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United States Patent [19]

(54) HTIMANIZED IMMUNOCI ORIIT INS

Queen et al.

[11] Patent Number:

5,530,101

[45] Date of Patent:

Jun. 25, 1996

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(List continued on next page.)

Primary Examiner—Lila Fcisee
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[57] ABSTRACT

Novel methods for producing, and compositions of, humanized immunoglobulins having one or more complementarity determining regions (CDR's) and possible additional amino acids from a donor immunoglobulin and a framework region from an accepting human immunoglobulin are provided. Each humanized immunoglobulin chain will usually comprise, in addition to the CDR's, amino acids from the donor immunoglobulin framework that arc, e.g., capable of interacting with the CDR's to effect binding affinity, such as one or more amino acids which are immediately adjacent to a CDR in the donor immunoglobulin or those within about 3 A as predicted by molecular modeling. The heavy and light chains may each be designed by using any one or all of various position criteria. When combined into an intact antibody, the humanized immunoglobulins of the present invention will be substantially non-immunogenic in humans and retain substantially the same affinity as the donor immunoglobulin to the antigen, such as a protein or other compound containing an epitope.

13 Claims, 55 Drawing Sheets

	HUMANI	ZED INIMIONOGEOBOLENS
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[21]	Appl. No.:	634,278
[22]	Filed:	Dec. 19, 1990
	Rel	ated U.S. Application Data
[63]	abandoned, Feb. 13, 198	n-in-part of Ser. No. 590,274, Sep. 28, 1990, and a continuation-in-part of Ser. No. 310,252, 19, abandoned, which is a continuation-in-part of 0,975, Dec. 28, 1988, abandoned.

	Jel. 140. 290,973, Dec.	20, 1900, availedned.
[51]	Int. Cl.6	A61K 39/395; C07K 16/28
[52]	U.S. CI	530/387.3 ; 530/387.1;
	53	0/388.22; 424/133.1; 424/143.1
1581	Rield of Search	424/85 8 133 1

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1	Q D	I	V Q	L M	T T	Q	S S	P	A S	I	M	S S	A A	S	P V	G	E D	K R	V	T
21 21	I	T T	C	S R	A A	S S	s o	S S	I	N	S T	Y W	M L	H A	W	F Y	Q Q	Q Q	K	P P
40 41	G G	T	S A	P P	K	L L	W	<u>M</u>	Y Y	T K	T A	S S	N S	L L	A E	S S	G G	V V	P	A S
60 61	R R	F F	SI	G G	S	G G	S	G G	T	S	Y F	S	L	T	I	S	R	M L	EQ	A P
80 81	E D	D	A	A A	T	Y Y	Y Y	C	H Q	Q	R Y	S N	T S	Y D	PS	L K	T M	F	G	S Q
100 101	G	T	K	L V	E	L	K K													

FIGURE 1A

1	Q Q	V	Q Q	L L	Q V	Q Q	S	G G	A A	E	L V	A K	K K	P	G G	AS	s s	V	K K	M V
21 21	s s	C	K	A A	S S	G G	Y G	T	F F	TS	S R	Y S	R A	M	H	W	V		Q Q	R A
41 41	P P	G G	Q Q	G G	L L	E E	W	I M	G G	Y G	I	N V	P P	S M	T F	G G	Y P	T P	E N	Y Y
61 61	N A	Q Q	K	F	K Q	D G	K R	A V	T	L	T	A A	D D	K	S S	S	S N	T T	A A	Y Y
81 81	M M	Q E	L	S	s s	L	T R	F S	E E	D D	S	A A	V F	Y Y	У <u>F</u>	C	A A	R G	G G	Y
100 101	G G	G I	V Y	F S	D P	Y E	W E	G Y	N Q	G G	T G	T	L V	T T	V V	S	S S			

FIGURE 1B

1	D E	I	V	L M	Ţ	Q Q	S	P	A A	S	L L	AS	V V	S	1-1 Q1	G G	QE	R	A A	T
21 21	I	S	CC	R	A A	S S	Q Q	S S	V	S S	T	S S	T	Ã Ā	N N	Ā	M M	H H	W	Y Y
41 41	Q Q	Q	K	טי טי	G G	0	P S	P	KR	L	L L	I	K K	Ā Ā	A A	S S	N N		2	S S
61 61	G G	V	ים ים	A	RR	मि वि	S	G G	S S	G G	FS	G	T	D E	Es fes	T	L	N T	I	H
81 81	P R	V L	E	ES	[1] [1]	ם	T	V A	T V	Y	Y	C	Q Q	H	S S	W	E	I	P	Ā
101 101	T	for fer	G	G Q	G G	F1 F1	K R	L V	E	I	K									

FIGURE 2A

FIGURE 2B

1	D D	I	V Q	M M	T	QQ	S	H	K	F	M L	S	T A	S S	V	G	D D	RR	V	S
21 21	I	T	C	K	A A	S S	Q Q	D D	V V	G G	S S	A A	V V	V V	W	# #	Q Q	Q	K	S
41 41	G G	Q K	S A	P P	К . <u>К</u>	L	L	I I	Y	W W	A A	S S	T	R R	H	T	G G	V V	ים ים	D S
61 61	R R	F	T	G G	S	G G	S	G G	T	D E	F	T	L	T	I	TS	N S	V L	Q	S
81 81	E	D D	L F	A A	D T	Y Y	F F	C	0	Q	Y	S S	I	F	D D	L L	T	मा मा	G G	A Q
101	G G	T	R K	L V	E	L	K K													

FIGURE 3A

1	Q Q	V V	Q Q	L	Q V	Q Q	S	D G	A	E	L V	V K	K	P P	G G	s.	S	V	K	I V
21 21	S	C	K K	V A	S	G G	Ā Ā	T	F	T T	D D	H	T	I	H	W	М <u>М</u>	K R	Q	R A
41	יט יט	E G	Q Q	G	-1-1-1	[1] [1]	W W =	71 13	G	Ž Ž	Ţ	Y	נם ונו	?	D D	G G	H	Ţ	R	× -
61 61	SA	E	K	F	K K	G	<u>K</u>	A A	T	L	T	A A	D D	K	S	A T	S	T	A	21 21
81 81	M M	H	L L	N S	S	L	TR	S S	E	D D	S	A A	<u>v</u>	Y Y	F	C	A	2, 2,	G	R 3
101	D D	S S	R R	[13 [21	R	N	G	F	A A	<u>Y</u>	W	GG	00	G G	Ţ	1-1	Y V	T.	V	S
121 121	AS						9													

FIGURE 3B

1 1	D D	I		L	T	Q Q	S S	P	A S	S S	L L	A	V A	S	Ľ V	G	Q D	R R	A V	T T
21 21	I	S T	C	R R	A A	S S	E E	S S	V	D D	N N	Y Y	G G	I	S S	F	M M	N N	W	F F
41 41	Q Q	Q Q	K	ח ח	G	Q K	P A	P	K	L L	L	I	Y	A A	A A	S S	N N	Q	G G	S S
61 61	G G	V	P	A S	R	ra ra	S S	G	S S	G G	S	G	T T	D D	F.	S	L	N N	I	H
8 1 8 1	PS	M	E Q	E	D D	D D	T	A A	M T	Y Y	F Y	C	QQ	00	S S	K K	E	V V	P	W
101	T T	F	G G	GQ	G G	T	K	L V	E E	I I	K K									

FIGURE 4A

1	E Q	V	Q Q	L	Q V	QQ	S	G	p A	E	L V	V K	K	0, 0,	G G	A S	S	V		I
21 21	S S	C	K K	A A	S	G	Ã Ā	T	F	T	D D	Ă Ā	N	M M	H	W	v	KR	Q Q	S A
41 41	7.	G	KQ	S	L	[1]	W W =	I	G	× ×	I	Š	P	Y	N N	G		T	G	> > -
61 61	N N	0	K	F	K	S S	K <u>K</u>	A	T	L	T	V A	D	NE	S	S	S N	T	A A	Y Y
81 81	M	D E	V	R	S S	L	TR	S S	E	D D	S	A A		Y	Ā Ā	C	A	R R	G	R R
101 101	24 0	A	M M	D D	X X	W	<u>G</u>	0	G	T T	S	v v	T	V V	S	S				

FIGURE 4B

1	Q D	I	V Q	L	T T	Q Q	S	P	AS	I	M L	S	A A	S	P	G	E	K R	V	T T
21 21	M I	T	C	S	G G	S	S S	S S	v v	S S	F	M M	Ā Ā	W	Y	Q Q	Q Q	R K	P	G
41 41	S K	SA	P	R	L	L	I	Y	D	T	S S	N	L L	A A	S S	G	V	P	V S	R
61 61	F	S	G G	S	GG	S	G	T	S	Y Y	S	L	T	I	S	RS	M	EQ	A	E
81 81	D D	A	A	T	>1 >1	Ϋ́	C	0	0	W	S S	T T	Ā Ā	P	ī L	T	म्ब ख	G G	A Q	G G
101 101	T	K K	Ľ	ELE	L	K														

FIGURE 5A

1	Q E	V V	Q	L L	K	QE	S S	G G	P. G	G G	L	v v	Q Q	P	S G	QQ	S S	L L	S R	I L
21 21	TS	C		V A	S S	G G	E E	S	v v	T T	S S	Ā Ā	G G	V	H H	W	ī	R	QQ	S A
41 41	ום ום	G G	K K	G	1	[1]	W	L	G G	v	<u> </u>	W	S S	G G	00	S	£1 F	D D	× -	N N
61 61	A A	A A	E fu	I	S S	R	F	T	I	S	KR	D D	N N	S	K	S	Q	V	ř.	for and
81 81	KQ	V M	N N	S	L	QQ	PA	A E	D D	T T	A	I	Y	Y Y	CC	A		A A	G	D D
101 101	Ā Ā	N	× ·	D	G G	fa fa	A A	Ž Ž	W	G	QQ	G	Ţ	L	V	T	v	S S	<u>A</u> S	

FIGURE 5B

1	D E	I	V	14	T	Q	S S	P	A G	T	L L	S S	V	S	D. P	G	DE	SR	V A	S
21 21	L	S	C	R	A A	S S	0	S S	I	S S	N N	N	L	H H	W	Y	Q	Q	K	SP
41 41	H	EQ	SA	P	R R	L	L	I	K K	Y	A A	S	Q	S S	I	S S	G	I	P	S
51 61	R	fa fa	S	G	S	G	S	G	T T	D	F	T	L	S	V	N S	G R	V L	E	T
81 81	E	D D	F	G A	M V	Y	F	C	00	00	S S	N N	S S	W	0, 0,	H H	T T	File	G G	GQ
101 101	G	T T	K	Ľ V	E	I	K													

FIGURE 6A

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E V Q L Q Q S G P E L V K P G A S M K I
Q V Q L V Q S G A E V K K P G S S V R V

21  S C K A S V Y S F T G Y T M N W V K Q S
21  S C K A S G Y S F T G Y T M N W V R Q A

41  H G Q N L E W I G L I N P Y N G G T S Y

41  P G K G L E W V G L I N P Y N G G T S Y

61  N Q K F K G K A T L T V D K S S N T A Y

61  N Q K F K G R V T V S L K P S F N Q A Y

81  M E L L S L T S A D S A V Y Y C T R R G

81  M E L S S L F S E D T A V Y Y C T R R G

81  M E L S S L F S E D T A V Y Y C T R R G

81  F R D Y S M D Y W G Q G T S V T V S S

101  F R D Y S M D Y W G Q G T L V T V S S
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FIGURE 6B

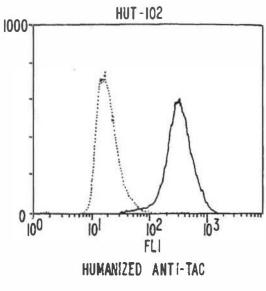


FIGURE 7A

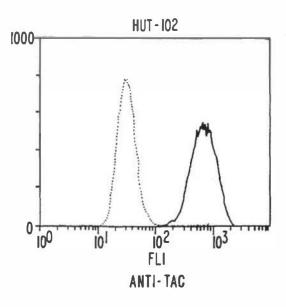


FIGURE 7B

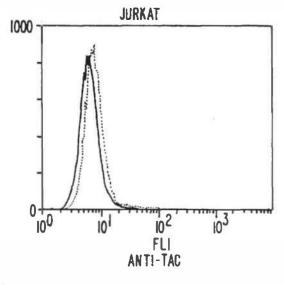


FIGURE 7C

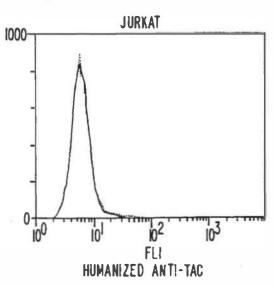


FIGURE 7D

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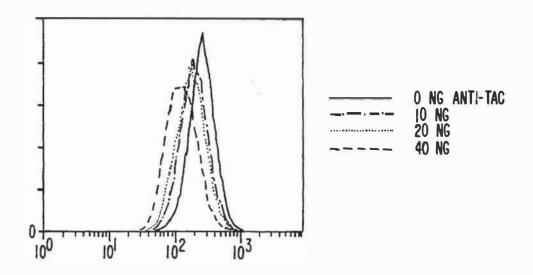


FIGURE 8A

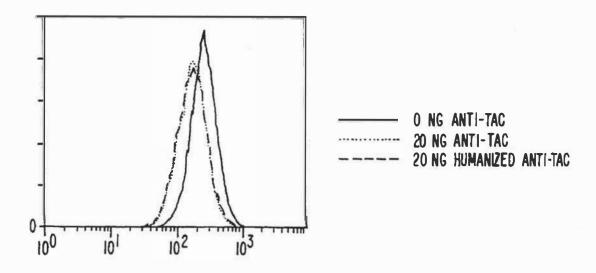


FIGURE 8B

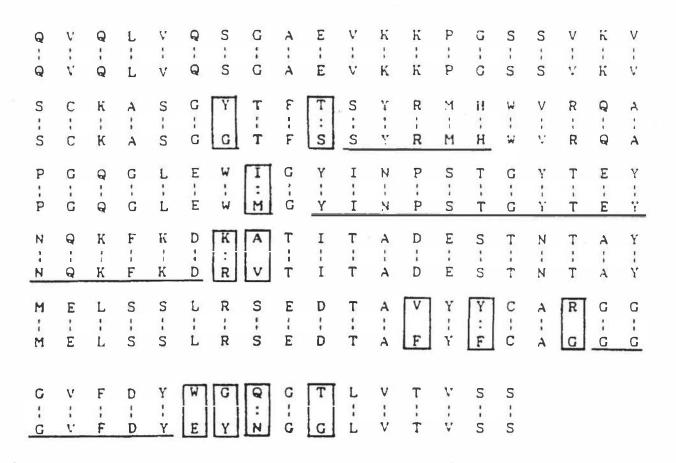


FIGURE 10A

FIGURE 10B

10	20	30	40	50	60	70
AGCTTCTAGA	TGGGATGGAG	CTGGATCTTT	CTCTTCCTCC	TGTCAGGTAC	CGCGGGCGTG	CACTCTCAGG
TCGAAGATCT	ACCCTACCTC	GACCTAGAAA	GAGAAGGAGG	ACAGTCCATG	GCGCCCGCAC	GTGAGAGTCC
80	90	100	110	120	130	140
			AGAAACCTGG			
AGGTCGAACA	GGTCAGACCC	CGACTTCAGT	TCTTTGGACC	GAGCTCGCAC	TTCCAGAGGA	CCTTCCGAAG
150	160		180	190	200	210
			CTGGGTAAGG			
ACCGCCCTGG	AAAAGATCGA	TGTCCTACGT	GACCCATTCC	GTCCGGGGAC	CTGTCCCAGA	CCTTACCTAC
220	230	240	250	260	270	280
GGATATATTA	ATCCGTCGAC	TGGGTATACT	GAATACAATC	AGAAOTTCAA	GGACAGGGTC	ACAATTACTG
CCTATATAAT	TAGGCAGCTG	ACCCATATGA	CTTATGTTAG	TCTTCAAGTT	CCTGTCCCAG	TGTTAATGAC
				10-200		
290	300			330	340	350
			AACTGAGCAG			
GTCTGCTTAG	GTGGTTATGT	CGGATGTACC	TTGACTCGTC	GGACTCTAGA	CTCCTGTGGC	GTAAGATAAA
				414.42		
360	370			400	410	420
			CGAATACAAT			
GACACGTCCC	CCACCCCCTC	AGAAACTGAT	GCTTATGTTA	CCTCCCGACC	AGTGTCAGAG	GAGTCCACTC
430	440					
TCCTTAAAAC						
AGGAATTTTG	GAGATCTGCT	ATA				

FIGURE 11A

10	20	30	40	50	60	70
CAAATCTAGA	TGGAGACCGA	TACCCTCCTG	CTATGGGTCC	TCCTGCTATG	GGTCCCAGGA	TCAACCGGAG
GTTTAGATCT	ACCTCTGGCT	ATGGGAGGAC	GATACCCAGG	AGGACGATAC	CCAGGGTCCT	AGTTGGCCTC
80	90		110	120	130	140
ATATTCAGAT				CGTCGGGGAT		
TATAAGTCTA	CTGGGTCAGA	GGTAGATGGG	AGAGACGATC	GCAGCCCCTA	TCCCAGTGGT	ATTGGACGAG
150	160		180	190	200	210
	AGTATAAGTT			AAGCCAGGCA		
ACGGTCGAGT	TCATATTCAA	TGTACGTGAC	CATGGTCGTC	TTCGGTCCGT	TTCGAGGGTT	CGAAGATTAC
220	230	THE RESIDENCE OF THE PERSON NAMED IN COLUMN 2 IS NOT THE PERSON NA	- Company of the Comp	260	270	280
TATACCACAT	CCAACCTGGC	TTCTGGAGTC	CCTTCTCGCT	TCATTGGCAG	TGGATCTGGG	ACCGAGTTCA
ATATGGTGTA	GGTTGGACCG	AAGACCTCAG	GGAAGAGCGA	AGTAACCGTC	ACCTAGACCC	TGGCTCAAGT
	•					
290	300	310	320	330	340	350
CCCTCACAAT	CAGCTCTCTG	CAGCCAGATG	ATTTCGCCAC	TTATTACTGC	CATCAAAGGA	GTACTTACCC
GGGAGTGTTA	GTCGAGAGAC	GTCGGTCTAC	TAAAGCGGTG	AATAATGACG	GTAGTTTCCT	CATGAATGGG
						38.
360	370	380	390	400		
ACTCACGTTC	CCTCACCCCA	CCAACCTCCA	GGTCAAACGT	AAGTACACTT	TTCTAGATAT	A
	GOICAGGGA	CCAAGGIGGA	GG I ONNHOG I			• •
			A CONTRACTOR OF THE PARTY OF TH	TTCATGTGAA		T

FIGURE 11B

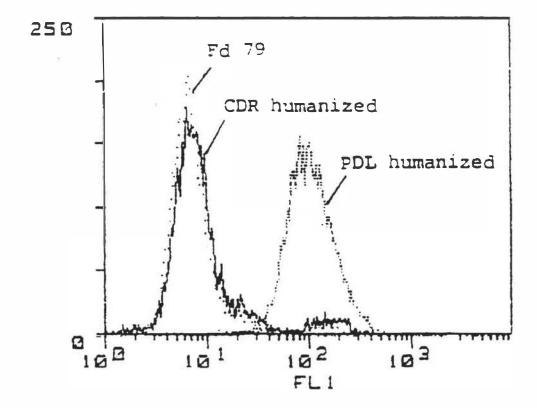


FIGURE 12

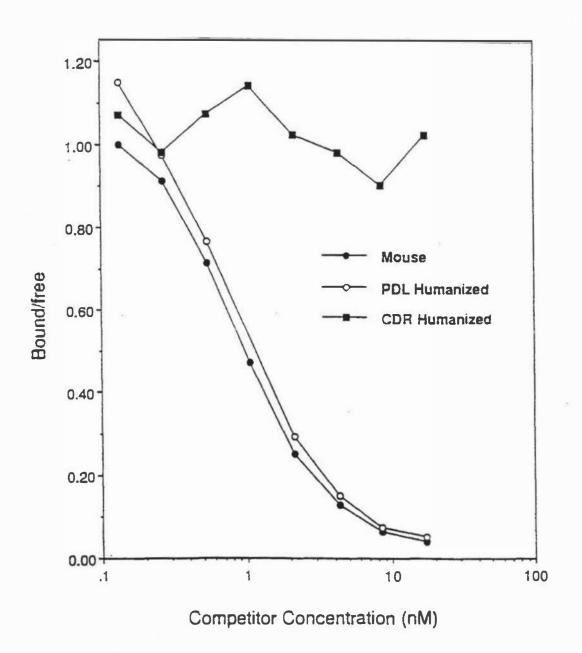


FIGURE 13

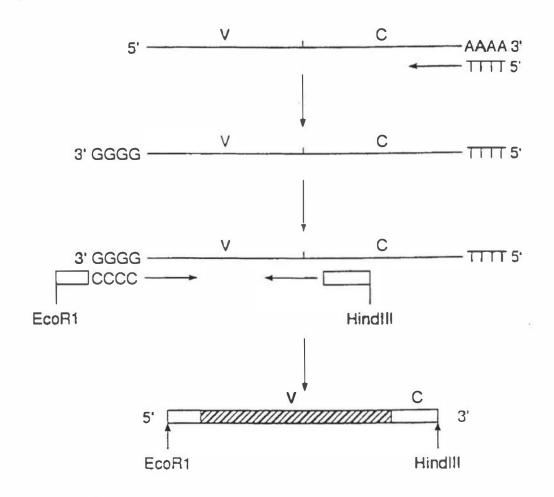


FIGURE 14

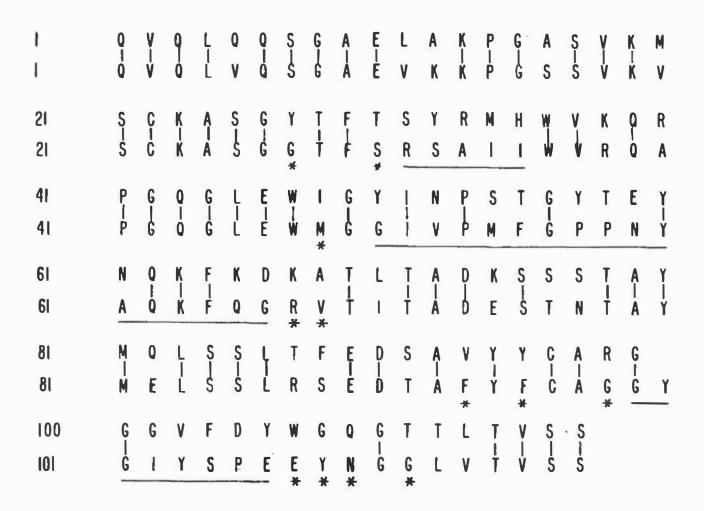


FIGURE 15

1	Q D		V Q	L M	Ţ	0	S	P I P	A S	I T	M L	S S	A A	S S	P V	G	E D	K R	V	T
2I 2I		Ţ 	C	S R	A	S S	S Q	S S	-	N	S	Y	M L	H	W	F Y	0	0	K	P I P
40 41	9	T K	S A	P	K 	[[W	! M *	Y	T K	T A	S	N S	L 	A E	S	G	V	P P	A S
60 61	R R	F	\$ *	G G	\$ \$	G	S S	G	Ţ	S E	Y F	s T	[[Ţ	1	S	R S	M	3	A P
80 81	E	0	A F	A	Ţ	Y	Y I Y	C	H Q	0	R Y	S N	T S	Y D	P S	L K	T M	F	G	S O
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FIGURE 16

