#### SOLID STATE PHYSICS, VOL. 57

## The Physics of Protein Crystallization

Peter G.  $Vekilov^1$  and Alexander A.  $Chernov^2$ 

<sup>1</sup>Department of Chemical Engineering, University of Houston, Houston, TX 77546, USA
<sup>2</sup>Universities Space Research Association, Marshall Space Flight Center, SD46
Huntsville, AL 35812, USA

I.	Introduction		
II.	Prot	eins and Protein Crystallization: Phenomenology	9
	1.	The Protein Molecules	9
	2.		9
	3.		13
III.	The	Protein Crystal 1	14
	4.		14
	5.		17
	6.	<del></del>	18
	7.		22
IV.	Inter		23
	8.		23
	9.		25
	10.	The Need for High Electrolyte Concentration in the Crystallization	
		of Hemoglobin C 3	30
	11.	Phase Diagram: Dense Liquid Phases	32
V.	Ther	modynamics and Crystallization Driving Force	34
	12.	Interactions and Thermodynamics	14
	13.		39
	14.	Human Hemoglobin C—Positive Enthalpy of Crystallization	1
	15.	Apoferritin—Athermal Crystallization Driven by the Release	
		of Water Molecules 4	5
	16.	Lysozyme—Negative Crystallization Enthalpy and Entropy 4	6
	17.	Solution Non-Ideality and the Crystallization Driving Force 4	7
VI.	Crys	tal Nucleation	0
	18.	Techniques for Nucleation Rates Determinations 5	1
	19.	Kinetics of the Nucleation Processes	4
	20.	Enhanced Crystal Nucleation Around the L-L Separation Boundary 6	2
	21.	Control of the Nucleation Rate of Protein Crystals	5
	22.	The Shape and Structure of the Critical Nucleus 6	9
/II.	Grov	th of Crystals: the Mesoscopic Lengthscales	7
	23.	General Background	7
	24.	Generation of Steps	9
	25.	Kinetics of Step Propagation, the Kinetic Coefficients	1
	26.	Impurity Effects on Step Propagation	0
	27	Interactions between Stens 0	1



VIII.	Molecular Processes at Steps		96
	28.	Density of Growth Sites	96
	29.	Higher-Order Neighbors and Kink Density	104
	30.	Molecular Potentials and Kink Density	106
	31.	Frequency of Molecular Attachment	108
	32.	Molecular-Level Parameters Underlying the Macroscopic Growth Kinetics	110
IX.	Impu	urities	111
	33.	Types of Impurities	111
	34.	Aging and Nucleation	112
	35.	Trapping of Impurities	113
Χ.	Non-	uniform Step Density and Step Pattern Formation	115
XI.	Crys	tal Perfection	123
	36.	Sub-Molecular Level Defects	124
	37.	Rotational and Translational Lattice Defects	124
	38.	Impurity-Related Defects	128
	39.	Linear and Planar Defects	134
	40.	Striations, Occlusions	134
	41.	Incorporation of Microcrystals	135
	42.	Mosaicity	137
	43.	Growth under Reduced Gravity Conditions	143
XII.	Conc	cluding Remarks	146
	Ackr	nowledgments	147

### I. Introduction

Proteins are the elementary building units of all living creatures and essential components for information and energy processing within the living systems. The correlation between the structure and the function for this group of natural compounds has been the focus of intense investigations for more than 50 years. (For a brief history, see Ref. 2, and for further and more recent developments see Refs. 3–6.) Currently, the two most broadly used methods of protein structure determinations are x-ray crystallography and Nuclear Magnetic Resonance (NMR). NMR is limited to proteins of molecular mass lower than 30,000–60,000 Da, involves rather long data collection times, <sup>7–9</sup> and thus is expected to

<sup>&</sup>lt;sup>9</sup> S. J. Glaser, T. Schulte-Herbrüggen, M. Sieveking, O. Schedletzky, N. C. Nielsen, O. W. Sørensen,



<sup>&</sup>lt;sup>1</sup> T. E. Creighton, *Proteins: Structure and Molecular Properties*, W. H. Freeman, New York (1993).

