

NEW TOOLS IN SYNTHESIS

Pro-Nucleotides - Recent Advances in the Design of Efficient Tools for the Delivery of Biologically Active Nucleoside Monophosphates

C. Meier*

Institut für Organische Chemie, Julius-Maximilians-Universität Würzburg, Am Hubland, 97074 Würzburg, Germany

Tel. +49-931-8885326; Fax +49-931-8884606; e-mail: meier@chemie.uni-wuerzburg.de

Received 10 September 1997

Dedicated to Professor Gernot Boche on the occasion of his 60th birthday

Abstract: A summary of the most recent advances to the design of pro-nucleotides will be presented. Approaches that have been designed to be activated by enzymes such as carboxyesterases [bis(POM)-, bis(POC)-, bis(SATE)-, bis(AB) phosphotriesters and the arylphosphoramidates] or by reductases [bis(SDTE) approach] will be discussed as well as the amino acids phosphoramidate diester concept with its still unknown delivery mechanism and the *cycloSal* approach that releases the nucleotides by an induced tandem reaction.

Nucleoside analogues, e.g. 2',3'-dideoxy-2',3'-dideoxythymidine **1** (d4T), 3'-azido-2',3'-dideoxythymidine **2** (AZT) or 5-fluoro-2'-deoxyuridine **3** (5-FdU), are structurally different as compared to the corresponding natural DNA or RNA nucleosides with regard to modification of the glycon as well as the aglycon residue. Due to this modified structure, these compounds are widely used as antiviral or antitumor drugs in chemotherapy (Figure 1)⁽¹⁾. Since the discovery of AZT **2** as the first nucleoside drug for the treatment of AIDS, considerable efforts have been made to develop new nucleoside analogues that would be more active, less toxic inhibitors of the HIV-1 reverse transcriptase (RT)⁽²⁾. The general mode of action of nucleoside analogues is the inhibition of the HIV-1 RT by acting as competitive inhibitors or as DNA chain terminators. To act as DNA chain termination agents or RT inhibitors, intracellular conversion of the nucleoside analogues into their 5'-mono-, 5'-di- or 5'-triphosphates is a prerequisite after cell penetration⁽³⁾. The enormous disparity in anti-HIV activity that is evident for a large number of dideoxynucleoside analogues belies their apparent structural similarity. Due to these structural differences as compared to natural nucleosides the metabolization to the corresponding dideoxynucleoside triphosphates is often inefficient and consequently the therapeutic efficacy is sometimes limited⁽⁴⁾. For example, in the case of the anti-HIV active dideoxynucleoside analogue d4T **1** (Stavudine, Zerit[®])⁽⁵⁾ the first phosphorylation to d4T 5'-monophosphate **4** catalyzed by thymidine kinase (TK) is the rate-limiting step in human cells⁽⁶⁾. More striking, however is 2',3'-dideoxyuridine triphosphate (ddUTP) which is one of the most powerful and selective inhibitors of HIV reverse transcriptase ($K_i = 0.05 \mu\text{M}$) while the parent nucleoside 2',3'-dideoxyuridine **5** (ddU) is virtually ineffective at blocking HIV infection in cultured cells. Biochemical and pharmacological studies in three different human T cell lines (CEM, ATH8 and Molt-4) showed that ddU **5** itself was not anabolized to the 5'-monophosphate, most apparently because it was a poor substrate for cellular nucleoside kinases because of the considerable substrate specificity of these enzymes⁽⁷⁾. In contrast, in a few cases the limited efficacy is also due to a catabolic enzymatic reaction. For example, 2',3'-dideoxyadenosine **6** (ddA) is rapidly intracellularly deaminated to dIdI **7** by adenosine deaminase (ADA)^(8,9). As a consequence, dIdI **7** has to be converted into its ultimate bioactive metabolite ddATP via ddAMP **8** by five enzymatic steps (5'-nucleotidase, adenylosuccinate synthase, adenylosuccinate lyase and

two nucleotide kinases)^(8,9). Finally, the resistance of the human immunodeficiency virus to the clinically used antiviral dideoxynucleoside AZT **2** (Zidovudine, Retrovir[®])⁽¹⁰⁾ is on the one hand directly related to multiple point mutations within the HIV-1 reverse transcriptase gene of the virus but may on the other hand be also due to lower susceptibility of resistant target cells to the drug related with a decreased activity or inability of the enzyme thymidine kinase to phosphorylate AZT **2** to the dideoxynucleoside monophosphate AZTMP **9**⁽¹¹⁾.

