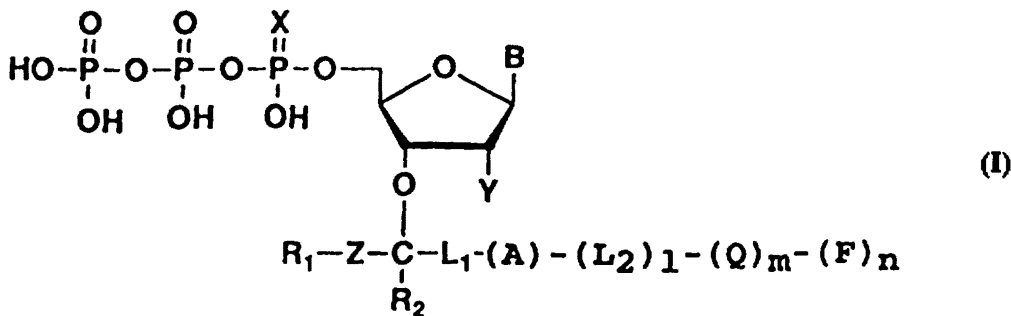


INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C07H 19/10, 19/20, C12Q 1/68	A1	(11) International Publication Number: WO 96/23807 (43) International Publication Date: 8 August 1996 (08.08.96)
(21) International Application Number: PCT/SE96/00096 (22) International Filing Date: 30 January 1996 (30.01.96) (30) Priority Data: 9500342-2 31 January 1995 (31.01.95) SE (71)(72) Applicant and Inventor: KWIATKOWSKI, Marek [SE/SE]; Lövsångarvägen 17, S-756 52 Uppsala (SE). (74) Agents: WIDÉN, Björn et al.; Pharmacia AB, Patent Dept., S-751 82 Uppsala (SE).	(81) Designated States: CA, JP, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report.</i>	

(54) Title: NOVEL CHAIN TERMINATORS, THE USE THEREOF FOR NUCLEIC ACID SEQUENCING AND SYNTHESIS AND A METHOD OF THEIR PREPARATION



(57) Abstract

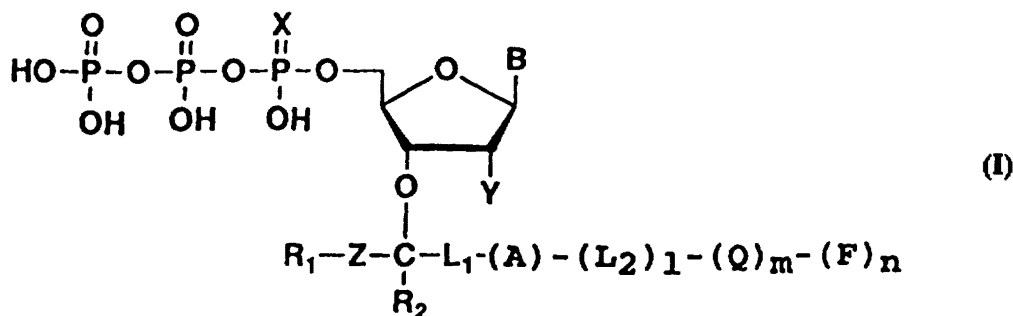
The invention relates to compounds of general structure (I) or salts thereof, wherein B is a nucleobase, X and Z independently are oxygen or sulphur, Y is hydrogen or hydroxy, which optionally may be protected, R₁ is hydrocarbyl, which optionally is substituted with a functional group, R₂ is hydrogen or hydrocarbyl, which optionally is substituted with a functional group, A is an electron withdrawing or electron donating group capable of moderating the acetal stability of compound (I), L₁ and L₂ are hydrocarbon linkers, which may be the same or different, L₂, when present, being either (i) connected to L₁ via the group A, or (ii) directly connected to L₁, the group A then being connected to one of linkers L₁ and L₂, F is a dye label, Q is a coupling group for F, and l, m and n independently are 0 or 1, with the proviso that l is 1 when m is 1, and l is 1 and m is 1 when n is 1. The compounds of formula (I) are useful as deactivatable chain extension terminators. The invention also relates to the use of the compounds (I) in

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(54) Title: NOVEL CHAIN TERMINATORS, THE USE THEREOF FOR NUCLEIC ACID SEQUENCING AND SYNTHESIS AND A METHOD OF THEIR PREPARATION



