

#### (12) United States Patent

Papadopoulos et al.

#### (54) MODIFIED CHIMERIC POLYPEPTIDES WITH IMPROVED PHARMACOKINETIC PROPERTIES AND METHODS OF USING THEREOF

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U.S.C. 154(b) by 489 days.

This patent is subject to a terminal disclaimer.

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- (60) Provisional application No. 60/138,133, filed on Jun. 8, 1999.
- (51) Int. Cl. A61K 38/18 (2006.01)C07K 14/71 (2006.01)C12N 15/62 (2006.01)
- (52) **U.S. Cl.** ...... **424/134.1**; 424/192.1; 514/2; 514/23; 530/350; 536/23.4
- (58) Field of Classification Search ...... None See application file for complete search history.

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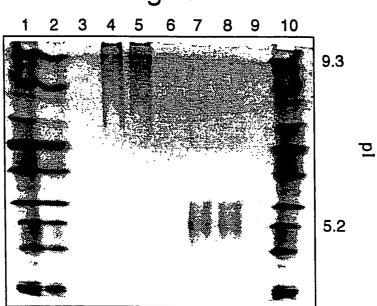
Primary Examiner—Christine J Saoud Assistant Examiner—Jon M Lockard (74) Attorney, Agent, or Firm-Valeta Gregg, Esq.

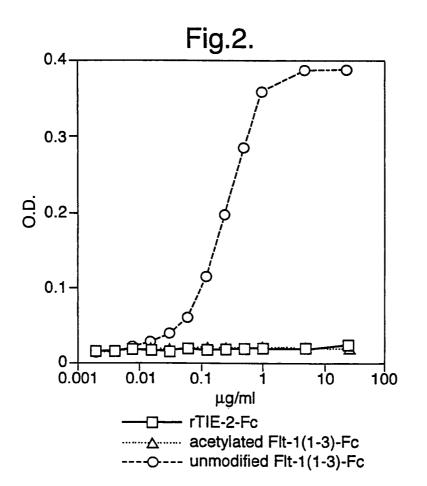
#### (57)**ABSTRACT**

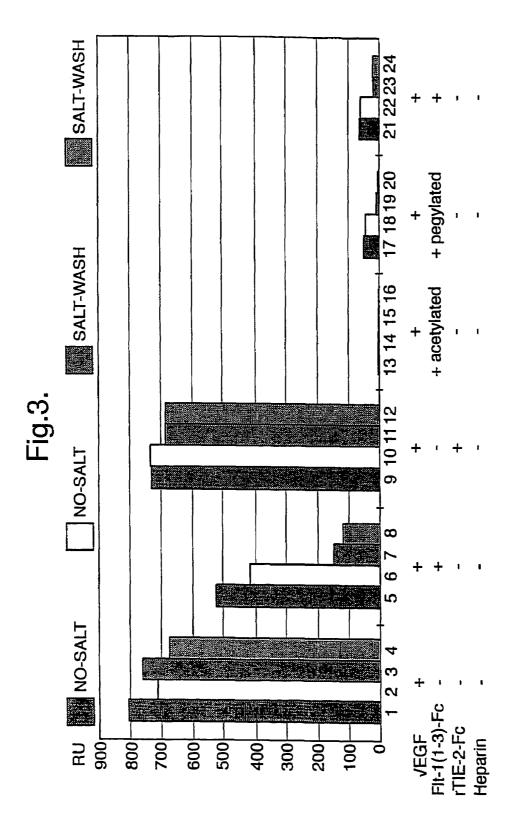
Modified chimeric polypeptides with improved pharmacokinetics are disclosed. Specifically, modified chimeric Flt1 receptor polypeptides that have been modified in such a way as to improve their pharmacokinetic profile are disclosed. Also disclosed are methods of making and using the modified polypeptides including but not limited to using the modified polypeptides to decrease or inhibit plasma leakage and/or vascular permeability in a mammal.

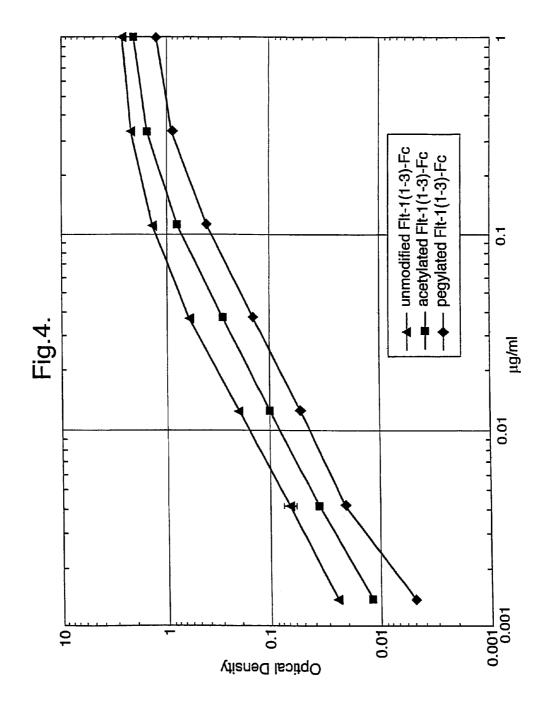
#### 3 Claims, 55 Drawing Sheets

Fig.1.









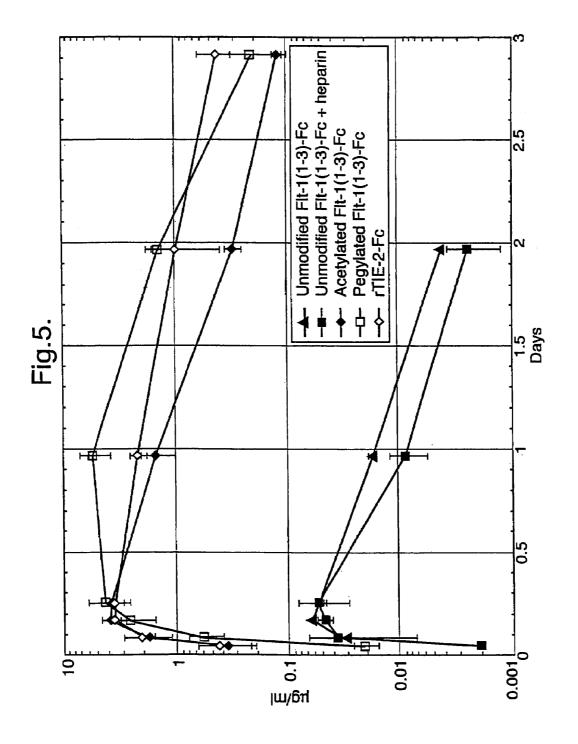


Fig.6A.

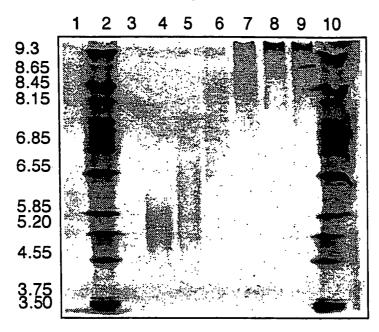
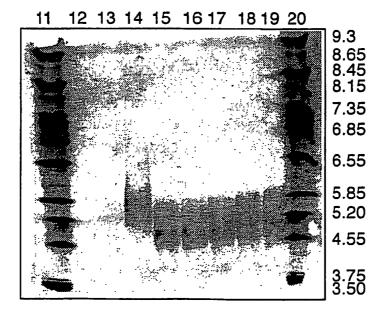
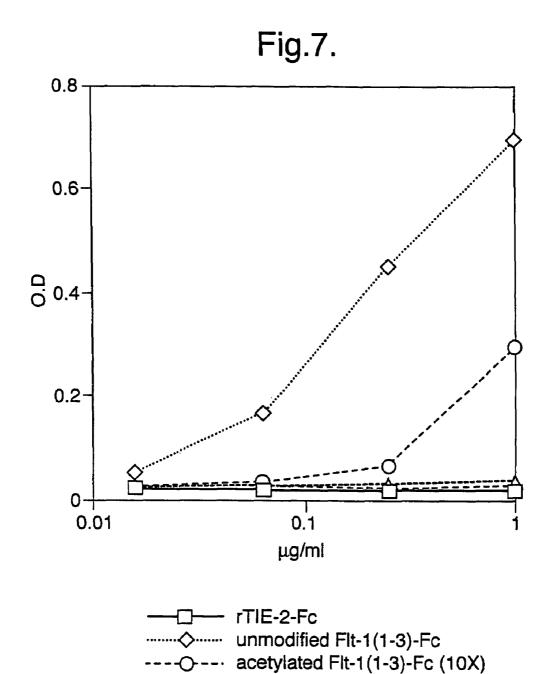


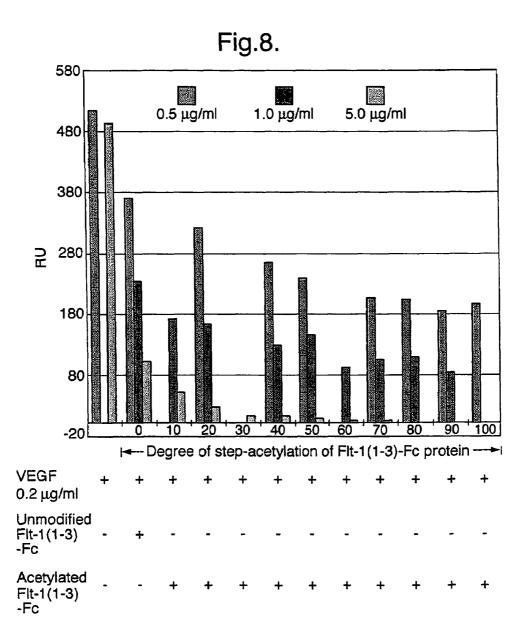
Fig.6B.

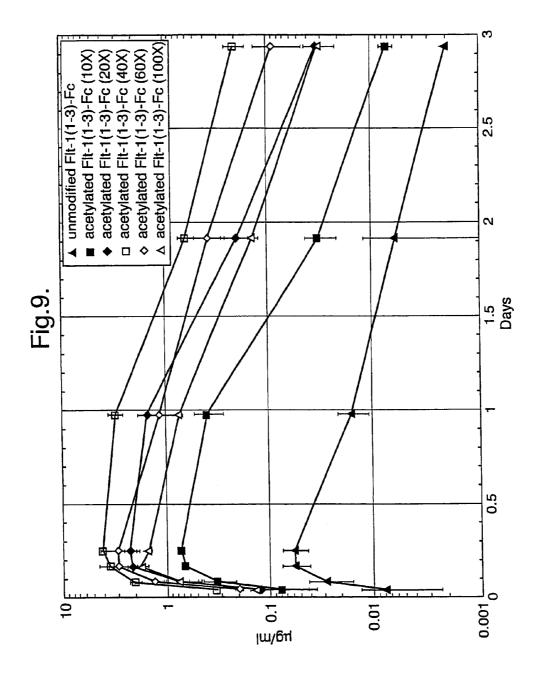




--- acetylated Flt-1(1-3)-Fc (20X)

--- = acetylated Flt-1(1-3)-Fc (30X)





# Fig.10A.

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### Fig.10B.

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# Fig.10C.

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May 20, 2008

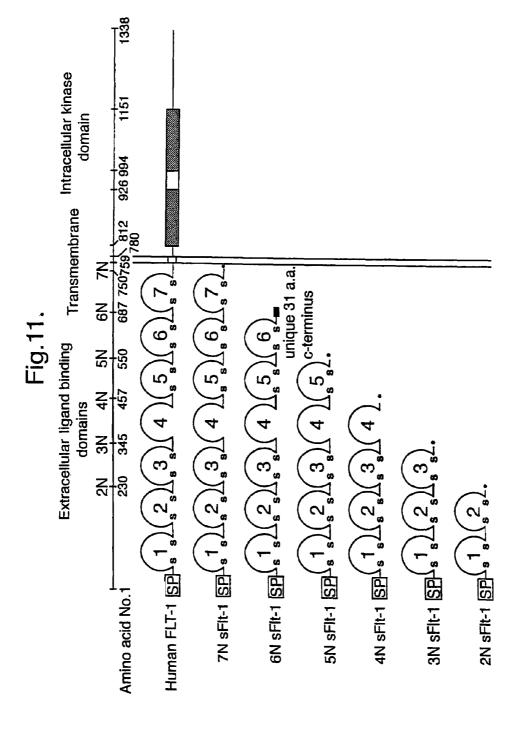
### Fig. 10D.

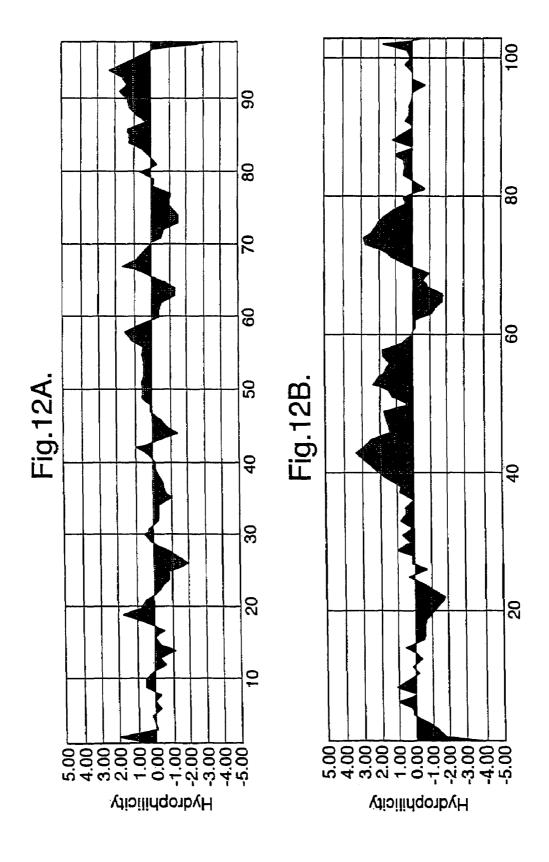
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### Fig.13A.

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		3	10.			320			330	ı		3	40			350			360
	*		*	*		*		*	•		*		*	*		*		•	*
CAC	ACT	GGC	TTC	TAC	AGC	TGC	: AAA	TAT	CTA	GCT	GTA	. cci	ACT	TCA	AAG	AAG	AAG	GAA	ACA
GTG	TGA	co	AAC	ATC	TCG	ACC	TTT	ATA	GAT	CGA	CAT	GGP	TGA	AGT	TTC	TIC	TIC	CII	TGT
His	Thr	: Gl <sub>3</sub>	Phe	Tyr	Ser	Cys	Lys	Tyr	Leu	ı Ala	Val	Pro	Thr	Ser	Lys	Lys	Lys	Glu	Thr>
		3	70			380			390	)		4	100			410			420
	*		*	4	•	*		*	•	+	*		*	*		*			*
																			AGT
																			TCA
Glu	ı Ser	: Ala	a Ile	Tyr	: Ile	: Phe	e Ile	Ser	Ası	Thi	. G12	Arg	y Pro	Phe	· Va.	. GII	1 Wei	' IYI	Ser>
									450							470			400
	_	4	130	_	_	440		_	450	,		4	160			470			480
~~			. ~			-		- -	י ייייייייייייייייייייייייייייייייייי	. ~~				. cmc	י אוראלי	, CC	• 110-1	. ~~	
																			GTT CAA
																			Val>
910	. 116		. 611		- 116	* ****	, 1 <sup>1</sup> 151,		311	1	ייייי	, 311		. 401			1		,
			190			500			510	נ			520			530			540
	*	7	*			*		*		k	*	•	*	•				*	*
ACC	TC	A CC	r aac	TA :	ACT	GT	r aci	TT	LAA!	A AAC	TT	r cci	A CT	CAD 1	AC'	r TI	TA E	cc	TAD 1
																			A CTA
																			q <b>zA</b> o

# Fig.13B.

		55	50		5	60			570			58	10		5	90			600
	*		*	*		*		*	*		*		*	*		•		*	•
GGA	AAA	CGC	ATA	ATC	TGG	GAC	agt	AGA	AAG	GGC	TTC	ATC	ATA	TCA	TAA	GCA	<b>DOA</b>	TAC	AAA
CCT	727.	GCG N	TAT	TAG	ACC	CIG	TCA	TCT	TTC	CCC	AAG	TAG	TAT	agt	TTA	CGT	TCC	atg	TTT
Gly	пåв	wrâ	me	TTE	rxb	qzA	Ser	Arg	Lys	GJA	Phe	Ile	Ile	Ser	Asn	Ala	Thr	Tyr	Lys>
		6:	LD.		,	20			<b>630</b>			-			,	^			
	*	٠.	*	*	•	*		*	630		*	64	*		•	550 *			660
GAA	ATA	GGG	CTT	CTG	ACC	TGT	GAA	GCA	ACA	GTC	AAT	GGG	ሮጀጥ	2345	ጥልጥ		202	אאר	ጥልጥ
CIT	TAT	$\alpha$	GAA	GAC	TGG	ACA	CIT	CGT	TGT	CAG	TTA	ccc	GTA	AAC	ATA	JIL	ılæı.	المالات حصت	ATA
Glu	Ile	$Gl_Y$	Leu	Leu	Mr	Cys	Glu	Ala	Thr	Val	Asn	Gly	His	Leu	Tvr	Lvs	Thr	Asn	Tvr>
												_				•			-2-
	_	6'	70		(	580			690			7(	00		•	710			720
CIT/C	202	~~	-			*		<b>k</b>	*		*		*	*		*		*	*
CIC	W.A.	CAT	COR	CAA	ACC	TAA	ACA	ATC	ATA	CAT	GTC	CAA	ATA	AGC	ACA	CCA	CGC	CCA	GIC
Ten	Thr	Hic	GCI	Cla	TGG	TTA	TGT	TAG	TAT	CTA	CAG	GIT	TAT	TCG	TGT	GGT	GCC	GGT	CAG
			мy	GIII	THE	ASN	THE	TTE	TTE	ASP	Val	GIN	Ile	Ser	Thr	Pro	Arg	Pro	Val>
		7:	30			740			750			7	50			770			780
	*		*	*		*		*	*		*	,,	*	*		*		*	*
AAA	TTA	CTT	AGA	GGC	CAT	ACT	CIT	GTC	CTC	AAT	TGT	ACT	GCT	ACC	ACT	ccc	TTG	AAC	ACG
TTT	AAT	GAA	TCT	CCC	GTA	TGA	GAA	CAG	GAG	TTA	ACA	TGA	CGA	TGG	TGA	GGG	AAC	TIG	TGC
Lys	Leu	Leu	Arg	Gly	His	Thr	Leu	Val	Leu	Asn	Cys	Thr	Ala	Thr	Thr	Pro	Leu	Asn	Thr>
		7.	90			800			810		_	8:	20			830			840
) ACIA	Catal	CAS	PUA:	y C.C.	m		, mr.c	~~~	*		*		*	*		*		*	*
TCT	CAA	راملت) محجر	ATC.	MCC.	TGG ACC	WEL	TAC	CCI	GAT	GAA	ATT	GAC	CAA	AGC	TAA	TCC	CAT	GCC	AAC
Arq	Val	Gln	Met	Thr	محدر	Ser	Three .	Dro	CIN	CIT	TAA	200	GIT	103	TTA	AGG	GIA	CGG	TTG Asn>
-							-1-		nop	GIU	-1-	rap	GIII	Ser	Asn	Ser	urs	WTG	ASII>
		8	50			860			870			8	в0			890			900
	*		*	*		*		*	*		*		*	*		*		*	*
ATA	IJC	TAC	AGT	GTT	CTT	ACT	ATT	GAC	AAA	ATG	CAG	AAC	AAA	GAC	AAA	GGA	CIT	TAT	ACT
TAT	AAG	ATG	TCA	CAA	GAA	TGA	TAA	CIG	TTT	TAC	GTC	TTG	TTT	CTG	TTT	CCT	GAA	ATA	TGA
TTE	rne	TYT	Ser	Val	Leu	Thr	Ile	Asp	Lys	Met	Gln	Asn	Lys	Asp	Lys	Cly	Leu	Tyr	Thr>
		9	10			920			620				40			050			
	*		*	*		*		•	930		*	9	40	*		950			960
TGT	CGT	GTA	AGG	AGT	GGA	CCA	TCA	TTC	AAA	TOT	GTT	AAC	ACC	ፈጋљ	CTIC	יים. ייים	מיתב	ጥይጥ	CPL
ACA	GCA	CAT	TCC	TCA	CCT	GGT	AGT	AAG	TTT	AGA	CAA	TTG	TGG	AGT	CAC	GTA	TAT	ATA	CTA
Cys	Arg	Val	Arg	Ser	Gly	Pro	Ser	Phe	Lys	Ser	Val	Asn	Thr	Ser	Val	His	Ile	Тут	Asp>
		9	70 *			980			990			10	00		1	010			1020
222	CCF -	CCC		-	C) C	~~~	***	*	*		*		*	*		*		*	*
Lalal	CCT	CCC	CCC	990	- CALC	000	AAA	TCT	TGT	GAC	AAA	ACT	CAC	ACA	TGC	CCA	CCG	TGC	CCA
Lys	Ala	Glv	Pro	Glv	Glu	Pro	Tare	Ser	מנים.	) CIG	Tim	TUR	ere.	1G1	ACG	GGT	GGC	ACG	GGT Pro>
-				3		-20	~20		-ys	asp	-ys	1115	nıs	INI	Cys	rro	PTO	Cys	LLO>
		10	30		1	040			1050			10	60		1	070			1080
	*		*	*		*		*	*		*		.*	*		*		*	*
GCA	CCT	GAA	CIC	CTG	GGG	GGA	CCC	TCA	GIC	TIC	CTC	TTC	000	CCA	AAA	. 000	AAG	GAC	ACC
CCT	GGA	CIT	GAG	GAC	ccc	CCI	GCC	AGT	CAG	AAG	GAG	AAG	GGG	GGT	LIL	GGG	TIC	CTO	TGC
-WIG	FIO	GIU	ren	Leu	Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr>

# Fig.13C.

		_	ت	<i>,</i>															
		109	•	*	11	.00		*	110		*	112	0 *	•	11	30	,	. 1	140
CTC	ATG	<b>STA</b>	TCC	CGG	ACC	CCT	GAG	GTC	ACA	TGC	GTG	CTG	GTG	GAC	GTG	AGC	CAC	AAE	GAC
														CIG					
Leu	Met	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	Val	Asp	Val	Ser	His (	Glu	Asp>
		115	0		11	.60		:	1170			118	30		11	.90		1	200
	*		*	*		*		*	*		*		*	*		*		*	*
														CAT					
														GTA					
Pro	Glu	Val	Lys	Phe	Asn	Trp	TYI	Val	Asp	GIĀ	Val	Glu	Val	His	ASN	ATA	гĀ2	ınr	ràe>
		12:	10		13	220			1230			124	40		1.	250		1	L260
	*		*	*		*		*	*		*		*	*		*		*	*
														GTC					
GGC	GCC	CIC	CIC	GIC	ATG	TTG	TOG	TGC	ATG	GCA	CAC	CAG	TCG	CAG	GAG	TGG	CAG	GAC	GIG
Pro	Arg	Glu	Glu	Gln	Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	Leu	Thr	Val	ren	His>
		12	70		13	280			1290			13	00	_	1	310		:	1320
CNC	~	m~~	~~~	* * *	000	*	CNC	m>-	* ~~~	mee	220	CIIV.	_ 	AAC	222	-	حىلات	~~~	
														TTG					
																			Ala>
U.M.	, up		DCu		GLJ	2,5	010	-1-		0,0	_,_								
		13	30		1	340		_	1350		_	13	60		1	370			1380
	*		*	*		*		*	*		*	. ~~	*	~~~	~~	C)C	CALAC	400 C	יים או
														GAA					
5000	TAG	CIU	TIT	Ties	TAG	COL	Tare	יומ	y Taro	Gla	. GIV	Dr.	. Am	CII	Pro	Glo	Val	Tvr	Thr>
PLU	116	GIU	nys	. 1111	TIC	. SEL	. Lys	, Ale	ı Dya	GLJ	GIL			GIG			·	-2-	
		13	90		1	400			1410			14	20		1	430		_	1440
	*		*	*		*		*	*		*		*	*				×	
																			AAA
														GAC					Lys>
reu	PIO	PIC	ser	Arg	ASE	GIU	L	. 111	Luys	, ASI	ı Gıı	ı vaı	. 561	. Leu		. cys			
		14	50		1	460			1470	)		14	80		1	490			1500
	*		ŧ	*		*		*	*	r	*		*	*		*		*	*
																			AAC
																			TTG
GIY	Pne	Tyr	Pro	ser .	AST	116	: Ale	ı va.	I GIL	1 121	) GI	ı ser	ASI	ı Gış	GII	l PIO	GIU	. ASI	Asn>
		15	10		1	520		_	1530	)		15	40	_		.550			1560
m> 0	*		*	*		*		*	, , , , , ,				· mmc	*		, mac	700	220	. מער
																			GAG
																			Leu>
171	Lys	***			, ,,,	, ,					, 01,								
	*	15	70 *	*	. 1	.580 *		*	1590	) •	•	16	*	•	. 1	.610		*	1620
ACC	GTG	GAC	AAG	AGC	AGG	TGG	CAC	CAC	GGG	AA C	GT	TIC	TO	A TGC	TCC	GTG	ATG	CAT	C GAG
TGG	CAC	CTG	TTC	TCG	TCC	: ACC	GT	GIV	c ccc	TI	CAC	AAC	AG7	DOA 1	AGC	CAC	TAC	GT	A CTC
Thr	Val	Asp	Lys	Ser	Arg	Tr	Gli	Gl	n Gly	Ası	ı Va	l Phe	e Sei	: Cys	s Ser	. Val	Met	His	s Glu>

U.S. Patent

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### Fig.13D.

1670 1630 1640 1650 1660 GCT CTG CAC AAC CAC TAC ACG CAG AAG AGC CTC TCC CTG TCT CCG GGT AAA TGA CGA GAC GTG TTG GTG ATG TGC GTC TTC TCG GAG AGG GAC AGA GGC CCA TTT ACT Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys \*\*\*>

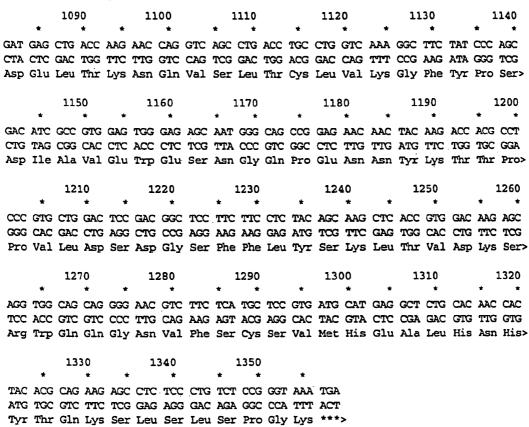
### Fig.14A.

			• •	9.	1 6	, · · ·													
		1	.0			20			30			4	10			50			60
	*		*	*		*		*	*		*		*	*		*		•	*
														CIC					
														GAG					
Met	Val	Ser	Tyr	Trp	Asp	Thr	Gly	Val	Leu	Leu	CA2	Ala	Leu	Leu	Ser	Cys	Leu	Leu	Leu>
		_																	
		7	0			80			90			10	00		1	.10			120
			-			*		*	*				*			*		•	
														AGT					
														TCA					
1111	GIĀ	Ser	Ser	Ser	GIA	GIĀ	AIG	PTO	Pne	AST	GIU	met	ıyı	Ser	GIU	TTE	PTO	GIU	TIE>
		1.	<b>.</b> n			40			150			4.			-	70			3.00
		13	*		-	140			150			T.	50		2	L70			180
ATA	רשר	באדע	ب حداد	CDP	CCA	<b>y</b> GG	CAC	<u>ст</u> .	CU.	y uan	CCC	WGC.		GTT	y.~~	ער_צ	<b>~</b> ™	220	בייים
														CAA					
														Val					
	*112	TIE C	1111	GIU	GTA	w	GIU	Trea	var	TTE	FLO	Cys	Arg	Val	THE	SeT	PLU	Yan	1167
		1	90		•	200			210			2	20			230			240
	*		*	*	•	*		*	*		*	_	*	*	•	*		*	*
ACT	GIT	ACT	TTA	AAA	AAG	TTT	CCA	CTT	GAC	ACT	TTG	ATC	CCT	GAT	GGA	AAA	CGC	ATA	ATC
														CTA					
		_										_						_	Ile>
				•	•				•					_		•	_		
		2	50		:	260			270			2	80			290			300
	*		*	*		*		*	*		*		*	*		*		*	*
TGG	GAC	AGT	AGA	AAG	GGC	TTC	ATC	ATA	TCA	AAT	GCA	ACG	TAC	AAA	GAA	ATA	GGG	CTT	CTG
ACC	CIG	TCA	TCT	TTC	CCG	AAG	TAG	TAT	AGT	TTA	CGT	TGC	ATG	TTT	CTT	TAT	ccc	GAA	GAC
Trp	Asp	Ser	Arg	Lys	Gly	Phe	Ile	Ile	Ser	Asn	Ala	Thr	Tyr	Lys	Glu	Ile	Gly	Leu	Leu>
		3:	10			320			330			3	40			350			360
	*		*	*		*		*	*		*		*	*		*		*	*
				٠.										TAT					
														ATA					
1111	Cys	GIU	AIA	Thr	VAI	Asn	GIĀ	HIS	Leu	TYT	гÃЕ	THE	Asn	ıyr	reu	THE	HIE	Arg	Gln>
		3'	70			380			390			٨	00			410			420
	*	٠	*	*	•	*		*	350		*	•	*	*	'	**		*	*
ACC	ААТ	ACA	ATC	ATA	GAT	GTC	CAA	ATA	AGC	ACA	CCA	CGC	CCA	GTC	AAA	מידיים	الملت	AGA	GGC
														CAG					
																			Gly>
					•													-	•
		4	30			440			450			4	60			470			480
	*		*	*		*		*	*		*		*	*		*		*	*
CAT	ACT	CTT	GTC	CTC	TAA	TGT	ACT	GCT	ACC	ACT	ccc	TTG	AAC	ACG	AGA	GTI	CAA	ATG	ACC
GTA	TGA	GAA	CAG	GAG	TTA	ACA	TGA	CGA	TGG	TGA	GGG	AAC	TTG	TGC	TCT	CAA	GTI	TAC	TGG
His	Thr	Leu	Val	Leu	Asn	Cys	Thr	Ala	Thr	Thr	Pro	Leu	Asn	Thr	Arg	Val	Gln	Met	Thr>
		4	90			500			510			5	20			530			540
m~~	*	m	*	*		*		*	*		*		*	*		*		*	*
	_													AAC					
																			CAA
ızb	ser	TAL	rro	ASP	GIU	TTE	ASP	GIN	ser	asn	ser	HIS	ALA	Asn	Ile	Phe	тут	ser	Val>