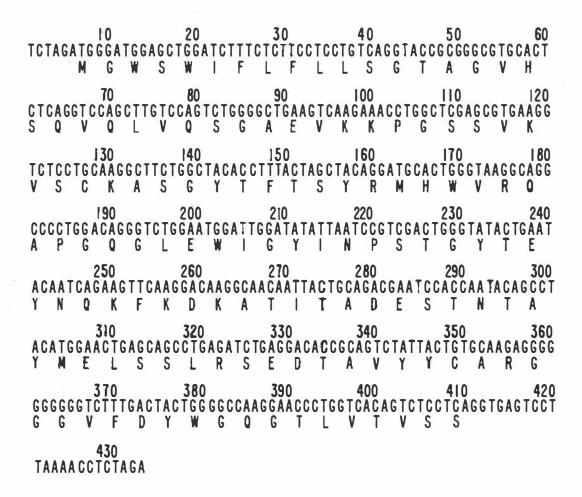


FIGURE 17

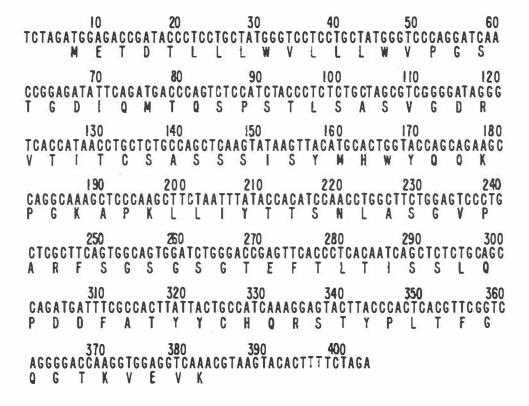


FIGURE 18

HESI5

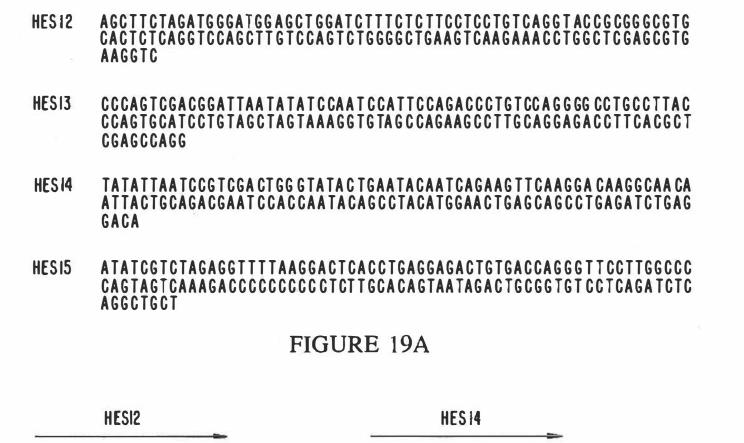


FIGURE 19B

HESI3



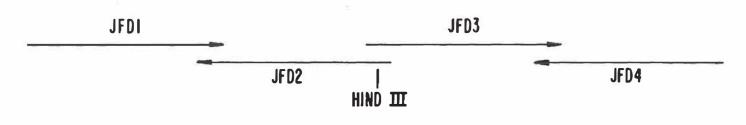


FIGURE 20B

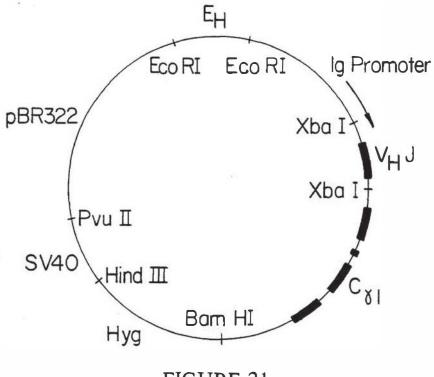


FIGURE 21

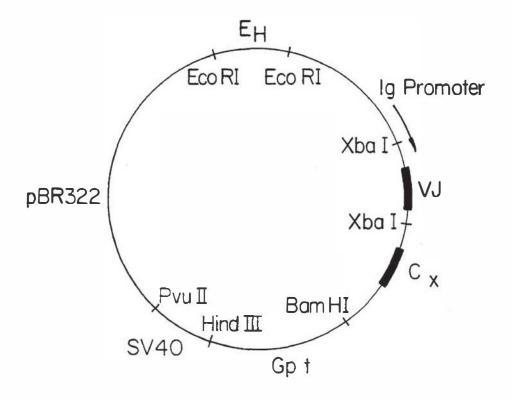


FIGURE 22

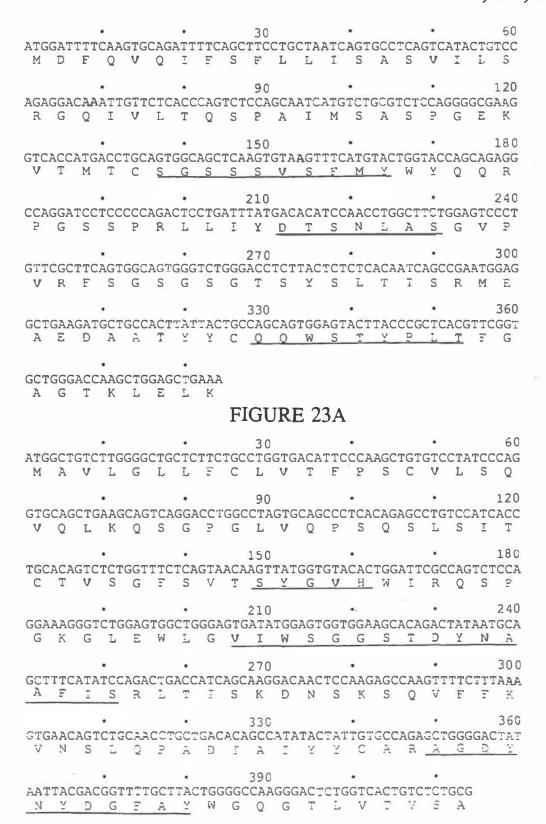
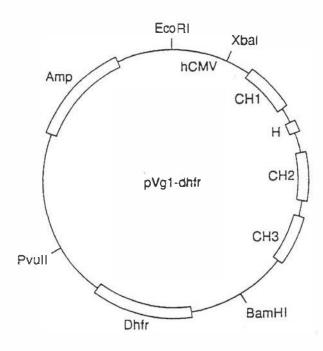


FIGURE 23B



Jun. 25, 1996

FIGURE 24A

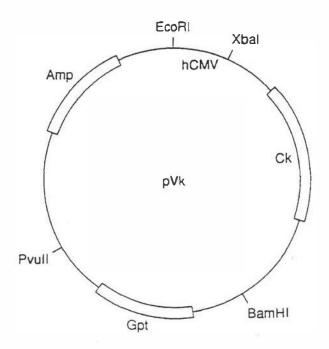


FIGURE 24B

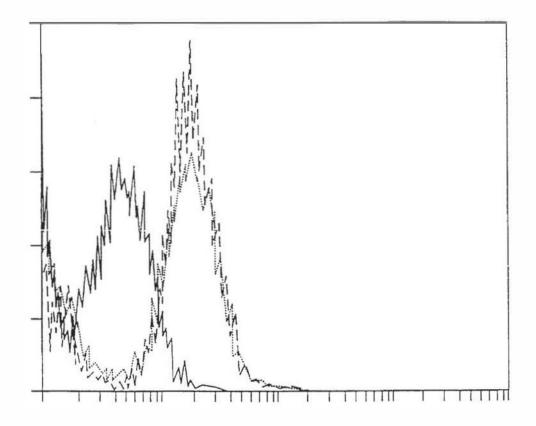


FIGURE 25

1	D I	Q Q	M	T	QQ	S	0, 0,	S S	S	L L	S S	<u>7</u>	S S	V	G	D	R	V	4
21 21	I T	C	Q S	A G	S	Q S	N S	V V	N	A S	Y	L M	Ň	W W	Ā Ā	Q	QQ	K	יט יט
4 <u>1</u> 40	G L	A A	P	K	L	L	I	>1 >1	G D	A	S S	T N	R L	E A	A S	G	V	P	S
61 60	R F	S S	G G	S	G	S	G	T	D D	- ×	T	FF	中	I	S S	S	L	Q	ם ם
81 80	E D E D	I	A A	T T	Y. Y.	Ā	C	00	Q	Y W	N S	N T	Ä	ום וח	D .	1313	*11 m	G G	Q Q
101 100	G T	K	V V	E E	V V	K													

FIGURE 26A

FIGURE 26B

vc13

10 20 30 40 50 60 TTCTGCTGGT ACCAGTACAT GAAACTTACA CTTGAGCTGC CACTGCAGGT GATGGTGACG

70 80 90 100 CGGTCACCCA CTGAGGCACT GAGGCTAGAT GGAGACTGGG TCATTTG

vcl4

10 20 30 40 CATGTACTGG TACCAGCAGA AGCCAGGAAA AGCTCCGAAA CTTCTGATTT ATGACACATC

70 80 90 100 110 CAACCTGGCT TCTGGAGTCC CTTCCCGCTT CAGTGGCAGT GGGTCTGGGA CCGATTACAC

130 CTTTACAATC TCTTCA

vc15

10 20 30 40 50 60 TGTGTCTAGA AAAGTGTACT TACGTTTTAC CTCGACCTTG GTCCCTTGAC CGAACGTGAG

70 80 90 100 110 120 CCGGTAAGTA CTCCACTGCT GGCAGTAATA AGTGGCTATA TCTTCCGGCT GAAGTGAAGA

130 GATTGTAAAG GTGTAAT

vc16

10 20 30 40 50 60 CACATCTAGA CCACCATGGA TTTTCAAGTG CAGATCTTCA GCTTCCTGCT AATCAGTGCC

80 90 100 TCAGTCATAC TGTCCAGAGG AGATATTCAA ATGACCCAGT CTCCATCT

FIGURE 27A

vc11

10 20 30 40 50 60
TAGTCTGTCG ACCCACCACT CCATATCACT CCCACCCACT CGAGTCCCTT TCCAGGAGCC

70 80 90 100 110 120
TGGCGGACCC AGTGTACACC ATAACTTGTT ACGGTGAAAC CACTGGCGGC ACAAGACAGT

130
CTCAGAGATC CTCCTGGC

vc12

10 20 30 40 50 60
TGGTGGGTCG ACAGACTATA ATGCAGCTTT CATATCCAGA TTTACCATCA GCAGAGACAA

70 80 90 100 110 120
CAGCAAGAAC ACACTGTATC TCCAAATGAA TAGCCTGCAA GCCGAGGACA CAGCCATATA

wps54

TTATTG

10 20 30 40 50 60
ACACTCTAGA CCACCATGGC TGTCTTGGGG CTGCTCTTCT GCCTGGTGAC ATTCCCAAGC

70 80 90 100 110 120
TGTGTCCTAT CCGCTGTCCA GCTGCTAGAG AGTGGTGGCG GTCTGGTGCA GCCAGGAGGA

130
TCTCTGAGAC

wos57

10 20 30 40 50 60
ACACTCTAGA AGTTAGGACT CACCTGAAGA GACAGTGACC AGAGTCCCTT GGCCCCAGTA

70 80 90 100 110
AGCAAAACCG TCGTAATTAT AGTCCCCAGC TCTGGCACAA TAATATATGG CTGTGTCC

FIGURE 27B

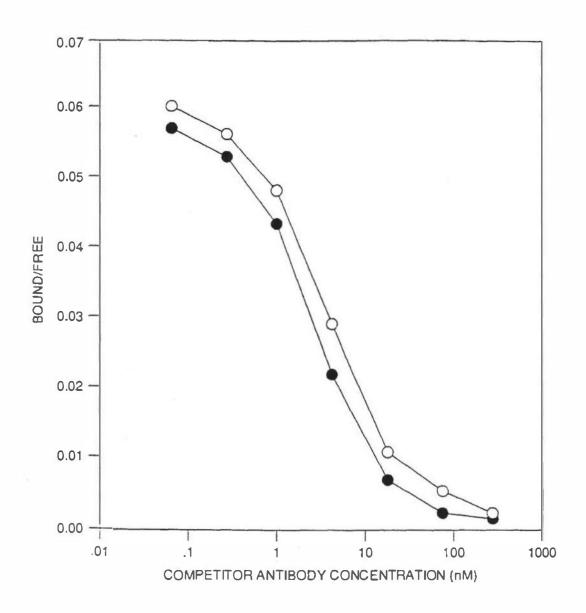


FIGURE 28

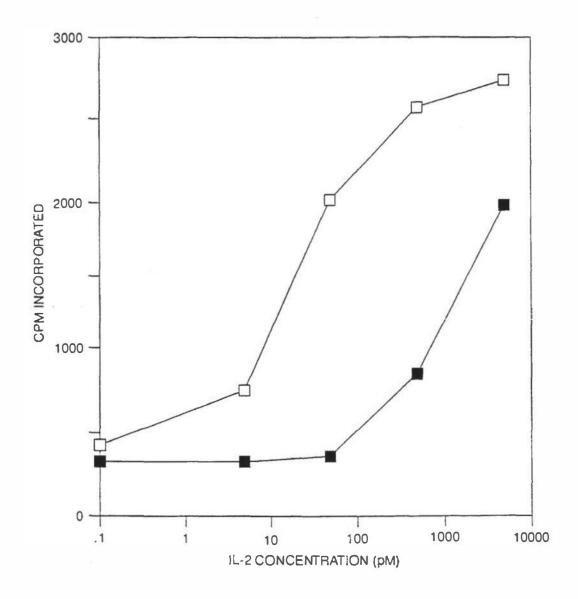


FIGURE 29

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	1 1	1 E E	M V	I Q	L	5 V L	EE	S	G	G G	10 G G	L	v	K Q	ים ים	15 G G	A G	S	L	KR	20 1	
	2 <u>1</u> 21	S S	C	A A	A A	25 S S	G G	[44 [44	T	F	30 S S	N N	Y Y	G G	i L	35 S	W	v v	2 2	Q Q	40 T A	
	41 41	SP	DG	R	R G	45 L L	E	W	v	A A	50 S S	I	52 \$ \$	a R R	G G	G G	G	55 R R	Ĩ	Ā Ā	S S	•
	60 60	60 P P	D D	N N	L L	K	65 G	R R	E E	T T	Ī	70 S S	R	E	D	A	K	75 N N	T	L L	Y Y	
	80 80	80 L L	00	82 M M	a S N	2 2 3	C L L	K Q	S A	85 E E	D D	T T	A A	L L	90 Y Y	Y	СС	L	R R	95 E E	G G	
	97 97	I	Y Y	Y	100 A A	а О	o Y	G G	d F F	k F	D D	V V	W	105 G G	T Q	G	F: 61	T	110 V V	I T	v	
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		1				5					10					15					20	
	1	D E	I	v	L M	T T 25	Q Q	S	P	A	S	L	AS	v	s s	L	G	QE	R	A A 35	T	v
	21	I L	S	CC	R R	A A	S S	27 Q Q	a S S	v v	S S	T T	S S	T	×- ×-	Ŋ	Ā	M M	H	W	ć.	
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	37 37	Q	Q	K	P. P.	G	Q	PS	P	KR	L	L L	Ī	K	Y	A	S	N	L	E	S	
					60					65					70					75		
	57 57	G G	V I	P P	A A	R	F	S	G G	S	G	FS	G	T . T	DE	fir fir	T.	1.1.1.1	N	Ţ	E S	
					80					85					90					95	_	
	77 77	P R	V L	[H [H]	2 5	E) [1]	D	Er fu	V A	T	ž	7.	C	Q	H	S	M	E3 E3	=	P	>1 >1	
					100)				105		107	,									
	97 97	T	111 111	GG	GQ	G	3.3	KR	<u>1</u>	[1] [1]	=======================================	K										