(57) Abstract

The invention relates to compounds of general structure (I) or salts thereof, wherein B is a nucleobase, X and Z independently are oxygen or sulphur, Y is hydrogen or hydroxy, which optionally may be protected, R₁ is hydrocarbyl, which optionally is substituted with a functional group, R₂ is hydrogen or hydrocarbyl, which optionally is substituted with a functional group, A is an electron withdrawing or electron donating group capable of moderating the acetal stability of compound (I), L₁ and L₂ are hydrocarbon linkers, which may be the same or different, L₂, when present, being either (i) connected to L₁ via the group A, or (ii) directly connected to L₁, the group A then being connected to one of linkers L₁ and L₂, F is a dye label, Q is a coupling group for F, and l, m and n independently are 0 or 1, with the proviso that l is 1 when m is 1, and l is 1 and m is 1 when n is 1. The compounds of formula (I) are useful as deactivatable chain extension terminators. The invention also relates to the use of the compounds (I) in nucleic acid synthesis and nucleic acid sequencing as well as to a method of preparing compounds of Formula (I).

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NOVEL CHAIN TERMINATORS, THE USE THEREOF FOR NUCLEIC ACID SEQUENCING AND SYNTHESIS AND A METHOD OF THEIR PREPARATION

The present invention relates to novel nucleic acid
5 chain extension terminators, their use in nucleic acid
sequencing and synthesis, respectively, as well as a
method for preparing such compounds.

Today, there are two predominant methods for DNA
sequence determination: the chemical degradation method
10 (Maxam and Gilbert, Proc. Natl. Acad. Sci. 74:560-564
(1977)), and the dideoxy chain termination method (Sanger
et al., Proc. Natl. Acad. Sci. 74:5463-5467 (1977)). Most
automated sequencers are based on the chain termination
method utilizing fluorescent detection of product
15 formation. In these systems either primers to which
deoxynucleotides and dideoxynucleotides are added are dye-
labelled, or the added dideoxynucleotides are
fluorescently labelled. As an alternative, dye labelled
deoxynucleotides can be used in conjunction with unlabeled
20 dideoxynucleotides. This chain termination method is based
upon the ability of an enzyme to add specific nucleotides
onto the 3' hydroxyl end of a primer annealed to a
template. The base pairing property of nucleic acids
determines the specificity of nucleotide addition. The
25 extension products are then separated electrophoretically
on a polyacrylamide gel and detected by an optical system
utilizing laser excitation.

Although both the chemical degradation method and the
dideoxy chain termination method are in widespread use,
30 there are many associated disadvantages. For example, the
methods require gel-electrophoretic separation. Typically,
only 400-800 base pairs can be sequenced from a single
clone. As a result, the systems are both time- and labor-
intensive. Methods avoiding gel separation have been
35 developed in attempts to increase the sequencing
throughput.

Sequencing by hybridization (SBH) methods have been
proposed by Crkvenjakov (Drmanac et al., Genomics 4:114

(1989); Strezoska et al., (Proc. Natl. Acad. Sci. USA 88:10089 (1991)), Bains and Smith (Bains and Smith, J. Theoretical Biol. 135:303 (1988)) and in US-A-5,202,231. This type of system utilizes the information obtained from
5 multiple hybridizations of the polynucleotide of interest, using short oligonucleotides to determine the nucleic acid sequence. These methods potentially can increase the sequence throughput because multiple hybridization reactions are performed simultaneously. To reconstruct the
10 sequence, however, an extensive computer search algorithm is required to determine the most likely order of all fragments obtained from the multiple hybridizations.

The SBH methods are problematic in several respects. For example, the hybridization is dependent upon the
15 sequence composition of the duplex of the oligonucleotide and the polynucleotide of interest, so that GC-rich regions are more stable than AT-rich regions. As a result, false positives and false negatives during hybridization detection are frequently present and complicate sequence
20 determination. Furthermore, the sequence of the polynucleotide is not determined directly, but is inferred from the sequence of the known probe, which increases the possibility for error.

Methods have also been proposed which detect the
25 addition or removal of single molecules from a DNA strand.

For example, Hyman E.D., Anal. Biochem., 174:423 (1988) discloses the addition of a nucleotide to a an immobilised DNA template/primer complex in the presence of a polymerase and determination of polymerisation reaction
30 by detecting the pyrophosphate liberated as a result of the polymerisation.

Jett et al., J. Biomol. Struct. Dyn., I, p. 301, 1989 discloses a method wherein a single stranded DNA or RNA molecule of labelled nucleotides, complementary to the
35 sequence to be determined, is suspended in a moving flow stream. Individual bases are then cleaved sequentially from the end of the suspended sequence and determined by a detector passed by the flow stream.

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