## Fig.14B.

		55	0		5	60			570			58	0		5	90			600
	*		*	*		*		<b>*</b> 1	*		*		*	*		*		<b>*</b> ·	*
													TAT						
													ATA						
Leu	Thr	Ile	Asp	Lys	Met	Gln	Asn	Lys	qaA	Lys	Gly	Leu	Tyr	Thr	Суѕ	Arg	Val	Arg	Ser>
		61	LO		6	20			630			64	lÓ		ŧ	550			660
	*		•	*		*		*	*		*		*	*	-	*		*	*
													TAT						
													ATA						
Gly	Pro	Ser	Phe	Lys	Ser	Val	Asn	Thr	Ser	Val	His	Ile	Tyr	Asp	Lys	Ala	Gly	Pro	Gly>
		6'	70			580			690			70	00			710			720
	*		*	. *		*		*	*		*	• •	*	*		•		*	*
GAG	ccc	AAA	TCT	TGT	GAC	AAA	ACT	CAC	ACA	TGC	CCA	ccc	TGC	CCA	GCA	CCT	GAA	CTC	CIG
													ACG						
Glu	Pro	Lys	Ser	Сув	Ązp	Lys	Thr	His	Thr	Cys	Pro	Pro	Cys	Pro	Ala	Pro	Glu	Leu	Leu>
		7	30	•		740			750			7.	- ^			770			780
	*.	′.	*	*		*		*	/3U		*	/(	50 *	*		*			, so
GGG	GGA	CCG	TCA	GTC	TTC	CTC	TTC	CCC	CCA	AAA	ccc	AAG	GAC	ACC	CTC	ATG	ATC	TCC	CGG
													CIG						
Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg>
		7	90			900			910							020			940
	*	<i>r</i> :	*	*		B00 *		*	810		•	5.	20	*	i	B30 *			840
ACC	CCT	GAG	GTC	ACA	TCC	GTG	GIG	GTG	GAC	CITC	AGC	CAC	GAA	GAC	رس	GAG	CIC	AAG	TTC
													CTT						
																			Phe>
												_							
		8:	50	_		860		_	870			В	30	_	;	890		_	900
ממ	TCC	mac.	~ _	~ ~ ~	~~~	~	CNC			220	~~		» ACA		~~~	~~	ana.		C) C
													TGT						
																			Gln>
	•	•										-2-		-3-					
	_	9:	10		:	920			930			9	40			950			960
mac.	220	300	,	m» c	~~	~~~	~m~	*	~~~	~~~		OTT-0	~~~	~~	<b>~</b>	~		~~~	880
													CTG						
																			Asn>
				-							•					-	-		
		9	70	_		980		_	990		_	10	00		1	010		_	1020
CCC	970	CNC	m> ~	200	m-c	*	~m~		220	888	-	ama.	~~~	~~~	~~~	* *	CNO	* * * * *	, CC
																			ACC TGG
																			Thr>
_	-		-	-	-	-				•		_	_					•	
		10:	30	_	1	040			1050			10	60		1	070			1080
3 m~	*		*	*	~~~	*	~~~	*	~		*	<b></b> -	•	*		*		*	*
																			ccc
																			GCC Arg>
-20	JEI	Lys	MIG	פעש	GIY	GIN	FIO	Αrg	GIU	FLO	GIN	AGT	TYE	THE	ren	PIO	210	Jei	wrg>

### Fig.14C.



# Fig.15A.

		1	10			20			30			4	0			50			60
	*		*	. *		*		*	*		*		*	*		*		*	•
4														CIC					
														GAG					
Met	Val	ser	Tyr	Trp	Asp	Thr	GIĀ	Val	Leu	Leu	Cys	Ala	Leu	Leu	Ser	Cys	Leu	Leu	ren>
			70			80			90			3.0	00		•	L10			120
			*	*		*		*	*		*	Δ,	*		•	*		•	+ -
ACA	GGA	TCT	AGT	TCC	GGA	CCT	AGA	ССТ	TTC	GT'A	GAG	ATG	TAC	AGT	GAA	ATC	ccc	GAA	ATT
														TCA					
														Ser					
					_	_													
		1	30		:	140			150			1	60			170			180
	. *		*	*		*		*	*		*		*	*		*		*	*
														GIT					
														CAA					
TTE	HIS	Met	Thr	Glu	GJĀ	Arg	Glu	Leu	Val	IIe	Pro	CAR	Arg	Val	Thr	ser	Pro	asn	Ile>
		4	.90			200			210			2	20			230			240
	*	_	*			*		*	210		•	-	+			*		*	*
ACT	GTT	ACT	TTA	AAA	AAG	TTT	CCA	CIT	GAC	ACT	TTG	ATC	CCI	GAT	GGA	AAA .	CGC	ATA	ATC
														CTA					
Thr	Val	Thr	Leu	Lys	Lys	Phe	Pro	Leu	Asp	Thr	Leu	Ile	Pro	Asp	Gly	Lys	Arg	Ile	: Ile>
		2	250			260			270			2	80			290		_	300
maa	*	~	*	*		*		* .			*		* .		-		~~		
														AAA :					
																	_		GAC Leu>
	ر س	,		, Lyc	·					, 1 NJ			,-	. <b>.</b>	, 01.				
		:	310			320			330	,		:	340			350			360
	*		*	1	t	*		*	*		*.		*	•	•	*		*	*
ACC	TGT	GA	A GCI	ACI	A GTO	: AA1	C GGG	CA	TTC	TAT	DAA 1	AC	AA A	TAT	CIC	AC	A CA!	r cc	A CAA
TGG	ACI	CT	rcg	TG	CAC	TT?	f CCC	GT	AAC	AT	TTC	TG	r TT	ETA E	CA(	G TG	CT	A GC	rgrr
Thr	CA	Gl	n Ala	Th	r Val	Ası	; GJ7	His	Let	ı Tyı	Lys	Th	r Ası	נעד ב	Lei	ı Thi	Hi	s Ar	g Gln>
			300			200			200				400			430			420
	*		370			380		*	390	,		•	400	,	ŧ	410			*
ACC	: AA	r ac	A ATY	C AT	A GA!	r GTY	CAZ	AT	A AGC	: AC	A CC	CG	c cc:	A GT	. AA	A TT.	A CT	T AG	A GGC
																			T COG
																			g Gly>
			430			440			450	)			460			470			480
	•		* 		* 	* 		*	,	•	*		*		* 	*		*	*
																			G ACC
																			C TGG t Thr>
****			- va	- 114	- A-	cy	~ 111						~ ~-	+*1	- ~1	y va	_ 51	_,	
			490			500			51	0			520			530			540
	*		*		*	*		*		*	*		*		*	*		*	* *
																			LA AGC
																			T TCG
Tr	se.	т Ту	r Pr	o As	p Gl	u Ly	's As	n Ly	s Ar	g Al	a Se	r Va	LI AT	g Ar	g Ar	g Il	e As	sp G	in Ser>

### Fig.15B.

		55	io		5	60			570			58	រេ			590			600
	*		*	*		*		*	•		*		*	*		*		*	*
TAA	TCC	CAT	GCC	AAC	ATA	TTC	TAC	agt	GIT	CIT	ACT	ATT	GAC	AAA	ATG	CAG	AAC	AAA	GAC
TTA	AGG	GTA	CCC	TTG	TAT	AAG	ATG	TCA	CAA	GAA	TGA	TAA	CIG	TTT	TAC	GTC	TTG	TTT	CTG
Asn	Ser	His	Ala	Asn	Ile	Phe	Tyr	Ser	Val	Leu	Thr	Ile	Asp	Lys	Met	Gln	Asn	Lys	y2b>
		6:	LO		ε	20			630			64	ın			650			660
	*		*	•		*		*	*		•	0,	*	*	`	*		*	*
AAA	GGA	CIT	TAT	ACT	TGT	CGT	GTA	AGG	AGT	GGA	CCA	TCA	TTC	AAA	TCT	GTT	AAC	ACC	TCA
TTT	CCT	GAA	ATA	TGA	ACA	GCA	CAT	TCC	TCA	CCT	GGI	agt	aag	TTT	AGA	CAA	TTG	TGG	AGT
Lys	Gly	Leu	Tyr	Thr	CAR	Arg	Val	Arg	Ser	Gly	Pro	Ser	Phe	Lys	Ser	Val	Asn	Thr	Ser>
		6'	70			580			690			7,	00			710			720
	*	·	*	*	`	*		*	*		*		*	*		710		*	720 *
GIG	CAT	ATA	TAT	GAT	AAA	GCA	GGC	CCG	GGC	GAG	ccc	AAA	TCT	TGT	GAC	AAA	ACT	CAC	ACA
CAC	GTA	TAT	ATA	CTA	TTT	CCT	CCC	GGC	CCCG	CIC	GGG	TTT	AGA	ACA	CIG	TTT	TGA	GTG	TGT
Val	His	Ile	Tyr	Asp	Lys	Ala	Gly	Pro	Gly	Glu	Pro	Lys	Ser	Сув	Asp	Lys	Thr	His	Thr>
		-	30			740			==0			_							
	*		*			740		*	750 *		*	7	60 *	*		770 *			780
TGC	CCA	CCG	TGC	CCA	GCA	CCT	GAA	CIC	CTG	GGG	GGA	CCG	TCA	GTC	TTC	CTC	TTC	CCC	CCA
ACG	GGT	GGC	ACG	GGT	CGT	GGA	CIT	GAG	GAC	ccc	CCT	GGC	AGT	CAG	AAG	GAG	AAG	GGG	GGT
Cys	Pro	Pro	Cys	Pro	Ala	Pro	Glu	Leu	Leu	Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro>
		-										_							
	*	,	90 *	*	,	800			810			8	20			830			B40
AAA	. ecc	AAG	GAC	ACC	CIC	ATG	ATC	TCC	CGG	ACC	CCT	വദാ	י. ה	ACA	ara-c	. Ga.c	מונים:	Calca -	ב
														TGT					
																			Asp>
		_																	
	*	8	50 *	*		860			870			8	80	_		890		_	900
GIG	AGC	CAC	GAA	GAC	CCT	GAG	GTC	AAG		חבה	. aksc	ጥልቦ	Callet *	GAC	GGC	, Gaz.	C TO	E CERC	ቸ የውጥ
CAC	TOG	GTG	CTT	CIG	GGA	CIC	CAG	TTC	AAG	TTG	ACC	ATG	CAC	CTG	CCG	CAC	CTC	CAC	GTA
																			His>
		_																	
	*	9	10			920			930		_	9	40			950		4	960
AAT	GCC	AAG	ACA	AAG	ccc	. C.C.C	GAG	. C.V.		ምል/	י אנה י	י איני	200	יי מחיי		יי כיוויכ	. C:00°	• »/2/	GTC
																			CAG
																			Val>
		_										_							
		9	70			980		•	990	l	_	10	00		. 1	1010		_	1020
CTC	ACC	GTC	CIG	CAC	CAG	GAC	are:	ت	- Taa:	. ממר	. 770	. GNG	- የመልር	ממי	n n	• • 330	. Call	· 4	AAC
GAG	TGG	CAG	GAC	GTG	GIC	CTG	ACC	GAC	TTA	. 000	TTC	CTC	ATY	TTC	ACC	י שישיר השיי	CAC	ACC	TTG
Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	Gly	Lys	Glu	Туз	Lys	Cys	Lys	Val	Se	Asn>
														_	-	-			
	*	10	30		1	.040			1050			10	60		. 1	1070		,	1080
ААА	GCC	CIC	CCA	GCC	CCC	ATY	GAC	נממ:	ייאב ו	ይጥ	. 44~-\ -	מממי	~~ ~	* ימתי	منحا	ב ריאים *	- ~~	. المارة *	A GAA
TTT	CCC	GAG	GGT	CGG	GGG	TAG	CTC	TT	TGG	TAC	AGG	Links Tours	CGC	dalai . www	ישטע	CAL	3 CCC	. cc	r CTT
Lys	Ala	Leu	Pro	Ala	Pro	Ile	Gli	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	/ Glr	n Pro	Ar	g Glu>

## Fig.15C.

	1090				11	.00		1	110		1120 1130							1140			
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CCA	CAG	GTG	TAC	ACC	خالت	ccc	CCA	ጥርር	സ്ത	ርልጥ	CAG	حكات	200	244	244	CAG	GTC	AGC	CTG		
																		TCG			
																			Leu>.		
														_,_							
		115	0		11	L60		3	170			118	30		13	190		1	.200		
	*		*	*		*		*	*		*		*	*		*		*	*		
ACC	TGC	CTG	GTC	AAA	GGC	TTC	TAT	CCC	AGC	GAC	ATC	GCC	GTG	GAG	TGG	GAG	AGC	TAA	GGG		
TGG	ACG	GAC	CAG	TTT	CCG	AAG	ATA	GGG	TCG	CIG	TAG	CGG	CAC	CTC	ACC	CTC	TCG	TTA	CCC		
Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser	Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly>		
	• •																				
		12:	70		1:	220			1230			12	40		1	250		1	L2'60		
	*		*	*		•		*	*		*		*	*		*		*	*		
CAG	CCG	GAG	AAC	AAC	TAC	AAG	ACC	ACG	CCT	ccc	GTG	CTG	GAC	TCC	GAC	GGC	TCC	TTC	TTC		
GTC	GGC	CIC	TTG	TTG	ATG	TTC	TGG	TGC	GGA	GGG	CAC	GAC	CIG	AGG	CTG	CCG	AGG	AAG	aag		
Gln	Pro	Glu	Asn	Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	qaA	Gly	Ser	Phe	Phe>		
		12	70		1	280		1290							1310			1320			
	*		*	*			*	*		*		*	*		*		*	*			
																		TCA			
GAG	ATG	TCG	TTC	GAG	TGG	CAC	CIG	TTC	TCG	TCC	ACC	GTC	GIC	ccc	TTG	CAG	AAG	TDA	ACG		
Leu	Tyr	Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp	Gln	Gln	G1y	Asn	Val	Phe	Ser	Cys>		
		13				340			1350			13	60		_	.370			1380		
	*		*			*		*	*		*		*	*		*		*	*		
																			CCC		
																			. GGC		
Ser	Val	Met	His	Glu	Ala	Leu	His	Asn	His	Tyr	Thr	Glr	Lys	Ser	Lev	Sex	Leu	Ser	Pro>		
~~-																					
		TGA																			
		ACT																			
GTZ	' Lys	***	>																		

# Fig.16A.

10						20			30			4	10			50	)				
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ATG (																					
TAC	CAG	TCG	ATG	ACC	CTG	TGG	CCC	CAG	GAC	GAC	ACG	CGC	GAC	GAG	TC	GA	A C	AC	GAA	فلاعا دده ۲	
Met '	Val	Ser	Tyr	TIP	Asp	Thr	Gly	Val	Leu	Leu	Cys	Ala	Leu	Leu	se	rc	ys I	_eu	Leu	بالثاب	•
			70			BO			90			3	00			11	D			120	
	*		*	*		*		*	*		*	_	*	*			*		*	*	
ACA	CCZ	ىلمكت	بلت	A.A.	GGT	A Th	AAA	מידים	AAA	ጥፋጋ	CCT	GAA	CTG	AGT	TT	A A	AA (	GC	ACC	CAG	
TGT	CCT	AGA	TCA	AGT	CCA	AGT	LalaL	AAT	LILL	CTA	GGA	CTT	GAC	TCA	. AA	TT	TT (	CCG	TGG	GTC	!
Thr	Glv	Ser	Ser	Ser	Gly	Ser	LVS	Leu	Lvs	Asp	Pro	Glu	Lev	Ser	Le	u L	vs (	Gly	Thr	Gln	>
					3		-4-		-,-								-	-			
	130					L <b>40</b>		150			160				170					180	
	*		*	*		*		R	*		*		*	1	:		*		*	•	
CAC	ATC	ATG	CAA	GCA	GGC	CAG	ACA	CTG	CAT	CTC	CAA	TGC	AGG	GGG	G)	AA G	CA	GCC	CAT	AA	1
GIG	TAG	TAC	GTT	CGT	CCC	GTC	TGT	GAC	GTA	GAG	GIT	ACG	TC	. cc	C	T C	GT	CGG	GTA	TT.	r
His	Ile	Met	Gln	Ala	Gly	Gln	Thr	Leu	His	Leu	Gln	Cys	Arg	Gly	/ G	Lu A	Ja	Ala	His	Ly	3>
		1	90			200			210			2	20			23	30		_	24	<b>.</b>
	*		*			*		*	*		*						. ~		_		
					ATG																
ACC	AGA	AAC	: GGA	CTT	TAC	CAC	TCA	TTC	CII	TCG	CIT	TCC	GA	2 10	3 T	AT :	Min.	111	For		
Trp	Ser	Leu	Pro	Glu	Met	Val	ser	Lys	GIU	ser	GIU	Arç	i re	n se	Z 1.	Te :	LIII.	гÃг	SET		-
		_				250			270				280			21	90			30	n
			:50			260		*	270		*	•	*		*	-	*		*		*
al Car	GC3	202	רע ע	י מפר	: AAA	CAA	TPT(	- 44.5c	י אכדו	אכיי	קובה י	AC	Turn	G AA	C A	CA (	CT	CAA	GCZ	AA A	С
JCJ.	200	1	1 dats 2 cocs		TIT	املت)	מת	. ACC	מישר ב	TGP	TAA	TG	3 AA	CTT	GT	GT (	CGA	GTI	· cc	TT	G
Cvs	Gly	Arc	. Ast	Gly	Lys	Gln	Phe	Cvs	Ser	Thr	Leu	Th	r Le	u As	n T	hr .	Ala	Glr	Ala	. As	n>
-2-	,	;	,	,	-2-			-4-													
		3	310			320			330	)			340			3	50			36	0
	*		*	•	,	*		*	•	•	*		*		*		*		*		*
CAC	ACT	GGG	TT	TAC	AGC	TGC	AA:	A TA	r CIX	GCT	r GT?	CC	T AC	T TO	A A	AG	aag	AA	GA:	A AC	A.
GTG	TGF	r cc	S AAC	TA E	TCG	ACC	TI	TA 1	A GA	CCI	A CA	GG	A TO	A AC	TI	TC	TTC	TT	CT	TT	T
His	Thr	Gly	/ Pho	e Tyr	: Ser	Cyl	Lys	Ty	r Le	ı Ale	a Val	l Pr	o Tr	ur S∈	r I	ys.	Lys	Ly	Gl	u Tr	ır>
																				4.	
		;	370			380			396				400			4	.10			42	*
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GAA	TC	r GC	TA A	C TA	TA T	/ Jala	r AT	r AG	T GA	r AC	A GG	r AG	A CC	71. 17.		TIA	CAL	MAT.	n var	C 47	27.
					A TAT																
GIU	Sez	c Ali	a II	е ту:	r Ile	e Pn	5 TT	e Se	I AS	ם יותו	r Gi	y Az	g Pi	O PI	ie (	/a1	GIU	rie:	c ly		
			430			440			45	n			460			4	170			4	80
	*		*		*	*		*		*	*		*		•	,	*		*		*
GAA			C GA	ል ልጥ	T AT	A CA	CAT	G AC	T GA	A GG	A AG	G GJ	G C	rc G	rc i	TT	CCC	TG	cœ	G G	TT
CIM	TAC	3 66	G CT	T TA	A TA	י פא	G TA	C TG	ACT	T CC	T TC	C C1	C G	AG C	AG 1	TAA	GGG	AC	G GC	:c c	AA
Glu	Il	e Pr	o Gl	u Il	e Il	e Hi	s Me	t Th	r Gl	u Gl	y Ar	g GI	u L	eu V	al :	Lle	Pro	э Су	ъ AI	g V	al>
											_										
490						500			51	0	520					:			5	40	
	*		*		*	*		*		*	*		*		*		*		*		*
ACC	TC	A CC	T AA	C AT	CAC	T GT	T AC	T TI	AA A'	A AA	G TT	T C	CA C	TT G	AC .	ACT	TT	AI	C CC	er G	AT
TGC	: AG	r GG	TT A	AT D	G TG	A CA	A TG	AA A	T TI	T TI	AA X	A G	er G	AA C	TG	TGA —	AA(	TA	re ex	غΑ.C	TA
Thi	: Se	r Pr	o As	n Il	e Th	r Va	l Th	r Le	u Ly	's Ly	rs Ph	e P	ro L	eu A	sp	ınr	re	u Il	e P	LOA	sp>

# Fig.16B.

		550		560				570				580			5			600	
	*		*	*		*		*	*		*		*	*		*		*	*
														TCA					
														AGT					
GIY	TĀZ	Arg	TTE	TTE	JTD.	Asp	ser	Arg	гуs	GIĀ	Pne	тте	116	Ser	ASN	ATS	THE	ıyr	n\e>
		6:	10		6	20			630			64	10		6	550			660
	*		*	*		*		*	*		*		*	*		*		*	*
GAA	ATA	GGG	CIT	CTG	ACC	TGT	GAA	GCA	ACA	GTC	AAT	GGG	CAT	TTG	TAT	AAG	ACA	AAC	TAT
CII	TAT	ccc	GAA	GAC	TGG	ACA	CTT	CGT	TGT	CAG	TTA	ccc	GTA	AAC	ATA	TTC	TGT	TTG	ATA
Glu	Ile	Gly	Leu	Leu	Thr	Cys	Glu	Ala	Thr	Val	Asn	Gly	His	Leu	Tyr	Lys	Thr	Asn	TYT>
		6	70			580			690			7	00			710			720
	*	·	*	*	•	*		* *			* *			•			*	*	
CTC	ACA	CAT	CGA	CAA	ACC	AAT	ACA	ATC	ATA	GAT	GTC	CAA	ATA	AGC	ACA	CCA	CGC	CCA	GTC
														TCG					
																			Val>
			_							_							•		
		7	30		•	740			750			7	60			770			780
	*		*	*		*		*	*		*		*	*		*		*	*
														ACC					
														TGG					
Lys	reo	Leu	Arg	GIY	HIS	Thi	ren	Vai	Leu	ASI	Cys	THI	ATA	The	Thi	PIO	Let	ASI	Thr>
		7	90			800			810			8	20			830			840
	*		*	*		*		*	*		*		*	*		*		*	*
AGA	GT	CA	ATC	ACC	TGG	AGT	TAC	CCT	GAT	GAA	AAA	LAA .	' AAC	AAC	GCT	TCC	GT	AGG	CGA
TCI	, CY	GM	TAC	TGG	ACC	TCA	ATG	GGA	CTA	CTI	TIT	TI	TTC	TTG	CGZ	AGG	CA	TC	GCT
Arg	[Va]	Glr	n Met	Thr	Trp	Ser	Tyr	Pro	Asp	Glu	Lys	Ası	Lys	Asn	Ala	Sex	Va.	LAr	arg>
		•	350			860			870	<b>1</b>			380			890			900
	*	•	*	•	,	*		*	6/6	,	*	•	*	*		*		*	*
CGA	ATT	CAC	CAZ	AGC	י אא	ישרי	י ראַיז	GCC	. AAC	· ATZ	אנייני	TAC	: AG1	ו בינים	املت	י אכיי	ידא י	r GA	C AAA
																			GTTT
																			p Lys>
												_							
		9	910			920			930	)		!	940	_		950			960
200	* -		*				~~~		~		* * 000	. ~	· *		. ~~			a Amm	- -
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																			e Lys>
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																			T TGT
																			A ACA
sei	va.	L AS	n Th	r Se	r Va.	His	s Ile	з Туг	r Asj	p Ly	s Ala	a Gl	y Pr	o Gly	/ G1	u Pr	o Ly	rs Se	r Cys>
		1	030			1040			105	0	1060							1080	
	*		*		*	*		* *		* *				*	*	*			
GA	C AA	A AC	T CA	C AC	A TG		A CC	G TG	c cc	A GC	A CC	T GA	A CT	CCI	G GG	G GG	A CC	C T	CTC
																			T CAG
Ası	o Ly	s Th	r Hi	s Th	r Cy	s Pr	o Pr	o Cy	s Pr	o Al	a Pr	o Gl	u Le	u Le	u Gl	A CJ	y Pi	co Se	er Val>

# Fig.16C.

	1090				11	00		1110				1120			1130				140			
,	*	* *				*		* -	*		*	112	*	•		*						
TTC C	TC	TTC	ccc	CCA	AAA	ccc	AAG	GAC	ACC	CTC	ATG	ATC	TCC	CGG	ACC	CCT	GAG	GTC	ACA			
AAG G																•						
Phe L	eu	Phe	Pro	Pro	Lys	Pro	Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr>			
		115				<b>.</b> .																
		115	*		11	.60			1170			118	30	_	11	.90		• 1	.200			
TGC G	TC:	GTG	ভোগে	GPC _	Calc	<b>7</b> CC	CAC		GZC -	<del>С</del> т	GAG.	CTY	A A C	- Abril	ממ	ancaca -	רארי	تىلت _	_			
ACG C																						
Cys V																						
													-			-						
	_	12:	LO		12	220			1230			124	-		12	250			260			
	*		*	*		*		*	*		*		*	•		*		*	*			
GGC G																						
Gly V																						
GIY V	aı	GIU	val	nis	Wali	AIG	гуs	1111	гуs	PLO	AIG	GIU	GIU	GIII	ıyı	ASII	ser	THE	Tyr>			
		12	70		1:	280			1290		1300		00		1	310	1320					
	*		*	*		*		*	*		*		*	*		*		*	*			
CGT G																						
GCA C																						
Arg (	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln	Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys>			
		13	30		1	340			1350			12	60		,	370			1380			
	*		*	*	-	*		*	*		*		*	*	-	*		*	*			
TGC A	AAG	GTC	TCC	AAC	AAA	GCC	CTC	CCA	GCC	ccc	ATC	GAG	AAA	ACC	ATC	TCC	AAA	GCC	AAA			
ACG :																						
Cys I	Lys	Val	Ser	Asn	Lys	Ala	Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys>			
			~~			400									_				- 440			
	*	13	90 *		1	400		*	1410			14	20		1	430			1440			
GGG (	CAG	ccc	CGA	GAA	CCA		GTC:				_	CCA	_	י רכים -	יבאים		Cutt	ACC	AAG			
ccc (																						
																			Lys>			
	_	14	.50	_	1	460		_	1470	)	_	14	80		1	490		_	1500			
ממ	ראם. "	CTY	. DCC	, Cutc	י ארכ	. W.	- Catto	. Cut.	- תמתי		· mm~	. m>u	· ~~	• • >		* • • • • •		· CITY	GAG			
TTG																						
																			Glu>			
						-			•	-												
		15	10		1	520			1530	)		15	40		j	.550			1560			
<b>6</b> 700 /	* ~~		•	*		*		*	•	' 	*		*			*		*	*			
																			TCC			
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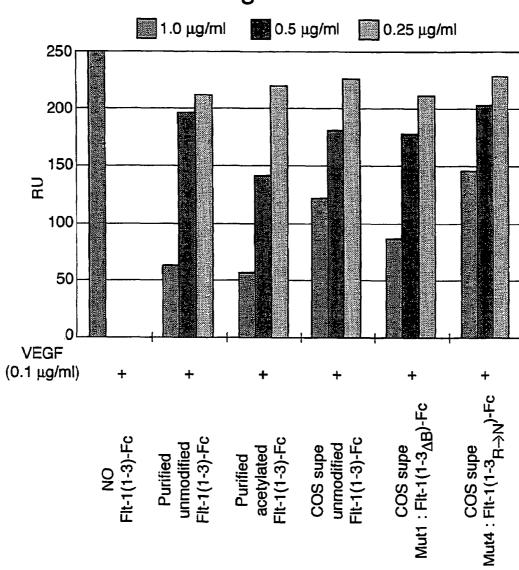
### Fig.16D.

CTC TCC CTG TCT CCG GGT AAA TGA
GAG AGG GAC AGA GGC CCA TTT ACT
Leu Ser Leu Ser Pro Gly Lys \*\*\*>

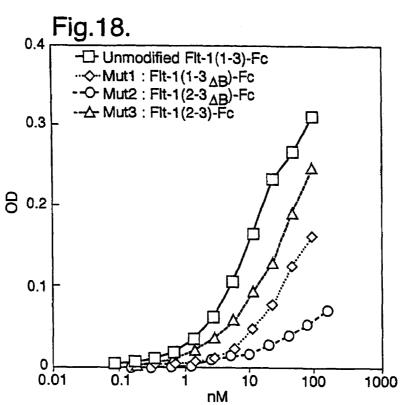
1700

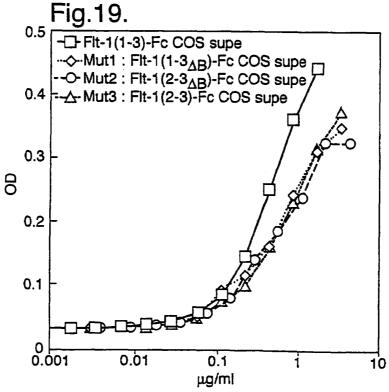
1690

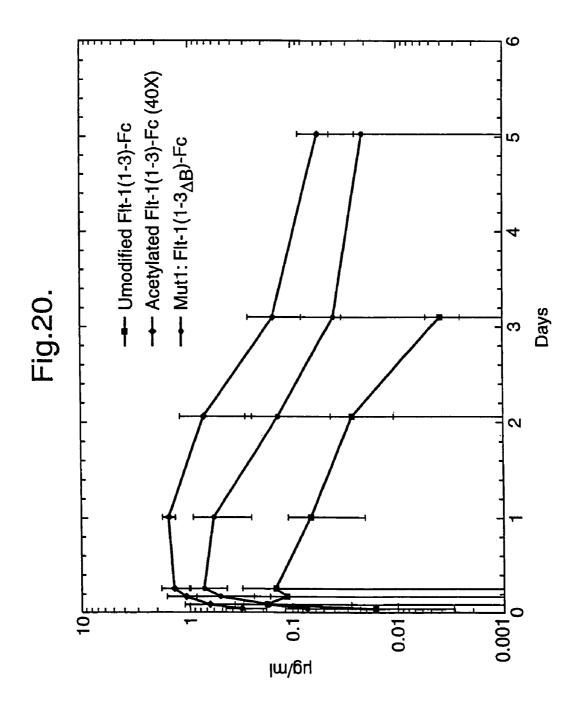
Fig.17.