FIGURE 30B

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U.S. Patent
                  Jun. 25, 1996 Sheet 33 of 55
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                     Q S D A E
Q S G A E
                I Q
              Q L
                  25
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                C.
                  S
                     GYT
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                               T
                                  DH
                                            H
                                              WMKQR
             K
                          TF
                   5
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                     G Y
                                              WMRQA
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                            G Y
G Y
                                  I
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                                       P
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             Q
               G
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   41
                G
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                                 70
                   K
                                         KE
           E
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S
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         S
              K
                E
                     GKATL
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              K
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S L T S E D S A V Y F C A R
S L R S E D T A V Y F C A R
         80
                             85
              82
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         MHLN
   80
         MEL
                S
   80
                         F A Y
F A Y
                        G
         D S R E R
D S R E R
                               YWGQGTL
   97
                   R N
                                 WGQGTLVTVS
   97
                     N
                        G
        113
   113
         A
   113
         S
                        FIGURE 30C
                   5
                               10
           I V M
         D
                   TQSHKFMSTSVGDRV
                                                        S
                                      ASVGD
              O M
                     Q S P
                            S
                               T
                                  L
                                    S
                                                         40
                                            35
                                            M H
   21
            T
             CK
                   A
                     SQDV
                               G
                                  S
                                    A
                                       V
                                         V
                                                         S
                                          VW
            T C
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                                       T
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                                               T
                                                         S
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              A
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                                                        80
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                               70
                                            I
           F T G S G S G
                            T
                               D
                                          T
            F T G
                  S
                            T
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                      G S
                          G
                                                        100
                  85
                     74 E
                          C
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A T
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   81
                                Q
                     107
                  105
                L E L K
   101
         G
              3
   101
            TK
```

FIGURE 30D

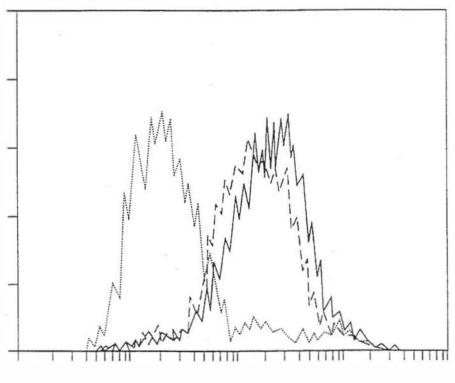


FIGURE 31A

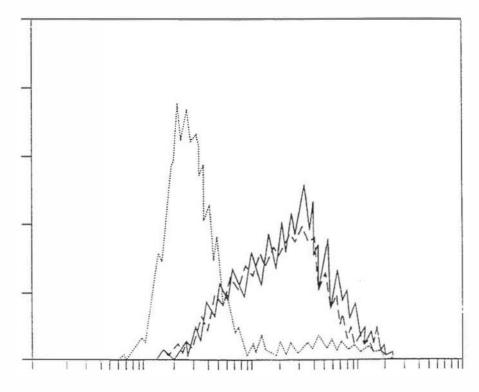


FIGURE 31B

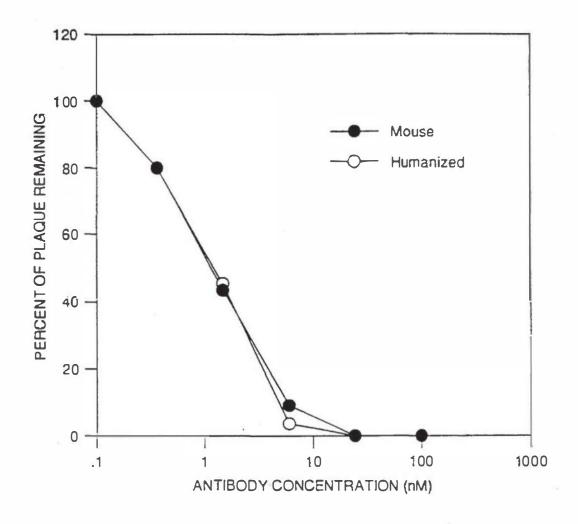


FIGURE 32A

5,530,101

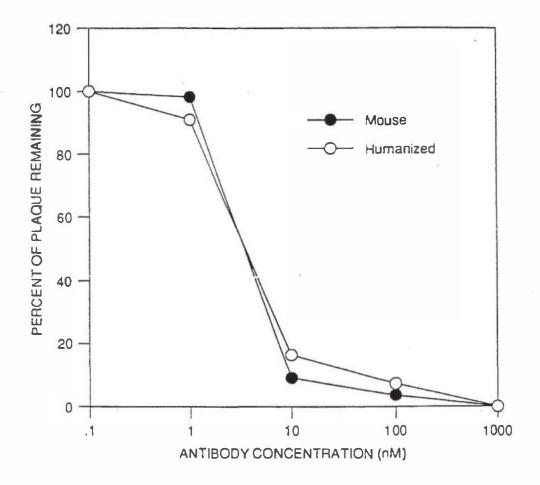


FIGURE 32B









FIGURE 34B

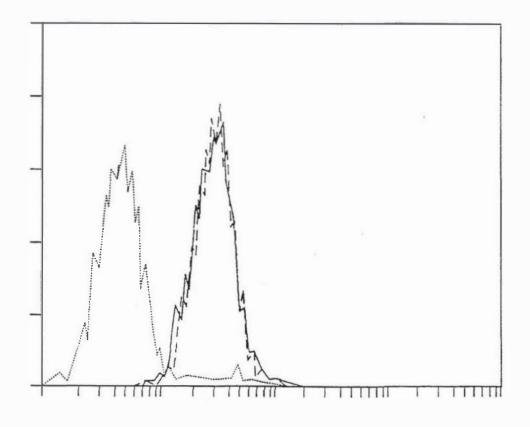


FIGURE 35

1	D	I	Q Q	M M	T	QQ	S	P	S S	TS	L L	S	A A	S	V	G	D	R	v v	T
21 21	I	T T	C	R R	A	S S	Q E	S S	V	I D	N N	Ā	G	I	TS	W	L M	A N	W W	E X
37 41	QQ	Q	K	P	G	G	A A	P ₁	K K	L	L L	M	Y	K A	A A	S S	S	L 0	E	S S
57 61	G	V	P P	S S	R	F	I S	G	S	G G	S	G	T	ED	fri fri	T	L	TT	I ī	S
77 81	S	L	QQ	P P	D D	D D	F	A A	T	Y Y	Y Y	CC	00	Q	Y S	N K	S	D	S P	K W
97 101	M	F	G	QQ	G	T	K	v	E	v <u>I</u>	K									

FIGURE 36A

FIGURE 36B

mai

10 20 30 40 50 60 TATATCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGAACTGCT 90 80 100 110 GGCGTCCACT CTCAGGTTCA GCTGGTGCAG TCTGGAGCTG AGGTGAAGAA GCCTGGGAGC

ma2

TCAGTGAAGG TT

10 20 30 40 50 60 110 80 90 100 CCTGCCTCAC CCAGTGCATG TTGTAGTCAG TGAAGGTGTA GCCAGAAGCT TTGCAGGAAA 130 CCTTCACTGA GCT

ma3

10 20 30 40 50 60 TGGTGGTACC GGCTACAACC AGAAGTTCAA GAGCAAGGCC ACAATTACAG CAGACGAGAG 80 90 100 TACTAACACA GCCTACATGG AACTOTOCAG COTGAGGTOT GAGGACACTG CA

ma4

10 20 30 40 50 60 TATATCTAGA GGCCATTCTT ACCTGAAGAG ACAGTGACCA GAGTCCCTTG GCCCCAGTAG 70 80 90 100 110 TOCATAGOGG GGOGCOCTOT TGCGCAGTAA TAGACTGCAG TGTCCTCAGA C

FIGURE 37A

mas 10 20 30 40 50 60 TATATCTAGA CCACCATGGA GAAAGACACA CTCCTGCTAT GGGTCCTGCT TCTCTGGGTT 70 80 90 100 110 120 CCAGGTTCCA CAGGTGACAT TCAGATGACC CAGTCTCCGA GCTCTCTGTC CGCATCAGTA GG ma6 10 20 30 40 50 60 TCAGAAGCTT AGGAGCCTTC CCGGGTTTCT GTTGGAACCA GTTCATAAAG CTAATGCCAT 70 30 90 100 110 AATTGTCGAC ACTTTCC-CTG GCTCTGCATG TGATGGTGAC CCTGTCTCCT ACTGATGCGG AC ma7 10 20 30 40 50 60 TCCTAAGCTT CTGATTTACG CTGCATCCAA CCAAGGCTCC GGGGTACCCT CTCGCTTCTC 70 80 90 100 110 AGGCAGTGGA TCTGGGACAG ACTTCACTCT CACCATTTCA TCTCTGCAGC CTGATGACT ma8

70 80 90 100 110 TCCACC-GAAC CTCCTTACTT TGCTGACAGT AATAGGTTGC GAAGTCATCA GGCTGCAG

FIGURE 37B

TATATCTAGA CTTTGGATTC TACTTACGTT TGATCTCCAC CTTGGTCCCT TGACCGAACG

10 20 30 40 50 60

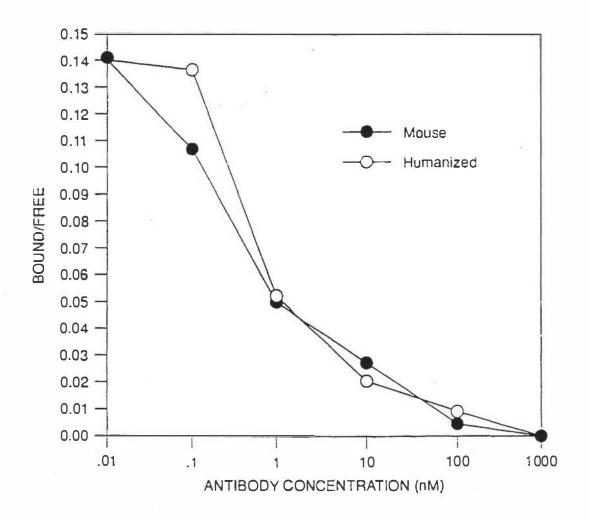


FIGURE 38

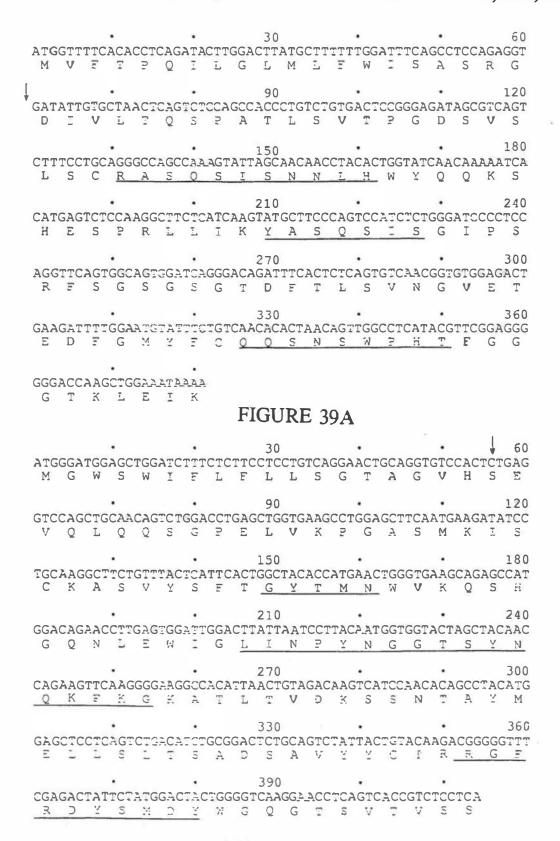


FIGURE 39B

1	E E	I	V V	L L	T	Q	S	P P	G G	T		S	L	S		G		R R	A A		
21 21	L	S	С	R	A A	S	0	S	V	S	S	G	Y	L	G	W	>- >+	Q	Q	K	
41 40	P P	G G	QQ	A A	P	R	L L	L	I	<u>Ү</u>	G	A A	S S	S	R	A	S	G	I	P	
61 60		R	F	S	G G	S	G G	S	G G	T	D D	F	T T	L L		I	S		L L		
81 80		E E		F	A A	V	Y Y	Ϋ́	СС	QQ	00	Y S	G N	S S	L W	G	R H	T	F	G	
101	QQ	G G	T T		v v			K K													

FIGURE 40A

FIGURE 40B

jb16

10 20 30 40 50 60
TAGATCTAGA CCACCATGGT TTTCACACCT CAGATACTAG GACTCATGCT CTTCTGGATT

70 80 90 100 110 120
TCAGCCTCCA GAGGTGAAAT TGTGCTAACT CAGTCTCCAG GCACCCTAAG CTTATCACCG

GGAGAAAGG

jb17

10 20 30 40 50 60
TAGACAGAAT TCACGCGTAC TTGATAAGTA GACGTGGAGC TTGTCCAGGT TTTTGTTGGT

70 80 90 100 110 120
ACCAGTGTAG GTTGTTGCTA ATACTTTGGC TGGCCCTGCA GGAAAGTGTA GCCCTTTCTC

CCGGTGAT

jb18

10 20 30 40 50 60
AAGAGAATTC ACGCGTCCCA GTCCATCTCT GGAATACCCG ATAGGTTCAG TGGCAGTGGA

70 80 90 100 110
TCAGGGACAG ATTTCACTCT CACAATAAGT AGGCTCGAGC CGGAAGATTT TGC

jb19

10 20 30 40 50 60
TAGATCTAGA GTTGAGAAGA CTACTTACGT TTTATTTCTA CCTTGGTCCC TTGTCCGAAC

70 80 90 100 110
GTATGAGGCC AACTGTTACT CTGTTGACAA TAATACACAG CAAAATCTTC CGGCTC

FIGURE 41A

jb20

10 20 30 40 50 60
TATATCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGAACTGCA

70 80 90 100 110 120
GGTGTCCACT CTCAAGTCCA ACTCGTACAG TCTGGAGCTG AGGTTAAAAA GCCTGGAAGT

130
TCAGTAAGAG TTTC

jb21

10 20 30 40 50 60
TATATAGGTA CCACCATTGT AAGGATTAAT AAGTCCAACC CACTCAAGTC CTTTTCCAGG

70 80 90 100 110 120
TGCCTGTCTC ACCCAGTTCA TGGTATACCC AGTGAATGAG TATCCGGAAG CTTTGCAGGA

130
AACTCTTACT GAAC

jb22

10 20 30 40 50 60
TATATAGGTA CCAGCTACAA CCAGAAGTTC AAGGGCACAG TTACAGTTC TTTGAAGCCT

70 80 90 100 110
TCATTTAACC AGGCCTACAT GGAGCTCAGT AGTCTGTTTT CTGAAGACAC TGCAGT

jb23

10 20 30 40 50 60
TATATCTAGA GGCCATTCTT ACCTGAGGAG ACGGTGACTA AGGTTCCTTG ACCCCAGTAG

70 80 90 100 110
TCCATAGAAT AGTCTCGAAA CCCCCGTCTT CTACAGTAAT AGACTGCAGT GTCTTC

FIGURE 41B

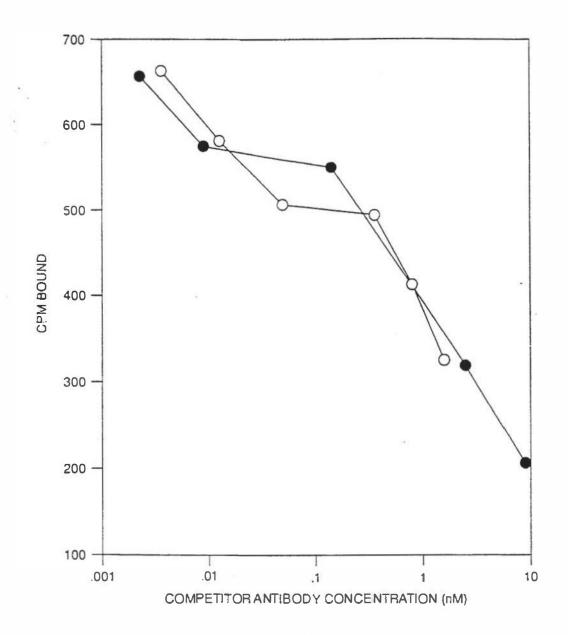


FIGURE 42

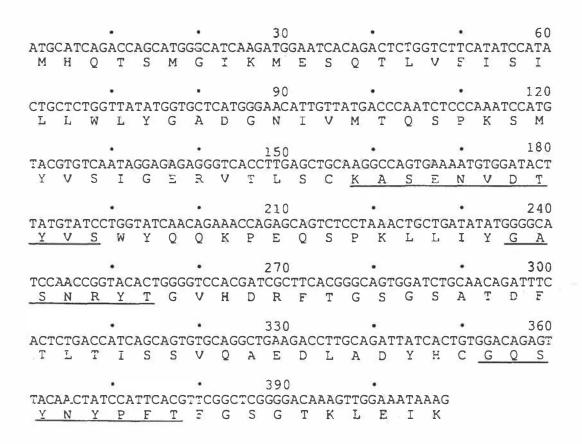


FIGURE 43A

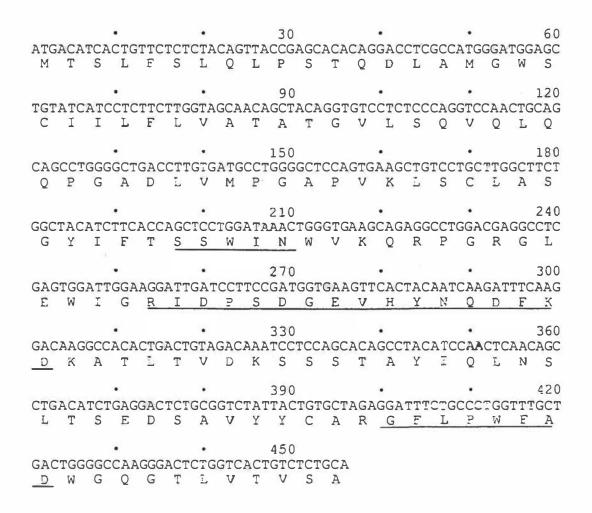


FIGURE 43B

1	D D	ī	Q	M M	T T	Q	S S	P	S	T T	L L	S S	A A	S S	V V	G G	D D	R R	V V	Ţ
21 21	I	T T	C	R K				S N				W Y	L V	A S	W	Y Y	Q	Q	K K	P
41 41	G G	K	A A	P	K	L	L	M <u>I</u>	Y	K G	A A	S S	S N	L R	E Y	S	G	V	ם, ם,	S
61 61	R R	F	I S	G				G G				T	L L	T	I	S	S	L L	Q	P P
81 81	D	D D	F	A	T T	Y Y	¥	C	G	Q	Q S	Y Y	N N	S	D Y	Sp	K F	M T	F	G G

FIGURE 44A

1	Q	V	Q	L L	V	Q	S	G	A A	E	V V	K	K K	P	G G	S S	S S	V	K	
21 21	S	C	K	A A	S	G G	G	T I	F F	S	R S	S S	A W	I	I N	W	V V	R R	Q	A A
41 41	חוח,	G G	Q	G	ī L	EJ EJ	W	M	G G	G R	I	V D	ים ים	M S	E D	G G	Dr [11]	P. V	N H	Y Y
61 61	A N	0	K D	F	Q K	G D	R	V	T	ĩ	T	A A	D D	田田	S	T	N	T	A A	>1 >1
81 81	M M	E	L	S	S	L	R R	S S	E	D	T	A A	F V	Y	E >	C	A A	G R	G	>1 [1,
101 101	G L	I	M Ā	ž S	PA	E D	EW	Y G	N O	G G	G	L	V	Ţ	V	S	S S			

FIGURE 44B

5,530,101

rhlo 10 20 30 40 50 60 TTTTTCTAG ACCACCATGG AGACCGATAC CCTCCTGCTA TGGGTCCTCC TGCTATGGGT 70 80 90 100 110 CCCAGGATCA ACCGGAGATA TTCAGATGAC CCAGTCTCCG TCGACCCTCT CTGCT rhll 10 20 30 40 50 60 TTTTAAGCTT GGGAGCTTTG CCTGGCTTCT GCTGATACCA GGATACATAA GTATCCACAT 80 90 100 110 TTTCACTGGC CTTGCAGGTT ATGGTGACCC TATCCCCGAC GCTAGCAGAG AGGTTCCACG rh12 10 20 30 40 50 60 TTTTAAGCTT CTAATTTATG GGGCATCCAA CCGGTACACT GGGGTACCTT CACGCTTCAG 90 80 100 110 TGGCAGTGGA TCTGGGACCG ATTTCACCCT CACAATCAGC TCTCTGCAGC CAGATGAT rh13 10 20 30 40 TTTTTTCTAG AGCAAAGTC TACTTACGTT TGACCTCCAC CTTGGTCCCC TGACCGAACG 70 80 90 100 110 120 TGAATGGATA GTTGTAACTC TGTCCGCAGT AATAAGTGGC GAAATCATCT GGCTCCAGAG

FIGURE 45A

rh20

10 20 30 40 50 60
TTTTTCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGTACCGCG

70 80 90 100 110
GGCGTGCACT CTCAGGTCCA GCTTGTCCAG TCTGGGGCTG AAGTCAAGAA ACCT

rh21

10 20 30 40 50 60
TTTTGAATTC TCGAGACCCT GTCCAGGGGC CTGCCTTACC CAGTTTATCC AGGAGCTAGT

70 80 90 100 110 120
AAAGATGTAG CCAGAAGCTT TGCAGGAGAC CTTCACGGAG CTCCCAGGTT TCTTGACTTC

A

rh22

10 20 30 40 50 60
TTTTGAATTC TCGAGTGGAT GGGAAGGATT GATCCTTCCG ATGGTGAAGT TCACTACAAT

70 80 90 100 110 120
CAAGATTTCA AGGACCGTGT TACAATTACA GCAGACGAAT CCACCAATAC AGCCTACATG

130
GAACTGAGCA GCCTGAG

rh23

10 20 30 40 50 60
TTTTTCTAGA GGTTTTAAGG ACTCACCTGA GGAGACTGTG ACCAGGGTTC CTTGGCCCCA

70 80 90 100 110 120
GTCAGCAAAC CAGGGCAGAA ATCCTCTTGC ACAGTAATAG ACTGCAGTGT CCTCTGATCT

130
CAGGCTGCTC AGTT

FIGURE 45B

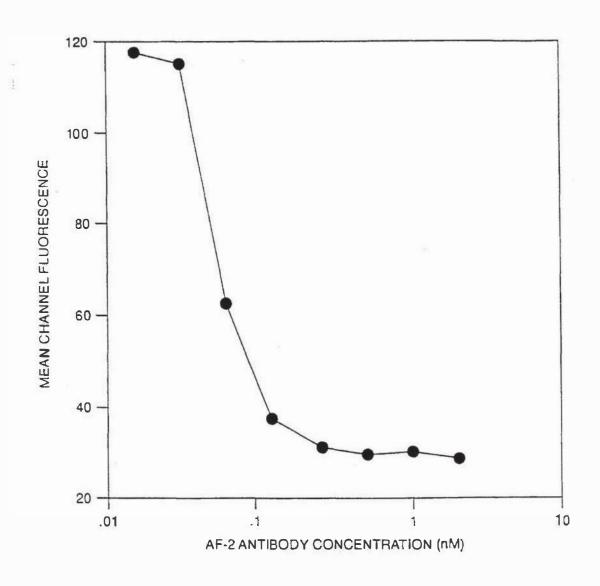


FIGURE 46

HUMANIZED IMMUNOGLOBULINS

CROSS-REFERENCE TO RELATED APPLICATIONS

This is a continuation-in-part application of commonly assigned patent application U.S. Ser. No. 07/590,274, filed Sep. 28, 1990 (now abandoned) and of U.S. Ser. No. 07/310,252, filed Feb. 13, 1989 (now abandoned), which is a continuation-in-part of U.S. Ser. No. 07/290,975, filed 10 Dec. 28, 1988 (now abandoned). All of these applications are specifically incorporated herein by reference.

FIELD OF THE INVENTION

The present invention relates generally to the combination of recombinant DNA and monoclonal antibody technologies for developing novel therapeutic agents and, more particularly, to the production of non-immunogenic antibodies having strong affinity for a predetermined antigen.

BACKGROUND OF THE INVENTION

The advent of monoclonal antibody technology in the mid 1970's heralded a new age of medicine. For the first time, researchers and clinicians had access to essentially unlimited quantities of uniform antibodies capable of binding to a predetermined antigenic site and having various immunological effector functions. These proteins, known as "monoclonal antibodies" were thought to hold great promise in, e.g., theremoval of harmful cells in vivo. Indeed, the clinical value of monoclonal antibodies seemed limitless for this use alone.

Unfortunately, the development of appropriate therapeutic products based on these proteins has been severely hampered by a number of drawbacks inherent in monoclonal antibody production. For example, most monoclonal antibodies are mouse derived, and thus do not fix human complement well. They also lack other important immunoglobulin functional characteristics when used in humans.

Perhaps most importantly, non-human monoclonal antibodies contain substantial stretches of amino acid sequences that will be-immunogenic when injected into a human patient. Numerous studies have shown that after injection of a foreign antibody, the immune response mounted by a patient can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, as increasing numbers of different mouse or other antigenic (to humans) monoclonal antibodies can be expected to be developed to treat various diseases, after one or several ireatments with any non-human antibodies, subsequent treauments, even for unrelated therapies, can be ineffective or even dangerous in themselves, because of cross-reactivity.

While the production of so called "chimeric antibodies" 55 (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. Moreover, efforts to immortalize human B-cells or generate human hybridomas capable of producing human immunoglobulins against a desired antigen have been generally unsuccessful, particularly with many important human antigens: Most recently, recombinant DNA technology has been utilized to produce immunoglobulins which have human framework regions combined with complementarity determining regions 65 (CDR's) from a donor mouse or rat immunoglobulin (see, e.g., EPO Publication No. 0239400, which is incorporated

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herein by reference). These new proteins arc called "reshaped" or "humanized" immunoglobulins and the process by which the donor immunoglobulin is converted into a human-like immunoglobulin by combining its CDR's with a human framework is called "humanization". Humanized antibodies are important because they bind to the same antigen as the original antibodies, but are less immunogenic when injected into humans.

However, a major problem with present humanization procedures has been a loss of affinity for the antigen (Jones et al., Nature, 321, 522–525 (1986)), in some instances as much as 10-fold or more, especially when the antigen is a protein (Verhoyen et al., Science, 239, 1534–1536 (1988)). Loss of any affinity is, of course, highly undesirable. At the least, it means that more of the humanized antibody will have to be injected into the patient, at higher cost and greater risk of adverse effects. Even more critically, an antibody with reduced affinity may have poorer biological functions, such as complement lysis, antibody-dependent cellular cytotoxicity, or virus neutralization. For example, the loss of affinity in the partially humanized antibody HuVHCAMP may have caused it to lose all ability to mediate complement lysis (see, Riechmann et al., Nature, 332, 323–327 (1988); Table 1).

Thus, there is a need for improved means for producing humanized antibodies specifically reactive with strong affinity to a predetermined antigen. These humanized immunoglobulins should remain substantially non-immunogenic in humans, yet be easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

SUMMARY OF THE INVENTION

The present invention provides novel methods for preparing humanized immunoglobulin chains having generally one or more complementarity determining regions (CDR's) from a donor immunoglobulin and a framework region from a human immunoglobulin. The preferred methods comprise first comparing the framework or variable region amino acid sequence of the donor immunoglobulin to corresponding sequences in a collection of human immunoglobulin chains, and selecting as the human immunoglobulin one of the more homologous sequences from the collection. The human immunoglobulin, or acceptor immunoglobulin, sequence is typically selected from a collection of at least 10 to 20 immunoglobulin variable region sequences, and usually will have the highest homology to the donor immunoglobulin sequence of any sequence in the collection. The human immunoglobulin framework sequence will typically have about 65 to 70% homology or more to the donor immunoglobulin framework sequences. The donor immunoglobulin may be either a heavy chain or light chain, and the human collection will contain the same kind of chain. A humanized light and heavy chain can be used to form a complete humanized immunoglobulin or antibody, having two light/ heavy chain pairs, with or without partial or full-length human constant regions.

To form the humanized variable region, amino acids in the human acceptor sequence will be replaced by the corresponding amino acids from the donor sequence if they are in the category

(1) the amino acid is in a CDR.

In another embodiment of the present invention, either in conjunction with the above comparison step or separately, additional amino acids in the acceptor immunoglobulin

chain may be replaced with amino acids from the CDR-donor immunoglobulin chain. More specifically, further optional substitutions of a human framework amino acid of the acceptor immunoglobulin with the corresponding amino acid from a donor immunoglobulin will be made at positions 5 which fall in one or more of the following categories:

- (2) the amino acid in the human framework region of the acceptor immunoglobulin is rare for that position and the corresponding amino acid in the donor immunoglobulin is common for that position in human immunoglobulin 10 sequences; or
- (3) the amino acid is immediately adjacent to one of the CDR's; or
- (4) the amino acid is predicted to be within about 3 Å of the CDR's in a three-dimensional immunoglobulin model and capable of interacting with the antigen or with the CDR's of the donor or humanized immunoglobulin.

Moreover, an amino acid in the acceptor sequence may optionally be replaced with an amino acid typical for human 20 sequences at that position if

(5) the amino acid in the acceptor immunoglobulin is rare for that position and the corresponding amino acid in the donor immunoglobulin is also rare, relative to other human sequences.

The humanized immunoglobulin chain will typically comprise at least about 3 amino acids from the donor immunoglobulin in addition to the CDR's, usually at least one of which is immediately adjacent to a CDR in the donor immunoglobulin. The heavy and light chains may each be designed by using any one or all three of the position criteria.

When combined into an intact antibody, the humanized light and heavy chains of the present invention will be substantially non-immunogenic in humans and retain substantially the same affinity as the donor immunoglobulin to the antigen (such as a protein or other compound containing an epitope). These affinity levels can vary from about 10 M⁻¹ or higher, and may be within about 4 fold, preferably within about 2 fold of the donor immunoglobulin. Ideally, the humanized antibodies will exhibit affinity levels at least about 60 to 90% of the donor immunoglobulin's original affinity to the antigen.

Once designed, the immunoglobulins, including binding fragments and other immunoglobulin forms, of the present invention may be produced readily by a variety of recombinant DNA or other techniques. Preferably, polynucleou des encoding the desired amino acid sequences are produced synthetically and by joining appropriate nucleic acid sequences, with ultimate expression in transfected cells. Notably, the methods of the present invention maximize the likelihood of producing humanized immunoglobulins with optimum binding characteristics without the need for producing intermediate forms that may display stepwise improvements in binding affinity. The humanized immunoglobulins will be particularly useful in treating human disorders susceptible to monoclonal antibody therapy, but find a variety of other uses as well.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1A and FIG. 1B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:1 and 2] and heavy chain (B) [SEQ ID NOS:3 and 4] variable regions of the mouse anti-Tac antibody (upper lines), compared with 65 the human Eu antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined.

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Residues in the Eu antibody framework replaced with mouse amino acids in the humanized antibody are double underlined. The number of the first position on each line is given on the left.

FIG. 2A and FIG. 2B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:46 and 47] and heavy chain (B) [SEQ ID NOS:48 AND 49] variable regions of the mouse Fd79 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 3A and FIG. 3B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:50 and 51] and heavy chain (B) [SEQ ID NOS:52 and 53] variable regions of the mouse Fd138-80 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 4A and FIG. 4B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:54 and 55] and heavy chain (B) [SEQ ID NOS:56 and 57] variable regions of the mouse M195 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in cach chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 5A and FIG. 5B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:58 and 59] and heavy chain (B) [SEQ ID NOS:60 and 61] variable regions of the mouse mik-β1 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 6A and FIG. 6B. Amino acid sequences (1-letter code) of the light chain (A) [SEQ ID NOS:62 and 63] and heavy chain (B) [SEQ ID NOS:64 and 65] variable regions of the mouse CMV5 antibody (upper lines), compared with the humanized antibody (lower lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the humanized antibody framework replaced with mouse amino acids or typical human amino acids are double underlined. The number of the first position on each line is given on the left.

FIG. 7A through FIG. 7D. Fluorocytometry of HUT-102 and Jurkat cells stained with anti-Tac antibody or humanized anti-Tac antibody followed respectively by fluorescein-conjugated goat anti-mouse Ig antibody or goat anti-human Ig antibody, as labeled. In each panel, the dotted curve shows the results when the first antibody was omitted, and the solid curve the results when the first and second (conjugated) antibodies were included as described.

FIG. 8A and FIG. 8B. (A) Fluorocytometry of HUT-102 cells stained with 0-40 ng of anti-Tac as indicated, then with biotinylated anti-Tac, and then with phycoeythrin-conjugated avidin. (B) Fluorocytometry of HUT-102 cells stained

with the indicated antibody, then with biotinylated anti-Tac, and then with phycoerythrin-conjugated avidin.

FIG. 9A and FIG. 9B. Schematic diagram of the plasmids pVgl (A) and pVk (B). The plasmid pVgl was constructed from the following fragments: an approximately 4850 base pair BamHI-EcoRI fragment from the plasmid pSV2hph containing the amp and hyg genes; a 630-pb fragment containing the human cytomegalovirus IEI gene promoter and enhancer flanked at the 5 and 3' by EcoRl and Xbal linkers respectively; and a 2800 bp XbaI-BamHI fragment containing the human gamma-1 constant region gene with 215 bp of the preceding intron and the poly(A) signal. The plasmid pVk was similarly constructed, with a 1530-bp human kappa constant region gene replacing the gamma-1 gene and the gpt replacing the hyg gene.

FIG. 10A and FIG. 10B. Amino acid sequences of the heavy (A) [SEQ ID NOS:5 and 6] and light (B) [SEQ ID NOS:7 and 8] chain variable regions of the PDL and CDR-only humanized anti-Tac antibodies. The PDL sequence is shown on the upper line, the CDR-only sequence below. Amino acid differences are boxed. Complementarity Determining Regions (CDR's) are underlined.

FIG. 11A and FIG. 11B. Double-stranded DNA sequence of fragments encoding the heavy (A) [SEQ ID NO:9] and light (B) [SEQ ID NO: 10] chain variable regions of the CDR-only humanized anti-Tac antibody including signal sequences. Oligonucleotides used for gene synthesis are marked by solid lines: above, for oligonucleotides from upper strand, and below, for oligonucleotides from lower strand. Restriction sites used for cloning are underlined.

FIG. 12. FACS analysis of HUT-102 cells stained with ³⁰ PDL and CDR-only humanized anti-Tac antibodies and negative control antibody Fd79.

FIG. 13. Competition by mouse, PDL humanized, and CDR-only humanized anti-Tac antibodies with binding of radioiodinated mouse anti-Tac antibody to HUT-102 cells.

FIG. 14. Scheme for anchored polymerase chain reaction (PCR) cloning of the heavy and light chain variable domain cDNAs. RNA was prepared from about 10' hybridoma cells using the hot phenol extraction method. Briefly, cells were 40 resuspended and vortexed in 1 ml of RNA extraction buffer (50 mM sodium acetate pH 5.2/1% SDS), extracted with 0.5 ml of phenol pH 5.2 at 65° C. for 15 min, followed by another 15 min on ice. The aqueous phase was recovered and precipitated twice with ethanol. cDNA was synthesized from 45 10 ug of total RNA using reverse transcriptase (BRL, Bethescda, Md.) and oligo-dT₁₂₋₁₈ (Pharmacia, Piscatway, N.J.) as primers. A poly(dG) tail was attached to the 3' end of the cDNA using terminal deoxynucleotide transferase (BRL) (E. Y. Loh et al., Science 243, 217 (1989)), the 50 variable domain genes (V) were amplified using AmpliTaq (Perkin Elmer-Cetus) with the primer mc045 (TAATCTA-GAATICCCCCCCCCCCCCCCC) [SEQ ID NO:11] that hybridized to the poly(dG) tails and primers that hybridized to the constant region genes (C). For the light chain, the 55 primer used was mc045 (TATAGAGCTCAAGCTTG-GATGGTGGGAAGAI'GGATACAGTI'GGTGC) [SEQ ID NO:12]. For the heavy chain, the primer used was mc047 (TATAGAGCTCAAGCITCCAGTGGATA-