<sup>&</sup>lt;sup>2</sup> A. McPherson, J. Cryst. Growth 110, 1-10 (1991).

<sup>&</sup>lt;sup>3</sup> M. F. Perutz and F. S. Mathews, J. Mol. Biol. 21, 199–202 (1966).

<sup>&</sup>lt;sup>4</sup> S. R. Simon, W. H. Konigsberg, W. Bolton, and M. F. Perutz, J. Mol. Biol. 28, 451-454 (1967).

<sup>&</sup>lt;sup>5</sup> A. McPherson, Crystallization of Biological Macromolecules, Cold Spring Harbor Laboratory Press, New York (1999).

<sup>&</sup>lt;sup>6</sup> A. McPherson, A. J. Malkin, and Y. G. Kuznetsov, Ann. Rev. Biomol. Struct. 20, 361–410 (2000).

<sup>&</sup>lt;sup>7</sup> K. Wütrich, Acta Cryst. Section D 51, 249-270 (1995).

<sup>&</sup>lt;sup>8</sup> W. S. Warren, *Science* **280**, 398–399 (1998).

contribute at most 20% of the data. On the other hand, due to recent advances in protein expression and isolation, <sup>10</sup> data collection on specialized high-intensity synchrotron beam lines, <sup>11,12</sup> crystallization screening techniques, <sup>13–18</sup> and computational methods for structure determination and refinement, x-ray crystallography is close to becoming a high-throughput method. 10

To resolve atoms that are, typically, 1.5–2 Å apart, the diffraction methods require single crystals that are as large as several tenths of a millimeter in all three dimensions, and have low defect contents and high compositional and structural uniformity. At present, most proteins can be enticed to produce some kind of microcrystals. However, the crystals are often of significantly lower quality and/or size than needed for the desired resolution. Often, further improvement in crystallization involves the application of elaborate, protein-specific techniques. A few non-exhaustive examples include genetic engineering of intermolecular contacts, <sup>19,20</sup> proteolysis, <sup>21</sup> utilization of the phase behavior of complex, multicomponent systems, <sup>22,23</sup> or even costly microgravity experiments aboard spacecraft.<sup>24-28</sup> Hence, claims that protein crystallization is a major rate-limiting step in structure determinations

<sup>&</sup>lt;sup>28</sup> E. H. Snell, A. Cassetta, J. R. Helliwell, T. J. Boggon, N. E. Chayen, E. Weckert, K. Hoelzer, K.



<sup>&</sup>lt;sup>10</sup> S. K. Burley, S. C. Almo, J. B. Bonanno, M. Capel, M. R. Chance, T. Gaasterland, D. Lin, A. Sali, F. W. Studier, and S. Swaminathan, Nature Genetics 23, 151-157 (1999).

<sup>11</sup> S.-H. Kim, Nature Struct. Biol. Synchrotron Supplement 643-645 (1998).

<sup>&</sup>lt;sup>12</sup> R. S. Service, *Science* **285**, 1342–1346 (1999).

<sup>&</sup>lt;sup>13</sup> N. E. Chaven, P. D. ShawStewart, and P. Baldock, Acta Crystallogr. Section D 50, 456–458 (1994).

<sup>&</sup>lt;sup>14</sup> B. Cudney, S. Pattel, K. Weisgraber, Y. Newhouse, and A. McPherson, Acta Crystallogr. Section D 50, 414-423 (1994).

<sup>&</sup>lt;sup>15</sup> G. L. Gilliland, M. Tung, D. M. Blakeslee, and J. E. Ladner, Acta Crystallogr. Section D 50, 408-413 (1994).

<sup>&</sup>lt;sup>16</sup> L. F. Kuyper and C. W. Carter Jr., J. Cryst. Growth 168, 155-169 (1996).

<sup>&</sup>lt;sup>17</sup> B. D. Prater, S. C. Tuller, and L. J. Wilson, J. Cryst. Growth 196, 674–684 (1999).

<sup>&</sup>lt;sup>18</sup> P. D. S. Stewart and P. F. M. Baldock, J. Cryst. Growth 196, 665-673 (1999).