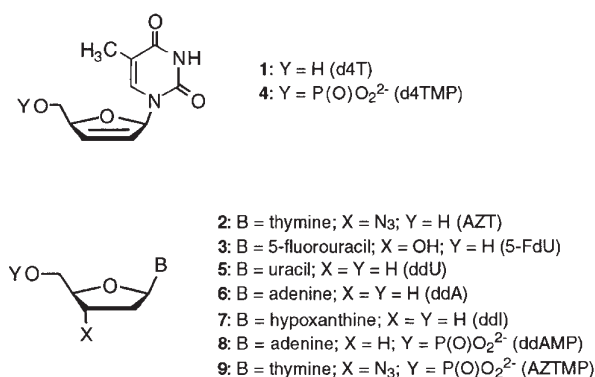
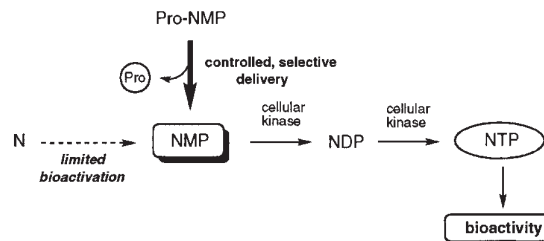


Figure 1. Examples of nucleosides and nucleotides used as antiviral and antitumor drugs

Consequently, direct administration of the nucleotides d4TMP **4**, ddAMP **8** and AZTMP **9** should bypass these limiting steps and hence has advantages for the biological activity. Unfortunately, because of the high polarity of the nucleoside monophosphates (nucleotides), these compounds are not able to easily penetrate cellular membranes or the blood-brain barrier. However, the phosphate moiety offers a suitable site to attach degradable lipophilic carrier residues. As a result, one effort to improve the therapeutic potential of nucleoside analogues is the delivery of the corresponding nucleotide from neutral, membrane-permeable prodrugs (*Pro-Nucleotide Approach*; Figure 2)⁽¹²⁾.



N: Dideoxynucleoside analogue

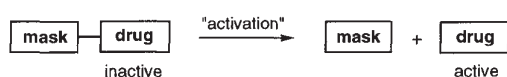
Figure 2. General principle of the pro-nucleotide approach of biologically active dideoxynucleotide analogues

So, a lipophilic phosphotriester may penetrate into the target cell where first partial and at the end complete hydrolysis delivers the nucleotide.

A suitable nucleotide prodrug has to fulfill two requirements: i) it has to be lipophilic enough for passive diffusion of the membrane and blood-brain barrier; ii) furthermore, it should be able to deliver the nucleoside hydrolytically or enzymatically leaving a non-toxic masking group⁽¹³⁾.

In principle, two different concepts for prodrug design are known: bipartate and tripartate prodrugs. In the former concept the drug is modified by a one-component masking group. In this form the drug is biologically inactive. After a simple cleavage of the mask, the active drug is liberated (Figure 3). In the latter concept, the drug is modified by a two-component masking group. Again, the drug is biologically inactive in this bound form. The mechanism of liberation involves a first chemical or enzymatic reaction under cleavage of part I of the masking moiety. The drug is still inactive but the effect of this first reaction is an activation of the remaining masking group II with the consequence of a spontaneous successive cleaving reaction releasing the now bioactive drug (Figure 3)⁽¹⁴⁾.

The bipartate prodrug concept



The tripartate prodrug concept

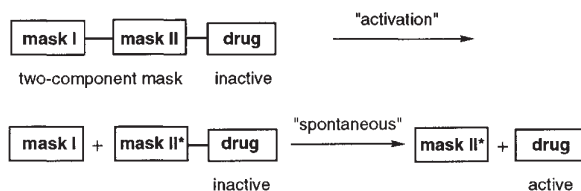


Figure 3. General principle of prodrugs bearing one-component or two-component masking groups

In the case of a nucleotide prodrug one should take into account that under physiological conditions two negatively charged phosphate oxygen's have to be masked in order to obtain a neutral, lipophilic phosphate ester. Consequently, not only one masking group is necessary but two. So, the efficient intracellular delivery of nucleotides from a prodrug requires the existence of a specific delivery mechanism or different rates of conversion of the prodrug to the drug intracellularly versus extracellularly. One comment with respect to toxic side events should be given. Neutral phosphorus derivatives with a good leaving

group attached to the phosphorus are known to be toxic suicide inhibitors of acetylcholinesterase⁽¹⁵⁾. The anti-acetylcholinesterase activity of phosphorus derivatives is an inverse function of the pK_a of the leaving group on the phosphorus atom and parallels the rate of spontaneous hydrolysis by P-O bond cleavage⁽¹⁶⁾. In order to circumvent the possible problem of anti-cholinesterase activity, neutral phosphate ester prodrugs should be designed to undergo heterolytic cleavage of the C-O bond rather than the P-O bond of the ester.

