

## TREATMENT OF ACUTE RENAL ALLOGRAFT REJECTION WITH OKT3 MONOCLONAL ANTIBODY<sup>1</sup>

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Eight cadaver donor renal allograft recipients, who had received azathioprine and prednisone from the day of transplantation, were treated with OKT3 monoclonal antibody (reactive with all mature peripheral blood T cells) at the time of diagnosis of acute rejection. In all cases, loss of essentially all detectable peripheral blood OKT3-reactive cells was noted within minutes after the initial 1- to 5-mg i.v. infusion. Chills and fever invariably occurred following the first or second infusion of monoclonal antibody, but were not noted during the subsequent 10- to 20-day course of therapy, suggesting rapid cell lysis as the etiology of this toxicity.

The established rejection episode was reversed in all cases within 2 to 7 days without addition of any therapy other than OKT3 antibody and despite continued lowering of the steroid dosages. During the subsequent 3- to 12-month follow-up period, further rejection episodes occurred in five of these patients, two of these were irreversible with conventional therapy so that six of the eight allografts continue with excellent renal function.

These preliminary observations suggest that homogeneity, limited dosage requirements, and ease of in vitro monitoring of dosage effects should markedly simplify the use of monoclonal antibody to T cell populations in human allograft recipients. This second generation of antilymphocyte preparations offers the potential for not only increased effectiveness but also the possibility of manipulating specific T cell subsets.

Although some heterologous antisera to human lymphocytes have proved to be effective in delaying or reversing allograft rejection (1, 2), the preparation of these agents has been difficult using conventional immunization techniques. Even the purified IgG fraction from animals immunized with lymphocytes contains not only a heterogeneous group of antibodies to T lymphocytes, but also antibodies reactive with other normal cells as well as extraneous antibodies reflecting the animal's previous immunological activity (3). Therefore, techniques of developing more specific reagents have been sought.

Based upon the recent demonstration by Kohler and Milstein (4) that monoclonal antibody to a specific membrane determinant can be reliably produced using cell hybridization techniques, Kung et al. (5) have produced a panel of monoclonal antibodies specifically reactive with human lymphocyte subpopulations. A phylogenetic screen of these reagents in our

laboratory revealed significant cross-reactivity of some of them with lymphocytes of subhuman primates. In order to evaluate the possible clinical role of monoclonal antibody as an immunosuppressive agent, we previously investigated in cynomolgus renal allograft recipients the effects of OKT4 antibody (6). This antibody is reactive with human T cells having major helper/inducer and T-T collaborative functions (7, 8). By using flow cytometry for monitoring of peripheral blood lymphocytes, we defined the dosage and timing of OKT4 antibody administration required to provide in vivo coating of this specific T cell population now in 10 cynomolgus recipients. With a dosage range of 0.5 to 1.0 mg/kg/day, coating of all reactive cells was observed and residual circulating Ab was usually detectable 24 hr after administration. When OKT4 therapy was started before transplantation, allograft survival was extended to as long as 7 weeks after a 1- to 2-week course of therapy (control survival 8 to 11 days).

Encouraged by the effectiveness, ease of administration, and lack of toxicity in this in vivo model, we have begun a trial of monoclonal antibody therapy in human renal allograft recipients. Although the ultimate goal is to evaluate only selected T cell subset suppression, in order to expose the patients in this initial study to the least risk of ineffective immunosuppression, we have tested OKT3 antibody which is reactive with all mature human T cells (9, 10).

### MATERIALS AND METHODS

Eight cadaver donor renal recipients, who had received azathioprine and prednisone from the time of transplantation, were treated with OKT3 monoclonal antibody at the time of diagnosis of acute rejection. Allograft rejection was suggested in these patients by deterioration in renal function and was confirmed in all patients by histopathological evaluation of tissue obtained by percutaneous needle biopsy.

