

BAUSCH Health

Mylan Pharmaceuticals Inc. et al.

v.

Bausch Health Ireland Limited

Patent Owner's Demonstratives | June 14, 2023

Case IPR2022-00722¹
U.S. Pat. No. 7,041,786

¹ IPR2023-00016 has been joined with this preceding.

Outline

- Claim 1 Would Not Have Been Obvious Over Currie and Li
- A POSA Would Not Have Selected Human Uroguanylin as a Lead Compound
- A POSA Would Not Have Been Motivated to Substitute Asp³ with Glu³ with Any Reasonable Expectation of Success
- Objective Evidence Supports the Nonobviousness of the Claims

Claim 1 Would Not Have Been Obvious Over Currie and Li

X. GROUND 1: CLAIM 1 WAS OBVIOUS OVER CURRIE AND LI

Currie teaches the human uroguanylin peptide activates the GC-C receptor, which is useful for providing laxative effect in the intestines (by drawing water into the intestines). EX1002, ¶¶126-29, 153. From Li, a skilled artisan would have known that making a Glu³ analog of human uroguanylin using known peptide synthesis methods was reasonably likely to result in an active peptide analog. *Id.*, ¶¶147-50, 154. Hence, a skilled artisan had good reason to make [Glu³]-human uroguanylin with a reasonable expectation of success. Claim 1 was obvious over Currie in view of Li. EX1002, ¶179.



BLAKE ROBERT
PETERSON, Ph.D.

Q. In your opinion, Li cannot be used to make cross-peptide activity comparisons, correct?

A. **That's correct.**

Q. In your opinion, the experiments in Li do not provide dose response curves and do not permit comparison of the affinity or potency of rat uroguanylin to human uroguanylin or opossum uroguanylin, correct?

A. **Correct.**

Obviousness of a New Chemical Compound Ordinarily Follows a Two-Part Analysis



First, Petitioner must establish that a POSA would have selected the asserted prior-art compound as a lead compound, or starting point, **“that would be most promising to modify in order to improve upon its ... activity and obtain a compound with better activity.”**

- “In determining whether a chemist would have selected a prior art compound as a lead, the analysis is guided by evidence of the **compound’s pertinent properties.**”
- “Absent a reason or motivation based on such prior art evidence, **mere structural similarity** between a prior art compound and the claimed compound **does not inform the lead compound selection.**”

Second, Petitioner must establish that **“the prior art would have supplied [a POSA] with a reason or motivation to modify a lead compound to make the claimed compound with a reasonable expectation of success.”**

A POSA Would Not Have Selected Human Uroguanylin as a Lead Compound

Federal Circuit: Lead Compound Selection

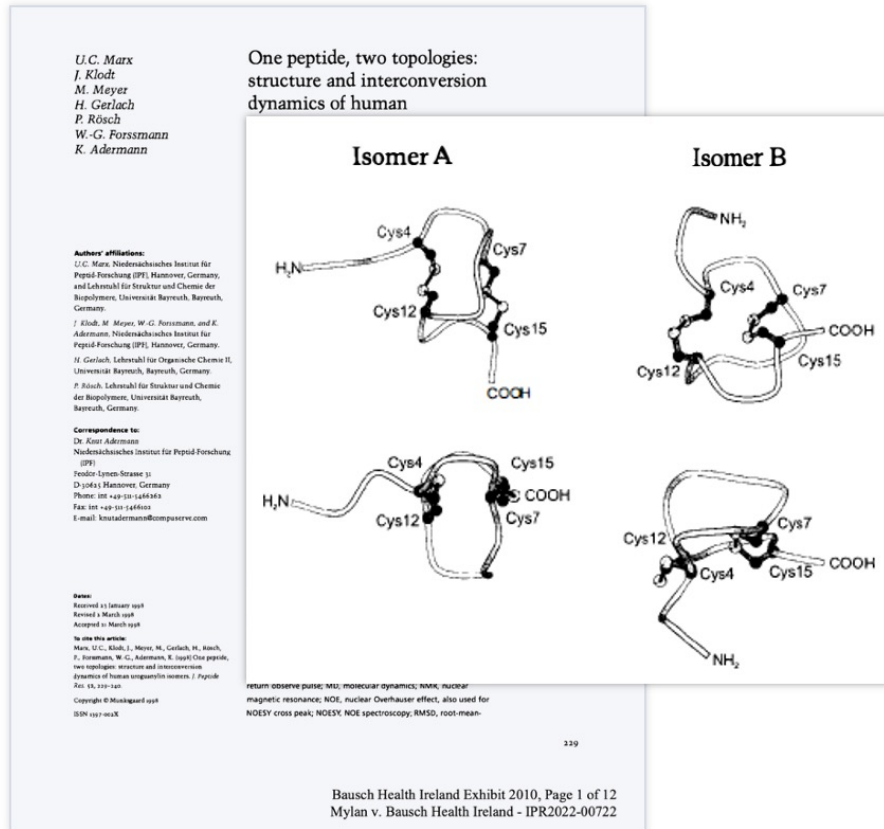


“[T]he analysis still requires the challenger to demonstrate . . . that one of ordinary skill in the art would have had a reason to select a proposed lead compound or compounds **over other compounds in the prior art.**”



“Potent and promising activity in the prior art trumps **mere structural relationships.**”

Human Uroguanylin Is Plagued by Topoisomerism

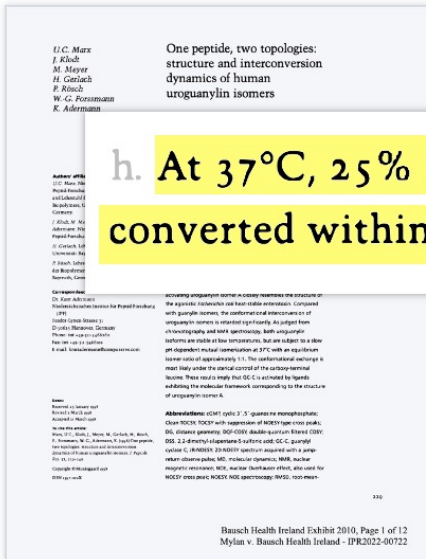


oisomers of uroguanylin are present in the body. In addition to the possible existence of other specific receptors for the GC-C-active isomer of uroguanylin, nothing is known about the structure and function of the peptide's B form that does not cause an increase of intracellular cGMP.

multiple-cysteine peptide. Our results demonstrate that, up to now, all published functional studies with bioactive guanylin reported in the literature have not been carried out with a single defined molecule but with a mixture of two peptides, one of these a ligand of GC-C, the other one with completely unknown biological properties. Because stereoisomers are present in syn-

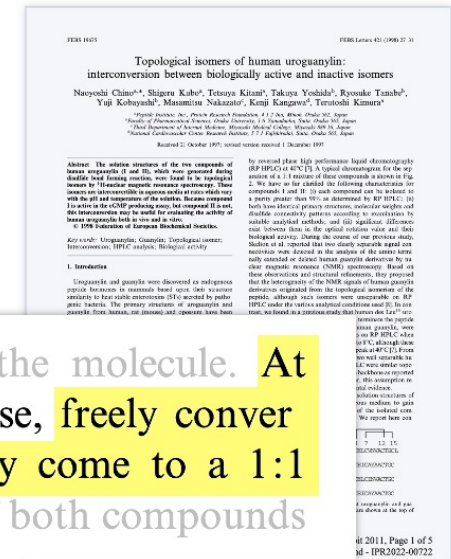
structures (Fig. 1). Both peptides are characterized by two disulfide bonds in relative positions 1 3 and 2 4, which are crucial for biological activity (2, 14, 15). The disulfides are

Topoisomerism Is Temperature and pH Dependent



h. At 37°C, 25% of both uroguanylin-16 isomers are interconverted within 24 h. This and conversion experiments

ionization state of functional group(s) in the molecule. At acidic pH, both compounds are, in one sense, freely convertible (same conversion rates) and eventually come to a 1:1 equilibrium ratio. In contrast, conversions of both compounds



MICHAEL SAMUEL EPSTEIN, M.D.

Q. And so if we're talking about the transit time from the time the patient ingests something all the way until the time they have a bowel movement, you agree that that transit time could be longer in a constipated patient?

THE WITNESS: It's -- it can be, yes. It can be longer if it's in a patient who's constipated primarily due to colonic transit.

Ex. 2010 at 236; Ex. 2011 at 30; Ex. 2070 at 45:18-46:4
Patent Owner's Resp. at 10; see also Sur-Reply at 19; Ex. 2024 ¶¶ 67-70.

Petitioner's Approach to Topoisomerism Leads Away from Plecanatide



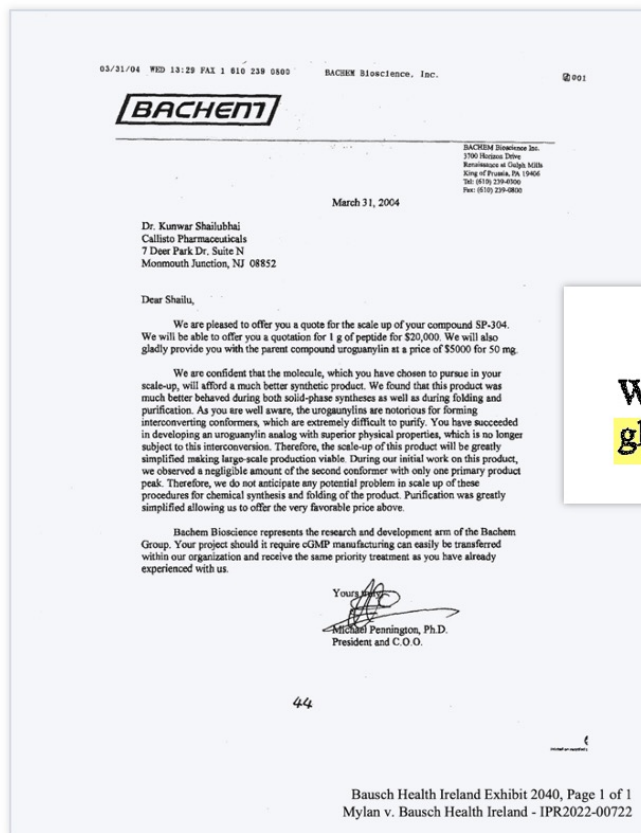
**BLAKE ROBERT
PETERSON, Ph.D.**

Q. In your opinion, in vivo topoisomeric conversion of human uroguanylin would have been easily avoided by administering it in a dosage form to time its release specifically for the intestines rather than the stomach, correct?

A. That's what's written here, yes.

- **This approach would not have resulted in plecanatide**

Topoisomerism Is More than an *In-Vivo* Problem



We are pleased to offer you a quote for the scale up of your compound SP-304. We will be able to offer you a quotation for 1 g of peptide for \$20,000. We will also gladly provide you with the parent compound uroguanylin at a price of \$5000 for 50 mg.

Ex. 2040

Patent Owner's Resp. at 32; see also Ex. 2024 ¶¶ 128-129

10

Carpick: STs Are "Long-Lived Superagonists"

INFECTION AND IMMUNITY, Nov. 1995, p. 4710-4715
0950-2688/95/14710-06\$02.00
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Vol. 61, No. 11

The *Escherichia coli* Heat-Stable Enterotoxin Is a Long-Lived Superagonist of Guanylin

BRUCE W. CARPICK AND JEAN GARIÉPY*

Department of Medical Biophysics, University of Toronto, and The Ontario Cancer Institute,
500 Sherbourne Street, Toronto, Ontario, Canada M4X 1K9

Received 7 July 1993/Returned for modification 18 August 1993/Accepted 31 August 1993

The mechanism by which bacterial heat-stable enterotoxins (ST I, ST_A) cause diarrhea in humans and animals has been linked to the activation of an intestinal membrane-bound guanylate cyclase. Guanylin, a recently discovered rat intestinal peptide, is homologous in structure to ST I and can activate guanylate cyclase present on the human colonic carcinoma cell line T84. To directly test the mechanistic association of guanylate cyclase activation with diarrhea, we synthesized guanylin and a guanylin analog termed N⁹P¹⁰ guanylin and compared their biological activities with those of a synthetic ST I analog, termed ST Ib(6-18). We report that guanylin is able to inhibit the binding of a radiolabeled ST I analog to rat intestinal cells but causes diarrhea in infant mice only at doses at least 4 orders of magnitude higher than that of ST Ib(6-18). In contrast, N⁹P¹⁰ guanylin was enterotoxic in mice at much lower doses than guanylin but proved to be a weaker inhibitor of radiolabeled ST I than guanylin in the receptor binding assay. The pattern of guanylate cyclase activation observed for ST Ib(6-18) and the two guanylin analogs parallels the results observed in the receptor binding assay rather than those observed in the diarrheal assay. Treatment of guanylin with chymotrypsin or luminal fluid derived from newborn mouse intestines resulted in a rapid loss of binding activity. Together, these results suggest that ST I enterotoxins may represent a class of long-lived superagonists of guanylin.

The heat-stable enterotoxins are a group of small homologous peptides elaborated by enterovirulent strains of bacteria (23, 29, 35). They are collectively responsible for a large proportion of worldwide cases of secretory diarrhea in humans and animals. These enterotoxins, abbreviated ST I (or ST_A), are known to bind to receptors located on the brush border surface of intestinal cells (13, 15, 17, 20, 22) and to cause an elevation of intracellular cyclic GMP (cGMP) levels (11, 16, 19, 26). It is generally believed that the binding of the enterotoxin to its receptor is coupled to the activation of a guanylate cyclase and that cGMP acts as the intracellular second messenger causing the eventual onset of diarrhea. Although more than one class of ST I receptors may exist (20, 24), research efforts in this field have focused on the cloning of one class of guanylate cyclases acting as ST I receptors (7, 8, 31, 33). The evidence linking the ST I-induced elevation of cGMP levels to secretory diarrhea has centered around the administration of a nonhydrolyzable analog of cGMP termed 8-bromo-cGMP to ligated rabbit intestines and to infant mice (16, 19). This treatment resulted in fluid accumulation in both animal models and suggested that an increased production of intracellular cGMP is a necessary step leading to watery diarrhea. Recently, a naturally occurring peptide termed guanylin was isolated from rat jejunum and was found to activate a particulate form of guanylate cyclase present on the human colonic carcinoma cell line T84 (6). This peptide was able to displace the binding of ¹²⁵I-labeled ST I to receptors on the surface of T84 cells (6). Guanylin is a 15-amino-acid peptide that is highly homologous in sequence to a region of ST I, abbreviated ST Ib(6-18), that codes for its receptor binding and enterotoxic properties (4, 14, 32, 39). In particular, identical residues are found at eight positions within the 13-amino-acid sequence of ST Ib(6-18) (Fig. 1). A major

difference between the two peptides is that ST Ib(6-18) has six cysteine residues participating in three intramolecular disulfide bridges within its sequence (14, 32) while guanylin has only four cysteines and two disulfide bridges (6) (Fig. 1). As a consequence of structural and functional similarities between guanylin and ST I, one would expect guanylin to cause diarrhea in mammals at a concentration relative to ST I that parallels its ability to inhibit the binding of radiolabeled ST I to intestinal cells. In this study, we report the synthesis of two analogs of guanylin and their ability to (i) inhibit the binding to rat cells of a radiolabeled ST I analog (4, 14), (ii) cause a diarrheal response in infant mice (16), and (iii) stimulate rat intestinal brush border guanylate cyclase (22).

MATERIALS AND METHODS

Preparation of analogs. Peptides were synthesized on an Applied Biosystems Model 430A automated peptide synthesizer using *tert*-butoxycarbonyl-protected amino acids coupled to a phenylacetamidomethyl resin support by classical solid phase methods (34). The peptides were cleaved from the resin by using anhydrous HF in the presence of anisole, dimethyl sulfide, and *p*-thiocresol. The reduced peptides were dissolved in a dilute aqueous solution (50 μM), pH 8.5, for 5 days to allow disulfide pairing to occur. The oxidized peptides were concentrated on a preparative C₁₈ column, eluted with 40% (vol/vol) acetonitrile in water, and lyophilized. The peptides were then purified by high-performance liquid chromatography (HPLC) on a semipreparative C₁₈ column with a linear gradient going from 10 to 40% (vol/vol) acetonitrile-0.08% (vol/vol) trifluoroacetic acid in water-0.1% (vol/vol) trifluoroacetic acid over a 60-min period. HPLC peaks were initially screened for their ability to inhibit the binding of a radiolabeled ST I analog, ¹²⁵I-⁹ST Ib(4-18), to rat villus cells (4). Only those peaks which exhibited inhibitory activity in this experiment were further analyzed. The homogeneity, amino acid composition, and

* Corresponding author.

4710

Bausch Health Ireland Exhibit 2060, Page 1 of 6
Mylan v. Bausch Health Ireland - IPR2022-00722

The *Escherichia coli* Heat-Stable Enterotoxin Is a Long-Lived Superagonist of Guanylin

The mechanism by which bacterial heat-stable enterotoxins (ST I, ST_A) cause diarrhea in humans and animals has been linked to the activation of an intestinal membrane-bound guanylate cyclase. Guanylin, a recently discovered rat intestinal peptide, is homologous in structure to ST I and can activate guanylate cyclase present on the human colonic carcinoma cell line T84. To directly test the mechanistic association of guanylate cyclase activation with diarrhea, we synthesized guanylin and a guanylin analog termed N⁹P¹⁰ guanylin and compared their biological activities with those of a synthetic ST I analog, termed ST Ib(6-18). We report that guanylin is able to inhibit the binding of a radiolabeled ST I analog to rat intestinal cells but causes diarrhea in infant mice only at doses at least 4 orders of magnitude higher than that of ST Ib(6-18). In contrast, N⁹P¹⁰ guanylin was enterotoxic in mice at much lower doses than guanylin but proved to be a weaker inhibitor of radiolabeled ST I than guanylin in the receptor binding assay. The pattern of guanylate cyclase activation observed for ST Ib(6-18) and the two guanylin analogs parallels the results observed in the receptor binding assay rather than those observed in the diarrheal assay. Treatment of guanylin with chymotrypsin or luminal fluid derived from newborn mouse intestines resulted in a rapid loss of binding activity. Together, these results suggest that ST I enterotoxins may represent a class of long-lived superagonists of guanylin.

Ex. 2060 at 4710
Patent Owner's Resp. at 14; Ex. 2025 ¶ 64

STs Are Not Plagued by Topoisomerism

U.C. Marx
J. Klodt
M. Meyer
H. Gerlach
P. Rösch
W.-G. Forssmann
K. Adermann

One peptide, two topologies:
structure and interconversion
dynamics of human
uroguanylin isomers

Authors' affiliations:

U.C. Marx, Niedersächsisches Institut für
Peptid-Forschung (IPF), Hannover, Germany,
and Lehrstuhl für Struktur und Chemie der
Biopolymere, Universität Bayreuth, Bayreuth,
Germany.

J. Klodt, M. Meyer, W.-G. Forssmann, and K.
Adermann, Niedersächsisches Institut für
Peptid-Forschung (IPF), Hannover, Germany.
H. Gerlach, Lehrstuhl für Organische Chemie II,
Universität Bayreuth, Bayreuth, Germany.
P. Rösch, Lehrstuhl für Struktur und Chemie
der Biopolymere, Universität Bayreuth,
Bayreuth, Germany.

Correspondence to:

Dr. Karin Adermann
Niedersächsisches Institut für Peptid-Forschung
(IPF)
Feldler-Lorenz-Strasse 31
D-30641 Hannover, Germany
Phone: Int +49-51-1464343
Fax: Int +49-51-146600
E-mail: karin.adermann@ipf.uni-hannover.de

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Res* 18, 229-240

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Key words: guanylin; heat-stable enterotoxin; isomerization;
solution structure; topological stereoisomer; uroguanylin

Abstract: The peptide hormone uroguanylin stimulates chloride
secretion via activation of intestinal guanylyl cyclase C (GC-C). It is
characterized by two disulfide bonds in a 1-3/2-4 pattern that
causes the existence of two topological stereoisomers of which
only one induces intracellular cGMP elevation. To obtain an
unambiguous structure-function relationship of the isomers, we
determined the solution structure of the separated uroguanylin
isoforms using NMR spectroscopy. Both isomers adopt well-defined
structures that correspond to those of the isomers of the related
peptide guanylin. Furthermore, the structure of the GC-C-
activating uroguanylin isomer A closely resembles the structure of
the agonistic Escherichia coli heat-stable enterotoxin. Compared
with guanylin isomers, the conformational interconversion of
uroguanylin isomers is retarded significantly. As judged from
chromatography and NMR spectroscopy, both uroguanylin
isoforms are stable at low temperatures, but are subject to a slow
pH-dependent mutual isomerization at 37°C with an equilibrium
isomer ratio of approximately 1:1. The conformational exchange is
most likely under the steric control of the carboxy-terminal
leucine. These results imply that GC-C is activated by ligands
exhibiting the molecular framework corresponding to the structure
of uroguanylin isomer A.