May 20, 2008







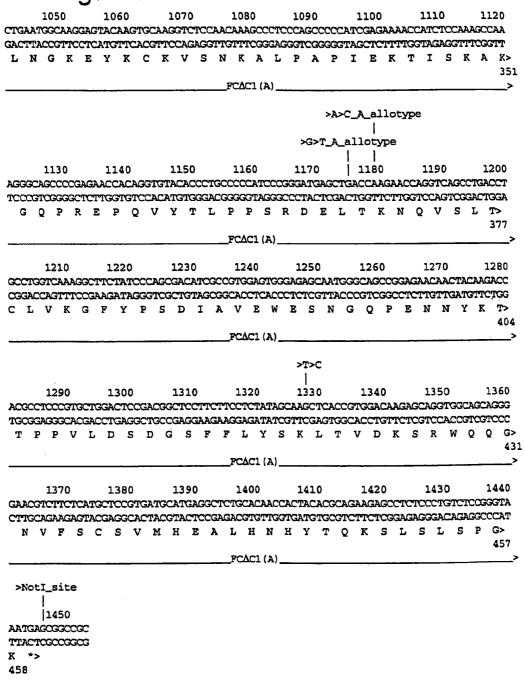
May 20, 2008

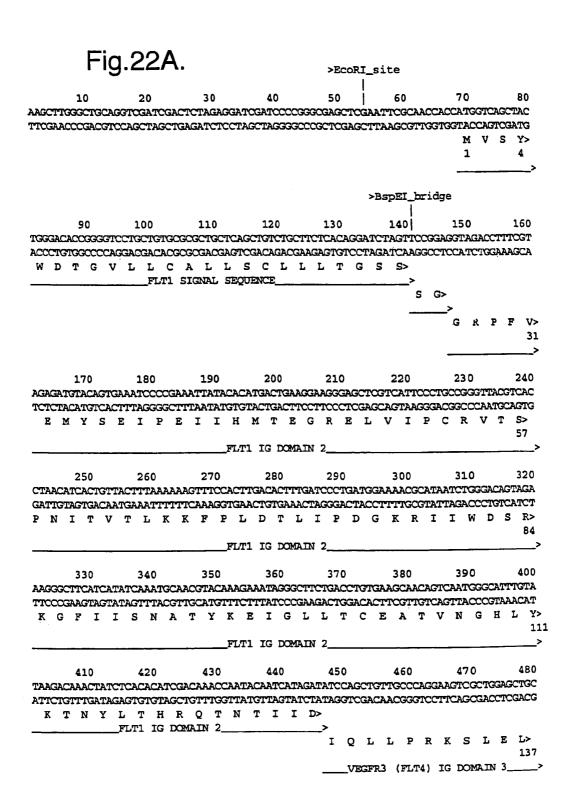
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CTAACATCACTGT GATTGTAGTGACA P N I T V  330 AAGGGCTTCATCA TTCCCGAAGTAGT	TACTITAAAAAAATTTTI T L K 340	AAGTTTCCAC MTCAAAGGTG K F P  350 AACGTACAAA	PTGACACTTT AACTGTGAAA L D T LHFLT1 D2 360 GAAATAGGGG	GATCCCTGA CTAGGGACT I P D 370 TTCTGACCT GAAGACTGGA	TGGAAAACGCF ACCTTTTGCGT G K R 380 GTGAAGCAAC	TAATCTGGC ATTAGACCC I I W  390 AGTCAATGGC	EACAGTAGA CTGTCATCA D S R3 84 400							
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CTAACATCACTGT GATTGTAGTGACA P N I T V  330 AAGGGCTTCATCA TTCCGAAGTAGT K G F I  410 TAAGACAAACTAT ATTCTGTTTGATA	TACTITAAAAA ATGAAATTTT T L K  340 TATCAAATGCI ATAGTITACGI I S N A  420 CTCACACATCI GAGTGTGTAGG	AGTTTCAC PTCAAAGGTG K F P  350 AACGTACAAA PTGCATGTTT T Y K  430 GACAAACCAA	TIGACACTTI AACTGIGAAA L D T L _HFLT1 D2  360 GAAATAGGGC CTITATCCCG E I G _HFLT1 D2  440 TACAATCATA ATGITAGIAA T I I	GATCCCTGA CTAGGGACT 370 TTCTGACCT AAGACTGGA L L T 450 AGATGTGGTT CTACACCAF	TGGAAACGCE ACCTTTTGCGT ACCTTTTGCGT AGACCAACL CACTTCGTTGT C E A T  460 ACTGAGTCCGTA	TAATCTGGC ATTAGACCC I I W  390 AGTCAATGGC CAGTTACCC V N G  470 ATCATGGAA	ACAGTAGE D S R 84 400 GCATTTGEE GTAAACA H L 1 1 486 TTGAACTA							
CTAACATCACTGT GATTGTAGTGACA P N I T V  330 AAGGGCTTCATCA TTCCGAAGTAGT K G F I  410 TAAGACAAACTAT ATTCTGTTTGATA	TACTITAAAAA ATGAAATTTT T L K  340 TATCAAATGCI ATAGTITACGI I S N A  420 CTCACACATCI GAGTGTGTAGG	AGTTTCAC PTCAAAGGTG  K F P  350 AACGTACAAA PTGCATGTTT  T Y K  430 GACAAACCAA CTGTTTGGTT  R Q T N	TIGACACTTI AACTGIGAAA L D T L _HFLT1 D2  360 GAAATAGGGC CTITATCCCG E I G _HFLT1 D2  440 TACAATCATA ATGITAGIAA T I I	GATCCCTGA CTAGGGACT 370 TTCTGACCT AAGACTGGA L L T 450 AGATGTGGTT CTACACCAF	TGGAAACGCA ACCITITGCGT ACCITITGCGT AGCAACAACA CACITCGTTGT C E A T  460	TAATCTGGC ATTAGACCC I I W  390 AGTCAATGGC CAGTTACCC V N G  470 ATCATGGAA	ACAGTAGE D S R 84 400 GCATTTGEE GTAAACA H L 1 1 486 TTGAACTA							

## Fig.21B.

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#### Fig.21C.



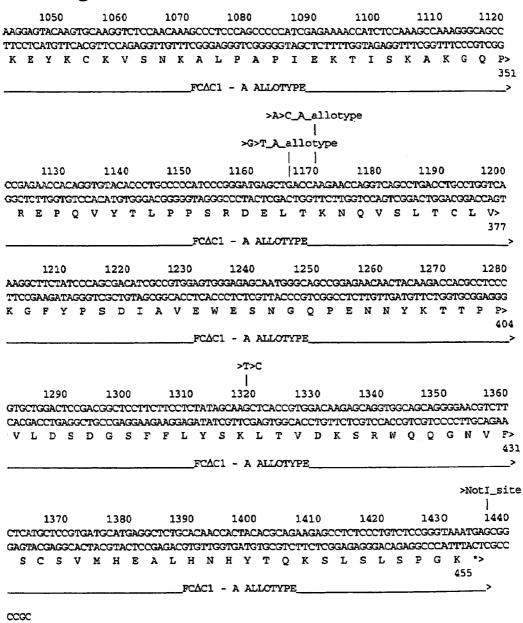


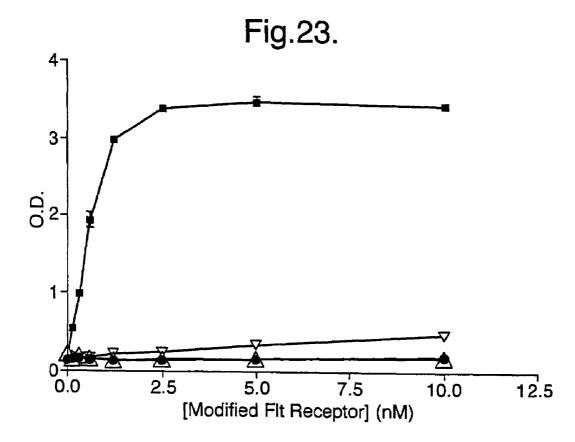
# Fig.22B.

GGTA		90 GAG			50 GT										530 TAAC		GGT		-							560 CCA
															attg N										_	
								V	GFR	13	(FL	T4)	IG	DO	MAIN	3_										<u></u> >
GGAZ	_	70 GGC				-									610 AACA											640 GAC
															TTGT Q Q											
								v	EGFF	23	(FI	.T4)	IG	DO	MAIN	3_										`
ATC	_	50 ACG													690 GCCA											720 .GCA
															CGG1											
								v	EGFI	R3	(FI	T4)	IG	DC	MAIN	3_										
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GGCG

# Fig.22C.





Flt1D2Flk1D3.FcdeltaC1(a)
 △Flt1D2VEGFR3D3.FcdeltaC1(a)
 ▼TIE2-Fc

■ Fit1(1-3)-Fc

# Fig.24A.

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# Fig.24B.

May 20, 2008

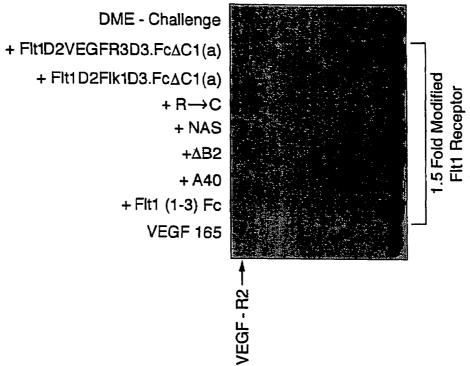
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# Fig.24C.

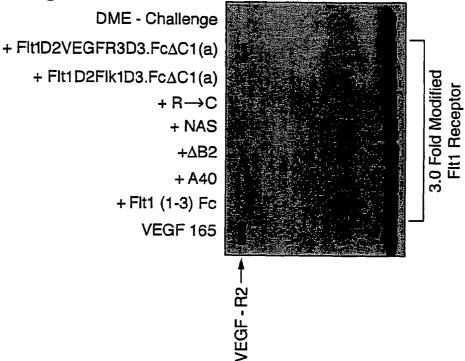
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TTT K	∞ G	TTC AAG F	* TAT ATA Y	ccc ccc gcc	AGC TCG S	GAC CTG D	ATC TAG	GCC CCC A	GTG CAC V	GAG CTC E	TGG ACC W	GAG CTC E	AGC TCG S	TAA ATT N	. ccc	CAG GTC	CCC GGC	GAG CIC	* AAC TTG
TTT K	∞ G	TTC AAG F	* TAT ATA Y	ccc ccc gcc	AGC TCG S	GAC CTG D	ATC TAG	GCC CCC A	GTG CAC V	GAG CTC E	TGG ACC W	GAG CTC E	AGC TCG S	TAA ATT N	. ccc	CAG GTC	CCC GGC	GAG CIC	* AAC TTG
TTT K	∞ G	TTC AAG F	* TAT ATA Y	ccc ccc gcc	AGC TCG S	GAC CTG D	TAG I	CCC CCC A h	GTG CAC V	GAG CIC E L A	TGG ACC W	GAG CTC E	AGC TCG S	TTA TTA N 395	G CCCC	CAG GTC Q	GGC P	GAG CTC E	* AAC TTG
TTT K 381_	G	TTC AAG F	* TAT ATA Y	CCC GGG P _385	AGC TCG S	GAC CTG D	ATC TAG I	A CGG	GTG V FCAC 1230	GAG CTC E LA	TGG ACC W	GAG E CTC E	AGC TCG S	TAA : ATT : N 295_	G GGG G G	CAG CTC Q 250	P GGC	GAG	* AAC TTG N> _400>
TTT K 381_ AAC TTG	G G TAC	AAG	* TAT ATA Y 10 * ACC	CCC P 385	AGC TCG S 1 CCT GGA	GAC CTG D 220 *	TAG I GIG	CTG	GTG CAC V FCAC 1230 * GAC CTG	GAG CTC E I A TCC AGG	TGG ACC W	GAG E 12 12 CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	AGC TCG S 40 * TCC	AAT N 395	GGG G G	CAG CTC Q 250	CCG P P TAC	GAG	* AAC TTG N> _400> 1260 * AAG TTC
TTT K 381_ AAC TTG N	TAC ATG	TTC AAG F 12 AAG	TATA Y 10 * ACC TGG	CCC GGG P _385 ACG	AGC TCG S	GAC CTG D 220 * CCC GGG	TAG I GIG	GCC CGG A h	GTG CAC V FCAC 1230 * GAC CTG	GAG CTC E L A TCC AGG	TGG ACC W	GAG E 12 12 GGC G	AGC TCG S 40 * TCC AGG	AAT N 395	GGG G G 1 TTC AAC	CAG CTC Q 250	GGC P TAC ATC	E AGC	* : AAC : TTG N> _400>  1260
TTT K 381_ AAC TTG N	TAC ATG	TTC AAG F 12 AAG	TATA Y 10 * ACC TGG	CCC GGG P _385 ACG	AGC TCG S	GAC CTG D 220 * CCC GGG	TAG I GIG	GCC CGG A h	GTG CAC V FCAC 1230 * GAC CTG	GAG CTC E L A TCC AGG	TGG ACC W	GAG E 12 12 GGC G	AGC TCG S 40 * TCC AGG	AAT N 395	GGG G G 1 TTC AAC	CAG CTC Q 250	GGC P TAC ATC	E AGC	* AAC TTG N> 400> 1260 * AAG TTC K> 420>
TTT K 381_ AAC TTG N	TAC ATG	TTC AAG F 12 AAG TTC K	TATA Y 10 * ACC TGG	CCC GGG P _385 ACG	AGC TCG S 11 CCT GGA P	GAC CTG D 220 * CCC GGG	TAG I GIG	CTG	GTG CAC V FCAC 1230 * GAC CTG	GAG CTC E 1 A TCC AGG S 1 A	TGG ACC W	12 GGC GGC G	AGC TCG S 40 * TCC AGG S	AAT N 395	GGG G G TTC G AAG	CAG CTC Q 250	GGC P TAC ATC	E AGC	* : AAC : TTG N> _400>  1260
TTT K 381 AAC TTG N 401	TAC ATG Y	TTC AAG F  12 AAG TTC K	TATA Y  10 * ACC TGG T	CCC GGG P_385 ACG TGC T_405	AGC TCG S 11 CCT GGA P 1	GAC CTG D 220 * CCC GGG P 280 *	TAG	GCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	CAC  CAC  V  FCAC  1230  *  GAC  D  1290  *  CAG  CAG	GAG CTC E AGG AGG S 1 A	TGG ACC W	GAG CTC E 12 GGC G 13	AGC TCG S 40 * TCC AGG S	TTA N 395 TTC	GGG G G TTC G AAG F	250 2 CTC C GAG L .310	GGC P TAC	GAGC CICC E	# AAC TTG N> _400> 1260 * AAG TTC R> _420> 1320 * CAT
AAC TIG N 401	TACO	TTC AAG F  12 AAG TTC K	* TATA Y 10 * ACC TGG T	CCC GGG P _385 ACG TGC T_405	AGC TOG S  1: CCT GGA P  1 AGC TOG	CACCCTG D 220 * CCCCGGGGP 280 * AGG	ATC TAG I GTG CAC V	CTG A A B CTG	CAC	GAG E I A TCC AGG S I A	ACC W GACC D ACC S	12 12 13 13 13 13	AGC TOG S 40 * TOC AGG S O0 * TTC	AAT N 395 TTC AAC F 415	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CAG Q 250 * CTC GAG L 310 * TCC GAG S S S S S S S S S S S S S S S S S S	GGC P TAC TAC Y CAC CAC CAC CAC CAC CAC CAC CAC CAC	E ASC TAC	* AAC TTG N> 400>  1260 * AAG TTC K> 420>  1320 * CAT GTA
AAC TTG N 401	TACO	TTC AAG F 12 AAG TTC K 12 GTG V	* TATT ATTA Y  10  * ACCC T  770  * GACC  CTGG  T	CCCC GGG P _385 ACG TGC TGC TCC _405	AGC TOG S 1. CCT GGA P 1. AGC TOG S	CACCCIGGD D * CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	ATC TAG I GIG CAC V TGG ACC W	CTG A A A A A CTG	GTG CAC V FFCAC 1230 * GAC 1230 * 1290 * GAC 1290 * CAG GCAG CAG CAG CAG CAG CAG CAG CAG CA	GAG CIC E I A TCC AGG S S I A	ACCOME GACCOME	12 COCG G 13 COCG C V	AGC S 40 * TCC AGC S O0 * TTCC F	AATIAN N 395 AAG	GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	CAGO Q 250 * COTO GAGG L 310 * TOTO S S	GGC P TAC : TAC Y CAC V	GAGC E  AGC S  TAC M	# AAC TTG N> 400> 1260 * AAG TTC K> 420> 1320 * GTA H>
AAC TTG N 401	TACO	TTCC AAG F 12 AAG K 12 CGGG CAC V	TATA Y  10  * ACC T  70  * GAC C C C C C C C C C C C C C C C C C C	CCC GGG P 385 ACG T 405	AGC TOG S  1 CCT GGA P  1 AGC TOG S	CAC CIG D 220 * CCC GGG P 280 * AGG TOCC R	ATC TAG I GIG CAC V TGG ACC W	CTG A A A A A CTG	GIGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	GAG CTC E 1 A TCC AGG S 11 A GGG G G 1 A	TGG ACC W GACC TGG ACC N AACC N AACC N AACC N	12 GGC G G 13 CGC CAG V	AGC S S AGG S F TTCC	TTA  N  395  TTC  TTC  415  TC2  435	GGG GGG GGG GGG GGG GGG GGGG GGGG GGGG GGGG	250 * CTC CTC CTC CTC S AGG	GGC P TAC : TAC Y CAC V	GAGC E  AGC S  TAC M	* AAC TTG N> 400>  1260 * AAG TTC K> 420>  1320 * CAT GTA
AAC TTG N 401	TACO	TTCC AAG F 12 AAG K 12 CGGG CAC V	* TATT ATTA Y  10  * ACCC T  770  * GACC  CTGG  T	CCC GGG P 385 ACG T 405	AGC TOG S  1 CCT GGA P  1 AGC TOG S	CACCCIGGD D * CCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	ATC TAG I GIG CAC V TGG ACC W	CTG A A A A A CTG	GTG CAC V FFCAC 1230 * GAC 1230 * 1290 * GAC 1290 * CAG GCAG CAG CAG CAG CAG CAG CAG CAG CA	GAG CTC E 1 A TCC AGG S 11 A GGG G G 1 A	TGG ACC W GACC TGG ACC N AACC N AACC N AACC N	12 COCG G 13 COCG C V	AGC S S AGG S F TTCC	TTA  N  395  TTC  TTC  415  TC2  435	GGG GGG GGG GGG GGG GGG GGGG GGGG GGGG GGGG	CAGO Q 250 * COTO GAGG L 310 * TOTO S S	GGC P TAC : TAC Y CAC V	GAGC E  AGC S  TAC M	# AAC TTG N> 400> 1260 * AAG TTC K> 420> 1320 * GTA H>
AAC TTG N 401 CTC GAG L 421	TACO	TTCC AAG F 122 AAG K 122 CAC V 13	* TATTA Y 10 * ACCC TGG T 770 * GACC CGG D	CCC GGG P 385 TGC T 405	AGC TOG S  1 CCCT GGA P  1 AGC TOG S	CAC CTG D 220 * CCC GGG P 280 * AGG TCC R 340 * TAC	ATC TAG I GTG CAC V TGG ACC W	GCCCGG A A CCTG GAC L CAC GAC C CAC C CAC C C C C C C C C C C	GTGCAC  V  FCAC  12300  * * GAC  CTG  D  * * * * * * * * * * * * * * * * *	GAG CTC E 1 A TCC AGG S 1 A GGGG CCC G 1 A	TGG ACC W GACC TGG ACC	12 COCC G G 13 COCC C C C C C C C C C C C C C C C C C	AGC TCG S S 40 * TCC S S S S S S S S S S S S S S S S S S	AAI  TTA  N  395  TTC  TC  TC  S  435	1 TGC	CAG	GGGC P TACC Y CACC CACC CACC CACC CACC CACC CA	GAGGE CTC E  E  AGC TCC  S  ATC  M	* AAC TTG N> 400>  1260 * AAG TTC K> 420>  1320 * GTA H> 440>
AAC TTG N 401 CTC GAG L 421	TACO TACO TACO TACO TACO TACO TACO TACO	TTIC AAG F 122 AAG K 122 CAC V 133 CAC V	* TATTA Y 10 * ACCC TGG T 770 * GACC CGG D	CCC GGG P P 385 TGC T TGC T AAG TTC K 425 TTC K AAG TTC K AAG TTC TTC TTC TTC TTC TTC TTC TTC TTC TT	AGC TOG S  1 CCCT GGA P  1 AGC TOG S	CAC CTG D 220 * CCC GGG P 280 * AGG TCC R 340 * TAC ATG	ATC TAG I GTG CAC V TGG ACC W	CAG	GTGCAC  V  FCAC  12300  * * GAC  CTG  D  * * * CAG  CAG  CTG  CAG  CAG  CAG  CAG  CAG	GAGG CTC E 1 A TCC AGG S S 1 A CCC G TA AGG CCC G TCC TCC TCC TCC TCC TCC TCC TCC	TGG ACC W GACC TGGAC  GACC TGGAC  GACC  GACG  GACC  GACG  GACC  GA	12 COCC G G 13 CAGC V 13 CAGC S AGGC	AGC TCG S S 40 * TCC S S S S S S S S S S S S S S S S S S	AATI TTA N 395 TTC: TTC: TC: TC: AG: AG: TC: AG: AG: AG: AG: AG: AG: AG: AG: AG: AG	1 COCC C C C C C C C C C C C C C C C C C	CAG	GGGC P  TACK Y  CACC V  AAA  TTACA T	GAGGE CTCC E  E  AGCC S  S  ATCC M  M  A TGA	* AAC TTG N> 400>  1260 * AAG TTC K> 420>  1320 * GTA H> 440>
AAC TTG N 401 CTC GAG L 421 GAG CTC	TACC TGG ATG A A A A A A A A A A A A A A A A	122 AAGS TOO K	* TATA Y 10 * ACCC T 70 * GACC * CTG D 30 * GACC GTG H	GGG P 385.  AGG TGG T AAGG TTG A425	AGC TOG S  1 CCCT GGA P  1 AGC TOG S	CAC CTG D 220 * CCC GGGG P 280 * AGG TCC ATG ATG Y	ATC TAG I GTG CAC V TGG ACC T T T	CACCO	GIGGE CAC  V  1230  * * * * * * * * * * * * * * * * * *	GAG CTC E I I A TCC AGG S I I A GGGG CC C	TGG ACC W ACC TO D AC	12 CTC E 12 CCG G CCG CCG CCG CCG CCG CCG CCG CCG	AGC TCG S 40 * TCCG AGG S 00 * TTCC AAGG F 60 * CTCG GAC L	AAI TTA AAG F15 AAG F415 AAG F415 AAG F15 AAG	1 TGC	CAG	GGGC P TAGE CAC V TAGE TAGE TAGE TAGE TAGE TAGE TAGE TAGE	GAGE CTC E E CAGE TOX S ATT M A TG A TG *	* AAC TTG N> 400> 1260 * AAG TTC R> 420> 1320 * TTC GTA H> 440>

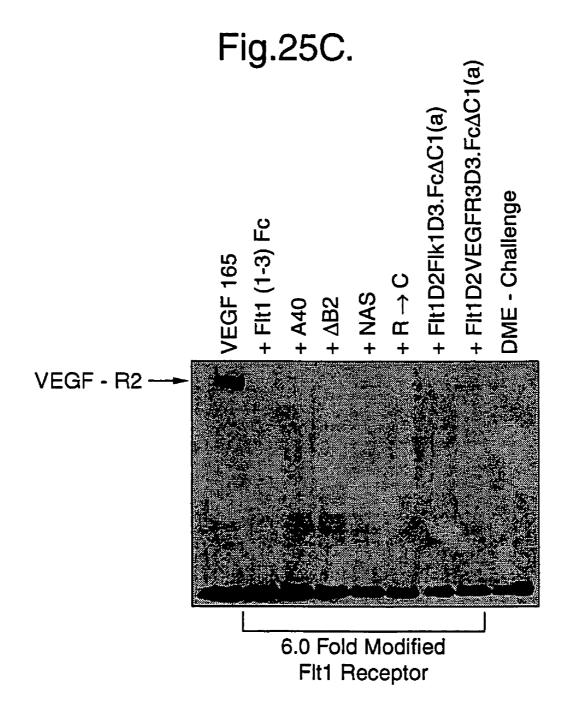
# Fig.25A.

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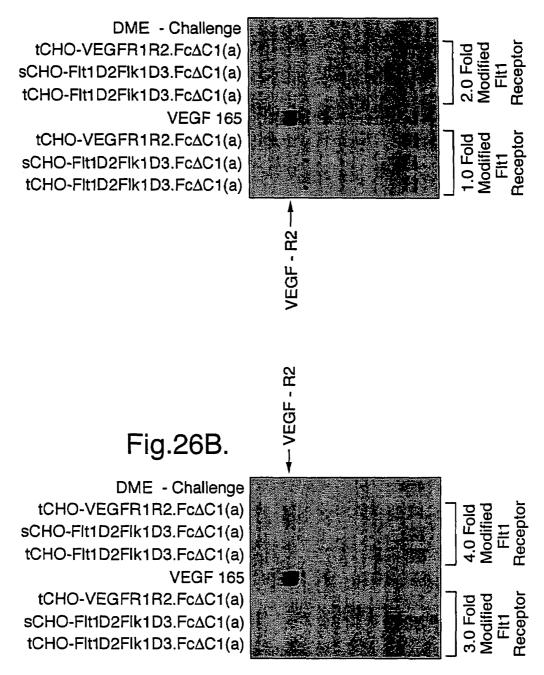


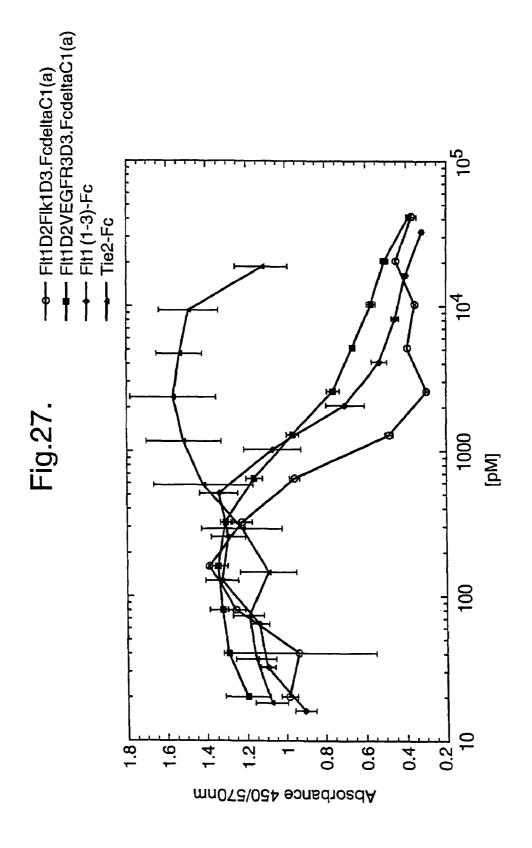
# Fig.25B.





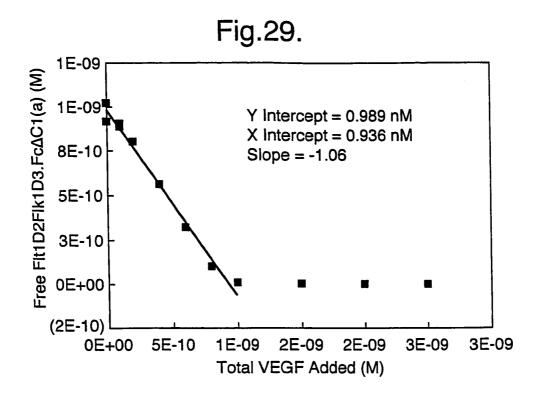
# Fig.26A.





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Binding Stoichio	toichiometry of hVEGF165 to FIt	metry of hVEGF165 to FIt1D2FIk1D3.FcΔC1(a) & VEGFR1R2-FcΔC1(a)
hVEGF165 (nM)	hVEGF165 (nM) VEGF/Flt1D2Flk1D3.FcΔC1(a)	VEGF/VEGFR1R2-Fc∆C1(a)
-	0.93	0.98
10	76.0	0.94
50	-	0.99
Average ± StDev	0.96 ± 0.03	0.97 ± 0.02



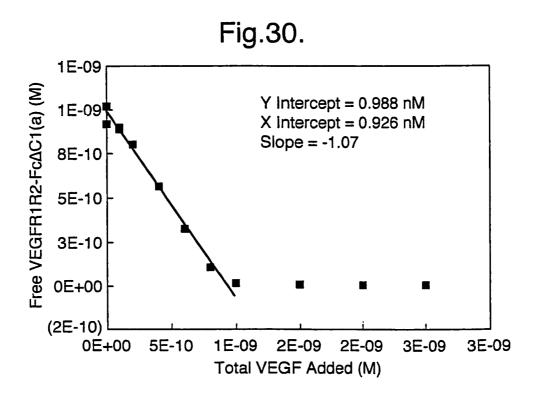
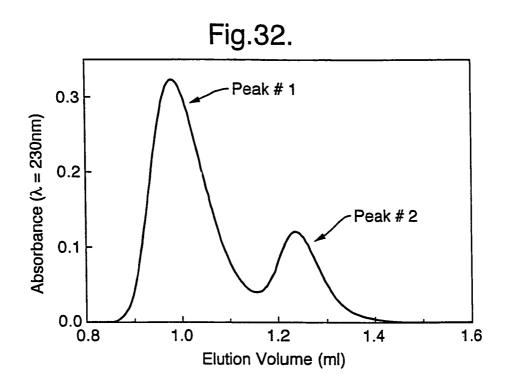
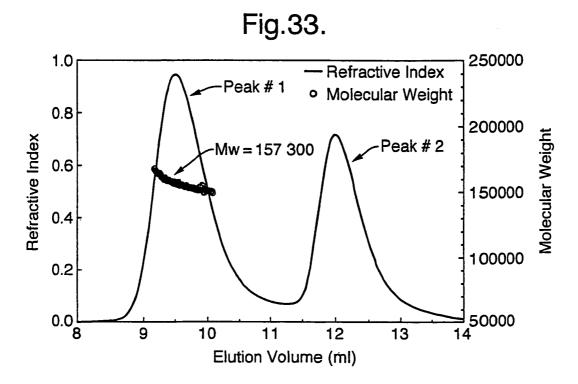
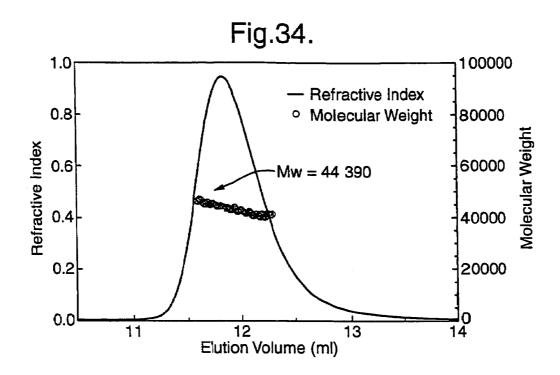


Fig.31. 2.5 Peak # 1 2.0 Absorbance ( $\lambda = 230 \text{ nm}$ ) 1.5 1.0 Peak # 2 0.5 0.0 1.2 1.4 1.6 1.8 2.0 1.0 Elution Volume (ml)







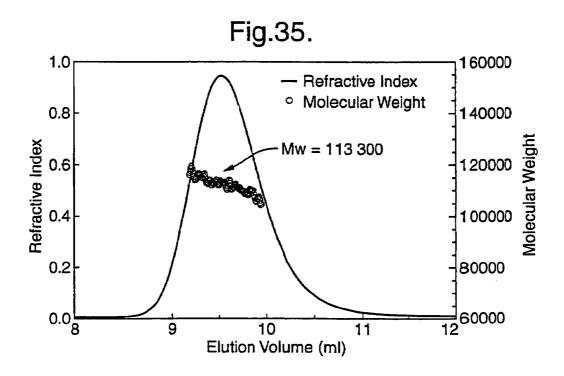


Fig.36.

