+49-214-30-81013)

Dr. Juergen Beunink (+49-214-30-65742)

Peter Dahlhoff (+49-214-30-33022)

Ilia Kürten (+49-214-30-35426)

Ute Menke (+49-214-30-33021)

Judith Nestmann (+49-214-30-66836)

Dr. Olaf Weber (+49-214-30-33567)

Forward-Looking Statements

This release may contain forward-looking statements based on current assumptions and forecasts made by Bayer Group or subgroup management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in Bayer's public reports which are available on the Bayer website at www.bayer.com. The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

REGENERON

Positive Interim Phase 2 Data Reported for VEGF Trap-Eye in Age-Related Macular Degeneration

March 27, 2007

Positive Interim Phase 2 Data Reported for VEGF Trap-Eye in Age-Related Macular Degeneration TARRYTOWN, N.Y. & LEVERKUSEN, Germany, Mar 27, 2007 (BUSINESS WIRE) -- Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) and Bayer HealthCare AG (NYSE: BAY) today announced positive preliminary data from a pre-planned interim analysis of a Phase 2 randomized study of their VEGF Trap-Eye in patients with the neovascular form of age-related macular degeneration (wet AMD). The VEGF Trap-Eye met its primary endpoint of a statistically significant reduction in retinal thickness after 12 weeks compared with baseline (all groups combined, decrease of 135 microns, $p < 0.0001$). Mean change from baseline in visual acuity, a key secondary endpoint of the study, also demonstrated statistically significant improvement (all groups combined, increase of 5.9 letters, $p < 0.0001$). Moreover, patients in the dose groups that received only a single dose, on average, demonstrated a decrease in excess retinal thickness ($p < 0.0001$) and an increase in visual acuity ($p = 0.012$) at 12 weeks. There were no drug-related serious adverse events, and treatment with the VEGF Trap-Eye was generally well-tolerated. The most common adverse events were those typically associated with intravitreal injections. Detailed data from this interim analysis will be presented at an upcoming scientific conference.

"These data support our efforts to develop the VEGF Trap as a potent blocker of VEGF in various diseases," said George D. Yancopoulos, M.D., Ph.D., President of Regeneron Research Laboratories. "Importantly, the VEGF Trap-Eye may offer the potential to improve vision in patients with wet AMD with dosing less frequently than every four weeks. Our Phase 3 program is being designed to test this possibility and further evaluate the safety and efficacy of various doses and dosing intervals of the VEGF Trap-Eye."

"We are very pleased with the outcome of this interim analysis and the findings support the potential of the VEGF Trap-Eye to improve the lives of patients suffering from wet AMD, which accounts for 90% of AMD related blindness," said Kemal Malik, M.D., member of the Bayer HealthCare Executive Committee, responsible for Global Development. "These results encourage us in our plans to foster next steps in development and to further study the VEGF Trap-Eye in additional eye diseases."

Based on these results, Regeneron and Bayer HealthCare AG plan to initiate the VEGF Trap-Eye Phase 3 program in the second half of 2007. The companies are collaborating on the global development of the VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare AG and Regeneron will jointly commercialize the VEGF Trap-Eye outside the United States, and Regeneron maintains exclusive rights in the United States.

The Phase 2 study is a 12-week, multi-center trial involving 150 patients who are randomized to 5 groups and treated with the VEGF Trap-Eye in one eye. Two groups received either 0.5 or 2.0 mg of VEGF Trap-Eye administered every four weeks, and three groups received a single dose of 0.5, 2.0, or 4.0 mg of VEGF Trap-Eye. Patients are monitored for safety, retinal thickness, and visual acuity over 12 weeks. Retinal thickness is determined by optical coherence tomography (OCT) scans read at an independent reading center. Visual acuity is defined as the total number of letters read correctly on the Early Diabetic Retinopathy Study (ETDRS) chart. Maintenance of vision is defined as losing fewer than 3 lines (equivalent to 15 letters) on the ETDRS chart.

The interim analysis was conducted on the first 78 patients who completed 12 weeks of study. As summarized above, overall, patients had a statistically significant improvement in retinal thickness and visual acuity. All but one patient maintained or improved vision at 12 weeks. Although the improvement in visual acuity was numerically larger in patients receiving injections every 4 weeks, there were no statistically significant differences across the five dose groups in either retinal thickness or visual acuity at 12 weeks.

About the VEGF Trap-Eye

Vascular endothelial growth factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye, which lead to the development of wet AMD. The VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related placental growth factor (PlGF). The VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. Blockade of VEGF, which can prevent abnormal blood vessel formation and vascular leak, has proven beneficial in the treatment of wet AMD.

About AMD

Age-related macular degeneration (AMD) is a leading cause of acquired blindness. Patients with condition can experience a loss of vision due to the development of abnormal, fragile blood vessels in the back of the eye. A particular type of AMD, called wet AMD, accounts for approximately 90% of AMD-related blindness. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe.

Macular degeneration is diagnosed as either dry (nonexudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating blind spots in central vision and can account for blindness in wet AMD patients.

About Regeneron Pharmaceuticals

Regeneron is a biopharmaceutical company that discovers, develops, and intends to commercialize therapeutic medicines for the treatment of serious medical conditions. Regeneron has therapeutic candidates for the potential treatment of cancer, eye diseases, and inflammatory diseases and has

preclinical programs in other diseases and disorders.

About Bayer HealthCare

Bayer HealthCare, a subsidiary of Bayer AG, is one of the world's leading, innovative companies in the healthcare and medical products industry and is based in Leverkusen, Germany. The company combines the global activities of the Animal Health, Consumer Care, Diabetes Care and Pharmaceuticals divisions. The pharmaceuticals business operates under the name Bayer Schering Pharma AG. Bayer HealthCare's aim is to discover and manufacture products that will improve human and animal health worldwide.

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of our drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict our ability to continue to develop or commercialize our drug candidates, competing drugs that are superior to our product candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including our agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2006. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

This news release contains forward-looking statements based on current assumptions and forecasts made by Bayer Group management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in our public reports filed with the Frankfurt Stock Exchange and with the U.S. Securities and Exchange Commission (including our Form 20-F). The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

Additional information about Regeneron and recent news releases are available on Regeneron's worldwide web site at www.regeneron.com

SOURCE: Regeneron Pharmaceuticals, Inc.

Regeneron Pharmaceuticals, Inc.
Charles Poole, 1.914.345.7640
Vice President, Investor Relations
charles.poole@regeneron.com

or

Media Relations:
Lauren Tortorete, 1.212.845.5609
ltortorete@biosector2.com

or

Bayer HealthCare AG
Dr. Jost Reinhard, +49 30 468 15062
Jost.Reinhard@schering.de

REGENERON

VEGF Trap-Eye Phase 2 Wet AMD Results Reported at ARVO Annual Meeting

May 9, 2007

VEGF Trap-Eye Phase 2 Wet AMD Results Reported at ARVO Annual Meeting Average Vision Gain of More Than 10 Letters at 12 Weeks in Highest Monthly Dose Group

Single Dose Results in Average Vision Gain at 12 Weeks

Phase 3 wet AMD Program to Begin in the Third Quarter of 2007

FORT LAUDERDALE, Fla.--(BUSINESS WIRE)--May 9, 2007--Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced positive results from several studies evaluating the VEGF Trap-Eye in the neovascular form of age-related macular degeneration (wet AMD) and diabetic macular edema (DME). These findings were presented this week at the annual meeting of the Association for Research in Vision and Ophthalmology (ARVO). The data reported at the meeting from a pre-planned interim analysis of a Phase 2 randomized study of the VEGF Trap-Eye in patients with wet AMD and a Phase 1 DME trial are available on the Regeneron website (www.regeneron.com on the Events page, under the Investor Relations heading).

"We are very encouraged by the preliminary observation in the Phase 2 wet AMD trial that the most intense dosing regimen studied, 2 milligrams delivered by intravitreal injection every 4 weeks, resulted in an average gain of more than 10 letters after 12 weeks of treatment," said George D. Yancopoulos, M.D., Ph.D., President of Regeneron Research Laboratories. "Perhaps equally important is that after receiving only a single dose of the VEGF Trap-Eye, patients on average had an improvement in the number of letters read both 8 and 12 weeks after treatment. Although significantly more clinical testing is required, the VEGF Trap-Eye may offer the potential to improve vision in patients with wet AMD with a regular dosing regimen that is less frequent than monthly."

Regeneron and Bayer HealthCare AG plan to initiate the VEGF Trap-Eye Phase 3 program in wet AMD in the third quarter of 2007. In the first Phase 3 trial, the companies currently plan to evaluate the VEGF Trap-Eye using 4 and 8 week dosing intervals in direct comparison with ranibizumab (Lucentis[®], a registered trademark of Genentech, Inc.) administered every 4 weeks according to its label. The companies are collaborating on the global development of the VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare AG and Regeneron will jointly commercialize the VEGF Trap-Eye outside the United States, and Regeneron maintains exclusive rights in the United States.

In the Phase 2 wet AMD trial, data were presented from a pre-planned interim analysis of the first 78 patients who completed 12 weeks of the study. The randomized, multi-center trial involves 150 patients who were randomized to 5 groups and treated with the VEGF Trap-Eye in one eye. Two groups received either 0.5 or 2.0 milligrams (mg) of VEGF Trap-Eye administered every 4 weeks, and three groups received a single dose of 0.5, 2.0, or 4.0 mg of VEGF Trap-Eye. Patients were monitored for safety, retinal thickness, and visual acuity over 12 weeks. The VEGF Trap-Eye met its primary endpoint of a statistically significant reduction in retinal thickness after 12 weeks compared with baseline (all groups combined, decrease of 135 microns, p less than 0.0001). Mean change in visual acuity, a key secondary endpoint of the study, also demonstrated a statistically significant improvement (all groups combined, increase of 5.9 letters, p less than 0.0001). There were no drug-related serious adverse events, and treatment with the VEGF Trap-Eye was generally well-tolerated. The most common adverse events were those typically associated with intravitreal injections. Interim data for all dose groups were presented at the ARVO meeting. The Phase 2 wet AMD study is now fully enrolled and results for all patients will be presented at a future scientific meeting.

Encouraging results were also presented from a Phase 1 study of VEGF Trap-Eye in DME. In this open-label safety study, the VEGF Trap-Eye was administered as a single 4.0 mg intravitreal injection to 5 patients with longstanding diabetes and multiple prior treatments for DME. The single injection resulted in a marked decrease in mean central retinal thickness and mean macular volume throughout the 6 week observation period. The VEGF Trap-Eye was generally well tolerated, and there were no drug-related serious adverse events. Adverse events were mostly related to the injection procedure.

About the VEGF Trap-Eye

Vascular endothelial growth factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye, which lead to the development of wet AMD. The VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related placental growth factor (PlGF). The VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. Blockade of VEGF, which can prevent abnormal blood vessel formation and vascular leak, has proven beneficial in the treatment of wet AMD. Blocking VEGF has been shown to be effective in patients with wet AMD; and a VEGF inhibitor, ranibizumab, has been approved for treatment of patients with this condition.

About AMD

Age-related macular degeneration (AMD) is a leading cause of acquired blindness. Patients with this condition can experience a loss of vision due to the development of abnormal, fragile blood vessels in the back of the eye. A particular type of AMD, called wet AMD, accounts for approximately 90% of AMD-related blindness. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe.

Macular degeneration is diagnosed as either dry (nonexudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating blind spots in central vision, and it can account for blindness in wet AMD patients.

About DME

Diabetic Macular Edema (DME) is the most prevalent cause of moderate vision loss in patients with diabetes. DME is a common complication of Diabetic Retinopathy (DR), a disease affecting the blood vessels of the retina. A leading cause of blindness in younger adults (under 50), DME occurs when fluid leaks into the center of the macula, the light-sensitive part of the retina responsible for sharp, direct vision. Fluid in the macula can cause severe vision loss or blindness.

Approximately 500,000 Americans currently suffer from DME, with 75,000 new cases arising each year. According to the American Diabetes Association, more than 18 million Americans currently suffer from diabetes and many other people are at risk for developing diabetes. With the incidence of diabetes steadily climbing, it is projected that up to 10 percent of all patients with diabetes will develop DME during their lifetime.

About Regeneron Pharmaceuticals

Regeneron is a biopharmaceutical company that discovers, develops, and intends to commercialize therapeutic medicines for the treatment of serious medical conditions. Regeneron has therapeutic candidates for the potential treatment of cancer, eye diseases, and inflammatory diseases and has preclinical programs in other diseases and disorders.

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of our drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict our ability to continue to develop or commercialize our drug candidates, competing drugs that are superior to our product candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including our agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-Q for the quarter ended March 31, 2007. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

Additional information about Regeneron and recent news releases are available on Regeneron's worldwide web site at www.regeneron.com

CONTACT: Regeneron Pharmaceuticals, Inc.

Charles Poole
Vice President, Investor Relations
1-914-345-7640
charles.poole@regeneron.com

or

Lauren Tortorete
Media Relations
1-212-845-5609
ltortorete@biosector2.com

SOURCE: Regeneron Pharmaceuticals, Inc.

REGENERON

Regeneron Reports Second Quarter Financial and Operating Results

August 1, 2007

Regeneron Reports Second Quarter Financial and Operating Results TARRYTOWN, N.Y.--(BUSINESS WIRE)--Aug. 1, 2007--Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced financial and operating results for the second quarter of 2007. The Company reported a net loss of \$26.8 million, or \$0.41 per share (basic and diluted), for the second quarter of 2007 compared with a net loss of \$23.6 million, or \$0.41 per share (basic and diluted), for the second quarter of 2006. The Company reported a net loss of \$56.7 million, or \$0.86 per share (basic and diluted), for the six months ended June 30, 2007 compared with a net loss of \$44.0 million, or \$0.77 per share (basic and diluted), for the same period in 2006.

At June 30, 2007, cash, restricted cash, and marketable securities totaled \$512.3 million compared with \$522.9 million at December 31, 2006. In the first quarter of 2007, the Company entered into non-exclusive license agreements with AstraZeneca UK Limited and Astellas Pharma Inc. with respect to the Company's Veloclmmune[®] technology for generating human monoclonal antibody product candidates, as described below. In connection with these agreements, AstraZeneca and Astellas each made an up-front payment to the Company of \$20.0 million in February and April 2007, respectively.

The Company's \$200.0 million of convertible notes, which bear interest at 5.5% per annum, mature in October 2008.

Current Business Highlights

Regeneron is currently focused on three clinical development programs: riloncept (IL-1 Trap) in various inflammatory indications, aflibercept (VEGF Trap) in oncology in collaboration with the sanofi-aventis Group, and the VEGF Trap-Eye in eye diseases in collaboration with Bayer HealthCare AG. The Company also is developing its pipeline of preclinical antibody candidates discovered utilizing its Veloclmmune technology.

Key planned milestones for the third quarter of 2007 include:

- FDA acceptance of the BLA submission for riloncept for CAPS and establishment of target completion date for FDA review of BLA.
- Reporting of results of the Phase 2 trial for the VEGF Trap-Eye in wet AMD.
- Initiation of the Phase 3 program for the VEGF Trap-Eye in wet AMD and receipt of a milestone payment from Bayer HealthCare upon initiation of the Phase 3 program.
- Initiation of the Phase 3 program for the VEGF Trap in oncology in combination with standard chemotherapy regimens.
- Full enrollment of 200 patients in the Phase 2 single-agent VEGF Trap study in advanced ovarian cancer, which was achieved in July.
- Reporting of results of an exploratory proof-of-concept trial of riloncept in gout and initiation of a safety and efficacy trial in gout.
- Completion of preparation for advancing our first human monoclonal antibody product candidate into clinical trials in the fourth quarter.

Riloncept - Inflammatory Diseases

The Company announced in June that it had completed the rolling submission of a Biologics License Application (BLA) to the U.S. Food and Drug Administration (FDA) for riloncept (IL-1 Trap) for the long-term treatment of Cryopyrin-Associated Periodic Syndromes (CAPS). CAPS is a spectrum of rare inherited inflammatory conditions, including Familial Cold Autoinflammatory Syndrome and Muckle-Wells Syndrome.

The FDA has previously granted Orphan Drug status and Fast Track designation to riloncept for the treatment of CAPS. Riloncept has also received Orphan Drug designation in the European Union for the treatment of CAPS.

Regeneron also is evaluating the potential use of rilonacept in other indications in which IL-1 may play a role. The Company is completing an exploratory proof of concept study of rilonacept in ten patients with chronic gout, and a safety and efficacy trial of rilonacept in patients with gout is planned to begin this quarter. The Company also expects to initiate an exploratory proof of concept study of rilonacept in another indication in the fourth quarter.

VEGF Trap - Eye Diseases

The VEGF Trap-Eye is a specially purified and formulated form of the VEGF Trap for use in intraocular applications. Regeneron and Bayer HealthCare plan to initiate the VEGF Trap-Eye Phase 3 program in the neovascular form of age-related macular degeneration (wet AMD) this quarter. The first Phase 3 trial will compare the VEGF Trap-Eye and Genentech, Inc.'s Lucentis[®] (ranibizumab), an anti-angiogenic agent approved for use in wet AMD. This Phase 3 trial will evaluate dosing intervals of four and eight weeks for the VEGF Trap-Eye, compared with ranibizumab dosing according to its label every four weeks. In May 2007, the companies announced positive preliminary results for a pre-planned interim analysis of the Phase 2 trial of the VEGF Trap-Eye in wet AMD. The companies expect to report full results of the Phase 2 trial in the third quarter. Regeneron and Bayer HealthCare plan to initiate a second Phase 3 trial in wet AMD around the end of 2007.

The companies are collaborating on the global development of the VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare and Regeneron will jointly commercialize the VEGF Trap-Eye outside the United States, and Regeneron maintains exclusive rights in the United States. The development program in eye disease is expected to total over \$250 million over the next several years, with the Company and Bayer HealthCare sharing the costs.

VEGF Trap - Oncology

Regeneron and sanofi-aventis are preparing to initiate a large Phase 3 program that will combine the VEGF Trap with standard chemotherapy regimens in five different advanced solid tumors: colorectal, non-small cell lung, prostate, pancreas and gastric cancer. The companies expect the first Phase 3 trial to begin in the current quarter. The development program in oncology is expected to total over \$400 million over the next several years, which will be funded by sanofi-aventis.

In June 2007, at the annual meeting of the American Society of Clinical Oncology (ASCO), Regeneron and sanofi-aventis announced interim results of two Phase 2 single-agent studies of the VEGF Trap in patients with advanced ovarian cancer (AOC) and non-small cell lung adenocarcinoma (NSCLA). The companies are also conducting a Phase 2 trial of the VEGF Trap in AOC patients with symptomatic malignant ascites (SMA).

The AOC study, selected for an oral presentation at ASCO, was an interim analysis of a Phase 2 randomized, double-blind, multi-center trial investigating two doses of the VEGF Trap used as a single agent in patients with recurrent platinum-resistant epithelial ovarian cancer. While the study remains blinded with regards to dose, the combined preliminary results of the two dose levels for 162 of a planned 200 patients demonstrated anti-tumor activity, as evidenced by an 8.0 percent partial response rate and 77 percent achievement of stable disease at 4 weeks in heavily pre-treated patients who had failed multiple other treatments. The VEGF Trap has been well tolerated, and the most common adverse events have been the typical class effect of anti-angiogenic agents. Of the 23 patients in the AOC study with evaluable baseline ascites, 7 patients (30 percent) experienced complete disappearance of the ascites, and 13 patients (57 percent) experienced no increase in ascites during treatment. The AOC study is ongoing and is now fully enrolled.

The second study, presented as a poster at ASCO, is a Phase 2 single-arm study conducted in patients with platinum-resistant and erlotinib-resistant adenocarcinoma of the lung. In this study, the preliminary results demonstrated activity in this heavily pre-treated patient base, as evidenced by a 3.7 percent partial response rate and 63 percent of patients achieving stable disease. The VEGF Trap has been well-tolerated in this trial as well. This study is ongoing and is now fully enrolled.

Sanofi-aventis has indicated that a first registration submission to a regulatory agency for the VEGF Trap is possible as early as 2008.

The companies have also initiated their first trial of the VEGF Trap in Japan, a Phase 1 safety and tolerability study in combination with S-1 in patients with advanced solid malignancies. In addition, currently underway or scheduled to begin are more than 12 studies to be conducted in conjunction with the National Cancer Institute (NCI) Cancer Therapy Evaluation Program (CTEP) evaluating the VEGF Trap as a single agent or in combination with chemotherapy regimens in a variety of cancer indications.

Monoclonal Antibodies

VelocImmune, Regeneron's novel technology for producing fully human monoclonal antibodies, is part of the Company's suite of proprietary, inter-related technology platforms that are designed to provide Regeneron with its next generation of therapeutic candidates. Regeneron plans to move its first new antibody product candidate into clinical trials in the fourth quarter of 2007, with plans to advance at least two antibody product candidates into human clinical trials each year going forward.

In 2007, Regeneron entered into non-exclusive license agreements with AstraZeneca and Astellas that will allow those companies to utilize VelocImmune technology in their internal research programs to discover human monoclonal antibody product candidates. Each of those companies made a \$20.0 million up-front, non-refundable payment and will make up to five additional annual payments of \$20.0 million, subject to the ability to terminate the agreement after making the first three additional payments. Upon commercialization of any antibody products discovered utilizing VelocImmune, the licensees will pay to Regeneron a mid-single-digit royalty on product sales.

Financial Results

Revenue

Regeneron's total revenue increased to \$22.2 million in the second quarter of 2007 from \$19.3 million in the same quarter of 2006 and to \$38.0 million for the first six months of 2007 from \$37.5 million for the same period of 2006. Contract research and development revenue in the first half of 2007 and 2006 principally related to the Company's VEGF Trap collaboration with sanofi-aventis in cancer indications. Contract manufacturing revenue in 2006 related to Regeneron's long-term manufacturing agreement with Merck & Co., Inc., which expired in October 2006. Technology licensing revenue in the first half of 2007 related to the Company's license agreements with AstraZeneca and Astellas.

Regeneron recognized contract research and development revenue of \$13.5 million in the second quarter of 2007 and \$25.3 million for the first six months of 2007 related to the Company's collaboration with sanofi-aventis, compared with \$14.8 million and \$28.7 million, respectively, for the same periods of 2006. Contract research and development revenue from the sanofi-aventis collaboration consisted of reimbursement of VEGF Trap development expenses plus recognition of amounts related to \$105.0 million of previously received and deferred up-front, non-refundable payments. Reimbursement of expenses decreased to \$11.3 million in the second quarter of 2007 from \$11.8 million in the comparable quarter of 2006, and to \$20.8 million in the first six months of 2007 from \$22.6 million in the same period of 2006, principally because costs related to the Company's manufacture of VEGF Trap clinical supplies were lower in 2007. With respect to the up-front payments from sanofi-aventis, \$2.2 million was recognized in the second quarter of 2007 compared to \$3.0 million in the same quarter of 2006, and \$4.5 million was recognized in the first six months of 2007 compared to \$6.1 million in the same period of 2006.

Sanofi-aventis also incurs VEGF Trap development expenses directly and these expenses are increasing because of the growing number of clinical trials sanofi-aventis is overseeing in the VEGF Trap oncology program. During the term of the collaboration, sanofi-aventis pays 100% of agreed-upon VEGF Trap development expenses incurred by both companies. Following commercialization of a VEGF Trap product by the collaboration, Regeneron, from its 50% share of VEGF Trap profits, will reimburse sanofi-aventis for 50% of the VEGF Trap development expenses previously paid by sanofi-aventis.

In connection with the Company's license agreements with AstraZeneca and Astellas, both of the \$20.0 million non-refundable, up-front payments received in February and April 2007, respectively, were deferred and are being recognized as revenue ratably over approximately the first year of each agreement. In the second quarter and for the first six months of 2007, the Company recognized \$6.3 million and \$8.4 million, respectively, of technology licensing revenue related to these agreements.

Bayer HealthCare Collaboration

In October 2006, the Company entered into a collaboration with Bayer HealthCare for the development and commercialization of the VEGF Trap-Eye outside the United States, and received a \$75.0 million up-front, non-refundable payment. In 2007, agreed upon VEGF Trap-Eye development expenses incurred by both companies under a global development plan will be shared as follows: Up to the first \$50.0 million will be shared equally; Regeneron is solely responsible for the next \$40.0 million; over \$90.0 million will be shared equally. Through June 30, 2007, reimbursements from Bayer HealthCare of our VEGF Trap-Eye development expenses totaled \$10.6 million. All payments received or receivable from Bayer HealthCare through June 30, 2007, totaling \$85.6 million, have been fully deferred and included in deferred revenue for financial statement purposes.

Expenses

Total operating expenses for the second quarter of 2007 were \$52.8 million, 21 percent higher than the same period in 2006, and \$102.2 million for the first six months of 2007, 23 percent higher than the same period in 2006. Operating expenses included non-cash compensation expense related to employee stock option awards (Stock Option Expense) of \$6.9 million in the second quarter of 2007 and \$13.5 million for the first six months of 2007, compared with \$4.6 million and \$8.5 million, respectively, for the same periods of 2006. The increase in total Stock Option Expense in 2007 was primarily due to the higher fair market value of the Company's Common Stock on the date of annual employee option grants made by the Company in December 2006 in comparison to the fair market value of the Company's Common Stock on the dates of annual employee option grants made in recent prior years.

Research and development (R&D) expenses increased to \$43.9 million in the second quarter of 2007 from \$34.4 million in the comparable quarter of 2006, and to \$85.1 million in the first six months of 2007 from \$66.5 million in the same period of 2006. In addition to the impact of Stock Option Expense, as described above, in the first half of 2007, the Company incurred higher R&D costs related to additional headcount and additional clinical manufacturing capacity, and higher costs related to preclinical development of new antibody candidates and clinical development of the VEGF Trap-Eye and rilonacept. These were partly offset by lower development expenses for the VEGF Trap cancer program.

General and administrative (G&A) expenses increased to \$8.9 million in the second quarter of 2007 from \$6.3 million in the comparable quarter of 2006, and to \$17.1 million in the first six months of 2007 from \$12.2 million in the same period of 2006. In addition to the impact of Stock Option Expense, as described above, in the first half of 2007, the Company incurred higher G&A costs related to additional headcount and higher fees for various professional services.

About Regeneron Pharmaceuticals

Regeneron is a biopharmaceutical company that discovers, develops, and intends to commercialize therapeutic medicines for the treatment of serious medical conditions. Regeneron has therapeutic candidates in clinical trials for the potential treatment of cancer, eye diseases, and inflammatory diseases, and has preclinical programs in other diseases and disorders.

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of our drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict our ability to continue to develop or commercialize our drug candidates, competing drugs that are superior to our product candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including our agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2006 and Form 10-Q for the quarter ended March 31, 2007. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

REGENERON PHARMACEUTICALS, INC.
CONDENSED BALANCE SHEETS (Unaudited)
(In thousands)

June 30, December 31,
2007 2006

ASSETS

Cash, restricted cash, and marketable securities	\$512,282	\$522,859	
Receivables	20,478	7,493	
Property, plant, and equipment, net	47,647	49,353	
Other assets	17,451	5,385	
	<u> </u>	<u> </u>	
Total assets	\$597,858	\$585,090	
	=====	=====	

LIABILITIES AND STOCKHOLDERS' EQUITY

Accounts payable and accrued expenses	\$34,905	\$21,471
Deferred revenue	183,617	146,995
Notes payable	200,000	200,000
Stockholders' equity	179,336	216,624
	<u> </u>	<u> </u>
Total liabilities and stockholders' equity	\$597,858	\$585,090
	=====	=====

REGENERON PHARMACEUTICALS, INC.
CONDENSED STATEMENTS OF OPERATIONS (Unaudited)
(In thousands, except per share data)

For the three months For the six months
ended June 30, ended June 30,
2007 2006 2007 2006

Revenues

Contract research and development	\$15,917	\$14,991	\$29,562	\$29,578
Contract manufacturing		4,267	7,899	
Technology licensing	6,278		8,421	
	<u> </u>	<u> </u>	<u> </u>	<u> </u>
	22,195	19,258	37,983	37,477
	<u> </u>	<u> </u>	<u> </u>	<u> </u>

Expenses

Research and development	43,864	34,398	85,099	66,482
Contract manufacturing		2,810	4,662	
General and administrative	8,935	6,299	17,137	12,245
	<u> </u>	<u> </u>	<u> </u>	<u> </u>
	52,799	43,507	102,236	83,389
	<u> </u>	<u> </u>	<u> </u>	<u> </u>

Loss from operations (30,604) (24,249) (64,253) (45,912)

Other income (expense)				
Investment income	6,841	3,684	13,584	7,165
Interest expense	(3,011)	(3,011)	(6,022)	(6,022)
	<u>3,830</u>	<u>673</u>	<u>7,562</u>	<u>1,143</u>

Net loss before cumulative effect of a change in accounting principle	(26,774)	(23,576)	(56,691)	(44,769)
Cumulative effect of adopting Statement of Financial Accounting Standards No. 123R ("SFAS 123R")			813	

Net loss	<u>(\$26,774)</u>	<u>(\$23,576)</u>	<u>(\$56,691)</u>	<u>(\$43,956)</u>
----------	-------------------	-------------------	-------------------	-------------------

Net loss per share amounts, basic and diluted:				
Net loss before cumulative effect of a change in accounting principle	(\$0.41)	(\$0.41)	(\$0.86)	(\$0.79)
Cumulative effect of adopting SFAS 123R			0.02	

Net loss	<u>(\$0.41)</u>	<u>(\$0.41)</u>	<u>(\$0.86)</u>	<u>(\$0.77)</u>
----------	-----------------	-----------------	-----------------	-----------------

Weighted average shares outstanding, basic and diluted	65,950	56,915	65,757	56,821
--	--------	--------	--------	--------

CONTACT: Regeneron Pharmaceuticals, Inc.

Investors:

Charles Poole, 914-345-7640
charles.poole@regeneron.com

or

Media:

Lauren Tortorete, 212-845-5609
ltortorete@biosector2.com

SOURCE: Regeneron Pharmaceuticals, Inc.

REGENERON

Regeneron Announces Positive Primary Endpoint Results from a Phase 2 Study of VEGF Trap-Eye in Age-related Macular Degeneration

October 1, 2007

Regeneron Announces Positive Primary Endpoint Results from a Phase 2 Study of VEGF Trap-Eye in Age-related Macular Degeneration TARRYTOWN, N.Y.--(BUSINESS WIRE)--Oct. 1, 2007--Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) and development partner, Bayer HealthCare AG (NYSE:BAY) of Leverkusen, Germany, today announced positive results from the full analysis of the primary 12-week endpoint of a Phase 2 study evaluating the VEGF Trap-Eye in the neovascular form of age-related macular degeneration (wet AMD). The VEGF Trap-Eye met the primary study endpoint of a statistically significant reduction in retinal thickness, a measure of disease activity, after 12 weeks of treatment compared with baseline (all five dose groups combined, mean decrease of 119 microns, $p < 0.0001$). The mean change from baseline in visual acuity, a key secondary endpoint of the study, also demonstrated statistically significant improvement (all groups combined, increase of 5.7 letters, $p < 0.0001$). Preliminary analyses at 16 weeks showed that the VEGF Trap-Eye, dosed monthly, achieved a mean gain in visual acuity of 9.3 to 10 letters (for the 0.5 and 2 mg dose groups, respectively). In additional exploratory analyses, the VEGF Trap-Eye, dosed monthly, reduced the proportion of patients with vision of 20/200 or worse (a generally accepted definition for legal blindness) from 14.3 percent at baseline to 1.6 percent at week 16; the proportion of patients with vision of 20/40 or better (part of the legal minimum requirement for an unrestricted driver's license in the U.S.) was likewise increased from 19.0 percent at baseline to 49.2 percent at 16 weeks. These findings were presented at the Retina Society Conference in Boston, MA. The data reported at the meeting are available on the Regeneron website (www.regeneron.com on the Events Page, under the Investor Relations heading).

In this double-masked, prospective, randomized, multi-center Phase 2 trial, 157 patients were randomized to five groups and treated with the VEGF Trap-Eye in one eye. Two groups received monthly doses of 0.5 or 2.0 milligrams (mg) of VEGF Trap-Eye and three groups received quarterly doses of 0.5, 2.0, or 4.0 mg of VEGF Trap-Eye (at baseline and week 12). Patients were monitored for safety, retinal thickness, and visual acuity. All five dose groups showed an improvement in retinal thickness and an increase in mean letters read versus baseline at all time points through week 12. There were no drug-related ocular or systemic serious adverse events (SAE) reported. Treatment with the VEGF Trap-Eye was generally well tolerated. The most common adverse events were those typically associated with intravitreal injections.

Preliminary week 16 results showed that retinal thickness for all groups combined continued to improve with a mean decrease of 159 microns versus baseline ($p < 0.0001$). The mean change from baseline in visual acuity also continued to improve (all groups combined, increase of 6.6 letters versus baseline, $p < 0.0001$). Patients receiving monthly doses of the VEGF Trap-Eye, either 0.5 or 2 mg, achieved mean decreases in retinal thickness of 160 and 183 microns, respectively, and mean improvements in visual acuity of 9.3 and 10 letters, respectively, at week 16. While quarterly dosing improved retinal thickness and visual acuity versus baseline at 12 and 16 weeks, the effect was not as robust as with monthly dosing. A single 2-mg dose maintained similar effect on visual acuity as 2 mg dosed monthly out to eight weeks (5.8 vs. 6.2 letters gained at 8 weeks, respectively). The table below summarizes preliminary 16-week results for patients in each dosing arm of the study.

"We are particularly encouraged by the decrease, following monthly treatment, in the proportion of patients with vision at the legally blind level of 20/200 or worse, as well as the proportion of patients whose vision improved to 20/40 or better," said George D. Yancopoulos, M.D., Ph.D., President of Regeneron Research Laboratories. "Our large Phase 3 program will help us determine the full impact of the VEGF Trap-Eye on visual acuity in these patient populations with significant unmet clinical needs."

"These results reaffirm the decision to study both the 0.5 mg and 2 mg monthly doses in the Phase 3 program," stated Jeffrey Heier, M.D., a clinical ophthalmologist at Ophthalmic Consultants of Boston, a primary investigator in the Phase 2 study, and chair of the steering committee for the Phase 3 VIEW 1 trial. "The quarterly dosing arms seemed to sustain their effect on visual acuity out to eight weeks, providing the rationale for exploring an eight-week dosing schedule in the Phase 3 program. Further improvement in visual acuity and dosing convenience continue to represent major unmet medical needs in the treatment of wet AMD."

	0.5 mg q4wk (n=32)	2 mg q4wk (n=31)	0.5 mg q12wk (n=32)	2 mg q12wk (n=31)	4 mg q12wk (n=31)
Retinal thickness (mean decrease in microns) at 16 wks	160	183	135	107	210
Visual acuity (mean letters gained) at 16 wks	9.3	10.0	5.6	4.3	3.9

% of patients who gained 15 or more letters at 16 wks	25%	39%	22%	19%	10%
---	-----	-----	-----	-----	-----

% of patients with 20/40 vision or better:

-At Baseline	16%	23%	22%	10%	16%
-At Week 16	44%	55%	31%	36%	32%

% of patients with 20/200 vision or less:

-At Baseline	19%	10%	9%	7%	19%
-At Week 16	3%	0%	13%	7%	13%

About the Phase 3 Program in Wet AMD

Regeneron and Bayer HealthCare AG initiated a Phase 3 global development program for the VEGF Trap-Eye in wet AMD in August of this year. In the first Phase 3 trial, the companies will evaluate the VEGF Trap-Eye using four- and eight-week dosing intervals in direct comparison with ranibizumab (Lucentis[®], a registered trademark of Genentech, Inc.) administered every four weeks according to its label. The Phase 3 wet AMD study is currently being enrolled. The companies are collaborating on the global development of the VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare will market the VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of the VEGF Trap-Eye. Regeneron maintains exclusive rights in the United States.

About the VEGF Trap-Eye

Vascular endothelial growth factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye, which lead to the development of wet AMD. The VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related placental growth factor (PlGF). The VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. Blockade of VEGF, which can prevent abnormal blood vessel formation and vascular leak, has proven beneficial in the treatment of wet AMD and a VEGF inhibitor, ranibizumab, has been approved for treatment of patients with this condition.

About Wet AMD

Age-related macular degeneration (AMD) is a leading cause of acquired blindness. Macular degeneration is diagnosed as either dry (nonexudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating blind spots in central vision, and it can account for blindness in wet AMD patients. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe.

About Regeneron Pharmaceuticals

Regeneron is a biopharmaceutical company that discovers, develops, and intends to commercialize therapeutic medicines for the treatment of serious medical conditions. Regeneron has therapeutic candidates for the potential treatment of cancer, eye diseases, and inflammatory diseases and has preclinical programs in other diseases and disorders. Additional information about Regeneron and recent news releases are available on Regeneron's worldwide web site at www.regeneron.com

Forward Looking Statement - Regeneron

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of our drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict our ability to continue to develop or commercialize our drug candidates, competing drugs that are superior to our product candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including our agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-Q for the quarter ended June 30, 2007. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

CONTACT: Regeneron Pharmaceuticals, Inc.

Investor Relations:

Charles Poole, 914-345-7640

charles.poole@regeneron.com

or

Corporate Communications:
Laura Lindsay, 914-345-7800
laura.lindsay@regeneron.com

or

Media Relations:
Lauren Tortorete, 212-845-5609
ltortorete@biosector2.com

SOURCE: Regeneron Pharmaceuticals, Inc.

REGENERON

Regeneron Reports Fourth Quarter and Full Year 2007 Financial and Operating Results

February 27, 2008

Regeneron Reports Fourth Quarter and Full Year 2007 Financial and Operating Results TARRYTOWN, N.Y.--(BUSINESS WIRE)--Feb. 27, 2008--Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced financial and operating results for the fourth quarter and full year 2007. The Company reported a net loss of \$13.1 million, or \$0.19 per share (basic and diluted), for the fourth quarter of 2007 compared with a net loss of \$31.0 million, or \$0.51 per share (basic and diluted), for the fourth quarter of 2006. The Company reported a net loss of \$105.6 million, or \$1.59 per share (basic and diluted), for the year ended December 31, 2007 compared with a net loss of \$102.3 million, or \$1.77 per share (basic and diluted), for the same period in 2006. In the fourth quarter of 2007, in connection with the Company's VEGF Trap-Eye collaboration with Bayer HealthCare, the Company recognized a cumulative catch-up of \$35.9 million of contract research and development revenue and \$10.6 million of additional research and development expense, as described below.

At December 31, 2007, cash, restricted cash, and marketable securities totaled \$846.3 million compared with \$522.9 million at December 31, 2006. In November 2007, the Company and the sanofi-aventis Group entered into a global, strategic collaboration to discover, develop, and commercialize fully human monoclonal antibodies and sanofi-aventis made an \$85.0 million up-front payment to Regeneron. In addition, in December 2007, sanofi-aventis purchased 12 million newly issued shares of Regeneron Common Stock at \$26.00 per share for proceeds to the Company of \$312.0 million.

The Company's \$200.0 million of convertible notes, which bear interest at 5.5 percent per annum, mature in October 2008.

Current Business Highlights

Regeneron has three late-stage clinical development programs: ARCALYST (rilonacept; also known as IL-1 Trap) in Cryopyrin-Associated Periodic Syndromes (CAPS), aflibercept (the VEGF Trap) in oncology in collaboration with the sanofi-aventis Group, and the VEGF Trap-Eye in eye diseases in collaboration with Bayer HealthCare. Regeneron has also initiated a Phase 2 trial of ARCALYST for the prevention of gout.

In addition, Regeneron has commenced a Phase 1 trial of its first fully human monoclonal antibody candidate, REGN88, an antibody targeting the interleukin-6 receptor (IL-6R) in rheumatoid arthritis, as part of its antibody collaboration with sanofi-aventis. The Company is developing a pipeline of preclinical antibody candidates utilizing its Veloclmmune[®] technology.

Regeneron achieved the following milestones in the fourth quarter of 2007:

- Entered into a global, strategic collaboration agreement with sanofi-aventis to discover, develop, and commercialize fully human monoclonal antibodies.
- Reported extended safety results from a Phase 3 trial of ARCALYST(TM) in patients with CAPS at the American College of Rheumatology (ACR) Annual Meeting in November 2007.
- Initiated a Phase 2 safety and efficacy trial of ARCALYST(TM) in the prevention of gout flares.
- Reported positive results from the extension phase of the Phase 2 trial of the VEGF Trap-Eye in age-related macular degeneration (wet AMD).
- Initiation by sanofi-aventis of the third and fourth Phase 3 oncology trials for aflibercept in combination with standard chemotherapy regimens.
- Initiated a Phase 1 clinical trial of REGN88 in rheumatoid arthritis.

ARCALYST (rilonacept; also known as IL-1 Trap) - Inflammatory Diseases

The Company announced in November 2007 that the action date for the FDA's priority review of the Biologics License Application (BLA) for ARCALYST for the long-term treatment of CAPS was set for February 29, 2008. CAPS is a group of rare inherited inflammatory conditions, including Familial Cold Auto-inflammatory Syndrome and Muckle-Wells Syndrome. The FDA previously granted Orphan Drug status and Fast Track designation to ARCALYST for the treatment of CAPS. ARCALYST has also received Orphan Drug designation in the European Union for the treatment of CAPS.

In the fourth quarter, Regeneron initiated a Phase 2 safety and efficacy trial of ARCALYST in the prevention of gout flares induced by the initiation of uric acid-lowering drug therapy used to control the disease. The Company had previously reported positive results from an exploratory proof-of-concept study of ARCALYST in ten patients with chronic active gout. In those patients, treatment with ARCALYST demonstrated a statistically significant reduction in patient pain scores in the single-blind, placebo-controlled study. Mean patients' pain scores, the key symptom measure in persistent gout, were reduced 41 percent ($p=0.025$) during the first two weeks of active treatment and reduced 56 percent (less than 0.004) after six weeks of active treatment. In this study, in which safety was the primary endpoint measure, treatment with ARCALYST was generally well-tolerated. Regeneron is evaluating the potential use of ARCALYST in other indications in which interleukin-1 (IL-1) may play a role.

Aflibercept (VEGF Trap) - Oncology

In December 2007, Regeneron and sanofi-aventis announced the initiation of the third and fourth Phase 3 trials in oncology that combine aflibercept with standard chemotherapy regimens. One trial is evaluating aflibercept in combination with folinic acid, 5-FU, and irinotecan in patients with 2nd line metastatic colorectal cancer. The other trial is evaluating aflibercept in combination with gemcitabine in patients with 1st line metastatic pancreatic cancer. In the first two Phase 3 trials initiated by the collaboration, aflibercept is being evaluated in combination with docetaxel/prednisone in patients with 1st line metastatic androgen independent prostate cancer and in combination with docetaxel in patients with 2nd line metastatic non-small cell lung cancer. All four trials are studying the current standard of chemotherapy care for the cancer being studied with or without aflibercept. In addition, currently underway are 10 studies being conducted in conjunction with the National Cancer Institute (NCI) Cancer Therapy Evaluation Program (CTEP) evaluating aflibercept as a single agent or in combination with chemotherapy regimens in a variety of cancer indications.

VEGF Trap-Eye - Eye Diseases

The VEGF Trap-Eye is a specially purified and formulated form of the VEGF Trap for use in intraocular applications. Regeneron and Bayer HealthCare initiated a Phase 3 global development program of the VEGF Trap-Eye in wet AMD in the third quarter of 2007. The first trial, known as VIEW 1 (VEGF Trap: Investigation of Efficacy and Safety in Wet age-related macular degeneration), is comparing the VEGF Trap-Eye and Genentech, Inc.'s Lucentis® (ranibizumab), an anti-angiogenic agent approved for use in wet AMD. The trial is evaluating dosing intervals of four and eight weeks for the VEGF Trap-Eye, compared with ranibizumab dosed every four weeks according to its label. Regeneron and Bayer HealthCare plan to initiate a second Phase 3 trial in wet AMD in the first half of 2008. This second trial will be conducted primarily in the European Union and other parts of the world outside the U.S.

In the fourth quarter of 2007, the companies announced positive results of the Phase 2 trial of the VEGF Trap-Eye in wet AMD. The VEGF Trap-Eye met the primary study endpoint of a statistically significant reduction in retinal thickness, a measure of disease activity, after 12 weeks of treatment compared with baseline (all five dose groups combined, mean decrease of 119 microns, less than 0.0001). In additional exploratory analyses, the VEGF Trap-Eye, dosed monthly, demonstrated improvements in visual acuity. The VEGF Trap-Eye reduced the proportion of patients with vision of 20/200 or worse (a generally accepted definition for legal blindness), from 14.3 percent at baseline to 1.6 percent at week 16. In a separate analysis, the proportion of patients with vision of 20/40 or better (part of the legal minimum requirement for an unrestricted driver's license in the U.S.) was increased from 19.0 percent at baseline to 49.2 percent at 16 weeks.

Regeneron and Bayer HealthCare are collaborating on the global development of the VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare will market the VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of the VEGF Trap-Eye. Regeneron maintains exclusive rights to the VEGF Trap-Eye in the United States.

Monoclonal Antibodies

In the fourth quarter of 2007, Regeneron and sanofi-aventis entered into a global, strategic collaboration agreement to discover, develop, and commercialize fully human monoclonal antibodies. The first therapeutic antibody to enter clinical development under the collaboration, REGN88, is an antibody to the Interleukin-6 receptor (IL-6R), which has started clinical trials in rheumatoid arthritis. The second is expected to be an antibody to Delta-like ligand-4 (DLI4), which is currently slated to start clinical development in mid-2008. Regeneron plans to advance two antibody product candidates into clinical development in 2008 and an additional two to three antibody product candidates each year thereafter beginning in 2009.

The collaboration is governed by a Discovery and Preclinical Development Agreement and a License and Collaboration Agreement. As part of the discovery agreement, sanofi-aventis made an \$85.0 million up-front payment to Regeneron. In addition, sanofi-aventis agreed to fund up to \$475.0 million of research over the next five years to identify and validate potential drug discovery targets and to develop fully human monoclonal antibodies against these targets. Sanofi-aventis has an option to extend the discovery agreement for up to an additional three years.

Sanofi-aventis has the exclusive option under the license agreement to co-develop antibodies arising from Regeneron's discovery efforts. Sanofi-aventis will fund the drug candidate development costs up-front and Regeneron will reimburse sanofi-aventis for half of the development costs from its share of future antibody profits from the collaboration.

For any product successfully developed as part of the collaboration, sanofi-aventis will take the lead in commercialization activities and Regeneron has worldwide co-promotion rights. In the United States, profits and losses from sales of collaboration antibodies will be shared equally. Outside the United States, profits will be split on a pre-determined sliding scale based on aggregate sales of collaboration antibodies with Regeneron's share ranging from 35 percent to 45 percent. Regeneron is responsible for 45 percent of losses outside the United States. In addition, Regeneron is entitled to receive up to a total of \$250.0 million of sales milestone payments when the collaboration antibodies achieve certain aggregate annual ex-U.S. sales levels, starting at \$1.0 billion.

In December 2007, sanofi-aventis also increased its ownership of Regeneron's outstanding Common Stock from approximately 4 percent to approximately 19 percent by purchasing 12 million newly issued shares of Regeneron Common Stock at \$26.00 per share for proceeds to the Company of \$312.0 million.

Earlier in 2007, Regeneron entered into non-exclusive license agreements with AstraZeneca and Astellas that will allow those companies to utilize VelocImmune technology in their internal research programs to discover human monoclonal antibody product candidates. Each of those companies made a \$20.0 million up-front, non-refundable payment and will make up to five additional annual payments of \$20.0 million, subject to the ability to

terminate the agreement after making the first three additional payments. Upon commercialization of any antibody products discovered utilizing VelocImmune, the licensees will pay to Regeneron a mid-single-digit royalty on product sales.

Financial Results

Revenue

Regeneron's total revenue increased to \$64.7 million in the fourth quarter of 2007 from \$10.3 million in the same quarter of 2006 and to \$125.0 million for the full year 2007 from \$63.4 million for the same period of 2006. Contract research and development revenue in the first three quarters of 2007 and the full-year 2006 principally related to the Company's aflibercept collaboration with sanofi-aventis in cancer indications. In the fourth quarter of 2007, the Company also recognized contract research and development revenue from the Company's VEGF Trap-Eye collaboration with Bayer HealthCare and its new collaboration with sanofi-aventis to discover, develop, and commercialize fully human monoclonal antibodies. Contract manufacturing revenue in 2006 related to Regeneron's long-term manufacturing agreement with Merck & Co., Inc., which expired in October 2006. Technology licensing revenue in 2007 related to the Company's license agreements with AstraZeneca and Astellas.

Regeneron recognized contract research and development revenue of \$12.6 million in the fourth quarter of 2007 and \$47.1 million for the full year 2007 related to the Company's aflibercept collaboration with sanofi-aventis, compared with \$9.1 million and \$47.8 million, respectively, for the same periods of 2006. Contract research and development revenue from the collaboration consisted of reimbursement of aflibercept development expenses incurred by the Company plus recognition of amounts related to \$105.0 million of previously received and deferred non-refundable, up-front payments. Reimbursement of expenses increased to \$10.5 million in the fourth quarter of 2007 and to \$38.3 million for the full year 2007 from \$6.8 million and \$36.4 million, respectively, in the comparable periods of 2006, principally due to higher preclinical and clinical development costs and, in the fourth quarter of 2007, higher costs related to the Company's manufacture of aflibercept clinical supplies. With respect to the \$105.0 million of up-front payments from sanofi-aventis, \$2.1 million was recognized in the fourth quarter of 2007 compared to \$2.2 million in the same quarter of 2006, and \$8.8 million was recognized in the full year 2007 compared to \$11.4 million in the same period of 2006.

Sanofi-aventis also incurs aflibercept development expenses directly and these expenses are increasing because of the growing number of clinical trials sanofi-aventis is overseeing in the aflibercept oncology program. During the term of the aflibercept collaboration, sanofi-aventis pays 100 percent of agreed-upon aflibercept development expenses incurred by both companies. Following commercialization of an aflibercept product, Regeneron, from its 50 percent share of aflibercept profits, will reimburse sanofi-aventis for 50 percent of aflibercept development expenses previously paid by sanofi-aventis.

In connection with the Company's VEGF Trap-Eye collaboration with Bayer HealthCare, the Company received a \$75.0 million non-refundable, up-front payment in October 2006 and a \$20.0 million milestone payment in August 2007. Through September 30, 2007, all payments received from Bayer HealthCare, including the up-front and milestone payments and cost-sharing reimbursements, were fully deferred and included in deferred revenue. In the fourth quarter of 2007, the Company commenced recognizing previously deferred payments from Bayer HealthCare and cost-sharing of the Company's and Bayer HealthCare's 2007 VEGF Trap-Eye development expenses in the Company's Statement of Operations through a cumulative catch-up. The \$75.0 million non-refundable, up-front license payment and \$20.0 million milestone payment are being recognized as contract research and development revenue over the related estimated performance period in accordance with Staff Accounting Bulletin No. 104, Revenue Recognition (SAB 104) and Emerging Issues Task Force 00-21, Accounting for Revenue Arrangements with Multiple Deliverables (EITF 00-21). In periods when the Company recognizes VEGF Trap-Eye development expenses that it incurs under the collaboration, the Company also recognizes, as contract research and development revenue, the portion of those VEGF Trap-Eye development expenses that are reimbursable from Bayer HealthCare. In periods when Bayer HealthCare incurs agreed upon VEGF Trap-Eye development expenses that benefit the collaboration and Regeneron, the Company also recognizes, as additional research and development expense, the portion of Bayer HealthCare's VEGF Trap-Eye development expenses that the Company is obligated to reimburse.

In the fourth quarter of 2007, the Company recorded a cumulative catch-up of \$35.9 million of contract research and development revenue from Bayer HealthCare, consisting of (i) \$15.9 million related to the \$75.0 million up-front licensing payment and the \$20.0 million milestone payment and (ii) \$20.0 million related to the portion of the Company's 2007 VEGF Trap-Eye development expenses that is reimbursable from Bayer HealthCare. In addition, in the fourth quarter of 2007, the Company recorded a cumulative catch-up of \$10.6 million of additional research and development expense related to the portion of Bayer HealthCare's 2007 VEGF Trap-Eye development expenses that the Company was obligated to reimburse.

In connection with the Company's antibody collaboration with sanofi-aventis, the Company recognized \$4.6 million of contract research and development revenue in the fourth quarter of 2007, which consisted of \$3.0 million for reimbursement of the Company's expenses under the collaboration's discovery agreement, \$0.7 million for reimbursement of the Company's REGN88 development expenses, and \$0.9 million related to the \$85.0 million non-refundable, up-front payment, which was deferred upon receipt in December 2007. Contract research and development revenue in connection with the antibody collaboration with sanofi-aventis is being recognized in accordance with SAB 104 and EITF 00-21.

Contract research and development revenue also includes \$1.0 million in the fourth quarter of 2007 and \$5.5 million for the full year 2007, compared to \$0.4 million and \$0.5 million, respectively, for the same periods of 2006, in connection with the Company's five-year grant from the National Institutes of Health (NIH), which was awarded to the Company in September 2006 as part of the NIH's Knockout Mouse Project.

In connection with the Company's license agreements with AstraZeneca and Astellas, both of the \$20.0 million non-refundable, up-front payments received in February and April 2007, respectively, were deferred and are being recognized as revenue ratably over approximately the first year of each agreement. In the fourth quarter and for the full year 2007, the Company recognized \$10.0 million and \$28.4 million, respectively, of technology licensing revenue related to these agreements.

Expenses

Total operating expenses for the fourth quarter of 2007 were \$76.3 million, 74 percent higher than the same period in 2006, and \$239.5 million for the full year 2007, 40 percent higher than for the same period of 2006. Operating expenses included non-cash compensation expense related to employee stock option awards (Stock Option Expense) of \$7.5 million in the fourth quarter of 2007 and \$28.0 million for the full year 2007, compared

with \$5.1 million and \$18.4 million, respectively, for the same periods of 2006. The increase in total Stock Option Expense in 2007 was primarily due to the higher fair market value of the Company's Common Stock on the date of annual employee option grants made by the Company in December 2006 in comparison to the fair market value of the Company's Common Stock on the dates of annual employee option grants made in recent prior years.

Research and development (R&D) expenses increased to \$64.8 million in the fourth quarter of 2007 from \$35.8 million in the comparable quarter of 2006, and to \$201.6 million for the full year 2007 from \$137.1 million for the same period of 2006. In addition to the impact of Stock Option Expense, as described above, in 2007, the Company incurred higher R&D costs primarily related to additional R&D headcount, clinical development costs for the VEGF Trap-Eye and ARCALYST, research and preclinical development costs for new antibody candidates, and costs to manufacture clinical supplies of ARCALYST and REGN88. Also, as described above, in the fourth quarter of 2007, the Company recorded a cumulative catch-up of \$10.6 million of additional research and development expense related to the Company's VEGF Trap-Eye collaboration with Bayer HealthCare.

General and administrative (G&A) expenses increased to \$11.4 million in the fourth quarter of 2007 from \$7.6 million in the comparable quarter of 2006, and to \$37.9 million in the full year 2007 from \$25.9 million in the same period of 2006. In addition to the impact of Stock Option Expense, as described above, in 2007, the Company incurred higher compensation expense due, in part, to additional headcount, higher recruitment and related costs associated with expanding the Company's headcount in 2007, and higher fees for various professional services.

Other Income

Investment income decreased to \$1.5 million in the fourth quarter of 2007 from \$5.5 million in the comparable quarter of 2006, and increased to \$20.9 million for the full year 2007 from \$16.5 million for the same period of 2006. In the fourth quarter and for the full year 2007, the Company recognized \$5.1 million and \$5.9 million, respectively, in charges related to certain marketable securities that were determined to be other-than-temporarily impaired in value. For the full-year 2007, the increase in investment income resulted primarily from higher balances of cash and marketable securities due, in part, to the up-front payment received from Bayer HealthCare in October 2006 and the receipt of \$174.6 million in net proceeds from the November 2006 public offering of 7.6 million shares of the Company's Common Stock, partly offset by the impairment charges previously described.

About Regeneron Pharmaceuticals

Regeneron is a biopharmaceutical company that discovers, develops, and intends to commercialize therapeutic medicines for the treatment of serious medical conditions. Regeneron has therapeutic candidates in clinical trials for the potential treatment of cancer, eye diseases, and inflammatory diseases, and has preclinical programs in other diseases and disorders.

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of our drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict our ability to continue to develop or commercialize our drug candidates, competing drugs that are superior to our product candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including our agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2006 and Form 10-Q for the quarter ended September 30, 2007. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

REGENERON PHARMACEUTICALS, INC.
CONDENSED BALANCE SHEETS (Unaudited)
(In thousands)

December 31, December 31,
2007 2006

ASSETS

Cash, restricted cash, and marketable securities	\$846,279	\$522,859	
Receivables	18,320	7,493	
Property, plant, and equipment, net	58,304	49,353	
Other assets	13,355	5,385	

Total assets	\$936,258	\$585,090	
--------------	-----------	-----------	--

LIABILITIES AND STOCKHOLDERS' EQUITY

Accounts payable and accrued expenses	\$39,232	\$21,471	
Deferred revenue	236,759	146,995	
Notes payable	200,000	200,000	
Stockholders' equity	460,267	216,624	

Total liabilities and

stockholders' equity \$936,258 \$585,090
=====

REGENERON PHARMACEUTICALS, INC.
CONDENSED STATEMENTS OF OPERATIONS (Unaudited)
(In thousands, except per share data)

For the three months For the year
ended December 31, ended December 31,
2007 2006 2007 2006

Revenues

Contract research and
development \$54,730 \$10,110 \$96,603 \$51,136
Contract manufacturing 236 12,311
Technology licensing 10,000 28,421

64,730 10,346 125,024 63,447

Expenses

Research and development 64,825 35,774 201,613 137,064
Contract manufacturing 430 8,146
General and administrative 11,439 7,628 37,865 25,892

76,264 43,832 239,478 171,102

Loss from operations (11,534) (33,486) (114,454) (107,655)

Other income (expense)

Investment income 1,473 5,525 20,897 16,548
Interest expense (3,010) (3,010) (12,043) (12,043)

(1,537) 2,515 8,854 4,505

Net loss before cumulative

effect of a change in
accounting principle (13,071) (30,971) (105,600) (103,150)

Cumulative effect of
adopting Statement of
Financial

Accounting Standards No.
123R ("SFAS 123R") 813

Net loss \$(13,071) \$(30,971) \$(105,600) \$(102,337)
=====

Net loss per share amounts,
basic and diluted:

Net loss before cumulative
effect of a change in
accounting principle \$(0.19) \$(0.51) \$(1.59) \$(1.78)
Cumulative effect of
adopting SFAS 123R 0.01

Net loss \$(0.19) \$(0.51) \$(1.59) \$(1.77)
=====

Weighted average shares

outstanding, basic and
diluted 67,754 61,229 66,334 57,970

CONTACT: Investor Relations

Charles Poole, 914-345-7640
charles.poole@regeneron.com

OR

Media Relations

Laura Lindsay, 914-345-7800
laura.lindsay@regeneron.com

OR

Kimberly Chen, 212-845-5634
kchen@biosector2.com

SOURCE: Regeneron Pharmaceuticals, Inc.

REGENERON

Regeneron and Bayer HealthCare Announce Encouraging 32-Week Follow-Up Results from a Phase 2 Study of VEGF Trap-Eye in Age-Related Macular Degeneration

April 28, 2008

Regeneron and Bayer HealthCare Announce Encouraging 32-Week Follow-Up Results from a Phase 2 Study of VEGF Trap-Eye in Age-Related Macular Degeneration TARRYTOWN, N.Y. & LEVERKUSEN, Germany--(BUSINESS WIRE)--April 28, 2008--Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) and Bayer HealthCare AG today announced that VEGF Trap-Eye dosed on a PRN (as-needed) dosing schedule maintained the statistically significant gain in visual acuity achieved after an initial, 12-week, fixed-dosing phase of a Phase 2 study in the neovascular form of Age-related Macular Degeneration (wet AMD). A full analysis of the 32-week results of the Phase 2 study will be presented today at the 2008 Association for Research in Vision and Ophthalmology (ARVO) meeting in Fort Lauderdale, Florida. The data being reported at the meeting are available on the Regeneron website (www.regeneron.com on the Investor Relations page, under the Presentations heading).

Study results showed that across all dose groups in the study population, the 6.6 mean letter gain in visual acuity achieved versus baseline at the week 16 evaluation visit, following 12 weeks of fixed dosing, was maintained out to week 32 (a 6.7 mean letter gain versus baseline; p less than 0.0001) using a PRN dosing schedule (where dosing frequency was determined by the physician's assessment of pre-specified criteria). The decrease in retinal thickness, an anatomical measure of treatment effect achieved with a fixed-dose schedule was also maintained for all dose groups combined at week 32 (a 137 micron mean decrease versus baseline, p less than 0.0001).

In this double-masked, prospective, randomized, multi-center Phase 2 trial, 157 patients were randomized to five dose groups and treated with VEGF Trap-Eye in one eye. Two groups initially received monthly doses of 0.5 or 2.0 milligrams (mg) of VEGF Trap-Eye for 12 weeks and three groups received quarterly doses of 0.5, 2.0, or 4.0 mg of VEGF Trap-Eye (at baseline and week 12). Following the initial 12-week fixed-dose phase of the trial, patients continued to receive therapy at the same dose on a PRN dosing schedule based upon the physician assessment of the need for re-treatment in accordance with pre-specified criteria. Patients were monitored for safety, retinal thickness, and visual acuity. These data represent the week 32 analysis from the 52-week study, which is continuing to follow patients.

Patients receiving monthly doses of VEGF Trap-Eye, either 0.5 or 2.0 mg, for 12 weeks followed by PRN dosing thereafter achieved mean improvements in visual acuity of 8.0 (p less than 0.01 versus baseline) and 10.1 letters (p less than 0.0001 versus baseline), respectively, and mean decreases in retinal thickness of 141 (p less than 0.0001 versus baseline) and 162 microns (p less than 0.0001 versus baseline) at week 32, respectively. While PRN dosing also maintained the improvements in retinal thickness and visual acuity achieved versus baseline following a fixed dosing regimen utilizing quarterly dosing at baseline and week 12, the results achieved with a quarterly fixed dosing regimen were generally not as robust as obtained with initial fixed monthly dosing.

VEGF Trap-Eye was generally safe and well tolerated and there were no drug-related serious adverse events. There was one reported case of culture-negative endophthalmitis/uveitis in the study eye, which was deemed not to be drug-related. The most common adverse events were those typically associated with intravitreal injections.

After the last fixed-dose administration at week 12, patients from all dose groups combined required, on average, only one additional injection over the following 20 weeks to maintain the visual acuity gain established during the fixed-dosing period. Notably, 55 percent of the patients who received 2.0 mg monthly for 12 weeks did not require any additional treatment throughout the next 20-week PRN dosing period. Moreover, 97 percent of the patients who received 2.0 mg monthly for 12 weeks did not require re-dosing at the week 16 evaluation visit, indicating that an 8-week dosing schedule may be feasible.

"Due to its high affinity for all isoforms of VEGF-A and PIGF, potent mediators of blood vessel overgrowth in wet AMD, as well as its long residence time in the eye, it is anticipated that VEGF Trap-Eye may be able to be dosed at a frequency less than once monthly, especially on a chronic basis, without compromising visual acuity," stated Quan Dong Nguyen, M.D., M.Sc.,* Assistant Professor of Ophthalmology, Wilmer Ophthalmological Institute, the Johns Hopkins University School of Medicine, Baltimore, MD and a primary investigator in the Phase 2 study. "These emerging Phase 2 clinical data seem to support the concept of durability of VEGF Trap-Eye."

In this study, treatment with VEGF Trap-Eye was associated with a reduction in the size of the choroidal neovascular membrane (CNV), the lesion that is the underlying cause of vision loss due to wet AMD. Patients initially treated with a 0.5 mg or 2.0 mg monthly fixed dose for 12 weeks, followed by PRN dosing thereafter, experienced 1.55 mm² and 2.52 mm² reductions in mean CNV size at 24 weeks (the most recently available analysis from the independent reading center) versus baseline, respectively. Patients treated initially with fixed quarterly dosing also experienced an overall reduction in CNV size.

"Regression in CNV size is generally not seen when treating wet AMD patients. The reduction in CNV size achieved thus far with VEGF Trap-Eye treatment highlights the potential clinical utility of this investigational treatment in patients suffering from this devastating condition," stated Jason Slakter, M.D., Clinical Professor of Ophthalmology, New York University School of Medicine, New York.

"These study results further increase our confidence in the design of our Phase 3 clinical program for VEGF Trap-Eye in wet AMD," said George D. Yancopoulos, M.D., Ph.D., President of Regeneron Research Laboratories. "These studies are evaluating the clinical efficacy and safety of VEGF Trap-Eye, using a monthly loading dose of 0.5 mg or 2.0 mg for 12 weeks, followed by a nine-month fixed-dosing regimen of 0.5 mg monthly, 2.0 mg monthly, or 2.0 mg every eight weeks. In the second year of the studies, all patients will be dosed on a PRN basis."

About the Phase 3 Program in Wet AMD

Regeneron and Bayer HealthCare initiated a Phase 3 global development program for VEGF Trap-Eye in wet AMD in August 2007. In two Phase 3 trials, the companies are evaluating VEGF Trap-Eye using four- and eight-week dosing intervals in direct comparison with ranibizumab (Lucentis[®], a registered trademark of Genentech, Inc.) administered every four weeks according to its label during the first year of the studies. PRN dosing will be evaluated during the second year of each study. The VIEW1 study is currently enrolling patients in the United States and Canada. The VIEW2 study has recently been initiated and will enroll patients in up to 200 centers in Europe, Asia Pacific, Japan, and Latin America. The companies are collaborating on the global development of VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About VEGF Trap-Eye

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye, which lead to the development of wet AMD. The VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. Blockade of VEGF, which can prevent abnormal blood vessel formation and vascular leak, has proven beneficial in the treatment of wet AMD and a VEGF inhibitor, ranibizumab, has been approved for treatment of patients with this condition.

About Wet AMD

Age-related Macular Degeneration (AMD) is a leading cause of acquired blindness. Macular degeneration is diagnosed as either dry (nonexudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating blind spots in central vision, and it can account for blindness in wet AMD patients. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe.

About Regeneron Pharmaceuticals, Inc.

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in clinical trials for the potential treatment of cancer, eye diseases, and inflammatory diseases, and has preclinical programs in other diseases and disorders. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Forward Looking Statement

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2007. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

* The assessment made by Dr. Nguyen does not necessarily imply endorsement by the Johns Hopkins University, the Johns Hopkins Hospital, or the Johns Hopkins Medical Institutions.

CONTACT: Regeneron Pharmaceuticals, Inc.

Investor Relations, 914-345-7640

invest@regeneron.com

or

Corporate Communications

Laura Lindsay, 914-345-7800

laura.lindsay@regeneron.com

or

Media Relations

Kimberly Chen, 212-845-5634

kchen@biosector2.com

SOURCE: Regeneron Pharmaceuticals, Inc.

REGENERON

Regeneron and Bayer HealthCare Announce VEGF Trap-Eye Achieved Durable Improvement in Vision over 52 Weeks in a Phase 2 Study in Patients with Age-related Macular Degeneration

August 19, 2008

Regeneron and Bayer HealthCare Announce VEGF Trap-Eye Achieved Durable Improvement in Vision over 52 Weeks in a Phase 2 Study in Patients with Age-related Macular Degeneration **Tarrytown, NY and Leverkusen, Germany (August 19, 2008)** – Regeneron Pharmaceuticals, Inc. (Nasdaq: **REGN**) and Bayer HealthCare AG today announced that patients with wet age-related macular degeneration (AMD) receiving VEGF Trap-Eye in a Phase 2 extension study on a PRN (as needed) dosing schedule continued to show highly significant improvements at 52 weeks in the primary and key secondary endpoints of retinal thickness (an anatomic measure of treatment effect) and vision gain. The 12-week primary endpoint results from the fixed-dosing period of the study were presented at the 2007 Retina Society conference in September 2007. The 32-week results of the Phase 2 study were presented at the 2008 Association for Research in Vision and Ophthalmology (ARVO) meeting in Fort Lauderdale, Florida in April 2008. A full analysis of the 52-week results of the Phase 2 study will be presented at the 2008 meeting of the Retina Society on September 26-28, 2008 in Scottsdale, Arizona.

In this double-masked, prospective, randomized, multi-center Phase 2 trial, 157 patients were randomized to five dose groups and treated with VEGF Trap-Eye in one eye. Two groups initially received monthly doses of 0.5 or 2.0 milligrams (mg) of VEGF Trap-Eye (at weeks 0, 4, 8, and 12) and three groups received quarterly doses of 0.5, 2.0, or 4.0 mg of VEGF Trap-Eye (at baseline and week 12). Following the initial 12-week fixed-dosing phase of the trial, patients continued to receive therapy at the same dose on a PRN dosing schedule based upon the physician assessment of the need for re-treatment in accordance with pre-specified criteria. Patients were monitored for safety, retinal thickness, and visual acuity. These data represent the final one-year analysis from the 52-week study.

Patients receiving four monthly doses of VEGF Trap-Eye, either 2.0 or 0.5 mg, for 12 weeks followed by PRN dosing thereafter, achieved mean improvements in visual acuity versus baseline of 9.0 letters ($p < 0.0001$) and 5.4 letters ($p = 0.085$), respectively, and mean decreases in retinal thickness versus baseline of 143 microns ($p < 0.0001$) and 125 microns ($p < 0.0001$) at week 52, respectively. During the subsequent PRN dosing phase, patients initially dosed on a 2.0 mg monthly schedule received, on average, only 1.6 additional injections and those initially dosed on a 0.5 mg monthly schedule received, on average, 2.5 injections.

For all dose cohorts combined, there was a 5.3 mean letter gain in visual acuity versus baseline at the week 52 evaluation visit ($p < 0.0001$). The mean decrease in retinal thickness for all dose groups combined at week 52 was 130 microns versus baseline ($p < 0.0001$). During the week 12 to week 52 PRN dosing period, patients from all dose groups combined received, on average, only two additional injections.

VEGF Trap-Eye was generally well tolerated and there were no drug-related serious adverse events. There was one reported case of culture-negative endophthalmitis/uveitis in the study eye and one arterial thrombotic event, neither of which was deemed to be drug-related. The most common adverse events were those typically associated with intravitreal injections.

"Based upon retinal physicians' feedback, there remains a significant unmet medical need for a treatment for wet AMD that can reliably improve visual acuity over time without the need for monthly intravitreal injections," said George D. Yancopoulos, M.D., Ph.D., President of Regeneron Research Laboratories. "We are excited about these study findings and the potential for VEGF Trap-Eye to fulfill this need pending the results of our ongoing Phase 3 clinical studies."

"The 52-week results underline that VEGF Trap-Eye has the potential to significantly reduce retinal thickness and improve vision," said Dr. Kemal Malik, member of the Bayer HealthCare Executive Committee responsible for product development. "The further development of this compound is important for millions of people worldwide who suffer from this devastating ocular disease."

About the Phase 3 Program in Wet AMD

Regeneron and Bayer HealthCare initiated a Phase 3 global development program for VEGF Trap-Eye in wet AMD in August 2007. In two Phase 3 trials, VIEW 1 and VIEW 2 (VEGF Trap-Eye: Investigation of Efficacy and Safety in Wet Age-related Macular Degeneration), the companies are evaluating VEGF Trap-Eye dosed 0.5 mg every 4 weeks, 2 mg every 4 weeks, or 2 mg every 8 weeks (following three monthly doses) in direct comparison with ranibizumab (Lucentis®, a registered trademark of Genentech, Inc.) administered 0.5 mg every four weeks according to its U.S. label during the first year of the studies. PRN dosing will be evaluated during the second year of each study. The VIEW1 study (http://www.regeneron.com/vegfrap_eye.html) is currently enrolling patients in the United States and Canada and the VIEW2 study (www.view2study.com) is currently enrolling patients in Europe, Asia Pacific, Japan, and Latin America. The companies are collaborating on the global development of VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About VEGF Trap-Eye

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye, which lead to the development of wet AMD. The VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. Blockade of VEGF, which can prevent abnormal blood vessel formation and vascular leak, has proven beneficial in the treatment of wet AMD.

About Wet AMD

Age-related Macular Degeneration (AMD) is a leading cause of acquired blindness. Macular degeneration is diagnosed as either dry (nonexudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating blind spots in central vision, and it can account for blindness in wet AMD patients. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe.

About Regeneron Pharmaceuticals, Inc.

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in clinical trials for the potential treatment of cancer, eye diseases, and inflammatory diseases, and has preclinical programs in other diseases and disorders. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Forward Looking Statement

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2007 and Form 10-Q for the quarter ending June 30, 2008. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

Contact Information:

Regeneron Pharmaceuticals, Inc.
Investor Relations
914-345-7640
invest@regeneron.com

Laura Lindsay
Corporate Communications
914-345-7800
laura.lindsay@regeneron.com

Lauren Tortorete
Media Relations
212-845-5609
ltortorete@biosector2.com

Bayer HealthCare
Astrid Kranz
+49 30 468-12057
astrid.kranz@bayerhealthcare.com

REGENERON

Regeneron Reports Full Year and Fourth Quarter 2008 Financial and Operating Results

February 26, 2009

Regeneron Reports Full Year and Fourth Quarter 2008 Financial and Operating Results TARRYTOWN, N.Y.--(BUSINESS WIRE)--Feb. 26, 2009-- Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced financial and operating results for the full year and fourth quarter 2008. The Company reported a net loss of \$82.7 million, or \$1.05 per share (basic and diluted), for the year ended December 31, 2008 compared with a net loss of \$105.6 million, or \$1.59 per share (basic and diluted), for the same period in 2007. The Company reported a net loss of \$31.5 million, or \$0.40 per share (basic and diluted), for the fourth quarter of 2008 compared with a net loss of \$13.1 million, or \$0.19 per share (basic and diluted), for the fourth quarter of 2007. In the fourth quarter of 2007, in connection with the Company's VEGF Trap-Eye collaboration with Bayer HealthCare, the Company recognized a cumulative catch-up of revenue and expenses that reduced the net loss for the quarter by \$25.3 million, as described below.

At December 31, 2008, cash, restricted cash, and marketable securities totaled \$527.5 million compared with \$846.3 million at December 31, 2007. During 2008, the Company retired the full \$200 million of its 5.5 percent Convertible Senior Subordinated Notes.

Current Business Highlights

ARCALYST® (rilonacept) – Inflammatory Diseases

The Company shipped \$10.7 million of ARCALYST® (rilonacept) Injection for Subcutaneous Use to its distributors in 2008. In February 2008, the Company received marketing approval from the U.S. Food and Drug Administration (FDA) for ARCALYST for the treatment of Cryopyrin-Associated Periodic Syndromes (CAPS), including Familial Cold Auto-inflammatory Syndrome (FCAS) and Muckle-Wells Syndrome (MWS) in adults and children 12 and older. ARCALYST, an interleukin-1 (IL-1) blocker, is the only therapy approved in the United States for patients with CAPS, a group of rare, inherited, auto-inflammatory conditions characterized by life-long, recurrent symptoms of rash, fever/chills, joint pain, eye redness/pain, and fatigue. Intermittent, disruptive exacerbations or flares can be triggered at any time by exposure to cooling temperatures, stress, exercise, or other unknown stimuli.

In March 2008, ARCALYST became available for prescription in the United States, and the Company transitioned the patients who participated in the CAPS pivotal study from clinical study drug to commercial supplies. The Company currently projects shipments of ARCALYST to its distributors to total approximately \$20-24 million in 2009.

The Company is in the process of initiating a Phase 3 clinical development program with ARCALYST for the treatment of gout. Two Phase 3 clinical trials will evaluate ARCALYST versus placebo for the prevention of gout flares in patients initiating urate-lowering drug therapy. The Company plans to initiate a Phase 3 clinical trial of ARCALYST for acute gout that will evaluate treatment with ARCALYST alone versus ARCALYST in combination with a non-steroidal anti-inflammatory drug (NSAID) versus an NSAID alone. The Phase 3 clinical development program will also include a separate safety study.

Aflibercept (VEGF Trap) – Oncology

Regeneron and collaborator sanofi-aventis are enrolling patients in four Phase 3 trials that combine aflibercept, an anti-angiogenesis agent, with standard chemotherapy regimens for the treatment of cancer. One trial is evaluating aflibercept as a 2nd line treatment for metastatic colorectal cancer (the VELOUR study) in combination with FOLFIRI (folinic acid (leucovorin), 5-fluorouracil, and irinotecan). A second trial is evaluating aflibercept as a 1st line treatment for metastatic pancreatic cancer in combination with gemcitabine (the VANILLA study). A third trial is evaluating aflibercept as a 1st line treatment for metastatic androgen-independent prostate cancer in combination with docetaxel/prednisone (the VENICE study). The fourth trial is evaluating aflibercept as a 2nd line treatment for metastatic non-small cell lung cancer in combination with docetaxel (the VITAL study). All four trials are studying the current standard of chemotherapy care for the cancer being studied with and without aflibercept. Each of the four Phase 3 trials is over one-third enrolled, and initial data from the Phase 3 program is expected in 2010. In addition, a Phase 2 study of aflibercept in 1st line metastatic colorectal cancer in combination with folinic acid (leucovorin), 5-fluorouracil, and oxaliplatin (the AFFIRM study) began recruiting patients in January 2009.

Aflibercept is also being studied in a Phase 2 single-agent study in advanced ovarian cancer (AOC) patients with symptomatic malignant ascites (SMA). This trial is now fully enrolled and we expect to have initial data from this trial by mid 2009.

VEGF Trap-Eye – Ophthalmologic Diseases

VEGF Trap-Eye is a specially purified and formulated form of VEGF Trap for use in intraocular applications. Regeneron and collaborator Bayer HealthCare are testing VEGF Trap-Eye in a Phase 3 program in patients with the neovascular form of age-related macular degeneration (wet AMD). Regeneron and Bayer HealthCare also initiated a Phase 2 study of VEGF Trap-Eye in patients with diabetic macular edema (DME) in late 2008.

The Phase 3 trials in wet AMD, known as VIEW 1 and VIEW 2 (VEGF Trap: Investigation of Efficacy and Safety in Wet age-related macular degeneration), are comparing VEGF Trap-Eye and ranibizumab (Lucentis®, a registered trademark of Genentech, Inc.), an anti-angiogenic agent approved for use in wet AMD. VIEW 1 is being conducted in North America and VIEW 2 is being conducted in Europe, Asia Pacific, Japan, and Latin America. The VIEW 1 and VIEW 2 trials are both evaluating dosing intervals of four and eight weeks for VEGF Trap-Eye compared with ranibizumab

dosed according to its U.S. label every four weeks over the first year. As needed dosing (PRN) with both agents will be evaluated in the second year of the studies. The VIEW 1 and VIEW 2 trials are expected to complete enrollment in 2009, and initial data are expected in late 2010.

The recently initiated Phase 2 DME study, known as the DA VINCI study, is a double-masked, randomized, controlled trial that is evaluating four different VEGF Trap-Eye regimens versus laser treatment. The study will be enrolling approximately 200 patients in the U.S., Canada, European Union, and Australia. The patients in the study will be treated for 52 weeks followed up by six additional months of safety evaluation. The primary efficacy endpoint is the change in best corrected visual acuity (BCVA) from baseline to week 24.

Monoclonal Antibodies

Regeneron and sanofi-aventis are collaborating on the discovery, development, and commercialization of fully human monoclonal antibodies generated by Regeneron using its *VelocImmune*[®] technology. The first therapeutic antibodies to enter clinical development under the collaboration are REGN88, an antibody to the interleukin-6 receptor (IL-6R) that is being evaluated in rheumatoid arthritis, and REGN475, an antibody to Nerve Growth Factor (NGF) that is being developed for the treatment of pain. In addition, a Phase 1 trial is in the process of being initiated for REGN421, an antibody to Delta-like ligand-4 (Dll4) that will be evaluated in oncology in patients with advanced malignancies. Over the course of the next several years, the Company and sanofi-aventis plan to advance an average of two to three new fully human antibodies into clinical development each year.

Financial Results

Revenues

Total revenues decreased to \$55.8 million in the fourth quarter of 2008 from \$64.7 million in the same quarter of 2007 and increased to \$238.5 million for the full year 2008 from \$125.0 million for the same period of 2007. The Company's revenue was comprised of contract research and development revenue, technology licensing revenue, and net product sales.

Contract Research and Development Revenue

Contract research and development revenue relates primarily to the Company's aflibercept and antibody collaborations with sanofi-aventis and the Company's VEGF Trap-Eye collaboration with Bayer HealthCare. Contract research and development revenue for the three months and years ended December 31, 2008 and 2007, consisted of the following:

	Three months ended		Year ended	
	December 31,		December 31,	
	2008	2007	2008	2007
<i>(In millions)</i>				
Contract research & development revenue				
Sanofi-aventis	\$37.6	\$17.2	\$154.0	\$51.7
Bayer HealthCare	3.0	35.9	31.2	35.9
Other	1.7	1.6	7.0	9.0
Total contract research & development revenue	\$42.3	\$54.7	\$192.2	\$96.6

For the three months and years ended December 31, 2008 and 2007, contract research and development revenue from sanofi-aventis consisted of the following:

	Three months ended		Year ended	
	December 31,		December 31,	
	2008	2007	2008	2007
<i>(In millions)</i>				
Aflibercept:				
Regeneron expense reimbursement	\$6.3	\$10.5	\$35.6	\$38.3
Recognition of deferred revenue related to up-front payments	2.5	2.1	8.8	8.8
Total aflibercept	8.8	12.6	44.4	47.1
Antibody:				
Regeneron expense reimbursement	25.5	3.7	97.9	3.7
Recognition of deferred revenue related to up-front payment	2.6	0.9	10.5	0.9
Other	0.7	—	1.2	—
Total antibody	28.8	4.6	109.6	4.6
Total sanofi-aventis contract research & development revenue	\$37.6	\$17.2	\$154.0	\$51.7

Sanofi-aventis' reimbursement of Regeneron's aflibercept expenses decreased for the three months and year ended December 31, 2008, compared to the same periods in 2007, primarily due to lower costs related to manufacturing aflibercept clinical supplies.

Revenue under the antibody collaboration increased for the three months and year ended December 31, 2008 compared to the same periods in 2007 due to the initiation of the collaboration in November 2007.

For the three months and years ended December 31, 2008 and 2007, contract research and development revenue from Bayer HealthCare consisted of the following:

	Three months ended		Year ended	
	December 31,		December 31,	
<i>(In millions)</i>	2008	2007	2008	2007
Cost-sharing of Regeneron VEGF Trap-Eye development expenses	\$0.5	\$20.0	\$18.8	\$20.0
Recognition of deferred revenue related to up-front and milestone payments	2.5	15.9	12.4	15.9
Total Bayer HealthCare contract research & development revenue	\$3.0	\$35.9	\$31.2	\$35.9

In connection with the Company's VEGF Trap-Eye collaboration with Bayer HealthCare, the Company received a \$75.0 million non-refundable, up-front payment in October 2006 and a \$20.0 million milestone payment in August 2007. Through September 30, 2007 all payments received from Bayer HealthCare, including the up-front and milestone payments and cost sharing reimbursements, were fully deferred and included in deferred revenue. In the fourth quarter of 2007, the Company commenced recognizing previously deferred payments from Bayer HealthCare and cost sharing of the Company's VEGF Trap-Eye development expenses in the Company's Statement of Operations through a cumulative catch-up. The \$75.0 million non-refundable, up-front license payment and \$20.0 million milestone payment are being recognized as contract research and development revenue over the related estimated performance period. In periods when the Company recognizes VEGF Trap-Eye development expenses that it incurs under the collaboration, the Company also recognizes, as contract research and development revenue, the portion of those VEGF Trap-Eye development expenses that is reimbursable from Bayer HealthCare. In periods when Bayer HealthCare incurs agreed upon VEGF Trap-Eye development expenses that benefit the collaboration and Regeneron, the Company also recognizes, as additional research and development expense, the portion of Bayer HealthCare's VEGF Trap-Eye development expenses that the Company is obligated to reimburse.

In the fourth quarter of 2007, the Company recorded a cumulative catch-up of \$35.9 million of contract research and development revenue from Bayer HealthCare. In addition, in the fourth quarter of 2007, the Company recorded a cumulative catch-up of \$10.6 million of additional research and development expense related to the portion of Bayer HealthCare's 2007 VEGF Trap-Eye development expenses that the Company was obligated to reimburse.

Under the terms of the Bayer HealthCare collaboration, in 2008, the first \$70.0 million of agreed-upon VEGF Trap-Eye development expenses incurred by the Company and Bayer HealthCare under a global development plan were shared equally, and the Company was solely responsible for up to the next \$30.0 million. During the fourth quarter of 2008, Regeneron was solely responsible for most of the collaboration's VEGF Trap-Eye development expenses. As a result, in the fourth quarter of 2008, the portion of the Company's VEGF Trap-Eye development expenses that were reimbursable from Bayer HealthCare, and recognized as contract research and development revenue, amounted to only \$0.5 million.

Technology Licensing Revenue

Regeneron has entered into non-exclusive license agreements with AstraZeneca and Astellas that allow those companies to utilize *VelocImmune*[®] technology in their internal research programs to discover human monoclonal antibodies. Each company made \$20.0 million annual, non-refundable payments in each of 2007 and 2008 and agreed to make up to four additional annual payments of \$20.0 million, subject to the ability to terminate their agreements after making two such additional payments. Upon receipt, these payments are deferred and are recognized as revenue ratably over approximately the ensuing year of each agreement. Regeneron will also receive a mid-single-digit royalty on sales of any antibodies discovered utilizing *VelocImmune*.

Net Product Sales

The Company shipped \$10.7 million of ARCALYST[®] (rilonacept) to its distributors in 2008 and recorded \$3.5 million and \$6.3 million in product sales revenue for the three months and year ended December 31, 2008. Revenue and deferred revenue from product sales are recorded net of applicable provisions for prompt pay discounts, product returns, estimated rebates payable under governmental programs (including Medicaid), distributor fees, and other sales-related costs. At December 31, 2008, \$4.0 million of ARCALYST net product sales was included in deferred revenue in the Company's financial statements.

Expenses

Total operating expenses for the fourth quarter of 2008 were \$90.4 million, 19 percent higher than the same period in 2007, and \$328.3 million for the full year 2008, 37 percent higher than for the same period of 2007. Average headcount increased to 903 for the fourth quarter of 2008 compared to 665 for the same period in 2007 and increased to 810 for the full year 2008 from 627 for the full year 2007, due primarily to the Company's expanding research and development activities principally in connection with the Company's antibody collaboration with sanofi-aventis.

Operating expenses included non-cash compensation expense related to employee stock option and restricted stock awards of \$7.8 million in the fourth quarter of 2008 and \$32.5 million for the full year of 2008, compared with \$7.5 million and \$28.1 million, respectively, for the same periods of 2007.

Research and development (R&D) expenses increased to \$76.3 million in the fourth quarter of 2008 from \$64.8 million in the comparable quarter of 2007, and to \$278.0 million for the full year 2008 from \$201.6 million for the same period of 2007. In the fourth quarter and for the full year of 2008, the Company incurred higher R&D costs primarily related to additional R&D headcount, clinical development costs for ARCALYST and REGN88, research and preclinical development costs associated with our antibody programs, and facility-related costs to support the Company's expanded R&D activities. In addition, for the full year of 2008, the Company incurred higher R&D costs related to clinical development of VEGF Trap-Eye and manufacturing supplies of our drug product candidates, especially our monoclonal antibodies. Also, as described above, commencing in the fourth quarter of 2007, the Company began recognizing as additional R&D expense, the portion of Bayer HealthCare's VEGF Trap-Eye development expenses that the Company is obligated to reimburse.

Selling, general, and administrative (SG&A) expenses increased to \$13.5 million in the fourth quarter of 2008 from \$11.4 million in the comparable quarter of 2007, and to \$49.4 million for the full year 2008 from \$37.9 million for the full year 2007. In 2008, the Company incurred \$5.2 million of selling expenses related to ARCALYST[®] (rilonacept) for the treatment of CAPS. In addition, the Company incurred higher compensation expense and recruitment costs associated with expanding the Company's SG&A headcount, higher professional fees related to various general corporate matters, and higher SG&A facility related costs.

Other Income and Expense

Investment income increased to \$2.6 million in the fourth quarter of 2008 from \$1.5 million in the comparable quarter of 2007, and decreased to \$18.2 million for the full year 2008 from \$20.9 million for the full year 2007. For the full year 2008, investment income decreased primarily due to lower yields on our cash and marketable securities. The Company recognized charges of \$0.2 million and \$5.1 million for the fourth quarters of 2008 and 2007, respectively, and \$2.5 million and \$5.9 million for the full year 2008 and 2007, respectively, related to certain marketable securities that were determined to be other-than-temporarily impaired. For the full year 2008, these charges were partially offset by realized gains of \$1.2 million on sales of marketable securities during the year.

During the second and third quarters of 2008, the Company repurchased \$82.5 million in principal amount of its 5.5 percent Convertible Senior Subordinated Notes. In connection with the repurchased notes, the Company recognized a \$0.9 million loss on early extinguishment of debt. The remaining \$117.5 million of these notes were repaid in full upon their maturity in October 2008.

Income Tax Expense

In the fourth quarter of 2008, the Company recognized a \$0.7 million income tax benefit, resulting from a provision in the Housing Assistance Tax Act of 2008 that allowed the Company to claim a refund for certain unused pre-2006 research tax credits. For the full year 2008, income tax expense was \$2.4 million and consisted primarily of alternative minimum tax, which resulted from the utilization of certain net operating loss carry-forwards, that would otherwise have expired over the next several years, to offset income for tax purposes.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST[®] (niloncept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in clinical trials for the potential treatment of cancer, eye diseases, inflammatory diseases, and pain, and has preclinical programs in other diseases and disorders. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2008. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

REGENERON PHARMACEUTICALS, INC. CONDENSED BALANCE SHEETS (Unaudited) (In thousands)

	December 31, 2008	December 31, 2007
ASSETS		
Cash, restricted cash, and marketable securities	\$527,461	\$846,279
Receivables	35,212	18,320
Property, plant, and equipment, net	87,853	58,304
Other assets	19,512	13,355
Total assets	\$670,038	\$936,258
LIABILITIES AND STOCKHOLDERS' EQUITY		
Accounts payable, accrued expenses, and other liabilities	\$41,261	\$39,232
Deferred revenue	209,925	236,759
Notes payable		200,000
Stockholders' equity	418,852	460,267
Total liabilities and stockholders' equity	\$670,038	\$936,258

REGENERON PHARMACEUTICALS, INC. CONDENSED STATEMENTS OF OPERATIONS (Unaudited) (In thousands, except per share data)

	For the three months ended December 31,		For the year ended December 31,	
	2008	2007	2008	2007
Revenues				
Contract research and development	\$42,294	\$54,730	\$192,208	\$96,603
Technology licensing	10,000	10,000	40,000	28,421
Net product sales	3,543		6,249	
	55,837	64,730	238,457	125,024
Expenses				
Research and development	76,314	64,825	278,016	201,613
Selling, general, and administrative	13,491	11,439	49,348	37,865
Cost of goods sold	631		923	
	90,436	76,264	328,287	239,478
Loss from operations	(34,599)	(11,534)	(89,830)	(114,454)
Other income (expense)				
Investment income	2,648	1,473	18,161	20,897
Interest expense	(295)	(3,010)	(7,752)	(12,043)
Loss on early extinguishment of debt			(938)	
	2,353	(1,537)	9,471	8,854
Net loss before income tax expense	(32,246)	(13,071)	(80,359)	(105,600)
Income tax expense (benefit)	(728)		2,351	
Net loss	\$ (31,518)	\$ (13,071)	\$ (82,710)	\$ (105,600)
Net loss per share amounts, basic and diluted	\$ (0.40)	\$ (0.19)	\$ (1.05)	\$ (1.59)
Weighted average shares outstanding, basic and diluted	79,190	67,754	78,827	66,334

Source: Regeneron Pharmaceuticals, Inc.

Regeneron Pharmaceuticals, Inc.

Investor Relations:

Peter Dworkin, 914-345-7640

peter.dworkin@regeneron.com

or

Media Relations:

Laura Lindsay, 914-345-7800

laura.lindsay@regeneron.com

or

Kelly Hershkowitz, 212-845-5624

khershkowitz@biobase.com

REGENERON

First Patient Enrolled in Regeneron and Bayer HealthCare VEGF Trap-Eye Phase 3 Program in Central Retinal Vein Occlusion

July 23, 2009

First Patient Enrolled in Regeneron and Bayer HealthCare VEGF Trap-Eye Phase 3 Program in Central Retinal Vein Occlusion TARRYTOWN, N.Y.--(BUSINESS WIRE)--Jul. 23, 2009-- Regeneron Pharmaceuticals, Inc. (NASDAQ:REGN) today announced that the first patient has been enrolled in the Phase 3 program of VEGF Trap-Eye for the treatment of central retinal vein occlusion (CRVO), a leading cause of blindness in adults. Regeneron received a \$20 million milestone payment from Bayer Healthcare that was triggered by the dosing of the first patient in the CRVO program. Regeneron also announced that enrollment in the Phase 2 DA VINCI study of VEGF Trap-Eye in diabetic macular edema (DME) has been completed and data are expected during the first half of 2010.

VEGF Trap-Eye, an investigational drug, is being developed by Regeneron and Bayer HealthCare AG for the potential treatment of eye diseases, including the neovascular form of age-related macular degeneration (wet AMD), DME, and CRVO.

