



Tumor necrosis factor- α in the pathogenesis and treatment of cancer

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The critical pathogenic role of tumor necrosis factor (TNF)a in inflammatory disorders such as rheumatoid arthritis and inflammatory bowel disease is well established. The role played by TNFa in both the treatment and pathogenesis of cancer remains less understood. Recent advances help to create a framework for understanding seemingly paradoxical effects of TNF α as both an anti-tumour agent and a mediator of tumour growth. High pharmacological doses of TNFa combined with chemotherapy can regress otherwise intractable tumours, and efforts continue to optimize delivery to avoid severe toxicities. Mounting evidence demonstrates that pathophysiological concentrations of endogenous TNFa act to promote tumour genesis and growth. The cellular and molecular pathways mediating these phenomena are starting to be clarified. Current data support the continued development of both TNF α and anti-TNF α therapy for clinical treatment of cancers in distinct settings.

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Abbreviations

CLL	chronic lymphocytic leukemia
GVHD	graft versus host disease
IL	interleukin
MDS	myelodysplastic syndrome
MMP	matrix metalloproteinase
NF-κB	nuclear factor-κB
TNF	tumor necrosis factor
TNFR	TNF receptor

Introduction

Tumor necrosis factor (TNF) α is a potent pleiotropic proinflammatory cytokine produced by macrophages, neutrophils, fibroblasts, keratinocytes, NK cells, T and B cells, and tumour cells. TNF α mediates host responses in acute and chronic inflammatory conditions, and is a mediator of protection from infection and malignancy. The biology of the TNE/TNE receptor (TNEP) system has previously been reviewed $[1^{\circ}, 2^{\circ \circ}, 3]$. TNF α mediates tumour regression, and recombinant $TNF\alpha$ is approved in Europe to be administered locoregionally at supraphysiological levels as a therapy for sarcoma [4]. The concept that the major effects of endogenous $TNF\alpha$ in cancer are opposite to the effects observed with high dose TNF α therapy has gained momentum recently [5^{••}]. Instead of causing tumour regression, cancer-derived TNF α can mediate tumour progression by causing the proliferation, invasion and metastasis of tumour cells (see also Update). The paradox that this cytokine is both a 'tumour necrosis factor' and a 'tumour-promoting factor' is lucidly explored by Balkwill [6^{••}]. This paradox can be explained partly by the differences in levels of $TNF\alpha$ in distinct settings. When TNFa is administered therapeutically in extremely high doses, it acts as a vasculotoxic tumour-regressing agent. However, when $TNF\alpha$ is produced by tumours and tumour-associated macrophages or stromal cells at physiological levels, it promotes tumour growth and additional macrophage recruitment, stimulating the elaboration of angiogenic and growth factors from infiltrating cells.

TNFα-mediated cellular signaling

The signaling pathways that determine whether a cell will respond to endogenous TNF by proliferating or by undergoing apoptosis continue to be elucidated. Micheau and Tschopp [7^{••}] reported a new model with evidence for the production of two distinct intracellular complexes after TNF α binds to TNFR1 in HT1080 fibrosarcoma cells. The first complex promotes cell survival by induction of nuclear factor- κB (NF- κB). The second complex, formed substantially later after receptor ligation, is cytoplasmic and spatially removed from the initial receptor complex. The second complex is pro-apoptotic but can be inhibited by anti-apoptotic gene products previously induced via the NF-kB pathway. This system represents a switch that can determine the fate of a given cell receiving a TNF α signal. That fate clearly will vary depending upon the presence or absence of key signaling factors resulting from cell-specific gene expression or the mutational status of the cell.

TNF α as a therapy for cancer

Systemically administered TNF- α was evaluated in the clinic in the 1980s as a therapy for solid tumours, and was found to have severe toxicities, most notably hypotension and organ failure [8]. Studies revealed that the maximally tolerated dose of TNF α was significantly lower than that required to cause anti-tumour effects. The delivery of

