

United States Patent [19]

Oueen et al.

[54] HUMANIZED IMMUNOGLOBULINS

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- [21] Appl. No.: 634,278
- [22] Filed: Dec. 19, 1990

Related U.S. Application Data

- [63] Continuation-in-part of Ser. No. 590,274, Sep. 28, 1990, abandoned, and a continuation-in-part of Ser. No. 310,252, Feb. 13, 1989, abandoned, which is a continuation-in-part of Ser. No. 290,975, Dec. 28, 1988, abandoned.
- Int. Cl.6 A61K 39/395; C07K 16/28 [51]
- U.S. CI. 530/387.3; 530/387.1; [52]
 - 530/388.22; 424/133.1; 424/143.1
- [58] 424/143.1; 530/387, 388.22, 387.1, 387.3

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[57] ABSTRACT

Novel methods for producing, and compositions of, humanized immunoglobulins having one or more complementarity determining regions (CDR's) and possible additional amino acids from a donor immunoglobulin and a framework region from an accepting human immunoglobulin are provided. Each humanized immunoglobulin chain will usually comprise, in addition to the CDR's, amino acids from the donor immunoglobulin framework that arc, c.g., capable of interacting with the CDR's to effect binding affinity, such as one or more amino acids which are immediately adjacent to a CDR in the donor immunoglobulin or those within about 3 Å as predicted by molecular modeling. The heavy and light chains may cach be designed by using any one or all of various position criteria. When combined into an intact antibody, the humanized immunoglobulins of the present invention will be substantially non-immunogenic in humans and retain substantially the same affinity as the donor immunoglobulin to the antigen, such as a protein or other compound containing an epitope.

13 Claims, 55 Drawing Sheets

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1	Q D	I I	V Q	L M	T T	QQ	S S	P P	A S	I T	M L	S S	A A	S S	P V	G G	E D	K R	vv	T T
21 21	I I	Т Т	C C	S R	A A	S S	S Q	S S	I I	N	S T	Y W	M L	H A	W W	F Y	Q Q	QQ	K K	P P
40 41	G G	T K	S A	P P	K K	L L	W L	I M	Y Y	T K	T A	S S	N S	L L	A E	S S	G G	v v	P P	A S
60 61	R R	F F	S I	GG	S S	G	S S	G G	T T	SE	Y F	S T	L L	Т	I	S S	R S	M L	Ê Q	A P
80 81	E D	DD	A F	A A	T T	Y Y	Y Y	C C	H Q	QQ	R Y	S N	T S	Y D	P S	L K	T M	F	GG	SQ
100 101	G G	Т	ĸ	L V	E	L V	K													

FIGURE 1A

1 1	Q Q	V V	Q Q	L L	Q V	Q Q	S S	G G	A A	E	L V	A K	K K	P P	G	AS	S S	v v	K K	M V
21 21	s S	C C	K K	A A	S S	G G	Y G	T T	F F	TS	S R	Y S	R A	M I	H I	W W	v v		Q Q	R A
41 41	P P	G G	Q Q	G G	L L	E E	W W	M	G G	Y G	I I	N V	P P	S M	T F	G G	Y P	T P	E N	Y Y
61 61	N A	Q Q	K K	F	K Q	D G	K R	A V	T T	L	T T	A A	D D	K E	S S	S T	S N	T T	A A	Y Y
81 81	M M	Q E	L L	S S	S S	L L	T R	F	E E	D	S T	A	V	Y	Y F	C C	A A	R G	G G	Y
100 101	G	G I	V Y	F	D P	Y E	W E	G Y	QN	G G	T G	T L	L V	T T	v v	S S	S S			

FIGURE 1B

1 1	D E	I	v v	L M	T T	Q Q	S S	P	A A	S T	L L	AS	v v	S	El Di	G G	QE	R R	A A	T T
21 21	I L	S S	СС	RR	A A	S S	Q Q	S S	v v	S S	T T	S S	T T	Y Y	N N	Y	M M	H H	W W	Y
41 41	Q Q	Q Q	ĸ	D: D:	G G	00	P S	P P	K R	L L	L L	I I	K	Ŷ	A A	S S	N N	L		S S
61 61	G G	V I	Dr Di	AA		In ter			S S	G G	FS	G G	T T	D E	Fr Fr	T T	L L	N T	I I	H S
81 81	PR	V L	E	FI S	[1] [1]	D D	T F	V A	T V	Y Y	Y Y	СС	Q Q	H H	S S	W W	E E	I I	P Di	Y
101 101		fig fai		GQ	G G	FIFI	K R	L V	EE	I I	K K									

FIGURE 2A

1 1	E	M V	I Q	1	V L	E	S S	G G	G G	G G	L L	V V	K		G G	<u>A</u> G	S S	Ľ L	к R	L L
21 21	S S	CC	A A	A	SS	GG	Щų	T	Ε	S	N N	Y	G G	L L	S S	W	v v	R R	QQ	T A
41 41	S P	D G	RX	RG	L L	6161	W W	V V	A A	S S	1	S S	RR	G G	G G	G	R. 7.	 T	>1 >1	S S
61 61	PL P	D D	N N	L L	K K	G G	R R	E E	T T	I	S S	R R	E N	D D	A S	K K	N N	T T	L L	Y Y
81 81	L L :	00	M M	S N	S S	L S	K Q	S A	E	D D	T T	A A	L L	Y Y	>1 >1	СС	्र <u>।</u>	R	61 64	G G
101 101	1	21.2	vi vi	A A	D 0	Y Y	G G	Li fi	F F	D D	v v	W W	GG	T Q	G G	T T	₽ t r]	v v	-	v v
121	S S	S S																		

FIGURE 2B

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1 1	D D	I	V Q	M M	T T	Q Q	S	H P	K S	F T	M L	S	T A	S S	V V	G G	D D	RR	V V	S
21 21	I I	T T	C C	K K	A A	S S	Q Q	D D	v v	G G	S S	A A	v v	v v	W	HE H	Q Q	QQ	K	S p,
41 41	G G	Q	S A	P P	K K	L L	L L	I	Y Y	W	A A	S S	T T	R R	H: HI	T T	G G	v v	D1 D1	D S
61 61	R R	F F	T T	G G	S S	G G	SS	G G	T T	DE	ΕF	T T	L L	T T	I	T S	NS	V L	Q Q	S Di
81 81	E D	D D	L F	A A	D T	Y Y	E E	C C	Q Q Q	Q Q	Y Y	S S	I I	E	D1 D1	L L	T T	In In	G G	Ą
101 101	G G	T T	R K	L V	E	L V	K K													

FIGURE 3A

1 1	Q Q	V V	Q Q	L L	Q V	Q Q	S S	D G	A A	E E	L V	V K	K K	P P	G	s.	S S	V V	K K	I V	
21 21	SS	C C	K K	V A	S S	G G	Y Y	T T	F F	T T	D D	H H	T T	I	H	W W	M 	K R	QQ	R A	
41 41	n, n	E G	Q Q	G	1-1-1	[1] [J]	W W	fei fui	G G	Y	I -	Y Y Y	n, D,	R R	D D	G G	H	T	R R	vi v	
61 61	S A	E	K K	F	K K	G G	K	A A	T T	L I	T T	A A	D D	K E	S S	A T	S N	T T	A A	21 21	
81 81	M M	H E	L L	N S	S S	L L	T R	S S	E	D D	S T	A A	V V	Y Y	En La	C	A A	R. R.	G	RR	
101 101	D	S S	R R	[F] [1]	R. P.	N	G G	F	A A	Y Y	W W	GG	QQ	G	T Ţ	1-1 1-1	V V	T C	V V	S	
121 121	AS						3														

FIGURE 3B

1

1	D D		V Q	L M	T T	QQ	S S	P	A S	S	L L	A S	V A	S S	L V	G G	Q D	R R	A V	T T
21 21	Ī	S T	СС	R R	A A	S S	E E	S S	v v	D D	N N	2 2	G G	I I	S S	F F	M M	N N	W W	년 년
41 41	Q Q	Q Q	K K	Pt D1	GG	QK	PA	PP	K K	L L	L L	I I	Y Y	A A	A A	S S	N N	Q Q	G G	S S
61 61	G G	v v	PP	AS	R R	[ո իս	s s	GG	S S	G	S S	GG	T T	DD	E E	S T	L L	N N	I I	H S
81 81	P S	M L	E Q	EP	D D	D D	T F	A A	M T	Y Y	F Y	CC	QQ	Q Q	S S	K K	E	v v	P P	W W
101 101	Т 	F F	GG	GQ	G G	Ē	K K	L V		I I	K K									

FIGURE 4A

1 1	EQ	v v	QQ	L	Q V	QQ	S	GG	PA	E	L V	V K	K K	D1 D1	G G	AS	S	v v	K K	I V
21 21		C C	K K	A A	S S	G G	Y Y	T T	F F	T T	D D	Y	N N	M M	H H	W W	v v	K R	Q Q	S A
41 41	1: P	GG	K Q	S G	<u>.</u> 1	EI EI	W W =	I I	GG	>: >	I I	~ ~	P	21 21	N N	G G		T T	G G	>1 >1
61 61	N N	QQ	K K	E E	K K	S S	K K	A A	T T	L I	T T	V A	D D	NE	SS	S T	S N	T T		Y Y
81 81	M M	D E	V	R S	S S	L Ľ	T R	S S	E	D D	S T	A A	v v	Y Y	Y Y	C C	A A	R R	G	R R
101 101	n, D.	AA	M	D D	Y Y	W W	00	Q Q	GG	T T	S L	v v	T T	v v	S S	S S				

FIGURE 4B

1	Q D	Ĩ	V Q	L M	T T	Q Q	S	P P	A S	I S	M L	S S	A A	S S	P V	GG	ED	K R	v v	T T
21 21	M I	T T	СС	S S	G G	S S	S S	S S	v v	S S	F	M M	Y Y	W W	¥1 ¥1	Q Q	Q Q	R K	P P	U U
41 41	S K	S A	Pi Pi	R K	Ŀ	L	I I	Y Y	D D	T T	S S	N N	L L	A A	S S	GG	v	P	v s	R
61 61	F	S S	G G	S S	GG	SS	G G	T T	S D	Y Y	S T	L F	T T	I	S S	R. S	M	ΞQ	A D	Ē
81 81	D D	A I	A	T T	>1 >1	¥ Y	C Ç	QQ	00	W W	S S	T T	¥ Y	Pp	L L	T	tel (a)	G G	A Q	G G
101 101	T T	K K	: V	[1] [1]	L V	X														

FIGURE 5A

1 1	QE	v v	QQ	L L	ĸ	QIE	S S	G G	n, G	G G	L L	v v	QQ	P	S G	QQ	S S	ī. L	SR	I L
21 21	T S	C C	T A	VA	S S	G G	E Li	S T	v v	T T	S S	Y Y	G G	v v	HE	W W	ī V	R R	QQ	S A
41 41		G G	K	GG	1-11	[1] [1]	W		G		1111	W W	S S	G G	6 6	S (7)	F+ F +	ם ס	>1 >1	N
61 61		A A	Ei fu	I	S S	R	L F	T T		S S	K R		N	S S	K K	S N	Q	V	1.1 >1	(rr)
81 81		V M	N N	S S	L	QQ	PA	A E	D	T Ţ	A A	I	21 21	Y Y	CC	A A	n. n.	A A	GG	D D
101 101	Y	N N	>1 >1	DD	U G	fer fer	A A	Y Y	W	G	Q O	G	T.	L	V V	T T	v	S	<u>a</u> S	

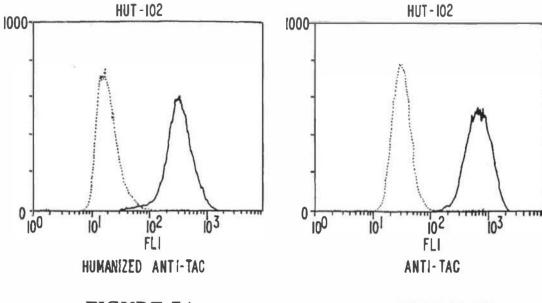
FIGURE 5B

D I V L T Q S P A T L S V T P G D S V S E I V L T Q S P G T L S L S P G E R A T 1 L S C R A S Q S I S N N L H W Y Q Q K S L S C <u>R A S Q S I S N N L H</u> W Y Q Q K P 21 21 41 H E S P R L L I K Y A S Q S I S G G Q A P R L L I <u>K Y A S Q S I S</u> G P S I IPD 41 R F S G S G S G T D F T L R F S G S G S G T D F T L 51 S T V N G V E T I S R L E P 61 E D F G M Y F C Q Q S N S W P H T F G G E D F A V Y Y C <u>Q Q S N S W P H T</u> F G Q 81 81 G T K L E I K G T K V E I K 101 101

FIGURE 6A

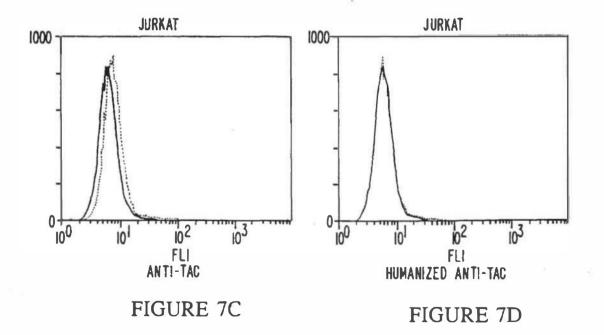
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21 21	SS	C C	K K	A A	S S	V G	1 >1	S S	F F	T T	GG	>1 >1	T T	M M	N N	W	v v	K R	QQ	S A
41 41	11: U1	G G	Q X	N G	-1 1-2	61 [1]	W W	1 V	G	L -	I	N N	D1 D1	21.21	N :J	G G	G	T Ţ	SS	>- >-
61 61	N N	00	K	Irs is	K K	G G	K R	A V	T T	L V	T S	V L	D K	Kp	S S	S [11	N N	T Q	A A	>1 >1
81 81	M	[6] [6]	L L	L S	S S	L	T F	S S	A E	D	S T	A A	v v	YY	Y Y	C C	<u>يت</u> 1.	R R	R R	G G
101 101	In fr	N. N.	0 0	24 24	S S	M	D D	>1 >1	W	GG	00	GG	5-1 E-1	S.J	v v	T	V V	S S	C C	

FIGURE 6B









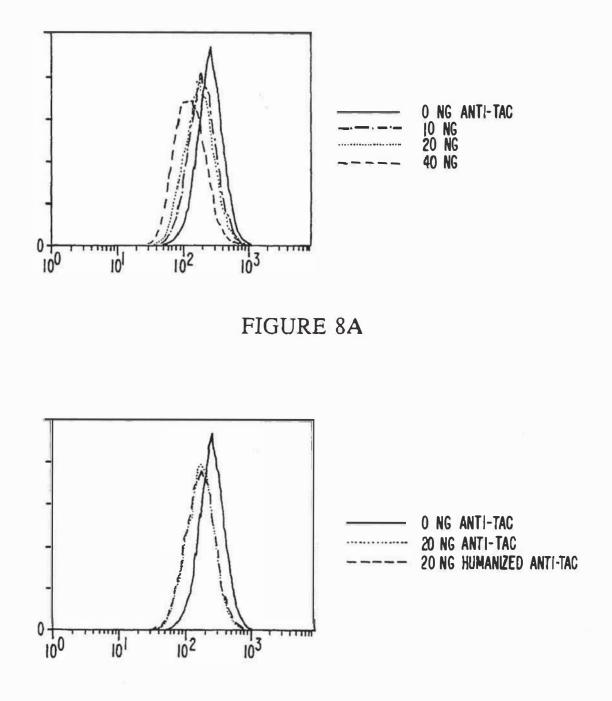


FIGURE 8B

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Amp

Pvull

Amp Amp PVull Hyg Kbal CHI Hyg Kbal CHI H

EcoRI

FIGURE 9A

FIGURE 9B

Gpt

pVk

BamHI

Ck

,Xbal



EcoRI

hCMV

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Q' Q'	V : V	Q Q	L L	v 1 v	Q Q	S S	G G	A : A	E - E	v I V	К К	K ¦ K	P P	0 C	S S	S S	V 	к - К	V ¦ V	
S S	C : C	К К	A ; A	S S	G G	Y G	T T	£ F	T : S	S ·· S	Y I V	R ¦ R	M	Н ; Н	3 3	V ¦ V	R R	Q' Q	A :- A	
P P	G : G	Q Q	G ¦ G	L ¦ L	E E	¥ ¥	I : M	G G	Y ¦ Y	I ¦ I	2 7.	Р Р	S S	т ; т	G ; G	Y † Y	Τ ; Τ	E E	Y	0
N N	Q Q	K 1 K	F F	K ¦ K	D I D	K : R	A V	т : т	I ¦ I	т : т	A ; A	D : D	E :-	S S	T ; T	и и	T : T	A A	Y ¦ Y	
M - M	E E	L ¦ L	S S	S S	ւ : Լ	R ¦ R	S S	E E	D D	т : т	A A	V F	Y ¦ Y	Y : F	с с	A A	R G	G G	G G	
с с	v v	F ¦ F	D D	Y 	W E	G Y	Q Z	G G	T G	L : L	v I V	Т : Т	V 	S S	S S					

FIGURE 10A

D : D	1 ; I	Q Q	M : M	T ¦ T	Q Q	S S	P : P	s s	T ¦ T	ե Լ	S S	А : А	s s	V ¦ V	G G	D : D	R ¦ R	v T V	T ¦ T
I ; I	T ; T	с с	s s	A A	s s	S S	s s	I ¦ I	s s	Y Y	M : M	н : н	W W	Y Y	Q Q	Q Q	К - К	P 0	G G
К ¦ К	A 	P :- P	к : К	L : L	L : L	I : M	Y ¦ Y	T 	`T : T	S S	и и	L : L	א א	S S	G I G	V F V	P P.	A : S	R ¦ R
F F	S I	G G	S S	G G	s s	G G	T ¦ T	E 14	F F	T T	և ։ Լ	T T	1	S S	S S	L : L	Q Q	P P	D D
D : D	년 년	A ¦ A	T ¦ T	Y Y	Y ¦ Y	с с	Н ¦ Н	Q Q	R ¦ R	S S	T T	Y ; Y	P P	L ; L	T ; T	ተ ተ	G G	Q Q	G G
T ; T	К К	v : v	E E	v t v	К ; К														

FIGURE 10B

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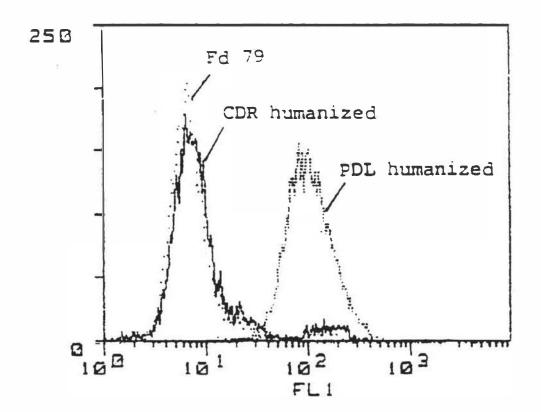
10	20	30	40	50	60	70
AGCTTCTAGA	TGGGATGGAG	CTGGATCTTT	CTCTTCCTCC	TGTCAGGTAC	CCCCCCCCTC	CACTCTCAGG
TCGAAGATCT	ACCCTACCTC	GACCTAGAAA	GAGAAGGAGG	ACAGTCCATG	GCGCCCGCAC	GTGAGAGTCC
80	90		110		130	140
TCCACCTTGT			AGAAACCTGG			
AGGTCGAACA	GGTCAGACCC	CGACTTCAGT	TCTTTGGACC	GAGCTCGCAC	TTCCAGAGGA	CGTTCCGAAG
150	160	170	180	190	200	210
TGGCGGGGACC	TTTTCTAGCT	ACAGGATGCA	CTGGGTAAGG	CAGGCCCCTG	GACAGGGTCT	GGAATGGATG
ACCGCCCTGG	AAAAGATCGA	TGTCCTACGT	GACCCATTCC	GTCCGGGGGAC	CTGTCCCAGA	CCTTACCTAC
220	230	240		260	270	280
GGATATATTA	ATCCGTCGAC	TGGGTATACT	GAATACAATC	AGAAOTTCAA	GGACAGGGTC	ACAATTACTG
CCTATATAAT	TAGGCAGCTG	ACCCATATGA	CTTATGTTAG	TCTTCAAGTT	CCTGTCCCAG	TGTTAATGAC
290	300			330	340	350
CAGACGAATC	CACCAATACA	GCCTACATGG	AACTGAGCAG	CCTGAGATCT	GAGGACACCG	CATTCTATTT
GTCTGCTTAG	GTGGTTATGT	CGGATGTACC	TTGACTCGTC	GGACTCTAGA	CTCCTGTGGC	GTAAGATAAA
360	370	380	390	400	410	420
CTGTGCAGGG			CGAATACAAT			
GACACGTCCC	CCACCCCCTC	AGAAACTGAT	GCTTATGTTA	CCTCCCGACC	AGTGTCAGAG	GAGTCCACTC
430	440					
TCCTTAAAAC	CTCTAGACGA	TAT				
	GAGATCTGCT					

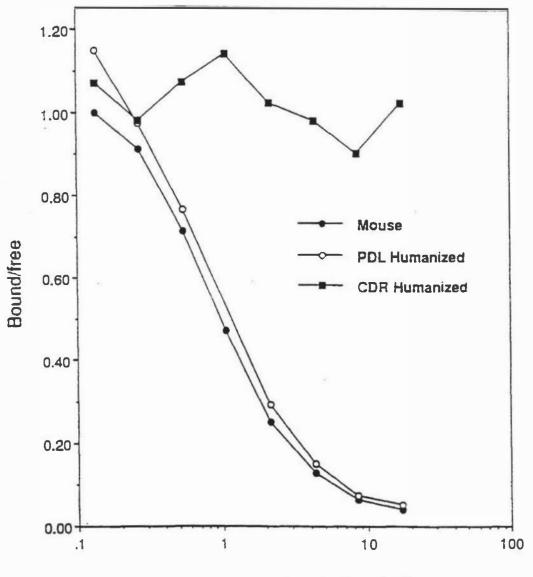
FIGURE 11A

BI Exhibit 1136

10	20	30	40	50	60	70
CAAATCTAGA	TGGAGACCGA	TACCCTCCTG	CTATGGGTCC	TCCTGCTATG	GGTCCCAGGA	TCAACCGGAG
GTTTAGATCT	ACCTCTGGCT	ATGGGAGGAC	GATACCCAGG	AGGACGATAC	CCAGGGTCCT	AGTTGGCCTC
80	90	100	110	120	130	140
ATATTCAGAT	GACCCAGTCT	CCATCTACCC	TCTCTGCTAG	CGTCGGGGAT	AGGGTCACCA	TAACCTGCTC
TATAAGTCTA	CTGGGTCAGA	GGTAGATCCG	AGAGACGATC	GCAGCCCCTA	TCCCAGTGGT	ATTGGACGAG
150	160				200	210
TGCCAGCTCA	AGTATAAGTT			AAGCCAGGCA		
ACGGTCGACT	TCATATTCAA	TGTACGTGAC	CATGGTCGTC	TTCGGTCCGT	TTCGAGGGTT	CGAAGATTAC
220	230	240	250	260	270	280
TATACCACAT	CCAACCTGGC	TTCTGGAGTC	CCTTCTCGCT	TCATTGGCAG	TCGATCTCCC	ACCGAGTTCA
ATATGGTGTA	GGTTGGACCG	AAGACCTCAG	GGAAGAGCGA	AGTAACCGTC	ACCTAGACCC	TGGCTCAAGT
	•					
290	300	310	320	330	340	350
CCCTCACAAT	CAGCTCTCTG	CAGCCAGATG	ATTTCGCCAC	TTATTACTGC	CATCAAAGGA	GTACTTACCC
GGGAGTGTTA	GTCGAGAGAC	GTCGGTCTAC	TAAAGCGGTG	AATAATGACG	GTAGTTTCCT	CATGAATGGG
360	370	380	390	400		
ACTCACGTTC	GGTCAGGGGA	CCAAGGTGGA	GGTCAAACGT	AAGTACACTT	TTCTAGATAT	Α
TGAGTGCAAG	CCAGTCCCCT	GGTTCCACCT	CCAGTTTGCA	TTCATGTGAA	AAGATCTATA	T

FIGURE 11B





Competitor Concentration (nM)

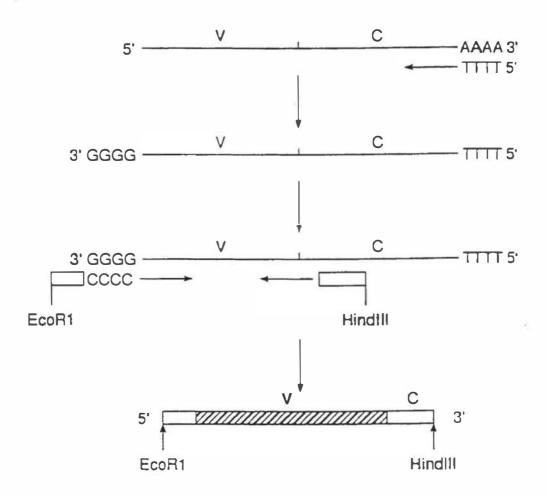


FIGURE 14

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1	0	V I V	9	L L	Q V	0	S S S	G	A I A	E I E	L V	A K	K I K	P I P	G	A S	S-S	V I V	K E K	M V
21 21	S S	C-C	K	A I A	S	G-G	Y G	T T	F	T S ≠	S R	Y S	R A	M I	H	W	V	K R	0	R A
41 41	₽ Î ₽	G G	0	G G	L L	E I E	W 1 W	 M *	GLG	Y G		N V	P P	S M	T F	G G	Y P	T P	E N	Y I Y
61 61	N A	0	K I K	F F	K Q	D G	K R *	A V *	T I T	L	T I T	A I A	D J D	K E	SIS	S T	S N	T I T	A I A	Y I Y
81 81	M I M	0 E	L L	S S	S S S	L L	T R	F S	E E	D D	S T	A I A	V F *	Y I Y	Y F *	C C	A I A	R G *	G G	Y
100 101	G								Q N *											

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1	Q D		V Q	L M	T T	0 0	S I S	P I P	A S	I T	M	S I S	A 1 A	S I S	P V	GIG	E D	K R	V I V	Ţ
21 21		T T	C I C	S R	A	S S	S Q	S S		N	S T	Y W	M	H	W I W	F Y	0 1 0	0	K I K	P I P
40 41	6 - 6	T K	S A	P I P	K K	L I L	W L	 	Y I Y	T K	T A	S I S	N S	L I L	A E	S S	G I G	V I V	P I P	A S
60 6 I					S - S												R		ε	A P
80 81	E D	D	A F	A I A	T I T	Y I Y	Y I Y	C C	H Q	0	R Y	S N	T S	Y D	P S	L K	T M	F I F	G	S O
100 101	G G G		K K	L V	E I E	Ł	K I K													

20 30 40 50 10 60 TCTAGATGGGATGGAGCTGGATCTTTCTCTTCTCCTCCTGTCAGGTACCGCGGGCGTGCACT M G W S W I F L F L L S G T A G V H 70 80 90 100 110 120 CTCAGGTCCĂGCTTGTCCĂĞTCTGGGGCŤĞAAGTCAAGĂĂACCTGGCŤCĞAGCGTGAAĞĞ S Q V Q L V Q S G A E V K K P G S S V K I 30 I 40 I 50 I 60 I 70 I 80 TCTCCTGCAAGGCTTCTGGCTACACCTTTACTAGCTACAGGATGCACTGGGTAAGGCAGG V S C K A S G Y T F T S Y R M H W V R O 200 220 210 240 190 230 CCCCTGG ACAGGGTCTGGAATGGATTGGATATATTAATCCGTCGACTGGGTATACTGAAT A P G O G L E W I G Y I N P S T G Y T E 260 270 280 250 290 300 A CAATCAGAAGTT CAAGGACAAGGCAACAATTACTG CAGACGAATCCACCAA TACAGCCT YN QKFKDKATITADESTNTA 310 320 330 340 350 360 ACATGGAACTGAGCAGCCTGAGATCTGAGGACACCGCAGTCTATTACTGTGCAAGAGGGG YMELSSLRSEDTAVYYCARG 370 380 390 400 410 420 GGGGGGTCTTTGACTACTGGGGCCAAGGAACCCTGGTCACAGTCTCCTCAGGTGAGTCČT G G V F D Y W G Q G T L V T V S S

430 TAAAACCTCTAGA

10 20 30 TCTAGATGGAGACCGATACCCTCCTGCTATGGGTCCTCCTGCTATGGGTCCCAGGATCAA METDTLLEWVLLLWVPGS CCGGAGATATTCAGATGACCCAGTCTCCATCTACCCTCTCTGCTAGCGTCGGGGATAGGG T G D I Q M T Q S P S T L S A S V G D R TCACCATAACCTGCTCTGCCAGCTCAAGTATAAGTTACATGCACTGGTACCAGCAGAAGC V T I T C S A S S S I S Y M H W Y Q Q K CAGGCAAAGCTCCCAAGCTTCTAATTTATACCACATCCAACCTGGCTTCTGGAGTCCCTG PGKAPKLLIYTTSNLASGVP CTCGCTTCÃGTGGCAGTGGATCTGGGACCGAGTTCACCCTCACAATCÃGCTCTCTGCÃĞČ A R F S G S G S G T E F T L T I S S L Q CAGATGATTTCGCCACTTATTACTGCCATCAAAGGAGTACTTACCCACTCACGTTCGGTC P D D F A T Y Y C H Q R S T Y P L T F G AGGGGACCAAGGTGGAGGTCAAACGTAAGTACACTITTCTAGA Q G T K V E V K

HES 12	AGCTTCTAGATGGGATG Cactctcaggtccagct AAGGTC	G A G C T G G A T C T T T C T C T T G T C C A G T C T G G G G C T G	T C C T C T G T C A G G T A C C G C G G G C G T G A A G T C A A G A A A C C T G G C T C G A G C G T G
HES13	CCCAGTCGACGGATTAA CCAGTGCATCCTGTAGC CGAGCCAGG	T A T A T C CA A T C C A T T C C T A G T A A A G G T G T A G C C A	AGACCCTGTCCAGGGG CCTGCCTTAC GAAGCCTTGCAGGAGACCTTCACGCT
HES 14			A T C A G A A G T T C A A G G A C A A G G C A A C A T G G A A C T G A G C A G C C T G A G A T C T G A G
HES15			AGACTGTGACCAGGGTTCCTTGGCCC AATAGACTGCGGTGTCCTCAGATCTC
		FIGURE 19A	
	HESI2		HES 14

HESI2 HESI3 HESI5 FIGURE 19B **U.S.** Patent

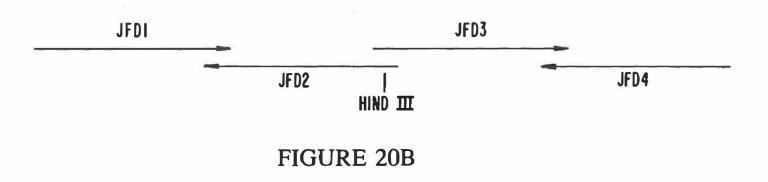
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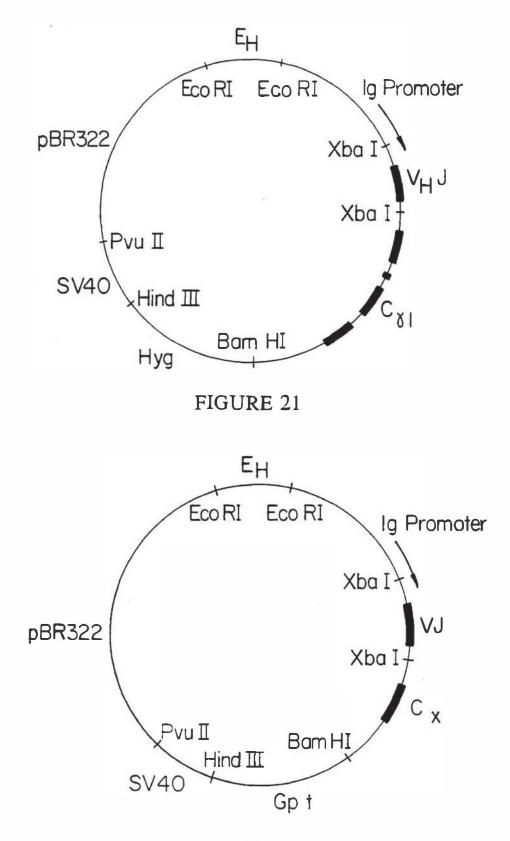
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JFDI	CAAATCTAGATGGAGACCGATACCCTCCTGCTATGGGTCCTCCTGCTATGGGTCCCAGGA TCAACCGGAGATATTCAGATGACCCAGTCTCCATCTACCCTCTCTGCTAGCGTCGGGGAT
JFD2	ATAAATTAGAAGCTT GGGAGCTTTGCCTGGCTTCTGCTGGTACCAGTGCATGTAACTTAT ACTTGAGCTGGCAGAGCAGGTTATGGTGACCCTATCCCCGACGCTAGCAGAGAG
JFD3	G C T C C C A A G C T T C T A A T T T A T A C C A C A T C C A A C C T G G C T C T G G A G T C C C T G C T C G C A G T G G C A G T G G A T C T G G G A C C G A G T T C A C C C T C A C A A T C A G C T C T C T G C A G C C A G A T G A T T T C

JFD4 TA TA TCTAGAAAAGTGTACTTACGTTTGACCTCCACCTTGGTCCCCTGACCGAACGTGAG TGGGTAAGTACTCCTTTGATGGCAGTAATAAGTGGCGAAATCATCTGGCTGCAGAGAGCT GA









50

120

180

300

360

. 30 . ATGGATTTTCAAGTGCAGATTTTCAGCTTCCTGCTAATCAGTGCCTCAGTCATACTGTCC M D F Q V Q I F S F L L I S A S V I L S . . 90 AGAGGACAAATTGTTCTCACCCAGTCTCCAGCAATCATGTCTGCGTCTCCAGGGGGGGAAG R G Q I V L T Q S P A I M S A S P G E K 150 . GTCACCATGACCTGCAGTGGCAGCTCAAGTGTAAGTTTCATGTACTGGTACCAGCAGAGG V T M T C <u>S G S S S V S F M Y</u> W Y Q Q R • 210 • 240 . . CCAGGATCCTCCCCAGACTCCTGATTTATGACACATCCAACCTGGCTTCTGGAGTCCCT PGSSPRLLIY<u>DTSNLAS</u>GVP • 270 • • GTTCGCTTCAGTGGCAGTGGGTCTGGGACCTCTTACTCTCACAATCAGCCGAATGGAG V R F S G S G S G T S Y S L T I S R M Ê • 330 •

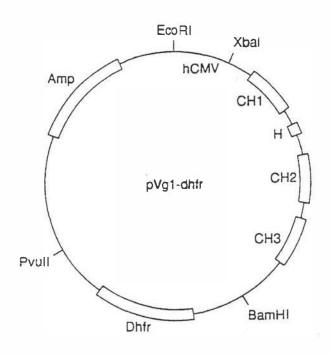
GCTGAAGATGCTGCCACTTATTACTGCCAGCAGTGGAGTACTTACCCGCTCACGTTCGGT A E D A A T Y Y C <u>Q Q W S T Y P L T</u> F G

GCTGGGACCAAGCTGGAGCTGAAA AGTKLELK

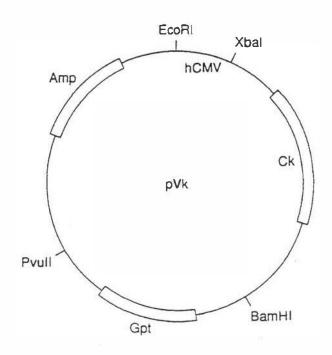
FIGURE 23A

30 60 ATGGCTGTCTTGGGGGCTGCTCTTCTGCCTGGTGACATTCCCAAGCTGTGTCCTATCCCAG MAVLGLLFCLVTFPSCVLSQ . 90 . . 120 GTGCAGCTGAAGCAGTCAGGACCTGGCCTAGTGCAGCCCTCACAGAGCCTGTCCATCACC V Q L K Q S G P G L V Q P S Q S L S I T . . 180 150 TGCACAGTCTCTGGTTTCTCAGTAACAAGTTATGGTGTACACTGGATTCGCCAGTCTCCA CTVSGFSVT<u>SYGVH</u>WIRQSP 210 . 240 . . GGAAAGGGTCTGGAGTGGCTGGGAGTGATATGGAGTGGTGGAAGCACAGACTATAATGCA G K G L E W L G V I W S G G S T D Y N A . • 270 300 . . GCTTTCATATCCAGACTGACCATCAGCAAGGACAACTCCAAGAGCCAAGTTTTCTTTAAA A F I S R L T I S K D N S K S Q V F F K • 360 . 330 . **GTGAACAGTCTGCAACCTGCTGACACAGCCATATACTATTGTGCCAGAGCTGGGGACTAT** VNSLQPADFAIYYCAR<u>AGDY</u> . . 390 . AATTACGACGGTTTTGCTTACTGGGGCCAAGGGACTCTGGTCACTGTCTCTGCG NYDGFAYWGQGTLVTVSA

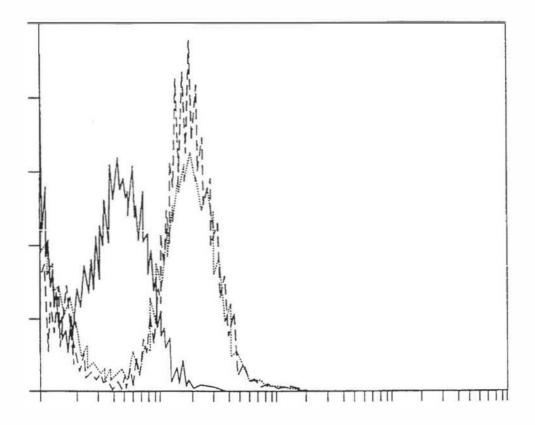
FIGURE 23B











1 1	D I D I	QQ	M	T T	20	S	Di Di	S S	S S	L L	S S	V A	S S	v v	GG	D D	R R	v v	t-i * i
21 21	I T I T	C C	QS	A G	S S	Q S	N S	v v	N	A S	Y F	L M	N Y	W W	Y	QQ	QQ	K K	0, 0,
4 <u>1</u> 40	G L G <u>K</u>	A A	P P	K K	í L	L L	I I	>1 >1	G D	A T	S S	T N	R L	E A	A S	GG	v v	P P	S
61 60	R F R F	S S	G G	S S	G G	S S	G G	T	D D	[u >1	T T	티티	ŢŢ	HI	S S	SS	L	QQ	Di Di
81 80	E D E D	I I	A A	T T	>1 >1	¥	C C	00	QQ	Y W	N S	N T	WY	ni Di	11 10	6+ 6 -	111 Ins	G G	Q Q
101 100	G T G T	K	v v	E	v v	K													

FIGURE 26A

1	A E	v v	QQ	L L	L L	E	S S	G G	G G	G G	L L	V V	QQ	P P	G G	G G	S S	L L	R R	111
21 21	S S	C C	A A	A A	S S	G G	Έ	T T	F	ST	A S	S Y	A G	M V	SH	W W	V V	R R	QQ	A A
41	Di Di	G	ĸ	GG	L L	[1] [1]	1 1 1 1 1	V V	A G	W	X	Y	E	N S	G	NG	0 5	X F	HD	¥ - × -
61 60	AN	D A	S A	V F	N <u>T</u>	G S	R R	E E	T T	I I	S S	R R	N D	D N	S S	K K	N	- - -	E E	>1 >1
81 80	L L	QQ	M M	N N	G S	L L	QQ	A A	ZE	V D	S T	A A	I	Y Y	>1 >1	C C	A A	R	D	A A
101 99	G G	0, D	¥ V	V	S N	D1 >1	T D	Fr G	[14] [14]	A A	H Y	W	G G	QQ	GG	E E	1.7 1-3	V V	E4 F4	V V
121	S S	SS																		

FIGURE 26B

vc13

10 20 30 40 50 60 TTCTGCTGGT ACCAGTACAT GAAACTTACA CTTGAGCTGC CACTGCAGGT GATGGTGACG 70 80 90 100 CGGTCACCCA CTGAGGCACT GAGGCTAGAT GGAGACTGGG TCATTTG

vcl4

10 20 30 40 50 60 CATGTACTGG TACCAGCAGA AGCCCAGGAAA AGCTCCGAAA CTTCTGATTT ATGACACATC 70 80 90 100 110 120 CAACCTGGCT TCTGGAGTCC CTTCCCSCTT CAGTGGCAGT GGGTCTGGGA CCGATTACAC 130 CTTTACAATC TCTTCA

vc15

10 20 30 40 50 60 TGTGTCTAGA AAAGTGTACT TACGTTTTAC CTCGACCTTG GTCCCTTGAC CGAACGTGAG 70 80 90 100 110 120 CCGGTAAGTA CTCCACTGCT GGCAGTAATA AGTGGCTATA TCTTCCGGCT GAAGTGAAGA 130 GATTGTAAAG GTGTAAT

vc16

10 20 30 40 50 60 CACATCTAGA CCACCATGGA TTTTCAAGTG CAGATCTTCA GCTTCCTGCT AATCAGTGCC 70 80 90 100 TCAGTCATAC TGTCCAGAGG AGATATTCAA ATGACCCAGT CTCCATCT

FIGURE 27A

vc11

10 20 30 40 50 60 TAGTCTGTCG ACCCACCACT CCATATCACT CCCACCCACT CGAGTCCCTT TCCAGGAGCC 70 80 90 100 110 120 TGGCGGACCC AGTGTACACC ATAACTTGTT ACGGTGAAAC CACTGGCGGC ACAAGACAGT 130 CTCAGAGATC CTCCTGGC

vc12

10 20 30 40 50 60 TGGTGGGTCG ACAGACTATA ATGCAGCTTT CATATCCAGA TTTACCATCA GCAGAGACAA 70 80 90 100 110 120 CAGCAAGAAC ACACTGTATC TCCAAATGAA TAGCCTGCAA GCCGAGGACA CAGCCATATA

TTATTG

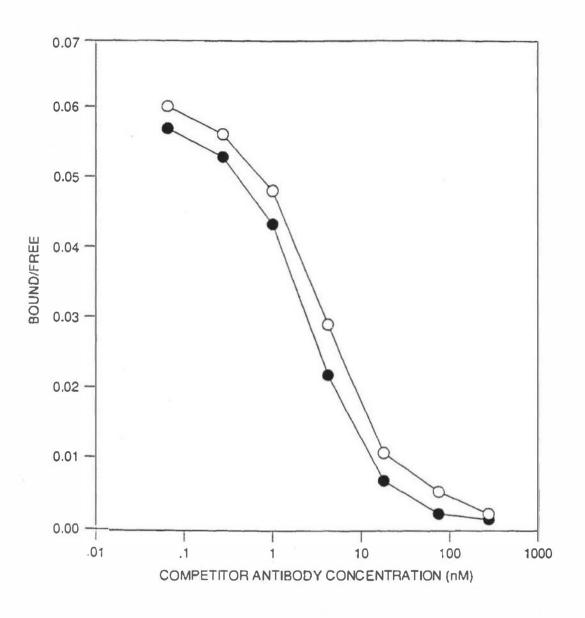
wps54

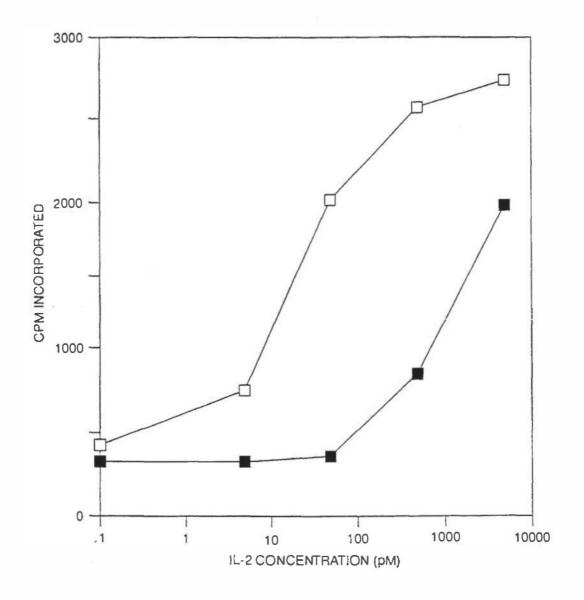
10 20 30 40 50 60 ACACTCTAGA CCACCATGGC TGTCTTGGGG CTGCTCTTCT GCCTGGTGAC ATTCCCAAGC 80 90 100 110 120 70 TGTGTCCTAT CCGCTGTCCA GCTGCTAGAG AGTGGTGGCG GTCTGGTGCA GCCAGGAGGA 130 TCTCTGAGAC

WOSS7

20 30 40 50 10 60 ACACTCTAGA AGTTAGGACT CACCTGAAGA GACAGTGACC AGAGTCCCTT GGCCCCAGTA 90 70 80 100 110 AGCAAAACCG TCGTAATTAT AGTCCCCAGC TCTGGCACAA TAATATATGG CTGTGTCC

FIGURE 27B





57 57

77 77

97 97 80

100

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	1	5	10	15	20
1 1	E M I E V Q	LVES			S L K L S L R L
2 <u>1</u> 21	S C A S C A		30 TFSN TFSN		40 V R Q T V R Q A
41 41	S D R P G K	45 RLEW	VASI	<u>52 a 5</u> S R G G G I	S R I Y S R I Y S
60 60	60 Р D N Р D N		70 RFTIS RFTIS	REDAKI	95 NTLY NTLY
80 80	80 82 LQM LQM	SSLX		90 A L Y Y C I A L Y Y C I	95 LREG LREG
97 97	ĬŸŸ IŸŸ		GFFDV	105 WGTGT WGQGT	110 T V I V L V T V
112 112	113 S S S S		FIGURE 30	A	
		-	1.0	15	20
1	l D I V	5 / L T Q S	10 SPASL SP <u>A</u> TL	AVSLG	QRAT
1	ΕΙV	25		SVSPG 30	e r a t 35
21 21	ISC LSC	RAS	TabcdQSVSTQSVST		M H W Y M H W Y
		40	45	50	55
37 37	Q Q K Q Q K	P G Q E P <u>G Q</u> S	P P K L L S P R L L	I K Y A S I K Y A S	N L E S N L E S
		60	65	70	75

FIGURE 30B

85

P V E E E D T V T Y Y C Q H S W E R L E S E D F A V Y Y C Q H S W E

105 107

G V P A R F S G S G F G T D F T L N I H G I P A R F S G S G S G T E F T L T I S

90

95

P P 21 2

5,530,101

1 1	1 Q Q	v v	QQ	⊢,1	5 Q V	QQ	SS	DG	A A	10 E E	1. V	V X	X	Di Di	15 13 0 0	<u>A</u> S	S S	V V	ĸ	20 I V
21 21	S S	СС	ĸ	V A	25 S S	GG	Y ¥	T	hi tri	30 T T	D D	H H	T	1-1-1	35 H H	W W	M	KR	QQ	40 R A
41 41	P. P.	ЕG	QQ	G G	45 1 L	E E	W W	fer far	G G	50 Y Y	I I	52 Y Y		R R	0 0	55 G G	H H	T T	RR	Y Y
60 60	60 S A	ΕE	K	In fu	ĸ	65 G G	K K	A A	T T	L I	70 T T	A A	D D	K E	SS	75 A T	SN	T	A A	Y
80 80	80 M M	H E	82 1 1	a N S	20 00 00		T R	S	85 E E	D D	S T	A A	v v	90 Y Y	Ing for	СС	A A	R R	95 G G	R R
97 9 7	D D	S S	R	100 E E	id R. R.	N N	0000	ы ту р.	A A	Y Y	W W	9 9	105 Q Q	GG	f · F	L L	V V	110 T T	v v	S
113 113																				
1	l D D	I.	V Q	M M	5 61 8-1	Q Q	SS	HP	ĸs	10 F T	M L	SS	T A	S S	15 V V	G G	ם ם	R	V V	20 S T
21 21	I.	T T	c	K K	25 A A	S S	Q Q	D D	v v	30 G G	S S	A A	V V	V V	35 W W	11: 11	QQ	QQ	X	40 S P
41 41	GG	QK	SA	n, n,	45 K X	L L	L	I	¥ ¥	50 W W	y Y	S S	T T	N. K.	55 :: ::	T T	G	V	D, D,	60 D S
61 61	P. R.	[11] [11]	E. F.	G G	65 S S	69	S	GG	T T	70 D E	նս նս	Ŧ	L L	E E	75 I I	F+ S	N 5	V I.	QQ	80 S Pi
		-	Ŧ	2	85 0 -	v	E	С	0	90 0	Y	S	I	[1]	95 Di Di	-1-1	Ŧ.	F	()	100 A Ç
81 51	5	Ð	} [iii	A		24.24	In In	CC	QQ	Q Q	5- 5-	S Ø	11	to to	01	1	E • F •	E. F.	0.0	Ç

FIGURE 30D

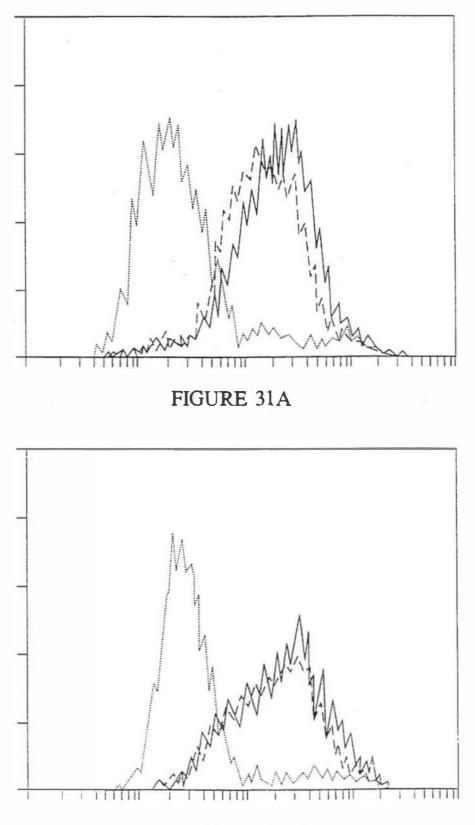


FIGURE 31B

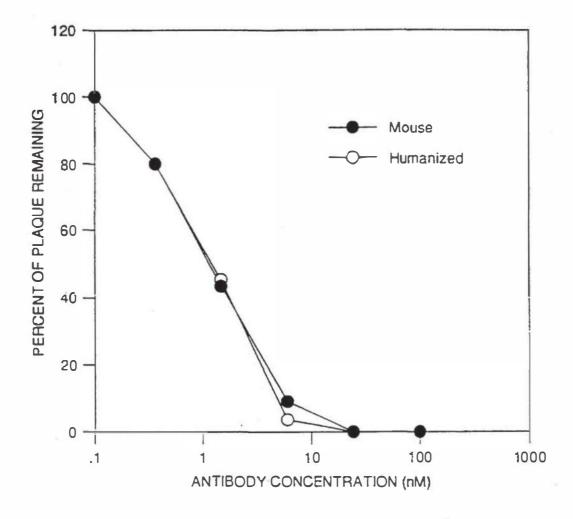


FIGURE 32A

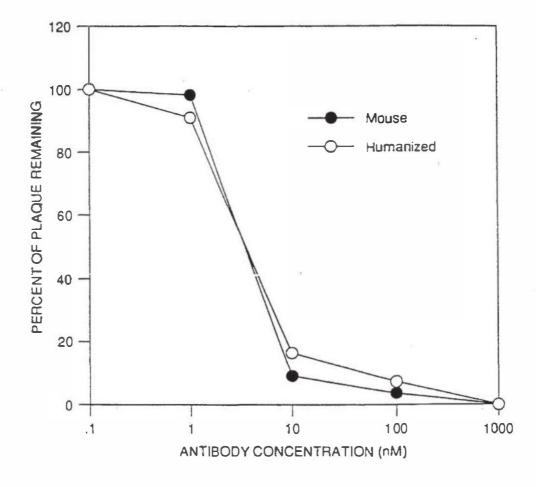
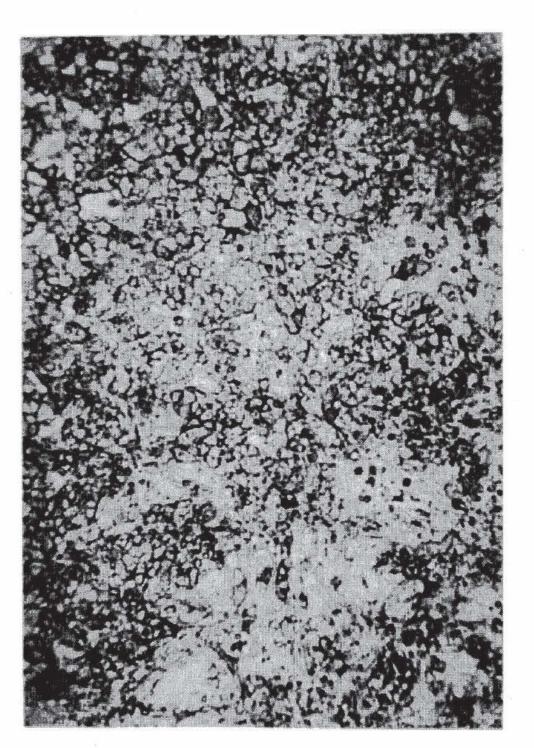


FIGURE 32B



FIGURE 33A



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1 8

FIGURE 34B

390

.