Abbreviations: cGMP, cyclic 3',5'-guanosine monophosphate;
Clean-TOCSY, TOCSY with suppression of NOESY-type cross peaks;
DG, distance geometry; DQF-COSY, double-quantum filtered COSY;
DSS, 2,2-dimethyl-silapentane-5-sulfonic acid; GC-C, guanylyl
cyclase C; IR-NOESY 2D NOESY spectrum acquired with a jump-
return observe pulse; MD, molecular dynamics; NMR, nuclear
magnetic resonance; NOE, nuclear Overhauser effect, also used for
NOESY cross peak; NOESY, NOE spectroscopy; RMSD, root-mean-

0.45 nm. The known higher activation potency of ST may be related to the additional disulfide bond which causes a higher rigidity of its three-dimensional structure and, thus, a possibly more efficient interaction with the receptor. Structure calculations of uroguanylin-16 with an ad-

Bausch Health Ireland Exhibit 2010, Page 1 of 12
Mylan v. Bausch Health Ireland - IPR2022-00722

Ex. 2010 at 235

Patent Owner's Resp. at 14, 33, 39; see also Ex. 2024 ¶¶ 145-146, 215; Ex. 2025 ¶¶ 46-47

STs Outperform Human Uroguanylin: Potency and Affinity

US005489670A

United States Patent [19] Patent Number: **5,489,670**
 Currie et al. [45] Date of Patent: **Feb. 6, 1996**

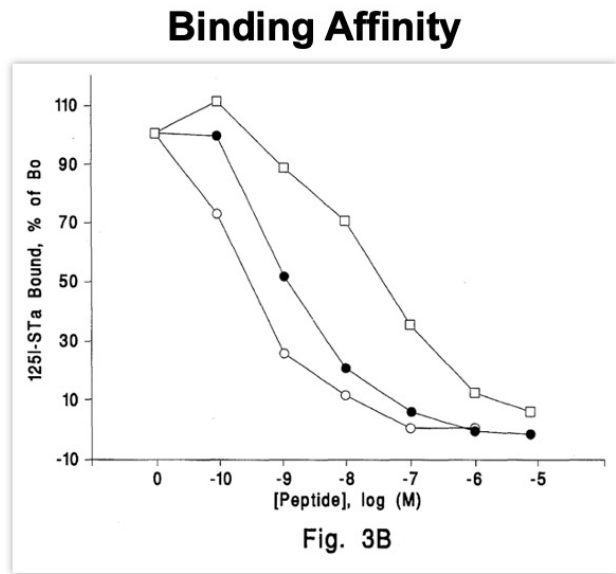
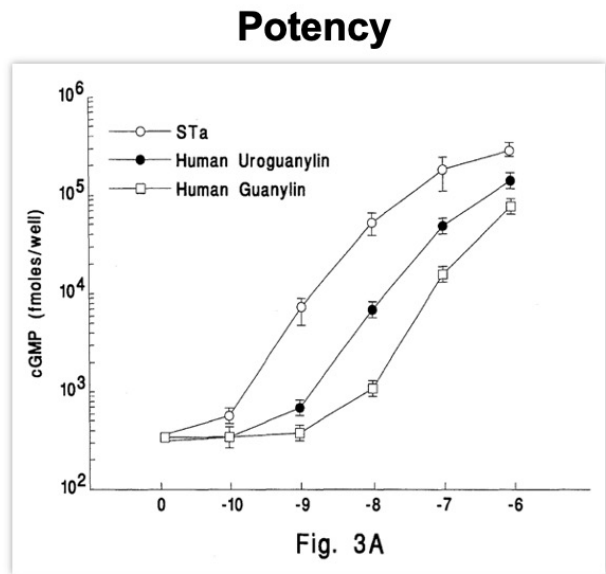
[54] **HUMAN UROGUANYLIN**
 [75] Inventors: Mark G. Currie, St. Charles; Yoshihiro Kina, Creve Coeur; Kam F. Fok, St. Louis; Christine E. Smith, Manchester, all of Mo.
 [73] Assignee: G. D. Searle & Co., Chicago, Ill.
 [21] Appl. No.: 145,940
 [22] Filed: Oct. 29, 1993
 [51] Int. Cl.²: A61K 30/00; C07K 5/00; C07K 7/00; C07K 4/00
 [52] U.S. Cl.: 530/226
 [58] Field of Search: 530/226
 [56] References Cited
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 5,140,102 8/1992 Currie 530/26
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 Schultz et al., *J. Biol. Chem.* 267: 16019-16021 (1992).
 Primary Examiner—Jill A. Warden
 Assistant Examiner—Sheila J. Huff
 Attorney Agent, or Firm—Dennis A. Bennett

ABSTRACT
 A novel peptide is disclosed which is useful for the control of intestinal fluid absorption and that has the following amino acid sequence

(SEQ ID NO: 1)
 Asp—Arg—Arg—Cys—Glu—Leu—Cys—Val—Asn—Val—
 Ala—Cys—Thr—Gly—Cys—Leu

2 Claims, 4 Drawing Sheets

MYLAN EXHIBIT - 1005
 Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722



BLAKE ROBERT PETERSON, Ph.D.

Q. Based upon Currie's disclosure, quantitatively, how much less effective is human uroguanylin than STa at stimulating cyclic GMP?

A. In this particular experiment, it appears to be about **tenfold less potent**.

Ex. 1005 at 6:13-15, Fig. 3A, 6:15-19, Fig. 3B; Ex. 2026 at 59:17-21
 Patent Owner's Resp. at 15-17, 34; see also Sur-Reply at 25; Ex. 2024 ¶¶ 93-94; Ex. 2025 ¶¶ 74-75

STs Outperform Human Uroguanylin: Potency and Affinity

Proc. Natl. Acad. Sci. USA
Vol. 94, pp. 2705-2710, March 1997
Pharmacology

Regulation of intestinal uroguanylin/guanylin receptor-mediated responses by mucosal acidity

F. KENT HAMRA^{1,2}, SAMMY L. EBER^{1,2}, DAVID T. CHEN¹, MARK G. CURRIE^{1,3}, AND LEONARD R. FORTE^{1,2}

¹Truman Veterans Affairs Medical Center and ²Departments of Pharmacology and Biochemistry and Molecular Biology Program, Missouri University, Columbia, MO 65212 and ³Neuro Research Development, St. Louis, MO 63107

Communicated by Philip Needleman, Monsanto Company, St. Louis, MO, January 2, 1997 (received for review March 27, 1996)

ABSTRACT Guanylin and uroguanylin are intestinal peptides that stimulate chloride secretion by activating a common set of receptor-guanylate cyclase signaling molecules located on the mucosal surface of enterocytes. High mucosal acidity, similar to the pH occurring within the fluid microclimate domain at the mucosal surface of the intestine, markedly enhances the cGMP accumulation responses of T84 human intestinal cells to uroguanylin. In contrast, a mucosal acidity of pH 5.0 renders guanylin essentially inactive. T84 cells were used as a model epithelium to further explore the concept that mucosal acidity imposes agonist selectivity for activation of the intestinal receptors for uroguanylin and guanylin, thus providing a rationale for the evolution of these related peptides. At an acidic mucosal pH of 5.0, uroguanylin is 100-fold more potent than guanylin, but at an alkaline pH of 8.0 guanylin is more potent than uroguanylin in stimulating intracellular cGMP accumulation and transepithelial chloride secretion. The relative affinities of uroguanylin and guanylin for binding to receptors on the mucosal surface of T84 cells is influenced dramatically by mucosal acidity, which explains the strong pH dependency of the cGMP and chloride secretion responses to these peptides. The guanylin-binding affinities for peptide-receptor interaction were reduced by 100-fold at pH 5 versus pH 8, whereas the affinities of uroguanylin for these receptors were increased 10-fold by acidic pH conditions. Deletion of the N-terminal acidic amino acids in uroguanylin demonstrated that these residues are responsible for the increase in binding affinities that are observed for uroguanylin at acidic pH. We conclude that guanylin and uroguanylin evolved distinctly different structures, which enables both peptides to regulate, in a pH-dependent fashion, the activity of receptors that control intestinal salt and water transport via cGMP.

Guanylin and uroguanylin are structurally related peptides that were isolated from intestinal mucosa and urine (1-5). A receptor for guanylin and uroguanylin that has been identified at the molecular level is a transmembrane form of guanylate cyclase, termed GC-C (6). This membrane protein was originally discovered as an intestinal receptor for the heat-stable toxin (ST) peptides, which are secreted intraluminally by enteric bacteria that cause traveler's diarrhea (7). Bacterial ST peptides are related in primary structure to uroguanylin and guanylin, thus acting as molecular mimics of the enteric peptide hormones (reviewed in refs. 8 and 9). Membrane receptor-guanylate cyclases are found on the luminal surface of enterocytes throughout the small and large intestine and in other epithelia (10-13). Binding of peptide agonists to an extracellular domain of the receptor activates the intracellular

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2705
Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722

catalytic domain producing the second messenger cGMP within target enterocytes (1-6). Intracellular cGMP stimulates transepithelial chloride secretion by regulating the phosphorylation state and chloride channel activity of the cystic fibrosis transmembrane conductance regulator, an apical protein that is located with the receptors for uroguanylin, guanylin, and ST peptides (14-16).

Isolation of uroguanylin from opossum urine (2) followed by the cloning of a cDNA that encodes opossum pre-uroguanylin (17) revealed that the uroguanylin and guanylin genes are evolutionarily related (18-20). Furthermore, the mRNAs and precursor proteins for both uroguanylin and guanylin are expressed together throughout the mucosa of small and large intestine along with their receptors (5, 11, 17-20). This raised a question of whether the differences in primary structure between guanylin and uroguanylin evolved to regulate intestinal salt and water transport through a cooperative mechanism using common receptor-guanylate cyclase signaling molecules located on the mucosal surface of the intestine.

During the isolation of uroguanylin, guanylin, and their prohormone precursors, we observed that acidic column reagents markedly attenuated the cGMP responses of T84 cells to guanylin, but enhanced the responses to uroguanylin (4, 5). This pH dependency for activation of guanylate cyclase was successfully used to detect guanylin and uroguanylin during their separation and purification from intestinal mucosa. The possibility was then considered that the primary structures of guanylin and uroguanylin could have evolved to regulate the enzymatic activity of a common set of receptors over the wide range of mucosal acidity that occurs within the intestinal lumen during digestion (21-24). We report here that high mucosal acidity rendered guanylin ineffective as a cGMP agonist and chloride secretagogue, whereas an acid pH markedly enhanced the potency of uroguanylin. A mucosal pH of 8.0 substantially increased the potency of guanylin but decreased the potency of uroguanylin. These changes in agonist potencies were explained by corresponding directional shifts in the affinities of guanylin and uroguanylin for binding to receptors at pH 5.0 versus 8.0. Uroguanylin and guanylin cooperatively regulate the guanylate cyclase activity of a common set of mucosal receptors in a pH-dependent fashion, thus providing an enteric signaling pathway for the intrinsic, paracrine regulation of intestinal salt and water transport.

MATERIALS AND METHODS

cGMP Accumulation Assay in T84 Cells. T84 cells were cultured in 24-well plastic dishes, and the cGMP levels were

Abbreviations: ST, heat-stable toxin.

¹Present address: Howard Hughes Medical Institute and Department of Pharmacology, University of Texas Southwest Medical Center, 5323 Harry Hines Boulevard, Dallas, TX 75235-9008.

²To whom reprint requests should be addressed at: Department of Pharmacology, School of Medicine, University of Missouri, Columbia, MO 65212, e-mail: Leonard.R.Forte@umc.missouri.edu.

MYLAN EXHIBIT - 1021

pared with the stimulation observed at pH 7.8 (Fig. 3). The rank order of potencies for agonist-mediated stimulation of chloride secretion was **ST > uroguanylin > guanylin at acidic pH and ST > guanylin > uroguanylin at an alkaline pH** (Fig. 3). The relative potencies of uroguanylin, guanylin, and ST-(5-17) in the stimulation of transepithelial chloride secretion across monolayers of T84 cells at acidic versus alkaline pH matched their relative potencies for stimulation of cGMP levels under these conditions.

***E. coli* ST-(5-17) binds with extraordinarily high affinities to the uroguanylin/guanylin receptors on the apical surface of T84 cells and potently stimulates cGMP production and chloride secretion at both alkaline and acidic pH. The interactions**

of the intestinal hormones, uroguanylin and guanylin. The remarkable potencies of ST peptides compared with the potencies of the enteric hormones is caused by higher affinities for ST binding to the intestinal receptors for uroguanylin and guanylin. Bacteria have created superagonist peptide toxins

Ex. 1021 at 2706-07, 2710

Patent Owner's Resp. at 16-17, 34; Ex. 2024 ¶¶ 96-98; Ex. 2025 ¶ 76

Currie Selected a Prior Art ST for Commercial Development



BLAKE ROBERT
PETERSON, Ph.D.

Q. Are you aware that Dr. Currie selected a prior art enterotoxin and modified it to make linaclotide which became marketed as Linzess?

A. Yes, I'm aware of that.

**A POSA Would Not Have Been Motivated
to Substitute Asp³ with Glu³ with Any
Reasonable Expectation of Success**

Marx: Isomerism Not Affected by N-Terminal Region

U.C. Marx
J. Klodt
M. Meyer
H. Gerlach
P. Rösch
W.-G. Forssmann
K. Adermann

One peptide, two topologies:
structure and interconversion
dynamics of human
uroguanylin isomers

Authors' affiliations:
U.C. Marx, Niedersächsisches Institut für
Peptid-Forschung (IPF), Hannover, Germany,
and Lehrstuhl für Struktur und Chemie der
Biopolymere, Universität Bayreuth, Bayreuth,
Germany.
J. Klodt, M. Meyer, W.-G. Forssmann, and K.
Adermann, Niedersächsisches Institut für
Peptid-Forschung (IPF), Hannover, Germany.
H. Gerlach, Lehrstuhl für Organische Chemie II,
Universität Bayreuth, Bayreuth, Germany.
P. Rösch, Lehrstuhl für Struktur und Chemie
der Biopolymere, Universität Bayreuth,
Bayreuth, Germany.

Correspondence to:
Dr. Karin Adermann
Niedersächsisches Institut für Peptid-Forschung
(IPF)
Feldler 12000-50000 31
D-30641 Hannover, Germany
Phone: Int +49-51-1464343
Fax: Int +49-51-1464302
E-mail: karin.adermann@ipf.uni-hannover.de

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Marx, U.C.; Klodt, J.; Meyer, M.; Gerlach, H.; Rösch,
P.; Forssmann, W.-G.; Adermann, K. 1691: One peptide,
two topologies: structure and interconversion
dynamics of human uroguanylin isomers. *J. Peptide
Res.* 18, 137-149
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ISSN 0377-024X

Key words: guanylin; heat-stable enterotoxin; isomerization;
solution structure; topological stereoisomer; uroguanylin

Abstract: The peptide hormone uroguanylin stimulates chloride
secretion via activation of intestinal guanylyl cyclase C (GC-C). It is
characterized by two disulfide bonds in a 1-3/2-4 pattern that
causes the existence of two topological stereoisomers of which
only one induces intracellular cGMP elevation. To obtain an
unambiguous structure-function relationship of the isomers, we
determined the solution structure of the separated uroguanylin
isoforms using NMR spectroscopy. Both isomers adopt well-defined
structures that correspond to those of the isomers of the related
peptide guanylin. Furthermore, the structure of the GC-C-
activating uroguanylin isomer A closely resembles the structure of
the agonistic Escherichia coli heat-stable enterotoxin. Compared
with guanylin isomers, the conformational interconversion of
uroguanylin isomers is retarded significantly. As judged from
chromatography and NMR spectroscopy, both uroguanylin
isoforms are stable at low temperatures, but are subject to a slow
pH-dependent mutual isomerization at 37°C with an equilibrium
isomer ratio of approximately 1:1. The conformational exchange is
most likely under the steric control of the carboxy-terminal
leucine. These results imply that GC-C is activated by ligands
exhibiting the molecular framework corresponding to the structure
of uroguanylin isomer A.

Abbreviations: cGMP, cyclic 3',5'-guanosine monophosphate;
Clean-TQSY, TQCSY with suppression of NOESY-type cross peaks;
DG, distance geometry; DOF-COSY, double-quantum filtered COSY;
DSS, 2,2-dimethyl-silapentane-5-sulfonic acid; GC-C, guanylyl
cyclase C; IR-NOESY, 2D NOESY spectrum acquired with a jump-
return observe pulse; MD, molecular dynamics; NMR, nuclear
magnetic resonance; NOE, nuclear Overhauser effect, also used for
NOESY cross peak; NOESY, NOE spectroscopy; RMSD, root-mean-

329

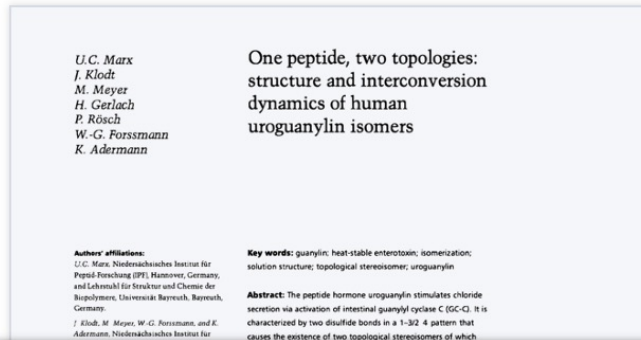
Bausch Health Ireland Exhibit 2010, Page 1 of 12
Mylan v. Bausch Health Ireland - IPR2022-00722

The slow development of the equilibrium between uroguanylin isomers at alkaline pH indicates that the ionization state of the isomeric molecules strongly influences the kinetics of transition between the isomers of uroguanylin 16 and uroguanylin 24. Thus, the terminal carboxyl, ionizable side-chains of Asp2, Asp3 and Glu5, or those groups able to form intrachain hydrogen bonds, may be involved in the control of stabilization of the two isomers. After 3 days at alkaline pH, both isoforms decomposed because of disulfide exchange. Comparison of the conversion of uroguanylin 16 isomers with the isomers of the N-terminally extended uroguanylin-24 resulted in identical kinetics for isoforms A and B at a pH of 4.5 and 7.7 for both peptides (Fig. 6C). This result clearly demonstrates that the isomerization is not affected by the amino-terminal region of uroguanylin. Corresponding to the

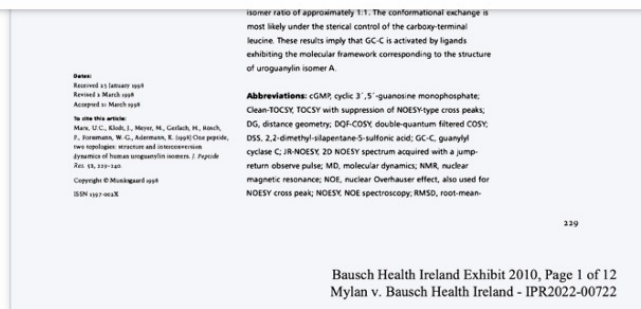
Ex. 2010 at 236

Patent Owner's Resp. at 39, 59; see also Sur-Reply at 26; Ex. 2024 ¶ 147

Marx: Disulfide Bonds Address Topoisomerism



peptide guanylin. Furthermore, the structure of the GC-C-activating uroguanylin isomer A closely resembles the structure of the agonistic *Escherichia coli* heat-stable enterotoxin. Compared



0.45 nm. The known higher activation potency of ST may be related to the additional disulfide bond which causes a higher rigidity of its three-dimensional structure and, thus, a possibly more efficient interaction with the receptor. Structure calculations of uroguanylin-16 with an additional distance restraint between protons that occupy the positions of fictitious sulfur atoms of a third disulfide bridge between residues 3 and 8 show that a third disulfide bridge is possible for the A form structure without distortion of the peptide backbone. The same calculation for the B form resulted in a higher overall energy of these structures and a slight violation of the additional fictitious distance restraint, indicating that a third disulfide bridge for the B form structure could be possible but needs a higher distortion of the peptide backbone (data not shown). A third disulfide bond apparently would lead to a preference of a structure similar to the A form isomer that was found for ST (22).

Ex. 2010 at 229, 235

Patent Owner's Resp. at 14, 39; see also Sur-Reply at 11; Ex. 2024 ¶¶ 90-91; Ex. 2025 ¶¶ 66-67

Petitioner's Conservative Substitution Theory Fails

promising modification. EX1002, ¶¶132-38, 142, 155. As Professor Peterson explains, a skilled artisan would have expected this particular conservative substitution to retain excellent receptor-activating activity. EX1002, ¶¶139-52

137. When preparing synthetic analogs to human uroguanylin, a skilled artisan would have had good reason to look first to making conservative changes to the peptide. The target receptor, as Currie discloses, is part of a “group of proteins that share structural characteristics relative to the enzymatic function of producing cyclic GMP, but differ quite remarkably in their selective activation by ligands.” EX1005, 1:7-11. Thus, as Currie notes, the guanylate cyclases are only selectively activated by their ligands. Moreover, as described above in Section VII, skilled artisans routinely began analog synthesis with conservative substitutions to avoid causing immunogenicity or ablating activity altogether.



A lead compound is “a compound in the prior art that would be most promising to modify in order to **improve upon its . . . activity and obtain a compound with better activity.**”

Pet. at 36; Ex. 1002 ¶ 137; *Otsuka Pharm. Co. v. Sandoz, Inc.*, 678 F.3d 1280, 1291 (Fed. Cir. 2012)
Patent Owner's Resp. at 3, 41-42; see also Sur-Reply at 12

Conservative Substitution Would Not Enhance Functionality



BLAKE ROBERT
PETERSON, Ph.D.

Q. Sure. Why would a person of ordinary skill in the art not expect a conservative substitution to enhance functionality?

THE WITNESS: A conservative substitution, generally speaking, has a minimal effect on biological activity, but in some cases can enhance biological activity, or reduce biological activity; but typically a person of ordinary skill in the art would make a conservative substitution to modulate biological activity.

- Dr. Peterson also admits that **“screening approaches...are not considered as rational”**

Jonson: Asp-Glu Is Not a Conservative Substitution

Protein Engineering vol.14 no.6 pp.397-402, 2001

A critical view on conservative mutations

Per Harald Jonson and Steffen B.Petersen¹

Biotechnology and Protein Engineering Group, Department of Life Sciences, Aalborg University, Sohngaardsholmsvej 49, DK-9000 Aalborg, Denmark

¹To whom correspondence should be addressed. E-mail: sp@bio.aau.dk

By analysing the surface composition of a set of protein 3D structures, complemented with predicted surface compositional information for homologous proteins, we have found significant evidence for a layer composition of protein structures. In the innermost and outermost parts of proteins there is a net negative charge, while the middle has a net positive charge. In addition, our findings indicate that the concept of conservative mutation needs substantial revision, e.g. very different spatial preferences were found for glutamic acid and aspartic acid. The alanine screening often used in protein engineering projects involves the substitution of residues to alanine, based on the assumption that alanine is a 'neutral' residue. However, alanine has a high negative correlation with all but the non-polar residues. We therefore propose the use of, for example, serine as a substitute for the residues that are negatively correlated with alanine.