GAC(CAT)GATGGGG(GC)TGT(TC)GTITTGGC) [SEQ 60 ID NO:13]. The sequence in parenthesis indicates a base degeneracy. The degeneracy was introduced so that the primer would be able to hybridize to most gamma chains. The amplified fragments were then digested with EcoRI and HindIII and cloned into pUC18 vector for sequencing.

FIG. 15. Comparison of sequences of anti-Tac heavy chain (upper lines) [SEQ ID NO:14] and Eu heavy chain

(lower lines) [SEQ ID NO:15]. The 1-letter code for amino acids is used. The first amino acid on each line is numbered at the left. Identical amino acids in the two sequences are connected by lines. The 3 CDRs are underlined. Other amino acid positions for which the anti-Tac amino acid rather than the Eu amino acid was used in the humanized anti-Tac heavy chain are denoted by an *.

FIG. 16. Comparison of sequences of anti-Tac light chain (upper lines) [SEQ ID NO:16] and Eu light chain (lower lines) [SEQ ID NO:17]. The single-letter code for amino acids is used. The first amino acid on each line is numbered at the left. Identical amino acids in the two sequences are connected by lines. The 3 CDRs are underlined. Other amino acid positions for which the anti-Tac amino acid rather than the Eu amino acid was used in the humanized anti-Tac heavy chain are denoted by an *.

FIG. 17. Nucleotide sequence of the gene for the humanized anti-Tac heavy chain variable region gene [SEQ ID NOS: 18 and 19]. The translated amino acid sequence for the part of the gene encoding protein is shown underneath the nucleotide sequence. The nucleotides TCTAGA at the beginning and end of the gene are Xba I sites. The mature heavy chain sequence begins with amino acid #20 Q.

FIG. 18. Nucleotide sequence of the gene for the humanized anti-Tac light chain variable region gene [SEQ ID NOS:20 and 21]. The translated amino acid sequence for the part of the gene encoding protein is shown underneath the nucleotide sequence. The nucleotides TCTAGA at the beginning and end of the gene are Xba I sites. The mature light chain sequence begins with amino acid #21 D.

FIG. 19A and FIG. 19B. (A) Sequences of the four oligonucleotides [SEQ ID NOS:22, 23, 24, and 25] used to synthesize the humanized anti-Tac heavy chain gene, printed 5' to 3'. (B) Relative positions of the oligonucleotides. The arrows point in the 3' direction for each oligonucleotide.

FIG. 20A and FIG. 20B. (A) Sequences of the four oligonucleotides [SEQ ID NOS:26, 27, 28, and 29] used to synthesize the humanized anti-Tac light chain gene, printed 5' to 3'. (B) Relative positions of the oligonucleotides. The arrows point in the 3' direction for each oligonucleotide. The position of a Hind III site in the overlap of JFD2 and JFD3 is shown.

FIG. 21. Schematic d'agram of the plasmid pHuGTACI used to express the humanized anti-Tac heavy chain. Relevant restriction sites are shown, and coding regions of the heavy chain are displayed as boxes. The direction of transcription from the immunoglobulin (Ig) promoter is shown by an arrow. E_H=heavy chain enhancer, Hyg=hygromycin resistance gene.

FIG. 22. Schematic diagram of the plasmid pHuLTAC used to express the humanized anti-Tac light chain. Relevant restriction sites are shown, and coding regions of the light chain are displayed as boxes. The direction of transcription from the Ig promoter is shown by an arrow.

FIG. 23A and FIG. 23B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:30 and 31] and heavy chain (B) [SEQ ID NOS:32 and 33] variable regions of the antibody mik- β 1. The CDR sequences are underlined. The mature light chain protein begins with amino acid 23 Q and the mature heavy chain protein with amino acid 20 Q, preceded by the respective signal sequences.

FIG. 24A and FIG. 24B. Schematic diagram of the plasmids pVgl-dhfr (A) and pVk (B). The plasmid pVgl-dhfr contains the following parts: an approximately 4200 base pair BamHI-EcoRI fragment containing the amp and

dhfr genes; a 630-bp fragment containing the human cytomegalovirus IE1 gene promoter and enhancer (Boshart et al., Cell 41, 521 (1985), which is incorporated herein by reference) flanked at the 5' and 3' ends by EcoRI and XbaI linkers respectively; and a 2800 bp Xbal-BamHI fragment 5 containing the human gamma-1 constant region gene with 215 bp of the preceding intron and the poly(A) signal. The plasmid pVk was similarly constructed, with a 1530-bp human kappa constant region gene replacing the gamma-1 gene and the gpt gene replacing the dhfr gene. The plasmids were constructed from the indicated parts using methods well-known in the art (see, Maniatis et al., Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1989) and commonly assigned PCT Publication No. WO 89/09622, published Oct. 19, 1989. For example, pVgl-dhfr was 15 constructed from the plasmid pVgl by replacing the Hind III-Bgl II fragment containing the hyg gene with a 660 bp fragment containing the dhfr genc and extending to a Bgl II site (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983)).

FIG. 25. Fluorocytometry of YTJB cells stained with (_____) Isotype matched control antibody, (- - -) humanized mik-β1 antibody, (. . .) chimeric mik-β1 antibody. Cells were suspended in FACS buffer (PBS+2% BSA+0.1% azide) at approximately 5×10⁶/ml. 100 ul of cell suspension was transferred to a polystyrene tube and incubated with 100 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with goat antihuman lg antibody on ice for another 30 min. Then the cells were washed and incubated with FITC labeled rabbit antigoat lg antibody for 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldehyde. Cells were analyzed on a FACSmatc (Becton Dickinson)

FIG. 26A and FIG. 26B. Amino acid sequences of the light chain (A) [SEQ ID NOS:34 and 35] and the heavy chain (B) [SEQ ID NOS:36 and 37] of the humanized mik- β l antibody, (lower lines) and human Lay antibody (upper lines), not including signal sequences. The three CDRs in each chain are underlined. Amino acids in the framework that have been replaced with mouse amino acids or consensus human amino acids in the humanized antibody are double underlined.

FIG. 27A and FIG. 27B. Oligonucleotides used in the construction of the humanized mik-β1 heavy chain (B) [SEQ ID NOS: 42, 43, 44, and 45] and light chain (A) [SEQ 45 ID NOS:38, 39, 40, and 41]. The following pairs of oligonucleotides were mixed, extended with sequenase and cut with the indicated enzymes before ligation into the pBluescriptII ks (+) vector: wps54 and vc11 with Xba I and Sal I, vc12 and wps57 with Xba I and Sal I, vc12 and wps57 with Xba I and Sal I, vc16 and vc13 with Xba I and Kpn I, vc14 and vc15 with Xba I and Kpn I. Then the wps54-vc11 and vc12-wps57 fragments were excised with Xba I and Sal I ligated together into the Xba I site of pVg1-dhfr; and the vc16-vc13 fragments and vc14-vc15 fragments were excised with Xba I and Kpn I and ligated together into the Xba I site of pVk.

FIG. 28. Competitive binding of labeled mik-β1 tracer to YTJB cells. About 10⁶ YTJB cells were incubated with 3.0 ng of radio-iodinated mouse mik-β1 antibody (6 μCi/μg) and varying amounts of either unlabeled mouse mik-β1 antibody (Θ) or humanized mik-β1 antibody (Θ) in 200 ul of binding buffer (PBS+10% fetal calf serum+0.1% NaN₃+10 μg/ml mouse monoclonal lg). After incubation for 2 hr at 0° C. the cells were washed twice with binding buffer without mouse Ig and collected by centrifugation. The radioactivity bound 65 to cells was measured and expressed as the ratio of bound/free cpm.

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FIG. 29. Inhibition of II_2 stimulated proliferation of human PHA blasts by humanized mik-β1+humanized anti-Tac antibodies. No antibody added (□), 2 ug cach of humanized mik-β1 and humanized anti-Tac added (■).

FIG. 30 A through FIG. 30D. Amino acid sequences of the heavy chain (A) [SEQ ID NOS:48 and 49] and the light chain (B) [SEQ ID NOS:46 and 47] of the murine and humanized Fd79 antibodies, and the heavy chain (C) (SEQ ID NOS:52 and 53] and the light chain (D) [SEQ ID NOS:50 and 51] of the murine and humanized Fd138-80 antibodies. The sequences of the murinc antibody as deduced from the cDNA (upper lines) are shown aligned with the humanized antibody sequences (lower lines). The humanized Fd79 and Fd138-80 framework sequences are derived from Pom antibody and Eu antibody, respectively. Residues are numbered according to the Kabat system (E. A. Kabat et al., Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, Md.) (1987). The three CDRs in each chain are boxed. Residues in the Pom or Eu framework that have been replaced with murine sequences or consensus human sequences are underlined.

FIG. 31A and FIG. 31B. Fluorocytometry of HSV-I infected Vero cells stained with Fd79 (A) and Fd138-80 (B) antibodies, (. .) Isotype matched control antibody, (. . .) humanized antibody, (____) chimeric antibody. Vero cells were infected with HSV-1 (Δ305 mutant (F strain)) at 3 pfu/cell overnight. Cells were trypsinized at 0.5 mg/ml for I minute, washed extensively with PBS and resuspended in FACS buffer (PBS+2% BSA+0.1% azide) at approximately 5×10°/ml. 100 ul of cell suspension was transferred to a polystyrene tube and incubated with 100 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with FITC labeled goat antihuman antibody (Cappel) on ice for another 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldchydc. Cells were analyzed on a FACSmate (Becton Dickinson).

FIG. 32A and FIG. 32B. Neutralization of HSV-1 by Fd79 (A) and Fd138-80 (B). Serial dilutions of antibodies were mixed with 100 pfn of virus and incubated at 37° C. for 1 hr. The viruses were then inoculated onto 6-well plates with confluent Vero cells and adsorbed at 37° C. for I hr. Cells were overlayed with 1% agarose in medium and incubated for 4 days. Plaques were stained with neutral red.

FIG. 33A and FIG. 33B. Immunostaining of infected Vero cell monolayers to examine protection of cells from viral spread in tissue culture by (A) murine or humanized Fd79, (B) murine or humanized Fd138-80. 24-well plates of confluent Vero cells were inoculated with virus at 0.1 pfu/cell and allowed to adsorb for 2 hrs. at 37° C. before adding 200 ul of 10 ug/ml antibodies in medium. At the end of 4 days, culture medium was removed and plates were dried by placing overnight in a 37° C. incubator. To detect viral antigens, each well was incubated with 200 ul of anti-gB antibody at 0.5 ug/ml for 1 hr. at 37° C., washed twice and incubated with 200 ul of peroxidase conjugated goat antimouse IgG (Cappel, 1:300 dilution) for 1 hr. at 37° C. The plates were washed and then developed with the substrate 3-amino-9-cthyl-carbazole (AEC) (Sigma, St. Louis, Mo.) for 15 minutes at room temperature. Reaction was stopped by rinsing with water and air dried.

FIG. 34A and FIG. 34B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:66 and 67] and heavy chain (B) [SEQ ID NOS:68 and 69] variable regions of the antibody M195. The CDR sequences are underlined. The mature light chain protein

begins with amino acid 21 D and the mature heavy chain protein with amino acid 20 E, preceded by the respective signal sequences.

FIG. 35. Fluorocytometry of U937 cells stained with (...) no antibody, (...) humanized M195 antibody, (---) chimeric M195 antibody. Cells were suspended in FACS buffer (PBS+2% FCS+0.1% azide) at approximately 5×10⁶⁷ ml. 100 ul of cell suspension was transferred to a polystyrene tube and incubated with 50 ng of purified antibody on ice for 30 min. The cells were washed with FACS buffer and incubated with FITC labeled goat anti-human lg antibody on ice for another 30 min. The cells were washed again and finally resuspended in PBS+1% paraformaldehydc. Cells were analyzed on a FACSmate (Bccton Dickinson).

FIG. 36A and FIG. 36B. Amino acid sequences of the 1s light chain (A) [SEQ ID NOS:70 and 71] and the heavy chain (B) [SEQ ID NOS:72 and 73] of the humanized M195 antibody (lower lines) and human Eu antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework that have 20 been replaced with mouse amino acids in the humanized antibody are double underlined.

FIG. 37A and FIG. 37B. Oligonucleotides used in the construction or the humanized M195 heavy chain (A; mal-4) [SEQ ID NOS:74, 75, 76, and 77] and light chain (B; ma5-8) ²⁵ [SEQ ID NOS:78, 79, 80, and 81]. The following pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: mal and ma2 with Xba I and Kpn I, ma3 and ma4 with Xba I and Kpn I, ma5 and ma6 with Xba I and 30 Hind III, ma7 and ma8 with Xba I and Hind III. Then the mal-ma2 and ma3-ma4 fragments were excised from pUC18 with Xba I and kpn I and ligated together into the Xba I site of pVgl-dhfr, and the ma5-ma6 and ma7-ma8 fragments were excised with Xba I and Hind III and ligated together into the Xba I site of pVk.

FIG. 38. Competitive binding of labeled M195 tracer to U937 cells. About 4×10⁵ U937 cells were incubated with 4.5 ng of radio-iodinated mouse M195 antibody (6 μci/μg) and varying amounts of either unlabeled mouse M195 antibody (Φ) or humanized M195 antibody (Φ) in 200 ul of binding buffer (PBS+2% fetal calf serum+0.1% sodium azide). After incubation for 2 hr at 0° C., the cells were washed twice with binding buffer and collected by centrifugation. The radioactivity bound to cells was measured and is expressed as the ratio of bound/free cpm.

FIG. 39A and FIG. 39B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS: 82 and 83] and heavy chain (B) [SEQ ID NOS:84 and 85] variable regions of the antibody CMV5. The CDR sequences are underlined. The start of the mature protein sequences are indicated by arrows, preceded by the respective signal sequences.

FIG. 40A and FIG. 40B. Amino acid sequences of the light chain (A) [SEQ ID NOS:86 and 87] and the heavy chain (B) [SEQ ID NOS:88 and 89] of the humanized CMV5 antibody (lower lines) and human Wol antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework replaced with mouse amino acids or typical human amino acids in the humanized antibody are double underlined.

FIG. 41A and FIG. 41B. Oligonucleotides used in the construction of the humanized CMV5 light chain (A; jbl6-65 jbl9) [SEQ ID NOS:90, 91, 92, and 93] and heavy chain (B; jb20-jb22) [SEQ ID NOS:94, 95, 96, and 97]. The following

pairs of oligonucleotides were mixed, extended with Klenow polymerase and cut with the indicated enzymes before ligation into pUC18: jb16 and jb17 with Xba I and EcoR I, jb18 and jb19 with Xba I and EcoR I, jb20 and jb21 with Xba I and Kpn I, jb22 and jb23 with Xba I and Kpn I. Then the jb16-jb17 and jb18-jb19 fragments were excised with Xba I and Mlu I and ligated together into the Xba I site of pVk; and the jb20-jb21 and jb22-jb23 fragments were excised with Xba I and Kpn I and ligated together into the Xba I site of pVg1-dhfr.

FIG. 42. Competitive binding of labeled CMV5 racer to CMV-infected cells. Increasing amounts of mouse (①) or humanized (o) CMV5 antibody was added to CMV-infected HEL cells with tracer radio-iodinated mouse CMV5, and the amount of tracer bound to the cells was determined.

FIG. 43A and FIG. 43B. Sequences of the cDNA and translated amino acid sequences of the light chain (A) [SEQ ID NOS:98 and 99] and heavy chain (B) [SEQ ID NOS:100 and 101] variable regions of the antibody AF2. The CDR sequences are underlined. The mature light chain protein begins with amino acid 30 N and the mature heavy chain protein with amino acid 36 Q, preceded by the respective signal sequences.

FIG. 44A and FIG. 44B. Amino acid sequences of the light chain (A) [SEQ ID NOS:102 and 103] and the heavy chain (B) [SEQ ID NOS:104 and 105] of the humanized AF2 antibody (lower lincs) and human Eu antibody (upper lines), not including signal sequences. The three CDR's in each chain are underlined. Residues in the framework that have been replaced with mouse amino acids or typical human amino acids in the humanized antibody are double underlined.

FIG. 45A and FIG. 45B. Oligonucleotides used in the construction of the humanized AF2 light chain (A; rh10-rh13) [SEQ ID NOS:106, 107, 108, and 109] and heavy chain (B; rh20-23) [SEQ ID NOS:110, 111, 112, and 113]. The following pairs of oligonucleotides were mixed, extended with Kienow polymerase and cut with the indicated enzymes before ligation into pUC18: rh10 and rh11 with Xba I and Hind III, rh12 and rh13 with Xba I and Hind III, rh20 and rh21 with Xba I and EcoR I, rh22 and rh23 with Xba I and EcoR I. Then the rh10-rh11 and rh12-rh13 fragments were excised with Xba I and Hind III and ligated together into then Xba I site of pVk; and the rh20-rh21 and rh22-rh23 fragments were excised with Xba I and Xho I and ligated together into the Xba I site of pVgI-dhfr.

FIG. 46. Fluorescence of HS294T cells incubated with γ-IFN plus varying concentrations of mouse AF2 antibody, and stained with an anti-HLA-D antibody.

DETAILED DESCRIPTION OF THE INVENTION

In accordance with the present invention, novel means of designing humanized immunoglobulins capable of specifically binding to a predetermined antigen with strong affinity are provided. These improved methods produce immunoglobulins that are substantially non-immunogenic in humans but have binding affinities of at least about 10⁸ M⁻¹, preferably 10⁹ M⁻¹ to 10¹⁰ M⁻¹, or stronger. The humanized immunoglobulins will have a human framework and have one or more complementary determining regions (CDR's), plus a limited number of other amino acids, from a donor immunoglobulin specifically reactive with an antigen. The immunoglobulins can be produced economically in large quantities and find use, for example, in the treatment of

various human disorders by a variety of techniques.

In order that the invention may be more completely understood, several definitions are set forth. As used herein, the term "immunoglobulin" refers to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma (IgG_{1,} IgG_{2,} lgG₃ IgG₄), delta, epsilon and mu constant region genes, as well as the myriad immunoglobulin variable region genes. Fulllength immunoglobulin "light chains" (about 25 Kd or 214 amino acids) are encoded by a variable region gene at the NH2-terminus (about 110 amino acids) and a kappa or lambda constant region gene at the COOH - terminus. Full-length immunoglobulin "heavy chains" (about 50 Kd or 446 amino acids), are similarly encoded by a variable region gene (about 116 amino acids) and one of the other aforementioned constant region genes, e.g., gamma (encoding about 330 amino acids).

One form of immunoglobulin constitutes the basic structural unit of an antibody. This form is a terramer and consists of two identical pairs of immunoglobulin chains, each pair having one light and one heavy chain. In each pair, the light and heavy chain variable regions are together responsible for binding to an antigen, and the constant regions are responsible for the antibody effector functions. In addition to antibodies, immunoglobulins may exist in a variety of other forms including, for example, Fv, Fab, and (Fab')2, as well as bifunctional hybrid antibodies (e.g., Lanzavecchia et al., Eur. J. Immunol. 17, 105 (1987)) and in single chains (e.g., Huston et al., Proc. Natl. Acad. Sci. U.S.A., 85, 5879-5883 (1988) and Bird et al., Science, 242, 423-426 (1988), which are incorporated herein by reference). (See, generally, Hood et al., "Immunology", Benjamin, N.Y., 2nd ed. (1984), and Hunkapiller and Hood, Nature, 323, 15-16 (1986), which are incorporated herein by reference).

An immunoglobulin light or heavy chain variable region consists of a "framework" region interrupted by three hypervariable regions, also called CDR's. The extent of the framework region and CDR's have been precisely defined (see, "Sequences of Proteins of Immunological Interest," E. Kabat et al., U.S. Department of Health and Human Services, (1983); which is incorporated herein by reference). The sequences of the framework regions of different light or heavy chains are relatively conserved within a species. As used herein, a "human framework region" is a framework region that is substantially identical (about 85% or more, usually 90-95% or more) to the framework region of a naturally occurring human immunoglobulin. The framework region of an antibody, that is the combined framework regions of the constituent light and heavy chains, serves to 50 position and align the CDR's. The CDR's are primarily responsible for binding to an epitope of an antigen.

Chimeric antibodies are antibodies whose light and heavy chain genes have been constructed, typically by genetic engineering, from immunoglobulin variable and constant region genes belonging to different species. For example, the variable segments of the genes from a mouse monoclonal antibody may be joined to human constant segments, such as gamma 1 and gamma 3. A typical therapeutic chimeric antibody is thus a hybrid protein composed of the variable or antigen-binding domain from a mouse antibody and the constant or effector domain from a human antibody (e.g., A.T.C.C. Accession No. CRL 9688 secretes an anti-Tac chimeric antibody), although other mammalian species may be used

As used herein, the term "humanized" immunoglobulin refers to an immunoglobulin comprising a human frame12

work region and one or more CDR's from a non-human (usually a mouse or rat) immunoglobulin. The non-human immunoglobulin providing the CDR's is called the "donor" and the human immunoglobulin providing the framework is called the "acceptor". Constant regions need not be present, but if they are, they must be substantially identical to human immunoglobulin constant regions, i.e., at least about 85-90%, preferably about 95% or more identical. Hence, all parts of a humanized immunoglobulin, except possibly the CDR's, are substantially identical to corresponding parts of natural human immunoglobulin sequences. A "humanized antibody" is an antibody comprising a humanized light chain and a humanized heavy chain immunoglobulin. For example, a humanized antibody would not encompass a typical chimeric antibody as defined above, e.g., because the entire variable region of a chimeric antibody is non-human. One says that the donor antibody has been "humanized", by the process of "humanization", because the resultant humanized antibody is expected to bind to the same antigen as the donor antibody that provides the CDR's.

It is understood that the humanized antibodies designed by the present method may have additional conservative amino acid substitutions which have substantially no effect on antigen binding or other immunoglobulin functions. By conservative substitutions is intended combinations such as gly, ala; val, ile, leu; asp, glu; asn, gln; ser, thr; lys, arg; and phe, tyr.

Humanized immunoglobulins, including humanized antibodies, have been constructed by means of genetic engineering. Most humanized immunoglobulins that have been previously described (Jones et al., op. cit.; Verhoeyen et al., op. cit.; Riechmann et al., op. cit.) have comprised a framework that is identical to the framework of a particular human immunoglobulin chain, the acceptor, and three CDR's from a non-human donor immunoglobulin chain. In one case (Ricchmann et al., op. cit.), two additional amino acids in the framework were changed to be the same as amino acids in other human framework regions. The present invention includes criteria by which a limited number of amino acids in the framework of a humanized immunoglobulin chain are chosen to be the same as the amino acids at those positions in the donor rather than in the acceptor, in order to increase the affinity of an antibody comprising the humanized immunoglobulin chain.

The present invention is based in part on the model that two contributing causes of the loss of affinity in prior means of producing humanized antibodies (using as examples mouse antibodies as the source of CDR's) arc:

- (1) When the mouse CDR's are combined with the human framework, the amino acids in the framework close to the CDR's become human instead of mouse. Without intending to be bound by theory, we believe that these changed amino acids may slightly distort the CDR's, because they create different electrostatic or hydrophobic forces than in the donor mouse antibody, and the distorted CDR's may not make as effective contacts with the antigen as the CDR's did in the donor antibody;
- (2) Also, amino acids in the original mouse antibody that are close to, but not part of, the CDR's (i.e., still part of the framework), may make contacts with the antigen that contacture to affinity. These amino acids are lost when the antibody is humanized, because all framework amino acids are made human.

To avoid these problems, and to produce humanized antibodies that have a very strong affinity for a desired

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antigen, the present invention uses one or more of the following principles for designing humanized immunoglobulins. Also, the criteria may be used singly, or when necessary in combination, to achieve the desired affinity or other characteristics.