GCTATGGACTACTGGGGTCA.AGG.AACCTCAGTCACCGTCTCCTCA <u>A M D Y</u> W G Q G T S V T V S S

120 . 90 . GTCCAGCTTCAGCAGTCAGGACCTGAGCTGGTGAAACCTGGGGCCTCAGTGAAGATATCC V Q L Q Q S G P E L V K P G A S V K I S • 150 . . 180 TGCAAGGCTTCTGGATACACATTCACTGACTACAACATGCACTGGGTGAAGCAGAGCCAT CKASGYTFT<u>DYNMH</u>WVKQSH 210 • . . 240 **GGAAAGAGCCTTGAGTGGATTGGATATATTTATCCTTACAATGGTGGTACTGGCTACAAC** G K S L E W I G Y I Y P Y N G G T G Y N • 270 • • 300 CAGAAGTTCAAGAGCAAGGCCACATTGACTGTAGACAATTCCTCCAGCACAGCCTACATG <u>Q K F K S K A T L T V D N S S S T A Y M</u> . 330 . 360 GACGTCCGCAGCCTGACATCTGAGGACTCTGCAGTCTATTACTGTGCAAGAGGGCGCCCC DVRSLTSEDSAVYICAR<u>GR</u>P

CCTATGGAGGAGGATGATACTGCAATGTATTTCTGTCAGCAAAGTAAGGAGGTTCCGTGG PMEEDDTAMYFCQQSKEVPW 390 ACGTTCGGTGGAGGCACCAAGCTGGAAATCAAA

TFGGGTKLEIK

.

330

. 270 GGGGTCCCTGCCAGGTTTAGTGGCAGTGGGTCTGGGACAGACTTCAGCCTCAACATCCAT G V P A R F S G S G S G T D F S L N I H

150

D I V L T Q S P A S L A V S L G Q R A T

30

ATGGAGAAAGACACACTCCTGCTATGGGTCCTGCTTCTCTGGGTTCCAGGTTCCACAGGT MEKDTLLLWVLLLWVPGSTG

180 ATCTCCTGCAGAGCCAGCGAAAGTGTTGATAATTATGGCATTAGTTTATGAACTGGTTC I S C R A S E S V D N Y G I S F M N W F

.

210 240

Q Q K P G Q P P K L L I Y <u>A A S N Q G S</u> . .

FIGURE 34A

30

ATGGGATGGAGCTGGATCTTTCTCTTCTCCTGTCAGGAACTGCAGGCGTCCACTCTGAG M G W S W I F L F L L S G T A G V H S E

90 . GACATTGTGCTGACCCAATCTCCAGCTTCTTTGGCTGTGTCTCTAGGGCAGAGGGCCACC

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60

120

300

360

60

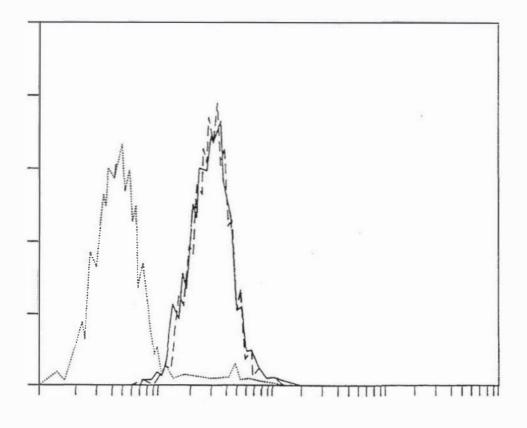


FIGURE 35

1	D I D I	Q Q	M M	T	QQ	S S	P P	S S	TS	L L	S S	A A	S S	v v	G G	D	RR	v v	T T
21 21	I T I T	C C	R R	A A	S S	QE	S S	v	I D	N N	Y	G	I	T S	WE	L M	A N	W W	Ξ Y
37 41		K K	P P	G	G	A A		K K	L L	L L	M	Y Y	K A	A A	S S	S N	L Q	E G	S S
57 61	G V G V	P P	S S	R	E fa	I S	G	S S	G	S	GG	T	E D	Ire Ire	T	L L	TT	I ī	S S
77 81	S I S I	QQ	Pi pi	D D	D D	ΕF	A A	T T	Y Y	Y Y	CC	QQ	QQ	¥ S	N K	S LI	D V	S P	K W
97 101	M E T E	G G	00	GG	T T	K K	v v		V I	K									

FIGURE 36A

1		v v		L	V	Q	S	G	A	E	V	K	K K	P	GG	S S	S S	v v	K K	v v
21			Q														v	R	Q	A
21	S		K	A	S	G	G	Ţ	H H	ST	D	S Y	N	M	I H	W	v	R	Q	A
41 41	D1 D1	GG	QQ	GG	t t	ि) वि	W W	M	G G	G		Š Š	0, 0,	M	E N	Q Q	A G	Di F.	N G	Y Y
61 61	AN	00	K K	F	QK	G S	R K	V A	T T	I I	T	A A	D D	EE	SS	T T	N N	T T	A A	Y Y
81 81	M M	64 (F)	L L	S S	S S	L L	R R	S S	EE	D	T T	A A	F	21 21	E ¥	C C	A A	G R	G	¥
101 100	GR	10	Y A	S M	ק ט	E Y	EW	¥G	N	G G	G	L	V V	E-I E-+	V V	S	S S			

FIGURE 36B

mal

10 20 30 40 50 60 TATATCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGAACTGCT 90 70 80 100 110 120 GECGTCCACT CTCAGGTTCA GCTGGTGCAG TCTGGAGCTG AGGTGAAGAA GCCTGGGAGC 130 TCAGTGAAGG TT

ma2

10 20 30 40 50 60 70 110 80 90 100 120 CCTGCCTCAC CCAGTGCATG TTGTAGTCAG TGAAGGTGTA GCCAGAAGCT TTGCAGGAAA 130 CCTTCACTGA GCT

ma3

10 20 30 40 50 60 TGGTGGTACC GGCTACAACC AGAAGTTCAA GAGCAAGGCC ACAATTACAG CAGACGAGAG 80 70 90 100 110 TACTAACACA GCCTACATGG AACTOTOCAG COTGAGGTOT GAGGACACTG CA

ma4

10 20 30 40 50 60 TATATCTAGA GGCCATTCTT ACCTGAAGAG ACAGTGACCA GAGTCCCTTG GCCCCAGTAG 70 80 90 100 110 TECATAGOGG GGOGCOCTOT TGOGCAGTAA TAGACTGOAG TGTCCTCAGA C

FIGURE 37A

10 20 30 40 50 60 TATATCTAGA CCACCATGGA GAAAGACACA CTCCTGCTAT GGGTCCTGCT TCTCTGGGTT 70 80 90 100 110 120 CCAGGTTCCA CAGGTGACAT TCAGATGACC CAGTCTCCGA GCTCTCTGTC CGCATCAGTA

GG

maб

10 20 30 40 50 60 TCAGAAGCTT AGGAGCCTTC CCGGGTTTCT GTTGGAACCA GTTCATAAAG CTAATGCCAT 70 30 90 100 110 120 AATTGTCGAC ACTTTCC-CTG GCTCTGCATG TGATGGTGAC CCTGTCTCCT ACTGATGCGG

AC

ma7

10 20 30 40 50 60 TCCTAAGCTT CTGATTTACG CTGCATCCAA CCAAGGCTCC GGGGTACCCT CTCGCTTCTC 70 80 90 100 110 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT

ma8

10 20 30 40 50 60 TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 80 90 100 110 70 TCCACC-GAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG

FIGURE 37B

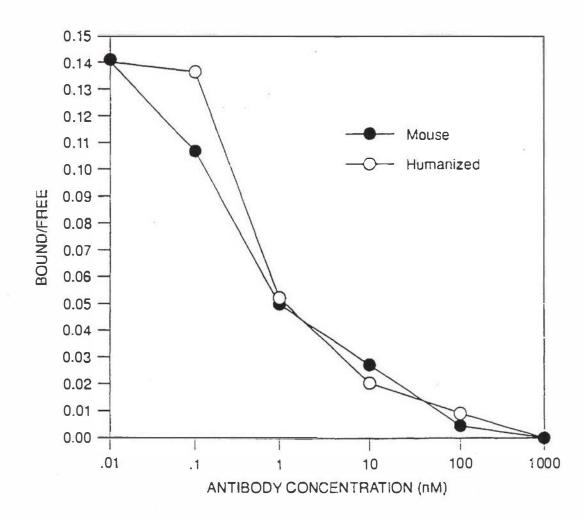


FIGURE 38

30 . 60 ATGGTTTTCACACCTCAGATACTTGGACTTATGCTTTTTTGGATTTCAGCCTCCAGAGGT M V F T P Q I L G L M L F W I S A S R G . . 90 . 120 GATATTGTGCTAACTCAGTCTCCAGCCACCCTGTCTGTGACTCCGGGAGATAGCGTCAGT DIVLTQSPATLSVTPGDSVS 180 . . . 150 CTTTCCTGCAGGGCCAGCCAAAGTATTAGCAACAACCTACACTGGTATCAACAAAAATCA L S C <u>R A S Q S I S N N L H</u> W Y Q Q K S • 210 • • 240 . CATGAGTCTCCAAGGCTTCTCATCAAGTATGCTTCCCAGTCCATCTCTGGGATCCCCTCC H E S P R L I K Y A S Q S I S G I P S 270 • 300 . . AGGTTCAGTGGCAGTGGATCAGGGACAGATTTCACTCTCAGTGTCAACGGTGTGGAGACT R F S G S G S G T D F T L S V N G V E T • 330 • • 360 . GAAGATTTTGGAATGTATTTCTGTCAACACACAACAGTTGGCCTCATACGTTCGGAGGG EDFGMYFC<u>QQSNSWPHT</u>FGG GGGACCAAGCTGGAAATAAAA GTKLEIK FIGURE 39A 60 . 30 . . ATGGGATGGAGCTGGATCTTTCTCTTCCTCCTGTCAGGAACTGCAGGTGTCCACTCTGAG M G W S W I F L F L L S G T A G V H S E 90 120 GTCCAGCTGCAACAGTCTGGACCTGAGCTGGTGAAGCCTGGAGCTTCAATGAAGATATCC V Q L Q Q S G P E L V K P G A S M K I S . 150 . 180 TGCAAGGCTTCTGTTTACTCATTCACTGGCTACACCATGAACTGGGTGAAGCAGAGCCAT CKASVYSFT<u>GYTMN</u>WVKQSH . 210 240 GGACAGAACCTTGAGTGGATTGGACTTATTAATCCTTACAATGGTGGTACTAGCTACAAC G Q N L E W I G L I N P Y N G G T S Y N . . 270 300 CAGAAGTTCAAGGGGAAGGCCACATTAACTGTAGACAAGTCATCCAACAGCCTACATG QKFKGKATLTVDKSSNTAYM . . 360 . 330 . GAGCTCCTCAGTCTGACATCTGCGGACTCTGCAGTCTATTACTGTACAAGACGGGGGTTT ELLSLTSADSAVYYCPR<u>RGF</u> 390 . .

CGAGACTATTCTATGGACTACTGGGGTCAAGGAACCTCAGTCACCGTCTCCTCA <u>R D Y S M D Y</u> W G Q G T S V T V S S

FIGURE 39B

11 2 1

47 of 139

1 1	E E	<u>T</u> I	v v	L L	T T							S S		S S	P P		E	R R		T T
21 21	L L		C C		A A		Q Q	S S	v	S I	S S	G N	Y N		G H	W W	>1 >4	QQ	Q Q	K K
41 40	P P	G G	QQ	A A	P P	R R	L L	L L	I I	Y K	G	A A	S S	S Q	RS	A T	E · S	G	I I	P P
61 60	D D	R	FF	S S	G G	SS	G G	S	G G	T T	D D	E F	T T	L L		I I	S S	R	L L	E
81 80	P P	E	D D	FF	A A	v v	Y Y	Y Y	C C	Q Q	00	¥ S	G N	S S	L W	Gp		T		G G
101 100	Q Q	G G	T T	K K	v v	ыы	I I	K K												

FIGURE 40A

1	Q Q	v v	QQ	L L	M V	Q Q	S S	G G	A A	E E	v v	K K	K K	P	G G	S S	S S	v v	R R	v v
21 21	S S	СС	K K	T A	S S	G G	G	T S	F F	V T	D G	Y Y	K T	G M	L N	W W	v v	R R	QQ	A A
41 41	Pi P	G G	K K	G G	L L	E	W	v v	G	Q L	I I	N N	L P	R	E N	NG	G	EI EI	V S	X
61 61	NN	P Q	G K	S It.	V K	V G	RR	v v	S T	v v	SS	L L	K K	₽ ₽	S S	în E	N	Q Q		H Y
81 81	M M	E	L L	S S	S S	L L	التا التا	S S	E	D D	T	A A	v v	Å	Y	C C	A Ţ	R. R.	ER	ž
101 100	G	[1. E	C	T	S R	D D	Y y	¥ S	Y M	⊻ D	Y Y	W W	G G	QQ	G	Ţ	1.1	V V	E-4 E-4	v V
121 118	S S	s s																		

FIGURE 40B

jb16

10 20 30 40 50 60 TAGATCTAGA CCACCATGGT TTTCACACCT CAGATACTAG GACTCATGCT CTTCTGGATT 70 80 90 100 110 120 TCAGCCTCCA GAGGTGAAAT TGTGCTAACT CAGTCTCCAG GCACCCTAAG CTTATCACCG

GGAGAAAGG

jb17

10 20 30 40 50 60 TAGACAGAAT TCACGCGTAC TTGATAAGTA GACGTGGAGC TTGTCCAGGT TTTTGTTGGT 70 80 90 100 110 120 ACCAGTGTAG GTTGTTGCTA ATACTTTGGC TGGCCCTGCA GGAAAGTGTA GCCCTTTCTC

CCGGTGAT

jb18

10 20 30 40 50 60 AAGAGAATTC ACGCGTCCCA GTCCATCTCT GGAATACCCG ATAGGTTCAG TGGCAGTGGA 70 80 90 100 110 TCAGGGACAG ATTTCACTCT CACAATAAGT AGGCTCGAGC CGGAAGATTT TGC

jb19

10 20 30 40 50 60 TAGATCTAGA GTTGAGAAGA CTACTTACGT TTTATTTCTA CCTTGGTCCC TTGTCCGAAC 70 80 90 100 110 GTATGAGGCC AACTGTTACT CTGTTGACAA TAATACACAG CAAAATCTTC CGGCTC

FIGURE 41A

jb20

30 40 20 50 10 60 TATATCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGAACTGCA 70 80 90 100 110 120 GGTGTCCACT CTCAAGTCCA ACTCGTACAG TCTCGAGCTG AGGTTAAAAA GCCTGGAAGT 130 TCAGTAAGAG TTTC

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jb21
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10 20 30 40 50 60 TATATAGGTA CCACCATTGT AAGGATTAAT AAGTCCAACC CACTCAAGTC CTTTTCCAGG 70 80 90 100 110 120 TGCCTGTCTC ACCCAGTTCA TGGTATACCC AGTGAATGAG TATCCGGAAG CTTTGCAGGA 130 AACTCTTACT GAAC

jb22

10 20 30 40 50 60 TATATAGGTA CCAGCTACAA CCAGAAGTTC AAGGGCACAG TTACAGTTC TTTGAAGCCT 70 80 90 100 110 TCATTTAACC AGGCCTACAT GGAGCTCAGT AGTCTGTTTT CTGAAGACAC TGCAGT

jb23

20 30 40 50 10 60 TATATCTAGA GGCCATTCTT ACCTGAGGAG ACGGTGACTA AGGTTCCTTG ACCCCAGTAG 70 80 90 100 110 TCCATAGAAT AGTCTCGAAA CCCCCGTCTT CTACAGTAAT AGACTGCAGT GTCTTC

FIGURE 41B

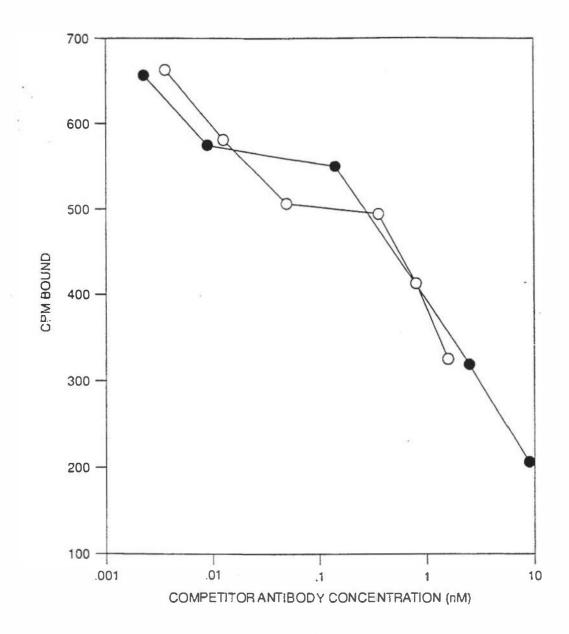


FIGURE 42

30 . . 60 . ATGCATCAGACCAGCATGGSCATCAAGATGGAATCACAGACTCTGGTCTTCATATCCATA M H Q T S M G I K M E S Q T L V F I S I 90 120 CTGCTCTGGTTATATGGTGCTCATGGGAACATTGTTATGACCCAATCTCCCAAATCCATG L L W L Y G A D G N I V M T Q S P K S M . 180 150 . . TACGTGTCAATAGGAGAGAGGGTCACCTTGAGCTGCAAGGCCAGTGAAAATGTGGATACT Y V S I G E R V T L S C <u>K A S E N V D T</u> 210 . 240 TATGTATCCTGGTATCAACAGAAACCAGAGCAGTCTCCTAAACTGCTGATATATGGGGGCA <u>YVS</u>WYQQKPEQSPKLLIY<u>GA</u> • • 300 • 270 TCCAACCGGTACACTGGGGTCCACGATCGCTTCACGGGCAGTGGATCTGCAACAGATTTC <u>SNRYT</u>GVHDRFTGSGSATDF • • • 360 . 330 ACTCTGACCATCAGCAGTGTGCAGGCTGAAGACCTTGCAGATTATCACTGTGGACAGAGT T L T I S S V Q A E D L A D Y H C <u>G Q S</u> . 390 TACAACTATCCATTCACGTTCGGCTCGGGGACAAAGTTGGAAATAAAG <u>YNYPFT</u> FGSGTKLEIK

FIGURE 43A

60 . 30 . ATGACATCACTGTTCTCTCTACAGTTACCGAGCACACAGGACCTCGCCATGGGATGGAGC M T S L F S L Q L P S T Q D L A M G W S . 90 . . . 120 TGTATCATCCTCTTCTTGGTAGCAACAGCTACAGGTGTCCTCTCCCAGGTCCAACTGCAG C I I L F L V A T A T G V L S Q V Q L Q • 150 • • 180 CAGCCTGGGGCTGACCTTGTGATGCCTGGGGCTCCAGTGAAGCTGTCCTGCTTGGCTTCT Q P G A D L V M P G A P V K L S C L A S • 210 • • . 240 GGCTACATCTTCACCAGCTCCTGGATAAACTGGGTGAAGCAGAGGCCTGGACGAGGCCTC GYIFT<u>SSWIN</u>WVKQRPGRGL . . 270 . 300 GAGTGGATTGGAAGGATTGATCCTTCCGATGGTGAAGTTCACTACAATCAAGATTTCAAG E W I G R I D P S D G E V H Y N Q D F K . 330 . 360 GACAAGGCCACACTGACTGTAGACAAATCCTCCAGCACAGCCTACATCCAACAGC D K A T L T V D K S S S T A Y I Q L N S 390 420 CTGACATCTGAGGACTCTGCGGTCTATTACTGTGCTAGAGGATTTCTGCCCTGGTTTGCT L T S E D S A V Y Y C A R <u>G F L P W F A</u> • 450 GACTGGGGCCAAGGGACTCTGGTCACTGTCTCTGCA

FIGURE 43B

D W G Q G T L V T V S A

1 1	D D	ī	QQ	M M	T T	QQ	S S	P P	S S	T T	L L	S S	A A	S S	v v	G G	D D	R R	v v	T
21 21	I	T T	C C	R K	A A	S S	Q E	S N	I V	N D	T T	W Y	L V	A S	W W	Y Y	QQ	QQ	K K	P P
41 41	GG	K K	A A	PP	K K	L L	L L	MI	Y Y	K G	A A	S S	S N	LR	E Y	S T	GG	v v	Di Di	S S
61 61	R R	F F	IS	G G	S	GG	S S	GG	T T	ED	F F	T T	L L	T T	I	S S	S S	L L	QQ	P P
81 81	D D	D D	Et iti	A	T T	Y Y	¥	C C	G	QQ	QS	Y Y	N N	S	D Y	S P.	K F	MT	FF	G G
100 100	QQ	G	T T	K	v v	E	v v	K												

FIGURE 44A

1 1		/ Q / Q	L L	v v	QQ	S S	G	A A	E	v v	K K	K K	P P	G G	S S	S S	v v	K	v v
21 21	S C S C	K K	A A	S S	G	Gv	T	F F	S T	RS	S S	A W	I T	I N	W	v v	R R	QQ	A A
41 41			G G	1,1	EI EI	W	M M	G G	GR	II TI	V D	D. D.	M S	F D		Di fil		N H	v v
61 61	A (N (2 K 2 D	F	Q K	G D	R R	v v	T T	ĩ I	T T	A A	D D	EI EI	S S	T T	NN	T T	A A	>1 >1
81 81	M E M E	E L	S S	S S	L L	R R	S S	E E	D D	T T	A A	E V	Y Y	(T > 1	C C	A A	GR	GG	>1 [11]
101 101	G I I. E	¥ W	S L.	D A	E D	EW	G	N O	G	G T	L L	v v	T T	v v	S	S S			

FIGURE 44B

rhlO

10 20 30 40 50 60 TTTTTTCTAG ACCACCATGG AGACCGATAC CCTCCTGCTA TGGGTCCTCC TGCTATGGGT 70 80 90 100 110 CCCAGGATCA ACCGGAGATA TTCAGATGAC CCAGTCTCCG TCGACCCTCT CTGCT

rhll

10 20 30 40 50 60 TTTTAAGCTT GGGAGCTTTG CCTGGCTTCT GCTGATACCA GGATACATAA GTATCCACAT 80 90 100 110 70 120 TTTCACTGGC CTTGCAGGTT ATGGTGACCC TATCCCCGAC GCTAGCAGAG AGGTTCCACG

rh12

10 20 30 40 50 60 TTTTAAGCTT CTAATTTATG GGGCATCCAA CCGGTACACT GGGGTACCTT CACGCTTCAG 100 80 90 70 110 TGGCAGTGGA TCTGGGACCG ATTTCACCCT CACAATCAGC TCTCTGCAGC CAGATGAT

rh13

10 20 30 40 50 60 TTTTTTCTAG AGCAAAAGTC TACTTACGTT TGACCTCCAC CTTGGTCCCC TGACCGAACG 70 80 90 100 110 120 TGAATGGATA GTTGTAACTC TGTCCGCAGT AATAAGTGGC GAAATCATCT GGCTCCAGAG

FIGURE 45A

rh20

10	20	30	40	50	60
TTTTTTCTAGA	CCACCATGGG	ATGGAGCTGG	ATCTTTCTCT	TCCTCCTGTC	AGGTACCGCG
70 GGCGTGCACT	80 CTCAGGTCCA	20	100 TCTGGGGCTG		ACCT

rh21

10 20 30 40 50 60 TTTTGAATTC TCGAGACCCT GTCCAGGGGC CTGCCTTACC CAGTTTATCC AGGAGCTAGT 80 90 70 100 110 120 AAAGATGTAG CCAGAAGCTT TGCAGGAGAC CTTCACGGAG CTCCCAGGTT TCTTGACTTC

Α

rh22

10 20 30 40 50 60 TTTTGAATTC TCGAGTGGAT GGGAAGGATT GATCCTTCCG ATGGTGAAGT TCACTACAAT 70 80 90 100 110 120 CAAGATTTCA AGGACCGTGT TACAATTACA GCAGACGAAT CCACCAATAC AGCCTACATG 130 GAACTGAGCA GCCTGAG

rh23

10 20 30 40 50 60 TTTTTCTAGA GGTTTTAAGG ACTCACCTGA GGAGACTGTG ACCAGGGTTC CTTGGCCCCA 70 80 90 100 110 120 GTCAGCAAAC CAGGGCAGAA ATCCTCTTGC ACAGTAATAG ACTGCAGTGT CCTCTGATCT 130 CAGGCTGCTC AGTT

FIGURE 45B

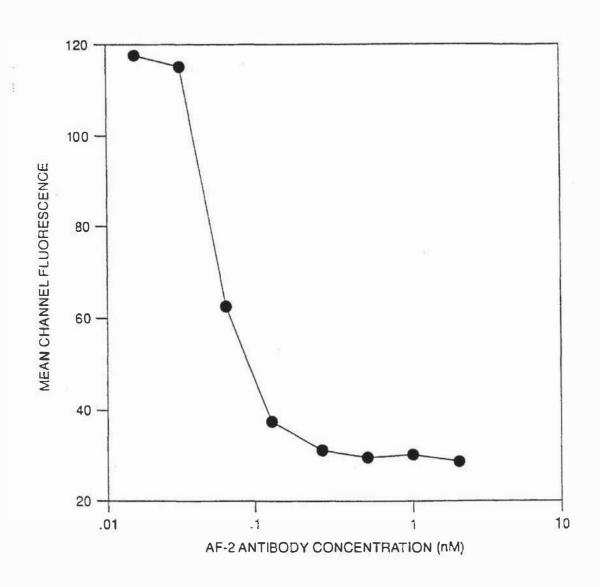


FIGURE 46

HUMANIZED IMMUNOGLOBULINS

CROSS-REFERENCE TO RELATED APPLICATIONS

This is a continuation-in-part application of commonly assigned patent application U.S. Ser. No. 07/590,274, filed Sep. 28, 1990 (now abandoned) and of U.S. Ser. No. 07/310,252, filed Feb. 13, 1989 (now abandoned), which is a continuation-in-part of U.S. Ser. No. 07/290,975, filed 10 Dec. 28, 1988 (now abandoned). All of these applications are specifically incorporated herein by reference.

FIELD OF THE INVENTION

The present invention relates generally to the combination of recombinant DNA and monoclonal antibody technologies for developing novel therapeutic agents and, more particularly, to the production of non-immunogenic antibodies having strong affinity for a predetermined antigen. 20

BACKGROUND OF THE INVENTION

The advent of monoclonal antibody technology in the mid 1970's heralded a new age of medicine. For the first time, 25 researchers and clinicians had access to essentially unlimited quantities of uniform antibodies capable of binding to a predetermined antigenic site and having various immunological effector functions. These proteins, known as "monoclonal antibodies" were thought to hold great promise in, 30 e.g., theremoval of harmful cells in vivo. Indeed, the clinical value of monoclonal antibodies seemed limitless for this use

Unfortunately, the development of appropriate therapeutic products based on these proteins has been severely 35 hampered by a number of drawbacks inherent in monoclonal antibody production. For example, most monoclonal antibodies are mouse derived, and thus do not fix human complement well. They also lack other important immunoglobulin functional characteristics when used in humans.

Perhaps most importantly, non-human monoclonal antibodies contain substantial stretches of amino acid sequences that will bc-immunogenic when injected into a human patient. Numerous studies have shown that after in jection of a foreign antibody, the immune response mounted by a 45 patient can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, as increasing numbers of different mouse or other antigenic (to humans) monoclonal antibodies can be expected to be developed to treat various diseases, after one 50 or several treatments with any non-human antibodies, subsequent treaunents, even for unrelated therapies, can be ineffective or even dangerous in themselves, because of cross-reactivity.

While the production of so called "chimeric antibodies" 55 (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. Moreover, efforts to immortalize human B-cells or generate human hybridomas capable of producing human immunoglobulins against a 60 desired antigen have been generally unsuccessful, particularly with many important human antigens: Most recently, recombinant DNA technology has been utilized to produce immunoglobulins which have human framework regions combined with complementarity determining regions 65 (CDR's) from a donor mouse or rat immunoglobulin (see, e.g., EPO Publication No. 0239400, which is incorporated

herein by reference). These new proteins arc called "reshaped" or "humanized" immunoglobulins and the process by which the donor immunoglobulin is converted into a human-like immunoglobulin by combining its CDR's with a human framework is called "humanization". Humanized antibodies are important because they bind to the same antigen as the original antibodies, but are less immunogenic when injected into humans.

However, a major problem with present humanization procedures has been a loss of affinity for the antigen (Jones ct al., Nature, 321, 522-525 (1986)), in some instances as much as 10-fold or more, especially when the antigen is a protein (Verhoyen et al., Science, 239, 1534-1536 (1988)). Loss of any affinity is, of course, highly undesirable. At the least, it means that more of the humanized antibody will have to be injected into the patient, at higher cost and greater risk of adverse effects. Even more critically, an antibody with reduced affinity may have poorer biological functions, such as complement lysis, anibody-dependent cellular cytotoxicity, or virus neutralization. For example, the loss of affinity in the partially humanized antibody HuVHCAMP may have caused it to lose all ability to mediate complement lysis (see, Riechmann et al., Nature, 332, 323-327 (1988); Table 1).

Thus, there is a need for improved means for producing humanized antibodies specifically reactive with strong affinity to a predetermined antigen. These humanized immunoglobulins should remain substantially non-immunogenic in humans, yet be easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

SUMMARY OF THE INVENTION

The present invention provides novel methods for prepating humanized immunoglobulin chains having generally one or more complementarity determining regions (CDR's) from a donor immunoglobulin and a framework region from a human immunoglobulin. The preferred methods comprise first comparing the framework or variable region amino acid sequence of the donor immunoglobulin to corresponding sequences in a collection of human immunoglobulin chains, and selecting as the human immunoglobulin one of the more homologous sequences from the collection. The human immunoglobulin, or acceptor immunoglobulin, sequence is typically selected from a collection of at least 10 to 20 immunoglobulin variable region sequences, and usually will have the highest homology to the donor immunoglobulin sequence of any sequence in the collection. The human immunoglobulin framework sequence will typically have about 65 to 70% homology or more to the donor immunoglobulin framework sequences. The donor immunoglobulin may be either a heavy chain or light chain, and the human collection will contain the same kind of chain. A humanized light and heavy chain can be used to form a complete humanized immunoglobulin or antibody, having two light/ heavy chain pairs, with or without partial or full-length human constant regions.

To form the humanized variable region, amino acids in the human acceptor sequence will be replaced by the corresponding amino acids from the donor sequence if they are in the category

(1) the amino acid is in a CDR.

In another embodiment of the present invention, either in conjunction with the above comparison step or separately, additional amino acids in the acceptor immunoglobulin

40

chain may be replaced with amino acids from the CDRdonor immunoglobulin chain. More specifically, further optional substitutions of a human framework amino acid of the acceptor immunoglobulin with the corresponding amino acid from a donor immunoglobulin will be made at positions 5 which fall in one or more of the following categories:

(2) the amino acid in the human framework region of the acceptor immunoglobulin is rare for that position and the corresponding amino acid in the donor immunoglobulin is common for that position in human immunoglobulin ¹⁰ sequences; or

(3) the amino acid is immediately adjacent to one of the CDR's; or

(4) the amino acid is predicted to be within about 3 Å of 15 the CDR's in a three-dimensional immunoglobulin model and capable of interacting with the antigen or with the CDR's of the donor or humanized immunoglobulin.

Moreover, an amino acid in the acceptor sequence may optionally be replaced with an amino acid typical for human $_{20}$ sequences at that position if

(5) the amino acid in the acceptor immunoglobulin is rare for that position and the corresponding amino acid in the donor immunoglobulin is also rare, relative to other human sequences. 25

The humanized immunoglobulin chain will typically comprise at least about 3 amino acids from the donor immunoglobulin in addition to the CDR's, usually at least one of which is immediately adjacent to a CDR in the donor immunoglobulin. The heavy and light chains may each be ³⁰ designed by using any one or all three of the position criteria.

When combined into an intact antibody, the humanized light and heavy chains of the present invention will be substantially non-immunogenic in humans and retain substantially the same affinity as the donor immunoglobulin to the antigen (such as a protein or other compound containing an epitope). These affinity levels can vary from about 10⁸ M^{-1} or higher, and may be within about 4 fold, preferably within about 2 fold of the donor immunoglobulin. Ideally, the humanized antibodies will exhibit affinity levels at least about 60 to 90% of the donor immunoglobulin's original affinity to the antigen.

Once designed, the immunoglobulins, including binding fragments and other immunoglobulin forms, of the present 45 invention may be produced readily by a variety of recombinant DNA or other techniques. Preferably, polynucleou des encoding the desired amino acid sequences are produced synthetically and by joining appropriate nucleic acid sequences, with ultimate expression in transfected cells. Notably, the methods of the present invention maximize the likelihood of producing humanized immunoglobulins with optimum binding characteristics without the need for producing intermediate forms that may display stepwise improvements in binding affinity. The humanized immuno-55 globulins will be particularly useful in treating human disorders susceptible to monoclonal antibody therapy, but find a variety of other uses as well.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1A and FIG. 1B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:1 and 2] and heavy chain (B) [SEQ ID NOS:3 and 4] variable regions of the mouse anti-Tac antibody (upper lines), compared with 65 the human Eu antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined.

Residues in the Eu antibody framework replaced with mouse amino acids in the humanized antibody are double underlined. The number of the first position on each line is given on the left.

FIG. 2A and FIG. 2B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:46 and 47] and heavy chain (B) [SEQ ID NOS:48 AND 49] variable regions of the mouse Fd79 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 3A and FIG. 3B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:50 and 51] and heavy chain (B) [SEQ ID NOS:52 and 53] variable regions of the mouse Fd138-80 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 4A and FIG. 4B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:54 and 55] and heavy chain (B) [SEQ ID NOS:56 and 57] variable regions of the mouse M195 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in cach chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 5A and FIG. 5B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:58 and 59] and heavy chain (B) [SEQ ID NOS:60 and 61] variable regions of the mouse mik- β 1 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 6A and FIG. 6B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:62 and 63] and heavy chain (B) [SEQ ID NOS:64 and 65] variable regions of the mouse CMV5 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 7A through FIG. 7D. Fluorocytometry of HUT-102 and Jurkat cells stained with anti-Tac antibody or humanized anti-Tac antibody followed respectively by fluorescein-conjugated goat anti-mouse Ig antibody or goat anti-human Ig antibody, as labeled. In each panel, the dotted curve shows the results when the first antibody was omitted, and the solid curve the results when the first and second (conjugated) antibodies were included as described.

FIG. 8A and FIG. 8B. (A) Fluorocytometry of HUT-102 cells stained with 0-40 ng of anti-Tac as indicated, then with biotinylated anti-Tac, and then with phycoerythrin-conjugated avidin. (B) Fluorocytometry of HUT-102 cells stained

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with the indicated antibody, then with biotinylated anti-Tac, and then with phycoerythin-conjugated avidin.

FIG. 9A and FIG. 9B. Schematic diagram of the plasmids pVgl (A) and pVk (B). The plasmid pVgl was constructed from the following fragments: an approximately 4850 base 5 pair BamHI-EcoRI fragment from the plasmid pSV2hph containing the amp and hyg genes; a 630-pb fragment containing the human cytomegalovirus IE1 gene promoter and enhancer flanked at the 5 and 3' by EcoRI and Xbal linkers respectively; and a 2800 bp Xbal-BamHI fragment 10 containing the human gamma-l constant region gene with 215 bp of the preceding intron and the poly(A) signal. The plasmid pVk was similarly constructed, with a 1530-bp human kappa constant region gene replacing the gamma-l gene and the gpt replacing the hyg gene.

FIG. 10A and FIG. 10B. Amino acid sequences of the heavy (A) [SEQ ID NOS:5 and 6] and light (B) [SEQ ID NOS:7 and 8] chain variable regions of the PDL and CDR-only humanized anti-Tac antibodies. The PDL sequence is shown on the upper line, the CDR-only sequence below. Amino acid differences are boxed. Complementarity Determining Regions (CDR's) are underlined.

FIG. 11A and FIG. 11B. Double-stranded DNA sequence of fragments encoding the heavy (A) [SEQ ID NO:9] and light (B) [SEQ ID NO:10] chain variable regions of the CDR-only humanized anti-Tac antibody including signal²⁵ sequences. Oligonucleotides used for gene synthesis are marked by solid lines: above, for oligonucleotides from upper strand, and below, for oligonucleotides from lower strand. Restriction sites used for cloning are underlined.

FIG. 12. FACS analysis of HUT-102 cells stained with ³⁰ PDL and CDR-only humanized anti-Tac antibodies and negative control antibody Fd79.

FIG. 13. Competition by mouse, PDL humanized, and CDR-only humanized anti-Tac antibodies with binding of radioiodinated mouse anti-Tac antibody to HUT-102 ceils. ³⁵

FIG. 14. Scheme for anchored polymerase chain reaction (PCR) cloning of the heavy and light chain variable domain cDNAs. RNA was prepared from about 10⁷ hybridoma cells using the hot phenol extraction method. Briefly, cells were 40 resuspended and vortexed in 1 ml of RNA extraction buffer (50 mM sodium acetate pH 5.2/1% SDS), extracted with 0.5 ml of phenol pH 5.2 at 65° C. for 15 min, followed by another 15 min on ice. The aqueous phase was recovered and precipitated twice with ethanol. cDNA was synthesized from 45 10 ug of total RNA using reverse transcriptase (BRL, Bethescda, Md.) and oligo-dT12-18 (Pharmacia, Piscatway, N.J.) as primers. A poly(dG) tail was attached to the 3' end of the cDNA using terminal deoxynucleotide transferase (BRL) (E. Y. Loh et al., Science 243, 217 (1989)), the 50 variable domain genes (V) were amplified using AmpliTaq (Perkin Elmer-Cetus) with the primer mc045 (TAATCTA-hybridized to the poly(dG) tails and primers that hybridized to the constant region genes (C). For the light chain, the 55 primer used was mc045 (TATAGAGCTCAAGCTTG-GATGGTGGGAAGAI'GGATACAGTI'GGTGC) [SEQ ID NO:12]. For the heavy chain, the primer used was mc047 (TATAGAGCTCAAGCITCCAGTGGATA-

GAC(CAT)GATGGGG(GC)TGT(TC)GTITTGGC) [SEQ 60 ID NO:13]. The sequence in parenthesis indicates a base degeneracy. The degeneracy was introduced so that the primer would be able to hybridize to most gamma chains. The amplified fragments were then digested with EcoRI and HindIII and cloned into pUC18 vector for sequencing. 65

FIG. 15. Comparison of sequences of anti-Tac heavy chain (upper lines) [SEQ ID NO:14] and Eu heavy chain

(lower lines) [SEQ ID NO:15]. The 1-letter code for amino acids is used. The first amino acid on each line is numbered at the left. Identical amino acids in the two sequences are connected by lines. The 3 CDRs are underlined. Other amino acid positions for which the anti-Tac amino acid rather than the Eu amino acid was used in the humanized anti-Tac heavy chain are denoted by an *.

FIG. 16. Comparison of sequences of anti-Tac light chain (upper lines) [SEQ ID NO:16] and Eu light chain (lower lines) [SEQ ID NO:17]. The single-letter code for amino acids is used. The first amino acid on each line is numbered at the left. Identical amino acids in the two sequences are connected by lines. The 3 CDRs are underlined. Other amino acid positions for which the anti-Tac amino acid rather than the Eu amino acid was used in the humanized anti-Tac heavy chain are denoted by an *.

FIG. 17. Nucleotide sequence of the gene for the humanized anti-Tac heavy chain variable region gene [SEQ ID NOS: 18 and 19]. The translated amino acid sequence for the part of the gene encoding protein is shown underneath the nucleotide sequence. The nucleotides TCTAGA at the beginning and end of the gene are Xba I sites. The mature heavy chain sequence begins with amino acid #20 Q.

FIG. 18. Nucleotide sequence of the gene for the humanized anti-Tac light chain variable region gene [SEQ ID NOS:20 and 21]. The translated amino acid sequence for the part of the gene encoding protein is shown underneath the nucleotide sequence. The nucleotides TCTAGA at the beginning and end of the gene are Xba I sites. The mature light chain sequence begins with amino acid #21 D.

FIG. 19A and FIG. 19B. (A) Sequences of the four oligonucleotides [SEQ ID NOS:22, 23, 24, and 25] used to synthesize the humanized anti-Tac heavy chain gene, printed 5' to 3'. (B) Relative positions of the oligonucleotides. The arrows point in the 3' direction for each oligonucleotide.

FIG. 20A and FIG. 20B. (A) Sequences of the four oligonucleotides [SEQ ID NOS:26, 27, 28, and 29] used to synthesize the humanized anti-Tac light chain gene, printed 5' to 3'. (B) Relative positions of the oligonucleotides. The arrows point in the 3' direction for each oligonucleotide. The position of a Hind III site in the overlap of JFD2 and JFD3 is shown.

FIG. 21. Schematic diagram of the plasmid pHuGTACI used to express the humanized anti-Tac heavy chain. Relevant restriction sites are shown, and coding regions of the heavy chain are displayed as boxes. The direction of transcription from the immunoglobulin (Ig) promoter is shown by an arrow. E_{ff} =heavy chain enhancer, Hyg=hygromycin resistance gene.

FIG. 22. Schematic diagram of the plasmid pHuLTAC used to express the humanized anti-Tac light chain. Relevant restriction sites are shown, and coding regions of the light chain are displayed as boxes. The direction of transcription from the Ig promoter is shown by an arrow.

FIG. 23A and FIG. 23B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:30 and 31] and heavy chain (B) [SEQ ID NOS:32 and 33] variable regions of the antibody mik- β I. The CDR sequences are underlined. The mature light chain protein begins with amino acid 23 Q and the mature heavy chain protein with amino acid 20 Q, preceded by the respective signal sequences.

FIG. 24A and FIG. 24B. Schematic diagram of the plasmids pVgl-dhfr (A) and pVk (B). The plasmid pVgldhfr contains the following parts: an approximately 4200 base pair BamHI-EcoRI fragment containing the amp and dhfr genes; a 630-bp fragment containing the human cytomegalovirus IEI gene promoter and enhancer (Boshart et al., Cell 41, 521 (1985), which is incorporated herein by reference) flanked at the 5' and 3' ends by EcoRI and Xbal linkers respectively; and a 2800 bp Xbal-BamHI fragment 5 containing the human gamma-1 constant region gene with 215 bp of the preceding intron and the poly(A) signal. The plasmid pVk was similarly constructed, with a 1530-bp human kappa constant region gene replacing the gamma-1 gene and the gpt gene replacing the dhfr gene. The plasmids were constructed from the indicated parts using methods well-known in the art (see, Maniatis et al., Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1989) and commonly assigned PCT Publication No. WO 89/09622, published Oct. 19, 1989. For example, pVgl-dhfr was 15 constructed from the plasmid pVgl by replacing the Hind III-Bgl II fragment containing the hyg gene with a 660 bp fragment containing the dhfr genc and extending to a Bgl II site (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983)). 20

FIG. 25. Fluorocytometry of YTJB cells stained with (____) Isotype matched control antibody, (- - -) humanized mik- β I antibody, (. . .) chimeric mik- β I antibody. Cells were suspended in FACS buffer (PBS+2% BSA+0.1% azide) at approximately 5×10⁶/ml. 100 ul of cell suspension was 25 transferred to a polystyrene tube and incubated with 100 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with goat antihuman Ig antibody on ice for another 30 min. Then the cells were washed and incubated with FTC labeled rabbit antigoat Ig antibody for 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldehyde. Cells were analyzed on a FACSmatc (Becton Dickinson)

FIG. 26A and FIG. 26B. Amino acid sequences of the light chain (A) [SEQ ID NOS:34 and 35] and the heavy 35 chain (B) [SEQ ID NOS:36 and 37] of the humanized mik- β l antibody, (lower lines) and human Lay antibody (upper lines), not including signal sequences. The three CDRs in each chain are underlined. Amino acids in the framework that have been replaced with mouse amino acids or consensus human amino acids in the humanized antibody are double underlined.

FIG. 27A and FIG. 27B. Oligonucleotides used in the construction of the humanized mik- β 1 heavy chain (B) [SEQ ID NOS: 42, 43, 44, and 45] and light chain (A) [SEQ 45 ID NOS:38, 39, 40, and 41]. The following pairs of oligonucleotides were mixed, extended with sequenase and cut with the indicated enzymes before ligation into the pBluescriptII ks (+) vector: wps54 and vc11 with Xba I and Sal I, vc12 and wps57 with Xba I and Sal I, vc16 and vc13 with 50 Xba I and Kpn I, vc14 and vc15 with Xba I and Kpn I. Then the wps54-vc11 and vc12-wps57 fragments were excised with Xba I and Sal I ligated together into the Xba I site of pVg1-dhfr; and the vc16-vc13 fragments and vc14-vc15 fragments were excised with Xba I and Kpn I and ligated 55 together into the Xba I site of pVk.

FIG. 28. Competitive binding of labeled mik- β l tracer to YTJB cells. About 10⁶ YTJB cells were incubated with 3.0 ng of radio-iodinated mouse mik- β I antibody (6 µCi/µg) and varying amounts of either unlabeled mouse mik- β I antibody 60 (•) or humanized mik- β I antibody (o) in 200 ul of binding buffer (PBS+10% fetal calf serum+0.1% NaN₃+10 µg/ml mouse monoclonal lg). After incubation for 2 hr at 0° C. the cells were washed twice with binding buffer without mouse Ig and collected by centrifugation. The radioactivity bound 65 to cells was measured and expressed as the ratio of bound/ free cpm.