is now accomplished by isolated limb perfusion in which the circulation of the limb is connected surgically to a bypass circuit into which drugs are administered [9[•]]. Isolated limb perfusion with $TNF\alpha$ in combination with chemotherapy is approved in Europe for the treatment of locally advanced unresectable soft-tissue sarcomas (for a summary of the clinical studies of isolated limb perfusion with TNFα, see [9[•]]). Positive clinical data have been reported with this TNFa therapy for in-transit melanoma and drug-resistant bony sarcomas [9[•]]. Although effective, this procedure is highly specialized and not widely used. The mechanism of action is believed to involve direct toxic effects on the tumour, without affecting normal vasculature. TNF α causes vessel regression, as well as hemorrhagic necrosis, because it induces endothelial cell apoptosis, resulting in a procoagulant endothelium. TNF α enhances vascular permeability, facilitating the uptake and accumulation of chemotherapeutic drugs such as melphalan and doxorubicin [9[•]], as well as antibodies [10]. TNF α alone is not effective when administered via isolated limb perfusion, suggesting that chemotherapy accumulation is a major mechanism of action. The specific effect of $TNF\alpha$ on angiogenic, but not normal, vasculature might be attributable to the deactivation of the angiogenesis-associated integrin $\alpha v\beta 3$. TNF α in combination with interferon- γ leads to endothelial apoptosis [11].

Many approaches are being used to target therapeutic TNF α directly to tumours, without causing systemic toxicity. These include gene therapy (TNFerade) [12] and TNF α conjugated to targeting peptides or single chain antibody fragments (see also Update) [13–15].

Role of TNF α in the pathogenesis of cancer

In contrast to the use of pharmacologic doses of TNF α as an anti-cancer therapeutic, mounting evidence suggests that TNFa acts to promote tumour growth and progression at physiologically relevant concentrations [16]. Some of the most detailed work regarding the role of $TNF\alpha$ as a tumour promoter was published in a series of papers from the laboratory of Fran Balkwill [16,17[•],18]. The authors used the induction of skin tumours in mice as a model system to understand the growth-promoting actions of TNF α and its signaling pathway. In this system, 9,10-dimethyl-1,2-benzanthracene (DMBA) is used as a topical DNA-damaging tumour inducer; 12-O-tetradecanoylphorbol-13-acetate (TPA) is then applied as a tumour promoter. TNF α was first implicated in tumour growth in this model by the observation that $TNF\alpha$ knockout mice are highly resistant to the generation of skin tumours by this method. The presence of functional TNFa has no effect on DNA mutation rates or tumour initiation, but instead profoundly influences tumour promotion. TPA was shown to induce $TNF\alpha$ in skin keratinocytes of wild-type mice. The production of

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scription pathway, such as granulocyte-macrophage colony-stimulating factor, matrix metalloproteinase (MMP)-3 and MMP-9, is important for the tumour-promoting effect. Anti-tumour effects were also observed following pharmacologic intervention with a neutralizing anti-mouse TNF α antibody. The potential role of TNF α in the growth of human skin cancers, initiated largely by UV radiation, is an important area for future investigation.

Clinical observations supporting a role for TNF α in tumor growth

$TNF\alpha$ plasma levels in cancer patients

Several reports have associated detection of abnormally high levels of TNF α protein in the blood of cancer patients with a wide range of tumour types [19], including pancreatic [20], kidney [21], breast [22], asbestosis induced lung [23] and prostate cancer [24]. Within groups of patients with the same tumour type, higher levels of TNF α have been correlated with advanced tumour stage, greater paraneoplastic complications and shorter survival time. However, circulating TNF α is not always detectable in cancer patients and can vary within individual patients over time and course of disease [25]. Regulation of TNFRs is critical to tumour cell responsiveness to TNF α , and tumour tissue levels of TNF α might be more relevant than blood levels in explaining pro-tumourigenic associations.

Endometriosis and ovarian cancer develop along a continuum of malignant transformation and promotion. Serum TNF α has been associated significantly with endometriomas and malignant, cystic and ovarian cancers, but not with benign tumours [26]. Inflammatory conditions associated with endometriosis and ovarian cancer include exposure to exogenous irritants and ovulation, accompanied by cell proliferation, oxidative stress, vascular permeability, and overproduction of prostaglandins, leukotrienes, TNFα, interleukin (IL)-6 and IL-1 [27^{••}]. mRNA for these cytokines is found in epithelial ovarian tumours and in related ascites. TNFa is prevalent in peritoneal fluid around endometrial foci. Attracted platelets and macrophages secrete vascular endothelial growth factor, MMP-9 and transforming growth factor-\u00df, and promote infiltration of ectopic endometrium and/or invasion and metastasis. The invasive tissue of endometriosis is surrounded by an ineffective immune response, just as ovarian tumours are surrounded by inflammatory cells impotent against the neoplasia. Chronic inflammation might invoke a switch from Th1-dominant to a Th2-dominant microenvironment linked to mutagenesis. Progression can be modified by immunosuppression, suggesting a therapeutic role for anti-inflammatory agents.