DLKTQSGSEMKKFLSTLTIDGVTRSDQGLYT<u>C</u>AASSGLMTKK<u>N</u>STFVRVH VVLSPSHGIELSVGEKLVL<u>NC</u>TARTELNVGIDFNWEYPSSKHQHKKLVNR KRIIWDSRKGFIIS<u>N</u>ATYKEIGLLT<u>C</u>EATVNGHLYKTNYLTHRQTNTIIL **GRPFVEMYSEIPEIIHMTEGRELVIP<u>C</u>RVTSP<u>N</u>ITVTLKKFPLDTLIP**DG

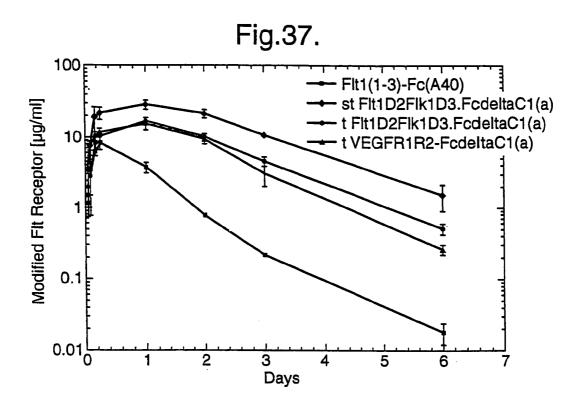
GKEYK<u>C</u>KVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQVSL

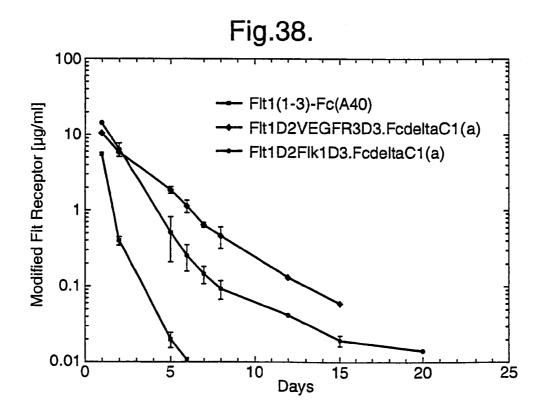
VSHEDPEVKENWYVDGVEVHNAKTKPREEQY<u>N</u>STYRVVSVLTVLHQDWLN

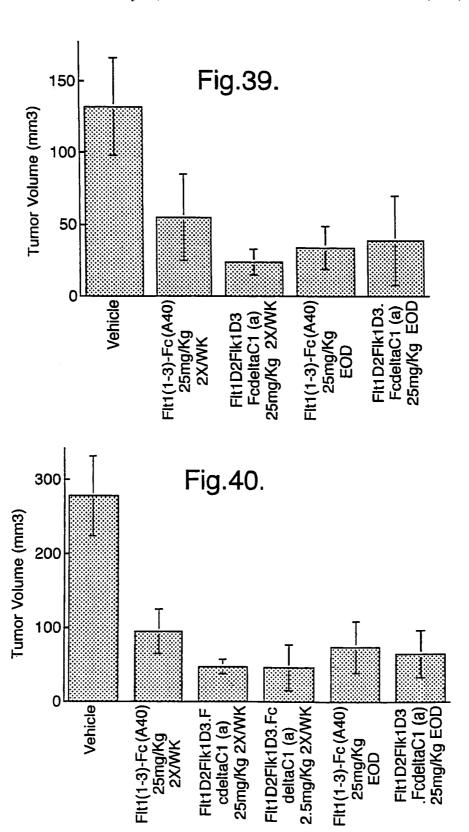
EKGPGDKTHT<u>C</u>PPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD

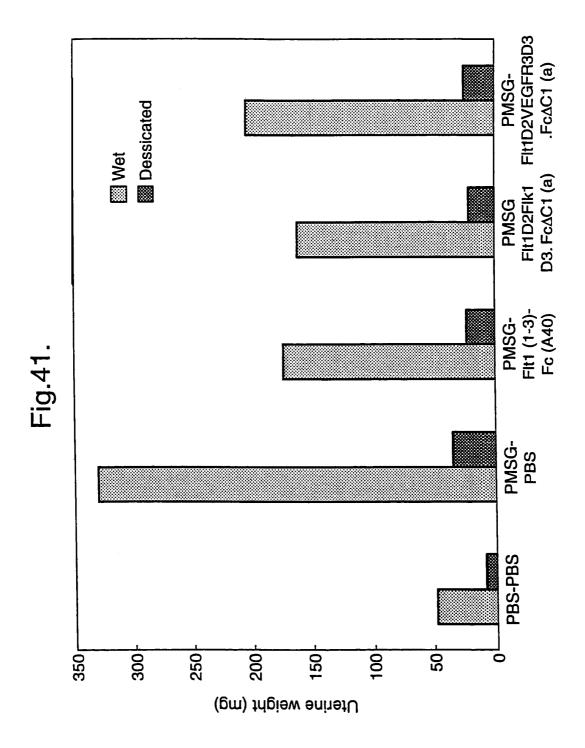
TCL VKGFYPSDIA VEWESNGQPENNYKTTPPVLDSDGSFFL YSKLTVDKS

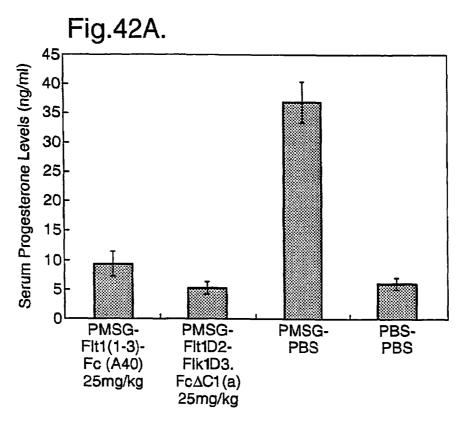
**RWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK** 

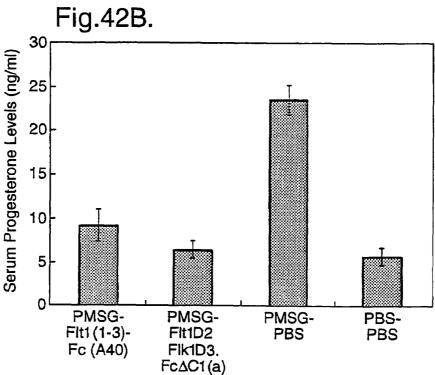












#### MODIFIED CHIMERIC POLYPEPTIDES WITH IMPROVED PHARMACOKINETIC PROPERTIES AND METHODS OF USING THEREOF

This application is a divisional of U.S. patent application Ser. No. 10/009,852, filed Dec. 6, 2001, now U.S. Pat. No. 7,070,959, which is a national stage application of International Application No. PCT/US00/14142, filed May 23, 2000, which claims priority of U.S. Provisional Application 10 Ser. No. 60/138,133, filed Jun. 8, 1999. The disclosures of these applications are herein specifically incorporated by reference in their entirety.

#### INTRODUCTION

The field of this invention is modified polypeptides with improved pharmacokinetics. Specifically, the field of this invention relates to Flt1 receptor polypeptides that have been modified in such a way as to improve their pharmacokinetic profile. The field of this invention also relates to methods of making and using the modified polypeptides including but not limited to using the modified polypeptides to decrease or inhibit plasma leakage and/or vascular permeability in a mammal.

#### BACKGROUND

The ability of polypeptide ligands to bind to cells and thereby elicit a phenotypic response such as cell growth, survival, cell product secretion, or differentiation is often mediated through transmembrane receptors on the cells. The extracellular domain of such receptors (i.e. that portion of the receptor that is displayed on the surface of the cell) is generally the most distinctive portion of the molecule, as it 35 provides the protein with its ligand binding characteristic. Binding of a ligand to the extracellular domain generally results in signal transduction which transmits a biological signal to intracellular targets. Often, this signal transduction acts via a catalytic intracellular domain. The particular array of sequence motifs of this catalytic intracellular domain determines its access to potential kinase substrates (Mohammadi, et al., 1990, Mol. Cell. Biol. 11:5068-5078; Fantl, et al., 1992, Cell 69:413-413). Examples of receptors that transduce signals via catalytic intracellular domains include 45 the receptor tyrosine kinases (RTKs) such as the Trk family of receptors which are generally limited to cells of the nervous system, the cytokine family of receptors including the tripartate CNTF receptor complex (Stahl & Yancopoulos, 1994, J. Neurobio. 25:1454-1466) which is also gener- 50 ally limited to the cells of the nervous system, G-protein coupled receptors such as the  $\beta_2$ -adrenergic receptor found on, for instance, cardiac muscle cells, and the multimeric IgE high affinity receptor Fc∈RI which is localized, for the most part, on mast cells and basophils (Sutton & Gould, 55 1993, Nature 366:421-428).

All receptors identified so far appear to undergo dimerization, multimerization, or some related conformational change following ligand binding (Schlessinger, J., 1988, Trend Biochem. Sci. 13:443-447; Ullrich & Schlessinger, 60 1990, Cell 61:203-212; Schlessinger & Ullrich, 1992, Neuron 9:383-391) and molecular interactions between dimerizing intracellular domains lead to activation of catalytic function. In some instances, such as platelet-derived growth factor (PDGF), the ligand is a dimer that binds two receptor 65 molecules (Hart, et al., 1988, Science, 240:1529-1531; Heldin, 1989, J. Biol. Chem. 264:8905-8912) while, for

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example, in the case of epidermal growth factor (EGF), the ligand is a monomer (Weber, et al., 1984, J. Biol. Chem. 259:14631-14636). In the case of the FceRI receptor, the ligand, IgE, exists bound to FceRI in a monomeric fashion and only becomes activated when antigen binds to the IgE/FceRI complex and cross-links adjacent IgE molecules (Sutton & Gould, 1993, Nature 366:421-428).

Often, the tissue distribution of a particular receptor within higher organisms provides insight into the biological function of the receptor. The RTKs for some growth and differentiation factors, such as fibroblast growth factor (FGF), are widely expressed and therefore appear to play some general role in tissue growth and maintenance. Members of the Trk RTK family (Glass & Yancopoulos, 1993, 15 Trends in Cell Biol. 3:262-268) of receptors are more generally limited to cells of the nervous system, and the Nerve Growth Factor family consisting of nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), neurotrophin-3 (NT-3) and neurotrophin-4/5 (NT-4/5), which bind the Trk RTK family receptors, promote the differentiation of diverse groups of neurons in the brain and periphery (Lindsay, R. M, 1993, in Neurotrophic Factors, S. E. Loughlin & J. H. Fallon, eds., pp. 257-284, San Diego, Calif., Academic Press). Fc∈RI is localized to a very limited 25 number of types of cells such as mast cells and basophils. Mast cells derive from bone marrow pluripotent hematopoietic stem cell lineage, but complete their maturation in the tissue following migration from the blood stream (See Janeway & Travers, 1996, in Immunobiology, 2d. Edition, M. Robertson & E. Lawrence, eds., pp. 1:3-1:4, Current Biology Ltd., London, UK, Publisher) and are involved in the allergic response.

Many studies have demonstrated that the extracellular domain of a receptor provides the specific ligand binding characteristic. Furthermore, the cellular environment in which a receptor is expressed may influence the biological response exhibited upon binding of a ligand to the receptor. For example, when a neuronal cell expressing a Trk receptor is exposed to a neurorophin which binds to that receptor neuronal survival and differentiation results. When the same receptor is expressed by a fibroblast, exposure to the neurotrophin results in proliferation of the fibroblast (Glass, et al., 1991, Cell 66:405-413).

A class of cell-derived dimeric mitogens with selectivity for vascular endothelial cells has been identified and designated vascular endothelial cell growth factor (VEGF). VEGF has been purified from conditioned growth media of rat glioma cells [Conn et al., (1990), Proc. Natl. Acad. Sci. U.S.A., 87. pp 2628-2632]; and conditioned growth media of bovine pituitary follicle stellate cells [Ferrara and Henzel, (1989), Biochem. Biophys. Res. Comm., 161, pp. 851-858; Gozpadorowicz et al., (1989), Proc. Natl. Acad. Sci. U.S.A., 86, pp. 7311-7315] and conditioned growth medium from human U937 cells [Connolly, D. T. et al. (1989), Science, 246, pp. 1309-1312]. VEGF is a dimer with an apparent molecular mass of about 46 kDa with each subunit having an apparent molecular mass of about 23 kDa. VEGF has some structural similarities to platelet derived growth factor (PDGF), which is a mitogen for connective tissue cells but not mitogenic for vascular endothelial cells from large

The membrane-bound tyrosine kinase receptor, known as Flt, was shown to be a VEGF receptor [DeVries, C. et al., (1992), Science, 255, pp. 989-991]. The Flt receptor specifically binds VEGF which induces mitogenesis. Another form of the VEGF receptor, designated KDR, is also known to bind VEGF and induce mitogenesis. The partial cDNA

sequence and nearly full length protein sequence of KDR is known as well [Terman, B. I. et al., (1991) Oncogene 6, pp. 1677-1683; Terman, B. I. et al., (1992) Biochem. Biophys. Res. Comm. 187, pp. 1579-1586].

Persistent angiogenesis may cause or exacerbate certain 5 diseases such as psoriasis, rheumatoid arthritis, hemangiomas, angiofibromas, diabetic retinopathy and neovascular glaucoma. An inhibitor of VEGF activity would be useful as a treatment for such diseases and other VEGF-induced pathological angiogenesis and vascular permeability conditions, such as tumor vascularization. The present invention relates to a VEGF inhibitor that is based on the VEGF receptor Flt1.

Plasma leakage, a key component of inflammation, occurs in a distinct subset of microvessels. In particular, in most organs plasma leakage occurs specifically in the venules. Unlike arterioles and capillaries, venules become leaky in response to numerous inflammatory mediators including histamine, bradykinin, and serotonin. One characteristic of inflammation is the plasma leakage that results from inter- 20 cellular gaps that form in the endothelium of venules. Most experimental models of inflammation indicate that these intercellular gaps occur between the endothelial cells of postcapillary and collecting venules (Baluk, P., et al., Am. J. Pathol. 1998 152:1463-76). It has been shown that certain 25 lectins may be used to reveal features of focal sites of plasma leakage, endothelial gaps, and finger-like processes at endothelial cell borders in inflamed venules (Thurston, G., et al., Am. J.

Physiol, 1996, 271: H2547-62). In particular, plant lectins 30 have been used to visualize morphological changes at endothelial cell borders in inflamed venules of, for example, the rat trachea. Lectins, such as conconavalin A and ricin, that bind focally to inflamed venules reveal regions of the subendothelial vessel wall exposed by gaps that correspond 35 to sites of plasma leakage (Thurston, G., et al., Am J Physiol, 1996, 271: H2547-62).

The properties of the microvessels are dynamic. Chronic inflammatory diseases, for example, are associated with microvascular remodeling, including angiogenesis and 40 microvessel enlargement. Microvessels can also remodel by acquiring abnormal phenotypic properties. In a murine model of chronic airway inflammation, airway capillaries acquire properties of venules, including widened vessel diameter, increased immunoreactivity for von Willebrand 45 factor, and increased immunoreactivity for P-selectin. In addition, these remodeled vessels leak in response to inflammatory mediators, whereas vessels in the same position in the airways of normal mice do not.

Certain substances have been shown to decrease or inhibit vascular permeability and/or plasma leakage. For example, mystixins are synthetic polypeptides that have been reported to inhibit plasma leakage without blocking endothelial gap formation (Baluk, P., et al., J. Pharmacol. Exp. Ther., 1998, 284: 693-9). Also, the beta 2-adrenergic receptor agonist 55 formoterol reduces microvascular leakage by inhibiting endothelial gap formation (Baluk, P. and McDonald, D. M., Am. J. Physiol., 1994, 266:L461-8).

The angiopoietins and members of the vascular endothelial growth factor (VEGF) family are the only growth factors 60 thought to be largely specific for vascular endothelial cells. Targeted gene inactivation studies in mice have shown that VEGF is necessary for the early stages of vascular development and that Ang-1 is required for later stages of vascular remodeling.

U.S. Pat. No. 6,011,003, issued Jan. 4, 2000, in the name of Metris Therapeutics Limited, discloses an altered, soluble

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form of FLT polypeptide being capable of binding to VEGF and thereby exerting an inhibitory effect thereon, the polypeptide comprising five or fewer complete immunoglobulin domains.

U.S. Pat. No. 5,712,380, issued Jan. 27, 1998 and assigned to Merck & Co., discloses vascular endothelial cell growth factor (VEGF) inhibitors that are naturally occurring or recombinantly engineered soluble forms with or without a C-terminal transmembrane region of the receptor for VEGF

Also assigned to Merck & Co. is PCT Publication No. WO 98/13071, published Apr. 2, 1998, which discloses gene therapy methodology for inhibition of primary tumor growth and metastasis by gene transfer of a nucleotide sequence encoding a soluble receptor protein which binds to VEGF.

PCT Publication No. WO 97/44453, published Nov. 27, 1997, in the name of Genentech, Inc., discloses novel chimeric VEGF receptor proteins comprising amino acid sequences derived from the vascular endothelial growth factor (VEGF) receptors Flt1 and KDR, including the murine homologue to the human KDR receptor FLK1, wherein said chimeric VEGF receptor proteins bind to VEGF and antagonize the endothelial cell proliferative and angiogenic activity thereof.

PCT Publication No. WO 97/13787, published Apr. 17, 1997, in the name of Toa Gosei Co., LTD., discloses a low molecular weight VEGF inhibitor usable in the treatment of diseases accompanied by neovascularization such as solid tumors. A polypeptide containing the first immunoglobulin-like domain and the second immunoglobulin-like domain in the extracellular region of a VEGF receptor FLT but not containing the sixth immunoglobulin-like domain and the seventh immunoglobulin-like domain thereof shows a VEGF inhibitory, activity.

Sharifi, J. et al., 1998, The Quarterly Jour. of Nucl. Med. 42:242-249, disclose that because monoclonal antibodies (MAbs) are basic, positively charged proteins, and mammalian cells are negatively charged, the electrostatic interactions between the two can create higher levels of background binding resulting in low tumor to normal organ ratios. To overcome this effect, the investigators attempted to improve MAb clearance by using various methods such as secondary agents as well as chemical and charge modifications of the MAb itself.

Jensen-Pippo, et al., 1996, Pharmaceutical Research 13:102-107, disclose that pegylation of a therapeutic protein, recombinant human granulocyte colony stimulating factor (PEG-G-CSF), results in an increase in stability and in retention of in vivo bioactivity when administered by the intraduodenal route.

Tsutsumi, et al., 1997, Thromb Haemost. 77:168-73, disclose experiments wherein the in vivo thrombopoietic activity of polyethylene glycol-modified interleukin-6 (MPEG-IL-6), in which 54% of the 14 lysine amino groups of IL-6 were coupled with PEG, was compared to that of native IL-6.

Yang, et al., 1995, Cancer 76:687-94, disclose that conjugation of polyethylene glycol to recombinant human interleukin-2 (IL-2) results in a compound, polyethylene glycolmodified IL-2 (PEG-IL-2) that retains the in vitro and in vivo activity of IL-2, but exhibits a markedly prolonged circulating half-life.

R. Duncan and F. Spreafico, Clin. Pharmacokinet. 27: 290-306, 296 (1994) review efforts to improve the plasma 65 half-life of asparaginase by conjugating polyethylene glycol.

PCT International Publication No. WO 99/03996 published Jan. 28, 1999 in the name of Regeneron Pharmaceu-

ticals, Inc. and The Regents of The University of California describes modified human noggin polypeptides having deletions of regions of basic amino acids. The modified human noggin polypeptides are described as retaining biological activity while having reduced affinity for heparin and superior pharmacokinetics in animal sera as compared to the unmodified human noggin.

#### SUMMARY OF THE INVENTION

The present invention is directed to VEGF antagonists with improved pharmacokinetic properties. A preferred embodiment is an isolated nucleic acid molecule encoding a fusion polypeptide capable of binding a VEGF polypeptide comprising (a) a nucleotide sequence encoding a VEGF receptor component operatively linked to (b) a nucleotide sequence encoding a multimerizing component, wherein the VEGF receptor component is the only VEGF receptor component of the fusion polypeptide and wherein the nucleotide sequence of (a) consists essentially of a nucleotide sequence encoding the amino acid sequence of Ig domain 2 of the extracellular domain of a first VEGF receptor and a nucleotide sequence encoding the amino acid sequence of Ig domain 3 of the extracellular domain of a second VEGF receptor

In a further embodiment, the isolated nucleic acid of the first VEGF receptor is Flt1.

In a further embodiment, the isolated nucleic acid of the second VEGF receptor is Flt1.

In yet another embodiment, the isolated nucleic acid of  $_{30}$  the second VEGF receptor is Flt4.

In another preferred embodiment, the nucleotide sequence encoding Ig domain 2 of the extracellular domain of the first VEGF receptor is upstream of the nucleotide sequence encoding Ig domain 3 of the extracellular domain of the 35 second VEGF receptor.

In still another preferred embodiment, the nucleotide sequence encoding Ig domain 2 of the extracellular domain of the first VEGF receptor is downstream of the nucleotide sequence encoding Ig domain 3 of the extracellular domain 40 of the second VEGF receptor.

In a preferred embodiment of the invention, the multimerizing component comprises an immunoglobulin domain.

In another embodiment, the immunoglobulin domain is selected from the group consisting of the Fc domain of IgG, 45 the heavy chain of IgG, and the light chain of IgG.

Preferred embodiments include an isolated nucleic acid molecule comprising a nucleotide sequence encoding a modified Flt1 receptor fusion polypeptide, wherein the coding region of the nucleic acid molecule consists of a nucleotide sequence selected from the group consisting of

- (a) the nucleotide sequence set forth in FIG. 13A-13D (SEQ ID NO:3):
- (b) the nucleotide sequence set forth in FIG. 14A-14C (SEQ ID NO:5):
- (c) the nucleotide sequence set forth in FIG. 15A-15C (SEQ ID NO:7);
- (d) the nucleotide sequence set forth in FIG. **16**A-**16**D (SEQ ID NO:9):
- (e) the nucleotide sequence set forth in FIG. **21**A-**21**C (SEQ 60 ID NO:11):
- (f) the nucleotide sequence set forth in FIG. 22A-22C (SEQ ID NO:13);
- (g) the nucleotide sequence set forth in FIG. 24A-24C (SEQ ID NO:15); and
- (h) a nucleotide sequence which, as a result of the degeneracy of the genetic code, differs from the nucleotide

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sequence of (a), (b), (c), (d), (e), (f), or (g) and which encodes a fusion polypeptide molecule having the biological activity of the modified Flt1 receptor fusion polypeptide.

In a further embodiment of the invention, a fusion polypeptide is encoded by the isolated nucleic acid molecules described above.

A preferred embodiment is a composition capable of binding a VEGF molecule to form a nonfunctional complex comprising a multimer of the fusion polypeptide.

Also preferred is a composition wherein the multimer is a dimer.

In yet another embodiment, the composition is in a carrier. Another embodiment is a vector which comprises the nucleic acid molecules described above, including an expression vector comprising a the nucleic acid molecules described wherein the nucleic acid molecule is operatively linked to an expression control sequence.

Other included embodiments are a host-vector system for the production of a fusion polypeptide which comprises the expression vector, in a suitable host cell; the host-vector system wherein the suitable host cell is a bacterial cell, yeast cell, insect cell, or mammalian cell; the host-vector system wherein the suitable host cell is *E. Coli*; the host-vector system wherein the suitable host cell is a COS cell; the host-vector system wherein the suitable host cell is a CHO cell

Another embodiment of the invention is a method of producing a fusion polypeptide which comprises growing cells of the host-vector system under conditions permitting production of the fusion polypeptide and recovering the fusion polypeptide so produced.

Additional embodiments include a fusion polypeptide encoded by the nucleic acid sequence set forth in FIG. 10A-10D (SEQ ID NO:1) or FIG. 24A-24C (SEQ ID NO:15), which has been modified by acetylation or pegylation wherein the acetylation is accomplished with at least about a 100 fold molar excess of acetylation reagent or wherein acetylation is accomplished with a molar excess of acetylation reagent ranging from at least about a 10 fold molar excess to about a 100 fold molar excess or wherein the pegylation is 10K or 20K PEG.

A preferred embodiment includes a method of decreasing or inhibiting plasma leakage in a mammal comprising administering to the mammal the fusion polypeptide described above, including embodiments wherein the mammal is a human, the fusion polypeptide is acetylated or the fusion polypeptide is pegylated.

A further embodiments is a fusion polypeptide which specifically binds the VEGF receptor ligand VEGF.

A preferred embodiment of the invention is a method of blocking blood vessel growth in a human comprising administering an effective amount of the fusion polypeptide described above.

Also preferred is a method of inhibiting VEGF receptor ligand activity in a mammal comprising administering to the mammal an effective amount of the fusion polypeptide described above.

Preferred embodiments of these methods are wherein the mammal is a human.

Further embodiments of the methods of the invention include attenuation or prevention of tumor growth in a human; attenuation or prevention of edema in a human, especially wherein the edema is brain edema; attenuation or prevention of ascites formation in a human, especially wherein the ascites is ovarian cancer-associated ascites.

Preferred embodiments of the invention include a fusion polypeptide capable of binding a VEGF polypeptide comprising (a) a VEGF receptor component operatively linked to (b) a multimerizing component, wherein the VEGF receptor component is the only VEGF receptor component in the fusion polypeptide and consists essentially of the amino acid sequence of Ig domain 2 of the extracellular domain of a first VEGF receptor and the amino acid sequence of Ig domain 3 of the extracellular domain of a second VEGF receptor.

In a further embodiment of the fusion polypeptide, the 10 first VEGF receptor is Flt1.

In yet a further embodiment of the fusion polypeptide, the second VEGF receptor is Flk1.

Still another embodiment of the fusion polypeptide is one in which the second VEGF receptor is Flt4.

Preferred embodiments include a fusion polypeptide wherein amino acid sequence of Ig domain 2 of the extracellular domain of the first VEGF receptor is upstream of the amino acid sequence of Ig domain 3 of the extracellular domain of the second VEGF receptor and a fusion polypeptide wherein the amino acid sequence of Ig domain 2 of the extracellular domain of the first VEGF receptor is downstream of the amino acid sequence of Ig domain 3 of the extracellular domain of the second VEGF receptor.

In yet another embodiment, the fusion polypeptide multimerizing component comprises an immunoglobulin domain including an embodiment wherein the immunoglobulin domain is selected from the group consisting of the Fc domain of IgG, the heavy chain of IgG, and the light chain of IgG.

Preferred embodiments include a fusion polypeptide comprising an amino acid sequence of a modified Flt1 receptor, wherein the amino acid sequence selected from the group consisting of (a) the amino acid sequence set forth in FIG. 13A-13D (SEQ ID NO:4); (b) the amino acid sequence set forth in FIG. 14A-14C (SEQ ID NO:6); (c) the amino acid sequence set forth in FIG. 15A-15C (SEQ ID NO:8); (d) the amino acid sequence set forth in FIG. 16A-16D (SEQ ID NO:10); (e) the amino acid sequence set forth in FIG. 21A-21C (SEQ ID NO;12); (f) the amino acid sequence set forth in FIG. 22A-22C (SEQ ID NO:14); and (g) the amino acid sequence set forth in FIG. 24A-24C (SEQ ID NO:16).

Another preferred embodiment is a method of decreasing or inhibiting plasma leakage in a mammal comprising administering to the mammal the fusion polypeptide 45 described above.

An alternative preferred embodiment is a method of inhibiting VEGF receptor ligand activity in a mammal comprising administering to the mammal an effective amount of the fusion polypeptide described above.

#### BRIEF DESCRIPTION OF THE FIGURES.

FIG. 1. IEF gel analysis of unmodified and acetylated Flt1(1-3)-Fc proteins. Unmodified Flt1(1-3)-Fc protein is 55 unable to enter the gel due to its >9.3 pl, whereas acetylated Flt1(1-3)-Fc is able to enter the gel and equilibrate at pl 5.2.

FIG. 2. Binding of unmodified Flt1(1-3)-Fc and acetylated Flt1(1-3)-Fc proteins to MATRIGEL® coated plates. Unmodified Flt1(1-3)-Fc proteins binds extensive to extracellular matrix components in Matrigel.RTM., whereas acetylated Flt1(1-3)-Fc does not bind.

FIG. 3. Binding of unmodified Flt1(1-3)-Fc, acetylated Flt1(i-3)-Fc, and pegylated Flt1(1-3)-Fc in a BIACORE<sup>TM</sup>-based assay. Acetylated (columns 13-16), pegylated (columns 17-20), and heparin-treated Flt1(1-3)-Fc (columns 21-24) are each able to completely compete 25 with the

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BIACORE<sup>TM</sup> chip-bound Flt1(1-3)-Fc for VEGF binding as compared to control (columns 1-4) and irrelevant protein (columns 5-8). Unmodified Flt1(1-3)-Fc (columns 5-6) appears to only partially compete with BIACORE<sup>TM</sup> chip-bound Flt1(1-3)-Fc for VEGF binding. However, washing the bound samples with 0.5M NaCl (columns 7-8) results in a binding profile similar to the modified forms of Flt1(1-3)-Fc, indicating that the unmodified protein is exhibiting non-specific binding to the chip that can be eliminated by the salt wash. However, washing the bound samples with 0.5M NaCl (columns 7-8) results in a binding profile similar to the modified forms of Flt1(1-3)-Fc, indicating that the unmodified protein is exhibiting non-specific binding to the chip that can be eliminated by the salt wash.

FIG. 4. Binding of unmodified Flt1(1-3)-Fc, acetylated Flt1(1-3)-Fc, and pegylated Flt1(1-3)-Fc to VEGF in an ELISA-based assay. Both pegylated and acetylated Flt1(1-3)-Fc proteins bind to VEGF with affinities approaching that of unmodified Flt1(1-3)-Fc.

FIG. **5**. Pharmacokinetic profiles of unmodified Flt1(1-3)-Fc, acetylated Flt1(1-3)-Fc, and pegylated Flt1(1-3)-Fc. Balb/c mice (23-28 g) were injected subcutaneously with 4 mg/kg of unmodified, acetylated, or pegylated Flt1(1-3)-Fc. The mice were tail bled at 1, 2, 4, 6, 24 hours, 2 days, and 3 days after injection of protein and the sera were assayed in a standard ELISA-based assay designed to detect Flt1(1-3)-Fc protein. The  $T_{max}$  for all of the Flt1(1-3)-Fc proteins between the 6 hour and 24 hour time points. The  $C_{max}$  for the different proteins was as follows: Unmodified:  $0.06~\mu$ g/ml- $0.15~\mu$ g/ml; acetylated:  $1.5~\mu$ g/ml- $4.0~\mu$ g/ml; and pegylated: approximately  $5~\mu$ g/ml.

FIG. 6A-6B. IEF gel analysis of unmodified and step-acetylated Flt1(1-3)-Fc proteins. Unmodified Flt1(1-3)-Fc protein is unable to enter the gel due to its >9.3 pl, whereas most of the step-acetylated Flt1(1-3)-Fc samples (30-100 fold excess samples) were able to migrate into the gel and equilibrate at pls ranging between 4.55-8.43, depending on the degree of acetylation.

FIG. 7. Binding of unmodified Flt1(1-3)-Fc and step-acetylated Flti (1-3)-Fc proteins to MATRIGEL® coated plates. As with the irrelevant control protein, rTie2-Fc, step-acetylated Flt1(1-3)-Fc (20 and 30 fold excess samples) does not exhibit any binding to the Matrigel coated plate, whereas the non-acetylated Flt1(1-3)-Fc protein exhibits significant binding. The 10 fold excess sample shows reduced binding, but the degree of acetylation is not enough to completely block binding to extracellular matrix components.

FIG. **8**. Binding of unmodified Flt1(1-3)-Fc and step-acetylated Flt1 (1-3)-Fc in a BlACORE<sup>TM</sup>-based assay. At a sub-stoichiometric ratio (0.5 μg/ml of either unmodified Flt1(1-3) or step-acetylated Flt1(1-3)-Fc vs. 0.2 μg/ml VEGF), there is not enough Flt1(1-3)-Fc (either unmodified or step-acetylated) in the solution to completely bind the VEGF. At 1.0 μg/ml, which approximates a 1:1 stoichiometric ratio, the both unmodified and step-acetylated Flt1 (1-3)-Fc are better able to compete for VEGF binding, but there is still insufficient Flt1(1-3)-Fc protein (either unmodified or step-acetylated) to completely saturate the available VEGF. However, at 5.0 μg/ml, which is several times greater than a 1:1 stoichiometric ratio, both the Flt1(1-3)-Fc and the step-acetylated Flt1(1-3)-Fc proteins are able to saturate the VEGF, regardless of the degree of acetylation.