FIG. 29. Inhibition of 11.-2 stimulated proliferation of human PHA blasts by humanized mik- β 1+humanized anti-Tac antibodies. No antibody added (\Box), 2 ug cach of humanized mik- β 1 and humanized anti-Tac added (\blacksquare).

FIG. 30A through FIG. 30D. Amino acid sequences of the heavy chain (A) [SEQ ID NOS:48 and 49] and the light chain (B) [SEQ ID NOS:46 and 47] of the murine and humanized Fd79 antibodies, and the heavy chain (C) (SEQ ID NOS:52 and 53] and the light chain (D) [SEQ ID NOS:50 and 51] of the murine and humanized Fd138-80 antibodies. The sequences of the murinc antibody as deduced from the cDNA (upper lines) are shown aligned with the humanized antibody sequences (lower lines). The humanized Fd79 and Fd138-80 framework sequences are derived from Pom antibody and Eu antibody, respectively. Residues are numbered according to the Kabat system (E. A. Kabat et al., Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, Md.) (1987). The three CDRs in each chain arc boxed. Residues in the Pom or Eu framework that have been replaced with mutine sequences or consensus human sequences are underlined.

FIG. 31A and FIG. 31B. Fluorocytometry of HSV-I infected Vero cells stained with Fd79 (A) and Fd138-80 (B) antibodies, (. .) Isotype matched control antibody, (. . .) humanized antibody, (_____) chimeric antibody. Vero cells were infected with HSV-1 (A305 mutant (F strain)) at 3 pfu/cell overnight. Cells were trypsinized at 0.5 mg/ml for I minute, washed extensively with PBS and resuspended in FACS buffer (PBS+2% BSA+0.1% azide) at approximately 5×10⁶/ml. 100 ul of cell suspension was transferred to a polystyrene tube and incubated with 100 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with FITC labeled goat antihuman antibody (Cappel) on ice for another 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldchydc. Cells were analyzed on a FACSmate (Becton Dickinson).

FIG. 32A and FIG. 32B. Neutralization of HSV-1 by Fd79 (A) and Fd138-80 (B). Serial dilutions of antibodies were mixed with 100 pfn of virus and incubated at 37° C. for 1 hr. The viruses were then inoculated onto 6-well plates with confluent Vero cells and adsorbed at 37° C. for I hr. Cells were overlayed with I% agarose in medium and incubated for 4 days. Plaques were stained with neutral rcd.

FIG. 33A and FIG. 33B. Immunostaining of infected Vero cell monolayers to examine protection of cells from viral spread in tissue culture by (A) murine or human'ized Fd79, (B) murine or humanized Fd138-80. 24-well plates of confluent Vero cells were inoculated with virus at 0.1 pfu/cell and allowed to adsorb for 2 hrs. at 37° C. before adding 200 ul of 10 ug/ml antibodies in medium. At the end of 4 days, culture medium was removed and plates were dried by placing overnight in a 37° C. incubator. To detect viral antigens, cach well was incubated with 200 ul of anti-gB antibody at 0.5 ug/ml for 1 hr. at 37° C., washed twice and incubated with 200 ul of peroxidase conjugated goat antimouse IgG (Cappel, 1:300 dilution) for 1 hr. at 37° C. The plates were washed and then developed with the substrate 3-amino-9-cthyl-carbazole (AEC) (Sigma, St. Louis, Mo.) for 15 minutes at room temperature. Reaction was stopped by rinsing with water and air dried.

FIG. 34A and FIG. 34B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:66 and 67] and heavy chain (B) [SEQ ID NOS:68 and 69] variable regions of the antibody M195. The CDR sequences are underlined. The mature light chain protein

begins with amino acid 21 D and the mature heavy chain protein with amino acid 20 E, preceded by the respective signal sequences.

FIG. 35. Fluorocytometry of U937 cells stained with (..) no antibody, (...) humanized M195 antibody, (---) chimeric 5 M195 antibody. Cells were suspended in FACS buffer (PBS+2% FCS+0.1% azide) at approximately 5×10^{67} ml. 100 ul of cell suspension was transferred to a polystyrene tube and incubated with 50 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with FITC labeled goat anti-human lg antibody on ice for another 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldehydc. Cells were analyzed on a FACSmate (Becton Dickinson).

FIG. 36A and FIG. 36B. Amino acid sequences of the 15 light chain (A) [SEQ ID NOS:70 and 71] and the heavy chain (B) [SEQ ID NOS:72 and 73] of the humanized M195 antibody (lower lines) and human Eu antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework that have 20 been replaced with mouse amino acids in the humanized antibody are double underlined.

FIG. 37A and FIG. 37B. Oligonucleotides used in the construction or the humanized M195 heavy chain (A; mal-4) [SEQ ID NOS:74, 75, 76, and 77] and light chain (B; ma5-8) ²⁵ [SEQ ID NOS:78, 79, 80, and 81]. The following pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: mal and ma2 with Xba 1 and Kpn I, ma3 and ma4 with Xba I and Kpn I, ma5 and ma6 with Xba I and ³⁰ Hind III, ma7 and ma8 with Xba 1 and Hind III. Then the mal-ma2 and ma3-ma4 fragments were excised from pUC18 with Xba I and kpn 1 and ligated together into the Xba I site of pVgl-dhfr, and the ma5-ma6 and ma7-ma8 fragments were excised with Xba I and Hind III and ligated together ³⁵ into the Xba I site of pVk.

FIG. 38. Competitive binding of labeled M195 tracer to U937 cells. About 4×10^5 U937 cells were incubated with 4.5 ng of radio-iodinated mouse M195 antibody (6 µci/µg) and varying amounts of either unlabeled mouse M195 antibody ⁴⁰ (•) or humanized M195 antibody (o) in 200 ul of binding buffer (PBS+2% fetal calf serum+0.1% sodium azide). After incubation for 2 hr at 0° C., the cells were washed twice with binding buffer and collected by centrifugation. The radio-activity bound to cells was measured and is expressed as the ⁴⁵ ratio of bound/free cpm.

FIG. 39A and FIG. 39B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS: 82 and 83] and heavy chain (B) [SEQ ID NOS:84 and 85] variable regions of the antibody CMV5. The CDR sequences are underlined. The start of the mature protein sequences are indicated by arrows, preceded by the respective signal sequences.

FIG. 40A and FIG. 40B. Amino acid sequences of the 55 light chain (A) [SEQ ID NOS:86 and 87] and the heavy chain (B) [SEQ ID NOS:88 and 89] of the humanized CMV5 antibody (lower lines) and human Wol antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework replaced with mouse amino acids or typical human amino acids in the humanized antibody are double underlined.

FIG. 41A and FIG. 41B. Oligonucleotides used in the construction of the humanized CMV5 light chain (A; jbl6-65 jbl9) [SEQ ID NOS:90, 91, 92, and 93] and heavy chain (B; jb20-jb22) [SEQ ID NOS:94, 95, 96, and 97]. The following

pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: jb16 and jb17 with Xba I and EcoR I, jb18 and jb19 with Xba I and EcoR I, jb20 and jb21 with Xba I and Kpn I, jb22 and jb23 with Xba I and Kpn I. Then the jb16-jb17 and jb18-jb19 fragments were excised with Xba I and Mlu I and ligated together into the Xba I site of pVk; and the jb20-jb21 and jb22-jb23 fragments were excised with Xba I and Kpn I and ligated together into the Xba I site of pVg1-dhfr.

FIG. 42. Competitive binding of labeled CMV5 water to CMV-infected cells. Increasing amounts of mouse () or humanized () CMV5 antibody was added to CMV-infected HEL cells with tracer radio-iodinated mouse CMV5, and the amount of tracer bound to the cells was determined.

FIG. 43A and FIG. 43B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:98 and 99] and heavy chain (B) [SEQ ID NOS:100 and 101] variable regions of the antibody AF2. The CDR sequences are underlined. The mature light chain protein begins with amino acid 30 N and the mature heavy chain protein with amino acid 36 Q, preceded by the respective signal sequences.

FIG. 44A and FIG. 44B. Amino acid sequences of the light chain (A) [SEQ ID NOS:102 and 103] and the heavy chain (B) [SEQ ID NOS: 104 and 105] of the humanized AF2 antibody (lower lincs) and human Eu antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework that have been replaced with mouse amino acids or typical human amino acids in the humanized antibody are double underlined.

FIG. 45A and FIG. 45B. Oligonucleotides used in the construction of the humanized AF2 light chain (A; rh10-rh13) [SEQ ID NOS:106, 107, 108, and 109] and heavy chain (B; rh20-23) [SEQ ID NOS:110, 111, 112, and 113]. The following pairs of oligonucleotides were mixed, extended with Kienow polymerase and cut with the indicated enzymes before ligation into pUC18: rh10 and rh11 with Xba I and Hind 111, rh12 and rh13 with Xba I and Hind 111, rh20 and rh21 with Xba I and EcoR I, rh22 and rh23 with Xba I and EcoR I. Then the rh10-rh11 and rh12-rh13 fragments were excised with Xba I and Hind 111 and ligated together into then Xba I site of pVk; and the rh20-rh21 and rh22-rh23 fragments were excised with Xba I and Xho I and ligated together into the Xba I site of pVgl-dhfr.

FIG. 46. Fluorescence of HS294T cells incubated with γ -IFN plus varying concentrations of mouse AF2 antibody, and stained with an anti-HLA-D antibody.

DETAILED DESCRIPTION OF THE INVENTION

In accordance with the present invention, novel means of designing humanized immunoglobulins capable of specifically binding to a predetermined antigen with strong affinity are provided. These improved methods produce immuno-globulins that are substantially non-immunogenic in humans but have binding affinities of at least about 10^8 M^{-1} , preferably 10^9 M^{-1} to 10^{10} M^{-1} , or stronger. The humanized immunoglobulins will have a human framework and have one or more complementary determining regions (CDR's), plus a limited number of other amino acids, from a donor immunoglobulins can be produced economically in large quantities and find use, for example, in the treatment of

various human disorders by a variety of techniques.

In order that the invention may be more completely understood, several definitions are set forth. As used herein, the term "immunoglobulin" refers to a protein consisting of one or more polypeptides substantially encoded by immu-5 noglobulin genes. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma (IgG1, IgG2, IgG3 IgG₄), delta, epsilon and mu constant region genes, as well as the myriad immunoglobulin variable region genes. Fulllength immunoglobulin "light chains" (about 25 Kd or 214 amino acids) are encoded by a variable region gene at the NH2-terminus (about 110 amino acids) and a kappa or lambda constant region gene at the COOH - terminus. Full-length immunoglobulin "heavy chains" (about 50 Kd or 446 amino acids), are similarly encoded by a variable region 15 gene (about 116 amino acids) and one of the other aforementioned constant region genes, e.g., gamma (encoding about 330 amino acids).

One form of immunoglobulin constitutes the basic structural unit of an antibody. This form is a tearamer and consists 20 of two identical pairs of immunoglobulin chains, each pair having one light and one heavy chain. In each pair, the light and heavy chain variable regions arc together responsible for binding to an antigen, and the constant regions are responsible for the antibody effector functions. In addition to 25 antibodies, immunoglobulins may exist in a variety of other forms including, for example, Fv, Fab, and (Fab')2, as well as bifunctional hybrid antibodies (e.g., Lanzavecchia et al., Eur. J. Immunol. 17, 105 (1987)) and in single chains (e.g., Huston et al., Proc. Natl. Acad. Sci. U.S.A., 85, 5879-5883 30 (1988) and Bird et al., Science, 242, 423-426 (1988), which are incorporated herein by reference). (See, generally, Hood et al., "Immunology", Benjamin, N.Y., 2nd ed. (1984), and Hunkapiller and Hood, Nature, 323, 15-16 (1986), which are incorporated herein by reference).

An immunoglobulin light or heavy chain variable region consists of a "framework" region interrupted by three hypervariable regions, also called CDR's. The extent of the framework region and CDR's have been precisely defined (see, "Sequences of Proteins of Immunological Interest," E. 40 Kabat et al., U.S. Department of Health and Human Services, (1983); which is incorporated herein by reference). The sequences of the framework regions of different light or heavy chains are relatively conserved within a species. As used herein, a "human framework region" is a framework 45 region that is substantially identical (about 85% or more, usually 90-95% or more) to the framework region of a naturally occurring human immunoglobulin. The framework region of an antibody, that is the combined framework regions of the constituent light and heavy chains, serves to 50 position and align the CDR's. The CDR's are primarily responsible for binding to an epitope of an antigen.

Chimeric antibodies are antibodies whose light and heavy chain genes have been constructed, typically by genetic engineering, from immunoglobulin variable and constant region genes belonging to different species. For example, the variable segments of the genes from a mouse monoclonal antibody may be joined to human constant segments, such as gamma 1 and gamma 3. A typical therapeutic chimeric antibody is thus a hybrid protein composed of the variable or antigen-binding domain from a mouse antibody and the constant or effector domain from a human antibody (e.g., A.T.C.C. Accession No. CRL 9688 secretes an anti-Tac chimeric antibody), although other mammalian species may be used.

As used herein, the term "humanized" immunoglobulin refers to an immunoglobulin comprising a human frame-

work region and one or more CDR's from a non-human (usually a mouse or rat) immunoglobulin. The non-human immunoglobulin providing the CDR's is called the "donor" and the human immunoglobulin providing the framework is called the "acceptor". Constant regions need not be present, but if they are, they must be substantially identical to human immunoglobulin constant regions, i.e., at least about 85-90%, preferably about 95% or more identical. Hence, all parts of a humanized immunoglobulin, except possibly the CDR's, are substantially identical to corresponding parts of natural human immunoglobulin sequences. A "humanized antibody" is an antibody comprising a humanized light chain and a humanized heavy chain immunoglobulin. For example, a humanized antibody would not encompass a typical chimeric antibody as defined above, e.g., because the entire variable region of a chimeric antibody is non-human. One says that the donor antibody has been "humanized", by the process of "humanization", because the resultant humanized antibody is expected to bind to the same antigen as the donor antibody that provides the CDR's.

It is understood that the humanized antibodies designed by the present method may have additional conservative amino acid substitutions which have substantially no effect on antigen binding or other immunoglobulin functions. By conservative substitutions is intended combinations such as gly, ala; val, ile, leu; asp, glu; asn, gln; ser, thr; lys, arg; and phe, tyr.

Humanized immunoglobulins, including humanized antibodies, have been constructed by means of genetic engineering. Most humanized immunoglobulins that have been previously described (Jones et al., op. cit.; Verhoeyen et al., op. cit.; Riechmann et al., op. cit.) have comprised a framework that is identical to the framework of a particular human immunoglobulin chain, the acceptor, and three CDR's from a non-human donor immunoglobulin chain. In one case (Ricchmann et al., op. cit.), two additional amino acids in the framework were changed to be the same as amino acids in other human framework regions. The present invention includes criteria by which a limited number of amino acids in the framework of a humanized immunoglobulin chain are chosen to be the same as the amino acids at those positions in the donor rather than in the acceptor, in order to increase the affinity of an antibody comprising the humanized immunoglobulin chain.

The present invention is based in part on the model that two contributing causes of the loss of affinity in prior means of producing humanized antibodies (using as examples mouse antibodies as the source of CDR's) arc:

- (1) When the mouse CDR's are combined with the human framework, the amino acids in the framework close to the CDR's become human instead of mouse. Without intending to be bound by theory, we believe that these changed amino acids may slightly distort the CDR's, because they create different electrostatic or hydrophobic forces than in the donor mouse antibody, and the distorted CDR's may not make as effective contacts with the antigen as the CDR's did in the donor antibody;
- (2) Also, amino acids in the original mouse antibody that are close to, but not part of, the CDR's (i.e., still part of the framework), may make contacts with the antigen that contrabute to affinity. These amino acids are lost when the antibody is humanized, because all framework amino acids are made human.

To avoid these problems, and to produce humanized antibodies that have a very strong affinity for a desired 5

antigen, the present invention uses one or more of the following principles for designing humanized immunoglobulins. Also, the criteria may be used singly, or when necessary in combination, to achieve the desired affinity or other characteristics.

A principle is that as acceptor, a framework is used from a particular human immunoglobulin that is unusually homologous to the donor immunoglobulin to be humanized, or use a consensus framework from many human antibodies. For example, comparison of the sequence of a mouse heavy 10 (or light) chain variable region against human heavy (or light) variable regions in a data bank (for example, the National Biomedical Research Foundation Protein Identification Resource) shows that the extent of homology to different human regions varies greatly, typically from about 15 40% to about 60-70%. By choosing as the acceptor immunoglobulin one of the human heavy (respectively light) chain variable regions that is most homologous to the heavy (respectively light) chain variable region of the donor immunoglobulin, fewer amino acids will be changed in going 20 from the donor immunoglobulin to the humanized immunoglobulin. Hence, and again without intending to be bound by theory, it is believed that there is a smaller chance of changing an amino acid near the CDR's that distorts their conformation. Moreover, the precise overall shape of a 25 humanized antibody comprising the humanized immunoglobulin chain may more closely resemble the shape of the donor antibody, also reducing the chance of distorting the CDR's.

Typically, one of the 3–5 most homologous heavy chain 30 variable region sequences in a representative collection of at least about 10 to 20 distinct human heavy chains will be chosen as acceptor to provide the heavy chain framework, and similarly for the light chain. Preferably, one of the 1–3 most homologous variable regions will be used. The selected 35 acceptor immunoglobulin chain will most preferably have at least about 65% homology in the framework region to the donor immunoglobulin.

In many cases, it may be considered preferable to use light and heavy chains from the same human antibody as acceptor 40 sequences, to be sure the humanized light and heavy chains will make favorable contacts with each other. In this case, the donor light and heavy chains will be compared only against chains from human antibodies whose complete sequence is known, e.g., the Eu, Lay, Pom, Wol, Sie, Gal, Ou 45 and WEA antibodies (Kabat et al., op. cit.; occasionally, the last few amino acids of a human chain are not known and must be deduced by homology to other human antibodies). The human antibody will be chosen in which the light and heavy chain variable regions sequences, taken together, are 50 overall most homologous to the donor light and heavy chain variable region sequences. Sometimes greater weight will be given to the heavy chain sequence. The chosen human antibody will then provide both light and heavy chain acceptor sequences. In practice, it is often found that the 55 human Eu antibody will serve this role.

Regardless of how the acceptor immunoglobulin is chosen, higher affinity may be achieved by selecting a small number of amino acids in the framework of the humanized immunoglobulin chain to be the same as the amino acids at 60 those positions in the donor rather than in the acceptor. A second principle is that the following categories define what amino acids may be selected from the donor. Preferably, at many or all amino acid positions in one of these categories, the donor amino acid will in fact be selected. 65

Category 1: The amino acid position is in a CDR is defined by Kabat et al., op. cit.

Category 2: If an amino acid in the framework of the human acceptor immunoglobulin is unusual (i e., "rare", which as used herein indicates an amino acid occurring at that position in less than about 20% but usually less than about 10% of human heavy (respectively light) chain V region sequences in a representative data bank), and if the donor amino acid at that position is typical for human sequences (i.e., "common", which as used herein indicates an amino acid occurring in more than about 25% but usually more than about 50% of sequences in a representative data bank), then the donor amino acid rather than the acceptor may be selected. This criterion helps ensure that an atypical amino acid in the human framework does not disrupt the antibody structure. Moreover, by replacing an unusual amino acid with an amino acid from the donor antibody that happens to be typical for human antibodies, the humanized antibody may be made less immunogenic.

All human light and heavy chain variable region sequences are respectively grouped into "subgroups" of sequences that are especially homologous to each other and have the same amino acids at certain critical positions (Kabat et al., op. cit.). When deciding whether an amino acid in a human acceptor sequence is "rare" or "common" among human sequences, it will often be preferable to consider only those human sequences in the same subgroup as the acceptor sequence.

Category 3: In the positions immediately adjacent to one or more of the 3 CDR's in the primary sequence of the humanized immunoglobulin chain, the donor amino acid(s) rather than acceptor amino acid may be selected. These amino acids are particularly likely to interact with the amino acids in the CDR's and, if chosen from the acceptor, to distort the donor CDR's and reduce affinity. Moreover, the adjacent amino acids may interact directly with the antigen (Amit et al., *Science*, 233, 747–753 (1986), which is incorporated herein by reference) and selecting these amino acids from the donor may be desirable to keep all the antigen contacts that provide affinity in the original antibody.

Category 4: A 3-dimensional model, typically of the original donor antibody, shows that certain amino acids outside of the CDR's are close to the CDR's and have a good probability of interacting with amino acids in the CDR's by hydrogen bonding, Van der Waals forces, hydrophobic interactions, etc. At those amino acid positions, the donor immunoglobulin amino acid rather than the acceptor immunoglobulin amino acid may be selected. Amino acids according to this criterion will generally have a side chain atom within about 3 angstrom units of some atom in the CDR's and must contain an atom that could interact with the CDR atoms according to established chemical forces, such as those listed above.

In the case of atoms that may form a hydrogen bond, the 3 angstroms is measured between their nuclei, but for atoms that do not form a bond, the 3 angstroms is measured between their Van der Waals surfaces. Hence, in the latter case, the nuclei must be within about 6 angstroms (3+sum of the Van der Waals radii) for the atoms to be considered capable of interacting. In many cases the nuclei will be from 4 or 5 to 6 Å apart. In determining whether an amino acid can interact with the CDRs, it is preferred not to consider the last 8 amino acids of heavy chain CDR 2 as part of the CDRs, because from the viewpoint of structure, these 8 amino acids behave more as part of the framework.

Amino acids in the framework that are capable of interacting with amino acids in the CDR's, and which therefore belong to Category 4, may be distinguished in another way. The solvent accessible surface area of each framework amino acid is calculated in two ways: (I) in the intact antibody, and (2) in a hypothetical molecule consisting of the antibody with its CDRs removed. A significant difference between these numbers of about 10 square angstroms or more shows that access of the framework amino acid to solvent is at least partly blocked by the CDRs, and therefore that the amino acid is making contact with the CDRs. Solvent accessible surface area of an amino acid may be calculated based on a 3-dimensional model of an antibody, using algorithms known in the art (e.g., Connolly, J. Appl. 10 Cryst. 16,548 (1983) and Lee and Richards, J. Mol. Biol. 55, 379 (1971), both of which are incorporated herein by reference). Framework amino acids may also occasionally interact with the CDR's indirectly, by affecting the conformation of another framework amino acid that in turn con- 15 tacts the CDR's.

The amino acids at several positions in the framework are known to be capable of interacting with the CDRs in many antibodies (Chothia and Lesk, J. Mol. Biol. 196, 901 (1987), Chothia et al., Nature 342, 877 (1989), and Tramontano et 20 al., J. Mol. Biol. 215, 175 (1990), all of which are incorporated herein by reference), notably at positions 2, 48, 64 and 71 of the light chain and 26-30, 71 and 94 of the heavy chain (numbering according to Kabat, op. cit.), and therefore these amino acids will generally be in Category 4. Typically, 25 humanized immunoglobulins, of the present invention will include donor amino acids (where different) in category 4 in addition to these. The amino acids at positions 35 in the light chain and 93 and 103 in the heavy chain are also likely to interact with the CDRs. At all these numbered positions, 30 choice of the donor amino acid rather than the acceptor amino acid (when they differ) to be in the humanized immunoglobulin is preferred. On the other hand, certain positions that may be in Category 4 such as the first 5 amino acids of the light chain may sometimes be chosen from the 35 acceptor immunoglobulin without loss of affinity in the humanized immunoglobulin.

Chothia and Lesk (op. cit.) define the CDRs differently from Kabat et al. (op. cit.). Notably, CDR1 is defined as including residues 26-32. Accordingly, Riechmann ct al., 40 (op. cit.) chose these amino acids from the donor immunoglobulins.

Computer programs to create models of proteins such as antibodies are generally available and well known to those shilled in the art (sec, Levy et al., *Biochemistry*, 28, 7168-7175 (1989); Bruccoleri et al., *Nature*, 335, 564–568 (1988); Chothia et al., *Science*, 233, 755–758 (1986), all of which are incorporated herein by reference). These do not form part of the invention. Indeed, because all antibodies have similar structures, the known antibody structures, which are 50 available from the Brookhaven Protein Data Bank, can be used if necessary as rough models of other antibodies. Commercially available computer programs can be used to display these models on a computer monitor, to calculate the distance between atoms, and to estimate the likelihood of 55 different amino acids interacing (see, Feirin et al., *J. Mol. Graphics*, 6, 13–27 (1988)).

In addition to the above categories, which describe when an amino acid in the humanized immunoglobulin may be taken from the donor, certain amino acids in the humanized 60 immunoglohulin may be taken from neither the donor nor acceptor, if then fall in:

Category 5: If the amino acid at a given position in the donor immunoglobulin is "rare" for human sequences, and the amino acid at that position in the acceptor immunoglo-55 bulin is also "rare" for human sequences, as defined above, then the amino acid at that position in the humanized

immunoglobulin may be chosen to be some amino acid "typical" of human sequences. A preferred choice is the amino acid that occurs most often at that position in the known human sequences belonging to the same subgroup as the acceptor sequence.

Humanized antibodies generally have at least three potential advantages over mouse or in some cases chimeric antibodies for use in human therapy:

- Because the effector portion is human, it may interact better with the other parts of the human immune system (e.g., destroy the target cells more efficiently by complement-dependent cytotoxicity (CDC) or antibody-dependent cellular cytotoxicity (ADCC)).
- 2) The human immune system should not recognize the framework or constant region of the humanized antibody as foreign, and therefore the antibody response against such an injected antibody should be less than against a totally foreign mouse antibody or a partially foreign chimeric antibody.
- 3) Injected mouse antibodies have been reported to have a half-life in the human circulation much shorter than the half-life of normal antibodies (D. Shaw et al., J. Immunol., 138, 4534-4538 (1987)). Injected humanized antibodies will presumably have a half-life more similar to naturally occurring human antibodies, allowing smaller and less frequent doses to be given.

In one aspect, the present invention is directed to designing humanized immunoglobulins that are produced by expressing recombinant DNA segments encoding the heavy and light chain CDR's from a donor immunoglobulin capable of binding to a desired antigen, such as the human IL-2 receptor, attached to DNA segments encoding acceptor human framework regions. Exemplary DNA sequences designed in accordance with the present invention code for the polypeptide chains comprising heavy and light chain CDR's with substantially human framework regions shown in FIG. 1A through FIG. 6B. Due to codon degeneracy and non-critical amino acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. In general, the criteria of the present invention find applicability to designing substantially any humanized immunoglobulin.

The DNA segments will typically further include an expression control DNA sequence operably linked to the humanized immunoglobulin coding sequences, including naturally-associated or hctcrologous promoter regions. Preferably, the expression control sequences will be eukaryotic promoter systems in vectors capable of transforming or transfecting eukaryotic host cells, but control sequences for prokaryotic hosts may also be used. Once the vector has been incorporated into the appropriate host, the host is maintained under conditions suitable for high level expression of the nucleotide sequences, and, as desired, the collection and purification of the humanized light chains, heavy chains, light/heavy chain dimers or intact antibodies, binding fragments or other immunoglobulin forms may follow (see, S. Beychok, Cells of Immunoglobulin Synthesis, Academic Press, N.Y., (1979), which is incorporated herein by reference).

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immortalized B-cells (see, Kabat op. cit. and WP87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to the predetermined antigen, such as the human IL-2 receptor, and produced by well known methods in any convenient mammalian source including, mice, rats, rabbits, or other vertebrates, capable of producing antibodies. Suitable source cclls for the constant region and framework DNA sequences, and host cells for immunoglobulin expression and secretion, can be obtained from a number of sources, 5 such as the American Type Culture Collection ("Catalogue of Cell Lines and Hybridomas," sixth edition (1988) Rockville, Md. U.S.A., which is incorporated herein by reference).

In addition to the humanized immunoglobulins specifi- 10 cally described herein, other "substantially homologous" modified immunoglobulins to the native sequences can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the framework regions can vary specifi- 15 cally from the sequences in FIG. 1A through FIG. 6B at the primary structure level by several amino acid substitutions, terminal and intermediate additions and deletions, and the like. Moreover, a variety of different human framework regions may be used singly or in combination as a basis for 20 the humanized immunoglobulins of the present invention. In general, modifications of the genes may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Gillman and Smith, Gene, 8, 81-97 (1979) and S. Roberts et al., Nature, 328, 731-734 25 (1987), both of which are incorporated herein by reference).

Substantially homologous immunoglobulin sequences are those which exhibit at least about 85% homology, usually at least about 90%, and preferably at least about 95% homology with a reference immunoglobulin protein. 30

Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic 35 cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors pVk and pVgl (FIGS. 9A and 9B) using sitedirected mutagenesis, such as after CH1 to produce Fab fragments or after the hinge region to produce (Fab')₂ 40 fragments. Single chain antibodies may be produced by joining VL and VH with a DNA linker (see, Huston et al., op. cit., and Bird et al., op. cit.). Also because like many genes, the immunoglobulin-related genes contain separate functional regions, each having one or more distinct biological 45 activitics, the genes may be fused to functional regions from other genes (e.g., enzymes, see, commonly assigned U.S. Pat. No. 5,004,692) to produce fusion proteins (e.g., immunotoxins) having novel properties. The nucleic acid sequences of the present invention capable of ultimately 50 expressing the desired humanized antibodies can be formed from a variety of different polynucleotides (genomic or cDNA, RNA, synthetic oligonucleotides, etc.) and components (e.g., V, J, D, and C regions), as well as by a variety of different techniques. Joining appropriate synthetic and 55 genomic sequences is presently the most common method of production, but cDNA sequences may also be utilized (see, European Patent Publication No. 0239400 and L. Reichmann et al., Nature, 332, 323-327 (1988), both of which are incorporated herein by reference).

As stated previously, the DNA sequences will be expressed in hosts after the sequences have been operably linked to (i.e., positioned to ensure the functioning of) an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes 65 or as an integral part of the host chromosomal DNA. Commonly, expression vectors will contain selection mark-

ers, e.g., tetracycline or neomycin, to permit detection of those cells transformed with the desired DNA sequences (see, e.g., U.S. Pat. No. 4,704,362, which is incorporated herein by reference).

E. coli is one prokaryotic host useful particularly for cloning the DNA sequences of the present invention. Other microbial hosts suitable for use include bacilli, such as Bacillus subtilus, and other enterobacteriaceae, such as Salmonella, Serratia, and various Pseudomonas species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of wellknown promoters will be present, such as the lactose promoter system, a tryptophan (p) promoter system, a betalactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing transcription and translation.

Other microbes, such as yeast, may also be used for expression. Saccharomyces is a preferred host, with suitable vectors having expression control sequences, such as promoters, including 3-phosphoglycerate kinase or other glycolytic enzymes, and an origin of replication, termination sequences and the like as desired.

In addition to microorganisms, mammalian tissue cell culture may also be used to express and produce the polypeptides of the present invention (see, Winnacker, "From Genes to Ciones," VCH Publishers, N.Y., N.Y. (1987), which is incorporated herein by reference). Eukaryotic cells are actually preferred, because a number of suitable host cell lines capable of secreting intact immunoglobulins have been developed in the art, and include the CHO cell lines, various COS cell lines, HeLa cells, preferably myeloma cell lines, etc, and transformed B-cells or hybridomas. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter, an enhancer (Queen et al., Immunol. Rev., 89, 49-68 (1986), which is incorporated herein by reference), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences are promoters derived from immunoglobulin genes, SV40, Adenovirus, cytomegalovirus, Bovine Papilloma Virus, and the like.

The vectors containing the DNA segments of interest (e.g., the heavy and light chain encoding sequences and expression control sequences) can be transferred into the host cell by well-known methods, which vary depending on the type of cellular host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cellular hosts. (See, generally, Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press, (1982), which is incorporated herein by reference.)

Once expressed, the whole antibodies, their dimers, individual light and heavy chains, or other immunoglobulin forms of the present invention, can be purified according to standard procedures of the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like (see, generally, R. Scopes, "Protein Purification", Springer-Verlag, N.Y. (1982)). Substantially pure immunoglobulins of at least about 90 to 95% homogeneity are preferred, and 98 to 99% or more homogeneity most preferred, for pharmaceutical uses. Once purified, partially or to homogeneity as desired, the polypeptides may then be used therapeutically (including extracorporeally) or in developing and performing assay procedures, immunofluorescent stainings, and the like. (See, generally, *Immunological Methods*, Vols. I and II, Lefkovits and Per-5 nis, eds., Academic Press, New York, N.Y. (1979 and 1981)).

The antibodies of the present invention will typically find use individually in treating substantially any disease susceptible to monoclonal antibody-based therapy. In particular, the immunoglobulins can be used for passive immuni- 10 zation or the removal of unwanted cells or antigens, such as by complement mediated lysis, all without substantial immune reactions (e.g., anaphylactic shock) associated with many prior antibodies. For example, where the cell linked to a disease has been identified as IL-2 receptor bearing, then 15 humanized antibodies that bind to the human IL-2 receptor are suitable (see, U.S. Ser. No. 085,707, entitled "Treating Human Malignancies and Disorders," which is incorporated herein by reference). For such a humanized immunoglobulin, typical disease states suitable for treatment include graft 20 versus host disease and mansplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia 25 gravis.

The method of producing humanized antibodies of the present invention can be used to humanize a variety of donor antibodies, especially monoclonal antibodies reactive with markers on cells responsible for a disease. For example, 30 suitable antibodies bind to antigens on T-cells, such as those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, Leukocyte Typing, Bernard et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by refer- 35 ence.

The antibodies of the present invention can also be used as separately administered compositions given in conjunction with chemotherapeutic or immunosuppressive agents. Possible agents include cyclosporin A or a purine analog 40 (e.g., methotrexate, 6-mercaptopurine, or the like), but numerous additional agents (e.g., cyclophosphamide, prednisone, etc.) well-known to those skilled in the art of medicine may also be utilized.

A preferred pharmaceutical composition of the present 45 invention comprises the use of the subject antibodies in immunotoxins. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. 50 The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells comprising a carcinoma. The two components are commonly chemically bonded together by any of a variety of well-known chemical procedures. For 55 example, when the cytotoxic agent is a protein and the second component is an intact immunoglobulin, the linkage may be by way of heterobifunctional cross-linkers, e.g., SPDP, carbodiimide, glutaraldehyde, or the like. Production of various immunotoxins is well-known with the art, and can 60 be found, for example in "Monoclonal Antibody-Toxin Conjugates: Aiming the Magic Bullet," Thorpe et al., Monoclonal Antibodies in Clinical Medicine, Academic Press, pp. 168-190 (1982), which is incorporated herein by reference. The components may also be linked genetically (see, 65 Chaudhary et al., Nature 339, 394 (1989), which is herein incorporated by reference).

A variety of cytotoxic agents arc suitable for use in immunotoxins. Cytotoxic agents can include radionuclides, such as lodine-131 or other isotopes of iodine, Yttrium-90, Rhenium-188, and Bismuth-212 or other alpha emitters; a number of chemotherapeutic drugs, such as vindesine, methotrexate, adtiamycin, and cisplatinum; and cytotoxic proteins such as ribosomal inhibiting proteins like pokeweed antiviral protein, Pseudomonas exotoxin A, ricin, diphtheria toxin, ricin A chain, etc., or an agent active at the cell surface, such as the phospholipase enzymes (e.g., phospholipase C). (See, generally, "Chimeric Toxins," Olsnes and Phil, *Pharmac. Ther.*, 25, 355–381 (1982), and "Monoclonal Antibodies for Cancer Detection and Therapy," eds. Baldwin and Byers, pp. 159–179, 224–266, Academic Press (1985), all of which are incorporated herein by reference.)

The delivery component of the immunotoxin will include the humanized immunoglobulins of the present invention. Intact immunoglobulins or their binding fragments, such as Fab, are preferably used. Typically, the antibodies in the immunotoxins will be of the human lgM or lgG isotypc, but other mammalian constant regions may be utilized as desired.

For diagnostic purposes, the antibodies may either be labeled or unlabeled. Unlabeled antibodies can be used in combination with other labeled antibodies (second antibodies) that are reactive with the humanized antibody, such as antibodies specific for human immunoglobulin constant regions. Alternatively, the antibodies can be directly labeled. A wide variety of labels may be employed, such as radionuclides, fluors, enzymes, enzyme substrates, enzyme cofactors, enzyme inhibitors, ligands (particularly haptens), etc. Numerous types of immunoassays are available and are well known to those skilled in the art.

Anti-II-2 Receptor Antibodies

To exert its biological effects, IL-2 interacts with a specific high-affinity membrane receptor (Greene, W., et al., Progress in Hematology XIV, E. Brown, Ed., Grune and Statton, New York (1986), at pgs. 283 ff and Waldmann, Ann. Rev. Biochem. 58, 875 (1989), both of which are incorporated herein by reference). The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide, being about 55 kD in size (see, Leonard, W., ct al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). A gene encoding this protein has been isolated, and predicts a 272 amino acid peptide, including a 21 amino acid signal peptide (see, Leonard, W., et al., Nature 311, 626 (1984)). The 219 NH₂-terminal amino acids of the p55 Tac protein apparently comprise an extracellular domain (see, Leonard, W., et al., Science, 230, 633-639 (1985), which is incorporated herein by reference).

Much of the elucidation of the human IL-2 receptor's structure and function is due to the development of specifically reactive monoclonal antibodies. In particular, one mouse monoclonal antibody, known as anti-Tac (Uchiyama, et al., *J. Immunol.* 126, 1393 (1981)) has been used to show that IL-2 receptors can be detected on T-cells, but also on cells of the monocyte-macrophage family, Kupffer cells of the liver, Langerhans' cells of the skin and, of course, activated T-cells. Importantly, resting T-cells, B-cells or circulating machrophages typically do not display the IL-2 receptor (Hermann, et al., *J. Exp. Med.* 162, 1111 (1985)).

The anti-Tac monoclonal antibody has also been used to define lymphocyte functions that require IL-2 interaction, and has been shown to inhibit various T-cell functions, including the generation of cytotoxic and suppressor T lymphocytes in cell culture. Also, based on studies with anti-Tac and other antibodies, a variety of disorders are now associated with improper IL-2 receptor expression by T-cells, in particular adult T-cell leukemia.

More recently, the IL-2 receptor has been shown to be an ideal target for novel therapeutic approaches to T-cell mediated diseases. It has been proposed that IL-2 receptor specific antibodies, such as the anti-Tac monoclonal antibody, can be used either alone or as an immunoconjugate 10 (e.g., with Ricin A, isotopes and the like) to effectively remove cells bearing the IL-2 receptor. These agents can, for cxample, theoretically eliminate IL-2 receptor-expressing leukemic cells, certain B-cells, or activated T-cells involved in a disease state, yct allow the retention of mature normal 15 T-cells and their precursors to ensure the capability of mounting a normal T-cell immune response as needed. In general, most other T-cell specific agents can destroy essentially all peripheral T-cells, which limits the agents' therapeutic efficacy. Overall, the use of appropriate monoclonal 20 antibodics specific for the IL-2 receptor may have therapeutic utility in autoimmune diseases, organ transplantation and any unwanted response by activated T-cells. Indeed, clinical trials have been initiated using, e.g., anti-Tac antibodies (see, generally, Waldmann, T., et al., Cancer Res. 45, 625 (1985), 25 Waldmann, T., Science 232, 727-732 (1986) and Kirkman ct al., Transplant. Proc. 21, 1766 (1989), all of which are incorporated herein by reference).

Unfortunately, the use of the anti-Tac and other nonhuman monoclonal antibodies have certain drawbacks, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans. 35

Perhaps more importantly, anti-Tac and other non-human monoclonal antibodies contain substantial stretches of amino acid sequences that will be immunogenic when injected into a human patient. Numerous studies have shown that, after injection of a foreign antibody, the immune response elicited by a patient against an antibody can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, as increasing numbers of different mouse or other antigenic (to humans) monoclonal antibodies can be expected to be developed to treat various diseases, after the first or several treatments with any different non-human antibodies, subsequent treatments even for unrelated therapies can be ineffective or even dangerous in themselves, because of crossreactivity.

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of human immunoglobulins reactive with the human IL-2 receptor, as with many human antigens, has been extremely difficult using typical human monoclonal antibody production techniques. Similarly, utilizing recombinant DNA technology to produce so-called "reshaped" or "humanized" antibodies (see, e.g., Riechmann et al., *Nature* 332, 323 (1988) and EPO Publication No. 0239460), provides uncertain results, in part due to unpredictable binding affinities.

Thus, there is a need for improved forms of human-like 65 immunoglobulins specific for the human IL-2 receptor that are substantially non-immunogenic in humans, yet easily

and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of T-cell mediated human disorders, the compositions containing human-like immunoglobulins specifically capable of blocking the binding of human IL-2 to its receptor and/or capable of binding to the p55 Tac protein on human IL-2 receptors. The immunoglobulins can have two pairs of light chain/heavy chain complexes, typically at least one chain comprising mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be used to produce human-like antibodies capable of binding to the human IL-2 receptor at affinity levels stronger than about 10⁸ M⁻¹.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for human-like immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The human-like immunoglobulins may be utilized alone in substantially pure form, or complexed with a cytotoxic agent, such as a radionuclide, a ribosomal inhibiting protein or a cytotoxic agent active at cell surfaces. All of these compounds will be particularly useful in treating T-cell mediated disorders. The human-like immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, human-like immunoglobulins specifically reactive with the IL-2 receptor on human T-cclls arc provided. These immunoglobulins, which have binding affinities of at least about 10^8 M⁻¹, and preferably 10^9 M⁻¹ to 10^{10} M⁻¹ or stronger, are capable of, e.g., blocking the binding of IL-2 to human IL-2 receptors. The human-like immunoglobulins will have a human-like framework and can have complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an epitope on p55 Tac protein. The immunoglobulis of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of T-cell mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope on the human IL-2 receptor, such as the anti-Tac monoclonal antibody. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human-like framework regions. Preferred DNA sequences, which on expression code for the polypeptide chains comprising the anti-Tac heavy and light chain hypervariable regions (with human-like framework regions), are included in FIGS. 15A and 16A, respectively. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. The antibodies of the present invention will typically find use individually in treating a T-cell mediated disease state. Generally, where the cell linked to a disease has been identified as IL-2 receptor bearing, then the human-like antibodies capable of blocking the binding of IL-2 to the 5 human IL-2 receptor are suitable.

For example, typical disease states suitable for treatment include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The human-like antibodies of the present invention may also be used in combination with other antibodies, particularly human monoclonal antibodies reactive with other markers on cells responsible for the disease. For example, suitable T-cell markers can include those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, *Leukocyte Typing*, Bemard, et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference.

The human-like antibodies and pharmaceutical compositions thereof of this invention are particularly useful for parenteral administration, i.e., subcutaneously, intramuscu-25 larly or intravenously. The compositions for parenteral administration will commonly comprise a solution of the antibody or a cocktail thereof dissolved in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers can be used, e.g., water, buffered water, 0.4% saline, 30 0.3% glycine and the like. These solutions are sterile and generally free of particulate matter. These compositions may be sterilized by conventional, well known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents, toxicity adjusting agents and the like, for example sodium acetate, sodium chloride, potassium chloride, calcium chloride, sodium lactate, human albumin, etc. The concentration of antibody in these formulations can vary widely, i.e., from less than about 0.5%, usually at or at least about 1% to as much as 15 or 20% by weight and will be selected primarily based on fluid volumes, viscosities, etc., in accordance with the particular mode of administration selected. 45

Thus, a typical pharmaccutical composition for injection could be made up to contain 1 ml sterile buffered water, and 1 to 50 mg of antibody. A typical composition for intravenous infusion could be made up to contain 250 ml of sterile Ringer's solution, and 150 mg of antibody. Actual methods for preparing parenterally administrable compositions will be known or apparent to those skilled in the art and are described in more detail in, for example, *Remington's Pharmaceutical Science*, 15th ed., Mack Publishing Company, Easton, Pa. (1980), which is incorporated herein by refer-55 ence.

The antibodies of this invention can be frozen or lyophilized for storage and reconstituted in a suitable carrier prior to use. This technique has been shown to be effective with conventional immune globulins and art-known lyoof philization and reconstitution techniques can be employed. It will be appreciated by those skilled in the art that lyophilization and reconstitution can lead to varying degrees of antibody activity loss (e.g., with conventional immune globulins, IgM antibodies tend to have greater activity loss 65 than IgG antibodies) and that use levels may have to be adjusted to compensate.

The compositions containing the present human-like antibodies or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from a disease, in an amount sufficient to cure or at least partially arrest the disease and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the infection and the general state of the patient's own immune system, but generally range from about 1 to about 200 mg of antibody per dose, with dosages of from 5 to 25 mg being more commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease states, that is life-threatening or potentially lifethreatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability of "foreign substance" rejections which are achieved by the present human-like antibodies of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these antibodies.

In prophylactic applications, compositions containing the present antibodies or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 0.1 to 25 mg per dose, especially 0.5 to 2.5 mg per dose. A preferred prophylactic use is for the prevention of kidney transplant rejection.

Single or multiple administrations of the compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the antibody(ies) of this invention sufficient to effectively treat the patient.

Human-like antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for T-cell typing, for isolating specific IL-2 receptor bearing cells or fragments of the receptor, for vaccine preparation, or the like.

Kits can also be supplied for use with the subject antibodies in the protection against or detection of a cellular activity or for the presence of a selected antigen. Thus, the subject antibody composition of the present invention may be provided, usually in a lyophilized form in a container, either alone or in conjunction with additional antibodies specific for the desired cell type. The antibodies, which may be conjugated to a label or toxin, or unconjugated, are included in the kits with buffers, such as Tris, phosphate, carbonate, etc., stabilizers, biocides, inert proteins, e.g., serum albumin, or the like, and a set of instructions for use. Generally, these materials will be present in less than about 5% wt. based on the amount of active antibody, and usually present in total amount of at least about 0.001% wt. based again on the antibody concentration. Frequently, it will be desirable to include an inert extender or excipient to dilute the active ingredients, where the excipient may be present in from about 1 to 99% wt. of the total composition. Where a second antibody capable of binding to the chimeric antibody is employed in an assay, this will usually be present in a separate vial. The second antibody is typically conjugated to a label and formulated in an analogous manner with the antibody formulations described above.

p75 Chain of IL-2 Receptor

The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide or alpha chain, being about 55 kD in size (see, Leonard, W., et al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). The second chain is known as the p75 or beta chain (Tsudo et al., Proc. Nat. Acad. Sci. USA, 83, 9694 (1986) and Sharon et al., Science 234, 859 (1986), both of which are incorporated herein by reference). The p55 or Tac chain and the p75 chain each independently bind IL-2 with low or intermediate affinity, while the IL-2 receptor complex of both chains binds IL-2 with high affinity. The p75 chain of the human IL-2 receptor will of ten be called herein simply the p75 protein.

The present invention provides novel compositions useful, for example, in the reatment of T-cell mediated human disorders, the compositions containing human-like immunoglobulins specifically capable of inhibiting the binding of 15 human IL-2 to its receptor and/or capable of binding to the p75 protein of human IL-2 receptors. The immunoglobulins can have two pairs of light chain/heavy chain complexes, typically at least one chain comprising mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse comple- 20 mentarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be used to produce human-like antibodies capable of binding to the p75 protein at affinity levels stronger than about 10^7 M^{-1} . These humanized immunoglobulins will also be capable of 25 blocking the binding of the CDR-donating mouse monoclonal antibody to p75.

The human-like immunoglobulins may be utilized alone in substantially pure form, or complexed with a cytotoxic agent, such as a radionuclide, a ribosomal inhibiting protein or a cytotoxic agent active at cell surfaces. All of these compounds will be particularly useful in treating T-cell mediated disorders. The human-like immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, human-like immunoglobulins specifically reactive with the p75 chain of the human 1L-2 receptor are provided. These immunoglo-40 bulins, which have binding affinities of at least 107 to 108 M⁻¹, and preferably 10⁹ M⁻¹ to 10¹⁰ M⁻¹ or stronger, are capable of, e.g., blocking the binding of IL-2 to human IL-2 receptors. The human-like immunoglobulins will have a human-like framework and can have complementarity deter-45 mining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an epitopc on p75 protein. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of T-cell 50 mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is diracted to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope on the human IL-2 receptor, such as the mik-β1 monoclonal antibody. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human-like framework regions. Exemplaty DNA sequences, which on expression code for the polypeptide chains comprising the mik-β1 heavy and light chain CDRs, arc included in FIG. 23A and FIG. 23B. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodics of the present invention will typically find use individually in treating a T-cell mediated disease state. Generally, where the cell linked to a disease has been identified as IL-2 receptor bearing, then the human-like antibodies capable of blocking the binding of IL-2 to the human IL-2 receptor are suitable.

For example, typical disease states suitable for treatment include graft-versus-host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The human-like antibodies of the present invention may also be used in combination with other antibodies, particularly human monoclonal antibodies reactive with other markers on cells responsible for the disease. For example, suitable T-cell markers can include those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, *Leukocyte Typing*, Bernard, et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference. A preferred use is the simultaneous treatment of a patient with a human-like antibody binding to p55 and a human-like anti-Tac plus humanized mik- β 1.

Human-like antibodi'es of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for T-cell typing, for isolating specific IL-2 receptor bearing cells or fragments of the receptor, for vaccine preparation, or the like.

Anti-HSV Antibodies

Herpes Simplex Virus types I and II (HSV-1 and HSV-2), are now estimated to be the second most frequent cause of sexually transmitted diseases in the world. Although completely accurate data are not available, infection estimates range from about 20 to 40% of the U.S. population.