In the attempt to identify the most effective and least toxic combination of conventional and OKT3 therapy, several dosage schedules were pursued. In the first two patients, azathioprine was administered in a dosage of 10 mg/kg on the day of transplantation and then maintained at 1 to 2 mg/kg/day unless the white blood count fell below 3000/mm<sup>3</sup>. Prednisone was begun at a daily dosage of 2 mg/kg. Beginning on the 5th postoperative day, the dosage was decreased by 10 mg/day to 0.8 mg/kg/day, after which the dosage was more slowly tapered to the maintenance dosage of 0.25 mg/kg/day. Following confirmation of the diagnosis of rejection, OKT3 antibody was administered by bolus i.v. injection in a total daily dosage of 1 to 2 mg for 10 days. In the next two patients, the azathioprine

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mg/kg/day during OKT3 therapy which was administered i.v. in a total daily dosage of 1 to 3 mg for 14 days. In the last four patients, the azathioprine and prednisone dosages were further reduced to 0.4 mg/kg/day during OKT3 therapy. In these patients OKT3 was administered daily for 14 to 20 days at a dosage of 4 to 5 mg/day (Fig. 1). After discontinuing OKT3 therapy, the azathioprine dosage was again increased to 1 to 2 mg/kg/day in the last six patients.

Prior to transplantation, during azathioprine and prednisone therapy, and at frequent intervals after institution of OKT3 antibody therapy, peripheral blood lymphocytes from buffy coat preparations were analyzed for OKT3-reactive cells using flow cytometry (11). Recipient serum was monitored by incubating sequentially diluted sera with normal human peripheral blood lymphocytes, followed by staining with fluoresceinated goat anti-mouse antibody, in order to detect and maintain a circulating level of OKT3 antibody. Effectiveness of therapy was judged by reversal of rejection defined as the day after which consistent improvement in renal function occurred. Percutaneous renal biopsies were performed on all patients after OKT3 therapy.

Toxicity was studied by daily monitoring of recipient complete blood count, blood urea nitrogen, and creatinine, weekly assays of hepatic function and urine protein excretion, and careful observation for any clinical evidence of serum sickness. Serial urine, salivary, and buffy coat specimens were cultured for viral activity as previously described (12).

#### RESULTS

The clinical course of a representative patient treated with OKT3 antibody is depicted in Figure 1. Following cadaver donor renal transplantation in this 41-year-old male, the serum creatinine level fell to normal levels by the 4th post-transplant day. Subsequently, the onset of rejection was suggested by the rising serum creatinine level which occurred in conjunction with decreased urinary output, weight gain, hypertension, and low-grade fever. Allograft biopsy confirmed the diagnosis on the 7th post-transplant day and the initial 5-mg dose of OKT3 antibody was infused i.v. after an i.d. skin challenge was observed to produce no reaction. Approximately 45 min later, an episode of shaking chills with fever to 101 C occurred. In addition, the patient complained of shortness of breath and diffuse wheezes were noted over the lung fields. These symptoms rapidly responded to acetaminophen and antihistamine therapy. The patient had no further chills, fever, or other adverse reactions with subsequent OKT3 infusions.

Sequential monitoring of peripheral blood lymphocytes was begun 15 min after the initial injection. As noted in Figure 1, there was essentially complete loss of OKT3-reactive cells from the peripheral circulation, a condition which persisted throughout the 14-day course of therapy. That this was not attributable to masking or modulation of OKT3 antigen was indicated by the failure of fluorescein conjugates of OKT4 and OKT8 monoclonal antibodies, which bind to other T cell antigens (11), to react with the residual cells. In addition, detectable antibody excess was present throughout the course of therapy during which a total dosage of 70 mg of OKT3 antibody was administered.

The serum creatinine level continued to rise for several days after institution of therapy but the patient's clinical condition rapidly improved with diuresis, weight loss, and improved control of blood pressure being noted within 20 hr of initiation of