A principle is that as acceptor, a framework is used from a particular human immunoglobulin that is unusually homologous to the donor immunoglobulin to be humanized, or use a consensus framework from many human antibodies. For example, comparison of the sequence of a mouse heavy 10 (or light) chain variable region against human heavy (or light) variable regions in a data bank (for example, the National Biomedical Research Foundation Protein Identification Resource) shows that the extent of homology to different human regions varies greatly, typically from about 15 40% to about 60-70%. By choosing as the acceptor immunoglobulin one of the human heavy (respectively light) chain variable regions that is most homologous to the heavy (respectively light) chain variable region of the donor immunoglobulin, fewer amino acids will be changed in going 20 from the donor immunoglobulin to the humanized immunoglobulin. Hence, and again without intending to be bound by theory, it is believed that there is a smaller chance of changing an amino acid near the CDR's that distorts their conformation. Moreover, the precise overall shape of a 25 humanized antibody comprising the humanized immunoglobulin chain may more closely resemble the shape of the donor antibody, also reducing the chance of distorting the CDR's.

Typically, one of the 3–5 most homologous heavy chain 30 variable region sequences in a representative collection of at least about 10 to 20 distinct human heavy chains will be chosen as acceptor to provide the heavy chain framework, and similarly for the light chain. Preferably, one of the 1–3 most homologous variable regions will be used. The selected 35 acceptor immunoglobulin chain will most preferably have at least about 65% homology in the framework region to the donor immunoglobulin.

In many cases, it may be considered preferable to use light and heavy chains from the same human antibody as acceptor 40 sequences, to be sure the humanized light and heavy chains will make favorable contacts with each other. In this case, the donor light and heavy chains will be compared only against chains from human antibodies whose complete sequence is known, e.g., the Eu, Lay, Pom, Wol, Sie, Gal, Ou 45 and WEA antibodies (Kabat et al., op. cit.; occasionally, the last few amino acids of a human chain are not known and must be deduced by homology to other human antibodies). The human antibody will be chosen in which the light and heavy chain variable regions sequences, taken together, are 50 overall most homologous to the donor light and heavy chain variable region sequences. Sometimes greater weight will be given to the heavy chain sequence. The chosen human antibody will then provide both light and heavy chain acceptor sequences. In practice, it is often found that the 55 human Eu antibody will serve this role.

Regardless of how the acceptor immunoglobulin is chosen, higher affinity may be achieved by selecting a small number of amino acids in the framework of the humanized immunoglobulin chain to be the same as the amino acids at 60 those positions in the donor rather than in the acceptor. A second principle is that the following categories define what amino acids may be selected from the donor. Preferably, at many or all amino acid positions in one of these categories, the donor amino acid will in fact be selected.

Category 1: The amino acid position is in a CDR is defined by Kabat et al., op. cit.

Category 2: If an amino acid in the framework of the human acceptor immunoglobulin is unusual (i e., "rare", which as used herein indicates an amino acid occurring at that position in less than about 20% but usually less than about 10% of human heavy (respectively light) chain V region sequences in a representative data bank), and if the donor amino acid at that position is typical for human sequences (i.e., "common", which as used herein indicates an amino acid occurring in more than about 25% but usually more than about 50% of sequences in a representative data bank), then the donor amino acid rather than the acceptor may be selected. This criterion helps ensure that an atypical amino acid in the human framework does not disrupt the antibody structure. Moreover, by replacing an unusual amino acid with an amino acid from the donor antibody that happens to be typical for human antibodies, the humanized antibody may be made less immunogenic.

All human light and heavy chain variable region sequences are respectively grouped into "subgroups" of sequences that are especially homologous to each other and have the same amino acids at certain critical positions (Kabat et al., op. cit.). When deciding whether an amino acid in a human acceptor sequence is "rare" or "common" among human sequences, it will often be preferable to consider only those human sequences in the same subgroup as the acceptor sequence.

Category 3: In the positions immediately adjacent to one or more of the 3 CDR's in the primary sequence of the humanized immunoglobulin chain, the donor amino acid(s) rather than acceptor amino acid may be selected. These amino acids are particularly likely to interact with the amino acids in the CDR's and, if chosen from the acceptor, to distort the donor CDR's and reduce affinity. Moreover, the adjacent amino acids may interact directly with the antigen (Amit et al., Science, 233, 747–753 (1986), which is incorporated herein by reference) and selecting these amino acids from the donor may be desirable to keep all the antigen contacts that provide affinity in the original antibody.

Category 4: A 3-dimensional model, typically of the original donor antibody, shows that certain amino acids outside of the CDR's are close to the CDR's and have a good probability of interacting with amino acids in the CDR's by hydrogen bonding, Van der Waals forces, hydrophobic interactions, etc. At those amino acid positions, the donor immunoglobulin amino acid rather than the acceptor immunoglobulin amino acid may be selected. Amino acids according to this criterion will generally bave a side chain atom within about 3 angstrom units of some atom in the CDR's and must contain an atom that could interact with the CDR atoms according to established chemical forces, such as those listed above.

In the case of atoms that may form a hydrogen bond, the 3 angstroms is measured between their nuclei, but for atoms that do not form a bond, the 3 angstroms is measured between their Van der Waals surfaces. Hence, in the latter case, the nuclei must be within about 6 angstroms (3+sum of the Van der Waals radii) for the atoms to be considered capable of interacting. In many cases the nuclei will be from 4 or 5 to 6 Å apart. In determining whether an amino acid can interact with the CDRs, it is preferred not to consider the last 8 amino acids of heavy chain CDR 2 as part of the CDRs, because from the viewpoint of structure, these 8 amino acids behave more as part of the framework.

Amino acids in the framework that are capable of interacting with amino acids in the CDR's, and which therefore belong to Category 4, may be distinguished in another way. The solvent accessible surface area of each framework

amino acid is calculated in two ways: (I) in the intact antibody, and (2) in a hypothetical molecule consisting of the antibody with its CDRs removed. A significant difference between these numbers of about 10 square angstroms or more shows that access of the framework amino acid to solvent is at least partly blocked by the CDRs, and therefore that the amino acid is making contact with the CDRs. Solvent accessible surface area of an amino acid may be calculated based on a 3-dimensional model of an antibody, using algorithms known in the art (e.g., Connolly, J. Appl. 10 Cryst. 16,548 (1983) and Lee and Richards, J. Mol. Biol. 55, 379 (1971), both of which are incorporated herein by reference). Framework amino acids may also occasionally interact with the CDR's indirectly, by affecting the conformation of another framework amino acid that in turn con- 15 tacts the CDR's.

The amino acids at several positions in the framework are known to be capable of interacting with the CDRs in many antibodies (Chothia and Lesk, J. Mol. Biol. 196, 901 (1987), Chothia et al., Nature 342, 877 (1989), and Tramontano et 20 al., J. Mol. Biol. 215, 175 (1990), all of which are incorporated herein by reference), notably at positions 2, 48, 64 and 71 of the light chain and 26-30, 71 and 94 of the heavy chain (numbering according to Kabat, op. cit.), and therefore these amino acids will generally be in Category 4. Typically, 25 humanized immunoglobulins, of the present invention will include donor amino acids (where different) in category 4 in addition to these. The amino acids at positions 35 in the light chain and 93 and 103 in the heavy chain are also likely to interact with the CDRs. At all these numbered positions, 30 choice of the donor amino acid rather than the acceptor amino acid (when they differ) to be in the humanized immunoglobulin is preferred. On the other hand, certain positions that may be in Category 4 such as the first 5 amino acids of the light chain may sometimes be chosen from the 35 acceptor immunoglobulin without loss of affinity in the humanized immunoglobulin.

Chothia and Lesk (op. cit.) define the CDRs differently from Kabat et al. (op. cit.). Notably, CDR1 is defined as including residues 26-32. Accordingly, Riechmann ct al., 40 (op. cit.) chose these amino acids from the donor immunoglobulins.

Computer programs to create models of proteins such as antibodies are generally available and well known to those shilled in the art (sec, Levy et al., *Biochemistry*, 28, 7168-7175 (1989); Bruccoleri et al., *Nature*, 335, 564–568 (1988); Chothia et al., *Science*, 233, 755–758 (1986), all of which are incorporated herein by reference). These do not form part of the invention. Indeed, because all antibodies have similar structures, the known antibody structures, which are so available from the Brookhaven Protein Data Bank, can be used if necessary as rough models of other antibodies. Commercially available computer programs can be used to display these models on a computer monitor, to calculate the distance between atoms, and to estimate the likelihood of 55 different amino acids interacting (see, Feirin et al., *J. Mol. Graphics*, 6, 13–27 (1988)).

In addition to the above categories, which describe when an amino acid in the humanized immunoglobulin may be taken from the donor, certain amino acids in the humanized 60 immunoglobulin may be taken from neither the donor nor acceptor, if then fall in:

Category 5: If the amino acid at a given position in the donor immunoglobulin is "rare" for human sequences, and the amino acid at that position in the acceptor immunoglo- 65 bulin is also "rare" for human sequences, as defined above, then the amino acid at that position in the humanized

immunoglobulin may be chosen to be some amino acid "typical" of human sequences. A preferred choice is the amino acid that occurs most often at that position in the known human sequences belonging to the same subgroup as the acceptor sequence.

Humanized antibodies generally have at least three potential advantages over mouse or in some cases chimeric antibodies for use in human therapy:

- Because the effector portion is human, it may interact better with the other parts of the human immune system (e.g., destroy the target cells more efficiently by complement-dependent cytotoxicity (CDC) or antibody-dependent cellular cytotoxicity (ADCC)).
- 2) The human immune system should not recognize the framework or constant region of the humanized antibody as foreign, and therefore the antibody response against such an injected antibody should be less than against a totally foreign mouse antibody or a partially foreign chimeric antibody.
- 3) Injected mouse antibodies have been reported to have a half-life in the human circulation much shorter than the half-life of normal antibodies (D. Shaw et al., J. Immunol., 138, 4534-4538 (1987)). Injected humanized antibodies will presumably have a half-life more similar to naturally occurring human antibodies, allowing smaller and less frequent doses to be given.

In one aspect, the present invention is directed to designing humanized immunoglobulins that are produced by expressing recombinant DNA segments encoding the heavy and light chain CDR's from a donor immunoglobulin capable of binding to a desired antigen, such as the human IL-2 receptor, attached to DNA segments encoding acceptor human framework regions. Exemplary DNA sequences designed in accordance with the present invention code for the polypeptide chains comprising heavy and light chain CDR's with substantially human framework regions shown in FIG. 1A through FIG. 6B. Due to codon degeneracy and non-critical amino acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. In general, the criteria of the present invention find applicability to designing substantially any humanized immunoglobulin.

The DNA segments will typically fruther include an expression control DNA sequence operably linked to the humanized immunoglobulin coding sequences, including naturally-associated or heterologous promoter regions. Preferably, the expression control sequences will be eukaryotic promoter systems in vectors capable of transforming or transfecting eukaryotic host cells, but control sequences for prokaryotic hosts may also be used. Once the vector has been incorporated into the appropriate host, the host is maintained under conditions suitable for high level expression of the nucleotide sequences, and, as desired, the collection and purification of the humanized light chains, heavy chains, light/heavy chain dimers or intact antibodies, binding fragments or other immunoglobulin forms may follow (see, S. Beychok, Cells of Immunoglobulin Synthesis, Academic Press, N.Y., (1979), which is incorporated herein by reference).

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immortalized B-cells (see, Kabat op. cit. and WP87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to the predetermined antigen, such as the human IL-2 receptor, and produced by well known methods in any convenient

mammalian source including, mice, rats, rabbits, or other vertebrates, capable of producing antibodies. Suitable source cells for the constant region and framework DNA sequences, and host cells for immunoglobulin expression and secretion, can be obtained from a number of sources, such as the American Type Culture Collection ("Catalogue of Cell Lines and Hybridomas," sixth edition (1988) Rockville, Md. U.S.A., which is incorporated herein by reference)

In addition to the humanized immunoglobulins specifi- 10 cally described herein, other "substantially homologous" modified immunoglobulins to the native sequences can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the framework regions can vary specifi- 15 cally from the sequences in FIG. 1A through FIG. 6B at the primary structure level by several amino acid substitutions, terminal and intermediate additions and deletions, and the like. Moreover, a variety of different human framework regions may be used singly or in combination as a basis for 20 the humanized immunoglobulins of the present invention. In general, modifications of the genes may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Gillman and Smith, Gene, 8, 81-97 (1979) and S. Roberts et al., Nature, 328, 731-734 25 (1987), both of which are incorporated herein by reference).

Substantially homologous immunoglobulin sequences are those which exhibit at least about 85% homology, usually at least about 90%, and preferably at least about 95% homology with a reference immunoglobulin protein.

Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic 35 cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors pVk and pVgl (FIGS. 9A and 9B) using sitedirected mutagenesis, such as after CH1 to produce Fab fragments or after the hinge region to produce (Fab')2 40 fragments. Single chain antibodies may be produced by joining VL and VH with a DNA linker (see, Huston et al., op. cit., and Bird et al., op. cit.). Also because like many genes, the immunoglobulin-related genes contain separate functional regions, each having one or more distinct biological 45 activities, the genes may be fused to functional regions from other genes (e.g., enzymes, see, commonly assigned U.S. Pat. No. 5,004,692) to produce fusion proteins (e.g., immunotoxins) having novel properties. The nucleic acid sequences of the present invention capable of ultimately 50 expressing the desired humanized antibodies can be formed from a variety of different polynucleotides (genomic or cDNA, RNA, synthetic oligonucleotides, etc.) and components (e.g., V, J, D, and C regions), as well as by a variety of different techniques. Joining appropriate synthetic and 55 genomic sequences is presently the most common method of production, but cDNA sequences may also be utilized (see, European Patent Publication No. 0239400 and L. Reichmann et al., Nature, 332, 323-327 (1988), both of which are incorporated herein by reference).

As stated previously, the DNA sequences will be expressed in hosts after the sequences have been operably linked to (i.e., positioned to ensure the functioning of) an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes 65 or as an integral part of the host chromosomal DNA. Commonly, expression vectors will contain selection mark-

ers, e.g., tetracycline or neomycin, to permit detection of those cells transformed with the desired DNA sequences (see, e.g., U.S. Pat. No. 4,704,362, which is incorporated herein by reference).

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E. coli is one prokaryonic host useful particularly for cloning the DNA sequences of the present invention. Other microbial hosts suitable for use include bacilli, such as Bacillus subtilus, and other enterobacteriaceae, such as Salmonella, Serratia, and various Pseudomonas species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of wellknown promoters will be present, such as the lactose promoter system, a tryptophan (pp) promoter system, a betalactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing transcription and translation.

Other microbes, such as yeast, may also be used for expression. Saccharomyces is a preferred host, with suitable vectors having expression control sequences, such as promoters, including 3-phosphoglycerate kinase or other glycolytic enzymes, and an origin of replication, termination sequences and the like as desired.

In addition to microorganisms, mammalian tissue cell culture may also be used to express and produce the polypeptides of the present invention (see, Winnacker, "From Genes to Ciones," VCH Publishers, N.Y., N.Y. (1987), which is incorporated herein by reference). Eukaryotic cells are actually preferred, because a number of suitable host cell lines capable of secreting intact immunoglobulins have been developed in the art, and include the CHO cell lines, various COS cell lines, HeLa cells, preferably myeloma cell lines, etc, and transformed B-cells or hybridomas. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter, an enhancer (Queen et al., Immunol. Rev., 89, 49-68 (1986), which is incorporated herein by reference), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences are promoters derived from immunoglobulin genes, SV40, Adenovirus, cytomegaloviius, Bovine Papilloma Virus, and the like.

The vectors containing the DNA segments of interest (e.g., the heavy and light chain encoding sequences and expression control sequences) can be transferred into the host cell by well-known methods, which vary depending on the type of cellular host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cellular hosts. (See, generally, Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press, (1982), which is incorporated herein by reference.)

Once expressed, the whole antibodies, their dimers, individual light and heavy chains, or other immunoglobulin forms of the present invention, can be purified according to standard procedures of the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like (see, generally, R. Scopes, "Protein Purification", Springer-Verlag, N.Y. (1982)). Substantially pure immunoglobulins of at least about 90 to 95% homogeneity are preferred, and 98 to 99% or more homogeneity most preferred, for pharmaceutical uses. Once puri-

fied, partially orto homogeneity as desired, the polypeptides may then be used therapeutically (including extracorporeally) or in developing and performing assay procedures, immunofluorescent stainings, and the like. (See, generally, *Immunological Methods*, Vols. I and II, Lefkovits and Pernis, eds., Academic Press, New York, N.Y. (1979 and 1981)).

The antibodies of the present invention will typically find use individually in treating substantially any disease susceptible to monoclonal antibody-based therapy. In particular, the immunoglobulins can be used for passive immuni- 10 zation or the removal of unwanted cells or antigens, such as by complement mediated lysis, all without substantial immune reactions (e.g., anaphylactic shock) associated with many prior antibodies. For example, where the cell linked to a disease has been identified as 11.-2 receptor bearing, then 15 humanized antibodies that bind to the human IL-2 receptor are suitable (see, U.S. Ser. No. 085,707, entitled "Treating Human Malignancies and Disorders," which is incorporated herein by reference). For such a humanized immunoglobulin, typical disease states suitable for treatment include graft versus host disease and mansplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia 25

The method of producing humanized antibodies of the present invention can be used to humanize a variety of donor antibodies, especially monoclonal antibodies reactive with markers on cells responsible for a disease. For example, 30 suitable antibodies bind to antigens on T-cells, such as those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, Leukocyte Typing, Bernard et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by referance.

The antibodies of the present invention can also be used as separately administered compositions given in conjunction with chemotherapeutic or immunosuppressive agents. Possible agents include cyclosporin A or a purinc analog 40 (e.g., methotrexate, 6-mercaptopurine, or the like), but numcrous additional agents (e.g., cyclophosphamide, prednisone, etc.) well-known to those skilled in the art of medicine may also be utilized.

A preferred pharmaceutical composition of the present 45 invention comprises the use of the subject antibodies in immunotoxins. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. 50 The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells comprising a carcinoma. The two components are commonly chemically bonded together by any of a variety of well-known chemical procedures. For 55 example, when the cytotoxic agent is a protein and the second component is an intact immunoglobulin, the linkage may be by way of heterobifunctional cross-linkers, e.g., SPDP, carbodiimide, glutaraldehyde, or the like. Production of various immunotoxins is well-known with the art, and can 60 be found, for example in "Monoclonal Antibody-Toxin Conjugates: Aiming the Magic Bullet," Thorpe et al., Monoclonal Antibodies in Clinical Medicine, Academic Press, pp. 168-190 (1982), which is incorporated herein by reference. The components may also be linked genetically (see, 65 Chaudhary et al., Nature 339, 394 (1989), which is herein incorporated by reference).

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A variety of cytotoxic agents are suitable for use in immunotoxins. Cytotoxic agents can include radionuclides, such as lodine-131 or other isotopes of iodine, Yttrium-90, Rhenium-188, and Bismuth-212 or other alpha emitters; a number of chemotherapeutic drugs, such as vindesine, methotrexate, adriamycin, and cisplatinum; and cytotoxic proteins such as ribosomal inhibiting proteins like pokeweed antiviral protein, Pscudomonas exotoxin A, ricin, diphtheria toxin, ricin A chain, etc., or an agent active at the cell surface, such as the phospholipase enzymes (e.g., phospholipase C). (See, generally, "Chimeric Toxins," Olsnes and Phil, *Pharmac. Ther.*, 25, 355–381 (1982), and "Monoclonal Antibodies for Cancer Detection and Therapy," eds. Baldwin and Byers, pp. 159–179, 224–266, Academic Press (1985), all of which are incorporated herein by reference.)

The delivery component of the immunotoxin will include the humanized immunoglobulins of the present invention. Intact immunoglobulins or their binding fragments, such as Fab, are preferably used. Typically, the antibodies in the immunotoxins will be of the human lgM or lgG isotypc, but other mammalian constant regions may be utilized as desired.

For diagnostic purposes, the antibodies may either be labeled or unlabeled. Unlabeled antibodies can be used in combination with other labeled antibodies (second antibodies) that are reactive with the humanized antibody, such as antibodies specific for human immunoglobulin constant regions. Alternatively, the antibodies can be directly labeled. A wide variety of labels may be employed, such as radionuclides, fluors, enzymes, enzyme substrates, enzyme cofactors, enzyme inhibitors, ligands (particularly haptens), etc. Numerous types of immunoassays are available and are well known to those skilled in the art.

Anti-II-2 Receptor Antibodies

To exert its biological effects, IL-2 interacts with a specific high-affinity membrane receptor (Greene, W., et al., Progress in Hematology XIV, E. Brown, Ed., Grune and Statton, New York (1986), at pgs. 283 ff and Waldmann, Ann. Rev. Biochem. 58, 875 (1989), both of which are incorporated herein by reference). The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide, being about 55 kD in size (see, Leonard, W., ct al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). A gene encoding this protein has been isolated, and predicts a 272 amino acid peptide, including a 21 amino acid signal peptide (see, Leonard, W., et al., Nature 311, 626 (1984)). The 219 NH₂-terminal amino acids of the p55 Tac protein apparently comprise an extracellular domain (see, Leonard, W., et al., Science, 230, 633-639 (1985), which is incorporated herein by reference).