Many strategies have been developed to achieve this goal. As a general motive, uncharged nucleotide triesters are used as membrane-permeable nucleotide precursors⁽¹²⁾. The major differences of these approaches are the delivery mechanisms to the nucleotides. First attempts have been made with simple dialkyl phosphotriesters. These compounds generally belong to the class of bipartate prodrug systems. After a first, sometimes selective hydrolysis of the phosphotriester via a nucleophilic reaction at the phosphorus center, the resulting phosphodiester is often extremely stable against a further chemical hydrolysis due to the charge at the phosphate which prevents a second nucleophilic reaction⁽¹⁷⁾. Even if the subsequent hydrolysis is possible, one should take the pseudorotation phenomenon into account that excludes a selective delivery of the nucleotide⁽¹⁸⁾. As a consequence, almost all approaches based on chemical hydrolysis reported so far were unable to deliver the nucleotide selectively except the *cycloSal* approach that will be discussed later. For this reason, the newer pro-nucleotide approaches are based on the concept of a tripartate prodrug system⁽¹⁴⁾ and are based on the general idea of a selective chemical or enzymatic reaction within the masking group which leads to a second, spontaneous successive reaction yielding the charged phosphate ester. These approaches utilize and exploit the differences in reducing potentials, enzyme activity, and pH value. The concepts working with enzymatic trigger processes (bis(POM)-, bis(POC)-, bis(DTE)-, bis(SATE)-, bis(AB)- and the arylphosphoramidate concept) as well as the *cycloSal* approach based on a pH-driven degradation have demonstrated the successful intracellular delivery of free nucleotides from lipophilic precursors.

It should be added, that it is not the intention of the author to give an entire overview of the pro-nucleotide field. The current review highlights some approaches that have been designed to deliver the nucleotide selectively by a special mechanism. With this selection, the author wants to point out that designing a delivery mechanism and bypassing a certain limiting metabolism the inherent biological potential of an already known nucleoside analogue could be used to a higher extent and consequently, the improvement in antiviral activity could be, in many cases, better than synthesizing new potential nucleoside analogues. Furthermore, this review is restricted to the delivery mechanism and so the synthesis of the compounds may be gleaned from the original literature.

Biographical Sketch



Chris Meier, born 1962 in Berlin, was trained in Chemistry at the University of Marburg. He passed his Diploma and Ph.D. thesis in the group of Prof. G. Boche in Organic Chemistry. Then he moved as post-doc fellow to the Pasteur-Institute in Paris where he started his work in nucleoside and oligonucleotide chemistry. In 1991 he joined the group of Prof. J.W. Engels at the University of Frankfurt/Main starting his Habilitation which he finished in July '96 and in May '97 he was appointed as an associate professor at the University of Würzburg.

Bis(POM)- and Bis(POC)-Nucleotides

The first approach reported by D. Farquhar et al. towards a new class of tripartate pro-nucleotides are the bis(*pivaloyloxymethyl*)- [POM] phosphotriesters of general type **10** (shown exemplified as the ddU derivative)^(19,20). This approach utilizes a carboxyesterase-catalyzed cleavage of the pivaloyl ester within the POM-masking group to yield the highly reactive hydroxymethyl phosphotriester **11** which subsequently eliminates spontaneously formaldehyde to give the mono(POM) phosphodiester **12**. The carboxyesterase which is used for this activation process may be more prevalent inside the cells. To obtain the free nucleotide, this enzymatic activation has to be repeated via **13** or, alternatively, a phosphodiesterase cleaves the phosphodiester **12** directly to yield the nucleotide (Figure 4).

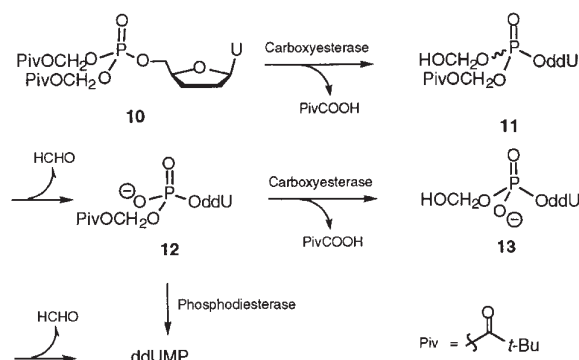


Figure 4. Bis(POM)-nucleotides **10** shown here as their 2',3'-dideoxyuridine derivatives and their degradation pathway