<sup>&</sup>lt;sup>19</sup> D. M. Lawson, P. J. Artymiuk, S. J. Yewdall, J. M. A. Smith, J. C. Livingstone, A. Trefry, A. Luzzago, S. Levi, P. Arosio, G. Cesareni, C. D. Thomas, W. V. Shaw, and P. M. Harrison, Nature 349, 541-544 (1991). <sup>20</sup> D. A. Doyle, J. M. Cabral, R. A. Pfuetzner, A. Kuo, J. M. Culbis, S. L. Cohen, B. T. Chait, and

R. MacKinnon, Science 280, 69-77 (1998).

<sup>&</sup>lt;sup>21</sup> S. L. Cohen, A. R. Ferre-D'Amare, S. K. Burley, and B. T. Chait, *Protein Sci.* 4, 1088–1099 (1995). <sup>22</sup> E. Pebay-Peyroula, G. Rummel, J. P. Rosenbusch, and E. M. Landau, *Science* **277**, 1676–1681 (1997).

<sup>&</sup>lt;sup>23</sup> E. M. Landau and J. P. Rosenbusch, *Proc. Natl. Acad. Sci. USA* 93, 14532–14535 (1996).

<sup>&</sup>lt;sup>24</sup> L. J. DeLucas and C. E. Bugg, Trends in Biotechnology 5, 188-193 (1987).

<sup>&</sup>lt;sup>25</sup> J. Dong, T. Boggon, N. Chayen, J. Raftery, R. Bi, and J. Helliwell, Acta Crystallogr. D Biol. Crystallogr. 55, 745-752 (1999).

<sup>&</sup>lt;sup>26</sup> D. C. Carter, B. S. Wright, T. Y. Miller, J. Chapman, P. D. Twigg, K. Keeling, K. Moody, M. White, J. Click, J. Ruble, J. X. Ho, L. Adhock-Downeey, G. Bunick, and J. Harp, J. Cryst. Growth 196, 602-609 (1999).

D. C. Carter, B. S. Wright, T. Y. Miller, J. Chapman, P. D. Twigg, K. Keeling, K. Moody, M. White, J. Click, J. Ruble, J. X. Ho, L. Adhock-Downeey, T. Dowling, C.-H. Chang, P. Ala, J. Rose, B. C. Wang, J.-P. Declerq, C. Evrard, J. Rosenberg, J.-P. Wery, D. Clawson, M. Wardell, W. Stallings, and A. Stevens, J. Cryst. Growth 196, 610-622 (1999).

abound, even in recent literature, 5,29-32 and reflect the fact that a determined effort that will eventually produce suitable crystals may take several years.

Beyond protein single crystal growth, progress in various biochemical and biomedical research and production tasks is impeded by lack of insight into protein nucleation and growth mechanisms. For instance, the slow dissolution rate of protein crystals is used to achieve sustained release of medications, such as insulin, interferon- $\alpha$ , or the human growth hormone. Work on the crystallization of other therapeutically active proteins—e.g., antibodies for foreign proteins—so they can be dispensed in a similar manner, is currently underway. If the administered dose consists of a few, larger, equidimensional crystallites, steady medication release rates can be maintained for longer periods than for doses composed of many smaller crystallites. To achieve such size distributions, crystal nucleation times must be short so that all crystals grow at the same decreasing supersaturation.

Other, more rare biomedical applications relying on insight into the formation of protein solid phases include situations where pathological conditions are related to the formation of crystals or other ordered solid aggregates in the human body. Two often-cited examples are the crystallization of hemoglobin C and the polymerization of hemoglobin S, which cause, respectively, CC and sickle cell diseases. Another example is formation of a denser liquid phase in the eye retina, which underlies the pathology of cataract formation.

<sup>&</sup>lt;sup>41</sup> C. R. Berland, G. M. Thurston, M. Kondo, M. L. Broide, J. Pande, O. Ogun, and G. B. Benedek, *Proc.* 



<sup>&</sup>lt;sup>29</sup> P. Weber, *Advances in Protein Chemistry*, Vol. 41, eds. C. B. Afinsen, F. M. Richards, J. T. Edsal, and D. S. Eisenberg, Academic Press, New York (1991).