Esophageal metaplasia (Barrett's oesophagus), in the setting of chronic inflammation can progress to adenocarcinoma (Barrett's adenocarcinoma). Tselepsis *et al.*

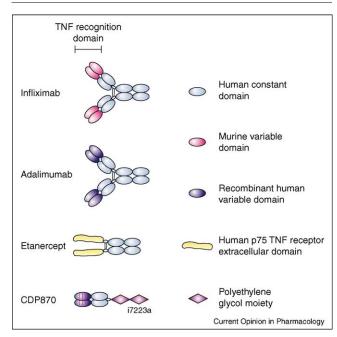
absent in normal gastric and esophageal squamous mucosae, but increased in sections from metaplasia through dysplasia to carcinoma. Elevated TNF α immunoreactivity in non-dysplastic Barrett's oesophagus is localized to mucosal regions of prominent lymphoid infiltrate where metaplastic stem cells are located. Within individual glands, the most dysplastic cells express TNF α , and TNFR1 levels are increased during disease progression, potentially amplifying the TNF α signal. This study demonstrated a novel signaling pathway for oncogene activation by an inflammatory cytokine.

In chronic lymphocytic leukemia (CLL), neoplastic lymphocytes release TNF α spontaneously *in vitro*, and leukaemic lymphocytes are more proliferative and viable when exposed to TNF α . TNF α in CLL patients correlated with extent of disease, serum β_2 -microglobulin, low hemoglobin and low platelets, as well as with karyotype. TNF α levels were predictive of survival, independent of staging, β_2 -microglobulin, hemoglobin, and white blood cell and platelet counts [29^{••}].

Anti-TNFα therapy in cancer

Tsimberidou and Giles [30] review potential clinical indications for agents that block or inactivate TNF α (Figure 1), specifically the soluble TNF α receptor fusion protein etanercept and the anti-TNF α antibody infliximab, in multiple myeloma, myelodysplastic syndrome

Figure 1



Multiple anti-TNF agents are currently marketed or in clinical development. Several anti-TNF biologics are illustrated above, including antibodies (infliximab and adalimumab), the modified antibody fragment CDP870, and the soluble receptor etanercept. The various domains present in the structures are illustrated for

(MDS), acute myelogenous leukemia and myelofibrosis [30]. The limited efficacy of monotherapy points to the need to optimize dose and schedule, to combine anti-TNF α agents with active biological or cytotoxic agents, and to understand individual patient proteomic patterns of disease and gene polymorphisms that could predict response.

Investigative and clinical uses of TNF α blockade in graft versus host disease (GVHD) are founded on the established role of upregulated TNF α as a consequence of conditioning regimens and as an effector molecule in multi-organ reactivity to allogeneic graft cells [31,32°]. Infliximab has shown activity in refractory acute and chronic GVHD [33]. With added immunosuppression and risk of serious infections, vigilance is required when using anti-TNF agents in patients with GVHD [34]. No studies yet have elucidated the kinetics, optimal dose and schedule of anti-TNF agents in GVHD [35].

Anti-TNF α agents can inhibit the excessive apoptosis in haematopoietic cells, which is suspected as the cause of cytopenias in MDS. The hyperproliferative marrow of MDS, with excessive apoptosis and overexpression of TNF α , compels intervention to modulate the dysregulated cytokine milieu in haematopoietic tissues. Therapeutic activity of infliximab suggests that the microenvironment of the marrow is more directly affected than are the dysplastic haematopoietic cells [36,37]. A logical next step would be to use a specific anti-TNF agent to modulate cell-cell signaling and stromal interactions in the marrow, combined with a cytotoxic agent known to be active against the myelodysplastic clone. Etanercept was tested in a Phase I clinical study in combination with IL-2 to determine if it could modulate the biological effects, and reduce toxicity, of high-dose IL-2 administration. TNFα bioactivity was inhibited, and the polymorphonuclear leukocyte chemotactic defect normally seen with IL-2 was not observed. Increases in levels of C-reactive protein, IL-6, IL-8 and IL-1 receptor antagonist were partially suppressed relative to historical controls [38].