This approach has been applied successfully to the delivery of 2',3'-dideoxyuridine monophosphate from bis(POM)-ddUMP⁽²¹⁾. Furthermore, this concept has been used for the delivery of the anti-HIV drug AZTMP⁽²²⁾, the anti-herpes and anti-HIV drug phosphonomethoxyethyladenine (PMEA)⁽²³⁾ and for the antitumor active drug 5-fluoro-2'-deoxyuridine monophosphate (5-FdUMP)⁽²⁴⁾. In the case of the phosphonate analogue PMEA a highly improved bioavailability was obtained. Moreover, the bis(POM) derivatives of (*R*)-9-(2-phosphonomethoxypropyl)adenine (PMPA) and (*R*)-9-(2-phosphonomethoxypropyl)diaminopurine (PMPDAP) were found to be 9- to 23-fold more active than the parental compounds⁽²³⁾. However, the cytotoxicity of the bis(POM) analogues was also increased by virtually similar degrees. Nevertheless, studies with radiolabelled bis(POM)-PMEA showed a considerable increase (100-fold) of the cellular uptake of PMEA by using the phosphonate diester⁽²³⁾. In an independent study, ten different PMEA prodrugs tested *in vivo* for oral bioavailability in cynomolgus monkeys. From the results of this study, bis(POM)-PMEA was selected for human clinical trials. In these clinical trials, the oral bioavailability of bis(POM)-PMEA from a single 500 mg dose was greater than 40% in fed subjects⁽²⁵⁾. The *in vitro* antitumor tests of the bis(POM)-5-FdUMP triester demonstrated high growth inhibition in cell lines that were resistant to the usually used drug 5-fluorouracil. Furthermore, this triester derivative showed *in vivo* bioactivity after intraperitoneal application against P 388 leukemia in mice. Beside the antitumor activity, these results verified also the penetration of the pro-nucleotide through cellular membranes⁽²⁴⁾. A drawback of this approach is the requirement of a second identical activation reaction because the intermediate phosphodiester is a significantly poorer substrate for the activating carboxyesterases. Moreover, the delivery of one molecule of the nucleotides results in the liberation of two equivalents of potentially toxic formaldehyde and pivalinic acid (Figure 4). Additionally, it has

been shown that the bis(POM) phosphotriesters **10** were chemically unstable and highly susceptible to serum-mediated hydrolysis, factors which limit their potential utility for intracellular drug delivery^(22,24a).

A modification of the bis(POM)-approach and a way to overcome the limitations observed with these compounds are bis(isopropylloxycarbonyloxymethyl)- [bis(POC)] nucleotides of general type **14** (shown as the PMPA derivative) that have been published very recently by A. Fridland et al.⁽²⁶⁾ and L. Naesens et al.⁽²⁷⁾. This modification uses a carbonate diester within the masking group. The mechanism of action is again a carboxyesterase-catalyzed cleavage of the isopropyl ester to give isopropanol and in the shown example the intermediate phosphonate diester **15** that fragmentates via **16** into carbon dioxide and formaldehyde and finally to the mono(POC) phosphonate ester **17**. The mono(POC) ester **17** is subsequently degraded to yield PMPA or generally the nucleotide after a second carboxyesterase activation or by phosphodiesterase cleavage (Figure 5).

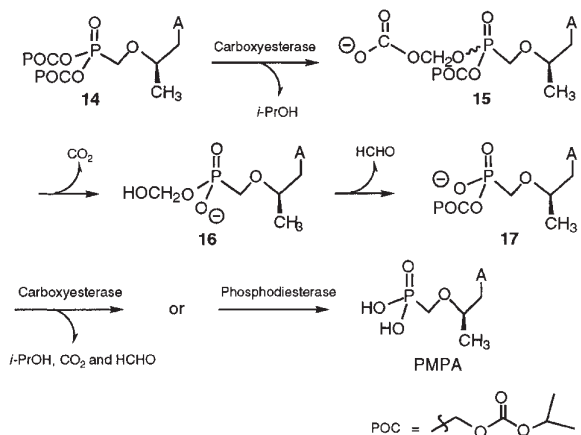


Figure 5. Bis(POC)-nucleotides **14** and their esterase-catalyzed degradation pathway to yield (*R*)-9-(2-phosphonylmethoxypropyl)-adenine as a representative drug

In contrast to the bis(POM)-approach, the bis(POC) modification avoids the formation of two equivalents of pivalinic acid that accumulate in the cells and potentially cause toxicity. The bis(POC)-approach was applied to the anti-HIV drug PMPA in order to increase the low bioavailability of this phosphonate derivative (Figure 5). Bis(POC)-PMPA is chemically stable at low pH and has shown 30% bioavailability in dogs with minimal toxicity in repeat 5-day dosage of 60 mg/kg/day. The anti-HIV activity of bis(POC)-PMPA **14** in human peripheral blood lymphocytes and in dendritic T-cell coculture system was 35- and 16-fold, respectively, higher than that of PMPA. Bis(POC)-PMPA **14** was non-toxic at concentrations that completely suppressed viral replication. Studies of the metabolism of [³H]-bis(POC)-PMPA showed that it was readily taken into the human cells, hydrolyzed to PMPA, and phosphorylated to the mono- and diphosphate derivatives. These results show that bis(POC)-PMPA **14** is a membrane-permeable form of PMPA and shows promise as a drug for the treatment of HIV infections⁽²⁶⁾.