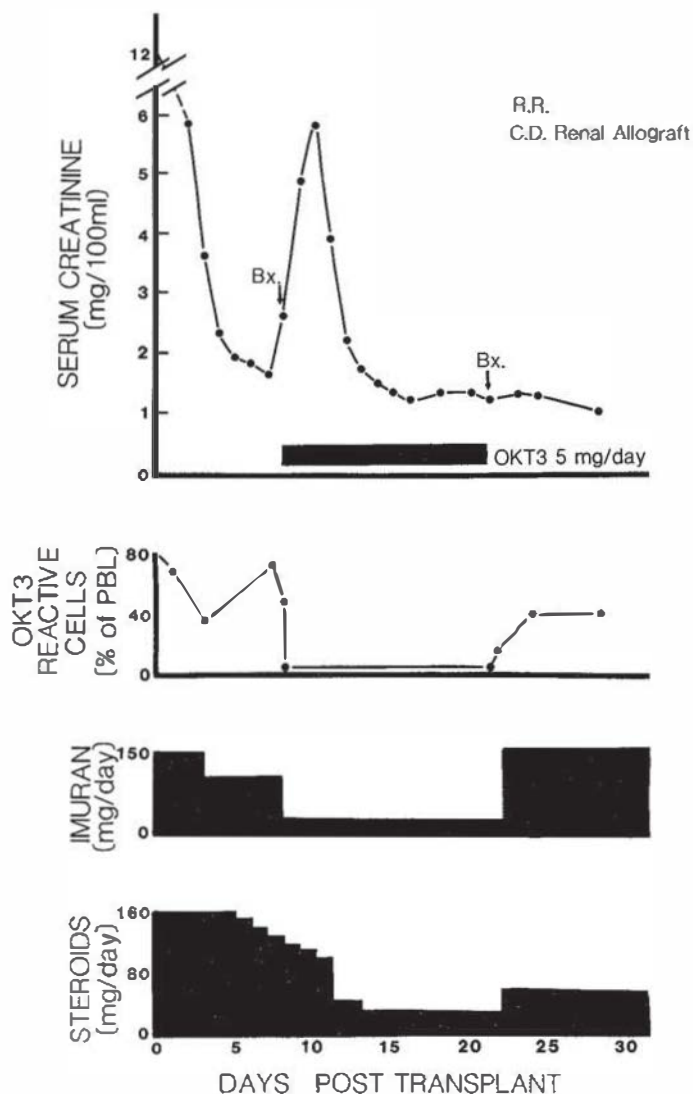


FIGURE 1. Clinical course of renal allograft recipient treated for acute rejection with OKT3 monoclonal antibody. A dramatic and sustained depletion of peripheral blood OKT3-reactive cells and return of normal renal function occurred despite rapid tapering of azathioprine and steroid dosages.

treatment. Continuous improvement in renal function began 72 hr after OKT3 treatment was initiated with the serum creatinine eventually stabilizing at 1.3 mg/100 ml. As depicted in Figure 1, the azathioprine and steroid dosages were rapidly tapered during this period. A second allograft biopsy performed on the last day of therapy showed essentially complete resolution of the histopathological findings of rejection (Fig. 2).

The initial results of treatment of the eight patients studied are summarized in Tables 1 and 2. In every instance, the rejection episode for which OKT3 therapy was instituted was reversed with steady improvement in allograft function beginning after 2 to 7 days of therapy. In the first three patients and the last patient treated, a subsequent rejection episode occurred beginning 2 to 6 weeks after cessation of OKT3 therapy while the patients were being maintained on azathioprine and prednisone. These episodes were easily reversed in three of these patients with increased steroids. The second rejection episode in patient 3, however, could not be reversed despite increased steroids, local irradiation, and acetaminophen D therapy. She



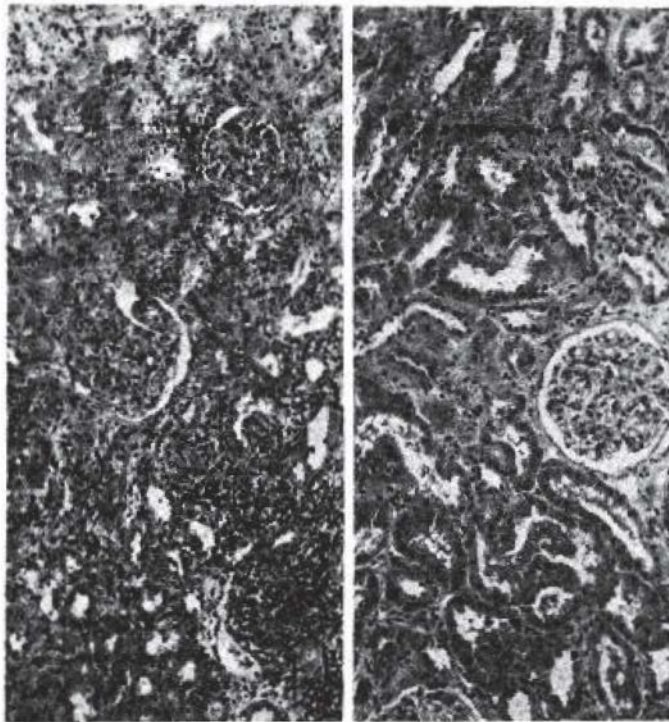


FIGURE 2. Histopathological picture of renal allograft biopsies before (left) and after (right) OKT3 monoclonal antibody therapy. Almost complete disappearance of the interstitial mononuclear infiltrate and reversal of endothelial damage is noted after treatment.