A large number of diseases, from asymptomatic to lifethreatening, are associated with HSV infection. Of particular clinical interest, encephalitis from HSV-1 infection and transmission of HSV-2 from a pregnant mother to her fetus are often fatal. Immunosuppressed patients are also subject to severe complications when infected with the virus.

More than 50 HSV polypeptides have been identified in HSV-infected cells, including at least seven major cell surface glycoproteins (see, Whitley, R., Chapt. 66, and Roizman and Sears, Chapt. 65, Virology, Eds. Fields et al., 2nd ed., Raven Press, N.Y., N.Y. (1990), which are incorporated herein by reference). The specific biologic functions of these glycoproteins are not well defined, although gB and gD have en shown to be associated with cell fusion activity (W. Cai et al., J. Virol. 62, 2596 (1988) and Fuller and Spear, Proc. Natl. Acad. Sci. USA 84, 5454 (1987)). gB and gD express both type-specific and type-common antigenic determinants. Oakes and Lausch demonstrated that monoclonal antibodies against gB and gE suppress replication of HSV-1 in trigeminal ganglia (Oakes and Lausch, J. Virol. 51, 656 (1984)). Dix et al. showed that anti-gC and gD antibodies protect mice against acute virus-induced neurological disease (Dix ct al., Infect. Immun. 34, 192 (1981)). Whitley and colleagues produced a panel of murine monoclonal antibodies against HSV-1 and showed that several of the antibodies protected mice against encephalitis and death following ocular inoculation with the virus (see, Koga et al., Virology 151, 385 (1986); Metcalf et al., Cur. Eye Res. 6, 173 (1987) and Metcalf et al., Intervirology 29, 39 1988), all

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of which are incorporated herein by reference). Clone Fd79 (anti-gB) prevented encephalitis even when immunization was delayed until 48 hours post-infection. Fd79 and Fd138-80 (anti-gD) significantly reduced the severity of epithelial keratitis and lowered the frequency of persistent viral infec-5 tion in an outbrcd mouse model.

Thus, there is a need for improved forms of humanized immunoglobulins specific for HSV antigens that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic ¹⁰ formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of HSV mediated human 15 disorders, the compositions containing humanized immunoglobulins specifically capable of blocking the binding of HSV to its receptors and/or capable of binding to the HSV specific proteins. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain 20 comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human 25 framework regions to produce humanized immunoglobulins capable of binding to the HSV surface proteins at affinity levels stronger than about 107 M-1. These humanized immunoglobulins will also be capable of blocking the binding of the CDR donating mouse monoclonal antibody to HSV. 30

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with an antiviral agent, such as acyclovir or a cytotoxic agent active at viral surfaces. All of these compounds will be particularly useful in treating HSV mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with HSV related 40 epitopes either directly on the virus or on infected cells are provided. These immunoglobulins, which have binding affinities to HSV specific antigens of at least about 107 M and preferably 108 M⁻¹ to 1010 M⁻¹ or stronger, are capable of, e.g., protecting cells from HSV transmission. The 45 humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an HSV protein, such as gB and gD proteins. The immunoglobulins 50 of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of HSV mediated disorders in human patients by a variety of techniques.

The HSVs are among the most intensively investigated of 55 all viruses, and the HSV virion structure has been shown to contain about 33 proteins. Humanized immunoglobulins utilizing CDR's from monoclonal antibodies reactive with these proteins, particularly the eight surface glycoproteins (e.g., gB, gC, gD, gE, gG, gH and gl), represent preferred 60 embodiments of the present invention (see, Spear, P. G., The Herpesviruses, vol. 3, pp. 315–356 (1984) (Roizman, B., ed), Plenum Press, N.Y., N.Y. and Spear, P. G., Immunochemistry of Viruses. The Basis for Serodiagnosis and Vaccines, pp. 425-446 (1985) (Neurath, A. R., eds.), 65 Amsterdam: Elsevier, both of which are incorporated herein by reference).

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of an HSV protein, such as monoclonal antibodies reactive with HSV gB and gD glycoproteins. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate humanized framework regions. Exemplary DNA sequences code for the polypeptide chains comprising the heavy and light chain hypervariable regions (with human framework regions) from monoclonal antibodies Fd79 and Fd138-80, shown in FIG. 30A through FIG. 30D. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating an HSV mediated disease state. For example, typical disease states suitable for treatment include any involving HSV infection. Specific diseases include neonatal herpes, herpes encephalitis, ocular herpes, genital herpes and disseminated herpes (see, Corey, L., Chapter 136, Harrison's Principles of Internal Medicine, 11th ed., McGraw-Hill Book Company, N.Y., N.Y. (1987), which is incorporated herein by reference).

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different HSV antigens. For example, suitable HSV antigens to which a cocktail of humanized immunoglobulins may react include gC, gE, gF, gG and gH (see, Rector, J. et al., Infect. Immun. 38, 168 (1982) and Fuller, A. et al., J. Virol. 63, 3435 (1989), both of which are incorporated herein by reference).

The antibodies can also be used as separately administered compositions given in conjunction with acyclovir or other antiviral agents. Typically, the agents may include idoxunidine or trifluorothymidine, but numerous additional agents (e.g., vidarabine) well-known to those skilled in the art for HSV treatment may also be utilized (sec, Corey, L., op. cit.).

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill cells infected by HSV Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing an HSV epitope.

The compositions containing the present humanized antibodies or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from HSV infection, in an amount sufficient to cure or at least partially arrest the disease and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the infection and the general state of the patient's own immune system, but generally range from about 1 to about 200 mg of antibody per dose, with dosages of from 5 to 25 mg being more commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease states, that is life-threatening or potentially lifethreatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability

of "foreign substance" rejections which are achieved by the present humanized immunoglobulins of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these antibodies.

In prophylactic applications, compositions containing the ⁵ present immunoglobulins or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and ¹⁰ general level of immunity, but generally range from 0.1 to 25 mg per dose. A preferred prophylactic use is for the prevention of herpes in immunocompromised patients, such as organ transplant recipients.

Single or multiple administrations of the compositions ¹⁵ can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the antibody(ics) of this invention sufficient to effectively treat the patient.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of HSV antigens, for isolating specific HSV infected cells or fragments of the virus, for vaccine preparation, or the like. 25

Anti-CD33 Antibodies

There are about 10,000-15,000 new cases of myeloid (also called non-lymphocytic or granulocytic) leukemia in the U.S. per ycar (Cancer Facts & Figures, American Cancer ³⁰ Society, 1987). There are two major forms of myeloid leukemia: acute myelogcnous leukemia (AML) and chronic myelogcnous leukemia (CML). Despite treatment with chemotherapy, long-term survival in patients with AML is less than 10-20% (Clarkson et al., CRC Critical Review in ³⁵ Oncology/Hematology 4, 221 (1986)), and survival with CML and related diseases such as chronic myelomonocytic leukemia (CMML), chronic monocytic leukemi'a (CMMOL) and myelodysplastic syndrome (MDS) is even lower.

The p67 protein or CD33 antigen is found on the surface ⁴⁰ of progenitors of mycloid cells and of the leukemic cells of most cases of AML, but not on lymphoid cells or non-hematopoietic cells (see, Leucocyte Typing III, ed. by A. J. McMichael, Oxford University Press, pp. 622–629 (1987), which is incorporated herein by reference). Antibodies that are known to bind to the CD33 antigen include L4B3, L1B2 and MY9 (Andrews et al., Blood 62, 124 (1983) and Griffin et al., Leukemia Research 8, 521 (1984), both of which are incorporated herein by reference).

Another antibody that binds to CD33 is M195 (Tanimoto ct al., Leukemia 3, 339 (1989) and Scheinberg et al., Leukemia 3, 440 (1989), both of which are incorporated herein by reference). The reactivity of M195 with a wide variety of cells and tissues was tested. Among normal cells, M195 was reported to bind only to some monocytes and myeloid progenitor cells. The research also reported that it does not bind to other hematopoietic cells or to nonhematopoietic tissues. M195 bound to cclls of most cases of AML and all cases of CML in myeloblastic phase.

A phase I clinical trial of M195 in AML has been conducted (Scheinberg et al., Proc. ASCO 9, 207 (1990)). M195 radiolabeled with iodine-131 was found to rapidly and specifically target leukemic cells in both the blood and bone marrow. 65

Unfortunately, the use of non-human monoclonal antibodies such as M195 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

Thus, there is a need for improved forms of humanized immunoglobulins specific for CD33 antigen that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of myeloid leukemiarelated human disorders, the compositions containing humanized immunoglobulins specifically capable of binding to CD33 antigen. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to the CD33 antigen at affinity levels stronger than about 107 M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to CD33.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleouides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such as cytosine arabinoside or daunorubicin active against leukemia cells, or complexed with a radionuclide such as iodine-131. All of these compounds will be particularly useful in treating leukemia and myeloid cellmediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with CD33 related epitopes are provided. These immunoglobulins, which have binding affinities to CD33 of at least about 107 M⁻¹, and preferably 10⁸ M⁻¹ to 10¹⁰ M⁻¹ or stronger, are capable of, e.g., destroying leukemia cells. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with CD33 antigen. In a preferred embodiment, one or more of the CDR's will come from the M195 antibody. Importantly, M195 docs not bind to the ultimate hematopoictic stem cells, so M195 used in therapy will minimally interact with and destroy those cells, which are critical for generating all blood cells. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of myeloid cell-mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of CD33 antigen, such as monoclonal antibodies M195, I.4B3, L1B2 or MY9. The DNA segments 5 encoding these regions will typically be joined to DNA segments encoding appropriate human framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody M195 are included in FIG. 10 34A and FIG. 34B. Due to codon degeneracy and noncritical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating hematologic malignancies. For ¹⁵ example, typical disease states suitable for treatment include AML, CML, CMML, CMMOL and MDS (see, generally, Hofibrand & Pettit, Essential Haematology, Blackwell Scientific Publications, Oxford (1980)). The antibodies may also be used for bone marrow ablation prior to bone marrow ²⁰ transplant.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different myeloid antigens. For example, suitable antigens to which a ²⁵ cocktail of humanized immunoglobulins may react include CD13, CD14, CD15, CD16 and CD34 (see, Leukocyte Typing III, op. cit., pp. 576-732).

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include cytosine arabinoside and daunorubicin, but numerous additional agents (e.g., 6-thioguanine) well-known to those skilled in the art for leukemia treatment may also be utilized (see, Hoffbrund & Pettit., op. cit.).

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill leukemia cells. Immunotoxins are characterized by two components and are particularly useful 40 for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a 45 CD33 epitope.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of CD33 antigens, for isolating specific myeloid cells, or the like.

It will be understood that although examples pertain to the M195 antibody, producing humanized antibodies with high binding affinity for the CD33 antigen is also contemplated using CDR's from L4B3, L1B2, MY9 or other monoclonal antibodies that bind to an epitope of CD33. 55

Anti-CMV Antibodies

Cytomegalovirus is a major pathogen of immunocompromised individuals, especially bone marrow transplant recipi-60 ents, organ transplant recipients, and AIDS patients (see, generally, Fields et al., Eds., Virology, 2nd ed., Raven Press, New York pp. 1981–2010 (1990), which is incorporated herein by reference). Approximately 15% of bone marrow transplant patients develop CMV pneumonia, with an 85% 65 mortality rate (Meyers, Rev. Inf. Dis. 11 (suppl. 7), S1691 (1989)). About 10% of AIDS patients develop severe CMV

disease; and congenitally acquired CMV, often with significant morbidity and mortality, affects 1% of newborns (Fields, op. cit.).

The drug gancielovir is effective against cettain forms of CMV infection, notably choriorctinitis and gastroenteritis, but is not very effective against CMV pneumonia, and it has serious toxicity. Use of pooled human imunoglobulin preparations has shown some beneficial effect for prophylaxis of CMV in bone marrow transplant patients (Meyers, op. cit.), and a combination of high-dose immune globulin and ganciclovir has been reported effective against CMV pneumonia (Emanuel et al., Trans. Proc. XIX (suppl. 7), 132 (1987)). However, the marginal effectiveness, variable potency and high cost of commercial human immune globulin remain serious problems. Hence, there is a great need for new drugs effective against CMV.

CMV is a member of the herpesvirus family of viruses, and as such, has a large double-stranded DNA core, a protein capsid, and an outer lipid envelope with viral glycoproteins on its surface. At least 8 proteins have been detected on the envelope of CMV (Britt et al., J. Virol. 62, 3309 (1988)) and others have been predicted to exist based on the DNA sequence of CMV (Chee et al., Nature 344, 774 (1990)). Murine monoclonal antibodies have been produced against two especially significant CMV glycoproteins: gB, also called p130/55 or gp55-116, and gH, also called p86 (Rasmussen et al., Virology 163, 308 (1988) and Britt et al., op. cit., both of which are incorporated herein by reference) and shown to neutralize infectivity of the virus. Three other neutralizing antibodies to gH are designated CMV5, CMV109 and CMV115. Human monoclonal antibodies to CMV have also been produced (Ehrlich et al., Hybridoma 6, 151 (1987)).

In animal models, murine monclonal antibodies have been shown effective in treating infections caused by various viruses, including members of the herpesvirus family (see, e.g., Metcalf et al., Intervirol. 29, 39 (1988)). Hence, such antibodies may be useful in treatment of CMV infections.

Unfortunately, the use of non-human monoclonal antibodies such as CMV5 and CMV115 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of human immunoglobulins reactive with CMV antigens, as with many antigens, is difficult using typical human monoclonal antibody production techniques. Moreover, the human antibodies produced may lack certain desirable properties, such as high binding affinity and the ability to neutralize all clinical CMV strains. Simularly, utilizing recombinant DNA technology to produce so-called "humanized" or "reshaped" antibodies (see, e.g., Riechmann et al., *Nature* 332, 323 (1988) and EPO Publication No. 0239400, which are incorporated herein by reference), provides uncertai'n results, in part due to unpredictable binding affinities.

Thus, there is a need for improved forms of humanized immunoglobulins specific for CMV antigen that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

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The present invention provides novel compositions useful, for example, in the treatment of CMV-mediated human disorders, the compositions containing humanized immunoglobulins specifically capable of blocking the binding of CMV to its receptors and/or capable of binding to CMV 5 antigens. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, 10 with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to CMV at affinity levels stronger than about 10⁷ M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse 15 monoclonal antibody to CMV.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, pref.²⁰ erably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can²⁵ be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such a acyclovir or ganciclovir active against CMV-infected cells, or complexed with a cytotoxic agent. All of these compounds will be particularly useful in treating CMV-mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with CMV and CMVinfected cells are provided. These immunoglobulins, which 40 have binding affinities to CMV specific antigens of at least about 10^7 M^{-1} , and preferably 10^8 M^{-1} to 10^{10} M^{-1} or stronger, are capable of, e.g., blocking CMV infection of cells. The humanized immunoglobulins will have a human framework and will have one or more complementarity 45 determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with a CMV antigen. In a preferred embodiment, one or more of the CDR's will come from the CMV5, or CMV109 or CMV115 antibodies. The immunoglobulins of the present 50 invention, which can be produced economically in large quantities, find use, for example, in the treatment of CMVmediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recom-55 binant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of a CMV antigen, such as monoclonal antibodies CMV5 or CMV115. The DNA segments encoding these regions will typically be joined to DNA segments 60 encoding appropriate human framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody CMV5 are included in FIG. 39A and FIG. 39B. Due to codon degeneracy and non-critical amino-65 acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immontalized B-cells (see, Kabat op. cit. and WP87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to CMV and produced in any convenient mammalian source, including, mice, rats, rabbits, or other vertebrate capable of producing antibodies by well known methods. Suitable source cells for the DNA sequences and host cells for immunoglobulin expression and secretion can be obtained from a number of sources, such as the American Type Culture Collection (*Catalogue of Cell Lines and Hybridomas*, Fifth edition (1985) Rockville, Md. U.S.A., which is incorporated herein by reference).

The antibodies of the present invention will typically find use individually in treating CMV-related disorders. For example, typical disease states suitable for treatment include CMV pneumonia, neonatal CMV infection, CMV mononucleosis and CMV-related chorioretinitis and gastroenteritis.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different CMV antigens. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include the gB and gH proteins.

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include acyclovir or ganciclovir, but numerous additional agents well-known to those skilled in the art for CMV treatment may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill CMV-infected cells, Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a CMV epitope. The two components are commonly chemically bonded together by any of a variety of well-known chemical procedures. For example, when the cytotoxic agent is a protein and the second component is an intact immunoglobulin, the linkage may be by way of heterobifunctional cross-linkers, e.g., SPDP, carbodiimide, glutaraldehyde, or the like. Production of various immunotoxins is well-known with the art, and can be found, for example in "Monoclonal Antibody-Toxin Conjugates: Aiming the Magic Bullet," Thorpe et al., Monoclonal Antibodies in Clinical Medicine, Academic Press, pp. 168-190 (1982), which is incorporated herein by reference. The components may also be linked genetically (see Chaudhary et al., Nature 339, 394 (1989)).

In prophylactic applications, compositions containing the present immunoglobulins or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 1 to 50 mg per dose. A preferred prophylactic use is for the prevention of CMV infection in immunocompromised patients, such as organ or bone marrow transplant recipients.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of CMV antigens, for isolating specific CMV-infected cells, or the like.

In particular, the same method may be used to produce a ⁵ humanized CMV109, CMV115 or other anti-CMV antibody as used to produce humanized CMV5 herein.

Anti-y-IFN Antibodies

In mammals, the immune response is mediated by several types of cells that interact specifically with foreign material, i.e., antigens. One of these cell types, B cells, is responsible for the production of antibodies. Another cell type, T cells, 15 include a wide variety of cellular subsets that destroy virally infected cells or control the in vivo function of both B cells and other hematopoictic cells, including T cells. A third cell type, macrophages, process and present antigens in conjunction with major histocompatibility complex (MHC) proteins 20 to T cells. Communication between these cell types is mediated in a complex manner by lymphokines, such as interleukins 1-6 and y-IFN (see, generally, Paul, W. E., ed., Fundamental Immunology, 2nd ed., Raven Press, New York (1989), which is incorporated herein by reference.)

One important lymphokine is γ -IFN, which is secreted by some T cells. In addition to its anti-viral activity, γ -JFN stimulates natual killer (NK) cells, activates macrophages, and stimulates the expression of MHC molecules on the surface of cells (Paul, op. cit., pp. 622-624). Hence γ -IFN generally serves to enhance many aspects of immune function, and is a logical candidate for a therapeutic drug in cases where such enhancement is desired, e.g., in treating cancer. Conversely, in disease states where the immune system is over-active, e.g., autoimmune diseases and organ transplant rejection. antagonists of γ -IFN may be used to treat the disease by neutralizing the stimulatory effects of γ -IFN.

One class of effective antagonists of y-IFN are monoclonal antibodies that bind to and neutralize it (see, e.g., Van der Meide et al., J. Gen. Virol, 67, 1059 (1986)). In in vitro 40 and in vivo mouse models of transplants, anti-y-IFN antibodies have been shown to delay or prevent rejection (Landolfo et al., Science 229, 176 (1985) and Rosenberg et al., J. fmmunol. 144, 4648 (1990), both of which are incorporated herein by reference). Treatment of mice prone 45 to develop a syndrome like systemic lupus erythematosus (SLE) with a monoclonal antibody to y-IFN significantly delayed onset of the disease (Jacob et al., J. Exp. Med. 166, 798 (1987)). Under some conditions, an anti-y-IFN antibody alleviated adjuvant arthriti's in rats (Jacob et al., J. Immuno). 50 142, 1500 (1989)), suggesting that anti-y-IFN may be effective against some cases of rheumatoid arthritis in human patients. Multiple sclerosis (MS) in patients is made worse by treatment with y-IFN (Panitch et al., Neurology 36 (suppl. I), 285 (1986)), so an anti-y-IFN antibody may 55 alleviate MS. Thus, an anti-y-IFN antibody may be effective in treating these and other autoimmune diseases.

For treatment of human patients, a murine monoclonal that binds to and neutralizes human γ -IFN (see, e.g., Yamamoto ct al., Microbiol. Immunol. 32, 339 (1988)) may be 60 used. Another murine monoclonal antibody designated AF2 that neutralizes human γ -IFN, and inhibits binding of γ -IFN to its cellular receptor, is disclosed hercin. Unfortunately, the use of non-human monoclonal antibodies such as AF2 have certain drawbacks in human treatment, particularly in 65 repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, have a relatively short circulating half-life in humans, and Jack other important immunoglobulin functional characteristics when used in humans.

The present invention provides novel compositions useful, for example, in the treatment of human autoimmune disorders, the compositions containing humanized immunoglobulins specifically capable of binding to y-IFN. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to γ -IFN at affinity levels stronger than about 10⁷ M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to y-IFN.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be unilized alone in substantially pure form, or together with a chemotherapeutic agent such as a non-steroidal anti-infiarmatory drug, a cortcosteroid, or an immunosuppressant. All of these compounds will be particularly useful in treating autoimmune disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with γ -IFN epitopes are provided. These immunoglobulins, which have binding affinities to γ -IFN of at least about 10⁷ M⁻¹, and preferably 10⁸ M⁻¹ to 10¹⁰ M⁻¹ or stronger, are capable of, e.g., neutralizing human γ -IFN. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with γ -IFN. In a preferred embodiment, one or more of the CDR's will come from the AF2 antibody. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of autoimmune disorders in human patients by a variety of techniques.

The antibodies of the present invention will typically find use individually in treating autoimmune conditions. For example, typical disease states suitable for treament include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with other lymphokines or lymphokine receptors. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include interleukins 1 through 10 and the p55 and p75 chains of the IL-2 receptor (see, Waldmann, Annu. Rev. Biochem. 58, 875 (1989) and Queen et al., Proc. Natl. Acad. 5 Sci. USA 86, 10029 (1989), both of which are incorporated herein by reference). Other antigens include those on cells responsible for the disease, e.g., the so-called "Clusters of Differentiation" (Leucocyte Typing III, ed. by A. J. McMichael, Oxford University Press (1987), which is incor-10 porated herein by reference).

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include non-steroidal anti-inflammatory agents (e.g., aspirin, ibuprofen), 15 steroids (e.g., prednisone) and immunosuppressants (e.g., cyclosporin A, cytoxan), but numerous additional agems well-known to those skilled in the art may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins²⁰ in inununotoxins, e.g., to kill γ -IFN -secreting cells. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second²⁵ component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a γ -IFN epitope.

Humanized antibodics of the present invention can further $_{30}$ find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of γ -IFN antigens, or the like.

The following examples are offered by way of illustration, not by limitation. 35

EXPERIMENTAL

EXAMPLE 1

Humanized anti-Tac antibody

Design of genes for humanized anti-Tac light and heavy chains

The sequence of the human antibody Eu (Sequences of Proteins of Immunological Interest, E. Kabat et al., U.S. Dept, of Health and Human Services, 1983) was used to provide the framework of the humanized antibody, because the amino acid sequence of the heavy chain variable region of anti-Tac is more homologous to the heavy chain of this antibody than to any other complete heavy chain variable region sequence in the National Biomedrical Foundation Protein Identification Resource.

To select the sequence of the humanized heavy chain, the ⁵⁵ anti-Tac heavy chain sequence (FIG. 1B., upper lines; see, commonly assigned U.S. Ser. No. 07/223.037 filed Sep. 28, 1988, and 07/181,862 filed Apr. 15, 1988, both of which are now abandoned and which are incorporated herein by reference) was aligned with the sequence of the Eu heavy chain ⁶⁰ (FIG. 1B., lower lines). At each position, the Eu amino acid was selected for the humanized sequence, unless that position fell in any one of four categories defined above, in which case the anti-Tac amino acid was selected:

 The position fell within a complementarity determining region (CDR), as defined by Kabat, et al., op. cit. (amino acids 31-35, 50-66, 99-106);

- (2) The Eu amino acid was rare for human heavy chains at that position, whereas the anti-Tac amino acid was common for human heavy chains at that position (amino acids 27, 93, 95, 98, 107–109, 111);
- (3) The position was immediately adjacent to a CDR in the amino acrd sequence of the anti-Tac heavy chain (amino acids 30 and 67); or
- (4) 3-dimensional modeling of the anti-Tac antibody suggested that the amino acid was physically close to the antigen bindring region (amino acids 48 and 68). Amino acid #27 is listed in category (2) because the acceptor Eu amino acid Gly is rare, and the donor anti-Tac amino acid Tyr is chemically similar to the amino acid Phc, which is common, but the substitution was actually made because #27 also fell in category (4). Although some amino acids fell in more than one of these categories, they are only listed in one. The amino acids in the humanized heavy and light chains are numbered according to the lower lines of FIG. 1A and FIG. 1B.

To select the sequence of the humanized light chain, the anti-Tac light chain sequence was aligned with the sequence of the Eu light chain (FIG. 1A, lower lines). The Eu amino acid was selected at each position for the humanized sequence, unless the position again fell into one of the categories (1)-(4):

- (1) CDR's (amino acids 24-34, 50-56, 89-97);
- (2) Anti-Tac amino acid more typical than Eu (amino acids 48 and 63);
- (3) Adjacent to CDR's (no amino acids; Eu and anti-Tac were already the same at all these positions); or
- (4) Possible 3-dimensional proximity to binding region (amino acid 60).
- The actual nucleotide sequence of the heavy and light chain genes were selected as follows:
- The nucleotide sequences code for the amino acid sequences chosen as described above;
- (2) 5' of these coding sequences, the nucleotide sequences code for a leader (signal) sequence, namely the leader of the light chain of the antibody MOPC 63 and the leader of the heavy chain of the antibody PCH 108A (Kabat et al., op. cit.). These leader sequences were chosen as typical of antibodies;
- (3) 3' of the coding sequences, the nucleotide sequences are the sequences that follow the mouse light chain J5 segment and the mouse heavy chain J2 segment, which are part of the anti-Tac sequences. These sequences are included because they contain splice donor signals; and
- (4) At each end of the sequence is an Xba I site to allow cutting at the Xba I sites and cloning into the Xba I site of a vector.

Construction of humanized light and heavy chain gencs

To synthesize the heavy chain, four oligonucleotides were synthesized using an Applied Biosystems 380B DNA synthesizer. Two of the oligonucleotides are part of each strand of the heavy chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing.

Together, the oligonuclootides cover the entire humanized heavy chain variable region with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

Each oligonucleotide was phosphorylated using ATP and T4 polynucleotide kinase by standard procedures (see, Maniatis, op. cit.). To anneal the phosphorylated oligonucleotides, they were suspended together in 40 ul of TA (33 mM

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Tris acetate, pH 7.9, 66 mM potassium acetate, 10 mM magnesium acetate) at a concentration of about 3.75 uM each, heated to 95° C. for 4 min. and cooled slowly to 4° C. To synthesize the complete gene from the oligonuclcotides by synthesizing the opposite strand of each oligonucleotide, the following components were added in a final volume of 100 ul:

10 ul	annealed oligonucleotides	
0.16 mM each	deoxyribonucleotide	
0.5mM	ATP	
0.5 mM	DTT	
100 ug/ml	BSA	
3.5 ug/ml	T4 g43 protein (DNA polymerase)	
25 ug/ml	T4 g44/62 protein (polymerasc	
	accessery protein)	
25 ug/ml	45 protein (polymerase accessory protein)	

The mixture was incubated at 37° C. for 30 min. Then 10 u of T4 DNA ligase was added and incubation at 37° C. 20 resumed for 30 min. The polymerase and ligase were inactivated by incubation of the reaction at 70° C. for 15 min. To digest the gene with Xba L to the reaction was added 50 ul of 2x TA containing BSA at 200 ug/ml and DTT at 1 mM, 43 ul of water, and 50 u of Xba I in 5 ul. The reaction 25 was incubated for 3 hr at 37° C., and run on a gel. The 431 bp Xba I fragment was purified from a gel and cloned into the Xba I site of the plasmid pUC19 by standard methods.

Four plasmid isolates were purified and sequenced using the dideoxy method. One of these had the correct sequence.

To synthesize the light chain, four oligonucleotides JFD1, 30 JFD2, JFD3, JFD4 were synthesized. Two of the oligonucleotides are part of each strand of the light chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing. Together, the oligonucleotides cover the entire humanized light chain variable region with 35 a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

The light chain gene was synthesized from these oligonucleotides in two parts. 0.5 ug each of JFD1 and JFD2 were 40 combined in 20 ul sequence buffer (40 mM Tris-HCl, pH 7.5, 20 mM magnesium chloride, 50 mM sodium chloride), heated at 70° C. for 3 min and allowed to cool slowly to 23° C. in order for the oligonucleotides to anneal. JFD3 and JFD4 were treated in the same way. Each reaction was made 45 10 mM in DTT and 0.5 mM in each deoxyribonucleotide and 6.5 u of sequenase (US Biochemicals) was added, in a final volume of 24 ul, and incubated for 1 hr at 37° C. to synthesize the opposite strands of the oligonucleotides. Xba I and Hind III were added to each reaction to digest the DNA (there is a Hind III site in the region where JFD2 and JFD3 50 overlap and therefore in each of the synthesized DNAs). The reactions were run on polyacrylamide gels, and the Xba I -Hind III fragments were purified and cloned into pUC18 by standard methods. Several plasmid isolates for each fragment were sequenced by the dideoxy method, and correct 55 ones chosen.

Construction of plasmids to express humanized light and heavy chains

The heavy chain Xba I fragment was isolated from the pUC19 plasmid in which it had been inserted and then 60 inserted into the Xba 1 site of the vector pVyl (see, commonly assigned U.S. Ser. No. 07/223,037 filed Sep. 28, 1988, now abandoned, which is incorporated herein by reference) in the correct orientation by standard methods, to produce the plasmid pHuGTAC1. This plasmid will express 65 high levels of a complete heavy chain when transfected into an appropriate host cell.

The two light chain Xba I - Hind III fragments were isolated from the pUCI8 plasmids in which they had been inserted. The vector plasmid pVkl (see, commonly assigned U.S. Ser. No. 07/223,037 filed Sep. 28, 1988, now abandoned, which is incorporated herein by reference) was cut with Xba I, dephosphorylated and ligated with the two fragments by standard methods. The desired reaction product has the circular form: vector - Xba I - fragment 1 - Hind III fragment 2 - Xba I - vector. Several plasmid isolates were analyzed by restriction mapping and sequencing, and one 10 with this form chosen. This plasmid, pHuLTAC, therefore contains the complete humanized light chain and will express high levels of the light chain when transfected into an appropriate host cell.

Synthesis and affinity of humanized antibody

The plasmids pHuGTACI and pHuLTAC were transfected into mouse Sp2/0 cells, and cells that integrated the plasmids were selected on the basis of resistance to mycophenolic acid and/or hygromycin B conferred by the gpt and hyg genes on the plasmids by standard methods. To verify that these cells secreted antibody that binds to the IL-2 receptor, supernatant from the cells was incubated with HUT-102 cells that are known to express the II-2 receptor. After washing, the cells were incubated with fluorcsceinconjugated goat anti-human antibody, washed, and analyzed for fluorescence on a FACSCAN cytofluorometer. The results (FIG. 7A), clearly show that the humanized antibody binds to these cells, but not to Jurkat T-cells that do not express the IL-2 receptor (FIG. 7D). As controls, the original mouse anti-Tac antibody was also used to stain these cells, giving similar results.

For the next experiments, cells producing the humanized antibody were injected into mice, and the resultant ascites collected. Humanized antibody was purified to substantial homogeneity from the ascites by passage through an affinity column of goat anti-human immunoglobulin antibody, prepared on an Afligel-10 support (Bio-Rad Laboratories, Inc., Richmond, Calif.) according to standard techniques. To determine the affinity of the humanized antibody relative to the original anti-Tac antibody, a competitive binding experiment was performed. About 5×10⁵ HUT-102 cells were incubated with known quantities (10-40 ng) of the anti-Tac antibody and the humanized anti-Tac antibody for 10 min at 4° C. Then 100 ng of biotinylated anti-Tac was added to the cells and incubated for 30 min at 4° C. This quantity of anti-Tac had previously been determined to be sufficient to saturate the binding sites on the cells, but not to be in large excess.

Then the cells were washed twice with 2 ml of phosphate buffered saline (PBS) containing 0.1% sodium azide. The cells were then incubated for 30 min at 4° C. with 250 ng of phycoerythrin-conjugated avidin, which bound to the biotinylated anti-Tac already bound to the cells. The cells were washed again as above, fixed in PBS containing 1% paraformaldehyde, and analyzed for fluorescence on a FAC-SCAN cytofluorometer.

Use of increasing amounts (10-40 ng) of the anti-Tac antibody as competitor in the first step decreased the amount of biotinylated anti-Tac that could bind to the cells in the second step, and therefore the amount of phycoerythrinconjugated avidin that bound in the last stcp, thus decreasing fluorescence (FIG. 8A). Equivalent amounts (20 ng) of anti-Tac, and humanized anti-Tac used as competitor decreased the fluorescence to approximately the same degree (FIG. 8B). This shows that these antibodies have approximately the same affinity, because if one had greater affinity, it would have more effectively competed with the biotinylated anti-Tac, thus decreasing fluorescence more.

EXAMPLE 2

A second humanized anti-Tac antibody

Higher level expression of the humanized anti-Tae antibody

Three new plasmid vectors were prepared for expression of the humanized antibodies. The plasmid pVgl (FIG. 9A) contains a human cytomegalovirus IE1 promoter and 10 enhancer (Boshart et al, Cell 41, 521 (1985), which is incorporated herein by reference), the human genomic Cyl segment including part of the preceding intron, and the hygromycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selec- 15 to bind to the IL-2 receptor was assessed by fluorescence tion. The plasmid pVk (FIG. 9B) is similar to pVgl but contains the human genomic Cx segment and the gpt gene. The plasmid pVgl-dhfr was constructed similarly to pVgI but contains a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1984), which 20 is incorporated herein by reference) in place of the hygromycin gene.

Xba I fragments containing the humanized anti-Tac light chain and heavy chain variable regions were excised respectively from the plasmids pHuLTAC and the pHuGTAC1 and 25 cloned into the Xba I sites of the plasmid vectors pVk and pVgl. To express the humanized anti-Tac antibody, the light chain encoding plasmid was introduced by electroporation into SP2/0 mouse myeloma cells followed by selection for gpt expression. Transfected cells expressing light chain were 30 then transfected with the plasmi'd encoding the heavy chain followed by selection for hygromycin B resistance. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody. Humanized antibody was purified from culture 35 supernatant of transfected cells by protein A sepharose chromatography.

Construction of the second humanized anti-Tac antibody To determine whether it was actually necessary to use the mouse anti-Tac amino acids in categories (2)-(4) in the 40 humanized anti-Tac antibody to retain binding affinity, a second humanized anti-Tac antibody was constructed. In the second antibody, only mouse anti-Tac amino acids in Category (1), i.e., in the CDR's themselves, were used, with all other amino acids coming from the human Eu framework. 45 For purposes of this discussion, the original humanized anti-Tac antibody will be called the "PDL humanized antibody," and the second humanized antibody will be called the "CDR-only humanized antibody." The amino acid sequences of the PDL and CDR-only humanized antibody 50 (variable regions) are compared in FIG. 10A and FIG. 10B.

The CDR-only humanized anti-Tac heavy and light chain variable (V) region gene segments were constructed in essentially the same manner as the light chain of the PDL humanized anti-Tac immunoglobulin, as described above. 55 Specifically, each V region gene segment was synthesized in two halves. For each half, two overlapping, opposite-strand oligonucleotides, approximately 110 to 130 bases in length (FIG. 11A and FIG. 11B), were annealed and extended with sequenase (U.S. Biochemicals). The resulting double strand 60 fragments were digested with either Xba I and Hind III (light chain) or Xba I and Sal I (heavy chain) and inserted into plasmid pUC19. Clones with the correct sequence were identified by DNA sequencing. Complete heavy and light chain genes were generated by inserting the V region halves 65 into the Xba I sites of pVg I and pVk respectively by three-fragment ligation.

The CDR-only humanized antibody was expressed in the same manner as the PDL humanized antibody, by transfecting first the light chain containing plasmid and then the heavy chain containing plasmid into SP2/0 cells. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody, which was purified by protein A sepharose chromatography. Antibody concentration was determined by ELISA using purified PDL humanized antibody as a standard. That the purified CDR-only humanized antibody is assembled into H2L2 telramers as expected was shown by analysis using reducing and non-reducing polyaciylamide gel electrophoresis.

The ability of the CDR-only humanized immunoglobulin staining. Approximately 3.4×10⁵ HUT-102 cells, which are known to highly express the IL-2 receptor on their surface, were incubated with 200 ng of either the PDL or CDR-only humanized antibody, washed, and then incubated with fluorescein-conjugated goat anti-human IgG antisera. Cell fluorescence was measured by flow cytometry with a FACScan (Becton Dickinson). As shown in FIG. 12, the PDL humanized antibody strongly stained the cells. However, staining by the CDR-only antibody was indistinguishable from staining by the negative control antibody humanized Fd79, which binds the gB glycoprotein of herpes simplex virus and not HUT-102 cells. Hence, by this assay, the CDR-only humanized antibody does not detectably bind the IL-2 receptor.

Binding of the PDL and CDR-only humanized anti-Tac antibodies to the IL-2 receptor were also compared in a competitive binding assay. Approximately 4×10⁵ HUT-102 cells were incubated with 1.5 ng of radioiodinated mouse anti-Tac antibody (7×10⁶ cpm/ug) and varying amounts of each humanized antibody (4 to 512 ng) in 200 ul total volume of binding buffer (RPMI 1040 medium, 10% fetal calf senim, 10 ug/ml murine IgG2a, 0.1% sodium azide). After incubation for 2 hours at 0° C. 800 ul of binding bull'er was added, cells were collected by centrifugation and radioactivity was measured. The relative binding by the two humanized antibodies and by mouse anti-Tac is shown in a plot of bound/free labelled antibody versus competitor concentration (FIG. 13). The PDL humanized anti-Tac antibody affinity for IL-2 receptor is essentially equal to that of the mouse anti-Tac antibody, because it competes about equally well. But competition by the CDR-only humanized anti-Tac ant body to IL-2 receptor was undetectable at the antibody concentrations used, indicating a binding affinity reduction of at least 100-fold as compared to the PDL humanized anti-Tac antibody. Because the sequences of the PDL and CDR humanized anti-Tac antibodies differ only at positions where mouse framework residues in categories (2)-(4) were used in the PDL molecule, we conclude that at least one of these mouse framework residues are essential for high affinity binding.

EXAMPLE 3

Construction of 5 other humanized antibodies

Cloning of heavy and light chain cDNAs

Five other humanized antibodies were designed and produced using the principles and categories disclosed herein. The antibodies arc Fd79 and Fd138-80 which respectively bind to the gB and gD glycoproteins of herpes simplex virus (Metcalf et al., Intervirology 29, 39 (1988)), MI95 (Tanimoto ct al., Leukemia 3, 339 (1989)) which binds to the CD33 antigen, mik-B1 (Tusdo et al., Proc. Natl. Acad. Sci.

USA 86, 982 (1989)) which binds to the p75 chain of the IL-2 receptor, and CMV5 which binds to the gH glycoprotein of cytomegalovirus.

cDNAs for the heavy chain and light chain variable domain genes of each antibody were cloned using anchored polymerase chain reactions (Loh et al., Science 243,219 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites 10 (Scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For each antibody, at least two heavy chain and two kappa clones were sequenced and found to have the same sequence. The deduced amino acid sequences of the mature light and heavy chain variable regions are shown in FIGS. 2A-6B, upper lines. Design of humanized antibodies

In order to retain high binding affinity of the humanized antibodies, the principles and categories described above 20 were utilized when designing the antibodies. Based on high sequence homology, human antibodies were selected to provide both the acceptor light and heavy chain human frameworks for the mouse antibodies, as follows: human Pom for Fd79, human Eu for Fd138-80, human Eu for 25 M195, human Lay for mik-\$1, and human Wol for CMV5.

The computer programs ABMOD and ENCAD (Levitt, J. Mol. Biol., 168, 595 (1983) and Zilber et al., Biochemistry 29, 10032 (1990), both of which are incorporated herein by reference) was used to construct a model of the variable region of each mouse antibody. The model was used to determine the amino acids in each framework that were close enough to the CDR's to potentially interact with them (category 4 above). For each antibody, the positions found to fall in the categories (1)-(5) defined above are given in Table 35 1, numbered as in FIGS. 2A-6B.

TABLE 1

Calegory	Light Chain	Heavy Chain	
	Fd79 Antibod	у	
1	24-38, 54-50, 93-100	31-35, 50-66, 99-111	
2 3	9, 45, 46, 83	82, 112	
	53	112	
4	53	97	
5	81		
	Fd138-80 Antib	ody	
1	24-34, 50-56, 89-97	31-35, 50-66, 99-110	
2	48, 63	93, 98, 111, 112,	
		113, 115	
3		30, 67, 98, 111	
4	36, 48, 87	27, 30, 37, 48, 67,	
		68, 98	
	M195 Antiboo	ly	
1	24-38, 54-60, 93-101	31-35, 50-66, 95-105	
2	10, 52, 67, 110	93, 95, 98, 106, 107	
		108, 110	
3	-	30, 67, 98, 106	
4	40. 52, 74	27, 30, 48, 68, 98	
	mik-β1 Δπτίδο	dy	
1	24-33, 49-55, 88-96	31-35, 50-65, 98-108	
2	13	84, 89, 90	
2 3 4	_	30, 49	
4	70	29, 30, 72, 73	
5	41	1	
	CMV5 Antibo	dy	
1	24-34, 50-56, 89-97	31-35, 50-66, 99-108	
2	_	69, 80	

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TABLE 1-continued

Category	Light Chain	Heavy Chain
3	49	30
4	49	24, 27, 28, 30, 97
5		5

In designing each humanized antibody, at each position the amino acid was selected to be the same as in the human acceptor sequence, unless the position fell in categories (1)-(4), in which case the amino acid from the mouse donor sequence was used, or in category (5), in which case an amino acid typical for human sequences at that position was used.

For the construction of genes for the humanized antibodies, nucleou'de sequences were selected that encode the protein sequences of the humanized heavy and light chains, including signal peptides typically from the mouse antibody chains, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included splice donor signals typical for immunoglobulin genes and an Xbal site at each end. Each gene was constructed from four overlapping synthetic oligonuclcotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal. The oligonucleotides were synthesized on an Applied Biosys-30 tems 380B DNA synthesizer. Each oligo was about 110-140 base long with a 15-20 base overlap. Double stranded DNA fragments were synthesized with Klenow or Taq polymerase or sequenase from each pair of oligonucleotides, digested with restriction enzymes, ligated to pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of pVg1 (heavy chains of Fd79 and Fd138-80) or pVg1-dhfr (heavy chains of M195, mik-B1, CMV5) or pVk (all light chains) expression vectors in the appropriate orientations to produce 40 the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.).

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Cloncs were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified

from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The

bound antibodies were eluted with 0.2M Glycine-HCl, pH 3.0 and neutralized with 1M Tris pH 8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

55 Properties of the humanized antibodies

The binding of the humanized antibodies to cell types expressing the corresponding antigens was tested: HSVinfected cells for Fd79 and Fd138-80, U937 cells for M195, YTJB cells for mik-β1 and CMV-infected cells for CMV5.

60 By fluorocytometry, the humanized antibodies bind approximately as well as the original mouse antibodies and the corresponding chimeric antibodies. Moreover, the humanized antibodies compete approximately as well as the corresponding mouse antibodies against the radiolabeled mouse antibodies for binding to the cells, so the humanized anti-

bodies have approximately the same binding affinity as the mouse antibodies, typically within about 2 fold or better, see, 25

TABLE 2

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Binding affinities of :	murine and humanized	anlibodies.	
Mouse K_a (M ⁻¹)	Humanized $K_a (M^{-1})$		
Fd79 (anti-gB)	1.1×10^{8}	5.3 × 10 ⁷	
Fd138-80 (anti-gD)	5.2×10^{7}	4.8×10^{7}	
			-

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other antibodies. In comparison to other monoclonal antibodics, the present humanized immunoglobulin can be more economically produced and contain 15 substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement. 20

EXAMPLE 4

Design of genes for anti-Tac human-like light and heavy chains

The sequence of the human antibody Eu (Sequences of Proteins of Immunological Interest, Kabat, E., et al., U.S. Dcpt. of Health and Human Services, 1983) was used to provide the framework of the humanized antibody, because the amino acid sequence of the heavy chain of anti-Tac is 30 more homologous to the heavy chain of this antibody than to any other heavy chain sequence in the National Biomedical Foundation Protein Identification Resource,

To select the sequence of the humanized heavy chain, the anti-Tac heavy chain sequence was aligned with the sequence of the Eu heavy chain (FIG. 15). At each position, the Eu amino acid was selected for the humanized sequence, unless that position fell in any one of the following categories, in which case the anti-Tac amino acid was selected.

- (1) The position fell within a complementarity determining region (CDR), as defined by Kabat, et al., op. cit. (amino acids 31-35, 50-66, 99-106);
- (2) The Eu amino acid was unusual for human heavy chains at that position, whereas the anti-Tac amino acid 45 was typical for human heavy chains at that position (amino acids 27, 93, 95, 98, 107-109, 111);
- (3) The position was immediately adjacent to a CDR in the amino acid sequence of the anti-Tac heavy chain (amino acids 30 and 67).
- (4) 3-dimensional modeling of the anti-Tac antibody suggested that the amino acid was physically close to the antigen binding region (amino acids 48 and 68). Some amino acids fell in more than one of these categories but are only listed in one.

To select the sequence of the humanized light chain, the anti-Tac light chain sequence was aligned with the sequence of the Eu light chain (FIG. 16). The Eu amino acid was selected at each position, unless the position again fcll into 60 one of the categories (1)-(4), (with light chain replacing heavy chain in the category definitions):

- (I) CDRs (amino acids 24-34, 50-56, 89-97).
- (2) Anti-Tac amino acid more typical than Eu (amino acids 48 and 63). 65
- (3) Adjacent to CDRs (no amino acids; Eu and anti-Tac were already the same at all these positions).

(4) Possible 3-dimensional proximity to binding region (amino acid 60).

The actual nucleotide sequence of the heavy (FIG. 17) and light chain (FIG. 18) genes were selected as follows:

- (1) the nucleotide sequences code for the amino acid sequences chosen as described above.
- (2) 5' of these coding sequences, the nucleou de sequences code for a leader (signal) sequence, namely the leader of the light chain of the antibody MOPC 63 and the leader of the heavy chain of the antibody PCH 108A (Kabat et al., op. cit.). These leader sequences were chosen as typical of antibodies.
- (3) 3' of the coding sequences, the nucleotide sequences are the sequences that follow the mouse light chain J5 segment and the mouse heavy chain J2 segment, which are part of the anti-Tac sequences. These sequences are included because they contain splice donor signals.
- (4) At each end of the sequence is an Xba I site to allow cutting at the Xba I sites and cloning into the Xba I site of a vector.

Construction of humanized light and heavy chain genes To synthesize the heavy chain, four oligonucleotides HES12, HES13, HES14, HES15 (FIG. 19A) were synthesized using an Applied Biosystems 380B DNA synthesizer. Two of the oligonucleotides are part of each strand of the heavy chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing (FIG. 19B). Together, the oligonucleotides cover the entire humanized heavy chain (FIG. 17) with a few extra nuclcotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

Each oligonucleotide was phosphorylated using ATP and T4 polynuclcotide kinase by standard procedures (see, Maniatis, op. cit.). To anneal the phosphorylated oligonucleotides, they were suspended together in 40 ul of TA (33 mM Tris acetate, pH 7.9, 66 mM potassium acetate, 10 mM magnesium acetate) at a concentration of about 3.75 uM each, heated to 95° C. for 4 min. and cooled slowly to 4° C. To synthesize the complete gene from the oligonucleotides by synthesizing the opposite strand of each oligonucleotide (FIG. 19B), the following components were added in a final volume of IOOul:

annealed oligonucleotides
deoxyrbonucleotide
ATP
DIT
BSA
T4 g43 protein (DNA polymerase)
T4 g44/62 protein (polymerase
accessory protein)
45 protein (polymerase accessory protein)

The mixture was incubated at 37° C. for 30 min. Then 10 U of T4 DNA ligase was added and incubation at 37° C. resumed for 30 min. The polymerase and ligasc were inactivated by incubation of the reaction at 70° C. for 15 min. To digest the gene with Xba I, to the reaction was added 50 ul of 2× TA containing BSA at 200 ug/ml and DTT at 1 mM, 43 ul of water, and 50 U of Xba I in 5 ul. The reaction was incubated for 3 hr at 37° C., and run on a gel. The 431 bp Xba I fragment was purified from a gel and cloned into the Xba I site of the plasmid pUCI9 by standard methods.

Four plasmid isolates were purified and sequenced using the dideoxy method. One of these bad the correct sequence (FIG. 17).

To synthesize the light chain, four oligonucleoi des JFDI, JFD2, JFD3, JFD4 (FIG. 20A) were synthesized. Two of the

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oligonucleotides are part of each strand of the light chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing (FIG. 20B). Together, the oligonucleotides cover the entire humanized light chain (FIG. 18) with a few extra nucleotides at each end to allow 5 cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

The light chain gene was synthesized from these oligonucleotides in two parts. 0.5 ug each of JFD1 and JFD2 were combined in 20 ul sequenase buller (40 mM Tris-HCl, pH 10 7.5, 20 mM magnesium chloride, 50 mM sodium chloride), heated at 70° C. for 3 min and allowed to cool slowly to 23° C. in order for the oligonucleotides to anneal. JFD3 and JFD4 were treated in the same way. Each reaction was made 10 mM in DTT and 0.5 mM in each deoxyribonucleotide and 15 6.5 U of sequenase (US Biochemicals) was added, in a final volume of 24 ul, and incubated for 1 hr at 37° C. to synthesize the opposite strands of the oligonucleotides. Xba I and Hind III were added to each reaction to digest the DNA (there is a Hind III site in the region where JFD2 and JFD3 20 overlap and therefore in each of the synthesized DNAs; FIG. 20B). The reactions were run on polyacrylamide gels, and the Xba I - Hind Ill fragments were purified and cloned into pUC18 by standard methods. Several plasmid isolates for each fragment were sequenced by the dideoxy method, and 25 correct ones chosen.