FIG. 9. Pharmacokinetic profiles of unmodified Flt1(1-3)-Fc and step-acetylated Flt1(1-3)-Fc. Balb/c mice (23-28 g) were injected subcutaneously with 4 mg/kg of unmodified or 10, 20, 40, 60 and 100 fold excess samples of step-

as follows: Unmodified Flt1(1-3)-Fc-0.15 µg/ml; 40 fold molar excess acetylated Flt1(1-3)-Fc—1.5 μg/ml; and Mut1: Flt1(1-3<sub>AB</sub>)-Fc  $-0.7 \mu g/ml$ . FIG. 21A-21C. Nucleotide (SEQ ID NO:11) and deduced

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acetylated Flt1(1-3)-Fc (3 mice for unmodified, 10, 20 and 40 fold excess samples and 2 mice for 60 and 100 fold excess samples). The mice were tail bled at 1, 2, 4, 6, 24 hours, 2 days and 3 days after injection. The sera were assayed in an ELISA-based assay designed to detect Flt1 (1-3)-Fc. The  $T_{max}$  for all of the Flt1(1-3)-Fc proteins tested was at the 6 hour time point but the  $C_{max}$  was as follows: Unmodified Flt1(1-3)-Fc: 0.06 µg/ml; 10 fold excess sample:—0.7 µg/ml, 20 fold excess sample—2 µg/ml, 40 fold excess sample—4 µg/ml, 60 fold excess sample—2 10 μg/ml, 100 fold excess sample—1 μg/ml.

amino acid sequence (SEQ ID NO:12) of the modified Flt1 receptor termed Flt1D2.Flk1D3.FcΔC1(a).

FIG. 10A-10D. Nucleic acid (SEQ ID NO:1) and deduced amino acid sequence (SEQ ID NO:2) of Flt1(1-3)-Fc.

FIG. 22A-22C. Nucleotide (SEQ ID NO:13) and deduced amino acid sequence (SEQ ID NO:14) of the modified Flt1 receptor termed Flt1D2.VEGFR3D3.Fc∆C1(a).

FIG. 11. Schematic diagram of the structure of Flt1.

FIG. 23. Extracellular Matrix (ECM) Assay. The results of this assay demonstrate that the Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1(a) proteins are considerably less sticky to the ECM as compared to the Flt1(1-3)-Fc protein.

FIG. 12A and 12B. Hydrophilicity analysis of the amino acid sequences of Ig domain 2 and Ig domain 3 of Flt1. FIG. 13A-13D. Nucleic acid (SEQ ID NO:3) and deduced

amino acid sequence (SEQ ID NO:4) of Mut1: Flt1(1-3<sub>A</sub><sup>B</sup>)-

FIG. 24A-24C. Nucleotide (SEQ ID NO:15) and deduced amino acid sequence (SEQ ID NO:16) of the modified Flt1 receptor termed VEGFR1R2-FcΔC1(a).

FIG. 14A-14C. Nucleic acid (SEQ ID NO:5) and deduced amino acid sequence (SEQ ID NO:6) of Mut2-Flt1(2-3<sub>A</sub><sup>B</sup>)-

FIG. 25A-25C. Phosphorylation assay. At a 1.5 molar excess of either Flt1(1-3)-Fc, Flt1(1-3)-Fc (A40) or transient Flt1D2Flk1D3.FcΔC1(a) there is complete blockage of receptor stimulation by these these is modified Flt1 receptors as compared to control media challenge. In contrast, transient Flt1D2VEGFR3D3.FcΔC1(a) does not show significant blockage at this molar excess, as compared with VEGF positive control challenge. Similar results are seen in FIG. 25B, where the modified Flt receptors are in a 3-fold molar excess to VEGF165 ligand. In FIG. 25C, where the modified Flt1 receptors are in a 6-fold molar excess to VEGF165 ligand, transient Flt1D2VEGFR3D3.FcΔC1(a) can now be shown to be partially blocking VEGF165-induced stimulation of cell-surface receptors.

FIG. 15A-15C. Nucleic acid (SEQ ID NO:7) and deduced amino acid sequence (SEQ ID NO:8) of Mut3: Flt1(2-3)-Fc. 25

> FIG. 26A-26B. Phosphorylation assay. Detection by Western blot of tyrosine phosphorylated VEGFR2(Flk1) by VEGF165 ligand stimulation shows that cell-surface recep-35 tors are not phosphorylated by challenge samples which have VEGF165 preincubated with 1 and 2 fold molar excess (FIG. 26A) or 3 and 4 fold molar excess (FIG. 26B) of either Flt1D2Flk1D3.FcΔC1(a), transient stable Flt1D2Flk1D3.FcΔC1(a), or transient VEGFR1R2-FcΔC1 (a). At all modified Flt1 receptor concentrations tested there is complete binding of VEGF165 ligand during the preincubation, resulting in no detectable stimulation of cellsurface receptors by unbound VEGF165 as compared to control media challenge.

FIG. 16A-16D. Nucleic acid (SEQ ID NO:9) and deduced

FIG. 27. MG/R2 Cell proliferation assay. The following modified Flt receptors  $\bar{\text{Flt1}}(1\text{--}3)\text{-Fc}$ ,  $\bar{\text{Flt1D2}}.\bar{\text{Flk1D3}}.\bar{\text{Fc}}\Delta C1$ (a) and Flt1D2.VEGFR3D3.FcΔC1(a), plus an irrelevant receptor termed Tie2-Fc as a negative control, were titrated from 40 nM to 20 pM and incubated on the cells for 1 hr at 37° C. Human recombinant VEGF165 in defined media was then added to all the wells at a concentration of 1.56 nM. The negative control receptor Tie2-Fc does not block VEGF165induced cell proliferation at any concentration whereas Flt1D2.Flk1D3.FcΔC1(a) blocks 1.56 nM VEGF165 with a half maximal dose of 0.8 nM. Flt1(1-3)-Fc and Flt1D2.VEGFR3D3.FcΔC1(a) are less effective in blocking VEGF165 in this assay with a half maximal dose of ~2 nM. VEGF165 alone gives a reading of 1.2 absorbance units and the background is 0.38 absorbance units.

amino acid sequence (SEQ ID NO:10) of Mut4: Flt1(1- $3_{R\rightarrow N}$ )-Fc.

> FIG. 28. BIACORE™ analysis of Binding Stoichiometry. Binding 20 stoichiometry was calculated as a molar ratio of bound VEGF165 to the immobilized Flt1D2Flk1D3.Fc.ΔC1 (a) or VEGFR1R2-FcΔC1(a), using the conversion factor of 1000 RU equivalent to 1 ng/ml. The results indicated binding stoichiometry of one VEGF165 dimeric molecule per one Flt1D2Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a)

FIG. 17. Binding of unmodified FIt1(1-3)-Fc, basic region deletion mutant Flt1(1-3)-Fc, and Flt1(1-3)<sub>R->N</sub> mutant proteins in a BIACORETM-based assay. At the sub-stoichiometric ratio (0.25 μg/ml Flt1(1-3)-Fc of unmodified, acetylated or genetically modified samples vs. 01. µg/ml VEGF), there is insufficient Flt1(1-3)-Fc protein to block binding of VEGF to the Flt1(1-3)-Fc immobilized on the BIACORETM chip. At 0.5 µg/ml of unmodified, acetylated or genetically modified Flt1(1-3)-Fc proteins, the stoichiometric ratio approximates 1:1 and there is an increased ability to block VEGF binding to the BIACORETM chip. At 1.0 μg/ml of unmodified, acetylated or genetically modified Flt1(1-3)-Fc proteins, which is approximately a 10:1 stoichiometric ratio, the FIt1(1-3)-Fc proteins are able to block binding of VEGF to the BIACORETM chip, but they are not equivalent. Unmodified, acetylated, and Mut1: Flt1 (1-3ΔB)-Fc are essentially equal in their ability to block VEGF binding, whereas Mut4: 45 Flt1(1-3R->N)-Fc is somewhat less efficient at blocking

FIG. 18. Binding of unmodified Flt1(1-3)-Fc, Mut1: Flt1  $(1-3_{AB})$ -Fc, Mut2: Flt1 $(2-3_{AB})$ -Fc, and Flt1(2-3) mutant proteins to Matrigel® coated plates. Unmodified Flt1(1-3)-Fc protein binds avidly to these wells, the Mut3: Flt1(2-3)-Fc protein binds somewhat more weakly, the Mut1: Flt1(1- $3_{AB}$ )-Fc protein binds -more weakly still, and the Mut2:  $\overline{\text{Flt1}}(2-3_{\Delta B})$ -Fc protein shows the best profile, binding more weakly than any of the other mutant proteins. The Mut4: Flt1(1-3<sub>R->N</sub>)-Fc glycosylation mutant protein shows only marginal benefit on the Matrigel assay.

FIG. 19. Binding of unmodified Flt1(1-3)-Fc, Mut1: Flt1  $(1-3_{\Delta B})$ -Fc, Mut2: Flt1 $(2-3_{\Delta B})$ -Fc, and Flt1(2-3) mutant proteins in an ELISA-based assay. At the concentrations tested, unmodified Flt1(1-3)-Fc, Mut1: Flt1(1-3<sub>AB</sub>)-Fc, Mut2: Flt1  $(2-3_{\Delta B})$ -Fc, and Flt1(2-3) mutant proteins bind VEGF simi-

FIG. 20. Pharmacokinetic profiles of unmodified Flt1(1- 65 3)-Fc, Mut1: Flt1(1-3 $_{\Delta B}$ )-Fc, Mut2: Flt1(2-3 $_{\Delta B}$ )-Fc, and Flt1(2-3) mutant proteins. the Cmax for these reagents was

FIG. **29** and FIG. **30**. Size Exclusion Chromatography Stoichiometry. Flt1D2Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a) at a concentration of 1 nM (estimated to be 1000 times higher than the KD of the Flt1D2Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a)/VEGF165 interaction) were mixed 5 with varied concentrations of VEGF165. After incubation, concentrations of the free Flt1D2Flk1D3.FcΔC1(a) in solution were measured. The data shows that the addition of 1 nM VEGF165 into the Flt1D2Flk1D3.FcΔC1(a) solution completely blocks Flt1D2Flk1D3.FcΔC1(a) binding to the VEGF165 surface. This result suggested the binding stoichiometry of one VEGF165 molecule per one Flt1D2Flk1D3.FcΔC1(a) molecule.

FIG. 31. Size Exclusion Chromatography (SEC) under native conditions. Peak #1 represents the Flt1D2Flk1D3.FcΔC1(a)/VEGF165 complex and peak #2 represents unbound VEGF165. Fractions eluted between 1.1 and 1.2 ml were combined and guanidinium hydrochloride (GuHCl)was added to a final concentration 4.5M to dissociate the complex.

FIG. **32**. Size Exclusion Chromatography (SEC) under dissociative conditions. To separate the components of the receptor-ligand complex and to determine their molar ratio, 50 .mu.l of dissociated complex was loaded onto a SUPER-OSE<sup>TM</sup> 12 PC 3.2/30 equilibrated in 6M GuHCl and eluted, 25 Peak #1 represents Flt1D2Flk1D3.FcΔC1(a) and peak #2 represents VEGF165.

FIG. 33, FIG. 34 and FIG. 35. Size Exclusion Chromatography (SEC) with On-Line Light Scattering. Size exclusion chromatography column with a MiniDawn on-line light 30 scattering detector (Wyatt Technology, Santa Barbara, Calif.) and refractive index (RI) detectors (Shimadzu, Kyoto, Japan) was used to determine the molecular weight (MW) of the receptor-ligand complex. As shown in FIG. 33, the elution profile shows two peaks. Peak #1 represents the 35 receptor-ligand complex and peak #2 represents the unbound VEGF165. MW was calculated from LS and RI signals. The same procedure was used to determine MW of the individual components of the receptor-ligand complex. The results of these determinations are as follows: MW of 40 the Flt1D2Flk1D3.FcΔC1(a)/VEGF165 complex at the peak position is 157 300 (FIG. 33), the MW of VEGF165 at the peak position is 44 390 (FIG. 34) and the MW of R1R2 at the peak is 113 300 (FIG. 35).

FIG. 36. Peptide mapping and glycosylation analysis. The 45 disulfide structures and glycosylation sites in Flt1D2.Flk1D3.FcΔC1(a) (SEQ ID NO:12) were determined by a peptide mapping method. There are a total of ten cysteines in Flt1D2.Flk1D3.FcΔC1(a); six of them belong to the Fc region. Cys27 is disulfide bonded to Cys76. Cys121 is disulfide bonded to Cys182. The first two cysteines in the Fc region (Cys211 and Cys214) form an intermolecular disulfide bond with the same two cysteines in another Fc chain. However, it can not be determined whether disulfide bonding is occurring between same cysteines (Cys211 to 55 Cys211, for example) or between Cys211 and Cys211. Cys216 is disulfide bonded to Cys306. Cys 352 is disulfide bonded to Cys410.

There are five possible N-linked glycosylation sites in Flt1D2.Flk1D3.Fc $\Delta$ C1(a) (SEQ ID NO:12) and are found to 60 be glycosylated to varying degrees. Complete glycosylation is observed at Asn33, Asn193, and Asn282. Partial glycosylation is observed on Asn65 and Asn120. Sites of glycosylation are highlighted by underline in the Figure.

FIG. 37. Pharmacokinetics of Flt1(1-3)-Fc (A40), 65 Flt1D2.Flk1D3.FcΔC1(a) and VEGFR1R2-FcΔC1(a). Balb/c mice were injected subcutaneously with 4 mg/kg of

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CHO transiently Flt1(1-3)-Fc (A40), expressed СНО Flt1D2.Flk1D3.FcΔC1(a), stably expressed Flt1D2.Flk1D3.FcΔC1(a), and CHO transiently expressed VEGFR1R2-Fc $\Delta$ C1(a). The mice were tail bled at 1, 2, 4, 6, 24 hrs, 2 days, 3 days and 6 days after injection. The sera were assayed in an ELISA designed to detect Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a). The Tmax for Flt1(1-3)-Fc (A40) was at 6 hrs while the Tmax for the transient and stable Flt1D2.Flk1D3.FcΔC1(a) and the transient VEGFR1R2-Fc∆C1(a) was 24 hrs. The Cmax for Flt1(1-3)-Fc (A40) was 8 µg/ml, For both transients (Flt1D2.Flk1D3.FcΔC1(a) and VEGFR1R2-FcΔC1 (a)) the Cmax was 18 µg/ml and the Cmax for the stable VEGFR1R2-FcΔC1(a) was 30 µg/ml.

FIG. **38**. Pharmacokinetics of Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1 (a). Balb/c mice were injected subcutaneously with 4 mg/kg of Flt1(1-3)-Fc (A40), CHO transiently expressed Flt1D2.Flk1D3.FcΔC1(a) and CHO transiently expressed Flt1D2.VEGFR3D3.FcΔC1(a). The mice were tail bled at 1, 2, 5, 6, 7, 8, 12, 15 and 20 days after injection. The sera were assayed in an ELISA designed to detect Flt1(1-3)-Fc, Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1 (a). Flt1(1-3)-Fc (A40) could no longer be detected in the serum after day 5 whereas Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1(a) are more

FIG. **39**. The Ability of Flt1D2.Flk1D3.FcΔC1(a) to Inhibit HT-1080 Fibrosarcoma Tumor Growth In Vivo. Every other day or 2 times per week treatment of SCID mice with Flt1D2.Flk1D3.FcΔC1(a) at 25 mg/Kg significantly decreases the growth of subcutaneous HT-1080 fibrosarcoma tumors.

FIG. 40. The Ability of Flt1D2.Flk1D3.Fc $\Delta$ C1(a) to Inhibit C6 Glioma Tumor Growth In Vivo. Every other day or 2 times a week treatment of SCID mice with Flt1D2.Flk1D3.Fc $\Delta$ C1(a) significantly decreases the growth of subcutaneous C6 glioma tumors at doses as low as 2.5 mg/Kg.

FIG. 41. VEGF-Induced Uterine Hyperpermeability. PMSG injected subcutaneously (5 IU) to induce ovulation in prepubertal female rats results in a surge of estradiol after 2 days which in turn causes an induction of VEGF in the uterus. This induction results in hyperpermeability of the uterus and an increase in uterine wet. Subcutaneous injection of Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1(a) at 25 mg/kg at 1 hr after PMSG injection results in about a 50% inhibition of the increase in uterine wet weight.

FIG. **42**A-**42**B. Assessment of Corpus Luteum Angiogenesis Using Progesterone as a Readout. PMSG was injected subcutaneously (5 IU) to induce ovulation in prepubertal female rats, resulting in a fully functioning corpus luteum containing a dense network of blood vessels that secretes progesterone into the blood stream to prepare the uterus for implantation. The induction of angiogenesis in the corpus luteum requires VEGF. Resting levels of progesterone are about 5 ng/ml and can be induced to 25-40 ng/ml after PMSG. Subcutaneous injection of Flt1(1-3)-Fc (A40) or Flt1D2.Flk1D3.FcΔC1(a) at 25 mg/kg or 5 mg/kg at 1 hr. after PMSG injection resulted in a complete inhibition of the progesterone induction on day 4.

### DETAILED DESCRIPTION OF THE INVENTION

It has been a long standing problem in the art to produce a receptor based VEGF antagonist that has a pharmacokinetic profile that is appropriate for consideration of the antagonist as a therapeutic candidate. Applicants describe herein, for the first time, a chimeric polypeptide molecule, capable of antagonizing VEGF activity, that exhibits improved pharmacokinetic properties as compared to other the known receptor-based VEGF antagonists. The chimeric polypeptide molecules described herein thus provide for the first time appropriate molecules for use in therapies in which antagonism of VEGF is a desired result.

The present invention provides for novel chimeric 15 polypeptide molecules formed by fusing a modified extracellular ligand binding domain of the Flt1 receptor to the Fc region of IgG.

The extracellular ligand binding domain is defined as the portion of a receptor that, in its native conformation in the 20 cell membrane, is oriented extracellularly where it can contact with its cognate ligand. The extracellular ligand binding domain does not include the hydrophobic amino acids associated with the receptor's transmembrane domain or any amino acids associated with the receptor's intracel- 25 lular domain. Generally, the intracellular or cytoplasmic domain of a receptor is usually composed of positively charged or polar amino acids (i.e. lysine, arginine, histidine, glutamic acid, aspartic acid). The preceding 15-30, predominantly hydrophobic or apolar amino acids (i.e. leucine, 30 valine, isoleucine, and phenylalanine) comprise the transmembrane domain. The extracellular domain comprises the amino acids that precede the hydrophobic transmembrane stretch of amino acids. Usually the transmembrane domain is flanked by positively charged or polar amino acids such as 35 lysine or arginine. von Heijne has published detailed rules that are commonly referred to by skilled artisans when determining which amino acids of a given receptor belong to the extracellular, transmembrane, or intracellular domains (See von Heijne, 1995, BioEssays 17:25-30). Alternatively, 40 websites on the Internet have become available to provide protein chemists with information about making predictions about protein domains.

The present invention provides for the construction of nucleic acid molecules encoding chimeric polypeptide mol- 45 ecules that are inserted into a vector that is able to express the chimeric polypeptide molecules when introduced into an appropriate host cell. Appropriate host cells include, but are not limited to, bacterial cells, yeast cells, insect cells, and mammalian cells. Any of the methods known to one skilled 50 in the art for the insertion of DNA fragments into a vector may be used to construct expression vectors encoding the chimeric polypeptide molecules under control of transcriptional/translational control signals. These methods may include in vitro recombinant DNA and synthetic techniques 5 and in vivo recombinations (genetic recombination) (See Sambrook, et al., Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory; Current Protocols in Molecular Biology, Eds. Ausubel, et al., Greene Publ. Assoc., Wiley-Interscience, NY).

Expression of nucleic acid molecules encoding the chimeric polypeptide molecules may be regulated by a second nucleic acid sequence so that the chimeric polypeptide molecule is expressed in a host transformed with the recombinant DNA molecule. For example, expression of the 65 chimeric polypeptide molecules described herein may be controlled by any promoter/enhancer element known in the

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art. Promoters which may be used to control expression of the chimeric polypeptide molecules include, but are not limited to, the long terminal repeat as described in Squinto et al., (1991, Cell 65:1-20); the SV40 early promoter region (Bernoist and Chambon, 1981, Nature 290:304-310), the CMV promoter, the M-MuLV 5' terminal repeat the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto, et al., 1980, Cell 22:787-797), the herpes thymidine kinase promoter (Wagner et al., 1981, Proc. Natl. Acad. Sci. U.S.A. 78:144-1445), the regulatory sequences of the metallothionine gene (Brinster et al., 1982, Nature 296:39-42); prokaryotic expression vectors such as the β-lactamase promoter (Villa-Kamaroff, et al., 1978, Proc. Natl. Acad. Sci. U.S.A. 75:3727-3731), or the tac promoter (DeBoer, et al., 1983, Proc. Natl. Acad. Sci. U.S.A. 80:21-25, see also "Useful proteins from recombinant bacteria" in Scientific American, 1980, 242:74-94); promoter elements from yeast or other fungi such as the Gal 4 promoter, the ADH (alcohol dehydrogenase) promoter, PGK (phosphoglycerol kinase) promoter, alkaline phosphatase promoter, and the following animal transcriptional control regions, which exhibit tissue specificity and have been utilized in transgenic animals: elastase I gene control region which is active in pancreatic acinar cells (Swift et al., 1984, Cell 38:639-646; Ornitz et al., 1986, Cold Spring Harbor Symp. Quant. Biol. 50:399-409; MacDonald, 1987, Hepatology 7:425-515); insulin gene control region which is active in pancreatic beta cells (Hanahan, 1985, Nature 315:115-122), immunoglobulin gene control region which is active in lymphoid cells (Grosschedl et al., 1984, Cell 38:647-658; Adames et al., 1985, Nature 318:533-538; Alexander et al., 1987, Mol. Cell. Biol. 7:1436-1444), mouse mammary tumor virus control region which is active in testicular, breast, lymphoid and mast cells (Leder et al., 1986, Cell 45:485-495), albumin gene control region which is active in liver (Pinkert et al., 1987, Genes and Devel. 1:268-276), alpha-fetoprotein gene control region which is active in liver (Krumlauf et al., 1985, Mol. Cell. Biol. 5:1639-1648; Hammer et al., 1987, Science 235:53-58); alpha 1-antitrypsin gene control region which is active in the liver (Kelsey et al, 1987, Genes and Devel. 1:161-171), beta-globin gene control region which is active in myeloid cells (Mogram et al., 1985, Nature 315:338-340; Kollias et al., 1986, Cell 46:89-94); myelin basic protein gene control region which is active in oligodendrocyte cells in the brain (Readhead et al., 1987, Cell 48:703-712); myosin light chain-2 gene control region which is active in skeletal muscle (Shani, 1985, Nature 314:283-286), and gonadotropic releasing hormone gene control region which is active in the hypothalamus (Mason et al., 1986, Science 234:1372-1378).

Thus, according to the invention, expression vectors capable of being replicated in a bacterial or eukaryotic host comprising chimeric polypeptide molecule-encoding nucleic acid as described herein, are used to transfect the host and thereby direct expression of such nucleic acids to produce the chimeric polypeptide molecules, which may then be recovered in a biologically active form. As used herein, a biologically active form capable of binding to VEGF.

Expression vectors containing the chimeric nucleic acid molecules described herein can be identified by three general approaches: (a) DNA-DNA hybridization, (b) presence or absence of "marker" gene functions, and (c) expression of inserted sequences. In the first approach, the presence of a foreign gene inserted in an expression vector can be detected by DNA-DNA hybridization using probes comprising

sequences that are homologous to the inserted chimeric polypeptide molecule sequences. In the second approach, the recombinant vector/host system can be identified and selected based upon the presence or absence of certain "marker" gene functions (e.g., thymidine kinase activity, resistance to antibiotics, transformation phenotype, occlusion body formation in baculovirus, etc.) caused by the insertion of foreign genes in the vector. For example, if the chimeric polypeptide molecule DNA sequence is inserted within the marker gene sequence of the vector, recombinants containing the insert can be identified by the absence of the marker gene function. In the third approach, recombinant expression vectors can be identified by assaying the foreign gene product expressed by the recombinant. Such assays can be based, for example, on the physical or functional properties of the chimeric polypeptide molecules.

Cells of the present invention may transiently or, preferably, constitutively and permanently express the chimeric polypeptide molecules.

The chimeric polypeptide molecules may be purified by any technique which allows for the subsequent formation of a stable, biologically active chimeric polypeptide molecule. For example, and not by way of limitation, the factors may be recovered from cells either as soluble proteins or as inclusion bodies, from which they may be extracted quantitatively by 8M guanidinium hydrochloride and dialysis (see, for example, Builder, et al., U.S. Pat. No. 5,663,304). In order to further purify the factors, conventional ion exchange chromatography, hydrophobic interaction chromatography, reverse phase chromatography or gel filtration may be used.

In one embodiment of the invention, the nucleotide sequence encoding the first component is upstream of the nucleotide sequence encoding the second component. In another embodiment of the invention, the nucleotide sequence encoding the first component is downstream of the nucleotide sequence encoding the second component. Further embodiments of the invention may be prepared in which the order of the first, second and third fusion polypeptide components are rearranged. For example, if the nucleotide sequence encoding the first component is designated 1, the nucleotide sequence encoding the second component is designated 2, and the nucleotide sequence of the third component is designated 3, then the order of the components in the isolated nucleic acid of the invention as read from 5' to 3' may be any of the following six combinations: 1,2,3; 1,3,2; 2,1,3; 2,3,1; 3,1,2; or 3,2,1.

The present invention also has diagnostic and therapeutic utilities. In particular embodiments of the invention, methods of detecting aberrancies in the function or expression of the chimeric polypeptide molecules described herein may be used in the diagnosis of disorders. In other embodiments, manipulation of the chimeric polypeptide molecules or agonists or antagonists which bind the chimeric polypeptide molecules may be used in the treatment of diseases. In further embodiments, the chimeric polypeptide molecule is utilized as an agent to block the binding of a binding agent to its target.

By way of example, but not limitation, the method of the invention may be useful in treating clinical conditions that are characterized by vascular permeability, edema or inflammation such as brain edema associated with injury, stroke or tumor; edema associated with inflammatory disorders such 65 as psoriasis or arthritis, including rheumatoid arthritis; asthma; generalized edema associated with burns; ascites

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and pleural effusion associated with tumors, inflammation or trauma; chronic airway inflammation; capillary leak syndrome; sepsis;

kidney disease associated with increased leakage of protein; and eye disorders such as age related macular degeneration and diabetic retinopathy.

An amino acid sequence analysis of Flt1(1-3)-Fc revealed the presence of an unusually high number (46) of the basic amino acid residue lysine. An IEF analysis of Flt1(1-3)-Fc showed that this protein has pl greater than 9.3, confirming the prediction that the protein is very basic. It was hypothesized that the basic nature of Flt1(1-3)-Fc protein was causing it to bind to extracellular matrix components and that this interaction might be the cause of the extremely short detectable circulating serum half-life exhibited by Flt1(1-3)-Fc when injected into mice. In order to test this hypothesis, Flt1(1-3)-Fc protein was acetylated at the lysine residues to reduce the basic charge. Acetylated Flt1(1-3)-Fc was then tested in the assays described infra.

The following examples are offered by way of illustration and not by way of limitation.

#### **EXAMPLES**

#### Example 1

#### Expression of Flt1(1-3)-Fc Protein in CHO K1 Cells

Using standard molecular biology techniques (see e.g., Molecular Cloning, A Laboratory Manual (Sambrook, et al., Cold Spring Harbor Laboratory), Current Protocols in Molecular Biology (Eds. Ausubel, et al., Greene Publ. Assoc., Wiley-Interscience, NY), the gene encoding Flt1(1-3)-Fc was inserted into the expression vector pEE14.1 (Lonza Biologics, plc) at a multiple cloning site downstream of the CMV promoter. CHO K1 cells were transfected with the pEE14.1/Flt1(1-3)-Fc DNA construct using lipofectamine (Gaithersburg, Md.). The transfected CHO K1 cells were grown in glutamine-free DMEM (JRH, Kansas City, Mo.) containing 25 µM methionine sulfoximine (MSX) from Sigma Inc., St. Louis, Mo., and high recombinant protein expressors were obtained by screening the CHO K1 cell supernatants from over 100 hand-picked colony isolates using a standard immunoassay which captures and detects human Fc. The selected hand-picked clone was amplified in the presence of 100 µM MSX followed by a second round of screening of the amplified clones. The highest producing clone had a specific productivity of recombinant Flt1(1-3)-Fc protein of 55 pg/cell/day.

The selected clone was expanded in 225 cm<sup>2</sup> T-flasks (Corning, Acton, Mass.) and then into 8.5 L roller bottles (Corning, Acton, Mass.) using the cell culture media described supra. Cells were removed from the roller bottles by standard trypsinization and put into 3.5 L of suspension medium. The suspension medium is comprised of glutamine-free ISCHO medium (Irvine Scientific, Santa Ana, Calif.) containing 5% fetal bovine serum (FBS from Hyclone Labs, Logan, Utah), 100 μM MSX and GS supplement (JRH Scientific, Kansas City, Mo.) in a 5 L Celligen bioreactor (New Brunswick Scientific, New Brunswick, N.J.) at a density of 0.3×10<sup>6</sup> cells/mL. After the cells reached a density of 3.6×10<sup>6</sup>/mL and were adapted to suspension they were transferred to a 60 L bioreactor (ABEC, Allentown, Pa.) at a density of  $0.5 \times 10^6$  cells/mL in 20 L of ISCHO medium with 5% fetal bovine serum. After two days an additional 20 L of ISCHO+5% fetal bovine serum was added

to the bioreactor. The cells were allowed to grow for an additional two days reaching a final density of  $3.1\times10^6$  cells/mL, and a final Flt1(1-3)-Fc concentration at harvest was 95 mg/L. At harvest the cells were removed by tangential flow filtration using 0.45  $\mu$ m Prostak Filters (Millipore, 5 Inc., Bedford, Mass.).

#### Example 2

### Purification of Flt1(1-3)-Fc Protein Obtained from CHO K1 Cells

Flt1(1-3)-Fc protein was initially purified by affinity chromatography. A Protein A column was used to bind, with high specificity, the Fc portion of the molecule. This affinity-purified protein was then concentrated and passed over a SEC column. The protein was then eluted into the formulation buffer. The following describes these procedures in detail.

#### Materials and Methods

All chemicals were obtained from J. T. Baker, Phillipsburg, N.J. with the exception of PBS, which was obtained as a 10.times. concentrate from Life Technologies, Gaithersburg, Md. Protein A Fast Flow and SUPERDEX<sup>TM</sup> 200 preparation grade resins were obtained from Pharmacia, Piscataway, N.J. Equipment and membranes for protein concentration were obtained from Millipore, Bedford, Mass.

Approximately 40 L of 0.45 µm-filtered CHO conditioned media containing Flt1(1-3)-Fc protein was applied to a 290 mL Protein A Fast Flow column (10 cm diameter) that had been equilibrated with PBS. The column was washed with PBS containing 350 mM NaCl and 0.02% CHAPS and the bound protein was eluted with 20 mM Citric Acid containing 10 mM Na<sub>2</sub>HPO<sub>4</sub>. The single peak in the elution was 35 collected and its pH was raised to neutrality with 1M NaOH. The eluate fractions was concentrated to approximately 9 mg/mL using 10K regenerated cellulose membranes by both tangential flow filtration and by stirred cell concentration. To remove aggregates and other contaminants, the concentrated 40 protein was applied to a column packed with Superdex 200 preparation grade resin (10 cm×55 cm) and run in PBS containing 5% glycerol. The main peak fractions were pooled, sterile filtered, aliquoted and stored at -80° C.

#### Example 3

#### Acetylation of Flt1(1-3)-Fc Protein

Two milligrams of Flt1(1-3)-Fc protein were acetylated as 50 described in the instruction manual provided with the sulfo-NHS-acetate modification kit (Pierce Chemical Co., Rockford, Ill., Cat.#26777).

#### Example 4

#### Characterization of Acetylated Flt1(1-3)-Fc Protein

(a.) IEF analysis: Flt1(1-3)-Fc and acetylated Flt1(1-3)-Fc were analyzed by standard IEF analysis. As shown in FIG. 60 1, Flt1(1-3)-Fc protein is not able to migrate into the gel and therefore must have a pl greater than 9.3, the highest pl in the standard. However, acetylated Flt1(1-3)-Fc is able to migrate into the gel and equilibrate at a pl of approximately 5.2. This result demonstrates that acetylation reduces the net positive charge of the protein and therefore its pl considerably.