Bis(SDTE)- and Bis(SATE)-Nucleotides

Two further approaches that are also based on enzymatic activation have been reported by J.-L. Imbach and G. Gosselin⁽²⁸⁾. They designed the bis(*S*-[2-hydroxyethylsulfidyl]-2-thioethyl)- [bis(SDTE)] **18** and the bis(*S*-acyl-2-thioethyl)- [bis(SATE)] nucleotides **19**. The former concept was constructed to take advantage of the greater reducing potential within the cells to liberate the nucleotide into the cytosol. After

the reductase cleavage of the disulfide bond in **18** with formation of thioethanol and thioethyl phosphotriester **20**, this compound eliminates spontaneously episulfide to yield the intermediate phosphodiester **21**. As in the case of the bis(POM)- (**10**) and the bis(POC) phosphotriester **14**, the bis(SDTE) derivative **18** requires a second identical enzyme-catalyzed activation step (Figure 6). The initial idea of this concept was, that the "soft" thiol-nucleophile in **20** will attack the "soft" electrophilic α -carbon atom rather than the "hard" phosphorus atom in the elimination reaction avoiding possible pseudorotation processes⁽¹⁸⁾ because all reactions take place within the masking group without involving the phosphate ester moiety. This concept belongs also to the class of tripartate prodrugs. It could be shown that the bis(SDTE) triester of the nucleotides ddUMP and AZTMP **9** were cleaved 30-fold faster by reductases in cell extracts than in culture medium^(28c). Furthermore, the bis(SDTE) approach has been applied to the nucleoside 5-FdU **3**⁽²⁹⁾ and to the phosphonate nucleotide PMEAs⁽³⁰⁾. Whereas an improvement in antiviral activity most probably due to an enhanced bioavailability for the bis(SDTE)-PMEA derivative **18** compared with PMEAs was found, the bis(SDTE)-5-FdUMP failed to show better antitumor potency as compared to the free nucleoside⁽²⁹⁾. The major limitation of this approach is the restricted chemical stability and high susceptibility to serum-mediated hydrolysis (Figure 6).

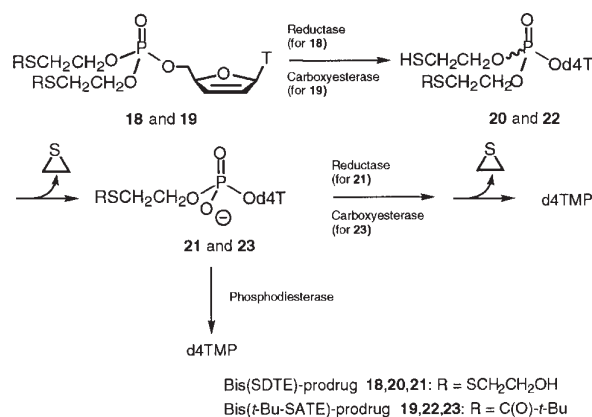


Figure 6. Bis(SDTE)- and bis(SATE)-nucleotides **18** and **21** as their 2',3'-dideoxy-2',3'-didehydrothymidine monophosphate (d4TMP) derivatives and the enzymatic delivery mechanism

In contrast to the bis(SDTE) approach, the bis(SATE) approach is similar to the bis(POM) concept with respect to the activation process. Again, carboxyesterases are used to trigger the nucleotide delivery. Here, the enzyme cleaves a thioester in **19** to yield a carboxylic acid (as in the bis(POM) approach pivalinic acid) as well as the above mentioned thioethyl phosphonate diester **22** which undergoes a fragmentation to episulfide and the phosphonate monoester **23** (Figure 6). Thus, the spontaneous, second step is identical in the bis(SDTE) and the bis(SATE) concepts. Nevertheless, again two independent activation steps are required to yield the free nucleotide. With these two approaches, two equivalents of episulfide, which has been shown to be chronically and acutely toxic in mice and rats, are released besides the nucleotide⁽³¹⁾. The toxicity and moreover also mutagenicity has also been observed *in vitro*⁽³²⁾. J.-L. Imbach and J.-P. Sommadoussi have shown that the introduction of the SDTE and the SATE group do not induce additional cytotoxicity of the phosphotriesters as compared to the parent nucleosides (e.g. AZT) in human myeloid colony-forming cells (human bone marrow cells)⁽³³⁾. It should be mentioned that the