Construction of plasmids to express humanized light and heavy chains

The heavy chain Xba I fragment was isolated from the pUC19 plasmid in which it had been inserted and then 30 inserted into the Xba I site of the vector $pV\gamma l$ in the correct orientation by standard methods, to produce the plasmid pHuGTAC1 (FIG. 21). This plasmid will express high levels of a complete heavy chain when transfected into an appropriate host cell. 35

The two light chain Xba I - Hind III fragments were isolated from the pUC18 plasmids in which they had been inserted. The vector plasmid pVxl was cut with Xba I, dephosphorylated and ligated with the two fragments by standard methods. The desired reaction product has the 40 circular form: vector - Xba I - fragment 1 - Hind III -fragment 2 - Xba I - vector. Several plasmid isolates were analyzed by restriction mapping and sequencing, and one with this form chosen. This plasmid, pHuLTAC (FIG. 22), therefore contains the complete humanized light chain (FIG. 45 **18**) and will express high levels of the light chain when transfected into an appropriate host cell. Synthesis and affinity of humanized antibody

The plasmids pHuGTACl and pHuLTAC were transfected into mouse Sp2/0 cells, and cells that integrated the 50 plasmids were selected on the basis of resistance to mycophenolic acid and/or hygromycin B conferred by the gpt and hyg genes on the plasmids (FIGS. 21, 22) by standard methods. To verify that these cells secreted antibody that binds to the IL-2 receptor, supernatant from the cells was 55 incubated with HUT-102 cells that are known to express the IL-2 receptor. After washing, the cells were incubated with fluorescein-conjugated goat anti-human antibody, washed, and analyzed for fluorescence on a FACSCAN cytofluorometcr. The results (FIG. 7A), clearly show that the humanized 60 antibody binds to these cells, but not to Jurkat T-cells that do not express the IL-2 receptor (FIG. 7D). As controls, the original mouse anti-Tac antibody was also used to stain these cells (FIG. 7B and FIG. 7C), giving similar results.

For further experiments, cells producing the humanized 65 antibody were injected into mice, and the resultant ascites collected. Humanized antibody was purified to substantial

homogeneity from the ascites by passage through an affinity column of goat ant human immunoglobulin anibody, prepared on an Affigel-10 support (Bio-Rad Laboratories, Inc., Richmond, Calif.) according to standard techniques. To determine the affinity of the humanized antibody relative to the original ant-Tac antibody, a competitive binding experiment was performed. About 5x105 HUT-102 cells were incubated with known quantities (10-40 ng) of the anti-Tac antibody and the humanized anti-Tac antibody for 10 min at 4° C. Then 100 ng of biotinylated anti-Tac was added to the cells and incubated for 30 min at 4° C. This quantity of anti-Tac had previously been determined to be sufficient to saturate the binding sites on the cells, but not to be in large excess. Then the cells were washed twice with 2 ml of phosphate buffered saline (PBS) containing 0.1% sodium azide. The cells were then incubated for 30 min at 4° C. with 250 ng of phycocrythrin-conjugated avidin, which bound to the biotinylated anti-Tac already bound to the cells. The cells were washed again as above, fixed in PBS containing 1% paraformaldehyde, and analyzed for fluorescence on a FAC-SCAN cytofluorometer.

Use of increasing amounts (10-40 ng) of the anti-Tac antibody as competitor in the first step decreased the amount of biotinylated anti-Tac that could bind to the cells in the second step, and therefore the amount of phycoerythrinconjugated avidin that bound in the last step, thus decreasing fluorescence (FIG. &A). Equivalent amounts (20 ng) of anti-Tac, and humanized anti-Tac used as competitor decreased the fluorescence to approximately the same degree (FIG. &B). This shows that these antibodies have approximately the same affinity, because if one had greater affinity, it would have more effectively competed with the biotinylated anti-Tac, thus decreasing fluorescence more. Biological properties of the humanized antibody

For optimal use in treatment of human disease, the humanized antibody should be able to destroy T-cells in the body that express the IL-2 receptor. One mechanism by which antibodies may destroy target cells is antibody-dependent cell-mediated cytotoxicity, abbreviated ADCC (Fundamental Immunology, Paul, W., Ed., Raven Press, New York (1984), at pg. 681), in which the antibody forms a bridge between the target cell and an effector cell such as a macrophage that can lyse the target. To determine whether the humanized antibody and the original mouse anti-Tac antibody can mediate ADCC, a chromium release assay was performed by standard methods. Specifically, human leukemia HUT-102 cells, which express the IL-2 receptor, were incubated with ⁵¹Cr to allow them to absorb this radionuclide. The HUT-102 cells were then incubated with an excess of either anti-Tac or humanized anti-Tac antibody. The HUT-102 cells were next incubated for 4 hrs with either a 30:1 or 100:1 ratio of effector cells, which were normal purified human peripheral blood mononuclear cells that had been activated by incubation for about 20 hrs with human recombinant IL-2. Release of ⁵¹Cr, which indicated lysis of the target HUT-102 cells, was measured and the background subtracted (Table 3). The results show that at either ratio of effector cells, anti-Tac did not lyse a significant number of the target cells (less than 5%), while the humanized antibody did (more than 20%). Hence, the humanized antibody is likely to be more efficacious than the original mouse antibody in treating T-cell leukemia or other T-cell mediated diseases.

TABLE 3

Percent ⁵¹ Cr rel	lease after ADCC		
		ector: et ratio	-
	30:1	100:1	
Antibody			
Anti-Tac Humanized anti-Tac	4% 24%	<1% 23%	10

Higher level expression of the humanized anti-Tac antibody

Two new plasmid vectors were prepared for expression of the humanized antibody. The plasmid pVgl (FIG. 9A) contains a human cytomegalovirus IE1 promoter and enhancer (Boshart et al., *Cell* 41, 521 (1985)), the human genomic C γ 1 segment including part of the preceding inkron, and the hygromycin gene (Blochlinger et al., *Mol. Cell. Biol.* 20 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 9B) is similar to pVgl but contains the human genomic C κ segment and the gpt gene.

Xba I fragments containing the humanized anti-Tac light 25 chain and heavy chain variable regions were excised respeclively from the plasmids pHuLTAC and the pHuGTACl and cloned into the Xba I sites of the plasmid vectors pVk and pVGI. To express the humanized anti-Tac antibody, the light chain encoding plasmid was introduced by electroporation 30 into SP2/0 mouse myeloma cells followed by selection for gpt expression. Transfected cells expressing light chain were then transfected with the plasmid encoding the heavy chain followed by selection for hygromycin B resistance. Transfected cells producing the highest levels of humanized 35 antibody as determined by ELISA were used for preparation of antibody. Humanized antibody was purified from culture supernatant of transfected cells by protein A sepharose chromatography.

From the foregoing, it will be appreciated that the humanlike immunoglobulins of the present invention offer numerous advantages of other human IL-2 receptor-specific antibodies. In comparison to anti-Tae mouse monoclonal antibodies, the present human-like immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

EXAMPLE 5

Design of genes for mikßl humanized light and heavy chains

To exert its biological effects, IL-2 interacts with a specific high-affinity membrane receptor (Greene, W., et al., 55 *Progress in Hematology XIV*, E. Brown, Ed., Grune and Statton, New York (1986), at pgs. 283 ff and Waldmann, *Ann. Rev. Biochem.* 58, 875 (1989), which is incorporated herein by reference). The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac 60 peptide or alpha chain, being about 55kD in size (see, Leonard, W., et al., *J. Biol. Chem.* 260, 1872 (1985), which is incorporated herein by reference). The second chain is known as the p75 or beta chain (Tsudo et al., *Proc. Nat. Acad. Sci. USA*, 83, 9694 (1986) and Sharon et al., *Science* 65 234, 859 (1986), both of which are incorporated herein by reference). The p55 or Tac chain and the p75 chain each

independently bind IL-2 with low or intermediate affinity, while the IL-2 receptor complex of both chains binds IL-2 with high affinity. The p75 chain of the human IL-2 receptor will often be called herein simply the p75 protein.

5 Much of the elucidation of the human IL-2 receptor's structure and function is due to the development of specifically reactive monoclonal antibodies. The antibody, mik-β1, binds to the p75 chain (Tsudo et al., *Proc. Nat. Acad. Sci. USA* 86, 1982 (1989), which is incorporated herein by reference).

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., *Science* 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC19 vector for sequencing. For mik- β 1, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG. 23A and FIG. 23B. Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVgl-dhfr (FIG. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic C, 1 segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVgl-dhfr but contains the human genomic Cx segment and the gpt gene. Derivatives of the mik-ßl heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained Xbal sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and Xbal sites (see, C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the CMV promoter and the pattial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappachain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric mik- β l antibody was shown to bind to YTJB cells, which express the p75 antigen, by flow eytometry (FIG. 25). Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (C. Queen et al., *Proc. Natl. Acad. Sci. USA* 86, 10029 (1989), which is incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDRs with the human framework be to introduce distortions into the CDRs that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence database (performed with the MicrorGenic Sequence Analysis Software (Beckman)), the

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antibody Lay was chosen to provide the framework sequences for humanization of mik- β 1.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the mik- β l variable region. ⁵ The model was used to determine the amino acids in the mik- β l framework that were close enough to the CDRs to potentially interact with them (category 4 below). To design the humanized light and heavy chain mik- β l variable regions, at each position the amino acid was chosen to be the same as in the Lay antibody, unless that position fell in one or more of five categories:

- (1) The position fell within a CDR,
- (2) The Lay amino acid was unusual for human antibodies at that position, whereas the mik-βI amino acid was ¹⁵ typical for human antibodies at that position.
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDRs).

For positions in these categories, the amino acid from the (mouse) mik- β 1 antibody was used. In addition, a position was in the fifth category if

(5) The Lay amino acid was highly unusual for human 25 antibodies at that position, and the mik-βl amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 4. $_{30}$ Some amino acids may be in more than one category. The final sequences of the humanized mik- β l light and heavy chain variable domains are shown in FIG. **26**A and FIG. **26**B, compared with the Lay sequences.

TABLE 4

Category	Light Chain	Heavy Chain
1	24-33, 49-55, 88-96	31-35, 50-65, 98- 108
2	13	84, 89, 90
3	30, 49	
4	70	29, 30, 72, 73
5	41	1

For the construction of genes for the humanized antibod- 45 ies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse mik-Bl chains (FIG. 23A and FIG. 23B), generally utilizing codons found in the mouse sequence. Several degenerate codons 50 were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an Xbal site at each end. Each gene was consulted from four overlapping synthetic oligonuclcotides. For each variable 55 domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 27A and FIG. 27B). The oligonucleotides were synthesized on an Applied Biosystems 60 380B DNA synthesizer. Each oligo was about 110-140 base long with about a 20 base overlap. Double stranded DNA fragments were synthesized with sequenase from each pair of oligonucleotides, digested with restriction enzymes, ligated to pBluescriptll KS (+) (Stratagene) vector and 65 sequenced. Two fragments with the respectively correct half-sequences were then ligated into the Xbal sites of the

pVgl-dhfr or pVk expression vectors. In vitro mutagenesis was used to change an Ala amino acid originally encoded by oligonucleotide wps54 to the Glu (E) at position 1 of the humanized heavy chain (FIG. 26B) by changing the nucleotides CT to AG. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by EHSA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody was eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized mik-βl antibody was characterized in comparison to the murine and chimeric antibodies. The humanized antibody bound to YTJB cells, which express p75 chain at a high level, in a fluorocytometric analysis in a manner similar to the chimeric antibody (FIG. 25), showing that it recognizes the same p75 protein.

The affinity of the humanized antibody was determined by competition with the radio-iodinated mouse mik- β 1 antibody (FIG. 28). The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in *Fundamental Immunology* (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference). The binding affinity of the humanized mik- β 1 antibody was within about 2-fold of the affinity of the mouse mik- β 1 antibody.

The ability of humanized mik-ßl plus humanized anti-Tac antibody to inhibit IL-2 stimulated proliferation of human lymphocytes was determined. Human mononuclear cells, collected from human blood by centrifugation on Ficoll-Paque (Pharmacia), were diluted to 2×10⁶ cells/ml in RPMI mcdium+10% fetal calf serum (FCS). A 1/200 volume of phytohemagglutinin P (Difeo) was added and the cells were incubated for 4 days. The cells were incubated an additional 4 days in RPMI+10% FCS+10 u/ml IL-2. 10° of these PHA activated blasts were then incubated with or without 2 µg each of humanized mik-Bl and humanized anti-Tac in 150 µl of RPMI+ 10% FCS in wells of a 96-well plate for 1 hr, to which various dilutions of IL-2 (Amgen) were then added in 50 µl medium. The cells were incubated 48 hr, 0.5 µCi methyl-3H-thymidine (Amersham, 82 Ci/mmol) was added, and the cells were incubated 24 hr. Cells were harvested with a cell harvester and radioactivity determined. The combination of the antibodies greatly inhibited proliferation of the cells in response to IL-2 (FIG. 29), suggesting a combination of the antibodies will have strong immunosuppressive properties. Humanized mik-Bl plus humanized anti-Tac inhibited proliferation much more strongly than did either antibody alone.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other p75 specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

EXAMPLE 6

Design of genes for Fd79 and Fd138-80 humanized light and heavy chains

Exemplary DNA sequences coding for the polypeptide chains comprising the heavy and light chain hypervariable regions (with human framework regions) from monoclonal antibodies Fd7Q and Fd138-80, are shown in FIG. 30A through FIG. 30D.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain regions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and con-15 tained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in FIG. 14). This method yields clones with authentic variable domain sequences, in contrast to other methods using mixed primers designed to anneal to the variable domain sequence (J. W. 20 Lanick et al., Bio/Technology 7, 934 (1989) and Y. L. Chiang et al., BioTech. 7, 360 (1989)). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For Fd79, two gamma-1 specific and 5 kappa specific clones were 25 sequenced. The two gamma-1 specific clones are identical in sequence. This heavy chain cDNA fragment encodes a signal peptide of 19 amino acids, a V region in mouse heavy chain subgroup IIIB, a D segment, and a J_{H} segment with 4 alterations compared to the genomic J_{H} sequence. The 30 deduced amino acid sequence is shown in FIG. 30A.

The five kappa specific clones belong to two groups. Two clones are identical and encode a kappa chain in which the conserved amino acid 23 cysteinc has been substituted by a tyrosine, probably representing the non-productive allele. 35 The other three clones have an identical sequence encoding a signal peptide sequence of 20 amino acids, a V region in mouse kappa chain subgroup III, and a $J_k 2$ segment with a single alteration compared to the genomic $J_k 2$ sequence (FIG. 30B). The validity of the heavy chain and the kappa 40 chain sequences was subsequently confirmed by the construction and expression of a chimeric antibody as discussed below.

The heavy chain and the kappa chain of Fd138-80 were cloned similarly. Three clones each of the heavy chain and 45 the kappa chain were sequenced. All three heavy chain clones have an identical sequence encoding a signal peptide sequence of 19 amino acids, a V region in mouse heavy chain subgroup II, a D segment and the J_{Fr} 3 segment (FIG. **30C**). The three kappa clones are also identical in sequence of 20 amino acids, a V region gene in mouse kappa chain subgroup V and the J_{FS} 5 segment (FIG. **30D**). Both chains shown no irregularities in coding sequence; their validity was subsequently confirmed by construction and expression 55 of a chimeric antibody.

Construction and expression of chimeric antibodies.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1 (FIG. 9A) contains a human cytomegalovirus IE1 60 promoter and enhancer (M. Boshart et al., Cell 4.1, 521 (1985)), the human genomic C_{y1} segment including part of the preceding intron, and the hygromycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 65 9B) is similar to pVg1 but contains the human genomic C_{y2} segment and the gpt gene. Derivatives of the Fd79 and

Fd138-80 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see., C. Queen et al., Proc, Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibodies, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric Fd79 and Fd138-80 antibodies were shown to bind to HSV-1 infected vero cells by flow cytometry. Vural neutralization assays also indicated that the chimeric antibodies retain the neutralization activities of the murine antibodies (data not shown, but see below for similar results with humanized antibodies).

Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (C. Queen et al., Proc. Natl, Acad. Sci, USA 86, 10029 (1989), which is incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDRs with the human framework be to introduce distortions into the CDRs that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Porn was chosen to provide the framework sequences for humanization of Fd79.

The computer program ENCAD (Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the Fd79 variable region. Inspection of the refined model of murine Fd79 revealed two amino acid residues in the framework that are close enough to have significant contacts with the CDR residues (Table 5). Lys in light chain position 49 has contacts with 3 amino acids in CDR2 of the light chain (L50 Tyr, L53 Asn, L55 Glu) and 2 amino acids in CDR3 of the heavy chain (H99 Asp, H100 Tyr). Leu in heavy chain position 93 also shows interactions with 2 amino acids in CDR2 of the heavy chain (H35 Ser, H37 Val) and an amino acid in CDR3 of the heavy chain (H100C Phe), Hence, L49 Lys and H93 Leu were retained in the construction of humanized Fd79, as their replacement with human Pom framework residues would be likely to introduce distortions into the CDRs. Also, 7 other residues in the Pom framework (5 in the light chain and 2 in the heavy chain) were substituted with common human residues (identical to the murine Fd79 sequence in 6 of the choices) because of their rare occurrence in other human antibodies. The elimination of unusual amino acids in the framework may further reduce immunogenicity. The murine Fd79 sequences and the corresponding humanized sequences are shown in FIG. 30A and FIG. 30B. Substituted residues in the Pom framework are underlined.

15

TABLE 5

Residue No. ¹	Amino Acid	Contacting CDR residues ²	
Fd79			
1,49	Lys	L50Y, L53N, L55E, H99D, H100Y	
H93 FdI38-80	Leu	H35S, H37V, H100CF	
L36	His	L34V, L89Q	
H27	Тут	H2H, H34I	
H30	Тут	H2H, H53R	
H48	Phc	H63F	
H66	Lys	H63F	
H67	Ala	H63F	

1. The amino acid residues are numbered according to the Kabat system (E. A. Kabat et al., Sequences of Proteins of Immunological Interest, National Institutes of Health, Bethesda, MD (1987)): the first letter (H or L) stands for 20 the heavy chain or light chain. The following number is the residue number. The last letter is the amino acid one letter code. 2. The hypervariable regions are defined according to Kabat: Light chain

CDR1: tesidue 24-34; CDR2: 50-56; CDR3: 89-97. Heavy chain CDR1: 31-35; CDR2: 50-65; CDR3: 95-102.

Similarly, the murine heavy chain and light chain 25 sequences of Fd138-80 were subjected to sequence homology search against the NBRF protein sequence database. The sequences of the human antibody Eu were selected to provide the framework sequences for humanized Fd138-80. Inspection of a computer-generated model of Fd138-80 30 revealed 6 amino acid residues in the framework that are close enough to have important contacts with CDR residues. The residues and their contacting counterparts are listed in Table 5; these murine residues were retained in the construction of humanized Fdl 38-80. Two other residues (L87 35 Phe and H37 Met) show significant contacts with L98 Phe, which is immediately adjacent to CDR3, so these two mouse residues were also retained. Eight amino acids in the Eu framework (2 in the light chain and 6 in the heavy chain) were substituted with the murine residues (which are also 40 consistent with the human consensus residues) because of their rare occurrence in other human antibodies. The murine Fd138-80 sequences and the corresponding humanized sequences are shown in FIG. 30C and FIG. 30D. Substituted residues in the Eu framework are underlined.

For the construction of genes for the humanized antibod- 45 ies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the signal peptides, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove unde- 50 sirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleou'des. For each variable domain gene, two pairs of overlapping oligonucleotides on 55 alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal. The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with a 15 base overlap. 60 Double stranded DNA fragments were synthesized with Klenow polymerase, digested with restriction enzymes, ligated to pUC18 vector and sequenced. The two fragments with the correct sequences were then ligated into the Xbal sites of pVgl or pVk expression vectors.

The synthetic genes were then cloned into the pVgl and pVk expression vectors. For each humanized antibody con-

structed, the heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibodies were purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibodies were eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer was 10 exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized Fd79 and Fd1 38-80 antibodies were characterized in comparison to the murine and chimeric antibodies. Both humanized antibodies bind to Vero cells infected with HSV-1 in a fluorocytometric analysis in a manner similar to the chimeric antibodies (FIG. 31A and FIG. 31B), suggesting that they recognize their respective viral antigens. To more quantitatively assess the binding activity, radioiodinated murine antibodies were bound to virally infected cells and Scatchard analysis performed.

The affinitics of the humanized antibodies were determined by competition with the iodinated antibodies. Vero cells infected with HSV-1 were used as source of gB and gD antigens. Increasing amounts of competitor antibody (mouse or humanized) were added to 1.5 ng of radioiodinated tracer mouse antibody (2uCi/ug) and incubated with 4×10^5 infected Vero cells in 0.2 ml of binding buffer (PBS+2%) FCS+0.1% azide) for 1 hr. at 4° C. Cells were washed and pelleted, and their radioactivities were measured. The concentrations of bound and free tracer antibody were calculated. The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in Fundamental Immunology (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference).

The measurements indicate that there is no significant loss of binding affinities in the humanized antibodies (Table 6). Specifically, there is an approximately 2-fold decrease in affinity in humanized Fd79 compared to the murine Fd79 (Ka of $5.3 \times 10^7 \text{ M}^{-1}$ vs. $1.1 \times 10^8 \text{ M}^{-1}$). The affinity of humanized Fd138-80 is comparable to that of the murine antibody (Ka of 4.8×107 M-1 vs 5.2×107 M-1).

TABLE 6

Binding affinities of murine and humanized antibodies.		antibodies.
Mouse K _a (M ⁻¹)	Humanized K _a (M ⁻¹)	
Fd79 (anti-gB) Fd1 38-80 (anti-gD)	1.1×10^{8} 5.2×10^{7}	5.3 × 10 ⁷ 4.8 × 10 ⁷

Murine Fd79 and Fd138-80 have been shown to neutralize HSV-1 in vitro without complement (J. Koga et al., Virology 151, 385 (1986)), so the neutralizing activities of the humanized antibodies were compared with the mouse antibodies, Serial dilutions of equal quantities of murine and humanized antibodies were incubated with virus for 1 hr. before inoculation onto Vero cells. After 4 days, cells were stained with neutral red to visualize plaques. Results from these plaque reduction assays indicated that both humanized Fd79 and Fd138-80 neutralize virus as efficiently as their murine counterparts (FIGS. 32A and B). Both humanized and murine Fd79 cause a 90% reduction of plaques at an antibody concentration of 10 nM (1.5 ug/ml), Similarly, humanized and murine Fd138-80 were able to cause a 90% plaque reduction at equivalent levels.

The antibodies were also investigated for their ability to protect cells from viral spread in tissue culture. Vero cells were inoculated with virus at 0.1 pfu/cell and allowed to adsorb for 2 hrs. at 37° C. before addition of 10 ug/ml antibody. After four days, cells were stained with an anti-gB 5 antibody for detection of viral antigens on infected cells. Results indicated that both murine and humanized Fd79 at 10 ug/ml protected culture cells from infection (FIG. 33A). However, neither murine nor humanized Fd138-80 were able to protect cells against viral spread (FIG. 33B), despite 10 their ability to neutralize virus before inoculation. Both gB and gD are thought to be associated with cell fusion and virus infectivity (W. Cai et al., J. Virol. 62, 2596 (1988) and A. O. Fuller and P. G. Spear, Proc. Natl. Acad. Sci. USA 84, 5454 (1987)). However, it is possible that Fd79 blocks both 15 the infectivity and cell fusion functions of gB, while Fd138-80 blocks only the infectivity function of gD, so virus can still spread cell-to-cell.

The binding, neutralization and protection results all indicate that the humanized Fd79 and Fd138-80 antibodies 20 have retained the binding activities and the biological properties of the murine monoclonal antibodies. The availability of humanized antibodies with specificity for HSV gB and gD, inter alia, provides an opportunity for studies of the in vivo potency and immunogenicity of humanized antibodies 25 in treating viral diseases. The recognition by Fd79 and Fd138-80 of type-conunon epitopes of gB and gD (J. Koga et al., Virology 151, 385 (1986)) expands the therapeutic potential to herpes simplex virus type 2 as well as type 1.

The use of a combination of two or more humanized 30 antibodies in therapy is important for reducing the development of antibody resistant strains. Combination therapy of humanized antibodies with other antiviral agents such as acyclovir provides further opportunities to combat diseases when chemotherapeutic agents alone have not been effec- 35 tivc. As Fd79 and Fd138-80 reduce the frequency of viral persistence in a murine ocular model (J. F. Metcalf et al., Cur. Eye Res. 6, 173 (1987)), the humanized antibodies, typically together with other antiviral agents, are capable of reducing episodes of recurrent genital infection, an area 40 where traditional anti-viral agents have not been effective (L. Corey et al., N. Engl. J. Med. 306, 1313 (1982)). Incorporation of the human constant domains can also enhance effector functions, such as antibody-dependent cellular cytotoxicity, leading to greater therapeutic efficiency in 45 human natients.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other HSV specific antibodies.

In comparison to mouse monoclonal antibodies, the 50 present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement. 55

EXAMPLE 7

Design of genes for M195 humanized light and heavy chains

The p67 protein or CD33 antigen is found on the surface of progenitors of myeloid cells and of the leukemic cells of most cases of AML, but not on lymphoid cells or nonhematopoietic cells (sec, Leucocyte Typing III, ed. by A. J. 65 McMichael, Oxford University Press, pp. 622–629 (1987), which is incorporated herein by reference). Antibodies that

are known to bind to the CD33 antigen include L4B3, L1B2 and MY9 (Andrews et al., Blood 62, 124 (1983) and Griffin et al., Leukemia Research 8, 521 (1984), both of which are incorporated herein by reference).

Another antibody that binds to CD33 is M195 (Tanimoto et al., Leukemia 3, 339 (1989) and Scheinberg et al., Leukemia 3, 440 (1989), both of which are incorporated herein by reference).

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindlII sites, and 5' primers that hybridized to the dG tails and contained EcoRl sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRl and HindIII and cloned into the pUC18 vector for sequencing. For M195, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequences. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG, 34A and FIG, 34B. Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVgl-dhfr (FIG. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cy1 segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl Acad. Sci. USA 80, 2495 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG, 24B) is similar to pVgl-dhft but contains the human genomic Ck segment and the gpt gene. Derivatives of the M195 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained Xbal sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (sec, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric M195 antibody was shown to bind to U937 cells, which express the CD33 antigen, by flow cytometry (FIG. 35).

Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antibody is to the original mutine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysi's Software (Beckman)), the antibody Eu was chosen to provide the framework sequences for humanization of M195.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the M195 variable region. The model was used to detennine the amino acids in the M195 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain M195 variable regions, at each position the amino acid was chosen to be the same as in the Eu antibody, unless that position fell in one or more of four categories:

- (1) The position fell within a CDR,
- (2) The Eu amino acid was unusual for human antibodies at that position, whereas the M195 amino acid was typical for human antibodies at that position,
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human 20 sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Eu light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these cat- 25 egories, the amino acid from the mouse M195 antibody was used: The amino acids in each category are shown in Table 7. Some amino acids may be in more than one category. The final sequences of the humanized M195 light and heavy chain variable domains arc shown in FIG. 36A and FIG. 30 36B, compared with the Eu sequences.

TABLE 7

Category	Light Chain	Heavy Chain	
1	24-38, 54-60, 93-101	31-35, 50-66, 99-105	
2	10, 52, 67, 110	93, 95, 98, 106, 107, 108, 110	
3	_	30, 67, 98, 106	
4	40, 52, 74	27, 30, 48, 68, 98	

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse M195 chains (FIG. 34A and FIG. 34B), generally utilizing codons 45 found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an Xbal site at each end. Each gene was constructed from four 50 overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 37A and FIG. 37B). The oligo- 55 the polypeptide chains comprising the heavy and light chain nucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction 60 enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the Xbal sites of the pVgl-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were 65 carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones wcre screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody was eluted with 0.2M Glycine-HCI, pH3.0 10 and neutralized with 1M Tris PH8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized M195 antibody was characterized in 15 comparison to the murine and chimeric antibodies. The humanized antibody bound to U937 cells in a fluorocytometric analysis in a manner similar to the chimeric antibody (FIG. 35), showing that it recognizes the same CD33 antigen

The affinity of the humanized antibody was determined by competition with the radio-iodinated mouse M195 antibody (FIG. 38). The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in Fundamental Immunology (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference). The mouse M195 had an affinity comparable to the published value (Tanimoto et al., op. cit.) and the humanized M195 antibody had an affinity the same as the mouse M195 to within experimental error.

Humanized M195 is useful to mediate antibody-dependent cellular cytotoxicity when human effector cells and human CD33-expressing cells are used. This is analogous to other humanized antibodies, such as reported by Junghans et al., Cancer Research 50, 1495 (1990), which is incorporated herein by reference.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other CD33 specific antibodies. In comparison to mouse monoclonal antibodics, the present 40 humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

EXAMPLE 8

Design of genes for CMV5 humanized light and heavy chains

Three neutralizing antibodies to the gH glycoprotein of human cytomegalovirus (CMV) are designated CMV5, CMV109 and CMV115.

Exemplary DNA sequences, which on expression code for CDR's of monoclonal antibody CMV5 are included in FIG. 39A and FIG. 39B. Due to codon degeneracy and noncritical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors pVk and pVgl-dhfr (FIG. 24A and 24B) using site-directed mutagencsis, such as after CHI to produce Fab fragments or after the hinge region to produce $(Fab')_2$ fragments.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable 5 domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoR I sites (scheme shown in FIG. IO 14). The PCR amplified fragments were digested with EcoR I and HindIII and cloned into the pUC18 vector for sequencing. For CMV5, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced 15 amino acid sequences are shown in FIG. 39A and FIG. 39B. Similarly, by using techniques, which are well-known in the art, cDNAs for the CMV109 and CMV115 antibodies may be obtained and their sequence determined. Construction and expression of chimeric antibody. 20

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVgl-dhfr (FIG. 24A) contains a human cytomegalovirus IEI promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cyl segment including part of 25 the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVgl-dhfr but contains the human genomic Ck segment and the gpt 30 gene. Derivatives of the CMV5 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained Xbal sites; the 3' primers hybridized to the last 15 nucleotides of the J 35 regions and contained splice donor signals and X bal sites (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the cytomegalovirus 40 promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse mycloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete 45 antibody were detected by ELISA. Purified chimeric CMV5 antibody was shown to bind to CMV-infected cells, which express the gH antigen, by immunostaining of CMV-infected human embryonic lung fibroblasts.

Computer modeling of humanized antibodies 50 In order to retain high binding affinity in the humanized amibodies, the general procedures of Queen et al. were followed (See, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which arc incorporated herein by reference). The more homologous a human anti- 55 body is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework 60 sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Wol was chosen to 65 provide the framework sequences for humanization of CMV5.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the CMV5 variable region. The model was used to determine the amino acids in the CMV5 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain CMV5 variable regions, at each position the amino acid was chosen to be the same as in the Wol antibody, unless that position fell in one or more of five categories:

(1) The position fell within a CDR,

- (2) The Wol amino acid was unusual for human antibodies at that position, whereas the CMV5 amino acid was typical for human antibodies at that position,
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Wol light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse CMV5 antibody was used. In addition, a position was in the fifth category if the Wol amino acid was highly unusual for human antibodi es at that position, and the CMV5 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 8. Some amino acids may be in more than one category. The final sequences of the humanized CMV5 light and heavy chain variable domains are shown in FIG. 40A and FIG. 40B, compared with the Wol sequences.

TABLE 8

INDEE 0		
Light Chain	Heavy Chain	
24-34, 50-56, 89-97	31-35, 50-66, 99-108	
	69,80	
	69, 80	
49	30	
	24, 27, 28, 30, 97	
	5	
	24-34, 50-56, 89-97	

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse CMV5 chains (FIG. 39A and FIG. 39B), generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an Xbal site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleoides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 41A and FIG. 41B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction

enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the Xbal sites of the pVgl-dbfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids are transfected into Sp2/0 mouse myeloma cells by electroporation and cells are selected for gpt expression. Clones are screened by 10 assaying human antibody production in the culture supernatant by ELISA, and antibody purified from the bestproducing clones. Antibody is purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody is 15 eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1 M Tris PH8.0. The buffer is exchanged into PBS by passing over a PD10 column (Pharmacia).

Humanized antibody was also produced by transient transfection. The heavy chain and light chain plasmids were 20 transfected into S194 cells (ATCC TIB 19) by the DEAEdextran method (Queen et al., Mol. Cell. Biol. 4, 1043 (1984), which is incorporated herein by reference), and humanized CMV5 antibody was purified from the media supernatant as above. Antibody was quantitated by ELISA 25 assay for human lg.

Properties of humanized antibodies.

The humanized CMV5 antibody was characterized in comparison to the mutinc and chimeric antibodies. The humanized CMV5 antibody was shown to bind about as well 30 as the mouse and chimeric antibodies to CMV antigen, by immunostaining of CMV-infected human embryonic lung (HEL) cells (ATCC CCL 137). HEL cells monolayers in 96-well plates were infected with CMV at 0.01 pfu/cell, incubated for 4 days, dried at 37° C. and stored wrapped at 35 4° C. 100 µl blotto (5% Carnation Instant Milk in PBS at pH 7.4) was added to each well and incubated at 37° C. for 30 min. The blotto was poured off and 75 µl of a series of 2-fold dilutions of mouse, chimeric and humanized CMV5 antibody was added to the wells. The plate was incubated 1 hr 40 at 37° C. and washed twice with blotto (each wash was left on for 10 min). Then 75 µl of diluted peroxidase (HRP) conjugated goat anti-mouse or anti-human IgG (Tago) was added to each well and incubated for 1 hr at 37° C. The plate was washed 2x with PBS and 150 µl of HRP substrate 45 solution was added to each well. Color was allowed to develop at room temperature. The plates were washed with water and air dried. The wells were examined under a microscope to determine the highest dilution of the antibodies that formed a colored precipitate on the CMV-infected 50 cells. For all three antibodies, 63 ng/ml was the least amount of antibody that produced a detectable precipitate, indicating that humanized CMV5 binds about as well as the mouse and chimeric antibodies.

To compare the affinities of mouse and humaniz.ed CMV5 55 in another way, a competition experiment was performed. Plates of CMV-infected HEL cells as above were incubated with blotto for 30 min at 37° C. The blotto was poured off and dilutions of mouse or humanized CMV5 were added to each well in 75 μ l of PBS. Then 125 μ l of radio-iodinated 60 mouse CMV5 (1 μ Ci/ μ g) in PBS, containing 28,000 cpm was added to each well and incubated at 37° C. for 2.5 hr. The plate was washed 5 times with PBS, and the contents of each well were solubilized with 200 μ l of 2% SDS and counted. Increasing concentrations of mouse and humanized 65 CMV5 inhibited binding of the radiolabeled CMV5 about equally well (FIG. 42), so humanized CMV5 has approximately the same binding affinity as mouse CV5. An irrelevant antibody did not compete in this assay.

The ability of humanized CMV5 to neuralize CMV is compared to that of mouse CMV5. Mouse and humanized CMV5 are successively diluted by 2-fold in 100 µl of DME medium+2% FCS in wells of a 96-well plate. 100 µl of CMV, which has been diluted to contain 100 tissue culture infectious dose-50% (TCID50) units, are added to each well and incubated for 60 min at 37° C. Each well of antibodyvitus mixture is added to a well of subconfluent HEL cells in a 96-well plate from which the medium has been removed. The cells are incubated for 5 days and cytopathic effect (CPE) is examined in each well under a microscope. The highest dilution of antibody that inhibits CPE by 90% is a measure of the neutralizing ability of the antibody. The humanized CMV5 antibody will neutralize CMV antibody approximately as well as the mouse CMV5 antibody.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other CMV specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

EXAMPLE 9

Design of genes for AF2 human-like light and heavy chains

This example is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of γ -IFN, such as monoclonal antibody AF2. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody AF2 are included in FIG. 43A and FIG. 43B. Due to codon degeneracy and non-critical aminoacid substitutions, other DNA sequences can be readily substituted for those sequences.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoR 1 sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoR 1 and HindIII and cloned into the pUC18 vector for sequencing. For AF2, two gamma-2b specific and two kappa specific clones were sequenced. The two gamma-2b clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG. 43A and FIG. 43B. Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVgl-dhfr (FIG. 24A) contains a human cytomcgalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic C γ l segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVgl-dhfr but contains the human genomic C κ segment and the gpt gene. Derivatives of the AF2 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained Xbal sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and Xbal sites (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cclls sclected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Chimeric AF2 antibody ¹⁵ was shown to bind to human γ -IFN by ELISA. Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were 20 followed (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce 25 affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence data- 30 base (performed with the MicroGenic Sequence Analysis Software (Beckman)), the antibody Eu was chosen to provide the framework sequences for humanization of AF2.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) ³⁵ was used to construct a model of the AF2 variable region. The model was used to determine the amino acids in the AF2 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain AF2 variable regions, at ⁴⁰ each position the amino acid was chosen to be the same as in the Eu antibody, unless that position fell in one or more of five categories:

- (1) The position fell within a CDR,
- (2) The Eu amino acid was unusual for human antibodies at that position, whereas the AF2 amino acid was typical for human antibodics at that position,
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino 50 acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., 55 op. cit.) as the Eu light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse AF2 antibody was 60 used. In addition, a position was in the fifth category if the Eu amino acid was highly unusual for human antibodies at that position, and the AF2 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used. 65

The amino acids in cach category are shown in Table 9. Some amino acids may be in more than one category. The final sequences of the humanized AF2 light and heavy chain variable domains are shown in FIG. 44A and FIG. 44B, compared with the Eu sequences.

TABLE 9			
Calegory	Light Chain	Heavy Chain	
1	24-34, 50-56, 89-97	31-35, 50-66, 99-106	
2	48	93, 95, 98, 107, 108, 109, 111	
3		30, 98, 107	
4	48, 70	27, 28, 30, 98, 107	
5	63		

For the construction of genes for the humanized antibodies, nucleonide sequences were selected that encode the protein sequences of the humanized heavy and light chains, plus typical immunoglobulin signal sequences, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric gencs and an Xbal site at each cnd. Each gene was constructed from four overlapping synthetic oligonuclcotides. For each variable domain genc, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 45A and FIG. 45B) The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction enzymes, ligated to the pUC18 vcctor and sequenced. Two fragments with the respectively correct half-sequences are then ligated into the XbaI sites of the pVg l-dhir or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions are carried out under conditions wellknown in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids are transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones are screened by assaying human antibody production in the culture supernatant by ELISA, and antibody purified from the best-producing clones. Antibody is purified by passing tissue culture supernatant over a column of staphylococcal protein A-Scpharosc CL-4B (Pharmacia). The bound antibody is eluted with 0.2M Glycine-HCI, pH3.0 and neutralized with 1M Tris PH8.0. The buffer is exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized AF2 antibody is characterized in comparison to the murine and chimeric antibodics. The humanized antibody will bind to γ -IFN in an ELISA assay in a manner similar to the mouse and chimeric antibodies, showing that it recognizes γ -IFN.

To compare the binding affinities of mouseAF2 antibody and humanized AF2 antibody, a competitive ELISA assay is performed. An ELISA plate is coated with human recombinant γ -IFN by adding 100 µI of a 500 ng/ml solution of γ -IFN in PBS to each well and incubating overnight at 4° C. Subsequent steps are carried out at room temperature. The γ -IFN solution is removed and 200 µl of ELISA buffer (0.1% Tween-20, 1% Bovine serum albumin in PBS) is added to each well and incubated for 1 hr. After removing the solution, varying amounts of competitor antibody (mouse AF2 or humanizedAF2) in 100 µl PBS is added to each well,

along with an amount of biotinylated AF2 predetermined to give a good ELISA response. The plate is incubated for 1 hr and then washed 3 times with ELISA buffer. An amount of horseradish peroxidase (HRP)-conjugated strepavidin predetermined to be in excess is added in 100 µl PBS to each 5 well and incubated for 30 min. The plate is washed 3 times in ELISA buffer, and 100 µl of substrate solution for HRP is added to each well. The plate is incubated for 10-30 min, and the optical density of each well is determined with an ELISA reader (BioRad). The decrease in optical density with 10 increasing concentrations of competitor antibodies mouse AF2 and humanized AF2 are plotted. Mouse AF2 and humanized AF2 will compete similarly, showing that their binding affinities for γ -IFN are approximately the same. The procedures used are well known in the art (e.g., Harlow and 15 Lane, op. cit.).

An important biological activity of γ -IFN is the induction of expression of class II HLA antigens on cells. To determine the ability of mouse and humanized AF2 to neutralize this activity, about 5×10⁴ HS294T cells (Basham et al., J. Immu- 20 nol. 130, 1492 (1983), which is incorporated herein by reference) are plated in 1.0 ml DMEM medium+10% FCS in each well of a 24-well plate. After overnight incubation, 0.1 nM interferon and varying amounts of mouse or humanized AF2 are added to the cells, and the plate is incubated for 25 72 hr. The cells are removed from the plate with 0.05M EDTA, stained with monoclonal antibody L243 from the

American Type Culture Collection (ATCC) against HLA-D antigen, washed, stained with FITC conjugated goat antimouse lg and analyzed with a FACScan (Becton-Dickinson). Increasing concentrations of mouse AF2 refuce fluorescence of the cells (FIG. 46), indicating the antibody is preventing induction of HLA-D by γ -IFN. The humanized AF2 will act similarly to mouse AF2 in this assay, showing that it neutralizes the biological activity of γ -IFN.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other γ -IFN specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference. Although the present invention has been described in some detail by way of illustration and example for purposes of clarity and understanding, it will be apparent that certain changes and modifications may be practiced within the scope of the appended claims.

```
SEQUENCE LISTING
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( I ) GENERAL INFORMATION:
```

```
( i i i ) NUMBER OF SEQUENCES: 113
```

```
( 2 ) INFORMATION FOR SEQ ID NO:3:
```

```
    (i) SEQUENCE CHARACTERISTICS:
    (A) LENGTH: 106 amino acids
    (B) TYPE: amino acid
    (C) STRANDEDNESS: single
    (D) TOPOLOGY: unknown
    (ii) MOLECULE TYPE process
```

```
( i i i ) HYPOTHETICAL: NO
```

```
( i x ) FEATURE:
```

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( A ) NAMERCEY: Protein
```

```
( B ) LOCATION: 1.106
```

(D) OTHER INFORMATION: /notc="Variable region of the mouse

```
anti-Tac antibody light chain."
```

```
( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:1:
```

G I n 1	llc Val	Leu Thr Glo S	Scr Pro Ala	llc Mci Scr 10	Ala Scr Pr 15	o Gly
Glu	Lys Val	Thr llc Thr 20	Cys Scr Ala 25	Ser Ser Ser	llc Scr Ty 30	r Mci
His	Ттр Phc 35	Glp Gln Lys	Pro Gly Thτ 40	Ser Pro Lys	Leu Trp II 45	с Тут
Tbr	Tbr Scr 50	Asa Leu Ala	Scr Gly Vai 55	Pro Ala Arg 60	Phc Scr GI	ly Scr
G 1 y 6 5	Scr Gly	Thr Scт Tyr 70	Scr Lcs Thr	11с Sст Атд 75	Mcı Glu Al	ia Glu 80
			Cys His Gln			

69		70
	-continued	
Phe Gly Ser Gly Thr Lys . 100	Lev Glu Leu 105	Lys
2) INFORMATION FOR SEQ ID NO:2:		
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 107 amino acids ('B) TYPE: amino acid (C) STRANDEDNESS: aingle (D) TOPOLOGY: urknown 		
(i i) MOLECULE TYPE: protein		
(1 i i) WYPOTHETICAL: NO		
(i x) FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1107 (D) OTHER INFORMATION: /note≃"Va Eu entribody light chaim."	siable region of the human	
(x i) SEQUENCE DESCRIPTION: SEQ ID NO:2:		
Aspile Olm Met Thr Gin 1 5	Ser Pro Ser	Thr Lev Ser Ala Ser Val Gly 10 15
Asp Arg Val The lic Thr 20	Cys Arg Ala 25	Scr Gin Scr llc Asn Thr Trp 30
Leu Ala Trp Tyr Gln Gin 35	Lys Pro Gly 40	Lys Ala Pro Lys Leu Leu Mei 45
Tyr Lys Ala Scr Scr Leu 50	Glu Scr Gly 55	Val Pro Scr Arg Phc Ilc Gly 60
Ser Gly Ser Gly Thr Olu 65 70	РЪс ТЬг Іси	Thr lle Ser Ser Leu Gln Pro 75 80
Asp Asp Phe Ala Thr Tyr 85	Tyr Cys Gln	Glu Tyr Asn Scr Asp Scr Lys 90 95
Mct Phc Oly Gln Gly Thr 100	Lys Val Glu 105	Val Lys
2) INFORMATION FOR SEQ ID NO:3:		
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 116 amino arids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: unknown 		
(i i) MOLECULE TYPE: procin		
(i i i) HYPOTHETICAL: NO		
(i x) FEATURE (A) NAME/KEY: Protein (B) LOCATION: 1116 (D) OFFIER INFORMATION: /houe="Va anti-Tac antibody heavy chaim."		
(x i) SEQUENCE DESCRIPTION: SEQ ID NO-3:		
GIn Val Glu Leu Glu Glu I S	Scr Gly Ala	Glu Leu Ala Lys Pro Gly Ala 10 15
Scr Val Lys Mci Scr Cys 20	Lys Ale Scr 25	Gly Tyr Thr Phe Thr Ser Tyr 30
Arg Mci His Trp Val Lys 35	GIN Arg Pro 40	Gly Gln Gly Lcu Glu Trp llc 45
Gly Tyr Ile Aso Pro Ser 50	Thr Gly Tyr 55	Thr Giu Tyr Aso Gin Lys Phe 60
Lys Asp Lys Ala Tbr Lcu 65 70	The Ala Asp	Lys Scr Scr Scr Thr Ala Tyr 75 80

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	71	-соп	tinued		72
Met Gin Len	Ser Ser Lev 85	υ Thτ Phe	Glu Asp 90	Scr Ala V	al Tyr Tyr Cys 95
Ala Arg Gly	y Gly Gly Val 100	Pbc Asp	Туг Тгр 105	Gly Gln G	ly Thr Thr Leu li0
Thr Val Sca 113					
2) INFORMATION FOR SEQ II) NG:4:				
(B) TYPE (C) STRA	FTH: 117 emino eciós				
(i i) MOLECULE TYP	E: protein				
(i i i) HYPOTHETICAL	.: NO				
(B)LOCA (D)OTHE	DIKEY: Protein 1710N: 1117 38 INFORMA TION: /note="\ Eu anibody heavy chain."	/ariable.region of the	human		
(xi) SEQUENCE DES	CRIPTION: SEQ 1D NG:4:				
Gln Val Glu 1	a Leu Val Gli 5	Ser Gly	Ala Glu 10	Val Lys L	ys Pro Gly Scr 15
Scr Val Ly:	s Val Scr Cy: 20	s Lys Ala	Scr Gly 25	Gly Thr 🖡	he Ser Arg Ser 30
Ala 11c 11c 35	c Trp Val Arg	Gln Ala 40	Ρτα Gly	Gln Gly L 4	
Giy Giy 110 50	e Val Pro Mei	Phe Gly 55	Pro Pro	Asn Tyr A 60	la Gln Lys Phe
Gln Gly Ary 65	g Val Thr II 70	Thr Ala	Asp Glu	Ser Thr A 75	sn Thr Ala Tyr 80
Mcı Glu Lei	u Scr Ser Lev 85	1 Arg Scr	Glu Asp 90	Thr Ala P	hc Tyr Phe Cys 95
Ala Gly Gl	y Tyr Gly llo 100	с Туг Ѕсг	Pro Glu 105	Glu Tyr A	sn Gly Gly Lcu 110
Val îbr Va Ji:					
2) INFORMATION FOR SEQ II	ጋ እርጉት				
(i) SEQUENCE CHA (A) LENC (B) TYPE (C) STRA					
(i i) MOLECULE TY	PE: procein				
(i i i) HYPOTHETICAL	.: NO				
(B)LOCA	EJKEY: Protein ATION: 1116 ER INFORMATION: /nate="\	/ariable region of the	PDL		

- D) OTHER INFORMATION: /nate="Variable region of the PDL humanived anti-The antibody heavy chaim."

(x i) SEQUENCE DESCRIPTION: SEQ 1D NO:5:

```
Gla Val Glu Lcu Val Gla Scr Gly Ala Gtu Val Lys Lys Pro Gly Scr
l 5 10 15
Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser Tyr
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Arg	B	M	сl		H i 3 5			Г	p	v	8	l	A	r	g	G	l	n	A 4		2	P	r 0	G	1	у	G	1 т		G	ļy		с 5	u	G	l	ų	Т	r p		[1	С
Gly	у	Т 5			11	с	1	A s	n	Р	۴	0	S	c	r		հ Տ		G	1	у	Т	y r	T	Ъ	r	G	1 .		T :		A	5	n	G	l	n	L	y s	0	Pł	c
L y s 6 5	S	A	s p	,	L)	5	1	A 1	a	T	h	r	1 7		C	Т	h	t	A	1	a	A	s p	G	1	Ш	S 7			TI	r	A	s	n	T	h	r	A	1 a		T y S (
Mc	1	G	1 0		LC	u		S c	r		с 5		L	с	U	A	r	90	S	C	r	G	1 0		2	p	Т	hı		AI	a	V	8	l	T	у	r	T 9			C)	s
Ali	8	A	r g		G I	У			y 0	G	1	У	۷	а	ł	P	b	c	A	s	р		уг 05		r	P	G	1 3	,	G	l n	G	; 1	у	T 1			L	CU	. '	V	1
Th	г	۷	a l		S o			Sc	г																																	

(2

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 116 amino acids
- (B) TYPE: amino pcid (C) STRANDEDNESS: single (D) TOPOLOGY: unknown

(i i) MOLECULE TYPE: prokin

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

(A) NAMEREY: Protein (B) LOCATION: 1..116 (D) OTHER INFORMATION: /note="Variable region of the CDR only humanized anti-Tac antibody beavy chain."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO.6

G 1 n 1	Val	Gln L	cu Val S	Gln Scr	Gly Ala	Giu Val 10	Lys Lys	Ρεφ Gly 1.5	Scr
Scr	Val	Lys V 2		Cys Lys	Ala Scr 25	Gly Gly	Thr Phc	Scr Scr 30	Туг
Arg	Mct	His T 35	rp Val	Arg Gln	Ala Pro 40	Gly Glo	Gly Lcu 45	Glu Trp	McL
Gly	Τγτ 50	llc A	sa Pro	Ser Thr 55	Gly Туг	Thr Glu	Tyr Asn 60	Gln Lys	Рыс
L y s 6 5	Asp	Arg V	al Thr	llc Thr 70	Ala Asp	Glu Scr 75	Thr Asn	Thr Ala	Туг 80
Mc L	Glu	Lcu S	er Ser 85	Leu Arg	Scr Glu	Asp Thr 90	Ala Phc	Tyr Phc 95	Cys
Ala	Gly		1 y G l y 0 0	Val Phc	Asp Tyr 105	Glu Tyr	Asn Gly	Giy Leu 110	Val
Thr	Val	Scr S 115	СГ						

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 106 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: unicown

(i i) MOLECULE TYPE: protein

(i i i) HYPOTHETICAL: NO

(i x)FEATURE: (A) NAMERKEY: Protein -continued

76

(x	i) SE	OL	JE.	ICE	D	E	SCA	ריונ	DC	N'	SE	0	D	NC	:7:													1															
													-																															
A :	S	р	1	1 c		G	1	۵	М	C	L	T 5		г	C	1	۵		S	СГ		P	го	-	5 0	T				L	, C	U	S	C	г	A		D	S	C	r	V 1		G
A	S	р	A	r g			3		1 2		r	I	I	с	J	b	r	(0	y s	1	S	C r		2 5		-	c	г	S	С	T	S	С	٢	1	1 0	C	S 3		F	Т	уг	M
н	i	S	ľ	ı þ		T 3		τ	G	1	n	G	l	n	L	y	s		P	r 0		G 4		I	- 3	s	1	1	a	P	F	0	L	у	s		с і 5	ц	L	C	ц	à	1 c	1
ΤI	b		T 5			S	c	ť	A	5	D	L	c	u	A	. 1	D		5		1	G	l y	١	/ a	1	1	I	0	A	1	a		r O	B	P	h	C	S	C	r	G	1 y	S
G .		у	S	C 1		G	1	у	Т	h	r	G	1	U		b O		0	r	hr		L	c u	1	C	F	1	: 1	C		5		S	c	T	L	C	u	G	1 :	n	P	ro	8
A	S	р	P	h c		A	1	2	1	h	r		у 5		Т	y	r	(C	y s		H	i s	C	G 1	п			~	S	C	T	Т	h	r	Т	у	r	Р	r (þ	L 9		1
PI	h	c	G	1 y		G	1	п	G 1			Т	h	r	L	y	s	1	V	a 1		G	lu			1) 5	L	. y	s															

- (A) LENGTH: 106 amino acids
- (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: unknown

(i i) MOLECULE TYPE: proterin

(i i i) HYPOTHETICAL: NO

- (i x) FEATURE:
 - (A) NAMErKEY: Protein (B) LOCATION: 1..106
 - (D) OTHER INFORMATION: /note="Variable region of the CDR-only burnanised anti-Tac antibody light chain."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:8:

A s p I	llc Gln	Mci Thr Gla S	Scr Pro Scr	Thr Leu Ser 10	Ala Scr Val Gly 15
Asp	Arg Val	Thr Ilc Thr 20	Cys Scr Ala 25	Ser Scr Scr	lle Ser Tyr Mei 30
His	Т гр Туг 35	Gin Gin Lys	Pro Gly Lys 40	Ala Pro Lys	Leu Leu Mei Tyr 45
Thr	The Scr 50	Asn Lcu Alo	Scr Gly Val 55	Pro Scr Arg 60	Phc llc Gly Scr
G 1 y 6 5	Scr Gly	The Glu Pbc 70	Thr Leu Thr	llc Scr Scr 75	Leu Gla Pro Asp 80
Asp	Pbc Ala	Thr Tyr Tyr 85	Cys His Gln	Arg Scr Thr 90	Tyr Pro Lou Thr 95
Pbc	Gly Gln	Gly Thr Lys	Val Glu Val 105	Lys	

(2) INFORMATION FOR SEQ 1D NO:9:

(i) SEQUENCE CHARACIERISTICS: (A) LENG'I'H: 443 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

-continued

78

(i i i) HYPOTHETICAL: NO

(i x) FEATURE

(A) NAMEREY: misc_feature

(B) LOCATION: 1.443

(D) OTHER INFORMATION: /notc="Sequence encoding heavy chain variable region of CDR-only humanized

anti-Tac antibody including signal sequence."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:9:

AGCTTCTAGA	TGGGATGGAG	CTGGATCTTT	CTCTTCCTCC	TGTCAGGTAC	CGCGGGGCGTG	6 0
CACTCTCAGG	TCCAGCTTGT	CCAGTCTGGG	GCTGAAGTCA	AGAAACCTGG	CTCGAGCGTG	120
AAGGTCTCCT	GCAAGGCTTC	TGGCGGGACC	TTTTCTAGCT	ACAGGATGCA	CTGGGTAAGG	1 8 0
CAGGCCCCTG	GACAGGGTCT	GGAATGGATG	GGATATATTA	ATCCGTCGAC	TGGGTATACT	240
GAATACAATC	AGAAGTTCAA	GGACAGGGTC	ACAATTACTG	CAGACGAATC	CACCAATACA	30 🕈
GCCTACATGG	AACTGAGCAG	CCTGAGATCT	GAGGACACCG	CATTCTATTT	CTGTGCAGGG	360
GGTGGGGGAG	TCTTTGACTA	CGAATACAAT	GGAGGGCTGG	TCACAGTCTC	CTCAGGTGAG	420
TCCTTAAAAC	CTCTAGACGA	TAT				443

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 411 base pairs (B) TYPE: nucleic acid

- (C) STRANDEDNESS. single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

(A) NAME/KEY: misc_fcature

- (B) LOCATION: 1..411
- (D) OTHER INFORMATION: /vouc="Sequence cocoding light chain variable region of the CDR-only humanized
 - anti-Tac antibody including signal sequence."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:10:

C A A A T C T A G A	TGGAGACCGA	TACCCTCCTG	CTATGGGTCC	TCCTGCTATG	GGTCCCAGGA	60
TCAACCGGAG	ATATTCAGAT	GACCCAGTCT	CCATCTACCC	TCTCTGCTAG	CGTCGGGGAT	120
AGGGTCACCA	TAACCTGCTC	TGCCAGCTCA	AGTATAAGTT	ACATGCACTG	GTACCAGCAG	180
AAGCCAGGCA	AAGCTCCCAA	GCTTCTAATG	TATACCACAT	CCAACCTGGC	TTCTGGAGTC	240
CCTTCTCGCT	TCATTGGCAG	TGGATCTGGG	ACCGAGTTCA	CCCTCACAAT	CAGCTCTCTG	300
CAGCCAGATG	ATTTCGCCAC	TTATTACTGC	CATCAAAGGA	GTACTTACCC	ACTCACGTTC	3 6 0
GGTCAGGGGA	CCAAGGTGGA	GGTCAAACGT	AAGTACACTT	TTCTAGATAT	A	411

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single

- (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: DNA

(i i i) HYPOTHEDCAL: NO

(i x) FEATURE:

(A) NAME/KEY: misc_(calure (B) LOCATION: 1..29

(D) OTHER INFOR MATION: /standard_name="Primes me045"

		79										80			
						-CON	tinued					112.57	NC 25		
(xi)SEQU	ENCE DESCR	IPMON:	SEQ 1D	NO:11:											
ΤΑΑΤCΤΑGΑΑ	ττςςςς	ccc	c cc	cccc	ссс										29
(2) INFORMATION F	OR SEQ ID N	0:12:													
()	ENCE CHARA A) IENGTH B) TYPE: m C) STRANE D) TOPOLO	l: 46 bes nelcic aci DEDNES:	c pairs id S: single												
(ii) MOLE	CUIE TYPE:	DNA													
(iii)HYPO	THETICAL: N	10													
(JRE: A) NAME/K B) LOCATH D) OTHER	DN: 1_46	6	standard	_namc="P	rimer me	045''								
(xi)SEQUI	ENCE DESCR	IPIION:	SEQ ID I	XO:12:											
TATAGAGCTC	AAGCTT	GGA	r GG	GGG	AAGA	TGG	ΑΤΑΟ	AGT	TGGT	GC					4 6
(2) INFORMATION F	or seq id n	0:13:													
	ENCE CHARA A)LENGTE B)TYPE: D C)STRAND D)TOPOLO	l: 50 bas ucliic aci DEDNES	c pairs id S: single												
(ii) MOLE	CULE TYPE:	DNA													
(iii)HYPO	THETICAL: N	10													
(IRE: A) NAMER B) LOCATIO D) OTHER	ON: 1_50	0	standard_	.namc="f	น์ตระ กอร	347"								
(xi) SEQU	ENCE DESCR	PIION:	SEQ ID I	NO:13:											
TATAGAGCTC	AAGCTT	CCA	G TG	GATA	GACH	GAT	GGGG	STG	TYGT	T T T G	ĢC				50
(2) INFORMATION F	OR SEQ ID N	0:14:													
(ENCE CHARA A) LENGTE B) TYPE: au C) STRAND D) TOPOLO	I: 116 an mino acio DEDNTES:	nino zcids 1 S: single												
(ii) MOLE	CULE TYPE:	protein													
(iii)HYPO	INENCAL: N	0													
(A) NAME/K B) LOCAID D) OTHER	ON: 111	I6 IATION: /	nolc="An	u'-l'ac he	vy chain	amino								
(xi)SEQU	ENCE DESCR	IPHON:	SEQ ID	NO:14:											
Gln Va 1	1 GID	Lcu	G L 1 5	Gla	Scr	G 1 y	Ala	G 1 u 1 0	Lcu	Ala	Lys	Pro	GIY 15	Ala	
Ser Va	1 Lys	Mc 1 20	Ser	Суş	Lys	Ala	Sст 25	Gly	Туг	Thr	Phc	T b r 3 0	Scr	Туг	
	l His 35	Trp	Val	Lys	Gla	Arg 40	Pro	Gly	Glɒ	Gly	Lсц 45	Glu	Τrp	11c	
СІу Ту	r llc	Asn	Pro	Scr	Tbr	G 1 y	Tyr	Thr	GΙυ	Туг	Asp	Gla	Ĺys	РЪс	

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						_				_			-		_
	50					5 5					60				
- y s 5	Asp	Lys	Ala	Thτ	L c u 7 0	Tbr	Ala	Asp	Lys	Ser 75	Scr	Scr	Thr	Ala	Туг 80
Mc i	Gln	Lcu	Scr	S c r 8 5	Lcu	Tbr	РЪс	Glu	Asp 90	Scr	Ala	Val	Туг	Туг 95	Суѕ
Ala	Arg	Gly	GI y 100	Gly	Val	РЬс	Asp			Gly		Gly	Th r 110	Thr	Lcu
Гаг	Val		Ser												

(2) INFORMATION FOR SEQ ID NO.15:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTI: 117 smino acids (B) TYPE: amino scid (C) STRANDEDNESS: single (D) TOPOLOCY: urknown

(i i) MOLECULE TYPE: protein

(i i i) HYPOTHETICAL: NO

(i x)FEATURE:
(A) NAME/KEY: Protein
(B) LOCATION: 1.117
(D) OTHER DIFFORMATION: /note="Eu heavy chain amino acid sequence."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:15:

G l n 1	V a 1	Gin	Lcu	V a l S	GIn	Sci	G 1 y	Ala	G 1 u 1 0	Val	Lys	Lys	Pro	GI y 15	Ser
Scr	Val	Lys	V a I 2 0	Scr	Суs	Lys	Ala	Sст 25	G 1 y	GIy	Thr	Рһс	Sст 30	Arg	Scr
Ala] c	11 c 35	Тгр	V a l	Arg	Glu	A I a 4 0	Pro	Gly	Glo	Gly	L c u 4 5	Glu	Trp	McL
Gly	G 1 y 5 0	Il c	V a 1	Pro	Mcl	Phc 55	G 1 y	Pro	Pro	Aso	Туг 60	Ala	Gln	Lys	Phe
G 1 n 6 5	Gly	Arg	Vəl	Thr	II c 70	Tbr	Ala	Λsp	Glu	Ser 75	Thr	A s n	Thr	Ala	Туг 80
Mci	Glu	Lcu	Scr	Sст 85	Lcu	Arg	Scr	Glu	Asp 90	Tbr	Ala	Pbc	Туг	Р b с 9 5	Cys
A 2	Gly	Gly	Тут 100	Gly	11 c	Туr	Scr	Pro 105	Glu	Giu	Туг	Asa	G I y 1 1 0	Gły	Lcu
Val	Thr	V a 1 1 1 5	Scr	Sci											

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 106 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(i i) MOLDCULE TYPE protein

(111) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: Proucin
- (B) LOCATION: 1..106
- (D) OTHER INFORMATION: /neuc="Anti-Tac light chain amino acid sequence."
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:16:

1	1	

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Gin lle Val	Leu Thr Gln Scr Pr	o Ala lle Met Ser	Ala Scr Pro Gly						
1	S	10	15						
Glu Lys Val	Thr llc Thr Cys Sc	r Ala Scr Scr Scr	11с Ser Туг Мен						
	20	25	30						
His Trp Pbc	Gin Gin Lys Pro Gi	y Thr Scr Pro Lys	Lcu Trp llc Tyr						
35	40		45						
Thr Thr Scr	Asn Leu Ala Scr Gl	y Val Pro Ala Arg	Phc Scr Gly Scr						
50	55	60							
Gly Ser Gly	Thr Scr Tyr Scr Le	u Thr llc ScrArg	Mci Glu Ala Glu						
65	70	75	80						
Asp Ala Ala	Thr Tyr Tyr Cys Hi	s Gln Arg Scr Thr	Tyr Pro Leu Thr						
	85	90	95						
Phe Gly Scr	Gly Thr Lys Lou Gl 100	u Leu Lys 105							

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 107 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: unknown

(i i) MOLECULE TYPE: protein

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

(A) NAME/KEY: Protein (B) LOCATION: 1..107 (D) OTHER INFORMATION: / nove="Ex light chain amine acid sequence."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Asp 1	ll¢ Glπ	Mcl Thr Gln S 5	Scr Рго Scr	Thr Leu Ser 10	r Ala Scr Val Gly 15
Asp	Arg Val	Thr Ilc Thr C 20	Cys Arg Ala 25	Scr Gln Scr	r llc Asn Thr Trp 30
Lcu	Ala Trp 35	Tyr Gla Gla L	Lys Pro Gly 40	Lys Ala Pro	D Lys Leu Leu Mei 45
Туг	Lys Ala 50	Scr Scr Lcu C	Glu Ser Gly 55	Val Pro Scr 60	r Arg Phe lle Gly
5 c r 6 5	Gly Scr	Gly Thr Glu F 70	he Thr Leu	Thr lic Scr 75	r Ser Leu Gln Pro 80
Asp	Asp Phc	Ala Thr Tyr 1 85	fyr Cys Gln	Gln Tyr Asn 90	n ScrAsp ScrLys 95
МсL	Phc Gly	Gln Gly Thr L 100	ys Val Glu 105	Val Lys	

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 433 base pairs (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(i i) MOIECULE TYPE: DNA

(i i i) HYPOTHETICAL: NO

(I &) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 6.410

(D) OTHER INFORMATION: /procluct="Humanized anti-Tac beavy

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chain	variable	region.	Sog	ID.	19"	
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(x i) SEQUENCE DESCRIPTION: SEQ ID NO:18:

85

тстя		GA TO ly Ti				ac Lo					47
		CAC His									95
		G G C G I y									143
		A G C S c r 5 0									191
		Т G G Т г р									239
		AAG Lys									287
		GCC Ala									335
		ТАС Туг									383
		СТ G L с и 1 3 0			GGT	GAGT	ССТ	ΤΑΑΑ	АССТ	СT	430

AGA

433

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUE, NCE CHARACTERISTICS: (A) LENGTH: 135 amino acids (B) TYPE: amino acid (D) TOPOLOGY: bincar

(i i) MOLECULE TYPE: proucin

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:19:

M c L I	Gly Trp	Ser Trp Ile 5	Pbc Leu Pbc Leu Leu 10	Scr Gly Thr Ala Gly 15
Val	Hlis Scr	GID Val GIn 20	Leu Val Gin Ser Gly 25	Ala Glu Vəl Lys Lys 30
Pro	Gly Scr 35	Scr Val Lys	Val Scr Cys Lys Ala 40	Scr Gly Tyr Thr Phc 45
Thr	Scr Tyr 50	Arg Met His	Trp Val Arg Glu Ala SS	Pro Gly Gln Gly Lcu 60
G 1 u 6 5	Trp Ilc	Gly Tyr Ile 70	Asπ Pro Scr Tbr Gly 75	Tyr Thr Glu Tyr Asm 80
Gln	Lys Phc	Lys Asp Lys 85	Ala Thr llc Thr Ala 90	Asp Glu Scr Thr Asn 95
ТЪг	Ala Tyr	Mci Glu Leu 100	Ser Ser Leu Arg Ser 105	Glu Asp Thr Ala Val 110
Туг	Tyr Cys 115	Ala Arg Gly	Gly Gly Val Pbc Asp 120	Tyr Trp Gly Glu Gly 125
Tbr	Lcu Val 130	Thr Val Scr	S c r 1 3 5	

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(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGIH: 403 base pairs (B) TYPE: nucleic acid (C) SIRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

(i i i) HYPOTHETICAL: NO

- (i x) FEATURE:
 - (A) NAME/KEY: CDS (B) LOCATION: 6.383
 - (D) OTHER INFORMATION: /product="Humanized anti-Tac light
 - chain variable region: Seg ID. 21"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:20;

ТСТА				CC G/ hr A						rp Va							47
				ACC Tbr													95
				GTC Val 35												1	4 3
				T A C T y r					CAG					AAA		I	91
				A T T 1 I c				TCC					GGA			2	39
		TTC		G G C G 1 y		Gly	тст				Phc	ACC				2	87
	тст			CCA Pro	Asp					Туг					Arg	3	35
AGT				CTC Lcu					Gly					Val		3	83
CGT	AAGT	ACA	СТТТ	115 TCTA	GA				120					125		4	03

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACIERISTICS: (A) LENGTH: 126 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Mci I	GIU TH	hr Asp	Thr Leu 5	Leu Leu	Trp Val Lcu 10	Lcu Lcu '	Frp Val Pro 15
Gly	Scr Th	hr Gly 20	Asp IIc	Gin Mci	Thr Gln Scr 25	Pro Scr	Thr Leu Ser 30
Ala	ScrV:	al Gly 35	Asp Arg	Val Thr 40	Itc The Cys	Ser Ala : 45	Ser Ser Ser
Ilc	Scr Ty 50	yr Mici	Ніз Тгр	Tyr Glo 55	Gin Lys Pro	Gly Lys 50	Ala Pro Lys

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			-continue	d					
Leu Leu Ile 65	Tyr Thr Thr Sc 70	r Asol	Leu Ala	Sct 75	Gly Ve	1 Рто	Ala	Атд 80	
Phc Scr Gly	ScrGlyScrG 85	y Thr C	Glu Phc 90	ТЪг	Lcu Ti	nr Ilc	Sc 1 95	Scr	
Lou Gin Pro	Asp Asp Phc Al 100		Туг Туг 105	Cys	His G	n Arg 110		Thr	
Туг Рго Цен 115		n Giy 120	The Lys	V a 1		Lys 5			
(2) INFORMATION F	OR SEQ ID NO:22:								
	ENCE CHARACTERISTICS: A) LENGTH: 126 base pairs B) TYPE: nucleic acid C) STRANDEDNESS: single D) TOPOLOGY: linear								
(i i) MOLE	CULE TYPE: DNA								
(iii)HYPO	THETICAL: NO								
	 A) NAME/KEY: misc_feaure B) LOCATION: 1126 D) OTHER INFORMATION: /sta / note="One of four olig synthesize the humanized gene." 	onuclentides use l anti-Tae heavy	cd to						
	ENCE DESCRIPTION: SEQ ID NO								
AGCTTCTAGA	TGGGATGGAG CTG	GATCTTT	CTCTTC	CTCC	TGTCA	GGTAC	CGCG	GGCGTG	6 0
	TCCAGCTTGT CCAG	GTCTGGG	GCTGAA	GTCA	AGAAA	CCTGG	CTCG	AGCGTG	120
AAGGTC									126
(2) INFORMATION	OR SEQ ID NO:23:								
	ENCE CHARACTERISTICS; (A) LENGTH: 129 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear								
(i i) MOLI	ecule type: dna								
(iii)HYPC	THETICAL: NO								
	<pre>URE: (A) NA.ME/KEY: mi%_fcature (B) LOCATION: 1129 (D) OTHER INFORMATION://su / nous="One of four oily synthesize the humanized gene."</pre>	convolcosidos uso	cd La						
(xi)SEQU	ENCE DESCRIPTION: SEQ ID NO):23:							
CCCAGTCGAC	GGATTAATAT ATC	CAATCCA	TTCCAG	ACCC	TGTCC	AGGGG	ССТБ	CCTTAC	6 0
CCAGTGCATC	CTGTAGCTAG TAA	AGGTGTA	GCCAGA	AGCC	TTGCA	GGAGA	ССТТ	CACGCT	120
C G A G C C A G G									129
(2) INFORMATION	FOR SEQ ID NO:24:								
	ENCE CHARACIERISTICS: (A) LENGTH: 124 base pairs								

(A) LENGTH: 124 base pairs
 (A) LENGTH: 124 base pairs
 (B) TYPE: nucleix acid
 (C) STRAINDEDNESS: single
 (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

-continued (i i i) HYPOTHETTCAL: NO (i x) FEATURE: (A) NAME/KEY: misc_fcalure (B) LOCATION: 1..124 (D) OTHER INFORMATION: /standard_name="Ofigo HES14" / note"Ooc of four oligonucleotides used to synthesiae the humanized anti-Tae heavy chain genc." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:24: TATATTAATC CGTCGACTGG GTATACTGAA TACAATCAGA AGTTCAAGGA CAAGGCAACA 60 ATTACTGCAG ACGAATCCAC CAATACAGCC TACATGGAAC TGAGCAGCCT GAGATCTGAG 120 GACA 124 (2) INFORMATION FOR SEO ID NO:25: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 128 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE; DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: mise_feature (B) LOCATION: 1..128 (D) OTHER INFORMATION: /standard_name="Oligo HESI5" / note="One of four ofigonucleotides used to synthesize the humanized anti-Tac heavy chain gene." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:25: ATATCGTCTA GAGGTTTTAA GGACTCACCT GAGGAGACTG TGACCAGGGT TCCTTGGCCC 60 CAGTAGTCAA AGACCCCCCC CCCTCTTGCA CAGTAATAGA CTGCGGTGTC CTCAGATCTC 120 128 AGGCTGCT (2) INFORMATION FOR SEQ ID NO:26: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL. NO (i x) FEATURE: (A) NAME/KEY: misc_fcaure (B) LOCATION: 1.120 (D) OTHER INFORMATION: /standard_manne="Oligo JFDI" / noic="One of four eligenvelootides used to synthesize the humanized anti-Tec light chain gene." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:26; CAAATCTAGA TGGAGACCGA TACCCTCCTG CTATGGGTCC TCCTGCTATG GGTCCCAGGA 60 TCAACCGGAG ATATTCAGAT GACCCAGTCT CCATCTACCC TCTCTGCTAG CGTCGGGGAT 120 (2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTTI: 114 base pairs

93 94 -continued (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) JOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAMEREY: misc_feature (B) LOCATION: 1..114 () OTHER INFORMATION: /standard_non="Oligo JFD2" / notc="One of four oligonucleotides used to synthesize the humanized anti-Tac light chain gene." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:27: ATAAATTAGA AGCTTGGGAG CTTTGCCTGG CTTCTGCTGG TACCAGTGCA TGTAACTTAT 60 ACTTGAGCTG GCAGAGCAGG TTATGGTGAC CCTATCCCCG ACGCTAGCAG AGAG 114 (2) INFORMATION FOR SEQ ID NO:28: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 123 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: mise_feature (B) LOCATION: 1..123 (D) OTHER INFORMATION: /standard_name="O'igo IFD3" / note="One of four ol'igonucleotides used to symbosize the humanized anti-Tac light chain gene." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:28: GCTCCCAAGC TTCTAATTTA TACCACATCC AACCTGGCTT CTGGAGTCCC TGCTCGCTTC 60 AGTGGCAGTG GATCTGGGAC CGAGTTCACC CTCACAATCA GCTCTCTGCA GCCAGATGAT 120 TTC 123 (2) INFORMATION FOR SEQ ID NO:29: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 122 base pairs (B) TYPE: puckie acid (C) SIRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: misc_fcaunc (B) LOCATION: 1..122 (D) OTHER INFORMATION: /standard_name="Oligo JFD4" / note"One of four oligonucleotides used to synthesize the humanized anti-Tac light chain gcac." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:29: TATATCTAGA AAAGTGTACT TACGTTTGAC CTCCACCTTG GTCCCCTGAC CGAACGTGAG 60 TGGGTAAGTA CTCCTTTGAT GGCAGTAATA AOTGGCGAAA TCATCTGGCT GCAGAGAGCT 120

122

104 of 139

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(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 384 base pairs

- (B) TYPE: suchi c acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: cDNA

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAMEREY: CDS (B) LOCATION: 1..384
- (D) OTHER INFORMATION: /product="Light chain variable region of mik-betal: Seq ID No. 31"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:30.

							AGT Scr		48
						10.00	С С А Р г о 3 О		96
							A G T S c r		144
							G G A G 1 y		192
							G G A G 1 y		240
							C T C L c u		288
							CAG GIn JIO	-	336
							GAG Glu		384

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 128 amino acids (B) TYPE: amino acid (D) TOPOLOGY: Incar

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION; SEQ ID NO:31:

Мсі 1	Азр Рыс	Gin Val Gl 5	n Ilc Phc Scr	Phe Lev Lev I 10	lc Ser Ala Scr 15
V a J	Ilc Lcu	Scr Arg Gl 20	y Gin lic Val 25		cr Pro Ala ile 30
Mcl	Scr Ala 35	Scr Pro Gl	y Glu Lys Val 40	Thr Met Thr C	уз Scт Gly Scт 45
Sсг	Scr Val 50	Ser Phe Me	t Tyr Trp Tyr 55	Gln Gln Arg P 60	ro Gly Ser Scr
Pr q 65	Arg Leu		r Asp Thr Scr O	Азл Lcu Ala S 75	cr Gly Val Pro 80

96

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Val	Arg	Ple	Scr	G 1 y 8 5	Scr	Gly	Scr	Gly	Thr 90	Ser	Туг	Scr	Lcu	ТЪг 95	11 c
Scr	Arg	Μcι	G I u 1 0 0	Ala	Glu	Asp	ΛIЩ	A I a 1 0 5	Трг	Туґ	Туг	Cys	G 1 15 1 1 0	GIR	Trp
Scr	Тbг	Туг 115	Pro	Lcu	ТЪг	Pbc	Gly 120	Ala	Gly	Tbr	Lys	L c u 1 2 5	Glu	Lcu	Lys

(2) INFORMATION FORSEQID NO:32:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 414 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS; single (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: CONA

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1.414 (D) THER INFORMATION: /product="Hcavy chain var. region of the anibody mik-beual: Seq1D 33"

(* i) SEQUENCE DESCRIPTION: SEQ ID NO:32:

							CCA Pro			48
							CTA Lcu 30			96
							TTC Phc			144
							AAG Lys			192
							ТАТ Туг			240
							AAG Lys		,	288
							G C C A 1 a 1 1 0			336
							GCT AI a			384
				TCT Scr						414

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISITCS: (A) LENGTH: 138 amino acids (B) TYPE; amino acid (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: prokin

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Mei Ala Val Leu Gly Leu Leu Phe Cys Leu Val Thr Phe Pro Ser Cys 1 5 10 15

5,530,101

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Val	Lcu	Scr	G 1 n 2 0	Val	Gln	Ĺcu	Lys G	1 ¤ 2 5	Scr (ЗІу	Pro	Glyl	Ссц V 30	al GID
Pro	Scr	G I ¤ 3 5	Scr	Lcu	Scr	1 1 c	Tbr C 40	y s	Thr V	/ æ 1	Scr	G1y 1 45	Phc S	cr Val
ТЬг	S c r 5 0	Туг	Gly	Val	His	Trp 55	Ilc A	7 B.	Gln S	Scr	P r D 6 0	Gly 1	Lys G	ly Lcu
G I u 6 5	Тгр	Lcu	G 1 y	Val	11c 70	Тгр	Scr G	1 y	Gly S	5 ст 7 5	Thr	Asp	Гуг А	sn Ala 8
Ala	Рһс	1 c	Scr	Arg 85	Lcu	ТЬг	llc S	r	Lys A 90	Asp	Asn	Scr I		crGln 95
/ ə l	Phc	Phe	L y s 100	Val	Asn	Scr	Leu G	1 n 0 5	Prq A	Ala	Asp		Ala I IJO	lc Tyr
yr	Cys	A I a 115	Arg	Ala	Gly	Asp	Tyr A l20	S N	Tyr /	Asp		Phc /	Ala T	уг Тгр
Gly	G I л I 3 0	Gly	Thr	Lcu	Val	Thr 135	Val S	Cr	Ala					

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACIERISTICS:

(A) LENGIE : 107 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

99

(D) TOPOLOGY: unicoown

(i i) MOLECULE TYPE: probin

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

(A) NAME/KEY: Protein (B) LOCATION: 1..107

(D) OTHER ENFORMATION: /note="Amino acid sequence of the light chain for humanc Lay antibody."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Asp 1	11с СІл	Mct Thr Gia 5	Scr Pro Scr	Scr Leu Scr 10	Val Ser Val Gly 15
Asp	Arg Vel	Thr Ile Thr 20	Cys Gln Ala 25	Ser Gln Asn	Val Asn Ala Tyr 30
Leu	Asn Trp 35	Tyr Gin Gin	Lys Pro Oly 40	Lcu Ala Pro	Lys Leu Leu lle 45
Туг	Gly Ala 50	Scr Thr Arg	Glu Ala Gly 55	Val Pro Scr 60	Arg Phe Ser Gly
Scr €5	Gly Scr	Gly Thr Asp 70	Phc Thr Phc	Thr Ilc Ser 75	Scr Lcu Gla Pro 80
Glu	Asp IIc	Alə Thr Tyr 85	Tyr Cys Gln	Gln Tyr Asn 90	Asn Trp Pro Pro 95
Thr	Phc Gly	Gln Gly Thr 100	Lys Val Glu 105	Val Lys	

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS: (A) IENGTH: 106 amino acids

(B) TYPE: antino acid

(C) STRANDEDNESS: single (D) TOPOLOOY: unknown

(i i) MOLECULE TYPE: protein

(i i i) HYPOTHETICAL: NO

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(i x) FEATURE:

(A) NAME/KEY: Protein

(B) LOCATION: 1_106

(D) OTHER INFORMATION: /note="Amima acid sequence of the

light chain of the humanized mik-betal antibody."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:35:

AspIIcGInMciThrGInScrProScrScrLeuScrAlaScrValGlyAspArgValThrDrThrCysScrGlyScrScrScrScrValScrValScrPhcMcLTyrTrpTyrGInGInGInLysProGlyLysAlaProLysLeuIcuIleTyrAspTyrScrAsnLcuAlaScrGlyLysAlaProLysLeuIleTyrAspThrScrGlyThrAsnLcuAlaScrGlyValProScrArgPhcScrGlyScrGlyScrGlyThrAspTyrTyrThrPhcScrArgPhcScrGlyScrGlyScrGlyThrAspTyrTyrThrPhcScrScrArgPhcScrGlyScrGlyScrGlyThrAspTyrTyrPhcThrPhcScrScrLeuGlyScrGlyScrGlyScrGlyThrAspTyrCysGluGluTyrScrScrLeuGlnProScrGlyScrGlyThrTyrCysGluGluTyrScrThrTyrProScr</t

(2) INFORMATION FOR SEQ ID NO:36:

(i) SEQUENCE CHARACIERISTICS:

- (A) LENGTH: 122 amino acids
 - (B) TYPE; amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: unknown

(i i) MOLECULE TYPE: protein

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

(A) NAME/KEY: Protein

(B) LOCATION: 1..122

(D) OTHER INFORMATION: /not="Amino acid sequence of the

heavy chain of the human Lay antibody."

(* i) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Ala 1	Val (31 թ	Lcu	LCU S	GIu	Scr	Gly	Giy	Giy L 10	cu	Val G	ln Pr	Gly IS	Gły
Ser	Lcu /	Arg	L c u 2 0	Scr	Cys	Ala	Ala	S c r 2 5	Gly F	° h c	Thr P	hc Sc 30	r Ala	Scr
Ala		Scr 35	Тгр	Val	Arg	Glu	A 1 a 4 0	Pro	Gly L	у 5	-	cu G1 5	и Тгр	Val
Ala	Тгр 1 50	Lyş	Туг	Glu	Asn	GIy 55	Aso	Asp	Lys F		Туг А 60	la As	p Scr	Val
A s n 6 5	Gly	Arg	Рас	ТЪг	1 L C 7 0	Scr	Arg	Asn		Scr 75	Lys A	sn Th	r Lcu	Туг 80
Lcu	GIDI	Mct		G I y 8 5	Lcu	Gln	Ala	Glx	Val 5 90	бст	Ala 1	Іс Ту	r Tyr 95	Cys
Ala	Arg	Asp	A I a 1 0 0	Gly	210	Tyr	Val	S c r 1 0 5	Pro 7	ն հ г	Рьс Р	hс А1 11	a His O	Trp
O 1 y	Gln	G I y 1 1 5	ТЬс	Lcu	Val	ТЪг	V a 1 1 2 0	Scr	Scr					

(2) INFORMATION FOR SEQ ID NO:37:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 119 smino acids (B) TYPE: animo acid

104

-continued

(C) STRANDEDNESS: single (D) TOPOLOGY: unknown

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( i i ) MOLECULE TYPE: protein
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(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

(A) NAME/KEY: Protein (B) LOCATION: 1..119

- (D) OTHER INFORMATION: /nou="Amino acid sequence of the heavy chain of the humanized mik-betal antibody."

(x i) SEQUENCE DESCRIPTION: SEQ 1D ND:37:

ցլա 1	V a I	Gln	Lcu	Lcu S	Glu	Scr	G 1 y	Gly	G 1 y 1 0	Lcu	Val	Gln	Pro	G 1 y 1 5	Gly
Scr	Lcu	Arg	L c u 2 0	Scr	C y s	Ala	Ala	Scr 25	Gly	Phc	Τhr	V a l	Thr 30	Sсг	Туг
Gly	Val	Ні в 35	Trp	V a I	Arg	Glɑ	Ala 40	Pro	G 1 y	Lys	Gly	L c u 4 5	Glu	Ттр	Val
Gly	V a 1 5 0	11c	Ϋ́r p	Scr	G 1 y	G 1 y 5 5	Scr	Thr	Asp	Туг	Asn 60	Ala	Ala	Ρhο	110
Ser 65	Arg	Phc	Thr	1 l c	Sсг 70	Arg	Asp	Asn	Scr	L y s 7 5	Asn	Thr	Lcu	Туг	L C U 8 O
Gln	Μcι	Asn	Scr	L C U 8 5	Gln	Ala	Glu	Asp	Т h г 90	Ala	11c	Туг	Туг	C y s 9 5	Ala
Arg	AIa	Gly	Asp 100	Tyr	Asn	Туг	Asp	G 1 y 1 0 5	Рbс	Ala	Тут	Тгр	G1 y 1] 0	Gln	G 1 y
Thr	Lcu	V a 1] I 5	Thr	Val	Sсг	Scr									

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACIERISTICS: (A) LEANGTH: 107 base pairs

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1.107
 - (D) OTHER INFORMATION: /standard_panc="Oligo vel3"

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:38:

TTCTGCTGGT	ACCAGTACAT	GAAACTTACA	CTTGAGCTGC	CACTGCAGGT	GATGGTGACG	6 0
CGGTCACCCA	CTGAGGCACT	GAGGCTAGAT	GGAGACTGGG	TCATTTG		107

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 136 base pairs

- (B) TYPE: nucleic acid (C) STRANDEDN'ESS: single
- (D) TOPOLOGY: linea:

(i i) MOLECULE TYPE: DNA

(i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: misc_fcature
- (B) LOCATION: 1...136
- (D) OTHER INFORMATION: /standard_name="Oligu vel4"

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5,5	JU	,,1	UI

		-	,550,101			
	105		-continued	1	06	
(xi)SEQU	ENCE DESCRIPTION: SI	EQ ID NO:39:				
CATGTACTGG	TACCAGCAGA	AGCCAGGAAA	AGCTCCGAAA	CTTCTGATTT	ATGACACATC	6 0
CAACCTGGCT	TCTGGAGTCC	CTTCCCGCTT	CAGTGGCAGT	GGGTCTOGGA	CCGATTACAC	120
CTTTACAATC	TCTTCA					136
(2) INFORMATION	FOR SEQ ID NO:40:					
	IENCE CHARACTERISTI (A) LENGTH: 137 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA					
(iii)HYP(OTHETICAL: NO					
	TURE: (A) NAME/KEY: mix_ (B) LOCATION; 1137 (D) OTHER INFORMA JENCE DESCRIPTION: SI	MON: /standard_damc="C	Digo vol5''			
			CTCGACCTTG			6 0
		GGCAGTAATA	AGTGGCTATA	TCTTCCGGCT	GAAGTGAAGA	120
GATTGTAAAG	GIGIAAI					137
	 (A) LENGTH: 108 basc (B) TYPE: nucleic acid (C) SIRANDEDNESS: (D) TOPOLOGY: linear ECULE TYPE: DNA 	single				
	UTHETICAL: NO					
	(A) NAME/KEY: misc_ (B) LGCATION; 1108 (D) OTHER INFORMA		Dligo vel6"			
(xi)SEQU	JENCE DESCRIPTION: S	EQ 1D NO:41:				
CACATCTAGA	CCACCATGGA	TTTTCAAGTG	CAGATCTTCA	GCTTCCTGCT	AATCAGTGCC	6 (
TCAGTCATAC	TGTCCAGAGG	AGATATTCAA	ATGACCCAGT	CTCCATCT		10
(2) INFORMATION	FOR SEQ ID NO:42:					
	UENCE CHARACTERIST (A) LENGTH: 138 base (B) TYPE: sucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: finest	single				
(ii) MOL	ECULE TYPE: DNA					
(iii)(FYP	OTHEDCAL: NO					
	(A) NAMERKEY: misc. (B) LOCATION: 1138		Oligo vell'			
(xi)SEQ	LIENCE DESCRIPTION: S	EQ ID NO:42;				
TAGTCTOTCG	ACCCACCACT	CCATATCACT	CCCACCCACT	CGAGTCCCTT	TCCAGGAGCC	6

			5,530,101			
n	107		-continued		108	
TGGCGGACCC	AGTGTACACC	ΛΤΑΑCTTGTT	ACGGTGAAAC	CACTGGCGGC	ACAAGACAGT	120
C T C Λ G Α G Λ T C	CTCCTGGC					138
(2) INFORMATION F	OR SEQ ID NO:43:					
	ENCE CHARACTERISTI((A) LENGTH: 126 base] (B) TYPE: nucleic acid (C) STRANDEDNESS: s (D) TOPOLOGY: linear	pairs				
(ii)MOLI	CULE TYPE: DNA					
(iii)HYPO	THETICAL NO					
(URE: (A) NAME/KBY: misc_f (B) LOCATION: 1,126 (D) OTHER INFORMAT		Ligo vol2"			
(xi)SEQU	ENCE DESCRIPTION: SE	Q ID NO:43:				
TGGTGCGTCG	ΑСΛGΛCΤΑΤΑ	ATGCAGCTTT	C A T A T C C A G A	ΤΤΤΑΟΟΑΤΟΛ	GCAGAGACAA	6 0
CAGCAAGAAC	ACACTGTATC	TCCAAA1'GAA	TAGCCTGCAA	GCCGAGGACA	CAGCCATATA	120
ΤΤΑΤΤΟ						126
(2) INFORMATION I	FOR SEQ ID NO:44;					
	ENCE CHARACIERISTI (A) LENGIH: 130 base (B) TYPE: nucleic acid (C) STRANDEDNESS: s (D) TOPOLOGY: Jincar	pairs				
(ii) MOLE	ECULE TYPE: DNA					
(iii)HYPO	THETICAL: NO		<i>x</i>			
	URE: (A) NAME/MBY: misc_1 (B) LOCATION: 1130 (D) OTHER INFORMAT		lligo ₩ps54"			
(xi)SEQU	ENCE DESCRIPTION: SE	Q ID NO:44.				
ΛΟΛΟΤΟΤΛΟΑ	CCACCATGGC	TGTCTTGGGG	CTGCTCTTCT	GCCTGGTGAC	ATTCCCAAGC	60
TGTGTCCTAT	CCGCTGTCCA	GCTGCTAGAG	AGTGGTGGCG	GTCTGGTGCA	G C C A G G A G G A	120
TCTCTGAGAC						130
(2) INFORMATION	FOR SEQ ID NO:45:					
	ENCE CHARACIERISTI (A) LENGTH: 118 base ((B) TYPE: puckele actid (C) S'IRANDEDNESS: s (D) TOPOLOGY: Tincar	Pairs				
(ii) MOL	ECULE TYPE: DNA					
(iii)HYPC	THETICAL: NO					
(i x) FEAT	URE: (A) NAME/KEY: misc_(B) LOCATION: 1118		Dligo wps57"			
(i x) FEAT	URE: (A) NAME/KEY: misc_1	TON: /standard_rame="C	Dligo wps57"			
(i x) FEAT (x i) SEQU	URE: (A) NAME/KEY: misc (B) LOCATION: 1118 (D) OTHER INFORMAT	"ION: /standard_rame="C EQ ID NO:45:		AGAGTCCCTT	GGCCCCAGTA	6 0

5,530,101

110

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(2) INF ORMATION FOR SEQ ID NO.46:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 111 amino acids

- (B) TYPE: amino acid
- (C) SIRANDEDNESS: single (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: poplide

(x i) SEQUENCE DESCRIPTION: SEQ 1D NO:46:

Asp 1	lic Val	Leu Thr Gla S		cr Lcu Ala Val O	Scr Leu Gly 15
Gln	Arg Ala	Thr llc Scr 20	Cys Arg Ala S 25	cr Gin Scr Val	Scr Thr Scr 30
Thr	Tyr Aso 35	Tyr Mci His	Trp Tyr Gla G 40	la Lys Pro Giy 45	Gln Pro Pro
L y s	Lcu Lcu 50	lle Lys Tyr	Ala Ser Asn L 55	cu Glu Scr Gly 60	Val Pro Ala
Arg 65	Phc Scr	Gly Scr Gly 70	Phe Gly Thr A	sp Phc Thr Lcu 75	Азя llc His 80
Pro	Val GIU	Glu Glu Asp 85		'yr Tyr Cys Glm O	His Ser Trp 95
Glu	lic Pro	Tyr Thr Phc 100	Gly Gly Gly T 105	hr Lys Leu Glu	IIC Lys IIO

(2) INFORMATION FOR SEQ ID NO:47:

(i) SEQUENCE CHARACIERISTICS: (A) LENGTH: 111 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO.47.