Keywords: amino acid properties/protein engineering/solvent accessibility/spatial contacts/structural preference

Introduction

Upon folding of a peptide chain into a 3D protein structure, some residues are transferred from a polar environment to a more non-polar environment in the interior of the folded protein. This transfer is driven by the thermodynamic properties of the amino acids and the solvent. Throughout molecular evolution nature has selected for suitable function and stability of the resulting protein. For small to medium sized proteins—in the folded structure—only a few residues are totally buried (Chothia, 1976; Miller *et al.*, 1987; Petersen *et al.*, 1998), whereas most residues are only partially buried. The variation in solvent accessibility is dependent on the properties of the residue in question and is reflected in the amino acid composition throughout the protein structure. These differences in the solvent accessibility profile have found wide applications in various structure prediction methods (Holbrook *et al.*, 1990; Rost and Sander, 1994; Thompson and Goldstein, 1996). Also, the use of environment specific substitution matrices (Donnelly *et al.*, 1994; Wako and Blundell, 1994) have proven valuable. The sequential neighbourhood of amino acids has been investigated previously (Vondervant *et al.*, 1986) and its use has been found in, for example, loop prediction (Wojcik *et al.*, 1999) and secondary structure prediction (Chou and Fasman, 1978; Chandross and Karplus, 1999; Jones, 1999). No significant correlation between residues sequential neighbour preference was discovered.

The spatial neighbourhood around individual residues has

also been previously investigated (Burley and Petsko, 1985; Bryant and Amzel, 1987; Miyazawa and Jernigan, 1993; Petersen *et al.*, 1999). Further, spatial contacts have been studied to derive contact potentials for the different amino acid interactions (Brocchieri and Karlin, 1995; Miyazawa and Jernigan, 1996, 1999). The common strategy is to study the number of contacts within a given distance cutoff. However, the literature seems devoid of investigations of distance-dependent contacts and also of reports utilizing the embedded information of the solvent accessibility of the residues involved.

A two-state prediction of solvent accessibility correlation between hydrophobicity, buried contact propensity and the location in the prediction window has been reported (Maschelli-Georgi *et al.*, 1999). However, it does not describe any correlation between individual residue distributions.

It is important to be able to discriminate between correctly folded and misfolded model structures. It has been pointed out that potential energy-based methods do not discriminate well between folded and misfolded structures. However, structural features such as buried polar surface (Overington *et al.*, 1992) and number of polar contacts (Bryant and Amzel, 1987; Golovanov *et al.*, 1999) have proven valuable.

In protein engineering the concept of conservative mutations is frequently used. The general idea is that a substitution of an amino acid with another amino acid with similar physico-chemical properties will not influence the stability and function of the protein. The present paper shows that the spatial preferences for similar residues can be dramatically different in protein structures under similar circumstances (in this context solvent accessibility).

The results of the neighbour analysis will be valuable in model validation, as a tool for structure prediction and especially as a guide in the search for stability enhancing mutations.

Methods

The sequences used are a subset of the 25% sequence identity set of non-homologous structures (Hobohm *et al.*, 1992; Hobohm and Sander, 1994) derived from the protein structure databank PDB (Bernstein *et al.*, 1977). Only single-chain protein sequences were used. The resulting dataset consisted of 336 single-chain sequences with a maximum pairwise sequence identity of 25%. The subset was expanded through the use of the corresponding HSSP-files (Dodge *et al.*, 1998). The total data set contained 8379 aligned sequences and 1 415 986 residues. This corresponds to 6.7% of all residues in version 34 of SWISS-PROT (Bairoch and Apweiler, 1997). The length of the sequences was between 64 and 1017 residues. The resolution of the X-ray structures used varied between 1.0 and 3.0 Å, with an average of 2.0 Å. Further, the subset contained 31 structures solved by NMR. However, all hydrogen-atom co-ordinates were discarded, to check for a possible bias introduced by the use of the homologous sequences the complete analysis was done with and without the aligned sequences. No significant differences were observed.

397

Bausch Health Ireland Exhibit 2035, Page 1 of 6
Mylan v. Bausch Health Ireland - IPR2022-00722

positive charge. In addition, our findings indicate that the concept of conservative mutation needs substantial revision, e.g. very different spatial preferences were found for glutamic acid and aspartic acid. The alanine screening

of the protein. The present paper shows that the spatial preferences for similar residues can be dramatically different in protein structures under similar circumstances (in this context solvent accessibility).

tryptophan–aspartic acid are shown. The common belief that a glutamic acid to aspartic acid mutation is conservative is contrary to the observations shown. The tryptophan–glutamic

Ex. 2035 at 397 (Abstract), 400
Patent Owner's Resp. at 20; see also Sur-Reply at 12; Ex. 2024 ¶¶ 153-156

Jonson Reflects a Systematic Study of Over 1.4M Residues

Protein Engineering, vol. 14 no. 6 pp. 397-402, 2001

A critical view on conservative mutations

Per Harald Jonson and Steffen B. Petersen¹

Biotechnology and Protein Engineering Group, Department of Life Sciences, Aalborg University, Sohngaardsholmsvej 49, DK-9000 Aalborg, Denmark

¹To whom correspondence should be addressed. E-mail: sp@bio.auc.dk

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397

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Bausch Health Ireland Exhibit 2035, Page 1 of 6
Mylan v. Bausch Health Ireland - IPR2022-00722

Ex. 2035 at 397
Sur-Reply at 12

23

Petitioner's Own Argument Leads Away from Plecanatide



BLAKE ROBERT
PETERSON, Ph.D.

Q. Based upon Li's teachings with respect to opossum uroguanylin, what, if any, modification to human uroguanylin would a person of ordinary skill in the art have been motivated to make?

A. **The person of ordinary skill in the art would have been motivated to study changing those aspartic acids at positions 2 and 3, the glutamic acids, based on the homology to a opossum and rat uroguanylin.**

- This compound is the **unclaimed SP-302** in the '786 patent

Li Teaches Modifications to Rat, Not Human, Uroguanylin

Purification, cDNA sequence, and tissue distribution of rat uroguanylin

REGULATORY PEPTIDES

Purification
Zhipin
Department of

Abstract
Guanylin, a pe
a combination of
within the rat int
assayed tissue of
Northern blot w
but virtually abs
origin for urogu
BY.

Keywords: GCC

1. Introduction
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tion of water;
because it ca
(STa), an 18

*Corresponding
966927; e-mail:
The abbreviation
guanylate cyclase
guanylate cyclase
3-isobutyl-1-methyl
acetic acid, BSA.

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P/IJ S0167-0115(96)02103-9

MYLAN EXHIBIT - 1006
Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722

The affinity of GCC for uroguanylin (opossum or human) is about 10-fold higher than its affinity for guanylin (rat or human) [28,29]. Thus, features that are found in uroguanylin, but not in guanylin, offer information about structural elements that specify the strength of the ligand/receptor interaction. Of particular interest are two residues that are basic or uncharged in guanylin but acidic in uroguanylin (stippled arrowheads), and one residue that contains an aromatic ring in guanylin but an acid amide in uroguanylin (solid arrowhead). At all three positions, our duodenal peptide follows the consensus sequence of uroguanylin rather than that of guanylin, and thus we would expect its affinity to be comparable to that of opossum or human uroguanylin. Dose/response curves with synthetic rat peptide will be required to test this idea directly. It will be particularly of interest to determine whether the three extra N-terminal amino acids that distinguish our purified rat peptide from all previously-purified uroguanylins have a significant effect on binding affinity.

T/E	I	A	T	D	E	C	E	L	C	I	N	V	A	C	T	G	C	rat uroguanylin
Q	E	D	C	E	L	C	I	N	V	A	C	T	G	C				opossum uroguanylin
N	D	D	C	E	L	C	V	N	V	A	C	T	G	C	L			human uroguanylin
P	N	T	C	E	I	C	A	Y	A	A	C	T	G	C				rat guanylin
S	H	T	C	E	I	C	A	F	A	A	C	A	G	C				opossum guanylin
P	G	T	C	E	I	C	A	Y	A	A	C	T	G	C				human guanylin
P	N	T	C	E	I	C	A	Y	A	A	C	T	G	C				mouse guanylin
N	T	F	Y	C	C	E	L	C	C	N	P	A	C	A	G	C	Y	E Coli STa

directly. It will be particularly of interest to determine whether the three extra N-terminal amino acids that distinguish our purified rat peptide from all previously-purified uroguanylins have a significant effect on binding affinity.

Li: Rat Uroguanylin Is Far Less Potent than Opossum Uroguanylin


REGULATORY PEPTIDES

Regulatory Peptides 68 (1997) 43–56

Purification, cDNA sequence, and tissue distribution of rat uroguanylin

Zhiping Li*, Ashley G. Perkins, Matthew F. Peters, Michael J. Campa, Michael F. Goy

Department of Physiology and Center for Gastrointestinal Biology and Disease at the University of North Carolina, Chapel Hill NC 27590, USA

Received 29 June 1996; revised 17 October 1996; accepted 31 October 1996

Abstract

Guanylin, a peptide purified from rat jejunum, is thought to regulate water and electrolyte balance in the intestine. We show here, using a combination of Northern blots, Western blots, and functional assays, that guanylin and its receptor (GCC) are not distributed in parallel within the rat intestine. To investigate the possibility that there might be a second intestinal peptide that serves as a ligand for GCC, we assayed tissue extracts for the ability to stimulate cyclic GMP synthesis in a GCC-expressing cell line. Duodenal extracts display a peak of biological activity that is not present in colon and that does not comigrate with guanylin or proguanylin. The activity co-purifies with a novel peptide (TATYDCELCENNACTGC) that has high homology with uroguanylin, a peptide initially purified from human and opossum urine. A rat uroguanylin cDNA clone was found to encode a propeptide whose C-terminus corresponds to our purified peptide. Northern blots with probes generated from this clone reveal that prouroguanylin mRNA is strongly expressed in proximal small intestine, but virtually absent from colon, corroborating our biochemical measurements. Taken together, these studies demonstrate an intestinal origin for uroguanylin, and show that within the intestine its distribution is complementary to that of guanylin. © 1997 Elsevier Science B.V.

Keywords: GCC; STa receptor; CFTR; Guanylin; Uroguanylin

1. Introduction

A considerable body of evidence supports a role for the cyclic GMP pathway in the control of ion transport in the gastrointestinal tract. Elevation of intracellular cyclic GMP levels in intestinal epithelial cells enhances secretion of chloride into the intestinal lumen [1], and diminishes absorption of sodium and chloride [2]. The combination of increased secretion and decreased absorption elevates the osmolarity of the lumen, and drives the luminal accumulation of water. This mechanism was initially identified because it can be induced by heat-stable enterotoxin (STa), an 18 amino acid peptide secreted by pathogenic strains of *Escherichia coli* [3–5]. Exposure to high levels of toxin, as occurs during acute bacterial infections, triggers non-physiological movement of electrolytes, and produces a watery diarrhea that can lead to dehydration and death.

When the STa receptor was cloned from a small intestinal cDNA library [6], it was found to belong to a family of receptors that contain endogenous guanylate cyclase (GC) activity. Two other members of this family are the natriuretic peptide receptors, GCA and GCB [7]. Because the STa receptor was the third such receptor cloned, it was named GCC. All members of this family contain: (a) an intracellular catalytic domain responsible for the conversion of GTP to cyclic GMP, (b) an intracellular regulatory domain that controls the activity of the catalytic domain, (c) a single transmembrane domain, and (d) an extracellular receptor domain that provides an agonist binding site [7].

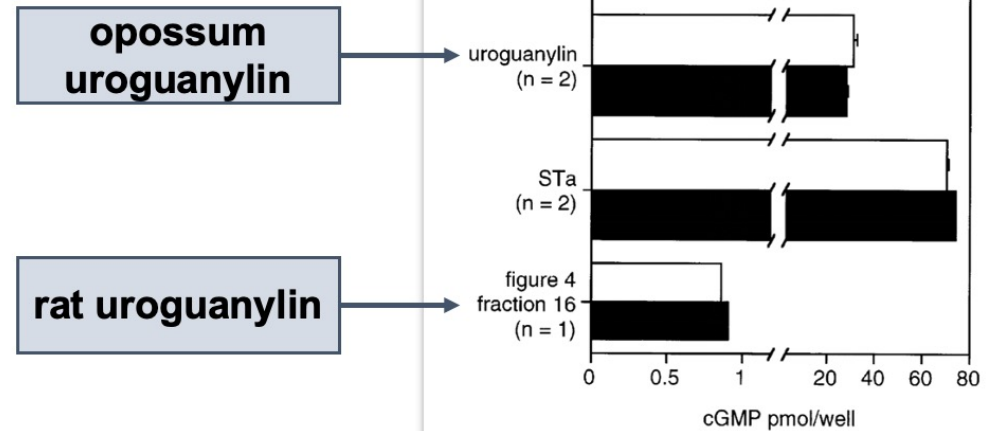
These findings led to the hypothesis that GCC serves as a receptor for one or more endogenous ligands in the GI

*Corresponding author. Tel.: +1 919 9666993; fax: +1 919 9666927; e-mail: zp@med.unc.edu
 The abbreviations used are: STa, heat-stable enterotoxin; GCA, guanylate cyclase type A; GCB, guanylate cyclase type B; GCC, guanylate cyclase type C; HBS, Hank's buffered salt solution; IBMX, 3-isobutyl-1-methylxanthine; TFA, trifluoroacetic acid; TGA, trichloroacetic acid; BSA, bovine serum albumin; RIA, radioimmunoassay.

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 PII S0167-0115(96)02103-9

MYLAN EXHIBIT - 1006
 Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722

Fig. 3. Preincubation at 37°C enhances the activity of HPLC column fractions, but not of synthetic guanylin or uroguanylin. The bars show the mean level of cyclic GMP (±SEM or range) in T84 cells after exposure to the indicated stimuli. Each stimulus was either held at 4°C (□) or incubated at 37°C (■) prior to applying it to the cells. The numbers used to identify the HPLC fractions correspond to (i) the figure in which the pertinent HPLC run is shown and (ii) the appropriate fraction number(s) from that column run.



Ex. 1006 at 49 (Fig. 3)
 Patent Owner's Resp. at 46-47; see also Sur-Reply at 13, fn. 3; Ex. 2024 ¶¶ 75, 79

Rat Uroguanylin Is Plagued by Topoisomerism

FEBS 19675 FEBS Letters 421 (1998) 27-31

**Topological isomers of human uroguanylin:
interconversion between biologically active and inactive isomers**

Naoyoshi Chino^{a,*}, Shigeru Kubo^b, Tetsuya Kitani^c, Takuya Yoshida^b, Ryosuke Tanabe^b,
Yuji Kobayashi^b, Masamitsu Nakazato^c, Kenji Kangawa^d, Terutoshi Kimura^a

^aPeptide Institute, Inc., Protein Research Foundation, 4-1-2 Ina, Minoh, Osaka 562, Japan
^bFaculty of Pharmaceutical Sciences, Osaka University, 1-6 Yamadaoka, Suita, Osaka 565, Japan
^cThird Department of Internal Medicine, Miyazaki Medical College, Miyazaki 889-16, Japan
^dNational Cardiovascular Center Research Institute, 5-7-1 Fujikurabi, Suita, Osaka 565, Japan

Received 21 October 1997; revised version received 1 December 1997

Abstract The solution structures of the two compounds of human uroguanylin (I and II), which were generated during disulfide bond forming reaction, were found to be topological isomers by ¹H-nuclear magnetic resonance spectroscopy. These isomers are interconvertible in aqueous media at rates which vary with the pH and temperature of the solution. Because compound I is active in the cGMP producing assay, but compound II is not, this interconversion may be useful for evaluating the activity of human uroguanylin both in vivo and in vitro.
© 1998 Federation of European Biochemical Societies.

Key words: Uroguanylin; Guanylin; Topological isomer; Interconversion; HPLC analysis; Biological activity

1. Introduction

Uroguanylin and guanylin were discovered as endogenous peptide hormones in mammals based upon their structure similarity to heat stable enterotoxins (STs) secreted by pathogenic bacteria. The primary structures of uroguanylin and guanylin from human, rat (mouse) and opossum have been reported as being comprised of 15 or 16 amino acid residues [1-5]; the human and rat peptide sequences are shown in Fig. 1. The sequence similarity among them is high and four Cys residues in all the peptides are conserved. These Cys residues participate in the formation of the two intramolecular disulfide linkages, one between Cys³ and Cys¹² and the other between Cys⁵ and Cys¹⁵. Uroguanylin and guanylin, as well as ST, are reported to be involved in the regulation of salt and water transport in the intestinal tract and kidney. In addition, these peptides are known to stimulate cGMP production by activating the guanylyl cyclase C in both enterocytes and T84 colon cancer cells. Therefore, endogenous uroguanylin and guanylin are suggested to play important roles in intestinal and renal dysfunction and salt dependent hypertension [6].

In our previous paper on the chemical synthesis of human uroguanylin using a two step selective disulfide forming method, two compounds (I and II) were found to be generated upon analyzing the second disulfide bond forming reaction

by reversed phase high performance liquid chromatography (RP HPLC) at 40°C [7]. A typical chromatogram for the separation of a 1:1 mixture of these compounds is shown in Fig. 2. We have so far clarified the following characteristics for compounds I and II: (i) each compound can be isolated to a purity greater than 99% as determined by RP HPLC; (ii) both have identical primary structures, molecular weights and disulfide connectivity patterns according to examination by suitable analytical methods; and (iii) significant differences exist between them in the optical rotation value and their biological activity. During the course of our previous study, Skellon et al. reported that two clearly separable signal components were detected in the analysis of the amino terminally extended or deleted human guanylin derivatives by nuclear magnetic resonance (NMR) spectroscopy. Based on these observations and structural refinements, they proposed that the heterogeneity of the NMR signals of human guanylin derivatives originated from the topological isomerism of the peptide, although such isomers were unseparable on RP HPLC under the various analytical conditions used [8]. In contrast, we found in a previous study that human des Leu¹⁶ uroguanylin and rat guanylin, both of which terminate the peptide chains at the fourth Cys residue like human guanylin, were detected as two base line separable peaks on RP HPLC when the analytical temperature was decreased to 8°C, although these peptides were eluted in a broad but single peak at 40°C [7]. From these observations, we assumed that the two well separable human uroguanylin compounds on RP HPLC were similar topological isomers with respect to the peptide backbone as reported for human guanylin derivatives. However, this assumption required definite confirmation by experimental evidence.

In the present study, we analyzed the solution structures of both compounds by NMR in an aqueous medium to gain further insight into the characteristics of the isolated compounds I and II of human uroguanylin. We report here con

Fig. 1. Primary structures of human and rat uroguanylin and guanylin. Two intramolecular disulfide linkages are shown at the top of the sequences.

*Corresponding author. Fax: +81 (727) 29 4124.
E-mail: chino@prt.or.jp

Abbreviations: ST, heat stable enterotoxin; cGMP, cyclic 3',5' guanine monophosphate; RP HPLC, reversed phase high performance liquid chromatography; CD, circular dichroism; NMR, nuclear magnetic resonance; NOE, nuclear Overhauser effect; NOESY, NOE spectroscopy; MD, molecular dynamics; RMSD, root mean square deviation; GdnHCl, guanidine hydrochloride; NEM, N-ethylmaleimide; TFA, trifluoroacetic acid; DMSO, dimethyl sulfoxide

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Bausch Health Ireland Exhibit 2011, Page 1 of 5
Mylan v. Bausch Health Ireland - IPR2022-00722

In the RP HPLC analyses of the two human uroguanylin isomers, we had already established that they are separable at 40°C. However, separations of the human des Leu¹⁶ uroguanylin isomers, as well as the rat guanylin isomers, were possible only at lower temperatures such as 8°C [7]. This separation characteristic has also been observed for a recently disclosed member of the uroguanylin and guanylin peptide family, rat uroguanylin 15 (unpublished result). Considering

N-terminal Amino Acids of Human Uroguanylin Are Required

Proc. Natl. Acad. Sci. USA
Vol. 94, pp. 2705-2710, March 1997
Pharmacology

Regulation of intestinal uroguanylin/guanylin receptor-mediated responses by mucosal acidity

F. KENT HAMRA^{1,2}, SAMMY L. EBERT¹, DAVID T. CHEN¹, MARK G. CURRIE^{1,3}, AND LEONARD R. FORTE^{1,4}

¹Trommsdorff Veterans Affairs Medical Center and ²Department of Pharmacology and Biochemistry and ³Molecular Biology Program, Missouri University, Columbia, MO 65212 and ⁴Biologic Research and Development, St. Louis, MO 63107

Communicated by Philip Needleman, Monsanto Company, St. Louis, MO, January 2, 1997 (received for review March 27, 1996)

ABSTRACT Guanylin and uroguanylin are intestinal peptides that stimulate chloride secretion by activating a common set of receptor-guanylate cyclase signaling molecules located on the mucosal surface of enterocytes. High mucosal acidity, similar to the pH occurring within the fluid microclimate domain at the mucosal surface of the intestine,

catalytic domain producing the second messenger cGMP within target enterocytes (1-6). Intracellular cGMP stimulates transepithelial chloride secretion by regulating the phosphorylation state and chloride channel activity of the cystic fibrosis transmembrane conductance regulator, an apical protein that is located with the receptors for uroguanylin, guanylin, and ST

mulation response to uroguanylin⁹⁸⁻¹⁰⁹. We conclude that the unique acidic amino acids at the N terminus of uroguanylin are required for the increased binding affinities, and accordingly, the enhanced potencies of uroguanylin in the stimulation of target cell responses under the acidic conditions of pH 5.0-5.5 maintained at the mucosal surface of T84 cells in this model epithelium.