stability of the bis(SATE) phosphotriesters could be adjusted by varying the thio ester residue: the lowest stability was obtained for thio esters of acetic acid whereas with increasing alkyl residue of the carboxylic acid the stability was increased. The optimized compounds are bis(SATE) compounds bearing two *S*-pivaloyl-2-thioethyl side chains as shown in Figure 6 because the highly lipophilic and sterically demanding *t*-butyl residue protects the masking group from too rapid cleavage as compared to the *S*-acetyl or *S*-*i*-propyl counterparts^(28d,30a,31). Again, J.-L. Imbach et al. demonstrated an increased stability of the bis(SATE) derivatives of AZTMP against degradation in culture medium as compared to cellular extracts⁽³⁴⁾. The bis(SATE) approach has been successfully applied to the thymidine kinase-bypass (TK-bypass) of d4T **1**⁽³⁵⁾ (shown in Figure 6) and AZT **2**⁽³⁴⁾, the adenosine deaminase-bypass (ADA-bypass) of the anti-HIV active nucleoside ddA **6**⁽³⁶⁾ as well as for an improvement of the bioavailability of the anti-herpes and anti-HIV drug PMEAs. Furthermore, the corresponding bis(SATE)-ddAMP triesters exhibited a higher stability than the dideoxynucleoside ddA **6** against acid-catalyzed depurination^(36a). It should be mentioned that the bis(SATE)-AZTMP derivatives demonstrated a 10-fold decrease in activity from the wild-type CEM/O cell line to the mutant TK⁻ CEM cell line⁽³⁴⁾. This result is puzzling but has also been observed by others. On the other hand, J. Cinatl, Jr. et al.^(11a) have shown that the bis(SATE)-AZTMP derivatives retained the biological activity of AZT **2** in AZT resistant Molt4/8 cells. Especially bis(*S*-pivaloyl-2-thioethyl)-AZTMP showed comparable cytotoxic and antiviral activity in sensitive and resistant cells. With these experiments, J. Cinatl et al. demonstrated that Molt4/8^rAZT cells exert resistance to the anti-HIV activity of the drug mainly owing to the lack of AZT phosphorylation and that resistance may be bypassed by using AZT monophosphate prodrugs. However, as for the bis(POM) phosphotriesters **10** reported by D. Farquhar, the intermediate mono(SATE) phosphonate esters **21** and **23**, respectively, are significantly lower substrates for the activating carboxyesterases due to the negative charge at the phosphonate residue in the vicinity of the enzyme cleavage site. Before hydrolysis takes place, the bis(SDTE) as well as the bis(SATE) phosphotriesters may serve as lipophilic precursors for a passive membrane permeation.

J.-L. Imbach et al. presented the comparison of the bis(SATE)-approach to the isomeric *S,S'*-bis(*O*-acyl-2-oxoethyl) phosphorodithiolates⁽³⁷⁾. Surprisingly, these derivatives proved as active as the original bis(SATE) triesters with the nucleosides d4T **1** and ddA **6**, although the delivery mechanism could not be the same. Further work is in progress to study the cleavage mechanism. Recently, the same group extended their prodrug design to bis(*S*-glycopyranosyl-2-*thioethyl*) [bis(SGTE)] nucleoside phosphotriesters⁽³⁸⁾. These compounds were designed to deliver the nucleotide by activation of glycosidases. However, no antiviral data have yet been published.

Bis(AB)-Nucleotides

One possibility to avoid the close vicinity of the negative charge of the intermediate phosphodiester and the enzyme cleavage site has been developed independently by S. Freeman et al.⁽³⁹⁾ and A. Glazier⁽⁴⁰⁾. The common motive of their approaches is to separate the phosphate group and the carboxyesterase cleaving site by a rigid spacer. Consequently, they are using bis(4'-acyloxybenzyl)- [bis(AB)] nucleotides **24** and **25**, respectively (shown as their AZTMP derivatives) instead of the *O*-acyloxymethyl or the *S*-acyl-2-thioethyl residue, respectively. As Freeman calculated a separation of about 4 Å, which she states is enough to avoid electrostatic disturbance of the anionic phosphate and the carboxyester residue⁽⁴¹⁾. The degradation mechanism of the bis(AB) nucleotides of type **24** is the following: the enzyme cleaves the ester moiety in the 4'-position of the aromatic ring to give

the 4'-hydroxybenzyl phosphotriester **26**. Again, a subsequent spontaneous fragmentation of **26** is induced by this enzymatic reaction resulting in the formation of phosphodiester **27** and the 4'-hydroxybenzyl cation **28** which is oxidized to yield the 4'-quinonemethide **29** or is quenched by water to yield 4'-hydroxybenzylalcohol **30**. Diester **27** could then be degraded after a second esterase activation or a phosphodiester cleavage to yield the nucleotide AZTMP **9** (Figure 7). The mechanism of delivery has been studied using methylphosphonate as model compound⁽⁴²⁾. Obviously, these compounds are also tripartate prodrugs.