returned to dialysis 2 months after transplantation. She subsequently developed severe cytomegalovirus infection and expired 3 months after transplantation.

Patient 4 is of particular interest because of the development during therapy of an antibody response to the OKT3 reagent. During the initial 10 days of therapy, peripheral blood T cell monitoring of this patient revealed essentially complete loss of cells reactive with OKT3 antibody, and allograft function steadily improved from a peak serum creatinine of 8.1 to 2.9 mg/100 ml. During the final 4 days of OKT3 therapy, however, large numbers of OKT3-reactive cells were repeatedly demonstrable in the patient's peripheral blood and no serum excess of OKT3 could be achieved even after the dosage of monoclonal antibody had been increased from 2 to 5 mg/day. At the time, the steadily falling serum creatinine again began to rise and allograft biopsy revealed extensive evidence of acute rejection. The immunosuppressive protocol was immediately changed to conventional therapy with high-dose steroids, local irradiation, and actinomycin D; but the rejection process continued, necessitating allograft nephrectomy 31 days after transplantation. Evaluation of serial serum samples from this patient by flow cytometry documented the appearance of anti-OKT3 antibody initially on day 10 of therapy with the titer peaking 5 days after therapy was discontinued. Characterization of these antibodies will be reported in detail elsewhere. No evidence of serum sickness or anaphylaxis was noted at any time in this patient.

Therefore, six of the eight allografts continue with excellent renal function 3 to 12 months after OKT3 treatment. All of the patients treated with OKT3 antibody developed chills and fever

TABLE 1. Cadaver donor renal allograft recipients treated for acute rejection with OKT3 monoclonal antibody

Patient	Age	Sex	Weight (kg)	Etiology of renal disease	HLA antigens matched	Follow-up (months)
1	16	Female	55	IgA nephropathy	0	12
2	52	Male	76	Nephrosclerosis	1	12
3	59	Female	50	Interstitial nephritis	1	3 <sup>a</sup>
4	41	Male	70	Chronic glomerulonephritis	1	8 <sup>b</sup>
5	41	Male	78	Chronic glomerulonephritis	2	6
6	39	Female	55	Dysplasia + focal sclerosing glomerulonephritis	2	6
7	52	Male	56	Chronic glomerulonephritis + diabetes	0	4
8	40	Female	51	Polycystic renal disease	1	3

<sup>a</sup> Expired with cytomegalovirus infection after second rejection episode.

<sup>b</sup> Maintained on dialysis after loss of allograft during second rejection episode.

TABLE 2. Results of treatment of acute rejection with OKT3 monoclonal antibody

Patient	Post-transplant day of rejection	Prerejection creatinine (mg/100 ml)	Peak creatinine	Days to reversal	Post-therapy creatinine	Total OKT3		Subsequent rejection episode
						Mg	Days	
1	6	1.6	5.4	4	1.3	13	10	Yes <sup>a</sup>
2	16	1.9	4.8	2	1.6	20	10	Yes <sup>a</sup>
3	8	1.6	4.3	2	0.9	18	15	Yes <sup>b</sup>
4	9	5.0	8.1 <sup>c</sup>	2	2.9	34	14	Yes <sup>b</sup>
5	7	1.6	5.8	2	1.3	70	14	No
6	6	2.9	8.5	5	0.9	57	14	No
7	6	3.0	8.1 <sup>c</sup>	4	1.2	90	17	No
8	9	10.4 <sup>c</sup>	11.3 <sup>c</sup>	7	1.1	100	20	Yes <sup>a</sup>

<sup>a</sup> Reversed with conventional immunosuppression.