This work was supported by the National Institutes of Health (F.K.H. and L.R.F.). The receptor for guanylin and uroguanylin that has been identified at the molecular level is a transmembrane form of guanylate cyclase, termed GC-C (6). This membrane protein was originally discovered as an intestinal receptor for the heat-stable toxin (ST) peptides, which are secreted intraluminally by enteric bacteria that cause traveler's diarrhea (7). Bacterial ST peptides are related in primary structure to uroguanylin and guanylin, thus acting as molecular mimics of the enteric peptide hormones (reviewed in refs. 8 and 9). Membrane receptor-guanylate cyclases are found on the luminal surface of enterocytes throughout the small and large intestine and in other epithelia (10-13). Binding of peptide agonists to an extracellular domain of the receptor activates the intracellular

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uroguanylin. These changes in agonist potencies were explained by corresponding directional shifts in the affinities of guanylin and uroguanylin for binding to receptors at pH 5.0 versus 8.0. Uroguanylin and guanylin cooperatively regulate the guanylate cyclase activity of a common set of mucosal receptors in a pH-dependent fashion, thus providing an enteric signaling pathway for the intrinsic, paracrine regulation of intestinal salt and water transport.

MATERIALS AND METHODS

cGMP Accumulation Assay in T84 Cells. T84 cells were cultured in 24-well plastic dishes, and the cGMP levels were

Abbreviations: ST, heat-stable toxin.
¹Present address: Howard Hughes Medical Institute and Department of Pharmacology, University of Texas Southwest Medical Center, 5323 Harry Hines Boulevard, Dallas, TX 75255-9005.
²To whom reprint requests should be addressed at: Department of Pharmacology, School of Medicine, University of Missouri, Columbia, MO 65212, e-mail: Leonard.R.Forste@umc.missouri.edu.

MYLAN EXHIBIT - 1021

Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722



BLAKE ROBERT PETERSON, Ph.D.

Q. Based on Hamra 1997, it is your opinion that the N-terminal amino acid residues of human uroguanylin are required for the increased binding affinities and enhanced potency for activation of receptors under acidic conditions, correct?

A. Correct.

Fiser: Asp Preferred Over Glu

FEBS Letters 397 (1996) 225-229

FEBS Letters 397 (1996) 225-229

Conservation of amino acids in multiple alignments: aspartic acid has unexpected conservation

András Fiser^{a,b}, István Simon^a, Geoffrey J. Barton^{a,*}

^aUniversity of Oxford, Laboratory of Molecular Biophysics, The Rex Richards Building, South Parks Road, Oxford OX1 3JQ, UK
^bInstitute of Enzymology, Biological Research Center, Hungarian Academy of Sciences, P.O. Box 7, Budapest H-1518, Hungary

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Abstract Analysis of the relationship between surface accessibility and amino acid conservation in multiple sequence alignments of homologous proteins confirms expected trends for hydrophobic amino acids, but reveals an unexpected difference between the conservation of Asp, Glu and Gln. Even when not in an active site, Asp is more highly conserved than Glu. There is a clear preference for conserved and buried Asp to be present in coil, but there is no tendency for Asp to conserve ϕ/ψ in the α region of the Ramachandran map. Glu does not show any preference to be conserved in a particular secondary structure. Analysis of recently derived substitution matrices (e.g. BLO-SUM) confirms that Glu tends to substitute more frequently with other amino acids than does Asp. Analysis of relative accessibility versus relative conservation for individual amino acid positions in alignments shows a negative correlation for all amino acid types. With the exception of Arg, Lys, Gln, Glu, Asp and Tyr, a relative conservation of >2 suggests the amino acid will have a relative accessibility of $<50\%$. Observation of conserved Cys, Gly or Asp in a reliable multiple alignment suggests a position important for the structure of the protein. Furthermore, the Asp is likely to be involved in polar interactions through its side-chain oxygen atoms. In contrast, Glu is the least conserved amino acid overall.

Key words: Conservation analysis; protein structure prediction; alignment; protein structure prediction

1. Introduction

Knowledge of protein sequences is growing much faster than knowledge of either three-dimensional structure or function. Accordingly, the interpretation of sequence data to identify structurally or functionally important residues is essential if the data are to be effective in furthering understanding of biological systems. Multiple sequence alignments of families of protein sequences are now used routinely to indicate residues of key importance to the function of the protein. A position in an alignment that has identical residues in all members of a protein family may have a key catalytic role.

A position where similar physico-chemical properties (e.g. hydrophobicity) are shared may suggest importance in stabilising the native conformation of the protein [1,2]. Identification of such conserved features in multiple alignments has been used to good effect to improve the accuracy of prediction of secondary structure and buried residues (helix and β -strand) (e.g. [3-7]).

Here we report a systematic study of residue conservation in multiple alignments where at least one protein is of known

tertiary structure. Our analysis complements that of Overington et al. [8] who considered only pairwise substitution frequencies for amino acids in structurally aligned families.

2. Materials and methods

2.1. Data base

A non-redundant set of 81 proteins was generated from the April 1993 release of the Brookhaven Protein Data Bank (PDB) [9]. The set was chosen in a two-step procedure. First, all pairs of chains (over 50 residues and resolution better than 2.5 Å) in the data bank were compared by calculating correlation coefficients between the dipeptide frequencies in each protein. A set of 101 protein chains was selected such that all pairs had a correlation of <0.8 . All pairs in this set were then compared by a rigorous sequence comparison method [10,11] followed by cluster analysis. This reduced the set to 81 protein chains that show no obvious sequence similarity (PDB code and chain identifier: 1SSC IACKA IALC IBBP A ICS5 IBCA IYK1 IFSR IIGK IIGP1 A IHD58 IHIP IHOE ILRD 4 IPAZ IPCY IPIH IPRC C IREI IREI IRNH ISN1 IRES ITRK A IWSY B 2508 A GADP ZALA A 2CAB 2CDM 2CDV 2CZF 2FXB 2G5L 2LH1 2LIV 2LTN A 2ORL L 2PAB A 2RNT 2RSP A 2S5C J 2SNL E 2SN6 2SGD B 2SH 2SVY 2T51 2ITGL A IADK IBCG ICIA IPRC IGAP B 3LZM IJGB J 451C 4BFI 4EDI 4FXN 4HBR A 4PEP 4PFR 4PTF 4TNC 4X7S 5CVT B 5EER 5RUB A 5RKN 4LHM 5TAN E 7PT1 RADH B 8ATC B 8CAT A 8DFR 8PAP 9R3A A 9WGA A).

Each protein in the set was compared by the Smith-Waterman algorithm [11,12] to the NBRF-PHR sequence data bank (Release 30) and all sequences that gave probability values of $<10^{-4}$ by a length-dependent scoring scheme (program SCANPS [ip.igodf@bio.ox.ac.uk/programscanps]) were multiply aligned with the query sequence by the algorithm of Barton and Sternberg [13]. This gave 81 alignments with between 3 and 499 sequences in each (median of 28 sequences).

2.2. Calculation of conservation and accessibility

Conservation scores based upon the physico-chemical properties of the amino acids were calculated for each position in each alignment according to Livignone and Barton [2]. Conservation scores range from 0 to 10 and represent the number of the properties: Hydrophobic, Positive, Negative, Polar, Charged, Small, Tiny, Aliphatic, Aromatic, Proline and their negations (e.g. not positive) that are shared at a position. The program AMAS [2], which calculates conservation values from a multiple alignment, may be run over the World Wide Web (<http://igodf.bio.ox.ac.uk/serve/amas-server.html>).

Although conservation scores are absolute, the relative importance of a conservation score is dependent on the overall similarity between the sequences in the multiple alignment. For example, in an alignment of 20 sequences that all share $>90\%$ pairwise identity conservation scores above 4 may be interesting. In contrast, if the pairwise identity is below 30% then lower conservation scores will be informative. Accordingly, in this study we normalised conservation scores by the average conservation for each alignment to give relative conservation scores C . We refer to a position as conserved if $C \geq 1$.

Accessible surface areas were calculated by the program DSSP [14] and converted to relative accessibilities by dividing by the accessibility of the residue in a Gly-X-Gly tripeptide [15]. Two relative accessibility classes were considered 0.4 \leq 0.25 (buried) and 0.25 $<$ 0.4 \leq 0.25 (exposed).

*Corresponding author. Fax: (44) (1865) 510454.

E-mail: gji@mol.bio.ox.ac.uk

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Bausch Health Ireland Exhibit 2036, Page 1 of 5
Mylan v. Bausch Health Ireland - IPR2022-00722

Abstract Analysis of the relationship between surface accessibility and amino acid conservation in multiple sequence alignments of homologous proteins confirms expected trends for hydrophobic amino acids, but reveals an unexpected difference between the conservation of Asp, Glu and Gln. Even

Since our data do not suggest a significant preference for $++ \phi/\psi$, the preferred conservation of Asp is likely to be due to differing side-chain interactions. The most obvious hypoth-

to differing side-chain interactions. The most obvious hypothesis is that since Glu has a higher proportion of non-polar atoms than Asp it can make more non-specific interactions and so there are fewer constraints on its environment. In

Why, then, is Asp most highly conserved when buried in coil? The short Asp side chain is restricted in mobility yet able to make strong polar interactions. It is possible that Asp may form a 'pin' that stabilises non-regular structures in loops.

Ex. 2036 at 225 (Abstract), 227, 228
Patent Owner's Resp. at 20-21, 50; Ex. 2024 ¶¶ 103, 147, 153, 214

WO '266: Retain Asp at 2 and 3 Positions of Human Uroguanylin

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(71) Applicant (for all designated States): MACIA CORPORATION, Department, P.O. Box 5110, Chicago, IL 60680-5110 (US)

(72) Inventors; and (75) Inventors/Applicants (as known): 154 MO 63017 (US); CU Mason Ridge Drive, St. Louis, MO 63017 (US)

(74) Agents: BENNETT, Dennis, A. et al.; Pharmacia Corporation, Corporate Patent Department, P.O. Box 5110, Chicago, IL 60680-5110 (US)

(54) Title: UROGUANYLIN AS AN INTESTINAL CANCER INHIBITING AGENT

(57) Abstract: Disclosed is a method of retarding the development of polyps and prevention, inhibition and treatment of cancer in the intestine of a subject by administration of a composition comprising a peptide with the active domain of uroguanylin, or any agonist peptide or compound binding to the guanylate cyclase receptor GC-C, in the intestine.

WO 01/25266 AI

Bausch Health Ireland Exhibit 2021, Page 1 of 55
Mylan v. Bausch Health Ireland - IPR2022-00722

FIG. 7

Asn Asp Asp Cys Glu Leu Cys Val Asn Val Ala Cys Thr Gly Cys Leu h UroG.
 Pro Gly Thr Cys Glu Ile Cys Ala Tyr Ala Ala Cys Thr Gly Cys h Gua
 Asp Ser Ser Asn Tyr Cys Cys Glu Leu Cys Cys Asn Pro Ala Cys Thr Gly Cys Tyr E. coli
 Leu Ile Ile Asp Cys Cys Glu Ile Cys Cys Asn Pro Ala Cys Phe Gly Cys Leu Asn V. cholerae



BLAKE ROBERT PETERSON, Ph.D.

intestinal pH. Two underlined (Asp-Asp) residues are believed to be important for regulating the functional activity of uroguanylin only at the acidic environment of the intestinal mucosa.

X₆-Asp- Asp- Cys- X₁- X₂- Cys- X₃- Asn- X₄- X₅- Cys- X₆- X₇- Cys-X₉

species. The functionally active domain in most of these peptides are highly conserved. Therefore, the physiological

agents as well. Thus, as long as the functionally active domains of these peptides are conserved, substitutions in the non-active domains may be achieved with no change in the activity of the peptides.

Q. WO 266 identifies the aspartate residues at positions 2 and 3 in human uroguanylin as important for regulating the functional activity of uroguanylin in the acidic environment of intestinal mucosa, correct?

THE WITNESS: I mean those amino acids are important for regulating the functional activity of uroguanylin in the acidic environment of intestinal mucosa, that's, that's correct.

Petitioner's Flawed pH Argument

UNITED STATES PATENT AND TRADEMARK OFFICE

BEFORE THE PATENT TRIAL AND APPEAL BOARD

MYLAN PHARMACEUTICALS INC.,
Petitioner,

v.

BAUSCH HEALTH IRELAND LIMITED,
Patent Owner.

Case IPR2022-00722
Patent US 7,041,786 B2

PETITION FOR INTER PARTES REVIEW

Moreover, Professor Peterson explains that the [Glu³]-substitution would have been expected to result in a protonated glutamate at a higher pH (pKa = 4.25 rather than aspartic acid's 3.65) for better activity in the less acidic environment of the intestinal lumen further away from the stomach. EX1002, ¶¶157-61; EX1012, 118, Table 5-1. In this way, the substitution would have been expected to apply the enhanced activity of human uroguanylin (relative to guanylin) more broadly to the intestines instead of being localized only proximate to the more acidic environment near the stomach. EX1002, ¶¶162-170; *see also* EX1002, ¶¶61-65; EX1016, E960,

Nelson: Free pKa Values Cannot Be Strictly Applied to Peptides

peptide R groups, and other environmental factors can affect the pK_a . The pK_a values for R groups listed in Table 5-1 can be a useful guide to the pH range in which a given group will ionize, but they cannot be strictly applied to peptides.

Lehninger Principles of Biochemistry

David L. Nelson
Professor of Biochemistry
University of Wisconsin-Madison

Michael M. Cox



BLAKE ROBERT
PETERSON, Ph.D.

Q. Nelson teaches that the pK_a values for the R groups listed in Table 5-1 of Nelson cannot be strictly applied to peptides, correct?

THE WITNESS: So, as I mentioned, it's not accurate to compare a free amino acid pK_a necessarily with a peptide amino acid pK_a , unless one does a systematic study where we're looking at systematic differences, such as aspartic acid versus glutamic acid in a given peptide sequence. If that makes sense.

Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722

Table 5-1

Properties and Conventions Associated with the Standard Amino Acids								
Amino acid	Abbreviated names	M_r	pK_a values			pI	Hydropathy index*	Occurrence in proteins (%) [†]
			pK_1 (–COOH)	pK_2 (–NH ₃ ⁺)	pK_R (R group)			
Nonpolar, aliphatic R groups								
Glycine	Gly G	75	2.34	9.60		5.97	–0.4	7.2
Alanine	Ala A	89	2.34	9.69		6.01	1.8	7.8
Valine	Val V	117	2.32	9.62		5.97	4.2	6.6
Leucine	Leu L	131	2.36	9.60		5.98	3.8	9.1
Isoleucine	Ile I	131	2.36	9.68		6.02	4.5	5.3
Methionine	Met M	149	2.28	9.21		5.74	1.9	2.3
Aromatic R groups								
Phenylalanine	Phe F	165	1.83	9.13		5.48	2.8	3.9
Tyrosine	Tyr Y	181	2.20	9.11	10.07	5.66	–1.3	3.2
Tryptophan	Trp W	204	2.38	9.39		5.89	–0.9	1.4
Polar, uncharged R groups								
Serine	Ser S	105	2.21	9.15		5.68	–0.8	6.8
Proline	Pro P	115	1.99	10.96		6.48	1.6	5.2
Threonine	Thr T	119	2.11	9.62		5.87	–0.7	5.9
Cysteine	Cys C	121	1.96	10.28	8.18	5.07	2.5	1.9
Asparagine	Asn N	132	2.02	8.80		5.41	–3.5	4.3
Glutamine	Gln Q	146	2.17	9.13		5.65	–3.5	4.2
Positively charged R groups								
Lysine	Lys K	146	2.18	8.95	10.53	9.74	–3.9	5.9
Histidine	His H	155	1.82	9.17	6.00	7.59	–3.2	2.3
Arginine	Arg R	174	2.17	9.04	12.48	10.76	–4.5	5.1
Negatively charged R groups								
Aspartate	Asp D	133	1.88	9.60	3.65	2.77	–3.5	5.3
Glutamate	Glu E	147	2.19	9.67	4.25	3.22	–3.5	6.3

*A scale combining hydrophobicity and hydrophilicity of R groups; it can be used to measure the tendency of an amino acid to seek an aqueous environment (– values) or a hydrophobic environment (+ values). See Chapter 12. From Kyte, J. & Doolittle, R.F. (1982) *J. Mol. Biol.* **157**, 105–132.
[†]Average occurrence in over 1150 proteins. From Doolittle, R.F. (1989) Redundancies in protein sequences. In *Prediction of Protein Structure and the Principles of Protein Conformation* (Fasman, G.D., ed) Plenum Press, NY, pp. 599–623.

Ex. 1012 at 127, 118, Ex. 2026 at 101:15-102:2
Patent Owner's Resp. at 52-53; Ex. 2024 ¶¶ 176-177

Side Chain pKa Requires Accounting for Peptide Environment



BLAKE ROBERT
PETERSON, Ph.D.

Q. Do you agree with Dr. Davies that a prediction as to the pKa of the carboxylic acid on the side chain of the aspartate or glutamate at the third position would require accounting for the environment in which these residues are found when incorporated into a peptide chain, correct?

A. **That's correct.**

When in a Peptide, Asp's pKa May Be Higher than Glu's pKa

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JMB Available online at www.sciencedirect.com  

The pKa Values of Acidic and Basic Residues Buried at the Same Internal Location in a Protein Are Governed by Different Factors

Michael J. Harms¹, Carlos A. Castañeda¹, Jamie L. Schlessman^{1,2}, Gloria R. Sue¹, Daniel G. Isom¹, Brian R. Cannon¹ and Bertrand Garcia-Moreno E.^{1*}

¹Department of Biophysics, Johns Hopkins University, 3400 North Charles Street, Baltimore, MD 21218, USA
²Department of Chemistry, United States Naval Academy, 572 Holloway Road, Annapolis, MD 21402, USA

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Introduction

A small fraction of ionizable residues in proteins are sequestered from water and buried in the protein interior.^{1–3} These internal ionizable groups are essential for catalysis,^{4–6} H⁺/e⁻ transport,^{7–10} and molecular recognition.¹¹ The pKa values of internal ionizable groups are usually different from the normal pKa values in water^{12–16} and are often tuned

for specific biological purposes.⁴ Understanding the determinants of these pKa values is important for quantitative description of the structural basis of function in a large variety of biological processes.

The shift in the pKa of an internal group relative to the normal pKa in water is governed by differences in the polarity and polarizability experienced by the charge in the two environments (A_{int}) and by Coulomb interactions with the charges of other ionizable groups. Structural reorganization of the protein coupled to the ionization of internal groups can also influence their pKa. One of the goals of this study was to examine the relative magnitude of these three determinants of the pKa values of internal groups.

The polarity and polarizability in the protein interior are usually lower than those in bulk water;

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Bausch Health Ireland Exhibit 2045, Page 1 of 14
 Mylan v. Bausch Health Ireland - IPR2022-00722

Table 1. Comparison of pKa values determined by linkage analysis and NMR spectroscopy

Variant	Residue	Linkage analysis		NMR	
		pKa	±	pKa	±
Δ+PHS/L38E	Glu38	7.0	0.3	7.0	0.1
Δ+PHS/L38E/E122Q	Glu38	—	—	6.2	0.1
Δ+PHS/L38D	Asp38	6.8	0.3	7.2	0.1
Δ+PHS/L38D/E122Q	Asp38	6.9	0.3	6.6	0.1

—, pKa could not be determined using linkage analysis because variant unfolds in an apparent three-state manner.

Intrachain Hydrogen Bonding Can Affect pKa



BLAKE ROBERT
PETERSON, Ph.D.

Q. How does the formation of intrachain hydrogen bonds affect the pKa of the involved amino acids?

THE WITNESS: Intrachain amino acids can -- intrachain interactions, I should say, can affect pKas, as they describe.

Q. In what ways does the formation of intrachain hydrogen bonds impact the modification of a lead compound for peptide drug development?

A. My answer is if an amino acid can form an intrachain amino acid, that could influence its pKa, and that might factor into drug development.

Petitioner's Arguments re Increased pH Are Unfounded

Proc. Natl. Acad. Sci. USA
Vol. 74, pp. 2705-2710, March 1997
Pharmacology

Regulation of intestinal uroguanylin/guanylin receptor-mediated responses by mucosal acidity

F. KENT HAMRA^{1,2}, SAMMY L. EBER¹, DAVID T. CHEN¹, MARK G. CURRIE^{1,3}, AND LEONARD R. FORTE^{1,4}

¹Truman Veterans Affairs Medical Center and ²Department of Pharmacology and Biochemistry and Molecular Biology Program, Missouri University, Columbia, MO 65212 and ³Basic Research and Development, St. Louis, MO 63107

Communicated by Philip Needleman, Monsanto Company, St. Louis, MO, January 2, 1997 (received for review March 27, 1996)

ABSTRACT Guanylin and uroguanylin are intestinal peptides that stimulate chloride secretion by activating a common set of receptor-guanylate cyclase signaling molecules located on the mucosal surface of enterocytes. High mucosal acidity, similar to the pH occurring within the fluid microclimate domain at the mucosal surface of the intestine, markedly enhances the cGMP accumulation responses of T84 human intestinal cells to uroguanylin. In contrast, a mucosal acidity of pH 5.0 renders guanylin essentially inactive. T84 cells were used as a model epithelium to further explore the concept that mucosal acidity imposes agonist selectivity for activation of the intestinal receptors for uroguanylin and guanylin, thus providing a rationale for the evolution of these related peptides. At an acidic mucosal pH of 5.0, uroguanylin is 100-fold more potent than guanylin, but at an alkaline pH of 8.0 guanylin is more potent than uroguanylin in stimulating intracellular cGMP accumulation and transepithelial chloride secretion. The relative affinities of uroguanylin and guanylin for binding to receptors on the mucosal surface of T84 cells is influenced dramatically by mucosal acidity, which explains the strong pH dependency of the cGMP and chloride secretion responses to these peptides. The guanylin-binding affinities for peptide-receptor interactions were increased by 100-fold at pH 5 versus pH 8, whereas the affinities of uroguanylin for these receptors were increased 10-fold by acidic pH conditions. Deletion of the N-terminal acidic amino acids in uroguanylin demonstrated that these residues are responsible for the increase in binding affinities that are observed for uroguanylin at acidic pH. We conclude that guanylin and uroguanylin evolved distinctly different structures, which enables both peptides to regulate, in a pH-dependent fashion, the activity of receptors that control intestinal salt and water transport via cGMP.