The Phase 3 program in CRVO consists of two multinational, one-year clinical studies. The COPERNICUS (Controlled Phase 3 Evaluation of Repeated Intravitreal administration of VEGF Trap-Eye In Central retinal vein occlusion: Utility and Safety) study is being led by Regeneron and the GALILEO (General Assessment Limiting Infiltration of Exudates in central retinal vein Occlusion with VEGF Trap-Eye) study is being led by Bayer HealthCare. Patients in both studies will receive six monthly intravitreal injections of either VEGF Trap-Eye at a dose of 2 milligrams (mg) or sham control injections. The primary endpoint of both studies is improvement in visual acuity versus baseline after six months of treatment. At the end of the initial six months, patients will be dosed on a PRN (as needed) basis for another six months. All patients will be eligible for rescue laser treatment. Results from both CRVO studies are expected in 2011.

In wet AMD, Regeneron and Bayer Healthcare are evaluating VEGF Trap-Eye in two ongoing Phase 3 studies, known as VIEW 1 and VIEW 2 (VEGF Trap: Investigation of Efficacy and Safety in Wet age-related macular degeneration). Enrollment in these trials is expected to be completed by the end of this year, and data are expected in late 2010.

Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States. Bayer HealthCare has exclusive rights to market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye.

About CRVO

Over 100,000 people in the United States are estimated to suffer from CRVO, a disease for which there is no current treatment that can be considered standard of care. CRVO is caused by obstruction of the central retinal vein that leads to a back up of blood and fluid in the retina, resulting in retinal injury and loss of vision. The retina can also become "ischemic" (starved for oxygen), resulting in the growth of new abnormal blood vessels that can cause further vision loss and more serious complications. Release of VEGF contributes to increased vascular permeability in the eye and abnormal new vessel growth. It is believed that anti-VEGF treatment may help decrease vascular permeability and edema and prevent the growth of abnormal new blood vessels in the retina in patients with CRVO.

About DME

Diabetic Retinopathy (DR) can lead to significant vision impairment and is a major complication of diabetes. Diabetic Macular Edema (DME) is a common complication of DR that involves fluid collection in the macula. DME is the most prevalent cause of moderate visual loss in patients with diabetes.

DME is a leading cause of adult blindness in the developed world. Severe visual loss is caused by a combination of fluid build-up around the retina and the unnatural growth of blood vessels in the back of the eye.

About VEGF Trap-Eye

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye and vascular permeability and edema. VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). Investigational VEGF Trap-Eye is a specific blocker of VEGF-A and PlGF that has been demonstrated in preclinical models to bind these growth factors with greater affinity than their natural receptors.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product in the United States, Regeneron has therapeutic candidates in clinical trials for the potential treatment of cancer, eye diseases, inflammatory diseases, and pain and has preclinical programs in other diseases and disorders. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Forward Looking Statement

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2008 and Form 10-Q for the quarter ended March 31, 2009. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

Source: Regeneron Pharmaceuticals, Inc.

Regeneron Pharmaceuticals, Inc.

Peter Dworkin, 914-345-7640

Investor Relations

peter.dworkin@regeneron.com

or

Laura Lindsay, 914-345-7800

Media Relations

laura.lindsay@regeneron.com

REGENERON

Regeneron Schedules November 22, 2010 Teleconference and Webcast to Discuss Results of Two Phase 3 Studies with VEGF Trap-Eye in Wet Age-related Macular Degeneration

November 19, 2010

TARRYTOWN, N.Y., Nov. 19, 2010 /PRNewswire-FirstCall/ -- Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced that it will hold a teleconference and webcast at 8:30 a.m. Eastern Time on Monday, November 22, to discuss results of two Phase 3 studies with VEGF Trap-Eye in Wet Age-related Macular Degeneration, VIEW 1 and VIEW 2, and its VEGF Trap-Eye program. A press release will be issued on Monday prior to the call.

Teleconference/Webcast Details

To participate in the live call on Monday, November 22, at 8:30 a.m. Eastern Time, please dial (877) 390-5538 for domestic callers and (408) 940-3843 for international callers, participant code 27197068. The live conference call is being webcast and it can be accessed on the "Newsroom" page of the Company's web site, www.regeneron.com. The webcast will be available for 30 days following the call.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration and central retinal vein occlusion), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Contact Information:

Michael Aberman, M.D.	Peter Dworkin
Investor Relations	Corporate Communications
914.345.7799	914.345.7640
michael.aberman@regeneron.com	peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

News Provided by Acquire Media

REGENERON

Regeneron and Bayer Start Phase 3 Trial to Extend Ophthalmology Research & Development Program for VEGF Trap-Eye in Asia

January 18, 2011

TARRYTOWN, N.Y., BERLIN and SINGAPORE, Jan. 18, 2011 /PRNewswire/ – Regeneron (Nasdaq: REGN) and Bayer HealthCare today announced initiation of a new Phase 3 clinical trial in collaboration with the Singapore Eye Research Institute (SERI) investigating the efficacy and safety of VEGF Trap-Eye (afibercept ophthalmic solution) in patients with choroidal neovascularisation (CNV) of the retina as a result of pathologic myopia. The trial has started in Japan and other Asian countries, including China, Korea, Singapore, and Taiwan.

Myopia is one of the most common eye conditions and is highly prevalent in Asian populations, including Singapore where 40% of adults have myopia and nearly 10% have high myopia. Myopic CNV is a complication of high myopia where abnormal blood vessels grow and leak blood and fluid into the retina as a result of degenerative changes in the retinal lining of the eye and is a potentially blinding condition. Currently, there is no well-established treatment for myopic CNV. VEGF Trap-Eye has previously met its primary efficacy endpoint in a Phase 3 trial for neovascular (wet) age-related macular degeneration (AMD).

Collaboration with the Singapore Eye Research Institute (SERI)

SERI has been appointed as the Asian reading center partner for this study. The Singapore Advanced Imaging Laboratory for Ocular Research (SAILOR) will serve as the first reading center for VEGF Trap-Eye studies in the region. SAILOR brings together an inter-disciplinary group of clinician researchers and scientists to collaborate on cutting-edge computer image research. SAILOR is the first clinical translational research unit to be located in Fusionopolis, a research and development complex in Singapore, and serves as a hub of translational research programs in ocular imaging among clinicians, scientists, computer scientists, and other experts. One of the major programs SAILOR has developed is a "tele-ophthalmic ocular imaging platform" to allow transfer and data capture of ocular images for diagnosis and screening. SAILOR will read the images for this myopic CNV trial from the different Asian sites.

"Myopia is a common problem in Singapore and Asia. In particular, myopic CNV, which affects certain groups of people with higher degrees of myopia, may lead to vision loss. There remains uncertainty regarding the best methods of treatment for myopic CNV and this new trial will go towards addressing this clinical need," said Prof. Wong Tien Yin, Director of SERI and Co-Director of SAILOR.

About mCNV

Myopic choroidal neovascularization is a disease of the retina where new, abnormal blood vessels grow into the retina in persons who are severely myopic (typically more than minus six diopters) and have pathological changes in the back of the eye. In myopic patients, the eyeball is too long, which puts strain on the retina and leads to those pathological changes. Anti-VEGF treatment has been shown to be effective in wet age-related macular degeneration, which is also characterised by the growth of new, abnormal blood vessels in the retina. Severe myopia is particularly common in Asia, with some scientists believing that it may be generally more common in Asians than in people from European descent. Myopic CNV (mCNV) is associated with high degrees of myopia and leads to progressive loss of the patient's sight, ending in blindness. In East Asia, the prevalence of myopia is significantly higher than in the West Asia, and seems to have an earlier onset. In Japan, mCNV is the second most common cause of blindness.

About the mCNV Trial

The Phase 3 myopic CNV trial, named MYRROR, will enroll approximately 250 patients and has started in Japan. Other Asian countries, including Singapore, China, Korea, and Taiwan, will join this clinical study throughout the year. Three out of four patients in the trial will receive an injection of VEGF Trap-Eye into the affected eye (and repeated injections on a PRN, as needed, basis, if required). One out of four patients will receive a sham procedure. The clinical outcome of the two treatment groups after 24 weeks will be assessed by a different team of doctors who are unaware of what treatment the patients received. From week 24 onward, sham patients may receive active treatment. The primary outcome measure of the trial is the mean change in vision (best corrected visual acuity) after 24 weeks, compared to baseline. Secondary outcome measures include the percentage of patients who gain or lose certain amounts of letters in the visual test, changes in retinal thickness from baseline, changes in the total mCNV lesion size, and vessel leakage as seen on an angiogram of the affected eye. The study is scheduled to run until June 2013.

About VEGF Trap-Eye

VEGF Trap-Eye is a fully human fusion protein, consisting of soluble VEGF receptors 1 and 2, that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. VEGF Trap-Eye is specially purified and contains iso-osmotic buffer concentrations, allowing for injection into the eye.

Bayer HealthCare and Regeneron are collaborating on the global development of VEGF Trap-Eye for the treatment of the neovascular form of age-related macular degeneration (wet AMD), diabetic macular edema (DME), central retinal vein occlusion (CRVO), and other eye diseases and disorders.

In November 2010, Regeneron and Bayer HealthCare announced positive top-line results from two parallel Phase 3 studies in patients with wet AMD, VIEW 1 and VIEW 2. In these trials, all regimens of VEGF Trap-Eye, including VEGF Trap-Eye dosed every two months, successfully met the primary endpoint compared to the current standard of care, ranibizumab dosed every month. The primary endpoint was statistical non-inferiority in the proportion of patients who maintained (or improved) vision over 52 weeks compared to ranibizumab. A generally favorable safety profile was observed for both VEGF Trap-Eye and ranibizumab. The incidence of ocular treatment emergent adverse events was balanced across all four

treatment groups in both studies. There were no notable differences in non-ocular adverse events among the study arms. Bayer HealthCare and Regeneron are planning to submit regulatory applications for marketing approval for the treatment of wet AMD in Europe and the U.S. in the first half of 2011.

Trials in other indications such as CRVO and DME are currently underway or in preparation.

Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About Singapore Eye Research Institute (SERI)

SERI is the national research institute for ophthalmic and vision research in Singapore. Serving as the research institute of the Singapore National Eye Centre, and affiliated to the Yong Loo Lin School of Medicine, National University of Singapore, as well the Duke-NUS Graduate Medical School, SERI undertakes vision research in collaboration with local clinical ophthalmic centers and biomedical research institutions, as well as major eye centers and research institutes throughout the world. For further information, kindly visit www.seri.com.sg.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration and central retinal vein occlusion), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

About Bayer HealthCare

The Bayer Group is a global enterprise with core competencies in the fields of health care, nutrition and high-tech materials. Bayer HealthCare, a subgroup of Bayer AG with annual sales of EUR 15,988 million (2009), is one of the world's leading, innovative companies in the healthcare and medical products industry and is based in Leverkusen, Germany. The company combines the global activities of the Animal Health, Consumer Care, Medical Care and Pharmaceuticals divisions. Bayer HealthCare's aim is to discover and manufacture products that will improve human and animal health worldwide. Bayer HealthCare has a global workforce of 53,400 employees and is represented in more than 100 countries. Find more information at www.bayerhealthcare.com.

Regeneron Forward Looking Statement

This news release includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties. These include, among others, risks and timing associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group, Bayer HealthCare, and Astellas to be canceled or terminated without any product success, and risks associated with third party intellectual property. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2009 and Form 10-Q for the quarter ended September 30, 2010. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Bayer Forward-Looking Statements

This release may contain forward-looking statements based on current assumptions and forecasts made by Bayer Group or subgroup management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in Bayer's public reports which are available on the Bayer website at www.bayer.com. The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

Your Contact at Bayer:

Doreen Schroeder, Tel. +49 30 468-11399

E-Mail: doreen.schroeder@bayer.com

Your Investor Relations Contact at Regeneron:

Michael Aberman, M.D. Tel. +1 (914) 345-7799

E-Mail: michael.aberman@regeneron.com

Your Media Contact at Regeneron:

Peter Dworkin, Tel. +1 (914) 345-7640

E-Mail: peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals

REGENERON

Regeneron to Webcast Investor Briefing on VEGF Trap-Eye Clinical Program on Sunday, February 13th at 9 am ET

February 9, 2011

TARRYTOWN, N.Y., Feb. 9, 2011 /PRNewswire/ -- Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced that it will webcast an investor briefing on Sunday, February 13 from 9 a.m. to 10:30 a.m. Eastern Time. At the investor briefing, principal investigators from the VEGF Trap-Eye clinical studies will recap presentations from the Bascom Palmer Eye Institute's Angiogenesis, Exudation and Degeneration 2011 meeting being held in Miami, Florida on Saturday, February 12.

The investigator presentations will provide additional data from the VIEW 1 and VIEW 2 Phase 3 trials in patients with the neovascular form of age-related macular degeneration (wet AMD), the COPERNICUS Phase 3 trial in macular edema due to central retinal vein occlusion (CRVO), and the DA VINCI Phase 2 trial in diabetic macular edema (DME). Regeneron reported positive top-line results from all these trials in the fourth quarter of 2010.

"It's a privilege to be able to release this important collection of VEGF Trap-Eye data at the Bascom Palmer Eye Institute's Eighth Annual Angiogenesis Meeting," said Philip J. Rosenfeld, M.D., Ph.D., Professor of Ophthalmology, University of Miami Miller School of Medicine and Course Co-Director of the Angiogenesis 2011 Meeting. "In particular, the Phase 3 results in wet AMD suggest that the VEGF Trap-Eye has the potential to address an important unmet need of providing optimal vision gain while reducing the burden of intravitreal injections and office visits for patients and their caregivers."

The webcast and slides may be accessed through the Company's web site, www.regeneron.com, on the Investor Relations page (<http://investor.regeneron.com>). An archived version of the presentation will be available after the live webcast through March 17, 2011.

About VEGF Trap-Eye

VEGF Trap-Eye is a recombinant fusion protein consisting of portions of human VEGF receptors 1 and 2 extracellular domains fused to the Fc portion of human IgG1 that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. VEGF Trap-Eye is specially purified and contains iso-osmotic buffer concentrations, allowing for injection into the eye.

Regeneron and Bayer HealthCare are collaborating on the global development of VEGF Trap-Eye for the treatment of wet AMD, DME, CRVO, myopic CNV, and other eye diseases and disorders. The companies plan to submit regulatory applications for marketing approval for VEGF Trap-Eye for the treatment of wet AMD in Europe and the U.S. in the first-half of 2011. Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

In November 2010, Regeneron and Bayer HealthCare announced positive top-line results from two parallel Phase 3 studies in patients with wet AMD, VIEW 1 and VIEW 2. In these trials, all regimens of VEGF Trap-Eye, including VEGF Trap-Eye dosed every two months, successfully met the primary endpoint compared to the current standard of care, ranibizumab dosed every month. The primary endpoint was statistical non-inferiority in the proportion of patients who maintained (or improved) vision over 52 weeks compared to ranibizumab. A generally favorable safety profile was observed for both VEGF Trap-Eye and ranibizumab. The most frequent ocular adverse events were conjunctival hemorrhage, macular degeneration, eye pain, retinal hemorrhage, and vitreous floaters and were balanced across all treatment groups in both studies. There were no notable differences in non-ocular adverse events among the study arms.

Trials in other indications such as CRVO and DME are currently underway or in preparation.

About Wet Age-Related Macular Degeneration (wet AMD)

Age-related macular degeneration (AMD) is a leading cause of acquired blindness. Macular degeneration is diagnosed as either dry (non-exudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating distortion and/or blind spots in central vision, and it can account for blindness in wet AMD patients. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe. It is estimated that more than 210,000 Americans are newly diagnosed with and treated for wet AMD each year.

About Central Retinal Vein Occlusion (CRVO)

Over 100,000 people in the United States and more than 66,000 people in key European countries are estimated to suffer from central retinal vein occlusion (CRVO). CRVO is caused by obstruction of the central retinal vein that leads to a back up of blood and fluid in the retina. This causes retinal injury and loss of vision. The retina can also become "ischemic" (starved for oxygen), resulting in the growth of new, inappropriate blood vessels that can cause further vision loss and more serious complications. Release of vascular endothelial growth factor (VEGF) contributes to increased vascular permeability in the eye and inappropriate new vessel growth. It is believed that anti-VEGF treatment may help decrease vascular permeability and edema and prevent the inappropriate growth of new blood vessels in the retina in patients with CRVO.

About Diabetic Macular Edema (DME)

Diabetic macular edema (DME) is the most prevalent cause of moderate vision loss in patients with diabetes. DME is a common complication of Diabetic Retinopathy (DR), a disease affecting the blood vessels of the retina. Clinically significant DME is a leading cause of blindness in younger adults (under 50). Clinically significant DME occurs when fluid leaks into the center of the macula, the light-sensitive part of the retina responsible for

sharp, direct vision. Fluid in the macula can cause severe vision loss or blindness.

Approximately 370,000 Americans currently suffer from clinically significant DME, with 95,000 new cases arising each year. According to the American Diabetes Association, more than 18 million Americans currently suffer from diabetes, and many other people are at risk for developing diabetes. With the incidence of diabetes steadily climbing, it is projected that up to 10 percent of all patients with diabetes will develop DME during their lifetime.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration and central retinal vein occlusion), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Regeneron Forward Looking Statement

This news release includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties. These include, among others, risks and timing associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or terminated without any product success, and risks associated with third party intellectual property. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2009 and Form 10-Q for the quarter ended September 30, 2010. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Investor Relations Contact:

Michael Aberman, M.D. Tel. +1 (914) 345-7799

E-Mail: michael.aberman@regeneron.com

Media Contact:

Peter Dworkin, Tel. +1 (914) 345-7640

E-Mail: peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

News Provided by Acquire Media

REGENERON

Regeneron and Bayer HealthCare Initiate Phase 3 Global Development Program For VEGF Trap-Eye In Wet Age-Related Macular Degeneration (AMD)

August 2, 2007

Regeneron and Bayer HealthCare Initiate Phase 3 Global Development Program For VEGF Trap-Eye In Wet Age-Related Macular Degeneration (AMD) TARRYTOWN, N.Y. & LEVERKUSEN, Germany--(BUSINESS WIRE)--Aug. 2, 2007--Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) and Bayer HealthCare AG (NYSE:BAY) announced today that the companies have initiated a Phase 3 study of the VEGF Trap-Eye in the neovascular form of age-related macular degeneration (wet AMD). The study will be a non-inferiority comparison of the VEGF Trap-Eye and ranibizumab (Lucentis®, a registered trademark of Genentech, Inc.), an anti-angiogenic agent approved for use in wet AMD. The study will be conducted pursuant to a Special Protocol Assessment from the U.S. Food and Drug Administration (FDA). This trial, known as VIEW 1 (VEGF Trap: Investigation of Efficacy and safety in Wet age-related macular degeneration), is the first study in the companies' Phase 3 global development program in wet AMD, which is planned to be carried out in the U.S., Europe, and other parts of the world.

"Age-related macular degeneration continues to be one of the leading causes of blindness in adults, and new therapies are essential to providing optimal patient care," stated Jeffrey Heier, M.D., a clinical ophthalmologist at Ophthalmic Consultants of Boston and chair of the steering committee for the trial. "The results of early phase studies of VEGF Trap-Eye suggest it has the potential to be a highly efficacious treatment with less frequent administration. If these results are confirmed in Phase 3 trials, it would be important for both patients and physicians and would be a significant advance in the treatment of these patients."

"The initiation of this Phase 3 trial represents a major milestone in the development of the VEGF Trap-Eye to treat wet AMD," said Avner Ingerman, M.D., vice president and ophthalmology team leader for Regeneron. "While this trial enables us to continue in our effort to improve the lives of patients suffering from wet AMD, it also signals the beginning of a larger, more global development program investigating the potential of VEGF Trap-Eye for the treatment of diabetic eye diseases and other eye diseases and disorders."

The randomized, double-masked Phase 3 study is expected to enroll approximately 1,200 patients in more than 200 centers throughout the United States and Canada. The study will evaluate the safety and efficacy of the VEGF Trap-Eye at doses of 0.5 milligrams (mg) and 2.0 mg administered at four-week dosing intervals and 2.0 mg at an eight-week dosing interval, compared to 0.5 mg of ranibizumab administered every four weeks, consistent with its labeled dosing schedule.

The primary endpoint of the study is the proportion of patients treated with the VEGF Trap-Eye who maintain or improve vision at the end of one year, compared to ranibizumab patients. Visual acuity is defined as the total number of letters read correctly on the Early Treatment Diabetic Retinopathy Study (ETDRS) chart. Maintenance of vision is defined as losing fewer than three lines (equivalent to 15 letters) on the ETDRS chart. After the first year of treatment, patients will continue to be treated and followed for another year.

In an analysis of interim data from the ongoing Phase 2 trial in wet AMD, where patients were treated with the VEGF Trap-Eye either monthly or quarterly, combined data for all patients demonstrated a statistically significant reduction in retinal thickness and improvement in visual acuity after 12 weeks, compared to baseline. There were no drug-related serious adverse events, and treatment with the VEGF Trap-Eye was generally well-tolerated. The most common adverse events were those typically associated with intravitreal injections. The interim results of this Phase 2 trial were presented at the annual meeting of the Association for Research in Vision and Ophthalmology (ARVO) this past May. The companies expect to report final primary endpoint results of the trial at a scientific meeting later this quarter.

Regeneron and Bayer HealthCare are collaborating on the global development of the VEGF Trap-Eye for the treatment of wet AMD, diabetic eye diseases, and other eye diseases and disorders. Bayer HealthCare will market the VEGF Trap-Eye outside the United States, where the parties will share equally in profits from any future sales of the VEGF Trap-Eye. Regeneron maintains exclusive rights to the VEGF Trap-Eye in the United States.

About the VEGF Trap-Eye

Vascular endothelial growth factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye, which lead to the development of wet AMD. The VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related placental growth factor (PlGF). The VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. Blockade of VEGF, which can prevent abnormal blood vessel formation and vascular leak, has proven beneficial in the treatment of wet AMD. Blocking VEGF has been shown to be effective in patients with wet AMD; and a VEGF inhibitor, ranibizumab, has been approved for treatment of patients with this condition.

About AMD

Age-related macular degeneration (AMD) is a leading cause of acquired blindness. Patients with this condition can experience a loss of vision due to the development of abnormal, fragile blood vessels in the back of the eye. A particular type of AMD, called wet AMD, accounts for approximately 90 percent of AMD-related blindness. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe.

Macular degeneration is diagnosed as either dry (nonexudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating blind spots in central vision, and it can lead to blindness in wet AMD patients.

About Regeneron Pharmaceuticals

Regeneron is a biopharmaceutical company that discovers, develops, and intends to commercialize therapeutic medicines for the treatment of serious medical conditions. Regeneron has therapeutic candidates for the potential treatment of cancer, eye diseases, and inflammatory diseases and has preclinical programs in other diseases and disorders. Additional information about Regeneron and recent news releases are available on Regeneron's worldwide web site at www.regeneron.com.

About Bayer HealthCare

The Bayer Group is a global enterprise with core competencies in the fields of health care, nutrition and high-tech materials. Bayer HealthCare, a subsidiary of Bayer AG, is one of the world's leading, innovative companies in the healthcare and medical products industry and is based in Leverkusen, Germany. The company combines the global activities of the Animal Health, Consumer Care, Diabetes Care and Pharmaceuticals divisions. The pharmaceuticals business operates under the name Bayer Schering Pharma AG. Bayer HealthCare's aim is to discover and manufacture products that will improve human and animal health worldwide. Find more information at www.bayerhealthcare.com.

Forward Looking Statement - Regeneron

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of our drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict our ability to continue to develop or commercialize our drug candidates, competing drugs that are superior to our product candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including our agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-Q for the quarter ended June 30, 2007. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

Forward-Looking Statements - Bayer HealthCare

This news release contains forward-looking statements based on current assumptions and forecasts made by Bayer Group management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in our annual and interim reports to the Frankfurt Stock Exchange and in our reports filed with the U.S. Securities and Exchange Commission (including our Form 20-F). The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

CONTACT: for Regeneron Pharmaceuticals, Inc.

Charles Poole, 914-345-7640
Investor Relations
charles.poole@regeneron.com

or

Laura Lindsay, 914-345-7800
Corporate Communications
laura.lindsay@regeneron.com

or

Lauren Tortorete, 212-845-5609
Media Relations
ltortorete@biosector2.com

SOURCE: Regeneron Pharmaceuticals, Inc.

REGENERON

Bayer and Regeneron Extend Development Program for VEGF Trap-Eye to Include Central Retinal Vein Occlusion

April 30, 2009

Bayer and Regeneron Extend Development Program for VEGF Trap-Eye to Include Central Retinal Vein Occlusion Two Phase 3 studies to start in the second half of this year

BERLIN and TARRYTOWN, N.Y., April 30 /PRNewswire-FirstCall/ -- Bayer HealthCare AG and Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced that the companies are extending their global development program for VEGF Trap-Eye, an investigational agent for the treatment of certain eye diseases, to include Central Retinal Vein Occlusion (CRVO). The companies plan to initiate a Phase 3 program evaluating the efficacy and safety of VEGF Trap-Eye in the treatment of CRVO in the second half of this year. CRVO is caused by obstruction of the central retinal vein that leads to a back up of blood and fluid in the retina, resulting in retinal injury and loss of vision. The retina can also become "ischemic" (starved for oxygen), resulting in the growth of abnormal new blood vessels that can cause further vision loss and more serious complications.

The Phase 3 program in CRVO will consist of two, multinational, one-year clinical studies which have been reviewed with regulatory authorities. These studies will expand the companies' global development collaboration for VEGF Trap-Eye, which already includes two ongoing Phase 3 studies in patients with the neovascular form of Age-related Macular Degeneration (wet AMD) and a Phase 2 study in patients with Diabetic Macular Edema (DME). Enrollment in the wet AMD and DME studies is expected to be completed later this year.

"Although CRVO is a leading cause of blindness, there is currently no treatment available that can be universally considered to be the standard of care, and there is no approved treatment to prevent the loss of vision or improve vision once it is lost," said Dr. Kemal Malik, Head of Global Development and member of the Bayer HealthCare Executive Committee. "Since the underlying biology of CRVO is related to edema and the growth of abnormal new blood vessels that are mediated by vascular endothelial growth factor (VEGF), we are hopeful that VEGF Trap-Eye may help address this significant unmet medical need."

About CRVO

Over 100,000 people in the United States are estimated to suffer from CRVO. CRVO is caused by obstruction of the central retinal vein that leads to a back up of blood and fluid in the retina, resulting in retinal injury and loss of vision. The retina can also become "ischemic" (starved for oxygen), resulting in the growth of new abnormal blood vessels that can cause further vision loss and more serious complications. Release of VEGF contributes to increased vascular permeability in the eye and abnormal new vessel growth. It is believed that anti-VEGF treatment may help decrease vascular permeability and edema and prevent the growth of abnormal new blood vessels in the retina in patients with CRVO.

About the Phase 3 CRVO Program

In the Phase 3 CRVO program for VEGF Trap-Eye, Regeneron and Bayer HealthCare will conduct two identical multinational clinical studies: COPERNICUS (COntrolled Phase 3 Evaluation of Repeated iNtravitreal administration of VEGF Trap-Eye In Central retinal vein occlusion: Utility and Safety) will be led by Regeneron and GALILEO (General Assessment Limiting Infiltration of Exudates in central retinal vein Occlusion with VEGF Trap-Eye) will be led by Bayer HealthCare. Enrollment will be initiated later in 2009.

Patients in both studies will receive 6 monthly intravitreal injections of either VEGF Trap-Eye at a dose of 2 milligrams (mg) or sham control injections. The primary endpoint of both studies is improvement in visual acuity versus baseline after 6 months of treatment. At the end of the initial 6 months, all patients will be dosed on a PRN (as needed) basis for another 6 months. All patients will be eligible for rescue laser treatment.

About VEGF Trap-Eye

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body whose normal role is to trigger formation of new blood vessels (angiogenesis) to support the growth of the body's tissues and organs. It has also been associated with the abnormal growth and fragility of new blood vessels in the eye and vascular permeability and edema. VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). Investigational VEGF Trap-Eye is a specific blocker of VEGF-A and PlGF that has been demonstrated in preclinical models to bind these growth factors with greater affinity than their natural receptors.

Regeneron and Bayer HealthCare are collaborating on the global development of VEGF Trap-Eye for the treatment of wet AMD, DME, CRVO, and other eye diseases and disorders. Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST[®] (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in clinical trials for the potential treatment of cancer, eye diseases, inflammatory diseases, and pain and has preclinical programs in other diseases and disorders. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

About Bayer HealthCare Pharmaceuticals

Bayer HealthCare Pharmaceuticals Inc. is the U.S.-based pharmaceuticals operation of Bayer HealthCare, an affiliate of Bayer AG. One of the world's

leading, innovative companies in the healthcare and medical products industry, Bayer HealthCare combines the global activities of the Animal Health, Consumer Care, Diabetes Care, and Pharmaceuticals divisions. In the United States, Bayer HealthCare Pharmaceuticals comprises the following business units: Women's Healthcare, Diagnostic Imaging, General Medicine, Hematology/Neurology, and Oncology. The company's aim is to discover and manufacture products that will improve human health worldwide by diagnosing, preventing and treating diseases.

Forward-Looking Statements - Bayer HealthCare AG

This release may contain forward-looking statements based on current assumptions and forecasts made by Bayer Group or subgroup management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in Bayer's public reports which are available on the Bayer website at www.bayer.com. The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

Forward Looking Statement -- Regeneron Pharmaceuticals, Inc.

This news release discusses historical information and includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties, such as risks associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or to terminate without any product success, risks associated with third party intellectual property, and other material risks. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2008. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise unless required by law.

SOURCE Regeneron Pharmaceuticals, Inc.; Bayer HealthCare AG

-0- 04/30/2009

/CONTACT: Anna Koch, Bayer HealthCare, +49-30-468-15942, anna.koch@bayerhealthcare.com, or Rose Talarico, +1-973-305-5302, rose.talarico@bayer.com, or Peter Dworkin, Investor Relations, +1-914-345-7640, peter.dworkin@regeneron.com, or Laura Lindsay, Media Relations, +1-914-345-7800, laura.lindsay@regeneron.com, or Olga Fleming, Media Relations, +1-212-845-5636, ofleming@biosector2.com, all of Regeneron Pharmaceuticals, Inc./

/Web Site: <http://www.regeneron.com>

<http://www.bayer.com/>

(REGN)

CO: Regeneron Pharmaceuticals, Inc.; Bayer HealthCare AG; Bayer HealthCare Pharmaceuticals Inc.

ST: Germany, New York

IN: HEA MTC

SU: TRI JVN

PR

-- NY08289 --

8289 04/30/2009 02:00 EDT <http://www.prnewswire.com>

[News](#)
[Events](#)
[Stock Information](#)
[Financial Information](#)
[Corporate Governance](#)
[FAQs](#)



REGENERON

◀ Back 

February 22, 2011 at 8:00 AM EST

REGENERON SUBMITS BIOLOGICS LICENSE APPLICATION TO FDA FOR VEGF TRAP-EYE FOR TREATMENT OF WET AGE-RELATED MACULAR DEGENERATION

TARRYTOWN, N.Y., Feb. 22, 2011 /PRNewswire/ – Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced that the company submitted a Biologics License Application (BLA) to the U.S. Food and Drug Administration (FDA) for VEGF Trap-Eye for the treatment of the neovascular form of age-related macular degeneration (wet AMD). Under the Prescription Drug User Fee Act (PDUFA), the goal for a standard review time from submission to FDA action is ten months. Regeneron's submission includes a request for Priority Review, which, if granted, would shorten the FDA's targeted goal for review time under PDUFA to six months.

"There have been significant advances in the treatment of wet AMD in recent years. However, the need for monthly intravitreal injections to obtain optimal vision gains has resulted in a significant burden for physicians, patients, and their caregivers," said Leonard S. Schleifer, M.D., Ph.D., President and Chief Executive Officer of Regeneron. "We are extremely proud to have conducted the largest global Phase 3 clinical program in patients with wet AMD, which demonstrated that patients treated with VEGF Trap-Eye 2 mg every two months, following three loading doses, were able to be dosed with fewer injections over one year without compromising efficacy. We look forward to working closely with the FDA to bring this potentially important new medicine to patients with wet AMD."

The VEGF Trap-Eye BLA is based on the positive results from two Phase 3 trials, the North American VIEW 1 trial and the global VIEW 2 trial. In these trials, all regimens of VEGF Trap-Eye, including VEGF Trap-Eye dosed 2 milligrams (mg) every two months (following three loading doses), successfully met the primary endpoint of non-inferiority, compared to the current standard of care, ranibizumab 0.5 mg dosed every month. The primary endpoint analysis was statistical non-inferiority in the proportion of patients who maintained (or improved) vision over 52 weeks compared to ranibizumab. A generally favorable safety profile was observed for both VEGF Trap-Eye and ranibizumab. The ocular adverse events were balanced across all treatment groups in both studies. There were no notable differences in non-ocular adverse events among the study arms.

About the VIEW Program

The VIEW (VEGF Trap-Eye: Investigation of Efficacy and Safety in Wet AMD) program consists of two randomized, double-masked, Phase 3 clinical trials evaluating VEGF Trap-Eye in the treatment of the neovascular form of age-related macular degeneration (wet AMD). The VIEW 1 study, which randomized 1217 patients, is being conducted in the United States and Canada by Regeneron under a Special Protocol Assessment (SPA) with the U.S. Food and Drug Administration. The VIEW 2 study, which randomized 1240 patients, is being conducted in Europe, Asia Pacific, Japan, and Latin America by Bayer HealthCare. The study designs are essentially identical. The primary endpoint evaluation was conducted at 52 weeks.

In each of the studies, VEGF Trap-Eye was evaluated for its effect on maintaining and improving vision when dosed as an intravitreal injection on a schedule of 0.5 mg monthly, 2.0 mg monthly, or 2.0 mg every two months (following three monthly loading doses), as compared with intravitreal ranibizumab administered 0.5 mg every month during the first year of the studies.

The primary endpoint of these non-inferiority studies was the proportion of patients treated with VEGF Trap-Eye who maintained visual acuity at the end of one year, compared to ranibizumab patients. Visual acuity was measured as a score based on the total number of letters read correctly on the Early Treatment Diabetic Retinopathy Study (ETDRS) eye chart, a standard chart used in research to measure visual acuity. Maintenance of vision was defined as losing fewer than three lines (equivalent to 15 letters) on the ETDRS eye chart.

The following table summarizes the VIEW 1 and VIEW 2 results for the primary and the first secondary endpoint pre-specified for testing:

	Ranibizumab 0.5mg monthly	VEGF Trap-Eye 0.5mg monthly	VEGF Trap-Eye 2mg monthly	VEGF Trap-Eye 2mg every 2 months
Maintenance of vision* (% patients losing <15 letters) at week 52 versus baseline				
VIEW 1	94.4%	95.9%**	95.1%**	95.1%**
VIEW 2	94.4%	96.3%**	95.6%**	95.6%**
Mean improvement in vision* (letters) at 52 weeks versus baseline (p-value versus ranibizumab 0.5mg monthly)***				
VIEW 1	8.1	6.9 (NS)	10.9 (p<0.01)	7.9 (NS)
VIEW 2	9.4	9.7 (NS)	7.6 (NS)	8.9 (NS)

*Visual acuity was measured as the total number of letters read correctly on the Early Treatment Diabetic Retinopathy Study (ETDRS) eye chart

**Statistically non-inferior based on a non-inferiority margin of 10%, using confidence interval approach (95.1% and 95% for VIEW 1 and VIEW 2, respectively)

*** Test for superiority

NS=not statistically significant

In the VIEW 1 and VIEW 2 trials, a generally favorable safety profile was observed for both VEGF Trap-Eye and ranibizumab. The incidence of ocular treatment emergent adverse events was balanced across all four treatment groups in both studies, with the most frequent events associated with the injection procedure, the underlying disease, and/or the aging process. The most frequent ocular adverse events were conjunctival hemorrhage, macular degeneration, eye pain, retinal hemorrhage, and vitreous floaters. The most frequent serious non-ocular adverse events were typical of those reported in this elderly population who receive intravitreal treatment for wet AMD; the most frequently reported events were falls, pneumonia, myocardial infarction, atrial fibrillation, breast cancer, and acute coronary syndrome. There were no notable differences among the study arms.

As-needed (PRN) dosing with both agents, with a dose administered at least every three months (but not more often than monthly), is being evaluated during the second year of VIEW 1 and VIEW 2. These studies are part of the global development program for VEGF Trap-Eye being conducted by Regeneron and Bayer HealthCare.

About VEGF Trap-Eye

VEGF Trap-Eye is a fusion protein consisting of portions of human VEGF receptors 1 and 2 extracellular domains fused to the Fc portion of human IgG1 that binds all forms of VEGF-A, along with the related Placental Growth Factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. VEGF Trap-Eye is specially purified and contains iso-osmotic buffer concentrations, allowing for injection into the eye.

Regeneron and Bayer HealthCare are collaborating on the development of VEGF Trap-Eye for the treatment of wet AMD, central retinal vein occlusion, diabetic macular edema, myopic choroidal neovascularisation, and other eye diseases and disorders. Bayer HealthCare intends to submit regulatory applications in the first half of 2011 for marketing approval in Europe. If approved by regulatory authorities, Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About Wet Age-Related Macular Degeneration (wet AMD)

Age-related macular degeneration (AMD) is a leading cause of acquired blindness. Macular degeneration is diagnosed as either dry (non-exudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating distortion and/or blind spots in central vision, and it can account for blindness in wet AMD patients. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe. It is estimated that more than 210,000 Americans are newly diagnosed with and treated for wet AMD each year.

About Central Retinal Vein Occlusion (CRVO)

Over 100,000 people in the United States and more than 66,000 people in key European countries are estimated to suffer from central retinal vein occlusion (CRVO). CRVO is caused by obstruction of the central retinal vein that leads to a back up of blood and fluid in the retina. This causes retinal injury and loss of vision. The retina can also become "ischemic" (starved for oxygen), resulting in the growth of new, inappropriate blood vessels that can cause further vision

loss and more serious complications. Release of vascular endothelial growth factor (VEGF) contributes to increased vascular permeability in the eye and inappropriate new vessel growth. It is believed that anti-VEGF treatment may help decrease vascular permeability and edema and prevent the inappropriate growth of new blood vessels in the retina in patients with CRVO.

About Diabetic Macular Edema (DME)

Diabetic macular edema (DME) is the most prevalent cause of moderate vision loss in patients with diabetes. DME is a common complication of Diabetic Retinopathy (DR), a disease affecting the blood vessels of the retina. Clinically significant DME is a leading cause of blindness in younger adults (under 50).

Clinically significant DME occurs when fluid leaks into the center of the macula, the light-sensitive part of the retina responsible for sharp, direct vision. Fluid in the macula can cause severe vision loss or blindness.

Approximately 370,000 Americans currently suffer from clinically significant DME, with 95,000 new cases arising each year. According to the American Diabetes Association, more than 18 million Americans currently suffer from diabetes, and many other people are at risk for developing diabetes. With the incidence of diabetes steadily climbing, it is projected that up to 10 percent of all patients with diabetes will develop DME during their lifetime.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration and central retinal vein occlusion), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Regeneron Forward Looking Statement

This news release includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties. These include, among others, risks and timing associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or terminated without any product success, and risks associated with third party intellectual property. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2010. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Investor Relations Contact:

Michael Aberman, M.D. Tel. +1 (914) 345-7799

E-Mail: michael.berman@regeneron.com

Media Contact:

Peter Dworkin, Tel. +1 (914) 345-7640

E-Mail: peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

News Provided by Acquire Media

Investor Relations

914.847.7741

invest@regeneron.com

Media Relations

media@regeneron.com

Investor [email alerts](#)

DOWNLOAD CENTER

- [2020 Proxy Statement](#)
- [2019 Annual Report](#)
- [2019 Responsibility Report](#)
- [2019 Form 10-K](#)
- [Senior Management Biographies](#)
- [Corporate Fact Sheet](#)
- [Regeneron Genetics Center Backgrounder](#)

Investor Relations

914.847.7741

invest@regeneron.com

Media Relations

media@regeneron.com

Investor [email alerts](#)



EMAIL
ALERTS



RSS



PRINT



SHARE



SEARCH

REGENERON

Regeneron and Bayer Announce Start of Phase 3 Clinical Program in Diabetic Macular Edema

April 8, 2011

Tarrytown, NY, USA, and Berlin, Germany, April 8, 2011 -- Regeneron Pharmaceuticals, Inc. (NASDAQ: **REGN**) and Bayer HealthCare today announced that they have initiated the first of two Phase 3 clinical trials evaluating the efficacy and safety of VEGF Trap-Eye (afibercept ophthalmic solution), an investigational new agent for the treatment of certain eye diseases, in the treatment of Diabetic Macular Edema (DME). The companies are extending their development program for VEGF Trap-Eye in DME after promising results in the global Phase 2 DME program.

The first Phase 3 trial in DME, named VIVID-DME, is being led by Bayer HealthCare and has started in Australia. The trial will also be conducted in Europe and Japan. A second study led by Regeneron, named VISTA-DME, is expected to begin later in 2011 in the United States, Canada, and other countries.

"Clinically significant DME is a leading cause of vision loss in adults under the age of 50 suffering from diabetes," said Dr. Kemal Malik, Head of Global Development and member of the Bayer HealthCare Executive Committee. "After reporting positive results from our global Phase 3 program (VIEW 1 and VIEW 2 studies) for the treatment of the neovascular form of age-related macular degeneration (wet AMD), we are pleased to start a Phase 3 program with VEGF Trap-Eye in DME which may help to address this significant unmet medical need."

The Phase 3 program in DME expands the companies' global development collaboration for VEGF Trap-Eye. The companies announced positive data for two Phase 3 studies in patients with wet AMD in November 2010 and for the first of two Phase 3 studies in patients with Central Retinal Vein Occlusion (CRVO) in December 2010.

About the Phase 3 DME Program

The VIVID-DME study (VEGF Trap-Eye In Vision Impairment Due to DME) has three study arms. In the first arm, patients will be treated every month with 2 milligrams (mg) of VEGF Trap-Eye. In the second arm, patients will be treated with 2mg of VEGF Trap-Eye every two months after a loading phase of monthly injections. In the third arm, the comparator arm, patients will be treated with macular laser photocoagulation. The primary endpoint is mean change in visual acuity from baseline as measured by the Early Treatment Diabetic Retinopathy Study (ETDRS) eye chart, a standard chart used in research to measure visual acuity. All patients will be followed for three years. The VISTA-DME study (VEGF Trap-Eye: Investigation of Safety, Treatment effect, and Anatomic outcomes in DME) is expected to begin later in 2011.

About Diabetic Macular Edema (DME)

DME is the most prevalent cause of moderate vision loss in patients with diabetes. DME is a common complication of Diabetic Retinopathy (DR), a disease affecting the blood vessels of the retina. Clinically significant DME is a leading cause of blindness in younger adults (under 50). Clinically significant DME occurs when fluid leaks into the center of the macula, the light-sensitive part of the retina responsible for sharp, direct vision. Fluid in the macula can cause severe vision loss or blindness.

According to figures from the World Health Organization, DME is the second leading cause of blindness in Western industrialized countries. In Europe, about 8% of the population is affected by diabetes. Approximately 370,000 Americans currently suffer from clinically significant DME, with 95,000 new cases arising each year. According to the American Diabetes Association, over 18 million Americans currently suffer from diabetes, and many more are at risk for developing diabetes. The incidence of diabetes is steadily climbing and it is projected that up to 10 percent of all patients with diabetes will develop DME during their lifetime.

About VEGF Trap-Eye

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body. Its normal role in a healthy organism is to trigger formation of new blood vessels (angiogenesis) supporting the growth of the body's tissues and organs. However, in certain diseases, such as diabetes, it is also associated with the growth of abnormal new blood vessels in the eye, which exhibit vascular permeability and lead to edema. VEGF Trap-Eye is a fully human, soluble VEGF receptor fusion protein that binds all forms of VEGF-A along with another vascular growth factor, the Placental Growth Factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of VEGF-A and PlGF that has been demonstrated in preclinical models to bind these growth factors with greater affinity than their natural receptors. Regeneron and Bayer HealthCare are collaborating on the global development of VEGF Trap-Eye. Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

Regeneron submitted a Biologics License Application for marketing approval in wet age-related macular degeneration (wet AMD) in the US in February 2011, and Bayer plans to submit a regulatory application outside the US in the first half of 2011. Phase 3 studies in central retinal vein occlusion (CRVO) and in patients with choroidal neovascularisation (CNV) of the retina as a result of pathologic myopia - a major eye disease common in Asia - are currently underway.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (riloncept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration and central retinal vein occlusion), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

About Bayer HealthCare

The Bayer Group is a global enterprise with core competencies in the fields of health care, nutrition and high-tech materials. Bayer HealthCare, a subgroup of Bayer AG with annual sales of EUR 16.913 billion (2010), is one of the world's leading, innovative companies in the healthcare and medical products industry and is based in Leverkusen, Germany. The company combines the global activities of the Animal Health, Consumer Care, Medical Care and Pharmaceuticals divisions. Bayer HealthCare's aim is to discover and manufacture products that will improve human and animal health worldwide. Bayer HealthCare has a global workforce of 55.700 employees (Dec 31, 2010) and is represented in more than 100 countries. Find more information at www.bayerhealthcare.com.

Regeneron Forward-Looking Statements

This news release includes forward-looking statements about Regeneron and its products, development programs, finances, and business, all of which involve a number of risks and uncertainties. These include, among others, risks and timing associated with preclinical and clinical development of Regeneron's drug candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that are superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or terminated without any product success, and risks associated with third party intellectual property. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission (SEC), including its Form 10-K for the year ended December 31, 2010. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Bayer Forward-Looking Statements

This release may contain forward-looking statements based on current assumptions and forecasts made by Bayer Group or subgroup management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in Bayer's public reports which are available on the Bayer website at www.bayer.com. The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

To learn more about Age-related macular degeneration (AMD), please visit:

www.bayerpharma.com.

Your Contact at Bayer:

Doreen Schroeder, Tel. +49 30 468-11399

E-Mail: doreen.schroeder@bayer.com

Your Investor Relations Contact at Regeneron:

Michael Aberman, MD., Tel. +1 (914) 345-7799

E-Mail: michael.aberman@regeneron.com

Your Media Contact at Regeneron:

Peter Dworkin, Tel. +1 (914) 345-7640

E-Mail: peter.dworkin@regeneron.com

REGENERON

FDA Grants Priority Review for VEGF Trap-Eye for the Treatment of Wet Age-Related Macular Degeneration

April 18, 2011

TARRYTOWN, N.Y., April 18, 2011 /PRNewswire/ -- Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced that the U.S. Food and Drug Administration (FDA) has accepted for review the Company's Biologics License Application (BLA) for VEGF Trap-Eye for the treatment of the neovascular form of age-related macular degeneration (wet AMD). The FDA also granted the Company's request for priority review of its BLA. A Priority Review designation is given to drugs that offer major advances in treatment, or provide a treatment where no adequate therapy exists. Under priority review, the target date for an FDA decision on the VEGF Trap-Eye BLA is August 20, 2011.

"We are very pleased that the FDA has chosen to grant priority review to VEGF Trap-Eye. We look forward to working closely with the FDA to achieve our goal of bringing a new treatment option that offers a major advance to patients with age-related macular degeneration," said Leonard S. Schleifer, M.D., Ph.D., President and Chief Executive Officer of Regeneron.

About VEGF Trap-Eye

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body. Its normal role in a healthy organism is to trigger formation of new blood vessels (angiogenesis) supporting the growth of the body's tissues and organs. However, in certain diseases, such as diabetes, it is also associated with the growth of abnormal new blood vessels in the eye, which exhibit vascular permeability and lead to edema. VEGF Trap-Eye is a fusion protein consisting of portions of human VEGF receptors 1 and 2 extracellular domains fused to the Fc portion of human IgG1. VEGF Trap-Eye binds all forms of VEGF-A, along with the related Placental Growth Factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. VEGF Trap-Eye is specially purified and contains iso-osmotic buffer concentrations, allowing for injection into the eye.

Regeneron and Bayer HealthCare are collaborating on the development of VEGF Trap-Eye for the treatment of wet AMD, central retinal vein occlusion, diabetic macular edema, myopic choroidal neovascularisation, and other eye diseases and disorders. Bayer HealthCare intends to submit a regulatory application outside of the United States in the first half of 2011. If approved by regulatory authorities, Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally in profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration and central retinal vein occlusion), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Forward Looking Statements

This news release includes forward-looking statements that involve risks and uncertainties relating to future events and the future financial performance of Regeneron, and actual events or results may differ materially from these forward-looking statements. These statements concern, and these risks and uncertainties include, among others, the nature, timing, and possible success and therapeutic applications of our product candidates and research and clinical programs now underway or planned, the likelihood and timing of possible regulatory approval and commercial launch of our late-stage product candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict our ability to continue to develop or commercialize our product and drug candidates, competing drugs that may be superior to our product and drug candidates, uncertainty of market acceptance of our product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including our agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or terminated without any product success, and risks associated with third party intellectual property. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission, including its Form 10-K for the year ended December 31, 2010. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Contact Information:

Michael Aberman, M.D.	Peter Dworkin
Investor Relations	Corporate Communications
914.345.7799	914.345.7640
michael.aberman@regeneron.com	peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

REGENERON

VEGF Trap-Eye Submitted for EU Marketing Authorization for Treatment of Wet Age-Related Macular Degeneration

June 7, 2011

TARRYTOWN, N.Y. and BERLIN, June 7, 2011 /PRNewswire/ – Regeneron Pharmaceuticals, Inc. (NASDAQ: REGN) and Bayer HealthCare today announced that Bayer HealthCare has submitted an application for marketing authorization in Europe for VEGF Trap-Eye for the treatment of the neovascular form of age-related macular degeneration (wet AMD). Regeneron and Bayer HealthCare are collaborating on the global development of VEGF Trap-Eye for the treatment of wet AMD, central retinal vein occlusion (CRVO), diabetic macular edema (DME), and myopic choroidal neovascularization (mCNV).

"The submission of VEGF Trap-Eye for EU marketing authorization represents a significant milestone in our goal to bring this potentially important new therapy to patients with wet AMD across the globe," said Leonard S. Schleifer, M.D., Ph.D., President and Chief Executive Officer of Regeneron.

The VEGF Trap-Eye submission is based on the positive results from two Phase 3 trials, the VIEW 1 study and the VIEW 2 study. In these trials, all regimens of VEGF Trap-Eye, including 2 mg VEGF Trap-Eye dosed every two months (following three loading doses), successfully met the primary endpoint of non-inferiority, compared to the current standard of care, ranibizumab 0.5 mg dosed every month. The primary endpoint analysis was statistical non-inferiority in the proportion of patients who maintained (or improved) vision over 52 weeks compared to ranibizumab at the dose that is currently known to provide the best possible efficacy. A generally favorable safety profile was observed for both VEGF Trap-Eye and ranibizumab.

The ocular adverse events were balanced across all treatment groups in both studies. There were no notable differences in non-ocular adverse events among the study arms.

Regeneron submitted a Biologics License Application (BLA) for marketing approval in wet AMD in the U.S. in February 2011 and received a Priority Review designation.

Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally the profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About the VIEW Program

The VIEW (VEGF Trap-Eye: Investigation of Efficacy and Safety in Wet AMD) program consists of two randomized, double-masked, Phase 3 clinical trials evaluating VEGF Trap-Eye in the treatment of the neovascular form of age-related macular degeneration (wet AMD). The VIEW 1 study, which randomized 1,217 patients, is being conducted in the United States and Canada by Regeneron under a Special Protocol Assessment (SPA) with the U.S. Food and Drug Administration. The VIEW 2 study, which randomized 1,240 patients, is being conducted in Europe, Asia Pacific, Japan, and Latin America by Bayer HealthCare. The study designs are essentially identical. The primary endpoint evaluation was conducted at 52 weeks.

In each of the studies, VEGF Trap-Eye was evaluated for its effect on maintaining and improving vision when dosed as an intravitreal injection on a schedule of 0.5 mg monthly, 2 mg monthly, or 2 mg every two months (following three monthly loading doses), as compared with intravitreal ranibizumab administered 0.5 mg every month during the first year of the studies. As-needed (PRN) dosing with both agents, with a dose administered at least every three months (but not more often than monthly) is being evaluated during the second year of each study.

About VEGF Trap-Eye

VEGF Trap-Eye is a fully human fusion protein, consisting of portions of VEGF receptors 1 and 2, that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). VEGF Trap-Eye is a specific and highly potent blocker of these growth factors. VEGF Trap-Eye is specially purified and contains iso-osmotic buffer concentrations, allowing for injection into the eye.

Regeneron and Bayer HealthCare are collaborating on the global development of VEGF Trap-Eye for the treatment of the neovascular form of age related macular degeneration (wet AMD), central retinal vein occlusion (CRVO), diabetic macular edema (DME), myopic choroidal neovascularization (mCNV), and other eye diseases and disorders.

Regeneron submitted a Biologics License Application (BLA) for marketing approval in wet AMD in the U.S. in February 2011 and received a Priority Review designation. Under Priority Review, the target date for an FDA decision on the VEGF Trap-Eye BLA is August 20, 2011.

In April 2011, Bayer HealthCare and Regeneron announced the initiation of a Phase 3 program in DME.

Bayer HealthCare will market VEGF Trap-Eye outside the United States, where the companies will share equally the profits from any future sales of VEGF Trap-Eye. Regeneron maintains exclusive rights to VEGF Trap-Eye in the United States.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration, central retinal vein occlusion, and diabetic macular edema), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

About Bayer HealthCare

The Bayer Group is a global enterprise with core competencies in the fields of health care, nutrition and high-tech materials. Bayer HealthCare, a subgroup of Bayer AG with annual sales of more than EUR 16.913 billion (2010), is one of the world's leading, innovative companies in the healthcare and medical products industry and is based in Leverkusen, Germany. The company combines the global activities of the Animal Health, Consumer Care, Medical Care and Pharmaceuticals divisions. Bayer HealthCare's aim is to discover and manufacture products that will improve human and animal health worldwide. Bayer HealthCare has a global workforce of 55.700 employees and is represented in more than 100 countries. Find more information at www.bayerhealthcare.com.

Regeneron Forward-Looking Statements

This news release includes forward-looking statements that involve risks and uncertainties relating to future events and the future financial performance of Regeneron, and actual events or results may differ materially from these forward-looking statements. These statements concern, and these risks and uncertainties include, among others, the nature, timing, and possible success and therapeutic applications of Regeneron's product candidates and research and clinical programs now underway or planned, the likelihood and timing of possible regulatory approval and commercial launch of Regeneron's late-stage product candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that may be superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including Regeneron's agreements with the sanofi-aventis Group and Bayer HealthCare, to be canceled or terminated without any product success, and risks associated with third party intellectual property and pending or future litigation relating thereto. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission, including its Form 10-K for the year ended December 31, 2010 and Form 10-Q for the quarter ended March 31, 2011. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Bayer Forward-Looking Statements

This release may contain forward-looking statements based on current assumptions and forecasts made by Bayer Group or subgroup management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in Bayer's public reports which are available on the Bayer website at www.bayer.com. The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

Your Contact at Bayer:

Doreen Schroeder, Tel. +49 30 468-11399

E-Mail: doreen.schroeder@bayer.com

Your Investor Relations Contact at Regeneron:

Michael Aberman, M.D., Tel. +1 (914) 345-7799

E-Mail: michael.aberman@regeneron.com

Your Media Contact at Regeneron:

Peter Dworkin, Tel. +1 (914) 345-7640

E-Mail: peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

News Provided by Acquire Media

REGENERON

Regeneron Announces EYLEA™ (afibercept ophthalmic solution) Receives Unanimous Recommendation for Approval for Treatment of Wet AMD from FDA Advisory Committee

June 17, 2011

TARRYTOWN, N.Y., June 17, 2011 /PRNewswire/ -- Regeneron Pharmaceuticals, Inc. (Nasdaq: **REGN**) today announced that the Dermatologic and Ophthalmic Drugs Advisory Committee of the U.S. Food and Drug Administration (FDA) has voted unanimously to recommend that the FDA approve EYLEA™, also known as VEGF Trap-Eye, for the treatment of the neovascular form of age-related macular degeneration (wet AMD) at a dose of 2 milligrams (mg) every eight weeks, following three initial doses given every four weeks.

The committee's recommendation will be considered by the FDA in its review of the Biologics License Application (BLA) for EYLEA, but the committee's recommendation is not binding on the FDA. Regeneron submitted a BLA for marketing approval in wet AMD in the U.S. in February 2011 and received a Priority Review designation. Under Priority Review, the target date for an FDA decision on the EYLEA BLA is August 20, 2011.

"The positive recommendation by the advisory committee is an important step toward providing wet AMD patients with a new treatment option that could potentially reduce the burden that exists with current therapies," said George D. Yancopoulos, M.D., Ph.D., President of Regeneron Research Laboratories. "We look forward to continuing to work with the FDA as it completes its evaluation of the EYLEA BLA."

About EYLEA

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body. Its normal role in a healthy organism is to trigger formation of new blood vessels (angiogenesis) supporting the growth of the body's tissues and organs. However, in certain diseases, such as age-related macular degeneration, it is also associated with the growth of abnormal new blood vessels in the eye, which exhibit vascular permeability and lead to edema.

EYLEA (afibercept ophthalmic solution), also known as VEGF Trap-Eye, is a fully human fusion protein, consisting of portions of VEGF receptors 1 and 2, that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). EYLEA is a specific and highly potent blocker of these growth factors. EYLEA is specially purified and contains iso-osmotic buffer concentrations, allowing for injection into the eye.

Regeneron and Bayer HealthCare are collaborating on the global development of EYLEA for the treatment of the neovascular form of age-related macular degeneration (wet AMD), central retinal vein occlusion (CRVO), diabetic macular edema (DME), and other eye diseases and disorders. Bayer submitted an application for marketing authorization in Europe in wet AMD in June 2011.

The EYLEA wet AMD regulatory submissions are based on the positive results from two Phase 3 trials, the VIEW 1 study and the VIEW 2 study. In these trials, all regimens of EYLEA, including 2 milligrams (mg) of EYLEA dosed every two months (following three loading doses), successfully met the primary endpoint of non-inferiority compared to the current standard of care, ranibizumab 0.5 mg dosed every month. The primary endpoint analysis was statistical non-inferiority in the proportion of patients who maintained (or improved) vision over 52 weeks compared to ranibizumab. A generally favorable safety profile was observed for both EYLEA and ranibizumab. The most frequent ocular adverse events were conjunctival hemorrhage, macular degeneration, eye pain, retinal hemorrhage, and vitreous floaters.

Bayer HealthCare will market EYLEA™ outside the United States, where the companies will share equally the profits from any future sales of EYLEA. Regeneron maintains exclusive rights to EYLEA in the United States.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (rilonacept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration, central retinal vein occlusion, and diabetic macular edema), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Regeneron Forward Looking Statement

This news release includes forward-looking statements that involve risks and uncertainties relating to future events and the future financial performance of Regeneron, and actual events or results may differ materially from these forward-looking statements. These statements concern, and these risks and uncertainties include, among others, the nature, timing, and possible success and therapeutic applications of Regeneron's product candidates and research and clinical programs now underway or planned, the likelihood and timing of possible regulatory approval and commercial launch of Regeneron's late-stage product candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize its product and drug candidates, competing drugs that may be superior to Regeneron's product and drug candidates, uncertainty of market acceptance of Regeneron's product and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including Regeneron's agreements with Sanofi and Bayer HealthCare, to be canceled or terminated without any product success, and risks associated with third party intellectual property and pending or future litigation relating thereto. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission, including its Form 10-K for the year ended December 31, 2010 and Form 10-Q for the quarter ended March 31, 2011. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Contact Information:

Michael Aberman, M.D.	Peter Dworkin
Investor Relations	Corporate Communications
914.345.7799	914.345.7640
michael.aberman@regeneron.com	peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

News Provided by Acquire Media

August 17, 2011

Regeneron Announces Clinical Presentations at ASRS 2011 Annual Meeting

TARRYTOWN, N.Y., Aug. 17, 2011 /PRNewswire/ -- Regeneron Pharmaceuticals, Inc. (Nasdaq: REGN) today announced that clinical data from four separate clinical studies of EYLEA™ (aflibercept injection) will be presented at the upcoming American Society of Retina Specialists (ASRS) meeting on Sunday, August 21 and Monday, August 22, 2011 in Boston, Massachusetts.

The presentations are:

- "Analysis of 2,457 Patients in the Phase 3 VIEW 1 and VIEW 2 Studies Comparing VEGF Trap-Eye and Ranibizumab in Neovascular AMD" will be presented by Jeffrey S. Heier, M.D. on Sunday, August 21 at 8:21 a.m.
- "One-year Results of the DA VINCI Study of VEGF Trap-Eye in DME" will be presented by Diana V. Do, M.D. on Sunday, August 21 at 2:48 p.m.
- "The 6-Month (Primary Endpoint) Results of the Phase 3 GALILEO Study: VEGF Trap-Eye in CRVO" will be presented by Jean-Francois Korobelnik, M.D. on Monday, August 22 at 8:20 a.m.
- "Trap-Eye in CRVO: 1-year Results of the Phase 3 COPERNICUS Study" will be presented by W. Lloyd Clark, M.D. on Monday, August 22 at 8:28 a.m.

About EYLEA™ (aflibercept injection)

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body. Its normal role in a healthy organism is to trigger formation of new blood vessels (angiogenesis) supporting the growth of the body's tissues and organs. However, in certain diseases, such as age-related macular degeneration, it is also associated with the growth of abnormal new blood vessels in the eye, which exhibit vascular permeability and lead to edema.

EYLEA, also known as VEGF Trap-Eye, is a fully human fusion protein, consisting of portions of VEGF receptors 1 and 2, that binds all forms of VEGF-A along with the related Placental Growth Factor (PlGF). EYLEA is a specific and highly potent blocker of these growth factors. EYLEA is specially purified and contains iso-osmotic buffer concentrations, allowing for injection into the eye.

Regeneron and Bayer HealthCare are collaborating on the global development of EYLEA for the treatment of neovascular age-related macular degeneration (wet AMD), central retinal vein occlusion (CRVO), diabetic macular edema (DME), and other eye diseases and disorders. Bayer submitted an application for marketing authorization in Europe in wet AMD in June 2011.

Bayer HealthCare will market EYLEA™ (aflibercept injection) outside the United States, where the companies will share equally the profits from any future sales of EYLEA. Regeneron maintains exclusive rights to EYLEA in the United States.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, develops, and commercializes medicines for the treatment of serious medical conditions. In addition to ARCALYST® (riloncept) Injection for Subcutaneous Use, its first commercialized product, Regeneron has therapeutic candidates in Phase 3 clinical trials for the potential treatment of gout, diseases of the eye (wet age-related macular degeneration, central retinal vein occlusion, and diabetic macular edema), and certain cancers. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on Regeneron's web site at www.regeneron.com.

Contact Information:

Michael Aberman, M.D.

Investor Relations

914.345.7799

michael.aberman@regeneron.com

Peter Dworkin

Corporate Communications

914.345.7640

peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

News Provided by Acquire Media

REGENERON

Regeneron Announces FDA Approval of EYLEA™ (aflibercept) Injection for the Treatment of Wet Age-Related Macular Degeneration: CORRECTED

November 18, 2011

In the news release, Regeneron Announces FDA Approval of EYLEA™ (aflibercept) Injection for the Treatment of Wet Age-Related Macular Degeneration, issued 18-Nov-2011 by Regeneron Pharmaceuticals, Inc. over PR Newswire, the third paragraph, second sentence, should read "EYLEA offers the potential of achieving the efficacy we've come to expect from current anti-VEGF agents, but with less frequent injections and monitoring." The complete, corrected release follows:

TARRYTOWN, N.Y., Nov. 18, 2011 /PRNewswire/ -- Regeneron Pharmaceuticals, Inc. (Nasdaq: **REGN**) today announced that the U.S. Food and Drug Administration (FDA) has approved EYLEA (aflibercept) Injection, known in the scientific literature as VEGF Trap-Eye, for the treatment of patients with neovascular (wet) Age-related Macular Degeneration (AMD) at a recommended dose of 2 milligrams (mg) every four weeks (monthly) for the first 12 weeks, followed by 2 mg every eight weeks (2 months).

The approval of EYLEA was granted under a Priority Review, a designation that is given to drugs that offer major advances in treatment, or provide a treatment where no adequate therapy exists. This approval was based upon the results of two Phase 3 clinical studies. In these studies, EYLEA dosed every eight weeks, following three initial monthly injections, was clinically equivalent to the standard of care, Lucentis® (ranibizumab injection) dosed every four weeks, as measured by the primary endpoint of maintenance of visual acuity (less than 15 letters of vision loss on an eye chart) over 52 weeks. The most common adverse reactions (frequency of 5% or more) reported in patients receiving EYLEA were conjunctival hemorrhage, eye pain, cataract, vitreous detachment, vitreous floaters, and increased intraocular pressure. The adverse event profile was similar to that seen with ranibizumab.

"The approval of EYLEA offers a much needed new treatment option for patients with wet AMD," said Jeffrey Heier, M.D., a clinical ophthalmologist and retinal specialist at Ophthalmic Consultants of Boston, Assistant Professor of Ophthalmology at Tufts School of Medicine, and Chair of the Steering Committee for the VIEW 1 trial. "EYLEA offers the potential of achieving the efficacy we've come to expect from current anti-VEGF agents, but with less frequent injections and monitoring. This may reduce the need for costly and time-consuming monthly office visits for patients and their caregivers."

"This approval is an important step forward for Regeneron and for patients suffering with wet AMD, the most common cause of blindness in the U.S. in older adults," said Leonard S. Schleifer, M.D., Ph.D., President and Chief Executive Officer of Regeneron. "We thank the patients and clinical investigators who participated in our clinical studies, the FDA, and the Regeneron employees who helped make this day possible. Now that EYLEA is approved, we plan to make EYLEA available to patients within the next few days."

About EYLEA™ (aflibercept) Injection

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body. Its normal role in a healthy organism is to trigger formation of new blood vessels (angiogenesis) supporting the growth of the body's tissues and organs. However, in certain diseases, such as wet age-related macular degeneration, it is also associated with the growth of abnormal new blood vessels in the eye, which exhibit abnormal increased permeability that leads to edema. Scarring and loss of fine-resolution central vision often results.

EYLEA, known in the scientific literature as VEGF Trap-Eye, is a recombinant fusion protein, consisting of portions of human VEGF receptors 1 and 2 extracellular domains fused to the Fc portion of human IgG1 and formulated as an iso-osmotic solution for intravitreal administration. EYLEA acts as a soluble decoy receptor that binds VEGF-A and placental growth factor (PlGF) and thereby can inhibit the binding and activation of these cognate VEGF receptors.

EYLEA is indicated for the treatment of patients with neovascular age-related macular degeneration (wet AMD). EYLEA is contraindicated in patients with ocular or periocular infections, active intraocular inflammation, or known hypersensitivity to aflibercept or to any of the excipients in EYLEA.

The recommended dose for EYLEA is 2 mg administered by intravitreal injection every four weeks (monthly) for the first 12 weeks (3 months), followed by 2 mg once every eight weeks (2 months). Although EYLEA may be dosed as frequently as 2 mg every four weeks (monthly), additional efficacy was not demonstrated when EYLEA was dosed every four weeks compared to every eight weeks.

There is a potential risk of arterial thromboembolic events (ATEs) following use of intravitreal VEGF inhibitors, including EYLEA, defined as nonfatal stroke, nonfatal myocardial infarction, or vascular death (including deaths of unknown cause). The incidence of ATEs with EYLEA in clinical trials was low (1.8%).

Serious adverse reactions related to the injection procedure have occurred in less than 0.1% of intravitreal injections with EYLEA and include endophthalmitis, traumatic cataract, and increased intraocular pressure.

About the VIEW 1 and VIEW 2 Clinical Studies

The safety and efficacy of EYLEA were assessed in two randomized, multi-center, double-masked, active-controlled studies in patients with wet AMD. A total of 2412 patients were treated and evaluable for efficacy (1817 with EYLEA) in the two studies (VIEW 1 and VIEW 2). In each study, patients were randomly assigned in a 1:1:1:1 ratio to one of four dosing regimens: 1) EYLEA administered 2 mg every eight weeks following three initial

monthly doses (EYLEA 2Q8); 2) EYLEA administered 2 mg every four weeks (EYLEA 2Q4); 3) EYLEA 0.5 mg administered every four weeks (EYLEA 0.5Q4); and 4) ranibizumab administered 0.5 mg every four weeks (ranibizumab 0.5Q4). Patient ages ranged from 49 to 99 years with a mean of 76 years.

In both studies, the primary efficacy endpoint was the proportion of patients who maintained vision, defined as losing fewer than 15 letters of visual acuity at week 52 compared to baseline. Data are available through week 52. Both the EYLEA™ (aflibercept) Injection 2Q8 and 2Q4 dosing groups were shown to have efficacy that was clinically equivalent to the ranibizumab 0.5Q4 group for the primary endpoint.

Select results of the VIEW 1 and VIEW 2 studies as described in the full Prescribing Information for the EYLEA 2 mg every four weeks and EYLEA 2 mg every eight weeks dosing groups as compared to ranibizumab dosed monthly group are shown below.

Efficacy Outcomes at Week 52 (Full Analysis Set with LOCF) in VIEW 1 and VIEW 2 Studies

	VIEW 1			VIEW 2		
	EYLEA 2 mg Q8 weeks(a)	EYLEA 2 mg Q4 weeks	ranibizu- mab 0.5 mg Q4 weeks	EYLEA 2 mg Q8 weeks(a)	EYLEA 2 mg Q4 weeks	ranibizu- mab 0.5 mg Q4 weeks
Full Analysis Set	N=301	N=304	N=304	N=306	N=309	N=291
Efficacy Outcomes						
Proportion of patients who maintained visual acuity (%) (

REGENERON

Regeneron and Bayer Initiate Phase 3 Clinical Program for the Treatment of Wet Age-Related Macular Degeneration in China

November 28, 2011

TARRYTOWN, N.Y. and BERLIN, Nov. 28, 2011 /PRNewswire/ -- Regeneron Pharmaceuticals, Inc. (NASDAQ: REGN) and Bayer HealthCare today announced that they have initiated a Phase 3 clinical trial evaluating the efficacy and safety of EYLEA™ (afibercept) Injection in the neovascular form of age-related macular degeneration (wet AMD) in China.

The new trial, named SIGHT, will include approximately 300 patients and will be the largest retinal trial conducted in China. SIGHT is being led by Bayer.

"Currently, only photodynamic therapy with verteporfin is approved as a treatment for wet AMD in China, and it is only approved for the subpopulation of patients with predominantly classic wet AMD," said Kemal Malik, M.D., Head of Global Development and member of the Bayer HealthCare Executive Committee. "After reporting positive data from our large VIEW program in wet AMD, we look forward to potentially bringing this new treatment to patients with wet AMD in China."

About the SIGHT Program

The SIGHT (VEGF Trap-Eye: Investigation of Efficacy and Safety in Chinese patients with wet AMD) program consists of a randomized, double-masked, Phase 3 clinical trial evaluating EYLEA (known in the scientific literature as VEGF Trap-Eye) in the treatment of the neovascular form of age-related macular degeneration (wet AMD). EYLEA will be evaluated for its effect on improving and maintaining vision when dosed as an intravitreal injection on a schedule of 2 milligrams (mg) every two months (following three initial monthly doses), as compared with Photodynamic Therapy (PDT) with verteporfin. After assessment of the primary endpoint at week 28, all patients, including those on PDT, will receive EYLEA treatment until the end of the study at week 52. The SIGHT study plans to randomize 300 patients.

About Wet AMD

Age-related Macular Degeneration (AMD) is a leading cause of acquired blindness. Macular degeneration is diagnosed as either dry (non-exudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction of the retina creating blind spots in central vision, and it can account for blindness in wet AMD patients. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe. In China, there were an estimated 540,000 newly diagnosed wet AMD patients over the age of 50 in 2010.

About EYLEA™ (afibercept) Injection For Intravitreal Injection

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body. Its normal role in a healthy organism is to trigger formation of new blood vessels (angiogenesis) supporting the growth of the body's tissues and organs. However, in certain diseases, such as wet age-related macular degeneration, it is also associated with the growth of abnormal new blood vessels in the eye, which exhibit abnormal increased permeability that leads to edema. Scarring and loss of fine-resolution central vision often results.

EYLEA, known in the scientific literature as VEGF Trap-Eye, is a recombinant fusion protein, consisting of portions of human VEGF receptors 1 and 2 extracellular domains fused to the Fc portion of human IgG1 and formulated as an iso-osmotic solution for intravitreal administration. EYLEA acts as a soluble decoy receptor that binds VEGF-A and placental growth factor (PlGF) and thereby can inhibit the binding and activation of these cognate VEGF receptors.

IMPORTANT PRESCRIBING INFORMATION

In this U.S. EYLEA is indicated for the treatment of patients with neovascular age-related macular degeneration (wet AMD).

The recommended dose for EYLEA is 2 mg administered by intravitreal injection every four weeks (monthly) for the first 12 weeks (3 months), followed by 2 mg once every eight weeks (2 months). Although EYLEA may be dosed as frequently as 2 mg every four weeks (monthly), additional efficacy was not demonstrated when EYLEA was dosed every four weeks compared to every eight weeks.

IMPORTANT SAFETY INFORMATION

EYLEA is contraindicated in patients with ocular or periocular infections, active intraocular inflammation, or known hypersensitivity to afibercept or to any of the excipients in EYLEA.

Intravitreal injections, including those with EYLEA, have been associated with endophthalmitis and retinal detachments. Proper aseptic injection technique must always be used when administering EYLEA. Patients should be instructed to report any symptoms suggestive of endophthalmitis or retinal detachment without delay and should be managed appropriately.

Acute increases in intraocular pressure have been seen within 60 minutes of intravitreal injection, including with EYLEA. Sustained increases in intraocular pressure have also been reported after repeated intravitreal dosing with VEGF inhibitors. Intraocular pressure and the perfusion of the optic nerve head should be monitored and managed appropriately.

There is a potential risk of arterial thromboembolic events (ATEs) following use of intravitreal VEGF inhibitors, including EYLEA, defined as nonfatal

stroke, nonfatal myocardial infarction, or vascular death (including deaths of unknown cause). The incidence of ATEs with EYLEA in clinical trials was low (1.8%).

Serious adverse reactions related to the injection procedure have occurred in less than 0.1% of intravitreal injections with EYLEA including endophthalmitis, traumatic cataract, and increased intraocular pressure.

The most common adverse reactions (greater than or equal to 5%) reported in patients receiving EYLEA were conjunctival hemorrhage, eye pain, cataract, vitreous detachment, vitreous floaters, and increased intraocular pressure.

Please see the full Prescribing Information for EYLEA, available online at www.regeneron.com/EYLEA-foi.pdf.

About the EYLEA™ (aflibercept) Injection Global Collaboration

Regeneron is collaborating with Bayer HealthCare on the global development of EYLEA. Bayer submitted an application for marketing authorization in Europe for wet AMD in June 2011.

Bayer HealthCare will market EYLEA outside the United States, where the companies will share equally the profits from any future sales of EYLEA. Regeneron maintains exclusive rights to EYLEA in the United States.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, invents, develops, manufactures, and commercializes medicines for the treatment of serious medical conditions. Regeneron markets two products, ARCALYST® (rilonacept) Injection For Subcutaneous Use and EYLEA™ (aflibercept) Injection. Regeneron also has completed several Phase 3 studies and is conducting an additional Phase 3 clinical trial for the product candidate ZALTRAP® (aflibercept) Concentrate for Intravenous Infusion. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on the Regeneron web site at www.regeneron.com.

About Bayer HealthCare

The Bayer Group is a global enterprise with core competencies in the fields of health care, nutrition and high-tech materials. Bayer HealthCare, a subgroup of Bayer AG with annual sales of EUR 16.913 billion (2010), is one of the world's leading, innovative companies in the healthcare and medical products industry and is based in Leverkusen, Germany. The company combines the global activities of the Animal Health, Consumer Care, Medical Care and Pharmaceuticals divisions. Bayer HealthCare's aim is to discover and manufacture products that will improve human and animal health worldwide. Bayer HealthCare has a global workforce of 55,700 employees (Dec 31, 2010) and is represented in more than 100 countries. Find more information at www.bayerhealthcare.com.

To learn more about wet Age-related Macular Degeneration (AMD), please visit www.bayerpharma.com/en/AMD

Regeneron Forward-Looking Statement

This news release includes forward-looking statements that involve risks and uncertainties relating to future events and the future performance of Regeneron, and actual events or results may differ materially from these forward-looking statements. These statements concern, and these risks and uncertainties include, among others, the nature, timing, and possible success and therapeutic applications of EYLEA and Regeneron's product candidates and research and clinical programs now underway or planned, the likelihood and timing of possible regulatory approval and commercial launch of Regeneron's late-stage product candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize EYLEA and other products and drug candidates, competing drugs that may be superior to EYLEA and Regeneron's products and drug candidates, uncertainty of market acceptance of EYLEA and Regeneron's products and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including Regeneron's agreements with Sanofi and Bayer HealthCare, to be canceled or terminated without any product success, and risks associated with third party intellectual property and pending or future litigation relating thereto. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission, including its Form 10-K for the year ended December 31, 2010 and Form 10-Q for the quarter ended September 30, 2011. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Bayer Forward-Looking Statement

This release may contain forward-looking statements based on current assumptions and forecasts made by Bayer Group or subgroup management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in Bayer's public reports which are available on the Bayer website at www.bayer.com. The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

Your Contact at Bayer:

Doreen Schroeder, Tel. +49 30 468-11399

E-Mail: doreen.schroeder@bayer.com

Your Investor Relations Contact at Regeneron:

Michael Aberman, MD., Tel. +1 (914) 847-7799

E-Mail: michael.aberman@regeneron.com

Your Media Contact at Regeneron:

Peter Dworkin, Tel. +1 (914) 847-7640

E-Mail: peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

News Provided by Acquire Media

REGENERON

Two Year Results of Phase 3 Studies with EYLEA™ (aflibercept) Injection in wet AMD Show Sustained Improvement in Visual Acuity

December 5, 2011

Patients in the EYLEA 2mg every eight week group achieved visual acuity gains similar to ranibizumab with 5 fewer injections, on average, over two years. Patients who required the most intense therapy received, on average, 1.4 fewer injections in the EYLEA 2mg every eight week group compared to ranibizumab in the second year.

TARRYTOWN, N.Y. and BERLIN, Dec. 5, 2011 /PRNewswire/ -- Regeneron Pharmaceuticals, Inc. (NASDAQ: REGN) and Bayer HealthCare today announced that in an integrated analysis of two parallel Phase 3 studies (VIEW 1 and VIEW 2) in patients with the neovascular form of age-related macular degeneration (wet AMD), patients treated with EYLEA™ (aflibercept) Injection For Intravitreal Injection showed a sustained improvement in visual acuity at 96 weeks versus baseline. The 52-week results (primary analyses) from these studies have previously been reported.

During the first year of the VIEW 1 and VIEW 2 studies, patients were treated with three different dosing regimens of EYLEA, 0.5 milligram (mg) every four weeks, 2mg every four weeks, and 2mg every eight weeks (following three initial monthly injections), compared to ranibizumab 0.5mg every four weeks. The EYLEA 2mg every eight week regimen was recently approved by the U.S. Food and Drug Administration (FDA), based on efficacy (maintenance of vision) that was clinically equivalent at one year to the monthly ranibizumab regimen. In the second year of the studies, patients were treated with the same dose per injection as in the first year and were evaluated monthly to determine need for retreatment. Patients were treated at least every 12 weeks. All year two analyses were considered exploratory.

In an integrated analysis of the VIEW 1 and VIEW 2 studies, the visual acuity gain from baseline in the EYLEA 2mg every eight week group at week 96 was 7.6 letters compared to 8.4 letters at week 52, with an average of 11.2 injections over two years and 4.2 injections during the second year. The visual acuity gain from baseline in the monthly ranibizumab group at week 96 was 7.9 letters compared to 8.7 letters at week 52, with an average of 16.5 injections over two years and 4.7 injections during the second year. The results of each of the VIEW 1 and VIEW 2 studies were consistent with the integrated analysis.

The overall fewer average number of injections in the second year in the EYLEA 2mg every eight week group compared to the ranibizumab group (4.2 versus 4.7) was driven by the fact that fewer patients needed more intense therapy in the EYLEA group and those patients required fewer injections.

The proportion of patients who required frequent injections (six or more) during the second year was lower in the EYLEA 2mg every eight week group compared to the ranibizumab group (15.9% versus 26.5%). In the 25% of patients who required the most intense therapy (the greatest number of injections), patients in the EYLEA 2mg every eight week group required an average of 1.4 fewer injections in the second year compared to the ranibizumab group (6.6 versus 8.0). In the 25% of patients in each group who had the fewest number of injections in the second year, the average number of injections was similar (approximately 3 for both groups, corresponding to the protocol-mandated minimum number of injections).

A generally favorable safety profile was observed for both EYLEA and ranibizumab. The incidence of ocular treatment emergent adverse events was balanced across all four treatment groups in both studies, with the most frequent events associated with the injection procedure, the underlying disease and/or the aging process. The most frequent ocular adverse events (greater than 10% of patients for the overall study population) were conjunctival hemorrhage, eye pain, retinal hemorrhage, and visual acuity reduced. The most frequent serious non-ocular adverse events were typical of those reported in this elderly population who receive intravitreal treatment for wet AMD; the most frequently reported events (greater than 1% of patients for the overall study population) were falls, pneumonia, myocardial infarction and atrial fibrillation. There were no notable differences among the study arms. The incidence of arterial thrombotic events as defined by the "Anti-Platelet Trialists" group criteria was 3.2% of patients for ranibizumab and 3.3% of patients in the combined EYLEA groups.

"These second year results confirm the sustainability of the vision gains achieved by EYLEA with a less than monthly dosing frequency. Importantly, the second year data demonstrated that for patients that needed more anti-VEGF treatment, this was achieved with fewer injections using EYLEA," said George D. Yancopoulos, M.D., Ph.D., Chief Scientific Officer of Regeneron and President of Regeneron Laboratories. "As a reminder, the recommended dose for EYLEA is 2mg every eight weeks following three initial monthly injections, which demonstrated visual acuity gains that were clinically equivalent to monthly ranibizumab. Retinal physicians and their wet AMD patients consider the predictable every eight week dosing regimen for EYLEA as a significant advance that helps overcome the challenges of monthly office visits."

Further results from year two of the studies will be presented at upcoming medical conferences.

About the VIEW Program

The VIEW (VEGF Trap: Investigation of Efficacy and Safety in Wet AMD) program consists of two randomized, double-masked, Phase 3 clinical trials evaluating EYLEA in the treatment of the neovascular form of age-related macular degeneration (wet AMD). The VIEW 1 study, which randomized 1217 patients, was conducted in the United States and Canada by Regeneron. The VIEW 2 study, which randomized 1240 patients, was conducted in Europe, Asia Pacific, Japan, and Latin America by Bayer HealthCare. The study designs are essentially identical. The primary endpoint evaluation was conducted at 52 weeks.

About Wet AMD

Age-related Macular Degeneration (AMD) is a leading cause of acquired blindness. Macular degeneration is diagnosed as either dry (non-exudative) or wet (exudative). In wet AMD, new blood vessels grow beneath the retina and leak blood and fluid. This leakage causes disruption and dysfunction

of the retina creating distortion and/or blind spots in central vision, and it can account for blindness in wet AMD patients. Wet AMD is the leading cause of blindness for people over the age of 65 in the U.S. and Europe.

About EYLEA™ (aflibercept) Injection For Intravitreal Injection

Vascular Endothelial Growth Factor (VEGF) is a naturally occurring protein in the body. Its normal role in a healthy organism is to trigger formation of new blood vessels (angiogenesis) supporting the growth of the body's tissues and organs. However, in certain diseases, such as wet age-related macular degeneration, it is also associated with the growth of abnormal new blood vessels in the eye, which exhibit abnormal increased permeability that leads to edema. Scarring and loss of fine-resolution central vision often results.

EYLEA™ (aflibercept) Injection, known in the scientific literature as VEGF Trap-Eye, is a recombinant fusion protein, consisting of portions of human VEGF receptors 1 and 2 extracellular domains fused to the Fc portion of human IgG1 and formulated as an iso-osmotic solution for intravitreal administration. EYLEA acts as a soluble decoy receptor that binds VEGF-A and placental growth factor (PlGF) and thereby can inhibit the binding and activation of these cognate VEGF receptors.

IMPORTANT PRESCRIBING INFORMATION

In the United States, EYLEA is indicated for the treatment of patients with neovascular age-related macular degeneration (wet AMD).

The recommended dose for EYLEA is 2 mg administered by intravitreal injection every four weeks (monthly) for the first 12 weeks (3 months), followed by 2 mg once every eight weeks (2 months). Although EYLEA may be dosed as frequently as 2 mg every four weeks (monthly), additional efficacy was not demonstrated when EYLEA was dosed every four weeks compared to every eight weeks.

IMPORTANT SAFETY INFORMATION

EYLEA is contraindicated in patients with ocular or periocular infections, active intraocular inflammation, or known hypersensitivity to aflibercept or to any of the excipients in EYLEA.

Intravitreal injections, including those with EYLEA, have been associated with endophthalmitis and retinal detachments. Proper aseptic injection technique must always be used when administering EYLEA. Patients should be instructed to report any symptoms suggestive of endophthalmitis or retinal detachment without delay and should be managed appropriately.

Acute increases in intraocular pressure have been seen within 60 minutes of intravitreal injection, including with EYLEA. Sustained increases in intraocular pressure have also been reported after repeated intravitreal dosing with VEGF inhibitors. Intraocular pressure and the perfusion of the optic nerve head should be monitored and managed appropriately.

There is a potential risk of arterial thromboembolic events (ATEs) following use of intravitreal VEGF inhibitors, including EYLEA, defined as nonfatal stroke, nonfatal myocardial infarction, or vascular death (including deaths of unknown cause). The incidence of ATEs with EYLEA in clinical trials was low (1.8%).

Serious adverse reactions related to the injection procedure have occurred in less than 0.1% of intravitreal injections with EYLEA including endophthalmitis, traumatic cataract, and increased intraocular pressure.

The most common adverse reactions (greater than or equal to 5%) reported in patients receiving EYLEA were conjunctival hemorrhage, eye pain, cataract, vitreous detachment, vitreous floaters, and increased intraocular pressure.

Please see the full Prescribing Information for EYLEA, available online at www.regeneron.com/EYLEA_fpi.pdf.

About the EYLEA™ (aflibercept) Injection Global Collaboration

Regeneron is collaborating with Bayer HealthCare on the global development of EYLEA. Bayer submitted an application for marketing authorization in Europe for wet AMD in June 2011.

Bayer HealthCare will market EYLEA outside the United States, where the companies will share equally the profits from any future sales of EYLEA. Regeneron maintains exclusive rights to EYLEA in the United States.

About Regeneron Pharmaceuticals

Regeneron is a fully integrated biopharmaceutical company that discovers, invents, develops, manufactures, and commercializes medicines for the treatment of serious medical conditions. Regeneron markets two products, ARCALYST® (rilonacept) Injection For Subcutaneous Use and EYLEA™ (aflibercept) Injection. Regeneron also has completed several Phase 3 studies and is conducting an additional Phase 3 clinical trial for the product candidate ZALTRAP® (aflibercept) Concentrate for Intravenous Infusion. Additional therapeutic candidates developed from proprietary Regeneron technologies for creating fully human monoclonal antibodies are in earlier stage development programs in rheumatoid arthritis and other inflammatory conditions, pain, cholesterol reduction, allergic and immune conditions, and cancer. Additional information about Regeneron and recent news releases are available on the Regeneron web site at www.regeneron.com.

About Bayer HealthCare

The Bayer Group is a global enterprise with core competencies in the fields of health care, nutrition and high-tech materials. Bayer HealthCare, a subgroup of Bayer AG with annual sales of more than EUR 16.913 billion (2010), is one of the world's leading, innovative companies in the healthcare and medical products industry and is based in Leverkusen, Germany. The company combines the global activities of the Animal Health, Consumer Care, Medical Care and Pharmaceuticals divisions. Bayer HealthCare's aim is to discover and manufacture products that will improve human and animal health worldwide. Bayer HealthCare has a global workforce of 55,700 employees and is represented in more than 100 countries. Find more information at www.bayerhealthcare.com.

Regeneron Forward-Looking Statement

This news release includes forward-looking statements that involve risks and uncertainties relating to future events and the future performance of Regeneron, and actual events or results may differ materially from these forward-looking statements. These statements concern, and these risks and uncertainties include, among others, the nature, timing, and possible success and therapeutic applications of EYLEA and Regeneron's product candidates and research and clinical programs now underway or planned, the likelihood and timing of possible regulatory approval and commercial launch of Regeneron's late-stage product candidates, determinations by regulatory and administrative governmental authorities which may delay or restrict Regeneron's ability to continue to develop or commercialize EYLEA and other products and drug candidates, competing drugs that may be superior to EYLEA and Regeneron's products and drug candidates, uncertainty of market acceptance of EYLEA and Regeneron's products and drug candidates, unanticipated expenses, the availability and cost of capital, the costs of developing, producing, and selling products, the potential for any license or collaboration agreement, including Regeneron's agreements with Sanofi and Bayer HealthCare, to be canceled or terminated without any product success, and risks associated with third party intellectual property and pending or future litigation relating thereto. A more complete description of these and other material risks can be found in Regeneron's filings with the United States Securities and Exchange Commission, including its Form 10-K for the year ended December 31, 2010 and Form 10-Q for the quarter ended September 30, 2011. Regeneron does not undertake any obligation to update publicly any forward-looking statement, whether as a result of new information, future events, or otherwise, unless required by law.

Bayer Forward-Looking Statements

This release may contain forward-looking statements based on current assumptions and forecasts made by Bayer Group or subgroup management. Various known and unknown risks, uncertainties and other factors could lead to material differences between the actual future results, financial situation, development or performance of the company and the estimates given here. These factors include those discussed in Bayer's public reports which are available on the Bayer website at www.bayer.com. The company assumes no liability whatsoever to update these forward-looking statements or to conform them to future events or developments.

Your Contact at Bayer:

Doreen Schroeder, Tel. +49 30 468-11399

E-Mail: doreen.schroeder@bayer.com

Your Investor Relations Contact at Regeneron:

Michael Aberman, M.D. Tel. +1 (914) 847-7799

E-Mail: michael.aberman@regeneron.com

Your Media Contact at Regeneron:

Peter Dworkin, Tel. +1 (914) 847-7640

E-Mail: peter.dworkin@regeneron.com

SOURCE Regeneron Pharmaceuticals, Inc.