<sup>&</sup>lt;sup>30</sup> A. Ducruix and R. Giege (Eds.), Crystallization of Nucleic Acids and Proteins. A Practical Approach, IRL Press, Oxford (1992).

<sup>&</sup>lt;sup>31</sup> N. E. Chayen, T. J. Boggon, A. Casseta, A. Deacon, T. Gleichmann, J. Habash, S. J. Harrop, J. R. Helliwell, Y. P. Neih, M. R. Peterson, J. Raftery, E. H. Snell, A. Hädener, A. C. Niemann, D. P. Siddons, V. Stojanoff, A. W. Thompson, T. Ursby, and M. Wulff, *Quart. Rev. Biophys.* 29, 227–278 (1996).

<sup>&</sup>lt;sup>32</sup> P. C. Weber, *Methods in Enzymology*, Vol. 276, eds. C. W. Carter Jr. and R. M. Sweet, Academic Press, New York (1997), 13–22.

<sup>&</sup>lt;sup>33</sup> J. Brange, Galenics of Insulin, Springer, Berlin (1987).

<sup>&</sup>lt;sup>34</sup> M. L. Long, J. B. Bishop, T. L. Nagabhushan, P. Reichert, G. D. Smith, and L. J. DeLucas, *J. Cryst. Growth* **168**, 233–243 (1996).

<sup>&</sup>lt;sup>35</sup> S. Matsuda, T. Senda, S. Itoh, G. Kawano, H. Mizuno, and Y. Mitsui, J. Biol. Chem. 264, 13381–13382 (1989)

S. Peseta, J. A. Langer, K. C. Zoon, and C. E. Samuel, *Annual Review of Biochemistry*, Vol. 56, eds.
 C. C. Richardson, P. D. Boyer, I. B. Dawid, and A. Meister, Annual Reviews, Palo Alto (1989), 727–778.
 P. Reichert, C. McNemar, N. Nagabhushan, T. L. Nagabhushan, S. Tindal, and A. Hruza, *Metal-Interferon-Alpha Crystals*, in: 5,441,734. (US Patent, 1995).

<sup>&</sup>lt;sup>38</sup> S. Charache, C. L. Conley, D. F. Waugh, R. J. Ugoretz, and J. R. Spurrell, *J. Clin. Invest.* 46, 1795–1811 (1967).

<sup>&</sup>lt;sup>39</sup> R. E. Hirsch, C. Raventos-Suarez, J. A. Olson, and R. L. Nagel, *Blood* 66, 775-777 (1985).

<sup>&</sup>lt;sup>40</sup> W. A. Eaton and J. Hofrichter, *Advances in Protein Chemistry*, Vol. 40, eds. C. B. Anfinsen, J. T. Edsal, F. M. Richards, and D. S. Eisenberg, Academic Press, San Diego (1990), 63–279.

Of all processes of molecular self-assembly in the biological world, the crystallization of the protein molecules is probably the best-studied example. Other examples include the formation of quaternary protein structures, protein and protein nucleic acid complexes whose assembly into structural units is often essential to chains of coupled functions, and assembly of viri and of complex intracellular organelles such as the ribosome. Similar to crystallization, the self-assembly processes are enabled by molecular recognition. Hence, insight into the molecular mechanisms of protein crystallization could provide guidance into these related areas.

In addition to its medical and biotechnological significance, protein crystallization has much in common and may provide an insight into crystallization phenomena that occur in a variety of systems, 45 such as water freezing in clouds and oceans, 46 magma solidification in the Earth's interior, 47 the pulling of semiconductor boules, 48 and so on. Given the resolution limits of modern surface characterization techniques, proteins are particularly attractive for studies of fundamental crystal growth mechanisms. For example, the size of the protein molecules (a few nanometers) and the time-scales for growth (up to a few seconds between sequential discrete molecular attachment events) are within the reach of current advanced experimental techniques. On the other hand, the molecular masses typical of most protein molecules still leave the thermal equilibration times relatively short. Thus, conclusions drawn from studies of protein model systems may still be meaningful for small molecule crystallization. In this regard, proteins could be a better model than, for instance, colloidal crystals. 49,50