Guanylin and uroguanylin are structurally related peptides that were isolated from intestinal mucosa and urine (1-5). A receptor for guanylin and uroguanylin that has been identified at the molecular level is a transmembrane form of guanylate cyclase, termed GC-C (6). This membrane protein was originally discovered as an intestinal receptor for the heat-stable toxin (ST) peptides, which are secreted intraluminally by enteric bacteria that cause traveler's diarrhea (7). Bacterial ST peptides are related in primary structure to uroguanylin and guanylin, thus acting as molecular mimics of the enteric peptide hormones (reviewed in refs. 8 and 9). Membrane receptor-guanylate cyclases are found on the luminal surface of enterocytes throughout the small and large intestine and in other epithelia (10-13). Binding of peptide agonists to an extracellular domain of the receptor activates the intracellular

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2705

Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722

catalytic domain producing the second messenger cGMP within target enterocytes (1-6). Intracellular cGMP stimulates transepithelial chloride secretion by regulating the phosphorylation state and chloride channel activity of the cystic fibrosis transmembrane conductance regulator, an apical protein that is located with the receptors for uroguanylin, guanylin, and ST peptides (14-16).

Isolation of uroguanylin from opossum urine (2) followed by the cloning of a cDNA that encodes opossum pre-uroguanylin (17) revealed that the uroguanylin and guanylin genes are evolutionarily related (18-20). Furthermore, the mRNAs and precursor proteins for both uroguanylin and guanylin are expressed together throughout the mucosa of small and large intestine along with their receptors (5, 11, 17-20). This raised a question of whether the differences in primary structure between guanylin and uroguanylin evolved to regulate intestinal salt and water transport through a cooperative mechanism using common receptor-guanylate cyclase signaling molecules located on the mucosal surface of the intestine.

During the isolation of uroguanylin, guanylin, and their prohormone precursors, we observed that acidic column reagents markedly attenuated the cGMP responses of T84 cells to guanylin, but enhanced the responses to uroguanylin (4, 5). This pH dependency for activation of guanylate cyclase was successfully used to detect guanylin and uroguanylin during their separation and purification from intestinal mucosa. The possibility was then considered that the primary structures of guanylin and uroguanylin could have evolved to regulate the enzymatic activity of a common set of receptors over the wide range of mucosal acidity that occurs within the intestinal lumen during digestion (21-24). We report here that high mucosal acidity rendered guanylin ineffective as a cGMP-agonist and chloride secretagogue, whereas an acid pH markedly enhanced the potency of uroguanylin. A mucosal pH of 8.0 substantially increased the potency of guanylin but decreased the potency of uroguanylin. These changes in agonist potencies were explained by corresponding directional shifts in the affinities of guanylin and uroguanylin for binding to receptors at pH 5.0 versus 8.0. Uroguanylin and guanylin cooperatively regulate the guanylate cyclase activity of a common set of mucosal receptors in a pH-dependent fashion, thus providing an enteric signaling pathway for the intrinsic, paracrine regulation of intestinal salt and water transport.

MATERIALS AND METHODS

cGMP Accumulation Assay in T84 Cells. T84 cells were cultured in 24-well plastic dishes, and the cGMP levels were

Abbreviations: ST, heat-stable toxin.

¹Present address: Howard Hughes Medical Institute and Department of Pharmacology, University of Texas Southwest Medical Center, 5323 Harry Hines Boulevard, Dallas, TX 75235-9086.

²To whom reprint requests should be addressed at: Department of Pharmacology, School of Medicine, University of Missouri, Columbia, MO 65212; e-mail: Leonard_R_Forte@medical.missouri.edu.

MYLAN EXHIBIT - 1021

secretion responses to these peptides. The guanylin-binding affinities for peptide-receptor interaction were reduced by 100-fold at pH 5 versus pH 8, whereas the affinities of uroguanylin for these receptors were increased 10-fold by acidic pH conditions. Deletion of the N-terminal acidic amino

Ex. 1021 at 2705 (Abstract)
Patent Owner's Resp. at 48; Ex. 2024 ¶ 44

Petitioner's Arguments re Increased pH Are Unfounded



MICHAEL SAMUEL
EPSTEIN, M.D.

in colonic mucosa contains uroguanylin
nylin peptides

AGENT HAMRA, WILLIAM J. KRAUSE, SAMMY L. EBER, RONALD H. FREEMAN,

Q. So are you aware of any pharmacologic therapies available as of January 17th, 2002 that acted only in the small intestine?

THE WITNESS: Not that I can answer for you.

Q. Is that a "no"?

THE WITNESS: I would say I don't know of any specifically on the top of my head.

resulting in phosphorylation of protein kinase A (PKA) and cyclic monophosphate (cGMP) (7, 18). All species of mammals and birds examined express GC-C-like receptor activity on the apical surface of enterocytes throughout the intestine (21, 22). The opossum kidney also expresses high levels of GC-C-like receptors located in the apical membrane of proximal tubular cells (14). Guanylin was first isolated from rat jejunum as a heat-stable, 15-amino acid peptide that activated GC-C in human intestinal T84 cells (7). Guanylin cDNAs encoding 116- to 116-amino acid precursors have been isolated from rat, human, and mouse intestine (19). Uroguanylin was initially purified as 13- to 15-amino acid peptides from

MYLAN EXHIBIT - 1019 (Corrected)

Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd.

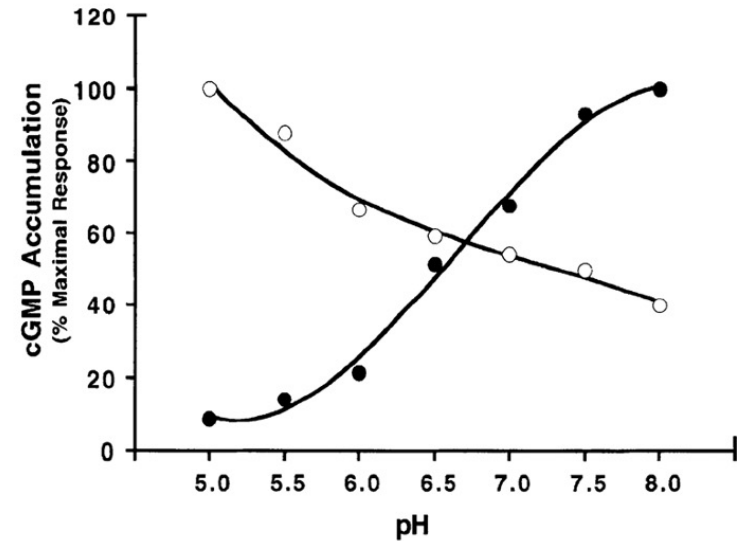


Fig. 1. Effects of medium pH on uroguanylin (○) and guanylin (●)-stimulated guanosine 3',5'-cyclic monophosphate (cGMP) accumulation in T84 cells. Vehicle, 30 nM synthetic opossum uroguanylin, and 30 nM synthetic opossum guanylin were suspended in buffered assay medium previously adjusted to pH values indicated, as described in MATERIALS AND METHODS. Levels of T84 cell cGMP accumulation (pmol/well, average of 3 wells) elicited by vehicle and peptides in this experiment when tested at pH 5.0 and pH 8.0, respectively, were as follows: basal (vehicle control) = 0.45 and 0.78, uroguanylin = 43.9 and 17.5, and guanylin = 0.85 and 10.0. Data are representative of 4 experiments with similar results.

Petitioner's Arguments re Increased pH Are Unfounded

PRP Pharmacology Research & Perspectives Open Access

ORIGINAL ARTICLE

Therapeutically targeting guanylate cyclase-C: computational modeling of plicanatide, a uroguanylin analog

Andrea Brancale¹, Kunwar Shailubhai², Salvatore Ferla¹, Antonio Ricci¹, Marcella Bassetto¹ & Gary S. Jacob²

¹School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, United Kingdom
²Syngene Pharmaceuticals, New York, New York

Keywords
guanylate cyclase C, linaclotide, molecular dynamics, plicanatide, uroguanylin

Correspondence
Andrea Brancale, Cardiff School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Redwood Building, King Edward VII Avenue, Cardiff, CF10 3NB.
Tel: +44 (0)29 2087 4480;
Fax: +44(29) 2087 4149;
E-mail: brancalea@cardiff.ac.uk

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doi: 10.1002/prp2.295

dx: 10.1002/prp2.295

Introduction
Chronic idiopathic constipation (CIC) and irritable bowel syndrome with constipation (IBS-C) are two of the most common conditions affecting the gastrointestinal (GI) tract, creating a burden on healthcare resources and leading to significant negative impact on quality of life (Hei delbaugh et al. 2015). These disorders are characterized by diminished stool frequency, straining and abdominal pain (IBS-C) or discomfort (CIC). CIC alone affects 14% of the population, while IBS-C affects 10% (Hei delbaugh et al. 2015). These disorders are characterized by diminished stool frequency, straining and abdominal pain (IBS-C) or discomfort (CIC). CIC alone affects 14% of the population, while IBS-C affects 10% (Hei delbaugh et al. 2015).

Abstract
Plicanatide is a recently developed guanylate cyclase C (GC-C) agonist and the first uroguanylin analog designed to treat chronic idiopathic constipation (CIC) and irritable bowel syndrome with constipation (IBS-C). GC-C receptors are found across the length of the intestines and are thought to play a key role in fluid regulation and electrolyte balance. Ligands of the GC-C receptor include endogenous agonists, uroguanylin and guanylin, as well as diarrhoeagenic, *Escherichia coli* heat stable enterotoxins (ST). Plicanatide mimics uroguanylin in its 2 disulfide bond structure and in its ability to activate GC-Cs in a pH dependent manner, a feature associated with the presence of acid sensing residues (Asp2 and Glu3). Linaclotide, a synthetic analog of STh (a 19 amino acid member of ST family), contains the enterotoxin's key structural elements, including the presence of three disulfide bonds. Linaclotide, like STh, activates GC-Cs in a pH independent manner due to the absence of pH sensing residues. In this study, molecular dynamics simulations compared the stability of plicanatide and linaclotide to STh. Three dimensional structures of plicanatide at various protonation states (pH 2.0, 5.0, and 7.0) were simulated with GROMACS software. Deviations from ideal binding conformations were quantified using root mean square deviation values. Simulations of linaclotide revealed a rigid conformer most similar to STh. Plicanatide simulations retained the flexible, pH dependent structure of uroguanylin. The most active conformers of plicanatide were found at pH 5.0, which is the pH found in the proximal small intestine. GC-C receptor activation in this region would stimulate intraluminal fluid secretion, potentially relieving symptoms associated with CIC and IBS-C.

Abbreviations
CIC, chronic idiopathic constipation; FGID, functional gastrointestinal disorder; GC-C, guanylate cyclase C; GI tract, gastrointestinal tract; IBS-C, irritable bowel syndrome with constipation; RMSD, root mean square deviation; ST, family of heat stable enterotoxin produced by enterotoxigenic *Escherichia coli* that include STh and STp; STh, 19 amino acid member of ST family.

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2017 | Vol. 5 | Iss. 2 | e00295
Page 1

Bausch Health Ireland Exhibit 2019, Page 1 of 10
Mylan v. Bausch Health Ireland - IPR2022-00722

As an analog of the pathological GC C agonist STh, linaclotide maintains many structural features of STh, including the presence of three disulfide bonds and an insensitivity to pH. MD simulations in this study show that the addition of a third intramolecular bond makes both STh and linaclotide insensitive to MD perturbations (Ozaki et al. 1991). The structural similarity of these two molecules is reflected by the low RMSD values of 1.28 Å for linaclotide. The amino acid substitutions that differentiate linaclotide from STh further enhance the pharmacokinetic stability and proteolytic resistance of linaclotide, allowing it to remain active across a longer portion of the small intestine (Bharucha and Waldman 2010; Harris and Crowell 2007). The absence of pH sensing amino acid residues would additionally give these molecules maximum biological activity across the range of pH environments in the GI tract. This lack of focused areas of

Aspartimide Formation Can Be Avoided Through Protecting Groups

Pepti

The Wave of t

Proceedings of the
and the Seventeenth
Peptide Symposium

Edited by

Michal Lebl
and
Richard A. Houghten

American Peptide Society

MYLAN EXHIBIT - 1022

Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722

This systematic investigation clearly showed that in our test system, no detectable amounts of aspartimide were formed if Hmb-backbone protection was applied in addition to standard OtBu-protection of the Asp side chain. However, the synthesis of all

compounds. The OMpe-protecting group showed a significant improvement with respect to aspartimide formation when compared to regular OtBu-protection. Most inter-

Ex. 1022 at 64

Patent Owner's Resp. at 56-57; Ex. 2024 ¶¶ 198-199

40

Aspartimide Formation Can Be Avoided Through Protecting Groups

Letters in Peptide Science, 7: 107-112, 2006.
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107

Base-induced side reactions in Fmoc-solid phase peptide synthesis: Minimization by use of piperazine as N^α-deprotection reagent*

John D. Wade**, Marc N. Mathieu, Mary Macris & Geoffrey W. Tregear
Howard Flory Institute, University of Melbourne, Parkville, VIC 3052, Australia

Received 1 November 1999; Accepted 24 November 1999

Key words: aspartimide formation, base-induced side reaction, Fmoc-solid phase peptide synthesis, N^α-deprotection reagent, piperazine

Summary

Base-induced aspartimide (cyclic imide) and subsequent base adduct formation in the Fmoc-solid phase synthesis of sensitive sequences are serious side reactions that are difficult to both anticipate and control. The effect of extended treatment of piperazine as N^α-Fmoc deprotection reagent on two sensitive peptide sequences was examined. For comparison, other bases were also investigated, including piperidine, 1-hydroxypiperidine, tetrabutylammonium fluoride, and 1,8-diazabicyclo[5.4.0]undec-7-ene. The results showed that all bases induced varying degrees of both aspartimide and, in some cases, base adduct formation, although piperazine caused the least side reaction. Use of N-(2-hydroxy-6-methoxybenzyl)peptide backbone amide protection was confirmed to confer complete protection against side reaction. In the absence of such protection, for all bases, the use of 1-hydroxybenzotriazole as additive had some, but not complete, beneficial effect in further reducing side reaction. Best results were obtained with piperazine containing 0.1M 1-hydroxybenzotriazole indicating that this reagent merits serious consideration for N^α-deprotection in the Fmoc-solid phase synthesis of base-sensitive sequences. A further advantage of this reagent is that it causes little racemisation of resin-bound C-terminal cysteine, an occasionally serious base-mediated problem in Fmoc-solid phase assembly.

Introduction

Aspartimide (cyclic imide) formation is a long-recognized side reaction that can occur both during solid phase peptide synthesis (SPPS) and storage of peptides, and may be either acid- or base-catalyzed [1]. Numerous studies on the mechanism of the reaction have shown it to be dependent on the nature of the acid or base, and the residue adjoining the carboxyl of the aspartate as well as the side chain protecting group used [1]. Imide formation was originally thought not to occur in Fmoc-SPPS. However, several recent studies have shown it to be a significant side reaction and one that is highly sequence and conformation dependent [2-4]. The problem is not confined

exclusively to Asp-X sequences, for there has also been a report of Asn-X cyclization [5]. An additional side reaction now known to be associated with sensitive Asp-X sequences is subsequent modification of the imide by nucleophilic base to produce a base adduct (Figure 1). Several palliative measures for controlling imide and adduct formation have been recommended. These include addition to the N^α-Fmoc deprotection reagent of choice, piperidine, of agents such as 1-hydroxybenzotriazole (HOBt), but none completely suppress side reaction [2,6,7]. Aspartyl side chain protecting groups other than the commonly employed *tert*-butyl ester have also been reported to give improved yields of α -aspartyl peptides through increased steric hindrance. These include 1-salamonyl and β -3-methylpent-3-yl esters [2,8]. However, these are either not entirely compatible with Fmoc-SPPS or commercially unavailable. The sole effective preventive measure to date is the use of Asp-X amide bond protec-

* A preliminary account of this work was presented at the 25th European Peptide Symposium, Budapest, Hungary, 1998.
** To whom correspondence should be addressed. E-mail: j.wade@hfl.inmehb.unimelb.au

MYLAN EXHIBIT - 1023

Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722

Conclusions

In the absence of prior information regarding the susceptibility of a new peptide sequence to modification during Fmoc-solid phase synthesis, it is recommended that – where feasible – Asp-X pairs be routinely protected with the Hmb moiety. Should this not be possible, then piperazine containing 0.1M HOBt is a practical and effective alternative.

Ex. 1023 at 111

Patent Owner's Resp. at 56-57; Ex. 2024 ¶¶ 198, 200

Aspartimide Formation Can Be Avoided Through Protecting Groups



alkyl amino acid prior to Fmoc-Asp(*Ot*Bu). Based on our study, the most effective combinations for minimization of aspartimide formation were (i) *t*Bu side-chain protection of aspartate, piperidine for removal of the Fmoc group, and either HOBt or Dnp as an additive to the piperidine solution; or (ii) 1-Ada side-chain protection of aspartate and DBU for removal of the Fmoc group.

Aspartimide Formation Can Be Avoided Through Protecting Groups



BLAKE ROBERT
PETERSON, Ph.D.

Q. How can aspartimide formation in peptides be reduced or avoided using protecting groups?

THE COURT REPORTER: I'm sorry, protecting?

THE WITNESS: Protecting groups.

THE COURT REPORTER: Thank you.

MR. HASFORD: Protecting groups.

THE WITNESS: One can reduce side reactions in chemical synthesis in general by using protecting groups in some cases. They -- they can limit the undesired pathways that lead to undesired products.

Objective Evidence Supports the Nonobviousness of the Claims

Federal Circuit: Objective Evidence of Nonobviousness



Objective evidence . . . must be considered before a conclusion on obviousness is reached and is not merely “icing on the cake.”

Plecanatide's Unexpected Stabilization Against Interconversion

STUDIES ON SP-304 THERMOSTABILITY, pH DEPENDENCY AND TOPOISOMERIC STABILITY

Study Number: SP-PI-004
Test Article: SP-304
Author: Kunwar Shalubhai
Senior Vice President, Discovery
Synergy Pharmaceuticals, Inc.
Study Dates: November 24, 2001 to December 25, 2001
Testing Facility: R&D Center, Synergy Pharmaceuticals, Inc., Norristown, PA
Final Report Date: February 15, 2008
Source Data: Synergy Pharmaceuticals Lab Notebook # 2, Pages: 170-185
Of Dr. Surendra Dheer

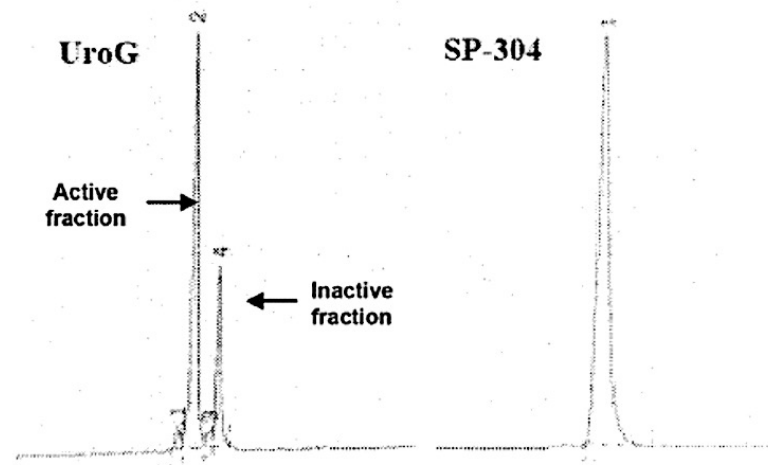
Synergy Pharmaceuticals, Inc.
420 Lexington Ave. - Suite 1609
New York, NY 10170

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HIGHLY CONFIDENTIAL INFORMATION TRUL00018260

PROTECTIVE ORDER MATERIAL Bausch Health Ireland Exhibit 2028, Page 1 of 18
Mylan v. Bausch Health Ireland - IPR2022-00722

Figure 1: HPLC Chromatographs of Uroguanylin and SP-304 Following 16-Hour Incubations at pH 3.0 in Aqueous Media at 37°C



Ex. 2028 at TRUL00018269 (Fig. 1)
Patent Owner's Resp. at 59; Ex. 2024 ¶¶ 206-215

Marx: Topoisomerism Not Affected by N-Terminal Region

U.C. Marx
J. Klodt
M. Meyer
H. Gerlach
P. Rösch
W.-G. Forssmann
K. Adermann

One peptide, two topologies:
structure and interconversion
dynamics of human
uroguanylin isomers

Authors' affiliations:

U.C. Marx, Niedersächsisches Institut für
Peptid-Forschung (IPF), Hannover, Germany,
and Lehrstuhl für Struktur und Chemie der
Biopolymere, Universität Bayreuth, Bayreuth,
Germany.

J. Klodt, M. Meyer, W.-G. Forssmann, and K.
Adermann, Niedersächsisches Institut für
Peptid-Forschung (IPF), Hannover, Germany.
H. Gerlach, Lehrstuhl für Organische Chemie II,
Universität Bayreuth, Bayreuth, Germany.
P. Rösch, Lehrstuhl für Struktur und Chemie
der Biopolymere, Universität Bayreuth,
Bayreuth, Germany.

Correspondence to:

Dr. Karl Adermann
Niedersächsisches Institut für Peptid-Forschung
(IPF)
Feldberg-Strasse 59
D-30645 Hannover, Germany
Phone: Int +49-51-1464343
Fax: Int +49-51-1464603
E-mail: karl.adermann@ipf-peptide.com

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two topologies: structure and interconversion
dynamics of human uroguanylin isomers. *J Peptide
Res* 18, 139–149

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ISSN 0367-0882

Key words: guanylin; heat-stable enterotoxin; isomerization;
solution structure; topological stereoisomer; uroguanylin

Abstract: The peptide hormone uroguanylin stimulates chloride
secretion via activation of intestinal guanylyl cyclase C (GC-C). It is
characterized by two disulfide bonds in a 1–3/2–4 pattern that
causes the existence of two topological stereoisomers of which
only one induces intracellular cGMP elevation. To obtain an
unambiguous structure-function relationship of the isomers, we
determined the solution structure of the separated uroguanylin
isoforms using NMR spectroscopy. Both isomers adopt well-defined
structures that correspond to those of the isomers of the related
peptide guanylin. Furthermore, the structure of the GC-C-
activating uroguanylin isomer A closely resembles the structure of
the agonistic Escherichia coli heat-stable enterotoxin. Compared
with guanylin isomers, the conformational interconversion of
uroguanylin isomers is retarded significantly. As judged from
chromatography and NMR spectroscopy, both uroguanylin
isoforms are stable at low temperatures, but are subject to a slow
pH-dependent mutual isomerization at 37°C with an equilibrium
isomer ratio of approximately 1:1. The conformational exchange is
most likely under the steric control of the carboxy-terminal
leucine. These results imply that GC-C is activated by ligands
exhibiting the molecular framework corresponding to the structure
of uroguanylin isomer A.

