Amino Acids and Peptides

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Synthesis of amino acids

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6.1 General

There is an abundant supply of L-enantiomers of most of the coded amino acids. These are made available through large-scale fermentative production in most cases, and also through processing of protein hydrolysates. The early sections of this chapter cover this aspect, However, laboratory synthesis methods are required for the provision of most of the other natural amino acids and for all other amino acids, so the main part of this chapter deals with established syntheses.

6.2 Commercial and research uses for amino acids

In addition to the provision of supplies of common amino acids, there are growing needs for routes to new amino acids, since pharmaceutically useful compounds of this class continue to be discovered, which must be free from toxic impurities and homochirally pure in this particular context. Important functions for close analogues of coded and other biologically significant amino acids include *enzyme inhibi-tion* and retarding the growth of undesirable organisms (fungistatic, antibiotic and other physiological properties, possessed either by the free amino acids or by peptides containing them). Free amino acids that perform in this way are α -amino isobutyric acid (an example of an α -methylated analogue of a coded amino acid), which has been proposed for the control of domestic wood-rotting fungi), and α -methyl-Dopa (α -methyl-3',4'-dihydroxy-L-phenylalanine), a well-known treatment for Parkinson's disease. Similar success for new therapeutic amino acids, based on their enzyme-inhibition properties, is indicated for amino acids with a minimal structural change such as the substitution of a side-chain hydrogen atom by a fluorine atom.

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6.3 Biosynthesis

6.3 Biosynthesis: isolation of amino acids from natural sources

Many examples of the discovery and isolation of amino acids from natural sources date from the early 1900s, though some were characterised several years before that (Greenstein and Winitz, 1961). Further new examples continue to be discovered, either as constituents of proteins, revealing new post-translational processes for higher organisms (Table 1.3 in Chapter 1), or in the free or bound form (from fungal or bacterial sources or from marine organisms).

6.3.1 Isolation of amino acids from proteins

Hydrolysis of proteins and separation of the resulting mixture is an obvious, and traditional way (Greenstein and Winitz, 1961) of obtaining moderate quantities of the coded and post-translationally modified $L-\alpha$ -amino acids. However, because of the availability of viable methods of industrial synthesis, hydrolysis of proteins no longer offers a sensible approach owing to its tedious and expensive nature and the fact that some amino acids are destroyed in the process (see Chapter 3).

6.3.2 Biotechnological and industrial synthesis of coded amino acids

Knowledge gained of biosynthetic routes to L- α -amino acids and isolation of the enzymes mediating the steps in these routes has been exploited for the industrial-scale manufacture of most of the coded L- α -amino acids. In some cases, the enzymatic production of near-analogues of the coded L- α -amino acids can also be achieved (Goldberg and Williams, 1991; Rozzell and Wagner, 1992).

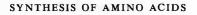
To illustrate the methods, a culture medium that contains indole, pyruvic acid, tyrosine phenollyase and an ammonium salt, as well as the usual buffers and salts, will accumulate L-tryptophan; or will produce an indole-substituted L-tryptophan if indole itself is replaced by a substituted indole. L-Dopa formed in a system employing tyrosinase from *Aspergillus terreus* provides a further example of this approach (Chattopadhyay and Das, 1990).

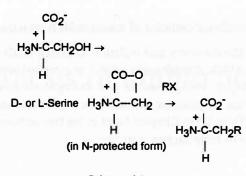
The crucial enzymes need not be isolated, since 'bio-reactors' containing microorganisms that are fed with the appropriate starting materials are often more convenient. L-Threonine from *Brevibacterium flavum*, L-lysine from *Corynebacterium glutamicum* (Eggeling, 1994) and use of plant-cell suspension cultures illustrated by L-Dopa from *Mucuna pruriens* (Wichers *et al.*, 1985) are examples. However, bioengineering of the whole organisms to be used in this way may need to be carefully optimised to achieve reasonable yields. The other main opportunity offered by biotechnological methods is the conversion of one amino acid into a less plentifully available amino acid, e.g. the conversion of L-tyrosine into L-Dopa using *Mucuna pruriens* (Wichers *et al.*, 1985).

For a limited range of amino acids, this approach is increasingly in competition

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Scheme 6.1.

with chemical synthesis, which can accomplish the necessary modifications in some cases more easily (Section 6.4). Examples of 'non-biotechnological' synthesis are provided by the industrial production of glutamic acid and lysine, conducted on a large scale (several thousand tons per year). DL-Glutamic acid is obtained from acrylonitrile, electrochemical reductive dimerisation and functional group modifications giving the DL compound. DL-Lysine is obtained from caprolactam, through its 3-amino-derivative, which is resolved (Scheme 6.6) with L-pyroglutamic acid before ring-opening to give L-lysine.

6.4 Synthesis of amino acids starting from coded amino acids other than glycine

With the easy availability of many of the natural amino acids, some general methods for the synthesis of more complex structures are based on the modification of simple natural amino acids. An important benefit from this approach is the fact that homochirality at the α -carbon atom can be preserved in reactions at side-chains that are in current use.

Thus, D- or L-serine can be converted through the Mitsunobu reaction into the homochiral α -amino- β -lactone, a chiral synthon amenable to ring-opening by organometallic reagents (Pansare and Vederas, 1989) to give β -substituted alanines (Scheme 6.1). β -Iodo-L-alanine (also obtained from L-serine) can be elaborated similarly into the general class of β -substituted alanines (L-serine \rightarrow H₃N⁺CH(CH₂I)CO₂⁻ \rightarrow H₃N⁺CH(CH₂R)CO₂⁻ (Jackson *et al.*, 1989)). L-Aspartic acid and L-glutamic acid serve the same function, electrophiles being substituted at the carbon atom next to the side-chain carboxy group after its deprotonation with lithium di-isopropylamide (Baldwin *et al.*, 1989). As shown in this composite example from a number of research papers, the side-chain carboxy group can be transformed into other functional groups, when one starts with suitably protected glutamates and aspartates (Scheme 6.2).

There are numerous other isolated examples of the conversion of a coded amino acid into another amino acid. These usually amount to applications of straight-

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