<sup>b</sup> Irreversible using conventional immunosuppression.

<sup>c</sup> On hemodialysis



to as high as 102 C within 1 hr of the first injection, and several were noted to have diffuse wheezing during this period. No reactions to subsequent injections were noted and no other evidence of toxicity could be identified.

The effective removal of OKT3-reactive cells (mature T lymphocytes) from the circulation within minutes after infusion of OKT3 antibody was clearly documented by flow cytometry analysis in each case. This dramatic lysis of cells occurred only with the first injection, the OKT3-reactive population then remaining depressed (except in patient 4) throughout the course of therapy.

#### DISCUSSION

A central problem in transplantation remains that of incompletely controlled rejection. Intense, nonspecific suppression of the recipient's immune system is produced with currently used agents but the level of clinical success and toxicity has changed little over the past decade. The only significant addition to this regimen has been the gradual acceptance of heterologous antilymphocyte preparations such as antilymphocyte globulin, antithymocyte globulin (ATG), etc. (13), with which a more selective suppression of the recipient's cellular immune responses is anticipated. However, with currently available agents, prepared by routine immunization techniques, only 5 to 10% of the total dose administered represents the actual therapeutic product. As hybridoma technology has developed, the possibility of producing anti-T cell monoclonal antibodies with effectiveness similar to currently used ATG preparations but which are active in much smaller quantities has been realized. Furthermore, the feasibility of using antibodies to suppress selected T cell subsets rather than the entire population can now be tested.

The results of our studies have begun to delineate the *in vivo* effects of such reagents. We have observed significant immunosuppression in subhuman primate renal allograft recipients receiving relatively minute quantities of monoclonal antibody (total recipient dosage: 17 to 56 mg compared with 50 to 100 mg/kg/day (14) when using heterologous ATG preparations). Moreover, the OKT4 antibody administered to these recipients has been shown to be directed only to the helper/inducer T cell subset, reacting with approximately 50 to 60% of human peripheral blood T cells and  $45 \pm 9\%$  of cynomolgus T cells. These observations demonstrate not only the immunosuppressive potency of monoclonal antibody but also that effective protocols may be developed in which the requirement for agents, which produce indiscriminate T cell depression, might be markedly reduced.

In our previous clinical evaluation of equine ATG, we have found the most definitive means of demonstrating effectiveness was in studies in which ATG alone was added to the immunosuppressive protocol at the time of diagnosis of acute rejection (2). Since only a single agent is being used to reverse a readily defined event, it is possible to elucidate immediately the effect of the added therapy without the need for long-term randomized trials. Thus, we have pursued a similar model for the evaluation of monoclonal antibody. We have selected OKT3 antibody rather than antibody to a T cell subpopulation in order to reduce the likelihood of inadequate suppression in this initial trial. Our most important clinical observation in the patients treated to date has been the initial reversal in all cases of the established rejection episode without addition of any

Whether improved long-term allograft survival can be achieved with such therapy remains to be established; however, some observations regarding the addition of monoclonal antibody to conventional therapy can be made. In the first four patients, the attempt was made to use the minimum total immunosuppression which would produce reversal of the rejection episode. Thus, the dosage of OKT3 antibody administered was limited to that (1 to 2 mg/day) which maintained a barely detectable serum antibody level. In each case, except patient 4 who developed anti-OKT3 antibodies, all clinical manifestations of rejection were abolished. However, post-therapy allograft biopsies showed persistent cellular infiltration and recurrent rejection requiring conventional high-dose steroid therapy appeared within 2 to 6 weeks. All of these patients subsequently suffered significant oral infection secondary to herpes simplex virus and cytomegalovirus pneumonitis was documented after the second rejection episode in two of the patients, one of whom subsequently died. These observations appear to emphasize the previously proposed concept that a modestly high-dose "net state of immunosuppression" over a prolonged period of time is probably of greater risk to patients than a limited, more intensive course of therapy (12). In the next four patients, therefore, the dosage of OKT3 antibody was sharply increased in an attempt to reverse more definitively the rejection activity during a limited period of therapy. In addition, as in our studies with ATG (14), we have continued to try to limit the dosages of azathioprine and prednisone administered during the period in which the patients received OKT3 antibody. To date, the clinical course in these patients has been much more satisfactory.