Abbreviations: cGMP, cyclic 3',5'-guanosine monophosphate;
Clean-TOCYSY, TOCSY with suppression of NOESY-type cross peaks;
DG, distance geometry; DQF-COSY, double-quantum filtered COSY;
DSS, 2,2-dimethyl-silapentane-5-sulfonic acid; GC-C, guanylyl
cyclase C; IR-NOESY 2D NOESY spectrum acquired with a jump-
return observe pulse; MD, molecular dynamics; NMR, nuclear
magnetic resonance; NOE, nuclear Overhauser effect, also used for
NOESY cross peak; NOESY, NOE spectroscopy; RMSD, root-mean-

139

Bausch Health Ireland Exhibit 2010, Page 1 of 12
Mylan v. Bausch Health Ireland - IPR2022-00722

The slow development of the equilibrium between uroguanylin isomers at alkaline pH indicates that the ionization state of the isomeric molecules strongly influences the kinetics of transition between the isomers of uroguanylin 16 and uroguanylin 24. Thus, the terminal carboxyl, ionizable side-chains of Asp2, Asp3 and Glu5, or those groups able to form intrachain hydrogen bonds, may be involved in the control of stabilization of the two isomers. After 3 days at alkaline pH, both isoforms decomposed because of disulfide exchange. Comparison of the conversion of uroguanylin 16 isomers with the isomers of the N-terminally extended uroguanylin-24 resulted in identical kinetics for isoforms A and B at a pH of 4.5 and 7.7 for both peptides (Fig. 6C). This result clearly demonstrates that the isomerization is not affected by the amino-terminal region of uroguanylin. Corresponding to the

Ex. 2010 at 236

Patent Owner's Resp. at 59; Sur-Reply at 26; see also Ex. 2024 ¶¶ 37, 147, 210-214

Fiser: Asp Preferred Over Glu for Stabilization

FEBS Letters 397 (1996) 225–229

Conservation of amino acids in multiple alignments: aspartic acid has unexpected conservation

András Fiser^{a,b}, István Simon^b, Geoffrey J. Barton^{a,*}

^aUniversity of Oxford, Laboratory of Molecular Biophysics, The Rex Richards Building, South Parks Road, Oxford OX1 3QU, UK
^bInstitute of Enzymology, Biological Research Center, Hungarian Academy of Sciences, P.O. Box 7, Budapest H-1518, Hungary

Received 18 July 1996; revised version received 25 September 1996

Abstract Analysis of the relationship between surface accessibility and amino acid conservation in multiple sequence alignments of homologous proteins confirms expected trends for hydrophobic amino acids, but reveals an unexpected difference between the conservation of Asp, Glu and Gln. Even when not in an active site, Asp is more highly conserved than Glu. There is a clear preference for conserved and buried Asp to be present in coil, but there is no tendency for Asp to conserve ϕ/ψ in the $\alpha+$ region of the Ramachandran map. Glu does not show any preference to be conserved in a particular secondary structure. Analysis of recently derived substitution matrices (e.g. BLO-SUM) confirms that Glu tends to substitute more frequently with other amino acids than does Asp. Analysis of relative accessibility versus relative conservation for individual amino acid positions in alignments shows a negative correlation for all amino acid types. With the exception of Arg, Lys, Gly, Gln, Asp and Tyr, a relative conservation of > 2 suggests the amino acid will have a relative accessibility of $< 50\%$. Observation of conserved Cys, Gly or Asp in a reliable multiple alignment suggests a position important for the structure of the protein. Furthermore, the Asp is likely to be involved in polar interactions through its side-chain oxygen atoms. In contrast, Glu is the least conserved amino acid overall.

Key words: Conservation analysis; Multiple sequence alignment; Protein structure prediction

1. Introduction

Knowledge of protein sequences is growing much faster than knowledge of either three-dimensional structure or function. Accordingly, the interpretation of sequence data to identify structurally or functionally important residues is essential if the data are to be effective in furthering understanding of biological systems. Multiple sequence alignments of families of protein sequences are now used routinely to indicate residues of key importance to the function of the protein. A position in an alignment that has identical residues in all members of a protein family may have a key catalytic role.

A position where similar physico-chemical properties (e.g. hydrophobicity) are shared may suggest importance in stabilizing the native conformation of the protein [1,2]. Identification of such conserved features in multiple alignments has been used to good effect to improve the accuracy of prediction of secondary structure and buried residues (α -helix and β -strand) (e.g. [3–7]).

Here we report a systematic study of residue conservation in multiple alignments where at least one protein is of known

tertiary structure. Our analysis complements that of Overington et al. [8] who considered only pairwise substitution frequencies for amino acids in structurally aligned families.

2. Materials and methods

2.1. Data base

A non-redundant set of 81 proteins was generated from the April 1993 release of the Brookhaven Protein Data Bank (PDB) [9]. The set was chosen in a two-step procedure. First, all pairs of chains (over 50 residues and resolution better than 2.5 Å) in the data bank were compared by calculating correlation coefficients between the dipeptide frequencies in each protein. A set of 101 protein chains was selected such that all pairs had a correlation of < 0.4 . All pairs in this set were then compared by a rigorous sequence comparison method [10,11] followed by cluster analysis. This reduced the set to 81 protein chains that show no obvious sequence similarity (PDB code and chain identifiers: 15SC, IACK, IALC, IBBP, A1CC5, IBCA, IFKI, IFSR, IGGK, IGPI, A1HDS8, IHIP, IHOE, ILRD, A1PAZ, IPCY, IPIH, IPRC, C1RP, IRHD, IRNH, ISN1, ITR5, ITPK, A1NSY, B1S08, A1GDP, A2AA, A1ZAB, ZCDA, XCDV, ZCZF, JFXB, ZD5, ZLH1, ZLIV, ZLTN, A1ZOR, L2PAB, A1ZNT, ZRSP, A1ZSC, J1SNL, E1SNG, ZSDG, B1ZSI, ZSYV, ZTK, ZITGL, A1ADK, IKSQ, ICLG, JPKC, JGAP, B1ZLM, JSGB, J451C, 48P1, 4FD1, 4FXN, 4HHR, A14EP, 4FFK, 4PFF, 4TNC, XGTS, SCVT, B1SEK, SRUS, A1SRN, 4LWH, STMN, E17P1, RADH, SATC, B1CAT, A1BFR, SPAP, 9R3A, A1PKCA).

Each protein in the set was compared by the Smith-Waterman algorithm [11,12] to the NBRF-PHR sequence data bank (Release 38) and all sequences that gave probability values of $< 10^{-4}$ by a length-dependent scoring scheme (program SCANSP ftp://goeff.bio.psu.ac/pub/program/scansp) were multiple aligned with the query sequence by the algorithm of Barton and Sternberg [13]. This gave 81 alignments with between 3 and 499 sequences in each (median of 28 sequences).

2.2. Calculation of conservation and accessibility

Conservation scores based upon the physico-chemical properties of the amino acids were calculated for each position in each alignment according to Livshits and Barton [2]. Conservation scores range from 0 to 19 and represent the number of the properties: Hydrophobic, Positive, Negative, Polar, Charged, Small, Tiny, Aliphatic, Aromatic, Proline and their negatives (e.g. not positive) that are shared at a position. The program AMAS [14], which calculates conservation values from a multiple alignment, may be run over the World Wide Web (<http://goeff.bio.psu.ac.uk/ukherve/rtt/amas-server.html>).

Although conservation scores are absolute, the relative importance of a conservation score is dependent on the overall similarity between the sequences in the multiple alignment. For example, in an alignment of 20 sequences that all share $> 90\%$ pairwise identity conservation scores above 8 may be interesting. In contrast, if the pairwise identity is below 30%, then lower conservation scores will be informative. Accordingly, in this study we normalized conservation scores by the average conservation for each alignment to give relative conservation scores *C*. We refer to a position as conserved if $C \geq 1$.

Accessible surface areas were calculated by the program DSSP [14] and converted to relative accessibilities by dividing by the accessibility of the residue in a Gly-X-Gly tripeptide [15]. Two relative accessibility classes were considered $0.4 \leq a \leq 0.25$ (buried) and $0.25 < a \leq 0.4$ (exposed).

Since our data do not suggest a significant preference for $\alpha+\phi/\psi$, the preferred conservation of Asp is likely to be due to differing side-chain interactions. The most obvious hypothesis is that since Glu has a higher proportion of non-polar atoms than Asp it can make more non-specific interactions and so there are fewer constraints on its environment. In

Why, then, is Asp most highly conserved when buried in coil? The short Asp side chain is restricted in mobility yet able to make strong polar interactions. It is possible that Asp may form a ‘pin’ that stabilises non-regular structures in loops.

*Corresponding author. Fax: (44) (1865) 510454.

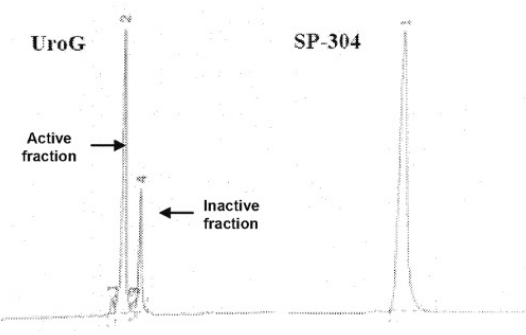
E-mail: gjb@bioch.ox.ac.uk

0164-5704/96/0025-0000 © 1996 Federation of European Biochemical Societies. All rights reserved.

PII S0164-5703(96)01181-7

Patent Owner Did Not Use Wild-Type Human Uroguanylin

Figure 1: HPLC Chromatographs of Uroguanylin and SP-304 Following 16-Hour Incubations at pH 3.0 in Aqueous Media at 37°C




ionization state of functional group(s) in the molecule. At acidic pH, both compounds are, in one sense, freely convertible (same conversion rates) and eventually come to a 1:1 equilibrium ratio. In contrast, conversions of both compounds

h. At 37°C, 25% of both uroguanylin-16 isomers are interconverted within 24 h. This and conversion experiments

Ex. 2028 at TRUL00018269 (Fig 1); Ex. 2010 at 236, Ex. 2011 at 30
Sur-Reply at 19; see also Patent Owner's Resp. at 59; Ex. 2024 ¶¶ 70, 131, 206-215

Petitioner's Uncorroborated Data Do Not Include Human Uroguanylin as a Comparator


Beschwerdekammern
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Chambres de recours

European Patent Office
 82581 MUNICH
 GERMANY
 Tel: +49 (0)89 2399-0
 Fax: +49 (0)89 2399-4485

Datum/Date
 11.06.13

Zeichen/Reference/Référence
 117839 OPPO01 Anmeldung Nr./Application No./Demande n°/Patent No./Brevet n°
 02721604.3 / 1379224

Anmelder/Applicant/Demandeur/Patenthaber/Propriétaire/Titulaire
 Synergy Pharmaceuticals, Inc.

Appeal number: **T1366/12-3.3.04**

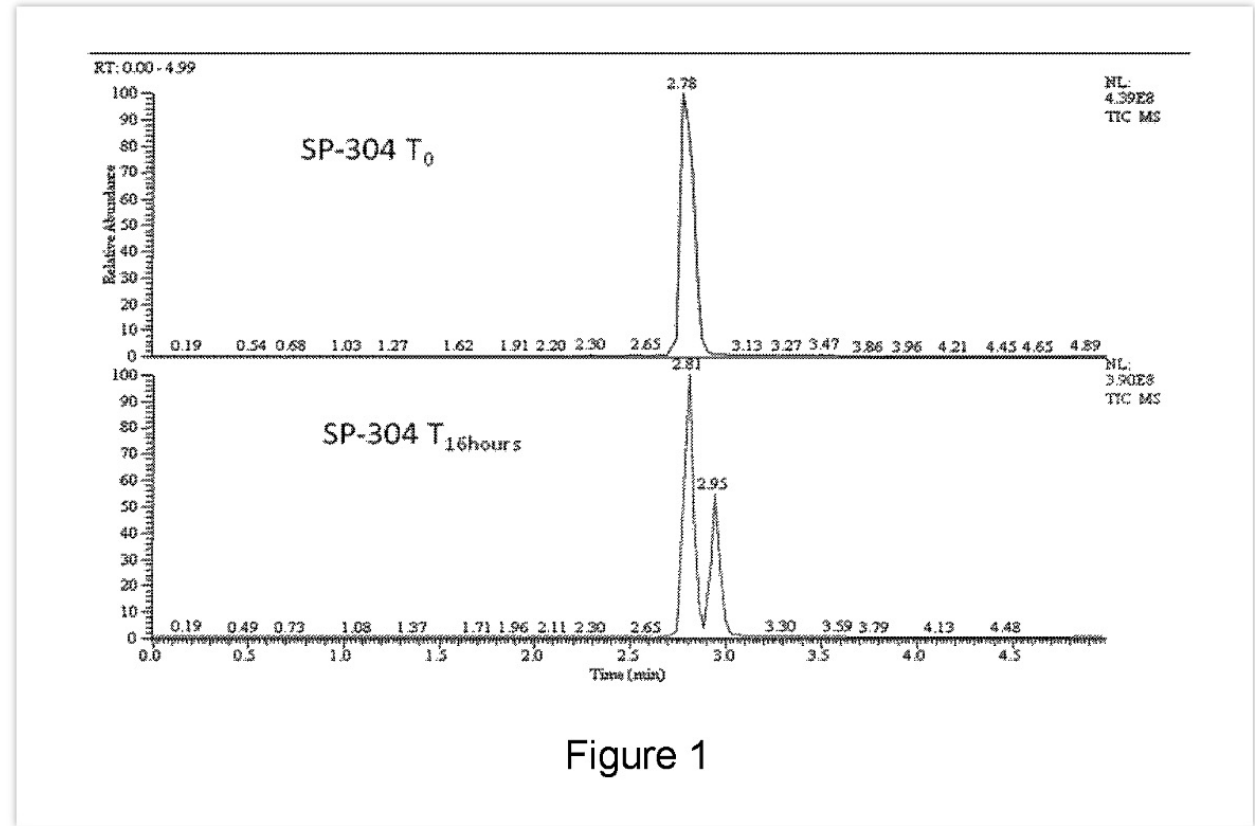
Please find enclosed a copy of the decision dated 04.06.13.

The Registrar P. Cremona
 Tel.: 089 / 2399 - 3341

Annex(es): Acknowledgement of receipt - EPO Form 2936

Registered letter with advice of delivery

EPO Form 3032 01/11 MYLAN EXHIBIT - 1067, Pg. 001
 Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd.
 IPR2022-00722



Ex. 1067 at 91
 Sur-Reply at 25-26; see also Patent Owner's Resp. at 59; Ex. 2024 ¶¶ 206-215

Plecanatide's Significantly and Unexpectedly Superior Potency

SP-304: STIMULATION OF INTRACELLULAR cGMP SYNTHESIS IN T84 CELLS

Study Number: SP-PH-001
 Test Article: SP-304
 Author: Kunsar Shalubhai
 Senior Vice President, Discovery
 Synergy Pharmaceuticals, Inc.
 Study Dates: September 2001 to November 2001
 Testing Facility: R&D Center, Synergy Pharmaceuticals, Inc., Norristown, PA
 Final Report Date: February 15, 2008
 Source Data: Synergy Pharmaceuticals Lab Notebook # 2; Pages: 70-91
 Dr. Surendra Dheer

Synergy Pharmaceuticals, Inc.
 420 Lexington Ave. - Suite 1609
 New York, NY 10170

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HIGHLY CONFIDENTIAL INFORMATION TRUL00018203

PROTECTIVE ORDER MATERIAL Bausch Health Ireland Exhibit 2027, Page 1 of 22
 Mylan v. Bausch Health Ireland - IPR2022-00722

Table 1. Effects of SP-304, Uroguanylin, and Other Test Peptides in the T84 cGMP Stimulation Bioassay

Test Material	Concentration		cGMP Levels (pmol/well) *	EC ₅₀ **	
	Molar	ng/mL		Molar	ng/mL
SP-301 (uroguanylin)	0	0	0	40 ⁻⁶ M 2.3x10 ⁻⁷ M	1668 383.6
	10 ⁻⁹ M	1.668	0		
	10 ⁻⁸ M	16.668	12		
	10 ⁻⁷ M	166.8	82		
	10 ⁻⁶ M	1668	205		
	10 ⁻⁵ M	16680	254		
SP-302	0	0	0	40 ⁻⁶ M 3.5x10 ⁻⁷ M	1696 593.6
	10 ⁻⁹ M	1.696	0		
	10 ⁻⁸ M	16.96	8		
	10 ⁻⁷ M	169.6	62		
	10 ⁻⁶ M	1696	185		
	10 ⁻⁵ M	16960	248		
SP-303	0	0	0	40 ⁻⁶ M 2.4x10 ⁻⁷ M	1682 403.7
	10 ⁻⁹ M	1.682	0		
	10 ⁻⁸ M	16.82	12		
	10 ⁻⁷ M	168.2	82		
	10 ⁻⁶ M	1682	195		
	10 ⁻⁵ M	16820	254		
SP-304	0	0	0	40 ⁻⁷ M 1.1x10 ⁻⁷ M	168.2 185.0
	10 ⁻⁹ M	1.682	0		
	10 ⁻⁸ M	16.82	17		
	10 ⁻⁷ M	168.2	149		
	10 ⁻⁶ M	1682	320		
	10 ⁻⁵ M	16820	315		

* cGMP levels in T84 cells after a 30-minute incubation.

** EC₅₀: median effective concentration (required to induce a 50% effect)

Reason for Amendment: To update the scientific notation for the EC50 molar value for each peptide to include the coefficient as well as the exponent and to update the corresponding ng/mL values. The new values were calculated using Prism 6 (Version 6.05) using a nonlinear regression curve fit (log[peptide concentration] versus cGMP level) using a least squares fit with no constraints)

Ex. 2027 at TRUL00018211-212, TRUL00018222

Sur-Reply at 26; see also Patent Owner's Resp. at 60-61; EX. 2024 ¶¶ 216-220; Ex. 2025 ¶¶ 97-99

Significantly Superior Potency Would Have Been Unexpected

US07041786B2

(12) **United States Patent** (10) **Patent No.:** **US 7,041,786 B2**
Shailubhai et al. (45) **Date of Patent:** **May 9, 2006**

(54) **GUANYLATE CYCLASE RECEPTOR AGONISTS FOR THE TREATMENT OF TISSUE INFLAMMATION AND CARCINOGENESIS**

(75) Inventors: **Kumar Shailubhai**, Blue Bell, PA (US); **Gregory Nieklerovich**, St. Louis, MO (US); **Gary S. Jacob**, Creve Coeur, MO (US)

(73) Assignee: **Callisto Pharmaceuticals**, New York, NY (US)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 362 days.

(21) Appl. No.: **10/107,814**

(22) Filed: **Mar. 28, 2002**

(65) **Prior Publication Data**
 US 2003/0073628 A1 Apr. 17, 2005

Related U.S. Application Data

(66) Provisional application No. 60/348,646, filed on Jan. 17, 2002.

(51) **Int. Cl.**
A61K 38/22 (2006.01)

(52) **U.S. Cl.**
 530/317; 530/300; 530/326; 514/10; 514/13

(58) **Field of Classification Search**
 530/317; 530/300; 326; 514/10; 13
 See application file for complete search history.

(56) **References Cited**

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FOREIGN PATENT DOCUMENTS

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 WO WO 02/098912 A3 12/2002

MYLAN EXHIBIT - 1001
 Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd. - IPR2022-00722



uroguanylin

Asp² & Asp³ → Glu² & Glu³

Asp² → Glu²

Asp³ → Glu³

TABLE 4

Peptide agonists evaluated for biological activity in the T84 cell bioassay.

SEQ ID NO.*	Compound Code	cGMP Level** (pmol/well)
1	SP301	205
6	SP302	225
7	SP303	195
20	SP304	315
14	SP306	0
4	SP310	0
21	SP316	275

*SEQ ID's for SP301, SP304 and SP316 are the precise amino acid sequences for these analogs as given in the text.


**Intracellular cGMP level observed in T84 cells following treatment with 1 micromolar solution of the respective peptide agonist for 30 minutes. The value observed for SP304 was statistically significant with a p > 0.5.

Peptides shown in Table 4 were custom synthesized and purified (>95% purity) using a published procedure (38).

Ex. 1001 at 15:53-54, 16 (Table 4)

Patent Owner's Resp. at 63-64; Sur-Reply at 18-19; EX. 2024 ¶¶ 221-222; Ex. 2025 ¶¶ 100-102

Currie's Data Are Internally Inconsistent


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GERMANY
Tel: +49 (0)89 2399-0
Fax: +49 (0)89 2399-4485

Zeichnungsnummer / Application No. / Demande n° / Patent No. / Brevet n°
 117839 OPPO01 02721604.3 / 1379224

Anmelder / Applicant / Demander / Patentinhaber / Propriétaire / Titulaire
 Synergy Pharmaceuticals, Inc.

Datum / Date
 11.06.13

Appeal number: **T1366/12-3.3.04**

Please find enclosed a copy of the decision dated 04.06.13.