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#### (b.) Binding to Extracellular Matrix Components

To test for binding to extracellular matrix components, Flt1(1-3)-Fc and acetylated Flt1(1-3)-Fc where tested in an assay designed to mimic the interaction with extracellular matrix components. In this assay, 96-well tissue culture plates are coated with Matrigel (Biocoat MATRIGEL® matrix thin layer 96 well plate, Catalog #40607, Becton Dickinson Labware, Bedford, Mass.). The plates are incubated with varying concentrations of either Flt1(1-3)-Fc, acetylated Flt1(1-3)-Fc, or rTie2-Fc (an irrelevant control) protein are added to the wells. The plates are incubated for 1-2 hours at either room temperature or 37° C. degrees and then detection of bound proteins is accomplished by adding a secondary alkaline phosphatase-conjugated anti-human Fc antibody to the wells. Finally, alkaline phosphatase substrate is added to the wells and optical density is measured. FIG. 2 shows the results of this assay. Like the irrelevant control protein rTie2-Fc, acetylated Flt1(1-3)-Fc does not exhibit any binding to the Matrigel coated plate, whereas the non-acetylated Flt1(1-3)-Fc protein exhibits significant binding. This result indicates that acetylation of basic amino acid residues is an effective way to interfere with the charge interactions that exist between positively charged proteins and the negatively charged extracellular matrix components they are exposed to in vivo.

#### Example 5

#### Pegylation of Flt1(1-3)-Fc Protein

Although pegylation (polyethylene glycol—PEG) of proteins has been shown to increase their in vivo potency by enhancing stability and bioavailability while minimizing immunogenicity (see references cited supra), it is counterintuitive that pegylating molecules that are too large to be filtered by the kidney glomeruli would improve their pharmacokinetic properties. Without being bound by theory, Applicants postulated that pegylation of the Flt1(1-3)-Fc molecules could improve the pharmacokinetic properties, possibly not by altering the positive charge or by decreasing the pl of Flt1(1-3)-Fc, but rather by physically shielding the positive charges from interacting with the extracellular matrix. Applicants decided to attempt to improve the pharmacokinetic properties of Flt1(1-3)-Fc molecules by attaching strands of 20K PEGs as described infra.

#### 45 Materials and Methods

Purified Flt1(1-3)-Fc derived from CHO cells (see supra) was used in the following pegylation experiments. Functionalized PEGs were obtained from Shearwater Polymers, Huntsville, Ala.; Bicine from Sigma, St Louis, Mo.; Superose 6 column from Pharmacia, Piscataway, N.J.; PBS as a 10x concentrate from Life Technologies, Gaithersburg, Md.; Glycerol from J. T. Baker, Phillipsburg, N.J.; and Bis-Tris precast gels from Novex, Calif.

20K PEG strands functionalized with amine-specific terminal moieties were used in small-scale reaction studies that were set-up to evaluate different reaction conditions in which the PEG:protein stoichiometry was varied. Based on these reactions and the analyses of samples on standard SDS-PAGE, Flt1(1-3)-Fc at a concentration of 1.5 mg/mL was reacted at pH 8.1 with 20K SPA-PEG (PEG succinimidyl propionate) molecules at a PEG-to-Flt1(1-3)-Fc monomer molar ratio of 1:6. The reaction was allowed to proceed at 8° C. overnight. For initial purification, the reaction products were applied to a 10 mm×30 cm Superose 6 column equilibrated with PBS containing 5% Glycerol. The column appeared to separate pegylated Flt1(1-3)-Fc molecules based on the extent of pegylation. Fractions corresponding to what appeared to be primarily mono-pegylated

and di-pegylated dimeric Flt1(1-3)-Fc, as judged by banding patterns on reducing and non-reducing SDS-PAGE gels were pooled. The protein concentration was determined by measuring absorbance at 280 nm. The pegylated Flt1(1-3)-Fc protein was sterile filtered, aliquoted and stored at  $-40^{\circ}$  C

#### Example 6

Binding of Unmodified, Acetylated, and Pegylated Flt1(1-3)-Fc in a BIACORE<sup>TM</sup>-Based Assay

Unmodified, acetylated, and pegylated Flt1(1-3)-Fc proteins were tested in a BIACORETM-based assay to evaluate their ability to bind to the Flt1 ligand, VEGF. In this assay,  $_{15}$ unmodified Flt1(1-3)-Fc protein was immobilized on the surface of a BIACORETM chip (see BIACORETM Instruction Manual, Pharmacia, Inc., Piscataway, N.J., for standard procedures) and a sample containing 0.2 µg/ml VEGF and either unmodified Flt1(1-3)-Fc, acetylated Flt1(1-3)-Fc or 20 pegylated Flt1(1-3)-Fc (each at 25 µg/ml) was passed over the Flt1(1-3)-Fc-coated chip. To minimize the effects of non-specific binding, the bound samples were washed with a 0.5M NaCl wash. In one sample, unmodified Flt1(1-3)-Fc was mixed with heparin. Heparin is a negatively charged 25 molecule and the Flt1(1-3)-Fc protein is a positively charged molecule, so when the two molecules are mixed together, they should interact through their respective charges. This essentially neutralizes Flt1(1-3)-Fc's inherent positive charge making the molecule behave as if it has been chemically or genetically modified so as to reduce its charge and its tendency to bind via charge interactions. As shown in FIG. 3, acetylated (columns 13-16), pegylated (columns 17-20), and heparin-treated Flt1(1-3)-Fc (columns 21-24) are each able to completely compete with the BIACORETM chip-bound Flt1(1-3)-Fc for VEGF binding as compared to control (columns 1-4) and irrelevant protein (columns 5-8). Unmodified Flt1(1-3)-Fc (columns 5-6) appeared to only partially compete with BIACORE™ chip-bound Flt1(1-3)-Fc for VEGF binding. However, washing the bound samples 40 with 0.5M NaCl (columns 7-8) resulted in a binding profile similar to the modified forms of Flt1(1-3)-Fc, indicating that the unmodified protein was exhibiting non-specific binding to the chip that could be eliminated by the salt wash.

#### Example 7

Binding of Unmodified, Acetylated, and Pegylated Flt1(1-3)-Fc in an ELISA-Based Assay

Unmodified, acetylated, and pegylated Flt1(1-3)-Fc proteins were tested in a standard ELISA-based assay to evaluate their ability to bind the Flt1 receptor ligand VEGF. As shown in FIG. 4, both pegylated and acetylated Flt1(1-3)-Fc proteins are capable of binding to VEGF, demonstrating that 55 modifying the protein either by pegylation or acetylation does not destroy its ability to bind its ligand.

#### Example 8

Pharmacokinetic Analysis of Unmodified Flt1(1-3)-Fc, Acetylated Flt1(1-3)-Fc, and Pegylated Flt1(1-3)-Fc

In vivo experiments were designed to assess the pharma-65 cokinetic profiles of unmodified Flt1(1-3)-Fc, acetylated Flt1(1-3)-Fc, and pegylated Flt1(1-3)-Fc protein. Balb/c

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mice (23-28 g; 3 mice/group) were injected subcutaneously with 4 mg/kg of unmodified, acetylated, or pegylated Flt1 (1-3)-Fc. The mice were tail bled at 1, 2, 4, 6, 24 hours, 2 days, and 3 days after injection of protein. The sera were sasayed in a standard ELISA-based assay designed to detect Flt1(1-3)-Fc protein. Briefly, the assay involves coating an ELISA plate with VEGF, binding the unmodified, acetylated, or pegylated Flt1(1-3)-Fc-containing sera, and reporting with an anti-Fc antibody linked to alkaline phosphatase.

10 As shown in FIG. 5, the Tmax for all of the Flt1(1-3)-Fc proteins was between the 6 hour and 24 hour time points. The Cmax for the different proteins was as follows: Unmodified: 0.06 μg/ml-0.15 μg/ml; acetylated: 1.5 μg/ml-4.0 μg/ml; and pegylated: approximately 5 μg/ml.

#### Example 9

#### Step-Acetylation of Flt1(1-3)-Fc

To determine what minimal amount of acetylation is necessary to eliminate binding to extracellular matrix components, an experiment was designed that acetylated the Flt1(1-3)-Fc protein in a step-wise fashion by using increasing amounts of molar excess of acetylation reagent in the acetylation reaction mixture. The range of molar excess was as follows: 0, 10, 20, 30, 40, 50, 60, 70, 80, 90, and 100 moles of acetylation reagent per 1 mole of Flt1(1-3)-Fc monomer. The reactions were performed as detailed in the instruction manual provided with the sulfo-NHS-Acetate modification kit (Pierce Chemical Co., Rockford, Ill., Cat.# 26777).

#### Example 10

#### Characterization of Step-Acetylated Flt1(1-3)-Fc

(a.) IEF analysis Unmodified Flt1(1-3)-Fc and step-acety-lated Flt1(1-3)-Fc proteins were analyzed by standard IEF analysis. As shown in FIG. 6A-6B, unmodified Flt1(1-3)-Fc protein was not able to migrate into the gel due to its extremely high pl (greater than 9.3). However, most of the step-acetylated Flt1(1-3)-Fc samples (30-100 fold molar excess samples) were able to migrate into the gel and equilibrate at pls ranging between 4.55-8.43, depending on the degree of acetylation of the protein. This result demonstrates that acetylation can change the positive charge of the protein in a dose-dependent manner and that reduction of the pl can be controlled by controlling the degree of acetylation.

(b.) Binding of Step-Acetylated Flt1(1-3)-Fc to Extracellular Matrix Components

To test for binding to extracellular matrix components, Flt1(1-3)-Fc and step-acetylated Flt1(1-3)-Fc where tested in the above-described assay designed to mimic the interaction with extracellular matrix components. Varying concentrations of either unmodified Flt1(1-3)-Fc, step-acetylated Flt1(1-3)-Fc (10, 20, and 30 fold molar excess samples), or rTie2-Fc (an irrelevant control) protein were added to the wells. The plates were incubated for 1-2 hours 60 at room temperature or 37° C. and then detection of bound proteins was accomplished by adding a secondary alkaline phosphatase-conjugated anti-human Fc antibody to the wells. Alkaline phosphatase substrate was subsequently added to the wells and optical density measured. FIG. 7 shows the results of this assay. Like the irrelevant control protein rTie2-Fc, step-acetylated Flt1(1-3)-Fc (20 and 30 fold molar excess samples) did not exhibit any significant

binding to the Matrigel coated plate, whereas the non-acetylated Flt1(1-3)-Fc protein exhibited significant binding. The binding is saturable, indicating that the Flt1(1-3)-Fc protein may be binding to specific sites, rather than a more general charge-mediated interaction that might not be saturable. The 10 fold molar excess sample showed reduced binding, but the degree of acetylation was not enough to completely block binding to extracellular matrix components. The 20 fold molar excess and higher samples displayed no detectable binding, despite the fact that by IEF analysis (FIG. 6A and 6B) the lower molar excess samples till had a large net positive charge. This result demonstrates that it is not necessary to completely acetylate all available basic amino acids in order to eliminate binding to extracellular matrix components.

- (c.) Binding of Step-Acetylated Flt1(1-3)-Fc in a Biacore-Based Assay.
- (c.) Binding of Step-Acetylated Flt1(1-3)-Fc in a BIA-  $_{20}$  CORETM-Based Assay

Unmodified and step-acetylated Flt1(1-3)-Fc proteins where tested in a BIACORETM-based assay to evaluate their ability to bind to the Flt1 ligand, VEGF. In this assay, unmodified Flt1(1-3)-Fc protein (0.5, 1.0, or 5.0 µg/ml) was 25 immobilized on the surface of a BIACORETM chip (see BIACORETM Instruction Manual, Pharmacia, Inc., Piscataway, N.J., for standard procedures) and a solution containing 0.2 µg/ml VEGF and either unmodified Flt1(1-3)-Fc (at either 0.5, 1.0, or 5.0 µg/ml) or 10 different step-acetylated 30 Flt1(1-3)-Fc samples (at 0.5, 1.0, or 5.0 µg/ml each) were passed over the Flt1 (1-3)-Fc-coated chip. As shown in FIG. 8, at a sub-stoichiometric ratio (0.5 μg/ml of either unmodified Flt1(1-3) or step-acetylated Flt1(1-3)-Fc vs. 0.2 1 μg/ml VEGF), there is not enough Flt1(1-3)-Fc (either unmodified 35 or step-acetylated) in the solution to completely bind the VEGF. At 1.0 µg/ml, which approximates a 1:1 stoichiometrie ratio, both unmodified and step-acetylated Flt1(1-3)-Fc are better able to compete for VEGF binding, but there is still insufficient Flt1(1-3)-Fc protein (either unmodified or 40 step-acetylated) to completely bind the available VEGF. However, at 5.0 µg/ml, which is several times greater than a 1:1 stoichiometrie ratio, both the Flt1(1-3)-Fc and the step-acetylated Flt1(1-3)-Fc proteins are able to bind the VEGF, regardless of the degree of acetylation. This clearly 45 demonstrates that acetylation does not alter Flt1(1-3)-Fc's ability to bind VEGF.

### (d.) Pharmacokinetic Analysis of Step-Acetylated Flt1(1-3)-Fc

In vivo experiments were designed to assess the pharmacokinetic profiles of unmodified Flt1(1-3)-Fc and stepacetylated Flt1(1-3)-Fc protein. Balb/c mice (23-28 g) were injected subcutaneously with 4 mg/kg of unmodified or 10, 20, 40, 60 and 100 fold molar excess samples of stepacetylated Flt1(1-3)-Fc (3 mice for unmodified, 10, 20 and 40 fold molar excess samples and 2 mice for 60 and 100 fold molar excess samples). The mice were tail bled at 1, 2, 4, 6, 24 hours, 2 days and 3 days after injection. The sera were assayed in an ELISA-based assay designed to detect Flt1 60 (1-3)-Fc (described supra). FIG. 9 details the results of this study. The Tmax for all of the Flt1(1-3)-Fc proteins tested was at the 6 hour time point but the Cmax was as follows: Unmodified Flt1(1-3)-Fc: 0.06 µg/ml; 10 fold molar excess sample:—0.7 μg/ml, 20 fold molar excess sample—2 μg/ml, 40 fold molar excess sample—4 μg/ml, 60 fold molar excess sample—2 μg/ml, 100 fold molar excess sample—1 μg/ml.

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This results demonstrates that acetylation or pegylation of Flt1(1-3)-Fc significantly improves its pharmacokinetic profile

#### Example 11

Construction of Flt1(1-3)-Fc Basic Region Deletion Mutant Designated Mut1: Flt1(1-3<sub>AB</sub>)-Fc

Based on the observation that acetylated Flt1(1-3)-Fc, which has a pl below 6, has much better pharmacokinetics than the highly positive unmodified Flt1(1-3)-Fc (pl>9.3), it was asked whether the difference in pharmacokinetics could be attributed to the net charge of the protein, Which made it stick to negatively charged extracellular matrix components, or whether there were perhaps specific locations on the surface of the Flt1(1-3)-Fc protein that constituted specific binding sites for extracellular matrix components. For example, many proteins are known to have heparin binding sites, often consisting of a cluster of basic residues. Sometimes these residues are found in a cluster on the primary sequence of the protein; some of the literature has identified "consensus sequences" for such heparin binding sites (see for example Hileman, et al., 1998, Bioessays 20(2):156-67). In other cases, the known crystal structure of a protein reveals a cluster of positively charged residues on the surface of a protein, but the residues come from different regions of the primary sequence and are only brought together when the protein folds into its tertiary structure. Thus it is difficult to deduce whether an isolated amino acid residue forms part of a cluster of basic residues on the surface of the protein. However, if there is a cluster of positively charged amino acid residues in the primary sequence, it is not unreasonable to surmise that the residues are spatially close to one another and might therefore be part of an extracellular matrix component binding site. Flt1 receptor. has been studied extensively and various domains have been described (see for example Tanaka et al., 1997, Jpn. J. Cancer Res 88:867-876). Referring to the nucleic acid and amino acid sequence set forth in FIG. 10A-10D of this application, one can identify the signal sequence for secretion which is located at the beginning of the sequence and extends to the glycine coded for by nucleotides 76-78. The mature protein begins with Ser-Lys-Leu-Lys, starting at nucleotide 79 of the nucleic acid sequence. Flt1 Ig domain 1 extends from nucleotide 79 to 393, ending with the amino acids Ser-Asp-Thr. Flt1 Ig domain 2 extends from nucleotide 394 to 687 (encoding Gly-Arg-Pro to Asn-Thr-Ile), and Flt1 Ig domain 3 extends from nucleotides 688 to 996 (encoding Ile-Asp-Val to Asp-Lys-Ala). There is a bridging amino acid sequence, Gly-Pro-Gly, encoded by nucleotides 997-1005, followed by the nucleotide sequence encoding human Fc (nucleotides 1006-1701 or amino acids Glu-Pro-Lys to Pro-Glv-Lvs-stop).

A more detailed analysis of the Flt1 amino acid sequence reveals that there is a cluster, namely, amino acid residues 272-281 (KNKRASVRR) of FIG. 10A-10D, in which 6 out of 10 amino acid residues are basic. This sequence is located in Flt1 Ig domain 3 of the receptor (see FIG. 11), which is not itself essential for binding of VEGF ligand, but which confers a higher affinity binding to ligand. An alignment of the sequence of Ig domain 3 with that of Ig domain 2 reveals that in this region, there is very poor alignment between the two Ig domains, and that there are about 10 additional amino acids in Ig domain 3. An analysis of the hydrophilicity profiles MACVECTOR™ computer software) of these two domains clearly indicates the presence of a hydrophilic

region in the protein (FIG. 12A-12B). These observations raised the possibility that the actual three dimensional conformation of Flt1 Ig domain 3 allowed for some type of protrusion that is not in Flt1 Ig domain 2. To test this hypothesis, the 10 additional amino acids were deleted and the resulting protein was tested to see whether the deletion would affect the pharmacokinetics favorably without seriously compromising the affinity of the receptor for VEGF. This DNA construct, which was constructed using standard molecular biology techniques (see e.g., Molecular Cloning, A Laboratory Manual (Sambrook, F et al., Cold Spring Harbor Laboratory), Current Protocols in Molecular Biology (Eds. Ausubel, et al., Greene Publ. Assoc., Wiley-Interscience, N.Y.) in the mammalian expression vector pMT21, (Genetics Institute, Inc., Cambridge, Mass.), is referred to as Mut1: Flt1(1-3 $\Delta$ B)-Fc. The Mut1: Flt1(1-3 $\Delta$ B)-Fc construct was derived from Flt1(1-3)-Fc by deletion of nucleotides 814-843 (set forth in FIG. 10A-10D), which deletes the highly basic 10-amino acid residue sequence Lys-Asn-Lys- 20 Arg-Ala-Ser-Val-Arg-Arg-Arg from Flt1 Ig domain 3.

The final DNA construct was sequence-verified using an ABI 373A DNA sequencer and Taq Dideoxy Terminator Cycle Sequencing Kit (Applied Biosystems, Inc., Foster City, Calif.). The sequence of Mut1:  $Flt1(1-3_{\Delta B})$ -Fc is set forth in FIG. 13A-13D.

#### Example 12

Construction of Flt1(1-3)-Fc Basic Region Deletion Mutant Designated Mut2: Flt1(2-3<sub>AB</sub>)-Fc

A second deletion mutant construct, designated Mut2: Flt1(2-3<sub>AB</sub>)-Fc, was derived from the Mut1: Flt1(1-3<sub>AB</sub>)-Fc construct by deletion of Flt1 Ig domain 1 encoded by nucleotides 79-393 (see FIG. 10A-10D); for convenience, nucleotides 73-78 (TCA GGT) were changed to TCC GGA. This introduced a restriction site (BspE1) without altering the associated amino acid sequence, Ser-Gly. This DNA construct, which was constructed using standard molecular biology techniques (see e.g., Molecular Cloning, A Laboratory Manual (Sambrook, et al., Cold Spring Harbor Laboratory), Current Protocols in Molecular Biology (Eds. 45 Ausubel, et al., Greene Publ. Assoc., Wiley-Interscience, NY) in the mammalian expression vector pMT21 (Genetics Institute, Inc., Cambridge, Mass.), was also sequence-verified using an ABI 373A DNA sequencer and Taq Dideoxy Terminator Cycle Sequencing Kit (Applied Biosystems, 50 Inc., Foster City, Calif.). The sequence of Mut2: Flt1(2-3 $_{AB}$ )-Fc is set forth in FIG. 14A-14C.

#### Example 13

Construction of Flt1(1-3)-Fc Deletion Mutant Designated Mut3: Flt1(2-3)-Fc

A third deletion mutate construct, designated Mut3: Flt1 (2-3)-Fc, was constructed the same way as the Mut2: Flt1  $(2-3_{\Delta B})$ -Fc construct, except that Flt1 Ig domain 3 was left intact (the basic region amino acids were not deleted). The construct was constructed using standard molecular biology techniques and the final construct was sequence-verified as described supra. The sequence of Mut3: Flt1(2-3)-Fc is set forth in FIG. **15A-15**C.

#### Example 14

Construction of Flt(1-3)-Fc Basic Region N-glycosylation Mutant Designated Mut4: Flt1(1-3<sub>R->N</sub>)-Fc

A final construct was made in which a N-glycosylation site was introduced into the middle of the basic region of Flt1 Ig domain 3. This construct was designated Mut4: Flt1(1-3 $_{R->N}$ )-Fc and was made by changing nucleotides 824-825 from GA to AC, consequently changing the coded Arg residue (AGA) into an Asn residue (AAC) (see FIG. 10A-10D). The resulting amino acid sequence is therefore changed from Arg-Ala-Ser to Asn-Ala-Ser, which matches the canonical signal (Asn-Xxx-Ser/Thr) for the addition of a N-glycosylation site at the Asn residue. The sequence of Mut4: Flt1(1-3 $_{R->N}$ )-Fc is set forth in FIG. 16A-16D.

#### Example 15

Characterization of Acetylated Flt1(1-3)-Fc Mut1: Flt1(1-3<sub>AB</sub>)-Fc, and Mut4: Flt1(1-3<sub>R->N</sub>)-Fc mutants

#### (a.) Binding to Extracellular Matrix Components

To determine whether the three modified proteins were more or less likely to have improved pharmacokinetic properties, Matrigel coated 96-well dishes (as described supra ) were incubated with varying concentrations of the mutant proteins and detected with anti-human Fc/alkalinephosphatase conjugated antibodies. As shown in FIG. 18, this experiment showed that while the unmodified Flt1(1-3)-Fc protein could bind avidly to these wells, the Mut3: Flt1(2-3)-Fc protein bound somewhat more weakly, the Mut1: Flt1(1-3 $_{\Delta B}$ )-Fc protein bound more weakly still, and the Mut2: Flt1(2-3<sub>AB</sub>)-Fc protein showed the best profile, binding more weakly than any of the other mutant proteins. The Mut4:  $Flt1(1-3_{R->N})$ -Fc glycosylation mutant protein showed only marginal benefit on the Matrigel assay. These results confirm the hypothesis that a linear sequence of positive amino acids can be deleted from the primary sequence resulting in a decrease in charge interaction with extracellular matrix components.

# (b.) Binding of Mut1: Flt1(1-3 $\Delta$ B)-Fc and Mut4: Flt1(1-3 $_{R->N}$ ) Fc in a BIACORETM-Based Assay.

Unmodified and acetylated Flt1(1-3)-Fc and genetically modified Mut1: Flt1(1-3 $\Delta$ B)-Fc and Mut4: Flt1(1-3<sub>R->N</sub>)-Fc proteins where tested in a BIACORETM-based assay to evaluate their ability to bind to the Flt1 ligand, VEGF. In this assay, unmodified Flt1(1-3)-Fc protein (0.25, 0.5, or 1.0 μg/ml) was immobilized on the surface of a BIACORETM chip (see BIACORETM Instruction Manual, Pharmacia, Inc., Piscataway, N.J., for standard procedures) and a solution containing 0.1 µg/ml VEGF and either purified or COS cell supernatant containing unmodified Flt1(1-3)-Fc (at approxi-55 mately (0.25, 0.5, or 1.0 μg/ml), purified acetylated Flt1(1-3)-Fc (at (0.25 0.5, or 1.0  $\mu g/ml$ ), COS cell supernatant containing Mut1: FIt1 (1-3 $\Delta$ B)-Fc. (at approximately (0.25, 0.5, or 1.0 μg/ml), or COS cell supernatant containing Mut4: Flt1(1-3<sub>R->N</sub>)-Fc (at approximately (0.25, 0.5, or 1.0  $\mu$ g/ml) were passed over the Flt1(1-3)-Fc-coated chip. As shown in FIG. 17, at the sub-stoichiometric ratio (0.25 μg/ml Flt1(1-3)-Fc of unmodified, acetylated or genetically modified samples vs. 01. µg/ml VEGF), there is insufficient Flt1 (1-3)-Fc protein to block binding of VEGF to the Flt1(1-3)-Fc immobilized on the BIACORETM chip. At 0.5 μg/ml of unmodified, acetylated or genetically modified Flt1(1-3)-Fc proteins, the stoichiometric ratio approximates 1:1 and there

is an increased ability to block VEGF binding to the BIA-CORETM chip. At 1.0 µg/ml of unmodified, acetylated or genetically modified Flt1(1-3)-Fc proteins, which is approximately a 10:1 stoichiometric ratio, the Flt1(1-3)-Fc proteins are able to block binding of VEGF to the BIA-CORETM chip, but they are not equivalent. Unmodified, acetylated, and Mut1: Flt1(1-3 $\Delta$ B)-Fc are essentially equal in their ability to block VEGF binding, whereas Mut4: Flt1(1-3 $_{R-N}$ )-Fc is somewhat less efficient at blocking binding. These results confirm the hypothesis that it is possible to reduce the non-specific binding of a positively charged molecule by genetically removing a linear sequence of predominantly negatively charged amino acids.

(c.) Binding of Mut1: Flt1(1-3 $_{\Delta B}$ )-Fc, Mut2: Flt1(2-3 $_{\Delta B}$ )-Fc, Mut3: Flt1(2-3)-Fc, and in an ELISA-Based Assay.

To determine whether the three mutant proteins could bind the Flt1 ligand VEGF, binding experiments were done in which 96-well plates coated with VEGF were incubated with varying concentrations of the respective mutant protein, and after washing, the amount bound was detected by incubating with an alkaline phosphatase conjugated antihuman Fc antibody and quantitated colorimetrically by the addition of an appropriate alkaline phosphatase substrate. As shown in FIG. 19, this experiment showed that all the mutant proteins could bind VEGF similarly, at the concentrations tested.

#### Example 16

Pharmacokinetic Analysis of Acetylated Flt1(1-3)-Fc, Mut1: Flt1(1-3 $_{\Delta B}$ )-Fc, and Unmodified Flt1(1-3)-Fc

In vivo experiments were designed to assess the pharmacokinetic profiles of unmodified Flt1(1-3)-Fc, Mut1: Flt1(1- $3_{AB}$ )-Fc, and 40 fold molar excess acetylated Flt1(1-3)-Fc protein. Balb/c mice (25-30 g) were injected subcutaneously with 4 mg/kg of unmodified Flt1(1-3)-Fc, 40 fold molar excess acetylated Flt1(1-3)-Fc, and Mut1: Flt1(1-3<sub>AB</sub>)-Fc 40 proteins (4 mice each). These mice were tail bled at 1, 2, 4, 6, 24 hours, 2 days, 3 days, and 5 days after injection. The sera were assayed in an ELISA designed to detect Flt1(1-3)-Fc protein which involves coating an ELISA plate with VEGF, binding the Flt1(1-3)-Fc and reporting with an 45 anti-Fc antibody linked to alkaline phosphatase. As shown in FIG. 20, the Cmax for these reagents was as follows: Unmodified Flt1(1-3)-Fc—0.15 μg/ml; 40 fold molar excess acetylated Flt1(1-3)-Fc—1.5  $\mu$ g/ml; and Mut1: Flt1(1-3 $_{\Delta B}$ )-Fc $\longrightarrow$ 0.7 µg/ml.

#### Example 17

#### Modified Flt1 Receptor Vector Construction

The rationale for constructing modified versions of the Flt1 receptor (also known as VEGFR1) was based on the observation that the protein sequence of Flt1 was highly basic, and was therefore likely to stick to extracellular matrix (ECM). The highly basic nature of Flt1 probably 60 explains why unmodified Flt1(1-3)-Fc (described supra) has poor pharmacokinetics that make it difficult to use as a therapeutic agent. As described supra, the chemically modified form of 40 fold molar excess acetylated Flt1(1-3)-Fc, hereinafter termed A40 exhibited a greatly improved pharmacokinetic (PK) profile over the non-acetylated Flt1(1-3)-Fc. Therefore, attempts were made to engineer DNA mol-

ecules that could be used to recombinantly express modified forms of a Flt1 receptor molecule that would possess the improved PK profile exhibited by A40 and still maintain the ability to bind tightly to VEGF.

It is known in the literature that the first Ig domain of Flt1 (which has a net charge of +5 at neutral pH) is not essential for tight binding to VEGF, so this domain was deleted. The third Ig domain (having a net charge of +11) is not essential for binding, but confers higher affinity for VEGF than the second Ig domain, so instead of deleting it entirely, it was replaced with the equivalent domains of the Flt1 receptor relatives Flk1 (also known as VEGFR2) and Flt4 (also known as VEGFR3). These chimeric molecules (denoted R1R2 (Flt1.D2.Flk1D3.FcΔC1(a) and VEGFR1R2-FcΔC1 and R1R3 (Flt1D2.VEGFR3D3-FcΔC1(a) VEGFR1R3-FcΔC1(a) respectively, wherein R1 and Flk1D2=Ig domain 2 of Flt1 (VEGFR1); R2 and Flk1D3=Ig domain 3 of Flk1 (VEGFR2); and R3 and VEGFR3D3=Ig domain 3 of Flt4 (VEGFR3)) were much less sticky to ECM, as judged by an in vitro ECM binding assay as described infra, had greatly improved PK as described infra. In addition, these molecules were able to bind VEGF tightly as described infra and block phosphorylation of the native Flk1 receptor expressed in endothelial cells as described infra.

(a) Construction of the Expression Plasmid pFlt1D2.Flk1D3.FcΔC1(a)

Expression plasmids pMT21 .Flt1(1-3).Fc (6519bp) and pMT21.Flk-1(1-3).Fc (5230bp) are plasmids that encode ampicillin resistance and Fc-tagged versions of Ig domains 1-3 of human Flt1 and human Flk1, respectively. These plasmids were used to construct a DNA fragment consisting of a fusion of Ig domain 2 of Flt1 with Ig domain 3 of Flk1, using PCR amplification of the respective Ig domains followed by further rounds of PCR to achieve fusion of the two domains into a single fragment. For Ig domain 2 of Flt1, the 5' and 3' amplification primers were as follows:

```
5': bsp/flt1D2
(5'-GACTAGCAGTCCGGAGGTAGACCTTTCGTAGAGATG-3')
3': Flt1D2-Flk1D3.as
(5'-CGGACTCAGAACCACATCTATGATTGTATTGGT-3')
```

The 5' amplification primer encodes a BspE1 restriction enzyme site upstream of Ig domain 2 of Flt1, defined by the amino acid sequence GRPFVEM (corresponding to amino acids 27-33 of FIG. 21A-21C). The 3' primer encodes the reverse complement of the 3' end of Flt1 Ig domain 2 fused directly to the 5' beginning of Flk1 Ig domain 3, with the fusion point defined as TIID of Flt1 (corresponding to amino acids 123-126 of FIG. 21A-21C) and continuing into VVLS (corresponding to amino acids 127-130 of FIG. 21A-21C) of Flk1.