A pilot study of etanercept in patients with refractory multiple myeloma reported tolerability but no objective responses, and some evidence of acceleration of disease soon after starting therapy [39]. TNF α was increased during treatment compared with pretreatment values; this is probable evidence of accumulation of the bound cytokine. More research on mRNA expression in the myeloma and stromal cells might determine if the bioactivity of increased TNFa was attenuated fully. Infliximab and etanercept differ in function: etanercept binds and blocks both TNF α and lymphotoxin (TNF β), whereas infliximab binds only to TNFa. Infliximab, but not etanercept, binds to soluble TNFa bound to cell-surface C.L.LL TNE C 1 ...1

infliximab; TNF α dissociates more rapidly from etanercept, releasing bioactive TNF α [40]. Given these differences and the expectation that blocking TNF α should show anti-myeloma activity [41], it could be worthwhile to investigate infliximab in this disease.

Potential risks of anti-TNF-α therapy

Because of the adaptive protective purpose of inflammation, pharmacological inhibition of this proinflammatory cytokine could have adverse effects in the host, unrelated to the target disease of the anti-TNFa therapy. The risk of reactivation of latent tuberculosis is addressed in the prescribing information for infliximab (REMICADE®; Centocor Inc, Malvern, PA). Preemptive systemic antifungal therapy is recommended for patients receiving infliximab for treatment of GVHD [34]. Smith and Skelton [42] reported cases of squamous cell carcinoma that became evident and grew rapidly during an initial period of etanercept therapy for rheumatoid arthritis. The tumours might have been present, but occult before disruption of immunological control. Etanercept could disable innate anti-tumour surveillance by blockade of both lymphotoxin α and the cytotoxic effects of TNFα. Additional proposed mechanisms include the inhibition of the Th1 cytokine pattern and impairment of cytotoxic T cells. All cases were in chronically UV-damaged actinic skin predisposed to tumourigenesis by long-term, low-level production of TNFa. No new squamous cell carcinomas developed in patients who continued treatment for more than one year, suggesting prolonged anti-TNFa therapy could be preventive of cutaneous malignancies.

Pharmacovigilance data on etanercept, infliximab and adalimumab were reviewed by the FDA in 2003, with a focus on lymphoproliferative disease in patients treated with these anti-TNF α agents, relative to the rate expected in populations with immune-mediated diseases [43]. The potential role of TNF α -blocking therapy in the development of malignancies is not known. A prospective study of 18 572 patients with rheumatoid arthritis treated with anti-TNF α therapy plus methotrexate reported an increased standard incidence ratio compared with patients not receiving methotrexate or biologics, but confidence intervals overlapped for all treatments [44]. Patients with highly active disease and/or chronic exposure to immunosuppressant therapies could have severalfold higher risk for development of lymphoma, thus caution should be exercised when considering anti-TNFa agents in patients either with a history of malignancy or who develop malignancy during treatment.

The FDA also reported on the risk of histoplasmosis [45], lymphoma [46] and/or listeriosis [47]. The Mayo clinic reviewed the safety of infliximab in 500 patients with Crohn's disease treated with infliximab [48]. The biolo-

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consideration of the benefit to risk ratio when prescribing anti-TNF therapies.

Anti-TNFa therapy in cancer supportive care

In addition to direct anti-tumour effects, anti-TNF α therapies are being tested for supportive care indications. These include cancer-related cachexia, fatigue, depression, amelioration of chemotherapy-induced toxicities and metastatic bone pain [49,50]. TNF α has long been associated with cachexia, and trials are underway testing anti-TNF α agents in this condition [51,52]. Ramesh and Reeves [53] recently demonstrated a role for TNF α in a mouse model of cisplatin-mediated renal injury. Treatment of animals with TNF α synthesis inhibitors or anti-TNF α antibodies prevented kidney damage in the model.

TNF α appears to play a complex role in side effects of radiotherapy, and anti-TNF α treatments could be useful in their management [54,55]. TNF α is also a known activator of osteoclasts [56] and mediator of neuropathic pain [57]. An intriguing pair of clinical cases, in which etanercept was used to treat refractory metastatic bone pain, suggest that anti-TNF α agents may ultimately play a role in controlling cancer pain [58°]. The potential value of anti-TNF α agents in these debilitating conditions presents broad opportunities to improve cancer care.

Conclusions

It is likely that there is no 'right' or 'wrong' answer to the question of whether TNF α is good or bad for the tumour. Research should continue to optimize the use of pharmacological TNF α to treat cancer, as well as the use of neutralizing therapies to inhibit the effects of pathophysiologically derived TNF α . As we increase our understanding of the complex biology of TNF α in differing contexts within oncology and hematology, these opposing approaches should prove more effective against cancer.