The Registrar P. Cremona
 Tel.: 089 / 2399 - 3341

Annex(es): Acknowledgement of receipt - EPO Form 2936

Registered letter with advice of delivery

EPO Form 3032 01/11 MYLAN EXHIBIT - 1067 Pg. 001
 Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd.
 IPR2022-00722

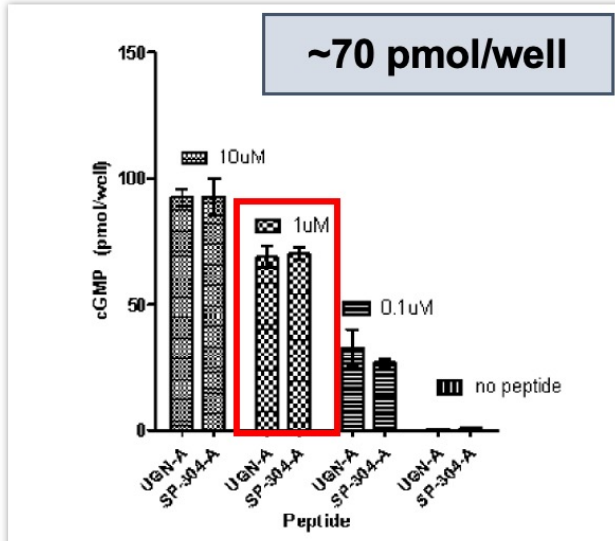


Figure 1

Concentration	p-value	Significance
10 µM	0.9889	ns
1 µM	0.8190	ns
0.1 µM	0.5402	ns

Table 1

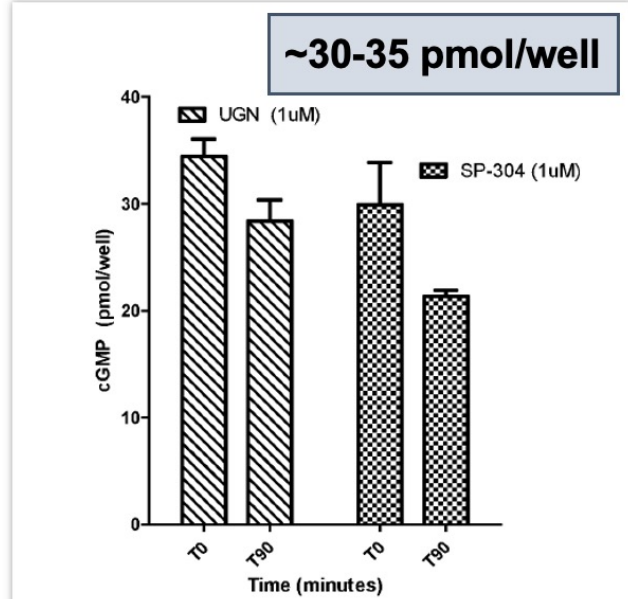
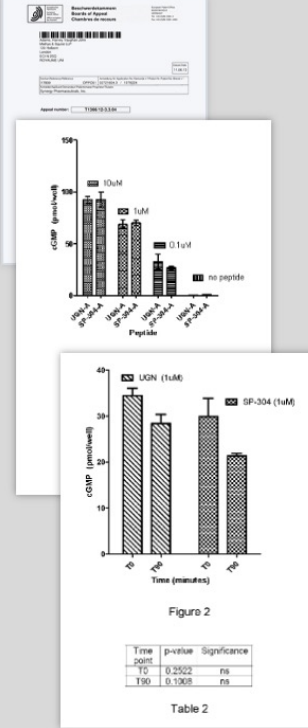


Figure 2

Time point	p-value	Significance
T0	0.2522	ns
T90	0.1008	ns

Table 2

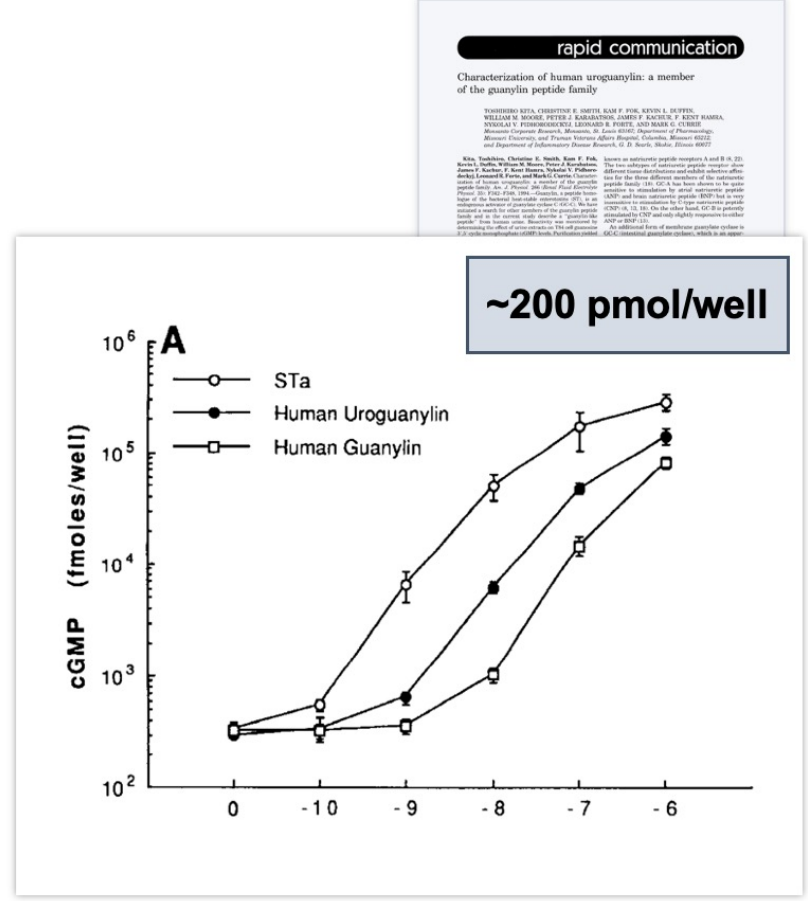
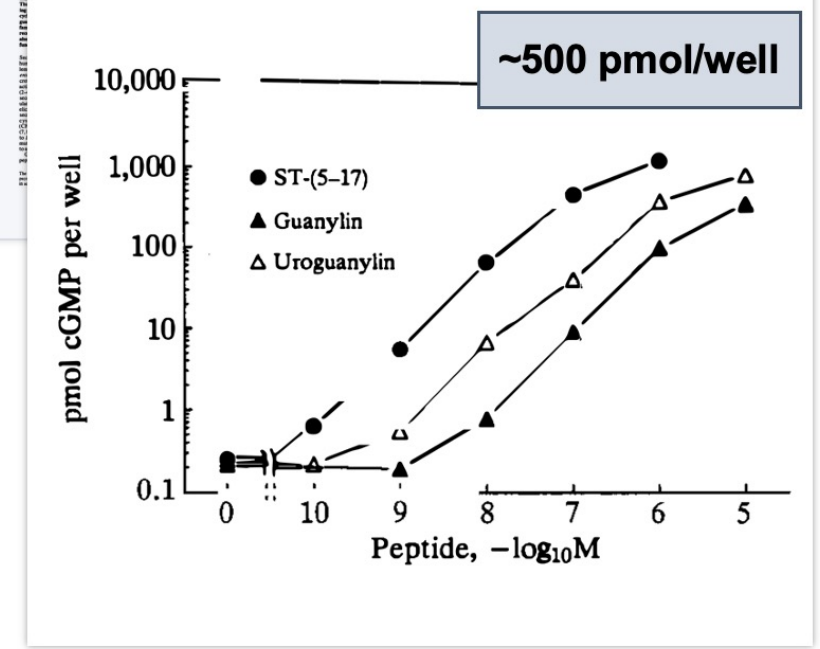
Currie's Data Are Inconsistent with the Literature



Uroguanylin: Structure and activity of a second endogenous peptide that stimulates intestinal guanylate cyclase

Y. Kato, M. Kato, T. Iijima, K. Fuyuki, S. Kato, L. Eise, Y. Nishida, Y. Yamaguchi, Y. Watanabe, M. H. Hamada, D. Cavall, J. A. Townsend, K. F. Hu, C. O'Connell, E. Sirtori, K. Nishida, A. S. B. S. Silva, and M. G. Currie

The Journal of Neuroscience, August 11, 2010; 30(32):10933-10942



Ex. 2012 at 10467 (Fig. 5); Ex. 2065 at 346 (Fig. 4A)
 Sur-Reply at 21-22

Plecanatide's Unexpected pH Sensitivity

STUDIES ON SP-304 THERMOSTABILITY, pH DEPENDENCY AND TOPOISOMERIC STABILITY

Study Number: SP-PH-004
 Test Article: SP-304
 Author: Kunwar Shailubhai, Senior Vice President, Discovery, Synergy Pharmaceuticals, Inc.
 Study Dates: November 24, 2001 to December 25, 2001
 Testing Facility: R&D Center, Synergy Pharmaceuticals, Inc., Norristown, PA
 Final Report Date: February 15, 2008
 Source Data: Synergy Pharmaceuticals Lab Notebook # 2; Pages: 170-185
 Of/Dr: Surendra Dheer

Synergy Pharmaceuticals, Inc.
 420 Lexington Ave. - Suite 1609
 New York, NY 10170

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HIGHLY CONFIDENTIAL INFORMATION TRUL00018260

Bausch Health Ireland Exhibit 2028, Page 1 of 18
 Mylan v. Bausch Health Ireland - IPR2022-00722

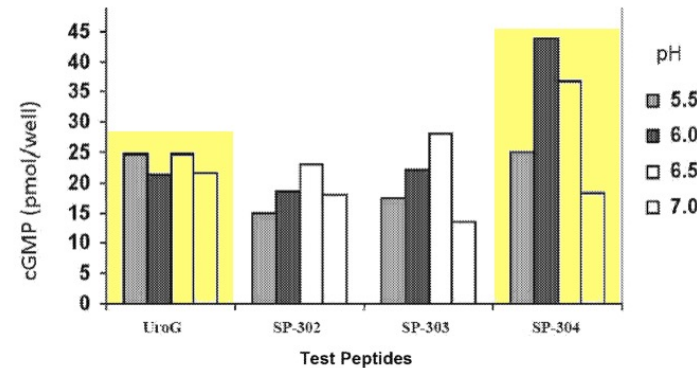
PROTECTIVE ORDER MATERIAL

Table 2: Raw Data (pH sensitivity assays)

Test Peptide	Concentration	cGMP Levels (pmol/well) *			
		pH 5.5	pH 6.0	pH 6.5	pH 7.0
Uroguanylin	0.1 μ M (1.67 μ g/mL)	24.6	21.36	24.6	21.72
SP-304	0.1 μ M (1.68 μ g/mL)	24.9	43.8	36.68	18.18
SP-302	0.1 μ M (1.67 μ g/mL)	14.80	18.6	22.98	17.94
SP-303	0.1 μ M (1.68 μ g/mL)	17.46	22.26	28.14	13.62

* cGMP levels in T84 cells after a 30-minute incubation at 37°C.

Figure 2: Effect of 30 Minute Incubations of 0.1 μ M Concentrations of SP-304, Uroguanylin, SP-302 and SP-303 on cGMP Production at Various Physiological pH Values in T84 Cells



Plecanatide's Unexpected pH Sensitivity



**SCOTT
WALDMAN, M.D.,
Ph.D.**

UNITED STATES PATENT AND TRADEMARK OFFICE
BEFORE THE PATENT TRIAL AND APPEAL BOARD

MYLAN PHARMACEUTICALS INC.,
Petitioner,
v.
BAUSCH HEALTH IRELAND LIMITED,
Patent Owner.

Case IPR2022-00722
U.S. Patent No. 7,041,786

DECLARATION OF SCOTT A. WALDMAN, M.D., PH.D., FDP, FAHA,
FNAI, FASPET

PROTECTIVE ORDER MATERIAL

Bausch Health Ireland Exhibit 2025, Page 1 of 64
Mylan v. Bausch Health Ireland - IPR2022-00722

110. In my opinion, these results are clinically significant and would have been unexpected to a person of ordinary skill in the art. Nothing in the prior art taught that the one amino acid substitution of plecanatide would produce this significant improvement. This increase in cGMP production is surprising, but additionally, plecanatide's increased pH sensitivity would have been entirely unexpected. It is also clinically meaningful. As discussed above, this unexpected pH sensitivity in the acidic areas of the intestines allows plecanatide's unexpectedly superior selective activity in the small intestine to provide fluid into the intestine to treat constipation, rather than in the colon, where its activity drops (pH 7) and is thus less likely to cause adverse effects, like diarrhea. In other words, plecanatide's targeted activity in the small intestine (not in the colon) advantageously reduces the diarrhea rate in patient populations.

Plecanatide's Significantly and Unexpectedly Superior Heat Stability

STUDIES ON SP-304 THERMOSTABILITY, pH DEPENDENCY AND TOPOISOMERIC STABILITY

Study Number: SP-PH-004
 Test Article: SP-304
 Author: Kunwar Shailubhai
 Senior Vice President, Discovery
 Synergy Pharmaceuticals, Inc.
 Study Dates: November 24, 2001 to December 25, 2001
 Testing Facility: R&D Center, Synergy Pharmaceuticals, Inc., Norristown, PA
 Final Report Date: February 15, 2008
 Source Data: Synergy Pharmaceuticals Lab Notebook # 2; Pages: 170-185
 Of/Dr: Surendra Dheer

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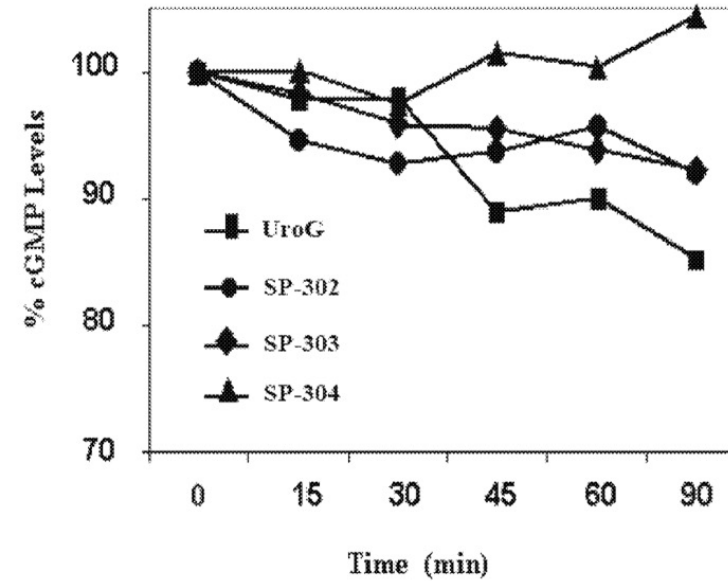
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PROTECTIVE ORDER MATERIAL

Bausch Health Ireland Exhibit 2028, Page 1 of 18
 Mylan v. Bausch Health Ireland - IPR2022-00722

Figure 3: Thermostability of SP-304, Uroguanylin, SP-302 and SP-303 Following Heat Treatment at 95°C in T84 Cells as a Function of Time



Plecanatide's Significantly and Unexpectedly Superior Binding Affinity

ANTICANCER RESEARCH 29: 3777-3784 (2009)

In Vivo Imaging of Human Colorectal Cancer Using Radiolabeled Analogs of the Uroguanylin Peptide Hormone

DIJIE LIU^{1,2}, DOUGLAS OYERBEY¹, LISA D. WATKINSON^{1,3}, SAID DAIBES-FIGUEROA^{1,3}, TIMOTHY J. HOFFMAN^{1,4,5}, LEONARD R. FORTI^{1,2,3}, WYNN A. VOLKERT^{1,2,3} and MICHAEL F. GIBLIN^{1,2,3,*}

¹Research Service, Harry S. Truman Memorial Veterans' Administration Hospital, Columbia, MO 65201; ²Departments of ³Radiology, ⁴Medical Pharmacology and Physiology, and ⁵Internal Medicine, and ⁶The Radiopharmaceutical Sciences Institute, University of Missouri-Columbia, Columbia, MO 65211, U.S.A.

Abstract. Background: Uroguanylin is an endogenous peptide agonist that binds to the guanylate cyclase C receptor (GC-C). GC-C is overexpressed in human colorectal cancer (CRC), and exposure of GC-C-expressing cells to GC-C agonists results in cell cycle arrest and/or apoptosis, highlighting the therapeutic potential of such compounds. This study describes the first use of radiolabeled uroguanylin analogs for in vivo detection of CRC. Materials and Methods: The peptides uroguanylin and E³-uroguanylin were N-terminally labeled with ¹¹¹In and used for in vivo biodistribution and SPECT imaging studies. In vivo experiments were carried out using SCID mice bearing T84 human colorectal cancer tumor xenografts. Results: Alteration of the position 3 aspartate residue to glutamate resulted in increased affinity for GC-C, with IC₅₀ values of 5.0±0.3 and 9.6±2.9 nM for E³-uroguanylin and DOTA-E³-uroguanylin, respectively. In vivo, ¹¹¹In-DOTA-E³-uroguanylin demonstrated tumor uptake of 1.17±0.23 and 0.61±0.07% ID/g at 1 and 4 h post injection, respectively. The specificity of tumor localization was demonstrated by co-injection of 3 mg/kg unlabeled E³-uroguanylin, which reduced tumor uptake by 69%. Uptake in kidney, however, was dramatically higher for the uroguanylin peptides than for previously characterized radiolabeled E³-cell heat-stable enterostatin (STE) analogs targeting GC-C, and was also

inhibited by coinjection of unlabeled peptide in a fashion not previously observed. Conclusion: Use of uroguanylin-targeting vectors for in vivo imaging of colorectal cancers expressing GC-C resulted in tumor uptake that paralleled that of higher affinity heat-stable enterostatin peptides, but also resulted in increased kidney uptake in vivo.

Guanylate cyclase C (GC-C) is a type 1 transmembrane glycoprotein expressed on brush border membranes of intestinal epithelial cells, as well as on transformed human colon cancer cell lines such as the T-84 cell line (1, 2). In the normal intestinal mucosa, GC-C receptors are expressed within the apical (luminal) face of epithelial cell membranes, and are therefore isolated from the bloodstream by cell-cell tight junctions (3-6). GC-C expression is maintained in transformed cells throughout the process of colorectal carcinogenesis, while expression of the endogenous GC-C ligands guanylin and uroguanylin is typically lost (7-9). Normally expressed at high levels within the lumen of the gut, GC-C is expressed on virtually all histologically confirmed primary and metastatic colorectal tumors examined in human patients, while normal tissues and other types of cancer express minimal or no GC-C receptors (4-6). GC-C receptors on colorectal tumors retain their ligand-binding capacity, and expression of GC-C receptors does not vary as a function of metastatic site or grade of these tumors (5). The unique expression of GC-C by metastatic cells of colorectal origin within lymph nodes of patients undergoing staging for colorectal cancer (CRC) forms the basis for a PCR-based diagnostic test that is currently undergoing clinical trials (10). GC-C expression has also formed the basis of development of ligand-based molecular agents for in vivo detection and therapy of colorectal cancer (9, 11-20). Uroguanylin is a 16 amino acid peptide with two disulfide bonds, and has nanomolar affinity for the GC-C receptor (21, 22). Secretion of the endogenous peptides guanylin and uroguanylin into the lumen of the gut by enterochromaffin cells plays a role in regulation of ion and fluid homeostasis

Correspondence to: Dr. Michael F. Giblin, Harry S. Truman Memorial VA Hospital, Research Service Room 4024, 803 Hospital Drive, Columbia, MO 65201, U.S.A. Tel: +1 5738146000 ext. 53669, Fax: +1 5738821693, e-mail: giblinm@health.missouri.edu

Key Words: Uroguanylin, E³-cell heat-stable enterostatin, guanylyl cyclase C, single photon-emitting computed tomography (SPECT), colorectal cancer, in vivo imaging.

0250-7005/2009 \$2.00+40

3777

Bausch Health Ireland Exhibit 2046, Page 1 of 7
Mylan v. Bausch Health Ireland - IPR2022-00722

Table I. Calculated and observed (M+H)⁺ values and IC₅₀ values (± SD) for characterized peptides.

Peptide	(M+H) ⁺ Calc.	(M+H) ⁺ Obs.	IC ₅₀ (nM)
Uroguanylin	1667.6	1667.7	39.8±14.9
DOTA-uroguanylin	2053.6	2053.9	34.5±3.3
E ³ -uroguanylin	1681.6	1681.6	5.0±0.3
DOTA-E ³ -uroguanylin	2067.6	2067.9	9.6±2.9

Ex. 2046 at 3779

Patent Owner's Resp. at 66; Sur-Reply at 19-20, 25; Ex. 2024 ¶¶ 241-242

Liu Did Not Use Wild-Type Human Uroguanylin

ANTICANCER RESEARCH 29: 3777-3784 (2009)

In Vivo Imaging of Human Colorectal Cancer Using Radiolabeled Analogs of the Uroguanylin Peptide Hormone

DIIE LIU^{1,2}, DOUGLAS OVERBEY¹, LISA D. WATKINSON^{1,2}, SAID DAIBES-FIGUEROA^{1,2}, TIMOTHY J. HOFFMAN^{1,2,3}, LEONARD R. FORTE^{1,2,3}, WYNN A. VOLKERI^{1,2,3} and MICHAEL F. GIBLIN^{1,2,3*}

¹Research Service, Harry S. Truman Memorial Veterans' Administration Hospital, Columbia, MO 63201

as a 0.05 N HCl solution. Wild-type human uroguanylin was obtained from the American Peptide Company, and E³-uroguanylin was kindly provided by Dr. Kunwar Shailubhai at Callisto Pharmaceuticals. All other reagents were purchased from Aldrich

Alteration of the position 3 aspartate residue to glutamate or glycine, while expression of the endogenous GC-C ligands guanylin and uroguanylin is typically lost (7-9).

Peptides were purified by RP-HPLC and characterized by MALDI-TOF MS and by a competitive displacement receptor binding assay utilizing T84 human colorectal cancer cells and ¹²⁵I-labeled F¹⁹-STh(1-19) (Table I, Figure 2).