News Provided by Acquire Media

Electronic Patent Application Fee Transmittal

Application Number:	16055847
Filing Date:	06-Aug-2018
Title of Invention:	USE OF A VEGF ANTAGONIST TO TREAT ANGIOGENIC EYE DISORDERS
First Named Inventor/Applicant Name:	George D. Yancopoulos
Filer:	Karl Bozicevic/Kimberly Zuehlke
Attorney Docket Number:	REGN-008CIPCON3

Filed as Large Entity

Filing Fees for Utility under 35 USC 111(a)

Description	Fee Code	Quantity	Amount	Sub-Total in USD(\$)
Basic Filing:				
Pages:				
Claims:				
Miscellaneous-Filing:				
Petition:				
Patent-Appeals-and-Interference:				
Post-Allowance-and-Post-Issuance:				
Extension-of-Time:				

Description	Fee Code	Quantity	Amount	Sub-Total in USD(\$)
Miscellaneous:				
RCE- 1ST REQUEST	1801	1	1300	1300
Total in USD (\$)				1300

Electronic Acknowledgement Receipt

EFS ID:	39874692
Application Number:	16055847
International Application Number:	
Confirmation Number:	3451
Title of Invention:	USE OF A VEGF ANTAGONIST TO TREAT ANGIOGENIC EYE DISORDERS
First Named Inventor/Applicant Name:	George D. Yancopoulos
Customer Number:	96387
Filer:	Karl Bozicevic/Kimberly Zuehlke
Filer Authorized By:	Karl Bozicevic
Attorney Docket Number:	REGN-008CIPCON3
Receipt Date:	30-JUN-2020
Filing Date:	06-AUG-2018
Time Stamp:	17:27:34
Application Type:	Utility under 35 USC 111(a)

Payment information:

Submitted with Payment	yes
Payment Type	CARD
Payment was successfully received in RAM	\$1300
RAM confirmation Number	E20206TH28125500
Deposit Account	
Authorized User	

The Director of the USPTO is hereby authorized to charge indicated fees and credit any overpayment as follows:

--	--	--	--	--	--

File Listing:

Document Number	Document Description	File Name	File Size(Bytes)/ Message Digest	Multi Part /.zip	Pages (if appl.)
1	Request for Continued Examination (RCE)	0725US04_2020-06-30_RCE_Transmittal_REGN-008CIPCON3.pdf	1352026 f8e8a41049f3ab7e908acc03fc3f9867c4d48497	no	3

Warnings:

Information:

2		0725US04_2020-06-30_Pre_Amend_REGN-008CIPCON3.pdf	35110 0a622d5db8ca446fc0854e9776c1a389205331fc	yes	4
---	--	---	---	-----	---

Multipart Description/PDF files in .zip description

Document Description	Start	End
Preliminary Amendment	1	1
Claims	2	2
Applicant Arguments/Remarks Made in an Amendment	3	4

Warnings:

Information:

3	Transmittal Letter	0725US04_2020-06-30_Supp_IDS_trans_REGN-008CIPCON3.pdf	50738 7ecc4b925a86e8289634c9990d7346430b192a84	no	2
---	--------------------	--	---	----	---

Warnings:

Information:

4	Information Disclosure Statement (IDS) Form (SB08)	0725US04_2020-06-30_Supp_IDS_SB08A_REGN-008CIPCON3.pdf	36091 9d7fa266519b1d2366fc68d67c6e10d2af7d5b7b	no	2
---	--	--	---	----	---

Warnings:

Information:

This is not an USPTO supplied IDS fillable form

5	Non Patent Literature	BayerNews_20080928_0448_e n.pdf	127802	no	5
			6d6478579d4b400aca428b8b3a577d4030 031a53		
Warnings:					
Information:					
6	Non Patent Literature	REGN_Press_Release_Mar_27_ 2007.pdf	1360202	no	2
			986d7edd7cdd3899ced99c0603f9540b36f 32357		
Warnings:					
Information:					
7	Non Patent Literature	REGN_Press_Release_May_9_2 007.pdf	1348579	no	2
			4aea4c46411524118a56a126ad8880d5628 dc9d1		
Warnings:					
Information:					
8	Non Patent Literature	REGN_Press_Release_Aug_1_2 007.pdf	2839120	no	5
			9d80051b30879bcf5163b4639fd18a64ca6 b2bb6		
Warnings:					
Information:					
9	Non Patent Literature	REGN_Press_Release_Oct_1_20 07.pdf	1457215	no	3
			f6338613f2aabca358d8b8103489fb0f6782 6e36		
Warnings:					
Information:					
10	Non Patent Literature	REGN_Press_Release_Feb_27_ 2008.pdf	3789068	no	6
			585e6714db309793c0e0f1d5dd74666c361 5a379		
Warnings:					
Information:					
11	Non Patent Literature	REGN_Press_Release_Apr_28_2 008.pdf	24012	no	2
			21d51e0e9eb9e2b27059dd2eaf647e2220e f6395		
Warnings:					
Information:					

12	Non Patent Literature	REGN_Press_Release_Aug_19_2008.pdf	23293 5d13932b21799c081aff58b6fa4fc0a8e6b4b825	no	2
Warnings:					
Information:					
13	Non Patent Literature	REGN_Press_Release_Feb_26_2009.pdf	3449787 c5f91432559098f3ad1fb2789fde8c0fd873e91e	no	5
Warnings:					
Information:					
14	Non Patent Literature	REGN_Press_Release_Jul_23_2009.pdf	1070186 5d38c7318355bbb4bd99263ad4d5c178ce9c0eb36	no	2
Warnings:					
Information:					
15	Non Patent Literature	REGN_Press_Release_Nov_19_2010.pdf	350695 b241755a11b1083bf843f27bbf27849ece1d60a6	no	1
Warnings:					
Information:					
16	Non Patent Literature	REGN_Press_Release_Jan_18_2011.pdf	1765835 834d1807fb38ebe24b62e6859d6c78251db8b218	no	3
Warnings:					
Information:					
17	Non Patent Literature	REGN_Press_Release_Feb_9_2011.pdf	1371797 aab4ca6426678222de6ccf81389495a08d533ce5	no	2
Warnings:					
Information:					
18	Non Patent Literature	REGN_Press_Release_Aug_2_2007.pdf	106077 a6c28091d1985f689be1eccf605d9cbff04cb0eb	no	2
Warnings:					
Information:					

19	Non Patent Literature	REGN_Press_Release_Apr_30_2009.pdf	107443 9b7019f9ac83ae628b74e4cf5d36232c548ea7e9	no	2
Warnings:					
Information:					
20	Non Patent Literature	REGN_Press_Release_Feb_22_2011.pdf	110830 1c15ab6a0cda244ad867d79f5f839abc7251f8963	no	6
Warnings:					
Information:					
21	Non Patent Literature	REGN_Press_Release_Apr_8_2011.pdf	1480611 a13f642ce8b29401bd55e0d61db1de29d171f413	no	2
Warnings:					
Information:					
22	Non Patent Literature	REGN_Press_Release_Apr_18_2011.pdf	17501 a824d1d3a227ce42763b0da65601593f396c765b	no	2
Warnings:					
Information:					
23	Non Patent Literature	REGN_Press_Release_Jun_7_2011.pdf	1463628 41e6c89cfa55d29dff0736ab0e82cebfbec32dea	no	2
Warnings:					
Information:					
24	Non Patent Literature	REGN_Press_Release_Jun_17_2011.pdf	19498 afef5a7b39b651100449db5b31c908297f673434	no	2
Warnings:					
Information:					
25	Non Patent Literature	REGN_Press_Release_Aug_17_2011.pdf	581399 8e96e29aa8c022ae65c25ab110c11f9144593e27	no	2
Warnings:					
Information:					

26	Non Patent Literature	REGN_Press_Release_Nov_18_2011.pdf	19527 8864153c02303fe0f6969e05c4cf59b74582f9f3	no	2
Warnings:					
Information:					
27	Non Patent Literature	REGN_Press_Release_Nov_28_2011.pdf	27840 42379aa75f28abacc9ffca7fea7a138a3fcd374f	no	3
Warnings:					
Information:					
28	Non Patent Literature	REGN_Press_Release_Dec_5_2011.pdf	32377 cc45ca0792fb752a70ed96149f7f07e1638ad13f	no	3
Warnings:					
Information:					
29	Fee Worksheet (SB06)	fee-info.pdf	30737 4afbeb931079935cf90907dae4d52d3e2f65aaa8	no	2
Warnings:					
Information:					
Total Files Size (in bytes):				24449024	
<p>This Acknowledgement Receipt evidences receipt on the noted date by the USPTO of the indicated documents, characterized by the applicant, and including page counts, where applicable. It serves as evidence of receipt similar to a Post Card, as described in MPEP 503.</p> <p><u>New Applications Under 35 U.S.C. 111</u> If a new application is being filed and the application includes the necessary components for a filing date (see 37 CFR 1.53(b)-(d) and MPEP 506), a Filing Receipt (37 CFR 1.54) will be issued in due course and the date shown on this Acknowledgement Receipt will establish the filing date of the application.</p> <p><u>National Stage of an International Application under 35 U.S.C. 371</u> If a timely submission to enter the national stage of an international application is compliant with the conditions of 35 U.S.C. 371 and other applicable requirements a Form PCT/DO/EO/903 indicating acceptance of the application as a national stage submission under 35 U.S.C. 371 will be issued in addition to the Filing Receipt, in due course.</p> <p><u>New International Application Filed with the USPTO as a Receiving Office</u> If a new international application is being filed and the international application includes the necessary components for an international filing date (see PCT Article 11 and MPEP 1810), a Notification of the International Application Number and of the International Filing Date (Form PCT/RO/105) will be issued in due course, subject to prescriptions concerning national security, and the date shown on this Acknowledgement Receipt will establish the international filing date of the application.</p>					

To: docket@bozpat.com,,
From: PAIR_eOfficeAction@uspto.gov
Cc: PAIR_eOfficeAction@uspto.gov
Subject: Private PAIR Correspondence Notification for Customer Number 96387

Jul 01, 2020 03:44:23 AM

Dear PAIR Customer:

Regeneron - Bozicevic, Field & Francis
201 REDWOOD SHORES PARKWAY
SUITE 200
REDWOOD CITY, CA 94065
UNITED STATES

The following USPTO patent application(s) associated with your Customer Number, 96387 , have new outgoing correspondence. This correspondence is now available for viewing in Private PAIR.

The official date of notification of the outgoing correspondence will be indicated on the form PTOL-90 accompanying the correspondence.

Disclaimer:

The list of documents shown below is provided as a courtesy and is not part of the official file wrapper. The content of the images shown in PAIR is the official record.

Application	Document	Mailroom Date	Attorney Docket No.
16055847	NOA	07/01/2020	REGN-008CIPCON3
	1449	07/01/2020	REGN-008CIPCON3

To view your correspondence online or update your email addresses, please visit us anytime at <https://sportal.uspto.gov/secure/myportal/privatepair>.

If you have any questions, please email the Electronic Business Center (EBC) at EBC@uspto.gov with 'e-Office Action' on the subject line or call 1-866-217-9197 during the following hours:

Monday - Friday 6:00 a.m. to 12:00 a.m.

Thank you for prompt attention to this notice,

UNITED STATES PATENT AND TRADEMARK OFFICE
PATENT APPLICATION INFORMATION RETRIEVAL SYSTEM

Under the Paperwork Reduction Act of 1995, no persons are required to respond to a collection of information unless it displays a valid OMB control number.

PATENT APPLICATION FEE DETERMINATION RECORD Substitute for Form PTO-875	Application or Docket Number 16/055,847	Filing Date 08/06/2018	<input type="checkbox"/> To be Mailed
---	--	---------------------------	---------------------------------------

ENTITY: LARGE SMALL MICRO

APPLICATION AS FILED - PART I

FOR	(Column 1) NUMBER FILED	(Column 2) NUMBER EXTRA	RATE (\$)	FEE (\$)
<input type="checkbox"/> BASIC FEE (37 CFR 1.16(a), (b), or (c))	N/A	N/A	N/A	
<input type="checkbox"/> SEARCH FEE (37 CFR 1.16(k), (i), or (m))	N/A	N/A	N/A	
<input type="checkbox"/> EXAMINATION FEE (37 CFR 1.16(o), (p), or (q))	N/A	N/A	N/A	
TOTAL CLAIMS (37 CFR 1.16(i))	minus 20 = *		x \$ 100 =	
INDEPENDENT CLAIMS (37 CFR 1.16(h))	minus 3 = *		x \$ 460 =	
<input type="checkbox"/> APPLICATION SIZE FEE (37 CFR 1.16(s))	If the specification and drawings exceed 100 sheets of paper, the application size fee due is \$310 (\$155 for small entity) for each additional 50 sheets or fraction thereof. See 35 U.S.C. 41(a)(1)(G) and 37 CFR 1.16(s).			
<input type="checkbox"/> MULTIPLE DEPENDENT CLAIM PRESENT (37 CFR 1.16(j))				
* If the difference in column 1 is less than zero, enter "0" in column 2.			TOTAL	

APPLICATION AS AMENDED - PART II

		(Column 1)		(Column 2)	(Column 3)	RATE (\$)	ADDITIONAL FEE (\$)
AMENDMENT	06/30/2020	CLAIMS REMAINING AFTER AMENDMENT		HIGHEST NUMBER PREVIOUSLY PAID FOR	PRESENT EXTRA		
	Total (37 CFR 1.16(i))	* 3	Minus	** 20	= 0	x \$ 100 =	0
	Independent (37 CFR 1.16(h))	* 1	Minus	*** 3	= 0	x \$ 460 =	0
<input type="checkbox"/> Application Size Fee (37 CFR 1.16(s))							
<input type="checkbox"/> FIRST PRESENTATION OF MULTIPLE DEPENDENT CLAIM (37 CFR 1.16(j))							
						TOTAL ADD'L FEE	0
AMENDMENT		CLAIMS REMAINING AFTER AMENDMENT		HIGHEST NUMBER PREVIOUSLY PAID FOR	PRESENT EXTRA		
	Total (37 CFR 1.16(i))	*	Minus	**	=	x \$ 0 =	
	Independent (37 CFR 1.16(h))	*	Minus	***	=	x \$ 0 =	
<input type="checkbox"/> Application Size Fee (37 CFR 1.16(s))							
<input type="checkbox"/> FIRST PRESENTATION OF MULTIPLE DEPENDENT CLAIM (37 CFR 1.16(j))							
						TOTAL ADD'L FEE	
* If the entry in column 1 is less than the entry in column 2, write "0" in column 3.						LIE	
** If the "Highest Number Previously Paid For" IN THIS SPACE is less than 20, enter "20".						/CHRISTINE V MOORE/	
*** If the "Highest Number Previously Paid For" IN THIS SPACE is less than 3, enter "3".							
The "Highest Number Previously Paid For" (Total or Independent) is the highest number found in the appropriate box in column 1.							

This collection of information is required by 37 CFR 1.16. The information is required to obtain or retain a benefit by the public which is to file (and by the USPTO to process) an application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This collection is estimated to take 12 minutes to complete, including gathering, preparing, and submitting the completed application form to the USPTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden, should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, P.O. Box 1450, Alexandria, VA 22313-1450. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. **SEND TO: Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450.**

If you need assistance in completing the form, call 1-800-PTO-9199 and select option 2.

INFORMATION DISCLOSURE STATEMENT BY APPLICANT			Application Number	16/055,847	
			Filing Date	August 6, 2018	
			First Named Inventor	George D. Yancopoulos	
			Art Unit	1647	
			Examiner Name	Jon McClelland Lockard	
Sheet	1	of	5	Attorney Docket Number	REGN-008CIPCON3

U.S. PATENT DOCUMENTS						
Examiner Initial*	Cite No.	Patent Number		Issue Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear
		Number-Kind Code (if known)				
	1					

U.S. PATENT APPLICATION PUBLICATIONS						
Examiner Initial*	Cite No.	Publication Number		Publication Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear
		Number-Kind Code (if known)				
	1	2019/0290725		2019-09-26	Vitti et al.	

FOREIGN PATENT DOCUMENTS							
Examiner Initial*	Cite No.	Foreign Document Number		Publication Date YYYY-MM-DD	Name of Patentee or Applicant of Cited Document	Pages, Columns, Lines, Where Relevant Passages or Relevant Figures Appear	T
		Country Code-Number-Kind Code (if known)					
	1	WO 2004/106378 A2		2004-12-09	Regeneron Pharmaceuticals, Inc.		
	2	WO 2005/000895 A2		2005-01-05	Regeneron Pharmaceuticals, Inc.		

NON PATENT LITERATURE DOCUMENTS			
Examiner Initials*	Cite No.	Include name of the author (in CAPITAL LETTERS), title of the article (when appropriate), title of the item (book, magazine, journal, serial, symposium, catalog, etc.), date, page(s), volume-issue number(s), publisher, city and/or country where published.	T
	1	BENZ et al. "CLEAR-IT-2: Interim Results Of The Phase II, Randomized, Controlled Dose- and Interval-ranging Study Of Repeated Intravitreal VEGF Trap Administration In Patients With Neovascular Age-related Macular Degeneration (AMD)" ARVO Annual Meeting Abstract (May 2007)	
	2	DO et al. "Results of a Phase 1 Study of Intravitreal VEGF Trap in Subjects with Diabetic Macular Edema: The CLEAR-IT DME Study" ARVO Annual Meeting Abstract (May 2007)	
	3	DO et al. "VEGF Trap-Eye Vision-specific Quality of Life through 52 Weeks in Patients with Neovascular AMD in CLEAR-IT 2: A Phase 2 Clinical Trial" ARVO Annual Meeting Abstract (April 2009)	
	4	HALLER et al., "VEGF Trap-Eye In CRVO: Primary Endpoint Results of the Phase 3 COPERNICUS Study" ARVO Annual Meeting Abstract (April 2011)	
	5	HEIER et al., "CLEAR-IT 2: Phase 2, Randomized Controlled Dose and Interval-Ranging Study of Intravitreal VEGF Trap Eye in Patients with Neovascular Age-Related Macular Degeneration: Predictive Factors for Visual Acuity" ARVO Annual Meeting Abstract (April 2009)	
	6	HEIER et al., "The 1-year Results of CLEAR-IT 2, a Phase 2 Study of Vascular Endothelial Growth Factor Trap-Eye Dosed As-needed After 12-week Fixed Dosing" Ophthalmology 2011;118:1098-1106 (June 2011)	
	7	HEIER et al., "The 1-year Results of CLEAR-IT 2, a Phase 2 Study of Vascular Endothelial Growth Factor Trap-Eye Dosed As-needed After 12-week Fixed Dosing: Erratum" Ophthalmology 2011;118:1700 (September 2011)	
	8	Information from ClinicalTrials.gov archive History of Changes for Study: NCT00320775 "Safety and Tolerability of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration" 70 pages, Latest version submitted June 8, 2011 on ClinicalTrials.gov (NCT00320775 2006-2011)	

Examiner Signature		Date Considered	
--------------------	--	-----------------	--

*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

INFORMATION DISCLOSURE STATEMENT BY APPLICANT			Application Number	16/055,847	
			Filing Date	August 6, 2018	
			First Named Inventor	George D. Yancopoulos	
			Art Unit	1647	
			Examiner Name	Jon McClelland Lockard	
Sheet	2	of	5	Attorney Docket Number	REGN-008CIPCON3

NON PATENT LITERATURE DOCUMENTS			
Examiner Initials*	Cite No.	T	
	9		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00320775 "Safety and Tolerability of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration" 10 pages, Latest version submitted March 16, 2015 on ClinicalTrials.gov (NCT00320775_2015)
	10		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00320788 "Safety and Efficacy of Repeated Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Wet Age-Related Macular Degeneration (AMD)" 71 pages, Latest version submitted December 1, 2011 on ClinicalTrials.gov (NCT00320788_2006-2011)
	11		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00320788 "Safety and Efficacy of Repeated Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Wet Age-Related Macular Degeneration (AMD)" 31 pages, Latest version submitted January 27, 2012 on ClinicalTrials.gov (NCT00320788_2012)
	12		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00320814 "Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema" 30 pages, Latest version submitted June 8, 2011 on ClinicalTrials.gov (NCT00320814_2006-2011)
	13		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00509795 "Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)" 318 pages, Latest version submitted December 1, 2011 on ClinicalTrials.gov (NCT00509795_2007-2011)
	14		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00509795 "Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)" 200 pages, Latest version submitted December 20, 2012 on ClinicalTrials.gov (NCT00509795_2012)
	15		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00527423 "Randomized, Single-Masked, Long-Term, Safety and Tolerability Study of VEGF Trap-Eye in AMD" 64 pages, Latest version submitted November 1, 2011 on ClinicalTrials.gov (NCT00527423_2007-2011)
	16		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00527423 "Randomized, Single-Masked, Long-Term, Safety and Tolerability Study of VEGF Trap-Eye in AMD" 42 pages, Latest version submitted June 10, 2013 on ClinicalTrials.gov (NCT00527423_2012-2013)
	17		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00637377 "Vascular Endothelial Growth Factor (VEGF) Trap-Eye: Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration (AMD) (VIEW 2)" 667 pages, Latest version submitted December 16, 2011 on ClinicalTrials.gov (NCT00637377_2008-2011)
	18		Information from ClinicalTrials.gov archive History of Changes for Study: NCT00637377 "Vascular Endothelial Growth Factor (VEGF) Trap-Eye: Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration (AMD) (VIEW 2)" 289 pages, Latest version submitted November 28, 2014 on ClinicalTrials.gov (NCT00637377_2012-2014)

Examiner Signature		Date Considered	
--------------------	--	-----------------	--

*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

INFORMATION DISCLOSURE STATEMENT BY APPLICANT			Application Number	16/055,847	
			Filing Date	August 6, 2018	
			First Named Inventor	George D. Yancopoulos	
			Art Unit	1647	
			Examiner Name	Jon McClelland Lockard	
Sheet	3	of	5	Attorney Docket Number	REGN-008CIPCON3

NON PATENT LITERATURE DOCUMENTS

Examiner Initials*	Cite No.	T
	19	Information from ClinicalTrials.gov archive History of Changes for Study: NCT00789477 "DME And VEGF Trap-Eye [Intravitreal Aflibercept Injection (IAI;EYLEA®;BAY86-5321)] INvestigation of Clinical Impact (DA VINCI)" 135 pages, Latest version submitted May 2, 2011 on ClinicalTrials.gov (NCT00789477_2008-2011)
	20	Information from ClinicalTrials.gov archive History of Changes for Study: NCT00789477 "DME And VEGF Trap-Eye [Intravitreal Aflibercept Injection (IAI;EYLEA®;BAY86-5321)] INvestigation of Clinical Impact (DA VINCI)" 53 pages, Latest version submitted August 28, 2014 on ClinicalTrials.gov (NCT00789477_2013-2014)
	21	Information from ClinicalTrials.gov archive History of Changes for Study: NCT00943072 "Vascular Endothelial Growth Factor (VEGF) Trap-Eye: Investigation of Efficacy and Safety in Central Retinal Vein Occlusion (CRVO)" 98 pages, Latest version submitted May 9, 2011 on ClinicalTrials.gov (NCT00943072_2009-2011)
	22	Information from ClinicalTrials.gov archive History of Changes for Study: NCT00943072 "Vascular Endothelial Growth Factor (VEGF) Trap-Eye: Investigation of Efficacy and Safety in Central Retinal Vein Occlusion (CRVO)" 64 pages, Latest version submitted April 16, 2013 on ClinicalTrials.gov (NCT00943072_2012-2013)
	23	MAJOR et al., "DA VINCI: DME and VEGF Trap-Eye: Investigation of Clinical Impact: Phase 2 Study in Patients with Diabetic Macular Edema (DME)" ARVO Annual Meeting Abstract (April 2010)
	24	NGUYEN et al., "Randomized, Double-masked, Active-controlled Phase 3 Trial of the Efficacy and Safety of Intravitreal VEGF Trap-Eye in Wet AMD: One-year Results of the VIEW 1 Study" ARVO Annual Meeting Abstract (April 2011)
	25	NGUYEN et al., "Results of a Phase I, Dose-Escalation, Safety, Tolerability, and Bioactivity Study of Intravitreal VEGF Trap in Patients with Neovascular Age-Related Macular Degeneration" ARVO Annual Meeting Abstract (May 2006)
	26	Regeneron SEC Form 10-K (February 27, 2008)
	27	Regeneron SEC Form 10-K (February 26, 2009)
	28	Regeneron SEC Form 10-K (February 17, 2011)
	29	Regeneron SEC Form 10-Q (May 8, 2006)
	30	Regeneron SEC Form 10-Q (August 8, 2006)
	31	Regeneron SEC Form 10-Q (November 6, 2006)
	32	Regeneron SEC Form 10-Q (May 4, 2007)
	33	Regeneron SEC Form 10-Q (August 3, 2007)
	34	Regeneron SEC Form 10-Q (April 30, 2009)
	35	Regeneron SEC Form 10-Q (November 3, 2009)
	36	Regeneron SEC Form 10-Q (April 29, 2010)
	37	Regeneron SEC Form 10-Q (July 28, 2010)
	38	Regeneron SEC Form 10-Q (October 28, 2010)
	39	Regeneron SEC Form 10-Q (May 3, 2011)
	40	Regeneron SEC Form 10-Q (July 28, 2011)
	41	Regeneron SEC Form 10-Q (October 27, 2011)

Examiner Signature		Date Considered	
--------------------	--	-----------------	--

*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

INFORMATION DISCLOSURE STATEMENT BY APPLICANT			Application Number	16/055,847	
			Filing Date	August 6, 2018	
			First Named Inventor	George D. Yancopoulos	
			Art Unit	1647	
			Examiner Name	Jon McClelland Lockard	
Sheet	4	of	5	Attorney Docket Number	REGN-008CIPCON3

NON PATENT LITERATURE DOCUMENTS			
Examiner Initials*	Cite No.	Include name of the author (in CAPITAL LETTERS), title of the article (when appropriate), title of the item (book, magazine, journal, serial, symposium, catalog, etc.), date, page(s), volume-issue number(s), publisher, city and/or country where published.	T
	42	Regeneron SEC Form 8-K Exhibit: "Press Release of Regeneron Pharmaceuticals, Inc. dated May 1, 2006" (May 2, 2006)	
	43	Regeneron SEC Form 8-K Exhibit: "Press Release of Regeneron Pharmaceuticals, Inc. dated May 3, 2006" (May 5, 2006)	
	44	Regeneron SEC Form 8-K Exhibit: "Slides presented at the Company's 2006 Annual Meeting of Shareholders held on June 9, 2006" (June 9, 2006)	
	45	Regeneron SEC Form 8-K Exhibit: "Press Release dated May 2, 2007" (May 3, 2007)	
	46	Regeneron SEC Form 8-K Exhibit: "Overheads for presentation at Regeneron's Annual Meeting of Shareholders to be held on June 8, 2007" (June 8, 2007)	
	47	Regeneron SEC Form 8-K Exhibit: "Press Release dated October 1, 2007" (October 1, 2007)	
	48	Regeneron SEC Form 8-K Exhibit: "Press Release dated November 6, 2007" (November 6, 2007)	
	49	Regeneron SEC Form 8-K Exhibit: "Press Release dated May 1, 2008" (May 2, 2008)	
	50	Regeneron SEC Form 8-K Exhibit: "Press Release dated November 4, 2008" (November 4, 2008)	
	51	Regeneron SEC Form 8-K Exhibit: "99(a) Slides that Regeneron Pharmaceuticals, Inc. intends to use in conjunction with meetings with investors at the J.P. Morgan 27th Annual Healthcare Conference in San Francisco on January 12-15, 2009." (January 9, 2009)	
	52	Regeneron SEC Form 8-K Exhibit: "Press Release dated April 30, 2009" (May 1, 2009)	
	53	Regeneron SEC Form 8-K Exhibit: "Press Release dated November 3, 2009." (November 4, 2009)	
	54	Regeneron SEC Form 8-K Exhibit: "Press Release Reporting Positive Results for VEGF Trap-Eye in Phase 3 Study in Central Retinal Vein Occlusion (CRVO) and in Phase 2 Study in Diabetic Macular Edema (DME) dated December 20, 2010." (December 20, 2010)	
	55	Regeneron SEC Form 8-K Exhibit: "Press Release dated February 17, 2011" (February 18, 2011)	
	56	Regeneron SEC Form 8-K Exhibit: "Press Release Reporting Positive Results for VEGF Trap-Eye in Second Phase 3 Study in Central Retinal Vein Occlusion, dated April 27, 2011" (April 27, 2011)	
	57	Regeneron SEC Form 8-K Exhibit: "Press Release dated May 3, 2011." (May 3, 2011)	
	58	Regeneron SEC Form 8-K Exhibit: "Press Release, dated June 17, 2011, Announcing that EYLEA™ (aflibercept ophthalmic solution) Received Unanimous Recommendation for Approval for Treatment of Wet AMD from FDA Advisory Committee." (June 21, 2011)	
	59	Regeneron SEC Form 8-K Exhibit: "Presentation entitled VEGF Trap-Eye in CRVO: 1-year Results of the Phase 3 COPERNICUS Study" (August 22, 2011)	
	60	Regeneron SEC Form 8-K Exhibit: "Press Release Announcing FDA Approval of EYLEA™ (aflibercept) Injection for the Treatment of Wet Age-Related Macular Degeneration, dated November 18, 2011" (November 21, 2011)	

Examiner Signature		Date Considered	
--------------------	--	-----------------	--

*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

INFORMATION DISCLOSURE STATEMENT BY APPLICANT			Application Number	16/055,847	
			Filing Date	August 6, 2018	
			First Named Inventor	George D. Yancopoulos	
			Art Unit	1647	
			Examiner Name	Jon McClelland Lockard	
Sheet	5	of	5	Attorney Docket Number	REGN-008CIPCON3

NON PATENT LITERATURE DOCUMENTS			
Examiner Initials*	Cite No.	Include name of the author (in CAPITAL LETTERS), title of the article (when appropriate), title of the item (book, magazine, journal, serial, symposium, catalog, etc.), date, page(s), volume-issue number(s), publisher, city and/or country where published.	T
	61	Regeneron Pharmaceuticals Inc., "CLEAR-IT-2: Interim Results Of The Phase II, Randomized, Controlled Dose-and Interval-ranging Study Of Repeated Intravitreal VEGF Trap Administration In Patients With Neovascular Age-related Macular Degeneration (AMD)" poster presented at the 2007 Association for Research in Vision and Ophthalmology meeting in Ft. Lauderdale, Florida (May 2007)	
	62	Regeneron Pharmaceuticals Inc., "An Exploratory Study of the Safety, Tolerability and Biological Effect of a Single Intravitreal Administration of VEGF Trap in Patients with Diabetic Macular Edema" poster presented at the 2007 Association for Research in Vision and Ophthalmology meeting in Ft. Lauderdale, Florida (May 2007)	
	63	Regeneron Pharmaceuticals Inc., "Optical Coherence Tomography Outcomes of a Phase 1, Dose-Escalation, Safety, Tolerability, and Bioactivity Study of Intravitreal VEGF Trap in Patients with Neovascular Age-Related Macular Degeneration: The CLEAR-IT 1 Study" poster presented at the 2007 Association for Research in Vision and Ophthalmology meeting in Ft. Lauderdale, Florida (May 2007)	
	64	Regeneron Pharmaceuticals Inc., "VIEW 1 Vascular Endothelial Growth Factor (VEGF) Trap-Eye 1-Year Results: Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration (AMD) " presented at Bascom Palmer Eye Institute's Angiogenesis, Exudation and Degeneration 2011 meeting in Miami, Florida (February 12, 2011)	
	65	Regeneron Pharmaceuticals Inc., "VIEW 2 Vascular Endothelial Growth Factor (VEGF) Trap-Eye 1-Year Results: Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration (AMD) " presented at Bascom Palmer Eye Institute's Angiogenesis, Exudation and Degeneration 2011 meeting in Miami, Florida (February 12, 2011)	
	66	Regeneron Pharmaceuticals Inc., "VEGF Trap-Eye CLEAR-IT 2 Final Primary Endpoint Results" presented at the 2007 Retina Society Conference in Boston, Massachusetts (September 30, 2007)	
	67	Regeneron 2008 Annual Report	
	68	Regeneron 2009 Annual Report and 10-K	
	69	Regeneron 2010 Annual Report and 10-K	
	70	RUDGE et al. "Clinical Development of VEGF Trap" In: Figg W.D., Folkman J. (eds) Angiogenesis (2008)	
	71	SCHMIDT-ERFURTH et al. "Primary Results of an International Phase III Study Using Intravitreal VEGF Trap-Eye Compared to Ranibizumab in Patients with Wet AMD (VIEW 2)" ARVO Annual Meeting Abstract (April 2011)	
	72	SLAKTER et al., "Influence of Baseline Angiographic Classification on Outcomes in the CLEAR-IT 2 Phase 2 Study of Intravitreal VEGF Trap-Eye in Neovascular Age-Related Macular Degeneration" ARVO Annual Meeting Abstract (April 2010)	
	73	SLAKTER et al., "A Phase 2, Randomized, Controlled Dose-and Interval-Ranging Study of Intravitreal VEGF Trap-Eye in Patients with Neovascular Age-Related Macular Degeneration: Optical Coherence Tomography (OCT) and Fluorescein Angiography (FA) Outcomes at 1 Year" ARVO Annual Meeting Abstract (April 2009)	

Examiner Signature		Date Considered	
--------------------	--	-----------------	--

*EXAMINER: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property
Organization
International Bureau



(43) International Publication Date
9 December 2004 (09.12.2004)

PCT

(10) International Publication Number
WO 2004/106378 A2

- (51) International Patent Classification⁷: **C07K 16/24**, A61K 31/7088, C07K 19/00, 14/71
- (74) Agent: **VALETA, Gregg**; Regeneron Pharmaceuticals, Inc., 777 Old Saw Mill River Road, Tarrytown, NY 10591 (US).
- (21) International Application Number: PCT/US2004/012540
- (81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.
- (22) International Filing Date: 23 April 2004 (23.04.2004)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
60/473,734 28 May 2003 (28.05.2003) US
60/492,865 6 August 2003 (06.08.2003) US
- (84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).
- (71) Applicants (for all designated States except US): **REGENERON PHARMACEUTICALS, INC.** [US/US]; 777 Old Saw Mill River Road, Tarrytown, NY 10591 (US). **THE SCHEPENS EYE RESEARCH INSTITUTE** [US/US]; 20 Staniford Street, Boston, MA 021114 (US).
- (72) Inventors; and
- (75) Inventors/Applicants (for US only): **WIEGAND, Stanley** [US/US]; 15 Fox Run Road, Croton on Hudson, NY 10520 (US). **CAO, Jingtai** [CN/US]; 308 N. Greeley Avenue, Chappaqua, NY 10514 (US). **CURSIEFEN, Claus** [DE/DE]; Nordliche Stadtmauerstr. 14, 91054 Erlangen (DE).
- Published:**
— without international search report and to be republished upon receipt of that report
- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.*



WO 2004/106378 A2

(54) Title: METHOD OF TREATING CORNEAL TRANSPLANT REJECTION

(57) Abstract: Methods of preventing, reducing, or treating corneal transplant rejection to improve transplant survival in a subject in need thereof comprising administering an agent capable of blocking or inhibiting vascular endothelial growth factor (VEGF) are provided. The methods are useful for inhibiting or preventing corneal transplant rejection in a human subject who is the recipient of a transplanted cornea.

METHOD OF TREATING CORNEAL TRANSPLANT REJECTION

BACKGROUND

Field of the Invention

[0001] The field of the invention is related to methods of using VEGF antagonists to reduce, prevent, or treat corneal transplant rejection, thus improving long-term transplant survival.

Description of Related Art

[0002] It has previously been reported that topical application of an anti-VEGF neutralizing antibody suppresses acute allograft rejection in a rat corneal transplant model (Yatoh et al. (1998) Transplantation 66(11):1519-24). As the leading cause of human corneal transplant failure is transplant rejection, there is a need for a therapeutic for use in preventing corneal transplant rejection in humans who receive a corneal transplant.

BRIEF SUMMARY OF THE INVENTION

[0003] The invention is based in part on the finding that administration of an agent capable of blocking or inhibiting vascular endothelial growth factor (VEGF) prevents corneal transplant rejection. The experiments, described below, conducted in an animal model of corneal transplantation show that long-term transplant survival is promoted by blocking VEGF-mediated activity.

[0004] In a first aspect, the invention features a method of improving transplant survival in a subject in need thereof, comprising administering to the subject an agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity, such that transplant survival is improved.

[0005] In specific embodiments, the agent capable of blocking, inhibiting, or ameliorating VEGF-mediated activity is a VEGF antagonist. The VEGF antagonist may be a polypeptide, an antibody, a small molecule, or a nucleic acid. More specifically, the VEGF antagonist includes a VEGF trap selected from the group consisting of acetylated Flt-1(1-3)-Fc, Flt-1(1-3_{R->N})-Fc, Flt-1(1-3_{ΔB})-Fc, Flt-1(2-3_{ΔB})-Fc, Flt-1(2-3)-Fc, Flt-1D2-VEGFR3D3-FcΔC1(a), Flt-1D2-Flk-1D3-FcΔC1(a), and VEGFR1R2-FcΔC1(a). In a specific and preferred embodiment, the VEGF trap is VEGFR1R2-FcΔC1(a) (also termed VEGF trap_{R1R2}) having the nucleotide sequence set forth in SEQ ID NO: 1 and the amino acid sequence set forth in SEQ ID NO: 2. The invention encompasses the use of a VEGF trap that is at least 90%, 95%, 98%, or at least 99%

homologous with the nucleotide sequence set forth in SEQ ID NO: 1 and/or the amino acid sequence set forth in SEQ ID NO:2.

[0006] In other embodiments, the agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity is a nucleic acid-based antagonist capable of interfering with the expression of VEGF. A specific example of this embodiment is one in which the nucleic acid-based antagonist is an aptamer, an siRNA, or an antisense molecule.

[0007] Administration of the agent may be by any method known in the art, including subcutaneous, intramuscular, intradermal, intraperitoneal, intravenous, intranasal, oral, or topical routes of administration. Preferable, administration to the subject in need of the agent is topical administration to the eye or subconjunctival administration. Administration may occur prior to or following corneal transplantation, preferably following surgery. Administration may also include a second agent, such as an immunosuppressive agent.

[0008] The subject to be treated is preferably a human subject who has or will receive a corneal transplant.

[0009] In a related second aspect, the invention features the use of an agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity in the preparation of a medicament for improving transplant survival in a mammalian subject.

[0010] In a third aspect, the invention features a method of preventing corneal transplant rejection in a subject in need thereof, comprising administering to the subject an agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity, such that corneal transplant rejection is prevented.

[0011] In a related fourth aspect, the invention features the use of an agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity in the preparation of a medicament for the treatment of corneal transplant rejection in a mammalian subject.

[0012] In a fifth aspect, the invention features a method of reducing the incidence of corneal transplant rejection in a subject in need thereof, comprising administering to the subject an agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity, such that the incidence of corneal transplant rejection is reduced.

[0013] In a related sixth aspect, the invention features the use of an agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity in the preparation of a medicament for reducing the incidence of corneal transplant rejection in a mammalian subject receiving a corneal transplant.

[0014] In a seventh aspect, the invention features a pharmaceutical composition comprising a VEGF antagonist, for example the VEGF trap VEGFR1R2-Fc Δ C1(a), in a pharmaceutically

acceptable carrier. Such pharmaceutical compositions may be liquid, gel, ointment, salve, slow release formulations or other formulations suitable for ophthalmic administration.

[0015] In an eighth aspect, the invention features an article of manufacture comprising packaging materials and a pharmaceutical agent contained within the packaging materials, wherein the pharmaceutical agent comprises at least one VEGF-specific fusion protein of the invention, and the packaging material comprises a label or package insert which indicates that the VEGF-specific fusion protein can be used for the treatment or prevention of corneal transplant rejection.

[0016] Other objects and advantages will become apparent from a review of the ensuing detailed description.

DETAILED DESCRIPTION

[0017] Before the present methods are described, it is to be understood that this invention is not limited to particular methods, and experimental conditions described, as such methods and conditions may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

[0018] As used in this specification and the appended claims, the singular forms “a”, “an”, and “the” include plural references unless the context clearly dictates otherwise. Thus for example, a reference to “a method” includes one or more methods, and/or steps of the type described herein and/or which will become apparent to those persons skilled in the art upon reading this disclosure and so forth.

[0019] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are now described. All publications mentioned herein are incorporated herein by reference in their entirety.

General Description

[0020] Experiments were undertaken to evaluate occurrence and time course of hem- and lymphangiogenesis after normal-risk corneal transplantation and to test whether pharmacologic strategies inhibiting both processes improve long-term graft survival. As described in the experimental section below, normal-risk allogeneic (C57BL/6 to BALB/c) and syngeneic (BALB/c to BALB/c) corneal transplantations were performed and occurrence and time course

of hem- and lymphangiogenesis after keratoplasty was observed using double immunofluorescence of corneal flatmounts (with CD31 as panendothelial and LYVE-1 as lymphatic vascular endothelial specific marker). A molecular trap designed to eliminate VEGF-A (“VEGF Trap_{R1R2}”; 12.5 mg/kg) was tested for its ability to inhibit both processes after keratoplasty and to promote long-term graft survival (intraperitoneal injections on the day of surgery and 3, 7, and 14 days later). The results show that no blood or lymph vessels were detectable immediately after normal-risk transplantation in either donor or host cornea, but hem- and lymphangiogenesis were clearly visible at day 3 after transplantation. Both vessel types reached donor tissue at one week after allo- and similarly after syngeneic grafting. Early postoperative trapping of VEGF-A significantly reduced both hem- and lymphangiogenesis and significantly improved long-term graft survival (78% versus 40%; $p < 0.05$). There is concurrent, VEGF-A-dependent hem- and lymphangiogenesis after normal-risk keratoplasty within the preoperatively avascular recipient bed. Inhibition of hem- and lymphangiogenesis (which mediate the efferent and afferent arms of an immune response) after normal-risk corneal transplantation improves long-term graft survival, establishing that early postoperative hem- and lymphangiogenesis are risk factors for graft rejection even in low-risk eyes.

Definitions

[0021] By the term “therapeutically effective dose” is meant a dose that produces the desired effect for which it is administered. The exact dose will depend on the purpose of the treatment, and will be ascertainable by one skilled in the art using known techniques (see, for example, Lloyd (1999) *The Art, Science and Technology of Pharmaceutical Compounding*).

[0022] By the term “blocker”, “inhibitor”, or “antagonist” is meant a substance that retards or prevents a chemical or physiological reaction or response. Common blockers or inhibitors include but are not limited to antisense molecules, antibodies, antagonists and their derivatives. More specifically, an example of a VEGF blocker or inhibitor is a VEGF receptor-based antagonist including, for example, an anti-VEGF antibody, or a VEGF trap such as VEGFR1R2-Fc Δ C1(a) (SEQ ID NOs:1-2). For a complete description of VEGF-receptor based antagonists including VEGFR1R2-Fc Δ C1(a), see PCT publication WO/00/75319, the contents of which is incorporated in its entirety herein by reference.

[0023] A “small molecule” is defined herein to have a molecular weight below about 500 Daltons, and may include chemical as well as peptide molecules.

VEGF Antagonists

[0024] In one aspect of the invention, VEGF-mediated activity is blocked or inhibited by the use

of VEGF receptor-based blockers of VEGF-mediated activity. A non-limiting example of a VEGF receptor-based blocker includes, but is not limited to, VEGFR1R2-Fc Δ C1(a). Other suitable receptor-based blockers include acetylated Flt-1(1-3)-Fc, Flt-1(1-3_{R→N})-Fc, Flt-1(1-3_{ΔB})-Fc, Flt-1(2-3_{ΔB})-Fc, Flt-1(2-3)-Fc, Flt-1D2-VEGFR3D3-Fc Δ C1(a), Flt-1D2-Flk-1D3-Fc Δ C1(a). For a complete description of these and other VEGF-receptor-based blockers, including pegylated receptor-based blockers, see PCT Publication No. WO/00/75319, the contents of which is incorporated in its entirety herein by reference.

[0025] In addition to the VEGF receptor-based blockers described in PCT Publication No. WO/00/75319, variants and derivatives of such VEGF receptor-based blockers are also contemplated by the invention. The sequence of the variants or derivatives may differ by a change which is one or more additions, insertions, deletions and/or substitutions of one or more nucleotides of the sequence set forth in SEQ ID NO:1. Changes to a nucleotide sequence may result in an amino acid change at the protein level, or not, as determined by the genetic code. Thus, nucleic acid according to the present invention may include a sequence different from the sequence shown in SEQ ID NO:1, yet encode a polypeptide with the same amino acid sequence as SEQ ID NO: 2. On the other hand, the encoded polypeptide may comprise an amino acid sequence which differs by one or more amino acid residues from the amino acid sequence shown in SEQ ID NO:2. Nucleic acid encoding a polypeptide which is an amino acid sequence variant or derivative of the sequence shown in SEQ ID NO:2 is further provided by the present invention. Nucleic acid encoding such a polypeptide may show at the nucleotide sequence and/or encoded amino acid level greater than about 90%, 95%, 98%, or 99% homology with the coding sequence shown in SEQ ID NO:1 and/or the amino acid sequence shown in SEQ ID NO:2. For amino acid "homology", this may be understood to be similarity (according to the established principles of amino acid similarity, e.g. as determined using the algorithm GAP (Genetics Computer Group, Madison, Wis.)) or identity. GAP uses the Needleman and Wunsch algorithm to align two complete sequences that maximizes the number of matches and minimizes the number of gaps. Generally, the default parameters are used, with a gap creation penalty=12 and gap extension penalty=4.

[0026] Individual components of the VEGF-specific fusion proteins of the invention may be constructed by molecular biological methods known to the art with the instructions provided by the instant specification. These components are selected from a first cellular receptor protein, such as, for example, VEGFR1; a second cellular receptor protein, such as, for example, VEGFR2; a multimerizing component, such as an Fc.

[0027] Specific embodiments of the VEGF-specific fusion proteins useful in the methods of the invention comprise a multimerizing component which allows the fusion proteins to associate,

e.g., as multimers, preferably dimers. Preferably, the multimerizing component comprises an immunoglobulin derived domain. Suitable multimerizing components are sequences encoding an immunoglobulin heavy chain hinge region (Takahashi et al. 1982 Cell 29:671-679); immunoglobulin gene sequences, and portions thereof.

[0028] The nucleic acid constructs encoding the fusion proteins useful in the methods of the invention are inserted into an expression vector by methods known to the art, wherein the nucleic acid molecule is operatively linked to an expression control sequence. Host-vector systems for the production of proteins comprising an expression vector introduced into a host cell suitable for expression of the protein are known in the art. The suitable host cell may be a bacterial cell such as *E. coli*, a yeast cell, such as *Pichia pastoris*, an insect cell, such as *Spodoptera frugiperda*, or a mammalian cell, such as a COS, CHO, 293, BHK or NS0 cell.

Antisense Nucleic Acids

[0029] In one aspect of the invention, VEGF-mediated activity is blocked or inhibited by the use of VEGF antisense nucleic acids. The present invention provides the therapeutic or prophylactic use of nucleic acids comprising at least six nucleotides that are antisense to a gene or cDNA encoding VEGF or a portion thereof. As used herein, a VEGF "antisense" nucleic acid refers to a nucleic acid capable of hybridizing by virtue of some sequence complementarity to a portion of an RNA (preferably mRNA) encoding VEGF. The antisense nucleic acid may be complementary to a coding and/or noncoding region of an mRNA encoding VEGF. Such antisense nucleic acids have utility as compounds that prevent VEGF expression, and can be used in the treatment or prevention of corneal transplant rejection. The antisense nucleic acids of the invention are double-stranded or single-stranded oligonucleotides, RNA or DNA or a modification or derivative thereof, and can be directly administered to a cell or produced intracellularly by transcription of exogenous, introduced sequences.

[0028] The VEGF antisense nucleic acids are of at least six nucleotides and are preferably oligonucleotides ranging from 6 to about 50 oligonucleotides. In specific aspects, the oligonucleotide is at least 10 nucleotides, at least 15 nucleotides, at least 100 nucleotides, or at least 200 nucleotides. The oligonucleotides can be DNA or RNA or chimeric mixtures or derivatives or modified versions thereof and can be single-stranded or double-stranded. In addition, the antisense molecules may be polymers that are nucleic acid mimics, such as PNA, morpholino oligos, and LNA. Other types of antisense molecules include short double-stranded RNAs, known as siRNAs, and short hairpin RNAs, and long dsRNA (>50 bp but usually ≥ 500 bp).

Short interfering RNAs

[0029] In another embodiment, VEGF-mediated activity is blocked by blocking VEGF expression. One method for inhibiting VEGF expression is the use of short interfering RNA (siRNA) through RNA interference (RNAi) or post-transcriptional gene silencing (PTGS) (see, for example, Ketting et al. (2001) *Genes Develop.* 15:2654-2659). siRNA molecules can target homologous mRNA molecules for destruction by cleaving the mRNA molecule within the region spanned by the siRNA molecule. Accordingly, siRNAs capable of targeting and cleaving homologous VEGF mRNA are useful for treating, reducing or preventing corneal transplant rejection.

Inhibitory Ribozymes

[0030] In aspect of the invention, corneal transplant rejection may be treated or prevented in a subject suffering from such disease by decreasing the level of VEGF activity by using ribozyme molecules designed to catalytically cleave gene mRNA transcripts encoding VEGF, preventing translation of target gene mRNA and, therefore, expression of the gene product.

[0031] Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by an endonucleolytic cleavage event. The composition of ribozyme molecules must include one or more sequences complementary to the target gene mRNA, and must include the well known catalytic sequence responsible for mRNA cleavage. For this sequence, see, e.g., U.S. Patent No. 5,093,246. While ribozymes that cleave mRNA at site-specific recognition sequences can be used to destroy mRNAs encoding VEGF, the use of hammerhead ribozymes is preferred. Hammerhead ribozymes cleave mRNAs at locations dictated by flanking regions that form complementary base pairs with the target mRNA. The sole requirement is that the target mRNA has the following sequence of two bases: 5'-UG-3'. The construction and production of hammerhead ribozymes is well known in the art. The ribozymes of the present invention also include RNA endoribonucleases (hereinafter "Cech-type ribozymes") such as the one that occurs naturally in *Tetrahymena thermophila* (known as the IVS, or L-19 IVS RNA). The Cech-type ribozymes have an eight base pair active site that hybridizes to a target RNA sequence where after cleavage of the target RNA takes place. The invention encompasses those Cech-type ribozymes that target eight base-pair active site sequences that are present in the gene encoding VEGF.

Generation of Antibodies to VEGF Proteins

[0032] In another aspect of the invention, the invention may be practiced with an anti-VEGF

antibody or antibody fragment capable of binding and blocking VEGF activity. Anti-VEGF antibodies are disclosed, for example, in US Patent No. 6,121,230, herein specifically incorporated by reference. The term "antibody" as used herein refers to a polypeptide comprising a framework region from an immunoglobulin gene or fragments thereof that specifically binds and recognizes an antigen. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon, and mu constant regions, as well as the myriad immunoglobulin variable region genes. Light chains are classified as either kappa or lambda. Heavy chains are classified as gamma, mu, alpha, delta, or epsilon, which in turn define the immunoglobulin classes, IgG, IgM, IgA, IgD, and IgE, respectively. Within each IgG class, there are different isotypes (eg. IgG₁, IgG₂, etc.). Typically, the antigen-binding region of an antibody will be the most critical in determining specificity and affinity of binding.

[0033] Antibodies exist as intact immunoglobulins, or as a number of well-characterized fragments produced by digestion with various peptidases. For example, pepsin digests an antibody below the disulfide linkages in the hinge region to produce F(ab)₂, a dimer of Fab which itself is a light chain joined to V_H-C_{H1} by a disulfide bond. The F(ab)₂ may be reduced under mild conditions to break the disulfide linkage in the hinge region, thereby converting the F(ab)₂ dimer into an Fab' monomer. The Fab' monomer is essentially Fab with part of the hinge region. While various antibody fragments are defined in terms of the digestion of an intact antibody, one of skill will appreciate that such fragments may be synthesized *de novo* either chemically or by using recombinant DNA methodology. Thus, the terms antibody, as used herein, also includes antibody fragments either produced by the modification of whole antibodies, or those synthesized *de novo* using recombinant DNA methodologies (e.g., single chain Fv)(scFv) or those identified using phase display libraries (see, for example, McCafferty et al. (1990) Nature 348:552-554).

[0034] Methods for preparing antibodies are known to the art. See, for example, Kohler & Milstein (1975) Nature 256:495-497; Harlow & Lane (1988) Antibodies: a Laboratory Manual, Cold Spring Harbor Lab., Cold Spring Harbor, NY). The genes encoding the heavy and light chains of an antibody of interest can be cloned from a cell, e.g., the genes encoding a monoclonal antibody can be cloned from a hybridoma and used to produce a recombinant monoclonal antibody. Gene libraries encoding heavy and light chains of monoclonal antibodies can also be made from hybridoma or plasma cells. Random combinations of the heavy and light chain gene products generate a large pool of antibodies with different antigenic specificity. Techniques for the production of single chain antibodies or recombinant antibodies (US 4,946,778; US 4,816,567) can be adapted to produce antibodies used in the fusion proteins and methods of the instant invention. Also, transgenic mice, or other organisms such as other mammals, may be

used to express human or humanized antibodies. Alternatively, phage display technology can be used to identify antibodies and heteromeric Fab fragments that specifically bind to selected antigens.

Antibody Screening and Selection

[0035] Screening and selection of preferred antibodies can be conducted by a variety of methods known to the art. Initial screening for the presence of monoclonal antibodies specific to a target antigen may be conducted through the use of ELISA-based methods, for example. A secondary screen is preferably conducted to identify and select a desired monoclonal antibody for use in construction of the multi-specific fusion proteins of the invention. Secondary screening may be conducted with any suitable method known to the art. One preferred method, termed "Biosensor Modification-Assisted Profiling" ("BiaMAP") is described in co-pending USSN 60/423,017 filed 01 Nov 2002, herein specifically incorporated by reference in its entirety. BiaMAP allows rapid identification of hybridoma clones producing monoclonal antibodies with desired characteristics. More specifically, monoclonal antibodies are sorted into distinct epitope-related groups based on evaluation of antibody:antigen interactions.

Treatment Population

[0036] A suitable subject for treatment by the method of the invention is a human who has received or will receive a corneal transplant. Corneal transplantation is the oldest, most successful and most commonly performed tissue transplantation, with nearly 40,000 transplantations a year alone in the US. When corneal grafts are placed into an avascular recipient bed (so-called normal-risk keratoplasty), 2-year graft survival rates approach 90% under cover of topical steroids, even without HLA-matching. This very successful outcome is attributed to corneal immune privilege, i.e. the phenomenon of suppressed corneal inflammation induced by an array of endogenous mechanisms downregulating alloimmune and inflammatory responses in the cornea and its bed. These mechanisms include the lack of both afferent lymphatic and efferent blood vessels in the normal-risk recipient cornea, lack of MHC II⁺ antigen presenting cells (APCs), FASL-expression on corneal epithelium and endothelium, and the anterior chamber associated immune privilege (ACAID) directed at graft antigens etc. (Streilein et al. (1999) *Transplant Proc.* 31:1472-1475).

[0037] In contrast, survival rates of cornea grafts placed into vascularized, not immune-privileged recipient beds (so called high-risk keratoplasty) drop significantly to below 50% (even with local and systemic immune suppression). Pre-existing corneal stromal blood vessels have been identified as strong risk factors for immune rejection after corneal transplantation, both in

the clinical setting as well as in the well-defined mouse model of corneal transplantation (Sano et al. (1995) Invest. Ophthalmol. Vis. Sci. 36:2176-85). Recently, in addition to blood vessels, biomicroscopically undetectable lymphatic vessels have been found in association with blood vessels in vascularized high-risk human corneas (Cursiefen et al. (2003) Cornea. 22:273-81) and it is likely that corneal lymphatic vessels enable effective access of donor and host APCs and antigenic material to regional lymph nodes where accelerated sensitisation to graft antigens occurs (Liu et al. (2002) J. Exp. Med. 195:259-68) even in the normal-risk setting (with a preoperatively avascular recipient bed), where mild corneal hemangiogenesis develops after keratoplasty. Outgrowth of new blood vessels from the limbal arcade towards the graft can be observed within the first postoperative year in about 50% of patients undergoing normal-risk keratoplasty, and in 10% of patients these new blood vessels even reach the interface or invade donor tissue (Cursiefen et al. (2001) Graefes Arch. clin. Exp. Ophthalmol. 39:514-21) at corneal suture sites, and then proceed centrally.

Methods of Administration

[0038] The invention provides methods of treatment comprising administering to a subject an effective amount of an agent of the invention. In a preferred aspect, the agent is substantially purified (*e.g.*, substantially free from substances that limit its effect or produce undesired side-effects). The subject is preferably an animal, *e.g.*, such as cows, pigs, horses, chickens, cats, dogs, etc., and is preferably a mammal, and most preferably human.

[0039] Various delivery systems are known and can be used to administer an active agent of the invention, *e.g.*, delivery systems suitable for topical administration, preferably topical administration directly to the eye, or subconjunctival administration, as well as other delivery systems such as those that utilize encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the compound, receptor-mediated endocytosis (see, *e.g.*, Wu and Wu, 1987, J. Biol. Chem. 262:4429-4432), construction of a nucleic acid as part of a retroviral or other vector, etc. Methods of introduction are preferably topical or subconjunctival, but may be enteral or parenteral including but are not limited to intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, and oral routes. The active agents may be administered by any convenient route, for example by absorption through epithelial (*e.g.* topical administration to the eye) or mucocutaneous linings (*e.g.*, oral mucosa, intestinal mucosa, etc.) or infusion or bolus injection, and may be administered together with other biologically active agents. Administration can be systemic or local. Administration can be acute or chronic (*e.g.* daily, weekly, monthly, etc.) or in combination or alteration with other agents. Pulmonary

administration can also be employed, *e.g.*, by use of an inhaler or nebulizer, and formulation with an aerosolizing agent.

[0040] In another embodiment, the active agent can be delivered in a vesicle, in particular a liposome (see Langer (1990) *Science* 249:1527-1533). In yet another embodiment, the active agent can be delivered in a controlled release system. In one embodiment, a pump may be used (see Langer (1990) *supra*). In another embodiment, polymeric materials can be used (see Howard et al. (1989) *J. Neurosurg.* 71:105). In another embodiment where the active agent of the invention is a nucleic acid encoding a protein, the nucleic acid can be administered *in vivo* to promote expression of its encoded protein, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, *e.g.*, by use of a retroviral vector (see, for example, U.S. Patent No. 4,980,286), or by direct injection, or by use of microparticle bombardment (*e.g.*, a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors or transfecting agents, or by administering it in linkage to a homeobox-like peptide which is known to enter the nucleus (see *e.g.*, Joliot et al., 1991, *Proc. Natl. Acad. Sci. USA* 88:1864-1868), etc. Alternatively, a nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination.

[0041] In a specific embodiment, it may be desirable to administer the pharmaceutical compositions of the invention locally to the area in need of treatment; this may be achieved, for example, and not by way of limitation, by topical administration, subconjunctival administration, local infusion during surgery, *e.g.*, by injection, by means of a catheter, or by means of an implant, said implant being of a porous, non-porous, or gelatinous material, including membranes, such as sialastic membranes, fibers, or commercial skin substitutes.

Cellular Transfection and Gene Therapy

[0042] The present invention encompasses the use of nucleic acids encoding the VEGF-specific fusion proteins of the invention for transfection of cells *in vitro* and *in vivo*. These nucleic acids can be inserted into any of a number of well-known vectors for transfection of target cells and organisms. The nucleic acids are transfected into cells *ex vivo* and *in vivo*, through the interaction of the vector and the target cell. Reintroduction of transfected cells may be accomplished by any method known to the art, including re-implantation of encapsulated cells. The compositions are administered (*e.g.*, by injection into a muscle) to a subject in an amount sufficient to elicit a therapeutic response. An amount adequate to accomplish this is defined as "a therapeutically effective dose or amount."

[0043] In another aspect, the invention provides a method of treating or preventing corneal transplant rejection in a human comprising transfecting a cell with a nucleic acid encoding a

VEGF-specific fusion protein of the invention, wherein the nucleic acid comprises an inducible promoter operably linked to the nucleic acid encoding the VEGF-specific fusion protein. For gene therapy procedures in the treatment or prevention of human disease, see for example, Van Brunt (1998) *Biotechnology* 6:1149-1154.

Pharmaceutical Compositions

[0044] Pharmaceutical compositions useful in the practice of the method of the invention include a therapeutically effective amount of an active agent, and a pharmaceutically acceptable carrier. The term “pharmaceutically acceptable” means approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, and more particularly, in humans. The term “carrier” refers to a diluent, adjuvant, excipient, or vehicle with which the therapeutic is administered. Such pharmaceutical carriers can be sterile liquids, such as water and oils, including those of petroleum, animal, vegetable or synthetic origin, such as peanut oil, soybean oil, mineral oil, sesame oil and the like. Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol and the like. The composition, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. These compositions can take the form of solutions, suspensions, emulsion, tablets, pills, capsules, powders, sustained-release formulations and the like. The composition can be formulated as a suppository, with traditional binders and carriers such as triglycerides. Oral formulation can include standard carriers such as pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, etc. Examples of suitable pharmaceutical carriers are described in “Remington’s Pharmaceutical Sciences” by E.W. Martin.

[0045] In a preferred embodiment, the composition is formulated in accordance with routine procedures as a pharmaceutical composition adapted for intravenous, subcutaneous, or intramuscular administration to human beings. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lidocaine to ease pain at the site of the injection. Where the composition is to be administered by infusion, it can be dispensed with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the composition is administered by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

[0046] The active agents of the invention can be formulated as neutral or salt forms. Pharmaceutically acceptable salts include those formed with free amino groups such as those

derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, etc., and those formed with free carboxyl groups such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2-ethylamino ethanol, histidine, procaine, etc.

[0047] The amount of the active agent of the invention that will be effective in the treatment or prevention of corneal transplant rejection can be determined by standard clinical techniques based on the present description. In addition, *in vitro* assays may optionally be employed to help identify optimal dosage ranges. The precise dose to be employed in the formulation will also depend on the route of administration, and the seriousness of the condition, and should be decided according to the judgment of the practitioner and each subject's circumstances. However, suitable dosage ranges for intravenous administration are generally about 50-5000 micrograms of active compound per kilogram body weight. Suitable dosage ranges for intranasal administration are generally about 0.01 pg/kg body weight to 1 mg/kg body weight. Effective doses may be extrapolated from dose-response curves derived from *in vitro* or animal model test systems.

[0048] For systemic administration, a therapeutically effective dose can be estimated initially from *in vitro* assays. For example, a dose can be formulated in animal models to achieve a circulating concentration range that includes the IC₅₀ as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Initial dosages can also be estimated from *in vivo* data, e.g., animal models, using techniques that are well known in the art. One having ordinary skill in the art could readily optimize administration to humans based on animal data.

[0049] Dosage amount and interval may be adjusted individually to provide plasma levels of the compounds that are sufficient to maintain therapeutic effect. One having skill in the art will be able to optimize therapeutically effective local dosages without undue experimentation.

[0050] The amount of compound administered will, of course, be dependent on the subject being treated, on the subject's weight, the severity of the affliction, the manner of administration, and the judgment of the prescribing physician. The therapy may be repeated intermittently while symptoms are detectable or even when they are not detectable. The therapy may be provided alone or in combination with other drugs.

Combination Therapies

[0051] In numerous embodiments, the VEGF blockers of the present invention may be administered in combination with one or more additional compounds or therapies or medical procedures. For example, suitable therapeutic agents for use in combination, either alternating or simultaneously, with the VEGF blockers may include topically administered immunosuppressive

agents such as corticosteroids, dexamethasone, cyclosporin A, or anti-metabolic agents or systemically administered immunosuppressive agents such as corticosteroids, dexamethasone, cyclosporin A, FK506, or anti-metabolic agents, as well as other agents effective to treat, reduce, or prevent corneal transplant rejection (see Barker, NH, *et al.*, (2000) Clin Exp Opthal 28:357-360). Other suitable therapeutic agents for use in combination, either alternating or simultaneously, with the VEGF blockers of the subject invention may include blockers that can block other VEGF family members such as VEGF-C and VEGF-D.

Kits

[0052] The invention also provides an article of manufacturing comprising packaging material and a pharmaceutical agent contained within the packaging material, wherein the pharmaceutical agent comprises at least one VEGF-specific fusion protein of the invention and wherein the packaging material comprises a label or package insert which indicates that the VEGF-specific fusion protein can be used for treating corneal transplant rejection.

[0053] Other features of the invention will become apparent in the course of the following descriptions of exemplary embodiments which are given for illustration of the invention and are not intended to be limiting thereof.

EXAMPLES

[0054] The following example is put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the methods and compositions of the invention, and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers used (e.g., amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees Centigrade, and pressure is at or near atmospheric.

Example 1: Inhibition of corneal lymphangiogenesis and angiogenesis after low-risk keratoplasty using VEGFR1R2-Fc Δ C1(a).

[0055] Mice and anesthesia. Six to 8 weeks old male C57BL/6 mice were used as donors and same-aged male BALB/c mice (Taconic, Germantown, NY) as recipients in the mouse model of normal-risk keratoplasty (Sonoda et al. (1992) Transplantation 54:694-704). For syngeneic transplantations, 6-8 weeks old male BALB/c mice were used both as donors as well as recipients. For the dose response studies, 8 weeks old male C57BL/6 mice were used. All animals were treated in accordance with the ARVO Statement for the Use of Animals in

Ophthalmic and Vision Research. Mice were anesthetized using a mixture of ketamine and xylazine (120 mg/kg body weight and 20 mg/kg body weight respectively).

[0056] Dose response of VEGF Trap_{R1R2}. Five different doses of VEGF-Trap_{R1R2} (SEQ ID NO:2) were tested in mice that received three interrupted intrastromal sutures (10-0 nylon, 50- μ m-diameter, Sharpoint, Surgical Specialties Corporation, Reading, PA). Gentamicine and ophthalmic ointment were applied immediately after surgery. Following surgery (day 0), mice received a single subcutaneous injection of VEGF Trap_{R1R2} (25 mg/kg, 12.5 mg/kg, 6.25 mg/kg, 2.5 mg/kg or 0.5 mg) or human Fc (12.5 mg/kg; control). Corneas were harvested on day 9 after suture placement, following an intravenous administration of an endothelial-specific fluorescein-conjugated lectin (*Lycopersicon esculentum*, Vector Laboratories, Burlingame, CA). The isolated corneas were flat-mounted on glass slides, and images of lectin-labeled vessels were captured using a Spot RT Digital camera (Diagnostic Instrument, Inc. Sterling Heights, MI) attached to a Nikon Microphot-FXA microscope (Nikon Inc. Garden City, NY). Scion Image 1.62c (Scion Corporation, Frederick, MD) was used to quantify the extent of corneal neovascularization.

[0057] Corneal transplantation in mice. Orthotopic corneal allografting in the mouse model of normal-risk keratoplasty was performed as described previously (Sonoda et al. (1992) *supra*). Donor corneas were excised by trephination using a 2.0 mm bore and cut with a curved vannis scissor. Until grafting, corneal tissue was placed in chilled phosphate-buffered saline. Recipients were anesthetized and the graft bed was prepared by trephining a 1.5 mm site in the central cornea of the right eye and discarding the excised cornea. The donor cornea was immediately applied to the bed and secured in place with 8 interrupted sutures (11-0 nylon, 70 μ m diameter needles, Arosurgical, Newport Beach, CA). Antibiotic ointment (Oxymycin, Pharmafair, Hauppauge, NY) was placed on the corneal surface and the eyelids sutured with 8-0 suture (Sharpoint, Reading, PA). Recipients of grafts in which bleeding developed in the immediate postoperative period were discarded from further evaluation. All grafted eyes were examined after 72 hours, and grafts with technical difficulties (hyphema, cataract, infection, loss of anterior chamber) were excluded from further consideration. Tarsorrhaphy and corneal sutures were removed after 7 days and grafts were then examined at least twice a week until week 8 post transplantation by slit-lamp microscopy and scored for opacity. The survival experiment was performed twice and comprised 10 and 12 mice per experiment in both groups, respectively. Clinical scores of corneal grafts for opacity were as follows: 0= clear; +1= minimal, superficial (nonstromal) opacity; pupil margin and iris vessels readily visible through the cornea; +2= minimal, deep (stroma) opacity; pupil margins and iris vessels visible; +3= moderate stromal opacity; only pupil margin visible; +4= intense stromal opacity; only a portion of pupil margin

visible; +5= maximum stromal opacity; anterior chamber not visible. Grafts with opacity scores of +2 or greater after 2 weeks were considered to have been rejected. Syngeneic transplantations were performed and evaluated in a similar manner.

[0058] Immunohistochemistry and morphometry of angiogenesis and lymphangiogenesis in the cornea. Briefly, corneal flat mounts were rinsed in PBS, fixed in acetone, rinsed in PBS, blocked in 2% bovine serum albumin, stained with FITC-conjugated CD31/PECAM-1 overnight (Santa Cruz Biotechnology, Santa Cruz, CA; 1:100), washed, blocked, stained with LYVE-1 (1:500; a lymphatic endothelium specific hyaluronic acid receptor (Cursiefen et al. (2002) Invest. Ophthalmol. Vis. Sci. 43:2127-35) washed, blocked, and stained with Cy3 (1:100; Jackson ImmunoResearch Laboratories, West Grove, PA) and analyzed using a Zeiss Axiophot microscope. Digital pictures of the flat mounts were taken using Spot Image Analysis system. Then the area covered by CD31⁺⁺⁺/LYVE-1⁻ blood vessels and CD31⁺/LYVE-1⁺⁺⁺ lymph vessels was measured morphometrically on these flat-mounts using NIH Image software. The total corneal area was outlined using the innermost vessel of the limbal arcade as the border. The total area of blood versus lymphatic neovascularization was then normalized to the total corneal area and the percentage of the cornea covered by each vessel type calculated.

[0059] Neutralization of VEGF-A using VEGF Trap_{R1R2}. The VEGF trap_{R1R2} (Regeneron Pharmaceuticals Inc, Tarrytown, NY (Holash et al. (2002) Proc. Natl. Acad. Sci. USA 99:11393-8, herein specifically incorporated by reference in its entirety) was used in the transplant survival experiment at a concentration of 12.5 mg/kg intraperitoneally (i.p.) at time of surgery (CHO hVEGFR1 [Ig domain 2] R2 [Ig domain 3]-Fc), and 3, 7, and 14 days after surgery. Human Fc-fragment given i.p. at same concentration and times was used in the control mice (sCHO h Fc).

[0060] Statistical analysis. Statistical significance was analyzed by Mann-Whitney's test. Differences were considered significant at $P < 0.05$. Each experiment was performed at least twice with similar results. Graphs were drawn using Graph Pad Prism, Version 3.02.

[0061] Results. Dose response of angiogenesis inhibition by VEGF Trap_{R1R2}. VEGF-Trap_{R1R2} at doses of either 25 mg/kg or 12.5 mg/kg completely inhibited suture-induced inflammatory corneal neovascularization. In contrast, doses of 6.25mg/kg and 2.5mg/kg produced ~50% and ~20% inhibition of corneal neovascularization, respectively, while the lowest dose tested, 0.5 mg/kg, had a negligible effect (<5% inhibition). Therefore, for subsequent experiments a dose of 12.5 mg/kg VEGF Trap_{R1R2} was chosen.

[0062] Rapid and parallel onset of hemangiogenesis and lymphangiogenesis after normal-risk allogeneic corneal transplantation. To determine whether the mild and temporary hemangiogenesis occurring after normal-risk keratoplasty is accompanied by lymphatic vessel outgrowth from the limbus into the normally alymphatic cornea, we studied the time course of

ingrowth of both vessel types at days 0, 3, 7, 14, 21, and 28 *after* allogeneic keratoplasty (only accepted grafts). Immediately *after* surgery, blood and lymphatic vessels were not detectable either in the host or in donor tissue using biomicroscopy and immunohistochemistry on corneal flat mounts. But, at day 3 after allografting, both methods revealed new blood vessels growing into the cornea already 1/3 to halfway towards the graft interface. By day 7 these vessels had usually reached the donor tissue, but they rarely invaded the donor tissue itself. Analyzing flatmounts stained with LYVE-1 as a lymphatic vessel specific marker showed that CD31⁺⁺⁺/LYVE-1⁻ blood vessels were regularly accompanied by LYVE-1⁺⁺⁺/CD31⁺ lymphatic vessels. Both vessel types reached the interface simultaneously at day 7. Thereafter, coincident with suture removal, both vessel types started to regress (if no immune rejection occurred; data not shown).

[0063] No difference in postkeratoplasty hem- and lymphangiogenesis between syngeneic and allogeneic corneal transplantation. To determine whether the simultaneous induction of hem- and lymphangiogenesis *after* normal-risk keratoplasty is primarily an effect of the surgical trauma, suturing and wound healing processes or secondary to early immunological rejection reactions, we compared speed and extent of both hem- and lymphangiogenesis occurring *after* keratoplasty between allogeneic (C57BL/6 into BALB/c) and syngeneic grafts (BALB/c into BALB/c) at day 3, 7, 14, 21, 28 after transplantation. In both groups, blood and lymphatic vessels grew out after keratoplasty and by day 3 reached about 1/3 to $\frac{1}{2}$ of the limbus-interface distance. At day 7 after syngeneic and allogeneic grafting both vessel types had reached the interface, before they started to regress thereafter. Furthermore, there was no significant difference in the hem- and lymphvascularized area, comparing syngeneic and allogeneic grafts at 3 days (allogeneic: hemvascularized area [HA] 25.2 \pm 4.1% and lymphvascularized area [LA] 22.2 \pm 9.4% versus syngeneic HA: 23 \pm 2.7% and LA 19.4 \pm 7.2%) and 7 days (allogeneic HA: 53.8 \pm 11.2% and LA: 37.9 \pm 6.2% versus syngeneic HA: 55.9 \pm 8.2% and LA: 38 \pm 22.7%) after surgery (n=8 mice per group per timepoint).

[0064] Neutralization of VEGF-A after normal-risk keratoplasty inhibits postoperative hemangiogenesis and lymphangiogenesis. Mice received either intraperitoneal injections of VEGF Trap_{R1R2} (12.5 mg/kg) at surgery and 3 days later, or in the controls the Fc-protein in the same dosage. At day 3 and 7 after surgery, the extent of hem- and lymphangiogenesis was compared between these two groups (n=6 mice per group per timepoint). At day 3 and day 7 after surgery, the hemvascularized area was significantly smaller in trap-treated mice (day 3: 15.8 \pm 4.0%; day 7: 25.2 \pm 13.3%) compared to mice just receiving the Fc-fragment (day 3: 25.8 \pm 4.4%; day 7: 48.3 \pm 12.8%; p<0.0001). This was also true for the lymphvascularized area

comparing Trap- ($9.5\pm 9.4\%$) and Fc-treated mice on day 3 ($21.5\pm 9.3\%$; $p<0.0001$). At day 7, the lymphovascularized area was smaller, but not significantly different in the Trap-group ($28.7\pm 20.3\%$) compared to the Fc-group ($51.5\pm 23.8\%$; $p=0.06$). In contrast to results obtained in corneal injury models neither hem- or lymphangiogenesis were completely inhibited by the VEGF Trap_{R1R2} following corneal transplantation. However, the number of lymphatic vessels reaching the graft-host interface (10.6 ± 0.6 versus 1.3 ± 1.5 vessels) and the number of hours where the interface was filled with draining lymphatic vessels were much larger in the Fc-treated compared to the Trap-treated group (3 ± 2 versus 0.2 ± 0.3 hours; not significant due to small sample size) at day 7. This might indicate that lymphovascularized area per se is less decisive for host sensitisation than the contact area with donor tissue.

[0065] Partial inhibition of early postoperative hem- and lymphangiogenesis by trapping VEGF-A after normal-risk surgery improves long-term graft survival.

Since hem- and lymphangiogenesis occurring *after* normal-risk keratoplasty peaked around day 7, and regressed thereafter, and since both vascular processes could be significantly inhibited by early postoperative neutralization of VEGF-A, we determined whether inhibition of postkeratoplasty hem- and lymphangiogenesis during this interval improves graft survival. The long-term survival of C57BL/6 grafts placed into avascular BALB/c recipient beds was compared between mice receiving an i.p. injection of 12.5 mg/kg VEGF Trap_{R1R2}, or Fc-fragment alone, at surgery and 3, 7, and 14 days later. Trapping of VEGF-A postoperatively caused a significantly improved long-term graft survival at 8 weeks (78%), compared to grafts in eyes of Fc-treated controls (40%; $p=0.044$; $n=22$ in both groups).

[0066] The present invention may be embodied in other specific forms without departing from the spirit or essential attributes thereof.

Claims**We claim,**

1. Use of an first agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity in the preparation of a medicament for treating or preventing corneal transplant rejection in a mammalian subject.
2. The use of claim 1, wherein the agent capable of blocking, inhibiting, or ameliorating VEGF-mediated activity is a VEGF antagonist.
3. The use of claim 2, wherein the VEGF antagonist is a polypeptide, an antibody, a small molecule, or a nucleic acid.
4. The use of claim 3, wherein the VEGF antagonist includes a VEGF trap selected from the group consisting of acetylated Flt-1(1-3)-Fc, Flt-1(1-3_{R→N})-Fc, Flt-1(1-3_{ΔB})-Fc, Flt-1(2-3_{ΔB})-Fc, Flt-1(2-3)-Fc, Flt-1D2-VEGFR3D3-FcΔC1(a), Flt-1D2-Flk-1D3-FcΔC1(a), and VEGFR1R2-FcΔC1(a).
5. The use of claim 4, wherein the VEGF trap is VEGFR1R2-FcΔC1(a).
6. The use of claim 3, wherein the VEGF antagonist is a nucleic acid selected from the group consisting of aptamer, an siRNA, or an antisense molecule.
7. The use of claim 1, wherein administration is subcutaneous, intramuscular, intradermal, intraperitoneal, intravenous, intranasal, oral, subconjunctival, or topical. Administration may also include a second agent, such as an immunosuppressive agent.
8. The use of claim 1, further comprising administering a second agent.
9. The use of claim 8, wherein the second agent is an immunosuppressive agent.
10. The use of claim 1, wherein the mammalian subject is a human.
11. The use of claim 10, wherein the human subject has received a corneal transplant.

12. A method of reducing the incidence of corneal transplant rejection in a subject in need thereof, comprising administering to the subject an agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity, such that the incidence of corneal transplant rejection is reduced.

13. A method of treating corneal transplant rejection in a subject in need thereof, comprising administering to the subject an agent capable of blocking, inhibiting, or ameliorating vascular endothelial growth factor (VEGF)-mediated activity, such that corneal transplant rejection is treated.

14. A pharmaceutical composition for prevention or treatment of corneal transplant rejection, comprising a vascular endothelial growth factor (VEGF) antagonist, and a pharmaceutically acceptable carrier.

15. The pharmaceutical composition of claim 14, in the form of a liquid, gel, ointment, salve, or ophthalmic solution.

16. An article of manufacturing comprising:

(a) packaging material; and

(b) a pharmaceutical agent contained within the packaging material;

wherein the pharmaceutical agent comprises at least one VEGF-specific fusion protein of the invention and wherein the packaging material comprises a label or package insert which indicates that the VEGF-specific fusion protein can be used to treat or prevent corneal transplant rejection in a mammalian subject.

SEQUENCE LISTING

<110> Regeneron Pharmaceuticals, Inc.
The Schepens Eye Research Institute

<120> Method of Treating Corneal Transplant
Rejection

<130> REG 713B-WO

<140> To be Assigned

<141> 2004-04-23

<160> 2

<170> FastSEQ for Windows Version 4.0

<210> 1

<211> 1377

<212> DNA

<213> homo sapiens

<400> 1

```

atggtcagct actgggacac cggggtcctg ctgtgcgcgc tgctcagctg tctgcttctc 60
acaggatcta gttccggaag tgataccggt agacctttcg tagagatgta cagtgaaatc 120
cccgaaatta tacacatgac tgaaggaagg gagctcgtca ttccctgccg ggttacgtca 180
cctaacatca ctgttacttt aaaaaagttt cactttgaca ctttgatccc tgatggaaaa 240
cgcataatct gggacagtag aaagggtctc atcatatcaa atgcaacgta caagaaata 300
gggctttctga cctgtgaagc aacagtcaat gggcatttgt ataagacaaa ctatctcaca 360
catcgacaaa ccaatacaat catagatgtg gttctgagtc cgtctcatgg aattgaacta 420
tctgttggag aaaagcttgt cttaaattgt acagcaagaa ctgaaactaa tgtggggatt 480
gacttcaact gggaataccc ttcttcgaag catcagcata agaaacttgt aaaccgagac 540
ctaaaaaacc agtctgggag tgagatgaag aaatttttga gcaccttaac tatagatggg 600
gtaaccggga gtgaccaagg attgtacacc tgtgcagcat ccagtgggct gatgaccaag 660
aagaacagca catttgtcag ggtccatgaa aaggacaaaa ctcacacatg cccaccgtgc 720
ccagcacctg aactcctggg gggaccgtca gtcttcctct tcccccaaa acccaaggac 780
accctcatga tctcccggac ccctgaggtc acatgctgtg tgggtggacgt gagccacgaa 840
gaccctgagg tcaagttcaa ctggtacgtg gacggcgtgg aggtgcataa tgccaagaca 900
aagccgcggg aggagcagta caacagcacg taccgtgtgg tcagcgtcct caccgtcctg 960
caccaggact ggctgaatgg caaggagtac aagtgcaagg tctccaacia agccctccca 1020
gccccatcg agaaaacat ctccaaagcc aaagggcagc cccgagaacc acaggtgtac 1080
accctgcccc catcccggga tgagctgacc aagaaccagg tcagcctgac ctgcctgggtc 1140
aaaggcttct atcccagcga catcgccgtg gagtgggaga gcaatgggca gccgggagaac 1200
aactacaaga ccacgcctcc cgtgctggac tccgacggct ctttcttctc ctacagcaag 1260
ctcaccgtgg acaagagcag gtggcagcag ggaacgtct tctcatgctc cgtgatgcat 1320
gaggctctgc acaaccacta cacgcagaag agcctctccc tgtctccggg taaatga 1377

```

<210> 2

<211> 458

<212> PRT

<213> homo sapiens

<400> 2

```

Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1           5           10           15
Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Ser Asp Thr Gly Arg Pro

```

			20					25					30			
Phe	Val	Glu	Met	Tyr	Ser	Glu	Ile	Pro	Glu	Ile	Ile	His	Met	Thr	Glu	
		35					40					45				
Gly	Arg	Glu	Leu	Val	Ile	Pro	Cys	Arg	Val	Thr	Ser	Pro	Asn	Ile	Thr	
		50					55				60					
Val	Thr	Leu	Lys	Lys	Phe	Pro	Leu	Asp	Thr	Leu	Ile	Pro	Asp	Gly	Lys	
65					70					75				80		
Arg	Ile	Ile	Trp	Asp	Ser	Arg	Lys	Gly	Phe	Ile	Ile	Ser	Asn	Ala	Thr	
				85					90					95		
Tyr	Lys	Glu	Ile	Gly	Leu	Leu	Thr	Cys	Glu	Ala	Thr	Val	Asn	Gly	His	
				100					105					110		
Leu	Tyr	Lys	Thr	Asn	Tyr	Leu	Thr	His	Arg	Gln	Thr	Asn	Thr	Ile	Ile	
				115					120					125		
Asp	Val	Val	Leu	Ser	Pro	Ser	His	Gly	Ile	Glu	Leu	Ser	Val	Gly	Glu	
							135					140				
Lys	Leu	Val	Leu	Asn	Cys	Thr	Ala	Arg	Thr	Glu	Leu	Asn	Val	Gly	Ile	
145					150					155				160		
Asp	Phe	Asn	Trp	Glu	Tyr	Pro	Ser	Ser	Lys	His	Gln	His	Lys	Lys	Leu	
				165					170					175		
Val	Asn	Arg	Asp	Leu	Lys	Thr	Gln	Ser	Gly	Ser	Glu	Met	Lys	Lys	Phe	
				180					185					190		
Leu	Ser	Thr	Leu	Thr	Ile	Asp	Gly	Val	Thr	Arg	Ser	Asp	Gln	Gly	Leu	
				195					200				205			
Tyr	Thr	Cys	Ala	Ala	Ser	Ser	Gly	Leu	Met	Thr	Lys	Lys	Asn	Ser	Thr	
							215						220			
Phe	Val	Arg	Val	His	Glu	Lys	Asp	Lys	Thr	His	Thr	Cys	Pro	Pro	Cys	
225					230					235					240	
Pro	Ala	Pro	Glu	Leu	Leu	Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	
					245					250				255		
Lys	Pro	Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	
				260					265					270		
Val	Val	Val	Asp	Val	Ser	His	Glu	Asp	Pro	Glu	Val	Lys	Phe	Asn	Trp	
				275										285		
Tyr	Val	Asp	Gly	Val	Glu	Val	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	
							295						300			
Glu	Gln	Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	
305							310				315				320	
His	Gln	Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	
				325						330				335		
Lys	Ala	Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	
				340						345				350		
Gln	Pro	Arg	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro	Pro	Ser	Arg	Asp	Glu	
				355					360					365		
Leu	Thr	Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	
							375						380			
Pro	Ser	Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	
385							390				395				400	
Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	
				405						410				415		
Leu	Tyr	Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp	Gln	Gln	Gly	Asn	
				420						425				430		
Val	Phe	Ser	Cys	Ser	Val	Met	His	Glu	Ala	Leu	His	Asn	His	Tyr	Thr	
				435						440				445		
Gln	Lys	Ser	Leu	Ser	Leu	Ser	Pro	Gly	Lys							
							455									
450																

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property
Organization
International Bureau



(43) International Publication Date
6 January 2005 (06.01.2005)

PCT

(10) International Publication Number
WO 2005/000895 A2

- (51) International Patent Classification⁷: **C07K 14/71**
- (21) International Application Number:
PCT/US2004/021059
- (22) International Filing Date: 29 June 2004 (29.06.2004)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
10/609,775 30 June 2003 (30.06.2003) US
- (71) Applicant (for all designated States except US): **REGENERON PHARMACEUTICALS, INC.** [US/US]; 777 Old Saw Mill River Road, Tarrytown, NY 10591 (US).
- (72) Inventors; and
- (75) Inventors/Applicants (for US only): **DALY, Thomas, J.** [US/US]; 4 Dolphin Road, New City, NY 10956 (US). **FANDL, James, P.** [US/US]; 40 Amanda's Way, LaGrangeville, NY 12540 (US). **PAPADOPOULOS, Nicholas, J.** [US/US]; 59 Heritage Lane, LaGrangeville, NY 12540 (US).
- (74) Agent: **VALETA, Gregg;** Regeneron Pharmaceuticals, Inc., 777 Old Saw Mill River Road, Tarrytown, NY 10591 (US).
- (81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.
- (84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).
- Published:**
— without international search report and to be republished upon receipt of that report
- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 2005/000895 A2

(54) Title: VEGF TRAPS AND THERAPEUTIC USES THEREOF

(57) Abstract: Nucleic acid molecules and multimeric proteins capable of binding vascular endothelial growth factor (VEGF). VEGF traps are disclosed which are therapeutically useful for treating VEGF-associated conditions and diseases, and are specifically designed for local administration to specific organs, tissues, and/or cells.

VEGF TRAPS AND THERAPEUTIC USES THEREOF**BACKGROUND OF THE INVENTION****Field of the Invention**

[0001] The invention encompasses fusion polypeptides capable of binding vascular endothelial cell growth factor (VEGF), VEGF family members, and splice variants with specifically desirable characteristics, as well as therapeutic methods of use.

BRIEF SUMMARY OF THE INVENTION

[0002] In a first aspect, the invention features an isolated nucleic acid molecule encoding a fusion polypeptide comprising receptor components $(R1R2)_X$ and/or $(R1R3)_Y$, wherein R1 is vascular endothelial cell growth factor (VEGF) receptor component Ig domain 2 of Flt-1 (Flt1D2), R2 is VEGF receptor component Ig domain 3 of Flk-1 (Flk1D3), and R3 is VEGF receptor component Ig domain 3 of Flt-4 (Flt1D3 or R3), and wherein $X \geq 1$ and $Y \geq 1$.

[0003] In a related second aspect, the invention features a monomeric VEGF trap or fusion polypeptide comprising VEGF receptor components $(R1R2)_X$ and/or $(R1R3)_Y$ wherein $X \geq 1$, $Y \geq 1$, and R1, R2, and R3 are as defined above. The VEGF receptor components R1, R2, and R3, may be connected directly to each other or connected via one or more spacer sequences. In one specific embodiment, the monomeric VEGF trap is $(R1R2)_X$, where $X=2$. In a more specific embodiment, the monomeric VEGF trap is SEQ ID NO:24, or a functionally equivalent amino acid variant thereof. The invention encompasses a monomeric VEGF trap consisting essentially of VEGF receptor components $(R1R2)_X$ and/or $(R1R3)_Y$, and functionally equivalent amino acid variants thereof.

[0004] In a third aspect, the invention features an isolated nucleic acid molecule encoding a fusion polypeptide comprising VEGF receptor components $(R1R2)_X$ and/or $(R1R3)_Y$, and a fusion partner (FP) component selected from the group consisting of a multimerizing component (MC), a serum protein, or a molecule capable of binding a serum protein. In a preferred embodiment, FP is a multimerizing component (MC) capable of interacting with a multimerizing component on another fusion polypeptide to form a multimeric structure, e.g., a dimer or trimer. Most preferably, the MC is selected from the group consisting of (i) a multimerizing component comprising a cleavable region (C-region), (ii) a truncated multimerizing component, (iii) an amino acid sequence between 1 to about 200 amino acids in length having at least one cysteine residue, (iv) a leucine zipper, (v) a helix loop motif, (vi) a coil-coil motif, and (vii) an immunoglobulin domain. Further encompassed are fusion polypeptides consisting essentially of $(R1R2)_X$ and/or $(R1R3)_Y$, and FP. In a preferred embodiment, the fusion polypeptide consists essentially of $(R1R2)_X$ and MC.

[0005] In a fourth aspect, the invention features a fusion polypeptide comprising VEGF receptor components $(R1R2)_X$ and/or $(R1R3)_Y$, and FP, as described above. The receptor components may be arranged in different orders, for example, $(R1R2)_X$ -FP; $(R1R2)_X$ -FP- $(R1R2)_X$; FP- $(R2R1)_X$, etc. The components of the fusion polypeptide may be connected directly to each other, or connected via a spacer sequence.

[0006] In a fifth aspect, the invention features a VEGF trap, comprising a multimer of two or more fusion polypeptides consisting of VEGF receptor components $(R1R2)_X$ and/or $(R1R3)_Y$, and FP, wherein the FP component is a multimerizing component (MC) comprising a C-region. The C-region may be naturally occurring or artificial, and may occur at any point within the multimerizing component, and functions to allow cleavage of a parent MC to a truncated MC. A VEGF trap composed of two or more fusion polypeptides having at least one truncated MC is termed a "truncated mini-trap."

[0007] The C-region may be created in MC by insertion, deletion, or mutation, such that an enzymatically or chemically cleavable site is created. The C-region may be created in any MC and at any position within the MC; preferably, the C-region is created in a full length Fc domain, or a fragment thereof, or a C_H3 domain. The C-region may be a site cleavable by an enzyme, such as, thrombin, ficin, pepsin, matrilysin, or prolidase or cleavable chemically by, for example, formic acid or $CuCl_2$.

[0008] In a sixth related aspect, the invention features a truncated VEGF mini-trap which is a multimeric protein comprising two or more fusion polypeptides consisting of $(R1R2)_X$ and/or $(R1R3)_Y$ and a multimerizing component which is a truncated by cleavage from a parent MC comprising a C-region (tMC).

[0009] In a seventh aspect, the invention features a fusion polypeptide consisting of VEGF receptor components $(R1R2)_X$ and/or $(R1R3)_Y$ and a MC, wherein the MC is an amino acid sequence between 1 to about 200 amino acids in length comprising at least one cysteine residue, wherein the at least one cysteine residue is capable of forming a disulfide bond with a cysteine residue present in the MC of another fusion polypeptide (cMC). In a preferred embodiment, cMC is an amino acid sequence between 1-50 amino acids in length comprising at least one cysteine residue. In a more preferred embodiment, cMC is an amino acid sequence between 1-15 amino acids in length comprising at least one amino acid. In an even more preferred embodiment, cMC is an amino acid sequence between 1-10 amino acids in length comprising 1-2 cysteine residues. One exemplification of this embodiment of the invention is shown in SEQ ID NO:27 having a signal sequence (1-26) followed by R1 (27-129) and R2 (130-231) components, followed by a nine amino acid sequence ending in a cysteine residue. In another embodiment, shown in SEQ ID NO:28, a signal sequence (1-26) is followed by R1 (27-129) and R2 (130-231) components, followed by a six amino acid sequence ending in a cysteine residue.

[0010] In an eighth aspect, the invention features a VEGF mini-trap, comprising a multimer of two or more fusion polypeptides consisting of $(R1R2)_X$ and/or $(R1R3)_Y$ and a cMC. In a more specific embodiment, the mini-trap is a dimer. One exemplification of this embodiment of the mini-trap of the invention is a dimer of the fusion polypeptide shown in SEQ ID NO:2, wherein each fusion polypeptide (R1R2-cMC) has a molecular weight of 23.0 kD and a pI of 9.22.

[0011] In another embodiment, cMC is 4 amino acids in length consisting of two cysteine residues, for example, XCXC (SEQ ID NO:3). In one exemplification of this embodiment of the invention, the mini-trap consists of the VEGF receptor components of the invention, and a cMC consisting of ACGC (SEQ ID NO:4). One exemplification of this embodiment of the mini-trap of the invention is

a dimer of the fusion polypeptide shown in SEQ ID NO:5, wherein each monomer has a molecular weight of 23.2 kD and a pI of 9.22. Another exemplification of this embodiment of the invention is shown in SEQ ID NO:26 having a signal sequence (1-26) followed by R1 (27-129) and R2 (130-231) components, followed by a nine amino acid sequence ending in CPPC.

[0012] In all embodiments of the VEGF trap of the invention (including truncated VEGF mini-trap, VEGF mini-traps, and monomeric VEGF mini-traps), a signal sequence (S) may be included at the beginning (or N-terminus) of the fusion polypeptide of the invention. The signal sequence may be native to the cell, recombinant, or synthetic. When a signal sequence is attached to the N-terminus of a first receptor component, thus a fusion polypeptide may be designated as, for example, S-(R1R2)_x.

[0013] The components of the fusion polypeptide may be connected directly to each other or be connected via spacers. In specific embodiments, one or more receptor and/or fusion partner components of the fusion polypeptide are connected directly to each other without spacers. In other embodiments, one or more receptor and/or fusion partner components are connected with spacers.

[0014] The invention encompasses vectors comprising the nucleic acid molecules of the invention, including expression vectors comprising the nucleic acid molecule operatively linked to an expression control sequence. The invention further encompasses host-vector systems for the production of a fusion polypeptide which comprise the expression vector, in a suitable host cell; host-vector systems wherein the suitable host cell is a bacterial, yeast, insect, mammalian cell; an *E. coli* cell, or a COS or CHO cell. Additional encompassed are VEGF traps of the invention modified by acetylation or pegylation. Methods for acetylating or pegylating a protein are well known in the art.

[0015] In a related ninth aspect, the invention features a method of producing a VEGF trap of the invention, comprising culturing a host cell transfected with a vector comprising a nucleic acid sequence of the invention, under conditions suitable for expression of the protein from the host cell, and recovering the fusion polypeptides so produced.

[0016] The VEGF traps of the invention are therapeutically useful for treating any disease or condition which is improved, ameliorated, or inhibited by removal, inhibition, or reduction of VEGF. A non-exhaustive list of specific conditions improved by inhibition or reduction of VEGF include, for example, undesirable plasma leakage or vascular permeability, undesirable blood vessel growth, e.g., such as in a tumor, edema associated with inflammatory disorders such as psoriasis or arthritis, including rheumatoid arthritis; asthma; generalized edema associated with burns; ascites and pleural effusion associated with tumors, inflammation or trauma; chronic airway inflammation; asthma; capillary leak syndrome; sepsis; kidney disease associated with increased leakage of protein; pancreatic ductal adenocarcinoma (PDAC) and eye disorders such as age related macular degeneration and diabetic retinopathy. The VEGF mini-trap is particularly useful in treatment of eye disorders, and as an adjuvant to eye surgeries, including glaucoma surgery; and the treatment of intra-ocular tumors, such as for example, uveal melanoma, retinoblastoma, via intravitreal delivery.