Much of the elucidation of the human IL-2 receptor's structure and function is due to the development of specifically reactive monoclonal antibodies. In particular, one mouse monoclonal antibody, known as anti-Tac (Uchiyama, et al., *J. Immunol.* 126, 1393 (1981)) has been used to show that IL-2 receptors can be detected on T-cells, but also on cells of the monocyte-macrophage family, Kupffer cells of the liver, Langerhans' cells of the skin and, of course, activated T-cells. Importantly, resting T-cells, B-cells or circulating machrophages typically do not display the IL-2 receptor (Herrmann, et al., *J. Exp. Med.* 162, 1111 (1985)).

The anti-Tac monoclonal antibody has also been used to define lymphocyte functions that require IL-2 interaction, and has been shown to inhibit various T-cell functions,

including the generation of cytotoxic and suppressor T lymphocytes in cell culture. Also, based on studies with anti-Tac and other antibodies, a variety of disorders are now associated with improper IL-2 receptor expression by T-cells, in particular adult T-cell leukemia.

More recently, the IL-2 receptor has been shown to be an ideal target for novel therapeutic approaches to T-cell mediated diseases. It has been proposed that IL-2 receptor specific antibodies, such as the anti-Tac monoclonal antibody, can be used either alone or as an immunoconjugate 10 (e.g., with Ricin A, isotopes and the like) to effectively remove cells bearing the IL-2 receptor. These agents can, for example, theoretically eliminate IL-2 receptor-expressing leukemic cells, certain B-cells, or activated T-cells involved in a disease state, yet allow the retention of mature normal 15 T-cells and their precursors to ensure the capability of mounting a normal T-cell immune response as needed. In general, most other T-cell specific agents can destroy essentially all peripheral T-cells, which limits the agents' therapeutic efficacy. Overall, the use of appropriate monoclonal 20 antibodics specific for the IL-2 receptor may have therapeutic utility in autoimmune diseases, organ transplantation and any unwanted response by activated T-cells. Indeed, clinical trials have been initiated using, e.g., anti-Tac antibodies (see, generally, Waldmann, T., et al., Cancer Res. 45, 625 (1985), 25 Waldmann, T., Science 232, 727-732 (1986) and Kirkman et al., Transplant. Proc. 21, 1766 (1989), all of which are incorporated herein by reference).

Unfortunately, the use of the anti-Tac and other non-human monoclonal antibodies have certain drawbacks, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

Perhaps more importantly, anti-Tac and other non-human monoclonal antibodies contain substantial stretches of amino acid sequences that will be immunogenic when injected into a human patient. Numerous studies have shown that, after injection of a foreign antibody, the immune response elicited by a patient against an antibody can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, as increasing numbers of different mouse or other antigenic (to humans) monoclonal antibodies can be expected to be developed to treat various diseases, after the first or several treatments with any different non-human antibodies, subsequent treatments even for unrelated therapies can be ineffective or even dangerous in themselves, because of cross-reactivity.

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of human immunoglobulins reactive with the human IL-2 receptor, as with many human antigens, has been extremely difficult using typical human monoclonal antibody production techniques. Similarly, utilizing recombinant DNA technology to produce so-called "reshaped" or "humanized" antibodies (see, c.g., Riechmann et al., *Nature* 332, 323 (1988) and EPO Publication No. 0239460), provides uncertain results, in part due to unpredictable binding affinities.

Thus, there is a need for improved forms of human-like 65 immunoglobulins specific for the human IL-2 receptor that are substantially non-immunogenic in humans, yet easily

and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of T-cell mediated human disorders, the compositions containing human-like immunoglobulins specifically capable of blocking the binding of human IL-2 to its receptor and/or capable of binding to the p55 Tac protein on human IL-2 receptors. The immunoglobulins can have two pairs of light chain/heavy chain complexes, typically at least one chain comprising mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be used to produce human-like antibodies capable of binding to the human IL-2 receptor at affinity levels stronger than about 10⁸ M⁻¹.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for human-like immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The human-like immunoglobulins may be utilized alone in substantially pure form, or complexed with a cytotoxic agent, such as a radionuclide, a ribosomal inhibiting protein or a cytotoxic agent active at cell surfaces. All of these compounds will be particularly useful in treating T-cell mediated disorders. The human-like immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, human-like immunoglobulins specifically reactive with the IL-2 receptor on human T-cells are provided. These immunoglobulins, which have binding affinities of at least about 10⁸ M⁻¹, and preferably 10⁹ M⁻¹ to 10¹⁰ M⁻¹ or stronger, are capable of, e.g., blocking the binding of IL-2 to human IL-2 receptors. The human-like immunoglobulins will have a human-like framework and can have complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an epitope on p55 Tac protein. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of T-cell mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope on the human IL-2 receptor, such as the anti-Tac monoclonal antibody. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human-like framework regions. Preferred DNA sequences, which on expression code for the polypept de chains comprising the anti-Tac heavy and light chain hypervariable regions (with human-like framework regions), are included in FIGS. 15A and 16A, respectively. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating a T-cell mediated disease state. Generally, where the cell linked to a disease has been identified as IL-2 receptor bearing, then the human-like antibodies capable of blocking the binding of IL-2 to the 5 human IL-2 receptor are suitable.

For example, typical disease states suitable for treatment include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The human-like antibodies of the present invention may also be used in combination with other antibodies, particularly human monoclonal antibodies reactive with other markers on cells responsible for the disease. For example, suitable T-cell markers can include those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, *Leukocyte Typing*, Bernard, et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference.

The human-like antibodies and pharmaceutical compositions thereof of this invention are particularly useful for parenteral administration, i.e., subcutaneously, intramuscularly or intravenously. The compositions for parenteral administration will commonly comprise a solution of the antibody or a cocktail thereof dissolved in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers can be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine and the like. These solutions are sterile and generally free of particulate matter. These compositions may be sterilized by conventional, well known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents, toxicity adjusting agents and the like, for example sodium acetate, sodium chloride, potassium chloride, calcium chloride, sodium lactate, human albumin, etc. The concentration of antibody in these formulations can vary widely, i.e., from less than about 0.5%, usually at or at least about 1% to as much as 15 or 20% by weight and will be selected primarily based on Huid volumes, viscosities, etc., in accordance with the particular mode of administration selected.

Thus, a typical pharmaceutical composition for injection could be made up to contain 1 ml sterile buffered water, and 1 to 50 mg of antibody. A typical composition for intravenous infusion could be made up to contain 250 ml of sterile Ringer's solution, and 150 mg of antibody. Actual methods for preparing parenterally administrable compositions will be known or apparent to those skilled in the art and are described in more detail in, for example, Remington's Pharmaceutical Science, 15th ed., Mack Publishing Company, Easton, Pa. (1980), which is incorporated herein by reference.

The antibodies of this invention can be frozen or lyophilized for storage and reconstituted in a suitable carrier prior to use. This technique has been shown to be effective with conventional immune globulins and art-known lyophilization and reconstitution techniques can be employed. It will be appreciated by those skilled in the art that lyophilization and reconstitution can lead to varying degrees of antibody activity loss (e.g., with conventional immune globulins, IgM antibodies tend to have greater activity loss 65 than IgG antibodies) and that use levels may have to be adjusted to compensate.

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The compositions containing the present human-like antibodies or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from a disease, in an amount sufficient to cure or at least partially arrest the disease and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the infection and the general state of the patient's own immune system, but generally range from about 1 to about 200 mg of antibody per dose, with dosages of from 5 to 25 mg being more commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease states, that is life-threatening or potentially lifethreatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability of "foreign substance" rejections which are achieved by the present human-like antibodies of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these antibodies.

In prophylactic applications, compositions containing the present antibodies or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 0.1 to 25 mg per dose, especially 0.5 to 2.5 mg per dose. A preferred prophylactic use is for the prevention of kidney transplant rejection.

Single or multiple administrations of the compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the antibody(ies) of this invention sufficient to effectively treat the patient.

Human-like antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for T-cell typing, for isolating specific IL-2 receptor bearing cells or fragments of the receptor, for vaccine preparation, or the like.

Kits can also be supplied for use with the subject antibodies in the protection against or detection of a cellular activity or for the presence of a selected antigen. Thus, the subject antibody composition of the present invention may be provided, usually in a lyophilized form in a container, either alone or in conjunction with additional antibodies specific for the desired cell type. The antibodies, which may be conjugated to a label or toxin, or unconjugated, are included in the kits with buffers, such as Tris, phosphate, carbonate, etc., stabilizers, biocides, inert proteins, e.g., serum albumin, or the like, and a set of instructions for use. Generally, these materials will be present in less than about 5% wt. based on the amount of active antibody, and usually present in total amount of at least about 0.001% wt. based again on the antibody concentration. Frequently, it will be desirable to include an inert extender or excipient to dilute the active ingredients, where the excipient may be present in from about 1 to 99% wt. of the total composition. Where a second antibody capable of binding to the chimeric antibody is employed in an assay, this will usually be present in a separate vial. The second antibody is typically conjugated to a label and formulated in an analogous manner with the antibody formulations described above.

p75 Chain of IL-2 Receptor

The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac peptide or alpha chain, being about 55 kD in size (see, Leonard, W., et al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). The second chain is known as the p75 or beta chain (Tsudo et al., Proc. Nat. Acad. Sci. USA, 83, 9694 (1986) and Sharon et al., Science 234, 859 (1986), both of 5 which are incorporated herein by reference). The p55 or Tac chain and the p75 chain each independently bind IL-2 with low or intermediate affinity, while the IL-2 receptor complex of both chains binds IL-2 with high affinity. The p75 chain of the human IL-2 receptor will often be called herein simply the p75 protein.

The present invention provides novel compositions useful, for example, in the reatment of T-cell mediated human disorders, the compositions containing human-like immunoglobulins specifically capable of inhibiting the binding of human IL-2 to its receptor and/or capable of binding to the p75 protein of human IL-2 receptors. The immunoglobulins can have two pairs of light chain/heavy chain complexes, typically at least one chain comprising mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse comple- 20 mentarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be used to produce human-like antibodies capable of binding to the p75 protein at affinity levels stronger than about 10⁷ M⁻¹. These humanized immunoglobulins will also be capable of 25 blocking the binding of the CDR-donating mouse monoclonal antibody to p75.

The human-like immunoglobulins may be utilized alone in substantially pure form, or complexed with a cytotoxic agent, such as a radionuclide, a ribosomal inhibiting protein or a cytotoxic agent active at cell surfaces. All of these compounds will be particularly useful in treating T-cell mediated disorders. The human-like immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, human-like immunoglobulins specifically reactive with the p75 chain of the human IL-2 receptor are provided. These immunoglobulins, which have binding affinities of at least 107 to 108 M⁻¹, and preferably 109 M⁻¹ to 10¹⁰ M⁻¹ or stronger, are capable of, e.g., blocking the binding of IL-2 to human IL-2 receptors. The human-like immunoglobulins will have a human-like framework and can have complementatity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an epitopc on p75 protein. The immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of T-cell mediated disorders in human patients by a variety of techniques.

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope on the human IL-2 receptor, such as the mik-\(\beta\)1 monoclonal antibody. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate human-like framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the mik-\(\beta\)1 heavy and light chain CDRs, are included in FIG. 23A and FIG. 23B. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodics of the present invention will typically find use individually in treating a T-cell mediated disease state.

Generally, where the cell linked to a disease has been identified as IL-2 receptor bearing, then the human-like antibodies capable of blocking the binding of IL-2 to the human IL-2 receptor are suitable.

For example, typical disease states suitable for treatment include graft-versus-host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

The human-like antibodies of the present invention may also be used in combination with other antibodies, particularly human monoclonal antibodies reactive with other markers on cells responsible for the disease. For example, suitable T-cell markers can include those grouped into the so-called "Clusters of Differentiation," as named by the First International Leukocyte Differentiation Workshop, Leukocyte Typing, Bernard, et al., Eds., Springer-Verlag, N.Y. (1984), which is incorporated herein by reference. A preferred use is the simultaneous treatment of a patient with a human-like antibody binding to p55 and a human-like antibody binding to p75 of the IL-2 receptor, i.e., humanized anti-Tac plus humanized mik-β1.

Human-like antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for T-cell typing, for isolating specific IL-2 receptor bearing cells or fragments of the receptor, for vaccine preparation, or the like.

Anti-HSV Antibodies

Herpes Simplex Virus types I and II (HSV-1 and HSV-2), are now estimated to be the second most frequent cause of sexually transmitted diseases in the world. Although completely accurate data are not available, infection estimates range from about 20 to 40% of the U.S. population.

A large number of diseases, from asymptomatic to lifethreatening, are associated with HSV infection. Of particular climical interest, encephalitis from HSV-1 infection and transmission of HSV-2 from a pregnant mother to her fetus are often fatal. Immunosuppressed patients are also subject to severe complications when infected with the virus.

More than 50 HSV polypeptides have been identified in HSV-infected cells, including at least seven major cell surface glycoproteins (see, Whitley, R., Chapt. 66, and Roizman and Sears, Chapt. 65, Virology, Eds. Fields et al., 2nd ed., Raven Press, N.Y., N.Y. (1990), which are incorporated herein by reference). The specific biologic functions of these glycoproteins are not well defined, although gB and gD have en shown to be associated with cell fusion activity (W. Cai et al., J. Virol. 62, 2596 (1988) and Fuller and Spear, Proc. Natl. Acad. Sci. USA 84, 5454 (1987)). gB and gD express both type-specific and type-common antigenic determinants. Oakes and Lausch demonstrated that monoclonal antibodies against gB and gE suppress replication of HSV-1 in trigeminal ganglia (Oakes and Lausch, J. Virol. 51, 656 (1984)). Dix et al. showed that anti-gC and gD antibodies protect mice against acute virus-induced neurological disease (Dix ct al., Infect. Immun. 34, 192 (1981)). Whitley and colleagues produced a panel of murine monoclonal antibodies against HSV-1 and showed that several of the antibodies protected mice against encephalitis and death following ocular inoculation with the virus (see, Koga et al., Virology 151, 385 (1986); Metcalf et al., Cur. Eye Res. 6, 173 (1987) and Metcalf et al., Intervirology 29, 39 1988), all

of which are incorporated herein by reference). Clone Fd79 (anti-gB) prevented encephalitis even when immunization was delayed until 48 hours post-infection. Fd79 and Fd138-80 (anti-gD) significantly reduced the severity of epithelial keratitis and lowered the frequency of persistent viral infection in an outbred mouse model.

Thus, there is a need for improved forms of humanized immunoglobulins specific for HSV antigens that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of HSV mediated human disorders, the compositions containing humanized immunoglobulins specifically capable of blocking the binding of HSV to its receptors and/or capable of binding to the HSV specific proteins. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to the HSV surface proteins at affinity levels stronger than about 107 M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR donating mouse monoclonal antibody to HSV.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with an antiviral agent, such as acyclovir or a cytotoxic agent active at viral surfaces. All of these compounds will be particularly useful in treating HSV mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with HSV related 40 epitopes either directly on the virus or on infected cells are provided. These immunoglobulins, which have binding affinities to HSV specific antigens of at least about 10⁷ M and preferably $10^8 \,\mathrm{M}^{-1}$ to $10^{10} \,\mathrm{M}^{-1}$ or stronger, are capable of, e.g., protecting cells from HSV transmission. The 45 humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with an HSV protein, such as gB and gD proteins. The immunoglobulins 50 of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of HSV mediated disorders in human patients by a variety of techniques.

The HSVs are among the most intensively investigated of 51 all viruses, and the HSV virion structure has been shown to contain about 33 proteins. Humanized immunoglobulins utilizing CDR's from monoclonal antibodies reactive with these proteins, particularly the eight surface glycoproteins (e.g., gB, gC, gD, gE, gG, gH and gl), represent preferred 60 embodiments of the present invention (see, Spear, P. G., The Herpesviruses, vol. 3, pp. 315–356 (1984) (Roizman, B., ed), Plenum Press, N.Y., N.Y. and Spear, P. G., Immunochemistry of Viruses. The Basis for Serodiagnosis and Vaccines, pp. 425–446 (1985) (Neurath, A. R., eds.), 65 Amsterdam: Elsevier, both of which are incorporated herein by reference).

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In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of an HSV protein, such as monoclonal antibodies reactive with HSV gB and gD glycoproteins. The DNA segments encoding these regions will typically be joined to DNA segments encoding appropriate humanized framework regions. Exemplary DNA sequences code for the polypeptide chains comprising the heavy and light chain hypervariable regions (with human framework regions) from monoclonal antibodies Fd79 and Fd138-80, shown in FIG. 30A through FIG. 30D. Due to codon degeneracy and non-critical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The antibodies of the present invention will typically find use individually in treating an HSV mediated disease state. For example, typical disease states suitable for treatment include any involving HSV infection. Specific diseases include neonatal herpes, herpes encephalitis, ocular herpes, genital herpes and disseminated herpes (see, Corey, L., Chapter 136, Harrison's Principles of Internal Medicine, 11th ed., McGraw-Hill Book Company, N.Y., N.Y. (1987), which is incorporated herein by reference).

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different HSV antigens. For example, suitable HSV antigens to which a cocktail of humanized immunoglobulins may react include gC, gE, gF, gG and gH (see, Rector, J. et al., Infect. Immun. 38, 168 (1982) and Fuller, A. et al., J. Virol. 63, 3435 (1989), both of which are incorporated herein by reference).

The antibodies can also be used as separately administered compositions given in conjunction with acyclovir or other antiviral agents. Typically, the agents may include idoxunidine or trifluorothymidine, but numerous additional agents (e.g., vidarabine) well-known to those skilled in the art for HSV treatment may also be utilized (sec, Corey, L., op. cit.).

A preferred pharmaceutical composition of the present invention compises the use of the subject immunoglobulins in immunotoxins to kill cells infected by HSV Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing an HSV epitope.

The compositions containing the present humanized antibodies or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In therapeutic application, compositions are administered to a patient already suffering from HSV infection, in an amount sufficient to cure or at least partially arrest the disease and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the infection and the general state of the patient's own immune system, but generally range from about 1 to about 200 mg of antibody per dose, with dosages of from 5 to 25 mg being more commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease states, that is life-threatening or potentially lifethreatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability

of "foreign substance" rejections which are achieved by the present humanized immunoglobulins of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these antibodies.

In prophylactic applications, compositions containing the present immunoglobulins or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 0.1 to 25 mg per dose. A preferred prophylactic use is for the prevention of herpes in immunocompromised patients, such as organ transplant recipients.

Single or multiple administrations of the compositions ¹⁵ can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the antibody(ics) of this invention sufficient to effectively treat the patient.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of HSV antigens, for isolating specific HSV infected cells or fragments of the virus, for vaccine preparation, or the like.

Anti-CD33 Antibodies

There are about 10,000-15,000 new cases of myeloid (also called non-lymphocytic or granulocytic) leukemia in the U.S. per year (Cancer Facts & Figures, American Cancer 30 Society, 1987). There are two major forms of myeloid leukemia: acute myelogcnous leukemia (AML) and chronic myelogcnous leukemia (CML). Desprite treatment with chemotherapy, long-term survival in patients with AML is less than 10-20% (Clarkson et al., CRC Critical Review in 35 Oncology/Hematology 4, 221 (1986)), and survival with CML and related diseases such as chronic myelomonocytic leukemia (CMML), chronic monocytic leukemia (CMMOL) and myelodysplastic syndrome (MDS) is even lower.

The p67 protein or CD33 antigen is found on the surface of progenitors of mycloid cells and of the leukemic cells of most cases of AML, but not on lymphoid cells or non-hematopoietic cells (see, Leucocyte Typing III, ed. by A. J. McMichael, Oxford University Press, pp. 622–629 (1987), which is incorporated herein by reference). Antibodies that are known to bind to the CD33 antigen include L4B3, L1B2 and MY9 (Andrews et al., Blood 62, 124 (1983) and Griffin et al., Leukemia Research 8, 521 (1984), both of which are incorporated herein by reference).