Bausch Health Ireland Exhibit 2046, Page 1 of 7
Mylan v. Bausch Health Ireland - IPR2022-00722

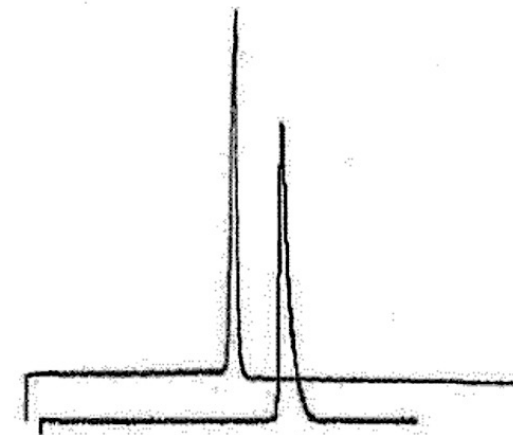




Figure 3. RP-HPLC chromatograms of purified ¹¹¹In-DOTA-E³-uroguanylin (top), ¹¹¹In-DOTA-uroguanylin (bottom).

Ex. 2046 at 3778-3780, Fig. 3
Sur-Reply at 19-20, 25; see also Patent Owner's Resp. at 66; Ex. 2024 ¶¶ 241-242

Patent Owner Did Not Use Wild-Type Human Uroguanylin

 **Beschwerdekammern
Boards of Appeal
Chambres de recours** European Patent Office
80299 MUNICH
GERMANY
Tel: +49 (0)89 2399-0
Fax: +49 (0)89 2399-4485


Adams, Harvey Vaughan John
Mathys & Squire LLP
120 Holborn
London
EC1N 2SQ
ROYAUME UNI

Datum/Date
11.06.13

Zeichen/Reference/Référence: 117839
Anmeldung Nr./Application No./Demande No./Requisito No./Requisito No.: 02721604.3 / 1379224
Anmelder/Applicant/Demandeur/Patenthaber/Proprietor/Titolare: Synergy Pharmaceuticals, Inc.

OPPO01

Appeal number:

Please find enclosed a copy of the decision dated 04.06.13.

The Registrar P. Cremona
Tel.: 089 / 2399 - 3341

Annex(es): Acknowledgement of receipt - EPO Form 2936

Registered letter with advice of delivery

EPO Form 3022 01/11 MYLAN EXHIBIT - 1067 Pg. 001
Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd.
IPR2022-00722

For completeness, we note that, because of the rapid isomeric interconversion of uroguanylin (and the much slower interconversion of E³-uroguanylin (SP-304)), it is impossible to compare the affinity of the active conformation (only) of uroguanylin to the affinity of the active conformation of E³-uroguanylin (SP-304). Testing of the activity or affinity of the active form (only) of uroguanylin is not possible, as it rapidly converts to, and eventually reaches equilibrium with, the inactive conformation.

The European Patent Office Did Not Credit Currie's Data



**BLAKE ROBERT
PETERSON , Ph.D**

Q. Okay. Are you aware that the European Patent Office did not credit Dr. Currie's results and declined to invalidate the plecanatide patent?

THE WITNESS: I believe that to be true, yes.

Federal Circuit: A Compound and All of Its Properties Are Inseparable



“From the standpoint of patent law, a compound and all of its properties are inseparable; they are one and the same thing.”

Federal Circuit: Permitting Reliance on Non-Prior Art



Permitting reliance on non-prior art as “evidence of the motivation of a POSITA to explore less frequent dosing regimens as of the priority date.”

Drs. Peterson and Epstein Are Not GCC Experts



**BLAKE ROBERT
PETERSON, Ph.D.**

Q. You have never been qualified by any court or by the U.S. Patent and Trademark Office as an expert in the biochemistry of GC-C receptors, correct?

A. That's correct.

Q. In connection with your work in this case, have you had any communications with Dr. Mark Currie?

A. No, I have not.

Q. Aside from your work on this case, have you ever had any communications with Dr. Mark Currie?

A. No, I have not.



**MICHAEL SAMUEL
EPSTEIN, M.D.**

Q. Do you consider yourself an expert in guanylin cyclase C receptors?

A. An expert, no.

Q. But you personally have never developed a GCC receptor agonist?

A. That is correct.

Dr. Peterson Has Never Conducted a T84 Cell Bioassay



**BLAKE ROBERT
PETERSON , Ph.D.**

Q. Have you yourself ever conducted a T84 cell bioassay?

THE WITNESS: I have conducted countless different whole cell assays. That particular specific assay I have not conducted in my laboratory, no.

Dr. Epstein's Lack of Expertise



**MICHAEL SAMUEL
EPSTEIN, M.D.**

- Q. Do you have a Ph.D. in chemistry?
A. **No, I do not.**
- Q. Do you have a master's in chemistry?
A. **No, I do not.**
- Q. Do you have a B.S. in chemistry?
A. **No.**
- Q. Do you have a Ph.D. in protein engineering?
A. **No, I do not.**
- Q. Do you have a master's in protein engineering?
A. **No, I do not.**
- Q. Do you have a B.S. in protein engineering?
A. **No, I do not.**
- Q. Do you have any degrees in pharmaceutical chemistry?
A. **No, I do not.**
- Q. Do you have any degrees in pharmacy?
A. **No, I do not.**
- Q. Do you have any degrees in clinical pharmacology?
A. **No, I do not.**

Dr. Epstein's Lack of Expertise in Statistics and Biostatistics



**BLAKE ROBERT
PETERSON, Ph.D.**

Q. You have never been qualified by any court or by the U.S. Patent and Trademark Office as an expert in statistics or biostatistics, correct?

THE WITNESS: I use statistics regularly in my research, but I'm not qualified by a court in an official certified sense, no.

Petitioner's Topoisomer "Reservoir" Argument Is Unavailing



BLAKE ROBERT
PETERSON, Ph.D.

Q. There is no reservoir of inactive topoisomer for a drug that does not exhibit topoisomeric interconversion, correct?

THE WITNESS: I mean if there's no interconversion, there is no reservoir. If that answers your question.

Petitioner's Reliance on Currie's Diarrhea Statements Are Misplaced



**BLAKE ROBERT
PETERSON, Ph.D.**

Q. Trulance is not approved for use in infants, correct?

THE WITNESS: I believe that's correct.

Q. Linzess is not approved for use in infants, correct?

THE WITNESS: I believe that's correct.

Q. Trulance is not approved for use in domestic animals, correct?

THE WITNESS: I'm not sure about the veterinary implications or indications of Trulance.

Q. Linzess is not approved for use in domestic animals, correct?

THE WITNESS: Again, I'm not sure if it's been approved for veterinary use.

HPLC Is Not Suitable for Manufacturing Scale



BLAKE ROBERT
PETERSON, Ph.D.

Q. High-performance liquid chromatography, or HPLC, is an analytical chemistry technique used to separate compounds in a chemical mixture, correct?

A. That is correct.

Q. HPLC can be used for research purposes, correct?

A. That is correct.

Q. HPLC can be used for manufacturing purposes, correct?

A. That is correct.

Q. What are some of the different considerations for using HPLC on a research scale versus using HPLC on a manufacturing scale?

THE WITNESS: HPLC is better suited to smaller scales, and so on a manufacturing scale, it might not be a cost-effective method.

Any Compound Is Toxic at High Levels



**MICHAEL SAMUEL
EPSTEIN, M.D.**

Q. Insulin is a peptide that the human body naturally produces; is that correct?

THE WITNESS: Yes.

Q. And it's possible to overdose on insulin; is that correct?

THE WITNESS: You may overdose on insulin.

Q. And an insulin overdose can lead to death; is that correct?

THE WITNESS: It may lead to hypoglycemia which, if left untreated, could be fatal.

Certain Drugs Can Slow Colonic Transit Times



MICHAEL SAMUEL
EPSTEIN, M.D.

THE WITNESS: I'm not sure exactly what you're asking. I'm a little bit lost in that. What are you asking? You mean something like that would cause the colon to slow down or something like that or...

Q. Correct.

A. Well, it could be longer if you take an antidiarrheal agent like Imodium or Lomotil that would slow the colon down. It's meant to do that. So it might slow down the transit through the colon.

Q. And opioids are another example of that would delay the transit time?

A. Those are actually opioids. So yes --

Q. Perfect.

A. -- opioids would do that.

“Medicine Is an Art, Not a Science”




MICHAEL SAMUEL
EPSTEIN, M.D.

Q. And how frequently do constipated patients have bowel movements?

A. That's actually an interesting question. I have patients who have a bowel movement every day and say they're constipated, and I have patients that have a bowel movement less than once a week and say they're constipated.

There's so many factors that goes into that, whether they're obstipated, whether they have tenesmus or, you know, pressure in the rectum, urgency or straining or incomplete evacuation or crampy discomfort. It really is very variable. And sometimes what we doctors think is not what the patient thinks. So it's more a matter of what does the patient feel, not so much what we think. That's why medicine is an art, not a science.

Claim 1 of the '786 Patent


 US007041786B2

(12) **United States Patent** (10) **Patent No.:** US 7,041,786 B2
Shailubhai et al. (45) **Date of Patent:** May 9, 2006

(54) **GUANYLATE CYCLASE RECEPTOR AGONISTS FOR THE TREATMENT OF TISSUE INFLAMMATION AND CARCINOGENESIS**

(75) **Inventors:** **Kunwar Shailubhai**, Ilaco, Ill., PA (US); **Gregory Nikiforovich**, St. Louis, MO (US); **Gary S. Jacob**, Creve Coeur, MO (US)

(73) **Assignee:** **Cellite Pharmaceuticals**, New York, NY (US)

(*) **Notice:** Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 362 days.

(21) **Appl. No.:** 10/107,814

(22) **Filed:** Mar. 28, 2002

(65) **Prior Publication Data**
 US 2003/0073628 A1 Apr. 17, 2003

Related U.S. Application Data

(60) Provisional application No. 60/348,646, filed on Jan. 17, 2002.

(51) **Int. Cl.** **A61K 38/12** (2006.01)

(52) **U.S. Cl.** **530/317; 530/300; 530/326; 514/10; 514/13**

(58) **Field of Classification Search** **530/317; 530/300; 326; 514/10; 13**
 See application file for complete search history.

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(Continued)

Primary Examiner—Stephen L. Rawlings
(74) Attorney, Agent, or Firm—Mintz, Levin, Cohn, Ferris, Glusky and Pappas, P.C.; Ivor R. Hitt

(57) **ABSTRACT**

A method of treatment of inflamed, pre-cancerous or cancerous tissue or polyps in a mammalian subject is disclosed. The treatment involves administration of a composition of at least one peptide agonist of a guanylate cyclase receptor and/or other small molecules that enhance intracellular production of cGMP. The at least one peptide agonist of a guanylate cyclase receptor may be administered either alone or in combination with an inhibitor of cGMP-dependent phosphodiesterase. The inhibitor may be a small molecule, peptide, protein or other compound that inhibits the degradation of cGMP. Without requiring a particular mechanism of action, this treatment may restore a healthy balance between proliferation and apoptosis in the subject's population of epithelial cells, and also suppress carcinogenesis. Thus, the method may be used to treat, inter alia, inflammation, including gastrointestinal inflammatory disorders, general organ inflammation and asthma, and carcinogenesis of the lung, gastrointestinal tract, bladder, testis, prostate and pancreas, or polyps.

6 Claims, No Drawings

1. A peptide consisting of the amino acid sequence of SEQ ID NO:20.

<400> SEQUENCE: 20

Asn Asp Glu Cys Glu Leu Cys Val Asn Val Ala Cys Thr Gly Cys Leu
 1 5 10 15

Ex. 1001 at claim 1; 35-36.
 Patent Owner's Resp. at 1

Evans: Isomer Activity Is Unpredictable

Clin Rheumatol (2001) (Suppl 1):S9-S14
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Clinical Rheumatology

Comparative Pharmacology of S(+)-Ibuprofen and (RS)-Ibuprofen

A. M. Evans
School of Pharmacy and Medical Sciences, University of South Australia, Adelaide, South Australia

Abstract: Racemic ibuprofen, which contains equal quantities of R(-)-ibuprofen and S(+)-ibuprofen, has been used as an anti-inflammatory and analgesic agent for over 30 years. Although the S(+)-enantiomer is capable of inhibiting cyclooxygenase (COX) at clinically relevant concentrations, R(-)-ibuprofen is not a COX inhibitor. The two enantiomers of ibuprofen are therefore different in terms of their pharmacological properties and may be regarded as two different 'drugs'. They also differ in terms of their metabolic profiles. For example, R(-)-ibuprofen becomes involved in pathways of lipid metabolism and is incorporated into triglycerides along with endogenous fatty acids. S(+)-ibuprofen does not appear to become involved in these unusual metabolic reactions, which is why S(+)-ibuprofen is regarded as being metabolically 'cleaner' than racemic ibuprofen. When racemic ibuprofen is given to humans, a substantial fraction of the dose of R(-)-ibuprofen (50%–60%) undergoes 'metabolic inversion' to yield S(+)-ibuprofen. On this basis, it has been argued that to obtain clinical effects that are comparable to those of a given dose of racemic ibuprofen, the dose of S(+)-ibuprofen would need to be about 75% of the dose of the racemate. However, this 'pharmacokinetic' rationale does not take into account the fact that inversion is not instantaneous, that there is variability in the extent of inversion between individuals, and that the kinetics of inversion may differ depending on the dosing situation. For example, the extent of inversion appears to be reduced when the racemate is given to patients experiencing acute pain. Recent studies have demonstrated that the clinical benefits of racemic ibuprofen can be derived from the administration of the single

S(+)-enantiomer at a dose that is half that of the racemate. For example, 200 mg of S(+)-ibuprofen has been found to be superior or at least equivalent to 400 mg of the racemate in the relief of dental pain. Possible explanations for this higher than expected efficacy of S(+)-ibuprofen are considered.

Keywords: Chirality; Cyclooxygenase; Enantiomers; Ibuprofen; Non-Steroidal Anti-Inflammatory Drugs; Pharmacokinetics

Introduction

If an object is symmetrical, then the mirror image of that object is spatially identical to the original. This is not the case for an asymmetrical object (one that cannot be divided into two identical halves). Try placing your right-hand into a left-handed glove, and you will immediately understand the importance of asymmetry in everyday life. Handedness, or chirality, also exists in the structure of organic molecules – usually in the form of a tetrahedral carbon atom covalently linked to four different substituents. A molecule containing one chiral carbon atom can exist as two non-superimposable mirror-image forms, or enantiomers. As the number of chiral carbon atoms within a molecule increases, so too does the number of stereoisomers. About 50% of all medicinal drugs used by humans contain an element of chirality within their chemical structure, and can therefore exist as two or more stereoisomers. About half of these (about 25% of all drugs) are actually used as mixtures of these stereoisomers – that is, as racemic mixtures [1]. An example of such a drug is Ibuprofen, which contains a single chiral carbon atom within its propionic acid side chain (Fig. 1). The two individual enantiomers of the molecule are referred to as R(-)-

Correspondence and offprints requests to: Associate Professor A.M. Evans, School of Pharmacy and Medical Sciences, University of South Australia, North Terrace, Adelaide 5000, South Australia. E-mail: alan.evans@unisa.edu.au

MYLAN EXHIBIT - 1066
Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd.
IPR2022-00722

[12–15]. However, the extent of chiral inversion varies between individuals – for example, in osteoarthritis patients being treated with racemic ibuprofen, the fractional inversion of R(-)-ibuprofen varied between 35% and 85% [13].

The second important feature that distinguishes R(-)-ibuprofen from S(+)-ibuprofen is its ability to interfere with normal lipid metabolism. In rats, the formation of

placebo. These results were not unexpected. However, 200 mg of S(+)-ibuprofen provided a more rapid onset of analgesic action than 400 mg of the racemate and provided better pain relief over the first 3 h after dosing.

HPLC Is Not Suitable for Manufacturing Scale



BLAKE ROBERT
PETERSON, Ph.D.

Q. High-performance liquid chromatography, or HPLC, is an analytical chemistry technique used to separate compounds in a chemical mixture, correct?

A. That is correct.

Q. HPLC can be used for research purposes, correct?

A. That is correct.

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Evans: Pure Preparations Are Preferred

Clin Rheumatol (2001) (Suppl 1):S9-S14
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Clinical Rheumatology

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A. M. Evans

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MYLAN EXHIBIT - 1066
Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd.
IPR2022-00722

In 1992, and again in 1996, I identified a range of potential advantages of administering profens, including ibuprofen, as enantiomerically pure preparations of the S-enantiomers [4,6]. These advantages included: reduced metabolic load, reduced chance of pharmacokinetic interactions with other drugs, avoidance of involvement in lipid metabolism and of the pharmacokinetic variability that arises from the metabolic inversion of R(-)-ibuprofen, and prevention of adverse events that may arise from the COX-independent actions of R(-)-ibuprofen. In addition, it was suggested that patient acceptability could be improved through the use of smaller doses. At that time, potential interactions

Hamra 1996: Opossum Uroguanylin Is More Potent than Human Uroguanylin

Opossum colonic mucosa contains uroguanylin and guanylin peptides

F. KENT HAMRA, WILLIAM J. KRAUSE, SAMMY L. EBER, RONALD H. FREEMAN, CHRISTINE E. SMITH, MARK G. CURRIE, AND LEONARD R. FORTE
The Truman Veterans Affairs Medical Center and Departments of Pharmacology, Anatomy, and Physiology, School of Medicine, Missouri University, Columbia 65212; and Searle Research and Development, St. Louis, Missouri 63167

Hamra, F. Kent, William J. Krause, Sammy L. Eber, Ronald H. Freeman, Christine E. Smith, Mark G. Currie, and Leonard R. Forte. Opossum colonic mucosa contains uroguanylin and guanylin peptides. *Am. J. Physiol.* 270 (Gastrointest. Liver Physiol. 33): G708-G716, 1996. Uroguanylin and guanylin are structurally related acetides that

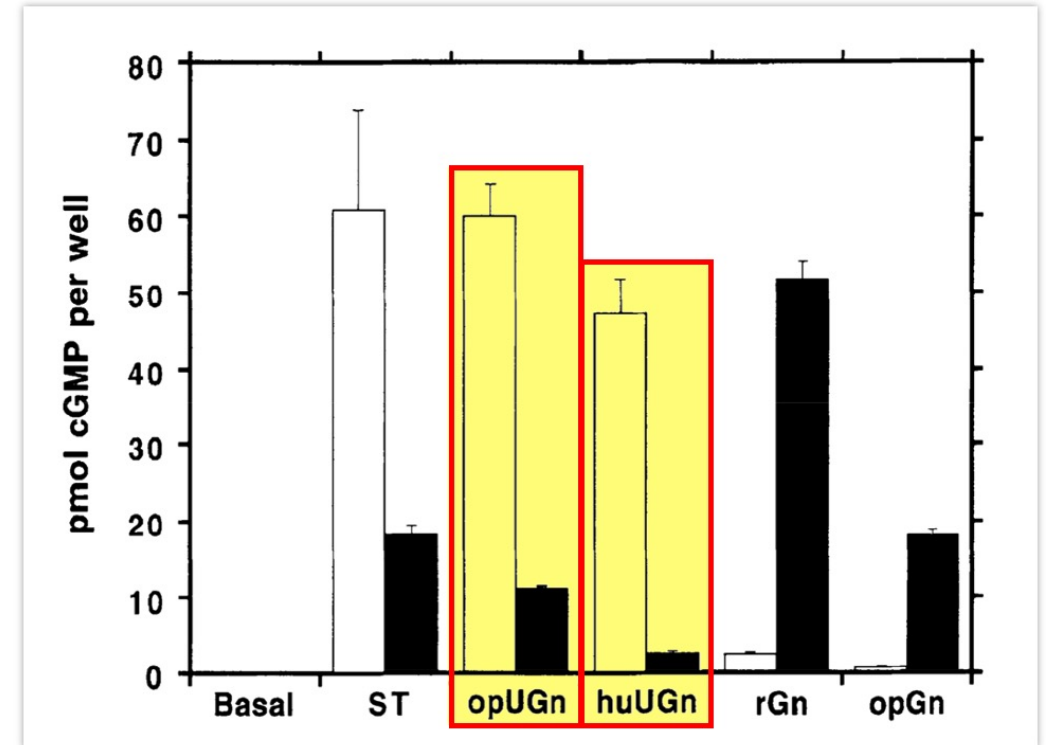
Fig. 2. Agonist-stimulated cGMP accumulation in T84 cells at pH 5.5 (open bars) and pH 8.0 (solid bars). Peptides and vehicle were suspended in HEPES and Dulbecco's modified Eagle's medium (DMEM) containing 50 mM sodium bicarbonate (pH 8.0), or 2-(*N*-morpholino)ethanesulfonic acid (MES) and DMEM at pH 5.5 (pH 5.5) for analysis in the T84 cell cGMP accumulation bioassay. Basal, vehicle control; ST, synthetic *E. coli* ST-(5-17); opGn, synthetic opossum guanylin; opUGn, synthetic opossum uroguanylin; huUGn, synthetic human uroguanylin; rGn, synthetic rat guanylin-(101-115). All peptides were tested at 30 nM except for *E. coli* ST-(5-17), which was tested at 3 nM. Error bars indicate standard error of the mean for 3 experiments.

intestine (21, 22). The opossum kidney also expresses high levels of GC-C-like receptors located in the apical membranes of proximal tubular cells (14). Guanylin was first isolated from rat jejunum as a heat-stable, 15-amino acid peptide that activated GC-C in human intestinal T84 cells (7). Guanylin cDNAs encoding 115- to 116-amino acid precursors have been isolated from rat, human, and mouse intestine (19). Uroguanylin was initially purified as 15- to 15-amino acid peptides from urine of opossums, humans, and rats could be derived from the kidney and/or from other tissues via filtration of uroguanylin from the circulation. In the current study, we have isolated uroguanylin and guanylin peptides from the colonic mucosa of opossums. Several independent analytical techniques were used to identify the bioactive peptides. Thus intestinal mucosa is a potential tissue source for uroguanylin and guanylin found in urine (10, 15, 20).

G708

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 Mylan Pharmaceuticals, Inc. v. Bausch Health Ireland, Ltd.



Ex. 1019 at G710 (Fig. 2)
 Patent Owner's Resp. at 48