Unacceptable *in vivo* toxicity of monoclonal antibody administration has not been observed. The chills, febrile response, and occasional wheezing noted on the first day of treatment have been found to be readily controlled with antihistamine and acetaminophen therapy. These symptoms have been noted with ATG treatment as well (15) and have been thought to be secondary to release of endogenous pyrogens following extensive lysis of peripheral blood lymphocytes. The observations in the patients treated with OKT3 antibody would tend to support such an explanation, for the chills and fever were noted only after the first infusion concomitant with the dramatic drop in the number of circulating T lymphocytes. Subsequent infusions were tolerated without similar incidents. It is hoped that later morbidity, primarily infection, can be minimized if an appropriate dosage of monoclonal antibody can be defined which will maximize control of rejection without the addition of other agents. It might then be anticipated that long-term morbidity would be less than with conventional therapy, since it is generally accepted that serious complications are most likely to occur when large doses of steroids are required to reverse rejection activity.

In conclusion, these observations suggest that the homogeneity, limited dosage requirements, and ease of *in vitro* monitoring of the effects of monoclonal antibody upon T cell populations in the peripheral blood will simplify their application to patient management. Although, to date, only a reagent reactive with all peripheral blood T cells has been evaluated in patients, the possibility of manipulating selected T cell subsets is now at hand.

#### LITERATURE CITED

1. Thomas F, Thomas J, Flora R, Mendez-Picon G, Peace K, Lee

2. Shield CF, Cosimi AB, Rubin RH, Tolloff-Rubin NE, Herrin J, Russell PS. Use of antithymocyte globulin for reversal of acute allograft rejection. *Transplantation* 1979; 28: 461.
3. Skamene E, Russell PS. A quantitative study of the binding of ALS to various cell types. *Clin Exp Immunol* 1971; 8: 195.
4. Kohler G, Milstein C. Continuous cultures of fused cells secreting antibody of predefined specificity. *Nature* 1975; 256: 495.
5. Kung PC, Talle MA, DeMaria ME, Butler MS, Lifter J, Goldstein G. Strategies for generating monoclonal antibodies defining human T lymphocyte differentiation antigens. *Transplant Proc* 1980; 12: 141.
6. Cosimi AB, Burton RC, Kung PC, et al. Evaluation in primate renal allograft recipients of monoclonal antibody to human T-cell sub-classes. *Transplant Proc* 1981; 13: 499.
7. Reinherz EL, Kung PC, Goldstein G, Schlossman SF. Further characterization of the human inducer T cell subset defined by monoclonal antibody. *J Immunol* 1979; 123: 2894.
8. Thomas Y, Sosman J, Irigoyen O, et al. Functional analysis of human T cell subsets defined by monoclonal antibodies. I. Collaborative T-T interactions in the immunoregulation of B cell differentiation. *J Immunol* 1980; 125: 2402.
9. Kung PC, Goldstein G, Reinherz EL, Schlossman SF. Monoclonal antibodies defining distinctive human T-cell surface antigens. *Science* 1979; 206: 347.
10. Chang TW, Kung PC, Gingras SP, Goldstein G. Does OKT3 monoclonal antibody react with an antigen-recognition structure on human T cells? *Proc Natl Acad Sci USA* 1981; 78: 1805.
11. Cosimi AB, Colvin RB, Burton RC, et al. Use of monoclonal antibodies to T-cell subsets for immunologic monitoring and treatment in recipients of renal allografts. *N Engl J Med* 1981; 305: 308.
12. Rubin RH, Cosimi AB, Hirsch MS, et al. Effects of antithymocyte globulin on cytomegalovirus in renal transplant recipients. *Transplantation* 1981; 31: 143.
13. Cosimi AB. The clinical value of anti-lymphocyte antibodies. *Transplant Proc* 1981; 13: 462.
14. Cosimi AB, Shield CF, Peters C, Burton R, Scott G, Russell PS. Prolongation of allograft survival by cyclosporin A. *Surg Forum* 1979; 30: 287.
15. Cosimi AB, Wortis HH, Delmonico FL, Russell PS. Randomized clinical trial of ATG in cadaver renal allograft recipients: Importance of T-cell monitoring. *Surgery* 1976; 80: 155.

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