[0017] Accordingly, in a tenth aspect, the invention features a therapeutic method for the treatment of a VEGF-related disease or condition, comprising administering a VEGF trap of the invention to a subject suffering from a VEGF-related disease or condition. Although any mammal

can be treated by the therapeutic methods of the invention, the subject is preferably a human patient suffering from or at risk of suffering from a condition or disease which can be improved, ameliorated, inhibited or treated with a VEGF trap.

[0018] In a eleventh aspect, the invention further features diagnostic and prognostic methods, as well as kits for detecting, quantitating, and/or monitoring VEGF with the mini-traps of the invention.

[0019] In a twelfth aspect, the invention features pharmaceutical compositions comprising a VEGF trap of the invention with a pharmaceutically acceptable carrier. Such pharmaceutical compositions may comprise a dimeric fusion polypeptide trap, or nucleic acids encoding the fusion polypeptide. The mini-traps of the invention find specific uses in conditions in which a VEGF trap with reduced serum half life (e.g., faster clearance), and/or increased tissue penetration due to smaller size is desirable. Specific applications for the VEGF mini-trap include, for example, diseases where local administration to a specific tissue or cell is desirable. Examples of such a condition or disease are ocular diseases of the eye.

[0020] Other objects and advantages will become apparent from a review of the ensuing detailed description.

DETAILED DESCRIPTION OF THE INVENTION

[0021] Before the present methods are described, it is to be understood that this invention is not limited to particular methods, and experimental conditions described, as such methods and conditions may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only the appended claims.

[0022] As used in this specification and the appended claims, the singular forms "a", "an", and "the" include plural references unless the context clearly dictates otherwise. Thus for example, a reference to "a method" includes one or more methods, and/or steps of the type described herein and/or which will become apparent to those persons skilled in the art upon reading this disclosure and so forth.

[0023] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are now described. All publications mentioned herein are incorporated herein by reference to describe the methods and/or materials in connection with which the publications are cited.

General Description

[0024] The invention encompasses a VEGF trap capable of binding and inhibiting VEGF activity which is a monomer or multimer of one or more fusion polypeptides. The molecules of the invention bind and inhibit the biological action of VEGF and/or the physiological reaction or response. For a description of VEGF-receptor-based antagonist VEGF traps Flt1D2.Flk1D3.Fc Δ C1(a) (SEQ ID NOs:7-8) and VEGFR1R2-Fc Δ C1(a) (SEQ ID NOs:9-10), see PCT WO/0075319, the contents of which is incorporated in its entirety herein by reference.

[0025] The mini-trap of the invention is smaller than the full sized trap, e.g., about 50 - 60 kD

versus 120 kD of the parent trap, and include monomeric traps consisting essentially of VEGF receptor domains (R1R2)_x, (R1R3)_y, or combinations thereof, traps generated by cleavage of a portion of a parent multimerized trap having a fusion partner component which is a multimerizing component (MC) containing a cleavage region (C-region); or by attaching a cysteine residue or amino acid sequence containing one or more cysteine residues to or between receptor component domains. In specific embodiments, the mini-trap of the invention is less than about 60 kD as measured by SDS-PAGE analysis; more preferably, about 50 kD; even more preferably about 20-30 kD; or is about 25 kD and capable of binding VEGF with an affinity comparable to a full-sized parent trap described in PCT/US00/14142.

Nucleic Acid Constructs and Expression

[0026] The present invention provides for the construction of nucleic acid molecules encoding fusion polypeptides capable of binding VEGF alone or multimerized VEGF traps. The nucleic acid molecules of the invention may encode wild-type R1, R2, and/or R3 receptor components, or functionally equivalent variants thereof. Amino acid sequence variants of the R1, R2 and/or R3 receptor components of the traps of the invention may also be prepared by creating mutations in the encoding nucleic acid molecules. Such variants include, for example, deletions from, or insertions or substitutions of, amino acid residues within the amino acid sequence of R1, R2 and/or R3. Any combination of deletion, insertion, and substitution may be made to arrive at a final construct, provided that the final construct possesses the ability to bind and inhibit VEGF.

[0027] These nucleic acid molecules are inserted into a vector that is able to express the fusion polypeptides when introduced into an appropriate host cell. Appropriate host cells include, but are not limited to, bacterial, yeast, insect, and mammalian cells. Any of the methods known to one skilled in the art for the insertion of DNA fragments into a vector may be used to construct expression vectors encoding the fusion polypeptides of the invention under control of transcriptional/translational control signals.

[0028] Expression of the nucleic acid molecules of the invention may be regulated by a second nucleic acid sequence so that the molecule is expressed in a host transformed with the recombinant DNA molecule. For example, expression may be controlled by any promoter/enhancer element known in the art. Promoters which may be used to control expression of the chimeric polypeptide molecules include, but are not limited to, a long terminal repeat (Squinto et al. (1991) Cell 65:1-20); SV40 early promoter region, CMV, M-MuLV, thymidine kinase promoter, the regulatory sequences of the metallothionein gene; prokaryotic expression vectors such as the b-lactamase promoter, or the tac promoter (see also Scientific American (1980) 242:74-94); promoter elements from yeast or other fungi such as Gal 4 promoter, ADH, PGK, alkaline phosphatase, and tissue-specific transcriptional control regions derived from genes such as elastase I.

[0029] Expression vectors capable of being replicated in a bacterial or eukaryotic host comprising the nucleic acid molecules of the invention are used to transfect the host and thereby direct expression of such nucleic acids to produce the fusion polypeptides of the invention, which form traps capable of binding to VEGF. Transfected cells may transiently or, preferably, constitutively

and permanently express the VEGF traps of the invention.

[0030] The traps of the invention may be purified by any technique which allows for the subsequent formation of a stable, biologically active trap. For example, and not by way of limitation, the factors may be recovered from cells either as soluble proteins or as inclusion bodies, from which they may be extracted quantitatively by 8M guanidinium hydrochloride and dialysis (see, for example, US Patent No. 5,663,304). In order to further purify the factors, conventional ion exchange chromatography, hydrophobic interaction chromatography, reverse phase chromatography or gel filtration may be used.

VEGF Receptor Components

[0031] The VEGF receptor components of the VEGF mini trap consist of the Ig domain 2 of Flt-1 (Flt1D2) (R1), the Ig domain 3 of Flk-1 (Flk1D3) (R2) (together, R1R2), and/or R1 and Ig domain 3 of Flt-4 (Flt1D3) (R3) (together, R1R3). The term "Ig domain" of Flt-1, Flt-4, or Flk-1 is intended to encompass not only the complete wild-type domain, but also insertional, deletional, and/or substitutional variants thereof which substantially retain the functional characteristics of the intact domain. It will be readily apparent to one of skill in the art that numerous variants of the above Ig domains can be obtained which will retain substantially the same functional characteristics as the wild-type domain.

[0032] The term "functional equivalents" when used in reference to R1, R2, or R3, is intended to encompass an R1, R2, or R3 domain with at least one alteration, e.g., a deletion, addition, and/or substitution, which retains substantially the same functional characteristics as does the wild type R1, R2, or R3 domain, that is, a substantially equivalent binding to VEGF. It will be appreciated that various amino acid substitutions can be made in R1, R2, or R3 without departing from the spirit of the invention with respect to the ability of these receptor components to bind and inactivate VEGF. The functional characteristics of the traps of the invention may be determined by any suitable screening assay known to the art for measuring the desired characteristic. Examples of such assays are described in the experimental section below which allow determination of binding characteristics of the traps for VEGF (K_d), as well as their half-life of dissociation of the trap-ligand complex ($T_{1/2}$). Other assays, for example, a change in the ability to specifically bind to VEGF can be measured by a competition-type VEGF binding assay. Modifications of protein properties such as thermal stability, hydrophobicity, susceptibility to proteolytic degradation, or tendency to aggregate may be measured by methods known to those of skill in the art.

[0033] The components of the fusion polypeptide may be connected directly to each other or be connected via spacers. Generally, the term "spacer" (or linker) means one or more molecules, e.g., nucleic acids or amino acids, or non-peptide moieties, such as polyethylene glycol, which may be inserted between one or more component domains. For example, spacer sequences may be used to provide a desirable site of interest between components for ease of manipulation. A spacer may also be provided to enhance expression of the fusion polypeptide from a host cell, to decrease steric hindrance such that the component may assume its optimal tertiary structure and/or interact appropriately with its target molecule. For spacers and methods of identifying desirable spacers, see,

for example, George et al. (2003) Protein Engineering 15:871-879, herein specifically incorporated by reference. A spacer sequence may include one or more amino acids naturally connected to a receptor component, or may be an added sequence used to enhance expression of the fusion polypeptides, provide specifically desired sites of interest, allow component domains to form optimal tertiary structures and/or to enhance the interaction of a component with its target molecule. In one embodiment, the spacer comprises one or more peptide sequences between one or more components which is (are) between 1-100 amino acids, preferably 1-25.

[0034] In the most specific embodiments, R1 is amino acids 27-126 of SEQ ID NO:8, or 1-126 of SEQ ID NO:8 (including the signal sequence 1-26); or amino acids 27-129 of SEQ ID NO:10, or 1-129 of SEQ ID NO:10 (including the signal sequence at 1-26). In the most specific embodiments, R2 is amino acids 127-228 of SEQ ID NO:8, or amino acids 130-231 of SEQ ID NO:10. In the most specific embodiments, R3 is amino acids 127-225 of SEQ ID NO: 13 (without a signal sequence). When, for example, R2 is placed at the N-terminus of the fusion polypeptide, a signal sequence may desirably precede the receptor component. The receptor component(s) attached to the multimerizing component may further comprise a spacer component, for example, the GPG sequence of amino acids 229-231 of SEQ ID NO:7.

Fusion Partner and Multimerizing Components

[0035] The fusion partner is any component that enhances the functionality of the fusion polypeptide. Thus, for example, an fusion partner may enhance the biological activity of the fusion polypeptide, aid in its production and/or recovery, or enhance a pharmacological property or the pharmacokinetic profile of the fusion polypeptide by, for example, enhancing its serum half-life, tissue penetrability, lack of immunogenicity, or stability. In preferred embodiments, the fusion partner is selected from the group consisting of a multimerizing component, a serum protein, or a molecule capable of binding a serum protein.

[0036] When the fusion partner is a serum protein or fragment thereof, it is selected from the group consisting of α -1-microglobulin, AGP-1, orosomucoid, α -1-acid glycoprotein, vitamin D binding protein (DBP), hemopexin, human serum albumin (hSA), transferrin, ferritin, afamin, haptoglobin, α -fetoprotein thyroglobulin, α -2-HS-glycoprotein, β -2-glycoprotein, hyaluronan-binding protein, syntaxin, C1R, C1q a chain, galectin3-Mac2 binding protein, fibrinogen, polymeric Ig receptor (PIGR), α -2-macroglobulin, urea transport protein, haptoglobin, IGFbps, macrophage scavenger receptors, fibronectin, giantin, Fc, α -1-antichymotrypsin, α -1-antitrypsin, antithrombin III, apolipoprotein A-I, apolipoprotein B, β -2-microglobulin, ceruloplasmin, complement component C3 or C4, CI esterase inhibitor, C-reactive protein, cystatin C, and protein C. In a more specific embodiment, fusion partner is selected from the group consisting of α -1-microglobulin, AGP-1, orosomucoid, α -1-acid glycoprotein, vitamin D binding protein (DBP), hemopexin, human serum albumin (hSA), afamin, and haptoglobin. The inclusion of a fusion partner component may extend the serum half-life of the fusion polypeptide of the invention when desired. See, for example, US Patent Nos. 6,423,512, 5,876,969, 6,593,295, and 6,548,653, herein specifically incorporated by

reference in their entirety, for examples of serum albumin fusion polypeptides. hSA is widely distributed throughout the body, particularly in the intestinal and blood components, and has an important role in the maintenance of osmolarity and plasma volume. It is slowly cleared in the liver, and typically has an *in vivo* half-life of 14-20 days in humans (Waldmann et al. (1977) Albumin, Structure Function and Uses; Pergamon Press; pp. 255-275).

[0037] When a fusion partner is a molecule capable of binding a serum protein, the molecule may be a synthetic small molecule, a lipid or liposome, a nucleic acid, including a synthetic nucleic acid such as an aptamer, a peptide, or an oligosaccharide. The molecule may further be a protein, such as, for example, Fc γ R1, Fc γ R2, Fc γ R3, polymeric Ig receptor (PIGR), ScFv, and other antibody fragments specific for a serum protein.

[0038] When the fusion partner is a multimerizing component (MC), it is any natural or synthetic sequence capable of interacting with another MC to form a higher order structure, e.g., a dimer, a trimer, etc. Suitable MCs may include a leucine zipper, including leucine zipper domains derived from c-jun or c-fos; sequences derived from the constant regions of kappa or lambda light chains; synthetic sequences such as helix-loop-helix motifs (Müller et al. (1998) FEBS Lett. 432:45-49), coil-coil motifs, etc., or other generally accepted multimerizing domains known to the art. In some embodiments, the fusion component comprises an immunoglobulin-derived domain from, for example, human IgG, IgM or IgA. In specific embodiments, the immunoglobulin-derived domain may be selected from the group consisting of the Fc domain of IgG, the heavy chain of IgG, and the light chain of IgG. The Fc domain of IgG may be selected from the isotypes IgG1, IgG2, IgG3, and IgG4, as well as any allotype within each isotype group. In one example of the VEGF trap of the invention, the multimerizing component is an IgG4 Fc domain (SEQ ID NO:29).

Generation of Truncated VEGF Mini-Traps

[0039] In one embodiment of the trap of the invention, a truncated VEGF mini-trap comprising two or more fusion polypeptides of the invention, is generated by subjecting a parent trap having C-region-containing MCs to conditions under which one or more of the C-region-containing MCs is (are) cleaved. The resulting truncated mini-trap may be a full and partial cleavage product of a parent trap.

[0040] The C-region-containing MC may be any MC capable of interacting with another MC to form a higher order structure, e.g., a dimer or a trimer. The C-region may be created within an MC at any desired location. In light of the guidance provided in the examples below, one of skill in the art would be able to select a desired site for creation of a C-region based on the desired properties of the resulting truncated traps, e.g., molecular weight, monomeric or dimeric, etc.

[0041] In a specific embodiment, the C-region is a thrombin cleavage site (LVPRGS) (SEQ ID NO:6) inserted into an Fc Δ C1 domain following the N-terminal CPPC sequence (SEQ ID NO:1). In this embodiment, a full-sized parent VEGF trap construct is expressed in a cell as an Fc-tagged protein, thus allowing capture and purification by, for example, a Protein A column. Following formation of a dimer and covalent bonding between one or both of the cysteine residues of the CPPC sequence

(SEQ ID NO:1), the dimer is exposed to thrombin under conditions which cleave one or both of the Fc Δ C1 domains such that truncated dimeric mini-traps are generated, having a molecular weight of approximately 50 kD – 90 kD, and has an affinity for VEGF comparable to that of the parent trap. The conditions of cleavage may be controlled by one of skill in the art to favor formation of the partial cleavage product or the fully cleaved product, the choice of cleavage conditions selected by desire for a particular product having specific properties such as molecular weight.

[0042] In a specific embodiment, the C-region is a thrombin cleavage site (LVPRGS) (SEQ ID NO:6) inserted into an Fc Δ C1 domain N-terminal to the CPPC sequence (SEQ ID NO:1). Following formation of a dimer and covalent bonding between one or both of the cysteine residues of the CPPC sequence (SEQ ID NO:1), the dimer is exposed to thrombin under conditions in which one or both of the Fc Δ C1 domain occur and truncated monomeric mini-traps are generated. The monomeric truncated mini-trap thus generated comprises a receptor component, and a small fragment of the Fc, and is approximately 25 kD in size and exhibits a reduced affinity for VEGF relative to the truncated dimeric trap and the full length parent trap. A similar monomeric trap produced as a recombinant protein has been shown to have a K_D of about 1 nM.

Generation of VEGF Mini-Traps

[0043] In one embodiment, the invention features VEGF mini-traps having one or more receptor component domains (R1R2)_X and/or (R1R3)_Y, wherein $X \geq 1$, $Y \geq 1$, and R1, R2, and R3 are as defined above, and optionally, a fusion partner which is preferably a MC domain which is an amino acid sequence between 1 to about 200 amino acids in length comprising at least one cysteine residue, wherein the at least one cysteine residue is capable of forming a disulfide bond with a cysteine residue present in the MC of another fusion polypeptide (cMC). The cMC may occur at the N-terminus or C-terminus of a fusion polypeptide, or between two receptor component domains. In one specific embodiment, cysteine is added to the C-terminus of a VEGF receptor component, e.g., R1R2_C, which allows the fusion polypeptide to form covalent dimers through formation of a covalent disulfide bond between the cysteine residue at the C-terminus of one fusion polypeptide and the cysteine residue at the C-terminus of another fusion polypeptide. In this exemplification, the mini-trap is a dimer of the fusion polypeptide shown in SEQ ID NO:2, wherein each fusion polypeptide (R1R2-cMC or R1R2_C) has a molecular weight of about 23.0 kD.

[0044] In another embodiment, the cMC is a sequence of 4 amino acids (XXXX) (SEQ ID NO:11) wherein X is any amino acid and the sequence comprises at least one cysteine residue. In a specific embodiment, the cMC is added to the C-terminus of a receptor component domain. In a more specific embodiment, the 4 amino acid sequence is ACGC (SEQ ID NO:4) and the cMC forms two disulfide bonds with the cysteine residues present in a second fusion polypeptide. As shown below (Table 2), both the exemplified mini-traps exhibit an affinity for VEGF comparable to the parent trap.

Therapeutic Uses

[0045] The VEGF mini-traps of the invention are therapeutically useful for treating any disease or

condition which is improved, ameliorated, inhibited or prevented by removal, inhibition, or reduction of VEGF. A non-exhaustive list of specific conditions improved by inhibition or reduction of VEGF include, clinical conditions that are characterized by excessive vascular endothelial cell proliferation, vascular permeability, edema or inflammation such as brain edema associated with injury, stroke or tumor; edema associated with inflammatory disorders such as psoriasis or arthritis, including rheumatoid arthritis; asthma; generalized edema associated with burns; ascites and pleural effusion associated with tumors, inflammation or trauma; chronic airway inflammation; capillary leak syndrome; sepsis; kidney disease associated with increased leakage of protein; and eye disorders such as age related macular degeneration and diabetic retinopathy.

[0046] The compositions of the invention are therapeutically useful for treating a wide variety of diseases associated with increased VEGF levels. For example, exaggerated Th2 inflammation and airway remodeling are characteristic in the pathogenesis of asthma (see, for example, Elias et al. (1999) *J. Clin. Invest.* 104:1001-6). Elevated VEGF levels have been detected in tissues and biologic samples from patients with asthma, which correlate directly with disease activity (Lee et al. (2001) *J. Allergy Clin. Immunol.* 107:1106-1108) and inversely with airway caliber and airway responsiveness. Further, VEGF has been postulated to contribute to asthmatic tissue edema.

[0047] Another disease associated with increased VEGF is pancreatic ductal adenocarcinoma (PDAC). This malignancy often exhibits enhanced foci of endothelial cell proliferation and frequently overexpresses VEGF (Ferrara (1999) *J. Mol. Med.* 77:527-543). PDAC is responsible for over 20% of deaths due to gastrointestinal malignancies, making it the fourth most common cause of cancer-related mortality in the U.S. and other industrialized countries. Experimental evidence supports an important role for VEGF in pancreatic cancer, thus a VEGF inhibitor has promise as a therapeutic to attenuate intrapancreatic tumor growth and regional and distal metastasis.

[0048] A smaller, non-glycosylated mini-trap expressed in *E. coli* (Example 4), a glycosylated mini-trap expressed in CHO cells (Example 5), or a receptor-based monomeric trap (Example 6) has optimized characteristics for local/intra-vitreous delivery, ie. a shorter serum half life for faster clearance and minimizing unwanted systemic exposure. In addition due to its smaller size, the mini-trap has the ability to penetrate through the inner-limiting membrane (ILM) in the eye, and diffuse through the vitreous to the retina/retinal pigment epithelial (RPE) layer which will help to treat retinal disease. Additionally, the mini-trap can be used for local administration for the treatment of ocular disease such as choroidal neovascularization, diabetic macular edema, proliferative diabetic retinopathy, corneal neovascularization/transplant rejection. Still further, the mini-trap can be used in any situation where transient (short-term) blocking of VEGF is required, e.g., to avoid chronic exposure to VEGF blockade, such as, for example, in the treatment of psoriasis.

[0049] A serious problem leading to failure following glaucoma surgery is early inflammation and angiogenesis, as well as too aggressive wound healing. Accordingly, the VEGF traps of the invention may be usefully employed as an adjuvant to glaucoma surgery to prevent early hem- and lymphangiogenesis and macrophage recruitment to the filterig bleb after glaucoma surgery, and improve surgical outcome.

Combination Therapies

[0050] In numerous embodiments, a VEGF trap may be administered in combination with one or more additional compounds or therapies, including a second VEGF trap molecule, a chemotherapeutic agent, surgery, catheter devices, and radiation. Combination therapy includes administration of a single pharmaceutical dosage formulation which contains a VEGF trap and one or more additional agents; as well as administration of a VEGF trap and one or more additional agent(s) in its own separate pharmaceutical dosage formulation. For example, a VEGF trap and a cytotoxic agent, a chemotherapeutic agent or a growth inhibitory agent can be administered to the patient together in a single dosage composition such as a combined formulation, or each agent can be administered in a separate dosage formulation. Where separate dosage formulations are used, the VEGF-specific fusion polypeptide of the invention and one or more additional agents can be administered concurrently, or at separately staggered times, i.e., sequentially.

[0051] The term "cytotoxic agent" as used herein refers to a substance that inhibits or prevents the function of cells and/or causes destruction of cells. The term is intended to include radioactive isotopes (e.g. I^{131} , I^{125} , Y^{90} and Re^{186}), chemotherapeutic agents, and toxins such as enzymatically active toxins of bacterial, fungal, plant or animal origin, or fragments thereof.

[0052] A "chemotherapeutic agent" is a chemical compound useful in the treatment of cancer. Examples of chemotherapeutic agents include alkylating agents such as thiotepa and cyclophosphamide (Cytoxan®); alkyl sulfonates such as busulfan, improsulfan and piposulfan; aziridines such as benzodopa, carboquone, meturedopa, and uredopa; ethylenimines and methylamelamines including altretamine, triethylenemelamine, trietylenephosphoramide, triethylenethiophosphoramide and trimethylolomelamine; nitrogen mustards such as chlorambucil, chlornaphazine, cholophosphamide, estramustine, ifosfamide, mechlorethamine, mechlorethamine oxide hydrochloride, melphalan, novembichin, phenesterine, prednimustine, trofosfamide, uracil mustard; nitrosureas such as carmustine, chlorozotocin, fotemustine, lomustine, nimustine, ranimustine; antibiotics such as aclacinomysins, actinomycin, authramycin, azaserine, bleomycins, cactinomycin, calicheamicin, carabycin, carminomycin, carzinophilin, chromomycins, dactinomycin, daunorubicin, detorubicin, 6-diazo-5-oxo-L-norleucine, doxorubicin, epirubicin, esorubicin, idarubicin, marcellomycin, mitomycins, mycophenolic acid, nogalamycin, olivomycins, peplomycin, potfirromycin, puromycin, quelamycin, rodorubicin, streptonigrin, streptozocin, tubercidin, ubenimex, zinostatin, zorubicin; anti-metabolites such as methotrexate and 5-fluorouracil (5-FU); folic acid analogues such as denopterin, methotrexate, pteropterin, trimetrexate; purine analogs such as fludarabine, 6-mercaptopurine, thiamiprine, thioguanine; pyrimidine analogs such as ancitabine, azacitidine, 6-azauridine, carmofur, cytarabine, dideoxyuridine, doxifluridine, enocitabine, floxuridine; androgens such as calusterone, dromostanolone propionate, epitio stanol, mepitio stanol, testolactone; anti-adrenals such as aminoglutethimide, mitotane, trilostane; folic acid replenisher such as frolic acid; aceglatone; aldophosphamide glycoside; aminolevulinic acid; amsacrine; bestrabucil; bisantrene; edatraxate; defofamine; demecolcine; diaziquone; elfornithine; elliptinium acetate; etoglucid; gallium nitrate; hydroxyurea; lentinan; lonidamine; mitoguazone; mitoxantrone; mopidamol; nitracrine; pentostatin; phenamet; pirarubicin; podophyllinic acid; 2-ethylhydrazide; procarbazine; PSK®;

razoxane; sizofiran; spirogermanium; tenuazonic acid; triaziquone; 2, 2',2"-trichlorotriethylamine; urethan; vindesine; dacarbazine; mannomustine; mitobronitol; mitolactol; pipobroman; gacytosine; arabinoside ("Ara-C"); cyclophosphamide; thiotepa; taxanes, e.g. paclitaxel (Taxol®, Bristol-Myers Squibb Oncology, Princeton, N.J.) and docetaxel (Taxotere®; Aventis Antony, France); chlorambucil; gemcitabine; 6-thioguanine; mercaptopurine; methotrexate; platinum analogs such as cisplatin and carboplatin; vinblastine; platinum; etoposide (VP-16); ifosfamide; mitomycin C; mitoxantrone; vincristine; vinorelbine; navelbine; novantrone; teniposide; daunomycin; aminopterin; xeloda; ibandronate; CPT-11; topoisomerase inhibitor RFS 2000; difluoromethylornithine (DMFO); retinoic acid; esperamicins; capecitabine; and pharmaceutically acceptable salts, acids or derivatives of any of the above. Also included in this definition are anti-hormonal agents that act to regulate or inhibit hormone action on tumors such as anti-estrogens including for example tamoxifen, raloxifene, aromatase inhibiting 4(5)-imidazoles, 4-hydroxytamoxifen, trioxifene, keoxifene, LY 117018, onapristone, and toremifene (Fareston); and anti-androgens such as flutamide, nilutamide, bicalutamide, leuprolide, and goserelin; and pharmaceutically acceptable salts, acids or derivatives of any of the above.

[0053] A "growth inhibitory agent" when used herein refers to a compound or composition which inhibits growth of a cell, especially a cancer cell either *in vitro* or *in vivo*. Examples of growth inhibitory agents include agents that block cell cycle progression (at a place other than S phase), such as agents that induce G1 arrest and M-phase arrest. Classical M-phase blockers include the vincas (vincristine and vinblastine), Taxol ®, and topo II inhibitors such as doxorubicin, epirubicin, daunorubicin, etoposide, and bleomycin. Those agents that arrest G1 also spill over into S-phase arrest, for example, DNA alkylating agents such as tamoxifen, prednisone, dacarbazine, mechlorethamine, cisplatin, methotrexate, 5-fluorouracil, and ara-C.

Methods of Administration

[0054] The invention provides methods of treatment comprising administering to a subject an effective amount of a VEGF trap of the invention. In a preferred aspect, the trap is substantially purified (*e.g.*, substantially free from substances that limit its effect or produce undesired side-effects). The subject is preferably a mammal, and most preferably a human.

[0055] Various delivery systems are known and can be used to administer an agent of the invention, *e.g.*, encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the compound, receptor-mediated endocytosis (*see, e.g.*, Wu and Wu, 1987, J. Biol. Chem. 262:4429-4432), construction of a nucleic acid as part of a retroviral or other vector, etc. Methods of introduction can be enteral or parenteral and include but are not limited to intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, intraocular, and oral routes. The compounds may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (*e.g.*, oral mucosa, rectal and intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local. Administration can be acute or chronic (*e.g.* daily, weekly, monthly, etc.) or in combination with other agents. Pulmonary administration can also be employed,

e.g., by use of an inhaler or nebulizer, and formulation with an aerosolizing agent.

[0056] In another embodiment, the active agent can be delivered in a vesicle, in particular a liposome, in a controlled release system, or in a pump. In another embodiment where the active agent of the invention is a nucleic acid encoding a protein, the nucleic acid can be administered *in vivo* to promote expression of its encoded protein, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, *e.g.*, by use of a retroviral vector (see, for example, U.S. Patent No. 4,980,286), by direct injection, or by use of microparticle bombardment, or coating with lipids or cell-surface receptors or transfecting agents, or by administering it in linkage to a homeobox-like peptide which is known to enter the nucleus (see *e.g.*, Joliot et al., 1991, Proc. Natl. Acad. Sci. USA 88:1864-1868), etc. Alternatively, a nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination.

[0057] In a specific embodiment, it may be desirable to administer the pharmaceutical compositions of the invention locally to the area in need of treatment; this may be achieved, for example, and not by way of limitation, by local infusion during surgery, topical application, *e.g.*, by injection, by means of a catheter, or by means of an implant, the implant being of a porous, non-porous, or gelatinous material, including membranes, such as sialastic membranes, fibers, or commercial skin substitutes.

[0058] A composition useful in practicing the methods of the invention may be a liquid comprising an agent of the invention in solution, in suspension, or both. The term "solution/suspension" refers to a liquid composition where a first portion of the active agent is present in solution and a second portion of the active agent is present in particulate form, in suspension in a liquid matrix. A liquid composition also includes a gel. The liquid composition may be aqueous or in the form of an ointment. Further, the composition can take the form of a solid article that can be inserted in the eye, such as for example between the eye and eyelid or in the conjunctival sac, where the VEGF trap is released. Release from such an article is usually to the cornea, either via the lacrimal fluid, or directly to the cornea itself, with which the solid article is generally in direct contact. Solid articles suitable for implantation in the eye are generally composed primarily of bioerodible or nonbioerodible polymers. An aqueous solution and/or suspension can be in the form of eye drops. A desired dosage of the active agent can be measured by administration of a known number of drops into the eye. For example, for a drop volume of 25 μ l, administration of 1-6 drops will deliver 25-150 μ l of the composition.

[0059] An aqueous suspension or solution/suspension useful for practicing the methods of the invention may contain one or more polymers as suspending agents. Useful polymers include water-soluble polymers such as cellulosic polymers and water-insoluble polymers such as cross-linked carboxyl-containing polymers. An aqueous suspension or solution/suspension of the present invention is preferably viscous or muco-adhesive, or even more preferably, both viscous or mucoadhesive.

[0060] In another embodiment, the composition useful in practicing the methods of the invention is an *in situ* gellable aqueous composition. Such a composition comprises a gelling agent in a concentration effective to promote gelling upon contact with the eye or with lacrimal fluid. Suitable

gelling agents include but are not limited to thermosetting polymers. The term "*in situ* gellable" as used herein includes not only liquids of low viscosity that form gels upon contact with the eye or with lacrimal fluid, but also includes more viscous liquids such as semi-fluid and thixotropic gels that exhibit substantially increased viscosity or gel stiffness upon administration to the eye.

Diagnostic and Screening Methods

[0061] The VEGF traps of the invention may be used diagnostically and/or in screening methods. For example, the trap may be used to monitor levels of VEGF during a clinical study to evaluate treatment efficacy. In another embodiment, the methods and compositions of the present invention are used to screen individuals for entry into a clinical study to identify individuals having, for example, too high or too low a level of VEGF. The traps can be used in methods known in the art relating to the localization and activity of VEGF, *e.g.*, imaging, measuring levels thereof in appropriate physiological samples, in diagnostic methods, etc.

[0062] The traps of the invention may be used in *in vivo* and *in vitro* screening assay to quantify the amount of non-bound VEGF present, *e.g.*, for example, in a screening method to identify test agents able to decrease the expression of VEGF. More generally, the traps of the invention may be used in any assay or process in which quantification and/or isolation of VEGF is desired.

Pharmaceutical Compositions

[0063] The present invention also provides pharmaceutical compositions comprising a VEGF mini-trap of the invention. Such compositions comprise a therapeutically effective amount of one or more mini-traps, and a pharmaceutically acceptable carrier. The term "pharmaceutically acceptable" means approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, and more particularly, in humans. The term "carrier" refers to a diluent, adjuvant, excipient, or vehicle with which the therapeutic is administered. Such pharmaceutical carriers can be sterile liquids, such as water and oils, including those of petroleum, animal, vegetable or synthetic origin, such as peanut oil, soybean oil, mineral oil, sesame oil and the like. Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol and the like. The composition, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. These compositions can take the form of solutions, suspensions, emulsion, tablets, pills, capsules, powders, sustained-release formulations and the like. Examples of suitable pharmaceutical carriers are described in "Remington's Pharmaceutical Sciences" by E.W. Martin.

[0064] The VEGF mini-trap of the invention can be formulated as neutral or salt forms. Pharmaceutically acceptable salts include those formed with free amino groups such as those derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, etc., and those formed with free carboxyl groups such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2-ethylamino ethanol, histidine, procaine, etc.

[0065] Further more, aqueous compositions useful for practicing the methods of the invention have ophthalmically compatible pH and osmolality. One or more ophthalmically acceptable pH adjusting agents and/or buffering agents can be included in a composition of the invention, including acids such as acetic, boric, citric, lactic, phosphoric and hydrochloric acids; bases such as sodium hydroxide, sodium phosphate, sodium borate, sodium citrate, sodium acetate, and sodium lactate; and buffers such as citrate/dextrose, sodium bicarbonate and ammonium chloride. Such acids, bases, and buffers are included in an amount required to maintain pH of the composition in an ophthalmically acceptable range. One or more ophthalmically acceptable salts can be included in the composition in an amount sufficient to bring osmolality of the composition into an ophthalmically acceptable range. Such salts include those having sodium, potassium or ammonium cations and chloride, citrate, ascorbate, borate, phosphate, bicarbonate, sulfate, thiosulfate or bisulfite anions.

[0066] The amount of the trap that will be effective for its intended therapeutic use can be determined by standard clinical techniques based on the present description. In addition, *in vitro* assays may optionally be employed to help identify optimal dosage ranges. Generally, suitable dosage ranges for intravenous administration are generally about 50-5000 micrograms of active compound per kilogram body weight. Suitable dosage ranges for intranasal administration are generally about 0.01 pg/kg body weight to 1 mg/kg body weight. Effective doses may be extrapolated from dose-response curves derived from *in vitro* or animal model test systems.

[0067] For systemic administration, a therapeutically effective dose can be estimated initially from *in vitro* assays. For example, a dose can be formulated in animal models to achieve a circulating concentration range that includes the IC₅₀ as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Initial dosages can also be estimated from *in vivo* data, e.g., animal models, using techniques that are well known in the art. One having ordinary skill in the art could readily optimize administration to humans based on animal data.

[0068] Dosage amount and interval may be adjusted individually to provide plasma levels of the compounds that are sufficient to maintain therapeutic effect. In cases of local administration or selective uptake, the effective local concentration of the compounds may not be related to plasma concentration. One having skill in the art will be able to optimize therapeutically effective local dosages without undue experimentation.

[0069] The amount of compound administered will, of course, be dependent on the subject being treated, on the subject's weight, the severity of the affliction, the manner of administration, and the judgment of the prescribing physician. The therapy may be repeated intermittently while symptoms are detectable or even when they are not detectable. The therapy may be provided alone or in combination with other drugs.

Cellular Transfection and Gene Therapy

[0070] The present invention encompasses the use of nucleic acids encoding the fusion polypeptides of the invention for transfection of cells *in vitro* and *in vivo*. These nucleic acids can be inserted into any of a number of well-known vectors for transfection of target cells and organisms. The nucleic acids are transfected into cells *ex vivo* and *in vivo*, through the interaction of the vector and the

target cell. The compositions are administered (e.g., by injection into a muscle) to a subject in an amount sufficient to elicit a therapeutic response. An amount adequate to accomplish this is defined as "a therapeutically effective dose or amount."

[0071] In another aspect, the invention provides a method of reducing VEGF levels in a human or other animal comprising transfecting a cell with a nucleic acid encoding a fusion polypeptide of the invention, wherein the nucleic acid comprises an inducible promoter operably linked to the nucleic acid encoding the fusion polypeptide or mini-trap. For gene therapy procedures in the treatment or prevention of human disease, see for example, Van Brunt (1998) *Biotechnology* 6:1149-1154.

Kits

[0072] The invention also provides an article of manufacturing comprising packaging material and a pharmaceutical agent contained within the packaging material, wherein the pharmaceutical agent comprises at least one VEGF trap composed of two or more fusion polypeptides of the invention, and wherein the packaging material comprises a label or package insert which indicates that the VEGF-specific fusion polypeptide can be used for treating a VEGF-mediated disease or condition.

Transgenic Animals

[0073] The invention includes transgenic non-human animals expressing a trap of the invention. A transgenic animal can be produced by introducing nucleic acid into the male pronuclei of a fertilized oocyte, e.g., by microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. Any of the regulatory or other sequences useful in expression vectors can form part of the transgenic sequence. A tissue-specific regulatory sequence(s) can be operably linked to the transgene to direct expression of the transgene to particular cells. A transgenic non-human animal expressing a fusion polypeptide or mini-trap of the invention is useful in a variety of applications, including as a means of producing such a fusion polypeptide.. Further, the transgene may be placed under the control of an inducible promoter such that expression of the fusion polypeptide or mini-trap may be controlled by, for example, administration of a small molecule.

Specific Embodiments

[0074] In the experiments described below, smaller VEGF traps were generated and their ability to bind VEGF was investigated. Such mini-traps are preferably uses in specific applications. For example, certain conditions or diseases may be preferably treated with local administration of a VEGF trap to a specific organ, tissue, or cell, rather than by systemic administration. In one exemplification of the mini-traps of the invention, a smaller VEGF trap was generated by directed cleavage of a dimerized VEGF trap having a cleavage region (C-region) generated in a Fc domain (Example 2). The truncated trap exhibited comparable affinity for VEGF and half-life as the full-sized parent trap. Examples 3-5 describe construction of fusion polypeptides having a VEGF receptor component and a multimerizing component consisting of one or two cysteine residues. Affinity measurements showed that the non-glycosylated fusion polypeptides expressed in *E. coli* or

the glycosylated polypeptides expressed in CHO cells had comparable binding affinity for VEGF as the full-sized parent trap. Example 6 further illustrates a monomeric VEGF trap consisting of (R1R2)₂ which is capable of binding and inhibiting VEGF. Example 7 describes the construction of a VEGF mini-trap (SEQ ID NO:26) exhibiting high affinity binding for VEGF comparable to the full length trap (SEQ ID NO:10).

[0075] Other features of the invention will become apparent in the course of the following descriptions of exemplary embodiments which are given for illustration of the invention and are not intended to be limiting thereof.

EXAMPLES

[0076] The following example is put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the methods and compositions of the invention, and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers used (e.g., amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees Centigrade, and pressure is at or near atmospheric.

Example 1. Construction of Flt1D2.Flk1D3.FcΔC1(a)

[0077] The construction of a parent VEGF trap, Flt1D2.Flk1D3.FcΔC1(a) (SEQ ID NOs:7-8), VEGFR1R2.FcΔC1(a) (SEQ ID NOs:9-10), and Flt1D2.VEGFR3D3.FcΔC1(a) (SEQ ID NOs:12-13) is described in detail in PCT publication WO/0075319, herein specifically incorporated by reference in its entirety. Also described in WO/0075319 are methods of constructing and expressing nucleic acid constructs encoding VEGF traps, methods of detecting and measuring VEGF trap binding to VEGF, methods of determining the stoichiometry of VEGF binding by BIAcore analysis, and pharmacokinetic analyses.

Example 2: Thrombin-cleaved dimeric VEGF mini-trap

[0078] The VEGFR1R2.FcΔC1(a) (SEQ ID NOs:9-10) construct was modified by insertion of a thrombin cleavage following the CPPC (SEQ ID NO:1) of the Fc domain. Purified VEGF trap (5 μg) was incubated with thrombin (Novagen) in 20 mM Tris-HCl, pH 8.4, 50 mM NaCl, 2.5 mM CaCl₂ for 16 hrs at 37° C. Controls included cleavage control protein (CCP) and parent VEGF trap protein incubated without thrombin. SDS-PAGE analysis (Tris-Glycine 4-20% gel; 5 μg protein per lane) verified correct cleavage (results not shown).

[0079] Affinity determination. The K_d of binding of each VEGF trap to hVEGF₁₆₅ was determined as described in WO/0075319, for the parent VEGF trap, uncleaved VEGF trap containing a thrombin cleavage site ("uncleaved VEGF trap"), cleaved VEGF mini-trap and recombinant monomeric R1R2-myc myc his. More specifically, the ability of the traps to block VEGF₁₆₅-dependent receptor phosphorylation was determined using primary human endothelial cells (HUVECs). VEGF₁₆₅ was incubated in the presence of varying concentrations of the test traps, and the mixture was added to

HUVECs to stimulate tyrosine phosphorylation of VEGFR2. At sub-stoichiometric concentrations of VEGF trap, unbound VEGF induced receptor phosphorylation. However, at a 1:1 molar ratio of greater of a VEGF trap to ligand, complete blocking of receptor signaling was observed, establishing that a single molecule of a trap dimer is capable of blocking a single molecule of human VEGF₁₆₅. Thus, the high binding affinity of the VEGF trap for VEGF results in formation of a complex that prevents VEGF from interaction with cell surface receptors. Equivalent results were obtained for identical phosphorylation inhibition experiments for the parent VEGF trap, uncleaved VEGF trap, and cleaved VEGF mini-trap. The results are shown in Table 1.

TABLE 1

Trap	Kinetic Dissociation Rate (1/s)	T _{1/2} (hr)
parent VEGF trap	5.51 x 10 ⁻³ ± 0.94%	3.5
uncleaved VEGF trap	4.93 x 10 ⁻³ ± 0.70%	3.9
cleaved VEGF mini-trap	5.46 x 10 ⁻³ ± 0.62%	3.53
R1R2-myc myc his monomer	6.74 x 10 ⁻³ ± 0.38%	0.028

Example 3. Construction of Plasmids Encoding VEGF Mini-Traps

[0080] VEGF mini-traps were constructed from a precursor of the parent VEGF trap, VEGFR1R2.FcΔC1(a) (SEQ ID NOs:9-10), in which the three amino acids glycine-alanine-proline served as a linker between the Flk1 D3 and FcΔC1(a). This plasmid, pTE115 was used in the construction of the VEGF mini-traps because the linker DNA sequence included a Srf I restriction endonuclease recognition sequence that facilitated engineering the VEGF trap. In all other respects, the VEGF trap encoded by pTE115 is identical to that of the VEGF trap, VEGFR1R2.FcΔC1(a) (SEQ ID NOs:9-10) described in detail in PCT publication WO/0075319.

[0081] Two VEGF mini-traps were constructed with multimerization domains consisting of either a single cysteine residue (R1R2_C) (SEQ ID NO:2) or the amino acids ACGC (SEQ ID NO:4) (R1R2_{ACGC}) (SEQ ID NO:5) added to the C-terminus of receptor components Flt1D2.Flk1D3. Both of these constructs are capable of forming homo-dimeric molecules stabilized by one (R1R2_C) or two (R1R2_{ACGC}) intermolecular disulfides.

[0082] The plasmid pTE517 was made by removing the 690 bp fragment generated by digestion of pTE115 DNA with Srf I and Not I and inserting the synthetic DNA fragment formed by annealing the oligos R1R2NC (SEQ ID NO:14) and R1R2CC (SEQ ID NO:15). The resulting plasmid encodes R1R2_C, which consists of the Flt1D2.Flk1D3 domains followed by a cysteine residue (SEQ ID NO:23). Similarly, the plasmid pTE518 was made by removing the 690 bp fragment generated by digestion of pTE115 DNA with Srf I and Not I, followed by ligation with the synthetic DNA fragment formed by annealing the oligos R1R2NACGC (SEQ ID NO:16) and R1R2CACGC (SEQ ID NO:17). The resulting plasmid encodes R1R2_{ACGC}, which consists of the Flt1D2.Flk1D3 domains followed by the amino acids ACGC (SEQ ID NO:25).

[0083] Plasmids were also constructed to direct the expression of these mini-traps in *E. coli*. The primers R1R2N-NcoI (SEQ ID NO:18) and R1R2CNotI (SEQ ID NO:19) were used to amplify a DNA fragment from pTE115 that encodes amino acids G30 to K231, relative to the parental VEGF trap (SEQ ID NO:10). Amplification of this sequence resulted in fusion of an initiating methionine

codon at the 5' end and fusion of the codon for cysteine, followed by a stop codon, at the 3' end (SEQ ID NO:2). This DNA fragment was then cloned into the Nco I and Not I sites of the *E. coli* expression plasmid pRG663 to yield pRG1102 such that expression of R1R2_C was dependent on transcription from the phage T7 Φ 1.1 promoter. Induction of gene expression from pRG1102 results in accumulation of R1R2_{cys} in the cytoplasm of the *E. coli* host strain RFJ238. Similarly, the primers R1R2N-Nco1 (SEQ ID NO:18) and R1R2ACGC-N ot1 (SEQ ID NO:20) were used to amplify a DNA fragment from pTE115 that encodes amino acids G30 to K231 (SEQ ID NO:10) resulting in fusion of an initiating methionine codon at the 5' end and fusion of codons for ACGC (SEQ ID NO:4), followed by a stop codon, at the 3' end (SEQ ID NO:5). This fragment was then cloned into the Nco I and Not I sites of the *E. coli* expression plasmid pRG663 to yield pRG1103 such that expression of R1R2_{ACGC} was dependent on transcription from the phage T7 Φ 1.1 promoter. Induction of gene expression from both pRG1102 and pRG1103 resulted in accumulation of R1R2_C or R1R2_{ACGC}, respectively, in the cytoplasm of the *E. coli* host strain RFJ238.

Example 4. Purification and characterization of VEGF mini-traps from *E. coli*

[0084] Both R1R2_C and R1R2_{ACGC} were expressed as cytoplasmic proteins in *E. coli* and were purified by the same method. Induction of the phage T7 Φ 1.1 promoter on either pRG1102 or pRG1103 in the *E. coli* K12 strain RFJ238 resulted in accumulation of the protein in the cytoplasm. After induction, cells were collected by centrifugation, resuspended in 50 mM Tris-HCl, pH 7.5, 20 mM EDTA, and lysed by passage through a Niro-Soavi cell homogenizer. Inclusion bodies were collected from lysed cells by centrifugation, washed once in distilled H₂O, then solubilized in 8 M guanidinium-HCl, 50 mM Tris-HCl, pH 8.5, 100 mM sodium sulfite, 10 mM sodium tetrathionate and incubated at room temperature for 16 hours. Clarified supernatant was fractionated on an S300 column equilibrated with 6 M guanidinium-HCl, 50 mM Tris-HCl, pH 7.5. Fractions containing R1R2_C were pooled and dialyzed against 6M Urea, 50 mM Tris-HCl, pH 7.5. Dialyzed protein was diluted to 2M Urea, 50 mM Tris-HCl, pH 8.5, 2 mM cysteine then stirred slowly for 7 days at 4°C. Refolded protein was dialyzed against 50 mM Tris-HCl, pH 7.5 then loaded onto an SP-sepharose column equilibrated with 50 mM Tris-HCl, pH 7.5 and eluted with a NaCl gradient from 0 to 1 M in 50 mM Tris-HCl, pH 7.5. Fractions containing R1R2_C were pooled, concentrated, and loaded onto a Superdex 200 column equilibrated with 50 mM Tris-HCl, pH 7.5, 150 mM NaCl. Fractions containing mini-trap dimer were collected and pooled. The molecular weight of purified mini-trap was estimated to be about 46 kD by SDS-PAGE.

[0085] BIAcore assays were conducted (as described in WO/0075319) to determine trap affinity for VEGF, and the results showed that the R1R2_C and R1R2_{ACGC} mini-traps had VEGF affinity comparable to the full length VEGF trap (Table 2).

TABLE 2

Trap	Kinetic Dissociation Rate (1/s)	T _{1/2} (hr)
VEGF trap	4.23 x 10 ⁻⁵	4.53
R1R2 _C	3.39 x 10 ⁻⁵	5.68
R1R2 _{ACGC}	3.41 x 10 ⁻⁵	5.65

Example 5. Expression of VEGF mini-traps in CHO K1

[0086] Expression of the VEGF mini-traps encoded by pTE517 and pTE518 is dependent on transcription from the human CMV-MIE promoter and results in secretion of the mini-traps into the culture medium when expressed in CHO cells. When expressed as secreted proteins in CHO K1, both mini-traps were found in the conditioned media and estimation of their molecular weight by SDS-PAGE suggested, as expected, that the proteins were glycosylated. Analysis by SDS-PAGE also indicated that the mini-traps were capable of forming homo-dimeric molecules stabilized by intermolecular disulfide(s) between the C-terminal cysteine(s). Specifically, the R1R2_C mini-trap efficiently formed covalent dimers when expressed as a secreted protein in CHO cells.

Example 6. Construction and expression of a single chain VEGF mini-trap

[0087] A VEGF mini-trap was also constructed that did not require a multimerization domain (SEQ ID NO:24). This mini-trap was constructed by direct fusion of one Flt1D2.Flk1D3 domain (R1R2) (amino acids 30-231 of SEQ ID NO:24) to a second Flt1D2.Flk1D3 domain (R1R2) (amino acids 234-435 of SEQ ID NO:24) with a Gly-Pro linker between the tandem receptor domains (amino acids 232-233 of SEQ ID NO:24).

[0088] To construct a gene encoding tandem Flt1D2.Flk1D3 domains, a DNA fragment was synthesized (Blue Heron Biotechnology) that encoded one Flt1D2.Flk1D3 domain that minimized DNA homology with the Flt1D2.Flk1D3 domain-encoding DNA found in pTE115. This synthetic DNA fragment was cloned as a Srf I-Not I fragment into the Srf I-Not I sites of pTE115 to yield pTE570, which expresses the R1R2-R1R2 VEGF mini-trap from the CMV-MIE promoter. When this plasmid is transfected into CHO K1 cells the R1R2-R1R2 VEGF mini-trap accumulates in the culture medium.

Example 7. Construction and expression of a VEGF mini-trap

[0089] A VEGF mini-trap was constructed as described above, by direct fusion of one Flt1D2.Flk1D3 domain (R1R2) (amino acids 30-231 of SEQ ID NO:26) with a C-terminal nine amino acid sequence terminating in CPPC. When this plasmid is transfected into CHO K1 cells the VEGF mini-trap of SEQ ID NO:26 is secreted into the culture medium. Subsequent purification by non-reducing SDS-PAGE electrophoresis as well as native light-scattering analysis identified a trap molecule with molecular weight approximately 64 kDa. This molecular weight indicates that a covalent dimer was formed between two fusion polypeptides of SEQ ID NO:26. Similar experiments were conducted with plasmids encoding the fusion polypeptides of SEQ ID NOS:27 and 28, and similarly showed these molecules formed homodimeric traps. Affinity determinations for human VEGF-165 binding to EGF traps composed of dimers of SEQ ID NO:10 and SEQ ID NO:26 are shown in Table 3.

TABLE 3

VEGF Trap	ka (1/Ms)	kd (1/s)	KD (M)
SEQ ID NO:10	$2.73 \times 10^{+7}$	1.79×10^{-5}	6.55×10^{-13}
SEQ ID NO:26	$2.00 \times 10^{+7}$	6.56×10^{-6}	3.28×10^{-13}
SEQ ID NO:26	$2.61 \times 10^{+7}$	5.77×10^{-6}	2.21×10^{-13}

We claim:

1. An isolated nucleic acid molecule encoding a fusion polypeptide consisting of components $(R1R2)_X$ or $(R1R3)_Y$, and a fusion partner (FP), wherein $X \geq 1$, $Y \geq 1$, R1 is vascular endothelial cell growth factor (VEGF) receptor component Ig domain 2 of Flt-1 and R2 is Ig domain 3 of Flk-1, R3 is Ig domain 3 of Flt-4.
2. The isolated nucleic acid of claim 1, wherein the fusion partner (FP) is a multimerizing component (MC) capable of interacting with another MC to form a multimeric structure.
3. The isolated nucleic acid of claim 3, wherein the MC is selected from the group consisting of (i) a multimerizing component comprising a cleavable region (C-region), (ii) a truncated multimerizing component, (iii) an amino acid sequence between 1 to about 200 amino acids in length having at least one cysteine residue, (iv) a leucine zipper, (v) a helix loop motif, (vi) a coil-coil motif, and (vii) an immunoglobulin domain.
4. A fusion polypeptide encoded by the nucleic acid molecule of claims 1 to 3.
5. The fusion polypeptide of claim 4, having the amino acid sequence of SEQ ID NO:26, 27, or 28.
6. A replicable expression vector capable in a transformed host cell comprising the nucleic acid molecule of claims 1 to 3.
7. A method of producing a VEGF fusion polypeptide, comprising the steps of introducing into a suitable expression system the expression vector of claim 6, and effecting expression of the VEGF fusion polypeptide.
8. A vascular endothelial cell growth factor (VEGF) trap, comprising a multimer of two or more fusion polypeptides of claim 4.
9. The VEGF trap of claim 8, which is a dimer.
10. A dimeric VEGF trap comprising two fusion polypeptides comprising the amino acid sequence of SEQ ID NO:26, 27, or 28.
11. A pharmaceutical composition comprising the fusion polypeptide of claims 8 or 9, and a pharmaceutically acceptable carrier.

12. A method of treating a disease or condition which is improved, ameliorated, or inhibited by removal or inhibition of vascular endothelial growth factor (VEGF), comprising administering the pharmaceutical composition of claim 11 to a subject in need thereof.
13. The method of claim 12, wherein the disease or condition is an ocular disease or condition.
14. The method of claim 13, wherein the ocular disease or condition is age related macular degeneration.
15. An isolated nucleic acid molecule encoding a fusion polypeptide consisting of receptor components (R1R2)_x or (R1R3)_y, and a multimerizing component (MC) capable of interacting with another MC to form a multimeric structure, wherein $X \geq 1$, $Y \geq 1$, R1 is vascular endothelial cell growth factor (VEGF) receptor component Ig domain 2 of Flt-1 and R2 is Ig domain 3 of Flk-1, R3 is Ig domain 3 of Flt-4, wherein the multimerizing component (MC) is selected from the group consisting of (i) a MC comprising a cleavable region (C-region), (ii) a truncated MC, (iii) an amino acid sequence between 1 to about 200 amino acids in length having at least one cysteine residue, (iv) a leucine zipper, (v) a helix loop motif, (vi) a coil-coil motif, and (vii) an immunoglobulin domain.
16. The isolated nucleic acid molecule of claim 15, wherein the receptor components are (R1R2)_x and the multimerizing component is an amino acid sequence between 1 to about 200 amino acids in length having at least one cysteine residue.
17. The isolated nucleic acid molecule of claim 16, wherein the receptor component is R1R2, X is 1, and the multimerizing component is an amino acid sequence 1-15 amino acids in length with 1-2 cysteine residues.
18. A fusion polypeptide capable of binding vascular endothelial growth factor (VEGF) encoded by the nucleic acid molecule of claims 15 to 17.
19. The fusion polypeptide of claim 18, comprising the amino acid sequence of SEQ ID NO:26, 27 or 28.
20. A fusion polypeptide consisting of receptor components (R1R2)_x or (R1R3)_y, and a multimerizing component (MC) capable of interacting with another MC to form a multimeric structure, wherein $X \geq 1$, $Y \geq 1$, R1 is vascular endothelial cell growth factor (VEGF) receptor component Ig domain 2 of Flt-1 and R2 is Ig domain 3 of Flk-1, R3 is Ig domain 3 of Flt-4, wherein the multimerizing component (MC) is selected from the group consisting of (i) a MC comprising a cleavable region (C-region), (ii) a truncated MC, (iii) an amino acid sequence between 1 to about 200 amino acids in length having at least one cysteine residue, (iv) a leucine zipper, (v) a helix loop motif, (vi) a coil-coil motif, and (vii) an immunoglobulin domain.

21. The fusion polypeptide of claim 20, wherein the receptor components are $(R1R2)_X$ and the multimerizing component is an amino acid sequence between 1 to about 200 amino acids in length having at least one cysteine residue.

22. The fusion polypeptide of claim 21, wherein the receptor component is $R1R2$, X is 1, and the multimerizing component is an amino acid sequence 1-15 amino acids in length with 1-2 cysteine residues.

23. A dimeric VEGF trap composed of two of the fusion polypeptides of claims 20 to 22.

24. An article of manufacturing comprising:

(a) packaging material; and

(b) a pharmaceutical agent contained within said packaging material;

wherein the pharmaceutical agent comprises at least one VEGF trap consisting of receptor components $(R1R2)_X$ or $(R1R3)_Y$, and a multimerizing component (MC) capable of interacting with another MC to form a multimeric structure, wherein $X \geq 1$, $Y \geq 1$, and wherein the packaging material comprises a label or package insert which indicates that said VEGF-specific fusion polypeptide can be used for treating a VEGF-mediated disease or condition.

SEQUENCE LISTING

<110> Daly, Thomas J.
 Fandl, James P.
 Papadopoulos, Nicholas J.

<120> VEGF TRAPS AND THERAPEUTIC USES THEREOF

<130> 710D2-WO

<140> to be assigned

<141> 2004-06-29

<150> 10/609,775

<151> 2003-06-30

<160> 29

<170> FastSEQ for Windows Version 4.0

<210> 1

<211> 4

<212> PRT

<213> homo sapiens

<400> 1

Cys Pro Pro Cys

1

<210> 2

<211> 200

<212> PRT

<213> homo sapiens

<400> 2

Met	Gly	Arg	Pro	Phe	Val	Glu	Met	Tyr	Ser	Glu	Ile	Pro	Glu	Ile	Ile
1				5					10					15	
His	Met	Thr	Glu	Gly	Arg	Glu	Leu	Val	Ile	Pro	Cys	Arg	Val	Thr	Ser
			20					25					30		
Pro	Asn	Ile	Thr	Val	Thr	Leu	Lys	Lys	Phe	Pro	Leu	Asp	Thr	Leu	Ile
		35					40					45			
Pro	Asp	Gly	Lys	Arg	Ile	Ile	Trp	Asp	Ser	Arg	Lys	Gly	Phe	Ile	Ile
	50					55					60				
Ser	Asn	Ala	Thr	Tyr	Lys	Glu	Ile	Gly	Leu	Leu	Thr	Cys	Glu	Ala	Thr
65					70					75					80
Val	Asn	Gly	His	Leu	Tyr	Lys	Thr	Asn	Tyr	Leu	Thr	Gln	Thr	Asn	Thr
			85						90					95	
Ile	Ile	Asp	Val	Val	Leu	Ser	Pro	Ser	His	Gly	Ile	Glu	Leu	Ser	Val
			100					105					110		
Gly	Glu	Lys	Leu	Val	Leu	Asn	Cys	Thr	Ala	Arg	Thr	Glu	Leu	Asn	Val
		115					120					125			
Gly	Ile	Asp	Phe	Asn	Trp	Glu	Tyr	Pro	Ser	Ser	Lys	His	Gln	His	Lys
	130					135					140				
Lys	Leu	Val	Asn	Arg	Asp	Leu	Lys	Thr	Gln	Ser	Gly	Ser	Glu	Met	Lys
145					150					155					160
Lys	Phe	Leu	Ser	Thr	Leu	Thr	Ile	Asp	Gly	Val	Thr	Arg	Ser	Asp	Gln
				165					170					175	
Gly	Thr	Cys	Ala	Ala	Ser	Ser	Gly	Leu	Met	Thr	Lys	Lys	Asn	Ser	Thr
			180					185						190	

Phe Val Arg Val His Glu Lys Cys
 195 200

<210> 3
 <211> 4
 <212> PRT
 <213> homo sapiens

<220>
 <221> VARIANT
 <222> 1, 3
 <223> Xaa = Any Amino Acid

<400> 3
 Xaa Cys Xaa Cys
 1

<210> 4
 <211> 4
 <212> PRT
 <213> homo sapiens

<400> 4
 Ala Cys Gly Cys
 1

<210> 5
 <211> 203
 <212> PRT
 <213> homo sapiens

<400> 5
 Met Gly Arg Pro Phe Val Glu Met Tyr Ser Glu Ile Pro Glu Ile Ile
 1 5 10 15
 His Met Thr Glu Gly Arg Glu Leu Val Ile Pro Cys Arg Val Thr Ser
 20 25 30
 Pro Asn Ile Thr Val Thr Leu Lys Phe Pro Leu Asp Thr Leu Ile
 35 40 45
 Pro Asp Gly Lys Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe Ile Ile
 50 55 60
 Ser Asn Ala Thr Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu Ala Thr
 65 70 75 80
 Val Asn Gly His Leu Tyr Lys Thr Asn Tyr Leu Thr Gln Thr Asn Thr
 85 90 95
 Ile Ile Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val
 100 105 110
 Gly Glu Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val
 115 120 125
 Gly Ile Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys
 130 135 140
 Lys Leu Val Asn Arg Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys
 145 150 155 160
 Lys Phe Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln
 165 170 175
 Gly Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 180 185 190
 Phe Val Arg Val His Glu Lys Ala Cys Gly Cys
 195 200

<210> 6

<211> 6
 <212> PRT
 <213> homo sapiens

<400> 6
 Leu Val Pro Arg Gly Ser
 1 5

<210> 7
 <211> 1453
 <212> DNA
 <213> homo sapiens

<400> 7
 aagcttggggc tgcaggtcga tgcactctag aggatcgatc cccggggcgag ctcgaattcg 60
 caaccaccat ggtcagctac tgggacaccg gggtcctgct gtgcgcgctg ctcagctgtc 120
 tgcttctcac aggatctagt tccggaggta gacctttcgt agagatgtac agtgaatcc 180
 ccgaaattat acacatgact gaaggaaggg agctcgtcat tocctgcccg gttacgtcac 240
 ctaacatcac tgttacttta aaaaagtctc cacttgacac tttgatccct gatggaaaac 300
 gcataatctg ggacagtaga aagggcttca tcatatcaaa tgcaacgtac aaagaaatag 360
 ggcttctgac ctgtgaagca acagtcaatg ggcatttcta taagacaac tatctcacac 420
 atcgacaaac caatacaatc atagatgtgg ttctgagtcg gtctcatgga attgaactat 480
 ctgttgagaa aaagcttgct ttaaattgta cagcaagaac tgaactaaat gtgggggattg 540
 acttcaactg ggaataccct tcttcgaagc atcagcataa gaaacttgta aaccgagacc 600
 taaaaaccca gtctgggagt gagatgaaga aatTTTTTgag caccttaact atagatgggtg 660
 taaccgggag tgaccaagga ttgtacacct gtgcagcatc cagtgggctg atgaccaaga 720
 agaacagcac atttgtcagg gtccatgaaa agggcccggg cgacaaaact cacacatgcc 780
 caccgtgccc agcacctgaa ctccctggggg gaccgtcagt cttcctcttc cccccaaaac 840
 ccaaggacac cctcatgata tcccggaccg ctgaggtcac atgcgtgggtg gtggacgtga 900
 gccacgaaga cctgaggtc aagtcaact ggtacgtgga cggcgtggag gtgcataatg 960
 ccaagacaaa gccgcgggag gagcagtaga acagcacgta ccgtgtggtc agcgtcctca 1020
 ccgtcctgca ccaggactgg ctgaatggca aggagtacaa gtgcaaggtc tccaacaaag 1080
 ccctcccagc ccccatcgag aaaaccatct ccaaagccaa agggcagccc cgagaaccac 1140
 aggtgtacac cctgccccca tcccgggatg agctgaccaa gaaccaggtc agcctgacct 1200
 gcctgggtcaa aggttctat cccagcgaca tcgccgtgga gtgggagagc aatgggcagc 1260
 cggagaacaa ctacaagacc acgcctcccg tgctggactc cgacggctcc ttcttctct 1320
 atagcaagct caccgtggac aagagcaggt ggcagcaggg gaacgtcttc tcatgctccg 1380
 tgatgcatga ggctctgcac aaccactaca cgcagaagag cctctccctg tctccgggta 1440
 aatgagcggc cgc 1453

<210> 8
 <211> 458
 <212> PRT
 <213> homo sapiens

<400> 8
 Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1 5 10 15
 Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Gly Arg Pro Phe Val Glu
 20 25 30
 Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu Gly Arg Glu
 35 40 45
 Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr Val Thr Leu
 50 55 60
 Lys Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys Arg Ile Ile
 65 70 75 80
 Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr Tyr Lys Glu
 85 90 95
 Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His Leu Tyr Lys
 100 105 110

Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile Asp Val Val
 115 120 125
 Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu Lys Leu Val
 130 135 140
 Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile Asp Phe Asn
 145 150 155 160
 Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu Val Asn Arg
 165 170 175
 Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe Leu Ser Thr
 180 185 190
 Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu Tyr Thr Cys
 195 200 205
 Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr Phe Val Arg
 210 215 220
 Val His Glu Lys Gly Pro Gly Asp Lys Thr His Thr Cys Pro Pro Cys
 225 230 235 240
 Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro
 245 250 255
 Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys
 260 265 270
 Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp
 275 280 285
 Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu
 290 295 300
 Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu
 305 310 315 320
 His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn
 325 330 335
 Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly
 340 345 350
 Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu
 355 360 365
 Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr
 370 375 380
 Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn
 385 390 395 400
 Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe
 405 410 415
 Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
 420 425 430
 Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr
 435 440 445
 Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
 450 455

<210> 9
 <211> 1377
 <212> DNA
 <213> homo sapiens

<400> 9
 atggtcagct actgggacac cggggctcctg ctgtgcgcgc tgctcagctg tctgcttctc 60
 acaggatcta gttccggaag tgataccggt agacctttcg tagagatgta cagtgaaatc 120
 cccgaaatta tacacatgac tgaaggaagg gagctcgtca ttccctgccg ggttacgtca 180
 cctaacaatca ctgttacttt aaaaaagttt ccaattgaca ctttgatccc tgatggaaaa 240
 cgcataatct gggacagtag aaagggttc atcatatcaa atgcaacgta caaagaaata 300
 gggcttctga cctgtgaagc aacagtcaat gggcatttgt ataagacaaa ctatctcaca 360
 catcgacaaa ccaatacaat catagatgtg gttctgagtc cgtctcatgg aattgaacta 420
 tctgttgag aaaagcttgt cttaaattgt acagcaagaa ctgaaactaaa tgtggggatt 480
 gacttcaact ggggaataccc ttcttcgaag catcagcata agaaacttgt aaaccgagac 540
 ctaaaaaccc agtctgggag tgagatgaag aaatTTTTga gcaccttaac tatagatggt 600

gtaaccgga gtgaccaagg attgtacacc tgtgcagcat ccagtgggct gatgaccaag 660
 aagaacagca catttgtcag ggtccatgaa aaggacaaaa ctcacacatg cccaccgtgc 720
 ccagcacctg aactcctggg gggaccgtca gtcttcctct tcccccaaa acccaaggac 780
 accctcatga tctcccggac ccctgaggtc acatgcgtgg tgggtggacgt gagccacgaa 840
 gaccctgagg tcaagttcaa ctggtacgtg gacggcgtgg aggtgcataa tgccaagaca 900
 aagccgcggg aggagcagta caacagcacg taccgtgtgg tcagcgtcct caccgtcctg 960
 caccaggact ggctgaatgg caaggagtac aagtgcaagg tctccaacaa agccctccca 1020
 gccccatcg agaaaaccat ctccaaagcc aaagggcagc cccgagaacc acaggtgtac 1080
 accctgcccc catcccggga tgagctgacc aagaaccagg tcagcctgac ctgctgtggc 1140
 aaaggcttct atcccagcga catcgccgtg gagtgggaga gcaatgggca gccggagaac 1200
 aactacaaga ccacgcctcc cgtgctggac tccgacggct ccttcttctc ctacagcaag 1260
 ctaccctggt acaagagcag gtggcagcag gggaacgtct tctcatgctc cgtgatgcat 1320
 gaggctctgc acaaccacta cacgcagaag agcctctccc tgtctccggg taaatga 1377

<210> 10
 <211> 458
 <212> PRT
 <213> homo sapiens

<400> 10
 Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1 5 10 15
 Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Ser Asp Thr Gly Arg Pro
 20 25 30
 Phe Val Glu Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu
 35 40 45
 Gly Arg Glu Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr
 50 55 60
 Val Thr Leu Lys Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys
 65 70 75 80
 Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr
 85 90 95
 Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His
 100 105 110
 Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile
 115 120 125
 Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu
 130 135 140
 Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile
 145 150 155 160
 Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu
 165 170 175
 Val Asn Arg Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe
 180 185 190
 Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu
 195 200 205
 Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 210 215 220
 Phe Val Arg Val His Glu Lys Asp Lys Thr His Thr Cys Pro Pro Cys
 225 230 235 240
 Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro
 245 250 255
 Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys
 260 265 270
 Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp
 275 280 285
 Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu
 290 295 300
 Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu
 305 310 315 320

His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn
 325 330 335
 Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly
 340 345 350
 Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu
 355 360 365
 Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr
 370 375 380
 Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn
 385 390 395 400
 Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe
 405 410 415
 Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn
 420 425 430
 Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr
 435 440 445
 Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
 450 455

<210> 11
 <211> 4
 <212> PRT
 <213> homo sapiens

<220>
 <221> VARIANT
 <222> 1, 2, 3, 4
 <223> Xaa = Any Amino Acid

<400> 11
 Xaa Xaa Xaa Xaa
 1

<210> 12
 <211> 1444
 <212> DNA
 <213> homo sapiens

<400> 12
 aagcttgggc tgcaggtcga tcgactctag aggatcgatc cccgggcgag ctcgaattcg 60
 caaccaccat ggtcagctac tgggacaccg gggctctgct gtgcgcgctg ctccagctgtc 120
 tgctttctcac aggatctagt tccggaggta gacctttcgt agagatgtac agtgaaatcc 180
 ccgaaattat acacatgact gaaggaaggg agctcgtcat tccctgcccg gttacgtcac 240
 ctaacatcac tgttácttta aaaaagtttc cacttgacac tttgatccct gatggaaaac 300
 gcataatctg ggacagtaga aagggcttca tcatatcaaa tgcaacgtac aaagaaatag 360
 ggctttctgac ctgtgaagca acagtcaatg ggcatttcta taagacaaac tatctcacac 420
 atcgacaaac caatacaatc atagatatcc agctggtgcc caggaagtcg ctggagctgc 480
 tggtagggga gaagctggtc ctcaactgca ccgtgtgggc tgagtttaac tcaggtgtca 540
 cctttgactg ggactaccca ggaagcagg cagagcgggg taagtgggtg cccgagcgac 600
 gctcccaaca gaccacaca gaactctcca gcatectgac catccacaac gtcagccagc 660
 acgacctggg ctcgatgtg tgcaaggcca acaacggcat ccagcgattt cgggagagca 720
 ccgaggtcat tgtgcatgaa aatggcccgg gcgacaaaac tcacacatgc ccaccgtgcc 780
 cagcacctga actoctgggg ggaccgtcag tcttctctct cccccaaaa cccaaggaca 840
 ccctcatgat ctcccggacc cctgaggtca catgcgtggt ggtggacgtg agccacgaag 900
 accctgaggt caagttcaac tggtagctgg acggcgtgga ggtgcataat gccaaagaca 960
 agccgcggga ggagcagtag aacagcagct accgtgtggt cagcgtcctc accgtcctgc 1020
 accagactg gctgaatggc aaggagtaca agtgcaaggt ctccaacaaa gccctcccag 1080
 cccccatcga gtaaaccatc tccaaagcca aagggcagcc ccgagaacca caggtgtaca 1140
 ccctgcccc atcccgggat gagctgacca agaaccaggt cagcctgacc tgcttggtca 1200
 aaggcttcta tcccagcgac atcgccgtgg agtgggagag caatgggcag ccggagaaca 1260

actacaagac cacgcctccc gtgctggact cgcagggctc cttcttctc tatagcaagc 1320
 tcaccgtgga caagagcagg tggcagcagg ggaacgtctt ctcattgctcc gtgatgcatg 1380
 aggctctgca caaccactac acgcagaaga gcctctcctt gtctccgggt aaatgagcgg 1440
 ccgc 1444

<210> 13
 <211> 455
 <212> PRT
 <213> homo sapiens

<400> 13
 Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1 5 10 15
 Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Gly Arg Pro Phe Val Glu
 20 25 30
 Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu Gly Arg Glu
 35 40 45
 Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr Val Thr Leu
 50 55 60
 Lys Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys Arg Ile Ile
 65 70 75 80
 Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr Tyr Lys Glu
 85 90 95
 Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His Leu Tyr Lys
 100 105 110
 Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile Asp Ile Gln
 115 120 125
 Leu Leu Pro Arg Lys Ser Leu Glu Leu Leu Val Gly Glu Lys Leu Val
 130 135 140
 Leu Asn Cys Thr Val Trp Ala Glu Phe Asn Ser Gly Val Thr Phe Asp
 145 150 155 160
 Trp Asp Tyr Pro Gly Lys Gln Ala Glu Arg Gly Lys Trp Val Pro Glu
 165 170 175
 Arg Arg Ser Gln Gln Thr His Thr Glu Leu Ser Ser Ile Leu Thr Ile
 180 185 190
 His Asn Val Ser Gln His Asp Leu Gly Ser Tyr Val Cys Lys Ala Asn
 195 200 205
 Asn Gly Ile Gln Arg Phe Arg Glu Ser Thr Glu Val Ile Val His Glu
 210 215 220
 Asn Gly Pro Gly Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro
 225 230 235 240
 Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys
 245 250 255
 Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val
 260 265 270
 Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp
 275 280 285
 Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr
 290 295 300
 Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp
 305 310 315 320
 Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu
 325 330 335
 Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg
 340 345 350
 Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys
 355 360 365
 Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp
 370 375 380
 Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys

385					390					395					400
Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	Ser
				405					410					415	
Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp	Gln	Gln	Gly	Asn	Val	Phe	Ser
			420					425					430		
Cys	Ser	Val	Met	His	Glu	Ala	Leu	His	Asn	His	Tyr	Thr	Gln	Lys	Ser
		435					440					445			
Leu	Ser	Leu	Ser	Pro	Gly	Lys									
	450					455									

<210> 14
 <211> 24
 <212> DNA
 <213> homo sapiens

<400> 14
 gggctggtga gagagagaga gagc 24

<210> 15
 <211> 28
 <212> DNA
 <213> homo sapiens

<400> 15
 ggccgctctc tctctctctc aacagccc 28

<210> 16
 <211> 23
 <212> DNA
 <213> homo sapiens

<400> 16
 gggcgcgatgc ggttggtgag agc 23

<210> 17
 <211> 27
 <212> DNA
 <213> homo sapiens

<400> 17
 ggccgctctc aacaaccgca tgcgccc 27

<210> 18
 <211> 36
 <212> DNA
 <213> homo sapiens

<400> 18
 gagagagacc atgggtagac ctttcgtaga gatgta 36

<210> 19
 <211> 48
 <212> DNA
 <213> homo sapiens

<400> 19
 agagaggcgg ccgctttatc aacacttttc atggaccctg acaaatgt 48

<210> 20
 <211> 57
 <212> DNA

```

<213> homo sapiens

<400> 20
agagaggcgg cgcgtttatc aacaaccgca tgccttttca tggaccctga caaatgt      57

<210> 21
<211> 39
<212> DNA
<213> homo sapiens

<400> 21
agttccggaa gtgccatggg tagacctttc gtagagatg      39

<210> 22
<211> 44
<212> DNA
<213> homo sapiens

<400> 22
agagaggcgg cgcgtggtat cacttctcgt gcacgcgcac gaag      44

<210> 23
<211> 235
<212> PRT
<213> homo sapiens

<400> 23
Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1          5          10          15
Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Ser Asp Thr Gly Arg Pro
 20          25          30
Phe Val Glu Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu
 35          40          45
Gly Arg Glu Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr
 50          55          60
Val Thr Leu Lys Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys
 65          70          75          80
Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr
 85          90          95
Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His
 100         105         110
Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile
 115         120         125
Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu
 130         135         140
Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile
 145         150         155         160
Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu
 165         170         175
Val Asn Thr Gln Ser Gly Ser Glu Met Lys Arg Asp Leu Lys Lys Phe
 180         185         190
Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu
 195         200         205
Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 210         215         220
Phe Val Arg Val His Glu Lys Gly Pro Gly Cys
 225         230         235

<210> 24
<211> 435

```

<212> PRT

<213> homo sapiens

<400> 24

```

Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1          5          10          15
Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Ser Asp Thr Gly Arg Pro
 20          25          30
Phe Val Glu Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu
 35          40          45
Gly Arg Glu Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr
 50          55          60
Val Thr Leu Lys Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys
 65          70          75          80
Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr
 85          90          95
Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His
 100         105         110
Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile
 115         120         125
Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu
 130         135         140
Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile
 145         150         155         160
Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu
 165         170         175
Val Asn Arg Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe
 180         185         190
Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu
 195         200         205
Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 210         215         220
Phe Val Arg Val His Glu Lys Gly Pro Gly Arg Pro Phe Val Glu Met
 225         230         235         240
Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu Gly Arg Glu Leu
 245         250         255
Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr Val Thr Leu Lys
 260         265         270
Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys Arg Ile Ile Trp
 275         280         285
Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr Tyr Lys Glu Ile
 290         295         300
Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His Leu Tyr Lys Thr
 305         310         315         320
Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile Asp Val Val Leu
 325         330         335
Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu Lys Leu Val Leu
 340         345         350
Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile Asp Phe Asn Trp
 355         360         365
Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu Val Asn Arg Asp
 370         375         380
Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe Leu Ser Thr Leu
 385         390         395         400
Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu Tyr Thr Cys Ala
 405         410         415
Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr Phe Val Arg Val
 420         425         430
His Glu Lys
 435
    
```

<210> 25
 <211> 238
 <212> PRT
 <213> homo sapiens

<400> 25
 Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1 5 10 15
 Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Ser Asp Thr Gly Arg Pro
 20 25 30
 Phe Val Glu Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu
 35 40 45
 Gly Arg Glu Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr
 50 55 60
 Val Thr Leu Lys Lys Phe Pro Leu Asn Thr Leu Ile Pro Asn Gly Lys
 65 70 75 80
 Ala Ile Ile Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr
 85 90 95
 Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His
 100 105 110
 Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile
 115 120 125
 Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu
 130 135 140
 Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile
 145 150 155 160
 Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu
 165 170 175
 Val Asn Arg Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe
 180 185 190
 Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu
 195 200 205
 Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 210 215 220
 Phe Val Arg Val His Glu Lys Gly Pro Gly Ala Cys Gly Cys
 225 230 235

<210> 26
 <211> 240
 <212> PRT
 <213> homo sapiens

<400> 26
 Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1 5 10 15
 Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Ser Asp Thr Gly Arg Pro
 20 25 30
 Phe Val Glu Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu
 35 40 45
 Gly Arg Glu Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr
 50 55 60
 Val Thr Leu Lys Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys
 65 70 75 80
 Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr
 85 90 95
 Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His
 100 105 110
 Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile
 115 120 125

Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu
 130 135 140
 Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile
 145 150 155 160
 Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu
 165 170 175
 Val Asn Arg Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe
 180 185 190
 Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu
 195 200 205
 Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 210 215 220
 Phe Val Arg Val His Glu Lys Asp Lys Thr His Thr Cys Pro Pro Cys
 225 230 235 240

<210> 27
 <211> 240
 <212> PRT
 <213> homo sapiens

<400> 27
 Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1 5 10 15
 Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Ser Asp Thr Gly Arg Pro
 20 25 30
 Phe Val Glu Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu
 35 40 45
 Gly Arg Glu Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr
 50 55 60
 Val Thr Leu Lys Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys
 65 70 75 80
 Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr
 85 90 95
 Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His
 100 105 110
 Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile
 115 120 125
 Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu
 130 135 140
 Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile
 145 150 155 160
 Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu
 165 170 175
 Val Asn Arg Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe
 180 185 190
 Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu
 195 200 205
 Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 210 215 220
 Phe Val Arg Val His Glu Lys Asp Lys Thr His Thr Ser Pro Pro Cys
 225 230 235 240

<210> 28
 <211> 237
 <212> PRT
 <213> homo sapiens

<400> 28
 Met Val Ser Tyr Trp Asp Thr Gly Val Leu Leu Cys Ala Leu Leu Ser
 1 5 10 15

Cys Leu Leu Leu Thr Gly Ser Ser Ser Gly Ser Asp Thr Gly Arg Pro
 20 25 30
 Phe Val Glu Met Tyr Ser Glu Ile Pro Glu Ile Ile His Met Thr Glu
 35 40 45
 Gly Arg Glu Leu Val Ile Pro Cys Arg Val Thr Ser Pro Asn Ile Thr
 50 55 60
 Val Thr Leu Lys Lys Phe Pro Leu Asp Thr Leu Ile Pro Asp Gly Lys
 65 70 75 80
 Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe Ile Ile Ser Asn Ala Thr
 85 90 95
 Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu Ala Thr Val Asn Gly His
 100 105 110
 Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg Gln Thr Asn Thr Ile Ile
 115 120 125
 Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu
 130 135 140
 Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val Gly Ile
 145 150 155 160
 Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys His Gln His Lys Lys Leu
 165 170 175
 Val Asn Arg Asp Leu Lys Thr Gln Ser Gly Ser Glu Met Lys Lys Phe
 180 185 190
 Leu Ser Thr Leu Thr Ile Asp Gly Val Thr Arg Ser Asp Gln Gly Leu
 195 200 205
 Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr
 210 215 220
 Phe Val Arg Val His Glu Lys Asp Lys Thr His Thr Cys
 225 230 235

<210> 29
 <211> 434
 <212> PRT
 <213> homo sapiens

<400> 29
 Ser Asp Thr Gly Arg Pro Phe Val Glu Met Tyr Ser Glu Ile Pro Glu
 1 5 10 15
 Ile Ile His Met Thr Glu Gly Arg Glu Leu Val Ile Pro Cys Arg Val
 20 25 30
 Thr Ser Pro Asn Ile Thr Val Thr Leu Lys Lys Phe Pro Leu Asp Thr
 35 40 45
 Leu Ile Pro Asp Gly Lys Arg Ile Ile Trp Asp Ser Arg Lys Gly Phe
 50 55 60
 Ile Ile Ser Asn Ala Thr Tyr Lys Glu Ile Gly Leu Leu Thr Cys Glu
 65 70 75 80
 Ala Thr Val Asn Gly His Leu Tyr Lys Thr Asn Tyr Leu Thr His Arg
 85 90 95
 Gln Thr Asn Thr Ile Ile Asp Val Val Leu Ser Pro Ser His Gly Ile
 100 105 110
 Glu Leu Ser Val Gly Glu Lys Leu Val Leu Asn Cys Thr Ala Arg Thr
 115 120 125
 Glu Leu Asn Val Gly Ile Asp Phe Asn Trp Glu Tyr Pro Ser Ser Lys
 130 135 140
 His Gln His Lys Lys Leu Val Asn Arg Asp Leu Lys Thr Gln Ser Gly
 145 150 155 160
 Ser Glu Met Lys Lys Phe Leu Ser Thr Leu Thr Ile Asp Gly Val Thr
 165 170 175
 Arg Ser Asp Gln Gly Leu Tyr Thr Cys Ala Ala Ser Ser Gly Leu Met
 180 185 190
 Thr Lys Lys Asn Ser Thr Phe Val Arg Val His Glu Lys Glu Ser Lys

		195					200				205				
Tyr	Gly	Pro	Pro	Cys	Pro	Pro	Cys	Pro	Ala	Pro	Glu	Phe	Leu	Gly	Gly
	210						215				220				
Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	Ile
225					230					235					240
Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	Gln	Glu
				245					250					255	
Asp	Pro	Glu	Val	Gln	Phe	Asn	Trp	Tyr	Val	Asp	Gly	Val	Glu	Val	His
			260					265					270		
Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln	Phe	Asn	Ser	Thr	Tyr	Arg
		275					280					285			
Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	Gly	Lys
	290					295					300				
Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Gly	Leu	Pro	Ser	Ser	Ile	Glu
305					310					315					320
Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro	Arg	Glu	Pro	Gln	Val	Tyr
				325					330					335	
Thr	Leu	Pro	Pro	Ser	Gln	Glu	Glu	Met	Thr	Lys	Asn	Gln	Val	Ser	Leu
			340					345					350		
Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Val	Glu	Trp
		355					360					365			
Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val
	370					375					380				
Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	Ser	Arg	Leu	Thr	Val	Asp
385					390					395					400
Lys	Ser	Arg	Trp	Gln	Glu	Gly	Asn	Val	Phe	Ser	Cys	Ser	Val	Met	His
			405						410					415	
Glu	Ala	Leu	His	Asn	His	Tyr	Thr	Gln	Lys	Ser	Leu	Ser	Leu	Ser	Leu
			420					425					430		
Gly	Lys														

CLEAR-IT-2: Interim Results of the Phase II, Randomized, Controlled Dose-and Interval-Ranging Study of Repeated Intravitreal VEGF Trap Administration in Patients With Neovascular Age-Related Macular Degeneration

M. S. Benz; Q. D. Nguyen; K. Chu; A. Cahn; I. Grimes; A. Ingerman; J. M. Cedarbaum

+ Author Affiliations & Notes

Investigative Ophthalmology & Visual Science May 2007, Vol.48, 4549. doi:

Abstract

Purpose:: To determine the safety, tolerability, and biological effect of repeated intravitreal (ITV) injection of VEGF Trap in patients with neovascular age-related macular degeneration (AMD).

Methods:: Five groups of patients with neovascular AMD were randomized in a balanced ratio to receive a series of ITV injections of one of 3 dose levels of VEGF Trap (0.5, 2 or 4 mg) into the study eye at 4- or 12-week intervals over a 12-week period. Measures of bioactivity included changes from baseline in best-corrected ETDRS visual acuity (BCVA), foveal thickness, and macular volume determined by optical coherence tomography, as well as total lesion and CNV area determined by fluorescein angiography. Dosing was continued beyond week 12 using a criteria-based schedule.

Results:: The study is currently ongoing and treatment assignments remain masked. No ocular serious adverse events, no identifiable intraocular inflammation or serious drug-related systemic adverse events have been reported to date.

Conclusions: Repeated intravitreal VEGF Trap administration according to the dosing regimens employed in this study appears to be safe and well tolerated. Safety and bioeffect data will be updated at time of presentation.

Clinical Trial: www.clinicaltrials.gov NCT00320788

Keywords: age-related macular degeneration • retina • clinical (human) or epidemiologic studies: outcomes/complications

© 2007, The Association for Research in Vision and Ophthalmology, Inc., all rights reserved. Permission to republish any abstract or part of an abstract in any form must be obtained in writing from the ARVO Office prior to publication.

Results of a Phase I Study of Intravitreal VEGF Trap in Subjects With Diabetic Macular Edema: The CLEAR-IT DME Study

D. V. Do; Q. D. Nguyen; D. J. Browning; J. A. Haller; K. Chu; J. Buskey; I. Grimes; A. Ingerman; J. Cederbaum; P. A. Campochiaro

+ Author Affiliations & Notes

Investigative Ophthalmology & Visual Science May 2007, Vol.48, 1430. doi:

Abstract

Purpose:: To determine the safety, tolerability, and bioactivity of a single dose (4.0mg) of intravitreal VEGF Trap in patients with diabetic macular edema (DME).

Methods:: Five patients with DME, foveal thickness $\geq 250\mu\text{m}$ measured by optical coherence tomography (OCT) and ETDRS best-corrected visual acuity (BCVA) of $\leq 20/40$ and $\geq 20/320$ were administered a single intravitreal injection of 4 mg VEGF Trap at day 0. Safety assessments included eye examinations, vital signs, and laboratory tests. Measures of bioactivity included changes from baseline in BCVA, centerpoint retinal thickness (CRT), and leakage on fluorescein angiography. Subjects were monitored for 6 weeks following VEGF Trap administration.

Results:: Mean patient age was 65.2 years (range=56-75); 4 were Type 2 diabetics. Mean duration of diabetes prior to treatment was 26 years. All had received prior treatment for DME. No severe ocular or serious systemic adverse events related to study drug were noted. Mean baseline BCVA was 69 letters and mean baseline CRT was $407\mu\text{m}$. Four patients had improvements in BCVA, ranging from 6 to 10 letters at 4 weeks post-injection. The mean decrease in centerpoint retinal thickness was $115\mu\text{m}$ at 4 weeks.

Conclusions:: A single intravitreal dose of VEGF Trap was well-tolerated and led to a reduction in CRT and an improvement in BCVA. Although the number of subjects is small, preliminary evidence for bioactivity of VEGF Trap in patients with DME was detected.

Additional studies are being planned to identify the potential therapeutic role of intravitreal VEGF Trap in DME.

Clinical Trial: www.clinicaltrials.gov NCT 00320814

Keywords: diabetic retinopathy • macula/fovea • clinical (human) or epidemiologic studies: treatment/prevention assessment/controlled clinical trials

© 2007, The Association for Research in Vision and Ophthalmology, Inc., all rights reserved. Permission to republish any abstract or part of an abstract in any form must be obtained in writing from the ARVO Office prior to publication.

VEGF Trap-Eye Vision-Specific Quality of Life Through 52 Weeks in Patients With Neovascular AMD in CLEAR-IT 2: A Phase 2 Clinical Trial

D. V. Do; CLEAR-IT 2 Investigators

+ Author Affiliations & Notes

Investigative Ophthalmology & Visual Science April 2009, Vol.50, 1887. doi:

Abstract

Purpose: : To examine the effects of VEGF Trap-Eye on patient-reported vision-specific Quality of Life (QoL) using the National Eye Institute Visual Function Questionnaire (NEI VFQ-25) in CLEAR-IT 2: a Phase 2 randomized trial in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Methods: : CLEAR-IT 2 was a randomized, double masked, multi-center Phase 2 trial. Patients received intravitreal injections of VEGF Trap-Eye 0.5 or 2.0 mg monthly or 0.5, 2.0, or 4.0 mg quarterly for 12 weeks, followed by PRN dosing based on results of OCT and clinical examination out to 52 weeks. The NEI-VFQ 25 was administered to patients at baseline, week 12, and week 52. QoL was assessed at 52 weeks by change in mean score from baseline. The NEI-VFQ 25 subscales are scored from 0-100; a positive difference represents improved functioning or reduced dependency. Pre-specified subscales included near activities, distance activities, and vision related dependency.

Results: : The mean overall change from baseline to 52 weeks in the total score of the NEI-VFQ 25 for all treatment groups combined (n=145) was +4.5. For all groups combined patients had mean changes of +5.7 for near activities, +3.4 for distance activities and +5.8 for vision related dependency. Patients receiving the 2.0 mg dose monthly for the first 12 weeks (2mg q4 group, n=28) had a change from baseline of +4.5 in the total score, +6.8 for near activities, +4.6 for distance activities, and +11.6 for vision related dependency at 52 weeks.

Conclusions: : Patients treated with multiple doses of VEGF Trap-Eye over a 52 week period had overall improvements in the patient-reported vision-specific QoL (total scores as well as subscales) as assessed by the NEI-VFQ 25. As a clinically meaningful change is often considered to be ± 5 points, the most notable improvements were seen in the 2mg q4 group for near activities and for vision related dependency.

Clinical Trial: : www.clinicaltrials.gov NCT00320788

Keywords: age-related macular degeneration • quality of life • clinical (human) or epidemiologic studies: treatment/prevention assessment/controlled clinical trials

© 2009, The Association for Research in Vision and Ophthalmology, Inc., all rights reserved. Permission to republish any abstract or part of an abstract in any form must be obtained in writing from the ARVO Office prior to publication.

VEGF Trap-Eye In CRVO: Primary Endpoint Results Of The Phase 3 COPERNICUS Study

[Julia A. Haller](#); [David S. Boyer](#); [Jeffrey S. Heier](#); [David M. Brown](#); [Lloyd Clark](#); [Robert Vitti](#)

+ Author Affiliations & Notes

Investigative Ophthalmology & Visual Science April 2011, Vol.52, 6643. doi:

Abstract

Purpose: : VEGF Trap-Eye is an intravitreally administered fusion protein that is designed to bind the pro-angiogenic factors VEGF-A and placental growth factor with higher affinity than their natural receptors. This study evaluated the efficacy and safety of VEGF Trap-Eye in patients with macular edema secondary to central retinal vein occlusion (CRVO) after 24 weeks of treatment.

Methods: : In this randomized, double-masked, controlled Phase 3 study, 114 patients received 6 monthly injections of 2 mg VEGF Trap-Eye and 73 patients received control sham injections. Visual acuity was measured as a score based on the total number of Early Treatment of Diabetic Retinopathy Study (ETDRS) letters read correctly. The primary endpoint was the proportion of patients who gained at least 15 ETDRS letters from baseline at 24 weeks. A key secondary endpoint was the mean change in best-corrected visual acuity from baseline at 24 weeks.

Results: : The primary endpoint was met in this study: 56.1% of patients receiving 2 mg VEGF Trap-Eye monthly gained at least 15 letters of vision from baseline, compared with 12.3% of patients receiving sham injections ($p < 0.0001$). Patients receiving VEGF Trap-Eye gained a mean of 17.3 letters of vision compared with a mean loss of 4.0 letters with sham injection ($p < 0.001$). The most common adverse events were those typically associated with intravitreal injections and/or the underlying disease. The proportions of patients who experienced serious ocular adverse events were 3.5% in the VEGF Trap-Eye group and 13.5% in the sham group. The incidence of non-ocular serious adverse events was generally well balanced between the treatment and sham groups.

Conclusions: : Dosing monthly with 2 mg VEGF Trap-Eye in patients with macular edema secondary to CRVO resulted in a statistically significant improvement in visual acuity compared with control sham treatment. VEGF Trap-Eye was generally well tolerated and had a generally favorable safety profile.

Clinical Trial: : <http://www.clinicaltrials.gov> NCT00943072

Keywords: vascular endothelial growth factor • visual acuity • vascular occlusion/vascular occlusive disease

© 2011, The Association for Research in Vision and Ophthalmology, Inc., all rights reserved. Permission to republish any abstract or part of an abstract in any form must be obtained in writing from the ARVO Office prior to publication.