For Ig domain 3 of Flk1, the 5' and 3' amplification primers were as follows:

```
5': Flt1D2-Flk1D3.s
(5'-ACAATCATAGATGTGGTTCTGAGTCCGTCTCATGG-3')

3': Flk1D3/apa/srf.as
(5'GATAATGCCCGGGCCCTTTTCATGGACCCTGACAAATG-3')
```

The 5' amplification primer encodes the end of Flt1 Ig domain 2 fused directly to the beginning of Flk1 Ig domain 3, as described above. The 3' amplification primer encodes the end of Flk1 Ig domain 3, defined by the amino acids

VRVHEK (corresponding to amino acids 223-228 of FIG. 21A-21C), followed by a bridging sequence that includes a recognition sequence for the restriction enzyme Srf1, and encodes the amino acids GPG. The bridging sequence corresponds to amino acids 229-231 of FIG. 21A-21C.

After a round of PCR amplification to produce the individual domains, the products were combined in a tube and subjected to a further round of PCR with the primers bsp/flt1D2 and Flk1D3/apa/srf.as (described supra) to produce the fusion product. This PCR product was subsequently digested with the restriction enzymes BspEI and SmaI and the resulting 614bp fragment was subcloned into the BspEI to SrfI restriction sites of the vector pMT21/ΔB2.Fc, to create the plasmid pMT21/Flt1D2.Flk1D3.Fc. The nucleotide sequence of the Flt1D2-Flk1D3 gene fusion insert was verified by standard sequence analysis. This plasmid was then digested with the restriction enzymes EcoRI and SrfI and the resulting 702bp fragment was transferred into the EcoRI to SrfI restriction sites of the plasmid pFlt1(1-3)B2-FcΔC1(a) to produce the plasmid pFlt1D2.Flk1D3.FcΔC1 (a). The complete DNA and deduced amino acid sequences of the Flt1D2.Flk1D3.FcΔC1(a) chimeric molecule is set forth in FIG. 21A-21C.

#### Construction of the Expression pFlt1D2VEGFR3D3FcΔC1(a)

The expression plasmid pMT21.Flt1(1-3).Fc (6519bp) encodes ampicillin resistance and an Fc-tagged version of Ig domains 1-3 of human Flt1 receptor. This plasmid was used to produce a DNA fragment containing Ig domain 2 of Flt1 by PCR. RNA from the cell line HEL921.7 was used to produce Ig domain 3 of Flk1, using standard RT-PCR methodology. A further round of PCR amplification was used to achieve fusion of the two Ig domains into a single fused fragment. For Ig domain 2 of Flt1, the 5' and 3'  $^{35}$ amplification primers were as follows:

```
5': bsp/flt1D2
(5'-GACTAGCAGTCCGGAGGTAGACCTTTCGTAGAGATG-3')
3': Flt1D2.VEGFR3D3.as
(TTCCTGGGCAACAGCTGGATATCTATGATTGTATTGGT)
```

The 5' amplification primer encodes a BspE1 restriction site upstream of Ig domain 2 of Flt1, defined by the amino 45 acid sequence GRPFVEM (corresponding to amino acids 27-33 of FIG. 22A-22C). The 3' amplification primer encodes the reverse complement of the end of Flt1 Ig domain 2 fused directly to the beginning of VEGFR3 Ig (corresponding to amino acids 123-126 of FIG. 22A-22C) and continuing into IQLL of VEGFR3 (corresponding to amino acids 127-130 of FIG. 22A-22C)

For Ig domain 3 of VEGFR3, the 5' and 3' primers used for RT-PCR were as follows:

```
5 ' R3D3 s
(ATCCAGCTGTTGCCCAGGAAGTCGCTGGAGCTGCTGGTA)
3': R3D3.as
(\verb|ATTTCATGCACAATGACCTCGGTGCTCTCCCGAAATCG|)
```

Both the 5' and 3' amplification primers match the sequence of VEGFR3. The 296bp amplification product of this RT-PCR reaction was isolated by standard techniques 65 and subjected to a second round of PCR to add suitable sequences to allow for fusion of the Flt1D2 with the Flk1D3

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domains and fusion of the Flk1D3 and Fc domains via a GPG bridge (see below). The amplification primers were as follows:

5':Flt1D2.VEGFR3D3.s  $(\verb|TCATAGATATCCAGCTGTTGCCCAGGAAGTCGCTGGAG|)$ 

3': VEGFR3D3/srf.as (GATAATGCCCGGGCCATTTTCATGCACAATGACCTCGGT)

The 5' amplification primer encodes the 3' end of Flt1 Ig domain 2 fused directly to the beginning (5' end) of VEGFR3 Ig domain 3, as described above. The 3' amplification primer encodes the 3' end of VEGFR3 Ig domain 3, defined by the amino acids VIVHEN (corresponding to amino acids 221-226 of FIG. 22A-22C), followed by a bridging sequence that includes a recognition sequence for Srf1, and encodes the amino acids GPG. The bridging sequence corresponds to amino acids 227-229 of FIG. 22A-22C.

After one round (for Flt1 Ig domain 2) or two rounds (for Flt4 Ig domain 3) of PCR to produce the individual Ig domains, the PCR products were combined in a tube and Plasmid 25 subjected to a further round of PCR amplification with the amplification primers bsp/flt1D2 and VEGFR3D3/srf.as described supra, to produce the fusion product. This PCR product was subsequently digested with the restriction enzymes BspEI and SmaI and the resulting 625bp fragment was subcloned into the BspEI to SrfI restriction sites of the vector pMT21/Flt1\DeltaB2.Fc (described supra), to create the plasmid pMT21/Flt1D2.VEGFR3D3.Fc. The sequence of the Flt1D2-VEGFR3D3 gene fusion insert was verified by standard sequence analysis. This plasmid was then digested with the restriction enzymes EcoRI and SrfI and the resulting 693bp fragment was subcloned into the EcoRI to SrfI restriction sites of the plasmid pFlt1(1-3)ΔB2-FcΔC1(a) to produce the plasmid designated pFlt1D2.VEGFR3D3.FcΔC1(a). The complete DNA deduced amino acid sequence of the Flt1D2.VEGFR3D3.FcΔC1(a) chimeric molecule is set forth in FIG. 22A-22C.

#### Example 18

Extracellular Matrix Binding (ECM) Binding Assay

ECM-coated plates (Becton Dickinson catalog #35-4607) domain 3, with the fusion point defined as TIID of Flt1 50 were rehydrated with warm DME supplemented with glutamine (2 mM), 100 U penicillin, 100 U streptomycin, and 10% BCS for at least 1 hr. before adding samples. The plates were then incubated for 1 hr. at room temperature with varying concentrations of Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1(a) starting at 10 nM with subsequent 2-fold dilutions in PBS plus 10% BCS. The plates were then washed 3 times with PBS plus 0.1% Triton-X and incubated with alkaline phosphatase-conjugated anti-human Fc antibody (Promega, 1:4000 in PBS plus 10% BCS) for 1 60 hr. at room temperature. The plates were then washed 4 times with PBS 0.1% Triton-X and alkaline phosphatase buffer/pNPP solution (Sigma) was added for color development. Plates were read at I=405-570 nm. The results of this experiment are shown in FIG. 23 and demonstrate that the Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1 (a) proteins are considerably less sticky to the ECM as compared to the Flt1(1-3)-Fc protein.

### Transient Expression of pFlt1D2.Flk1D3.FcΔC1(a) in CHO-K1 (E1A) cells

A large scale (2 L) culture of *E. coli* DH10B cells carrying the pFlt1D2.Flk1D3.Fc $\Delta$ C1(a) plasmid described supra in Example 17(a) was grown overnight in Terrific Broth (TB) plus 100 µg/ml ampicillin. The next day, the plasmid DNA was extracted using a QIAgen ENDOFREETM Megaprep kit following the manufacturer's protocol. The concentration of the purified plasmid DNA was determined by standard techniques using a UV spectrophotometer and fluorometer. The plasmid DNA was verified by standard restriction enzyme digestion of aliquots using the restriction enzymes EcoRI plus NotI and Asel. All restriction enzyme digest fragments corresponded to the predicted sizes when analyzed on a 1% agarose gel.

Forty 15 cm petri plates were seeded with CHO-K1/E1A cells at a density of 4×106 cells/plate. Plating media was Gibco Ham's F-12 supplemented with 10% HYCLONETM 20 Fetal Bovine Serum (FBS), 100 U penicillin/100 U streptomycin and glutamine (2 mM). The following day each plate of cells was transfected with 6 ug of the pFlt1D2.Flk1D3.FcΔC1(a) plasmid DNA using Gibco Optimem and Gibco Lipofectamine in 12 ml volume, following 25 the manufacturer's protocol. Four hours after adding the transfection mix to the cells, 12 ml/plate of Optimem supplemented with 10% FBS was added. Plates were incubated at 37° C. in a 5% CO<sub>2</sub> incubator overnight. The following day the media was removed from each plate and 25 ml expression media (Gibco CHO-S-SFM II supplemented with glutamine (2 mM) and 1 mM sodium butyrate) was added. The plates were incubated at 37° C. for 3 days. After 3 days of incubation, the media was aspirated from each plate and centrifuged at 400 rpm in a swinging bucket rotor to pellet cells. The supernatant was decanted into sterile 1 L bottles and purification of the expressed protein was performed as described infra.

#### Example 20

### Construction pVEGFR1R2-FcΔC1C(a) Expression Vector

The pVEGFR1R2.FcΔC1(a) expression plasmid was constructed by insertion of DNA encoding amino acids SDT (corresponding to amino acids 27-29 of FIG. 24A-24C) between Flt1d2-Flk1d3-FcΔC1(a) amino acids 26 and 27 of FIG. 21A-21C (GG) and removal of DNA encoding amino acids GPG corresponding to amino acids 229-231 of Figure. The SDT amino acid sequence is native to the Flt1 receptor and was added back in to decrease the likelihood of heterogeneous N-terminal processing. The GPG (bridging sequence) was removed so that the Flt1 and Flk1 Ig domains were fused directly to one another. The complete DNA and deduced amino acid sequences of the pVEGFR1R2.FcΔC1 (a) chimeric molecule is set forth in FIG. 24A-24C.

#### Example 21

#### Cell Culture Process Used to Produce Modified Flt1 Receptors

### (a) Cell Culture Process Used to Produce Flt1D2.Flk1D3.Fc $\Delta$ C1(a)

The process for production of Flt1D2.Flk1D3.FcΔC1(a) 65 protein using the expression plasmid pFlt1D2.Flk1D3.FcΔC1(a) described supra in Example 1

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involves suspension culture of recombinant Chinese hamster ovary (CHO K1/E1A) cells which constitutively express the protein product. The cells are grown in bioreactors and the protein product is isolated and purified by affinity and size exclusion chromatography. The process is provided in greater detail below.

#### Cell Expansion

Two confluent T-225 cm² flasks containing the Flt1D2.Flk1D3.Fc $\Delta$ C1(a) expressing cell line were expanded by passaging cells into eight T-225 cm² flasks in medium (GMEM +10% serum, GIBCO) and incubated at 37° C. and 5% CO $_2$ . When the flasks approached confluence (approximately 3 to 4 days) the cells were detached using trypsin. Fresh medium was added to protect the cells from further exposure to the trypsin. The cells were centrifuged and resuspended in fresh medium then transferred to eight 850 cm² roller bottles and incubated at 37° C. and 5% CO $_2$  until confluent.

#### Suspension Culture in Bioreactors

Cells grown in roller bottles were trypsinized to detach them from the surface and washed with suspension culture medium. The cells are aseptically transferred to a 5 L bioreactor (New Brunswick Celligen Plus) where the cells are grown in 3.5 L of suspension culture. The suspension culture medium was a glutamine-free low glucose modification of IS-CHO (Irvine Scientific) to which 5% fetal bovine serum (Hyclone), GS supplement (Life Technologies) and 25 µM methionine sulfoximine (Sigma) was added. The pH was controlled at 7.2 by addition of carbon dioxide to the inlet gas or by addition of a liquid solution of sodium carbonate to the bioreactor. Dissolved oxygen level was maintained at 30% of saturation by addition of oxygen or nitrogen to the inlet gas and temperature controlled at 37° C. When a density of  $4\times10^6$  cells/mL was reached the cells were transferred to a 40 L bioreactor containing the same medium and setpoints for controlling the bioreactor. The temperature setpoint was reduced to 34° C. to slow cell growth and increase the relative rate of protein expression.

## Cell Culture Process Used to Produce Flt1D2.VEGFR3D3.Fc $\Delta$ C1(a)

The same methodologies as described supra for  $^{45}$  Flt1D2.Flk1D3.Fc $\Delta C1(a)$  were used to produce Flt1D2.VEGFR3D3.Fc $\Delta C1(a)$ .

#### Example 22

#### Harvest and Purification of Modified Flt1 Receptors

#### (a) Harvest and Purification of Flt1D2.Flk1D3.FcΔC1(a)

The product protein was aseptically harvested from the bioreactor while retaining cells using Millipore Prostak tangential-flow filtration modules and a low-shear mechanical pump (Fristam). Fresh medium was added to the bioreactor to replace that removed during the harvest filtration. Approximately 40 L of harvest filtrate was then loaded onto a 400 mL column containing Protein A SEPHAROSETM resin (Amersham Pharmacia). After loading the resin was washed with buffer containing 10 mM sodium phosphate, 500 mM sodium chloride, pH 7.2 to remove any unbound contaminating proteins. Flt1D2.Flk1D3.Fc $\Delta$ C1(a) protein was eluted with a pH 3.0 citrate buffer. The eluted protein was neutralized by addition of Tris base and frozen at -20° C.

Several frozen lots of Flt1D2.Flk1D3.FcΔC1(a) protein from the Protein A step above -were thawed, pooled and concentrated using a Millipore 30 kD nominal molecular weight cutoff (NMWCO) tangential flow filtration membrane. The protein was transferred to a stirred cell concentrator (Millipore) and further concentrated to 30 mg/mL using a 30 kD NMWCO membrane. The concentrated protein was loaded onto a size exclusion column packed with Superdex 200 resin (Amersham Pharmacia) that was equilibrated with phosphate buffered saline plus 5% glyc- 10 VEGF165 as compared to control media challenge. erol. The same buffer was used to run the column. The fractions corresponding to Flt1D2.Flk1D3.FcΔC1(a) dimer were pooled, sterile filtered through a 0.22 micron filter, aliquoted and frozen.

### (b) Harvest and Purification of Flt1D2.VEGFR3D3.Fc $\Delta$ C1 $^{15}$

same methodologies as described supra for Flt1D2.Flk1D3.FcΔC1(a) were used to harvest and purify Flt1D2.VEGFR3D3.FcΔC1(a).

#### Example 23

#### Phosphorylation Assay for Transiently Expressed VEGFR2

Primary human umbilical vein endothelial cells (HU-VECs), passage 4-6, were starved for 2 hrs in serum-free DME high glucose media. Samples containing 40 ng/ml (1 nM) human VEGF165, which is a ligand for the VEGF receptors Flt1, Flk1 and Flt4(VEGFR3) were prepared and 30 were preincubated for 1 hr. at room temperature with varying amounts of the modified Flt1 receptors Flt1(1-3)-Fc, Flt1(1-3)-Fc (A40),Flt1D2Flk1D3.Fc∆C1(a) Flt1D2VEGFR3D3.FcΔC1(a) in serum-free DME-high glucose media containing 0.1% BSA. Cells were challenged for 35 minutes with the samples prepared above +/- VEGF165, followed by whole cell lysis using complete lysis buffer. Cell lysates were immunoprecipitated with an antibody directed against the C-terminus of VEGFR2 receptor. The immunoprecipitated lysates were loaded onto 4-12% SDS-PAGE 40 Novex gel and then transferred to PVDF membrane using standard transfer methodologies. Detection of phosphorylated VEGFR2 was done by immunoblotting with the antiphospho Tyrosine mAb called 4G10 (UBI) and developed using ECL-reagent (Amersham). FIGS. 25A-25C and 26A-45 26B show the results of this experiment. FIG. 25A-25C reveals that detection by Western blot of tyrosine phosphorylated VEGFR2(Flk1) by VEGF165 ligand stimulation shows that cell-surface receptors are phosphorylated to varying levels depending on which modified Flt1 receptor is 50 used during the preincubations with VEGF. As is seen in FIG. 25A, at a 1.5 molar excess of either Flt1(1-3)-Fc, Flt1(1-3)-Fc (A40) or transient Flt1D2Flk1D3.FcΔC1(a) there is complete blockage of receptor stimulation by these three modified Flt1 receptors as compared to control media 55 challenge. In contrast, transient Flt1D2VEGFR3D3.Fc∆C1 (a) does not show significant blockage at this molar excess, as compared with VEGF positive control challenge. Similar results are seen in FIG. 25B, where the modified Flt receptors are in a 3-fold molar excess to VEGF165 ligand. In FIG. 60 25C, where the modified Flt1 receptors are in a 6-fold molar VEGF165 ligand, transient Flt1D2VEGFR3D3.FcΔC1(a) can now be shown to be partially blocking VEGF165-induced stimulation of cell-surface receptors.

In FIG. 26A-26B, detection by Western blot of tyrosine phosphorylated VEGFR2(Flk1) by VEGF165 ligand stimulation shows that cell-surface receptors are not phosphorylated by challenge samples which have VEGF165 preincubated with 1 and 2 fold molar excess (FIG. 26A) or 3 and 4 fold molar excess (FIG. 26B) of either transient Flt1D2Flk1D3.FcΔC1(a), stable Flt1D2Flk1D3.FcΔC1(a), or transient VEGFR1R2-FcΔC1(a). At all modified Flt1 receptor concentrations tested there is complete binding of VEGF165 ligand during the preincubation, resulting in no detectable stimulation of cell-surface receptors by unbound

#### Example 24

#### Cell Proliferation Bioassay

The test cell population is MG87 cells that have been stably transfected with a expression plasmid that contains a DNA insert encoding the VEGFR2(Flk1) extracellular domain fused to the TrkB intracellular kinase domain, thus producing a chimeric molecule. The reason the TrkB intracellular kinase domain was used rather than the native VEGFR2(Flk1) intracellular kinase domain is that the intracellular kinase domain of VEGFR2(Flk1) does not cause a strong proliferative response when stimulated by VEGF165 in these cells. It is known that MG87 cells containing full length TrkB receptor give a robust proliferative response when stimulated with BDNF, so the TrkB intracellular kinase domain was engineered to replace the intracellular kinase domain of VEGFR2(Flk1) to take advantage of this proliferative response capability.

5×10<sup>3</sup> cells/well were plated in a 96 well plate and allowed to settle for 2 hrs at 37° C. The following modified Flt receptors Flt1(1-3)-Fc, Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1(a), plus an irrelevant receptor termed Tie2-Fc as a negative control, were titrated from 40 nM to 20 pM and incubated on the cells for 1 hr at 37° C. Human recombinant VEGF165 in defined media was then added to all the wells at a concentration of 1.56 nM. The plates were incubated for 72 hrs at 37° C. and then MTS (Owen's reagent, Promega) added and the plates were incubated for an additional for 4 hrs. Finally, the plates were read on a spectrophotometer at 450/570 nm. The results of this experiment are shown in FIG. 27. The control receptor Tie2-Fc does not block VEGF165-induced cell proliferation at any concentration whereas Flt1D2.Flk1D3.FcΔC1(a) blocks 1.56 nM VEGF165 with a half maximal dose of 0.8 nM. Flt1(1-3)-Fc and Flt1D2.VEGFR3D3.Fc $\Delta$ C1(a) are less effective in blocking VEGF165 in this assay with a half maximal dose of ~2 nM. VEGF165 alone gives a reading of 1.2 absorbance units and the background is 0.38 absorbance

#### Example 25

#### Binding Stoichiometry of Modified Flt Receptors to VEGF165

#### (a) BIACORETM Analysis

The stoichiometry of Flt1D2Flk1D3.FcΔC1(a) and VEGFR1R2-Fc∆C1(a) interaction with human VEGF165 was determined by measuring either the level of VEGF saturation binding to the Flt1D2Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a) surfaces or measuring concentration of VEGF165 needed to completely prevent binding of Flt1D2Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a) to VEGF BIACORE™ chip surface.

Example 26

Determination of the Binding Stoichiometry of Flt1D2Flk1D3.FcΔC1(a)/VEGF165 Complex by Size Exclusion Chromatography

Flt1D2Flk1D3.FcΔC1(a)/VEGF165 Complex Preparation

VEGF165 (concentration=3.61 mg/ml) was mixed with CHO cell transiently expressed Flt1D2.Flk1D3.FcΔC1(a) (concentration=0.9 mg/ml) in molar ratio of 3:1 (VEGF165: Flt1D2.Flk1D3.FcΔC1(a)) and incubated overnight at 4° C.

(a) Size Exclusion Chromatography (SEC) Under Native Conditions

To separate the complex from excess of unbound VEGF165, 50 µl of the complex was loaded on a Pharmacia SUPEROSE™ 12 PC 3.2/30 which was equilibrated in PBS buffer. The sample was eluted with the same buffer at flow rate 40 µl/min. at room temperature. The results of this SEC are shown in FIG. 31. Peak #1 represents the complex and peak #2 represents unbound VEGF165. Fractions eluted between 1.1 and 1.2 ml were combined and guanidinium hydrochloride (GuHCl)was added to a final concentration 4.5M to dissociate the complex.

(b) Size Exclusion Chromatography (SEC) Under Dissocia-

To separate the components of the receptor-ligand complex and to determine their molar ratio, 50 µl of dissociated complex as described supra was loaded onto a SUPER-OSETM 12 PC 3.2/30 equilibrated in 6M GuHCl and eluted with the same solution at a flow rate 40 ul/min, at room temperature. The results of this SEC are shown in FIG. 32.

Complex Stoichiometry

The stoichiometry of the receptor-ligand complex was determined from the peak area or the peak height of the VEGF165 components. Concentrations of Flt1D2Flk1D3.FcΔC1(a) corresponding to the peak height or peak area, respectively, were obtained from the standard curves for VEGF165 and Flt1D2Flk1D3.FcΔC1(a). To obtain a standard curve, four different concentrations (0.04 mg/ml -0.3 mg/ml) of either component were injected onto a Pharmacia SEPHAROSE™ 12 PC 3.2/30 column equilibrated in 6M guanidinium chloride and eluted with the same solution at flow rate 40 µl/min. at room temperature. The standard curve was obtained by plotting peak area or peak height vs protein concentration. The molar ratio of VEGF165:Flt1D2Flk1D3.FcΔC1(a) determined from the peak area of the components was 1.16. The molar ratio of VEGF165:Flt1D2Flk1D3.FcΔC1(a) determined from the peak height of the components was 1.10.

### Example 27

Determination of the Stoichiometry of the Flt1D2Flk1D3.FcΔC1(a)/VEGF165 Complex by Size Exclusion Chromatography with On-Line Light Scattering

Complex Preparation

VEGF165 was mixed with CHO transiently expressed curves. The results of this experiment are set forth in FIG. 65 Flt1D2.Flk1D3.FcΔC1(a) protein in molar ratio of 3:1 (VEGF165:Flt1D2Flk1D3.Fc $\Delta C1(a))$  and incubated overnight at 4° C.

Modified Flt receptors Flt1D2Flk1D3.FcΔC1(a) and VEGFR1R2-FcΔC1(a), were captured with an anti-Fc specific antibody that was first immobilized on a BIACORETM chip using amine-coupling chemistry. A blank antibody surface was used as a negative control. VEGF165 was injected at a concentration of 1 nM, 10 nM, and 50 nM over the Flt1D2Flk1D3.FcΔC1(a) and VEGFR1R2-FcΔC1(a) surfaces at 10 µl/min for one hour. A real-time binding signal was recorded and saturation binding was achieved at the end of each injection. Binding stoichiometry was calculated as a molar ratio of bound VEGF165 to the immobilized Flt1D2Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a), using the conversion factor of 1000 RU equivalent to 1 ng/ml. The results indicated binding stoichiometry of one  $\overline{\text{VEGF165}}^{15}$ dimeric molecule per one Flt1 D2Flk1 D3.FcΔC1(a) or VEGFR1R2-Fc ΔC1(a) molecule (FIG. 28).

In solution, Flt1D2Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a) at a concentration of 1 nM (estimated to be 1000 20 times higher than the KD of the Flt1 D2Flk1 D3.FcΔC1(a) or VEGFR1 R2-FcΔC1(a)NEGF165 interaction) were mixed with varied concentrations of VEGF165. After one concentrations of incubation. the free Flt1D2Flk1D3.FcΔC1(a) in solution were measured as a binding signal to an amine-coupled VEGF165 surface. A calibration curve was used to convert Flt1D2Flk1D3.FcΔC1(a) BIACORE™ binding signal to its molar concentration. The data showed that the addition of 1 30 nM VEGF165 into the Flt1D2Flk1D3.FcΔC1(a) solution completely blocked Flt1D2Flk1D3.FcΔC1(a) binding to the VEGF165 surface. This result suggested the binding stoichiometry of one VEGF165 molecule per one Flt1D2Flk1D3.FcΔC1(a) molecule (FIG. 29 and FIG. 30). 35 (c) Calculation of Flt1D2Flk1D3.FcΔC1(a):VEGF165 When the concentration of Flt1D2Flk1D3.FcΔC1(a) was plotted as a function of added concentration of VEGF165, the slope of the linear portion was 1.06 for Flt1D2Flk1D3.FcΔC1(a) and -1,07 for VEGFR1R2-FcΔC1 <sub>40</sub> (a). The magnitude of the slope, very close to negative one, was indicative that one molecule of VEGF165 bound to one molecule of either Flt1D2Flk1D3.FcΔC1(a) or VEGFR1R2- $Fc\Delta C1(a)$ .

## (b) Size Exclusion Chromatography

Flt1D2Flk1D3.FcΔC1(a) was mixed with a 3-fold excess of VEGF165 and the receptor-ligand complex was purified using a Pharmacia SUPEROSETM 6 size exclusion chromatography column. The receptor-ligand complex was then incubated in a buffer containing 6M guanidine hydrochloride in order to dissociate it into its component proteins. Flt1D2Flk1D3.FcΔC1(a) was separated from VEGF165 using SUPEROSE™ 6 size exclusion chromatography column run in 6M guanidium chloride. In order to determine complex stoichiometry, several injections Flt1D2Flk1D3.FcΔC1(a) and VEGF165 were made and peak height or peak integrated intensity was plotted as a function of the concentration of injected protein. The calibration was done under condition identical to one used in separating components of Flt1D2Flk1D3.FcΔC1(a)VEGF complex. Quantification of the Flt1D2Flk1D3.FcΔC1(a)/ VEGF complex composition was based on the calibration 28, which shows the ratio of VEGF165 to Flt1D2Flk1D3.FcΔC1(a) in a complex to be 1:1.

(a) Size Exclusion Chromatography (SEC) with On-Line Light Scattering

Size exclusion chromatography column with a MiniDawn on-line light scattering detector (Wyatt Technology, Santa Barbara, Calif.) and refractive index (RI) detectors (Shi-5 madzu, Kyoto, Japan) was used to determine the molecular weight (MW) of the receptor-ligand complex. Samples were injected onto a SUPEROSETM 12 HR 10/30 column (Pharmacia) equilibrated in PBS buffer and eluted with the same buffer at flow rate 0.5 ml/min. at room temperature. As  $\,^{10}$ shown in FIG. 33, the elution profile shows two peaks. Peak #1 represents the receptor-ligand complex and peak #2 represents the unbound VEGF165. MW was calculated from LS and RI signals. The same procedure was used to determine MW of the individual components of the receptor- 15 ligand complex. The results of these determinations are as follows: MW of the Flt1D2Flk1D3.FcΔC1(a)/VEGF-165 complex at the peak position is 157 300 (FIG. 33), the MW of VEGF165 at the peak position is .44 390 (FIG. 34) and the MW of R1R2 at the peak is 113 300 (FIG. 35).

These data indicated that the stoichiometry of the  $Flt1D2Flk1D3.Fc\Delta C1(a)/VEGF$  complex is 1:1 as its corresponds to the sum of molecular weights for  $Flt1D2Flk1D3.Fc\Delta C1(a)$  and VEGF165. Importantly, this method conclusively proved that the  $Flt1D2Flk1D3.Fc\Delta C1(a)/VEGF165$  complex was indeed composed of only one molecule of VEGF165 ligand and only one molecule of the  $Flt1D2Flk1D3.Fc\Delta C1(a)$ .

#### Example 28

## Peptide Mapping of Flt1D2.Flk1D3.FcΔC1(a)

The disulfide structures and glycosylation sites in Flt1D2.Flk1D3.Fc $\Delta$ C1(a) were determined by a peptide mapping method. In this method, the protein was first cleaved with trypsin. Tryptic fragments were analyzed and identified by HPLC coupled with mass spectrometry, in addition to an N-terminal sequencing technique.

Reduction of the tryptic digest was employed to help identify disulfide-bond-containing fragments. Treatment of the tryptic digest with PNGase F (Glyko, Novato, Calif.) was employed to help identify fragments with N-linked glycosylation sites. The results are summarized in the accompanying FIG. **36**.

There are a total of ten cvsteines in Flt1D2.Flk1D3.FcΔC1(a); six of them belong to the Fc region. Cys27 has been confirmed to be disulfide bonded to Cys76. Cys121 is confirmed to be disulfide bonded to Cys 50 182. The first two cysteines in the Fc region (Cys211 and Cys214) form an intermolecular disulfide bond with the same two cysteines in another Fc chain. However, because these two cysteines can not be separated enzymatically from each other, it can not be determined whether disulfide 55 bonding is occurring between same cysteines (Cys211 to Cys211, for example) or between Cys211 and Cys214. Cys216 is confirmed to be disulfide bonded to Cys306. Cys 352 is confirmed to be disulfide bonded to Cys410.

There are five possible N-linked glycosylation sites in 60 Flt1D2.Flk1D3.Fc $\Delta$ C1(a). All five of them are found to be glycosylated to varying degrees. Complete glycosylation was observed at Asn33 (amino acid sequence NIT), Asn193 (amino acid sequence NST), and Asn282 (amino acid sequence NST). In addition, partial glycosylation is 65 observed on Asn65 and Asn120. Sites of glycosylation are highlighted by underline in the FIG. 36.

## Example 29

#### Pharmacokinetic Analysis of Modified Flt Receptors

(a) Pharmacokinetic Analysis of Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.FcΔC1(a) and VEGFR1R2-FcΔC1(a)

Balb/c mice (25-30 g) were injected subcutaneously with 4 mg/kg of Flt1(1-3)-Fc (A40), CHO transiently expressed Flt1D2.Flk1D3.FcΔC1(a), CHO stably Flt1D2.Flk1D3.FcΔC1(a), and CHO transiently expressed VEGFR1R2-Fc $\Delta$ C1(a). The mice were tail bled at 1, 2, 4, 6, 24 hrs, 2 days, 3 days and 6 days after injection. The sera were assayed in an ELISA designed to detect Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.Fc $\Delta$ C1(a) or VEGFR1R2-Fc $\Delta$ C1(a). The ELISA involves coating an ELISA plate with VEGF165, binding the detect Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.FcΔC1(a) or VEGFR1R2-FcΔC1(a) and reporting with an anti-Fc antibody linked to horse radish peroxidase. The results of this experiments are shown in FIG. 37. The  $T_{max}$  for Flt1(1-3)-Fc (A40) was at 6 hrs while the T<sub>max</sub> for the transient and stable Flt1D2.Flk1D3.FcΔC1 (a) and the transient VEGFR1R2-FcΔC1(a) was 24 hrs. The  $C_{max}$  for Flt1(1-3)-Fc (A40) was 8 µg/ml. For both transients (Flt1D2.Flk1D3.Fc $\Delta$ C1(a) and VEGFR1R2-Fc $\Delta$ C1(a)) the  $C_{max}$  was 18  $\mu$ g/ml and the  $C_{max}$  for the stable VEGFR1R2-Fc $\Delta$ C1(a) was 30  $\mu$ g/ml.