Update

Recent results further support the anti-tumor effectiveness of TNF α when delivered by gene therapy approaches. Hecht *et al.* [59] reported preliminary evidence for dose-dependent improvement in progressionfree survival in a phase I/II trial of TNFerade in combination with radiation and 5-flourouracil in pancreatic cancer patients. Zarovni *et al.* [60] demonstrated substantially improved anti-tumor efficacy in animal models with tumor vessel-targeted peptide–TNF α fusion proteins. The molecules were delivered by intramuscular injection of cDNA expression vectors.

Additional evidence demonstrating the pro-metastatic effects of $\text{TNF}\alpha$ has been published recently. Mochizuki *et al.* [61] reported that $\text{TNF}\alpha$ promoted metastasis of

wall in a mouse model. Hagemann *et al.* [62] showed that macrophage-induced invasion by human breast cancer cells was mediated by MMPs and was dependent upon TNFR activity.

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References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- 1. Palladino MA, Bahjat FR, Theodorakis EA, Moldawer LL:
- Anti-TNF-alpha therapies: the next generation. Nat Rev Drug Discov 2003, 2:736-746.

A comprehensive review of the biology of TNF α , the role of TNF α in human disease and animal models, and the use and comparison of TNF α antagonists approved and in clinical development.

- 2. Younes A, Kadin ME: Emerging applications of the tumor
- necrosis factor family of ligands and receptors in cancer therapy. J Clin Oncol 2003, 21:3526-3534.

This comprehensive, thoughtful and provocative review focuses on clinical applications of four TNF family member-pairs for cancer treatment: CD30/CD30 ligand, CD40/CD40 ligand, receptor activator of nuclear factor-kB (RANK)/RANK ligand, and TNF-related apoptosis-inducing ligand (TRAIL)/Apo-2L receptors. Structures, roles and pathways are clearly and succinctly illustrated. The 252 references (a tour de force) provide the best foundation currently available on which to build understanding of the biology of these molecules relevant to the growth and, potentially, the interventional eradication of cancer cells.

- Younes A, Aggarwall BB: Clinical implications of the tumor necrosis factor family in benign and malignant hematologic disorders. Cancer 2003, 98:458-467.
- Eggermont AM, ten Hagen TL: Tumor necrosis factor-based isolated limb perfusion for soft tissue sarcoma and melanoma: ten years of successful antivascular therapy. *Curr Oncol Rep* 2003, 5:79-80.
- Dranoff G: Cytokines in cancer pathogenesis and cancer
 therapy. Nat Rev Cancer 2004, 4:11-22.

An excellent review of the conflicting role of cytokines in inhibiting tumour development and progression, as well as in promoting tumour growth and mediating invasion and metastasis. The cell sources and roles for endogenous cytokines in tumour progression, as well a listing of cytokines as cancer therapy, are presented in table form.

6. Balkwill F: Tumor necrosis factor or tumor promoting factor?

Cytokine Growth Factor Rev 2002, 13:135-141.
 An excellent and very thorough review of evidence for both the pro- and anti-tumour effects of TNFα.

7. Micheau O, Tschopp J: Induction of TNF receptor I-mediated
 apoptosis via two sequential signaling complexes.

Cell 2003, **114**:181-190. Provides a new model for the cellular and molecular mechanisms determining the fate of cells following TNF α binding.

- Lejeune FJ, Ruegg C, Lienard D: Clinical applications of TNF-alpha in cancer. Curr Opin Immunol 1998, 10:573-580.
- Eggermont AM, de Wilt JH, ten Hagen TL: Current uses of isolated
 limb perfusion in the clinic and a model system for new strategies. *Lancet Oncol* 2003, 4:429-437.

A review of the use of drugs, including $\text{TNF}\alpha$, with delivery via isolated limb perfusion. Includes a summary of response rates for the treatment of in-transit melanoma metastases and soft-tissue sarcomas, and reviews preclinical research being performed to advance the clinical applications of this technology.