CLEAR-IT 2: Phase 2, Randomized, Controlled Dose-and Interval-Ranging Study of Intravitreal VEGF Trap Eye in Patients With Neovascular Age-Related Macular Degeneration: Predictive Factors for Visual Acuity Outcome at One Year

J. S. Heier; CLEAR-IT 2 Investigators

+ Author Affiliations & Notes

Investigative Ophthalmology & Visual Science April 2009, Vol.50, 1255. doi:

Abstract

Purpose: : To evaluate potential predictive factors for visual acuity outcomes in patients with neovascular AMD after repeated intravitreal injections of VEGF Trap-Eye over one year.

Methods: : CLEAR-IT 2 was a double-masked, multi-center Phase 2 trial in AMD patients randomized to receive VEGF Trap-Eye 0.5 or 2.0 mg monthly or 0.5, 2.0, or 4.0 mg quarterly (at baseline and week 12) with monthly reassessment and PRN dosing to 1 year. Subgroups of patients were identified based on age, baseline best-corrected visual acuity (BCVA), baseline lesion size, and previous treatment for neovascular AMD.

Results: : Data through one year for all dose groups combined (n=157) demonstrated significant improvement in BCVA (mean increase of 5.3 letters, $p < 0.0001$.) Analyses of the subgroups identified above generated the following results for all treatment groups combined: Patients that were ≤ 75 years old (n=53) gained an average of 8.26 (± 13.55) letters as opposed to those > 75 years old (n=104) who gained 3.73 (± 13.31) letters ($p = 0.046$). Those with a BCVA ≤ 54 letters at baseline (n=65) gained an average of 7.54 (± 15.44) letters as opposed to those with a BCVA at baseline > 54 letters (n=92) who gained

3.65 (± 11.8) letters ($p=0.076$). Those who began the study with a lesion size ≤ 4 DA ($n=124$) gained an average of 5.52 (± 13.63) letters, while those who began the study with lesions >4 DA gained 4.27 (± 13.23) letters ($p=0.63$). Patients who were treatment naïve at the start of the study ($n=137$) gained an average of 4.8 (± 13.7) letters, while those who received previous treatment ($n=20$) gained an average of 8.4 (± 12.09) letters ($p=0.27$).

Conclusions: : In this study, ≤ 75 years of age at baseline predicted greater visual acuity gains following multiple treatments of VEGF Trap-Eye. In addition, worse baseline visual acuity was suggestive of better visual acuity outcomes. Baseline lesion size and previous treatment status did not significantly affect treatment responses.

Clinical Trial: : www.clinicaltrials.gov NCT00320788

Keywords: age-related macular degeneration • choroid: neovascularization • vascular endothelial growth factor

© 2009, The Association for Research in Vision and Ophthalmology, Inc., all rights reserved. Permission to republish any abstract or part of an abstract in any form must be obtained in writing from the ARVO Office prior to publication.

The 1-year Results of CLEAR-IT 2, a Phase 2 Study of Vascular Endothelial Growth Factor Trap-Eye Dosed As-needed After 12-week Fixed Dosing

Jeffrey S. Heier, MD,¹ David Boyer, MD,² Quan Dong Nguyen, MD, MSc,³ Dennis Marcus, MD,⁴ Daniel B. Roth, MD,⁵ George Yancopoulos, MD, PhD,⁶ Neil Stahl, PhD,⁶ Avner Ingerman, MD, MSc,⁶ Robert Vitti, MD, MBA,⁶ Alyson J. Berliner, MD, PhD,⁶ Ke Yang, PhD,⁶ David M. Brown, MD,⁷ for the CLEAR-IT 2 Investigators

Objective: To evaluate anatomic outcomes and vision, injection frequency, and safety during the as-needed (PRN) treatment phase of a study evaluating a 12-week fixed dosing period followed by PRN dosing to week 52 with vascular endothelial growth factor (VEGF) Trap-Eye for neovascular (wet) age-related macular degeneration (AMD).

Design: Multicenter, randomized, double-masked trial.

Participants: We included 159 patients with subfoveal choroidal neovascularization (CNV) secondary to wet AMD.

Methods: Patients were randomly assigned to 1 of 5 intravitreal VEGF Trap-Eye treatment groups: 0.5 mg or 2 mg every 4 weeks or 0.5, 2, or 4 mg every 12 weeks during the fixed-dosing period (weeks 1–12). From weeks 16 to 52, patients were evaluated monthly and were retreated PRN with their assigned dose (0.5, 2, or 4 mg).

Main Outcome Measures: Change in central retinal/lesion thickness (CR/LT), change in total lesion and CNV size, mean change in best-corrected visual acuity (BCVA), proportion of patients with 15-letter loss or gain, time to first PRN injection, reinjection frequency, and safety at week 52.

Results: The decrease in CR/LT at week 12 versus baseline remained significant at weeks 12 to 52 (–130 μm from baseline at week 52) and CNV size regressed from baseline by 2.21 mm^2 at 48 weeks. After achieving a significant improvement in BCVA during the 12-week, fixed-dosing phase for all groups combined, PRN dosing for 40 weeks maintained improvements in BCVA to 52 weeks (5.3-letter gain; $P < 0.0001$). The most robust improvements and consistent maintenance of visual acuity generally occurred in patients initially dosed with 2 mg every 4 weeks for 12 weeks, demonstrating a gain of 9 letters at 52 weeks. Overall, a mean of 2 injections was administered after the 12-week fixed-dosing phase, and the mean time to first reinjection was 129 days; 19% of patients received no injections and 45% received 1 or 2 injections. Treatment with VEGF Trap-Eye was generally safe and well tolerated, with few ocular or systemic adverse events.

Conclusions: PRN dosing with VEGF Trap-Eye at weeks 16–52 maintained the significant anatomic and vision improvements established during the 12-week fixed-dosing phase with a low frequency of reinjections. Repeated dosing with VEGF Trap-Eye was well tolerated over 52 weeks of treatment.

Financial Disclosure(s): Proprietary or commercial disclosure may be found after the references. *Ophthalmology* 2011;118:1098–1106 © 2011 by the American Academy of Ophthalmology.



Vascular endothelial growth factor (VEGF) is a critical regulator of normal ocular vasculogenesis and angiogenesis during development.^{1–3} Vascular endothelial growth factor also plays a central role in the abnormal growth of new blood vessels in the retina, as well as in vascular leakage that causes retinal edema and thickening, both of which characterize diseases such as neovascular (wet) age-related macular degeneration (AMD) and diabetic retinopathy that lead to loss of retinal function.^{3–6} Of the various members

of the VEGF gene family, VEGF-A and placental growth factor (PlGF) are the factors implicated in pathologic angiogenesis and the pathogenesis of AMD (*Invest Ophthalmol Vis Sci* 50 [Suppl]: 2943,2009).^{7–9}

An improved understanding of the pivotal role of VEGF in pathologic angiogenesis has resulted in the development and use of intravitreal anti-VEGF therapies in wet AMD and other eye diseases that have an angiogenesis-based etiology.^{10–12} Pegaptanib, an oligonucleotide aptamer,

and ranibizumab, a humanized monoclonal antibody fragment, are anti-VEGF therapies currently available for intravitreal treatment of neovascular AMD. In pivotal trials, pegaptanib mainly slowed loss of visual acuity, whereas ranibizumab improved visual function in a substantial proportion of patients.^{13–16} Bevacizumab is an off-label intravitreal anti-VEGF therapy that has also been shown in less rigorous studies to improve visual function.^{17–20}

The beneficial results with ranibizumab were obtained with a fixed-dosing regimen requiring an injection of ranibizumab 0.5 or 0.3 mg every month for 2 years in the pivotal phase 3 studies, Minimally Classic/Occult Trial of the Anti-VEGF Antibody Ranibizumab in the Treatment of Neovascular Age-Related Macular Degeneration (MARINA) and Anti-VEGF Antibody for the Treatment of Predominantly Classic Choroidal Neovascularization in Age-Related Macular Degeneration (ANCHOR).^{13,15} Several studies were undertaken to examine different dosing regimens for ranibizumab. In a large, randomized study in which patients were initially treated with a 3-month loading regimen of ranibizumab and then dosed at regular quarterly intervals, the initial gains after the loading regimen were not maintained at a year.²¹ A 40-patient uncontrolled, open-label, single-site, Prospective Optical Coherence Tomography Imaging of Patients Treated with intra-Ocular ranibizumab (PrONTO) trial evaluated an as-needed (PRN) dosing regimen (based on monthly evaluation of changes in retinal thickness and edema using optical coherence tomography [OCT]) after 3 consecutive monthly injections. Visual acuity outcomes at 12 and 24 months were comparable with those observed in the pivotal phase 3 studies and were attained with fewer intravitreal injections.^{22,23} However, in a larger randomized study, the gains in visual acuity after an initial 3-month loading regimen of ranibizumab were not maintained with subsequent protocol-defined retreatment.²⁴

Vascular endothelial growth factor Trap-Eye (VEGF Trap-Eye) is a potent, specific VEGF antagonist that binds and inactivates circulating VEGF and VEGF in the extravascular space. It was developed specifically as an ultrapurified, isoosmotic solution for ophthalmic use.²⁵ Consisting of extracellular portions of VEGF receptors 1 and 2 fused to the Fc portion of human immunoglobulin G, VEGF Trap-Eye binds both VEGF-A or PlGF and forms an inert 1:1 complex with the growth factor dimers.^{24,25} Thus, VEGF Trap-Eye has broader anti-VEGF activity compared with pegaptanib, which binds only the VEGF-A₁₆₅ isoform,²⁶ and ranibizumab, which neutralizes all active isoforms of VEGF-A, but not PlGF.²⁷ Because VEGF Trap-Eye contains only human sequences, its potential for immunogenicity is low. A key differentiating feature of VEGF Trap-Eye is its picomolar affinity for VEGF ligands, which is substantially higher than that of the natural receptors or anti-VEGF monoclonal antibodies.^{25,28,29} The clinical relevance of the higher binding affinity of VEGF Trap-Eye remains unknown, but it is thought that it might lead to more persistent VEGF blockade and a theoretically longer dosing interval between injections to maintain visual acuity relative to currently available anti-VEGF treatments.³⁰

The clinical efficacy of VEGF-Trap Eye was initially demonstrated in the CLinical Evaluation of Anti-angiogenesis in

the Retina Intravitreal Trial (CLEAR-IT 1), a 6-week phase 1 sequential cohort, single-ascending-dose (0.05 to 4 mg) study of intravitreal VEGF Trap-Eye in patients with neovascular AMD.³¹ The efficacy and safety of repeated dosing with VEGF Trap-Eye were subsequently examined in the phase 2 CLEAR-IT 2 study, which consisted of an initial 12-week fixed dosing period with 1 of 5 monthly or quarterly regimens of VEGF Trap-Eye, followed by PRN dosing from weeks 16 to 52. Detailed results to 16 weeks for the fixed-dosing phase, including the primary endpoint at week 12, are presented in the accompanying article (Brown et al., in this issue, pp 1089-97). At 12 weeks, treatment with VEGF Trap-Eye resulted in a significant reduction in central retinal/lesion thickness (CR/LT) of $-119 \mu\text{m}$ and a significant improvement in mean BCVA of 5.7 letters for all groups combined, and gains of >8 letters in the monthly dosing groups. The finding that improvements in visual acuity and retinal thickness were greater in the monthly dosing groups than in the quarterly dosing groups at week 12, support the need for an initial intensive monthly loading dose phase. Patients were treated with a PRN dosing regimen through week 52 to explore whether the high affinity of VEGF Trap-Eye for VEGF-A and PlGF could translate into the maintenance of initial visual acuity gains through 1 year with less frequent intravitreal injections. Results of the continued dosing phase of the CLEAR-IT 2 study are reported herein.

Materials and Methods

Study Design

The primary objectives of the study were to assess the effect of intravitreal VEGF Trap-Eye on CR/LT and to assess the ocular and systemic safety and tolerability of repeated doses of VEGF Trap-Eye in patients with choroidal neovascularization (CNV) associated with wet AMD. A key secondary objective was to assess the effect of VEGF Trap-Eye on BCVA.

This study was a double-masked, prospective, randomized, dose- and interval-ranging study in which 5 groups of approximately 30 patients each were assigned to a fixed-dose of intravitreal VEGF Trap-Eye in the study eye during the first 12 weeks of dosing, followed by PRN dosing from weeks 16 to 52 (Fig 1, available online at <http://aajournals.org>). The VEGF Trap-Eye regimens were 0.5 mg or 2 mg at 4-week intervals (0.5q4 or 2q4 on day 1 and at weeks 4, 8, and 12 for a total of 4 treatments) or 0.5, 2, or 4 mg every 12 weeks (0.5q12, 2q12, or 4q12 on day 1 and week 12 for a total of 2 treatments). During the PRN dosing phase beginning at week 16, patients received the same dose of VEGF Trap-Eye (0.5, 2, or 4 mg) as received during the fixed-dosing phase (Fig 2).

The study protocol was approved by the ethics committee at every institution and was conducted according to the recommendations of Good Clinical Practice and the Declaration of Helsinki. The study was compliant with the rules and regulations under the Health Insurance Portability and Accountability Act of 1996. All patients provided written informed consent to participate in the study. The CLEAR-IT 2 study is registered with ClinicalTrials.gov (NCT00320788).

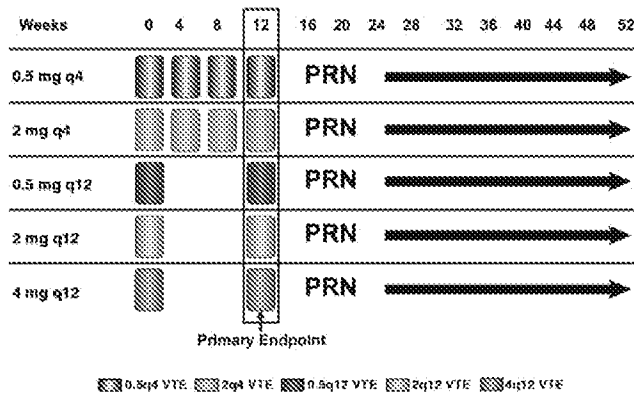


Figure 2. Study schedule. During the 12-week fixed dosing phase, patients in the monthly dosing groups received 0.5 or 2 mg of VEGF Trap-Eye every 4 weeks on day 0 and at weeks 4, 8, and 12 for a total of 4 doses; those in the quarterly dosing groups received 0.5, 2, or 4 mg of VEGF Trap-Eye every 12 weeks on day 0 and at week 12 for a total of 2 doses. Beginning at week 16 and continuing to week 52, patients were assessed every 4 weeks and, if needed, were retreated with the same dose of VEGF Trap-Eye as in the fixed dosing phase. The primary study endpoint was assessed at week 12. q = every; PRN = as-needed; VEGF = vascular endothelial growth factor; VTE = VEGF Trap-Eye; 0.5q4 = 0.5 mg every 4 weeks; 2q4 = 2 mg every 4 weeks; 0.5q12 = 0.5 mg every 12 weeks; 2q12 = 2 mg every 12 weeks; 4q12 = 4 mg every 12 weeks.

Patient Population

The study enrolled patients >50 years old who had a diagnosis of subfoveal CNV secondary to wet AMD and central retinal thickness $\geq 300 \mu\text{m}$, Early Treatment of Diabetic Retinopathy Study (ETDRS) BCVA of 73 to 34 letters, loss of ≥ 5 ETDRS letters in BCVA over the preceding 6 months for previously treated patients with minimally classic or occult lesions, linear diameter of lesion $\leq 5400 \mu\text{m}$ by fluorescein angiography (FA), subretinal hemorrhage sparing the fovea and comprising $\leq 50\%$ of total lesion, area of scar $\leq 25\%$ of total lesion, and sufficient clarity of ocular media to allow retinal photography.

Key exclusion criteria were history of vitreous hemorrhage in preceding 4 weeks; aphakia or pseudophakia with absence of a posterior capsule (unless as a result of a yttrium aluminum garnet capsulotomy); significant subfoveal atrophy or scarring; presence of other causes of CNV in either eye; previous treatments for AMD in the study eye within 12 weeks for photodynamic therapy, 8 weeks for pegaptanib sodium, or 24 weeks for intravitreal or juxtasclear steroids; no other treatments for AMD (thermal laser, surgery, or intraocular/systemic anti-VEGF therapy) were allowed; any retinal vascular disease other than CNV in either eye; active ocular inflammation or infection; history of trabeculectomy or pars plana vitrectomy; history of myocardial infarction, stroke, transient ischemic attack, symptomatic peripheral vascular disease, or treatment for congestive heart failure within the last 6 months; and other conditions or laboratory abnormalities that might interfere with patient participation in the study.

Retreatment Criteria

During the fixed-dosing phase, 1 eye was designated as the study eye and all evaluations were conducted on that eye as described previously (Brown et al, in this issue, pp 1089-97). Beginning at week 16, patients were evaluated every 4 weeks to determine the need for continued dosing. After week 16, the study eye was reinjected with VEGF Trap-Eye if any of the following changes

were observed by the evaluating practitioner: An increase in CR/LT $\geq 100 \mu\text{m}$ as measured by OCT; a loss of ≥ 5 ETDRS letters in conjunction with recurrent fluid as indicated by OCT; persistent fluid as indicated by OCT; new-onset classic neovascularization; new or persistent leak on FA; or new macular hemorrhage.

Endpoints and Assessments

Assessments were performed at scheduled clinic visits on days 1 and 7, at week 4, and every 4 weeks thereafter to week 52. At each visit, patients underwent a full ophthalmologic examination, including visual acuity testing, indirect ophthalmoscopy and slit lamp examination, intraocular pressure (IOP) measurement, and OCT. Fundus photography and FA were performed at baseline and at weeks 4, 12, 24, 36, and 48.

The primary efficacy endpoint was reduction of CR/LT from baseline to 12 weeks, at the end of the fixed-dosing phase. The variables assessed during the PRN dosing phase included change from baseline in CR/LT and mean change from baseline in CNV size determined by FA at 48 weeks, the last mandatory FA in the study; BCVA at 52 weeks; proportions of patients with avoidance of moderate vision loss (loss of < 15 letters); stabilization or improvement in visual acuity (gain of ≥ 0 letters); and significant vision gain (gain of ≥ 15 letters) at 52 weeks; time to first reinjection after week 12; and mean number of injections over the PRN period.

The CR/LT was determined from Stratus OCT (Version 4.0 or higher; CarlZeiss, Jena, Germany) scans read at a masked independent central reading center (Digital OCT Reading Center, Cleveland, OH). The CR/LT was defined as the distance between the inner limiting membrane and posterior border of retinal pigment epithelial/choriocapillaris complex including any subretinal fluid and thickness of any observable choroidal neovascular membrane or scar tissue in central 1 mm of posterior pole scan.

Changes in the size of the total lesion and the CNV component were evaluated with FA. The CNV size was defined as the area of visible CNV (classic or occult) with angiographic evidence of late leakage or pooling of dye. Angiographic images were transmitted to the masked reading center for review (Digital Angiography Reading Center, New York, NY). At least 1 designated photographer was certified by the Reading Center before enrollment of the first patient at each site.

Certified examiners assessed BCVA by using the ETDRS protocol at 4 m. Examiners were masked to treatment assignment, and performed no other study assessments.

Safety assessments included IOP (measured preinjection and approximately 30 minutes postinjection), ophthalmologic examinations for ocular toxicity, adverse events (AEs), serious AEs (SAEs), clinical laboratory tests, and vital signs.

Statistical Analysis

Efficacy assessments were made on the full analysis set, which included all patients who received study treatment and had a baseline and ≥ 1 postbaseline assessment. Safety assessments were performed on all patients who received study treatment.

The primary analysis was a paired comparison *t* test of the change in CR/LT from baseline to week 12 for all groups combined. If this was significant, an analysis of covariance was done on the 5 individual groups. A similar analysis was done for all continuous measures at all time points. Missing values were imputed using the last-observation-carried-forward method for continuous measures. The durability of the effect was assessed by evaluating all of the endpoints out to week 52. All of the analyses shown below were done using the same methods at week 12 and week 52 (week 48 for FA parameters).

Table 2. Baseline Demographic and Clinical Characteristics

Characteristic	All Treated Patients (n = 157)
Age, y (mean [range])	78.3 (53–94)
Gender (%M:%F)	38:62
Disease duration, months (mean [range])	3.9 (0–67)
Previous treatment	20 (12.7%)
Lesion size (mean ± SD) in disc area	3.11 ± 2.12
Lesion type (n [%])	
Predominantly classic	60 (38.2)
Minimally classic	37 (23.6)
Occult lesions	60 (38.2)
Disease status (mean [range])	
Central retinal/lesion thickness (μm)	456 (186–1316)
Foveal thickness (μm)	327 (116–1081)
Best-corrected visual acuity (ETDRS letters)	56 (27–83)

ETDRS = Early Treatment of Diabetic Retinopathy Study; F = Female; M = Male; SD = standard deviation.

Results

Patient Disposition

Of 159 patients who were randomized, 157 were treated and 134 (85.4%) completed 52 weeks in the study (Table 1 available online at <http://aaojournal.org>). For the 23 patients (14.6%) who were withdrawn before completion of 52 weeks, 6 (3.8%) withdrawals were at the request of the patient.

Baseline Characteristics

The study population was representative of the AMD population in the United States. Patients ranged in age from 53 to 94 years (mean, 78.3) and the majority were women (62%; Table 2). The mean time from diagnosis was 3.9 months (range, 0–67), and 12.7% of patients had received previous treatment. The distribution of CNV lesions was 38.2% predominantly classic, 23.6% minimally classic, and 38.2% occult with no classic. The treatment groups were well-balanced overall for baseline disease severity, but mean CR/LT and mean foveal thickness were somewhat higher in the 4 mg q12wk group (Table 3).

Change in Central Retinal/Lesion Thickness

The primary efficacy endpoint of the study was mean change in CR/LT at week 12. In all treatment groups combined, the significant decrease from baseline in CR/LT observed at week 12 (–119 μm) was maintained to week 52 (–130 μm; $P < 0.0001$) after 40 weeks of PRN dosing with VEGF Trap-Eye (Fig 3A). The de-

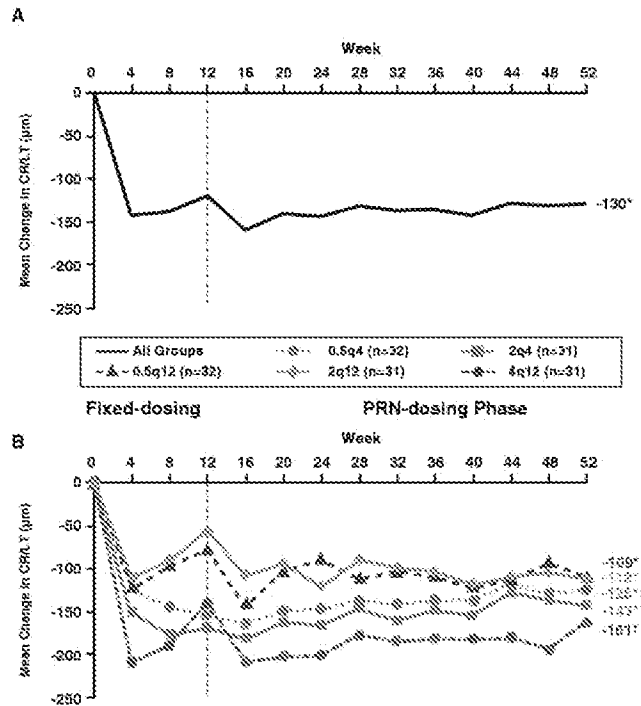


Figure 3. Mean change in central retinal/lesion (CR/LT) thickness in (A) all treatment groups combined and (B) individual treatment groups. The CR/LT was measured with optical coherence tomography. Change in CR/LT from baseline at 12 weeks was the primary study endpoint. In the combined treatment group, a significant ($*P < 0.0001$) decrease in CR/LT at week 12 was maintained to week 52 (–130 μm). The decrease in CR/LT in individual treatment groups was maintained between weeks 12 and 52 with PRN dosing was significant ($*P < 0.0001$; $†P = 0.0002$) compared with baseline values. The last-observation-carried-forward method was used to impute missing data. CR/LT = central retina/lesion thickness; PRN = as-needed; 0.5q4 = 0.5 mg every 4 weeks; 2q4 = 2 mg every 4 weeks; 0.5q12 = 0.5 mg every 12 weeks; 2q12 = 2 mg every 12 weeks; 4q12 = 4 mg every 12 weeks.

crease in CR/LT was also maintained in all individual dosing groups after PRN dosing and was significant compared with baseline values. The greatest decreases in CR/LT at week 52 versus baseline occurred in the 4 mg q12wk group (–161 μm; $P = 0.0002$) and the 2 mg q4wk group (–143 μm; $P < 0.0001$; Fig 3B).

Change in Angiographic Measures

Fluorescein angiography (FA) was performed at baseline and at weeks 4, 12, 24, 36, and 48. In all groups combined and in each treatment group, there were no significant changes in total lesion

Table 3. Baseline Disease Status by Treatment Group

Mean (Range)	0.5q4 (n = 32)	2q4 (n = 31)	0.5q12 (n = 32)	2q12 (n = 31)	4q12 (n = 31)	All Groups (n = 157)
CR/LT (μm)	434 (282–710)	453 (232–960)	442 (186–762)	447 (265–948)	507 (240–1316)	456 (186–1316)
Foveal thickness (μm)	329 (212–509)	307 (171–524)	319 (116–559)	334 (186–762)	360 (177–1081)	327* (116–1081)
BCVA (ETDRS letters)	54 (27–76)	58 (32–83)	56 (30–72)	57 (32–72)	53 (28–80)	56 (27–83)

BCVA = best-corrected visual acuity; CR/LT = central retina/lesion thickness; ETDRS = Early Treatment of Diabetic Retinopathy Study; q = every. *In all groups (n = 157), 25 patients at week 52 showed foveal thickness measurements of <150 μm.

size from baseline to week 12 and week 48 (data not shown). The decrease in total lesion size for the 2 mg q4wk group at week 12 (-0.75 mm^2 ; $-0.30 \text{ disc area [DA]}$) and week 48 (-1.75 mm^2 ; -0.69 DA ; $P < 0.04$) achieved significance.

The area of CNV (defined as classic and/or occult CNV demonstrating angiographic evidence of late leakage or pooling of dye) was also measured. In all treatment groups combined, PRN treatment with VEGF Trap-Eye resulted in a consistent decrease in the CNV size versus baseline at weeks 12 and 48, with a decrease of -2.21 mm^2 (-0.87 DA) at week 48 ($P < 0.001$; Fig 4A, available online at <http://aaojournal.org>). All treatment groups other than the 0.5 mg q12wk group experienced a decrease in active CNV size at 48 weeks (-1.42 to -3.41 mm^2 ; -0.56 to -1.34 DA), with the greatest reduction in the 2 mg q4wk group (-3.41 mm^2 , -1.34 DA ; $P < 0.001$; Fig 4B).

Change in Visual Acuity

The significant improvement from baseline in BCVA that was noted at 12 weeks was maintained through the PRN dosing phase to week 52. The combining of all treatment groups showed a mean gain of 5.7 letters at week 12 and 5.3 letters at week 52 ($P < 0.0001$ vs baseline; Fig 5A). The greatest improvement in BCVA occurred

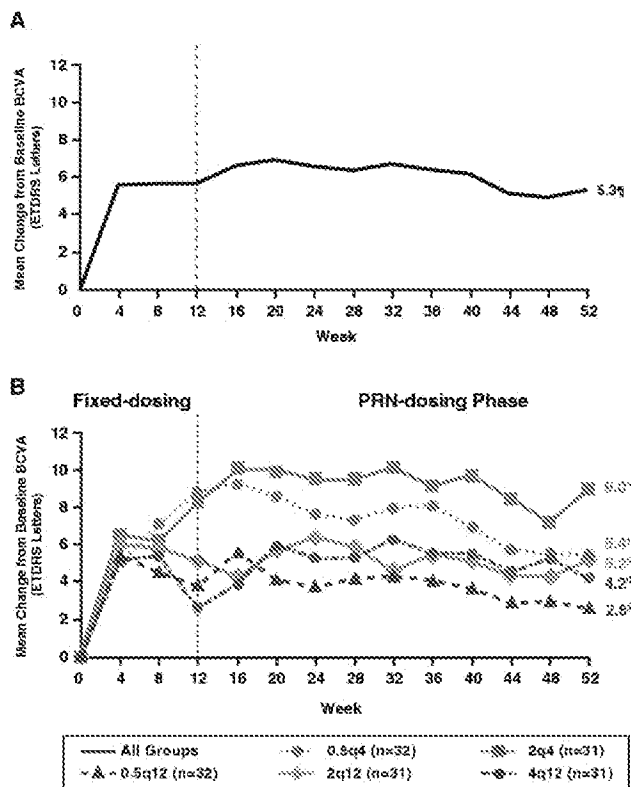


Figure 5. Mean change in best-corrected visual acuity (BCVA) in (A) all treatment groups combined and (B) individual treatment groups. The BCVA was assessed with the Early Treatment of Diabetic Retinopathy Study protocol at 4 m. Significant improvements from baseline in BCVA were noted in all treatment groups combined at week 12 (5.7 letters) and were maintained to week 52 (5.3 letters; $^{\dagger}P < 0.0001$). The 2 mg q4wk group showed the greatest gain in BCVA at 12 weeks, which was maintained to 52 weeks (9.0 letters; $^*P < 0.0001$; $^{\ddagger}P = 0.085$; $^{\S}P = 0.0412$; $^{\P}P = 0.0154$; and $^{\ddagger\ddagger}P = 0.344$ for individual groups versus baseline). The last-observation-carried-forward method was used to impute missing data. ETDRS = Early Treatment of Diabetic Retinopathy Study; PRN = as-needed.

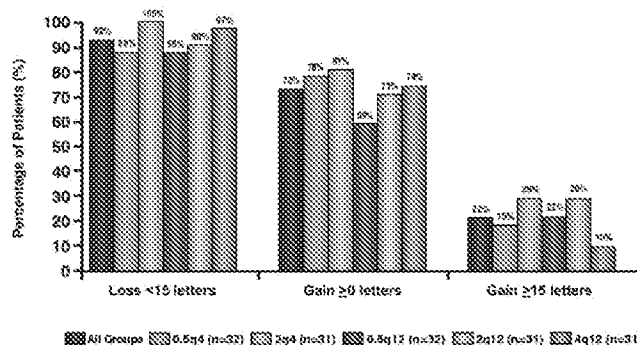


Figure 6. Visual acuity changes at week 52. The proportions of patients who avoided moderate vision loss (≥ 15 letters), had an improvement in visual acuity (gain of ≥ 0 letters), or had a significant vision gain (≥ 15 letters) in the treatment groups combined and individual dosing groups at week 52 are shown. In the treatment groups combined, only 8% of patients experienced moderate loss of vision, whereas 22% showed a significant gain in vision of ≥ 15 letters at week 52 after 40 weeks of as-needed (PRN) dosing. The last-observation-carried-forward method was used to impute missing data. 0.5q4 = 0.5 mg every 4 weeks; 2q4 = 2 mg every 4 weeks; 0.5q12 = 0.5 mg every 12 weeks; 2q12 = 2 mg every 12 weeks; 4q12 = 4 mg every 12 weeks.

in the 2 mg q4wk treatment group, with a mean increase of 8.3 letters at week 12 and of 9.0 letters at week 52 ($P < 0.0001$ vs baseline; Fig 5B). Visual acuity improvements compared with baseline were also maintained in all other treatment groups at week 52.

Frequency of Patients with Visual Acuity Changes

In all treatment groups combined, moderate loss of vision (loss of ≥ 15 letters) was avoided in 98% of patients at week 12 and 92% at week 52 after treatment with VEGF Trap-Eye. In the individual treatment groups, 88% to 100% of patients avoided moderate loss of vision at week 52 (Fig 6). Overall, 12 patients experienced moderate vision loss at week 52, including 4 in the 0.5 mg q4wk group, 4 in the 0.5 mg q12wk group, 3 in the 2 mg q12wk group, and 1 in the 4 mg q12wk group. None of the patients in the 2 mg q4wk group experienced moderate vision loss.

Stabilization or improvement in visual acuity (gain of ≥ 0 letters) occurred in 74.5% of patients in all treatment groups combined at week 12 and in 73% of patients at week 52. In the individual treatment groups, the proportion of patients experiencing stable or improved visual acuity ranged between 59% and 81%, with the highest proportion in the 2 mg q4wk group. Overall, these proportions remained steady from week 12 to week 52.

The frequency of patients in all treatment groups combined with a significant gain in vision (≥ 15 letters) was 18.5% at week 12 and 22% at week 52. Groups treated with 2 mg, either monthly or quarterly, had the highest frequency of patients with significant visual gain (26% and 16%, respectively, at week 12, and 29% in each group at week 52).

The proportion of patients with $\le 20/200$ vision in all treatment groups combined was 11.5% at baseline and remained stable at 10.8% at week 52. The proportion of patients with $\ge 20/40$ vision increased from baseline (15%) to week 52 (41%) for all groups combined, with the 0.5mg q4wk and 2mg q4wk groups increasing from 13% and 16% to 47% and 45%, respectively (Fig 7, available online at <http://aaojournal.org>).

Table 4. Retreatment Outcomes with VEGF Trap-Eye

Treatment Regimen	Mean No. of Injections over PRN Phase (Weeks 12–52)	Mean No. of Days to First Injection over PRN Phase (Weeks 12–52)	Median No. of Days to First Injection over PRN Phase (Months 3–12)
0.5 mg q4	2.52	102	85
2 mg q4	1.55	160	150
0.5 mg q12	1.84	133	86
2 mg q12	2.48	113	86
4 mg q12	1.7	138	111
All groups	2.01	129	110

PRN = as-needed.

Reinjection Outcomes

Beginning at week 16, patients were evaluated monthly for the need for reinjection. Over the 40-week PRN dosing phase, the mean number of reinjections received for all groups combined was 2.01, ranging from 1.55 injections in the 2 mg q4wk group to 2.52 in the 0.5 mg q4wk group (Table 4). During the PRN dosing phase, 29 patients (19%) did not receive any injections of VEGF Trap-Eye, 45% received 1 to 2 injections, and only 5% of patients received ≥ 5 injections (Fig 8). The most common reason for retreatment was the presence of persistent fluid on OCT examination (63%); in 26% of patients who needed retreatment, a new or persistent leak was noted on FA, and 25% of patients had a loss of BCVA of ≥ 5 ETDRS letters with recurrent fluid on OCT (Table 5 available online at <http://aaojournal.org>).

The average time from the last mandatory injection at week 12 to the first PRN injection was 129 days for all treatment groups combined. The longest initial treatment-free interval was in the 2 mg q4wk group (160 days; Table 4). This calculation accounts for the 29 patients who were not reinjected by assigning them a reinjection time at 52 weeks and is therefore an underestimate of the true mean. Kaplan–Meier analysis of the time to first reinjection showed a median time to reinjection for all groups combined of 110 days. The longest median time to reinjection was 150 days in the 2 mg q4wk group (Fig 9, available online at <http://aaojournal.org>; Table 4).

Safety

The types and frequencies of ocular AEs occurring in the study eye were consistent with AEs previously reported with intravitreal

anti-VEGF treatment and were generally related to the intravitreal injection procedure (Table 6). In all treatment groups combined, conjunctival hemorrhage (38.2%) was the most frequently reported AE in the study eye. Most ocular AEs in the study eye were mild (58%), with 4 events considered severe (conjunctival hemorrhage, reduced visual acuity, uveitis, and increased IOP). Seven patients (4.5%) had an increase in IOP in the study eye that was considered related to study treatment, but the increased IOP was not considered an SAE and did not lead to withdrawal of patients from the study. Most ocular events that occurred in the fellow eye were mild (41.4%) or moderate (12.7%); the most frequently reported AE in the fellow eye was vitreous detachment (8.9%).

The number of patients with systemic AEs was similar among treatment groups. The most commonly reported systemic AEs were urinary tract infection (10.2%), bronchitis (9.6%) and upper respiratory tract infection (9.6%). A total of 58 systemic SAEs were reported in 35 patients, but none of the events was deemed to be related to study treatment. Two deaths occurred during the study, one from pancreatic carcinoma and the other from preexisting pulmonary hypertension. Four ocular SAEs were reported: Culture-negative endophthalmitis/uveitis (0.5 mg q4wk group) and decreased visual acuity (0.5 mg q12wk group) in the study eye and increased IOP (4 mg q12wk group) and retinal detachment (0.5 mg q12wk group) in the fellow eye. Stroke was reported in 1 patient in the 0.5 mg q4wk group who had a history of stroke. No cases of vascular death were reported in this study.

Adverse events resulted in withdrawal of 7 patients from the

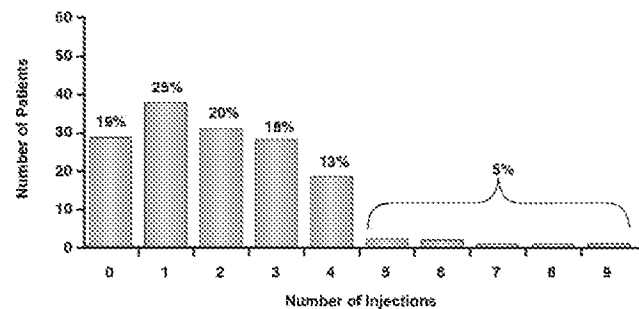


Figure 8. Number of injections after loading phase. Numbers of patients receiving 0 to 9 injections of VEGF Trap-Eye during the as-needed dosing period are shown. Overall, 19% of patients required no retreatment, 45% received 1 or 2 retreatments, and only 5% required 5 to 9 retreatments. VEGF = vascular endothelial growth factor.

Table 6. Ocular Adverse Events in the Study Eye (Frequency $\geq 5\%$ in All Groups Combined*)

Adverse Event	Number (n)	Percent (%)
Conjunctival hemorrhage	60	38.2
Increased IOP (transient postinjection)	29	18.5
Refraction disorder	26	16.6
Retinal hemorrhage	22	14.0
Visual acuity reduced (patient reported)	21	13.4
Vitreous detachment	18	11.5
Eye pain	15	9.6
Vitreous floaters	14	8.9
Detachment of retinal pigment epithelium	12	7.6
Retinal edema	10	6.4
Visual disturbance	8	5.1
Blepharitis	8	5.1
Subretinal fibrosis	8	5.1

IOP = intraocular pressure.

*Patients receiving treatment with VEGF Trap-Eye (n = 157).

study: Three ocular (retinal edema and retinal hemorrhage in the study eye and increased IOP in the fellow eye) and 4 systemic (non-Hodgkin's lymphoma, hip fracture, colon cancer, and bronchitis). There were no differences in frequency of AEs or SAEs between the treatment groups.

Discussion

Among patients with neovascular AMD, PRN dosing with VEGF Trap-Eye maintained efficacy established during a 12-week monthly or quarterly fixed-dosing phase for an additional 40 weeks. For all groups combined, a clinically significant improvement in visual acuity achieved at 12 weeks (5.7-letter gain) was maintained to 52 weeks (5.3-letter gain), accompanied by a decrease in CR/LT ($-119 \mu\text{m}$ at week 12 and $-130 \mu\text{m}$ at week 52). Among patients in all treatment groups combined, 22% experienced a gain of ≥ 15 letters and only 8% of patients had a loss of ≥ 15 letters at the end of the study period. These improvements were achieved with an average of only 2 additional injections of VEGF Trap-Eye over the 40-week PRN phase; notably, 44% of patients required either no retreatment or only 1 reinjection, suggesting a long duration of effect in these patients. Repeated intravitreal dosing of VEGF Trap-Eye was generally safe and well tolerated. The overall safety profile was similar to that previously reported with other intravitreal anti-VEGF agents.

Anti-VEGF treatment offers the hope of improved vision and is now the standard of care for neovascular AMD; however, the need for frequent monitoring and treatment places a significant burden on both the patient and the clinician. In addition, the injection procedure itself has associated risks, which, although rare, may be increased with repeated injections. Hence, a less frequent dosing regimen that is maximally effective in preserving or improving visual acuity is a desirable goal of anti-VEGF treatment for AMD.

With VEGF Trap-Eye, the improvements in visual acuity achieved with a 12-week fixed dosing schedule were maintained to week 52 with an average of 2 injections over the 9-month period after fixed dosing. In the 2 mg q4wk group, $\geq 50\%$ of patients remained injection free for 150 days after week 12. The best visual acuity outcomes to date with ranibizumab have been achieved with 3 consecutive monthly injections, followed by continuous monthly injections.^{13,15,32} Less frequent dosing with ranibizumab using both fixed and PRN schedules has been investigated. In the 2-year Phase 3b, Multicenter, Randomized, Double-masked, Sham Injection-controlled Study of the Safety and Efficacy of Ranibizumab (PIER), study a regimen of 3 consecutive monthly injections followed by fixed quarterly injections provided vision gains at 3 months that were comparable with those in the pivotal clinical studies; however, these gains declined with quarterly dosing and returned to baseline levels at 12 months.²¹ Results of a phase 3b study designed to evaluate the long-term effect of ranibizumab in patients with all subtypes of neovascular AMD (The Safety Assessment of Intravitreal Lucentis for AMD [SAILOR]) suggested that treating patients with ranibizumab on a PRN basis was less effective than monthly

dosing, although retreatment criteria and follow-up schedules in this study were less well-defined and investigator determined.³³ In the open-label HORIZON extension study, patients received PRN dosing with ranibizumab after 2 years of monthly dosing, resulting in a decline in visual acuity gain at years 3 and 4 (Invest Ophthalmol Vis Sci 50 [Suppl]:3093, 2009).

The 52-week results of the CLEAR-IT 2 study suggest that the efficacy of VEGF Trap-Eye, established with an initial fixed-dosing regimen, is maintained with PRN dosing and that VEGF Trap-Eye has an extended duration of action. The average time from last mandatory injection to first PRN injection for all groups combined was >4 months and for the 2 mg q4wk group, >5 months. The high binding affinity of VEGF Trap-Eye, its presumed long intravitreal half-life, and activity against multiple VEGF family members, including PIGF, may contribute to more sustained and comprehensive VEGF suppression than may be achieved with currently used anti-VEGF agents. Based on its binding affinity and estimated intravitreal half-life in humans, a mathematical model has predicted that, after intravitreal injection, VEGF Trap-Eye would maintain biological activity for 10 to 12 weeks, whereas ranibizumab would maintain such activity for 30 days, supporting the concept of less frequent dosing with VEGF Trap-Eye.³⁰

By the end of the fixed-dosing phase at week 12, VEGF Trap-Eye showed a significant improvement in visual acuity from baseline, and this efficacy was maintained to week 52. However, patients randomized to monthly injections during the fixed dosing phase had a trend toward improved visual acuity outcomes at 52 weeks compared with those randomized to quarterly injections. This difference was apparent at 12 weeks and was maintained throughout the 40-week PRN phase, despite the fact that all patients were treated as often as necessary during this phase. Therefore, initial sequential monthly loading doses seem to provide better control of neovascular leakage and lead to superior gains in visual acuity that can be subsequently maintained with less frequent dosing.

Ongoing phase 3 studies (VEGF Trap-Eye: Investigation of Efficacy and Safety in wet Age-Related Macular Degeneration, VIEW-1 and VIEW-2) are evaluating VEGF Trap-Eye doses of 0.5 and 2 mg every 4 weeks and 2 mg every 8 weeks after 3 initial monthly doses, compared with ranibizumab 0.5 mg every 4 weeks. The rationale for the 2 mg q8wk dose was based on an observation from the fixed-dosing phase of CLEAR-IT 2: At 8 weeks, the improvement in BCVA after a single 2-mg dose was similar to that obtained with 2 mg dosed monthly to 8 weeks and that the effect in visual acuity gain for the single dose groups began to wane by week 12.

After the first year of treatment on the above schedule, patients in the VIEW-1 and -2 studies will be evaluated monthly and treated on a PRN basis. To prevent undertreatment, the PRN dosing schedule during the second year of the study does not allow any patient to remain untreated for >12 weeks. Although many patients in the CLEAR-IT 2 study did maintain initial visual acuity gains for >12 weeks, the design of the phase 3 program is meant to optimize visual improvements for all patients by using this capped

PRN dosing schedule. These studies should provide further information on the duration and extent of the clinical benefit of VEGF Trap-Eye.

In conclusion, PRN dosing of VEGF Trap-Eye after 12 weeks of monthly or quarterly fixed dosing maintained clinically and statistically significant improvements in vision and retinal thickness to week 52 in patients with neovascular AMD, with a low frequency of reinjection. VEGF Trap-Eye was generally well-tolerated, with a safety profile similar to that reported with other intravitreally administered anti-VEGF agents.

Acknowledgment. Technical writing and editorial assistance was provided by Meher Dustoor, PhD.

References

- Maharaj AS, D'Amore PA. Roles for VEGF in the adult. *Microvasc Res* 2007;74:100–13.
- Tammela T, Enholm B, Alitalo K, Paavonen K. The biology of vascular endothelial growth factors. *Cardiovasc Res* 2005; 65:550–63.
- Penn J, Madan A, Caldwell R, et al. Vascular endothelial growth factor in eye disease. *Prog Retin Eye Res* 2008;27: 331–71.
- Grisanti S, Tatar O. The role of vascular endothelial growth factor and other endogenous interplayers in age-related macular degeneration. *Prog Retin Eye Res* 2008;27:372–90.
- Krzystolik MG, Afshari MA, Adamis AP, et al. Prevention of experimental choroidal neovascularization with intravitreal anti-vascular endothelial growth factor antibody fragment. *Arch Ophthalmol* 2002;120:338–46.
- Aiello L, Avery R, Arrigg P, et al. Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. *N Engl J Med* 1994;331:1480–7.
- Rakic J, Lambert V, Devy L, et al. Placental growth factor, a member of the VEGF family, contributes to the development of choroidal neovascularization. *Invest Ophthalmol Vis Sci* 2003;44:3186–93.
- Olsson A, Dimberg A, Kreuger J, Claesson-Welsh L. VEGF receptor signalling—in control of vascular function. *Nat Rev Mol Cell Biol* 2006;7:359–71.
- Boyer DS, Antoszyk AN, Awh CC, et al, MARINA Study Group. Subgroup analysis of the MARINA study of ranibizumab in neovascular age-related macular degeneration. *Ophthalmology* 2007;114:246–52.
- Waisbourd M, Loewenstein A, Goldstein M, Leibovitch I. Targeting vascular endothelial growth factor: a promising strategy for treating age-related macular degeneration. *Drugs Aging* 2007;24:643–62.
- van Wijngaarden P, Qureshi S. Inhibitors of vascular endothelial growth factor (VEGF) in the management of neovascular age-related macular degeneration: a review of current practice. *Clin Exp Optom* 2008;91:427–37.
- Ip M, Scott I, Brown G, et al. Anti-vascular endothelial growth factor pharmacotherapy for age-related macular degeneration: a report by the American Academy of Ophthalmology. *Ophthalmology* 2008;115:1837–46.
- Brown DM, Kaiser PK, Michels M, et al, ANCHOR Study Group. Ranibizumab versus verteporfin for neovascular age-related macular degeneration. *N Engl J Med* 2006;355:1432–44.
- Wong TY, Chakravarthy U, Klein R, et al. The natural history and prognosis of neovascular age-related macular degeneration: a systematic review of the literature and meta-analysis. *Ophthalmology* 2008;115:116–26.
- Rosenfeld PJ, Brown DM, Heier JS, et al, MARINA Study Group. Ranibizumab for neovascular age-related macular degeneration. *N Engl J Med* 2006;355:1419–31.
- Gragoudas ES, Adamis AP, Cunningham ET Jr, et al, VEGF Inhibition Study in Ocular Neovascularization Clinical Trial Group. Pegaptanib for neovascular age-related macular degeneration. *N Engl J Med* 2004;351:2805–16.
- Gunther J, Altaweel M. Bevacizumab (Avastin) for the treatment of ocular disease. *Surv Ophthalmol* 2009;54:372–400.
- Spaide R. Ranibizumab according to need: a treatment for age-related macular degeneration. *Am J Ophthalmol* 2007; 143:679–80.
- Park JE, Chen HH, Winer J, et al. Placenta growth factor: potentiation of vascular endothelial growth factor bioactivity, in vitro and in vivo, and high affinity binding to Flt-1 but not to Flk-1/KDR. *J Biol Chem* 1994;269:25646–54.
- Avery RL, Pieramici DJ, Rabena MD, et al. Intravitreal bevacizumab (Avastin) for neovascular age-related macular degeneration. *Ophthalmology* 2006;113:363–72.
- Regillo CD, Brown DM, Abraham P, et al, PIER Study Group. Randomized, double-masked, sham-controlled trial of ranibizumab for neovascular age-related macular degeneration: PIER Study year 1. *Am J Ophthalmol* 2008;145:239–48.
- Fung AE, Lalwani GA, Rosenfeld PJ, et al. An optical coherence tomography-guided, variable dosing regimen with intravitreal ranibizumab (Lucentis) for neovascular age-related macular degeneration. *Am J Ophthalmol* 2007;143:566–83.
- Lalwani GA, Rosenfeld PJ, Fung AE, et al. A variable-dosing regimen with intravitreal ranibizumab for neovascular age-related macular degeneration: year 2 of the PRONTO Study. *Am J Ophthalmol* 2009;148:43–58.
- Rudge J, Holash J, Hylton D, et al. VEGF Trap complex formation measures production rates of VEGF, providing a biomarker for predicting efficacious angiogenic blockade. *Proc Natl Acad Sci U S A* 2007;104:18363–70.
- Holash J, Davis S, Papadopoulos N, et al. VEGF-Trap: a VEGF blocker with potent antitumor effects. *Proc Natl Acad Sci U S A* 2002;99:11393–8.
- Lee JH, Canny MD, De Erkenez A, et al. A therapeutic aptamer inhibits angiogenesis by specifically targeting the heparin binding domain of VEGF165. *Proc Natl Acad Sci U S A* 2005;102:18902–7.
- Lowe J, Araujo J, Yang J, et al. Ranibizumab inhibits multiple forms of biologically active vascular endothelial growth factor in vitro and in vivo. *Exp Eye Res* 2007;85:425–30.
- Konner J, Dupont J. Use of soluble recombinant decoy receptor vascular endothelial growth factor trap (VEGF Trap) to inhibit vascular endothelial growth factor activity. *Clin Colorectal Cancer* 2004;4(suppl):S81–5.
- Dixon JA, Oliver SC, Olson JL, Mandava N. VEGF Trap-Eye for the treatment of neovascular age-related macular degeneration. *Expert Opin Investig Drugs* 2009;18:1573–80.
- Stewart MW, Rosenfeld PJ. Predicted biological activity of intravitreal VEGF Trap. *Br J Ophthalmol* 2008;92:667–8.
- Nguyen QD, Shah SM, Browning DJ, et al. A phase I study of intravitreal vascular endothelial growth factor Trap-Eye in patients with neovascular age-related macular degeneration. *Ophthalmology* 2009;116:2141–8.
- Mitchell P, Korobelnik JF, Lanzetta P, et al. Ranibizumab (Lucentis) in neovascular age-related macular degeneration: evidence from clinical trials. *Br J Ophthalmol* 2010;94:2–13.
- Boyer DS, Heier JS, Brown DM, et al. A Phase IIIb study to evaluate the safety of ranibizumab in subjects with neovascular age-related macular degeneration. *Ophthalmology* 2009; 116:1731–9.

Footnotes and Financial Disclosures

Originally received: October 29, 2010.

Final revision: March 14, 2011.

Accepted: March 15, 2011.

Manuscript no. 2010-1500.

¹ Ophthalmic Consultants of Boston, Boston, Massachusetts.

² Retina Vitreous Association Medical Group, Beverly Hills, California.

³ The Wilmer Eye Institute, The Johns Hopkins University School of Medicine, Baltimore, Maryland.

⁴ Southeast Retina Center, Augusta, Georgia.

⁵ Retina-Vitreous Center, New Brunswick, New Jersey.

⁶ Regeneron Pharmaceuticals, Inc., Tarrytown, New York.

⁷ Retina Consultants of Houston, The Methodist Hospital, Houston, Texas.

Presented at: The Retina Society, October, 2009; American Academy of Ophthalmology Annual Meeting, October, 2009; International Symposium on Ocular Pharmacology and Therapeutics, December 2009; Association for Research in Vision and Ophthalmology Annual Meeting, April, 2010.

Financial Disclosure(s):

The authors have made the following disclosures:

Jeffrey S. Heier – Regeneron (C,S), Alcon (C), Genentech (C), Glaxo-smithkline (C), Paloma (C), Neovista (C), Oraya (C).

David Boyer – Alcon (C,L), Genentech (C,L), Regeneron (C), Novartis (C), Pfizer (C), Eyetech (C), Allergan (C,L).

Quan Dong Nguyen – Genentech (S), Regeneron (S), Novartis (S), Pfizer (S), Lux Biosciences (S), Macu Sight (S), Bausch & Lomb (C).

Dennis Marcus – Genentech (C,S), Regeneron (S), Allergan (S), Neovista (S), Pfizer (S), Ophthotech (S), Alimaera (S).

Daniel B. Roth – Regeneron (C), Allergan (C), Notal Vision (C).

George Yancopoulos – Regeneron (E,O).

Neil Stahl – Regeneron Pharmaceuticals (E).

Avner Ingerman – Regeneron Pharmaceuticals (E).

Robert Vitti – Regeneron Pharmaceuticals (E).

Alyson J. Berliner – Regeneron Pharmaceuticals (E).

Ke Yang – Regeneron Pharmaceuticals (E, O).

David M. Brown – Thrombogenics(S), Molecular Partners (C,S), Schering Plough (S), Paloma (C,S), Alimaera (S), Ophthotech (S).

Supported by Regeneron Pharmaceuticals, Inc. and Bayer HealthCare AG. The sponsors participated in the design of the study, conducting the study, data collection, data management, data analysis, interpretation of the data, and the preparation, review and approval of the manuscript.

Correspondence:

David M. Brown, MD, Retina Consultants of Houston, The Methodist Hospital, 6560 Fannin, Suite 750, Houston, TX 77030. E-mail: dmbmd@houstonretina.com.

initial work were those that were underrepresented or excluded. Indeed, eyes at the extreme of axial length and keratometry, high astigmatism, postrefractive patients, and eyes with previous surgery (e.g., penetrating keratoplasty) are notorious for difficulty in estimation of the postoperative IOL position. Analysis of these groups may demonstrate significant improvement to a very difficult problem.

Another question, and the most difficult to address, is how to address the postoperative standard of care for our patients. We believe that there is enough evidence at this time to suggest evaluating the postoperative refraction of the first eye in most patients. Refractive stability after cataract surgery occurs within a few weeks, and thus, waiting for the second eye should be considered. Our current practice is to wait at least 3 to 4 weeks between surgeries, which allows for analysis of the refractive result among other postoperative factors. Bilateral simultaneous surgery, which has been advocated by some investigators, would obviously preclude a patient from these potential advantages. Finally, we thank the authors of these 2 large retrospective studies, which will further enhance our collective outcomes as we strive for postoperative refractive perfection.

References

1. Preferred Practice Patterns. American Academy of Ophthalmology. Cataract in the Adult Eye, 2006.
2. Norrby S. Sources of error in intraocular lens power calculation. *J Cataract Refract Surg* 2008;34:368–76.
3. Aristodemou P, Cartwright NE, Sparrow JM, Johnston RL. Intraocular lens formula constant optimization and partial coherence interferometry biometry: refractive outcomes in 8108 eyes after cataract surgery. *J Cataract Refract Surg* 2011; 37:50–62.
4. Covert DJ, Henry CR, Koenig SB. Intraocular lens power selection in the second eye of patients undergoing bilateral, sequential cataract extraction. *Ophthalmology* 2010;117: 49–54.
5. Olsen T. Use of fellow eye data in the calculation of intraocular lens power for the second eye. *Ophthalmology* 2011;118: 1710–5.
6. Aristodemou P, Cartwright NE, Sparrow JM, Johnston RL. First eye prediction error improves second eye refractive outcome: results in 2129 patients after bilateral sequential cataract surgery. *Ophthalmology* 2011;118:1701–9.
7. User group for Laser Interference Biometry. Optimized IOL Constants for the Zeiss IOL Master. <http://augenklinil.uniwuerzburg.de/ulib/cl.htm>. Accessed May 17, 2011.

Erratum

With apologies from the authors, the legend for Figure 5 in “The 1-year Results of CLEAR-IT 2, a Phase 2 Study of Vascular Endothelial Growth Factor Trap-Eye Dosed As-needed After 12-week Fixed Dosing” (*Ophthalmology* 2011;118:1098-106) contained errors in the order of some of the *P* values. Below is the corrected legend with changes in boldface.

Figure 5. Mean change in best-corrected visual acuity (BCVA) in (A) all treatment groups combined and (B) individual treatment groups. The BCVA was assessed with the Early Treatment of Diabetic Retinopathy Study protocol at 4 m. Significant improvements from baseline in BCVA were noted in all treatment groups combined at week 12 (5.7 letters) and were maintained to week 52 (5.3 letters; $^{\text{¶}}P < 0.0001$). The 2 mg q4wk group showed the greatest gain in BCVA at 12 weeks, which was maintained to 52 weeks (9.0 letters; $^*P < 0.0001$; $^{\dagger}P = 0.085$; $^{\ddagger}P = \mathbf{0.344}$; $^{\S}P = \mathbf{0.0412}$; and $^{\parallel}P = \mathbf{0.0154}$ for individual groups versus baseline). The last-observation-carried-forward method was used to impute missing data. ETDRS = Early Treatment of Diabetic Retinopathy Study; PRN = as-needed.

History of Changes for Study: NCT00320775

Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

[Latest version \(submitted March 16, 2015\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification
3	<input type="radio"/>	<input type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input type="radio"/>	<input type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input type="radio"/>	<input type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: June 8, 2011 (v8)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: June 2011

Overall Status: Completed

Study Start: June 2005

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that: April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that: June 8, 2011

Met QC Criteria:

Last Update Posted: June 10, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: The purpose of this trial is to assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C. Part A is a dose escalation. Part B was terminated early. The (one) subject who received Macugen is not discussed in this website. Part C had

subjects receive one of two doses of VEGF Trap (0.15 mg or 4.0 mg).

This is the first study in which human subjects received intravitreal injections of VEGF Trap in a study eye.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Parallel Assignment

Number of Arms: 3

Masking: None (Open Label)

Allocation: Randomized

Enrollment: 51 [Actual]

Arms and Interventions

Arms	Assigned Interventions
------	------------------------

Arms	Assigned Interventions
<p>Experimental: Part A</p> <p>Part A: An open label study in which six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye. The total volume of each injection will be 100 µL. Enrollment in new dose levels will not begin until all patients in the preceding dose level have completed Visit 5 (Day 15).</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part B</p> <p>Part B: A controlled, prospective, randomized, double-masked study in which up to 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive a single ITV injection of 2.0 mg/eye VEGF Trap (or the MTD if reached prior to 2.0 mg) followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later. Enrollment into Part B will begin 2 weeks after the last subject to receive the 2.0 mg/eye dose in Part A has been observed for 15 days and it has been determined that the safety profile of VEGF Trap at this dose level is adequate to support expansion of dosing at this dose level. The dose of pegaptanib sodium will be 0.3 mg, according to the package insert.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part C</p> <p>Part C: A controlled, prospective, randomized, double-masked study in which approximately 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap. Initiation of Part C is contingent upon the 4.0 mg dose being adequately tolerated in Part A.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect
From baseline to Day 43

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on excess central retinal/lesion thickness
From baseline to Day 43
3. Best-corrected Early Treatment of Diabetic Retinopathy Study (ETDRS) visual acuity
From baseline to Day 43
4. Extent of CNV leakage

From baseline to Day 43

5. Anti-VEGF Trap antibodies in the systemic circulation

From baseline to Day 43

6. Plasma levels of VEGF Trap

From baseline to Day 43

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by optical coherence tomography (OCT).
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Retina Centers, PC
Tuscon, Arizona, United States, 85704

United States, California

Loma Linda University Health Care
Loma Linda, California, United States, 92354

United States, Illinois

University of Chicago
Chicago, Illinois, United States, 60637

United States, Maryland

Johns Hopkins Hospital School of Medicine
Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates
Charlotte, North Carolina, United States, 28120

United States, Oklahoma

Dean A. McGee Eye Institute
Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC
Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

Retina-Vitreous Associates, P.C.
Nashville, Tennessee, United States, 37203

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320775

Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

Latest version (submitted March 16, 2015) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification
3	<input type="radio"/>	<input type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input type="radio"/>	<input type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input type="radio"/>	<input type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: January 25, 2011 (v7)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: January 2011

Overall Status: Completed

Study Start: June 2005

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that: April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that: January 25, 2011

Met QC Criteria:

Last Update Posted: January 26, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: The purpose of this trial is to assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C. Part A is a dose escalation. Part B was terminated early. The (one) subject who received Macugen is not discussed in this website. Part C had

subjects receive one of two doses of VEGF Trap (0.15 mg or 4.0 mg).

This is the first study in which human subjects received intravitreal injections of VEGF Trap in a study eye.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Parallel Assignment

Number of Arms: 3

Masking: None (Open Label)

Allocation: Randomized

Enrollment: 51 [Actual]

Arms and Interventions

Arms	Assigned Interventions
------	------------------------

Arms	Assigned Interventions
<p>Experimental: Part A</p> <p>Part A: An open label study in which six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye. The total volume of each injection will be 100 µL. Enrollment in new dose levels will not begin until all patients in the preceding dose level have completed Visit 5 (Day 15).</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part B</p> <p>Part B: A controlled, prospective, randomized, double-masked study in which up to 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive a single ITV injection of 2.0 mg/eye VEGF Trap (or the MTD if reached prior to 2.0 mg) followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later. Enrollment into Part B will begin 2 weeks after the last subject to receive the 2.0 mg/eye dose in Part A has been observed for 15 days and it has been determined that the safety profile of VEGF Trap at this dose level is adequate to support expansion of dosing at this dose level. The dose of pegaptanib sodium will be 0.3 mg, according to the package insert.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part C</p> <p>Part C: A controlled, prospective, randomized, double-masked study in which approximately 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap. Initiation of Part C is contingent upon the 4.0 mg dose being adequately tolerated in Part A.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect
From baseline to Day 43

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on excess central retinal/lesion thickness
From baseline to Day 43
3. Best-corrected Early Treatment of Diabetic Retinopathy Study (ETDRS) visual acuity
From baseline to Day 43
4. Extent of CNV leakage

From baseline to Day 43

5. Anti-VEGF Trap antibodies in the systemic circulation

From baseline to Day 43

6. Plasma levels of VEGF Trap

From baseline to Day 43

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by optical coherence tomography (OCT).
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Retina Centers, PC
Tuscon, Arizona, United States, 85704

United States, California

Loma Linda University Health Care
Loma Linda, California, United States, 92354

United States, Illinois

University of Chicago
Chicago, Illinois, United States, 60637

United States, Maryland

Johns Hopkins Hospital School of Medicine
Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates
Charlotte, North Carolina, United States, 28120

United States, Oklahoma

Dean A. McGee Eye Institute
Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC
Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

Retina-Vitreous Associates, P.C.
Nashville, Tennessee, United States, 37203

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320775

Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

[Latest version \(submitted March 16, 2015\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification
3	<input type="radio"/>	<input type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input type="radio"/>	<input type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: January 26, 2010 (v6)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: January 2010

Overall Status: Completed

Study Start: June 2005

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that January 26, 2010

Met QC Criteria:

Last Update Posted: January 27, 2010 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: The purpose of this trial is to assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C. Part A is a dose escalation. Part B was terminated early. The (one) subject who received Macugen is not discussed in this website. Part C had

subjects receive one of two doses of VEGF Trap (0.15 mg or 4.0 mg).

This is the first study in which human subjects received intravitreal injections of VEGF Trap in a study eye.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Parallel Assignment

Number of Arms: 3

Masking: None (Open Label)

Allocation: Randomized

Enrollment: 51 [Actual]

Arms and Interventions

Arms	Assigned Interventions
------	------------------------

Arms	Assigned Interventions
<p>Experimental: Part A</p> <p>Part A: An open label study in which six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye. The total volume of each injection will be 100 µL. Enrollment in new dose levels will not begin until all patients in the preceding dose level have completed Visit 5 (Day 15).</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part B</p> <p>Part B: A controlled, prospective, randomized, double-masked study in which up to 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive a single ITV injection of 2.0 mg/eye VEGF Trap (or the MTD if reached prior to 2.0 mg) followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later. Enrollment into Part B will begin 2 weeks after the last subject to receive the 2.0 mg/eye dose in Part A has been observed for 15 days and it has been determined that the safety profile of VEGF Trap at this dose level is adequate to support expansion of dosing at this dose level. The dose of pegaptanib sodium will be 0.3 mg, according to the package insert.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part C</p> <p>Part C: A controlled, prospective, randomized, double-masked study in which approximately 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap. Initiation of Part C is contingent upon the 4.0 mg dose being adequately tolerated in Part A.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect
From baseline to Day 43

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on excess central retinal/lesion thickness
From baseline to Day 43
3. Best-corrected Early Treatment of Diabetic Retinopathy Study (ETDRS) visual acuity
From baseline to Day 43
4. Extent of CNV leakage

From baseline to Day 43

5. Anti-VEGF Trap antibodies in the systemic circulation

From baseline to Day 43

6. Plasma levels of VEGF Trap

From baseline to Day 43

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by optical coherence tomography (OCT).
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Retina Centers, PC
Tuscon, Arizona, United States, 85704

United States, California

Loma Linda University Health Care
Loma Linda, California, United States, 92354

United States, Illinois

University of Chicago
Chicago, Illinois, United States, 60637

United States, Maryland

Johns Hopkins Hospital School of Medicine
Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates
Charlotte, North Carolina, United States, 28120

United States, Oklahoma

Dean A. McGee Eye Institute
Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC
Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

Retina-Vitreous Associates, P.C.
Nashville, Tennessee, United States, 37203

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320775

Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

[Latest version \(submitted March 16, 2015\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification
3	<input type="radio"/>	<input type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input type="radio"/>	<input type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: April 29, 2009 (v5)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: April 2009

Overall Status: Completed

Study Start: June 2005

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that: April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that: April 29, 2009

Met QC Criteria:

Last Update Posted: April 30, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: The purpose of this trial is to assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C. Part A is a dose escalation. Part B was terminated early. The (one) subject who received Macugen is not discussed in this website. Part C had

subjects receive one of two doses of VEGF Trap (0.15 mg or 4.0 mg).

This is the first study in which human subjects received intravitreal injections of VEGF Trap in a study eye.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Parallel Assignment

Number of Arms: 3

Masking: None (Open Label)

Allocation: Randomized

Enrollment: 51 [Actual]

Arms and Interventions

Arms	Assigned Interventions
------	------------------------

Arms	Assigned Interventions
<p>Experimental: Part A</p> <p>Part A: An open label study in which six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye. The total volume of each injection will be 100 µL. Enrollment in new dose levels will not begin until all patients in the preceding dose level have completed Visit 5 (Day 15).</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part B</p> <p>Part B: A controlled, prospective, randomized, double-masked study in which up to 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive a single ITV injection of 2.0 mg/eye VEGF Trap (or the MTD if reached prior to 2.0 mg) followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later. Enrollment into Part B will begin 2 weeks after the last subject to receive the 2.0 mg/eye dose in Part A has been observed for 15 days and it has been determined that the safety profile of VEGF Trap at this dose level is adequate to support expansion of dosing at this dose level. The dose of pegaptanib sodium will be 0.3 mg, according to the package insert.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part C</p> <p>Part C: A controlled, prospective, randomized, double-masked study in which approximately 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap. Initiation of Part C is contingent upon the 4.0 mg dose being adequately tolerated in Part A.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect
From baseline to Day 43

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on excess central retinal/lesion thickness
From baseline to Day 43
3. Best-corrected Early Treatment of Diabetic Retinopathy Study (ETDRS) visual acuity
From baseline to Day 43
4. Extent of CNV leakage

From baseline to Day 43

5. Anti-VEGF Trap antibodies in the systemic circulation

From baseline to Day 43

6. Plasma levels of VEGF Trap

From baseline to Day 43

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by optical coherence tomography (OCT).
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Retina Centers, PC

Tuscon, Arizona, United States, 85704

United States, California

Loma Linda University Health Care

Loma Linda, California, United States, 92354

United States, Illinois

University of Chicago

Chicago, Illinois, United States, 60637

United States, Maryland

Johns Hopkins Hospital School of Medicine

Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates

Charlotte, North Carolina, United States, 28120

United States, Oklahoma

Dean A. McGee Eye Institute

Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC

Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

Retina-Vitreous Associates, P.C.

Nashville, Tennessee, United States, 37203

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320775

Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

[Latest version \(submitted March 16, 2015\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification
3	<input type="radio"/>	<input type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
4	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input type="radio"/>	<input type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input type="radio"/>	<input type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: January 23, 2009 (v4)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: January 2009

Overall Status: Completed

Study Start: June 2005

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that: April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that: January 23, 2009

Met QC Criteria:

Last Update Posted: January 27, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: The purpose of this trial is to assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Parallel Assignment

Number of Arms: 3

Masking: None (Open Label)

Allocation: Randomized

Enrollment: 51 [Actual]

Arms and Interventions

Arms	Assigned Interventions
------	------------------------

Arms	Assigned Interventions
<p>Experimental: Part A</p> <p>Part A: An open label study in which six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye. The total volume of each injection will be 100 µL. Enrollment in new dose levels will not begin until all patients in the preceding dose level have completed Visit 5 (Day 15).</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> Aflibercept

Arms	Assigned Interventions
<p>Sham Comparator: Part B</p> <p>Part B: A controlled, prospective, randomized, double-masked study in which up to 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive a single ITV injection of 2.0 mg/eye VEGF Trap (or the MTD if reached prior to 2.0 mg) followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later. Enrollment into Part B will begin 2 weeks after the last subject to receive the 2.0 mg/eye dose in Part A has been observed for 15 days and it has been determined that the safety profile of VEGF Trap at this dose level is adequate to support expansion of dosing at this dose level. The dose of pegaptanib sodium will be 0.3 mg, according to the package insert.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Arms	Assigned Interventions
<p>Part C</p> <p>Part C: A controlled, prospective, randomized, double-masked study in which approximately 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap. Initiation of Part C is contingent upon the 4.0 mg dose being adequately tolerated in Part A.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect
From baseline to Day 43

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on excess central retinal/lesion thickness
From baseline to Day 43
3. Best-corrected Early Treatment of Diabetic Retinopathy Study (ETDRS) visual acuity
From baseline to Day 43
4. Extent of CNV leakage

From baseline to Day 43

5. Anti-VEGF Trap antibodies in the systemic circulation

From baseline to Day 43

6. Plasma levels of VEGF Trap

From baseline to Day 43

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by optical coherence tomography (OCT).
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Retina Centers, PC
Tuscon, Arizona, United States, 85704

United States, California

Loma Linda University Health Care
Loma Linda, California, United States, 92354

United States, Illinois

University of Chicago
Chicago, Illinois, United States, 60637

United States, Maryland

Johns Hopkins Hospital School of Medicine
Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates
Charlotte, North Carolina, United States, 28120

United States, Oklahoma

Dean A. McGee Eye Institute
Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC
Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

Retina-Vitreous Associates, P.C.
Nashville, Tennessee, United States, 37203

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320775

Safety and Tolerability of Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Neovascular Age-Related Macular Degeneration (AMD)

Latest version (submitted March 16, 2015) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	April 28, 2006	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	October 3, 2006	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification

Version	A	B	Submitted Date	Changes
3	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
4	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input type="radio"/>	<input type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input type="radio"/>	<input type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: July 25, 2007 (v3)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability of Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Neovascular Age-Related Macular Degeneration (AMD)

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: July 2007

Overall Status: Active, not recruiting

Study Start: June 2005

Primary Completion:

Study Completion:

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that July 25, 2007

Met QC Criteria:

Last Update Posted: July 27, 2007 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring:

Study Description

Brief Summary: The purpose of this trial is to assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model:

Number of Arms:

Masking: (masked roles unspecified)

Allocation: N/A

Enrollment: 96

Arms and Interventions

Intervention Details:

Drug: VEGF Trap

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on excess central retinal/lesion thickness
3. Best-corrected Early Treatment of Diabetic Retinopathy Study (ETDRS) visual acuity
4. Extent of CNV leakage
5. Anti-VEGF Trap antibodies in the systemic circulation
6. Plasma levels of VEGF Trap

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by optical coherence tomography (OCT).
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Locations: **United States, Arizona**

Tuscon, Arizona, United States, 85704

United States, California

Loma Linda, California, United States, 92354

United States, Illinois

Chicago, Illinois, United States, 60637

United States, Maryland

Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte, North Carolina, United States, 28120

United States, Ohio

Cleveland, Ohio, United States, 44195

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

Nashville, Tennessee, United States, 37203

United States, Wisconsin

Madison, Wisconsin, United States, 53705

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

History of Changes for Study: NCT00320775

Safety and Tolerability of Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Neovascular Age-Related Macular Degeneration (AMD)

Latest version (submitted March 16, 2015) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	April 28, 2006	None (earliest Version on record)
2	<input checked="" type="radio"/>	<input checked="" type="radio"/>	October 3, 2006	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
4	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input type="radio"/>	<input type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input type="radio"/>	<input type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: October 3, 2006 (v2)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability of Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Neovascular Age-Related Macular Degeneration (AMD)

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: April 2006

Overall Status: Recruiting

Study Start: June 2005

Primary Completion:

Study Completion:

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that October 3, 2006

Met QC Criteria:

Last Update Posted: October 4, 2006 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring:

Study Description

Brief Summary: The purpose of this trial is to assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model:

Number of Arms:

Masking: (masked roles unspecified)

Allocation: N/A

Enrollment: 96

Arms and Interventions

Intervention Details:

Drug: VEGF Trap

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on excess central retinal/lesion thickness
3. Best-corrected Early Treatment of Diabetic Retinopathy Study (ETDRS) visual acuity
4. Extent of CNV leakage
5. Anti-VEGF Trap antibodies in the systemic circulation
6. Plasma levels of VEGF Trap

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by optical coherence tomography (OCT).
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Central Contact: Regeneron

Email: VEGF.Trap@regeneron.com

Locations: **United States, Arizona**

[Recruiting]

Tuscon, Arizona, United States, 85704

United States, California

[Recruiting]

Loma Linda, California, United States, 92354

United States, Illinois

[Recruiting]

Chicago, Illinois, United States, 60637

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21287

United States, North Carolina

[Recruiting]

Charlotte, North Carolina, United States, 28120

United States, Ohio

[Recruiting]

Cleveland, Ohio, United States, 44195

United States, Oklahoma

[Recruiting]

Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

[Recruiting]

Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Wisconsin

[Recruiting]

Madison, Wisconsin, United States, 53705

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320775

Safety and Tolerability of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Latest version (submitted March 16, 2015) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input checked="" type="radio"/>	<input checked="" type="radio"/>	April 28, 2006	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	October 3, 2006	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
4	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input type="radio"/>	<input type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input type="radio"/>	<input type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: April 28, 2006 (v1)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: April 2006

Overall Status: Recruiting

Study Start: June 2005

Primary Completion:

Study Completion: June 2006

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that April 28, 2006

Met QC Criteria:

Last Update Posted: May 3, 2006 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring:

Study Description

Brief Summary: To assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C.

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model:

Number of Arms:

Masking: (masked roles unspecified)

Allocation: N/A

Enrollment: 96

Arms and Interventions

Intervention Details:

Drug: VEGF Trap

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on: excess central retinal/lesion thickness, best-corrected ETDRS visual acuity, extent of CNV leakage, anti-VEGF Trap antibodies in the systemic circulation, plasma levels of VEGF Trap

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by OCT.
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Central Contact: Regeneron

Email: VEGF.Trap@regeneron.com

Locations: **United States, Arizona**

[Recruiting]

Tuscon, Arizona, United States, 85704

United States, California

[Recruiting]

Loma Linda, California, United States, 92354

United States, Illinois

[Recruiting]

Chicago, Illinois, United States, 60637

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21287

United States, North Carolina

[Recruiting]

Charlotte, North Carolina, United States, 28120

United States, Ohio

[Recruiting]

Cleveland, Ohio, United States, 44195

United States, Oklahoma

[Recruiting]

Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

[Recruiting]

Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Wisconsin

[Recruiting]

Madison, Wisconsin, United States, 53705

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

History of Changes for Study: NCT00320775

Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

[Latest version \(submitted March 16, 2015\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Outcome Measures, Conditions, Study Status, Eligibility, Study Description and Study Identification
3	<input type="radio"/>	<input type="radio"/>	<u>July 25, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Study Design, Arms and Interventions, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
5	<input type="radio"/>	<input type="radio"/>	<u>April 29, 2009</u>	Study Status, Arms and Interventions, Study Description and Sponsor/Collaborators
6	<input type="radio"/>	<input type="radio"/>	<u>January 26, 2010</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status
9	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>March 16, 2015</u>	Sponsor/Collaborators, Study Status and References

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320775

Submitted Date: March 16, 2015 (v9)

Study Identification

Unique Protocol ID: VGFT-OD-0502

Brief Title: Safety and Tolerability Study of Intravitreal VEGF-Trap Administration in Patients With Neovascular AMD

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: March 2015

Overall Status: Completed

Study Start: June 2005

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that March 16, 2015

Met QC Criteria:

Last Update Posted: March 18, 2015 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party: Sponsor

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: The purpose of this trial is to assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This study consists of three parts, Part A, Part B and Part C. Part A is a dose escalation. Part B was terminated early. The (one) subject who received Macugen is not discussed in this website. Part C had

subjects receive one of two doses of VEGF Trap (0.15 mg or 4.0 mg).

This is the first study in which human subjects received intravitreal injections of VEGF Trap in a study eye.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Parallel Assignment

Number of Arms: 3

Masking: None (Open Label)

Allocation: Randomized

Enrollment: 51 [Actual]

Arms and Interventions

Arms	Assigned Interventions
------	------------------------

Arms	Assigned Interventions
<p>Experimental: Part A</p> <p>Part A: An open label study in which six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye. The total volume of each injection will be 100 µL. Enrollment in new dose levels will not begin until all patients in the preceding dose level have completed Visit 5 (Day 15).</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part B</p> <p>Part B: A controlled, prospective, randomized, double-masked study in which up to 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive a single ITV injection of 2.0 mg/eye VEGF Trap (or the MTD if reached prior to 2.0 mg) followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later. Enrollment into Part B will begin 2 weeks after the last subject to receive the 2.0 mg/eye dose in Part A has been observed for 15 days and it has been determined that the safety profile of VEGF Trap at this dose level is adequate to support expansion of dosing at this dose level. The dose of pegaptanib sodium will be 0.3 mg, according to the package insert.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> Aflibercept

Arms	Assigned Interventions
<p>Active Comparator: Part C</p> <p>Part C: A controlled, prospective, randomized, double-masked study in which approximately 30 subjects meeting eligibility criteria will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap. Initiation of Part C is contingent upon the 4.0 mg dose being adequately tolerated in Part A.</p>	<p>Drug: VEGF Trap</p> <p>Part A: Six successive cohorts of 3-6 patients each with neovascular AMD will receive a single intravitreal (ITV) injection of 0.05, 0.15, 0.5, 1.0, 2.0, or 4.0 mg of VEGF Trap into the study eye.</p> <p>Part B: Up to 30 subjects will be randomly assigned in a 1:1 ratio to receive a single of 2.0 mg/eye VEGF Trap followed by 1 sham injection six weeks later, or an initial dose of 0.3 mg pegaptanib sodium into the study eye, followed by a second dose six weeks later.</p> <p>Part C: Approximately 30 subjects will be randomly assigned in a 1:1 ratio to receive up to two ITV injections of either 0.15 or 4.0 mg/eye VEGF Trap.</p> <p>After completion of Visit 8 (Day 57), patients from all parts of the study, may be eligible to continue in Open-label Extension and will receive 4.0 mg of VEGF Trap.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, bioeffect
From baseline to Day 43

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on excess central retinal/lesion thickness
From baseline to Day 43
3. Best-corrected Early Treatment of Diabetic Retinopathy Study (ETDRS) visual acuity
From baseline to Day 43
4. Extent of CNV leakage

From baseline to Day 43

5. Anti-VEGF Trap antibodies in the systemic circulation

From baseline to Day 43

6. Plasma levels of VEGF Trap

From baseline to Day 43

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal/lesion thickness $\geq 250\mu\text{m}$ as measured by optical coherence tomography (OCT).
- ETDRS best-corrected visual acuity of:
 - 20/40 (73 letters) or worse
- Clear ocular media and clear lens(es) to permit good quality stereoscopic fundus photography.

Exclusion Criteria:

- Prior treatment with VEGF Trap, bevacizumab or ranibizumab.
- Any investigational agent within 12 weeks of Visit 2 (Day 1).
- Presence of other causes of CNV.
- Active ocular infection.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Retina Centers, PC
Tuscon, Arizona, United States, 85704

United States, California

Loma Linda University Health Care
Loma Linda, California, United States, 92354

United States, Illinois

University of Chicago
Chicago, Illinois, United States, 60637

United States, Maryland

Johns Hopkins Hospital School of Medicine
Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates
Charlotte, North Carolina, United States, 28120

United States, Oklahoma

Dean A. McGee Eye Institute
Oklahoma City, Oklahoma, United States, 73104

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC
Philadelphia, Pennsylvania, United States, 19107

United States, Tennessee

Retina-Vitreous Associates, P.C.
Nashville, Tennessee, United States, 37203

IPDSharing

Plan to Share IPD:

References

Citations: **[Study Results]** Nguyen QD, Campochiaro PA, Shah SM, Browning DJ, Hudson HL, Sonkin PL, Hariprasad SM, Kaiser PK, Slakter J, Haller JA, Do DV, Mieler W, Chu K, Ingerman A, Vitti R, Berliner AJ, Cedarbaum J; Clear-It 1 Investigators. Evaluation of very high- and very low-dose intravitreal aflibercept in patients with neovascular age-related macular degeneration. J Ocul Pharmacol Ther. 2012 Dec;28(6):581-8. doi: 10.1089/jop.2011.0261. Epub 2012 Jul 9. PubMed 22775078

[Study Results] Do DV, Schmidt-Erfurth U, Gonzalez VH, Gordon CM, Tolentino M, Berliner AJ, Vitti R, Rückert R, Sandbrink R, Stein D, Yang K, Beckmann K, Heier JS. The DA VINCI Study: phase 2 primary results of VEGF Trap-Eye in patients with diabetic macular edema. Ophthalmology. 2011 Sep;118(9):1819-26. doi: 10.1016/j.ophtha.2011.02.018. Epub 2011 May 5. PubMed 21546089

Links: URL: <http://www.ncbi.nlm.nih.gov/pubmed/22775078>

Description: Related Info

URL: <http://www.ncbi.nlm.nih.gov/pubmed/21546089>

Description: Related Info

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Latest version (submitted January 27, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 1, 2006</u>	Contacts/Locations and Study Status
3	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input type="radio"/>	<input type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: December 1, 2011 (v9)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: December 2011

Overall Status: Completed

Study Start: April 2006

Primary Completion: July 2007 [Actual]

Study Completion: June 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Certification/Extension November 30, 2010

First Submitted:

Certification/Extension November 30, 2010

First Submitted that

Met QC Criteria:

Certification/Extension December 2, 2010 [Estimate]

First Posted:

Last Update Submitted that December 1, 2011

Met QC Criteria:

Last Update Posted: December 8, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party: Sponsor

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: This study examines the effect of intravitreally administered VEGF Trap in patients with wet AMD.

The purpose of this trial is to assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of intravitreal (ITV) injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms: 1

Masking: Triple (Participant, Care Provider, Investigator)

Allocation: Randomized

Enrollment: 159 [Actual]

Arms and Interventions

Arms	Assigned Interventions
<p>1</p> <p>This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of ITV injections of VEGF Trap into the study eye at 4- or 12-week intervals over a 12-week period.</p> <p>Beginning with Week 16, all patients will be evaluated every 4 weeks for continued dosing for up to one year. The fellow eye may also be eligible for treatment with VEGF Trap beginning at this</p>	<p>Drug: VEGF Trap</p> <p>Subjects in Group A will receive 2 ITV injections of 0.5 mg VEGF Trap at 12-week intervals; Group B subjects will receive 4 ITV injections of 0.5 mg VEGF Trap at 4-week intervals; Group C subjects will receive 2 injections of 2.0 mg VEGF Trap at 12-week intervals; Group D subjects will receive 4 injections of 2.0 mg VEGF Trap at 4-week intervals, and Group E subjects will receive 2 injections of 4.0 mg VEGF Trap at 12-week intervals.</p> <p>Other Names:</p> <ul style="list-style-type: none">• Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety, biological effect (optical coherence tomography [OCT], fluorescein angiography, visual acuity)
From baseline to week 12

Secondary Outcome Measures:

2. Pharmacokinetics, immunogenicity, quality of life
From baseline to week 12

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness ≥ 300 μm as measured by OCT.
- Early Treatment of Diabetic Retinopathy Study (ETDRS) best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - Photodynamic therapy (PDT) within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).
 - Juxtasceral steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
 - Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.
 - Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Associated Retina Consultants

Phoenix, Arizona, United States, 85020

Retina Centers, PC

Tucson, Arizona, United States, 85704

United States, California

Retina Vitreous Associates Medical Group

Beverly Hills, California, United States, 90211

Loma Linda University Health Care

Loma Linda, California, United States, 92354

United States, Georgia

Southeast Retina Center

Augusta, Georgia, United States, 30909

United States, Illinois

University of Chicago

Chicago, Illinois, United States, 60637

United States, Indiana

Midwest Eye Institute

Indianapolis, Indiana, United States, 46280

United States, Maryland

Johns Hopkins Hospital School of Medicine

Baltimore, Maryland, United States, 21287

United States, Massachusetts

Ophthalmic Consultants of Boston

Boston, Massachusetts, United States, 02114

New England Retina Consultants PC

West Springfield, Massachusetts, United States, 10189

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates

Charlotte, North Carolina, United States, 28210

United States, Oklahoma

Dean A. McGee Eye Institute

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Retina Northwest PC

Portland, Oregon, United States, 97210

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC

Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

Black Hills Regional Eye Institute

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Retina-Vitreous Associates, P.C.

Nashville, Tennessee, United States, 37203

United States, Texas

Vitreoretinal Consultants Scurlock Tower Texas Medical Center

Houston, Texas, United States, 77030

Medical Center Ophthalmology

San Antonio, Texas, United States, 78240

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Latest version (submitted January 27, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 1, 2006</u>	Contacts/Locations and Study Status
3	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input type="radio"/>	<input type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: April 20, 2011 (v8)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: April 2011

Overall Status: Completed

Study Start: April 2006

Primary Completion: July 2007 [Actual]

Study Completion: June 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Certification/Extension November 30, 2010

First Submitted:

Certification/Extension November 30, 2010

First Submitted that

Met QC Criteria:

Certification/Extension December 2, 2010 [Estimate]

First Posted:

Last Update Submitted that April 20, 2011

Met QC Criteria:

Last Update Posted: April 28, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: This study examines the effect of intravitreally administered VEGF Trap in patients with wet AMD.

The purpose of this trial is to assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of intravitreal (ITV) injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms: 1

Masking: Triple (Participant, Care Provider, Investigator)

Allocation: Randomized

Enrollment: 159 [Actual]

Arms and Interventions

Arms	Assigned Interventions
<p>1</p> <p>This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of ITV injections of VEGF Trap into the study eye at 4- or 12-week intervals over a 12-week period.</p> <p>Beginning with Week 16, all patients will be evaluated every 4 weeks for continued dosing for up to one year. The fellow eye may also be eligible for treatment with VEGF Trap beginning at this</p>	<p>Drug: VEGF Trap</p> <p>Subjects in Group A will receive 2 ITV injections of 0.5 mg VEGF Trap at 12-week intervals; Group B subjects will receive 4 ITV injections of 0.5 mg VEGF Trap at 4-week intervals; Group C subjects will receive 2 injections of 2.0 mg VEGF Trap at 12-week intervals; Group D subjects will receive 4 injections of 2.0 mg VEGF Trap at 4-week intervals, and Group E subjects will receive 2 injections of 4.0 mg VEGF Trap at 12-week intervals.</p> <p>Other Names:</p> <ul style="list-style-type: none">• Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety, biological effect (optical coherence tomography [OCT], fluorescein angiography, visual acuity)
From baseline to week 12

Secondary Outcome Measures:

2. Pharmacokinetics, immunogenicity, quality of life
From baseline to week 12

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness ≥ 300 μm as measured by OCT.
- Early Treatment of Diabetic Retinopathy Study (ETDRS) best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - Photodynamic therapy (PDT) within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).
 - Juxtasceral steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
 - Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.
 - Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Associated Retina Consultants

Phoenix, Arizona, United States, 85020

Retina Centers, PC

Tucson, Arizona, United States, 85704

United States, California

Retina Vitreous Associates Medical Group

Beverly Hills, California, United States, 90211

Loma Linda University Health Care

Loma Linda, California, United States, 92354

United States, Georgia

Southeast Retina Center

Augusta, Georgia, United States, 30909

United States, Illinois

University of Chicago

Chicago, Illinois, United States, 60637

United States, Indiana

Midwest Eye Institute

Indianapolis, Indiana, United States, 46280

United States, Maryland

Johns Hopkins Hospital School of Medicine

Baltimore, Maryland, United States, 21287

United States, Massachusetts

Ophthalmic Consultants of Boston

Boston, Massachusetts, United States, 02114

New England Retina Consultants PC

West Springfield, Massachusetts, United States, 10189

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates

Charlotte, North Carolina, United States, 28210

United States, Oklahoma

Dean A. McGee Eye Institute

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Retina Northwest PC

Portland, Oregon, United States, 97210

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC

Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

Black Hills Regional Eye Institute

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Retina-Vitreous Associates, P.C.

Nashville, Tennessee, United States, 37203

United States, Texas

Vitreoretinal Consultants Scurlock Tower Texas Medical Center

Houston, Texas, United States, 77030

Medical Center Ophthalmology

San Antonio, Texas, United States, 78240

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Latest version (submitted January 27, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 1, 2006</u>	Contacts/Locations and Study Status
3	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input type="radio"/>	<input type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: November 30, 2010 (v7)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: November 2010

Overall Status: Completed

Study Start: April 2006

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Certification/Extension November 30, 2010

First Submitted:

Certification/Extension November 30, 2010

First Submitted that

Met QC Criteria:

Certification/Extension December 2, 2010 [Estimate]

First Posted:

Last Update Submitted that November 30, 2010

Met QC Criteria:

Last Update Posted: December 2, 2010 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: This study examines the effect of intravitreally administered VEGF Trap in patients with wet AMD.

The purpose of this trial is to assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of intravitreal (ITV) injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms: 1

Masking: Triple (Participant, Care Provider, Investigator)

Allocation: Randomized

Enrollment: 159 [Actual]

Arms and Interventions

Arms	Assigned Interventions
<p>1</p> <p>This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of ITV injections of VEGF Trap into the study eye at 4- or 12-week intervals over a 12-week period.</p> <p>Beginning with Week 16, all patients will be evaluated every 4 weeks for continued dosing for up to one year. The fellow eye may also be eligible for treatment with VEGF Trap beginning at this</p>	<p>Drug: VEGF Trap</p> <p>Subjects in Group A will receive 2 ITV injections of 0.5 mg VEGF Trap at 12-week intervals; Group B subjects will receive 4 ITV injections of 0.5 mg VEGF Trap at 4-week intervals; Group C subjects will receive 2 injections of 2.0 mg VEGF Trap at 12-week intervals; Group D subjects will receive 4 injections of 2.0 mg VEGF Trap at 4-week intervals, and Group E subjects will receive 2 injections of 4.0 mg VEGF Trap at 12-week intervals.</p> <p>Other Names:</p> <ul style="list-style-type: none">• Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety, biological effect (optical coherence tomography [OCT], fluorescein angiography, visual acuity)
From baseline to week 12

Secondary Outcome Measures:

2. Pharmacokinetics, immunogenicity, quality of life
From baseline to week 12

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness ≥ 300 μm as measured by OCT.
- Early Treatment of Diabetic Retinopathy Study (ETDRS) best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - Photodynamic therapy (PDT) within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).
 - Juxtasceral steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
 - Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.
 - Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Associated Retina Consultants

Phoenix, Arizona, United States, 85020

Retina Centers, PC

Tucson, Arizona, United States, 85704

United States, California

Retina Vitreous Associates Medical Group

Beverly Hills, California, United States, 90211

Loma Linda University Health Care

Loma Linda, California, United States, 92354

United States, Georgia

Southeast Retina Center

Augusta, Georgia, United States, 30909

United States, Illinois

University of Chicago

Chicago, Illinois, United States, 60637

United States, Indiana

Midwest Eye Institute

Indianapolis, Indiana, United States, 46280

United States, Maryland

Johns Hopkins Hospital School of Medicine

Baltimore, Maryland, United States, 21287

United States, Massachusetts

Ophthalmic Consultants of Boston

Boston, Massachusetts, United States, 02114

New England Retina Consultants PC

West Springfield, Massachusetts, United States, 10189

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates

Charlotte, North Carolina, United States, 28210

United States, Oklahoma

Dean A. McGee Eye Institute

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Retina Northwest PC

Portland, Oregon, United States, 97210

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC

Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

Black Hills Regional Eye Institute

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Retina-Vitreous Associates, P.C.