(b) Pharmacokinetic Analysis of Flt1(1-3)-Fc (A40),
 30 Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1
 (a)

Balb/c mice (25-30 g) were injected subcutaneously with 4 mg/kg of Flt1(1-3)-Fc (A40), CHO transiently expressed Flt1D2.Flk1D3.FcΔC1(a) and CHO transiently expressed 35 Flt1D2.VEGFR3D3.FcΔC1(a). The mice were tail bled at 1, 2, 5, 6, 7, 8, 12, 15 and 20 days after injection. The sera were assayed in an ELISA designed to detect Flt1(1-3)-Fc, Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1 (a). The ELISA involves coating an ELISA plate with 165, binding the Flt1(1-3)-Fc, Flt1D2.Flk1D3.FcΔC1(a) or Flt1D2.VEGFR3D3.FcΔC1(a) and reporting with an anti-Fc antibody linked to horse radish peroxidase. Flt1(1-3)-Fc (A40) could no longer be detected in the serum after day 5 Flt1D2.Flk1D3.FcΔC1(a) whereas. Flt1D2.VEGFR3D3.FcΔC1(a) were detectable for 15 days or more. The results of this experiment are shown in FIG. 38.

#### Example 30

# Evaluation of the Ability of Flt1D2.Flk1D3.FcΔC1(a) to Inhibit Tumor Growth in Vivo

To evaluate the ability of Flt1D2.Flk1D3.FcΔC1(a) to inhibit tumor growth in vivo a model in which tumor cell suspensions are implanted subcutaneously on the right flank of male severe combined immunodeficiency (SCID) mice was employed. Two cell lines, the human HT-1080 fibrosarcoma cell line (ATCC accession no. CCL-121) and the rat C6 glioma cell line (ATCC accession no. CCL-121) and the rat C6 glioma cell line (ATCC accession no. CCL-107), each of which exhibit distinctly different morphologies and growth characteristics, were used in the assay. The first dose of Flt1D2.Flk1D3.FcΔC1(a) (at 25 mg/Kg or as indicated in FIGS. 39 and 40) was given on the day of tumor implantation. Animals subsequently received subcutaneous injections of Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.FcΔC1(a) or vehicle either every other day (EOD) or two times per week

(2x/wk) for a period of 2 weeks. After 2 weeks, animals were perfused with fixative, tumors were removed and samples were blinded. Tumor volume was determined by measuring the length and width of visible subcutaneous tumors. Both of Flt1(1-3)-Fc (A40) and Flt1D2.Flk1D3.Fc $\Delta$ C1(a) significantly reduced the growth of tumors formed by HT-1080 and C6 cells. The results of these experiments are shown in FIG. **39** and FIG. **40**.

#### Example 31

The Effect of VEGF165 and Modified Flt Receptors in Female Reproductive System

The stereotypic pattern of vascular remodeling which 15 occur in the uterus and ovary over the course of the reproductive cycle has been well characterized, making these tissues particularly well suited to the study of mechanisms which regulate angiogenesis, vascular remodeling and vascular regression. Indeed, in situ hybridization studies in 2 the reproductive tissues provided the first clear evidence that VEGF acts as a mediator of physiological angiogenesis in mature rodents, as well as humans and non-human primates (Phillips et al, 1990; Ravindranath et al, 1992; Shweiki et al, 1993; Kamat et al, 1995). As cyclic angiogenesis and 25 vascular remodeling are prominent features of the normal ovary and uterus, it is not surprising that abnormal blood vessel growth and/or vascular dysfunction have been found to characterize many pathological conditions which affect these organs. Furthermore, these pathogenic vascular abnor- 30 malities are thought to be caused or perpetuated by the dysregulated expression of one or more angiogenic or antiangiogenic factors, most prominently VEGF.

For example, abnormal angiogenesis is characteristic of polycystic ovary disease, endometriosis and endometrial 35 the carcinoma, and in each case VEGF is over expressed in the affected tissue (Kamat et al, 1995; Shifren et al, 1996; Guidi et al, 1996; Donnez et al, 1998). Overexpression of VEGF is also thought to play a pathogenic role in the establishment of systemic vascular hyperpermeability in ovarian hyper- 40 stimulation syndrome (McClure et al, 1994; Levin et al, 1998) and preeclampsia (Baker et al, 1995; Sharkey et al, 1996). In addition, VEGF has been implicated as the permeability factor responsible for the production of ascites associated with ovarian carcinoma and other tumors (Senger 45 et al, 1983; Boocock et al, 1995). Agents which effectively neutralize the biological actions of VEGF can reasonably be anticipated to be of therapeutic benefit in the above and related conditions.

Angiogenesis and vascular remodeling are also hallmarks 50 of blastocyst implantation and placental development (Findlay, 1986). VEGF is strongly expressed both in the maternal decidua and in embryonic trophoblasts, where it is thought to first stimulate expansion and hyperpermeability of the uterine vasculature during the peri-implantation period and 55 subsequently mediate formation of both the maternal and embryonic components of the placental vasculature (Shweiki et al, 1993; Cullinan-Bove and Koos, 1993; Chakraborty et al, 1995; Das et al, 1997). VEGF is also required for luteal angiogenesis and associated progesterone 60 secretion necessary to prepare the uterus for implantation (Ferrara et al, 1998). Thus, agents which inhibit the biological actions of VEGF may prove to be useful as contraceptive agents (by preventing implantation), or as an abortifacients in the early stages of gestation. The latter application might find particular use as a non-surgical intervention for the termination of ectopic pregnancies.

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While the expression of VEGF receptors is largely confined to the vascular endothelium in normal reproductive tissues, Flt1 is also expressed by trophoblasts in the placenta in both humans and animals (Clark et al, 1996; He et al, 1999) where it has been proposed to play a role in trophoblast invasion. Interestingly, both Flt1 and KDR (Flk1) are expressed by choriocarcinoma cell line BeWo (Charnock-Jones et al, 1994), and VEGF has been shown to promote DNA synthesis and tyrosine phosphorylation of MAP kinase in these cells. Furthermore, primary and metastatic ovarian carcinomas not only to express high levels of VEGF, but—in addition to the vascular endothelium—the tumor cells themselves express KDR and/or Flt1 (Boocock et al, 1995). These findings suggest that VEGF may not only be critically involved in the generation and maintenance of tumor vasculature, but that at least in some tumors of reproductive origin VEGF may subserve an autocrine role, directly supporting the survival and proliferation of the tumor cells. Thus agents which block the actions of VEGF may have particularly beneficial applications to the treatment of tumors of reproductive origin.

#### Methods and Results

(a) Assessment of VEGF-Induced Uterine Hyperpermeability

Pregnant mare's serum gonadotrophin (PMSG) was injected subcutaneously (5 IU) to induce ovulation in prepubertal female rats. This results in a surge of estradiol after 2 days which in turn causes an induction of VEGF in the uterus. It is reported that this induction results in hyperpermeability of the uterus and an increase in uterine wet weight 6 hrs. later and, therefore, could potentially be blocked by modified Flt receptors Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1 (a). In this in vivo model, the normal weight of the rat uterus is about 50 mg and this can be induced to 300-350 mg by PMSG. Desiccation of the tissue reveals that this is all water weight. Subcutaneous injection of Flt1(1-3)-Fc (A40), Flt1D2.Flk1D3.FcΔC1(a) and Flt1D2.VEGFR3D3.FcΔC1 (a) at 25 mg/kg at 1 hr. after PMSG injection results in about a 50% inhibition of the increase in uterine wet weight. Increasing the dose of modified Flt receptor does not further reduce the increase in wet weight suggesting that there is a VEGF-independent component to this model. The results of this experiment are shown in FIG. 41.

## (a) Assessment of Corpus Luteum Angiogenesis Using Progesterone as a Readout

Pregnant mare's serum gonadotrophin (PMSG) is injected subcutaneously (5 IU) to induce ovulation in prepubertal female rats. This results in a fully functioning corpus luteum containing a dense network of blood vessels after 4 days that allows for the secretion of progesterone into the blood stream in order to prepare the uterus for implantation. The induction of angiogenesis in the corpus luteum requires VEGF; therefore, blocking VEGF would result in a lack of new blood vessels and thus a lack of progesterone secreted into the blood stream. In this in vivo model, resting levels of progesterone are about 5 ng/ml and this can be induced to a level of 25-40 ng/ml after PMSG. Subcutaneous injection of Flt1(1-3)-Fc (A40) or Flt1D2.Flk1D3.FcΔC1(a) at 25 mg/kg or 5 mg/kg at 1 hr. after PMSG injection results in a complete inhibition of the progesterone induction on day 4. The results of this experiment are shown in FIG. 42A-42B.

#### Example 33

## Pharmacokinetic Analysis of Flt1(1-3)-Fc (A40) and Pegylated Flt1(1-3)-Fc

Flt1(1-3)-Fc was PEGylated with either 10 kD PEG or 20 kD PEG and tested in balb/c mice for their pharmacokinetic profile. Both PEGylated forms of Flt1(1-3)-Fc were found to have much better PK profiles than Flt1(1-3)-Fc (A40), with the Tmax occurring at 24 hrs. for the PEGylated molecules 10 as opposed to 6 hrs. for Flt1(1-3)-Fc (A40).

#### Example 34

#### VEGF165 ELISA to Test Affinity of Modified Flt1 Receptor Variants

10 pM of VEGF165 was incubated overnight at room temperature with modified Flt1 receptor variants ranging from 160 pM to 0.1 pM. The modified Flt1 receptor variants 20 used in this experiment were Flt1(1-3)-Fc, Flt1(1-3)-Fc (A40), transiently expressed Flt1D2VEFGFR3D3-FcΔC1(a), Flt1-(1-siently expressed Flt1D2VEFGFR3D3-FcΔC1(a)

40

 $3_{NAS}$ )-Fc, Flt1(1- $3_{R->C}$ )-Fc and Tie2-Fc. Flt1(1- $3_{NAS}$ )-Fc is a modified version of Flt1(1-3)-Fc in which the highly basic amino acid sequence KNKRASVRRR is replaced by NAS-VNGSR, resulting in the incorporation of two new glycosylation sites and a net reduction of five positive charges, both with the purpose of reducing the unfavorable effects of this sequence on PK. Flt1(1-3<sub>R->C</sub>)-Fc is a modification in which a single arginine (R) residue within the same basic amino acid sequence is changed to a cysteine (C) (KNK RASVRRR->KNKCASVRRR) to allow for pegylation at that residue, which could then shield the basic region from exerting its unfavorable effects on PK. After incubation the solution was transferred to a plate containing a capture antibody for VEGF165 (R&D). The amount of free VEGF165 was then determined using an antibody to report free VEGF165. This showed that the modified Flt1 receptor variant with the highest affinity. for VEGF165 (determined as the lowest amount of free VEGF165) was Flt1D2Flk1D3.FcΔC1(a), followed by Flt1(1-3)-Fc and Flt1 (1-3)-Fc (A40) and then by  $Flt1(1-3_{R->C})$ -Fc,  $Flt1(1-3_{NAS})$ -Fc and Flt1D2VEFGFR3D3-FcΔC1(a). Tie2Fc has no affinity for VEGF165.

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Lys	Lys	Phe	Pro	Leu	Asp 70	Thr	Leu	Ile	Pro	Asp 75	Gly	Lys	Arg	Ile	Ile 80	
Trp	Asp	Ser	Arg	Lув 85	Gly	Phe	Ile	Ile	Ser 90	Asn	Ala	Thr	Tyr	Lys 95	Glu	
Ile	Gly	Leu	Leu 100	Thr	CÀa	Glu	Ala	Thr 105	Val	Asn	Gly	His	Leu 110	Tyr	Lys	

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	ctg Leu															144		
	cat His 50															192		
	atg Met															240		
	gga Gly															288		
	caa Gln															336		
Pro	act Thr	Ser 115	Lys	Lys	Lys	Glu	Thr 120	Glu	Ser	Āla	Ile	Tyr 125	Ile	Phe	Ile	384		
Ser	gat Asp 130	Thr	Gly	Arg	Pro	Phe 135	Val	Glu	Met	Tyr	Ser 140	Glu	Ile	Pro	Glu	432		
	ata Ile															480		
	tca Ser															528		
	atc Ile															576		
Ile	ata Ile	Ser 195	Asn	Ala	Thr	Tyr	Lys 200	Glu	Ile	Gly	Leu	Leu 205	Thr	Cys	Glu	624		
_	aca Thr 210	_				_		_							-	672		
Gln 225		Asn	Thr	Ile	Ile 230	Asp	Val	Gln	Ile	Ser 235	Thr	Pro	Arg	Pro	Val 240	720		
Lys	tta Leu	Leu	Arg	Gly 245	His	Thr	Leu	Val	Leu 250	Asn	CAa	Thr	Ala	Thr 255	Thr	768		
Pro	ttg Leu	Asn	Thr 260	Arg	Val	Gln	Met	Thr 265	Trp	Ser	Tyr	Pro	Asp 270	Glu	ГХа	816		
	aag Lys															864		
	aac Asn 290															912		
	aaa Lys															960		

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					aat Asn											1248
					gtg Val											1296
					gag Glu											1344
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					gag Glu											1536
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Cya	Leu	Leu	Leu 20	Thr	Gly	Ser	Ser	Ser 25	Gly	Ser	Lys	Leu	30 Lys	Asp	Pro	
	_		_	_												

Glu Leu Ser Leu Lys Gly Thr Gln His Ile Met Gln Ala Gly Gln Thr

											_	COII	CIII	ueu	
		35					40					45			
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Glu 65	Met	Val	Ser	Lys	Glu 70	Ser	Glu	Arg	Leu	Ser 75	Ile	Thr	Lys	Ser	Ala 80
Cys	Gly	Arg	Asn	Gly 85	ГÀа	Gln	Phe	Сув	Ser 90	Thr	Leu	Thr	Leu	Asn 95	Thr
Ala	Gln	Ala	Asn 100	His	Thr	Gly	Phe	Tyr 105	Ser	Cys	Lys	Tyr	Leu 110	Ala	Val
Pro	Thr	Ser 115	Lys	Lys	Lys	Glu	Thr 120	Glu	Ser	Ala	Ile	Tyr 125	Ile	Phe	Ile
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Ile 145	Ile	His	Met	Thr	Glu 150	Gly	Arg	Glu	Leu	Val 155	Ile	Pro	CAa	Arg	Val 160
Thr	Ser	Pro	Asn	Ile 165	Thr	Val	Thr	Leu	Lys 170	Lys	Phe	Pro	Leu	Asp 175	Thr
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Ala	Thr 210	Val	Asn	Gly	His	Leu 215	Tyr	Lys	Thr	Asn	Tyr 220	Leu	Thr	His	Arg
Gln 225	Thr	Asn	Thr	Ile	Ile 230	Asp	Val	Gln	Ile	Ser 235	Thr	Pro	Arg	Pro	Val 240
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Pro	Leu	Asn	Thr 260	Arg	Val	Gln	Met	Thr 265	Trp	Ser	Tyr	Pro	Asp 270	Glu	Lys
Asn	Lys	Asn 275	Ala	Ser	Val	Arg	Arg 280	Arg	Ile	Asp	Gln	Ser 285	Asn	Ser	His
Ala	Asn 290	Ile	Phe	Tyr	Ser	Val 295	Leu	Thr	Ile	Asp	300 TÀs	Met	Gln	Asn	Lys
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Pro	ГЛа	Ser	Сув 340	Asp	ràa	Thr	His	Thr 345	CAa	Pro	Pro	CAa	Pro 350	Ala	Pro
Glu	Leu	Leu 355	Gly	Gly	Pro	Ser	Val 360	Phe	Leu	Phe	Pro	Pro 365	ràs	Pro	Lys
Asp	Thr 370	Leu	Met	Ile	Ser	Arg 375	Thr	Pro	Glu	Val	Thr 380	CAa	Val	Val	Val
Asp 385	Val	Ser	His	Glu	390	Pro	Glu	Val	Lys	Phe 395	Asn	Trp	Tyr	Val	Asp 400
				405					410					415	Tyr
Asn	Ser	Thr	Tyr 420	Arg	Val	Val	Ser	Val 425	Leu	Thr	Val	Leu	His 430	Gln	Asp
Trp	Leu	Asn 435	Gly	Lys	Glu	Tyr	Lys 440	Сув	Lys	Val	Ser	Asn 445	Lys	Ala	Leu
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Thr	Thr	Pro 515	Pro	Val	Leu	Asp	Ser 520	Asp	Gly	Ser	Phe	Phe 525	Leu	Tyr	Ser	
Lys	Leu 530	Thr	Val	Asp	Lys	Ser 535	Arg	Trp	Gln	Gln	Gly 540	Asn	Val	Phe	Ser	
Cys 545	Ser	Val	Met	His	Glu 550	Ala	Leu	His	Asn	His 555	Tyr	Thr	Gln	Lys	Ser 560	
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														act Thr		254
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acc tgt gca gca tcc agt ggg ctg atg acc aag aag aac agc aca ttt Thr Cys Ala Ala Ser Ser Gly Leu Met Thr Lys Lys Asn Ser Thr Phe 210 215 220	734
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gat gag ctg acc aag aac cag gtc agc ctg acc tgc ctg gtc aaa ggc Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly 370 375 380	1214
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gag aac aac tac aag acc acg cct ccc gtg ctg gac tcc gac ggc tcc Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser 400 405 410	1310
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Met	Tyr	Ser 35	Glu	Ile	Pro	Glu	Ile 40	Ile	His	Met	Thr	Glu 45	Gly	Arg	Glu
Leu	Val 50	Ile	Pro	СЛа	Arg	Val 55	Thr	Ser	Pro	Asn	Ile 60	Thr	Val	Thr	Leu
Lys 65	Lys	Phe	Pro	Leu	Asp 70	Thr	Leu	Ile	Pro	Asp 75	Gly	Lys	Arg	Ile	Ile 80
Trp	Asp	Ser	Arg	Lув 85	Gly	Phe	Ile	Ile	Ser 90	Asn	Ala	Thr	Tyr	Lys 95	Glu
Ile	Gly	Leu	Leu 100	Thr	CAa	Glu	Ala	Thr 105	Val	Asn	Gly	His	Leu 110	Tyr	Lys
Thr	Asn	Tyr 115	Leu	Thr	His	Arg	Gln 120	Thr	Asn	Thr	Ile	Ile 125	Asp	Val	Val
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Asp	Leu	Lys	Thr 180	Gln	Ser	Gly	Ser	Glu 185	Met	Lys	ГÀа	Phe	Leu 190	Ser	Thr
Leu	Thr	Ile 195	Asp	Gly	Val	Thr	Arg 200	Ser	Asp	Gln	Gly	Leu 205	Tyr	Thr	Cys
Ala	Ala 210	Ser	Ser	Gly	Leu	Met 215	Thr	Lys	Lys	Asn	Ser 220	Thr	Phe	Val	Arg
Val 225	His	Glu	Lys	Gly	Pro 230	Gly	Asp	Lys	Thr	His 235	Thr	CAa	Pro	Pro	Cys 240
Pro	Ala	Pro	Glu	Leu 245	Leu	Gly	Gly	Pro	Ser 250	Val	Phe	Leu	Phe	Pro 255	Pro
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Pro 385	Ser	Asp	Ile	Ala	Val 390	Glu	Trp	Glu	Ser	Asn 395	Gly	Gln	Pro	Glu	Asn 400
Asn	Tyr	Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe

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Leu 7	Tyr	Ser	Lys 420	Leu	Thr	Val	Asp	Lys 425	Ser	Arg	Trp	Gln	Gln 430	Gly	Asn	
Val I	Phe	Ser 435	Cys	Ser	Val	Met	His 440	Glu	Ala	Leu	His	Asn 445	His	Tyr	Thr	
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ctc a Leu S 15																158
gta g Val (																206
agg g Arg (																254
act t Thr I																302
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Met	Tyr	Ser 35	Glu	Ile	Pro	Glu	Ile 40	Ile	His	Met	Thr	Glu 45	Gly	Arg	Glu		
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Ile	Gly	Leu	Leu 100	Thr	Cys	Glu	Ala	Thr 105	Val	Asn	Gly	His	Leu 110	Tyr	Lys
Thr	Asn	Tyr 115	Leu	Thr	His	Arg	Gln 120	Thr	Asn	Thr	Ile	Ile 125	Asp	Ile	Gln
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Pro	Ala	Pro	Ile 340	Glu	ГÀв	Thr	Ile	Ser 345	ГÀв	Ala	ГÀв	Gly	Gln 350	Pro	Arg
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Thr	Thr	Pro	Pro	Val 405	Leu	Asp	Ser	Asp	Gly 410	Ser	Phe	Phe	Leu	Tyr 415	Ser
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L> LE 2> TY	NGTH PE:	I: 45 PRT	8	sa <u>r</u>	oiens	3									
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Val	Glu 35	Met	Tyr	Ser	Glu	Ile 40	Pro	Glu	Ile	Ile	His 45	Met	Thr	Glu	
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Thr	Leu	Lys	Lys	Phe 70	Pro	Leu	Asp	Thr	Leu 75	Ile	Pro	Asp	Gly	Eys	
Ile	Ile	Trp	Asp 85	Ser	Arg	ГЛа	Gly	Phe 90	Ile	Ile	Ser	Asn	Ala 95	Thr	
Lys	Glu	Ile 100	Gly	Leu	Leu	Thr	Сув 105	Glu	Ala	Thr	Val	Asn 110	Gly	His	
Tyr	Lys 115	Thr	Asn	Tyr	Leu	Thr	His	Arg	Gln	Thr	Asn 125	Thr	Ile	Ile	
	Val 290  cag Gln  cag Gln  cag Gln  gcc Ala  cpro acc Thr 370  agc Ser  tac Tyr  ttc Phe  aag Lys 450  0> SE 1> LE 23 OR  Val Leu  Val Arg 50  Thr  Ile  Lys	gtg gac Val Asp 290  cag tac Gln Tyr  cag gac Gln Asp gcc ctc Ala Leu  ccc cga Pro Arg 355  acc aag Thr Lys 370  agc gac Ser Asp  tac aag Tyr Lys  tac agc Tyr Ser  ttc tca Phe Ser 435  aag agc Lys Ser 450  0> SEQ III 1> LENGTH 2> TYPE: 3> ORGANI  0> SEQUEN Val Ser  Leu Leu  Val Glu 35  Arg Glu 50  Thr Leu  Ile Ile  Lys Glu	gtg gac ggc Val Asp Gly 290  cag tac aac Gln Tyr Asn  cag gac tgg Gln Asp Trp  gcc ctc cca Ala Leu Pro 340  ccc cga gaa Pro Arg Glu 355  acc aag acc Thr Lys Asn 370  agc gac atc Ser Asp Ile  tac aag acc Tyr Lys Thr  tac agc aag Tyr Ser Lys 420  ttc tca tgc Phe Ser Cys 435  aag agc ctc Lys Ser Leu 450  O> SEQ ID NO O> LENGTH: 4E 2> TYPE: PRT 3> ORGANISM: O> SEQUENCE: Val Ser Tyr  Leu Leu Leu 20  Val Glu Met 35  Arg Glu Leu 50  Thr Leu Lys Ile Ile Trp  Lys Glu Ile Lys Glu Ile Ino	gtg gac ggc gtg Val 290  cag tac aac agc Gln Tyr Asn Ser  cag gac tgg ctg Gln Asp Trp Leu 325  gcc ctc cca gcc Ala Leu Pro Ala 340  ccc cga gaa cca Pro Arg Glu Pro 355  acc aag aac cag Thr Lys Asn Gln 370  agc gac atc gcc Ser Asp Ile Ala  tac aag acc acg Tyr Lys Thr Thr 405  tac agc aag ctc Tyr Ser Lys Leu 420  ttc tca tgc tcc Phe Ser Cys Ser 435  aag agc ctc tcc Lys Ser Leu Ser 450  0> SEQ ID NO 16 1> LENGTH: 458 2> TYPE: PRT 3> ORGANISM: Home 0> SEQUENCE: 16  Val Ser Tyr Trp 5  Leu Leu Leu Thr 20  Val Glu Met Tyr 35  Arg Glu Leu Val 50  Thr Leu Lys Lys Lys Lys Ile Ile Trp Asp 85  Lys Glu Ile Gly 100	gtg gac ggc gtg gag yal Asp Gly Val Glu 290  cag tac aac agc acg Gln Tyr Asn Ser Thr 310  cag gac tgg ctg aat Gln Asp Trp Leu Asn 325  gcc ctc cca gac cca cag Pro Arg Glu Pro Gln 355  acc aag aac cag gtc Thr Lys Asn Gln Val 370  agc gac atc gcc gtg Ser Asp 11e Ala Val 370  tac aag acc acg ct Tyr Lys Thr Thr Thr Fro tac agc aag ctc acc Tyr Ser Lys Leu Thr 420  ttc tca tgc tcc gtg Phe Ser Cys Ser Val 435  aag agc ctc tcc gtg Phe Ser Cys Ser Val 435  aag agc ttc tcc tcu tca tgc tcc gtg Phe Ser Cys Ser Val 435  aag agc ttc tcc ctg Lys Ser Leu Ser Leu 450  O SEQ ID NO 16 16 LENGTH: 458 2> TYPE: PRT 3> ORGANISM: Homo sar 0> SEQUENCE: 16  Val Ser Tyr Trp Asp 5  Leu Leu Leu Thr Gly 20  Val Glu Met Tyr Ser 35  Arg Glu Leu Val Ile 50  Thr Leu Lys Lys Phe 70  Ile Ile Trp Asp Ser  Lys Glu Ile Gly Leu Lou Gly Leu 100 Cly Leu 110 Cly Cly Cly 110 Cly Cly Cly 110 Cly 110 Cly Cly 110 Cl	## 275  ## 275	275	275	275   280   280   291   295   281   281   290   295   295   281   295		275   280   280   392   393   393   394   381   380   395   315   310		Second   S	275   280   281   282   285   382	275   280   281   381

Asp Val Val Leu Ser Pro Ser His Gly Ile Glu Leu Ser Val Gly Glu

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Tyr	Thr 210	Cys	Ala	Ala	Ser	Ser 215	Gly	Leu	Met	Thr	Lys 220	Lys	Asn	Ser	Thr	
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Pro	Ala	Pro	Glu	Leu 245	Leu	Gly	Gly	Pro	Ser 250	Val	Phe	Leu	Phe	Pro 255	Pro	
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Val	Val	Val 275	Asp	Val	Ser	His	Glu 280	Asp	Pro	Glu	Val	Lys 285	Phe	Asn	Trp	
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Glu 305	Gln	Tyr	Asn	Ser	Thr 310	Tyr	Arg	Val	Val	Ser 315	Val	Leu	Thr	Val	Leu 320	
His	Gln	Asp	Trp	Leu 325	Asn	Gly	Lys	Glu	Tyr 330	Lys	Cys	Lys	Val	Ser 335	Asn	
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Leu	Tyr	Ser	Lys 420	Leu	Thr	Val	Asp	Lys 425	Ser	Arg	Trp	Gln	Gln 430	Gly	Asn	
Val	Phe	Ser 435	Cha	Ser	Val	Met	His 440	Glu	Ala	Leu	His	Asn 445	His	Tyr	Thr	
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Na	11110 70	Glu		Arg	Glu	ьeu	va⊥	Ile 25	Pro	Cys	Arg	val	Thr 30	ser	Pro	
Met			20		Leu											

Asp	Gly 50	Lys	Arg	Ile	Ile	Trp 55	Asp	Ser	Arg	Lys	Gly 60	Phe	Ile	Ile	Ser
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Asn	Gly	His	Leu	Tyr 85	Lys	Thr	Asn	Tyr	Leu 90	Thr	His	Arg	Gln	Thr 95	Asn
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Val	Gly	Glu 115	ГЛа	Leu	Val	Leu	Asn 120	Cys	Thr	Ala	Arg	Thr 125	Glu	Leu	Asn
Val	Gly 130	Ile	Asp	Phe	Asn	Trp 135	Glu	Tyr	Pro	Ser	Ser 140	Lys	His	Gln	His
Lys 145	ГЛа	Leu	Val	Asn	Arg 150	Asp	Leu	Lys	Thr	Gln 155	Ser	Gly	Ser	Glu	Met 160
ГÀа	ГÀа	Phe	Leu	Ser 165	Thr	Leu	Thr	Ile	Asp 170	Gly	Val	Thr	Arg	Ser 175	Asp
Gln	Gly	Leu	Tyr 180	Thr	CAa	Ala	Ala	Ser 185	Ser	Gly	Leu	Met	Thr 190	Lys	Lys
Asn	Ser	Thr 195	Phe	Val	Arg	Val	His 200	Glu	Lys	Gly	Pro	Gly 205	Asp	Lys	Thr
His	Thr 210	CAa	Pro	Pro	CAa	Pro 215	Ala	Pro	Glu	Leu	Leu 220	Gly	Gly	Pro	Ser
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Thr	Pro	Glu	Val	Thr 245	CÀa	Val	Val	Val	Asp 250	Val	Ser	His	Glu	Asp 255	Pro
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ГÀа	Thr	Lys 275	Pro	Arg	Glu	Glu	Gln 280	Tyr	Asn	Ser	Thr	Tyr 285	Arg	Val	Val
Ser	Val 290	Leu	Thr	Val	Leu	His 295	Gln	Asp	Trp	Leu	Asn 300	Gly	ГÀв	Glu	Tyr
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Pro	Pro	Ser	Arg 340	Asp	Glu	Leu	Thr	Lys 345	Asn	Gln	Val	Ser	Leu 350	Thr	CAa
Leu	Val	Lys 355	Gly	Phe	Tyr	Pro	Ser 360	Asp	Ile	Ala	Val	Glu 365	Trp	Glu	Ser
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Ser 385	Asp	Gly	Ser	Phe	Phe 390	Leu	Tyr	Ser	Lys	Leu 395	Thr	Val	Asp	Lys	Ser 400
Arg	Trp	Gln	Gln	Gly 405	Asn	Val	Phe	Ser	Cys 410	Ser	Val	Met	His	Glu 415	Ala
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#### We claim:

- 1. A method of inhibiting vascular endothelial growth factor (VEGF) activity in a mammal, comprising:
  - administering a pharmaceutical composition to the mammal, wherein the pharmaceutical composition comprises
  - (a) a VEGF antagonist, and
  - (b) a pharmaceutically acceptable carrier
  - wherein the VEGF antagonist comprises a dimeric fusion polypeptide comprising two fusion polypeptides, each fusion polypeptide comprising:
  - (i) a VEGF receptor component consisting of an immunoglobulin-like (Ig) domain 2 of a first VEGF receptor human Flt1 and Ig domain 3 of a second VEGF receptor human Flk1 or human Flt4; and
  - (ii) a multimerizing component, wherein VEGF activity is inhibited.
- 2. The method of claim 1, wherein the mammal is a human.

- 10 3. A method of inhibiting tumor growth in a mammal, comprising:
  - administering a pharmaceutical composition to the mammal, wherein the pharmaceutical composition comprises
  - (a) a VEGF antagonist, and
  - (b) a pharmaceutically acceptable carrier
  - wherein the VEGF antagonist comprises a dimeric fusion polypeptide comprising two fusion polypeptides, each fusion polypeptide comprising:
  - (i) a VEGF receptor component consisting of an immunoglobulin-like (Ig) domain 2 of a first VEGF receptor human Flt1 and Ig domain 3 of a second VEGF receptor human Flk1 or human Flt4; and
  - (ii) a multimerizing component, wherein tumor growth is inhibited.

\* \* \* \* \*