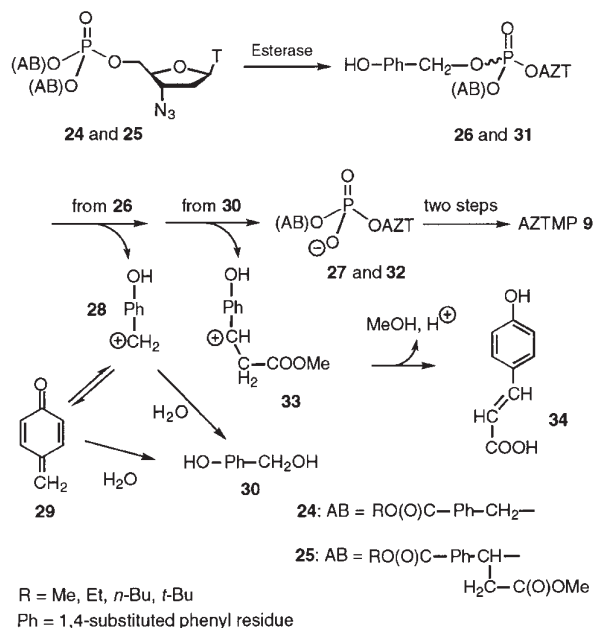


Figure 7. Delivery of 3'-azido-2',3'-dideoxythymidine monophosphate (AZTMP **9**) from bis(AB)-nucleotides **24** and **25**

S. Freeman et al. applied their approach to the delivery of AZTMP^(39,41) and obtained *in vitro* antiviral activity against HIV-1 and SIV that was comparable to AZT **2**. Unfortunately, she has not tested the compounds in thymidine kinase-deficient cells which could prove the direct intracellular delivery of AZTMP **9**. Recently, S. Freeman applied their bis(AB) approach to the intracellular delivery of foscarnet (PFA) from various precursors⁽⁴³⁾. A possible limitation of the approach is the generation of the reactive benzylation **28** in close vicinity of the active site of the enzyme because such cations are known to be potential alkylating agents that could react with side chains of amino acids⁽⁴⁴⁾.

This possible problem was faced by the approach reported by A. Glazier using the bis(AB) derivatives **25** (Figure 7)⁽⁴⁵⁾. Although the basic principle of the nucleotide delivery from **25** is essentially the same (carboxyesterase-mediated activation to give **32** via **31**) as from triesters **24**, the difference to the Freeman concept is the introduction of a methylmethoxycarbonyl group in the benzylic position of the AB residue. The rationale for this substitution at the benzylic position is the possibility of a fast elimination reaction via the fleeting 4'-quinonemethide **29** due to the electron-withdrawing methoxycarbonyl group that facilitates proton abstraction by decreasing the intrinsic barrier to proton removal. The transition state for proton removal is further stabilized by resonance delocalisation into the aromatic ring. The adjacent carbonyl group may also facilitate proton tautomerisation by acting by intramolecular general base catalysis. Firstly, this concept has been proven with bis(4'-acetoxy- α -methylmethoxycarbonyl-

benzyl)methylphosphonate. As expected, the final products after enzymatic deesterification were methylphosphonic acid, acetic acid and trans-4'-hydroxycinnamic acid **34**. Without addition of esterase no detectable decomposition was noted. Secondly, Glazier demonstrated convincingly, that after two hours of incubation with porcine liver esterase a nucleoside phosphotriester bearing the anti-herpes drug acyclovir monophosphate (ACVMP) has nearly completely been converted to methanol, acetic acid, trans-4'-hydroxycinnamic acid **34** as well as ACVMP as sole reaction products⁽⁴⁵⁾. This concept has been applied to the delivery of AZTMP⁽⁴⁰⁾ and ACVMP⁽⁴⁵⁾. However, a limitation of this approach may be the very short degradation half-lives of the compounds ($t_{1/2} = 16$ min). Furthermore, the bis(AB) derivatives reported by A. Glazier et al. were highly lipophilic (log Pa value = 1.07 to 4.18; depending on the carboxylic acid residue) making systemic applications problematic⁽⁴⁵⁾. Nevertheless, the bis(AB)-ACVMP derivatives showed promising *in vivo* activity against Herpes virus type-1 infections without toxic side effects using concentrations up to 100 mg/kg body weight⁽⁴⁵⁾. Furthermore, no mutagenicity was found for the prodrugs in the Ames test with or without microsomal activation⁽⁴⁵⁾. However, the compounds showed only a limited improvement in antiviral activity *in vitro*⁽⁴⁵⁾. As acyclovir, the prodrugs showed no toxicity *in vitro* ($\text{CD}_{50} > 100 \mu\text{M}$). Moreover, bis(AB) esters of PMEA were prepared and were found to be more potent *in vitro* than PMEA itself⁽⁴⁰⁾.