Nashville, Tennessee, United States, 37203

United States, Texas

Vitreoretinal Consultants Scurlock Tower Texas Medical Center

Houston, Texas, United States, 77030

Medical Center Ophthalmology

San Antonio, Texas, United States, 78240

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Latest version (submitted January 27, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 1, 2006</u>	Contacts/Locations and Study Status
3	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input type="radio"/>	<input type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: April 28, 2009 (v6)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: April 2009

Overall Status: Completed

Study Start: April 2006

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that: April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that: April 28, 2009

Met QC Criteria:

Last Update Posted: April 29, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: This study examines the effect of intravitreally administered VEGF Trap in patients with wet AMD.

The purpose of this trial is to assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of intravitreal (ITV) injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms: 1

Masking: Triple (Participant, Care Provider, Investigator)

Allocation: Randomized

Enrollment: 159 [Actual]

Arms and Interventions

Arms	Assigned Interventions
------	------------------------

Arms	Assigned Interventions
<p>1</p> <p>This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of ITV injections of VEGF Trap into the study eye at 4- or 12-week intervals over a 12-week period.</p> <p>Beginning with Week 16, all patients will be evaluated every 4 weeks for continued dosing for up to one year. The fellow eye may also be eligible for treatment with VEGF Trap beginning at this</p>	<p>Drug: VEGF Trap</p> <p>Subjects in Group A will receive 2 ITV injections of 0.5 mg VEGF Trap at 12-week intervals; Group B subjects will receive 4 ITV injections of 0.5 mg VEGF Trap at 4-week intervals; Group C subjects will receive 2 injections of 2.0 mg VEGF Trap at 12-week intervals; Group D subjects will receive 4 injections of 2.0 mg VEGF Trap at 4-week intervals, and Group E subjects will receive 2 injections of 4.0 mg VEGF Trap at 12-week intervals.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety, biological effect (optical coherence tomography [OCT], fluorescein angiography, visual acuity)
From baseline to week 12

Secondary Outcome Measures:

2. Pharmacokinetics, immunogenicity, quality of life
From baseline to week 12

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness ≥ 300 μm as measured by OCT.
- Early Treatment of Diabetic Retinopathy Study (ETDRS) best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - Photodynamic therapy (PDT) within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).
 - Juxtасcleral steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
 - Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.
 - Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Associated Retina Consultants

Phoenix, Arizona, United States, 85020

Retina Centers, PC

Tucson, Arizona, United States, 85704

United States, California

Retina Vitreous Associates Medical Group

Beverly Hills, California, United States, 90211

Loma Linda University Health Care

Loma Linda, California, United States, 92354

United States, Georgia

Southeast Retina Center

Augusta, Georgia, United States, 30909

United States, Illinois

University of Chicago

Chicago, Illinois, United States, 60637

United States, Indiana

Midwest Eye Institute

Indianapolis, Indiana, United States, 46280

United States, Maryland

Johns Hopkins Hospital School of Medicine

Baltimore, Maryland, United States, 21287

United States, Massachusetts

Ophthalmic Consultants of Boston

Boston, Massachusetts, United States, 02114

New England Retina Consultants PC

West Springfield, Massachusetts, United States, 10189

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates

Charlotte, North Carolina, United States, 28210

United States, Oklahoma

Dean A. McGee Eye Institute

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Retina Northwest PC

Portland, Oregon, United States, 97210

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC

Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

Black Hills Regional Eye Institute

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Retina-Vitreous Associates, P.C.

Nashville, Tennessee, United States, 37203

United States, Texas

Vitreoretinal Consultants Scurlock Tower Texas Medical Center

Houston, Texas, United States, 77030

Medical Center Ophthalmology

San Antonio, Texas, United States, 78240

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Latest version (submitted January 27, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 1, 2006</u>	Contacts/Locations and Study Status
3	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input type="radio"/>	<input type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: January 23, 2009 (v5)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of VEGF-Trap in Patients With Wet AMD

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: January 2009

Overall Status: Completed

Study Start: April 2006

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that: April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that: January 23, 2009

Met QC Criteria:

Last Update Posted: January 27, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: This study examines the effect of intravitreally administered VEGF Trap in patients with wet AMD.

The purpose of this trial is to assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of intravitreal (ITV) injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms: 1

Masking: Triple (Participant, Care Provider, Investigator)

Allocation: Randomized

Enrollment: 159 [Actual]

Arms and Interventions

Arms	Assigned Interventions
------	------------------------

Arms	Assigned Interventions
<p>1</p> <p>This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of ITV injections of VEGF Trap into the study eye at 4- or 12-week intervals over a 12-week period.</p> <p>Beginning with Week 16, all patients will be evaluated every 4 weeks for continued dosing for up to one year. The fellow eye may also be eligible for treatment with VEGF Trap beginning at this</p>	<p>Drug: VEGF Trap</p> <p>Subjects in Group A will receive 2 ITV injections of 0.5 mg VEGF Trap at 12-week intervals; Group B subjects will receive 4 ITV injections of 0.5 mg VEGF Trap at 4-week intervals; Group C subjects will receive 2 injections of 2.0 mg VEGF Trap at 12-week intervals; Group D subjects will receive 4 injections of 2.0 mg VEGF Trap at 4-week intervals, and Group E subjects will receive 2 injections of 4.0 mg VEGF Trap at 12-week intervals.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • Aflibercept

Outcome Measures

Primary Outcome Measures:

1. Safety, biological effect (optical coherence tomography [OCT], fluorescein angiography, visual acuity)
From baseline to week 12

Secondary Outcome Measures:

2. Pharmacokinetics, immunogenicity, quality of life
From baseline to week 12

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness ≥ 300 μm as measured by OCT.
- Early Treatment of Diabetic Retinopathy Study (ETDRS) best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - Photodynamic therapy (PDT) within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).
 - Juxtasclear steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
 - Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.
 - Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Associated Retina Consultants

Phoenix, Arizona, United States, 85020

Retina Centers, PC

Tucson, Arizona, United States, 85704

United States, California

Retina Vitreous Associates Medical Group

Beverly Hills, California, United States, 90211

Loma Linda University Health Care

Loma Linda, California, United States, 92354

United States, Georgia

Southeast Retina Center

Augusta, Georgia, United States, 30909

United States, Illinois

University of Chicago

Chicago, Illinois, United States, 60637

United States, Indiana

Midwest Eye Institute

Indianapolis, Indiana, United States, 46280

United States, Maryland

Johns Hopkins Hospital School of Medicine

Baltimore, Maryland, United States, 21287

United States, Massachusetts

Ophthalmic Consultants of Boston

Boston, Massachusetts, United States, 02114

New England Retina Consultants PC

West Springfield, Massachusetts, United States, 10189

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates

Charlotte, North Carolina, United States, 28210

United States, Oklahoma

Dean A. McGee Eye Institute

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Retina Northwest PC

Portland, Oregon, United States, 97210

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC

Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

Black Hills Regional Eye Institute

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Retina-Vitreous Associates, P.C.

Nashville, Tennessee, United States, 37203

United States, Texas

Vitreoretinal Consultants Scurlock Tower Texas Medical Center

Houston, Texas, United States, 77030

Medical Center Ophthalmology

San Antonio, Texas, United States, 78240

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Wet Age-Related Macular Degeneration (AMD)

[Latest version \(submitted January 27, 2012\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 1, 2006</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification
4	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input type="radio"/>	<input type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: July 24, 2007 (v4)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Wet Age-Related Macular Degeneration (AMD)

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: July 2007

Overall Status: Active, not recruiting

Study Start: April 2006

Primary Completion:

Study Completion:

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that July 24, 2007

Met QC Criteria:

Last Update Posted: July 26, 2007 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring:

Study Description

Brief Summary: This study examines the effect of intravitreally administered VEGF Trap in patients with wet AMD.

The purpose of this trial is to assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of intravitreal (ITV) injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms:

Masking: Double (masked roles unspecified)

Allocation: Randomized

Enrollment: 150

Arms and Interventions

Intervention Details:

Drug: VEGF Trap

Outcome Measures

Primary Outcome Measures:

1. Safety, biological effect (optical coherence tomography [OCT], fluorescein angiography, visual acuity)

Secondary Outcome Measures:

2. Pharmacokinetics, immunogenicity, quality of life

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness ≥ 300 μm as measured by OCT.
- Early Treatment of Diabetic Retinopathy Study (ETDRS) best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - Photodynamic therapy (PDT) within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).

- Juxtapapillary steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
- Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.
- Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director

Locations: **United States, Arizona**

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

United States, California

Beverly Hills, California, United States, 90211

Irvine, California, United States, 92697

Loma Linda, California, United States, 92354

Menlo Park, California, United States, 94025

Palm Springs, California, United States, 92262

United States, Florida

Ft. Myers, Florida, United States, 33912

Lakeland, Florida, United States, 33805

Oakland Park, Florida, United States, 33334

Orlando, Florida, United States, 32803

United States, Georgia

Augusta, Georgia, United States, 30909

United States, Illinois

Chicago, Illinois, United States, 60637

Glenview, Illinois, United States, 60025

United States, Indiana

Indianapolis, Indiana, United States, 46280

United States, Maryland

Baltimore, Maryland, United States, 21204

Baltimore, Maryland, United States, 21287

United States, Massachusetts

Boston, Massachusetts, United States, 02114

Peabody, Massachusetts, United States

West Springfield, Massachusetts, United States, 10189

United States, Michigan

Ann Arbor, Michigan, United States, 48105

United States, New Jersey

Toms River, New Jersey, United States, 08755

United States, New York

Great Neck, New York, United States, 11021

New York, New York, United States, 10032

United States, North Carolina

Charlotte, North Carolina, United States, 28210

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Portland, Oregon, United States, 97210

United States, Pennsylvania

Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Nashville, Tennessee, United States, 37203

United States, Texas

Houston, Texas, United States, 77030

San Antonio, Texas, United States, 78240

United States, Wisconsin

Madison, Wisconsin, United States, 53705

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Wet Age-Related Macular Degeneration (AMD)

[Latest version \(submitted January 27, 2012\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 1, 2006</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
3	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>October 3, 2006</u>	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification
4	<input type="radio"/>	<input type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input type="radio"/>	<input type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: October 3, 2006 (v3)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Wet Age-Related Macular Degeneration (AMD)

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: August 2006

Overall Status: Recruiting

Study Start: April 2006

Primary Completion:

Study Completion:

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that October 3, 2006

Met QC Criteria:

Last Update Posted: October 4, 2006 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring:

Study Description

Brief Summary: This study examines the effect of intravitreally administered VEGF Trap in patients with wet AMD.

The purpose of this trial is to assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of intravitreal (ITV) injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms:

Masking: Double (masked roles unspecified)

Allocation: Randomized

Enrollment: 150

Arms and Interventions

Intervention Details:

Drug: VEGF Trap

Outcome Measures

Primary Outcome Measures:

1. Safety, biological effect (optical coherence tomography [OCT], fluorescein angiography, visual acuity)

Secondary Outcome Measures:

2. Pharmacokinetics, immunogenicity, quality of life

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness $\geq 300 \mu\text{m}$ as measured by OCT.
- Early Treatment of Diabetic Retinopathy Study (ETDRS) best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - Photodynamic therapy (PDT) within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).

- Juxtapapillary steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
- Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.
- Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Central Contact: Regeneron

Email: VEGF.Trap@regeneron.com

Locations: **United States, Arizona**

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tucson, Arizona, United States, 85704

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Not yet recruiting]

Irvine, California, United States, 92697

[Not yet recruiting]

Loma Linda, California, United States, 92354

[Not yet recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Palm Springs, California, United States, 92262

United States, Florida

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Recruiting]

Lakeland, Florida, United States, 33805

[Recruiting]

Oakland Park, Florida, United States, 33334

[Not yet recruiting]

Orlando, Florida, United States, 32803

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

United States, Illinois

[Not yet recruiting]

Chicago, Illinois, United States, 60637

[Not yet recruiting]

Glenview, Illinois, United States, 60025

United States, Indiana

[Recruiting]

Indianapolis, Indiana, United States, 46280

United States, Maryland

[Not yet recruiting]

Baltimore, Maryland, United States, 21204

[Recruiting]

Baltimore, Maryland, United States, 21287

United States, Massachusetts

[Recruiting]

Boston, Massachusetts, United States, 02114

[Recruiting]

Peabody, Massachusetts, United States

[Recruiting]

West Springfield, Massachusetts, United States, 10189

United States, Michigan

[Not yet recruiting]

Ann Arbor, Michigan, United States, 48105

United States, New Jersey

[Recruiting]

Toms River, New Jersey, United States, 08755

United States, New York

[Not yet recruiting]

Great Neck, New York, United States, 11021

[Recruiting]

New York, New York, United States, 10032

United States, North Carolina

[Recruiting]

Charlotte, North Carolina, United States, 28210

United States, Oklahoma

[Not yet recruiting]

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

[Recruiting]

Portland, Oregon, United States, 97210

United States, Pennsylvania

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Houston, Texas, United States, 77030

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Wisconsin

[Not yet recruiting]

Madison, Wisconsin, United States, 53705

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of VEGF Trap in Patients With Wet AMD

Latest version (submitted January 27, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	April 28, 2006	None (earliest Version on record)
2	<input checked="" type="radio"/>	<input checked="" type="radio"/>	August 1, 2006	Contacts/Locations and Study Status
3	<input type="radio"/>	<input type="radio"/>	October 3, 2006	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input type="radio"/>	<input type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: August 1, 2006 (v2)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of VEGF Trap in Patients With Wet AMD

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: August 2006

Overall Status: Recruiting

Study Start: April 2006

Primary Completion:

Study Completion:

First Submitted: April 28, 2006

First Submitted that: April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that: August 1, 2006

Met QC Criteria:

Last Update Posted: August 3, 2006 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring:

Study Description

Brief Summary: The effect of intravitreally administered VEGF Trap in patients with wet AMD.

To assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of ITV injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms:

Masking: Double (masked roles unspecified)

Allocation: Randomized

Enrollment: 150

Arms and Interventions

Intervention Details:

Drug: VEGF Trap

Outcome Measures

Primary Outcome Measures:

1. Safety, biological effect (OCT, fluorescein angiography, visual acuity)

Secondary Outcome Measures:

2. Pharmacokinetics, immunogenicity, quality of life

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness $\geq 300 \mu\text{m}$ as measured by OCT.
- ETDRS best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - PDT within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).
 - Juxta-scleral steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
 - Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.

- Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Central Contact: Regeneron

Email: VEGF.Trap@regeneron.com

Locations: **United States, Arizona**

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tuscon, Arizona, United States, 85704

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Not yet recruiting]

Irvine, California, United States, 92697

[Not yet recruiting]

Loma Linda, California, United States, 92354

[Not yet recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Palm Springs, California, United States, 92262

United States, Florida

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Recruiting]

Lakeland, Florida, United States, 33805

[Recruiting]

Oakland Park, Florida, United States, 33334

[Not yet recruiting]

Orlando, Florida, United States, 32803

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

United States, Illinois

[Not yet recruiting]

Chicago, Illinois, United States, 60637

[Not yet recruiting]

Glenview, Illinois, United States, 60025

United States, Indiana

[Recruiting]

Indianapolis, Indiana, United States, 46280

United States, Maryland

[Not yet recruiting]

Baltimore, Maryland, United States, 21204

[Recruiting]

Baltimore, Maryland, United States, 21287

United States, Massachusetts

[Recruiting]

Boston, Massachusetts, United States, 02114

[Recruiting]

Peabody, Massachusetts, United States

[Recruiting]

West Springfield, Massachusetts, United States, 10189

United States, Michigan

[Not yet recruiting]

Ann Arbor, Michigan, United States, 48105

United States, New Jersey

[Recruiting]

Toms River, New Jersey, United States, 08755

United States, New York

[Not yet recruiting]

Great Neck, New York, United States, 11021

[Recruiting]

New York, New York, United States, 10032

United States, North Carolina

[Recruiting]

Charlotte, North Carolina, United States, 28210

United States, Oklahoma

[Not yet recruiting]

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

[Recruiting]

Portland, Oregon, United States, 97210

United States, Pennsylvania

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Houston, Texas, United States, 77030

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Wisconsin

[Not yet recruiting]

Madison, Wisconsin, United States, 53705

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00320788

Safety and Efficacy of Repeated Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Wet Age-Related Macular Degeneration (AMD)

[Latest version \(submitted January 27, 2012\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 1, 2006</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>October 3, 2006</u>	Conditions, Study Description, Contacts/Locations, Eligibility, Outcome Measures, Study Status and Study Identification
4	<input type="radio"/>	<input type="radio"/>	<u>July 24, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>January 23, 2009</u>	Recruitment Status, Study Status, Arms and Interventions, Study Design, Contacts/Locations, Outcome Measures, Oversight, Sponsor/Collaborators and Study Identification
6	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Study Status
7	<input type="radio"/>	<input type="radio"/>	<u>November 30, 2010</u>	Study Status
8	<input type="radio"/>	<input type="radio"/>	<u>April 20, 2011</u>	Study Status
9	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Sponsor/Collaborators, Study Status
10	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 27, 2012</u>	Arms and Interventions, Study Status, Outcome Measures, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, References, Contacts/Locations, Eligibility, Study Description and Study Identification

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320788

Submitted Date: January 27, 2012 (v10)

Study Identification

Unique Protocol ID: VGFT-OD-0508

Brief Title: Safety and Efficacy of Repeated Intravitreal Administration of Vascular Endothelial Growth Factor (VEGF) Trap in Patients With Wet Age-Related Macular Degeneration (AMD)

Official Title: A Randomized, Controlled Study of the Safety, Tolerability and Biological Effect of Repeated Intravitreal Administration of VEGF Trap in Patients With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: January 2012

Overall Status: Completed

Study Start: April 2006

Primary Completion: June 2008 [Actual]

Study Completion: August 2008 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Results First Submitted: December 16, 2011

Results First Submitted that January 27, 2012

Met QC Criteria:

Results First Posted: March 1, 2012 [Estimate]

Certification/Extension November 30, 2010

First Submitted:

Certification/Extension November 30, 2010

First Submitted that

Met QC Criteria:

Certification/Extension December 2, 2010 [Estimate]

First Posted:

Last Update Submitted that January 27, 2012

Met QC Criteria:

Last Update Posted: March 1, 2012 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party: Sponsor

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: This study examines the effect of intravitreally administered VEGF Trap in patients with wet AMD.

The purpose of this trial is to assess the ocular and systemic safety and tolerability of repeated intravitreal doses of VEGF Trap in patients with subfoveal choroidal neovascularization (CNV) due to AMD.

Detailed Description: This is a double masked, prospective, randomized study in which five groups of approximately 30 patients meeting the eligibility criteria will be randomly assigned in a balanced ratio to receive a series of intravitreal (IVT) injections of VEGF Trap into the study eye at 4- or 12 -week intervals over a 12-week period.

After Week 12, patients will be evaluated every 4 weeks. Patients will remain on study or may be eligible to enter a long-term extension study, in which they will continue to receive VEGF Trap.

Conditions

Conditions: Macular Degeneration

Keywords: Neovascular Age-Related Macular Degeneration

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 2

Interventional Study Model: Parallel Assignment

Number of Arms: 5

Masking: Triple (Participant, Care Provider, Investigator)

Allocation: Randomized

Enrollment: 159 [Actual]

Arms and Interventions

Arms	Assigned Interventions
Experimental: aflibercept injection (VEGF Trap-Eye, BAY86-5321) 0.5mg q4	Biological: aflibercept injection (VEGF Trap-Eye, BAY86-5321) Participants received 0.5 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 4 week intervals through Week 12 Other Names: <ul style="list-style-type: none">• VEGF Trap-Eye• BAY86-5321
Experimental: aflibercept injection (VEGF Trap-Eye, BAY86-5321) 0.5mg q12	Biological: aflibercept injection (VEGF Trap-Eye, BAY86-5321) Participants received 0.5 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 12 week intervals through Week 12. Other Names: <ul style="list-style-type: none">• VEGF Trap-Eye• BAY86-5321

Arms	Assigned Interventions
<p>Experimental: aflibercept injection (VEGF Trap-Eye, BAY86-5321) 2.0mg q4</p>	<p>Biological: aflibercept injection (VEGF Trap-Eye, BAY86-5321) Participants received 2.0 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 4 week intervals through Week 12</p> <p>Other Names:</p> <ul style="list-style-type: none"> • VEGF Trap-Eye • BAY86-5321
<p>Experimental: aflibercept injection (VEGF Trap-Eye, BAY86-5321) 2.0mg q12</p>	<p>Biological: aflibercept injection (VEGF Trap-Eye, BAY86-5321) Participants received 2.0 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 12 week intervals through Week 12.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • VEGF Trap-Eye • BAY86-5321
<p>Experimental: aflibercept injection (VEGF Trap-Eye, BAY86-5321) 4.0mg q12</p>	<p>Biological: aflibercept injection (VEGF Trap-Eye, BAY86-5321) Participants received 4.0 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 12 week intervals through Week 12.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • VEGF Trap-Eye • BAY86-5321

Outcome Measures

[See Results Section.]

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Subfoveal CNV secondary to AMD.
- Central retinal (including lesion) thickness ≥ 300 μm as measured by Optical Coherence Tomography (OCT).
- Early Treatment of Diabetic Retinopathy Study (ETDRS) best-corrected visual acuity of 73 letters to 34 letters.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Day 1.
- Aphakia.
- Significant subfoveal atrophy or scarring.
- Prior treatment with the following in the study eye:
 - Subfoveal thermal laser therapy.
 - Submacular surgery or other surgical intervention for the treatment of AMD.
 - Extrafoveal laser coagulation treatment within 12 weeks prior to Day 1.
 - Photodynamic therapy (PDT) within 12 weeks prior to Visit 2 (Day 1).
 - Pegaptanib sodium (Macugen) within 8 weeks of Visit 2 (Day 1).
 - Juxtasclear steroids or anecortave acetate within 24 weeks (6 months) prior to Visit 2 (Day 1).
 - Intravitreal administration of triamcinolone acetonide or other steroids within 24 weeks prior to Visit 2 (Day 1), unless no visible residue of drug substance can be seen in the vitreous cavity using indirect ophthalmoscopy.
 - Prior systemic or intravitreal treatment with VEGF Trap, ranibizumab (Lucentis) or bevacizumab (Avastin).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase II study.

Contacts/Locations

Study Officials: Clinical Trial Management
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Arizona**

Associated Retina Consultants
Phoenix, Arizona, United States, 85020

Retina Centers, PC
Tucson, Arizona, United States, 85704

United States, California

Retina Vitreous Associates Medical Group
Beverly Hills, California, United States, 90211

Loma Linda University Health Care
Loma Linda, California, United States, 92354

United States, Georgia

Southeast Retina Center
Augusta, Georgia, United States, 30909

United States, Illinois

University of Chicago
Chicago, Illinois, United States, 60637

United States, Indiana

Midwest Eye Institute
Indianapolis, Indiana, United States, 46280

United States, Maryland

Johns Hopkins Hospital School of Medicine
Baltimore, Maryland, United States, 21287

United States, Massachusetts

Ophthalmic Consultants of Boston
Boston, Massachusetts, United States, 02114

New England Retina Consultants PC
West Springfield, Massachusetts, United States, 10189

United States, North Carolina

Charlotte Eye, Ear, Nose & Throat Associates
Charlotte, North Carolina, United States, 28210

United States, Oklahoma

Dean A. McGee Eye Institute
Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Retina Northwest PC
Portland, Oregon, United States, 97210

United States, Pennsylvania

Retina Diagnostic and Treatment Assoc., LLC
Philadelphia, Pennsylvania, United States, 19107

United States, South Dakota

Black Hills Regional Eye Institute
Rapid City, South Dakota, United States, 57701

United States, Tennessee

Retina-Vitreous Associates, P.C.
Nashville, Tennessee, United States, 37203

United States, Texas

Vitreoretinal Consultants Scurlock Tower Texas Medical Center
Houston, Texas, United States, 77030

Medical Center Ophthalmology
San Antonio, Texas, United States, 78240

IPDSharing

Plan to Share IPD:

References

Citations:

Links: URL: <http://www.ncbi.nlm.nih.gov/pubmed/21640257>

Description: Primary endpoint results of a phase II study of vascular endothelial growth factor trap-eye in wet age-related macular degeneration.

URL: <http://www.ncbi.nlm.nih.gov/pubmed/21640258>

Description: The 1-year results of CLEAR-IT 2, a phase 2 study of vascular endothelial growth factor trap-eye dosed as-needed after 12-week fixed dosing.

Available IPD/Information:

Study Results

Participant Flow

Recruitment Details	The study was conducted at 29 study sites in the United States. Recruitment period: May 2006 to April 2007.
Pre-assignment Details	A total of 301 participants were screened; 159 participants were randomized; 157 participants were included in both the Safety Analysis Set (SAF) and the Full Analysis Set (FAS) as all received study treatment, had baseline assessments and at least 1 post-baseline assessment.

Reporting Groups

	Description
Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 0.5mg q4	Participants received 0.5 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 4 week intervals through Week 12.
Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 0.5mg q12	Participants received 0.5 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 12 week intervals through Week 12.

Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 2.0mg q4	Participants received 2.0 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 4 week intervals through Week 12.
Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 2.0mg q12	Participants received 2.0 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 12 week intervals through Week 12.
Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 4.0mg q12	Participants received 4.0 mg of aflibercept injection (VEGF Trap-Eye, BAY86-5321) at 12 week intervals through Week 12.

Overall Study

	Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 0.5mg q4	Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 0.5mg q12	Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 2.0mg q4	Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 2.0mg q12	Aflibercept Injection (VEGF Trap-Eye, BAY86-5321) 4.0mg q12
Started	32	32	32	32	31
Participants Received Treatment (SAF)	32	32	31	31	31
Completed	26	26	29	27	26
Not Completed	6	6	3	5	5
Adverse Event	0	0	0	1	0
Death	0	0	1	0	1
Lost to Follow-up	0	2	1	0	0
Decision by the Sponsor	1	1	0	0	1
Other	1	2	1	1	1
Physician Decision	1	1	0	0	0
Protocol Violation	0	0	0	0	1
Withdrawal by Subject	3	0	0	3	1

☐ Participants Randomized

☐ One participant in this group did not receive treatment and was not included in the SAF and FAS

Baseline Characteristics

Reporting Groups

	Description
Aflibercept Injection 0.5mg q4	Participants received 0.5 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 0.5mg q12	Participants received 0.5 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 2.0mg q4	Participants received 2.0 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 2.0mg q12	Participants received 2.0 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 4.0mg q12	Participants received 4.0 mg of aflibercept injection at 12 week intervals through Week 12.

Baseline Measures

		Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
Overall Number of Participants		32	32	31	31	31	157
Age Continuous Mean (Standard Deviation) Unit of measure: years	Number Analyzed	32 Participants	32 Participants	31 Participants	31 Participants	31 Participants	157 Participants
		79.6 (0 to 0)	78.5 (0 to 0)	74.9 (0 to 0)	79.6 (0 to 0)	78.3 (0 to 0)	78.2 (0 to 0)
Sex: Female, Male Measure type: Count of Participants Unit of measure: Participants	Number Analyzed	32 Participants	32 Participants	31 Participants	31 Participants	31 Participants	157 Participants
	Female	17 53.12% (0 to 0)	25 78.12% (0 to 0)	20 64.52% (0 to 0)	16 51.61% (0 to 0)	20 64.52% (0 to 0)	98 62.42%
	Male	15 46.88% (0 to 0)	7 21.88% (0 to 0)	11 35.48% (0 to 0)	15 48.39% (0 to 0)	11 35.48% (0 to 0)	59 37.58%

		Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
Ethnicity (NIH/OMB) Measure type: Count of Participants Unit of measure: Participants	Number Analyzed	32 Participants	32 Participants	31 Participants	31 Participants	31 Participants	157 Participants
	Hispanic or Latino	0 0% (0 to 0)	1 3.12% (0 to 0)	2 6.45% (0 to 0)	1 3.23% (0 to 0)	0 0% (0 to 0)	4 2.55%
	Not Hispanic or Latino	32 100% (0 to 0)	31 96.88% (0 to 0)	29 93.55% (0 to 0)	30 96.77% (0 to 0)	31 100% (0 to 0)	153 97.45%
	Unknown or Not Reported	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0%
Race (NIH/OMB) Measure type: Count of Participants Unit of measure: Participants	Number Analyzed	32 Participants	32 Participants	31 Participants	31 Participants	31 Participants	157 Participants
	American Indian or Alaska Native	0 0% (0 to 0)	0 0% (0 to 0)	1 3.23% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	1 0.64%
	Asian	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0%
	Native Hawaiian or Other Pacific Islander	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0%
	Black or African American	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0%
	White	32 100% (0 to 0)	32 100% (0 to 0)	30 96.77% (0 to 0)	31 100% (0 to 0)	31 100% (0 to 0)	156 99.36%

		Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
	More than one race	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0%
	Unknown or Not Reported	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0% (0 to 0)	0 0%
Central Retinal/Lesion Thickness (CR/LT) Mean (Standard Deviation) Unit of measure: μm	Number Analyzed	32 Participants	32 Participants	31 Participants	31 Participants	31 Participants	157 Participants
		442.2 (0 to 0)	442.6 (0 to 0)	453.3 (0 to 0)	447.0 (0 to 0)	497.5 (0 to 0)	456.4 (0 to 0)
Best Corrected Visual Acuity (BCVA) ^[1] Mean (Standard Deviation) Unit of measure: letters read	Number Analyzed	32 Participants	32 Participants	31 Participants	31 Participants	31 Participants	157 Participants
		54.1 (0 to 0)	55.6 (0 to 0)	57.9 (0 to 0)	57.2 (0 to 0)	53.0 (0 to 0)	55.5 (0 to 0)
		^[1] Measure Description: BCVA as Measured by 4 Meter Early Treatment Diabetic Retinopathy Study (ETDRS) eye charts/measures					

Outcome Measures

1. Primary Outcome Measure:

Measure Title	Mean Change of CR/LT From Baseline at Week 12
Measure Description	CR/LT measured in micrometers (μm); lower individual values represent better outcomes.
Time Frame	Baseline and at Week 12

Analysis Population Description

Full Analysis Set (FAS) used for analysis, Last Observation Carried Forward (LOCF)

Reporting Groups

	Description
Aflibercept Injection 0.5mg q4	Participants received 0.5 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 0.5mg q12	Participants received 0.5 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 2.0mg q4	Participants received 2.0 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 2.0mg q12	Participants received 2.0 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 4.0mg q12	Participants received 4.0 mg of aflibercept injection at 12 week intervals through Week 12.
Total	

Measured Values

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
Overall Number of Participants Analyzed	32	32	31	31	31	157
Mean Change of CR/LT From Baseline at Week 12 Measure Type: Mean (Standard Deviation) Unit of Measure: μm	-153.5 (0 to 0)	-75.6 (0 to 0)	-169.2 (0 to 0)	-56.3 (0 to 0)	-139.8 (0 to 0)	-118.8 (0 to 0)

2. Secondary Outcome Measure:

Measure Title	Mean Percent Change of CR/LT From Baseline at Week 12
Measure Description	CR/LT measured in micrometers (μm); a more negative percentage represents a better outcome
Time Frame	Baseline and at Week 12

Analysis Population Description

FAS used for analysis, LOCF

Reporting Groups

	Description

Aflibercept Injection 0.5mg q4	Participants received 0.5mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 0.5mg q12	Participants received 0.5mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 2.0mg q4	Participants received 2.0mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 2.0mg q12	Participants received 2.0mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 4.0mg q12	Participants received 4.0mg of aflibercept injection at 12 week intervals through Week 12.
Total	

Measured Values

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
Overall Number of Participants Analyzed	32	32	31	31	31	157
Mean Percent Change of CR/LT From Baseline at Week 12 Measure Type: Mean (Standard Deviation) Unit of Measure: percent change	-32.4 (0 to 0)	-15.2 (0 to 0)	-33.2 (0 to 0)	-10.3 (0 to 0)	-21.1 (0 to 0)	-22.5 (0 to 0)

3. Secondary Outcome Measure:

Measure Title	Mean Change in Best Corrected Visual Acuity (BCVA) as Measured by Early Treatment Diabetic Retinopathy Study (ETDRS) From Baseline at Week 12
Measure Description	Defined study baseline range of ETDRS Best Corrected Visual Acuity of: letter score of 73 to 25 (20/40 to 20/320) in the study eye; a higher score represents better functioning
Time Frame	Baseline and at week 12

Analysis Population Description

FAS used for analysis, LOCF

Reporting Groups

	Description
Aflibercept Injection 0.5mg q4	Participants received 0.5 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 0.5mg q12	Participants received 0.5 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 2.0mg q4	Participants received 2.0 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 2.0mg q12	Participants received 2.0 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 4.0mg q12	Participants received 4.0 mg of aflibercept injection at 12 week intervals through Week 12.
Total	

Measured Values

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
Overall Number of Participants Analyzed	32	32	31	31	31	157
Mean Change in Best Corrected Visual Acuity (BCVA) as Measured by Early Treatment Diabetic Retinopathy Study (ETDRS) From Baseline at Week 12 Measure Type: Mean (Standard Deviation) Unit of Measure: letters read	8.8 (0 to 0)	3.8 (0 to 0)	8.3 (0 to 0)	5.2 (0 to 0)	2.6 (0 to 0)	5.7 (0 to 0)

4. Secondary Outcome Measure:

Measure Title	Percentage of Participants Who Gained at Least 15 Letters of Vision in the ETDRS Letter Score From Baseline at Week 12
Measure Description	Defined study baseline range of ETDRS Best Corrected Visual Acuity of: letter score of 73 to 25 (20/40 to 20/320) in the study eye; a higher score represents better functioning
Time Frame	At Week 12

Analysis Population Description

FAS used for analysis, LOCF

Reporting Groups

	Description
Aflibercept Injection 0.5mg q4	Participants received 0.5 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 0.5mg q12	Participants received 0.5 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 2.0mg q4	Participants received 2.0 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 2.0mg q12	Participants received 2.0 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 4.0mg q12	Participants received 4.0 mg of aflibercept injection at 12 week intervals through Week 12.
Total	

Measured Values

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
Overall Number of Participants Analyzed	32	32	31	31	31	157
Percentage of Participants Who Gained at Least 15 Letters of Vision in the ETDRS Letter Score From Baseline at Week 12 Measure Type: Number Unit of Measure: percentage of participants	18.8 (0 to 0)	21.9 (0 to 0)	25.8 (0 to 0)	16.1 (0 to 0)	9.7 (0 to 0)	18.5 (0 to 0)

5. Post-Hoc Outcome Measure:

Measure Title	Mean Change of CR/LT From Baseline at Week 16
Measure Description	CR/LT measured in micrometers (µm); lower individual values represent better outcomes
Time Frame	Baseline and at Week 16

Analysis Population Description

FAS used for analysis, LOCF

Reporting Groups

	Description
Aflibercept Injection 0.5mg q4	Participants received 0.5 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 0.5mg q12	Participants received 0.5 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 2.0mg q4	Participants received 2.0mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 2.0mg q12	Participants received 2.0mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 4.0mg q12	Participants received 4.0mg of aflibercept injection at 12 week intervals through Week 12.
Total	

Measured Values

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
Overall Number of Participants Analyzed	32	32	31	31	31	157
Mean Change of CR/LT From Baseline at Week 16 Measure Type: Mean (Standard Deviation) Unit of Measure: μm	-163.3 (0 to 0)	-139.6 (0 to 0)	-182.7 (0 to 0)	-107.4 (0 to 0)	-208.6 (0 to 0)	-160.2 (0 to 0)

6. Post-Hoc Outcome Measure:

Measure Title	Mean Change in BCVA as Measured by ETDRS From Baseline at Week 16
Measure Description	Defined study baseline range of ETDRS Best Corrected Visual Acuity of: letter score of 73 to 25 (20/40 to 20/320) in the study eye; a higher score represents better functioning
Time Frame	Baseline and at Week 16

Analysis Population Description

FAS used for analysis, LOCF

Reporting Groups

	Description
Aflibercept Injection 0.5mg q4	Participants received 0.5 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 0.5mg q12	Participants received 0.5 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 2.0mg q4	Participants received 2.0 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 2.0mg q12	Participants received 2.0 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 4.0mg q12	Participants received 4.0 mg of aflibercept injection at 12 week intervals through Week 12.
Total	

Measured Values

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12	Total
Overall Number of Participants Analyzed	32	32	31	31	31	157
Mean Change in BCVA as Measured by ETDRS From Baseline at Week 16 Measure Type: Mean (Standard Deviation) Unit of Measure: letters read	9.3 (0 to 0)	5.6 (0 to 0)	10.0 (0 to 0)	4.3 (0 to 0)	3.9 (0 to 0)	6.6 (0 to 0)

Reported Adverse Events

Time Frame	Adverse events (AEs) considered related to study treatment were followed until resolution or until the event was considered chronic or stable.
Adverse Event Reporting Description	Safety was assessed through reported AEs, clinical laboratory test results, vital signs, and ophthalmic examinations

Reporting Groups

	Description
Aflibercept Injection 0.5mg q4	Participants received 0.5 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 0.5mg q12	Participants received 0.5 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 2.0mg q4	Participants received 2.0 mg of aflibercept injection at 4 week intervals through Week 12.
Aflibercept Injection 2.0mg q12	Participants received 2.0 mg of aflibercept injection at 12 week intervals through Week 12.
Aflibercept Injection 4.0mg q12	Participants received 4.0 mg of aflibercept injection at 12 week intervals through Week 12.

All-Cause Mortality

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)
Total	/	/	/	/	/

Serious Adverse Events

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)
Total	11/	5/	10/	7/	2/

Cardiac disorders

ANGINA PECTORIS ^{At}	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
ATRIAL FIBRILLATION ^{At}	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
ATRIOVENTRICULAR BLOCK COMPLETE ^{At}	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	1/31 (3.23%)
BRADYCARDIA ^{At}	0/32 (0%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	1/31 (3.23%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
CARDIAC FAILURE CONGESTIVE At	1/32 (3.12%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)
CORONARY ARTERY DISEASE At	0/32 (0%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)
Eye disorders					
RETINAL DETACHMENT A [1]†	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
UVEITIS A [2]†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
VISUAL ACUITY REDUCED A [1]†	0/32 (0%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
Gastrointestinal disorders					
DYSKINESIA OESOPHAGEAL At	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
INTESTINAL OBSTRUCTION At	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
SMALL INTESTINAL OBSTRUCTION At	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
VOLVULUS At	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
General disorders					
ASTHENIA At	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
NON-CARDIAC CHEST PAIN At	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
Infections and infestations					
BRONCHITIS At	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	1/31 (3.23%)	0/31 (0%)
CELLULITIS At	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
GASTROENTERITIS At	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
PNEUMONIA At	2/32 (6.25%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)
URINARY TRACT INFECTION At	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	1/31 (3.23%)	0/31 (0%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
Injury, poisoning and procedural complications					
FALL A†	1/32 (3.12%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
HIP FRACTURE A†	0/32 (0%)	1/32 (3.12%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
SKIN LACERATION A†	0/32 (0%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	0/31 (0%)
SYNOVIAL RUPTURE A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
Investigations					
INTRAOCULAR PRESSURE INCREASED A [1]†	0/32 (0%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	1/31 (3.23%)
Musculoskeletal and connective tissue disorders					
BACK PAIN A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
OSTEOARTHRITIS A†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
SPINAL OSTEOARTHRITIS A†	0/32 (0%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	0/31 (0%)
Neoplasms benign, malignant and unspecified (incl cysts and polyps)					
BRAIN NEOPLASM MALIGNANT A†	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
COLON CANCER A†	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
METASTASES TO LUNG A†	0/32 (0%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	0/31 (0%)
NON-HODGKIN'S LYMPHOMA A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
PANCREATIC CARCINOMA A†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
SQUAMOUS CELL CARCINOMA A†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
THYROID NEOPLASM A†	0/32 (0%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	0/31 (0%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
TRANSITIONAL CELL CARCINOMA A†	0/32 (0%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	0/31 (0%)
Nervous system disorders					
CAROTID ARTERY OCCLUSION A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
CEREBROVASCULAR ACCIDENT A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
COORDINATION ABNORMAL A†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
DYSARTHRIA A†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
SPINAL CORD DISORDER A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
TRANSIENT ISCHAEMIC ATTACK A†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
Renal and urinary disorders					
RENAL FAILURE A†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
STRESS INCONTINENCE A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
URINARY RETENTION A†	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
Respiratory, thoracic and mediastinal disorders					
CHRONIC OBSTRUCTIVE PULMONARY DISEASE A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
PNEUMONIA ASPIRATION A†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
PULMONARY EMBOLISM A†	0/32 (0%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	0/31 (0%)
PULMONARY HYPERTENSION A†	0/32 (0%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	1/31 (3.23%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
Surgical and medical procedures					
PILONIDAL SINUS REPAIR A†	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
Vascular disorders					
DEEP VEIN THROMBOSIS A†	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	0/31 (0%)

† Indicates events were collected by systematic assessment.

A Term from vocabulary, MedDRA 10.0

[1] Fellow Eye

[2] Study Eye

Other Adverse Events

Frequency Threshold Above Which Other Adverse Events are Reported: 5%

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)	Affected/At Risk (%)
Total	26/	25/	27/	22/	22/
Blood and lymphatic system disorders					
ANAEMIA A†	0/32 (0%)	1/32 (3.12%)	3/31 (9.68%)	0/31 (0%)	0/31 (0%)
Cardiac disorders					
ANGINA PECTORIS A†	2/32 (6.25%)	0/32 (0%)	1/31 (3.23%)	1/31 (3.23%)	0/31 (0%)
BRADYCARDIA A†	1/32 (3.12%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)
Eye disorders					
BLEPHARITIS A [1]†	2/32 (6.25%)	0/32 (0%)	2/31 (6.45%)	3/31 (9.68%)	1/31 (3.23%)
CATARACT A [1]†	0/32 (0%)	1/32 (3.12%)	1/31 (3.23%)	1/31 (3.23%)	2/31 (6.45%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
CATARACT NUCLEAR ^A [1]†	3/32 (9.38%)	0/32 (0%)	2/31 (6.45%)	1/31 (3.23%)	0/31 (0%)
CATARACT SUBCAPSULAR ^A [1]†	0/32 (0%)	2/32 (6.25%)	0/31 (0%)	1/31 (3.23%)	2/31 (6.45%)
CHOROIDAL NEOVASCULARISATION ^A [1]†	0/32 (0%)	1/32 (3.12%)	2/31 (6.45%)	1/31 (3.23%)	0/31 (0%)
CONJUNCTIVAL HAEMORRHAGE ^A [1]†	2/32 (6.25%)	0/32 (0%)	2/31 (6.45%)	2/31 (6.45%)	1/31 (3.23%)
DETACHMENT OF RETINAL PIGMENT EPITHELIUM ^A [1]†	2/32 (6.25%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	1/31 (3.23%)
DRY EYE ^A [1]†	1/32 (3.12%)	0/32 (0%)	1/31 (3.23%)	2/31 (6.45%)	0/31 (0%)
EYE INFLAMMATION ^A [2]†	2/32 (6.25%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	1/31 (3.23%)
EYE IRRITATION ^A [2]†	5/32 (15.62%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	1/31 (3.23%)
EYE PAIN ^A [2]†	5/32 (15.62%)	2/32 (6.25%)	4/31 (12.9%)	1/31 (3.23%)	3/31 (9.68%)
FOREIGN BODY SENSATION IN EYES ^A [2]†	3/32 (9.38%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
LACRIMATION DECREASED ^A [2]†	0/32 (0%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	2/31 (6.45%)
LACRIMATION INCREASED ^A [1]†	0/32 (0%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	2/31 (6.45%)
MACULAR DEGENERATION ^A [2]†	0/32 (0%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	2/31 (6.45%)
MACULOPATHY ^A [2]†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	2/31 (6.45%)	1/31 (3.23%)
PHOTOPSIA ^A [2]†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	2/31 (6.45%)	3/31 (9.68%)
PUNCTATE KERATITIS ^A [2]†	0/32 (0%)	0/32 (0%)	1/31 (3.23%)	2/31 (6.45%)	2/31 (6.45%)
REFRACTION DISORDER ^A [1]†	1/32 (3.12%)	4/32 (12.5%)	1/31 (3.23%)	1/31 (3.23%)	3/31 (9.68%)
RETINAL DEPIGMENTATION ^A [2]†	2/32 (6.25%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
RETINAL HAEMORRHAGE ^A [1]†	0/32 (0%)	2/32 (6.25%)	1/31 (3.23%)	3/31 (9.68%)	0/31 (0%)
RETINAL OEDEMA ^A [1]†	1/32 (3.12%)	2/32 (6.25%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
RETINAL PIGMENT EPITHELIOPATHY ^A [1]†	2/32 (6.25%)	3/32 (9.38%)	0/31 (0%)	1/31 (3.23%)	1/31 (3.23%)
SUBRETINAL FIBROSIS ^A [2]†	2/32 (6.25%)	1/32 (3.12%)	1/31 (3.23%)	2/31 (6.45%)	2/31 (6.45%)
VISION BLURRED ^A [2]†	2/32 (6.25%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	1/31 (3.23%)
VISUAL ACUITY REDUCED ^A [1]†	1/32 (3.12%)	1/32 (3.12%)	4/31 (12.9%)	1/31 (3.23%)	1/31 (3.23%)
VISUAL DISTURBANCE ^A [2]†	2/32 (6.25%)	1/32 (3.12%)	0/31 (0%)	2/31 (6.45%)	3/31 (9.68%)
VITREOUS DETACHMENT ^A [1]†	4/32 (12.5%)	1/32 (3.12%)	2/31 (6.45%)	3/31 (9.68%)	4/31 (12.9%)
VITREOUS FLOATERS ^A [1]†	2/32 (6.25%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	1/31 (3.23%)
Gastrointestinal disorders					
CONSTIPATION ^A †	5/32 (15.62%)	1/32 (3.12%)	0/31 (0%)	0/31 (0%)	1/31 (3.23%)
DIARRHOEA ^A †	1/32 (3.12%)	1/32 (3.12%)	0/31 (0%)	1/31 (3.23%)	2/31 (6.45%)
FLATULENCE ^A †	0/32 (0%)	2/32 (6.25%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
NAUSEA ^A †	3/32 (9.38%)	0/32 (0%)	0/31 (0%)	1/31 (3.23%)	1/31 (3.23%)
General disorders					
CHEST PAIN ^A †	2/32 (6.25%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
Immune system disorders					
SEASONAL ALLERGY ^A †	4/32 (12.5%)	2/32 (6.25%)	0/31 (0%)	0/31 (0%)	1/31 (3.23%)
Infections and infestations					
BRONCHITIS ^A †	5/32 (15.62%)	2/32 (6.25%)	1/31 (3.23%)	1/31 (3.23%)	3/31 (9.68%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
CELLULITIS ^{At}	0/32 (0%)	0/32 (0%)	0/31 (0%)	2/31 (6.45%)	1/31 (3.23%)
CYSTITIS ^{At}	2/32 (6.25%)	0/32 (0%)	1/31 (3.23%)	1/31 (3.23%)	0/31 (0%)
GASTROENTERITIS VIRAL ^{At}	0/32 (0%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	1/31 (3.23%)
HERPES ZOSTER ^{At}	0/32 (0%)	2/32 (6.25%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
INFLUENZA ^{At}	0/32 (0%)	2/32 (6.25%)	2/31 (6.45%)	0/31 (0%)	2/31 (6.45%)
NASOPHARYNGITIS ^{At}	3/32 (9.38%)	1/32 (3.12%)	3/31 (9.68%)	1/31 (3.23%)	2/31 (6.45%)
PNEUMONIA ^{At}	3/32 (9.38%)	0/32 (0%)	2/31 (6.45%)	1/31 (3.23%)	1/31 (3.23%)
RHINOVIRUS INFECTION ^{At}	0/32 (0%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)
SINUSITIS ^{At}	2/32 (6.25%)	1/32 (3.12%)	2/31 (6.45%)	1/31 (3.23%)	3/31 (9.68%)
TOOTH ABSCESS ^{At}	1/32 (3.12%)	1/32 (3.12%)	1/31 (3.23%)	2/31 (6.45%)	0/31 (0%)
UPPER RESPIRATORY TRACT INFECTION ^{At}	3/32 (9.38%)	1/32 (3.12%)	2/31 (6.45%)	6/31 (19.35%)	3/31 (9.68%)
URINARY TRACT INFECTION ^{At}	1/32 (3.12%)	3/32 (9.38%)	6/31 (19.35%)	1/31 (3.23%)	2/31 (6.45%)
VULVOVAGINAL MYCOTIC INFECTION ^{At}	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	2/31 (6.45%)	1/31 (3.23%)
Injury, poisoning and procedural complications					
FALL ^{At}	2/32 (6.25%)	1/32 (3.12%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
JOINT SPRAIN ^{At}	0/32 (0%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	2/31 (6.45%)
Investigations					
BLOOD CREATININE INCREASED ^{At}	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	2/31 (6.45%)	1/31 (3.23%)
BLOOD GLUCOSE INCREASED ^{At}	0/32 (0%)	2/32 (6.25%)	1/31 (3.23%)	1/31 (3.23%)	0/31 (0%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
BLOOD UREA INCREASED ^{A†}	0/32 (0%)	1/32 (3.12%)	0/31 (0%)	2/31 (6.45%)	0/31 (0%)
INTRAOCULAR PRESSURE INCREASED ^{A†††}	1/32 (3.12%)	3/32 (9.38%)	3/31 (9.68%)	2/31 (6.45%)	1/31 (3.23%)
Metabolism and nutrition disorders					
DEHYDRATION ^{A†}	1/32 (3.12%)	1/32 (3.12%)	2/31 (6.45%)	0/31 (0%)	1/31 (3.23%)
GOUT ^{A†}	1/32 (3.12%)	2/32 (6.25%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
HYPERCHOLESTEROLAEMIA ^{A†}	4/32 (12.5%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
Musculoskeletal and connective tissue disorders					
BACK PAIN ^{A†}	1/32 (3.12%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)
OSTEOARTHRITIS ^{A†}	0/32 (0%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	1/31 (3.23%)
Nervous system disorders					
DEMENTIA ALZHEIMER'S TYPE ^{A†}	0/32 (0%)	2/32 (6.25%)	0/31 (0%)	0/31 (0%)	0/31 (0%)
DIZZINESS ^{A†}	1/32 (3.12%)	0/32 (0%)	2/31 (6.45%)	1/31 (3.23%)	0/31 (0%)
HEADACHE ^{A†}	1/32 (3.12%)	3/32 (9.38%)	1/31 (3.23%)	0/31 (0%)	1/31 (3.23%)
SINUS HEADACHE ^{A†}	1/32 (3.12%)	2/32 (6.25%)	2/31 (6.45%)	0/31 (0%)	2/31 (6.45%)
Psychiatric disorders					
ANXIETY ^{A†}	0/32 (0%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)
DEPRESSION ^{A†}	0/32 (0%)	1/32 (3.12%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)
INSOMNIA ^{A†}	2/32 (6.25%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	0/31 (0%)
Respiratory, thoracic and mediastinal disorders					
COUGH ^{A†}	2/32 (6.25%)	1/32 (3.12%)	2/31 (6.45%)	1/31 (3.23%)	2/31 (6.45%)

	Aflibercept Injection 0.5mg q4	Aflibercept Injection 0.5mg q12	Aflibercept Injection 2.0mg q4	Aflibercept Injection 2.0mg q12	Aflibercept Injection 4.0mg q12
DYSPNOEA A†	2/32 (6.25%)	0/32 (0%)	1/31 (3.23%)	2/31 (6.45%)	0/31 (0%)
PHARYNGOLARYNGEAL PAIN A†	1/32 (3.12%)	0/32 (0%)	0/31 (0%)	0/31 (0%)	2/31 (6.45%)
Skin and subcutaneous tissue disorders					
RASH A†	2/32 (6.25%)	0/32 (0%)	1/31 (3.23%)	0/31 (0%)	1/31 (3.23%)
Vascular disorders					
HYPERTENSION A†	6/32 (18.75%)	1/32 (3.12%)	2/31 (6.45%)	0/31 (0%)	3/31 (9.68%)
HYPOTENSION A†	0/32 (0%)	0/32 (0%)	2/31 (6.45%)	0/31 (0%)	0/31 (0%)

† Indicates events were collected by systematic assessment.

A Term from vocabulary, MedDRA 10.0

[1] Fellow Eye

[2] Study Eye

Limitations and Caveats

This is a phase 2 study with small numbers of patients per group limiting the conclusions that can be drawn from the resulting data.

More Information

Certain Agreements:

Principal Investigators are NOT employed by the organization sponsoring the study.

There IS an agreement between the Principal Investigator and the Sponsor (or its agents) that restricts the PI's rights to discuss or publish trial results after the trial is completed.

Results Point of Contact:

Name/Official Title: Clinical Trials Administrator

Organization: Regeneron Pharmaceuticals, Inc.

Phone:

Email: clinicaltrials@regeneron.com

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320814

Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Latest version (submitted June 8, 2011) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>September 6, 2006</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2009</u>	Recruitment Status, Study Status, Outcome Measures, Arms and Interventions, Oversight, Contacts/Locations, Study Design and Sponsor/Collaborators

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
5	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>June 8, 2011</u>	Study Status

Compare

Comparison Format: Merged
 Side-by-Side

Scroll up to access the controls

Study NCT00320814

Submitted Date: June 8, 2011 (v5)

Study Identification

Unique Protocol ID: VGFT-OD-0512

Brief Title: Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of a Single Intravitreal Administration of VEGF Trap in Patients With Diabetic Macular Edema

Secondary IDs:

Study Status

Record Verification: June 2011

Overall Status: Completed

Study Start: April 2006

Primary Completion: August 2006 [Actual]

Study Completion: August 2007 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that June 8, 2011

Met QC Criteria:

Last Update Posted: June 10, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: To assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with diabetic macular edema.

Detailed Description: This is an open label study. Initially, 5 patients with DME will receive an ITV injection of VEGF Trap into the study eye. Additional patients may be enrolled at the same or additional dose levels. Patients will be observed for six weeks following the injection for assessments of ocular and systemic safety.

Conditions

Conditions: Diabetic Macular Edema

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Single Group Assignment

Number of Arms: 1

Masking: None (Open Label)

Allocation: Non-Randomized

Enrollment: 5 [Actual]

Arms and Interventions

Arms	Assigned Interventions
Experimental: VEGF Trap-Eye single IVT injection of 4.0 mg of VEGF Trap-Eye into the study eye on Day 1	Drug: VEGF Trap-Eye single IVT injection of 4.0 mg of VEGF Trap-Eye into the study eye on Day 1

Outcome Measures

Primary Outcome Measures:

1. To assess the ocular and systemic safety and tolerability of a single intravitreal (IVT) injection of VEGF Trap-Eye in patients with diabetic macular edema (DME)
Assessments for safety and tolerability are performed at each visit (Visit 1 - Visit 10)

Secondary Outcome Measures:

2. To obtain a preliminary assessment of the effect of a single dose of VEGF Trap-Eye on central retinal thickness (CRT) at the center point as determined by optical coherence tomography (OCT)
Assessments for CRT are performed at each visit (Visit 1 - Visit 10) by means of OCT.
3. To obtain a preliminary assessment of the effect of a single IVT administration of VEGF Trap-Eye on visual acuity
Assessments for visual acuity are performed at each visit (Visit 1 - Visit 10).

Eligibility

Minimum Age: 18 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Diagnosis of diabetes mellitus (type 1 or type 2).
- Best corrected E-ETDRS visual acuity score of ≥ 24 letters (i.e., 20/320 or better) and ≤ 73 letters (i.e., 20/40 or worse).
- On clinical exam, definite retinal thickening due to diabetic macular edema involving the center of the macula.
- Retinal Thickness at the center point ≥ 250 microns.
- Media clarity, pupillary dilation, and patient cooperation sufficient for adequate fundus photographs.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Visit 2 (Day 1).
- Macular edema due to causes other than diabetic macular edema. An eye should be considered ineligible: (1) if the macular edema is considered to be related to cataract extraction or (2) clinical exam and/or OCT suggests that vitreoretinal interface disease (e.g., a taut posterior hyaloid or epiretinal membrane) is the primary cause of the macular edema.
- An ocular condition is present such that, in the opinion of the investigator, visual acuity would not improve from resolution of macular edema (e.g., foveal atrophy, pigmentary changes, dense subfoveal hard exudates, nonretinal condition).
- An ocular condition is present (other than diabetes) that, in the opinion of the investigator, might affect macular edema or alter visual acuity during the course of the study (e.g., vein occlusion, age-related macular degeneration, uveitis or other ocular inflammatory disease, neovascular glaucoma, Irvine-Gass Syndrome, etc.).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase 1 study.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Maryland**

Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte, North Carolina, United States, 28210

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320814

Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Latest version (submitted June 8, 2011) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>September 6, 2006</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2009</u>	Recruitment Status, Study Status, Outcome Measures, Arms and Interventions, Oversight, Contacts/Locations, Study Design and Sponsor/Collaborators

Version	A	B	Submitted Date	Changes
4	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 25, 2011</u>	Study Status
5	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320814
Submitted Date: January 25, 2011 (v4)

Study Identification

Unique Protocol ID: VGFT-OD-0512

Brief Title: Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of a Single Intravitreal Administration of VEGF Trap in Patients With Diabetic Macular Edema

Secondary IDs:

Study Status

Record Verification: January 2011

Overall Status: Completed

Study Start: April 2006

Primary Completion: August 2006 [Actual]

Study Completion: August 2007 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that January 25, 2011

Met QC Criteria:

Last Update Posted: January 26, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: To assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with diabetic macular edema.

Detailed Description: This is an open label study. Initially, 5 patients with DME will receive an ITV injection of VEGF Trap into the study eye. Additional patients may be enrolled at the same or additional dose levels. Patients will be observed for six weeks following the injection for assessments of ocular and systemic safety.

Conditions

Conditions: Diabetic Macular Edema

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Single Group Assignment

Number of Arms: 1

Masking: None (Open Label)

Allocation: Non-Randomized

Enrollment: 5 [Actual]

Arms and Interventions

Arms	Assigned Interventions
Experimental: VEGF Trap-Eye single IVT injection of 4.0 mg of VEGF Trap-Eye into the study eye on Day 1	Drug: VEGF Trap-Eye single IVT injection of 4.0 mg of VEGF Trap-Eye into the study eye on Day 1

Outcome Measures

Primary Outcome Measures:

1. To assess the ocular and systemic safety and tolerability of a single intravitreal (IVT) injection of VEGF Trap-Eye in patients with diabetic macular edema (DME)

Assessments for safety and tolerability are performed at each visit (Visit 1 - Visit 10)

Secondary Outcome Measures:

2. To obtain a preliminary assessment of the effect of a single dose of VEGF Trap-Eye on central retinal thickness (CRT) at the center point as determined by optical coherence tomography (OCT)

Assessments for CRT are performed at each visit (Visit 1 - Visit 10) by means of OCT.

3. To obtain a preliminary assessment of the effect of a single IVT administration of VEGF Trap-Eye on visual acuity

Assessments for visual acuity are performed at each visit (Visit 1 - Visit 10).

Eligibility

Minimum Age: 18 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Diagnosis of diabetes mellitus (type 1 or type 2).
- Best corrected E-ETDRS visual acuity score of ≥ 24 letters (i.e., 20/320 or better) and ≤ 73 letters (i.e., 20/40 or worse).
- On clinical exam, definite retinal thickening due to diabetic macular edema involving the center of the macula.
- Retinal Thickness at the center point ≥ 250 microns.
- Media clarity, pupillary dilation, and patient cooperation sufficient for adequate fundus photographs.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Visit 2 (Day 1).
- Macular edema due to causes other than diabetic macular edema. An eye should be considered ineligible: (1) if the macular edema is considered to be related to cataract extraction or (2) clinical exam and/or OCT suggests that vitreoretinal interface disease (e.g., a taut posterior hyaloid or epiretinal membrane) is the primary cause of the macular edema.
- An ocular condition is present such that, in the opinion of the investigator, visual acuity would not improve from resolution of macular edema (e.g., foveal atrophy, pigmentary changes, dense subfoveal hard exudates, nonretinal condition).
- An ocular condition is present (other than diabetes) that, in the opinion of the investigator, might affect macular edema or alter visual acuity during the course of the study (e.g., vein occlusion, age-related macular degeneration, uveitis or other ocular inflammatory disease, neovascular glaucoma, Irvine-Gass Syndrome, etc.).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase 1 study.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Maryland**

Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte, North Carolina, United States, 28210

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320814

Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

[Latest version \(submitted June 8, 2011\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>September 6, 2006</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 5, 2009</u>	Recruitment Status, Study Status, Outcome Measures, Arms and Interventions, Oversight, Contacts/Locations, Study Design and Sponsor/Collaborators

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
5	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320814

Submitted Date: January 5, 2009 (v3)

Study Identification

Unique Protocol ID: VGFT-OD-0512

Brief Title: Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of a Single Intravitreal Administration of VEGF Trap in Patients With Diabetic Macular Edema

Secondary IDs:

Study Status

Record Verification: January 2009

Overall Status: Completed

Study Start: April 2006

Primary Completion: August 2006 [Actual]

Study Completion: August 2007 [Actual]

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that January 5, 2009

Met QC Criteria:

Last Update Posted: January 6, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: No

Study Description

Brief Summary: To assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with diabetic macular edema.

Detailed Description: This is an open label study. Initially, 5 patients with DME will receive an ITV injection of VEGF Trap into the study eye. Additional patients may be enrolled at the same or additional dose levels. Patients will be observed for six weeks following the injection for assessments of ocular and systemic safety.

Conditions

Conditions: Diabetic Macular Edema

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Single Group Assignment

Number of Arms: 1

Masking: None (Open Label)

Allocation: Non-Randomized

Enrollment: 5 [Actual]

Arms and Interventions

Arms	Assigned Interventions
Experimental: VEGF Trap-Eye single IVT injection of 4.0 mg of VEGF Trap-Eye into the study eye on Day 1	Drug: VEGF Trap-Eye single IVT injection of 4.0 mg of VEGF Trap-Eye into the study eye on Day 1

Outcome Measures

Primary Outcome Measures:

1. To assess the ocular and systemic safety and tolerability of a single intravitreal (IVT) injection of VEGF Trap-Eye in patients with diabetic macular edema (DME)
Assessments for safety and tolerability are performed at each visit (Visit 1 - Visit 10)

Secondary Outcome Measures:

2. To obtain a preliminary assessment of the effect of a single dose of VEGF Trap-Eye on central retinal thickness (CRT) at the center point as determined by optical coherence tomography (OCT)
Assessments for CRT are performed at each visit (Visit 1 - Visit 10) by means of OCT.
3. To obtain a preliminary assessment of the effect of a single IVT administration of VEGF Trap-Eye on visual acuity
Assessments for visual acuity are performed at each visit (Visit 1 - Visit 10).

Eligibility

Minimum Age: 18 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Diagnosis of diabetes mellitus (type 1 or type 2).
- Best corrected E-ETDRS visual acuity score of ≥ 24 letters (i.e., 20/320 or better) and ≤ 73 letters (i.e., 20/40 or worse).
- On clinical exam, definite retinal thickening due to diabetic macular edema involving the center of the macula.
- Retinal Thickness at the center point ≥ 250 microns.
- Media clarity, pupillary dilation, and patient cooperation sufficient for adequate fundus photographs.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Visit 2 (Day 1).
- Macular edema due to causes other than diabetic macular edema. An eye should be considered ineligible: (1) if the macular edema is considered to be related to cataract extraction or (2) clinical exam and/or OCT suggests that vitreoretinal interface disease (e.g., a taut posterior hyaloid or epiretinal membrane) is the primary cause of the macular edema.
- An ocular condition is present such that, in the opinion of the investigator, visual acuity would not improve from resolution of macular edema (e.g., foveal atrophy, pigmentary changes, dense subfoveal hard exudates, nonretinal condition).
- An ocular condition is present (other than diabetes) that, in the opinion of the investigator, might affect macular edema or alter visual acuity during the course of the study (e.g., vein occlusion, age-related macular degeneration, uveitis or other ocular inflammatory disease, neovascular glaucoma, Irvine-Gass Syndrome, etc.).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase 1 study.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Maryland**

Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte, North Carolina, United States, 28210

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320814

Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Latest version (submitted June 8, 2011) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>September 6, 2006</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2009</u>	Recruitment Status, Study Status, Outcome Measures, Arms and Interventions, Oversight, Contacts/Locations, Study Design and Sponsor/Collaborators

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
5	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status

Compare

Comparison Format: Merged
 Side-by-Side

Scroll up to access the controls

Study NCT00320814

Submitted Date: September 6, 2006 (v2)

Study Identification

Unique Protocol ID: VGFT-OD-0512

Brief Title: Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of a Single Intravitreal Administration of VEGF Trap in Patients With Diabetic Macular Edema

Secondary IDs:

Study Status

Record Verification: September 2006

Overall Status: Active, not recruiting

Study Start: April 2006

Primary Completion:

Study Completion:

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that September 6, 2006

Met QC Criteria:

Last Update Posted: September 8, 2006 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring:

Study Description

Brief Summary: To assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with diabetic macular edema.

Detailed Description: This is an open label study. Initially, 5 patients with DME will receive an ITV injection of VEGF Trap into the study eye. Additional patients may be enrolled at the same or additional dose levels. Patients will be observed for six weeks following the injection for assessments of ocular and systemic safety.

Conditions

Conditions: Diabetic Macular Edema

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Single Group Assignment

Number of Arms:

Masking: None (Open Label)

Allocation: Non-Randomized

Enrollment: 5

Arms and Interventions

Intervention Details:

Drug: VEGF Trap

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, Bioeffect

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on: central retinal thickness, visual acuity and anti-VEGF Trap antibodies in the systemic circulation

Eligibility

Minimum Age: 18 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Diagnosis of diabetes mellitus (type 1 or type 2).
- Best corrected E-ETDRS visual acuity score of ≥ 24 letters (i.e., 20/320 or better) and ≤ 73 letters (i.e., 20/40 or worse).

- On clinical exam, definite retinal thickening due to diabetic macular edema involving the center of the macula.
- Retinal Thickness at the center point \geq 250 microns.
- Media clarity, pupillary dilation, and patient cooperation sufficient for adequate fundus photographs.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Visit 2 (Day 1).
- Macular edema due to causes other than diabetic macular edema. An eye should be considered ineligible: (1) if the macular edema is considered to be related to cataract extraction or (2) clinical exam and/or OCT suggests that vitreoretinal interface disease (e.g., a taut posterior hyaloid or epiretinal membrane) is the primary cause of the macular edema.
- An ocular condition is present such that, in the opinion of the investigator, visual acuity would not improve from resolution of macular edema (e.g., foveal atrophy, pigmentary changes, dense subfoveal hard exudates, nonretinal condition).
- An ocular condition is present (other than diabetes) that, in the opinion of the investigator, might affect macular edema or alter visual acuity during the course of the study (e.g., vein occlusion, age-related macular degeneration, uveitis or other ocular inflammatory disease, neovascular glaucoma, Irvine-Gass Syndrome, etc.).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase 1 study.

Contacts/Locations

Locations: **United States, Maryland**

Baltimore, Maryland, United States, 21287

United States, North Carolina

Charlotte, North Carolina, United States, 28210

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00320814

Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Latest version (submitted June 8, 2011) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>April 28, 2006</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>September 6, 2006</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2009</u>	Recruitment Status, Study Status, Outcome Measures, Arms and Interventions, Oversight, Contacts/Locations, Study Design and Sponsor/Collaborators

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>January 25, 2011</u>	Study Status
5	<input type="radio"/>	<input type="radio"/>	<u>June 8, 2011</u>	Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00320814

Submitted Date: April 28, 2006 (v1)

Study Identification

Unique Protocol ID: VGFT-OD-0512

Brief Title: Phase 1 Study of VEGF Trap in Patients With Diabetic Macular Edema

Official Title: An Exploratory Study of the Safety, Tolerability and Biological Effect of a Single Intravitreal Administration of VEGF Trap in Patients With Diabetic Macular Edema

Secondary IDs:

Study Status

Record Verification: April 2006

Overall Status: Recruiting

Study Start: April 2006

Primary Completion:

Study Completion:

First Submitted: April 28, 2006

First Submitted that April 28, 2006

Met QC Criteria:

First Posted: May 3, 2006 [Estimate]

Last Update Submitted that April 28, 2006

Met QC Criteria:

Last Update Posted: May 3, 2006 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators:

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring:

Study Description

Brief Summary: To assess the ocular and systemic safety and tolerability of a single intravitreal injection of VEGF Trap in patients with diabetic macular edema.

Detailed Description: This is an open label study. Initially, 5 patients with DME will receive an ITV injection of VEGF Trap into the study eye. Additional patients may be enrolled at the same or additional dose levels. Patients will be observed for six weeks following the injection for assessments of ocular and systemic safety.

Conditions

Conditions: Diabetic Macular Edema

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 1

Interventional Study Model: Single Group Assignment

Number of Arms:

Masking: None (Open Label)

Allocation: Non-Randomized

Enrollment: 5

Arms and Interventions

Intervention Details:

Drug: VEGF Trap

Outcome Measures

Primary Outcome Measures:

1. Safety and tolerability, Bioeffect

Secondary Outcome Measures:

2. The effect of VEGF Trap administration on: central retinal thickness, visual acuity and anti-VEGF Trap antibodies in the systemic circulation

Eligibility

Minimum Age: 18 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

- Diagnosis of diabetes mellitus (type 1 or type 2).
- Best corrected E-ETDRS visual acuity score of ≥ 24 letters (i.e., 20/320 or better) and ≤ 73 letters (i.e., 20/40 or worse).

- On clinical exam, definite retinal thickening due to diabetic macular edema involving the center of the macula.
- Retinal Thickness at the center point \geq 250 microns.
- Media clarity, pupillary dilation, and patient cooperation sufficient for adequate fundus photographs.

Exclusion Criteria:

- History of any vitreous hemorrhage within 4 weeks prior to Visit 2 (Day 1).
- Macular edema due to causes other than diabetic macular edema. An eye should be considered ineligible: (1) if the macular edema is considered to be related to cataract extraction or (2) clinical exam and/or OCT suggests that vitreoretinal interface disease (e.g., a taut posterior hyaloid or epiretinal membrane) is the primary cause of the macular edema.
- An ocular condition is present such that, in the opinion of the investigator, visual acuity would not improve from resolution of macular edema (e.g., foveal atrophy, pigmentary changes, dense subfoveal hard exudates, nonretinal condition).
- An ocular condition is present (other than diabetes) that, in the opinion of the investigator, might affect macular edema or alter visual acuity during the course of the study (e.g., vein occlusion, age-related macular degeneration, uveitis or other ocular inflammatory disease, neovascular glaucoma, Irvine-Gass Syndrome, etc.).
- Presence of any other condition or laboratory abnormality, which, in the opinion of the Investigator, would interfere with the assessment of disease status/progression or jeopardize the patient's appropriate participation in this Phase 1 study.

Contacts/Locations

Central Contact: Regeneron

Email: VEGF.Trap@regeneron.com

Locations: **United States, Maryland**

[Not yet recruiting]

Baltimore, Maryland, United States, 21287

United States, North Carolina

[Recruiting]

Charlotte, North Carolina, United States, 28210

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00509795

Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

[Latest version \(submitted December 20, 2012\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795
Submitted Date: December 1, 2011 (v15)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: December 2011

Overall Status: Completed

Study Start: August 2007

Primary Completion: September 2010 [Actual]

Study Completion: July 2011 [Actual]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Certification/Extension January 5, 2011

First Submitted:

Certification/Extension January 5, 2011

First Submitted that

Met QC Criteria:

Certification/Extension January 10, 2011 [Estimate]

First Posted:

Last Update Submitted that December 1, 2011

Met QC Criteria:

Last Update Posted: December 8, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party: Sponsor

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1217 [Actual]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	Drug: VEGF Trap-Eye 0.5 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 2	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 3	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 8 weeks (including one additional 2.0 mg dose at week 4) during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Arms	Assigned Interventions
Active Comparator: 4	Drug: ranibizumab 0.5 mg administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Outcome Measures

Primary Outcome Measures:

1. The proportion of subjects who maintain vision at Week 52, where a subject is classified as maintaining vision if the subject has lost fewer than 15 letters on the ETDRS chart compared to baseline (i.e. prevention of moderate vision loss)
Week 52

Secondary Outcome Measures:

2. Mean change from baseline in BCVA as measured by ETDRS letter score at Week 52
Week 52
3. The proportion of subjects who gain at least 15 letters of vision at Week 52
Week 52
4. Mean change from baseline in total NEI VFQ-25 score at Week 52
Week 52
5. Mean change from baseline in CNV area at Week 52
Week 52

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women ≥ 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents in the study eye.
4. Total lesion size > 12 disc areas (30.5 mm^2 , including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up $> 50\%$ of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.

15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Study Officials: Robert Vitti, MD
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Alabama**

Birmingham, Alabama, United States, 35205

Birmingham, Alabama, United States, 35223

United States, Arizona

Phoenix, Arizona, United States, 85014

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

Tucson, Arizona, United States, 85710

United States, California

Beverly Hills, California, United States, 90211

Campbell, California, United States, 95008

Fullerton, California, United States, 92835

Glendale, California, United States, 91203

Irvine, California, United States, 92697

La Jolla, California, United States, 92037

Loma Linda, California, United States, 92354

Los Angeles, California, United States, 90033

Los Angeles, California, United States, 90048

Menlo Park, California, United States, 94025
Mountain View, California, United States, 94040
Oakland, California, United States, 94609
Palm Springs, California, United States, 92262
Pasadena, California, United States, 91105
Poway, California, United States, 92064
Sacramento, California, United States, 95819
San Diego, California, United States, 92120
San Francisco, California, United States, 94107
Santa Ana, California, United States, 92705
Torrance, California, United States, 90503
Ventura, California, United States, 93003
Westlake Village, California, United States, 91361
Yorba Linda, California, United States, 92887

United States, Colorado

Aurora, Colorado, United States, 80045
Denver, Colorado, United States, 80205
Denver, Colorado, United States, 80230

United States, Connecticut

Bridgeport, Connecticut, United States, 06606
Hamden, Connecticut, United States, 06518
New Haven, Connecticut, United States, 06510
New London, Connecticut, United States, 06320

United States, Florida

Altamonte Springs, Florida, United States, 32701

Boynton Beach, Florida, United States, 33426

Fort Myers, Florida, United States, 33907

Ft. Lauderdale, Florida, United States, 33351

Ft. Myers, Florida, United States, 33912

Gainesville, Florida, United States, 32610

Jacksonville, Florida, United States, 32224

Miami, Florida, United States, 33136

Miami, Florida, United States, 33143

Mount Dora, Florida, United States, 32757

Orlando, Florida, United States, 32803

Orlando, Florida, United States, 32806

Ocala, Florida, United States, 34472

Palm Beach Gardens, Florida, United States, 33410

Pensacola, Florida, United States, 32503

Sarasota, Florida, United States

Stuart, Florida, United States, 34994

Tampa, Florida, United States, 33612

Winter Haven, Florida, United States, 33880

United States, Georgia

Augusta, Georgia, United States, 30909

United States, Hawaii

Aiea, Hawaii, United States, 96701

Honolulu, Hawaii, United States, 96813

United States, Illinois

Oak Brook, Illinois, United States, 60523

United States, Indiana

Fort Wayne, Indiana, United States, 46804

Indianapolis, Indiana, United States, 46202

Indianapolis, Indiana, United States, 46260

Indianapolis, Indiana, United States, 46280

New Albany, Indiana, United States, 47150

United States, Iowa

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

Wichita, Kansas, United States, 67214

United States, Kentucky

Louisville, Kentucky, United States, 40202

Louisville, Kentucky, United States, 40207

Paducah, Kentucky, United States, 42001

United States, Louisiana

New Orleans, Louisiana, United States, 70115

New Orleans, Louisiana, United States, 70121

Shreveport, Louisiana, United States, 71105

United States, Maine

Bangor, Maine, United States, 04401

Portland, Maine, United States, 04102

United States, Maryland

Baltimore, Maryland, United States, 21209

Baltimore, Maryland, United States, 21287

Chevy Chase, Maryland, United States, 20815

Hagerstown, Maryland, United States, 21740

Towson, Maryland, United States, 21204

United States, Massachusetts

Boston, Massachusetts, United States, 02111

Boston, Massachusetts, United States, 02114

Boston, Massachusetts, United States, 02215

Boston, Massachusetts, United States

Peabody, Massachusetts, United States, 01960

United States, Michigan

Ann Arbor, Michigan, United States, 48105

Battle Creek, Michigan, United States, 49015

Detroit, Michigan, United States, 48202

Grand Rapids, Michigan, United States, 49525

Jackson, Michigan, United States, 49201

Royal Oak, Michigan, United States, 48073

Southfield, Michigan, United States, 48034

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

Edina, Minnesota, United States, 55435

Minneapolis, Minnesota, United States, 55404

Rochester, Minnesota, United States, 55905

United States, Missouri

Florissant, Missouri, United States, 63031

Kansas City, Missouri, United States, 64108

Kansas City, Missouri, United States, 64111

Springfield, Missouri, United States, 65804

St. Louis, Missouri, United States, 63110

United States, Montana

Missoula, Montana, United States, 59801

United States, Nebraska

Lincoln, Nebraska, United States, 68506

Omaha, Nebraska, United States, 68131

United States, Nevada

Las Vegas, Nevada, United States, 89144

United States, New Jersey

Lawrenceville, New Jersey, United States, 08648

New Brunswick, New Jersey, United States, 08901

Northfield, New Jersey, United States, 08225

Teaneck, New Jersey, United States, 07666

Toms River, New Jersey, United States, 08753

United States, New Mexico

Albuquerque, New Mexico, United States, 87106

United States, New York

Albany, New York, United States, 12206

Brooklyn, New York, United States, 11223

Lynbrook, New York, United States, 11563
New York, New York, United States, 10003
New York, New York, United States, 10021
New York, New York, United States, 10032
Poughkeepsie, New York, United States, 12601
Rochester, New York, United States, 14620
Rochester, New York, United States, 14642
Slingerlands, New York, United States, 12159
Syracuse, New York, United States, 13224

United States, North Carolina

Asheville, North Carolina, United States, 28803
Charlotte, North Carolina, United States, 28210
Raleigh, North Carolina, United States, 27607
Southern Pines, North Carolina, United States, 28387
Winston-Salem, North Carolina, United States, 27157

United States, Ohio

Cincinnati, Ohio, United States, 45202
Cincinnati, Ohio, United States, 45242
Columbus, Ohio, United States, 43215
Toledo, Ohio, United States, 43608

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Ashland, Oregon, United States, 97520

Portland, Oregon, United States, 97210

Portland, Oregon, United States, 97227

Salem, Oregon, United States, 97302

United States, Pennsylvania

Kingston, Pennsylvania, United States, 18704

Philadelphia, Pennsylvania, United States, 19104

Philadelphia, Pennsylvania, United States, 19107

Philadelphia, Pennsylvania, United States, 19124

Pittsberg, Pennsylvania, United States, 15231

Pittsburgh, Pennsylvania, United States, 15212

Pittsburgh, Pennsylvania, United States, 15213

West Mifflin, Pennsylvania, United States, 15122

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

Charleston, South Carolina, United States, 29414

Columbia, South Carolina, United States, 29223

Greenville, South Carolina, United States, 29605

West Columbia, South Carolina, United States, 29169

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Memphis, Tennessee, United States, 38119

Memphis, Tennessee, United States, 38120

Nashville, Tennessee, United States, 37203

United States, Texas

Abilene, Texas, United States, 79606

Austin, Texas, United States, 78705

Corpus Cristi, Texas, United States, 78413

Dallas, Texas, United States, 75390

DeSoto, Texas, United States, 75115

Ft. Worth, Texas, United States, 76102

Ft. Worth, Texas, United States, 76104

Galveston, Texas, United States, 77555

Houston, Texas, United States, 77030

McAllen, Texas, United States, 78503

Odessa, Texas, United States, 79761

San Antonio, Texas, United States, 78240

United States, Utah

Salt Lake City, Utah, United States, 84107

Salt Lake City, Utah, United States, 84132

United States, Vermont

Burlington, Vermont, United States, 05401

United States, Virginia

Charlottesville, Virginia, United States, 22908

Fairfax, Virginia, United States, 22031

Richmond, Virginia, United States, 23221

United States, Washington

Seattle, Washington, United States, 98104

Silverdale, Washington, United States, 98383

United States, Wisconsin

Madison, Wisconsin, United States, 53715

Madison, Wisconsin, United States, 58705

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

Vancouver, British Columbia, Canada, V5Z 3N9

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

London, Ontario, Canada, N6A 4G5

Mississauga, Ontario, Canada, L4W 1W9

Ottawa, Ontario, Canada, K1H8L6

Toronto, Ontario, Canada, M4N3M5

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

Montreal, Quebec, Canada, H1T 2M4

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00509795

Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: May 4, 2011 (v14)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: April 2011

Overall Status: Active, not recruiting

Study Start: August 2007

Primary Completion: September 2010 [Actual]

Study Completion: July 2011 [Anticipated]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Certification/Extension January 5, 2011

First Submitted:

Certification/Extension January 5, 2011

First Submitted that

Met QC Criteria:

Certification/Extension January 10, 2011 [Estimate]

First Posted:

Last Update Submitted that May 4, 2011

Met QC Criteria:

Last Update Posted: May 6, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1217 [Actual]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	Drug: VEGF Trap-Eye 0.5 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 2	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 3	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 8 weeks (including one additional 2.0 mg dose at week 4) during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Arms	Assigned Interventions
Active Comparator: 4	Drug: ranibizumab 0.5 mg administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Outcome Measures

Primary Outcome Measures:

1. The proportion of subjects who maintain vision at Week 52, where a subject is classified as maintaining vision if the subject has lost fewer than 15 letters on the ETDRS chart compared to baseline (i.e. prevention of moderate vision loss)
Week 52

Secondary Outcome Measures:

2. Mean change from baseline in BCVA as measured by ETDRS letter score at Week 52
Week 52
3. The proportion of subjects who gain at least 15 letters of vision at Week 52
Week 52
4. Mean change from baseline in total NEI VFQ-25 score at Week 52
Week 52
5. Mean change from baseline in CNV area at Week 52
Week 52

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women ≥ 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents in the study eye.
4. Total lesion size > 12 disc areas (30.5 mm^2 , including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up $> 50\%$ of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.

15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Study Officials: Robert Vitti, MD
Study Director
Regeneron Pharmaceuticals

Locations: United States, Alabama

Birmingham, Alabama, United States, 35205

Birmingham, Alabama, United States, 35223

United States, Arizona

Phoenix, Arizona, United States, 85014

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

Tucson, Arizona, United States, 85710

United States, California

Beverly Hills, California, United States, 90211

Campbell, California, United States, 95008

Fullerton, California, United States, 92835

Glendale, California, United States, 91203

Irvine, California, United States, 92697

La Jolla, California, United States, 92037

Loma Linda, California, United States, 92354

Los Angeles, California, United States, 90033

Los Angeles, California, United States, 90048

Menlo Park, California, United States, 94025
Mountain View, California, United States, 94040
Oakland, California, United States, 94609
Palm Springs, California, United States, 92262
Pasadena, California, United States, 91105
Poway, California, United States, 92064
Sacramento, California, United States, 95819
San Diego, California, United States, 92120
San Francisco, California, United States, 94107
Santa Ana, California, United States, 92705
Torrance, California, United States, 90503
Ventura, California, United States, 93003
Westlake Village, California, United States, 91361
Yorba Linda, California, United States, 92887

United States, Colorado

Aurora, Colorado, United States, 80045
Denver, Colorado, United States, 80205
Denver, Colorado, United States, 80230

United States, Connecticut

Bridgeport, Connecticut, United States, 06606
Hamden, Connecticut, United States, 06518
New Haven, Connecticut, United States, 06510
New London, Connecticut, United States, 06320

United States, Florida

Altamonte Springs, Florida, United States, 32701

Boynton Beach, Florida, United States, 33426

Fort Myers, Florida, United States, 33907

Ft. Lauderdale, Florida, United States, 33351

Ft. Myers, Florida, United States, 33912

Gainesville, Florida, United States, 32610

Jacksonville, Florida, United States, 32224

Miami, Florida, United States, 33136

Miami, Florida, United States, 33143

Mount Dora, Florida, United States, 32757

Orlando, Florida, United States, 32803

Orlando, Florida, United States, 32806

Ocala, Florida, United States, 34472

Palm Beach Gardens, Florida, United States, 33410

Pensacola, Florida, United States, 32503

Sarasota, Florida, United States

Stuart, Florida, United States, 34994

Tampa, Florida, United States, 33612

Winter Haven, Florida, United States, 33880

United States, Georgia

Augusta, Georgia, United States, 30909

United States, Hawaii

Aiea, Hawaii, United States, 96701

Honolulu, Hawaii, United States, 96813

United States, Illinois

Oak Brook, Illinois, United States, 60523

United States, Indiana

Fort Wayne, Indiana, United States, 46804

Indianapolis, Indiana, United States, 46202

Indianapolis, Indiana, United States, 46260

Indianapolis, Indiana, United States, 46280

New Albany, Indiana, United States, 47150

United States, Iowa

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

Wichita, Kansas, United States, 67214

United States, Kentucky

Louisville, Kentucky, United States, 40202

Louisville, Kentucky, United States, 40207

Paducah, Kentucky, United States, 42001

United States, Louisiana

New Orleans, Louisiana, United States, 70115

New Orleans, Louisiana, United States, 70121

Shreveport, Louisiana, United States, 71105

United States, Maine

Bangor, Maine, United States, 04401

Portland, Maine, United States, 04102

United States, Maryland

Baltimore, Maryland, United States, 21209
Baltimore, Maryland, United States, 21287
Chevy Chase, Maryland, United States, 20815
Hagerstown, Maryland, United States, 21740
Towson, Maryland, United States, 21204

United States, Massachusetts

Boston, Massachusetts, United States, 02111
Boston, Massachusetts, United States, 02114
Boston, Massachusetts, United States, 02215
Boston, Massachusetts, United States
Peabody, Massachusetts, United States, 01960

United States, Michigan

Ann Arbor, Michigan, United States, 48105
Battle Creek, Michigan, United States, 49015
Detroit, Michigan, United States, 48202
Grand Rapids, Michigan, United States, 49525
Jackson, Michigan, United States, 49201
Royal Oak, Michigan, United States, 48073
Southfield, Michigan, United States, 48034
West Bloomfield, Michigan, United States, 48322

United States, Minnesota

Edina, Minnesota, United States, 55435
Minneapolis, Minnesota, United States, 55404
Rochester, Minnesota, United States, 55905

United States, Missouri

Florissant, Missouri, United States, 63031

Kansas City, Missouri, United States, 64108

Kansas City, Missouri, United States, 64111

Springfield, Missouri, United States, 65804

St. Louis, Missouri, United States, 63110

United States, Montana

Missoula, Montana, United States, 59801

United States, Nebraska

Lincoln, Nebraska, United States, 68506

Omaha, Nebraska, United States, 68131

United States, Nevada

Las Vegas, Nevada, United States, 89144

United States, New Jersey

Lawrenceville, New Jersey, United States, 08648

New Brunswick, New Jersey, United States, 08901

Northfield, New Jersey, United States, 08225

Teaneck, New Jersey, United States, 07666

Toms River, New Jersey, United States, 08753

United States, New Mexico

Albuquerque, New Mexico, United States, 87106

United States, New York

Albany, New York, United States, 12206

Brooklyn, New York, United States, 11223

Lynbrook, New York, United States, 11563
New York, New York, United States, 10003
New York, New York, United States, 10021
New York, New York, United States, 10032
Poughkeepsie, New York, United States, 12601
Rochester, New York, United States, 14620
Rochester, New York, United States, 14642
Slingerlands, New York, United States, 12159
Syracuse, New York, United States, 13224

United States, North Carolina

Asheville, North Carolina, United States, 28803
Charlotte, North Carolina, United States, 28210
Raleigh, North Carolina, United States, 27607
Southern Pines, North Carolina, United States, 28387
Winston-Salem, North Carolina, United States, 27157

United States, Ohio

Cincinnati, Ohio, United States, 45202
Cincinnati, Ohio, United States, 45242
Columbus, Ohio, United States, 43215
Toledo, Ohio, United States, 43608

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Ashland, Oregon, United States, 97520

Portland, Oregon, United States, 97210

Portland, Oregon, United States, 97227

Salem, Oregon, United States, 97302

United States, Pennsylvania

Kingston, Pennsylvania, United States, 18704

Philadelphia, Pennsylvania, United States, 19104

Philadelphia, Pennsylvania, United States, 19107

Philadelphia, Pennsylvania, United States, 19124

Pittsberg, Pennsylvania, United States, 15231

Pittsburgh, Pennsylvania, United States, 15212

Pittsburgh, Pennsylvania, United States, 15213

West Mifflin, Pennsylvania, United States, 15122

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

Charleston, South Carolina, United States, 29414

Columbia, South Carolina, United States, 29223

Greenville, South Carolina, United States, 29605

West Columbia, South Carolina, United States, 29169

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Memphis, Tennessee, United States, 38119

Memphis, Tennessee, United States, 38120

Nashville, Tennessee, United States, 37203

United States, Texas

Abilene, Texas, United States, 79606

Austin, Texas, United States, 78705

Corpus Cristi, Texas, United States, 78413

Dallas, Texas, United States, 75390

DeSoto, Texas, United States, 75115

Ft. Worth, Texas, United States, 76102

Ft. Worth, Texas, United States, 76104

Galveston, Texas, United States, 77555

Houston, Texas, United States, 77030

McAllen, Texas, United States, 78503

Odessa, Texas, United States, 79761

San Antonio, Texas, United States, 78240

United States, Utah

Salt Lake City, Utah, United States, 84107

Salt Lake City, Utah, United States, 84132

United States, Vermont

Burlington, Vermont, United States, 05401

United States, Virginia

Charlottesville, Virginia, United States, 22908

Fairfax, Virginia, United States, 22031

Richmond, Virginia, United States, 23221

United States, Washington

Seattle, Washington, United States, 98104

Silverdale, Washington, United States, 98383

United States, Wisconsin

Madison, Wisconsin, United States, 53715

Madison, Wisconsin, United States, 58705

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

Vancouver, British Columbia, Canada, V5Z 3N9

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

London, Ontario, Canada, N6A 4G5

Mississauga, Ontario, Canada, L4W 1W9

Ottawa, Ontario, Canada, K1H8L6

Toronto, Ontario, Canada, M4N3M5

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

Montreal, Quebec, Canada, H1T 2M4

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00509795

Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795
Submitted Date: April 18, 2011 (v13)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: April 2011

Overall Status: Active, not recruiting

Study Start: August 2007

Primary Completion: September 2010 [Actual]

Study Completion: July 2011 [Anticipated]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Certification/Extension January 5, 2011

First Submitted:

Certification/Extension January 5, 2011

First Submitted that

Met QC Criteria:

Certification/Extension January 10, 2011 [Estimate]

First Posted:

Last Update Submitted that April 18, 2011

Met QC Criteria:

Last Update Posted: April 27, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1217 [Actual]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	Drug: VEGF Trap-Eye 0.5 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 2	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 3	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 8 weeks (including one additional 2.0 mg dose at week 4) during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Arms	Assigned Interventions
Active Comparator: 4	Drug: ranibizumab 0.5 mg administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Outcome Measures

Primary Outcome Measures:

1. The proportion of subjects who maintain vision at Week 52, where a subject is classified as maintaining vision if the subject has lost fewer than 15 letters on the ETDRS chart compared to baseline (i.e. prevention of moderate vision loss)
Week 52

Secondary Outcome Measures:

2. Mean change from baseline in BCVA as measured by ETDRS letter score at Week 52
Week 52
3. The proportion of subjects who gain at least 15 letters of vision at Week 52
Week 52
4. Mean change from baseline in total NEI VFQ-25 score at Week 52
Week 52
5. Mean change from baseline in CNV area at Week 52
Week 52

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women ≥ 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents in the study eye.
4. Total lesion size > 12 disc areas (30.5 mm^2 , including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up $> 50\%$ of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.