Glulle Val Met Thr Gln Ser Pro Ala Thr Leu Ser Vai Ser Pro Gly 1 5 10 15 Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gln Ser Val Ser Thr Ser 20 25 30 Thr Tyr Asn Tyr Mei His Trp Tyr Gln Gln Lys Pro Gly Gln Scr Pro 35 40 45 Arg Leu leu lle Lys Tyr Ala Ser Asn Leu Glu Ser Gly Ile Pro Ala 50 55 60 Arg Phe Ser Gly Ser Gly Thr Glu Phe Thr Leu Thr Ile Ser65707580 Arg Leu Glu Ser Glu Asp Phe Ala Val Tyr Tyr Cys Glu llis Ser Trp 85 - 90 95 Glu IIc Pro Tyr Thr Phc Gly Gln Gly Thr Arg Val Glu IIc Lys 100 105 110

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BI Exhibit 1136

(2) INFORMATION FOR SEQ ID NO:48:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTI-1: 122 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(i i) MOLECULE TYPE: pepuide

			111				-con	tinued				_			
(xi)5	EQUENC	E DESCA	UPTION:	SEQ ID	NO:48:										
G 1 u 1	Mcı	11 c	Lcu	Val 5	Glu	Ser	Gly	Gly	G I y 1 0	Lcu	Val	Lys	Pro	G 1 y 1 5	Ala
Scr	Lcu	Lys	<u>L с и</u> 2 0	Scr	Cys	Ala	Ala	<mark>Sсг</mark> 25	Gly	Рьс	Tbr	Phe	S c r 3 0	Asø	Туг
G 1 y	Leu	S c r 3 5	Ттр	Val	Arg	Gla	Thr 40	Scr	Asp	Arg	Arg	<u>і</u> си 45	Glu	Тгр	Val
Ala	Sст 50	llc	Scr	Arg	G 1 y	Gly 55	GIy	Агд	1 i c	Tyr	Sст 60	Ρτο.	Asp	Asn	Lcu
L y s 6 5	Gly	Агд	Phc	Thr	[] c 7 0	Scr	Arg	Glu	Asp	Ala 75	Lys	Asn	Thr	Lcu	Туг 80
Leu	Gln	Мсі	Scr	S c r 8 5	Lcu	Lys	Scr	Glu	Asp 90	Thr	Ala	Lcu	Туг	Туг 95	Суs
Lcu	Arg	Glu	G 1 y 1 0 0	llc	Туг	Туг	Ala	А <i>в р</i> 105	Тут	Gly	Phc	Phc	Asp 110	Val	Ттр
Gly	Thr	G 1 y 1 1 5	Thr	Τhr	Val	1 1 c	V a 1 1 2 0	Scr	Scr						
) INFORMAT	SEQUENC (A (B (C	E CHAR) LENGTI) TYPE: 2) STRAN	ACIERIS H: 122 an mino acu DEDNES	nin o acids } S: single											
(ii)) TOPOLO LE TYPE		r											
(xi)	EQUEN	E DESCI	RIPLION:	SEQ ID	NO:49:										
GIU I	Val	Glw	Lcu	Leu 5	Giu	Ser	Gly	Gly	G l y 10	Lcu	Val	Glı	Pro	G I y 1 5	Gly
Scr	Lcu	Arg	L c u 2 0	Scr	Cys	Ala	Ala	Ser 25	Gly	Phc	Thr	РЬс	Sсг 30	Asn	Tyr
Gly	Lcu	S c r 3 5	Тгр	Val	Arg	Gla	A] a 4 0	Pro	Gly	Lys	G 1 y	L c u 4 5	Glu	Тгр	Val
Ala	S c r 5 0	Ilc	Scr	Arg	Gly	G 1 y S 5	Gly	Arg	llc	Tyr	Sст 60	Pro	Asp	Ası	Lcu
L y s 6 5	Gly	Агд	Phe	Thr	11c 70	Scr	Аrg	Asn	Asp	Ser 75	Lys	A s n	Thr	Lcu	Туг 80
Lcu	Gln	Mcl	Asn	Scr 85	Lcu	Gln	Ala	Głu	Asp 90	Τμτ	Ala	Lcu	Туг	Туг 95	Суs
Lcu	Arg	GΙu	G I y 1 0 0	110	Туг	Туг	Ala	Asp 105	Tyr	G 1 y	Phc	Phc	Asp 110		
G 1 y	Gln	G I y 1 1 5	Thr	Lcu	Val	Tbr	Val 120	Scr	Sст						
) INFORMAT	ION FOR	SEQ ID	NO:50:												
(i):	(A	CE CHAR) LENGT		nino acids											

- (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: populde

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:50:

```
Asp lle Val Mei Thr Gin Ser His Lys Phr Mei Ser Thr Ser Val Gly
I 5 10 15
Asp Arg Val Ser Ile Thr Cys Lys Ala Ser Gln Asp Val Gly Ser Ala
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5,53	20	101
J,J.	ω,	101

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	<u>.</u>	-

			2 0					25					30		
V a 1	Val	Trp 35	His	Gln	Gln	Lys	Scr 40	GIy	GIn	Ser	Pro	L y s 4 5	Lcu	Lcu	llc
Tyr	T r p 5 0	Ala	Scr	Tbr	Arg	H i s 5 5	Thr	Gly	Val	Pro	Asp 60	Arg	Phc	Tbr	Gly
Sет 65	Gly	Scr	GІу	Тһг	Asp 70	Phc	Thr	Lси	Tbr	11c 75	Тbг	Asn	Val	Gin	Sст 80
Glu	Asp	Lcu	Ala	A s p 8 5	Туг	Fhc	Суs	Gln	Gіп 90	Туг	Sсг	i l c	Phe	Pro 95	Lcu
Thr	Phc	G 1 y	A I a 1 0 0	Gly	Tbr	Arg	Lcu	GIu IOS	Lcu	Lys					

(2) INFORMATION FOR SEQ ID NO:51:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 107 amino acids

- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: luncar
- (i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO.51:

Asp
111cG1nMciThr
5G1nSerProSerThr
10LeuSerAlaSerVal
15G1y
15AspArgValThr
20IicThrCysLysAla
25SerG1nAspValG1y
30SerAlaValValTrp
35HisG1nG1nCysLysAla
25SerG1nAspValG1y
30SerAlaValYalTrp
35HisG1nG1nCysLysAla
40SerG1nAspValLeuLeuIcuYalTrp
35AlaSerTbrArgPro
55G1yLysAlaPro
40LysLeuLeuIcuIcuIcuYalSerGly
35SerTbrArgPro
55G1yValPro
60SerArgPhcTbrG1ySerG1y
50SerGly
70ThrG1u
70PhcThrLeuThrIcuThrFro
60SerSerLeuG1n
80Pro
80AspAspPhc
80Ala
70ThrLys
70PhcG1n
75G1n
75SerIcu
80Icu
80Pro
80SerLeu
95AspPhc
80G1y
700PhrLys
85Val
80G1u
80G1u
80Val
80LysVal<

(2) INFORMATION FOR SEQ ID NO:52:

(i) SEQUEINCE CHARACTERISTICS:

(A) LENGTH: 121 antito acds
(B) TYPE: antito acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(i i) MOLECULE TYPE: pepuide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO 52:

GlnValGlnLeuGlnGlnSerAspAlaGluLeuValLysProGlyAla11Lys1CSerCysLysValSerGlyTyrThrProGlyLysProSerValLys1CSerCysLysValSerGlyTyrThrProThrProGlyLysAspHisThr1HisTrpMetLysGlnArgProGluGlnGlyLeuGluTrpPheGlyTyr1CTyrProArgAspGlyHisThrArgGlyLysPheGlyTyr1CTyrProArgAspGlyHisThrArgGlyLysPheGlyTyr1CThrLeuThrAlzAspLysSerAlaSerPheGlyTyr1CThrLeuThrAlzAspLysSerAlaSerAlaTyrGlyTyrLysAlaThrLeuThrAlzAspLysSerAlaSerAlaTyrGlyLysAlaThrLeuThrAlzAspLysSerAlaSerThrAlaTyrGlyLysAlaThrLeuThrAl

			115									1	16		
							-con	tinued							
				8 5					90					95	
Ala	Arg	Gly	Arg 100	Asp	Scr	Arg	Glu	Arg 105	Asn	Gly	Phc	Ala	Туг 110	Тгр	G 1 y
Gln	Gly	Thr 115	Lcu	Val	Tbr	Val	Scr 120	Ala							
INFORMAT	ION FOR	SEQ ID 1	NO:53:												
(i)\$	(B (C) LENGT) TYPE: :) STRAN	ACTERIS H: 121 an imino ació DEDNES OGY: Inc	níno acids) S: single											
(ii)	MOLECU	le type	peplide												
(xi)5	SEQUENC	CE DESC	RIFIION:	SEQ ID	NO:53:										
G 1 n 1	V a]	Gln	Lcu	V a 1 5	Gln	Sсг	Gly	Ala	G 1 u 1 0	V a 1	Ĺys	Lys	Рго	G l y 1 5	Scr
Sсг	Val	Lys	V a 1 2 0	Scr	Cys	Lys	Ala	<u>Sс</u> г 25	Gly	Туг	ТЪг	Phc	т h r 3 0	Asp	H i s
Thr	110	His 35	Тгр	Mci	Arg	Glı	A 1 a 4 0	Pro	G 1 y	Gln	Gly	L C U 4 5	Glu	Тгр	Рьс
Gly	Туг 50	I I c	Tyr	Pro	Arg	Asp 55	Gly	His	Thr	Arg	Tyr 60	Scr	Glu	Ľуş	Phc
Lys 65	Gly	Lys	Ala	Thr	1 1 c 7 0	Thr	Ala	Asp	Glu	S c r 7 5	Thr	Asn	Thr	Ala	Туг 80
Mct	Glu	Lcu	Sст	Sст 85	Lcu	Arg	Scr	Glu	A s p 9 0	Thr	Ala	Val	Туг	Phc 95	Cys
Ala	Arg	G 1 y	Arg 100	Asp	Scr	Αrg	Glu	Arg 105	Ası	Gly	Phc	Ala	Туг 110	Тгр	Gly
Gli	G 1 y	Thr 115	Lсц	Val	Thr	V a 1	Sсг 120	Sст							
INFORMAI	ION FOR	SEQ ID	NO:54:												
(i):	(B (C) LENGT) TYPE: :) STRAN	ACTERIS H: 111 an amino ació DEDNES OGY: linc	nino acids d S: single											

(i i) MOLECULE TYPE: populdo

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:54:

Asp 1	llc Val	Lcu Thr S	Gln Scr	Pro Ala	Scr Lcu 10	Ala Val	ScrLcuGly 15
Gln	Arg Ala	Thr Ilc 20	Scr Cys	Arg Ala 25	Scr Glu	Ser Val	Asp Asn Tyr 30
G 1 y	lle Ser 35	Phe Mcı	Asn Trp	Phc Gla 40	Gln Lys	Pro Gly 45	GIn Pro Pro
Lys	Leu Leu 50	і і с Туг	Ala Ala 55	Scr Азп	Gln Gly	Scr Gly 60	Val Pro Ala
Ar 5 6 5	Phc Scr	Gly Scr	Gly Scr 70	Gly Thr	Asp Phc 75	Scr Lcu	Asılle His 80
Pro	Мск Сін	Glu Asp 85	Asp Thr	Ala Mei	Tyr Phe 90	Cys Gln	Glos Scr Lys 95
Glu	Val Pro	Trp Thr 100	Phc Gly	G 1 y G 1 y 1 0 5	Thr Lys	Lcu Glu	Ilc Lys 110

118

(2) INFORMATION FOR SEQ ID NO:55:

(i) SEQUENCE CHARACTERISTICS:

(A) LE.NGTH: 111 amino acids

(B) TYPE: amino acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide

(x i) SEQUE, NCE DESCRIPTION: SEQ ID NO:55:

Asp
1IIcGInMciThr
SGInScrProScrScrLeuScrAlaScrVal
ISGIyAspArgValTbr10TbrCysArgAlaScrGIuScrValAspAsnTyrGlyIIcSerPbcMclAsnTrpPhcGInGInLysScrValAspAsnTyrGlyIIcSerPbcMclAsnTrpPhcGInGInLysProGlyLysAlaProLysLeuIcuIIcTyrAIaAlaScrAsnGInGlyScrGlyValProLysLeuIcuIIcTyrAIaAlaScrAsnGInGlyScrGlyValProScrLeuIcuIIcTyrAIaAlaScrAsnGInGlyScrGlyValProScrLeuIcuIIcTyrAIaAlaScrAsnGInGlyScrGlyValProScrScrLcuGInScrGlyScrGlyThrAspPbcThrAspPccScr

(2) INFORMATION FOR SEQ ID NO:56:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 116 amino acids (B) TYPE: amino acid

- (C) STRANDEDN'ESS: single (D) 10POLOGY: linear
- (i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:56:

GluValGlnLeuGlnGlnScrGlyProGluLeuValLysProGlyAla11LysLysLicSerCysLysAlaSerGlyTyrThrPheThrAspTyrAsnMciHisTrpValLysGloScrHisGlyLysScrLeuGluTrpIleGlyTyrJrrHisTrpValLysGloScrHisGlyLysScrLeuGluTrpIleGlyTyrJrrIleTyrProTyrAsnGlyGlyTyrGlyTyrAshGluTrpIleGlyTyrIleTyrProTyrAsnGlyGlyThrGlyTyrAshGluTrpIleGlyTyrIleTyrProTyrAsnGlyGlyThrGlyTyrAshGluTrpIleGlyTyrIleTyrProTyrAsnGlyGlyTyrAshGluTyrAshScrScrLysAshCluTyrProGlyTyrAlaThrLysAsnAsnScrScrScrScrThrAlaTyrScrLysScrLysAlaThrLouThrScrGluAsnScrScr<

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(2) INFORMATION FOR SEQ ID NO:57:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 116 amino acids
 (B) TYPE amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

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	119	-contrinued	120
		-contribution	
(xi) SEQUE	NCE DESCRIPTION: SEQ ID NO:57:		
Glo Va 1	i Gln 1.cu Val Glo 5	Ser Giy Ala Giu Va 10	l Lys Lys Pro Gly Sc 15
Scr Va	l Lys Val Scr Cys 20	Lys Ala Ser Gly Ty 25	r Thr Phe Thr Asp Ty 30
Asn Mc	l His Trp Val Arg 35	Gln Ala Pro Gly Gl 40	n Gly Leu Glu Trp 11 45
Gly Ty 50	т Ile Туг Рго Туг	Asn Gly Gly Thr Gl 55	y Tyr Asn Gin Lys Ph 60
1. ys 5 c 6 5	r Lys Ala Thr Ilc 70	Thr Ala Asp Glu Se 75	r Thr Asn Thr Ala Ty 80
Mei Gl	u Leu Ser Ser Leu 85	Arg Ser Glu Asp Th 90	r Ala Val Tyr Tyr Cy 95
Ala Ar;	g Gly Arg Pro Ala 100	Mel Asp Tyr Trp GI 105	y Gin Gly Thr Leu Va J10
Thr Va	l Ser Ser II5		
(1	NCE CHARACTERISTICS: A) LENGTH: 106 amino acids B) TYPE: amino acid C) STRANDEDNESS: single D) TOPOLOGY: bincor		
(i i) MOLEC	CULE TYPE: popuide		
(xi)SEQUE	NCE DESCRIPTION: SEQ ID NO:58		
		Ser Pro Ala Ile Me	L Scr Ala Scr Pro Gl
Gln Ila 1	5	1 D	I 5
		l0 CysSerGlySerSe 25	15
1	s Val Thr Mcı Thr 20	Cys Ser Gly Ser Se 25	15 1 Ser Val Ser Phec Me
l Glu Ly Tyr Trj	s Val Thr Mcl Thr 20 p Tyr Gla Gln Arg	Cys Ser Gly Ser Se 25 Pro Gly Ser Ser Pr	IS r Ser Val Ser Phe Me 30 o Arg Leu Leu lle Ty 45
l Glu Ly Tyr Trj Asp Th 50	s Val Thr Mcı Thr 20 p Tyr Gla Gln Arg 35 r Ser Asa Leu Ala	Cys Ser Gly Ser Se 25 Pro Gly Ser Scr Pr 40 Ser Gly Val Pro Va 55	IS r Ser Val Ser Phe Me 30 o Arg Leu Leu lle Ty 45 I Arg Phe Ser Gly Se
l GluLy TyrTrj AspThi 50 GlySc 65	s Val Thr Mcl Thr 20 p Tyr Gla Gln Arg 35 r Ser Asa Leu Ala r Gly Thr Ser Tyr 70	Cys Ser Gly Ser Se 25 Pro Gly Ser Ser Pr 40 Ser Gly Val Pro Va Ser Leu Thr ile Se 75	15 r Ser Val Ser Phic Me 30 o Arg Leu Leu lle Ty 45 l Arg Phe Ser Gly Se 60 r Arg Mei Glu Ala Gl

(2) INFORMATION FOR SEQ ID NO:59.

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 106 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: slogle (D) TOPOLOGY: lincer (i i) MOLECUIE TYPE: pepide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:59:

Asp lie Giv Mei Thr Gin Ser Pro Ser Ser Leu Ser Aia Ser Val Gly 1 5 10 15 Asp Arg Val Thr IIc Thr Cys Ser Gly Ser Ser Ser Val Ser Phe Mei 20 25 30

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Туг	Тгр	Туг 35	GΙπ	Gla	Lys	Ρτυ	G I y 4 0	Lys	Ala	Ρτο	Lys	Lси 45	Lcu	llc	Туг
Asp	Т b т 50	Scr	Абл	Lcu	Ala	S c r 5 5	Gly	Val	Pro	Scr	Arg 60	Phc	Scr	Gly	Scr
G I y 6 5	Scr	G 1 y	Thr	Asp	Туг 70	Thr	РЬс	Thr	11c	Scr 75	Scr	Lcu	GIn	Pro	G I u 8 0
Asp	110	Ala	Thr	Туг 85	Туг	Cys	Glu	Glɒ	Т г р 90	Scr	Tbr	Туг	Pro	Lси 95	Thr
Рbс	Gly	Gin	G l y 100	Tbr	Lys	Val	Glu	V a l 105	Lys						

(2) INFORMATION FOR SEQ ID NO:60.

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 119 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: funcar

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(i i) MOLECULE TYPE: populde

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:60:

 G1n
 Va1
 G1n
 Leu
 Lys
 G1n
 Ser
 G1y
 Pro
 G1y
 Leu
 Va1
 G1n
 Pro
 Ser
 G1y
 Pro
 G1n
 Leu
 Va1
 Ser
 G1y
 Pro
 G1y
 Leu
 Va1
 G1n
 Pro
 Ser
 G1n
 Pro
 Ser
 G1y
 Pro
 G1n
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 Ser
 Tyr
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 G1n
 Ser
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(2) INFORMATION FOR SEQ ID NO.61:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGIH: 119 summo acids
 (B) TYPE: amino acid
 (C) STRANDEDN'ESS: single
 (D) TOPOLOGY: linear

(1 i) MOLECULE TYPE: pepude

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:61:

GIu i	Val	Gln Leu	Lcu Glu S	Scr Gly (Gly Gly Leu 10	Val Glu	Pro Gly Gly 15
Scr	Lcu	Arg Leu 20	Scr Cys	Ala Ala	Ser Gly Phe 25	Thr Val	Thi Scr Tyr 30
G 1 y	V a 1	His Trp 35	Val Arg	GIn Ala 40	Ρτο Gly Lys	Giy Leu 45	Glu Trp Val
GIy	V a i 5 0	ІІс Ттр	Scr Gly	Gly Scr 55	Thr Asp Tyr	Asa Ala 60	Ala Phc IIc
Scr 65	Arg	Phc Thr	11c Scr 70	Arg Asp	Asn Scr Lys 75	Asn Thr	Leu Tyr Leu 80

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							-cont	inued							
Gln	Μcι	Asn	Sсг	L c u 8 5	Gln	Ala	Glu	Asp	T h r 9 0	Aln	110	Tyr	Tyr	Су8 95	Ala
Arg	Ala	Gly	Asp 100	Туг	Asn	Туг	Asp	G 1 y 1 0 5	Рћс	Ala	Туг	Тгр	G I y I 1 0	Gln	G 1 y
Thr	Ĺси	V a 1 1 1 5	Tbr	Val	Scr	Scr									
(2) INFORMAT	ION FOR	SEQID	ND:62:												
(i)5	(A) (B) (C)) LENGT) TYPE :	ACIERIS H.: 107 am amino acid DEDNES OGY: linc	nino acida 1 S: single											
(ii)?	NOLECUI	LETYPE	: poplide												
(xi)\$	SEQUENC	E DESC	RIPTION:	SEQ ID	NO62:										
Asp I	1 1 c	Val	Lcu	Thr 5	Gln	Scr	Pro	Ala	T h r 0	L c u	Scr	Vəl	Thr	Pro 15	Gly
Asp	Scr	V a 1	Sc 1 20	Lcu	Scr	Ċys	Arg	A I a 2 5	Scr	Gin	Scr	ll c	Sсг 30	Asu	Asn
Lcu	His	Т г р 3 5	Туг	Gin	Gln	Lys	Sсг 40	His	Glu	Sсг	Pro	Arg 45	L c u	Lcu	Ilc
Lys	Ту г 50	Ala	Scī	Gln	Scr	11c 55	Scr	Gly	11 c	Pro	Sсг 60	Arg	РЪс	Sсг	Gly
S c r 6 5	Gly	Scr	Gly	Thr	Asp 70	Pbc	Thr	Lcu	Scr	Val 75	Asn	GIy	Val	Glu	T b r 8 0
Glu	Asp	Phc	Gly	Mc 1 85	Tyr	Phe	Суѕ	Gln	G 1 10 9 0	Scr	Asn	Scr	Тгр	Pro 95	His
Thr	Pbc	G 1 y	G I y 100	Gly	Tbr	Lys	Lcu	G l u 1 D 5	1 1 c	Lys					
2) INFORMAT	ion for	SEQ ID	NO:63:												
(i)	(A (B (C) LENGI) TYPE: :) STRAN	ACIERIS TI: 107 an amino acid DEDNES OGY: linc	nino acid: d S: singlic	5										
(ii)	MOLECU	LE TYPE	i: peptide												
(xi):	SEQUENC	TE DESC	RIPTION:	SEQ ID											
G l u 1	11 c	Val	Lcu	Tbr S	Glπ	Scr	Pro	Gly	Tbr 10	Lcu	Scr	Lcu	Sст	Pro 15	GIy
Głu	Arg	Ala	Thr 20	Ĺcu	Scr	Суз	Arg	A 1 a 2 5	Scт	GIn	Scr	llc	Sст 30	Asn	Asn
Lcu	His	Trp 35	Тут	Gln	G 1 n	Lys	Рго 40	Gly	G1 n	Alz	Pro	Arg 45	Lcu	Lcu	11 c
Lys	Туг 50	Ala	Scr	Gln	Scr	11c 55	Ser	G 1 y	I 1 c	Pro	Asp 60	Arg	Phc	Scr	Gly
S c r 6 5	Gly	Scr	Gly	Thr	Asp 70	Phc	Thr	Lcu	Thr	11c 75	Scr	Arg	Leu	Glu	Fr • 80
Glu	Asp	Pbc	Ala	V a 1 8 5	Туг	Туг	Cys	Gln	G 1 n 9 0	Scr	Asn	Ѕсг	Trp	Рго 95	His
ТЪг	Phc	Gły	G i n 100	Gly	Tβr	Lys	V a l	G I u 1 0 5	IIC	Lys					

(2) INFORMATION FOR SEQ ID NO:64:

COL	114 11	110	d -
$-\infty$			S .

(i) SEQUENCE CHARACIERISTICS: (A) LENGTH: 119 amino acids (B) TYPE: amino acid (C) STRANDEDNESS; single (D) TOPOLOGY: linear (i 1) MOLECULE TYPE: pepilde (x i) SEQUENCE DESCRIPTION: SEQ ID NO.64: Glu Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Lys Pro Gly Ala 1 5 10 15 Ser Mei Lys lie Ser Cys Lys Ala Ser Val Tyr Ser Phe Thr Gly Tyr 20 25 30 Thr McL Asn Trp Val Lys Glp Ser His Gly Gln Asn Leu Glu Trp flc 35 40 45 Gly Leu lic Asn Pro Tyr Asn Gly Gly Thr Scr Tyr Asn Gln Lys Phe 50 55 60 Lys Gly Lys Ala Thr Leu Thr Val Asp Lys Scr Scr Ast Thr Ala Tyr 65 70 75 80 Mei Glu Leu Leu Ser Leu Thr Ser Ala Asp Ser Ala Val Tyr Tyr Cys 85 90 95 Thr Arg Arg Gly Phc Arg Asp Tyr Scr Mci Asp Tyr Trp Gly Gln Gly 100 105 110 Thr Scr Val Thr Val Scr Scr 115 (2) INFORMATION FOR SEQ ID NO:65: (i) SEQUENCE CHARACTERUSTICS: (A) LENGTH: 119 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: pepude (x i) SEQUENCE DESCRIPTION: SEQ ID NO:65: Gin Val Gin Leu Val Gin Ser Gly Ala Giu Val Lys Lys Pro Gly Ser 1 5 10 is Ser Val Arg Val Ser Cys Lys Ala Ser Gly Tyr Ser Phe Thr Gly Tyr 20 25 30 Thr Mcl Asp Trp Val Arg Gin Ala Pro Gly Lys Gly Leu Glu Trp Val 35 40 45 Gly Leu Ile Asn Pro Tyr Asn Gly Gly Thr Ser Tyr Asn Gln Lys Phe 50 55 60 Lys Gly Arg Val Thr Val Ser Leu Lys Pro Ser Phe Aso Gln Ala Tyr 65 70 75 80 Mci Glu Lou Sor Sor Lou Pho Sor Glu Asp Thr Ala Val Tyr Tyr Cys 85 90 95 Arg Arg Gly Phe Arg Asp Tyr Ser Met Asp Tyr Trp Gly Gln Gly 100 105 110

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(2) INFORMATION FOR SEQ ID NO:66:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 393 base pairs (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

Thr Leu Val Thr Val Ser Ser

(D) TOPOLOGY: linear

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(ii) MGLECULE TYPE: CDNA	(i	ì) MGLECULE TYPE: CDNA	
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(i x) FEATURE:

(A) NA. ME/KEY: CDS (B) LOCATION: 1,393

(x i) SEQUENCE DESCRIPTION: SEQ 1D NO:66:

Mcl	G A G G I u								Val					Val		4 8	
1 6.6 T	тсс	ACA	GGT	GAC	ΑΤΤ	бтб	стб	ACC		тст	CCA	GCT	тст	15 TTG	GCT	9.6	
	Scr																
	TCT Scr															144	
	G A T A s p 5 0															192	
	CAG Gln															240	
	GTC Val															288	
	A A C A s n															336	
	CAA Gln															384	
	АТС 11с 130															393	

(2) INFORMATIGN FOR SEQ ID NO:67:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 13) amino acids (B) TYPE: amino acid (D) TOPOLOGY: lincar

(i i) MOLECULE TYPE: procin

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:67:

Мсі 1	Glu	Lys Asp	Thr Leu 5	Leu Leu	Trp Val Lcu 10	Lcu Lcu Trp	Val Pro 15
Gly	Scr	Thr Gly 20	Asp 11c	Val Lcu	The Gln Scr 25	Pre Ala Scr 30	Lcu Ala
Val	Scr	Լշս Gly 35	Gln Arg	Ala Thr 40	llc Ser Cys	Arg Ala Scr 45	Glu Scr
Val	A s p 5 0	Asn Tyr	Gly Ilc	Scr Pbc 55	Ме с Азв Тер	Pèc Gla Gla 60	Lys Pro
G I y 6 5		Pro Pro	Lys Lcu 70	Lcu îlc	Tyr Ala Ala 75	Sct Asn Glo	Gly Ser 80
Gly	Val	Pro Ala	Arg Phc 85	Scr Gly	Scr Gly Scr 90	Gly Thr Asp	Phc Scr 95
Lcu	Αςπ	J1c His 100	Pro Mci		Азр Азр Тbr 105	Ala Mci Tyr 110	
Gln	Gln	ScrLys 115	Glu Val	Pro Trp 120	Тыг Рыс Сіу	Gly Gly Thr 125	Lys Lcu

G	L	U	1	1	C	Lys
			1	3	0	

(2) INFORMATION FOR SEQ ID NO.68;

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 405 base pairs (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPGLOGY: linear

(i i) MOLECULE TYPE: cDNA

(i x) FEATURE

(A) NAME/KEY: CDS (B) LOCATION: 1.405

(x i) SEQUENCE DESCRIPTION: SEQ ID NO.68:

ATG	GGA	TGG	AĠC	TGG	ATC	ТТТ	СТС	TTC	CTC	CTG	TCA	GGA	ACT	GCA	GGC	4 8
Mcl	Gly	Тгр	Scr	Тгр	llc	Pbc	Lcu	Pbc	Lcu	Lcu	Scr	Gly	Thr	Ala	Gly	
1				٢					10					15		
GTC	CAC	тст	GAG	GTC	CAG	СТТ	CAG	CAG	TCA	GGA	ССТ	GAG	CTG	GTG	AAA	96
Val	His	Scr	Glu	V a 1	Gln	Lcu	Gln	GIn	Scr	Gly	Pro	Glu	Lcu	Val	Lys	
			2 0					25					30			
CCT	GGG	GCC	TCA	GTG	AAG	ΛТΑ	тсс	ТGС	AAG	GCT	TCT	GGA	TAC	ACA	TTC	144
Pro	Gly	Ala	Scr	Val	Lys	1 1 c	Scr	Cys	Lys	Ala	Scr	Gly	Tyr	Thr	Phe	
		3 5					4 0					4 5				
ACT	GAC	TAC	AAC	ATG	CAC	тGG	GTG	AAG	CAG	AGC	САТ	GGA	AAG	AGC	СТТ	192
Thr	Asp	Туг	Asn	Mcı	His	Trp	Val	Lys	Glu	Scr	His	Gly	Lys	Scr	Lcu	
	50					55					60					
GAG	TGG	АТТ	GGA	ТАТ	АТТ	ТАТ	сст	TAC	AAT	GGT	GGT	ACT	GGC	TAC	AAC	240
Glu	Тгр	1 l c	Gly	Туг	Ilc	Tyr	Pro	Туг	Asn	Gly	Gly	Thr	Gly	Туг	Asn	
6 5					70					7 5					8 0	
CAG	AAG	ттс	AAG	AGC	AAG	GCC	ACA	TTG	ACT	GTA	GAC	ΑΑΤ	TCC	TCC	AGC	288
Glu	Lys	Phc	Lys		Lys	Ala	Thr	Lcu		Val	Asp	Asn	Scr		Scr	
				85					90					95		
ACA	GCC	TAC	ATG	GAC	GTC	CGC	AGC	CTG	ACA	тст	GAG	GAC	TCT	GCA	GTC	3 3 6
Thr	Ala	Туг	Mc 1	Asp	Val	Arg	Scr	Lcu	Τhτ	Scr	Glu	Asp	Scr	Ala	Val	
			100					105					110			
TAT	TAC	TGT	GCA	AGA	GGG	CGC	ccc	GCT	ATG	GAC	TAC	TGG	GGT	CAA	GGA	384
Tyr	Туг	Cys	Ala	Arg	Gly	Arg	Pro	Ala	McL	Asp	Туг	Trp	Gly	Gin	Gly	
		115					120					125				
ACC	TCA	GTC	ACC	GTC	TCC	TCA										405
Thr	Scr	Val	Thr	Val	Scr	Scr										-
	1 3 0					1 3 5										1.4

(2) INFORMATION FOR SEQ ID NO:69:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 135 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO. 69:

M c L 1	Gly	Trp	Scr Trp 5	llc Phe	Leu Phe	Lcu Lcu 10	Scr Gly	Thr Ala (15	Gly
V a 1	His	Scr	Glu Val 20	Gln Lcu	Gln Gln 25	Scr Gly	Pro Glu	Lcu Val 30	Lys
Pro	Gly	A 1 a 3 5	Scr Val	Lys llc	Ser Cys 40	Lys Ala	ScrGly 45	Tyr Thr	Pbc
Tbr	Asp 50	Туг	Азя Мсі	His Trp 55	Val Lys	Gin Scr	His Gly 60	Lys Scr	Lcu

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Glu Trp llc Gly Tyr IIc 65 70	Туг Pro Tyr Asn Gl 7	
Gln Lys Phc Lys Scr Lys . 85		
Tbr Ala Tyr Mci Asp Val 100	Arg Scr Lcu Thr Sc 105	r Glu Asp Scr Ala Val 110
Tyr Tyr Cys Ala Arg Gly . 115	Arg Pro Ala Mct As 120	p Tyr Trp Gly Gla Gly 125
Thr Ser Val Thr Val Ser 130	S c r 1 3 5	
(2) INFORMATION FOR SEQ 1D NO.70:		
 (1) SEQUENCE CHARACTERISTICS: (A) LENGTH: 107 amino acids (B) TYPE; amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 		
(i i) MOLECULE TYPE: peptide		
(x i) SEQUENCE DESCRIPTION: SEQ ID	NO:70:	
Asplic Gin McL Thr 1 5	Glm Scr Pro Scr T 1	hr Leu Ser Ala Ser Val Gly 0 15
Asp Arg Val Thr llc 20	Thr Cys Arg Ala S 25	er Gin Ser Ile Asn Thr Trp 30
Lcu Ala Trp Tyr Glm 35	Gla Lys Pro Gly L 40	ys Ala Pro Lys Leu Leu Mei 45
Tyr Lys Ala Scr Scr 50	Leu Glu Ser Gly V 55	al Pro Scr Arg Phc llc Gly 60
Scr Gly Scr Gly Thr 65	Glu Phc Thr Lcu T 70	hr lle Ser Ser Lev Glm Pro 75 80
Asp Asp Pbc Ala Thr 85	Tyr Tyr Cys Gln G 9	
Met Phe Gly Gln Gly 100	Tbr Lys Val Glu V 105	al Lys
(2) INFORMATION FOR SEQ ID NO:71;		
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 111 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 		
(i i) MOLECUIE TYPE: populide		
(x i) SEQUENCE DESCRIPTION: SEQ \square	NO:71:	
Asp lle Gla Mei Thr 1 5		ст Leu Ser Ala Ser Val Gly 0 l5
Asp Arg Val Thr IIc 20	Thr Cys Arg Ala S 25	cr GIu Scr Val Asp Asn [†] Tyr 30
Gly Ile Scr Phc Mct 35	Азл Тгр РЪс Gln G 40	lπ Lys Pro Gly Lys Ala Pro 45
Lys Lov Lou ilo Tyr 50	Ala Ala ScrAsnG 55	lm Gly Scr Gly Val Pro Scr 60
Arg Phc Scr Gly Scr 65	Gly Ser Gly Thr A 70	sp Phc Thr Leu Thr Ile Ser 75 80
Ser Lev Glu Pro Asp 85		yr Tyr Cys Glm Gln Scr Lys 0 95

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(2) INFORMATION FOR SEQ ID NO:72:

(i) SEQUENCE CHARACIERISIICS: (A) 1ENGTH: 117 amino acids

(B) TYPE: amino scid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(; i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:72:

GlaValGlnLeuValGlnSerGlyAlaGluValLysLysProGlySerSerValLysValSerCysLysAlaSerGlyGlyThrPheSerArgSerAlaIIcJisTrpValArgGlnAlaSerGlyGlyGlyThrPheSerArgSerAlaIIcJisTrpValArgGlnAlaProGlyGlnGlyLeuGluTrpMetGlyGiyIIcValProMeiPheGlyProAsnTyrAlaGlnLysPheGinGiyArgValThrProMeiPheGluSerThrAlaGluLysGinGiyArgValThrAngGlyProAsnTyrAlaFyrAlaTyrGinGiyArgSerLeuArgSerGluAspGiuSerThrAsnThrAlaFyrGinGiyArgSerLeuArgSerGiuAspThrAlaPheTyrFyrFyrGinCiyFinArgSerCiyFinAngFinAlaFyrFyrFyrFyrFyrFyrFyrGinCiyFinFinFinFin

(2) INFORMATION FOR SEQ ID NO:73:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTII: 116 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: bicar

(i i) MOLECULE TYPE: popuide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:73:

Gln 1	Val Gln	Lcu Val S	Gln Ser	Gly Ala	Glu Val 10	Lys Lys	Pro Gly IS	Scr
Scr	Val Lys	Vəl Scr 20	Cys Lys	Ala Scr 25	Gly Tyr	Thr Phc	Тыт Азр 30	Туг
Asn	Mct His 35	Trp Val	Arg Gln	Ala Pro 40	Gly Gln	Gly Lcu 45	Glu Trp	1 1 c
Gly	Tyr llc 50	Туг Рго	Tyr Asm SS	Gly Gly	Thr Gly	Tyr Asn 60	Gl¤ Lys	РЬс
L y s 6 5	Scr Lys	Ala Thr	llc Thr 70	Ala Asp	Glu Scr 75	Thr Asn	Thr Ala	Туг 80
Μcι	Glu Lcu	Scr Scr 85	Lcu Arg	Ser Glu	Asp Thr 90	Ala Val	Tyr Tyr 95	Cys
Ala	Arg Gly	Arg Pro 100	Ala Mici	Asp Tyr 105	Trp Gly	Glm_Gly	Thr Leu 110	V 8 1
Tbr	Val Scr 115							

(2) INFORMATION FOR SEQ ID NO:74:

			5,530,101			
	135				136	
			-continued			
	ENCE CHARACTERISTI (A) LENGTH: 132 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
	ECULE TYPE: DNA (oligo	onucleolide)				
	ENCE DESCRIPTION: SI					
			ATCTTTCTCT	TCCTCCTGTC	AGGAACTGCT	6
					GCCTGGGAGC	12
TCAGTGAAGG						I 3
(2) INFORMATION	OR SEQ ID NO:75:					
	ENCE CHARACIERISTI (A)LENOTH: 133 base (B)TYPE: nucleic acid (C)S'IRANDEDNESS: (D)TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA (oligo	onucleo ide)				
(xi)SEQU	ENCE DESCRIPTION: SI	EQ ID NO:75:				
AGCCGGTACC	ACCATTGTAA	GGATAAATAT	ATCCAATCCA	TTCCAGGCCT	TGGCCAGGAG	6
CCTGCCTCAC	CCAGTGCATG	TTGTAGTCAG	T G A A G G T G T A	GCCAGAAGCT	TTGCAGGAAA	12
CCTTCACTGA	GCT					13
(2) INFORMATION	OR SEO ID NO.76					
	ENCE CHARACTERISTI	CS.				
	(A) LENGTH: 112 base (B) TYPE: nucleic acid					
1	C) STRAN DEDNESS:	single				
	(D) TOPOLOGY: linear					
(ii) MOL	ECULE TYPE: DNA (oligo	onucleo ide)				
(xi)SEQU	ENCE DESCRIPTION: SI	EQ ID NO:76:				
TGGTGGTACC	GGCTACAACC	ΑGΑΑGTTCΑΛ	GAGCAAGGCC	ACAATTACAG	CAGACGAGAG	б
ΤΑCΤΑΑCΑCΑ	GCCTACATGG	AACTCTCCAG	CCTGAGGTCT	GAGGACACTG	CA	1
(2) INFORMATION	OR SEQ ID NO:77:					
	ENCE CHARACTERIS ¹¹ (A) LENGTH: 111 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (O) TOPOLOGY: linear	Pairs				
(ii) MOLI	ECULE TYPE: DNA (oligo	nucleolide)				
(x i) SEQU	ENCE DESCRIPTION: S	D NO:77:				
TATATCTAGA	GGCCATTCTT	ACCTGAAGAG	ACAGTGACCA	GAGTCCCTTG	GCCCCAGTAG	6
TCCATAGCGG	GGCGCCCTCT	TGCGCAGTAA	TAGACTGCAG	TGTCCTCAGA	С	1 1
(2) INFORMATION	FOR SEQ ID NO:78:					
	ENCE CHARACIEIUST					

(T) SEQUENCE CHARACI BIOS INCS: (A) LENGTH: 122 base pairs (B) TYPE: nucleic acid (C) STRANDEDN'ESS: single (D) TOPOLOGY: l'uncar

(i i) MOLECULE TYPE: DNA (oligonucleatide)

5	53	0	11	11
э,	22	υ,	11	JI

continued continued (i i) SEQUENCE DESCRIPTION SEQ D NOTE: CALATACETAGA CCACCATOGA GAAAGACACA CICCEGETAT OGGETCETGET TETETEGGGETT GG 12 GG 12 (i) SEQUENCE DESCRIPTION SEQ D NOTE: 12 (i) SEQUENCE DESCRIPTION SEQ D NOTE: 12 (i) SEQUENCE DESCRIPTION: SEQ D NOTE: 13 (i) SEQUENCE DESCRIPTION: SEQ D NOTE: 14 (i) SEQUENCE DESCRIPTION: SEQ D NOTE: <td< th=""><th></th><th>137</th><th></th><th>,050,101</th><th>1</th><th>20</th><th></th></td<>		137		,050,101	1	20	
TATATCTAGA CCACCATGGA GAAAGACACA CTCCTGCTAT GGGTCCTGT TCCTGGGTT 6 CCAGGTTCCA CAGGTGACAT TCAGATGACC CAGTCTGCGA GCTCTGTC CGCATCAGTA 12 GG 12 (2) INFORMATION FOR EQUID NO.29: (1) SEQUENCE GIAACTESISTICE (1) SEQUENCE DIACTESISTICE (1) SEQUENCE DISCRIPTION SEQUENCIAL (1) SEQUENCE DISCRI		157		-continued	1	30	
CCAGGUTICCA CAGUTGACAI TCAGATGACC CAGUTCCGGA GETETETE CGCATCAGIA I2 GG 12 (2) INFORMATION FOR EQ LD NO.79: (1) ISQUENCE CHARACTERSTICS (1) ISQUENCE DESCRIPTION EQU DNO.79: CCAGAAGCIT AGGAGCETTC CCGGGGITICT GITGGAACCA OTTCATAAAG CTAATGCCAI (1) SQUENCE DESCRIPTION EQU DNO.79: CCAGAAGCIT AGGAGCETTC CCGGGGITICT GITGGAACCA OTTCATAAAG CTAATGCCAI (1) SQUENCE DESCRIPTION EQU DNO.79: CCAGAAGCIT AGGAGCETTC CCGGGGITICT GITGGAACCA OTTCATAAAG CTAATGCCAI (2) INFORMATION FOR EQU DNO.79: (1) SQUENCE DESCRIPTION EQU DNO.79: (1) SQUENCE DESCRIPTION EQU DNO.79: (1) SQUENCE DESCRIPTION EQU DNO.79: (2) INFORMATION FOR EQU DNO.79: (1) SQUENCE DESCRIPTION EQU DNO.79: (2) INFORMATION FOR EQU DNO.79: (1) SQUENCE DESCRIPTION EQU DNO.79: (2) INFORMATION FOR EQU DNO.81: (1) SQUENCE DESCRIPTION EQU DNO.79: (2) INFORMATION FOR EQU DNO.81: (1) SQUENCE DESCRIPTION: SQU DNO.81: (1) SQUENCE DESCRIPTION: SQU DNO.81: (1) SQUENCE DESCRIPTION: SQU DNO.81: (1) SQUENCE CITACTTE TACTTACCTT TCATCCCAC CTTGGTCCCT TGACCGAACG 6 TCCACCGAAC CTCCTACTT TCATCACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 6 (2) INFORMATION FOR SQU DNO.81: (1) SQUENCE CITACTTE TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 6 (1) SQUENCE CITACTTE TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 6 (2) INFORMATION FOR SQU DNO.81: (1) SQUENCE CITACTTE TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 6 (2) INFORMATION FOR SQU DNO.82: (1) SQUENCE CITACTTE TACTTACCTTACTT TGCTGCCACG AATAGGTTCC ACGTCACG 11] (2) INFORMATION FOR SQU DNO.82: (1) SQUENCE CITACTTE TCACCTCACGT TATCTACCCCC CTGGTCCCT TGACCGAACG 6 (2) INFORMATION FOR SQUENCES SAUGHE (1) SQUENCE CITACTTE TCACCTCACGT TGCCCCCCCCCCCCCCCCCCCCCCCCC	(x i) SE(UENCE DESCRIPTION: S	EQ ID NO:78:				
GG 12 JINFORMATION FOR SEQ DINOSE: (1) SEQUENCE CHARACTERISTICS: (1) SEQUENCE CHARACTERISTICS: (1) SEQUENCE DESCRIPTION: SEQ DINOSE: (1) SEQUENCE CHARACTERISTICS: (1) SEQUE	ΤΑΤΑΤĊΤΑΘΑ	CCACCATGGA	GAAAGACACA	CTCCTGCTAT	GGGTCCTGCT	TCTCTGGGTT	6 0
<pre>(1) INFORMATION FOR SEQ ID NO.59: (1) SEQUENCE CHARACTERISTICE: (1) SEQUENCE CHARACTERISTICE: (1) SEQUENCE DESCRIPTION: SEQ ID NO.59: TEAGAAAGETT AGGAGECTTE CEGGGGTTEET GTTGGAACCCA GTTCATAAAG CTAATGECAT 6 AATTGTEGAC ACTITEGETG GETETGGCATG TGATGGTGAC CETGTEET ACTGATGCGG 12 AC 12 (2) INFORMATION FOR SEQ ID NO.82: (1) SEQUENCE CLARACTERISTICE: (1) SEQUENCE CLARACTERISTICE: (1) SEQUENCE CLARACTERISTICE: (1) SEQUENCE DESCRIPTION: SEQ ID NO.80: TECTAAGETT CTGGACAGA ACTITEACGTT CACCAATGECTE GTGGGGTACCCT CTGGGTGACC (1) SEQUENCE CLARACTERISTICE: (1) SEQUENCE CLARACTERISTICE: (1) SEQUENCE DESCRIPTION: SEQ ID NO.80: TECTAAGETT CTGGACAGA ACTICACTCT CACCAATGECTE GGGGTACCCT CTGGGTGACCT 11 (2) INFORMATION FOR SEQ ID NO.81: (1) SEQUENCE DESCRIPTION: SEQ ID NO.81: (2) INFORMATION FOR SEQ ID NO.81: (1) SEQUENCE DESCRIPTION: SEQ ID NO.81: (2) INFORMATION FOR SEQ ID NO.81: (1) SEQUENCE DESCRIPTION: SEQ ID NO.81: (1) N</pre>	CCAGGTTCCA	CAGGTGACAT	TCAGATGACC	CAGTCTCCGA	GCTCTCTGTC	CGCATCAGTA	120
<pre>(i) SEQUENCE CHARACTERISTICS: (A) LENOTH 22 bas pairs (E) TYPE works and (C) STANDEDRESS mage (i) TOTOLOGY here (ii) NOLECLE TYPE: NA (digeneticide) (x i) SEQUENCE DESCRIPTON SEQ D NO3P. TEAGAAGGET AGGAGCETT C CCGGGGTTECT GITGGAACCA GITCATAAAG CTAATGCCAT 6 AATTGTCGAC ACTITCGCTG GETCTGCATG TGATGGTGAC CCTGTCTCT ACTGATGCGG 12 AC (x i) SEQUENCE DESCRIPTON SEQ D NO3P. (i) SEQUENCE CLARACTERISTICS (A) LENOTE 10 Marginet (i) SEQUENCE DESCRIPTON SEQ D NO3P. (i) SEQUENCE DESCRIPTON SEQ D NO3P. (c) STANDEDRESS mage (i) TYPE: INA(digeneticide) (x i) SEQUENCE DESCRIPTON SEQ D NO3P. TCCTAAGCTT CTGATTACG CTGCATCCAA CCAAGGETCC GGGGTACCCT CTGGCTTETC 6 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT 11 (2) RFORMATION FOR SEQ D NO3E. (i) SEQUENCE DESCRIPTION SEQ D NO3P. (c) STANDEDRESS mage (i) SEQUENCE DESCRIPTION SEQ D NO3P. (c) STGANGEDRESS mage (i) SEQUENCE DESCRIPTION SEQ D NO3P. (c) STGANGENESS MARGENET (i) SEQUENCE DESCRIPTION SEQ D NO3P. (i) SEQUENCE DESCRIPTION SEQUENCED SECUENCES. (i) SEQUENCE DESCRIPTION SEQUENCED SECUENCES. (i) SEQUENCE DESCRIPTION SEQUENCES. (i) SEQUENCE DESCRIPTION SEQUE</pre>	GG						122
<pre>(i) SEQUENCE CHARACTERISTICS: (A) LENOTH 22 bas pairs (E) TYPE works and (C) STANDEDRESS mage (i) TOTOLOGY here (ii) NOLECLE TYPE: NA (digeneticide) (x i) SEQUENCE DESCRIPTON SEQ D NO3P. TEAGAAGGET AGGAGCETT C CCGGGGTTECT GITGGAACCA GITCATAAAG CTAATGCCAT 6 AATTGTCGAC ACTITCGCTG GETCTGCATG TGATGGTGAC CCTGTCTCT ACTGATGCGG 12 AC (x i) SEQUENCE DESCRIPTON SEQ D NO3P. (i) SEQUENCE CLARACTERISTICS (A) LENOTE 10 Marginet (i) SEQUENCE DESCRIPTON SEQ D NO3P. (i) SEQUENCE DESCRIPTON SEQ D NO3P. (c) STANDEDRESS mage (i) TYPE: INA(digeneticide) (x i) SEQUENCE DESCRIPTON SEQ D NO3P. TCCTAAGCTT CTGATTACG CTGCATCCAA CCAAGGETCC GGGGTACCCT CTGGCTTETC 6 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT 11 (2) RFORMATION FOR SEQ D NO3E. (i) SEQUENCE DESCRIPTION SEQ D NO3P. (c) STANDEDRESS mage (i) SEQUENCE DESCRIPTION SEQ D NO3P. (c) STGANGEDRESS mage (i) SEQUENCE DESCRIPTION SEQ D NO3P. (c) STGANGENESS MARGENET (i) SEQUENCE DESCRIPTION SEQ D NO3P. (i) SEQUENCE DESCRIPTION SEQUENCED SECUENCES. (i) SEQUENCE DESCRIPTION SEQUENCED SECUENCES. (i) SEQUENCE DESCRIPTION SEQUENCES. (i) SEQUENCE DESCRIPTION SEQUE</pre>							
<pre>(A LENDIE UZ Exc pair) (A LENDIE UZ Exc pair) (B) TYPE suble said (C) STRANDEDHESS saigh (C) TROUGON: Base (i i) NOLECLE TYPE: DNA (Digenstroade) (x i) SEQUENCE DESCRPTION: SEQ D NO.79: TECAGAA GGTT AGGAGGCCTTC CCGGGGTTCT GTTGGAACCA GTTCATAAAG CTAATGCCAT 6 AATTGTCGAC ACTTTCGCTG GCTCTGCATG TGATGGTGAC CCTGTCTCT ACTGATGCCGG 12 AC 12 () NFORMATION FOR SEQ D NO.80: (i) SEQUENCE DESCRPTION: SEQ D NO.80: (c) STRANDEDHESE saigh () D TOPOLOGY. Here (i) SEQUENCE DESCRPTION: SEQ D NO.80: (i) SEQUENCE DESCRPTION: SEQ D NO.80: (c) STRANDEDHESE saight () D TOPOLOGY. Here (i) SEQUENCE DESCRPTION: SEQ D NO.80: (c) STRANDEDHESE saight () D TOPOLOGY. Here (i) SEQUENCE DESCRPTION: SEQ D NO.80: (c) STRANDEDHESE saight (c) STRANDEDHESE SA</pre>	(2) INFORMATION	N FOR SEQ ID NO:79:					
<pre>(::)SQUENCE DESCRIPTION: SEQ D NO.7% TCAGAAGGTT AGGAGGCCTTC CGGGGTTCTC GTGGGAACCA GTTCATAAAG CTAATGCCAT 6 AATTGTCGAC ACTITCGCTG GCTCTGCATG TGATGGTGAC CCTGTCTCT ACTGATGCGG 12 AC 12 (2)NFORMATION FOR SEQ D NO.8% (i)SQUENCE CLARACTERSTICS:</pre>	(i) SEC	(A) LENGTH: 122 base (B) TYPE: nucleic acid (C) STRANDEDNESS:	pair; single				
TAGAAACTT AGGAGCCTTC CCGGGTTTCT GTTGGAACCA GTTCATAAAG CTAATGCCAT 6 AATTGTCGAC ACTTTCGGTG GCTCTGGATG TGATGGTGAC CCTGTCTCT ACTGATGCGG 12 AC 12 (2) NFORMATION FOR SQ (D NO30: (i) SEQUENCE CHARACTERSTICS (i) SEQUENCE CHARACTERSTICS (i) NOLECULE TYPE: DNA (bigeneckolds) (c) TTWOLOCY: New (ii) NOLECULE TYPE: DNA (bigeneckolds) (c) STRUMENESS segle NO30: TCCTAAGCTT CTGGATTACG CTGCATCCAA CCAAGGCTCC GGGGTACCCT CTCGCTTCTC 6 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT 11 (2) NFORMATION FOR SQ (D NO31: (i) SEQUENCE CHARACTERISTICS: (i) NFORMATION FOR SQ (D NO31: TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 6 TCCACGGGAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG 11 (2) NFORMATION FOR SQ (D NO32: (i) SEQUENCE CHARACTERISTICS: (i) NOLECULE TYPE: DNA (i) NUMERACT CDS (i) NUMERACTERISTICS: (i) NUMERACTERISTICS:	(ii) MO	LECULE TYPE: DNA (olig	onuc)colide)				
AATTGTCGAC ACTTTCGCTG GCTCTGCATG TGATGGTGAC CCTGTCTCCT ACTGATGCGG 12 AC 12 (2) NFORMATION FOR SEQ ID NOSE: (i) SEQUENCE CHARACTERISTICS (i) INFORMATION FOR SEQ ID NOSE: (i) TTFC muche add (i) TTFC muche	(x i) SE(QUENCE DESCRIPTION: S	EQ ID NO:79:				
AC 12 (2) INFORMATION FOR SEQ ID NOAD: (i) SEQUENCE CHARACTERISTICS (i) STORE CHARACTERISTICS (i) STORE OF A CONTRACT CONTRACTOR SEQ ID NOAD: (i) STORE OF A CONTRACT CONTRACTOR SEQ ID NOAD: (c) STRANDED NESS: Single (c) STRANDED N	TCAGAAGCT	r aggagccttc	CCGGGTTTCT	GTTGGAACCA	GTTCATAAAG	CTAATGCCAT	6 0
<pre>(2) INFORMATION FOR SEQ ID NOAD: (i) SEQUENCE CHARACTERSTICS (A) LENGTH: 10 base print (E) TYTE: numble tod (C) TYTE: numble tod (C) TYTE: DNA (objectedide) (c) TYTE: DNA (object</pre>	AATTGTCGAG	C ACTTTCGCTG	GCTCTGCATG	TGATGGTGAC	CCTGTCTCCT	ACTGATGCGG	120
<pre>(i) SEQUENCE CHARACTERISTICS (A) LENGTH: 10 Hase pair (B) TTRE subdit ad (C) STRANDEDNESS single (C) TOPOLOGY: Hear (ii) MOLECULE TYPE: DNA (bigoacteoide) (xi) SEQUENCE DESCRIPTION: SEQUE NOMO. TCCTAAGGETT CTGGATTTACG CTGGATCCAA CCAAGGGETCE GGGGTACCCT CTGGETTETC 6 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTCA TCTCTGCAGG CTGATGACT 11 (2) INFORMATION FOR SEQUE NO.31: (i) SEQUENCE CHARACTERISTICS; (A) LENGTH: 118 base pair (i) SEQUENCE DESCRIPTION: SEQUE NO.38: (ii) SEQUENCE DESCRIPTION: SEQUE NO.38: (iii) MOLECULE TYPE: DNA (bigoacteoide)) (xi) SEQUENCE DESCRIPTION: SEQUENCESE: (i) SEQUENCE DESCRIPTION: SEQUENCES</pre>	AC						122
<pre>(i) SEQUENCE CHARACTERISTICS (A) LENGTH 10 has pair (B) TTME under and (C) STRANDEDNESS single (C) STRANDEDNESS single (C) I) TOPOLOGY: Near (ii) MOLECULE TYPE: DNA (biguesteoide) (c) I) SEQUENCE DESCRIPTION: SEQU NOMO. TECTAAGECTT CTGATTTACG CTGCATCCAA CCAAGGETCE GEGEGTACCCT CTEEETTETC AGGEGAGTEGGA TETEGGGACAG ACTTCACTET CACCATTTCA TETETEGCAGE CTGATGACT 11 (2) INFORMATION FOR SEQU D NO.31: (i) SEQUENCE CHARACTERISTICS; (A) LENGTH: 118 base pairs (C) STRANDEDN ESS single (C) TOPOLOGY: Incom (C) STRANDEDN ESS single (C) TOPOLOGY: Incom (ii) MOLECULE TYPE: DNA (biguesteoide)) (x i) SEQUENCE DESCRIPTION: SEQ ID NO.31: TATATETAGA CTTTEGGATTE TACTTACGTT TGATETECAC CTTEGGTECCT TGAECGAACG (i) SEQUENCE DESCRIPTION: SEQ ID NO.31: TATATETAGA CTTTEGGATTE TACTTACGTT TGATETECAC CTTEGGTECCT TGAECGAACG (i) SEQUENCE DESCRIPTION: SEQ ID NO.31: (i) SEQUENCE DESCRIPTION: SEQ ID NO.32: (i) SEQUENCE DESCRIPTION: SEQ ID NO.33: (i) ID TTME under base pairs (i) SEQUENCE DESCRIPTION: SEQ ID NO.33: (i) ID NOLECULE TYPE: DNA (biguestedide)) (C) TTMENDES single (C) STRANDEDNESS single (C) STR</pre>							
<pre>(i) LENGTH: 19 base pair:</pre>	(2) INFORMATION	N FOR SEQ ID NO:80:					
<pre>(xi)SEQUENCE DESCRIPTION: SEQUE NOMO. TECTAAGETT CTGATTTAEG CTGCATECAA CCAAGGETEE GGGGTAECET CTEGETTETE 6 AGGEAGTGGA TETEGGGAEAG ACTTEACTET CACEATTTEA TETETGGAEC CTGATGAET 11 (2)INFORMATION FOR SEQ ID NO.31:</pre>	(i) SE((A) LENGTH: 119 base (B) TYPE: ancleic acid (C) STRANDEDNESS:	pair: single				
TCCTAAGCTT CTGATTTACG CTGCATCCAA CCAAGGCTCC GGGGTACCCT CTCGCTTCTC 6 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT 11 (2) NFORMATION FOR SEQ ID NO.81: (i) SEQUENCE CHARACTERISTICS; (A) LENGTH: 118 base pair (B) TYPE: molecule TYPE: DNA (okgoouclooide) (xi) SEQUENCE DESCRIPTION: SEQ ID NO.81: TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 6 TCCACGGAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG 11 (2) INFORMATION FOR SEQ ID NO.82: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 38 base pair (B) TYPE: muleie add (C) STRANDEDNESS: single (I) SEQUENCE CHARACTERISTICS: (A) LENGTH: 38 base pair (B) TYPE: muleie add (C) STRANDEDNESS: single (C) STRANESSEN (S) STRANESSEN	(i i) MC	LECULE TYPE: DNA (olig	connelcolide)				
AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT 11 (2) INFORMATION FOR SEQ ID NO.81: (A) LENGTH: 118 base pairs (B) TYPE: muchic atid (C) STRANDEDNE'SS single (D) TOPOLOGY: Human (I i MOLECULE TYPE: DNA (økgonucleonide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO.81: TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 11 (2) INFORMATION FOR SEQ ID NO.82: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 31 base pairs (B) TYPE: muchic atid (C) STRANDEDNE'SS. single (D) TOPOLOGY: Human (I) SEQUENCE CHARACTERISTICS: (A) LENGTH: 31 base pairs (B) TYPE: muchic atid (C) STRANDEDNE'SS. single (D) TOPOLOGY: Human (I) SEQUENCE CHARACTERISTICS: (A) LENGTH: 31 base pairs (B) TYPE: muchic atid (C) STRANDEDNE'SS. single (D) TOPOLOGY: Human (I) SEQUENCE CHARACTERISTICS: (A) SAMEKEY: CDS (B) LOCATION: 1.331 (X 1) SEQUENCE DESCRIPTION: SEQ ID NO.82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA	(x i) SEG	QUENCE DESCRIPTION: S	EQID NO:80.				
<pre>(2) INFORMATION FOR SEQ ID NO.81:</pre>	TCCTAAGCT	T CTGATTTACG	CTGCATCCAA	CCAAGGCTCC	GGGGTACCCT	CTCGCTTCTC	6 0
<pre>(i)SEQUENCE CHARACTERISTICS:</pre>	AGGCAGTGG	A TCTGGGACAG	ACTTCACTCT	CACCATTTCA	TCTCTGCAGC	CTGATGACT	119
<pre>(i) SEQUENCE CHARACTERISTICS:</pre>							
<pre>(A) LENGTH: 118 base pairs (B) TYPE: medicin acid (C) STRANDEDNESS: single (D) TOPOLOGY: Finear (i i) MOLECULE TYPE: DNA (% genucleoidde) (x i) SEQUENCE DESCRIPTION: SEQ ID NO.81: TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 6 TCCACGGGAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG 1 1 (2) INFORMATION FOR SEQ ID NO.82:</pre>							
<pre>(xi)SEQUENCE DESCRIPTION: SEQ ID NO:81: TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG 6 TCCACGGAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG 11 (2)DNFORMATION FOR SEQ ID NO:82: (i)SEQUENCE CHARACTERISTICS: (A)LENOTH: 38 hase pairs (B) TYPE: nucleic add (C) STRANDEDNESS: single (D) TOPOLOGY: Bincar (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAMERCEY: CDS (B) LOCATION: 1.381 (xi)SEQUENCE DESCRIPTION: SEQ ID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA 4</pre>	(1) SE((A) LENGTH: 118 base (B) TYPE: nucleic acid (C) STRANDEDNESS	e pairs				
TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG TCCACGGAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG 11 (2) INFORMATION FOR SEQ ID NO:82: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 381 base pair: (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: incom (ii) MOLECULE TYPE: cDNA (iii) MOLECULE TYPE: cDNA (iii) PEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1.381 (xii) SEQUENCE DESCRIPTION: SEQ ID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA	(ii) MC	LECULE TYPE: DNA (di)	gonucleotide)				
TCCACGGAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG (2) INFORMATION FOR SEQ ID NO:82: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 381 base pairi (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: incar (i) MOLECULE TYPE: cDNA (i) x) FEATURE: (A) NAMEREY: CDS (B) LOCATION: 1:381 (x) SEQUENCE DESCRIPTION: SEQ ID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA	(xi)SE(QUENCE DESCRIPTION: S	EQ ID NO:81:				
 (2) INFORMATION FOR SEQ ID NO:82: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 381 base pairi (B) TYPE: nucleic atid (C) STRANDEDNESS: single (D) TOPOLOGY: incar (i i) MOLECULE TYPE: cDNA (i x) FEATURE: (A) NAMEREY: CDS (B) LOCATION: 1.381 (x i) SEQUENCE DESCRIPTION: SEQ ID NO:82: 	тататстас	A CTTTGGATTC	TACTTACGTT	TGATCTCCAC	CTTGGTCCCT	TGACCGAACG	60
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 381 base pairi (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: incare (i i) MOLECULE TYPE: cDNA (i x) FEATURE: (A) NAMERCEY: CDS (B) LOCATION: 1.381 (x i) SEQUENCE DESCRIPTION: SEQ ID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA	TCCACGGAA	C CTCCTTACTT	TGCTGACAGT	AATAGGTTGC	GAAGTCATCA	GGCTGCAG	1 1 8
 (A) LENGTH: 381 base pairi (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: incar (i i) MOLECULE TYPE: cDNA (i x) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1.381 (x i) SEQUENCE DESCRIPTION: SEQ ID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA 4	(2) INFORMATIO	N FOR SEQ ID NO:82:					
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1-381 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT FGG ATT TCA 4	(i)SE	(A) LENGTH: 381 bas (B) TYPE: nucleic acid (C) STRANDEDNESS	e pair: l single				
(A) NAME/KEY: CDS (B) LOCATION: 1.381 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT FGG ATT TCA 4	(ii)M0	OLECULE TYPE: cDNA					
(x i) SEQUENCE DESCRIPTION: SEQID NO:82: ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA 4	(i x) FE	(A) NAME/KEY: CDS					
ATG GTT TTC ACA CCT CAG ATA CTT GGA CTT ATG CTT TTT TGG ATT TCA 4	(I) CE						
						G ATT TCA	
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			G G T G I y 2 0													9 (
			G G A G I y													Ι44
			A A C A s n													19:
			ATC IIc													240
			G G C G 1 y													28
			АСТ Т h r 1 0 0													33
			CAT Nis													38
2)1) SEQUE	NCE CHA NCE CHA N) LENC B) TYPE	RACTER TH: 127	u\$11CS; amino sci cid	ds										
(2)1) SEQUE	NCE CHA	RACTER TH: 127	u\$11CS; amino sci cid	ds										13
(2)1	(i) (ii)) SEQUE (/ () () MOLE(NCE CHA) LENC) TYPE) TOPO CULE TYPE	ARACTER ITH: 127 :: am'mo a LOGY: L E: protein	uSIICS: amino sci cid ncar											
	(i) (ii) (xi)) SEQUE (/ (1 (1) MOLEC	NCE CHA) LENC B) TYPE D) TOPO CULE TYPE NCE DES	ARACTER 3TH: 127 2: animo a 3LOGY: 15 2E: protein CRIPTIO	u\$11CS; amino sof oid near N: SEQ II	D NO:83:		GLV	I.c.u	Mei	Leu	Pho	Ten	LLC	Ser	3
	(i) (ii) (xi)) SEQUE (/ (1 (1) MOLEC	NCE CHA) LENC) TYPE) TOPO CULE TYPE	ARACTER 3TH: 127 2: animo a 3LOGY: 15 2E: protein CRIPTIO	u\$11CS; amino sof oid near N: SEQ II	D NO:83:		Gły	Lcu 10	Mc i	Leu	Phc	Тгр	I L c 1 5	Scr	đ
Víc L I	(i (ii (xi Vat) SEQUE (/ (1) MOLEC) SEQUE P h c	NCE CHA) LENC B) TYPE D) TOPO CULE TYPE NCE DES	ARACTER ITH: 127 I: animo a ULOGY: 15 PE: protein CRIPTIO P r o 5	U\$11CS; attino sci cid ncar N: SEQ II G I n	D NO:83: 1 1 c	Lcu		10					15		.3
Mci I A 1 z	(ii (xi) Val Scr) SEQUE (/ (1) MOLEC) SEQUE P h c A r g	NCE CHA) LENC) TYPE) TOPO CULE TYPE NCE DES Thr Gly	ARACTER TH: 127 :: animo a LOGY: Is PE: protein CRIPTIO P r o 5 A s p	U\$11CS: amino sof cid ncar N: SEQ II G I n I I c	D NO:83: 1 1 c V a 1	L c u L c u	Thr 25	10 G1n	S.c.r	Pro	Ala	Tbr 30	l 5 Lcu	Scr	13
Víci I A 1 a V a I	(i (ii (xi Val Scr Tbr) SEQUE (/ (1)) MOLEO Phe Arg Pro 35	NCE CHA) LENC) TYPE) TOPO CULE TYPE NCE DES T h r G 1 y 2 0	ARACTER DTH: 127 :: animo a iLOGY: Is PE: protein CRIPTIO P r o 5 A s p A s p	u\$19CS: antino sof cid moar N: SEQ 11 G J n J I c S c r	D NO:83: 11c Va1 Vat	Lcu Lcu Scr 40	Thr 25 Leu	IO Gln Scr	S.cr Cys	Pro Arg	Ala Ala 45	Tbr 30 Scr	l 5 L c u G 1 n	S c r	.3
Víci I A 1 z V a I I T c	(ii (xi) Val Scr Tbr Scr 50) SEQUE (1) (1) (1) (1) (1) SEQUE Pho Arg Pro 35 Asn	NCE CHA A) LENC B) TYPE D) TOPO CULE TYPE NCE DES T h r G 1 y 2 0 G 1 y	ARACTER TH: 127 :: an/mo a ULOGY: 15 PE: protein CRIPTIO P r o 5 A s p A s p L c u	u\$11CS; attnino act cid ncar N: SEQ II G J n I I c S c r H i s	D NO:83: 1 1 c V a 1 V a 1 T r p 5 5	L c u L c u S c r 4 0 T y r	Thr 25 Leu Gln	J O G l n S c r G l n	S.cr Cys Lys	Pro Arg Ser 60	Ala Ala 45 His	Tbr 30 Scr Glu	l S L c u G 1 n S c r	Scr Scr Pro	
Vic L I Val I I C 6 5	(i (xi) Val Scr Tbr Scr 50 Lcu) SEQUE (1) (1) (1) MOLEC) SEQUE Phc Arg Pro 35 Asn Lcu	A LENCE CHA A LENC B TYPE D TOPO CULE TYPE NCE DES T h r G 1 y 2 0 G 1 y A s n	ARACTER TH: 127 :: an/mo a ULOGY: 15 PE: protein CRIPTIO Pro S Asp Asp Lcu Lys	us 11 CS: atmino sof cid ncar N: SEQ II G I n I I c S c r H i s T y r 7 0	D NO:83: 11 c V a 1 V a 1 T r p 5 5 A t a	L c u L c u S c r 4 0 T y r S c r	Thr 25 Leu Gln Gln	i 0 G 1 n S c r G I n S c r	S.cr Cys Lys IIc 75	Pro Arg Scr 60 Scr	Ala Ala 45 His Gly	Tbr 30 Scr Glu Ilc	l 5 L c u G 1 n S c r P r o	Scr Scr ₽rœ Scr 80	.3
Vic L I A 1 z V a I I I C A r g A r g	(i (xi) Val Scr Tbr Scr 50 Lcu Phc) SEQUE (1) (1) (1) MOLEO) SEQUE Pho Arg Pro 35 Asn Lcu Scr	A) LENC) LENC) TOPO CULE TYPE CULE TYPE NCE DES T h r G l y 2 0 G l y A s n [i c	ARACTER TH: 127 :: an/mo a ULOGY: 15 PE: protein CRIPTIO Pro 5 Asp Asp Lcu Lys Scr 85	us 11 CS: atmino sof cid ncar N: SEQ II G I n I I c S c r H i s T y r 7 0 G I y	D NO:83: 1 1 c V a 1 V a 1 T r p 5 5 A t a S c r	L c u L c u S c r 4 0 T y r S c r G 1 y	Thr 25 Leu Gln Chr	IO GIN Scr GIN Scr Asp 90	S.cr Cys Lys Ilc 75 Phc	Pro Arg Scr 60 Scr Thr	Ala 45 His Gly Leu	Tbr 30 Scr Glu Ilc Scr	IS Leu GIR Scr Pro Val 95	Ser Ser Pro Ser 80 Astu	

(2) INFORMATION FOR SEQ ID NO:84:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 414 base pairs
 (B) TYPE: nucleic-acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: cDNA

(i x) FEATURE:

(A) NAME/KEY: CDS (B) LOCATION: 1.414

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	(xi) SEQUE	NCE DES	CRIPT10	N: SEQ 11	D NO:84:						
				Т G G Т г р 5								48
-				GTC Val		-						96
				АТG Мсι								144
				ATG Mci								192
				СТТ Lсu								240
				G G G G G 1 y 8 5								288
				G A G G I u								336
				A G A A r g							ТGG Тгр	384
				TCA Ser								414

(2) INFORMATION FOR SEQ ID NO:85:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTM: 138 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:SS:

Me I 1	G 1	уТ	гр	Sc	г	TI	р 5	1	lc		P	ac	L	C	u	P	ac		cu IO		e u	S	r	GI	у	т	h r		1 2	G	ly
Val	Ηi	s S	CΓ		u 0	V	a 1	G	1 п	5	L	c u	G	1	п		l n 2 5	S	сг	G	l y	P	0	G	lu		с и 3 О	v	a 1	L	y s
Pro	Gl		la 35	So	r	Mo	E	L	y s	5		C		c 4		C	y s	L	y s	A	la	S	г		15	T	уг	S	c r	р	h c
Thr	G 1 S		уг	TI	аг	Mo	: L	A	S T			5 5	V	a	1	L	y s	G	1 1	S	сг		is 50	GI	у	G	In	A	ş n	L	сц
G I u 6 5	Тґ	ρl	l c	GI	у	L	υ		1 0		A	s n	P	r	0	Т	ут	A	\$ n		1 y 7 5	G	у	T	1 1	S	сг	Т	Уr		s n 8 0
Gln	Ly	s P	h c	Ly	s		l y 8 5	L	у	S	A	la	Т	h	r	L	сц		ы г 9 0		a l	Α :	s p	L	y s	S	ст		c r 9 5	A	\$ n
Thr	AI	a T	ут	М с 1 С		G	u	L	CI	u	L	сu	S	С	ĩ		с и) 5	Т	b r	S	ст	A	ła	A	s p		c r 1 0	A	la	v	al
Tyr	Ту		ys 15	T	г	A	r g	A	Γį	ŝ	G	ly		ь 2		A	rg	A	s p	Т	уг	S	cr	M d		A	s p	Т	уг	Т	r p
Gly	G 1 1 3		ly	ТІ	1 1	S	1 3	v	8			br 35	v	۵	1	S	c f	S	C T												

(2) INFORMATION FOR SEQ ID NO:86:

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(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 108 amino acids

- (B) TYPE: ann'uno acid (C) STRANDEDNESS: single
- (D) TOPOLOGY: luncar

(i i) MOLECULE TYPE: peplide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:86:

GiuileValLeuThrGinSerProGiyThrLeuSerLeuSerProGiyGiuArgAlaThrLeuSerCysArgAlaSerGinSerValSerSerGiyGiuArgAlaThrLeuSerCysArgAlaSerGinSerValSerSerGiyTyrLeuGiyTrpTyrGinGinLysProGiyGinAlaProArgLeuLeuLeuGiyTrpTyrGinGinLysProGiyGinAlaProArgLeuLeuLeuGiyAiaSerSerArgAlaThrGiyGinAlaProArgLeuLeuLeuGiyAiaSerSerArgAlaThrGiyGinGinGiyArgLeuGiuSerGiyAiaSerSerArgArgArgPhcArgLeuGiuSerArgLeuGiuGiySerGiySerGiyThrAspPhcThrLeuThrLeuThrSerArgLeuGiuGiySerGiyThrAspPhcThrLuuThrLuuThrLuuSerLeuSerLeuSerGiySerGiy

(2) INFORMATION FOR SEQ ID NO:87:

(i)SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 107 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single

(D) TOPOLOGY: lincar

(1 i) MOLECULE TYPE: popude

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:87:

GluIleVaiLeuThrGinSerProGlyThrLeuSerLeuSerProGlyGluArgAlaThrLeuSerCysArgAlaSerGinSerleuSerAsnAsnLeuHisTrgTyrGinGinLysProGlyGinAlaProArgLeuLeulieLeuHisTrgTyrGinGinLysProGlyGinAlaProArgLeuLeulieLysTyrAisSerGinSerlieSerGiyGinAlaProArgLeuLeulieLysTyrAisSerGinSerlieSerGiyGinAlaProArgLeuLeulieLysTyrAisSerGinSerlieSerGiylieProAspArgLeuLeulieSerGiySerGinSerlieSerGiylieProAspArgLeuLeulieSerGiySerGinSerLieSerGiylieProAspArgLeuGiyProSerGiySerGinSerLieThrLeuThrLeuThrlieProAspArgLeuGiyProGiuAsp

(2) INFORMATION FOR SEQ ID NO:88:

(i) SEQU'ENCE CHARACTERISTICS:
 (A) LENGTH: 122 amino acids
 (B) TYPE: amino acid

(C) STRANDEDNESS: single (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: pepilde

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:88:

Gin Val Gin Leo Mei Gin Ser Gly Ala Giu Val Lys Lys Pro Gly Ser

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Scr	Val	Arg	Val 20	Scr	Суѕ	Lys	Th r	Scr 25	G1 y	Gly	Thr	Pbc	V a I 3 0	Asp	Туг
Lys	G 1 y	L c u 3 5	Тгр	Va I	Arg	Gln	A I a 4 0	Pro	Gly	Lys	Gly	L C U 4 5	Glu	Тгр	V a 1
Gly	Gl 1 50	llc	Pro	Lcu	Arg	Pbc 55	Asn	G 1 y	Glu	Vəl	L y s 6 0	Asp	Pro	Gly	Scr
Val 65	Val	Arg	Val	Scr	Val 70	Scr	Lcu	Lys	Pro	Sст 75	Рһс	Asn	Gle	Ala	H i s 80
McL	Glu	Lcu	Scr	Scr 85	Lcu	Phc	Scr	Glu	Asp 90	Tbr	Ala	Val	Tyr	Туг 95	Cys
Ala	Arg	Glu	Туг 100	Gly	Phc	Αsp	Thr	Scr l O S	Asp	Туг	Туг	Тут	Туг 110	Туг	Тгр
Gly	Glu	GI y 115	Tbr	Lcu	Val	Tbr	Val I20	Scr	Sст						
NFORMATIO	N FOR	SEQ 1D I	NO:89:												
(i)\$E	(A) (B) (C)	LENGT TYPE: a STRAN	ACTERIS H: 119 an m ino acid DEDNES: DGY: linc	imo acids 1 S: single											

(i i) MOCECULE TYPE: popuide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:89:

G 1 n 1	Val Glm	Leu Val G 5	In Ser Gly Ala	Glu Val Lys L 10	ys Pro Gly Scr IS
Sст	Val Arg	y 21 Ser C 20	ys Lys Ala Scr 25	Gly Tyr Scr P	bc Thr Gly Tyr 30
Tbr	Mici Asa 35	a Trp Vel A	rg Gln Ala Pra 40		.cu Glu Trp Val 15
Gly	Leu Ile 50	c Asn Pro T	yr Asn Gly Gly 55	Thr Ser Tyr A 60	sn Gln Lys Phe
Lys 65	Gly Arg	-	al Ser Leu Lys O	Pro Scr Phc A 75	Asn Gln Ale Tyr 80
McL	Glu Lcu	u Scr Scr L 85	cu Phe Ser Glu	Asp Thr Ala V 90	al Tyr Tyr Cys 95
Thr	Arg Arg	g Gly Рhс А 100	rg Asp Tyr Ser 105	Мсі Аѕр Туг Т	rp Gly Gla Gly 110
Thr	Leu Val 115	1 Thr Val S 5	сг Sст		

(2) INTORMATION FOR SEQ ID NO:90:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 129 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA (oligonucleotide)

(x i 1 SEQUENCE DESCRIPTION: SEQ ID NO:90:

TAGATCTAGA CCACCATGGT TTTCACACCT CAGATACTCA GACTCATGCT CT1CTGGATT 60 TCAGCCTCCA GAGGTGAAAT TGTGCTAACT CAGTCTCCAG GCACCCTAAG CTTATCACCG 120 GGAGAAAGG 129

			5,530,101			
	147		-continued		148	
(2) INFORMATION I	FORSEQ ID NO:91:					
	JENCE CHARACTERISTI (A) LENGIH: 128 basc (B) TYPE: nucleic acid (C) STRANDEDNESS: 1 (D) TOPOLOGY: linear	pairs				
(ii) MOLI	ECULE TYPE: DNA (oiigo	nucleotide)				
(x i) SEQU	IENCE DESCRIPTION: SE	EQ ID NO:91:			÷	
Ϋ́AGACAGAAT	TCACGCGTAC	T T G A T A A G T A	GACGTGGAGC	T T G T C C A G G T	T T T T G T T G G T	60
ACCAGTGTAG	GTTGTTGCTA	ATACTTTGGC	TGGCCCTGCA	GGAAAGTGTA	GCCCTTTCTC	120
CCGGTGAT						128
(2) INFORMATION I	FOR SEQ ID NO:92:					
	ENCE CHARUACIERISTI (A) LENG'III: 113 base (B) TYPE: nucleic acid (C) STRANDEDN'ESS: : (D) TOPOLOGY: linear	pairs				
(ii)MOL	ECULE TYPE: DNA (of ag	onucleolide)				
(xi)SEQU	ENCE DESCRIPTION: SE	EQ ID NO:92;				
AAGAGAATTC	ACGCGTCCCA	GTCCATCTCT	GGAATACCCG	A T A G G T T C A G	TGGCAGTGGA	6 0
TCAGGGACAG	ATTTCACTCT	CACAATAAGT	AGGCTCGAGC	CGGAAGATTT	TGC	113
(2) INFORMATION I	FOR SEQ ID NO:93:					
	IENCE CHARACIERIST (A) LENGTH: 116 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pa`ws				
(ii)MOL	ECULE TYPE: DNA (oligo	vnuclcutide)				
(xi)SEQL	JENCE DESCRIPTION: SI	EQ ID NO:93:				
ΤΑ G Α T C T A G A	GTTGAGAAGA	CTACTTACGT	ΤΤΤΑΤΤΤΟΤΑ	CCTTGGTCCC	TTGTCCGAAC	6 0
GTATGAGGCC	A A C T G T T A C T	CTGTTGACAA	TAATACACAG	CAAAATCTTC	CGGCTC	116
(2) INFORMATION	FOR SEQ ID NO:94.					
	DENCE CHARACIERISTI (A) LENGTH: 134 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA (oligo	anucleolide)				
(x i) SEQU	JENCE DESCRIPTION: SI	EQ ID NO:94:				
TATATCTAGA	CCACCATGGG	ATGGAGCTGG	ATCTTTCTCT	TCCTCCTGTC	AGGAACTGCA	60
GGTGTCCACT	CTCAAGTCCA	ACTGGTACAG	TCTGGAGCTG	AGGTTAAAAA	GCCTGGAAGT	120
TCAGTAAGAG	TTTC					I 3 4
(2) INFORMATION	FOR SEQ ID NO:95:					

(i) SEQUENCE CHARACIERISTICS:
 (A) LENGTH: 134 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

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(ii) MGL	ECULE TYPE: DNA (o	ligonuclcolide)										
(xi)SEQU	ENCE DESCREPTION	: SEQ ID NO:95:										
TATATAGGTA	CCACCATGG	G ATGGAG	CTGG	ATC	тттс	TCT	тсст	сстб	тс .	AGGAA	CTGCA	6 0
тосстотстс	ACCCAGTTC	A TOGTAT	ACCC	AGT	GAAT	GAG	ΤΑΤΟ	CGGA	AG	стттс	CAGGA	120
AACTCTTACT	GAAC											134
(2) INFORMATION	FOR SEQ ID NO. 96:											
	VENCE CHARACTER (A) LENGTH: 116 b (B) TYPE: nucleic at (C) STRANDEDNE! (D) TOPOLOGY: Lin	asc pair; cið SS: single										
(ii) MOL	ECULE TYPE: DNA (o	oligonucle dide)										
(xi)SEQU	ENCE DESCRIPTION	: SEQ ID NO:96:										
T A T A T A G G T A	C C A G C T A C A	A CCAGAA	GTTC	AAC	GGCA	GAG	ΤΤΑΟ	AGTT	тс	TTTG#	AGCCT	6 0
T C A T T T A A C C	A G G C C T A C A	T GGAGC1	r c a g t	AGI	CTGT	ттт	СТСА	AGAC	AC	TGCAC	бт	116
(2) INFORMATION	FOR SEQ ID NO:97:			-								
	IENCE CHARACTERI (A) IENGTH: 116 b (B) TYPE: nucleic at (C) STRANDEDNE: (C) TOPOLOGY: lin ECULE TYPE: DNA (c)	asc pairs cid SS: single car										
(xi)SEQ	JENCE DESCRIPTION	I: SEQ ID NO. 97:										
ΤΑΤΑΤΟΤΑΘΑ	GGCCATTCT	T ACCTGA	GGAG	ACO	GGTGA	СТА	AGGI	TCCT	ΤG	ACCCC	CAGTAG	6 0
TCCATAGAAT	A G T C T C G A A	A CCCCC	JTCTT	GTA	CAGT	ΑΑΤ	AGAO	TGCA	GТ	GTCT	0.01	116
(2) INFORMATION	FOR SEQ ID NO:98:											
	JENCE CHARACIERI (A) LENGTH: 408 b (B) TYPE: nocleic a (C) STRANDEDNE (D) TOPOLOGY: in	nase pairs cid SS: single										
(ii) MOL	ECULE TYPE: cDNA											
	URE: (A) NAME/KEY: CI (B) LOCATION: 1.4											
(xi)SEQ	UENCE DESCRIPTION	* SEQ ID NO:98:										
ATG CAT CA Mict His GI	n Thr Scr				Mc ı					Leu		4 8
1	5				1 0					15		
TTC ATA TC Phe lle Se												9 (
ATG ACC CA Mct Thr Gi 3	n Scr Pro											144
ACC TTG AG Thr Lcu Sc S												19:
TAT CAA CA Tyr Gln GI 65												24

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					G G G G 1 y											288
					C 7 G L c u											336
					G G А G 1 у											384
					GΛΛ GΙυ											408
(2)1	NFORMA	TION FO	RSEQI	D NO:99:												
	(i	()	A) LENG B) TYPE		anúno aci cid	ids										
	(i i) MOLEC	ULE TY	PE: protes'	n											
	(x i) SEQUE	NCE DES	CRIPTIO	N: SEQ I	D NO:99;										
Μcι 1	His	G 1 1	Thr	Scr 5	Μcι	GΙγ	110	Ĺys	Мсі 10	Glu	Scr	GIn	Tbr	L C U 1 5	V a 1	
РЬс	IIc	Scr	Ilc 20	Lcu	Lcμ	Ттр	Lcυ	Тут 25	Gly	Ala	Asp	Gly	A s n 3 0	Il c	Val	
Мсі	Thr	G 1 n 3 5	Scr	Pro	Lys	Scr	Mc 1 40	Туг	Val	Ser	11 c	G I y 4 5	Glu	Arg	Val	
ТЪг	Lcu 50	Sст	Суз	Lys	Ala	S c 1 5 5	GΙψ	Asn	Val	Asp	Т b т 60	Туг	Val	Scr	Trp	
Тут 65	Gla	GIn	Lys	Рто	G 1 u 7 0	Gln	Scr	Pro	Lys	L c u 7 5	Lси	llc	Тут	G 1 y	A 1 a 8 0	
Sст	Asm	Λтg	Тут	Т ћ т 8 5	Gly	Val	Рто	Λsp	Arg 90	Phc	Thr	GIy	Scr	G 1 y 9 5	Scr	
Ala	Thr	Asp	Р b с 100	Thr	Lси	Thr	1 J c	Scr 105	Scr	Vel	Gln	Ala	G l u I I 0	Asp	Lcu	
A 1 a	A s p	T y r 1 1 5	His	Cys	Gly	Gla	Scr 120	Туг	Asn	Туг	Pro	Р h с 1 2 5	Thr	Pbc	Gly	
Scr	G l y I 3 0	Thr	Lys	Lcu	Glu	1 I c 1 3 5	Lys									
(2)	FORMA	TION FC	or seq I	D NO:100):											
	(i	()	A) LENG B) TYPE C) STRA	STH: 456 : nucleic	base Poin acid ESS: singl											
	(i i) MOLE(OULE TY	PE: cDNA												
	(i x		A) NAM	E/KEY: (
	1 - :			ATION: 1		ר אים ומ	1.									
ATC					DN: SEQ I			ΤΤΛ	666	AGC		CAG	GAC	CTC	666	48
					Scr											48
					ATC Ilc											96

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								C A G G I n								144
								TGC Cys								192
								AAG Lys								2 4 0
								TCC Scr								288
								СТ G L с u 1 0 5								336
								C T G L c u								384
								C C C P r a								432
			GTC Val													456
(2)1			OR SEQ I													
	(i	(A) LEN B) TYP D) TOP	GTH: 152 E: amino :	am'ino ac acid	abi										
			CULE TY			0.10.10										
Mic L I			L c u					Lcu	Pro 10	Scr	Thr	Gln	Asp	L c u 1 5	Ala	
Мсı	G 1 y	Тгр	S c r 2 0	Суз	11 c	11 c	Leu	Phc 25	Lcu	Val	Ata	Thr	A I n 3 0	Ύbr	G 1 y	
Val	Lcu	S c r 3 5		Vat	GIR	Гсв	Gln 40	Glp	Pro	Gly	Ala	A s p 4 5	Lcu	V a l	Mcı	
Pro	Gly 50		Pro	Val	Lys	L c u 5 5		Cys	Lcu	Ala	Sст 60		Туг	11c	Phc	
Thr 65	Scr	Sст	Trp]] c	Asn 70		Val	Lys	Gln	Arg 75		G 1 y	Arg	Gly	L с и 8 0	

 Val
 Leu
 Ser
 Gin
 Val
 Gin
 Leu
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 Gin
 Gin
 Gin
 Gin
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 Asp
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 Val
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 Leu
 Val
 Met

 Pro
 Gin
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 Aia
 Ser
 Gin
 Aia
 Ser
 Gin
 Tyr
 Iic
 Pho

 Thr
 Ser
 Ser
 Trp
 Iic
 Asn
 Trp
 Val
 Lys
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 Arg
 Pro
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 Arg
 Gin
 Arg
 Pro
 Gin
 Arg
 Gin
 Lucu
 Arg
 Gin
 Lucu
 Arg
 Gin
 Lucu
 Arg
 Gin
 Lucu
 Arg
 Gin
 Arg
 Pro
 Gin
 Arg
 Gin
 Lucu
 Arg
 Gin
 Arg
 Gin
 Arg
 Gin
 Lucu
 Arg
 Gin
 Arg

(2) INFORMATION FOR SEQ ID NO:102:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 107 amino acids

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(B) TYPE: amino acid (C) STRANDEDNESS: single (O) TOPOLOGY: linear

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:102:

(i i) MOLECULE TYPE: peptide

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-continued
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Asp
1IleGlnMeiThr
5GlnSerProSerThr
10LeuSerAlaSerVal
15SerVal
16GlnAspArgValThrIleThrCysArg
40AlaSerGlnSerJleAsnTbrTrpLeuAlaTrp
35TyrGlnGlnLysArg
40AlaSerGlnSerJleAsnTbrTrpLeuAlaTrp
35TyrGlnGlnLysProGlyLysAlaProLysLeuMetTyrSoAlaSerSerLeuGlnSerGlyLysAlaProLysLeuMetSerSoAlaSerSerLeuGlnSerGlyValProSerArgPheIleMetSerSoSerGlySerGlySerGlyValProSerArgPheIleGlySerSoSerGlySerGlySerGlySerAlaSerAlaProSerSoSerGlySerGlyPheSerSerAlaProSerAlaSerSerGlyTrySerTryCysGlyTryIleSerAlaSerAlaAspSerGlyTryTyrCys

(2) INFORMATION FOR SEQ ID NO:103:

(i) SEQUENCE CHARACIERISTICS:

(A) LENGTH: 107 amino acids

(B) TYPE: wnino sold (C) STRANDEDNESS: single

(D) TOPOLOGY: l'incar

(i i) MOLECULE TYPE: pe pide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:103:

Asp
IIIcGInMetThr
SGInSerProSerThr
IOLeuSerAIaSerValGIyAspArgValThr
20IIcThrCysLysAlaSerGIuAsnValAsnValAsp
30ThrTyrValSerTrp
3STyrGInGInCysLysAlaSerGIuAsnValAsp
30ThrTyrValSerTrp
3STyrGInGInLysProGIyLysAlaProLysLeuLeuIIcTyrG1y
50AlaSerAsnArgTyr
55ThrGIyValProSerArgPhcSerGIySerG1y
50SerGIySerAsnArgTyr
70ThrLeuThrLysLeuIcuIcuSerGIy
65SerGIyThrAspPhcAlaThrAspPhcSerGIyAspAspPhcAlaThrAspPhcThrLeuThrLucThrSerGISerGISerGIy
50SerGIyThrAspPhcThrLucThrLucLucLucIcuSerSerGIy
50SerGIyThrAspThrLucThrLucThrSerSer

(2) INFORMATION FOR SEQ ID NO:104:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 117 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) JOPOLOGY: linear

(i i) MOLECULE TYPE: peplide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:104:

```
Gin Val Gin Leu Val Gin Ser Giy Ala Giu Val Lys Lys Pro Giy Ser

I 5 10 10 15

Ser Val Lys Val Ser Cys Lys Ala Ser Giy Giy Thr Phe Ser Arg Ser
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	157	-continued	158	
	2 0	2 5	3 0	
Ala II	cllcTrpVal 35	Arg Gln Ala Pro Gly C 40	Glu Gly Leu Glu Trp 45	Мсь
GI y GI 50		Mei Phe Gly Pro Pro 4 55	Asn Tyr Ala Gin Lys 60	РЪс
Gln Gl 65		llc Thr Ala Asp Glu 5 70		Туг 80
McL G1	u Leu Ser Ser 1 85	Leu Arg Ser Glu Asp 7 90	Thr Ala Phc Туг Рыс 95	Cys
Ala Gl	y Oly Tyr Gly IOO	ile Tyr Ser Pro Glu (105	Glu Tyr Asn Gly Gly IIO	Lcu
Val Tb	r Val Ser Ser 115			
(i) SEQU ((GR SEQ ID NO:105: ENCE CHARACIERISTICS: A) LENGTH: 117 anino acids B) TYPE: amino acid C) STRANDEDNESS: single			
	D) TOPOLOGY: littear		N	
(xi)SEQU	ENCE DESCRIPTION: SEQ ID N	O:105:		
Gln Va I	l Gio Leu Val 5	Gln Scr Gly Ala Glu ' 10	Val Lys Lys Pro Gly 15	Scr
Scr Va	l Lys Val Ser 20	Cys Lys Ala Scr Gly 25	Tyr lle Phe Thr Ser 30	Scr
Ттр I	c Asn Trp Vai 35	Arg Gln Ala Pro Gly (40	Gin Giy Lev Glu Trp 45	Μcι
Gly As 5 (Ser Asp Gly Glu Val 55	His Tyr Asn Gln Asp 60	Pbc
LysA: 65		lle Thr Ala Asp Glu 70	Ser Thr Asn Thr Ala 75	Туг 80
Mc i G	u Leu Ser Ser 85	Leu Arg Ser Glu Asp 90	Thr Ala Vəl Tyr Tyr 95	Cys
Ala A	g Gly Phe Leu 100	Pro Trp Pbc Ala Asp 105	Trp Gly Gla Gly The 110	Lcu

Val Thr Val Scr Scr 115

(2) INFORMATION FOR SEQ 1D NO:106-.

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 115 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA (oligenucleolide)

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:106:

TTTTTTCTAG	ACCACCATGG	AGACCGATAC	CCTCCTGCTA	TGGGTCCTCC	TGCTATGGGT	e	60
CCCAGGATCA	ACCGGAGATA	TTCAGATGAC	CCAGTCTCCG	TCGACCCTCT	CTGCT	1.1	15
						÷ .	

(2) INFORMATION FOR SEQ ID NO:107:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 120 hase pairs (B) TYPE: buckeic acid

150			5,530,101		160		
	159		-continued		100		
	(C) STRANDEDNESS: a (D) TOPOLOOY: linear	single					
(ii) MO1.	ECULE TYPE: DNA (oligo	nuclcouide)					
(xi)SEQ	JENCE DESCRIPTION: SE	Q ID NO:107;					
TTTTAAGCTT	GGGAGCTTTG	CCTGGCTTCT	GCTGATACCA	GGATACATAA	GTATCCACAT	6 0	
TTTCACTGGC	CTTGCAGGTT	ATGGTGACCC	TATCCCCGAC	GCTAGCAGAG	AGGGTCGACG	120	
(2) INFORMATION	FOR SEQ ID NO:108:						
2.0	UENCE CHARAC TERISTI (A) LENGTH: 118 base 1 (B) TYPE: nucleic acid (C) STRANDEDNESS; s (D) TOPOLOGY: linear	pairs					
(ii) MOL	ECULE TYPE: DNA (oligo	nuclcotide)					
(x i) SEQ	UENCE DESCRIPTION: SE	Q ID NO:108:					
T T T T A A G C T 'l'	CTAATTTATG	GGGCATCCAA	CCGGTACACT	GGGGTACCTT	CACGCTTCAG	6 0	
TGGCAGTGGA	TCTGGGACCG	A T T T C A C C C T	CACAATCAGC	TCTCTGCAGC	CAGATGAT	1 8	
(2) INFORMATION	FOR SEQ ID NO:109:						
	UENCE CHARACIERISTI (A) LENGTH: 120 base (B) TYPE: nucleic acid (C) SIRANDEDNESS: s (D) TOPOLOGY. linear	pairs					
(ii) MQL	ECULE TYPE: DNA (oligo	nucle@Eide)					
(x i) SEQ	UENCE DESCRIPTION: SE	EQ ID NO:109:					
TTTTTTCTAG	AGCAAAAGTC	TACTTACGTT	TGACCTCCAC	CTTGGTCCCC	TGACCGAACG	6 0	
TGAATGGATA	GTTGTAACTC	TGTCCGCAGT	A A T A A G T G G C	GAAATCATCT	GGCTGCAGAG	120	
(2) INFORMATION	FOR SEQ ID NO:110:						
(i) SEQ	UENCE CHARACTERISTI (A) LENGTH: 114 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOFOLOGY: linear	pairs	£				
(ii) MOI	ECULE TYPE: DNA (oligo	nuclcolide)					
(xi)SEQ	UENCE DESCRIPTION: SE	EQ ID NO:110:					
TTTTTCTAGA	CCACCATGGG	ATGGAGCTGG	A T C T T T C T C T	TCCTCCTGTC	AGGTACCGCG	6 0	
G G C G T G C A C T	CTCAGGTCCA	GCTTGTCCAG	TCTGGGGCTG	AAGTCAAGAA	ACCT	114	
(2) INFORMATION	FOR SEQ ID NO:111:						
(i)SEQ	UENCE CHARACTERISTI (A) LENGTH: 121 base (B) TYPE nucleic acid (C) STRANDEDNESS: 9 (D) TOPOLOGY: linear	paurs					
(ii) MOL	ECULE TYPE: DNA (oligo	onucleoside)					
(xi)SEQ	UENCE DESCRIPTION: SE	EQ ID NO:111:					
TTTTGAATTC	TCGAGACCCT	GTCCAGGGGC	CTGCCTTACC	CAGTTTATCC	AGGAGCTAGT	6 0	
AAAGATGTAG	CCAGAAGCTT	TGCAGGAOAC	CTTCACGGAG	CTCCCAGGTT	T C T T G A C T T C	120	

BI Exhibit 1136

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(2) INFORMATION FOR SEQ ID NO:112:

(i) SEQUENCE CHARACIERISTICS:

- (A) LENGIH: 137 base pairs
 - (B) TYPE: nuclei's acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
 - D / 10/02001. 200

(i i) MOLECUI.E TYPE: DNA (eligenucicelide)

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:112:

TTTTGAATTC	TCGAGTGGAT	GGGAAGGATT	GATCCTTCCG	ATGGTGAAGT	ΤΟΑΟΤΑΟΑΑΤ	60
CAAGATTTCA	AGGACCGTGT	ΤΑCΑΑΤΤΑCΑ	GCAGACGAAT	CCACCAATAC	AGCCTACATG	120
GAACTGAGCA	GCCTGAG					137

(2) INFORMATION FOR SEQ ID NO:113:

(i) SEQUENCE CHARACIERISTICS:
 (A) LENGIH: 134 base pairs
 (B) TYPE: puckele acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(i i) MOLECUTE TYPE: DNA (oligonucleolide)

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:113:

TTTTTCTAGA GGTTTTAAGG ACTCACCTGA GGAGACTGTG ACCAGGGTTC CTTGGCCCCA 60 GTCAGCAAAC CAGGGCAGAA ATCCTCTTGC ACAGTAATAG ACTGCAGTGT CCTCTGATCT 120 CAGGCTGCTC AGTT 134

What is claimed is:

1. A humanized immunoglobulin having complementarity determining regions (CDRs) from a donor immunoglobulin and heavy and light chain variable region frameworks from human acceptor immunoglobulin heavy and light chain frameworks, which humanized immunoglobulin specifically ⁴⁰ binds to a human interleukin-2 receptor with an affinity constant of at least 10^7 M^{-1} and no greater than about two-fold that of the donor immunoglobulin heavy chain variable region framework is 65% to 95% identical to the sequence of the donor immunoglobulin heavy chain variable region framework.

2. A humanized immunoglobulin according to claim 1 which is an antibody comprising two light chain/heavy chain dimers. 50

3. A humanized mimunoglobulin according to claim 1, which specifically binds to a human interleukin-2 receptor with an affinity of at least 10^8 M^{-1} .

4. A humanized immunoglobulin according to claim 1, wherein said human interleukin-2 receptor is the p55 Tac ⁵⁵ interleukin-2 receptor.

5. A humanized immunoglobulin according to claim 1, wherein said human interleukin-2 receptor is the p75 interleukin-2 receptor.

6. A humanized immunoglobulin according to claim 1,

³⁵ wherein said donor immunoglobulin is the anti-Tac antibody.

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7. A humanized immunoglobulin according to claim 1, wherein said acceptor immunoglobulin heavy and light chain frameworks are from the same human antibody.

8. A humanized immunoglobulin according to claim 7, wherein said human antibody is the Eu human antibody.

9. A humanized immunoglobulin according to claim **1** which is substantially pure.

10. A humanized immunoglobulin according to claim 1 that competitively inhibits the binding of interleukin-2 to said human interleukin-2 receptor.

11. A recombinant immunoglobulin which specifically binds to the p55 chain of the human IL-2 receptor, wherein the amino acid sequence of the mature light chain variable region is:

DI QMTQSPSTLSASVGDRVTITCSASSSIS YMHWYQQKPGKAPKLLIYTTSNLASGVPAR FS GS GSGTEFTLTISSLQPDDFATYYCHQR ST YP LTFGQGTKVEVK.

12. A recombinant immunoglobulin which specifically binds to the p55 chain of the human IL-2 receptor, wherein the amino acid sequence of the mature heavy chain variable region is:

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Q	V	Q	L	v	Q	S	G	Α	Ε	V	К	К	Р	G	S	S	V	К	v	S	С	К	Α	S	G	Υ	Т	F	Т
S	Y	R	М	Н	W	۷	R	Q	Α	P	G	Q	G	L	Ε	W	I	G	Y	I	N	Р	S	т	G	γ	Т	Е	Y
N	Q	К	F	K	D	К	Α	Т	I	Т	Α	D	E	S	Т	N	Т	Α	γ	М	E	L	S	S	Ł	R	S	E	D
Т	Α	v	Y	Y	С	Α	R	G	G	G	v	F	D	Y	W	G	Q	G	Т	Ł	v	Т	v	S	S.				

13. An immunoglobulin which specifically binds to the p55 chain of the human IL-2 receptor, said immunoglobulin having two pairs of heavy and light chains wherein:
 each light chain comprises a complete human kappa chain constant region sequence and a variable region sequence:

D 1 Q M T Q S P S T L S A S V G D R V T I T C S A S S S I S Y M H W Y Q Q K P G K A P K L L I Y T T S N L A S G V P A R F S G S G S G T E F T L T I S S L Q P D D F A T Y Y C H Q R S T Y P L T F G Q G T K V E V K;

and each heavy chain comprises a complete human γ chain constant region sequence and a variable region sequence:

Q V Q L V Q S G A E V K K P G S S V K V S C K A S G Y T F T S Y R M H W V R Q A P G Q G L E W I G Y I N P S T G Y T E Y N Q K F K D K A T I T A D E S T N T A Y M E L S S L R S E D T A V Y Y C A R G G G V F D Y W G Q G T L V T V S S.

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