 Folli S, Pelegrin A, Chalandon Y, Yao X, Buchegger F, Lienard D, Lejeune F, Mach JP: Tumor-necrosis factor can enhance radio-antibody uptake in human colon carcinoma xenografts by increasing vascular permeability. *Int J Cancer* 1993,

- 11. Ruegg C, Yilmaz A, Bieler G, Bamat J, Chaubert P, Lejeune FJ: Evidence for the involvement of endothelial cell integrin alphaVbeta3 in the disruption of the tumor vasculature induced by TNF and IFN-gamma. *Nat Med* 1998, **4**:408-414.
- Senzer N, Mani S, Rosemurgy A, Nemunaitis J, Cunningham C, Guha C, Bayol N, Gillen M, Chu K, Rasmussen C *et al.*: TNFerade biologic, an adenovector with a radiation-inducible promoter, carrying the human tumor necrosis factor alpha gene: a phase I study in patients with solid tumors. *J Clin Oncol* 2004, **22**:592-601.
- 13. Curnis F, Sacchi A, Corti A: Improving chemotherapeutic drug penetration in tumors by vascular targeting and barrier alteration. *J Clin Invest* 2002, **110**:475-482.
- 14. Curnis F, Gasparri A, Sacchi A, Longhi R, Corti A: Coupling tumor necrosis factor-alpha with alphaV integrin ligands improves its antineoplastic activity. *Cancer Res* 2004, **64**:565-571.
- Borsi L, Balza E, Carnemolla B, Sassi F, Castellani P, Berndt A, Kosmehl H, Biro A, Siri A, Orecchia P *et al.*: Selective targeted delivery of TNFalpha to tumor blood vessels. *Blood* 2003, 102:4384-4392.
- Szlosarek PW, Balkwill FR: Tumour necrosis factor alpha: a potential target for the therapy of solid tumours. Lancet Oncol 2003, 4:565-573.
- Scott KA, Moore RJ, Arnott CH, East N, Thompson RG, Scallon BJ,
 Shealy DJ, Balkwill FR: An anti-tumor necrosis factor-alpha antibody inhibits the development of experimental skin tumors. *Mol Cancer Ther* 2003, 2:445-451.

This paper establishes the anti-tumour effects of an anti-TNF antibody in a well-defined model system.

- Arnott CH, Scott KA, Moore RJ, Robinson SC, Thompson RG, Balkwill FR: Expression of both TNF-alpha receptor subtypes is essential for optimal skin tumour development. Oncogene 2003.
- Mantovani G, Maccio A, Mura L, Massa E, Mudu MC, Mulas C, Lusso MR, Madeddu C, Dessi A: Serum levels of leptin and proinflammatory cytokines in patients with advanced-stage cancer at different sites. J Mol Med 2000, 78:554-561.
- Karayiannakis AJ, Syrigos KN, Polychronidis A, Pitiakoudis M, Bounovas A, Simopoulos K: Serum levels of tumor necrosis factor-alpha and nutritional status in pancreatic cancer patients. *Anticancer Res* 2001, 21:1355-1358.
- Yoshida N, Ikemoto S, Narita K, Sugimura K, Wada S, Yasumoto R, Kishimoto T, Nakatani T: Interleukin-6, tumour necrosis factor alpha and interleukin-1beta in patients with renal cell carcinoma. Br J Cancer 2002, 86:1396-1400.
- Leek RD, Landers R, Fox SB, Ng F, Harris AL, Lewis CE: Association of tumour necrosis factor alpha and its receptors with thymidine phosphorylase expression in invasive breast carcinoma. Br J Cancer 1998, 77:2246-2251.
- Partanen R, Koskinen H, Hemminki K: Tumour necrosis factor-alpha (TNF-alpha) in patients who have asbestosis and develop cancer. Occup Environ Med 1995, 52:316-319.
- Pfitzenmaier J, Vessella R, Higano CS, Noteboom JL, Wallace D Jr, Corey E: Elevation of cytokine levels in cachectic patients with prostate carcinoma. *Cancer* 2003, 97:1211-1216.
- Bossola M, Muscaritoli M, Bellantone R, Pacelli F, Cascino A, Sgadari A, Battaglia F, Piccioni E, Scambia G, Doglietto GB et al.: Serum tumour necrosis factor-alpha levels in cancer patients are discontinuous and correlate with weight loss. *Eur J Clin Invest* 2000, **30**:1107-1112.
- Darai E, Detchev R, Hugol D, Quang NT: Serum and cyst fluid levels of interleukin (IL) -6, IL-8 and tumour necrosis factoralpha in women with endometriomas and benign and malignant cystic ovarian tumours. *Hum Reprod* 2003, 18:1681-1685.
- 27. Ness RB: Endometriosis and ovarian cancer: thoughts on
 shared pathophysiology. Am J Obstet Gynecol 2003, 189:280-294.

In this excellent and comprehensive review, the author develops and explores a unifying concept of local hormonal and inflammatory conditions existent within a causal pathway of malignant transformation.

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