15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Study Officials: Robert Vitti, MD
Study Director
Regeneron Pharmaceuticals

Locations: United States, Alabama

Birmingham, Alabama, United States, 35205

Birmingham, Alabama, United States, 35223

United States, Arizona

Phoenix, Arizona, United States, 85014

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

Tucson, Arizona, United States, 85710

United States, California

Beverly Hills, California, United States, 90211

Campbell, California, United States, 95008

Fullerton, California, United States, 92835

Glendale, California, United States, 91203

Irvine, California, United States, 92697

La Jolla, California, United States, 92037

Loma Linda, California, United States, 92354

Los Angeles, California, United States, 90033

Los Angeles, California, United States, 90048

Menlo Park, California, United States, 94025
Mountain View, California, United States, 94040
Oakland, California, United States, 94609
Palm Springs, California, United States, 92262
Pasadena, California, United States, 91105
Poway, California, United States, 92064
Sacramento, California, United States, 95819
San Diego, California, United States, 92120
San Francisco, California, United States, 94107
Santa Ana, California, United States, 92705
Torrance, California, United States, 90503
Ventura, California, United States, 93003
Westlake Village, California, United States, 91361
Yorba Linda, California, United States, 92887

United States, Colorado

Aurora, Colorado, United States, 80045
Denver, Colorado, United States, 80205
Denver, Colorado, United States, 80230

United States, Connecticut

Bridgeport, Connecticut, United States, 06606
Hamden, Connecticut, United States, 06518
New Haven, Connecticut, United States, 06510
New London, Connecticut, United States, 06320

United States, Florida

Altamonte Springs, Florida, United States, 32701

Boynton Beach, Florida, United States, 33426

Fort Myers, Florida, United States, 33907

Ft. Lauderdale, Florida, United States, 33351

Ft. Myers, Florida, United States, 33912

Gainesville, Florida, United States, 32610

Jacksonville, Florida, United States, 32224

Miami, Florida, United States, 33136

Miami, Florida, United States, 33143

Mount Dora, Florida, United States, 32757

Orlando, Florida, United States, 32803

Orlando, Florida, United States, 32806

Ocala, Florida, United States, 34472

Palm Beach Gardens, Florida, United States, 33410

Pensacola, Florida, United States, 32503

Sarasota, Florida, United States

Stuart, Florida, United States, 34994

Tampa, Florida, United States, 33612

Winter Haven, Florida, United States, 33880

United States, Georgia

Augusta, Georgia, United States, 30909

United States, Hawaii

Aiea, Hawaii, United States, 96701

Honolulu, Hawaii, United States, 96813

United States, Illinois

Oak Brook, Illinois, United States, 60523

United States, Indiana

Fort Wayne, Indiana, United States, 46804

Indianapolis, Indiana, United States, 46202

Indianapolis, Indiana, United States, 46260

Indianapolis, Indiana, United States, 46280

New Albany, Indiana, United States, 47150

United States, Iowa

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

Wichita, Kansas, United States, 67214

United States, Kentucky

Louisville, Kentucky, United States, 40202

Louisville, Kentucky, United States, 40207

Paducah, Kentucky, United States, 42001

United States, Louisiana

New Orleans, Louisiana, United States, 70115

New Orleans, Louisiana, United States, 70121

Shreveport, Louisiana, United States, 71105

United States, Maine

Bangor, Maine, United States, 04401

Portland, Maine, United States, 04102

United States, Maryland

Baltimore, Maryland, United States, 21209

Baltimore, Maryland, United States, 21287

Chevy Chase, Maryland, United States, 20815

Hagerstown, Maryland, United States, 21740

Towson, Maryland, United States, 21204

United States, Massachusetts

Boston, Massachusetts, United States, 02111

Boston, Massachusetts, United States, 02114

Boston, Massachusetts, United States, 02215

Boston, Massachusetts, United States

Peabody, Massachusetts, United States, 01960

United States, Michigan

Ann Arbor, Michigan, United States, 48105

Battle Creek, Michigan, United States, 49015

Detroit, Michigan, United States, 48202

Grand Rapids, Michigan, United States, 49525

Jackson, Michigan, United States, 49201

Royal Oak, Michigan, United States, 48073

Southfield, Michigan, United States, 48034

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

Edina, Minnesota, United States, 55435

Minneapolis, Minnesota, United States, 55404

Rochester, Minnesota, United States, 55905

United States, Missouri

Florissant, Missouri, United States, 63031

Kansas City, Missouri, United States, 64108

Kansas City, Missouri, United States, 64111

Springfield, Missouri, United States, 65804

St. Louis, Missouri, United States, 63110

United States, Montana

Missoula, Montana, United States, 59801

United States, Nebraska

Lincoln, Nebraska, United States, 68506

Omaha, Nebraska, United States, 68131

United States, Nevada

Las Vegas, Nevada, United States, 89144

United States, New Jersey

Lawrenceville, New Jersey, United States, 08648

New Brunswick, New Jersey, United States, 08901

Northfield, New Jersey, United States, 08225

Teaneck, New Jersey, United States, 07666

Toms River, New Jersey, United States, 08753

United States, New Mexico

Albuquerque, New Mexico, United States, 87106

United States, New York

Albany, New York, United States, 12206

Brooklyn, New York, United States, 11223

Lynbrook, New York, United States, 11563
New York, New York, United States, 10003
New York, New York, United States, 10021
New York, New York, United States, 10032
Poughkeepsie, New York, United States, 12601
Rochester, New York, United States, 14620
Rochester, New York, United States, 14642
Slingerlands, New York, United States, 12159
Syracuse, New York, United States, 13224

United States, North Carolina

Asheville, North Carolina, United States, 28803
Charlotte, North Carolina, United States, 28210
Raleigh, North Carolina, United States, 27607
Southern Pines, North Carolina, United States, 28387
Winston-Salem, North Carolina, United States, 27157

United States, Ohio

Cincinnati, Ohio, United States, 45202
Cincinnati, Ohio, United States, 45242
Columbus, Ohio, United States, 43215
Toledo, Ohio, United States, 43608

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Ashland, Oregon, United States, 97520

Portland, Oregon, United States, 97210

Portland, Oregon, United States, 97227

Salem, Oregon, United States, 97302

United States, Pennsylvania

Kingston, Pennsylvania, United States, 18704

Philadelphia, Pennsylvania, United States, 19104

Philadelphia, Pennsylvania, United States, 19107

Philadelphia, Pennsylvania, United States, 19124

Pittsberg, Pennsylvania, United States, 15231

Pittsburgh, Pennsylvania, United States, 15212

Pittsburgh, Pennsylvania, United States, 15213

West Mifflin, Pennsylvania, United States, 15122

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

Charleston, South Carolina, United States, 29414

Columbia, South Carolina, United States, 29223

Greenville, South Carolina, United States, 29605

West Columbia, South Carolina, United States, 29169

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Memphis, Tennessee, United States, 38119

Memphis, Tennessee, United States, 38120

Nashville, Tennessee, United States, 37203

United States, Texas

Abilene, Texas, United States, 79606

Austin, Texas, United States, 78705

Corpus Cristi, Texas, United States, 78413

Dallas, Texas, United States, 75390

DeSoto, Texas, United States, 75115

Ft. Worth, Texas, United States, 76102

Ft. Worth, Texas, United States, 76104

Galveston, Texas, United States, 77555

Houston, Texas, United States, 77030

McAllen, Texas, United States, 78503

Odessa, Texas, United States, 79761

San Antonio, Texas, United States, 78240

United States, Utah

Salt Lake City, Utah, United States, 84107

Salt Lake City, Utah, United States, 84132

United States, Vermont

Burlington, Vermont, United States, 05401

United States, Virginia

Charlottesville, Virginia, United States, 22908

Fairfax, Virginia, United States, 22031

Richmond, Virginia, United States, 23221

United States, Washington

Seattle, Washington, United States, 98104

Silverdale, Washington, United States, 98383

United States, Wisconsin

Madison, Wisconsin, United States, 53715

Madison, Wisconsin, United States, 58705

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

Vancouver, British Columbia, Canada, V5Z 3N9

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

London, Ontario, Canada, N6A 4G5

Mississauga, Ontario, Canada, L4W 1W9

Ottawa, Ontario, Canada, K1H8L6

Toronto, Ontario, Canada, M4N3M5

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

Montreal, Quebec, Canada, H1T 2M4

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00509795

Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: January 5, 2011 (v12)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: January 2011

Overall Status: Active, not recruiting

Study Start: August 2007

Primary Completion: December 2011 [Anticipated]

Study Completion: December 2011 [Anticipated]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Certification/Extension January 5, 2011

First Submitted:

Certification/Extension January 5, 2011

First Submitted that

Met QC Criteria:

Certification/Extension January 10, 2011 [Estimate]

First Posted:

Last Update Submitted that January 5, 2011

Met QC Criteria:

Last Update Posted: January 10, 2011 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	Drug: VEGF Trap-Eye 0.5 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 2	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 3	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 8 weeks (including one additional 2.0 mg dose at week 4) during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Arms	Assigned Interventions
Active Comparator: 4	Drug: ranibizumab 0.5 mg administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Outcome Measures

Primary Outcome Measures:

1. The proportion of subjects who maintain vision at Week 52, where a subject is classified as maintaining vision if the subject has lost fewer than 15 letters on the ETDRS chart compared to baseline (i.e. prevention of moderate vision loss)
Week 52

Secondary Outcome Measures:

2. Mean change from baseline in BCVA as measured by ETDRS letter score at Week 52
Week 52
3. The proportion of subjects who gain at least 15 letters of vision at Week 52
Week 52
4. Mean change from baseline in total NEI VFQ-25 score at Week 52
Week 52
5. Mean change from baseline in CNV area at Week 52
Week 52

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women ≥ 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents in the study eye.
4. Total lesion size > 12 disc areas (30.5 mm^2 , including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up $> 50\%$ of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.

15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Study Officials: Robert Vitti, MD
Study Director
Regeneron Pharmaceuticals

Locations: United States, Alabama

Birmingham, Alabama, United States, 35205

Birmingham, Alabama, United States, 35223

United States, Arizona

Phoenix, Arizona, United States, 85014

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

Tucson, Arizona, United States, 85710

United States, California

Beverly Hills, California, United States, 90211

Campbell, California, United States, 95008

Fullerton, California, United States, 92835

Glendale, California, United States, 91203

Irvine, California, United States, 92697

La Jolla, California, United States, 92037

Loma Linda, California, United States, 92354

Los Angeles, California, United States, 90033

Los Angeles, California, United States, 90048

Menlo Park, California, United States, 94025
Mountain View, California, United States, 94040
Oakland, California, United States, 94609
Palm Springs, California, United States, 92262
Pasadena, California, United States, 91105
Poway, California, United States, 92064
Sacramento, California, United States, 95819
San Diego, California, United States, 92120
San Francisco, California, United States, 94107
Santa Ana, California, United States, 92705
Torrance, California, United States, 90503
Ventura, California, United States, 93003
Westlake Village, California, United States, 91361
Yorba Linda, California, United States, 92887

United States, Colorado

Aurora, Colorado, United States, 80045
Denver, Colorado, United States, 80205
Denver, Colorado, United States, 80230

United States, Connecticut

Bridgeport, Connecticut, United States, 06606
Hamden, Connecticut, United States, 06518
New Haven, Connecticut, United States, 06510
New London, Connecticut, United States, 06320

United States, Florida

Altamonte Springs, Florida, United States, 32701

Boynton Beach, Florida, United States, 33426

Fort Myers, Florida, United States, 33907

Ft. Lauderdale, Florida, United States, 33351

Ft. Myers, Florida, United States, 33912

Gainesville, Florida, United States, 32610

Jacksonville, Florida, United States, 32224

Miami, Florida, United States, 33136

Miami, Florida, United States, 33143

Mount Dora, Florida, United States, 32757

Orlando, Florida, United States, 32803

Orlando, Florida, United States, 32806

Ocala, Florida, United States, 34472

Palm Beach Gardens, Florida, United States, 33410

Pensacola, Florida, United States, 32503

Sarasota, Florida, United States

Stuart, Florida, United States, 34994

Tampa, Florida, United States, 33612

Winter Haven, Florida, United States, 33880

United States, Georgia

Augusta, Georgia, United States, 30909

United States, Hawaii

Aiea, Hawaii, United States, 96701

Honolulu, Hawaii, United States, 96813

United States, Illinois

Oak Brook, Illinois, United States, 60523

United States, Indiana

Fort Wayne, Indiana, United States, 46804

Indianapolis, Indiana, United States, 46202

Indianapolis, Indiana, United States, 46260

Indianapolis, Indiana, United States, 46280

New Albany, Indiana, United States, 47150

United States, Iowa

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

Wichita, Kansas, United States, 67214

United States, Kentucky

Louisville, Kentucky, United States, 40202

Louisville, Kentucky, United States, 40207

Paducah, Kentucky, United States, 42001

United States, Louisiana

New Orleans, Louisiana, United States, 70115

New Orleans, Louisiana, United States, 70121

Shreveport, Louisiana, United States, 71105

United States, Maine

Bangor, Maine, United States, 04401

Portland, Maine, United States, 04102

United States, Maryland

Baltimore, Maryland, United States, 21209
Baltimore, Maryland, United States, 21287
Chevy Chase, Maryland, United States, 20815
Hagerstown, Maryland, United States, 21740
Towson, Maryland, United States, 21204

United States, Massachusetts

Boston, Massachusetts, United States, 02111
Boston, Massachusetts, United States, 02114
Boston, Massachusetts, United States, 02215
Boston, Massachusetts, United States
Peabody, Massachusetts, United States, 01960

United States, Michigan

Ann Arbor, Michigan, United States, 48105
Battle Creek, Michigan, United States, 49015
Detroit, Michigan, United States, 48202
Grand Rapids, Michigan, United States, 49525
Jackson, Michigan, United States, 49201
Royal Oak, Michigan, United States, 48073
Southfield, Michigan, United States, 48034
West Bloomfield, Michigan, United States, 48322

United States, Minnesota

Edina, Minnesota, United States, 55435
Minneapolis, Minnesota, United States, 55404
Rochester, Minnesota, United States, 55905

United States, Missouri

Florissant, Missouri, United States, 63031

Kansas City, Missouri, United States, 64108

Kansas City, Missouri, United States, 64111

Springfield, Missouri, United States, 65804

St. Louis, Missouri, United States, 63110

United States, Montana

Missoula, Montana, United States, 59801

United States, Nebraska

Lincoln, Nebraska, United States, 68506

Omaha, Nebraska, United States, 68131

United States, Nevada

Las Vegas, Nevada, United States, 89144

United States, New Jersey

Lawrenceville, New Jersey, United States, 08648

New Brunswick, New Jersey, United States, 08901

Northfield, New Jersey, United States, 08225

Teaneck, New Jersey, United States, 07666

Toms River, New Jersey, United States, 08753

United States, New Mexico

Albuquerque, New Mexico, United States, 87106

United States, New York

Albany, New York, United States, 12206

Brooklyn, New York, United States, 11223

Lynbrook, New York, United States, 11563
New York, New York, United States, 10003
New York, New York, United States, 10021
New York, New York, United States, 10032
Poughkeepsie, New York, United States, 12601
Rochester, New York, United States, 14620
Rochester, New York, United States, 14642
Slingerlands, New York, United States, 12159
Syracuse, New York, United States, 13224

United States, North Carolina

Asheville, North Carolina, United States, 28803
Charlotte, North Carolina, United States, 28210
Raleigh, North Carolina, United States, 27607
Southern Pines, North Carolina, United States, 28387
Winston-Salem, North Carolina, United States, 27157

United States, Ohio

Cincinnati, Ohio, United States, 45202
Cincinnati, Ohio, United States, 45242
Columbus, Ohio, United States, 43215
Toledo, Ohio, United States, 43608

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Ashland, Oregon, United States, 97520

Portland, Oregon, United States, 97210

Portland, Oregon, United States, 97227

Salem, Oregon, United States, 97302

United States, Pennsylvania

Kingston, Pennsylvania, United States, 18704

Philadelphia, Pennsylvania, United States, 19104

Philadelphia, Pennsylvania, United States, 19107

Philadelphia, Pennsylvania, United States, 19124

Pittsberg, Pennsylvania, United States, 15231

Pittsburgh, Pennsylvania, United States, 15212

Pittsburgh, Pennsylvania, United States, 15213

West Mifflin, Pennsylvania, United States, 15122

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

Charleston, South Carolina, United States, 29414

Columbia, South Carolina, United States, 29223

Greenville, South Carolina, United States, 29605

West Columbia, South Carolina, United States, 29169

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Memphis, Tennessee, United States, 38119

Memphis, Tennessee, United States, 38120

Nashville, Tennessee, United States, 37203

United States, Texas

Abilene, Texas, United States, 79606

Austin, Texas, United States, 78705

Corpus Cristi, Texas, United States, 78413

Dallas, Texas, United States, 75390

DeSoto, Texas, United States, 75115

Ft. Worth, Texas, United States, 76102

Ft. Worth, Texas, United States, 76104

Galveston, Texas, United States, 77555

Houston, Texas, United States, 77030

McAllen, Texas, United States, 78503

Odessa, Texas, United States, 79761

San Antonio, Texas, United States, 78240

United States, Utah

Salt Lake City, Utah, United States, 84107

Salt Lake City, Utah, United States, 84132

United States, Vermont

Burlington, Vermont, United States, 05401

United States, Virginia

Charlottesville, Virginia, United States, 22908

Fairfax, Virginia, United States, 22031

Richmond, Virginia, United States, 23221

United States, Washington

Seattle, Washington, United States, 98104

Silverdale, Washington, United States, 98383

United States, Wisconsin

Madison, Wisconsin, United States, 53715

Madison, Wisconsin, United States, 58705

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

Vancouver, British Columbia, Canada, V5Z 3N9

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

London, Ontario, Canada, N6A 4G5

Mississauga, Ontario, Canada, L4W 1W9

Ottawa, Ontario, Canada, K1H8L6

Toronto, Ontario, Canada, M4N3M5

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

Montreal, Quebec, Canada, H1T 2M4

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00509795

Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: December 1, 2009 (v11)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: December 2009

Overall Status: Active, not recruiting

Study Start: August 2007

Primary Completion: December 2011 [Anticipated]

Study Completion: December 2011 [Anticipated]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that December 1, 2009

Met QC Criteria:

Last Update Posted: December 2, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	Drug: VEGF Trap-Eye 0.5 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 2	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 3	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 8 weeks (including one additional 2.0 mg dose at week 4) during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Active Comparator: 4	Drug: ranibizumab 0.5 mg administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Outcome Measures

Primary Outcome Measures:

1. The proportion of subjects who maintain vision at Week 52, where a subject is classified as maintaining vision if the subject has

lost fewer than 15 letters on the ETDRS chart compared to baseline (i.e. prevention of moderate vision loss)

Week 52

Secondary Outcome Measures:

2. Mean change from baseline in BCVA as measured by ETDRS letter score at Week 52

Week 52

3. The proportion of subjects who gain at least 15 letters of vision at Week 52

Week 52

4. Mean change from baseline in total NEI VFQ-25 score at Week 52

Week 52

5. Mean change from baseline in CNV area at Week 52

Week 52

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents in the study eye.
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Study Officials: Robert Vitti, MD
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Alabama**

Birmingham, Alabama, United States, 35205

Birmingham, Alabama, United States, 35223

United States, Arizona

Phoenix, Arizona, United States, 85014

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

Tucson, Arizona, United States, 85710

United States, California

Beverly Hills, California, United States, 90211

Campbell, California, United States, 95008

Fullerton, California, United States, 92835

Glendale, California, United States, 91203

Irvine, California, United States, 92697

La Jolla, California, United States, 92037

Loma Linda, California, United States, 92354

Los Angeles, California, United States, 90033

Los Angeles, California, United States, 90048

Menlo Park, California, United States, 94025

Mountain View, California, United States, 94040

Oakland, California, United States, 94609

Palm Springs, California, United States, 92262

Pasadena, California, United States, 91105

Poway, California, United States, 92064

Sacramento, California, United States, 95819

San Diego, California, United States, 92120

San Francisco, California, United States, 94107

Santa Ana, California, United States, 92705

Torrance, California, United States, 90503

Ventura, California, United States, 93003

Westlake Village, California, United States, 91361

Yorba Linda, California, United States, 92887

United States, Colorado

Aurora, Colorado, United States, 80045

Denver, Colorado, United States, 80205

Denver, Colorado, United States, 80230

United States, Connecticut

Bridgeport, Connecticut, United States, 06606

Hamden, Connecticut, United States, 06518

New Haven, Connecticut, United States, 06510

New London, Connecticut, United States, 06320

United States, Florida

Altamonte Springs, Florida, United States, 32701

Boynton Beach, Florida, United States, 33426

Fort Myers, Florida, United States, 33907

Ft. Lauderdale, Florida, United States, 33351

Ft. Myers, Florida, United States, 33912

Gainesville, Florida, United States, 32610

Jacksonville, Florida, United States, 32224

Miami, Florida, United States, 33136

Miami, Florida, United States, 33143
Mount Dora, Florida, United States, 32757
Orlando, Florida, United States, 32803
Orlando, Florida, United States, 32806
Oscala, Florida, United States, 34472
Palm Beach Gardens, Florida, United States, 33410
Pensacola, Florida, United States, 32503
Sarasota, Florida, United States
Stuart, Florida, United States, 34994
Tampa, Florida, United States, 33612
Winter Haven, Florida, United States, 33880

United States, Georgia

Augusta, Georgia, United States, 30909

United States, Hawaii

Aiea, Hawaii, United States, 96701
Honolulu, Hawaii, United States, 96813

United States, Illinois

Oak Brook, Illinois, United States, 60523

United States, Indiana

Fort Wayne, Indiana, United States, 46804
Indianapolis, Indiana, United States, 46202
Indianapolis, Indiana, United States, 46260
Indianapolis, Indiana, United States, 46280
New Albany, Indiana, United States, 47150

United States, Iowa

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

Wichita, Kansas, United States, 67214

United States, Kentucky

Louisville, Kentucky, United States, 40202

Louisville, Kentucky, United States, 40207

Paducah, Kentucky, United States, 42001

United States, Louisiana

New Orleans, Louisiana, United States, 70115

New Orleans, Louisiana, United States, 70121

Shreveport, Louisiana, United States, 71105

United States, Maine

Bangor, Maine, United States, 04401

Portland, Maine, United States, 04102

United States, Maryland

Baltimore, Maryland, United States, 21209

Baltimore, Maryland, United States, 21287

Chevy Chase, Maryland, United States, 20815

Hagerstown, Maryland, United States, 21740

Towson, Maryland, United States, 21204

United States, Massachusetts

Boston, Massachusetts, United States, 02111

Boston, Massachusetts, United States, 02114

Boston, Massachusetts, United States, 02215

Boston, Massachusetts, United States

Peabody, Massachusetts, United States, 01960

United States, Michigan

Ann Arbor, Michigan, United States, 48105

Battle Creek, Michigan, United States, 49015

Detroit, Michigan, United States, 48202

Grand Rapids, Michigan, United States, 49525

Jackson, Michigan, United States, 49201

Royal Oak, Michigan, United States, 48073

Southfield, Michigan, United States, 48034

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

Edina, Minnesota, United States, 55435

Minneapolis, Minnesota, United States, 55404

Rochester, Minnesota, United States, 55905

United States, Missouri

Florissant, Missouri, United States, 63031

Kansas City, Missouri, United States, 64108

Kansas City, Missouri, United States, 64111

Springfield, Missouri, United States, 65804

St. Louis, Missouri, United States, 63110

United States, Montana

Missoula, Montana, United States, 59801

United States, Nebraska

Lincoln, Nebraska, United States, 68506

Omaha, Nebraska, United States, 68131

United States, Nevada

Las Vegas, Nevada, United States, 89144

United States, New Jersey

Lawrenceville, New Jersey, United States, 08648

New Brunswick, New Jersey, United States, 08901

Northfield, New Jersey, United States, 08225

Teaneck, New Jersey, United States, 07666

Toms River, New Jersey, United States, 08753

United States, New Mexico

Albuquerque, New Mexico, United States, 87106

United States, New York

Albany, New York, United States, 12206

Brooklyn, New York, United States, 11223

Lynbrook, New York, United States, 11563

New York, New York, United States, 10003

New York, New York, United States, 10021

New York, New York, United States, 10032

Poughkeepsie, New York, United States, 12601

Rochester, New York, United States, 14620

Rochester, New York, United States, 14642

Slingerlands, New York, United States, 12159

Syracuse, New York, United States, 13224

United States, North Carolina

Asheville, North Carolina, United States, 28803

Charlotte, North Carolina, United States, 28210

Raleigh, North Carolina, United States, 27607

Southern Pines, North Carolina, United States, 28387

Winston-Salem, North Carolina, United States, 27157

United States, Ohio

Cincinnati, Ohio, United States, 45202

Cincinnati, Ohio, United States, 45242

Columbus, Ohio, United States, 43215

Toledo, Ohio, United States, 43608

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Ashland, Oregon, United States, 97520

Portland, Oregon, United States, 97210

Portland, Oregon, United States, 97227

Salem, Oregon, United States, 97302

United States, Pennsylvania

Kingston, Pennsylvania, United States, 18704

Philadelphia, Pennsylvania, United States, 19104

Philadelphia, Pennsylvania, United States, 19107

Philadelphia, Pennsylvania, United States, 19124

Pittsberg, Pennsylvania, United States, 15231
Pittsburgh, Pennsylvania, United States, 15212
Pittsburgh, Pennsylvania, United States, 15213
West Mifflin, Pennsylvania, United States, 15122
Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

Charleston, South Carolina, United States, 29414
Columbia, South Carolina, United States, 29223
Greenville, South Carolina, United States, 29605
West Columbia, South Carolina, United States, 29169

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Memphis, Tennessee, United States, 38119
Memphis, Tennessee, United States, 38120
Nashville, Tennessee, United States, 37203

United States, Texas

Abilene, Texas, United States, 79606
Austin, Texas, United States, 78705
Corpus Cristi, Texas, United States, 78413
Dallas, Texas, United States, 75390
DeSoto, Texas, United States, 75115

Ft. Worth, Texas, United States, 76102

Ft. Worth, Texas, United States, 76104

Galveston, Texas, United States, 77555

Houston, Texas, United States, 77030

McAllen, Texas, United States, 78503

Odessa, Texas, United States, 79761

San Antonio, Texas, United States, 78240

United States, Utah

Salt Lake City, Utah, United States, 84107

Salt Lake City, Utah, United States, 84132

United States, Vermont

Burlington, Vermont, United States, 05401

United States, Virginia

Charlottesville, Virginia, United States, 22908

Fairfax, Virginia, United States, 22031

Richmond, Virginia, United States, 23221

United States, Washington

Seattle, Washington, United States, 98104

Silverdale, Washington, United States, 98383

United States, Wisconsin

Madison, Wisconsin, United States, 53715

Madison, Wisconsin, United States, 58705

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

Vancouver, British Columbia, Canada, V5Z 3N9

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

London, Ontario, Canada, N6A 4G5

Mississauga, Ontario, Canada, L4W 1W9

Ottawa, Ontario, Canada, K1H8L6

Toronto, Ontario, Canada, M4N3M5

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

Montreal, Quebec, Canada, H1T 2M4

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

History of Changes for Study: NCT00509795

Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: September 12, 2009 (v10)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: September 2009

Overall Status: Active, not recruiting

Study Start: August 2007

Primary Completion: December 2011 [Anticipated]

Study Completion: December 2011 [Anticipated]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that September 12, 2009

Met QC Criteria:

Last Update Posted: September 15, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	Drug: VEGF Trap-Eye 0.5 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 2	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 3	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 8 weeks (including one additional 2.0 mg dose at week 4) during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Active Comparator: 4	Drug: ranibizumab 0.5 mg administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Outcome Measures

Primary Outcome Measures:

1. The proportion of subjects who maintain vision at Week 52, where a subject is classified as maintaining vision if the subject has

lost fewer than 15 letters on the ETDRS chart compared to baseline (i.e. prevention of moderate vision loss)

Week 52

Secondary Outcome Measures:

2. Mean change from baseline in BCVA as measured by ETDRS letter score at Week 52

Week 52

3. The proportion of subjects who gain at least 15 letters of vision at Week 52

Week 52

4. Mean change from baseline in total NEI VFQ-25 score at Week 52

Week 52

5. Mean change from baseline in CNV area at Week 52

Week 52

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents in the study eye.
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Alabama**

Birmingham, Alabama, United States, 35205

Birmingham, Alabama, United States, 35223

United States, Arizona

Phoenix, Arizona, United States, 85014

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

Tucson, Arizona, United States, 85710

United States, California

Beverly Hills, California, United States, 90211

Campbell, California, United States, 95008

Fullerton, California, United States, 92835

Glendale, California, United States, 91203

Irvine, California, United States, 92697

La Jolla, California, United States, 92037

Loma Linda, California, United States, 92354

Los Angeles, California, United States, 90033

Los Angeles, California, United States, 90048

Menlo Park, California, United States, 94025

Mountain View, California, United States, 94040

Oakland, California, United States, 94609

Palm Springs, California, United States, 92262

Pasadena, California, United States, 91105

Poway, California, United States, 92064

Sacramento, California, United States, 95819

San Diego, California, United States, 92120

San Francisco, California, United States, 94107

Santa Ana, California, United States, 92705

Torrance, California, United States, 90503

Ventura, California, United States, 93003

Westlake Village, California, United States, 91361

Yorba Linda, California, United States, 92887

United States, Colorado

Aurora, Colorado, United States, 80045

Denver, Colorado, United States, 80205

Denver, Colorado, United States, 80230

United States, Connecticut

Bridgeport, Connecticut, United States, 06606

Hamden, Connecticut, United States, 06518

New Haven, Connecticut, United States, 06510

New London, Connecticut, United States, 06320

United States, Florida

Altamonte Springs, Florida, United States, 32701

Boynton Beach, Florida, United States, 33426

Fort Myers, Florida, United States, 33907

Ft. Lauderdale, Florida, United States, 33351

Ft. Myers, Florida, United States, 33912

Gainesville, Florida, United States, 32610

Jacksonville, Florida, United States, 32224

Miami, Florida, United States, 33136

Miami, Florida, United States, 33143
Mount Dora, Florida, United States, 32757
Orlando, Florida, United States, 32803
Orlando, Florida, United States, 32806
Oscala, Florida, United States, 34472
Palm Beach Gardens, Florida, United States, 33410
Pensacola, Florida, United States, 32503
Sarasota, Florida, United States
Stuart, Florida, United States, 34994
Tampa, Florida, United States, 33612
Winter Haven, Florida, United States, 33880

United States, Georgia

Augusta, Georgia, United States, 30909

United States, Hawaii

Aiea, Hawaii, United States, 96701
Honolulu, Hawaii, United States, 96813

United States, Illinois

Oak Brook, Illinois, United States, 60523

United States, Indiana

Fort Wayne, Indiana, United States, 46804
Indianapolis, Indiana, United States, 46202
Indianapolis, Indiana, United States, 46260
Indianapolis, Indiana, United States, 46280
New Albany, Indiana, United States, 47150

United States, Iowa

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

Wichita, Kansas, United States, 67214

United States, Kentucky

Louisville, Kentucky, United States, 40202

Louisville, Kentucky, United States, 40207

Paducah, Kentucky, United States, 42001

United States, Louisiana

New Orleans, Louisiana, United States, 70115

New Orleans, Louisiana, United States, 70121

Shreveport, Louisiana, United States, 71105

United States, Maine

Bangor, Maine, United States, 04401

Portland, Maine, United States, 04102

United States, Maryland

Baltimore, Maryland, United States, 21209

Baltimore, Maryland, United States, 21287

Chevy Chase, Maryland, United States, 20815

Hagerstown, Maryland, United States, 21740

Towson, Maryland, United States, 21204

United States, Massachusetts

Boston, Massachusetts, United States, 02111

Boston, Massachusetts, United States, 02114

Boston, Massachusetts, United States, 02215

Boston, Massachusetts, United States

Peabody, Massachusetts, United States, 01960

United States, Michigan

Ann Arbor, Michigan, United States, 48105

Battle Creek, Michigan, United States, 49015

Detroit, Michigan, United States, 48202

Grand Rapids, Michigan, United States, 49525

Jackson, Michigan, United States, 49201

Royal Oak, Michigan, United States, 48073

Southfield, Michigan, United States, 48034

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

Edina, Minnesota, United States, 55435

Minneapolis, Minnesota, United States, 55404

Rochester, Minnesota, United States, 55905

United States, Missouri

Florissant, Missouri, United States, 63031

Kansas City, Missouri, United States, 64108

Kansas City, Missouri, United States, 64111

Springfield, Missouri, United States, 65804

St. Louis, Missouri, United States, 63110

United States, Montana

Missoula, Montana, United States, 59801

United States, Nebraska

Lincoln, Nebraska, United States, 68506

Omaha, Nebraska, United States, 68131

United States, Nevada

Las Vegas, Nevada, United States, 89144

United States, New Jersey

Lawrenceville, New Jersey, United States, 08648

New Brunswick, New Jersey, United States, 08901

Northfield, New Jersey, United States, 08225

Teaneck, New Jersey, United States, 07666

Toms River, New Jersey, United States, 08753

United States, New Mexico

Albuquerque, New Mexico, United States, 87106

United States, New York

Albany, New York, United States, 12206

Brooklyn, New York, United States, 11223

Lynbrook, New York, United States, 11563

New York, New York, United States, 10003

New York, New York, United States, 10021

New York, New York, United States, 10032

Poughkeepsie, New York, United States, 12601

Rochester, New York, United States, 14620

Rochester, New York, United States, 14642

Slingerlands, New York, United States, 12159

Syracuse, New York, United States, 13224

United States, North Carolina

Asheville, North Carolina, United States, 28803

Charlotte, North Carolina, United States, 28210

Raleigh, North Carolina, United States, 27607

Southern Pines, North Carolina, United States, 28387

Winston-Salem, North Carolina, United States, 27157

United States, Ohio

Cincinnati, Ohio, United States, 45202

Cincinnati, Ohio, United States, 45242

Columbus, Ohio, United States, 43215

Toledo, Ohio, United States, 43608

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Ashland, Oregon, United States, 97520

Portland, Oregon, United States, 97210

Portland, Oregon, United States, 97227

Salem, Oregon, United States, 97302

United States, Pennsylvania

Kingston, Pennsylvania, United States, 18704

Philadelphia, Pennsylvania, United States, 19104

Philadelphia, Pennsylvania, United States, 19107

Philadelphia, Pennsylvania, United States, 19124

Pittsberg, Pennsylvania, United States, 15231
Pittsburgh, Pennsylvania, United States, 15212
Pittsburgh, Pennsylvania, United States, 15213
West Mifflin, Pennsylvania, United States, 15122
Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

Charleston, South Carolina, United States, 29414
Columbia, South Carolina, United States, 29223
Greenville, South Carolina, United States, 29605
West Columbia, South Carolina, United States, 29169

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Memphis, Tennessee, United States, 38119
Memphis, Tennessee, United States, 38120
Nashville, Tennessee, United States, 37203

United States, Texas

Abilene, Texas, United States, 79606
Austin, Texas, United States, 78705
Corpus Cristi, Texas, United States, 78413
Dallas, Texas, United States, 75390
DeSoto, Texas, United States, 75115

Ft. Worth, Texas, United States, 76102

Ft. Worth, Texas, United States, 76104

Galveston, Texas, United States, 77555

Houston, Texas, United States, 77030

McAllen, Texas, United States, 78503

Odessa, Texas, United States, 79761

San Antonio, Texas, United States, 78240

United States, Utah

Salt Lake City, Utah, United States, 84107

Salt Lake City, Utah, United States, 84132

United States, Vermont

Burlington, Vermont, United States, 05401

United States, Virginia

Charlottesville, Virginia, United States, 22908

Fairfax, Virginia, United States, 22031

Richmond, Virginia, United States, 23221

United States, Washington

Seattle, Washington, United States, 98104

Silverdale, Washington, United States, 98383

United States, Wisconsin

Madison, Wisconsin, United States, 53715

Madison, Wisconsin, United States, 58705

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

Vancouver, British Columbia, Canada, V5Z 3N9

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

London, Ontario, Canada, N6A 4G5

Mississauga, Ontario, Canada, L4W 1W9

Ottawa, Ontario, Canada, K1H8L6

Toronto, Ontario, Canada, M4N3M5

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

Montreal, Quebec, Canada, H1T 2M4

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

History of Changes for Study: NCT00509795

Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

[Latest version \(submitted December 20, 2012\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: April 28, 2009 (v9)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Vascular Endothelial Growth Factor(VEGF)Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: April 2009

Overall Status: Recruiting

Study Start: August 2007

Primary Completion: December 2011 [Anticipated]

Study Completion: December 2011 [Anticipated]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that April 28, 2009

Met QC Criteria:

Last Update Posted: April 29, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	Drug: VEGF Trap-Eye 0.5 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 2	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Experimental: 3	Drug: VEGF Trap-Eye 2.0 mg VEGF Trap-Eye administered every 8 weeks (including one additional 2.0 mg dose at week 4) during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.
Active Comparator: 4	Drug: ranibizumab 0.5 mg administered every 4 weeks during the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.

Outcome Measures

Primary Outcome Measures:

1. The proportion of subjects who maintain vision at Week 52, where a subject is classified as maintaining vision if the subject has

lost fewer than 15 letters on the ETDRS chart compared to baseline (i.e. prevention of moderate vision loss)

Week 52

Secondary Outcome Measures:

2. Mean change from baseline in BCVA as measured by ETDRS letter score at Week 52

Week 52

3. The proportion of subjects who gain at least 15 letters of vision at Week 52

Week 52

4. Mean change from baseline in total NEI VFQ-25 score at Week 52

Week 52

5. Mean change from baseline in CNV area at Week 52

Week 52

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents in the study eye.
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Central Contact: Regeneron

Telephone: 866-549-8439

Email: VIEW1study@rtp.ppd.com

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Alabama**

[Recruiting]

Birmingham, Alabama, United States, 35205

[Recruiting]

Birmingham, Alabama, United States, 35223

United States, Arizona

[Recruiting]

Phoenix, Arizona, United States, 85014

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tucson, Arizona, United States, 85704

[Recruiting]

Tucson, Arizona, United States, 85710

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Recruiting]

Campbell, California, United States, 95008

[Recruiting]

Fullerton, California, United States, 92835

[Terminated]

Glendale, California, United States, 91203

[Recruiting]

Irvine, California, United States, 92697

[Recruiting]

La Jolla, California, United States, 92037

[Recruiting]

Loma Linda, California, United States, 92354

[Withdrawn]

Los Angeles, California, United States, 90033

[Recruiting]

Los Angeles, California, United States, 90048

[Recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Mountain View, California, United States, 94040

[Recruiting]

Oakland, California, United States, 94609

[Recruiting]

Palm Springs, California, United States, 92262

[Recruiting]

Pasadena, California, United States, 91105

[Withdrawn]

Poway, California, United States, 92064

[Recruiting]

Sacramento, California, United States, 95819

[Active, not recruiting]

San Diego, California, United States, 92120

[Terminated]

San Francisco, California, United States, 94107

[Recruiting]

Santa Ana, California, United States, 92705

[Recruiting]

Torrance, California, United States, 90503

[Withdrawn]

Ventura, California, United States, 93003

[Recruiting]

Westlake Village, California, United States, 91361

[Recruiting]

Yorba Linda, California, United States, 92887

United States, Colorado

[Terminated]

Aurora, Colorado, United States, 80045

[Recruiting]

Denver, Colorado, United States, 80205

[Withdrawn]

Denver, Colorado, United States, 80205

[Withdrawn]

Denver, Colorado, United States, 80230

United States, Connecticut

[Recruiting]

Bridgeport, Connecticut, United States, 06606

[Withdrawn]

Hamden, Connecticut, United States, 06518

[Active, not recruiting]

New Haven, Connecticut, United States, 06510

[Recruiting]

New London, Connecticut, United States, 06320

United States, Florida

[Recruiting]

Altamonte Springs, Florida, United States, 32701

[Recruiting]

Boynton Beach, Florida, United States, 33426

[Recruiting]

Fort Myers, Florida, United States, 33907

[Withdrawn]

Ft. Lauderdale, Florida, United States, 33351

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Withdrawn]

Gainesville, Florida, United States, 32610

[Recruiting]

Jacksonville, Florida, United States, 32224

[Withdrawn]

Miami, Florida, United States, 33136

[Recruiting]

Miami, Florida, United States, 33143

[Terminated]

Mount Dora, Florida, United States, 32757

[Recruiting]

Orlando, Florida, United States, 32803

[Recruiting]

Orlando, Florida, United States, 32806

[Recruiting]

Ocala, Florida, United States, 34472

[Recruiting]

Palm Beach Gardens, Florida, United States, 33410

[Recruiting]

Pensacola, Florida, United States, 32503

[Withdrawn]

Sarasota, Florida, United States

[Recruiting]

Stuart, Florida, United States, 34994

[Recruiting]

Tampa, Florida, United States, 33612

[Recruiting]

Winter Haven, Florida, United States, 33880

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

United States, Hawaii

[Recruiting]

Aiea, Hawaii, United States, 96701

[Withdrawn]

Honolulu, Hawaii, United States, 96813

United States, Illinois

[Recruiting]

Oak Brook, Illinois, United States, 60523

United States, Indiana

[Recruiting]

Fort Wayne, Indiana, United States, 46804

[Recruiting]

Indianapolis, Indiana, United States, 46202

[Terminated]

Indianapolis, Indiana, United States, 46260

[Recruiting]

Indianapolis, Indiana, United States, 46280

[Recruiting]

New Albany, Indiana, United States, 47150

United States, Iowa

[Recruiting]

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

[Recruiting]

Wichita, Kansas, United States, 67214

United States, Kentucky

[Recruiting]

Louisville, Kentucky, United States, 40202

[Recruiting]

Louisville, Kentucky, United States, 40207

[Terminated]

Paducah, Kentucky, United States, 42001

United States, Louisiana

[Recruiting]

New Orleans, Louisiana, United States, 70115

[Recruiting]

New Orleans, Louisiana, United States, 70121

[Terminated]

Shreveport, Louisiana, United States, 71105

United States, Maine

[Recruiting]

Bangor, Maine, United States, 04401

[Recruiting]

Portland, Maine, United States, 04102

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21209

[Recruiting]

Baltimore, Maryland, United States, 21287

[Recruiting]

Chevy Chase, Maryland, United States, 20815

[Recruiting]

Hagerstown, Maryland, United States, 21740

[Recruiting]

Towson, Maryland, United States, 21204

United States, Massachusetts

[Withdrawn]

Boston, Massachusetts, United States, 02111

[Recruiting]

Boston, Massachusetts, United States, 02114

[Recruiting]

Boston, Massachusetts, United States, 02215

[Withdrawn]

Boston, Massachusetts, United States

[Recruiting]

Peabody, Massachusetts, United States, 01960

United States, Michigan

[Recruiting]

Ann Arbor, Michigan, United States, 48105

[Recruiting]

Battle Creek, Michigan, United States, 49015

[Active, not recruiting]

Detroit, Michigan, United States, 48202

[Recruiting]

Grand Rapids, Michigan, United States, 49525

[Recruiting]

Jackson, Michigan, United States, 49201

[Recruiting]

Royal Oak, Michigan, United States, 48073

[Recruiting]

Southfield, Michigan, United States, 48034

[Withdrawn]

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

[Withdrawn]

Edina, Minnesota, United States, 55435

[Recruiting]

Minneapolis, Minnesota, United States, 55404

[Recruiting]

Rochester, Minnesota, United States, 55905

United States, Missouri

[Recruiting]

Florissant, Missouri, United States, 63031

[Recruiting]

Kansas City, Missouri, United States, 64108

[Withdrawn]

Kansas City, Missouri, United States, 64111

[Recruiting]

Springfield, Missouri, United States, 65804

[Recruiting]

St. Louis, Missouri, United States, 63110

United States, Montana

[Recruiting]

Missoula, Montana, United States, 59801

United States, Nebraska

[Recruiting]

Lincoln, Nebraska, United States, 68506

[Withdrawn]

Omaha, Nebraska, United States, 68131

United States, Nevada

[Recruiting]

Las Vegas, Nevada, United States, 89144

United States, New Jersey

[Recruiting]

Lawrenceville, New Jersey, United States, 08648

[Terminated]

New Brunswick, New Jersey, United States, 08901

[Recruiting]

Northfield, New Jersey, United States, 08225

[Withdrawn]

Teaneck, New Jersey, United States, 07666

[Recruiting]

Toms River, New Jersey, United States, 08753

United States, New Mexico

[Recruiting]

Albuquerque, New Mexico, United States, 87106

United States, New York

[Recruiting]

Albany, New York, United States, 12206

[Recruiting]

Brooklyn, New York, United States, 11223

[Recruiting]

Lynbrook, New York, United States, 11563

[Recruiting]

New York, New York, United States, 10003

[Recruiting]

New York, New York, United States, 10021

[Recruiting]

New York, New York, United States, 10032

[Terminated]

Poughkeepsie, New York, United States, 12601

[Recruiting]

Rochester, New York, United States, 14620

[Recruiting]

Rochester, New York, United States, 14642

[Recruiting]

Slingerlands, New York, United States, 12159

[Recruiting]

Syracuse, New York, United States, 13224

United States, North Carolina

[Recruiting]

Asheville, North Carolina, United States, 28803

[Recruiting]

Charlotte, North Carolina, United States, 28210

[Recruiting]

Raleigh, North Carolina, United States, 27607

[Terminated]

Southern Pines, North Carolina, United States, 28387

[Recruiting]

Winston-Salem, North Carolina, United States, 27157

United States, Ohio

[Withdrawn]

Cincinnati, Ohio, United States, 45202

[Recruiting]

Cincinnati, Ohio, United States, 45242

[Terminated]

Columbus, Ohio, United States, 43215

[Terminated]

Toledo, Ohio, United States, 43608

United States, Oklahoma

[Recruiting]

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

[Recruiting]

Ashland, Oregon, United States, 97520

[Recruiting]

Portland, Oregon, United States, 97210

[Recruiting]

Portland, Oregon, United States, 97227

[Recruiting]

Salem, Oregon, United States, 97302

United States, Pennsylvania

[Recruiting]

Kingston, Pennsylvania, United States, 18704

[Active, not recruiting]

Philadelphia, Pennsylvania, United States, 19104

[Recruiting]

Philadelphia, Pennsylvania, United States, 19107

[Recruiting]

Philadelphia, Pennsylvania, United States, 19124

[Recruiting]

Pittsberg, Pennsylvania, United States, 15231

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15212

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15213

[Recruiting]

West Mifflin, Pennsylvania, United States, 15122

[Withdrawn]

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

[Recruiting]

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

[Recruiting]

Charleston, South Carolina, United States, 29414

[Recruiting]

Columbia, South Carolina, United States, 29223

[Recruiting]

Greenville, South Carolina, United States, 29605

[Recruiting]

West Columbia, South Carolina, United States, 29169

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Terminated]

Memphis, Tennessee, United States, 38119

[Terminated]

Memphis, Tennessee, United States, 38120

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Abilene, Texas, United States, 79606

[Recruiting]

Austin, Texas, United States, 78705

[Active, not recruiting]

Corpus Cristi, Texas, United States, 78413

[Recruiting]

Dallas, Texas, United States, 75390

[Terminated]

DeSoto, Texas, United States, 75115

[Recruiting]

Ft. Worth, Texas, United States, 76102

[Recruiting]

Ft. Worth, Texas, United States, 76104

[Withdrawn]

Galveston, Texas, United States, 77555

[Recruiting]

Houston, Texas, United States, 77030

[Withdrawn]

Houston, Texas, United States, 77030

[Terminated]

Houston, Texas, United States, 77030

[Recruiting]

McAllen, Texas, United States, 78503

[Recruiting]

Odessa, Texas, United States, 79761

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Utah

[Recruiting]

Salt Lake City, Utah, United States, 84107

[Recruiting]

Salt Lake City, Utah, United States, 84132

United States, Vermont

[Recruiting]

Burlington, Vermont, United States, 05401

United States, Virginia

[Recruiting]

Charlottesville, Virginia, United States, 22908

[Recruiting]

Fairfax, Virginia, United States, 22031

[Recruiting]

Richmond, Virginia, United States, 23221

United States, Washington

[Recruiting]

Seattle, Washington, United States, 98104

[Recruiting]

Silverdale, Washington, United States, 98383

United States, Wisconsin

[Recruiting]

Madison, Wisconsin, United States, 53715

[Recruiting]

Madison, Wisconsin, United States, 58705

[Recruiting]

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

[Recruiting]

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

[Recruiting]

Vancouver, British Columbia, Canada, V5Z 3N9

[Recruiting]

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

[Recruiting]

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

[Recruiting]

London, Ontario, Canada, N6A 4G5

[Recruiting]

Mississauga, Ontario, Canada, L4W 1W9

[Recruiting]

Ottawa, Ontario, Canada, K1H8L6

[Recruiting]

Toronto, Ontario, Canada, M4N3M5

[Active, not recruiting]

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

[Recruiting]

Montreal, Quebec, Canada, H1T 2M4

[Recruiting]

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

[Recruiting]

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00509795

Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1) (VIEW1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: March 3, 2009 (v8)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: March 2009

Overall Status: Recruiting

Study Start: August 2007

Primary Completion: October 2010 [Anticipated]

Study Completion: January 2012 [Anticipated]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that March 3, 2009

Met QC Criteria:

Last Update Posted: March 5, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	<p data-bbox="829 512 1057 552">Drug: VEGF Trap-Eye</p> <p data-bbox="873 569 1500 1570">VGFT-OD-0605 is a double-masked, randomized, Phase III study. Subjects will be rand. assigned in a 1:1:1:1 ratio to 1 of 4 dosing regimens: 1) 2mg VEGF Trap-Eye adm. every 4 wks (2Q4), 2) 0.5mg VEGF Trap-Eye adm. every 4 wks (0.5Q4), 3) 2mg VEGF Trap-Eye adm. every 8 wks (2Q8), and 4) 0.5mg ranibizumab adm. every 4 wks (RQ4); subj. assigned to (2Q8) will receive the 2mg injection every 4 wks to wk 8 & then a sham inject at interim 4-wk visits (when study drug is not to be adm) during the first 52 wks of the study. VEGF Trap-Eye will be supplied in sealed 3mL single use vials each with a "withdrawable" vol. of approx. 0.5 mL at a concentration of 10mg/mL or 40 mg/mL. VEGF Trap-Eye will be withdrawn using aseptic technique through an 18 "gauge" needle attached to a 1 mL syringe. The syringe needle will then be removed and replaced with a 30 gauge needle to be used for the ITV inject. The inject. vol. will be 50µL (0.05 mL) for the 0.5mg & 2mg doses of VEGF Trap-Eye and 0.5 mg ranibizumab</p>

Arms	Assigned Interventions
Experimental: 2	<p data-bbox="829 191 1057 226">Drug: VEGF Trap-Eye</p> <p data-bbox="873 247 1500 1245"> VGFT-OD-0605 is a double-masked, randomized, Phase III study. Subjects will be rand. assigned in a 1:1:1:1 ratio to 1 of 4 dosing regimens: 1) 2mg VEGF Trap-Eye adm. every 4 wks (2Q4), 2) 0.5mg VEGF Trap-Eye adm. every 4 wks (0.5Q4), 3) 2mg VEGF Trap-Eye adm. every 8 wks (2Q8), and 4) 0.5mg ranibizumab adm. every 4 wks (RQ4); subj. assigned to (2Q8) will receive the 2mg injection every 4 wks to wk 8 & then a sham inject at interim 4-wk visits (when study drug is not to be adm) during the first 52 wks of the study. VEGF Trap-Eye will be supplied in sealed 3mL single use vials each with a "withdrawable" vol. of approx. 0.5 mL at a concentration of 10mg/mL or 40 mg/mL. VEGF Trap-Eye will be withdrawn using aseptic technique through an 18 "gauge" needle attached to a 1 mL syringe. The syringe needle will then be removed and replaced with a 30 gauge needle to be used for the ITV inject. The inject. vol. will be 50µL (0.05 mL) for the 0.5mg & 2mg doses of VEGF Trap-Eye and 0.5 mg ranibizumab </p>

Arms	Assigned Interventions
Experimental: 3	<p>Drug: VEGF Trap-Eye</p> <p>VGFT-OD-0605 is a double-masked, randomized, Phase III study. Subjects will be rand. assigned in a 1:1:1:1 ratio to 1 of 4 dosing regimens: 1) 2mg VEGF Trap-Eye adm. every 4 wks (2Q4), 2) 0.5mg VEGF Trap-Eye adm. every 4 wks (0.5Q4), 3) 2mg VEGF Trap-Eye adm. every 8 wks (2Q8), and 4) 0.5mg ranibizumab adm. every 4 wks (RQ4); subj. assigned to (2Q8) will receive the 2mg injection every 4 wks to wk 8 & then a sham inject at interim 4-wk visits (when study drug is not to be adm) during the first 52 wks of the study. VEGF Trap-Eye will be supplied in sealed 3mL single use vials each with a "withdrawable" vol. of approx. 0.5 mL at a concentration of 10mg/mL or 40 mg/mL. VEGF Trap-Eye will be withdrawn using aseptic technique through an 18 "gauge" needle attached to a 1 mL syringe. The syringe needle will then be removed and replaced with a 30 gauge needle to be used for the ITV inject. The inject. vol. will be 50µL (0.05 mL) for the 0.5mg & 2mg doses of VEGF Trap-Eye and 0.5 mg ranibizumab</p>
Active Comparator: 4	<p>Drug: Comparator</p> <p>0.5 mg ranibizumab adm. every 4 wks (RQ4);</p>

Outcome Measures

Primary Outcome Measures:

1. Primary measure will be visual acuity changes compared to baseline.
Monthly

Secondary Outcome Measures:

2. Secondary measures will be angiographic and anatomical changes compared to baseline.
as dictated by protocol

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: Yes

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. CNV must be at least 50% of total lesion size.
5. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
6. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
7. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Prior treatment with anti-VEGF agents as follows:
 - o Prior treatment with anti-VEGF therapy in the study eye is not allowed.
 - o Prior treatment with anti-VEGF therapy in the fellow eye with an investigational agent (not FDA approved, e.g. bevacizumab) is allowed up to 3 months prior to first dose in the study, and such treatment will not be allowed during the study. Prior treatment with an FDA/Health Canada approved anti-VEGF therapy in the fellow eye is allowed.

- o Prior systemic anti-VEGF therapy, investigational or FDA/Health Canada approved, is only allowed up to 3 months prior to first dose, and will not be allowed during the study
- 4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
- 5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
- 6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
- 7. Scar, fibrosis, or atrophy involving the center of the fovea.
- 8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
- 9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
- 10. Presence of other causes of CNV, including pathologic myopia (spherical equivalent of -8 diopters or more negative, or axial length of 25 mm or more), ocular histoplasmosis syndrome, angioid streaks, choroidal rupture, or multifocal choroiditis in the study eye.
- 11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
- 12. Prior vitrectomy in the study eye.
- 13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
- 14. Any history of macular hole of stage 2 and above in the study eye.
- 15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Central Contact: Regeneron

Telephone: 866-549-8439

Email: VIEW1study@rtp.ppd.com

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Alabama**

[Recruiting]

Birmingham, Alabama, United States, 35205

[Recruiting]

Birmingham, Alabama, United States, 35223

United States, Arizona

[Recruiting]

Phoenix, Arizona, United States, 85014

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tucson, Arizona, United States, 85704

[Recruiting]

Tucson, Arizona, United States, 85710

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Recruiting]

Campbell, California, United States, 95008

[Recruiting]

Fullerton, California, United States, 92835

[Terminated]

Glendale, California, United States, 91203

[Recruiting]

Irvine, California, United States, 92697

[Recruiting]

La Jolla, California, United States, 92037

[Recruiting]

Loma Linda, California, United States, 92354

[Withdrawn]

Los Angeles, California, United States, 90033

[Recruiting]

Los Angeles, California, United States, 90048

[Recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Mountain View, California, United States, 94040

[Recruiting]

Oakland, California, United States, 94609

[Recruiting]

Palm Springs, California, United States, 92262

[Recruiting]

Pasadena, California, United States, 91105

[Withdrawn]

Poway, California, United States, 92064

[Recruiting]

Sacramento, California, United States, 95819

[Active, not recruiting]

San Diego, California, United States, 92120

[Terminated]

San Francisco, California, United States, 94107

[Recruiting]

Santa Ana, California, United States, 92705

[Recruiting]

Torrance, California, United States, 90503

[Withdrawn]

Ventura, California, United States, 93003

[Recruiting]

Westlake Village, California, United States, 91361

[Recruiting]

Yorba Linda, California, United States, 92887

United States, Colorado

[Terminated]

Aurora, Colorado, United States, 80045

[Recruiting]

Denver, Colorado, United States, 80205

[Withdrawn]

Denver, Colorado, United States, 80205

[Withdrawn]

Denver, Colorado, United States, 80230

United States, Connecticut

[Recruiting]

Bridgeport, Connecticut, United States, 06606

[Withdrawn]

Hamden, Connecticut, United States, 06518

[Active, not recruiting]

New Haven, Connecticut, United States, 06510

[Recruiting]

New London, Connecticut, United States, 06320

United States, Florida

[Recruiting]

Altamonte Springs, Florida, United States, 32701

[Recruiting]

Boynton Beach, Florida, United States, 33426

[Recruiting]

Fort Myers, Florida, United States, 33907

[Withdrawn]

Ft. Lauderdale, Florida, United States, 33351

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Withdrawn]

Gainesville, Florida, United States, 32610

[Recruiting]

Jacksonville, Florida, United States, 32224

[Withdrawn]

Miami, Florida, United States, 33136

[Recruiting]

Miami, Florida, United States, 33143

[Terminated]

Mount Dora, Florida, United States, 32757

[Recruiting]

Orlando, Florida, United States, 32803

[Recruiting]

Orlando, Florida, United States, 32806

[Recruiting]

Ocala, Florida, United States, 34472

[Recruiting]

Palm Beach Gardens, Florida, United States, 33410

[Recruiting]

Pensacola, Florida, United States, 32503

[Withdrawn]

Sarasota, Florida, United States

[Recruiting]

Stuart, Florida, United States, 34994

[Recruiting]

Tampa, Florida, United States, 33612

[Recruiting]

Winter Haven, Florida, United States, 33880

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

United States, Hawaii

[Recruiting]

Aiea, Hawaii, United States, 96701

[Withdrawn]

Honolulu, Hawaii, United States, 96813

United States, Illinois

[Recruiting]

Oak Brook, Illinois, United States, 60523

United States, Indiana

[Recruiting]

Fort Wayne, Indiana, United States, 46804

[Recruiting]

Indianapolis, Indiana, United States, 46202

[Terminated]

Indianapolis, Indiana, United States, 46260

[Recruiting]

Indianapolis, Indiana, United States, 46280

[Recruiting]

New Albany, Indiana, United States, 47150

United States, Iowa

[Recruiting]

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

[Recruiting]

Wichita, Kansas, United States, 67214

United States, Kentucky

[Recruiting]

Louisville, Kentucky, United States, 40202

[Recruiting]

Louisville, Kentucky, United States, 40207

[Terminated]

Paducah, Kentucky, United States, 42001

United States, Louisiana

[Recruiting]

New Orleans, Louisiana, United States, 70115

[Recruiting]

New Orleans, Louisiana, United States, 70121

[Terminated]

Shreveport, Louisiana, United States, 71105

United States, Maine

[Recruiting]

Bangor, Maine, United States, 04401

[Recruiting]

Portland, Maine, United States, 04102

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21209

[Recruiting]

Baltimore, Maryland, United States, 21287

[Recruiting]

Chevy Chase, Maryland, United States, 20815

[Recruiting]

Hagerstown, Maryland, United States, 21740

[Recruiting]

Towson, Maryland, United States, 21204

United States, Massachusetts

[Withdrawn]

Boston, Massachusetts, United States, 02111

[Recruiting]

Boston, Massachusetts, United States, 02114

[Recruiting]

Boston, Massachusetts, United States, 02215

[Withdrawn]

Boston, Massachusetts, United States

[Recruiting]

Peabody, Massachusetts, United States, 01960

United States, Michigan

[Recruiting]

Ann Arbor, Michigan, United States, 48105

[Recruiting]

Battle Creek, Michigan, United States, 49015

[Active, not recruiting]

Detroit, Michigan, United States, 48202

[Recruiting]

Grand Rapids, Michigan, United States, 49525

[Recruiting]

Jackson, Michigan, United States, 49201

[Recruiting]

Royal Oak, Michigan, United States, 48073

[Recruiting]

Southfield, Michigan, United States, 48034

[Withdrawn]

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

[Withdrawn]

Edina, Minnesota, United States, 55435

[Recruiting]

Minneapolis, Minnesota, United States, 55404

[Recruiting]

Rochester, Minnesota, United States, 55905

United States, Missouri

[Recruiting]

Florissant, Missouri, United States, 63031

[Recruiting]

Kansas City, Missouri, United States, 64108

[Withdrawn]

Kansas City, Missouri, United States, 64111

[Recruiting]

Springfield, Missouri, United States, 65804

[Recruiting]

St. Louis, Missouri, United States, 63110

United States, Montana

[Recruiting]

Missoula, Montana, United States, 59801

United States, Nebraska

[Recruiting]

Lincoln, Nebraska, United States, 68506

[Withdrawn]

Omaha, Nebraska, United States, 68131

United States, Nevada

[Recruiting]

Las Vegas, Nevada, United States, 89144

United States, New Jersey

[Recruiting]

Lawrenceville, New Jersey, United States, 08648

[Terminated]

New Brunswick, New Jersey, United States, 08901

[Recruiting]

Northfield, New Jersey, United States, 08225

[Withdrawn]

Teaneck, New Jersey, United States, 07666

[Recruiting]

Toms River, New Jersey, United States, 08753

United States, New Mexico

[Recruiting]

Albuquerque, New Mexico, United States, 87106

United States, New York

[Recruiting]

Albany, New York, United States, 12206

[Recruiting]

Brooklyn, New York, United States, 11223

[Recruiting]

Lynbrook, New York, United States, 11563

[Recruiting]

New York, New York, United States, 10003

[Recruiting]

New York, New York, United States, 10021

[Recruiting]

New York, New York, United States, 10032

[Terminated]

Poughkeepsie, New York, United States, 12601

[Recruiting]

Rochester, New York, United States, 14620

[Recruiting]

Rochester, New York, United States, 14642

[Recruiting]

Slingerlands, New York, United States, 12159

[Recruiting]

Syracuse, New York, United States, 13224

United States, North Carolina

[Recruiting]

Asheville, North Carolina, United States, 28803

[Recruiting]

Charlotte, North Carolina, United States, 28210

[Recruiting]

Raleigh, North Carolina, United States, 27607

[Terminated]

Southern Pines, North Carolina, United States, 28387

[Recruiting]

Winston-Salem, North Carolina, United States, 27157

United States, Ohio

[Withdrawn]

Cincinnati, Ohio, United States, 45202

[Recruiting]

Cincinnati, Ohio, United States, 45242

[Terminated]

Columbus, Ohio, United States, 43215

[Terminated]

Toledo, Ohio, United States, 43608

United States, Oklahoma

[Recruiting]

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

[Recruiting]

Ashland, Oregon, United States, 97520

[Recruiting]

Portland, Oregon, United States, 97210

[Recruiting]

Portland, Oregon, United States, 97227

[Recruiting]

Salem, Oregon, United States, 97302

United States, Pennsylvania

[Recruiting]

Kingston, Pennsylvania, United States, 18704

[Active, not recruiting]

Philadelphia, Pennsylvania, United States, 19104

[Recruiting]

Philadelphia, Pennsylvania, United States, 19107

[Recruiting]

Philadelphia, Pennsylvania, United States, 19124

[Recruiting]

Pittsberg, Pennsylvania, United States, 15231

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15212

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15213

[Recruiting]

West Mifflin, Pennsylvania, United States, 15122

[Withdrawn]

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

[Recruiting]

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

[Recruiting]

Charleston, South Carolina, United States, 29414

[Recruiting]

Columbia, South Carolina, United States, 29223

[Recruiting]

Greenville, South Carolina, United States, 29605

[Recruiting]

West Columbia, South Carolina, United States, 29169

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Terminated]

Memphis, Tennessee, United States, 38119

[Terminated]

Memphis, Tennessee, United States, 38120

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Abilene, Texas, United States, 79606

[Recruiting]

Austin, Texas, United States, 78705

[Active, not recruiting]

Corpus Cristi, Texas, United States, 78413

[Recruiting]

Dallas, Texas, United States, 75390

[Terminated]

DeSoto, Texas, United States, 75115

[Recruiting]

Ft. Worth, Texas, United States, 76102

[Recruiting]

Ft. Worth, Texas, United States, 76104

[Withdrawn]

Galveston, Texas, United States, 77555

[Recruiting]

Houston, Texas, United States, 77030

[Withdrawn]

Houston, Texas, United States, 77030

[Terminated]

Houston, Texas, United States, 77030

[Recruiting]

McAllen, Texas, United States, 78503

[Recruiting]

Odessa, Texas, United States, 79761

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Utah

[Recruiting]

Salt Lake City, Utah, United States, 84107

[Recruiting]

Salt Lake City, Utah, United States, 84132

United States, Vermont

[Recruiting]

Burlington, Vermont, United States, 05401

United States, Virginia

[Recruiting]

Charlottesville, Virginia, United States, 22908

[Recruiting]

Fairfax, Virginia, United States, 22031

[Recruiting]

Richmond, Virginia, United States, 23221

United States, Washington

[Recruiting]

Seattle, Washington, United States, 98104

[Recruiting]

Silverdale, Washington, United States, 98383

United States, Wisconsin

[Recruiting]

Madison, Wisconsin, United States, 53715

[Recruiting]

Madison, Wisconsin, United States, 58705

[Recruiting]

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

[Recruiting]

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

[Recruiting]

Vancouver, British Columbia, Canada, V5Z 3N9

[Recruiting]

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

[Recruiting]

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

[Recruiting]

London, Ontario, Canada, N6A 4G5

[Recruiting]

Mississauga, Ontario, Canada, L4W 1W9

[Recruiting]

Ottawa, Ontario, Canada, K1H8L6

[Recruiting]

Toronto, Ontario, Canada, M4N3M5

[Active, not recruiting]

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

[Recruiting]

Montreal, Quebec, Canada, H1T 2M4

[Recruiting]

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

[Recruiting]

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00509795

Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1) (VIEW1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: January 22, 2009 (v7)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: January 2009

Overall Status: Recruiting

Study Start: August 2007

Primary Completion: October 2010 [Anticipated]

Study Completion: January 2012 [Anticipated]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that January 22, 2009

Met QC Criteria:

Last Update Posted: January 26, 2009 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	<p data-bbox="829 512 1057 554">Drug: VEGF Trap-Eye</p> <p data-bbox="873 569 1500 1570">VGFT-OD-0605 is a double-masked, randomized, Phase III study. Subjects will be rand. assigned in a 1:1:1:1 ratio to 1 of 4 dosing regimens: 1) 2mg VEGF Trap-Eye adm. every 4 wks (2Q4), 2) 0.5mg VEGF Trap-Eye adm. every 4 wks (0.5Q4), 3) 2mg VEGF Trap-Eye adm. every 8 wks (2Q8), and 4) 0.5mg ranibizumab adm. every 4 wks (RQ4); subj. assigned to (2Q8) will receive the 2mg injection every 4 wks to wk 8 & then a sham inject at interim 4-wk visits (when study drug is not to be adm) during the first 52 wks of the study. VEGF Trap-Eye will be supplied in sealed 3mL single use vials each with a "withdrawable" vol. of approx. 0.5 mL at a concentration of 10mg/mL or 40 mg/mL. VEGF Trap-Eye will be withdrawn using aseptic technique through an 18 "gauge" needle attached to a 1 mL syringe. The syringe needle will then be removed and replaced with a 30 gauge needle to be used for the ITV inject. The inject. vol. will be 50µL (0.05 mL) for the 0.5mg & 2mg doses of VEGF Trap-Eye and 0.5 mg ranibizumab</p>

Arms	Assigned Interventions
Experimental: 2	<p data-bbox="829 191 1057 226">Drug: VEGF Trap-Eye</p> <p data-bbox="873 247 1500 1247"> VGFT-OD-0605 is a double-masked, randomized, Phase III study. Subjects will be rand. assigned in a 1:1:1:1 ratio to 1 of 4 dosing regimens: 1) 2mg VEGF Trap-Eye adm. every 4 wks (2Q4), 2) 0.5mg VEGF Trap-Eye adm. every 4 wks (0.5Q4), 3) 2mg VEGF Trap-Eye adm. every 8 wks (2Q8), and 4) 0.5mg ranibizumab adm. every 4 wks (RQ4); subj. assigned to (2Q8) will receive the 2mg injection every 4 wks to wk 8 & then a sham inject at interim 4-wk visits (when study drug is not to be adm) during the first 52 wks of the study. VEGF Trap-Eye will be supplied in sealed 3mL single use vials each with a "withdrawable" vol. of approx. 0.5 mL at a concentration of 10mg/mL or 40 mg/mL. VEGF Trap-Eye will be withdrawn using aseptic technique through an 18 "gauge" needle attached to a 1 mL syringe. The syringe needle will then be removed and replaced with a 30 gauge needle to be used for the ITV inject. The inject. vol. will be 50µL (0.05 mL) for the 0.5mg & 2mg doses of VEGF Trap-Eye and 0.5 mg ranibizumab </p>

Arms	Assigned Interventions
Experimental: 3	<p>Drug: VEGF Trap-Eye</p> <p>VGFT-OD-0605 is a double-masked, randomized, Phase III study. Subjects will be rand. assigned in a 1:1:1:1 ratio to 1 of 4 dosing regimens: 1) 2mg VEGF Trap-Eye adm. every 4 wks (2Q4), 2) 0.5mg VEGF Trap-Eye adm. every 4 wks (0.5Q4), 3) 2mg VEGF Trap-Eye adm. every 8 wks (2Q8), and 4) 0.5mg ranibizumab adm. every 4 wks (RQ4); subj. assigned to (2Q8) will receive the 2mg injection every 4 wks to wk 8 & then a sham inject at interim 4-wk visits (when study drug is not to be adm) during the first 52 wks of the study. VEGF Trap-Eye will be supplied in sealed 3mL single use vials each with a "withdrawable" vol. of approx. 0.5 mL at a concentration of 10mg/mL or 40 mg/mL. VEGF Trap-Eye will be withdrawn using aseptic technique through an 18 "gauge" needle attached to a 1 mL syringe. The syringe needle will then be removed and replaced with a 30 gauge needle to be used for the ITV inject. The inject. vol. will be 50µL (0.05 mL) for the 0.5mg & 2mg doses of VEGF Trap-Eye and 0.5 mg ranibizumab</p>
Active Comparator: 4	<p>Drug: Comparator</p> <p>0.5 mg ranibizumab adm. every 4 wks (RQ4);</p>

Outcome Measures

Primary Outcome Measures:

1. Primary measure will be visual acuity changes compared to baseline.

Monthly

Secondary Outcome Measures:

2. Secondary measures will be angiographic and anatomical changes compared to baseline.

as dictated by protocol

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: Yes

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. CNV must be at least 50% of total lesion size.
5. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
6. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
7. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Prior treatment with anti-VEGF agents as follows:
 - o Prior treatment with anti-VEGF therapy in the study eye is not allowed.
 - o Prior treatment with anti-VEGF therapy in the fellow eye with an investigational agent (not FDA approved, e.g. bevacizumab) is allowed up to 3 months prior to first dose in the study, and such treatment will not be allowed during the study. Prior treatment with an FDA/Health Canada approved anti-VEGF therapy in the fellow eye is allowed.

- o Prior systemic anti-VEGF therapy, investigational or FDA/Health Canada approved, is only allowed up to 3 months prior to first dose, and will not be allowed during the study
- 4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
- 5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
- 6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
- 7. Scar, fibrosis, or atrophy involving the center of the fovea.
- 8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
- 9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
- 10. Presence of other causes of CNV, including pathologic myopia (spherical equivalent of -8 diopters or more negative, or axial length of 25 mm or more), ocular histoplasmosis syndrome, angioid streaks, choroidal rupture, or multifocal choroiditis in the study eye.
- 11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
- 12. Prior vitrectomy in the study eye.
- 13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
- 14. Any history of macular hole of stage 2 and above in the study eye.
- 15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Central Contact: Regeneron

Telephone: 866-549-8439

Email: VIEW1study@rtp.ppd.com

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Alabama**

[Recruiting]

Birmingham, Alabama, United States, 35205

[Recruiting]

Birmingham, Alabama, United States, 35223

United States, Arizona

[Recruiting]

Phoenix, Arizona, United States, 85014

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tucson, Arizona, United States, 85704

[Recruiting]

Tucson, Arizona, United States, 85710

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Recruiting]

Campbell, California, United States, 95008

[Recruiting]

Fullerton, California, United States, 92835

[Terminated]

Glendale, California, United States, 91203

[Recruiting]

Irvine, California, United States, 92697

[Recruiting]

La Jolla, California, United States, 92037

[Recruiting]

Loma Linda, California, United States, 92354

[Withdrawn]

Los Angeles, California, United States, 90033

[Recruiting]

Los Angeles, California, United States, 90048

[Recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Mountain View, California, United States, 94040

[Recruiting]

Oakland, California, United States, 94609

[Recruiting]

Palm Springs, California, United States, 92262

[Recruiting]

Pasadena, California, United States, 91105

[Withdrawn]

Poway, California, United States, 92064

[Recruiting]

Sacramento, California, United States, 95819

[Active, not recruiting]

San Diego, California, United States, 92120

[Terminated]

San Francisco, California, United States, 94107

[Recruiting]

Santa Ana, California, United States, 92705

[Active, not recruiting]

Torrance, California, United States, 90503

[Withdrawn]

Ventura, California, United States, 93003

[Recruiting]

Westlake Village, California, United States, 91361

[Recruiting]

Yorba Linda, California, United States, 92887

United States, Colorado

[Terminated]

Aurora, Colorado, United States, 80045

[Recruiting]

Denver, Colorado, United States, 80205

[Withdrawn]

Denver, Colorado, United States, 80205

[Withdrawn]

Denver, Colorado, United States, 80230

United States, Connecticut

[Recruiting]

Bridgeport, Connecticut, United States, 06606

[Withdrawn]

Hamden, Connecticut, United States, 06518

[Active, not recruiting]

New Haven, Connecticut, United States, 06510

[Recruiting]

New London, Connecticut, United States, 06320

United States, Florida

[Recruiting]

Altamonte Springs, Florida, United States, 32701

[Recruiting]

Boynton Beach, Florida, United States, 33426

[Recruiting]

Fort Myers, Florida, United States, 33907

[Withdrawn]

Ft. Lauderdale, Florida, United States, 33351

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Withdrawn]

Gainesville, Florida, United States, 32610

[Recruiting]

Jacksonville, Florida, United States, 32224

[Withdrawn]

Miami, Florida, United States, 33136

[Recruiting]

Miami, Florida, United States, 33143

[Recruiting]

Mount Dora, Florida, United States, 32757

[Recruiting]

Orlando, Florida, United States, 32803

[Recruiting]

Orlando, Florida, United States, 32806

[Recruiting]

Ocala, Florida, United States, 34472

[Recruiting]

Palm Beach Gardens, Florida, United States, 33410

[Recruiting]

Pensacola, Florida, United States, 32503

[Withdrawn]

Sarasota, Florida, United States

[Recruiting]

Stuart, Florida, United States, 34994

[Recruiting]

Tampa, Florida, United States, 33612

[Recruiting]

Winter Haven, Florida, United States, 33880

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

United States, Hawaii

[Recruiting]

Aiea, Hawaii, United States, 96701

[Withdrawn]

Honolulu, Hawaii, United States, 96813

United States, Illinois

[Recruiting]

Oak Brook, Illinois, United States, 60523

United States, Indiana

[Recruiting]

Fort Wayne, Indiana, United States, 46804

[Recruiting]

Indianapolis, Indiana, United States, 46202

[Terminated]

Indianapolis, Indiana, United States, 46260

[Recruiting]

Indianapolis, Indiana, United States, 46280

[Recruiting]

New Albany, Indiana, United States, 47150

United States, Iowa

[Recruiting]

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

[Recruiting]

Wichita, Kansas, United States, 67214

United States, Kentucky

[Recruiting]

Louisville, Kentucky, United States, 40202

[Recruiting]

Louisville, Kentucky, United States, 40207

[Terminated]

Paducah, Kentucky, United States, 42001

United States, Louisiana

[Recruiting]

New Orleans, Louisiana, United States, 70115

[Recruiting]

New Orleans, Louisiana, United States, 70121

[Terminated]

Shreveport, Louisiana, United States, 71105

United States, Maine

[Recruiting]

Bangor, Maine, United States, 04401

[Recruiting]

Portland, Maine, United States, 04102

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21209

[Recruiting]

Baltimore, Maryland, United States, 21287

[Recruiting]

Chevy Chase, Maryland, United States, 20815

[Recruiting]

Hagerstown, Maryland, United States, 21740

[Recruiting]

Towson, Maryland, United States, 21204

United States, Massachusetts

[Withdrawn]

Boston, Massachusetts, United States, 02111

[Recruiting]

Boston, Massachusetts, United States, 02114

[Recruiting]

Boston, Massachusetts, United States, 02215

[Withdrawn]

Boston, Massachusetts, United States

[Recruiting]

Peabody, Massachusetts, United States, 01960

United States, Michigan

[Recruiting]

Ann Arbor, Michigan, United States, 48105

[Recruiting]

Battle Creek, Michigan, United States, 49015

[Active, not recruiting]

Detroit, Michigan, United States, 48202

[Recruiting]

Grand Rapids, Michigan, United States, 49525

[Recruiting]

Jackson, Michigan, United States, 49201

[Recruiting]

Royal Oak, Michigan, United States, 48073

[Recruiting]

Southfield, Michigan, United States, 48034

[Withdrawn]

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

[Withdrawn]

Edina, Minnesota, United States, 55435

[Recruiting]

Minneapolis, Minnesota, United States, 55404

[Recruiting]

Rochester, Minnesota, United States, 55905

United States, Missouri

[Recruiting]

Florissant, Missouri, United States, 63031

[Recruiting]

Kansas City, Missouri, United States, 64108

[Withdrawn]

Kansas City, Missouri, United States, 64111

[Recruiting]

Springfield, Missouri, United States, 65804

[Recruiting]

St. Louis, Missouri, United States, 63110

United States, Montana

[Recruiting]

Missoula, Montana, United States, 59801

United States, Nebraska

[Recruiting]

Lincoln, Nebraska, United States, 68506

[Withdrawn]

Omaha, Nebraska, United States, 68131

United States, Nevada

[Recruiting]

Las Vegas, Nevada, United States, 89144

United States, New Jersey

[Recruiting]

Lawrenceville, New Jersey, United States, 08648

[Terminated]

New Brunswick, New Jersey, United States, 08901

[Recruiting]

Northfield, New Jersey, United States, 08225

[Withdrawn]

Teaneck, New Jersey, United States, 07666

[Recruiting]

Toms River, New Jersey, United States, 08753

United States, New Mexico

[Recruiting]

Albuquerque, New Mexico, United States, 87106

United States, New York

[Recruiting]

Albany, New York, United States, 12206

[Recruiting]

Brooklyn, New York, United States, 11223

[Recruiting]

Lynbrook, New York, United States, 11563

[Recruiting]

New York, New York, United States, 10003

[Recruiting]

New York, New York, United States, 10021

[Recruiting]

New York, New York, United States, 10032

[Terminated]

Poughkeepsie, New York, United States, 12601

[Recruiting]

Rochester, New York, United States, 14620

[Recruiting]

Rochester, New York, United States, 14642

[Recruiting]

Slingerlands, New York, United States, 12159

[Recruiting]

Syracuse, New York, United States, 13224

United States, North Carolina

[Recruiting]

Asheville, North Carolina, United States, 28803

[Recruiting]

Charlotte, North Carolina, United States, 28210

[Recruiting]

Raleigh, North Carolina, United States, 27607

[Terminated]

Southern Pines, North Carolina, United States, 28387

[Recruiting]

Winston-Salem, North Carolina, United States, 27157

United States, Ohio

[Withdrawn]

Cincinnati, Ohio, United States, 45202

[Recruiting]

Cincinnati, Ohio, United States, 45242

[Terminated]

Columbus, Ohio, United States, 43215

[Recruiting]

Toledo, Ohio, United States, 43608

United States, Oklahoma

[Recruiting]

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

[Recruiting]

Ashland, Oregon, United States, 97520

[Recruiting]

Portland, Oregon, United States, 97210

[Recruiting]

Portland, Oregon, United States, 97227

[Recruiting]

Salem, Oregon, United States, 97302

United States, Pennsylvania

[Recruiting]

Kingston, Pennsylvania, United States, 18704

[Active, not recruiting]

Philadelphia, Pennsylvania, United States, 19104

[Recruiting]

Philadelphia, Pennsylvania, United States, 19107

[Recruiting]

Philadelphia, Pennsylvania, United States, 19124

[Recruiting]

Pittsberg, Pennsylvania, United States, 15231

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15212

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15213

[Recruiting]

West Mifflin, Pennsylvania, United States, 15122

[Withdrawn]

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

[Recruiting]

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

[Recruiting]

Charleston, South Carolina, United States, 29414

[Recruiting]

Columbia, South Carolina, United States, 29223

[Recruiting]

Greenville, South Carolina, United States, 29605

[Recruiting]

West Columbia, South Carolina, United States, 29169

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Recruiting]

Memphis, Tennessee, United States, 38119

[Terminated]

Memphis, Tennessee, United States, 38120

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Abilene, Texas, United States, 79606

[Recruiting]

Austin, Texas, United States, 78705

[Active, not recruiting]

Corpus Cristi, Texas, United States, 78413

[Recruiting]

Dallas, Texas, United States, 75390

[Terminated]

DeSoto, Texas, United States, 75115

[Recruiting]

Ft. Worth, Texas, United States, 76102

[Recruiting]

Ft. Worth, Texas, United States, 76104

[Withdrawn]

Galveston, Texas, United States, 77555

[Recruiting]

Houston, Texas, United States, 77030

[Withdrawn]

Houston, Texas, United States, 77030

[Terminated]

Houston, Texas, United States, 77030

[Recruiting]

McAllen, Texas, United States, 78503

[Recruiting]

Odessa, Texas, United States, 79761

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Utah

[Recruiting]

Salt Lake City, Utah, United States, 84107

[Recruiting]

Salt Lake City, Utah, United States, 84132

United States, Vermont

[Recruiting]

Burlington, Vermont, United States, 05401

United States, Virginia

[Recruiting]

Charlottesville, Virginia, United States, 22908

[Recruiting]

Fairfax, Virginia, United States, 22031

[Recruiting]

Richmond, Virginia, United States, 23221

United States, Washington

[Recruiting]

Seattle, Washington, United States, 98104

[Recruiting]

Silverdale, Washington, United States, 98383

United States, Wisconsin

[Recruiting]

Madison, Wisconsin, United States, 53715

[Recruiting]

Madison, Wisconsin, United States, 58705

[Recruiting]

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

[Recruiting]

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

[Recruiting]

Vancouver, British Columbia, Canada, V5Z 3N9

[Recruiting]

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

[Recruiting]

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

[Recruiting]

London, Ontario, Canada, N6A 4G5

[Recruiting]

Mississauga, Ontario, Canada, L4W 1W9

[Recruiting]

Ottawa, Ontario, Canada, K1H8L6

[Active, not recruiting]

Toronto, Ontario, Canada, M4N3M5

[Active, not recruiting]

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

[Recruiting]

Montreal, Quebec, Canada, H1T 2M4

[Recruiting]

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

[Recruiting]

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00509795

Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1) (VIEW1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: June 26, 2008 (v6)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: June 2008

Overall Status: Recruiting

Study Start: August 2007

Primary Completion:

Study Completion:

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that June 26, 2008

Met QC Criteria:

Last Update Posted: June 30, 2008 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Arms	Assigned Interventions
Experimental: 1	Biological: VEGF Trap-Eye
Experimental: 2	Biological: VEGF Trap-Eye
Experimental: 3	Biological: VEGF Trap-Eye
Active Comparator: 4	Drug: Comparator

Outcome Measures

Primary Outcome Measures:

1. Primary measure will be visual acuity changes compared to baseline.

Secondary Outcome Measures:

2. Secondary measures will be angiographic and anatomical changes compared to baseline.

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.

3. Active primary subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. CNV must be at least 50% of total lesion size.
5. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
6. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
7. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Prior treatment with anti-VEGF agents as follows:
 - o Prior treatment with anti-VEGF therapy in the study eye is not allowed.
 - o Prior treatment with anti-VEGF therapy in the fellow eye with an investigational agent (not FDA approved, e.g. bevacizumab) is allowed up to 3 months prior to first dose in the study, and such treatment will not be allowed during the study. Prior treatment with an FDA/Health Canada approved anti-VEGF therapy in the fellow eye is allowed.
 - o Prior systemic anti-VEGF therapy, investigational or FDA/Health Canada approved, is only allowed up to 3 months prior to first dose, and will not be allowed during the study
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.

10. Presence of other causes of CNV, including pathologic myopia (spherical equivalent of -8 diopters or more negative, or axial length of 25 mm or more), ocular histoplasmosis syndrome, angioid streaks, choroidal rupture, or multifocal choroiditis in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Central Contact: Regeneron

Telephone: 866-549-8439

Email: VIEW1study@rtp.ppd.com

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Alabama**

[Recruiting]

Birmingham, Alabama, United States, 35205

[Recruiting]

Birmingham, Alabama, United States, 35223

United States, Arizona

[Recruiting]

Phoenix, Arizona, United States, 85014

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tucson, Arizona, United States, 85704

[Recruiting]

Tucson, Arizona, United States, 85710

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Recruiting]

Campbell, California, United States, 95008

[Recruiting]

Fullerton, California, United States, 92835

[Not yet recruiting]

Glendale, California, United States, 91203

[Recruiting]

Irvine, California, United States, 92697

[Recruiting]

La Jolla, California, United States, 92037

[Recruiting]

Loma Linda, California, United States, 92354

[Not yet recruiting]

Los Angeles, California, United States, 90033

[Not yet recruiting]

Los Angeles, California, United States, 90048

[Recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Mountain View, California, United States, 94040

[Recruiting]

Oakland, California, United States, 94609

[Recruiting]

Orange, California, United States, 92868

[Recruiting]

Palm Springs, California, United States, 92262

[Recruiting]

Pasadena, California, United States, 91105

[Not yet recruiting]

Poway, California, United States, 92064

[Recruiting]

Sacramento, California, United States, 95819

[Not yet recruiting]

San Diego, California, United States, 92120

[Recruiting]

San Francisco, California, United States, 94107

[Recruiting]

Santa Ana, California, United States, 92705

[Not yet recruiting]

Ventura, California, United States, 93003

[Recruiting]

Westlake Village, California, United States, 91361

[Not yet recruiting]

Yorba Linda, California, United States, 92887

United States, Colorado

[Recruiting]

Aurora, Colorado, United States, 80045

[Recruiting]

Denver, Colorado, United States, 80205

United States, Connecticut

[Recruiting]

Bridgeport, Connecticut, United States, 06606

[Not yet recruiting]

New Haven, Connecticut, United States, 06510

[Recruiting]

New London, Connecticut, United States, 06320

United States, Florida

[Recruiting]

Altamonte Springs, Florida, United States, 32701

[Recruiting]

Boynton Beach, Florida, United States, 33426

[Recruiting]

Fort Myers, Florida, United States, 33907

[Not yet recruiting]

Ft. Lauderdale, Florida, United States, 33351

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Not yet recruiting]

Gainesville, Florida, United States, 32610

[Recruiting]

Jacksonville, Florida, United States, 32224

[Not yet recruiting]

Miami, Florida, United States, 33136

[Not yet recruiting]

Miami, Florida, United States, 33143

[Not yet recruiting]

Mount Dora, Florida, United States, 32757

[Recruiting]

Orlando, Florida, United States, 32803

[Recruiting]

Orlando, Florida, United States, 32806

[Recruiting]

Oscala, Florida, United States, 34472

[Recruiting]

Palm Beach Gardens, Florida, United States, 33410

[Recruiting]

Pensacola, Florida, United States, 32503

[Recruiting]

Stuart, Florida, United States, 34994

[Recruiting]

Tampa, Florida, United States, 33612

[Recruiting]

Winter Haven, Florida, United States, 33880

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

United States, Hawaii

[Recruiting]

Aiea, Hawaii, United States, 96701

United States, Illinois

[Suspended]

Chicago, Illinois, United States, 60637

[Recruiting]

Glenview, Illinois, United States, 90026

[Recruiting]

Oak Brook, Illinois, United States, 60523

United States, Indiana

[Recruiting]

Fort Wayne, Indiana, United States, 46804

[Recruiting]

Indianapolis, Indiana, United States, 46202

[Recruiting]

Indianapolis, Indiana, United States, 46260

[Recruiting]

Indianapolis, Indiana, United States, 46280

[Recruiting]

New Albany, Indiana, United States, 47150

United States, Iowa

[Recruiting]

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

[Recruiting]

Wichita, Kansas, United States, 67214

United States, Kentucky

[Recruiting]

Louisville, Kentucky, United States, 40202

[Recruiting]

Louisville, Kentucky, United States, 40207

[Recruiting]

Paducah, Kentucky, United States, 42001

United States, Louisiana

[Recruiting]

New Orleans, Louisiana, United States, 70115

[Recruiting]

New Orleans, Louisiana, United States, 70121

[Recruiting]

Shreveport, Louisiana, United States, 71105

United States, Maine

[Recruiting]

Bangor, Maine, United States, 04401

[Recruiting]

Portland, Maine, United States, 04102

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21209

[Recruiting]

Baltimore, Maryland, United States, 21287

[Recruiting]

Chevy Chase, Maryland, United States, 20815

[Recruiting]

Hagerstown, Maryland, United States, 21740

[Recruiting]

Towson, Maryland, United States, 21204

United States, Massachusetts

[Not yet recruiting]

Boston, Massachusetts, United States, 02111

[Recruiting]

Boston, Massachusetts, United States, 02114

[Not yet recruiting]

Boston, Massachusetts, United States, 02215

[Recruiting]

Peabody, Massachusetts, United States, 01960

United States, Michigan

[Recruiting]

Ann Arbor, Michigan, United States, 48105

[Recruiting]

Battle Creek, Michigan, United States, 49015

[Not yet recruiting]

Detroit, Michigan, United States, 48202

[Recruiting]

Grand Rapids, Michigan, United States, 49525

[Recruiting]

Jackson, Michigan, United States, 49201

[Recruiting]

Royal Oak, Michigan, United States, 48073

[Recruiting]

Southfield, Michigan, United States, 48034

United States, Minnesota

[Recruiting]

Minneapolis, Minnesota, United States, 55404

[Recruiting]

Rochester, Minnesota, United States, 55905

United States, Missouri

[Recruiting]

Florissant, Missouri, United States, 63031

[Recruiting]

Kansas City, Missouri, United States, 64108

[Recruiting]

Springfield, Missouri, United States, 65804

[Recruiting]

St. Louis, Missouri, United States, 63110

United States, Montana

[Recruiting]

Missoula, Montana, United States, 59801

United States, Nebraska

[Recruiting]

Lincoln, Nebraska, United States, 68506

United States, Nevada

[Recruiting]

Las Vegas, Nevada, United States, 89144

United States, New Jersey

[Recruiting]

Lawrenceville, New Jersey, United States, 08648

[Recruiting]

New Brunswick, New Jersey, United States, 08901

[Recruiting]

Northfield, New Jersey, United States, 08225

[Recruiting]

Toms River, New Jersey, United States, 08753

United States, New Mexico

[Recruiting]

Albuquerque, New Mexico, United States, 87106

United States, New York

[Recruiting]

Albany, New York, United States, 12159

[Recruiting]

Albany, New York, United States, 12206

[Recruiting]

Brooklyn, New York, United States, 11223

[Recruiting]

Lynbrook, New York, United States, 11563

[Recruiting]

New York, New York, United States, 10003

[Recruiting]

New York, New York, United States, 10021

[Recruiting]

New York, New York, United States, 10032

[Recruiting]

Poughkeepsie, New York, United States, 12601

[Recruiting]

Rochester, New York, United States, 14620

[Recruiting]

Rochester, New York, United States, 14642

[Recruiting]

Slingerlands, New York, United States, 12159

United States, North Carolina

[Recruiting]

Asheville, North Carolina, United States, 28803

[Recruiting]

Charlotte, North Carolina, United States, 28210

[Recruiting]

Raleigh, North Carolina, United States, 27607

[Recruiting]

Southern Pines, North Carolina, United States, 28387

[Recruiting]

Winston-Salem, North Carolina, United States, 27157

United States, Ohio

[Not yet recruiting]

Cincinnati, Ohio, United States, 45202

[Recruiting]

Cincinnati, Ohio, United States, 45242

[Recruiting]

Columbus, Ohio, United States, 43215

[Recruiting]

Toledo, Ohio, United States, 43608

United States, Oklahoma

[Recruiting]

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

[Not yet recruiting]

Ashland, Oregon, United States, 97520

[Recruiting]

Portland, Oregon, United States, 97210

[Recruiting]

Portland, Oregon, United States, 97227

[Recruiting]

Salem, Oregon, United States, 97302

United States, Pennsylvania

[Recruiting]

Kingston, Pennsylvania, United States, 18704

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19104

[Recruiting]

Philadelphia, Pennsylvania, United States, 19107

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19124

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15212

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15213

[Not yet recruiting]

Pittsburgh, Pennsylvania, United States, 15219

[Recruiting]

West Mifflin, Pennsylvania, United States, 15122

United States, Rhode Island

[Recruiting]

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

[Not yet recruiting]

Charleston, South Carolina, United States, 29414

[Recruiting]

Columbia, South Carolina, United States, 29223

[Recruiting]

Greenville, South Carolina, United States, 29605

[Not yet recruiting]

Mt. Pleasant, South Carolina, United States, 29464

[Recruiting]

West Columbia, South Carolina, United States, 29169

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Recruiting]

Memphis, Tennessee, United States, 38119

[Terminated]

Memphis, Tennessee, United States, 38120

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Abilene, Texas, United States, 79606

[Recruiting]

Austin, Texas, United States, 78705

[Not yet recruiting]

Corpus Christi, Texas, United States, 78413

[Not yet recruiting]

Dallas, Texas, United States, 75390

[Recruiting]

DeSoto, Texas, United States, 75115

[Recruiting]

Ft. Worth, Texas, United States, 76102

[Recruiting]

Ft. Worth, Texas, United States, 76104

[Not yet recruiting]

Galveston, Texas, United States, 77555

[Recruiting]

Houston, Texas, United States, 77030

[Recruiting]

McAllen, Texas, United States, 78503

[Recruiting]

Odessa, Texas, United States, 79761

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Utah

[Recruiting]

Salt Lake City, Utah, United States, 84107

[Recruiting]

Salt Lake City, Utah, United States, 84132

United States, Vermont

[Recruiting]

Burlington, Vermont, United States, 05401

United States, Virginia

[Recruiting]

Charlottesville, Virginia, United States, 22908

[Recruiting]

Fairfax, Virginia, United States, 22031

[Recruiting]

Richmond, Virginia, United States, 23221

United States, Washington

[Recruiting]

Seattle, Washington, United States, 98104

[Recruiting]

Silverdale, Washington, United States, 98383

United States, Wisconsin

[Recruiting]

Madison, Wisconsin, United States, 53715

[Recruiting]

Madison, Wisconsin, United States, 58705

[Recruiting]

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

[Recruiting]

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

[Recruiting]

Vancouver, British Columbia, Canada, V5Z 3N9

[Recruiting]

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

[Not yet recruiting]

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

[Recruiting]

London, Ontario, Canada, N6A 4G5

[Recruiting]

Mississauga, Ontario, Canada, L4W 1W9

[Not yet recruiting]

Ottawa, Ontario, Canada, K1H8L6

[Not yet recruiting]

Toronto, Ontario, Canada, M4N3M5

[Not yet recruiting]

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

[Recruiting]

Montreal, Quebec, Canada, H1T 2M4

[Recruiting]

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

[Recruiting]

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00509795

Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: March 13, 2008 (v5)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: March 2008

Overall Status: Recruiting

Study Start: August 2007

Primary Completion:

Study Completion:

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that March 13, 2008

Met QC Criteria:

Last Update Posted: March 17, 2008 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms:

Masking: Double (masked roles unspecified)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Intervention Details:

Drug: VEGF Trap-Eye

Outcome Measures

Primary Outcome Measures:

1. Primary measure will be visual acuity changes compared to baseline

Secondary Outcome Measures:

2. Secondary measures will be angiographic and anatomical changes compared to baseline.

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. CNV must be at least 50% of total lesion size.
5. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
6. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.

7. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Prior treatment with anti-VEGF agents as follows:
 - o Prior treatment with anti-VEGF therapy in the study eye is not allowed.
 - o Prior treatment with anti-VEGF therapy in the fellow eye with an investigational agent (not FDA approved, e.g. bevacizumab) is allowed up to 3 months prior to first dose in the study, and such treatment will not be allowed during the study. Prior treatment with an FDA/Health Canada approved anti-VEGF therapy in the fellow eye is allowed.
 - o Prior systemic anti-VEGF therapy, investigational or FDA/Health Canada approved, is only allowed up to 3 months prior to first dose, and will not be allowed during the study
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV, including pathologic myopia (spherical equivalent of -8 diopters or more negative, or axial length of 25 mm or more), ocular histoplasmosis syndrome, angioid streaks, choroidal rupture, or multifocal choroiditis in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.

14. Any history of macular hole of stage 2 and above in the study eye.

15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Central Contact: Regeneron

Telephone: 866-549-8439

Email: VIEW1study@rtp.ppd.com

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Alabama**

[Not yet recruiting]

Birmingham, Alabama, United States, 35205

[Not yet recruiting]

Birmingham, Alabama, United States, 35223

United States, Arizona

[Recruiting]

Phoenix, Arizona, United States, 85014

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tucson, Arizona, United States, 85704

[Not yet recruiting]

Tucson, Arizona, United States, 85710

[Not yet recruiting]

Tucson, Arizona, United States, 85711

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Not yet recruiting]

Fullerton, California, United States, 92835

[Not yet recruiting]

Glendale, California, United States, 91203

[Not yet recruiting]

Irvine, California, United States, 92697

[Not yet recruiting]

La Jolla, California, United States, 92037

[Recruiting]

Loma Linda, California, United States, 92354

[Not yet recruiting]

Los Angeles, California, United States, 90033

[Not yet recruiting]

Los Angeles, California, United States, 90048

[Not yet recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Mountain View, California, United States, 94040

[Recruiting]

Oakland, California, United States, 94609

[Recruiting]

Palm Springs, California, United States, 92262

[Recruiting]

Pasadena, California, United States, 91105

[Not yet recruiting]

Poway, California, United States, 92064

[Not yet recruiting]

Sacramento, California, United States, 95819

[Not yet recruiting]

San Diego, California, United States, 92120

[Recruiting]

San Francisco, California, United States, 94107

[Not yet recruiting]

San Marino, California, United States, 91108

[Recruiting]

Santa Ana, California, United States, 92705

[Not yet recruiting]

Ventura, California, United States, 93003

[Recruiting]

Westlake Village, California, United States, 91361

[Not yet recruiting]

Yorba Linda, California, United States, 92887

United States, Colorado

[Not yet recruiting]

Aurora, Colorado, United States, 80045

[Not yet recruiting]

Denver, Colorado, United States, 80205

[Not yet recruiting]

Denver, Colorado, United States, 80230

United States, Connecticut

[Not yet recruiting]

Bridgeport, Connecticut, United States, 06606

[Recruiting]

Hamden, Connecticut, United States, 06518

[Not yet recruiting]

New Haven, Connecticut, United States, 06520

[Recruiting]

New London, Connecticut, United States, 06320

United States, Florida

[Recruiting]

Altamonte Springs, Florida, United States, 32701

[Recruiting]

Boynton Beach, Florida, United States, 33426

[Recruiting]

Delray Beach, Florida, United States, 33484

[Recruiting]

Fort Myers, Florida, United States, 33907

[Not yet recruiting]

Ft. Lauderdale, Florida, United States, 33334

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Not yet recruiting]

Gainesville, Florida, United States, 32610

[Not yet recruiting]

Jacksonville, Florida, United States, 32216

[Not yet recruiting]

Jacksonville, Florida, United States, 32224

[Not yet recruiting]

Margate, Florida, United States, 33063

[Not yet recruiting]

Miami, Florida, United States, 33125

[Not yet recruiting]

Miami, Florida, United States, 33136

[Not yet recruiting]

Mount Dora, Florida, United States, 32757

[Not yet recruiting]

Ocala, Florida, United States, 34472

[Recruiting]

Orlando, Florida, United States, 32806

[Recruiting]

Orlando, Florida, United States, 38203

[Recruiting]

Palm Beach Gardens, Florida, United States, 33410

[Recruiting]

Pensacola, Florida, United States, 32503

[Recruiting]

Sarasota, Florida, United States, 34239

[Not yet recruiting]

Stuart, Florida, United States, 34994

[Recruiting]

Sunrise, Florida, United States, 33351

[Not yet recruiting]

Tampa, Florida, United States, 33607

[Recruiting]

Tampa, Florida, United States, 33612

[Recruiting]

Winter Haven, Florida, United States, 33880

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

[Not yet recruiting]

Marietta, Georgia, United States, 30060

United States, Hawaii

[Not yet recruiting]

Aiea, Hawaii, United States, 96701

[Not yet recruiting]

Honolulu, Hawaii, United States, 96813

United States, Illinois

[Not yet recruiting]

Chicago, Illinois, United States, 60611

[Not yet recruiting]

Chicago, Illinois, United States, 60637

[Recruiting]

Oak Brook, Illinois, United States, 60523

[Not yet recruiting]

Springfield, Illinois, United States, 62701

[Not yet recruiting]

Wheaton, Illinois, United States, 60187

United States, Indiana

[Not yet recruiting]

Ft. Wayne, Indiana, United States, 46804

[Not yet recruiting]

Indianapolis, Indiana, United States, 46202

[Not yet recruiting]

Indianapolis, Indiana, United States, 46260

[Recruiting]

Indianapolis, Indiana, United States, 46280

[Recruiting]

New Albany, Indiana, United States, 47150

United States, Iowa

[Not yet recruiting]

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

[Recruiting]

Wichita, Kansas, United States, 67214

United States, Kentucky

[Not yet recruiting]

Louisville, Kentucky, United States, 40202

[Not yet recruiting]

Louisville, Kentucky, United States, 40207

[Recruiting]

Paducah, Kentucky, United States, 42001

United States, Louisiana

[Not yet recruiting]

Metairie, Louisiana, United States, 70002

[Recruiting]

New Orleans, Louisiana, United States, 70115

[Not yet recruiting]

New Orleans, Louisiana, United States, 70121

[Not yet recruiting]

Shreveport, Louisiana, United States, 71105

United States, Maine

[Recruiting]

Bangor, Maine, United States, 04401

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21209

[Not yet recruiting]

Baltimore, Maryland, United States, 21287

[Recruiting]

Chevy Chase, Maryland, United States, 20815

[Not yet recruiting]

Hagerstown, Maryland, United States, 21740

[Recruiting]

Towson, Maryland, United States, 21204

United States, Massachusetts

[Not yet recruiting]

Boston, Massachusetts, United States, 02111

[Recruiting]

Boston, Massachusetts, United States, 02114

[Not yet recruiting]

Boston, Massachusetts, United States, 02215

[Recruiting]

Peabody, Massachusetts, United States, 01960

United States, Michigan

[Not yet recruiting]

Ann Arbor, Michigan, United States, 48105

[Not yet recruiting]

Battle Creek, Michigan, United States, 49015

[Not yet recruiting]

East Lansing, Michigan, United States, 48823

[Not yet recruiting]

Grand Blanc, Michigan, United States, 48439

[Recruiting]

Grand Rapids, Michigan, United States, 49525

[Not yet recruiting]

Jackson, Michigan, United States, 49201

[Recruiting]

Royal Oak, Michigan, United States, 48073

[Recruiting]

Southfield, Michigan, United States, 48034

[Not yet recruiting]

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

[Recruiting]

Edina, Minnesota, United States, 55435

[Recruiting]

Minneapolis, Minnesota, United States, 55404

[Not yet recruiting]

Rochester, Minnesota, United States, 55905

United States, Missouri

[Not yet recruiting]

Florissant, Missouri, United States, 63031

[Recruiting]

Kansas City, Missouri, United States, 64108

[Not yet recruiting]

Kansas City, Missouri, United States, 64111

[Not yet recruiting]

Springfield, Missouri, United States, 65804

[Not yet recruiting]

St. Louis, Missouri, United States, 63104

[Recruiting]

St. Louis, Missouri, United States, 63110

United States, Montana

[Recruiting]

Missoula, Montana, United States, 59801

United States, Nebraska

[Recruiting]

Lincoln, Nebraska, United States, 68506

[Recruiting]

Omaha, Nebraska, United States, 68124

[Recruiting]

Omaha, Nebraska, United States, 68131

United States, Nevada

[Recruiting]

Las Vegas, Nevada, United States, 89144

United States, New Hampshire

[Not yet recruiting]

Lebanon, New Hampshire, United States, 03756

United States, New Jersey

[Recruiting]

Lawrenceville, New Jersey, United States, 08648

[Not yet recruiting]

Newark, New Jersey, United States, 07103

[Not yet recruiting]

Northfield, New Jersey, United States, 08225

[Recruiting]

Northfield, New Jersey, United States, 08225

[Recruiting]

Teaneck, New Jersey, United States, 07666

[Recruiting]

Toms River, New Jersey, United States, 08753

United States, New Mexico

[Recruiting]

Albuquerque, New Mexico, United States, 87106

United States, New York

[Not yet recruiting]

Albany, New York, United States, 12206

[Not yet recruiting]

Bronxville, New York, United States, 10708

[Not yet recruiting]

Bronx, New York, United States, 10467

[Not yet recruiting]

Brooklyn, New York, United States, 11223

[Not yet recruiting]

Great Neck, New York, United States, 11021

[Recruiting]

Lynbrook, New York, United States, 11563

[Recruiting]

New York, New York, United States, 10003

[Recruiting]

New York, New York, United States, 10021

[Not yet recruiting]

New York, New York, United States, 10032

[Recruiting]

Rochester, New York, United States, 14618

[Not yet recruiting]

Rochester, New York, United States, 14642

[Recruiting]

Slingerlands, New York, United States, 12159

United States, North Carolina

[Recruiting]

Asheville, North Carolina, United States, 28803

[Recruiting]

Charlotte, North Carolina, United States, 28210

[Not yet recruiting]

Hendersonville, North Carolina, United States, 28791

[Not yet recruiting]

Raleigh, North Carolina, United States, 27607

[Not yet recruiting]

Raleigh, North Carolina, United States, 27612

[Recruiting]

Southern Pines, North Carolina, United States, 28387

[Recruiting]

Winston-Salem, North Carolina, United States, 27157

United States, North Dakota

[Not yet recruiting]

Fargo, North Dakota, United States, 58103

United States, Ohio

[Not yet recruiting]

Canton, Ohio, United States, 44718

[Not yet recruiting]

Cincinnati, Ohio, United States, 45202

[Recruiting]

Cincinnati, Ohio, United States, 45242

[Recruiting]

Cleveland, Ohio, United States, 44122

[Not yet recruiting]

Cleveland, Ohio, United States, 44195

[Not yet recruiting]

Columbus, Ohio, United States, 43215

[Not yet recruiting]

Toledo, Ohio, United States, 43606

[Recruiting]

Toledo, Ohio, United States, 43608

[Not yet recruiting]

Toledo, Ohio, United States, 43615

United States, Oklahoma

[Not yet recruiting]

Oklahoma City, Oklahoma, United States, 73104

[Not yet recruiting]

Tulsa, Oklahoma, United States, 74104

United States, Oregon

[Recruiting]

Ashland, Oregon, United States, 97520

[Not yet recruiting]

Portland, Oregon, United States, 97210

[Recruiting]

Portland, Oregon, United States, 97210

[Not yet recruiting]

Portland, Oregon, United States, 97227

[Not yet recruiting]

Salem, Oregon, United States, 97302

United States, Pennsylvania

[Not yet recruiting]

Camp Hill, Pennsylvania, United States, 17011

[Not yet recruiting]

Campbell, Pennsylvania, United States, 95008

[Not yet recruiting]

East Stroudsburg, Pennsylvania, United States, 18301

[Recruiting]

Kingston, Pennsylvania, United States, 18704

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19104

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19107

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19124

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15212

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15213

[Not yet recruiting]

Pittsburgh, Pennsylvania, United States, 15219

[Recruiting]

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

[Recruiting]

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

[Recruiting]

Columbia, South Carolina, United States, 29204

[Recruiting]

Columbia, South Carolina, United States, 29223

[Not yet recruiting]

Greenville, South Carolina, United States, 29605

[Not yet recruiting]

Rock Hill, South Carolina, United States, 29732

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Recruiting]

Memphis, Tennessee, United States, 38119

[Not yet recruiting]

Memphis, Tennessee, United States, 38120

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Abilene, Texas, United States, 79606

[Recruiting]

Austin, Texas, United States, 78705

[Recruiting]

DeSoto, Texas, United States, 75115

[Recruiting]

Ft. Worth, Texas, United States, 76102

[Not yet recruiting]

Ft. Worth, Texas, United States, 76104

[Not yet recruiting]

Galveston, Texas, United States, 77555

[Recruiting]

Houston, Texas, United States, 77030

[Recruiting]

McAllen, Texas, United States, 78503

[Not yet recruiting]

Odessa, Texas, United States, 79761

[Recruiting]

San Antonio, Texas, United States, 78133

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Utah

[Not yet recruiting]

Ogden, Utah, United States, 84403

[Recruiting]

Salt Lake City, Utah, United States, 84107

[Not yet recruiting]

Salt Lake City, Utah, United States, 84132

United States, Vermont

[Not yet recruiting]

Burlington, Vermont, United States, 05401

United States, Virginia

[Not yet recruiting]

Charlottesville, Virginia, United States, 22908

[Recruiting]

Richmond, Virginia, United States, 23221

United States, Washington

[Recruiting]

Seattle, Washington, United States, 98104

[Not yet recruiting]

Seattle, Washington, United States, 98195

[Recruiting]

Silverdale, Washington, United States, 98383

[Not yet recruiting]

Spokane, Washington, United States, 99203

United States, Wisconsin

[Not yet recruiting]

Madison, Wisconsin, United States, 53715

[Not yet recruiting]

Madison, Wisconsin, United States, 58705

[Not yet recruiting]

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

[Not yet recruiting]

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

[Not yet recruiting]

Vancouver, British Columbia, Canada, V5Z 3N9

[Not yet recruiting]

Victoria, British Columbia, Canada, V8X 5G6

Canada, Nova Scotia

[Not yet recruiting]

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

[Not yet recruiting]

London, Ontario, Canada, N6A 4G5

[Not yet recruiting]

Mississauga, Ontario, Canada, L4W 1W9

[Not yet recruiting]

Toronto, Ontario, Canada, M5C 2T2

[Recruiting]

Toronto, Ontario, Canada, M5T 2S8

Canada, Quebec

[Not yet recruiting]

Montreal, Quebec, Canada, H1T 2M4

[Not yet recruiting]

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

[Not yet recruiting]

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00509795

Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
4	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: December 4, 2007 (v4)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: December 2007

Overall Status: Recruiting

Study Start: August 2007

Primary Completion:

Study Completion:

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that December 4, 2007

Met QC Criteria:

Last Update Posted: December 6, 2007 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms:

Masking: Double (masked roles unspecified)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Intervention Details:

Drug: VEGF Trap-Eye

Outcome Measures

Primary Outcome Measures:

1. Primary measure will be visual acuity changes compared to baseline

Secondary Outcome Measures:

2. Secondary measures will be angiographic and anatomical changes compared to baseline.

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.

6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents.
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV, including pathologic myopia (spherical equivalent of -8 diopters or more negative, or axial length of 25 mm or more), ocular histoplasmosis syndrome, angioid streaks, choroidal rupture, or multifocal choroiditis in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Central Contact: Regeneron

Telephone: 866-549-8439

Email: VIEW1study@rtp.ppd.com

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Alabama**

[Not yet recruiting]

Birmingham, Alabama, United States, 35205

[Not yet recruiting]

Birmingham, Alabama, United States, 35223

United States, Arizona

[Recruiting]

Phoenix, Arizona, United States, 85014

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tucson, Arizona, United States, 85704

[Not yet recruiting]

Tucson, Arizona, United States, 85710

[Not yet recruiting]

Tucson, Arizona, United States, 85711

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Not yet recruiting]

Fullerton, California, United States, 92835

[Not yet recruiting]

Glendale, California, United States, 91203

[Not yet recruiting]

Irvine, California, United States, 92697

[Not yet recruiting]

La Jolla, California, United States, 92037

[Recruiting]

Loma Linda, California, United States, 92354

[Not yet recruiting]

Los Angeles, California, United States, 90033

[Not yet recruiting]

Los Angeles, California, United States, 90048

[Not yet recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Mountain View, California, United States, 94040

[Recruiting]

Oakland, California, United States, 94609

[Recruiting]

Palm Springs, California, United States, 92262

[Recruiting]

Pasadena, California, United States, 91105

[Not yet recruiting]

Poway, California, United States, 92064

[Not yet recruiting]

Sacramento, California, United States, 95819

[Not yet recruiting]

San Diego, California, United States, 92120

[Recruiting]

San Francisco, California, United States, 94107

[Not yet recruiting]

San Marino, California, United States, 91108

[Recruiting]

Santa Ana, California, United States, 92705

[Not yet recruiting]

Ventura, California, United States, 93003

[Recruiting]

Westlake Village, California, United States, 91361

[Not yet recruiting]

Yorba Linda, California, United States, 92887

United States, Colorado

[Not yet recruiting]

Aurora, Colorado, United States, 80045

[Not yet recruiting]

Denver, Colorado, United States, 80205

[Not yet recruiting]

Denver, Colorado, United States, 80230

United States, Connecticut

[Not yet recruiting]

Bridgeport, Connecticut, United States, 06606

[Recruiting]

Hamden, Connecticut, United States, 06518

[Not yet recruiting]

New Haven, Connecticut, United States, 06520

[Recruiting]

New London, Connecticut, United States, 06320

United States, Florida

[Recruiting]

Altamonte Springs, Florida, United States, 32701

[Recruiting]

Boynton Beach, Florida, United States, 33426

[Recruiting]

Delray Beach, Florida, United States, 33484

[Recruiting]

Fort Myers, Florida, United States, 33907

[Not yet recruiting]

Ft. Lauderdale, Florida, United States, 33334

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Not yet recruiting]

Gainesville, Florida, United States, 32610

[Not yet recruiting]

Jacksonville, Florida, United States, 32216

[Not yet recruiting]

Jacksonville, Florida, United States, 32224

[Not yet recruiting]

Margate, Florida, United States, 33063

[Not yet recruiting]

Miami, Florida, United States, 33125

[Not yet recruiting]

Miami, Florida, United States, 33136

[Not yet recruiting]

Mount Dora, Florida, United States, 32757

[Not yet recruiting]

Ocala, Florida, United States, 34472

[Recruiting]

Orlando, Florida, United States, 32806

[Recruiting]

Orlando, Florida, United States, 38203

[Recruiting]

Palm Beach Gardens, Florida, United States, 33410

[Recruiting]

Pensacola, Florida, United States, 32503

[Recruiting]

Sarasota, Florida, United States, 34239

[Not yet recruiting]

Stuart, Florida, United States, 34994

[Recruiting]

Sunrise, Florida, United States, 33351

[Not yet recruiting]

Tampa, Florida, United States, 33607

[Recruiting]

Tampa, Florida, United States, 33612

[Recruiting]

Winter Haven, Florida, United States, 33880

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

[Not yet recruiting]

Marietta, Georgia, United States, 30060

United States, Hawaii

[Not yet recruiting]

Aiea, Hawaii, United States, 96701

[Not yet recruiting]

Honolulu, Hawaii, United States, 96813

United States, Illinois

[Not yet recruiting]

Chicago, Illinois, United States, 60611

[Not yet recruiting]

Chicago, Illinois, United States, 60637

[Recruiting]

Oak Brook, Illinois, United States, 60523

[Not yet recruiting]

Springfield, Illinois, United States, 62701

[Not yet recruiting]

Wheaton, Illinois, United States, 60187

United States, Indiana

[Not yet recruiting]

Ft. Wayne, Indiana, United States, 46804

[Not yet recruiting]

Indianapolis, Indiana, United States, 46202

[Not yet recruiting]

Indianapolis, Indiana, United States, 46260

[Recruiting]

Indianapolis, Indiana, United States, 46280

[Recruiting]

New Albany, Indiana, United States, 47150

United States, Iowa

[Not yet recruiting]

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

[Recruiting]

Wichita, Kansas, United States, 67214

United States, Kentucky

[Not yet recruiting]

Louisville, Kentucky, United States, 40202

[Not yet recruiting]

Louisville, Kentucky, United States, 40207

[Recruiting]

Paducah, Kentucky, United States, 42001

United States, Louisiana

[Not yet recruiting]

Metairie, Louisiana, United States, 70002

[Recruiting]

New Orleans, Louisiana, United States, 70115

[Not yet recruiting]

New Orleans, Louisiana, United States, 70121

[Not yet recruiting]

Shreveport, Louisiana, United States, 71105

United States, Maine

[Recruiting]

Bangor, Maine, United States, 04401

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21209

[Not yet recruiting]

Baltimore, Maryland, United States, 21287

[Recruiting]

Chevy Chase, Maryland, United States, 20815

[Not yet recruiting]

Hagerstown, Maryland, United States, 21740

[Recruiting]

Towson, Maryland, United States, 21204

United States, Massachusetts

[Not yet recruiting]

Boston, Massachusetts, United States, 02111

[Recruiting]

Boston, Massachusetts, United States, 02114

[Not yet recruiting]

Boston, Massachusetts, United States, 02215

[Recruiting]

Peabody, Massachusetts, United States, 01960

United States, Michigan

[Not yet recruiting]

Ann Arbor, Michigan, United States, 48105

[Not yet recruiting]

Battle Creek, Michigan, United States, 49015

[Not yet recruiting]

East Lansing, Michigan, United States, 48823

[Not yet recruiting]

Grand Blanc, Michigan, United States, 48439

[Recruiting]

Grand Rapids, Michigan, United States, 49525

[Not yet recruiting]

Jackson, Michigan, United States, 49201

[Recruiting]

Royal Oak, Michigan, United States, 48073

[Recruiting]

Southfield, Michigan, United States, 48034

[Not yet recruiting]

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

[Recruiting]

Edina, Minnesota, United States, 55435

[Recruiting]

Minneapolis, Minnesota, United States, 55404

[Not yet recruiting]

Rochester, Minnesota, United States, 55905

United States, Missouri

[Not yet recruiting]

Florissant, Missouri, United States, 63031

[Recruiting]

Kansas City, Missouri, United States, 64108

[Not yet recruiting]

Kansas City, Missouri, United States, 64111

[Not yet recruiting]

Springfield, Missouri, United States, 65804

[Not yet recruiting]

St. Louis, Missouri, United States, 63104

[Recruiting]

St. Louis, Missouri, United States, 63110

United States, Montana

[Recruiting]

Missoula, Montana, United States, 59801

United States, Nebraska

[Recruiting]

Lincoln, Nebraska, United States, 68506

[Recruiting]

Omaha, Nebraska, United States, 68124

[Recruiting]

Omaha, Nebraska, United States, 68131

United States, Nevada

[Recruiting]

Las Vegas, Nevada, United States, 89144

United States, New Hampshire

[Not yet recruiting]

Lebanon, New Hampshire, United States, 03756

United States, New Jersey

[Recruiting]

Lawrenceville, New Jersey, United States, 08648

[Not yet recruiting]

Newark, New Jersey, United States, 07103

[Not yet recruiting]

Northfield, New Jersey, United States, 08225

[Recruiting]

Northfield, New Jersey, United States, 08225

[Recruiting]

Teaneck, New Jersey, United States, 07666

[Recruiting]

Toms River, New Jersey, United States, 08753

United States, New Mexico

[Recruiting]

Albuquerque, New Mexico, United States, 87106

United States, New York

[Not yet recruiting]

Albany, New York, United States, 12206

[Not yet recruiting]

Bronxville, New York, United States, 10708

[Not yet recruiting]

Bronx, New York, United States, 10467

[Not yet recruiting]

Brooklyn, New York, United States, 11223

[Not yet recruiting]

Great Neck, New York, United States, 11021

[Recruiting]

Lynbrook, New York, United States, 11563

[Recruiting]

New York, New York, United States, 10003

[Recruiting]

New York, New York, United States, 10021

[Not yet recruiting]

New York, New York, United States, 10032

[Recruiting]

Rochester, New York, United States, 14618

[Not yet recruiting]

Rochester, New York, United States, 14642

[Recruiting]

Slingerlands, New York, United States, 12159

United States, North Carolina

[Recruiting]

Asheville, North Carolina, United States, 28803

[Recruiting]

Charlotte, North Carolina, United States, 28210

[Not yet recruiting]

Hendersonville, North Carolina, United States, 28791

[Not yet recruiting]

Raleigh, North Carolina, United States, 27607

[Not yet recruiting]

Raleigh, North Carolina, United States, 27612

[Recruiting]

Southern Pines, North Carolina, United States, 28387

[Recruiting]

Winston-Salem, North Carolina, United States, 27157

United States, North Dakota

[Not yet recruiting]

Fargo, North Dakota, United States, 58103

United States, Ohio

[Not yet recruiting]

Canton, Ohio, United States, 44718

[Not yet recruiting]

Cincinnati, Ohio, United States, 45202

[Recruiting]

Cincinnati, Ohio, United States, 45242

[Recruiting]

Cleveland, Ohio, United States, 44122

[Not yet recruiting]

Cleveland, Ohio, United States, 44195

[Not yet recruiting]
Columbus, Ohio, United States, 43215

[Not yet recruiting]
Toledo, Ohio, United States, 43606

[Recruiting]
Toledo, Ohio, United States, 43608

[Not yet recruiting]
Toledo, Ohio, United States, 43615

United States, Oklahoma

[Not yet recruiting]
Oklahoma City, Oklahoma, United States, 73104

[Not yet recruiting]
Tulsa, Oklahoma, United States, 74104

United States, Oregon

[Recruiting]
Ashland, Oregon, United States, 97520

[Not yet recruiting]
Portland, Oregon, United States, 97210

[Recruiting]
Portland, Oregon, United States, 97210

[Not yet recruiting]
Portland, Oregon, United States, 97227

[Not yet recruiting]
Salem, Oregon, United States, 97302

United States, Pennsylvania

[Not yet recruiting]
Camp Hill, Pennsylvania, United States, 17011

[Not yet recruiting]

Campbell, Pennsylvania, United States, 95008

[Not yet recruiting]

East Stroudsburg, Pennsylvania, United States, 18301

[Recruiting]

Kingston, Pennsylvania, United States, 18704

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19104

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19107

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19124

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15212

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15213

[Not yet recruiting]

Pittsburgh, Pennsylvania, United States, 15219

[Recruiting]

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

[Recruiting]

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

[Recruiting]

Columbia, South Carolina, United States, 29204

[Recruiting]

Columbia, South Carolina, United States, 29223

[Not yet recruiting]

Greenville, South Carolina, United States, 29605

[Not yet recruiting]

Rock Hill, South Carolina, United States, 29732

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Recruiting]

Memphis, Tennessee, United States, 38119

[Not yet recruiting]

Memphis, Tennessee, United States, 38120

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Abilene, Texas, United States, 79606

[Recruiting]

Austin, Texas, United States, 78705

[Recruiting]

DeSoto, Texas, United States, 75115

[Recruiting]

Ft. Worth, Texas, United States, 76102

[Not yet recruiting]

Ft. Worth, Texas, United States, 76104

[Not yet recruiting]

Galveston, Texas, United States, 77555

[Recruiting]

Houston, Texas, United States, 77030

[Recruiting]

McAllen, Texas, United States, 78503

[Not yet recruiting]

Odessa, Texas, United States, 79761

[Recruiting]

San Antonio, Texas, United States, 78133

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Utah

[Not yet recruiting]

Ogden, Utah, United States, 84403

[Recruiting]

Salt Lake City, Utah, United States, 84107

[Not yet recruiting]

Salt Lake City, Utah, United States, 84132

United States, Vermont

[Not yet recruiting]

Burlington, Vermont, United States, 05401

United States, Virginia

[Not yet recruiting]

Charlottesville, Virginia, United States, 22908

[Recruiting]

Richmond, Virginia, United States, 23221

United States, Washington

[Recruiting]

Seattle, Washington, United States, 98104

[Not yet recruiting]

Seattle, Washington, United States, 98195

[Recruiting]

Silverdale, Washington, United States, 98383

[Not yet recruiting]

Spokane, Washington, United States, 99203

United States, Wisconsin

[Not yet recruiting]

Madison, Wisconsin, United States, 53715

[Not yet recruiting]

Madison, Wisconsin, United States, 58705

[Not yet recruiting]

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

[Not yet recruiting]

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

[Not yet recruiting]

Vancouver, British Columbia, Canada, V5Z 3N9

[Not yet recruiting]

Victoria, British Columbia, Canada, V8X 5G6

Canada, Nova Scotia

[Not yet recruiting]

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

[Not yet recruiting]

London, Ontario, Canada, N6A 4G5

[Not yet recruiting]

Mississauga, Ontario, Canada, L4W 1W9

[Not yet recruiting]

Toronto, Ontario, Canada, M5C 2T2

[Recruiting]

Toronto, Ontario, Canada, M5T 2S8

Canada, Quebec

[Not yet recruiting]

Montreal, Quebec, Canada, H1T 2M4

[Not yet recruiting]

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

[Not yet recruiting]

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00509795

Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: November 14, 2007 (v3)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: August 2007

Overall Status: Recruiting

Study Start: August 2007

Primary Completion:

Study Completion:

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that November 14, 2007

Met QC Criteria:

Last Update Posted: November 16, 2007 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms:

Masking: Double (masked roles unspecified)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Intervention Details:

Drug: VEGF Trap-Eye

Outcome Measures

Primary Outcome Measures:

1. Primary measure will be visual acuity changes compared to baseline

Secondary Outcome Measures:

2. Secondary measures will be angiographic and anatomical changes compared to baseline.

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.

6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents.
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV, including pathologic myopia (spherical equivalent of -8 diopters or more negative, or axial length of 25 mm or more), ocular histoplasmosis syndrome, angioid streaks, choroidal rupture, or multifocal choroiditis in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Central Contact: Regeneron

Telephone: 866-549-8439

Email: VIEW1study@ppdi.com

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Alabama**

[Not yet recruiting]

Birmingham, Alabama, United States, 35205

[Not yet recruiting]

Birmingham, Alabama, United States, 35223

United States, Arizona

[Recruiting]

Phoenix, Arizona, United States, 85014

[Recruiting]

Phoenix, Arizona, United States, 85020

[Recruiting]

Tucson, Arizona, United States, 85704

[Not yet recruiting]

Tucson, Arizona, United States, 85710

[Not yet recruiting]

Tucson, Arizona, United States, 85711

United States, California

[Recruiting]

Beverly Hills, California, United States, 90211

[Not yet recruiting]

Fullerton, California, United States, 92835

[Not yet recruiting]

Glendale, California, United States, 91203

[Not yet recruiting]

Irvine, California, United States, 92697

[Not yet recruiting]

La Jolla, California, United States, 92037

[Recruiting]

Loma Linda, California, United States, 92354

[Not yet recruiting]

Los Angeles, California, United States, 90033

[Not yet recruiting]

Los Angeles, California, United States, 90048

[Not yet recruiting]

Menlo Park, California, United States, 94025

[Recruiting]

Mountain View, California, United States, 94040

[Recruiting]

Oakland, California, United States, 94609

[Recruiting]

Palm Springs, California, United States, 92262

[Recruiting]

Pasadena, California, United States, 91105

[Not yet recruiting]

Poway, California, United States, 92064

[Not yet recruiting]

Sacramento, California, United States, 95819

[Not yet recruiting]

San Diego, California, United States, 92120

[Recruiting]

San Francisco, California, United States, 94107

[Not yet recruiting]

San Marino, California, United States, 91108

[Recruiting]

Santa Ana, California, United States, 92705

[Not yet recruiting]

Ventura, California, United States, 93003

[Recruiting]

Westlake Village, California, United States, 91361

[Not yet recruiting]

Yorba Linda, California, United States, 92887

United States, Colorado

[Not yet recruiting]

Aurora, Colorado, United States, 80045

[Not yet recruiting]

Denver, Colorado, United States, 80205

[Not yet recruiting]

Denver, Colorado, United States, 80230

United States, Connecticut

[Not yet recruiting]

Bridgeport, Connecticut, United States, 06606

[Recruiting]

Hamden, Connecticut, United States, 06518

[Not yet recruiting]

New Haven, Connecticut, United States, 06520

[Recruiting]

New London, Connecticut, United States, 06320

United States, Florida

[Recruiting]

Altamonte Springs, Florida, United States, 32701

[Recruiting]

Boynton Beach, Florida, United States, 33426

[Recruiting]

Delray Beach, Florida, United States, 33484

[Recruiting]

Fort Myers, Florida, United States, 33907

[Not yet recruiting]

Ft. Lauderdale, Florida, United States, 33334

[Recruiting]

Ft. Myers, Florida, United States, 33912

[Not yet recruiting]

Gainesville, Florida, United States, 32610

[Not yet recruiting]

Jacksonville, Florida, United States, 32216

[Not yet recruiting]

Jacksonville, Florida, United States, 32224

[Not yet recruiting]

Margate, Florida, United States, 33063

[Not yet recruiting]

Miami, Florida, United States, 33125

[Not yet recruiting]

Miami, Florida, United States, 33136

[Not yet recruiting]

Mount Dora, Florida, United States, 32757

[Not yet recruiting]

Ocala, Florida, United States, 34472

[Recruiting]

Orlando, Florida, United States, 32806

[Recruiting]

Orlando, Florida, United States, 38203

[Recruiting]

Palm Beach Gardens, Florida, United States, 33410

[Recruiting]

Pensacola, Florida, United States, 32503

[Recruiting]

Sarasota, Florida, United States, 34239

[Not yet recruiting]

Stuart, Florida, United States, 34994

[Recruiting]

Sunrise, Florida, United States, 33351

[Not yet recruiting]

Tampa, Florida, United States, 33607

[Recruiting]

Tampa, Florida, United States, 33612

[Recruiting]

Winter Haven, Florida, United States, 33880

United States, Georgia

[Recruiting]

Augusta, Georgia, United States, 30909

[Not yet recruiting]

Marietta, Georgia, United States, 30060

United States, Hawaii

[Not yet recruiting]

Aiea, Hawaii, United States, 96701

[Not yet recruiting]

Honolulu, Hawaii, United States, 96813

United States, Illinois

[Not yet recruiting]

Chicago, Illinois, United States, 60611

[Not yet recruiting]

Chicago, Illinois, United States, 60637

[Recruiting]

Oak Brook, Illinois, United States, 60523

[Not yet recruiting]

Springfield, Illinois, United States, 62701

[Not yet recruiting]

Wheaton, Illinois, United States, 60187

United States, Indiana

[Not yet recruiting]

Ft. Wayne, Indiana, United States, 46804

[Not yet recruiting]

Indianapolis, Indiana, United States, 46202

[Not yet recruiting]

Indianapolis, Indiana, United States, 46260

[Recruiting]

Indianapolis, Indiana, United States, 46280

[Recruiting]

New Albany, Indiana, United States, 47150

United States, Iowa

[Not yet recruiting]

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

[Recruiting]

Wichita, Kansas, United States, 67214

United States, Kentucky

[Not yet recruiting]

Louisville, Kentucky, United States, 40202

[Not yet recruiting]

Louisville, Kentucky, United States, 40207

[Recruiting]

Paducah, Kentucky, United States, 42001

United States, Louisiana

[Not yet recruiting]

Metairie, Louisiana, United States, 70002

[Recruiting]

New Orleans, Louisiana, United States, 70115

[Not yet recruiting]

New Orleans, Louisiana, United States, 70121

[Not yet recruiting]

Shreveport, Louisiana, United States, 71105

United States, Maine

[Recruiting]

Bangor, Maine, United States, 04401

United States, Maryland

[Recruiting]

Baltimore, Maryland, United States, 21209

[Not yet recruiting]

Baltimore, Maryland, United States, 21287

[Recruiting]

Chevy Chase, Maryland, United States, 20815

[Not yet recruiting]

Hagerstown, Maryland, United States, 21740

[Recruiting]

Towson, Maryland, United States, 21204

United States, Massachusetts

[Not yet recruiting]

Boston, Massachusetts, United States, 02111

[Recruiting]

Boston, Massachusetts, United States, 02114

[Not yet recruiting]

Boston, Massachusetts, United States, 02215

[Recruiting]

Peabody, Massachusetts, United States, 01960

United States, Michigan

[Not yet recruiting]

Ann Arbor, Michigan, United States, 48105

[Not yet recruiting]

Battle Creek, Michigan, United States, 49015

[Not yet recruiting]

East Lansing, Michigan, United States, 48823

[Not yet recruiting]

Grand Blanc, Michigan, United States, 48439

[Recruiting]

Grand Rapids, Michigan, United States, 49525

[Not yet recruiting]

Jackson, Michigan, United States, 49201

[Recruiting]

Royal Oak, Michigan, United States, 48073

[Recruiting]

Southfield, Michigan, United States, 48034

[Not yet recruiting]

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

[Recruiting]

Edina, Minnesota, United States, 55435

[Recruiting]

Minneapolis, Minnesota, United States, 55404

[Not yet recruiting]

Rochester, Minnesota, United States, 55905

United States, Missouri

[Not yet recruiting]

Florissant, Missouri, United States, 63031

[Recruiting]

Kansas City, Missouri, United States, 64108

[Not yet recruiting]

Kansas City, Missouri, United States, 64111

[Not yet recruiting]

Springfield, Missouri, United States, 65804

[Not yet recruiting]

St. Louis, Missouri, United States, 63104

[Recruiting]

St. Louis, Missouri, United States, 63110

United States, Montana

[Recruiting]

Missoula, Montana, United States, 59801

United States, Nebraska

[Recruiting]

Lincoln, Nebraska, United States, 68506

[Recruiting]

Omaha, Nebraska, United States, 68124

[Recruiting]

Omaha, Nebraska, United States, 68131

United States, Nevada

[Recruiting]

Las Vegas, Nevada, United States, 89144

United States, New Hampshire

[Not yet recruiting]

Lebanon, New Hampshire, United States, 03756

United States, New Jersey

[Recruiting]

Lawrenceville, New Jersey, United States, 08648

[Not yet recruiting]

Newark, New Jersey, United States, 07103

[Not yet recruiting]

Northfield, New Jersey, United States, 08225

[Recruiting]

Northfield, New Jersey, United States, 08225

[Recruiting]

Teaneck, New Jersey, United States, 07666

[Recruiting]

Toms River, New Jersey, United States, 08753

United States, New Mexico

[Recruiting]

Albuquerque, New Mexico, United States, 87106

United States, New York

[Not yet recruiting]

Albany, New York, United States, 12206

[Not yet recruiting]

Bronxville, New York, United States, 10708

[Not yet recruiting]

Bronx, New York, United States, 10467

[Not yet recruiting]

Brooklyn, New York, United States, 11223

[Not yet recruiting]

Great Neck, New York, United States, 11021

[Recruiting]

Lynbrook, New York, United States, 11563

[Recruiting]

New York, New York, United States, 10003

[Recruiting]

New York, New York, United States, 10021

[Not yet recruiting]

New York, New York, United States, 10032

[Recruiting]

Rochester, New York, United States, 14618

[Not yet recruiting]

Rochester, New York, United States, 14642

[Recruiting]

Slingerlands, New York, United States, 12159

United States, North Carolina

[Recruiting]

Asheville, North Carolina, United States, 28803

[Recruiting]

Charlotte, North Carolina, United States, 28210

[Not yet recruiting]

Hendersonville, North Carolina, United States, 28791

[Not yet recruiting]

Raleigh, North Carolina, United States, 27607

[Not yet recruiting]

Raleigh, North Carolina, United States, 27612

[Recruiting]

Southern Pines, North Carolina, United States, 28387

[Recruiting]

Winston-Salem, North Carolina, United States, 27157

United States, North Dakota

[Not yet recruiting]

Fargo, North Dakota, United States, 58103

United States, Ohio

[Not yet recruiting]

Canton, Ohio, United States, 44718

[Not yet recruiting]

Cincinnati, Ohio, United States, 45202

[Recruiting]

Cincinnati, Ohio, United States, 45242

[Recruiting]

Cleveland, Ohio, United States, 44122

[Not yet recruiting]

Cleveland, Ohio, United States, 44195

[Not yet recruiting]
Columbus, Ohio, United States, 43215

[Not yet recruiting]
Toledo, Ohio, United States, 43606

[Recruiting]
Toledo, Ohio, United States, 43608

[Not yet recruiting]
Toledo, Ohio, United States, 43615

United States, Oklahoma

[Not yet recruiting]
Oklahoma City, Oklahoma, United States, 73104

[Not yet recruiting]
Tulsa, Oklahoma, United States, 74104

United States, Oregon

[Recruiting]
Ashland, Oregon, United States, 97520

[Not yet recruiting]
Portland, Oregon, United States, 97210

[Recruiting]
Portland, Oregon, United States, 97210

[Not yet recruiting]
Portland, Oregon, United States, 97227

[Not yet recruiting]
Salem, Oregon, United States, 97302

United States, Pennsylvania

[Not yet recruiting]
Camp Hill, Pennsylvania, United States, 17011

[Not yet recruiting]

Campbell, Pennsylvania, United States, 95008

[Not yet recruiting]

East Stroudsburg, Pennsylvania, United States, 18301

[Recruiting]

Kingston, Pennsylvania, United States, 18704

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19104

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19107

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19124

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15212

[Recruiting]

Pittsburgh, Pennsylvania, United States, 15213

[Not yet recruiting]

Pittsburgh, Pennsylvania, United States, 15219

[Recruiting]

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

[Recruiting]

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

[Recruiting]

Columbia, South Carolina, United States, 29204

[Recruiting]

Columbia, South Carolina, United States, 29223

[Not yet recruiting]

Greenville, South Carolina, United States, 29605

[Not yet recruiting]

Rock Hill, South Carolina, United States, 29732

United States, South Dakota

[Recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Recruiting]

Memphis, Tennessee, United States, 38119

[Not yet recruiting]

Memphis, Tennessee, United States, 38120

[Recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Recruiting]

Abilene, Texas, United States, 79606

[Recruiting]

Austin, Texas, United States, 78705

[Recruiting]

DeSoto, Texas, United States, 75115

[Recruiting]

Ft. Worth, Texas, United States, 76102

[Not yet recruiting]

Ft. Worth, Texas, United States, 76104

[Not yet recruiting]

Galveston, Texas, United States, 77555

[Recruiting]

Houston, Texas, United States, 77030

[Recruiting]

McAllen, Texas, United States, 78503

[Not yet recruiting]

Odessa, Texas, United States, 79761

[Recruiting]

San Antonio, Texas, United States, 78133

[Recruiting]

San Antonio, Texas, United States, 78240

United States, Utah

[Not yet recruiting]

Ogden, Utah, United States, 84403

[Recruiting]

Salt Lake City, Utah, United States, 84107

[Not yet recruiting]

Salt Lake City, Utah, United States, 84132

United States, Vermont

[Not yet recruiting]

Burlington, Vermont, United States, 05401

United States, Virginia

[Not yet recruiting]

Charlottesville, Virginia, United States, 22908

[Recruiting]

Richmond, Virginia, United States, 23221

United States, Washington

[Recruiting]

Seattle, Washington, United States, 98104

[Not yet recruiting]

Seattle, Washington, United States, 98195

[Recruiting]

Silverdale, Washington, United States, 98383

[Not yet recruiting]

Spokane, Washington, United States, 99203

United States, Wisconsin

[Not yet recruiting]

Madison, Wisconsin, United States, 53715

[Not yet recruiting]

Madison, Wisconsin, United States, 58705

[Not yet recruiting]

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

[Not yet recruiting]

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

[Not yet recruiting]

Vancouver, British Columbia, Canada, V5Z 3N9

[Not yet recruiting]

Victoria, British Columbia, Canada, V8X 5G6

Canada, Nova Scotia

[Not yet recruiting]

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

[Not yet recruiting]

London, Ontario, Canada, N6A 4G5

[Not yet recruiting]

Mississauga, Ontario, Canada, L4W 1W9

[Not yet recruiting]

Toronto, Ontario, Canada, M5C 2T2

[Recruiting]

Toronto, Ontario, Canada, M5T 2S8

Canada, Quebec

[Not yet recruiting]

Montreal, Quebec, Canada, H1T 2M4

[Not yet recruiting]

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

[Not yet recruiting]

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00509795

Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: August 17, 2007 (v2)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: August 2007

Overall Status: Recruiting

Study Start: August 2007

Primary Completion:

Study Completion:

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that August 17, 2007

Met QC Criteria:

Last Update Posted: August 21, 2007 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms:

Masking: Double (masked roles unspecified)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Intervention Details:

Drug: VEGF Trap-Eye

Outcome Measures

Primary Outcome Measures:

1. Primary measure will be visual acuity changes compared to baseline

Secondary Outcome Measures:

2. Secondary measures will be angiographic and anatomical changes compared to baseline.

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.

6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents.
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV, including pathologic myopia (spherical equivalent of -8 diopters or more negative, or axial length of 25 mm or more), ocular histoplasmosis syndrome, angioid streaks, choroidal rupture, or multifocal choroiditis in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Central Contact: Regeneron

Email: vegf.trap@regeneron.com

Study Officials: Avner Ingerman, MD

Study Director

Regeneron Pharmaceuticals

Locations: **United States, Alabama**

[Not yet recruiting]

Birmingham, Alabama, United States, 35205

[Not yet recruiting]

Birmingham, Alabama, United States, 35223

United States, Arizona

[Not yet recruiting]

Phoenix, Arizona, United States, 85014

[Not yet recruiting]

Phoenix, Arizona, United States, 85020

[Not yet recruiting]

Tucson, Arizona, United States, 85704

[Not yet recruiting]

Tucson, Arizona, United States, 85710

[Not yet recruiting]

Tucson, Arizona, United States, 85711

United States, California

[Not yet recruiting]

Beverly Hills, California, United States, 90211

[Not yet recruiting]

Fullerton, California, United States, 92835

[Not yet recruiting]

Glendale, California, United States, 91203

[Not yet recruiting]
Irvine, California, United States, 92697

[Not yet recruiting]
La Jolla, California, United States, 92037

[Not yet recruiting]
Loma Linda, California, United States, 92354

[Not yet recruiting]
Los Angeles, California, United States, 90033

[Not yet recruiting]
Los Angeles, California, United States, 90048

[Not yet recruiting]
Menlo Park, California, United States, 94025

[Not yet recruiting]
Mountain View, California, United States, 94040

[Not yet recruiting]
Oakland, California, United States, 94609

[Not yet recruiting]
Palm Springs, California, United States, 92262

[Not yet recruiting]
Pasadena, California, United States, 91105

[Not yet recruiting]
Poway, California, United States, 92064

[Not yet recruiting]
Sacramento, California, United States, 95819

[Not yet recruiting]
San Diego, California, United States, 92120

[Not yet recruiting]
San Francisco, California, United States, 94107

[Not yet recruiting]

San Marino, California, United States, 91108

[Not yet recruiting]

Santa Ana, California, United States, 92705

[Not yet recruiting]

Ventura, California, United States, 93003

[Not yet recruiting]

Westlake Village, California, United States, 91361

[Not yet recruiting]

Yorba Linda, California, United States, 92887

United States, Colorado

[Not yet recruiting]

Aurora, Colorado, United States, 80045

[Not yet recruiting]

Denver, Colorado, United States, 80205

[Not yet recruiting]

Denver, Colorado, United States, 80230

United States, Connecticut

[Not yet recruiting]

Bridgeport, Connecticut, United States, 06606

[Not yet recruiting]

Hamden, Connecticut, United States, 06518

[Not yet recruiting]

New Haven, Connecticut, United States, 06520

[Not yet recruiting]

New London, Connecticut, United States, 06320

United States, Florida

[Not yet recruiting]

Altamonte Springs, Florida, United States, 32701

[Not yet recruiting]

Boynton Beach, Florida, United States, 33426

[Not yet recruiting]

Delray Beach, Florida, United States, 33484

[Not yet recruiting]

Fort Myers, Florida, United States, 33907

[Not yet recruiting]

Ft. Lauderdale, Florida, United States, 33334

[Not yet recruiting]

Ft. Myers, Florida, United States, 33912

[Not yet recruiting]

Gainesville, Florida, United States, 32610

[Not yet recruiting]

Jacksonville, Florida, United States, 32216

[Not yet recruiting]

Jacksonville, Florida, United States, 32224

[Not yet recruiting]

Margate, Florida, United States, 33063

[Not yet recruiting]

Miami, Florida, United States, 33125

[Not yet recruiting]

Miami, Florida, United States, 33136

[Not yet recruiting]

Mount Dora, Florida, United States, 32757

[Not yet recruiting]

Ocala, Florida, United States, 34472

[Not yet recruiting]

Orlando, Florida, United States, 32806

[Not yet recruiting]

Orlando, Florida, United States, 38203

[Not yet recruiting]

Palm Beach Gardens, Florida, United States, 33410

[Not yet recruiting]

Pensacola, Florida, United States, 32503

[Not yet recruiting]

Sarasota, Florida, United States, 34239

[Not yet recruiting]

Stuart, Florida, United States, 34994

[Not yet recruiting]

Sunrise, Florida, United States, 33351

[Not yet recruiting]

Tampa, Florida, United States, 33607

[Not yet recruiting]

Tampa, Florida, United States, 33612

[Not yet recruiting]

Winter Haven, Florida, United States, 33880

United States, Georgia

[Not yet recruiting]

Augusta, Georgia, United States, 30909

[Not yet recruiting]

Marietta, Georgia, United States, 30060

United States, Hawaii

[Not yet recruiting]

Aiea, Hawaii, United States, 96701

[Not yet recruiting]

Honolulu, Hawaii, United States, 96813

United States, Illinois

[Not yet recruiting]

Chicago, Illinois, United States, 60611

[Not yet recruiting]

Chicago, Illinois, United States, 60637

[Not yet recruiting]

Oak Brook, Illinois, United States, 60523

[Not yet recruiting]

Springfield, Illinois, United States, 62701

[Not yet recruiting]

Wheaton, Illinois, United States, 60187

United States, Indiana

[Not yet recruiting]

Ft. Wayne, Indiana, United States, 46804

[Not yet recruiting]

Indianapolis, Indiana, United States, 46202

[Not yet recruiting]

Indianapolis, Indiana, United States, 46260

[Not yet recruiting]

Indianapolis, Indiana, United States, 46280

[Not yet recruiting]

New Albany, Indiana, United States, 47150

United States, Iowa

[Not yet recruiting]

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

[Not yet recruiting]

Wichita, Kansas, United States, 67214

United States, Kentucky

[Not yet recruiting]

Louisville, Kentucky, United States, 40202

[Not yet recruiting]

Louisville, Kentucky, United States, 40207

[Not yet recruiting]

Paducah, Kentucky, United States, 42001

United States, Louisiana

[Not yet recruiting]

Metairie, Louisiana, United States, 70002

[Not yet recruiting]

New Orleans, Louisiana, United States, 70115

[Not yet recruiting]

New Orleans, Louisiana, United States, 70121

[Not yet recruiting]

Shreveport, Louisiana, United States, 71105

United States, Maine

[Not yet recruiting]

Bangor, Maine, United States, 04401

United States, Maryland

[Not yet recruiting]

Baltimore, Maryland, United States, 21209

[Not yet recruiting]

Baltimore, Maryland, United States, 21287

[Not yet recruiting]

Chevy Chase, Maryland, United States, 20815

[Not yet recruiting]

Hagerstown, Maryland, United States, 21740

[Not yet recruiting]

Towson, Maryland, United States, 21204

United States, Massachusetts

[Not yet recruiting]

Boston, Massachusetts, United States, 02111

[Not yet recruiting]

Boston, Massachusetts, United States, 02114

[Not yet recruiting]

Boston, Massachusetts, United States, 02215

[Not yet recruiting]

Peabody, Massachusetts, United States, 01960

United States, Michigan

[Not yet recruiting]

Ann Arbor, Michigan, United States, 48105

[Not yet recruiting]

Battle Creek, Michigan, United States, 49015

[Not yet recruiting]

East Lansing, Michigan, United States, 48823

[Not yet recruiting]

Grand Blanc, Michigan, United States, 48439

[Not yet recruiting]

Grand Rapids, Michigan, United States, 49525

[Not yet recruiting]

Jackson, Michigan, United States, 49201

[Not yet recruiting]

Royal Oak, Michigan, United States, 48073

[Not yet recruiting]

Southfield, Michigan, United States, 48034

[Not yet recruiting]

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

[Not yet recruiting]

Edina, Minnesota, United States, 55435

[Not yet recruiting]

Minneapolis, Minnesota, United States, 55404

[Not yet recruiting]

Rochester, Minnesota, United States, 55905

United States, Missouri

[Not yet recruiting]

Florissant, Missouri, United States, 63031

[Not yet recruiting]

Kansas City, Missouri, United States, 64108

[Not yet recruiting]

Kansas City, Missouri, United States, 64111

[Not yet recruiting]

Springfield, Missouri, United States, 65804

[Not yet recruiting]

St. Louis, Missouri, United States, 63104

[Not yet recruiting]

St. Louis, Missouri, United States, 63110

United States, Montana

[Not yet recruiting]

Missoula, Montana, United States, 59801

United States, Nebraska

[Not yet recruiting]

Lincoln, Nebraska, United States, 68506

[Not yet recruiting]

Omaha, Nebraska, United States, 68124

[Not yet recruiting]

Omaha, Nebraska, United States, 68131

United States, Nevada

[Not yet recruiting]

Las Vegas, Nevada, United States, 89144

United States, New Hampshire

[Not yet recruiting]

Lebanon, New Hampshire, United States, 03756

United States, New Jersey

[Not yet recruiting]

Lawrenceville, New Jersey, United States, 08648

[Not yet recruiting]

Newark, New Jersey, United States, 07103

[Not yet recruiting]

Northfield, New Jersey, United States, 08225

[Not yet recruiting]

Teaneck, New Jersey, United States, 07666

[Not yet recruiting]

Toms River, New Jersey, United States, 08753

United States, New Mexico

[Not yet recruiting]

Albuquerque, New Mexico, United States, 87106

United States, New York

[Not yet recruiting]

Albany, New York, United States, 12206

[Not yet recruiting]

Bronxville, New York, United States, 10708

[Not yet recruiting]

Bronx, New York, United States, 10467

[Not yet recruiting]

Brooklyn, New York, United States, 11223

[Not yet recruiting]

Great Neck, New York, United States, 11021

[Not yet recruiting]

Lynbrook, New York, United States, 11563

[Not yet recruiting]

New York, New York, United States, 10003

[Not yet recruiting]

New York, New York, United States, 10021

[Not yet recruiting]

New York, New York, United States, 10032

[Not yet recruiting]

Rochester, New York, United States, 14618

[Not yet recruiting]

Rochester, New York, United States, 14642

[Not yet recruiting]

Slingerlands, New York, United States, 12159

United States, North Carolina

[Not yet recruiting]

Asheville, North Carolina, United States, 28803

[Recruiting]

Charlotte, North Carolina, United States, 28210

[Not yet recruiting]

Hendersonville, North Carolina, United States, 28791

[Not yet recruiting]

Raleigh, North Carolina, United States, 27607

[Not yet recruiting]

Raleigh, North Carolina, United States, 27612

[Not yet recruiting]

Southern Pines, North Carolina, United States, 28387

[Not yet recruiting]

Winston-Salem, North Carolina, United States, 27157

United States, North Dakota

[Not yet recruiting]

Fargo, North Dakota, United States, 58103

United States, Ohio

[Not yet recruiting]

Canton, Ohio, United States, 44718

[Not yet recruiting]

Cincinnati, Ohio, United States, 45202

[Not yet recruiting]

Cincinnati, Ohio, United States, 45242

[Not yet recruiting]

Cleveland, Ohio, United States, 44122

[Not yet recruiting]

Cleveland, Ohio, United States, 44195

[Not yet recruiting]

Columbus, Ohio, United States, 43215

[Not yet recruiting]

Toledo, Ohio, United States, 43606

[Not yet recruiting]

Toledo, Ohio, United States, 43608

[Not yet recruiting]

Toledo, Ohio, United States, 43615

United States, Oklahoma

[Not yet recruiting]

Oklahoma City, Oklahoma, United States, 73104

[Not yet recruiting]

Tulsa, Oklahoma, United States, 74104

United States, Oregon

[Not yet recruiting]

Ashland, Oregon, United States, 97520

[Not yet recruiting]

Portland, Oregon, United States, 97210

[Not yet recruiting]

Portland, Oregon, United States, 97227

[Not yet recruiting]

Salem, Oregon, United States, 97302

United States, Pennsylvania

[Not yet recruiting]

Camp Hill, Pennsylvania, United States, 17011

[Not yet recruiting]

Campbell, Pennsylvania, United States, 95008

[Not yet recruiting]

East Stroudsburg, Pennsylvania, United States, 18301

[Not yet recruiting]

Kingston, Pennsylvania, United States, 18704

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19104

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19107

[Not yet recruiting]

Philadelphia, Pennsylvania, United States, 19124

[Not yet recruiting]

Pittsburgh, Pennsylvania, United States, 15212

[Not yet recruiting]

Pittsburgh, Pennsylvania, United States, 15213

[Not yet recruiting]

Pittsburgh, Pennsylvania, United States, 15219

[Not yet recruiting]

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

[Not yet recruiting]

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

[Not yet recruiting]

Columbia, South Carolina, United States, 29204

[Not yet recruiting]

Columbia, South Carolina, United States, 29223

[Not yet recruiting]

Greenville, South Carolina, United States, 29605

[Not yet recruiting]

Rock Hill, South Carolina, United States, 29732

United States, South Dakota

[Not yet recruiting]

Rapid City, South Dakota, United States, 57701

United States, Tennessee

[Not yet recruiting]

Memphis, Tennessee, United States, 38119

[Not yet recruiting]

Memphis, Tennessee, United States, 38120

[Not yet recruiting]

Nashville, Tennessee, United States, 37203

United States, Texas

[Not yet recruiting]

Abilene, Texas, United States, 79606

[Not yet recruiting]

Austin, Texas, United States, 78705

[Not yet recruiting]

DeSoto, Texas, United States, 75115

[Not yet recruiting]

Ft. Worth, Texas, United States, 76102

[Not yet recruiting]

Ft. Worth, Texas, United States, 76104

[Not yet recruiting]

Galveston, Texas, United States, 77555

[Recruiting]

Houston, Texas, United States, 77030

[Not yet recruiting]

McAllen, Texas, United States, 78503

[Not yet recruiting]

Odessa, Texas, United States, 79761

[Not yet recruiting]

San Antonio, Texas, United States, 78133

[Not yet recruiting]

San Antonio, Texas, United States, 78240

United States, Utah

[Not yet recruiting]

Ogden, Utah, United States, 84403

[Not yet recruiting]

Salt Lake City, Utah, United States, 84107

[Not yet recruiting]

Salt Lake City, Utah, United States, 84132

United States, Vermont

[Not yet recruiting]

Burlington, Vermont, United States, 05401

United States, Virginia

[Not yet recruiting]

Charlottesville, Virginia, United States, 22908

[Not yet recruiting]

Richmond, Virginia, United States, 23221

United States, Washington

[Not yet recruiting]

Seattle, Washington, United States, 98104

[Not yet recruiting]

Seattle, Washington, United States, 98195

[Not yet recruiting]

Silverdale, Washington, United States, 98383

[Not yet recruiting]

Spokane, Washington, United States, 99203

United States, Wisconsin

[Not yet recruiting]

Madison, Wisconsin, United States, 53715

[Not yet recruiting]

Madison, Wisconsin, United States, 58705

[Not yet recruiting]

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

[Not yet recruiting]

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

[Not yet recruiting]

Vancouver, British Columbia, Canada, V5Z 3N9

[Not yet recruiting]

Victoria, British Columbia, Canada, V8X 5G6

Canada, Nova Scotia

[Not yet recruiting]

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

[Not yet recruiting]

London, Ontario, Canada, N6A 4G5

[Not yet recruiting]

Mississauga, Ontario, Canada, L4W 1W9

[Not yet recruiting]

Toronto, Ontario, Canada, M5C 2T2

[Not yet recruiting]

Toronto, Ontario, Canada, M5T 2S8

Canada, Quebec

[Not yet recruiting]

Montreal, Quebec, Canada, H1T 2M4

[Not yet recruiting]

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

[Not yet recruiting]

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

ClinicalTrials.gov archive

History of Changes for Study: NCT00509795

Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Latest version (submitted December 20, 2012) on ClinicalTrials.gov

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status

Version	A	B	Submitted Date	Changes
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input type="radio"/>	<input type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format:

- Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: July 31, 2007 (v1)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Double-Masked Study of Efficacy and Safety of IVT VEGF Trap-Eye in Subjects With Wet AMD (VIEW 1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: July 2007

Overall Status: Not yet recruiting

Study Start: August 2007

Primary Completion:

Study Completion:

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Last Update Submitted that July 31, 2007

Met QC Criteria:

Last Update Posted: August 1, 2007 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party:

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase III, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Neovascular Age-Related Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms:

Masking: Double (masked roles unspecified)

Allocation: Randomized

Enrollment: 1200 [Anticipated]

Arms and Interventions

Intervention Details:

Drug: VEGF Trap-Eye

Outcome Measures

Primary Outcome Measures:

1. Primary measure will be visual acuity changes compared to baseline

Secondary Outcome Measures:

2. Secondary measures will be angiographic and anatomical changes compared to baseline.

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Key Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women \geq 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. ETDRS best-corrected visual acuity of: 20/40 to 20/320 (letter score of 73 to 25) in the study eye.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.

6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents.
4. Total lesion size > 12 disc areas (30.5 mm², including blood, scars and neovascularization) as assessed by FA in the study eye.
5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV, including pathologic myopia (spherical equivalent of -8 diopters or more negative, or axial length of 25 mm or more), ocular histoplasmosis syndrome, angioid streaks, choroidal rupture, or multifocal choroiditis in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Study Officials: Avner Ingerman, MD
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Alabama**

Birmingham, Alabama, United States, 35205

Birmingham, Alabama, United States, 35223

United States, Arizona

Phoenix, Arizona, United States, 85014

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

Tucson, Arizona, United States, 85710

Tucson, Arizona, United States, 85711

United States, California

Beverly Hills, California, United States, 90211

Fullerton, California, United States, 92835

Glendale, California, United States, 91203

Irvine, California, United States, 92697

La Jolla, California, United States, 92037

Loma Linda, California, United States, 92354

Los Angeles, California, United States, 90033

Los Angeles, California, United States, 90048

Menlo Park, California, United States, 94025

Mountain View, California, United States, 94040

Oakland, California, United States, 94609

Palm Springs, California, United States, 92262

Pasadena, California, United States, 91105
Poway, California, United States, 92064
Sacramento, California, United States, 95819
San Diego, California, United States, 92120
San Francisco, California, United States, 94107
San Marino, California, United States, 91108
Santa Ana, California, United States, 92705
Ventura, California, United States, 93003
Westlake Village, California, United States, 91361
Yorba Linda, California, United States, 92887

United States, Colorado

Aurora, Colorado, United States, 80045
Denver, Colorado, United States, 80205
Denver, Colorado, United States, 80230

United States, Connecticut

Bridgeport, Connecticut, United States, 06606
Hamden, Connecticut, United States, 06518
New Haven, Connecticut, United States, 06520
New London, Connecticut, United States, 06320

United States, Florida

Altamonte Springs, Florida, United States, 32701
Boynton Beach, Florida, United States, 33426
Delray Beach, Florida, United States, 33484
Fort Myers, Florida, United States, 33907

Ft. Lauderdale, Florida, United States, 33334
Ft. Myers, Florida, United States, 33912
Gainesville, Florida, United States, 32610
Jacksonville, Florida, United States, 32216
Jacksonville, Florida, United States, 32224
Margate, Florida, United States, 33063
Miami, Florida, United States, 33125
Miami, Florida, United States, 33136
Mount Dora, Florida, United States, 32757
Ocala, Florida, United States, 34472
Orlando, Florida, United States, 32806
Orlando, Florida, United States, 38203
Palm Beach Gardens, Florida, United States, 33410
Pensacola, Florida, United States, 32503
Sarasota, Florida, United States, 34239
Stuart, Florida, United States, 34994
Sunrise, Florida, United States, 33351
Tampa, Florida, United States, 33607
Tampa, Florida, United States, 33612
Winter Haven, Florida, United States, 33880

United States, Georgia

Augusta, Georgia, United States, 30909
Marietta, Georgia, United States, 30060

United States, Hawaii

Aiea, Hawaii, United States, 96701

Honolulu, Hawaii, United States, 96813

United States, Illinois

Chicago, Illinois, United States, 60611

Chicago, Illinois, United States, 60637

Oak Brook, Illinois, United States, 60523

Springfield, Illinois, United States, 62701

Wheaton, Illinois, United States, 60187

United States, Indiana

Ft. Wayne, Indiana, United States, 46804

Indianapolis, Indiana, United States, 46202

Indianapolis, Indiana, United States, 46260

Indianapolis, Indiana, United States, 46280

New Albany, Indiana, United States, 47150

United States, Iowa

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

Wichita, Kansas, United States, 67214

United States, Kentucky

Louisville, Kentucky, United States, 40202

Louisville, Kentucky, United States, 40207

Paducah, Kentucky, United States, 42001

United States, Louisiana

Metairie, Louisiana, United States, 70002

New Orleans, Louisiana, United States, 70115

New Orleans, Louisiana, United States, 70121

Shreveport, Louisiana, United States, 71105

United States, Maine

Bangor, Maine, United States, 04401

United States, Maryland

Baltimore, Maryland, United States, 21209

Baltimore, Maryland, United States, 21287

Chevy Chase, Maryland, United States, 20815

Hagerstown, Maryland, United States, 21740

Towson, Maryland, United States, 21204

United States, Massachusetts

Boston, Massachusetts, United States, 02111

Boston, Massachusetts, United States, 02114

Boston, Massachusetts, United States, 02215

Peabody, Massachusetts, United States, 01960

United States, Michigan

Ann Arbor, Michigan, United States, 48105

Battle Creek, Michigan, United States, 49015

East Lansing, Michigan, United States, 48823

Grand Blanc, Michigan, United States, 48439

Grand Rapids, Michigan, United States, 49525

Jackson, Michigan, United States, 49201

Royal Oak, Michigan, United States, 48073

Southfield, Michigan, United States, 48034

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

Edina, Minnesota, United States, 55435

Minneapolis, Minnesota, United States, 55404

Rochester, Minnesota, United States, 55905

United States, Missouri

Florissant, Missouri, United States, 63031

Kansas City, Missouri, United States, 64108

Kansas City, Missouri, United States, 64111

Springfield, Missouri, United States, 65804

St. Louis, Missouri, United States, 63104

St. Louis, Missouri, United States, 63110

United States, Montana

Missoula, Montana, United States, 59801

United States, Nebraska

Lincoln, Nebraska, United States, 68506

Omaha, Nebraska, United States, 68124

Omaha, Nebraska, United States, 68131

United States, Nevada

Las Vegas, Nevada, United States, 89144

United States, New Hampshire

Lebanon, New Hampshire, United States, 03756

United States, New Jersey

Lawrenceville, New Jersey, United States, 08648

Newark, New Jersey, United States, 07103

Northfield, New Jersey, United States, 08225

Teaneck, New Jersey, United States, 07666

Toms River, New Jersey, United States, 08753

United States, New Mexico

Albuquerque, New Mexico, United States, 87106

United States, New York

Albany, New York, United States, 12206

Bronxville, New York, United States, 10708

Bronx, New York, United States, 10467

Brooklyn, New York, United States, 11223

Great Neck, New York, United States, 11021

Lynbrook, New York, United States, 11563

New York, New York, United States, 10003

New York, New York, United States, 10021

New York, New York, United States, 10032

Rochester, New York, United States, 14618

Rochester, New York, United States, 14642

Slingerlands, New York, United States, 12159

United States, North Carolina

Asheville, North Carolina, United States, 28803

Charlotte, North Carolina, United States, 28210

Hendersonville, North Carolina, United States, 28791

Raleigh, North Carolina, United States, 27607

Raleigh, North Carolina, United States, 27612

Southern Pines, North Carolina, United States, 28387

Winston-Salem, North Carolina, United States, 27157

United States, North Dakota

Fargo, North Dakota, United States, 58103

United States, Ohio

Canton, Ohio, United States, 44718

Cincinnati, Ohio, United States, 45202

Cincinnati, Ohio, United States, 45242

Cleveland, Ohio, United States, 44122

Cleveland, Ohio, United States, 44195

Columbus, Ohio, United States, 43215

Toledo, Ohio, United States, 43606

Toledo, Ohio, United States, 43608

Toledo, Ohio, United States, 43615

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

Tulsa, Oklahoma, United States, 74104

United States, Oregon

Ashland, Oregon, United States, 97520

Portland, Oregon, United States, 97210

Portland, Oregon, United States, 97227

Salem, Oregon, United States, 97302

United States, Pennsylvania

Camp Hill, Pennsylvania, United States, 17011
Campbell, Pennsylvania, United States, 95008
East Stroudsburg, Pennsylvania, United States, 18301
Kingston, Pennsylvania, United States, 18704
Philadelphia, Pennsylvania, United States, 19104
Philadelphia, Pennsylvania, United States, 19107
Philadelphia, Pennsylvania, United States, 19124
Pittsburgh, Pennsylvania, United States, 15212
Pittsburgh, Pennsylvania, United States, 15213
Pittsburgh, Pennsylvania, United States, 15219
Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

Columbia, South Carolina, United States, 29204
Columbia, South Carolina, United States, 29223
Greenville, South Carolina, United States, 29605
Rock Hill, South Carolina, United States, 29732

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Memphis, Tennessee, United States, 38119
Memphis, Tennessee, United States, 38120

Nashville, Tennessee, United States, 37203

United States, Texas

Abilene, Texas, United States, 79606

Austin, Texas, United States, 78705

DeSoto, Texas, United States, 75115

Ft. Worth, Texas, United States, 76102

Ft. Worth, Texas, United States, 76104

Galveston, Texas, United States, 77555

Houston, Texas, United States, 77030

McAllen, Texas, United States, 78503

Odessa, Texas, United States, 79761

San Antonio, Texas, United States, 78133

San Antonio, Texas, United States, 78240

United States, Utah

Ogden, Utah, United States, 84403

Salt Lake City, Utah, United States, 84107

Salt Lake City, Utah, United States, 84132

United States, Vermont

Burlington, Vermont, United States, 05401

United States, Virginia

Charlottesville, Virginia, United States, 22908

Richmond, Virginia, United States, 23221

United States, Washington

Seattle, Washington, United States, 98104

Seattle, Washington, United States, 98195

Silverdale, Washington, United States, 98383

Spokane, Washington, United States, 99203

United States, Wisconsin

Madison, Wisconsin, United States, 53715

Madison, Wisconsin, United States, 58705

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

Vancouver, British Columbia, Canada, V5Z 3N9

Victoria, British Columbia, Canada, V8X 5G6

Canada, Nova Scotia

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

London, Ontario, Canada, N6A 4G5

Mississauga, Ontario, Canada, L4W 1W9

Toronto, Ontario, Canada, M5C 2T2

Toronto, Ontario, Canada, M5T 2S8

Canada, Quebec

Montreal, Quebec, Canada, H1T 2M4

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links:

Available IPD/Information:

[Scroll up to access the controls](#)

[Scroll to the Study top](#)

[U.S. National Library of Medicine](#) | [U.S. National Institutes of Health](#) | [U.S. Department of Health & Human Services](#)

History of Changes for Study: NCT00509795

Vascular Endothelial Growth Factor VEGF Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

[Latest version \(submitted December 20, 2012\) on ClinicalTrials.gov](#)

- A study version is represented by a row in the table.
- Select two study versions to compare. One each from columns A and B.
- Choose either the "Merged" or "Side-by-Side" comparison format to specify how the two study versions are to be displayed. The Side-by-Side format only applies to the Protocol section of the study.
- Click "Compare" to do the comparison and show the differences.
- Select a version's Submitted Date link to see a rendering of the study for that version.
- The yellow A/B choices in the table indicate the study versions currently compared below. A yellow table row indicates the study version currently being viewed.
- Hover over the "Recruitment Status" to see how the study's recruitment status changed.
- Study edits or deletions are displayed in ~~red~~.
- Study additions are displayed in green.

Study Record Versions

Version	A	B	Submitted Date	Changes
1	<input type="radio"/>	<input type="radio"/>	<u>July 31, 2007</u>	None (earliest Version on record)
2	<input type="radio"/>	<input type="radio"/>	<u>August 17, 2007</u>	Recruitment Status, Study Status and Contacts/Locations

Version	A	B	Submitted Date	Changes
3	<input type="radio"/>	<input type="radio"/>	<u>November 14, 2007</u>	Contacts/Locations and Study Status
4	<input type="radio"/>	<input type="radio"/>	<u>December 4, 2007</u>	Study Status and Contacts/Locations
5	<input type="radio"/>	<input type="radio"/>	<u>March 13, 2008</u>	Study Status and Eligibility
6	<input type="radio"/>	<input type="radio"/>	<u>June 26, 2008</u>	Contacts/Locations, Arms and Interventions, Study Design, Study Status, Outcome Measures and Study Identification
7	<input type="radio"/>	<input type="radio"/>	<u>January 22, 2009</u>	Contacts/Locations, Study Status, Arms and Interventions, Outcome Measures, Eligibility and Sponsor/Collaborators
8	<input type="radio"/>	<input type="radio"/>	<u>March 3, 2009</u>	Study Status and Contacts/Locations
9	<input type="radio"/>	<input type="radio"/>	<u>April 28, 2009</u>	Outcome Measures, Arms and Interventions, Study Status, Eligibility, Conditions and Study Identification
10	<input type="radio"/>	<input type="radio"/>	<u>September 12, 2009</u>	Recruitment Status, Study Status and Contacts/Locations
11	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2009</u>	Study Status, Contacts/Locations and Sponsor/Collaborators
12	<input type="radio"/>	<input type="radio"/>	<u>January 5, 2011</u>	Study Status
13	<input type="radio"/>	<input type="radio"/>	<u>April 18, 2011</u>	Study Status, Study Design
14	<input type="radio"/>	<input type="radio"/>	<u>May 4, 2011</u>	Study Status
15	<input type="radio"/>	<input type="radio"/>	<u>December 1, 2011</u>	Recruitment Status, Study Status, Sponsor/Collaborators
16	<input type="radio"/>	<input type="radio"/>	<u>April 13, 2012</u>	Arms and Interventions, Outcome Measures, Study Status, More Information, Reported Adverse Events, Baseline Characteristics, Participant Flow, Eligibility, Study Description and Study Identification
17	<input type="radio"/>	<input type="radio"/>	<u>December 17, 2012</u>	Reported Adverse Events, Outcome Measures, Baseline Characteristics, Participant Flow, More Information and Study Status
18	<input checked="" type="radio"/>	<input checked="" type="radio"/>	<u>December 20, 2012</u>	More Information, Outcome Measures, References and Study Status

Compare

Comparison Format: Merged
 Side-by-Side

[Scroll up to access the controls](#)

Study NCT00509795

Submitted Date: December 20, 2012 (v18)

Study Identification

Unique Protocol ID: VGFT-OD-0605

Brief Title: Vascular Endothelial Growth Factor VEGF Trap-Eye:Investigation of Efficacy and Safety in Wet Age-Related Macular Degeneration(AMD) (VIEW1)

Official Title: A Randomized, Double Masked, Active Controlled Phase III Study of the Efficacy, Safety, and Tolerability of Repeated Doses of Intravitreal VEGF Trap in Subjects With Neovascular Age-Related Macular Degeneration

Secondary IDs:

Study Status

Record Verification: December 2012

Overall Status: Completed

Study Start: August 2007

Primary Completion: September 2010 [Actual]

Study Completion: July 2011 [Actual]

First Submitted: July 31, 2007

First Submitted that July 31, 2007

Met QC Criteria:

First Posted: August 1, 2007 [Estimate]

Results First Submitted: December 16, 2011

Results First Submitted that April 13, 2012

Met QC Criteria:

Results First Posted: April 16, 2012 [Estimate]

Certification/Extension January 5, 2011

First Submitted:

Certification/Extension January 5, 2011

First Submitted that

Met QC Criteria:

Certification/Extension January 10, 2011 [Estimate]

First Posted:

Last Update Submitted that December 20, 2012

Met QC Criteria:

Last Update Posted: December 28, 2012 [Estimate]

Sponsor/Collaborators

Sponsor: Regeneron Pharmaceuticals

Responsible Party: Sponsor

Collaborators: Bayer

Oversight

U.S. FDA-regulated Drug:

U.S. FDA-regulated Device:

Data Monitoring: Yes

Study Description

Brief Summary: This study is a phase 3, double-masked, randomized, study of the efficacy and safety of VEGF Trap-Eye in patients with neovascular age-related macular degeneration. Approximately 1200 patients will be randomized in the US and Canada.

Detailed Description:

Conditions

Conditions: Macular Degeneration

Keywords:

Study Design

Study Type: Interventional

Primary Purpose: Treatment

Study Phase: Phase 3

Interventional Study Model: Parallel Assignment

Number of Arms: 4

Masking: Quadruple (Participant, Care Provider, Investigator, Outcomes Assessor)

Allocation: Randomized

Enrollment: 1217 [Actual]

Arms and Interventions

Arms	Assigned Interventions
Active Comparator: ranibizumab 0.5mg Q4	Biological: ranibizumab Participants received a 0.5mg dose of ranibizumab via intravitreal (IVT) injection every 4 weeks for the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks. Other Names: <ul style="list-style-type: none">• Lucentis

Arms	Assigned Interventions
Experimental: aflibercept injection 2.0mg Q4	<p>Biological: aflibercept injection (VEGF Trap-Eye, BAY86-5321)</p> <p>Participants received a 2.0mg dose of aflibercept injection every 4 weeks for the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • VEGF Trap-Eye • BAY86-5321
Experimental: aflibercept injection 0.5mg Q4	<p>Biological: aflibercept injection (VEGF Trap-Eye, BAY86-5321)</p> <p>Participants received a 0.5mg dose of aflibercept injection every 4 weeks for the first year. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • VEGF Trap-Eye • BAY86-5321
Experimental: aflibercept injection 2.0mg Q8	<p>Biological: aflibercept injection (VEGF Trap-Eye, BAY86-5321)</p> <p>Participants received a 2.0mg dose of aflibercept injection every 8 weeks (including one additional 2.0 mg dose at Week 4) for the first year and were to receive sham injections at interim monthly visits. Thereafter a dose may be administered as frequently as every 4 weeks, but no less frequently than every 12 weeks.</p> <p>Other Names:</p> <ul style="list-style-type: none"> • VEGF Trap-Eye • BAY86-5321

Outcome Measures

Eligibility

Minimum Age: 50 Years

Maximum Age:

Sex: All

Gender Based:

Accepts Healthy Volunteers: No

Criteria: Inclusion Criteria:

1. Signed Informed Consent.
2. Men and women ≥ 50 years of age.
3. Active primary or recurrent subfoveal CNV lesions secondary to AMD, including juxtafoveal lesions that affect the fovea as evidenced by FA in the study eye.
4. Early Treatment Diabetic Retinopathy Study (ETDRS) Best Corrected Visual Acuity (BCVA) of: letter score of 73 to 25 (20/40 to 20/320) in the study eye at 4 meters.
5. Willing, committed, and able to return for ALL clinic visits and complete all study-related procedures.
6. Able to read, (or, if unable to read due to visual impairment, be read to verbatim by the person administering the informed consent or a family member. See Appendix J.4) understand and willing to sign the informed consent form.

Key

Exclusion Criteria:

1. Any prior ocular (in the study eye) or systemic treatment or surgery for neovascular AMD except dietary supplements or vitamins.
2. Any prior or concomitant therapy with another investigational agent to treat neovascular AMD in the study eye, except dietary supplements or vitamins.
3. Any prior treatment with anti-VEGF agents in the study eye.
4. Total lesion size > 12 disc areas (30.5 mm^2 , including blood, scars and neovascularization) as assessed by FA in the study eye.

5. Subretinal hemorrhage that is either 50% or more of the total lesion area, or if the blood is under the fovea and is 1 or more disc areas in size in the study eye. (If the blood is under the fovea, then the fovea must be surrounded 270 degrees by visible CNV.)
6. Scar or fibrosis, making up > 50% of total lesion in the study eye.
7. Scar, fibrosis, or atrophy involving the center of the fovea.
8. Presence of retinal pigment epithelial tears or rips involving the macula in the study eye.
9. History of any vitreous hemorrhage within 4 weeks prior to Visit 1 in the study eye.
10. Presence of other causes of CNV in the study eye.
11. History or clinical evidence of diabetic retinopathy, diabetic macular edema or any other vascular disease affecting the retina, other than AMD, in either eye.
12. Prior vitrectomy in the study eye.
13. History of retinal detachment or treatment or surgery for retinal detachment in the study eye.
14. Any history of macular hole of stage 2 and above in the study eye.
15. Any intraocular or periocular surgery within 3 months of Day 1 on the study eye, except lid surgery, which may not have taken place within 1 month of day 1, as long as its unlikely to interfere with the injection.

Contacts/Locations

Study Officials: Robert Vitti, MD
Study Director
Regeneron Pharmaceuticals

Locations: **United States, Alabama**

Birmingham, Alabama, United States, 35205

Birmingham, Alabama, United States, 35223

United States, Arizona

Phoenix, Arizona, United States, 85014

Phoenix, Arizona, United States, 85020

Tucson, Arizona, United States, 85704

Tucson, Arizona, United States, 85710

United States, California

Beverly Hills, California, United States, 90211
Campbell, California, United States, 95008
Fullerton, California, United States, 92835
Glendale, California, United States, 91203
Irvine, California, United States, 92697
La Jolla, California, United States, 92037
Loma Linda, California, United States, 92354
Los Angeles, California, United States, 90033
Los Angeles, California, United States, 90048
Menlo Park, California, United States, 94025
Mountain View, California, United States, 94040
Oakland, California, United States, 94609
Palm Springs, California, United States, 92262
Pasadena, California, United States, 91105
Poway, California, United States, 92064
Sacramento, California, United States, 95819
San Diego, California, United States, 92120
San Francisco, California, United States, 94107
Santa Ana, California, United States, 92705
Torrance, California, United States, 90503
Ventura, California, United States, 93003
Westlake Village, California, United States, 91361
Yorba Linda, California, United States, 92887

United States, Colorado

Aurora, Colorado, United States, 80045

Denver, Colorado, United States, 80205

Denver, Colorado, United States, 80230

United States, Connecticut

Bridgeport, Connecticut, United States, 06606

Hamden, Connecticut, United States, 06518

New Haven, Connecticut, United States, 06510

New London, Connecticut, United States, 06320

United States, Florida

Altamonte Springs, Florida, United States, 32701

Boynton Beach, Florida, United States, 33426

Fort Myers, Florida, United States, 33907

Ft. Lauderdale, Florida, United States, 33351

Ft. Myers, Florida, United States, 33912

Gainesville, Florida, United States, 32610

Jacksonville, Florida, United States, 32224

Miami, Florida, United States, 33136

Miami, Florida, United States, 33143

Mount Dora, Florida, United States, 32757

Orlando, Florida, United States, 32803

Orlando, Florida, United States, 32806

Oscala, Florida, United States, 34472

Palm Beach Gardens, Florida, United States, 33410

Pensacola, Florida, United States, 32503

Sarasota, Florida, United States

Stuart, Florida, United States, 34994

Tampa, Florida, United States, 33612

Winter Haven, Florida, United States, 33880

United States, Georgia

Augusta, Georgia, United States, 30909

United States, Hawaii

Aiea, Hawaii, United States, 96701

Honolulu, Hawaii, United States, 96813

United States, Illinois

Oak Brook, Illinois, United States, 60523

United States, Indiana

Fort Wayne, Indiana, United States, 46804

Indianapolis, Indiana, United States, 46202

Indianapolis, Indiana, United States, 46260

Indianapolis, Indiana, United States, 46280

New Albany, Indiana, United States, 47150

United States, Iowa

Iowa City, Iowa, United States, 52242-1091

United States, Kansas

Wichita, Kansas, United States, 67214

United States, Kentucky

Louisville, Kentucky, United States, 40202

Louisville, Kentucky, United States, 40207

Paducah, Kentucky, United States, 42001

United States, Louisiana

New Orleans, Louisiana, United States, 70115

New Orleans, Louisiana, United States, 70121

Shreveport, Louisiana, United States, 71105

United States, Maine

Bangor, Maine, United States, 04401

Portland, Maine, United States, 04102

United States, Maryland

Baltimore, Maryland, United States, 21209

Baltimore, Maryland, United States, 21287

Chevy Chase, Maryland, United States, 20815

Hagerstown, Maryland, United States, 21740

Towson, Maryland, United States, 21204

United States, Massachusetts

Boston, Massachusetts, United States, 02111

Boston, Massachusetts, United States, 02114

Boston, Massachusetts, United States, 02215

Boston, Massachusetts, United States

Peabody, Massachusetts, United States, 01960

United States, Michigan

Ann Arbor, Michigan, United States, 48105

Battle Creek, Michigan, United States, 49015

Detroit, Michigan, United States, 48202

Grand Rapids, Michigan, United States, 49525

Jackson, Michigan, United States, 49201

Royal Oak, Michigan, United States, 48073

Southfield, Michigan, United States, 48034

West Bloomfield, Michigan, United States, 48322

United States, Minnesota

Edina, Minnesota, United States, 55435

Minneapolis, Minnesota, United States, 55404

Rochester, Minnesota, United States, 55905

United States, Missouri

Florissant, Missouri, United States, 63031

Kansas City, Missouri, United States, 64108

Kansas City, Missouri, United States, 64111

Springfield, Missouri, United States, 65804

St. Louis, Missouri, United States, 63110

United States, Montana

Missoula, Montana, United States, 59801

United States, Nebraska

Lincoln, Nebraska, United States, 68506

Omaha, Nebraska, United States, 68131

United States, Nevada

Las Vegas, Nevada, United States, 89144

United States, New Jersey

Lawrenceville, New Jersey, United States, 08648

New Brunswick, New Jersey, United States, 08901

Northfield, New Jersey, United States, 08225

Teaneck, New Jersey, United States, 07666

Toms River, New Jersey, United States, 08753

United States, New Mexico

Albuquerque, New Mexico, United States, 87106

United States, New York

Albany, New York, United States, 12206

Brooklyn, New York, United States, 11223

Lynbrook, New York, United States, 11563

New York, New York, United States, 10003

New York, New York, United States, 10021

New York, New York, United States, 10032

Poughkeepsie, New York, United States, 12601

Rochester, New York, United States, 14620

Rochester, New York, United States, 14642

Slingerlands, New York, United States, 12159

Syracuse, New York, United States, 13224

United States, North Carolina

Asheville, North Carolina, United States, 28803

Charlotte, North Carolina, United States, 28210

Raleigh, North Carolina, United States, 27607

Southern Pines, North Carolina, United States, 28387

Winston-Salem, North Carolina, United States, 27157

United States, Ohio

Cincinnati, Ohio, United States, 45202

Cincinnati, Ohio, United States, 45242

Columbus, Ohio, United States, 43215

Toledo, Ohio, United States, 43608

United States, Oklahoma

Oklahoma City, Oklahoma, United States, 73104

United States, Oregon

Ashland, Oregon, United States, 97520

Portland, Oregon, United States, 97210

Portland, Oregon, United States, 97227

Salem, Oregon, United States, 97302

United States, Pennsylvania

Kingston, Pennsylvania, United States, 18704

Philadelphia, Pennsylvania, United States, 19104

Philadelphia, Pennsylvania, United States, 19107

Philadelphia, Pennsylvania, United States, 19124

Pittsberg, Pennsylvania, United States, 15231

Pittsburgh, Pennsylvania, United States, 15212

Pittsburgh, Pennsylvania, United States, 15213

West Mifflin, Pennsylvania, United States, 15122

Wyomissing, Pennsylvania, United States, 19610

United States, Rhode Island

Providence, Rhode Island, United States, 02903-4928

United States, South Carolina

Charleston, South Carolina, United States, 29414

Columbia, South Carolina, United States, 29223

Greenville, South Carolina, United States, 29605

West Columbia, South Carolina, United States, 29169

United States, South Dakota

Rapid City, South Dakota, United States, 57701

United States, Tennessee

Memphis, Tennessee, United States, 38119

Memphis, Tennessee, United States, 38120

Nashville, Tennessee, United States, 37203

United States, Texas

Abilene, Texas, United States, 79606

Austin, Texas, United States, 78705

Corpus Cristi, Texas, United States, 78413

Dallas, Texas, United States, 75390

DeSoto, Texas, United States, 75115

Ft. Worth, Texas, United States, 76102

Ft. Worth, Texas, United States, 76104

Galveston, Texas, United States, 77555

Houston, Texas, United States, 77030

McAllen, Texas, United States, 78503

Odessa, Texas, United States, 79761

San Antonio, Texas, United States, 78240

United States, Utah

Salt Lake City, Utah, United States, 84107

Salt Lake City, Utah, United States, 84132

United States, Vermont

Burlington, Vermont, United States, 05401

United States, Virginia

Charlottesville, Virginia, United States, 22908

Fairfax, Virginia, United States, 22031

Richmond, Virginia, United States, 23221

United States, Washington

Seattle, Washington, United States, 98104

Silverdale, Washington, United States, 98383

United States, Wisconsin

Madison, Wisconsin, United States, 53715

Madison, Wisconsin, United States, 58705

Milwaukee, Wisconsin, United States, 53226

Canada, Alberta

Calgary, Alberta, Canada, T3E 7MB

Canada, British Columbia

Vancouver, British Columbia, Canada, V5Z 3N9

Victoria, British Columbia, Canada, V8V 1B3

Canada, Nova Scotia

Halifax, Nova Scotia, Canada, B3H 2Y9

Canada, Ontario

London, Ontario, Canada, N6A 4G5

Mississauga, Ontario, Canada, L4W 1W9

Ottawa, Ontario, Canada, K1H8L6

Toronto, Ontario, Canada, M4N3M5

Toronto, Ontario, Canada, M5C 2T2

Canada, Quebec

Montreal, Quebec, Canada, H1T 2M4

Montreal, Quebec, Canada, H3A 1A1

Canada, Saskatchewan

Regina, Saskatchewan, Canada, S4T 1A5

IPDSharing

Plan to Share IPD:

References

Citations:

Links: URL: [http://www.aajournal.org/article/S0161-6420\(12\)00865-2/abstract](http://www.aajournal.org/article/S0161-6420(12)00865-2/abstract) #

Description: Related Info

Available IPD/Information:

Study Results

Participant Flow

Recruitment Details	The study was conducted at 164 sites in the United States and Canada. Recruitment period: 02 Aug 2007 to 15 Sep 2009.
---------------------	---

Pre-assignment Details	2063 patients were screened, 1217 randomized, and 1215 included in the Safety Analysis Set (SAF). The Full Analysis Set (FAS) included 1210 patients with at least 1 post-baseline assessment. The Per Protocol Set (PPS) included 1089 patients who received ≥ 9 doses of study drug and attended ≥ 9 scheduled visits during the first year.
------------------------	--

Reporting Groups

	Description
Ranibizumab 0.5mg Q4	Patients received a 0.5mg dose of ranibizumab via intravitreal (IVT) injection every 4 weeks for the first year.
IAI (EYLEA, VEGF Trap-Eye) 2.0mg Q4	Patients received a 2.0mg dose of Intravitreal Aflibercept Injection (IAI, EYLEA, VEGF Trap-Eye) every 4 weeks for the first year.
IAI (EYLEA, VEGF Trap-Eye) 0.5mg Q4	Patients received a 0.5mg dose of Intravitreal Aflibercept Injection (IAI, EYLEA, VEGF Trap-Eye) every 4 weeks for the first year.
IAI (EYLEA, VEGF Trap-Eye) 2.0mg Q8	Patients received a 2.0mg dose of Intravitreal Aflibercept Injection (IAI, EYLEA, VEGF Trap-Eye) every 8 weeks (including one additional 2.0 mg dose at Week 4) for the first year and received sham injections at interim monthly visits.

Overall Study