The preceding factors led to the emergence of macromolecular crystallization as a distinct area of research in the early 1980s. Since then, the field has benefited from concepts and methods developed in other research areas. For instance, the application of direct light scattering and other methods used to probe colloids led to quantitative measurements of molecular interactions and crystal nucleation in protein solutions. Fluid dynamics analyses were applied to characterize the convective—diffusive supply fields in the solutions from which the crystals



<sup>&</sup>lt;sup>42</sup> N. Ban, P. Nissen, J. Hansen, P. B. Moore, and T. A. Steitz, *Science* **289**, 905-920 (2000).

<sup>&</sup>lt;sup>43</sup> P. Nissen, J. Hansen, N. Ban, P. B. Moore, and T. A. Steitz, *Science* 289, 920-930 (2000).

<sup>&</sup>lt;sup>44</sup> K. Wild, I. Sinning, and S. Cusack, *Science* **294**, 598–601 (2001).

<sup>&</sup>lt;sup>45</sup> F. Rosenberger, Cryst. Res. Technol. **34**, 163-165 (1999).

<sup>&</sup>lt;sup>46</sup> D. W. Oxtoby, J. Phys.: Condensed Matter 4, 7627-7650 (1992).

<sup>&</sup>lt;sup>47</sup> O. Melnik and R. S. J. Sparks, *Nature* **402**, 37–41 (1999).

<sup>&</sup>lt;sup>48</sup> H.-W. Ren, X. Q. Shen, and T. Nishinaga, J. Cryst. Growth 166, 217-221 (1995).

<sup>&</sup>lt;sup>49</sup> P. N. Pusey and W. Van Megen, *Nature* **320**, 340–343 (1986).

<sup>&</sup>lt;sup>50</sup> W. Van Megen and S. M. Underwood, *Nature* **362**, 616–619 (1993).

<sup>&</sup>lt;sup>51</sup> Z. Kam and J. Hofrichter, *Biophys. J.* **50**, 1015–1020 (1986).

<sup>&</sup>lt;sup>52</sup> A. George and W. W. Wilson, Acta Crystallogr. Section D 50, 361-365 (1994).

<sup>&</sup>lt;sup>53</sup> M. Muschol and F. Rosenberger, J. Chem. Phys. 103, 10424–10432 (1995).

<sup>&</sup>lt;sup>54</sup> F. Rosenberger, P. G. Vekilov, M. Muschol, and B. R. Thomas, J. Cryst. Growth 167, 1-27 (1996).

# DOCKET

## Explore Litigation Insights



Docket Alarm provides insights to develop a more informed litigation strategy and the peace of mind of knowing you're on top of things.

## **Real-Time Litigation Alerts**



Keep your litigation team up-to-date with **real-time** alerts and advanced team management tools built for the enterprise, all while greatly reducing PACER spend.

Our comprehensive service means we can handle Federal, State, and Administrative courts across the country.

## **Advanced Docket Research**



With over 230 million records, Docket Alarm's cloud-native docket research platform finds what other services can't. Coverage includes Federal, State, plus PTAB, TTAB, ITC and NLRB decisions, all in one place.

Identify arguments that have been successful in the past with full text, pinpoint searching. Link to case law cited within any court document via Fastcase.

## **Analytics At Your Fingertips**



Learn what happened the last time a particular judge, opposing counsel or company faced cases similar to yours.

Advanced out-of-the-box PTAB and TTAB analytics are always at your fingertips.

### API

Docket Alarm offers a powerful API (application programming interface) to developers that want to integrate case filings into their apps.

### **LAW FIRMS**

Build custom dashboards for your attorneys and clients with live data direct from the court.

Automate many repetitive legal tasks like conflict checks, document management, and marketing.

### **FINANCIAL INSTITUTIONS**

Litigation and bankruptcy checks for companies and debtors.

## **E-DISCOVERY AND LEGAL VENDORS**

Sync your system to PACER to automate legal marketing.