Nucleoside Arylphosphoramidates

Another novel class of a membrane-soluble enzyme triggered nucleotide delivery concepts are the phosphoramidate derivatives of type **35** recently reported by C. McGuigan et al.⁽⁴⁶⁾. The basic structure of these compounds (shown with the anti-HIV drug d4T **1**) is depicted in Figure 8. In contrast to the previously mentioned phosphotriester approaches, in this case the structural motive involves a phosphate moiety that is linked to the nucleoside, a phenyl group and, through a phosphoramidate linkage, also to the methyl ester of α -amino acid ester. This concept has been applied to the wide variety of nucleotides active against several viruses: d4TMP⁽⁴⁷⁾, AZTMP⁽⁴⁸⁾, 3'-thiacytidine monophosphate (3TCMP)⁽⁴⁹⁾, ddAMP⁽⁵⁰⁾ and 2',3'-dideoxy-2',3'-dideoxyadenosine monophosphate (d4AMP)^(50,51).

In *in vitro* antiviral tests, the d4T containing phosphoramidate was found to be slightly active as the parent nucleoside in wild-type CEM/O and MT-4 cells. More importantly and in contrast to d4T **1**, the d4TMP phosphoramidate completely retained the biological activity in mutant thymidine kinase-deficient CEM/TK⁻ cells⁽⁴⁷⁾ and suppresses HIV-1 infection in natural peripheral blood lymphocytes and freshly isolated monocyte/macrophages. This result proves that d4TMP **4** was delivered intracellularly making the observed activity of the compound almost entirely independent of thymidine kinase and consequently these compounds serve as efficient tools for the TK-bypass. The d4T-phosphoramidate was found to be equally active against HIV-1 and HIV-2 replication, and has also proved inhibitory to other retroviruses including simian immunodeficiency virus (SIV) and feline immunodeficiency virus (FIV), Visna virus and Moloney murine sarcoma virus in cell culture⁽⁵²⁾. An interesting structure-activity relationship (SAR) has been found with these compounds: first, the presence of an α -amino acid ester is essentially important for the biological activity. In contrast to α -amino acid bearing compounds, simple alkyl amine derivatives exhibited a complete loss of antiviral activity⁽⁵³⁾. The same was observed for derivatives bearing β -amino acid side groups. Among the natural α -amino acids L-alanine was found to be the most effective⁽⁵⁴⁾ while the enantiomeric D-alanine showed a 30-fold decrease in bioactivity⁽⁵⁵⁾. This interesting SAR points to an

Explore Litigation Insights

Docket Alarm provides insights to develop a more informed litigation strategy and the peace of mind of knowing you're on top of things.

Real-Time Litigation Alerts



Keep your litigation team up-to-date with **real-time alerts** and advanced team management tools built for the enterprise, all while greatly reducing PACER spend.

Our comprehensive service means we can handle Federal, State, and Administrative courts across the country.

Advanced Docket Research



With over 230 million records, Docket Alarm's cloud-native docket research platform finds what other services can't. Coverage includes Federal, State, plus PTAB, TTAB, ITC and NLRB decisions, all in one place.

Identify arguments that have been successful in the past with full text, pinpoint searching. Link to case law cited within any court document via Fastcase.

Analytics At Your Fingertips



Learn what happened the last time a particular judge, opposing counsel or company faced cases similar to yours.

Advanced out-of-the-box PTAB and TTAB analytics are always at your fingertips.

API

Docket Alarm offers a powerful API (application programming interface) to developers that want to integrate case filings into their apps.

LAW FIRMS

Build custom dashboards for your attorneys and clients with live data direct from the court.

Automate many repetitive legal tasks like conflict checks, document management, and marketing.

FINANCIAL INSTITUTIONS

Litigation and bankruptcy checks for companies and debtors.

E-DISCOVERY AND LEGAL VENDORS

Sync your system to PACER to automate legal marketing.