Another antibody that binds to CD33 is M195 (Tanimoto ct al., Leukemia 3, 339 (1989) and Scheinberg et al., Leukemia 3, 440 (1989), both of which are incorporated herein by reference). The reactivity of M195 with a wide variety of cells and tissues was tested. Among normal cells, M195 was reported to bind only to some monocytes and myeloid progenitor cells. The research also reported that it does not bind to other hematopoietic cells or to nonhematopoietic tissues. M195 bound to cells of most cases of AML and all cases of CML in myeloblastic phase.

A phase I clinical trial of M195 in AML has been conducted (Scheinberg et al., Proc. ASCO 9, 207 (1990)). M195 radiolabeled with iodine-131 was found to rapidly and specifically target leukemic cells in both the blood and bone

Unfortunately, the use of non-human monoclonal antibodies such as M195 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

Thus, there is a need for improved forms of humanized immunoglobulins specific for CD33 antigen that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

The present invention provides novel compositions useful, for example, in the treatment of myeloid leukemiarelated human disorders, the compositions containing humanized immunoglobulins specifically capable of binding to CD33 antigen. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to the CD33 antigen at affinity levels stronger than about 107 M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to CD33.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such as cytosine arabinoside or daunorubicin active against leukemia cells, or complexed with a radionuclide such as iodine-131. All of these compounds will be particularly useful in treating leukemia and myeloid cellmediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with CD33 related epitopes are provided. These immunoglobulins, which have binding affinities to CD33 of at least about 107 M⁻¹, and preferably 108 M⁻¹ to 1010 M⁻¹ or stronger, are capable of, e.g., destroying leukemia cells. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with CD33 antigen. In a preferred embodiment, one or more of the CDR's will come from the M195 antibody. Importantly, M195 docs not bind to the ultimate hematopoictic stem cells, so M195 used in therapy will minimally interact with and destroy those cells, which are critical for generating all blood cells. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of myeloid cell-mediated disorders in human patients by a variety of techniques.

disease; and congenitally acquired CMV, often with significant morbidity and mortality, affects 1% of newborns (Fields, op. cit.).

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In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of CD33 antigen, such as monoclonal antibodies M195, I.4B3, L1B2 or MY9. The DNA segments 5 encoding these regions will typically be joined to DNA segments encoding appropriate human framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody M195 are included in FIG. 34A and FIG. 34B. Due to codon degeneracy and noncritical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

The drug ganciclovir is effective against certain forms of CMV infection, notably choriorctinitis and gastroenteritis, but is not very effective against CMV pneumonia, and it has serious toxicity. Use of pooled human imunoglobulin preparations has shown some beneficial effect for prophylaxis of CMV in bone marrow transplant patients (Meyers, op. cit.), and a combination of high-dose immune globulin and ganciclovir has been reported effective against CMV pneumonia (Emanuel et al., Trans. Proc. XIX (suppl. 7), 132 (1987)). However, the marginal effectiveness, variable potency and high cost of commercial human immune globulin remain serious problems. Hence, there is a great need for new drugs effective against CMV.

The antibodies of the present invention will typically find use individually in treating hematologic malignancies. For example, typical disease states suitable for treatment include AML, CML, CMML, CMMOL and MDS (see, generally, Hofibrand & Pettit, Essential Haematology, Blackwell Scientific Publications, Oxford (1980)). The antibodies may also be used for bone marrow ablation prior to bone marrow ²⁰ transplant.

CMV is a member of the herpesvirus family of viruses, and as such, has a large double-stranded DNA core, a protein capsid, and an outer lipid envelope with viral glycoproteins on its surface. At least 8 proteins have been detected on the envelope of CMV (Britt et al., J. Virol. 62, 3309 (1988)) and others have been predicted to exist based on the DNA sequence of CMV (Chee et al., Nature 344, 774 (1990)). Murine monoclonal antibodies have been produced against two especially significant CMV glycoproteins: gB, also called p130/55 or gp55-116, and gH, also called p86 (Rasmussen et al., Virology 163, 308 (1988) and Britt et al., op. cit., both of which are incorporated herein by reference) and shown to neutralize infectivity of the virus. Three other neutralizing antibodies to gH are designated CMV5, CMV109 and CMV115. Human monoclonal antibodies to CMV have also been produced (Ehrlich et al., Hybridoma 6, 151 (1987)).

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different myeloid antigens. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include CD13, CD14, CD15, CD16 and CD34 (see, Leukocyte Typing III, op. cit., pp. 576-732).

In animal models, murine monclonal antibodies have been shown effective in treating infections caused by various viruses, including members of the herpesvirus family (see, e.g., Metcalf et al., Intervirol. 29, 39 (1988)). Hence, such antibodies may be useful in treatment of CMV infections.

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include cytosine arabinoside and daunorubicin, but numerous additional agents (e.g., 6-thioguanine) well-known to those skilled in the art for leukemia treatment may also be utilized (see, Hoffbrund & Pettit., op. cit.).

Unfortunately, the use of non-human monoclonal antibodies such as CMV5 and CMV115 have certain drawbacks in human treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, do not fix human complement well, and lack other important immunoglobulin functional characteristics when used in humans.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill leukemia cells. Immunotoxins are characterized by two components and are particularly useful 40 for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a 45

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of human immunoglobulins reactive with CMV antigens, as with many antigens, is difficult using typical human monoclonal antibody production techniques. Moreover, the human antibodies produced may lack certain desirable properties, such as high binding affinity and the ability to neutralize all clinical CMV strains. Similarly, utilizing recombinant DNA technology to produce so-called "humanized" or "reshaped" antibodies (see, e.g., Riechmann et al., Nature 332, 323 (1988) and EPO Publication No. 0239400, which are incorporated herein by reference), provides uncertain results, in part due to unpredictable binding affinities.

Humanized antibodies of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of CD33 antigens, for isolating specific myeloid cells, or the like.

CD33 epitope.

Thus, there is a need for improved forms of humanized immunoglobulins specific for CMV antigen that are substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic formulation and other uses. The present invention fulfills these and other needs.

It will be understood that although examples pertain to the M195 antibody, producing humanized antibodies with high binding affinity for the CD33 antigen is also contemplated using CDR's from L4B3, L1B2, MY9 or other monoclonal antibodies that bind to an epitope of CD33.

Anti-CMV Antibodies

Cytomegalovirus is a major pathogen of immunocompromised individuals, especially bone marrow transplant recipients, organ transplant recipients, and AIDS patients (see, generally, Fields et al., Eds., Virology, 2nd ed., Raven Press, New York pp. 1981–2010 (1990), which is incorporated herein by reference). Approximately 15% of bone marrow transplant patients develop CMV pneumonia, with an 85% 65 mortality rate (Meyers, Rev. Inf. Dis. 11 (suppl. 7), \$1691 (1989)). About 10% of AIDS patients develop severe CMV

The present invention provides novel compositions useful, for example, in the treatment of CMV-mediated human disorders, the compositions containing humanized immunoglobulins specifically capable of blocking the binding of CMV to its receptors and/or capable of binding to CMV antigens. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to CMV at affinity levels stronger than about 10? M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse $\,^{15}$ monoclonal antibody to CMV.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized alone in substantially pure form, or together with a chemotherapeutic agent such a acyclovir or ganciclovir active against CMV-infected cells, or complexed with a cytotoxic agent. All of these compounds will be particularly useful in treating CMV-mediated disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with CMV and CMVinfected cells are provided. These immunoglobulins, which 40 have binding affinities to CMV specific antigens of at least about 107 M⁻¹, and preferably 108 M⁻¹ to 10¹⁰ M⁻¹ or stronger, are capable of, e.g., blocking CMV infection of cells. The humanized immunoglobulins will have a human framework and will have one or more complementarity 45 determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with a CMV antigen. In a preferred embodiment, one or more of the CDR's will come from the CMV5, or CMV109 or CMV115 antibodies. The immunoglobulins of the present 50 invention, which can be produced economically in large quantities, find use, for example, in the treatment of CMVmediated disorders in human patients by a variety of tech-

In one aspect, the present invention is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of a CMV antigen, such as monoclonal antibodies CMV5 or CMVI15. The DNA segments encoding these regions will typically be joined to DNA segments 60 encoding appropriate human framework regions. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody CMV5 are included in FIG. 39A and FIG. 39B. Due to codon degeneracy and non-critical aminosaid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below.

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immortalized B-cells (see, Kabat op. cit. and WP87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to CMV and produced in any convenient mammalian source, including, mice, rats, rabbits, or other vertebrate capable of producing antibodies by well known methods. Suitable source cells for the DNA sequences and host cells for immunoglobulin expression and secretion can be obtained from a number of sources, such as the American Type Culture Collection (Catalogue of Cell Lines and Hybridomas, Fifth edition (1985) Rockville, Md. U.S.A., which is incorporated herein by reference).

The antibodies of the present invention will typically find use individually in treating CMV-related disorders. For example, typical disease states suitable for treatment include CMV pneumonia, neonatal CMV infection, CMV mononucleosis and CMV-related chorioretinitis and gastroenteritis

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with different CMV antigens. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include the gB and gH proteins.

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include acyclovir or ganciclovir, but numerous additional agents well-known to those skilled in the art for CMV treatment may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in immunotoxins to kill CMV-infected cells, Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for delivering the toxic agent to a particular cell type, such as cells expressing a CMV epitope. The two components are commonly chemically bonded together by any of a variety of well-known chemical procedures. For example, when the cytotoxic agent is a protein and the second component is an intact immunoglobulin, the linkage may be by way of heterobifunctional cross-linkers, e.g., SPDP, carbodiimide, glutaraldehyde, or the like. Production of various immunotoxins is well-known with the art, and can be found, for example in "Monoclonal Antibody-Toxin Conjugates: Aiming the Magic Bullet," Thorpe et al., Monoclonal Antibodies in Clinical Medicine, Academic Press, pp. 168-190 (1982), which is incorporated herein by reference. The components may also be linked genetically (see Chaudhary et al., Nature 339, 394 (1989)).

In prophylactic applications, compositions containing the present immunoglobulins or a cocktail thereof are administered to a patient not already in a disease state to enhance the patient's resistance. Such an amount is defined to be a "prophylactically effective dose." In this use, the precise amounts again depend upon the patient's state of health and general level of immunity, but generally range from 1 to 50 mg per dose. A preferred prophylactic use is for the prevention of CMV infection in immunocompromised patients, such as organ or bone marrow transplant recipients.

Humanized antibodies of the piesent invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of CMV antigens, for isolating specific CMV-infected cells, or the like.

In particular, the same method may be used to produce a 5 humanized CMV109, CMV115 or other anti-CMV antibody as used to produce humanized CMV5 herein.

Anti-y-IFN Antibodies

In mammals, the immune response is mediated by several types of cells that interact specifically with foreign material, i.e., antigens. One of these cell types, B cells, is responsible for the production of antibodies. Another cell type, T cells, include a wide variety of cellular subsets that destroy virally infected cells or control the in vivo function of both B cells and other hematopoictic cells, including T cells. A third cell type, macrophages, process and present antigens in conjunction with major histocompatibility complex (MHC) proteins to T cells. Communication between these cell types is mediated in a complex manner by lymphokines, such as interleukins 1-6 and γ-IFN (see, generally, Paul, W. E., ed., Fundamental Immunology, 2nd ed., Raven Press, New York (1989), which is incorporated herein by reference.)

One important lymphokine is γ -IFN, which is secreted by some T cells. In addition to its anti-viral activity, γ -IFN stimulates natual killer (NK) cells, activates macrophages, and stimulates the expression of MHC molecules on the surface of cells (Paul, op. cit., pp. 622-624). Hence γ -IFN generally serves to enhance many aspects of immune function, and is a logical candidate for a therapeutic drug in cases where such enhancement is desired, e.g., in treating cancer. Conversely, in disease states where the immune system is over-active, e.g., autoimmune diseases and organ transplant rejection. antagonists of γ -IFN may be used to treat the disease by neutralizing the stimulatory effects of γ -IFN.

One class of effective antagonists of y-IFN are monoclonal antibodies that bind to and neutralize it (see, e.g., Van der Meide et al., J. Gen. Virol, 67, 1059 (1986)). In in vitro 40 and in vivo mouse models of transplants, anti-y-IFN antibodies have been shown to delay or prevent rejection (Landolfo et al., Science 229, 176 (1985) and Rosenberg et al., J. fmmunol. 144, 4648 (1990), both of which are incorporated herein by reference). Treatment of mice prone 45 to develop a syndrome like systemic lupus erythematosus (SLE) with a monoclonal antibody to y-IFN significantly delayed onset of the disease (Jacob et al., J. Exp. Med. 166, 798 (1987)). Under some conditions, an anti-γ-IFN antibody alleviated adjuvant arthriti's in rats (Jacob et al., J. Immunol. 142, 1500 (1989)), suggesting that anti-γ-IFN may be effective against some cases of rheumatoid arthritis in human patients. Multiple sclerosis (MS) in patients is made worse by treatment with γ-IFN (Panitch et al., Neurology 36 (suppl. I), 285 (1986)), so an anti-y-IFN antibody may 55 alleviate MS. Thus, an anti-y-IFN antibody may be effective in treating these and other autoimmune diseases.

For treatment of human patients, a murine monoclonal that binds to and neutralizes human γ -IFN (see, e.g., Yamamoto ct al., Microbiol. Immunol. 32, 339 (1988)) may be 60 used. Another murine monoclonal antibody designated AF2 that neutralizes human γ -IFN, and inhibits binding of γ -IFN to its cellular receptor, is disclosed herein. Unfortunately, the use of non-human monoclonal antibodies such as AF2 have certain drawbacks in human treatment, particularly in 65 repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, have a relatively short

circulating half-life in humans, and lack other important immunoglobulin functional characteristics when used in humans.

The present invention provides novel compositions useful, for example, in the treatment of human autoimmune disorders, the compositions containing humanized immunoglobulins specifically capable of binding to γ-IFN. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to γ-IFN at affinity levels stronger than about 10⁷ M⁻¹. These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to γ-IFN.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be unilized alone in substantially pure form, or together with a chemotherapeutic agent such as a non-steroidal anti-inflammatory drug, a corticosteroid, or an immunosuppressant. All of these compounds will be particularly useful in treating autoimmune disorders. The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

In accordance with the present invention, humanized immunoglobulins specifically reactive with γ -IFN epitopes are provided. These immunoglobulins, which have binding affinities to γ -IFN of at least about $10^7~M^{-1}$, and preferably $10^8~M^{-1}$ to $10^{10}~M^{-1}$ or stronger, are capable of, e.g., neutralizing human γ -IFN. The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with γ -IFN. In a preferred embodiment, one or more of the CDR's will come from the AF2 antibody. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of autoimmune disorders in human patients by a variety of techniques.

The antibodies of the present invention will typically find use individually in treating autoimmune conditions. For example, typical disease states suitable for treatment include graft versus host disease and transplant rejection in patients undergoing an organ transplant, such as heart, lungs, kidneys, liver, etc. Other diseases include autoimmune diseases, such as Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, and myasthenia gravis.

Any humanized immunoglobulins of the present invention may also be used in combination with other antibodies, particularly humanized antibodies reactive with other lym-

phokines or lymphokine receptors. For example, suitable antigens to which a cocktail of humanized immunoglobulins may react include interleukins 1 through 10 and the p55 and p75 chains of the IL-2 receptor (see, Waldmann, Annu. Rev. Biochem. 58, 875 (1989) and Queen et al., Proc. Natl. Acad. 5 Sci. USA 86, 10029 (1989), both of which are incorporated herein by reference). Other antigens include those on cells responsible for the disease, e.g., the so-called "Clusters of Differentiation" (Leucocyte Typing III, ed. by A. J. McMichael, Oxford University Press (1987), which is incorporated herein by reference).

The antibodies can also be used as separately administered compositions given in conjunction with chemotherapeutic agents. Typically, the agents may include non-steroidal anti-inflammatory agents (e.g., aspirin, ibuprofen), 15 steroids (e.g., prednisone) and immunosuppressants (e.g., cyclosporin A, cytoxan), but numerous additional agems well-known to those skilled in the art may also be utilized.

A preferred pharmaceutical composition of the present invention comprises the use of the subject immunoglobulins in inununotoxins, e.g., to kill γ -IFN -secreting cells. Immunotoxins are characterized by two components and are particularly useful for killing selected cells in vitro or in vivo. One component is a cytotoxic agent which is usually fatal to a cell when attached or absorbed. The second component, known as the "delivery vehicle," provides a means for deliveting the toxic agent to a particular cell type, such as cells expressing a γ -IFN epitope.

Humanized antibodics of the present invention can further find a wide variety of utilities in vitro. By way of example, the antibodies can be utilized for detection of γ -IFN antigens, or the like.

The following examples are offered by way of illustration, not by limitation.

EXPERIMENTAL

EXAMPLE 1

Humanized anti-Tac antibody

Design of genes for humanized anti-Tac light and heavy chains

The sequence of the human antibody Eu (Sequences of Proteins of Immunological Interest, E. Kabat et al., U.S. Dept. of Health and Human Services, 1983) was used to provide the framework of the humanized antibody, because the amino acid sequence of the heavy chain variable region of anti-Tac is more homologous to the heavy chain of this antibody than to any other complete heavy chain variable region sequence in the National Biomedical Foundation Protein Identification Resource.

To select the sequence of the humanized heavy chain, the anti-Tac heavy chain sequence (FIG. 1B., upper lines; see, commonly assigned U.S. Ser. No. 07/223.037 filed Sep. 28, 1988, and 07/181,862 filed Apr. 15, 1988, both of which are now abandoned and which are incorporated herein by reference) was aligned with the sequence of the Eu heavy chain (FIG. 1B., lower lines). At each position, the Eu amino acid was selected for the humanized sequence, unless that position fell in any one of four categories defined above, in which case the anti-Tac amino acid was selected:

 The position fell within a complementarity determines ing region (CDR), as defined by Kabat, et al., op. cit. (amino acids 31-35, 50-66, 99-106); 38

- (2) The Eu amino acid was rare for human heavy chains at that position, whereas the anti-Tac amino acid was common for human heavy chains at that position (amino acids 27, 93, 95, 98, 107-109, 111);
- (3) The position was immediately adjacent to a CDR in the amino acid sequence of the anti-Tac heavy chain (amino acids 30 and 67); or
- (4) 3-dimensional modeling of the anti-Tac antibody suggested that the amino acid was physically close to the antigen bindring region (amino acids 48 and 68). Amino acid #27 is listed in category (2) because the acceptor Eu amino acid Gly is rare, and the donor anti-Tac amino acid Tyr is chemically similar to the amino acid Phc, which is common, but the substitution was actually made because #27 also fell in category (4). Although some amino acids fell in more than one of these categories, they are only listed in one. The amino acids in the humanized heavy and light chains are numbered according to the lower lines of FIG. 1A and FIG. 1B.

To select the sequence of the humanized light chain, the anti-Tac light chain sequence was aligned with the sequence of the Eu light chain (FIG. 1A, lower lines). The Eu amino acid was selected at each position for the humanized sequence, unless the position again fell into one of the categories (1)-(4):

- (1) CDR's (amino acids 24-34, 50-56, 89-97);
- Anti-Tac amino acid more typical than Eu (amino acids 48 and 63);
- (3) Adjacent to CDR's (no amino acids; Eu and anti-Tac were already the same at all these positions); or
- (4) Possible 3-dimensional proximity to binding region (amino acid 60).
- The actual nucleotide sequence of the heavy and light chain genes were selected as follows:
- The nucleotide sequences code for the amino acid sequences chosen as described above;
- (2) 5' of these coding sequences, the nucleotide sequences code for a leader (signal) sequence, namely the leader of the light chain of the antibody MOPC 63 and the leader of the heavy chain of the antibody PCH 108A (Kabat et al., op. cit.). These leader sequences were chosen as typical of antibodies;
- (3) 3' of the coding sequences, the nucleotide sequences are the sequences that follow the mouse light chain J5 segment and the mouse heavy chain J2 segment, which are part of the anti-Tac sequences. These sequences are included because they contain splice donor signals; and
- (4) At each end of the sequence is an Xba I site to allow cutting at the Xba I sites and cloning into the Xba I site of a vector.

Construction of humanized light and heavy chain gencs

To synthesize the heavy chain, four oligonucleotides were synthesized using an Applied Biosystems 380B DNA synthesizer. Two of the oligonucleotides are part of each strand of the heavy chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing.

Together, the oligonucleotides cover the entire humanized heavy chain variable region with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

Each oligonucleotide was phosphorylated using ATP and T4 polynucleotide kinase by standard procedures (see, Maniatis, op. cit.). To anneal the phosphorylated oligonucleotides, they were suspended together in 40 ul of TA (33 mM)

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Tris acetate, pH 7.9, 66 mM potassium acetate, 10 mM magnesium acetate) at a concentration of about 3.75 uM each, heated to 95° C. for 4 min. and cooled slowly to 4° C. To synthesize the complete gene from the oligonucleotides by synthesizing the opposite strand of each oligonucleotide, the following components were added in a final volume of 100 ul:

annealed oligonucleotides 0.16 mM each deoxyribonucleotide 0.5mM ATP 0.5 mM DTT 100 ug/ml BSA 3.5 ug/ml T4 g43 protein (DNA polymerase) T4 g44/62 protein (polymerasc 25 ug/ml accessery protein) 25 ug/ml 45 protein (polymerase accessory protein)

The mixture was incubated at 37° C. for 30 min. Then 10 u of T4 DNA ligase was added and incubation at 37° C. resumed for 30 min. The polymerase and ligase were inactivated by incubation of the reaction at 70° C. for 15 min. To digest the gene with Xba I, to the reaction was added 50 ul of 2×TA containing BSA at 200 ug/ml and DTT at 1 mM, 43 ul of water, and 50 u of Xba I in 5 ul. The reaction was incubated for 3 hr at 37° C., and run on a gel. The 431 bp Xba I fragment was purified from a gel and cloned into the Xba I site of the plasmid pUC19 by standard methods.

Four plasmid isolates were purified and sequenced using the dideoxy method. One of these had the correct sequence.

To synthesize the light chain, four oligonucleotides JFD1, 30 JFD2, JFD3, JFD4 were synthesized. Two of the oligonucleotides are part of each strand of the light chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing. Together, the oligonucleotides cover the entire humanized light chain variable region with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

The light chain gene was synthesized from these oligonucleotides in two parts. 0.5 ug each of JFD1 and JFD2 were combined in 20 ul sequence buffer (40 mM Tris-HCl, pH 7.5, 20 mM magnesium chloride, 50 mM sodium chloride), heated at 70° C. for 3 min and allowed to cool slowly to 23° C. in order for the oligonucleotides to anneal. JFD3 and JFD4 were treated in the same way. Each reaction was made 10 mM in DTT and 0.5 mM in each deoxyribonucleotide and 6.5 u of sequenase (US Biochemicals) was added, in a final volume of 24 ul, and incubated for 1 hr at 37° C. to synthesize the opposite strands of the oligonucleotides. Xba I and Hind III were added to each reaction to digest the DNA (there is a Hind III site in the region where JFD2 and JFD3 50 overlap and therefore in each of the synthesized DNAs). The reactions were run on polyacrylamide gels, and the Xba I -Hind III fragments were purified and cloned into pUC18 by standard methods. Several plasmid isolates for each fragment were sequenced by the dideoxy method, and correct 55

Construction of plasmids to express humanized light and heavy chains

The heavy chain Xba I fragment was isolated from the pUC19 plasmid in which it had been inserted and then 60 inserted into the Xba I site of the vector pVγ1 (see, commonly assigned U.S. Ser. No. 07/223,037 filed Sep. 28, 1988, now abandoned, which is incorporated herein by reference) in the correct orientation by standard methods, to produce the plasmid pHuGTAC1. This plasmid will express 65 high levels of a complete heavy chain when transfected into an appropriate host cell.

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The two light chain Xba I - Hind III fragments were isolated from the pUCl 8 plasmids in which they had been inserted. The vector plasmid pVkl (see, commonly assigned U.S. Ser. No. 07/223,037 filed Sep. 28, 1988, now abandoned, which is incorporated herein by reference) was cut with Xba I, dephosphorylated and ligated with the two fragments by standard methods. The desired reaction product has the circular form: vector - Xba I - fragment 1 - Hind III fragment 2 - Xba I - vector. Several plasmid isolates were analyzed by restriction mapping and sequencing, and one with this form chosen. This plasmid, pHuLTAC, therefore contains the complete humanized light chain and will express high levels of the light chain when transfected into an appropriate host cell.

Synthesis and affinity of humanized antibody

The plasmids pHuGTACl and pHuLTAC were transfected into mouse Sp2/0 cells, and cells that integrated the plasmids were selected on the basis of resistance to mycophenolic acid and/or hygromycin B conferred by the gpt and hyg genes on the plasmids by standard methods. To verify that these cells secreted antibody that binds to the 11-2 receptor, supernatant from the cells was incubated with HUT-102 cells that are known to express the II-2 receptor. After washing, the cells were incubated with fluoresceinconjugated goat anti-human antibody, washed, and analyzed for fluorescence on a FACSCAN cytofluorometer. The results (FIG. 7A), clearly show that the humanized antibody binds to these cells, but not to Jurkat T-cells that do not express the IL-2 receptor (FIG. 7D). As controls, the original mouse anti-Tac antibody was also used to stain these cells, giving similar results.

For the next experiments, cells producing the humanized antibody were injected into mice, and the resultant ascites collected. Humanized antibody was purified to substantial homogeneity from the ascites by passage through an affinity column of goat anti-human immunoglobulin antibody, prepared on an Affigel-10 support (Bio-Rad Laboratories, Inc., Richmond, Calif.) according to standard techniques. To determine the affinity of the humanized antibody relative to the original anti-Tac antibody, a competitive binding experiment was performed. About 5×10⁵ HUT-102 cells were incubated with known quantities (10-40 ng) of the anti-Tac antibody and the humanized anti-Tac antibody for 10 min at 4° C. Then 100 ng of biotinylated anti-Tac was added to the cells and incubated for 30 min at 4° C. This quantity of anti-Tac had previously been determined to be sufficient to saturate the binding sites on the cells, but not to be in large

Then the cells were washed twice with 2 ml of phosphate buffered saline (PBS) containing 0.1% sodium azide. The cells were then incubated for 30 min at 4° C. with 250 ng of phycoerythrin-conjugated avidin, which bound to the biotinylated anti-Tac already bound to the cells. The cells were washed again as above, fixed in PBS containing 1% paraformaldehyde, and analyzed for fluorescence on a FAC-SCAN cytofluorometer.

Use of increasing amounts (10-40 ng) of the anti-Tac antibody as competitor in the first step decreased the amount of biotinylated anti-Tac that could bind to the cells in the second step, and therefore the amount of phycocrythrin-conjugated avidin that bound in the last step, thus decreasing fluorescence (FIG. 8A). Equivalent amounts (20 ng) of anti-Tac, and humanized anti-Tac used as competitor decreased the fluorescence to approximately the same degree (FIG. 8B). This shows that these antibodies have approximately the same affinity, because if one had greater affinity, it would have more effectively competed with the biotinylated anti-Tac, thus decreasing fluorescence more.

EXAMPLE 2

A second humanized anti-Tac antibody

Higher level expression of the humanized anti-Tae anti-

Three new plasmid vectors were prepared for expression of the humanized antibodies. The plasmid pVgl (FIG. 9A) contains a human cytomegalovirus IE1 promoter and 10 enhancer (Boshart et al., Cell 41, 521 (1985), which is incorporated herein by reference), the human genomic Cyl segment including part of the preceding intron, and the hygromycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selec- 15 to bind to the IL-2 receptor was assessed by fluorescence tion. The plasmid pVk (FIG. 9B) is similar to pVgl but contains the human genomic Cx segment and the gpt gene. The plasmid pVgl-dhfr was constructed similarly to pVgl but contains a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1984), which 20 is incorporated herein by reference) in place of the hygromycin gene.

Xba I fragments containing the humanized anti-Tac light chain and heavy chain variable regions were excised respectively from the plasmids pHuLTAC and the pHuGTAC1 and 25 cloned into the Xba I sites of the plasmid vectors pVk and pVgl. To express the humanized anti-Tac antibody, the light chain encoding plasmid was introduced by electroporation into SP2/0 mouse myeloma cells followed by selection for gpt expression. Transfected cells expressing light chain were 10 then transfected with the plasmi'd encoding the heavy chain followed by selection for hygromycin B resistance. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody. Humanized antibody was purified from culture 35 supernatant of transfected cells by protein A sepharose chromatography.

Construction of the second humanized anti-Tac antibody

To determine whether it was actually necessary to use the mouse anti-Tac amino acids in categories (2)-(4) in the 40 humanized anti-Tac antibody to retain binding affinity, a second humanized anti-Tac antibody was constructed. In the second antibody, only mouse anti-Tac amino acids in Category (1), i.e., in the CDR's themselves, were used, with all other amino acids coming from the human Eu framework. 45 For purposes of this discussion, the original humanized anti-Tac antibody will be called the "PDL humanized antibody," and the second humanized antibody will be called the "CDR-only humanized antibody." The amino acid sequences of the PDL and CDR-only humanized antibody 50 (variable regions) are compared in FIG. 10A and FIG. 10B.

The CDR-only humanized anti-Tac heavy and light chain variable (V) region gene segments were constructed in essentially the same manner as the light chain of the PDL humanized anti-Tac immunoglobulin, as described above. 55 Specifically, each V region gene segment was synthesized in two halves. For each half, two overlapping, opposite-strand oligonucleotides, approximately 110 to 130 bases in length (FIG. 11A and FIG. 11B), were annealed and extended with sequenase (U.S. Biochemicals). The resulting double strand 60 fragments were digested with either Xba I and Hind III (light chain) or Xba I and Sal I (heavy chain) and inserted into plasmid pUC19. Clones with the correct sequence were identified by DNA sequencing. Complete heavy and light chain genes were generated by inserting the V region halves 65 into the Xba I sites of pVg I and pVk respectively by three-fragment ligation.

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The CDR-only humanized antibody was expressed in the same manner as the PDL humanized antibody, by transfecting first the light chain containing plasmid and then the heavy chain containing plasmid into SP2/0 cells. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody, which was purified by protein A sepharose chromatography. Antibody concentration was determined by ELISA using purified PDL humanized antibody as a standard. That the purified CDR-only humanized antibody is assembled into H2L2 telramers as expected was shown by analysis using reducing and non-reducing polyacrylamide gel electrophoresis.

The ability of the CDR-only humanized immunoglobulin staining. Approximately 3.4×10⁵ HUT-102 cells, which are known to highly express the IL-2 receptor on their surface, were incubated with 200 ng of either the PDL or CDR-only humanized antibody, washed, and then incubated with fluorescein-conjugated goat anti-human IgG antisera. Cell fluorescence was measured by flow cytometry with a FACScan (Becton Dickinson). As shown in FIG. 12, the PDL humanized antibody strongly stained the cells. However, staining by the CDR-only antibody was indistinguishable from staining by the negative control antibody humanized Fd79, which binds the gB glycoprotein of herpes simplex virus and not HUT-102 cells. Hence, by this assay, the CDR-only humanized antibody does not detectably bind the IL-2 receptor.

Binding of the PDL and CDR-only humanized anti-Tac antibodies to the IL-2 receptor were also compared in a competitive binding assay. Approximately 4x105 HUT-102 cells were incubated with 1.5 ng of radioiodinated mouse anti-Tac antibody (7×106 cpm/ug) and varying amounts of each humanized antibody (4 to 512 ng) in 200 ul total volume of binding buffer (RPMI 1040 medium, 10% fetal calf senim, 10 ug/ml murine IgG2a, 0.1% sodium azide). After incubation for 2 hours at 0° C. 800 ul of binding buffer was added, cells were collected by centrifugation and radioactivity was measured. The relative binding by the two humanized antibodies and by mouse anti-Tac is shown in a plot of bound/free labelled antibody versus competitor concentration (FIG. 13). The PDL humanized anti-Tac antibody affinity for IL-2 receptor is essentially equal to that of the mouse anti-Tac antibody, because it competes about equally well. But competition by the CDR-only humanized anti-Tac antibody to IL-2 receptor was undetectable at the antibody concentrations used, indicating a binding affinity reduction of at least 100-fold as compared to the PDL humanized anti-Tac antibody. Because the sequences of the PDL and CDR humanized anti-Tac antibodies differ only at positions where mouse framework residues in categories (2)-(4) were used in the PDL molecule, we conclude that at least one of these mouse framework residues are essential for high affinity binding.

EXAMPLE 3

Construction of 5 other humanized antibodies

Cloning of heavy and light chain cDNAs

Five other humanized antibodies were designed and produced using the principles and categories disclosed herein. The antibodies arc Fd79 and Fd138-80 which respectively bind to the gB and gD glycoproteins of herpes simplex virus (Metcalf et al., Intervirology 29, 39 (1988)), M195 (Tanimoto ct al., Leukemia 3, 339 (1989)) which binds to the CD33 antigen, mik-\$1 (Tusdo et al., Proc. Natl. Acad. Sci.

USA 86, 982 (1989)) which binds to the p75 chain of the IL-2 receptor, and CMV5 which binds to the gH glycoprotein of cytomegalovirus.

cDNAs for the heavy chain and light chain variable domain genes of each antibody were cloned using anchored polymerase chain reactions (Loh et al., Science 243,219 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (Scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For each antibody, at least two heavy chain and two kappa clones were sequenced and found to have the same sequence. The deduced amino acid sequences of the mature light and heavy chain variable regions are shown in FIGS. 2A-6B, upper lines. Design of humanized antibodies

In order to retain high binding affinity of the humanized antibodies, the principles and categories described above were utilized when designing the antibodies. Based on high sequence homology, human antibodies were selected to provide both the acceptor light and heavy chain human frameworks for the mouse antibodies, as follows: human Pom for Fd79, human Eu for Fd138-80, human Eu for M195, human Lay for mik-\(\beta_1 \), and human Wol for CMV5.

The computer programs ABMOD and ENCAD (Levitt, J. Mol. Biol., 168, 595 (1983) and Zilber et al., Biochemistry 29, 10032 (1990), both of which are incorporated herein by reference) was used to construct a model of the variable region of each mouse antibody. The model was used to determine the amino acids in each framework that were close enough to the CDR's to potentially interact with them (category 4 above). For each antibody, the positions found to fall in the categories (1)–(5) defined above are given in Table 1, numbered as in FIGS. 2A–6B.

TABLE

Category	Light Chain	Heavy Chain
	Fd79 Antibod	у
1	24-38, 54-50, 93-100	31-35, 50-66, 99-111
	9, 45, 46, 83	82, 112
2	53	112
4	53	97
5	81	
	Fd138-80 Antib	ody
1	24-34, 50-56, 89-97	31-35, 50-66, 99-110
2	48, 63	93, 98, 111, 112,
		113, 115
3		30, 67, 98, 111
4	36, 48, 87	27, 30, 37, 48, 67,
		68, 98
	M195 Antiboo	
1	24-38, 54-60, 93-101	31-35, 50-66, 95-105
2	10, 52, 67, 110	93, 95, 98, 106, 107
		108, 110
3	_	30, 67, 98, 106
4	40. 52, 74	27, 30, 48, 68, 98
	mik-β1 Λπτίδο	dy
1	24-33, 49-55, 88-96	31-35, 50-65, 98-108
2	13	84, 89, 90
2 3 4	_	30, 49
4	70	29, 30, 72, 73
5	41	1
	CMV5 Antibo	dy
1	24-34, 50-56, 89-97	31-35, 50-66, 99-108
2		69. 80

TABLE 1-continued

Category	Light Chain	Heavy Chain
3	49	30
4	49	24, 27, 28, 30, 9
5		5

In designing each humanized antibody, at each position the amino acid was selected to be the same as in the human acceptor sequence, unless the position fell in categories (1)–(4), in which case the amino acid from the mouse donor sequence was used, or in category (5), in which case an amino acid typical for human sequences at that position was used.

For the construction of genes for the humanized antibodies, nucleou'de sequences were selected that encode the protein sequences of the humanized heavy and light chains, including signal peptides typically from the mouse antibody chains, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included splice donor signals typical for immunoglobulin genes and an Xbal site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal. The oligonucleotides were synthesized on an Applied Biosys-30 tems 380B DNA synthesizer. Each oligo was about 110-140 base long with a 15-20 base overlap. Double stranded DNA fragments were synthesized with Klenow or Taq polymerase or sequenase from each pair of oligonucleotides, digested with restriction enzymes, ligated to pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of pVg1 (heavy chains of Fd79 and Fd138-80) or pVg1-dhfr (heavy chains of M195, mik-\$1, CMV5) or pVk (all light chains) expression vectors in the appropriate orientations to produce - 40 the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.).

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibodies were eluted with 0.2M Glycine-HCl, pH 3.0 and neutralized with 1M Tris pH 8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

55 Properties of the humanized antibodies

The binding of the humanized antibodies to cell types expressing the corresponding antigens was tested: HSV-infected cells for Fd79 and Fd138-80, U937 cells for M195, YTJB cells for mik-β1 and CMV-infected cells for CMV5.

60 By fluorocytometry, the humanized antibodies bind approximately as well as the original mouse antibodies and the corresponding chimeric antibodies. Moreover, the humanized antibodies compete approximately as well as the corresponding mouse antibodies against the radiolabeled mouse antibodies for binding to the cells, so the humanized antibodies have approximately the same binding affinity as the mouse antibodies, typically within about 2 fold or better, see,

e.g., Table 2.

TABLE 2

Binding affinities of murine and humanized antibodies.		
Mouse K _a (M ⁻¹)	Humanized $K_a (M^{-1})$	
Fd79 (anti-gB) Fd138-80 (anti-gD)	1.1×10^8 5.2×10^7	5.3 × 10 ⁷ 4.8 × 10 ⁷

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other antibodies. In comparison to other monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement

EXAMPLE 4

Design of genes for anti-Tac human-like light and heavy chains

The sequence of the human antibody Eu (Sequences of Proteins of Immunological Interest, Kabat, E., et al., U.S. Dept. of Health and Human Services, 1983) was used to provide the framework of the humanized antibody, because the amino acid sequence of the heavy chain of anti-Tac is more homologous to the heavy chain of this antibody than to any other heavy chain sequence in the National Biomedical Foundation Protein Identification Resource.

To select the sequence of the humanized heavy chain, the anti-Tac heavy chain sequence was aligned with the sequence of the Eu heavy chain (FIG. 15). At each position, the Eu amino acid was selected for the humanized sequence, unless that position fell in any one of the following categories, in which case the anti-Tac amino acid was selected.

- (1) The position fell within a complementarity determining region (CDR), as defined by Kabat, et al., op. cit. (amino acids 31-35, 50-66, 99-106);
- (2) The Eu amino acid was unusual for human heavy chains at that position, whereas the anti-Tac amino acid 45 was typical for human heavy chains at that position (amino acids 27, 93, 95, 98, 107-109, 111);
- (3) The position was immediately adjacent to a CDR in the amino acid sequence of the anti-Tac heavy chain (amino acids 30 and 67).
- (4) 3-dimensional modeling of the anti-Tac antibody suggested that the amino acid was physically close to the antigen binding region (amino acids 48 and 68). Some amino acids fell in more than one of these categories but are only listed in one.

To select the sequence of the humanized light chain, the anti-Tac light chain sequence was aligned with the sequence of the Eu light chain (FIG. 16). The Eu amino acid was selected at each position, unless the position again fell into one of the categories (1)–(4), (with light chain replacing heavy chain in the category definitions):

- (I) CDRs (amino acids 24-34, 50-56, 89-97).
- (2) Anti-Tac amino acid more typical than Eu (amino acids 48 and 63).
- (3) Adjacent to CDRs (no amino acids; Eu and anti-Tac were already the same at all these positions).

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(4) Possible 3-dimensional proximity to binding region (amino acid 60).

The actual nucleotide sequence of the heavy (FIG. 17) and light chain (FIG. 18) genes were selected as follows:

- the nucleotide sequences code for the amino acid sequences chosen as described above.
- (2) 5' of these coding sequences, the nucleot de sequences code for a leader (signal) sequence, namely the leader of the light chain of the antibody MOPC 63 and the leader of the heavy chain of the antibody PCH 108A (Kabat et al., op. cit.). These leader sequences were chosen as typical of antibodies.
- (3) 3' of the coding sequences, the nucleotide sequences are the sequences that follow the mouse light chain J5 segment and the mouse heavy chain J2 segment, which are part of the anti-Tac sequences. These sequences are included because they contain splice donor signals.
- (4) At each end of the sequence is an Xba I site to allow cutting at the Xba I sites and cloning into the Xba I site of a vector.

Construction of humanized light and heavy chain genes

To synthesize the heavy chain, four oligonucleotides HES12, HES13, HES14, HES15 (FIG. 19A) were synthesized using an Applied Biosystems 380B DNA synthesizer. Two of the oligonucleotides are part of each strand of the heavy chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing (FIG. 19B). Together, the oligonucleotides cover the entire humanized heavy chain (FIG. 17) with a few extra nucleotides at each end to allow cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

Each oligonucleotide was phosphorylated using ATP and T4 polynucleotide kinase by standard procedures (see, Maniatis, op. cit.). To anneal the phosphorylated oligonucleotides, they were suspended together in 40 ul of TA (33 mM Tris acetate, pH 7.9, 66 mM potassium acetate, 10 mM magnesium acetate) at a concentration of about 3.75 uM each, heated to 95° C. for 4 min. and cooled slowly to 4° C. To synthesize the complete gene from the oligonucleotides by synthesizing the opposite strand of each oligonucleotide (FIG. 19B), the following components were added in a final volume of I00ul:

10 wl	annealed oligonucleotides
0.16 mM each	deoxyr:bonucleotide
0.5mM	ATP
0.5MM	DIT
100 ug/m!	BSA
3.5 ug/m1	T4 g43 protein (DNA polymerase)
25 ug/ml	T4 g44/62 protein (polymerase
	accessory protein)
25 ug/ml	45 protein (polymerase accessory protein)

The mixture was incubated at 37° C. for 30 min. Then 10 U of T4 DNA ligase was added and incubation at 37° C. resumed for 30 min. The polymerase and ligasc were inactivated by incubation of the reaction at 70° C. for 15 min. To digest the gene with Xba I, to the reaction was added 50 ul of 2× TA containing BSA at 200 ug/ml and DTT at 1 mM, 43 ul of water, and 50 U of Xba I in 5 ul. The reaction was incubated for 3 hr at 37° C., and run on a gel. The 431 bp Xba I fragment was purified from a gel and cloned into the Xba I site of the plasmid pUCI9 by standard methods.

Four plasmid isolates were purified and sequenced using the dideoxy method. One of these bad the correct sequence (FIG. 17).

To synthesize the light chain, four oligonucleot des JFDI, JFD2, JFD3, JFD4 (FIG. 20A) were synthesized. Two of the

oligonucleotides are part of each strand of the light chain, and each oligonucleotide overlaps the next one by about 20 nucleotides to allow annealing (FIG. 20B). Together, the oligonucleotides cover the entire humanized light chain (FIG. 18) with a few extra nucleotides at each end to allow 5 cutting at the Xba I sites. The oligonucleotides were purified from polyacrylamide gels.

The light chain gene was synthesized from these oligonucleotides in two parts. 0.5 ug each of JFD1 and JFD2 were combined in 20 ul sequenase buffer (40 mM Tris-HCl, pH 10 7.5, 20 mM magnesium chloride, 50 mM sodium chloride), heated at 70° C. for 3 min and allowed to cool slowly to 23° C. in order for the oligonucleotides to anneal. JFD3 and JFD4 were treated in the same way. Each reaction was made 10 mM in DTT and 0.5 mM in each deoxyribonucleotide and 15 6.5 U of sequenase (US Biochemicals) was added, in a final volume of 24 ul, and incubated for 1 hr at 37° C. to synthesize the opposite strands of the oligonucleotides. Xba I and Hind III were added to each reaction to digest the DNA (there is a Hind III site in the region where JFD2 and JFD3 20 overlap and therefore in each of the synthesized DNAs; FIG. 20B). The reactions were run on polyacrylamide gels, and the Xba I - Hind Ill fragments were purified and cloned into pUC18 by standard methods. Several plasmid isolates for each fragment were sequenced by the dideoxy method, and 25 correct ones chosen.

Construction of plasmids to express humanized light and heavy chains

The heavy chain Xba I fragment was isolated from the pUC19 plasmid in which it had been inserted and then 30 inserted into the Xba I site of the vector pVyl in the correct orientation by standard methods, to produce the plasmid pHuGTAC1 (FIG. 21). This plasmid will express high levels of a complete heavy chain when transfected into an appropriate host cell.

The two light chain Xba I - Hind III fragments were isolated from the pUC18 plasmids in which they had been inserted. The vector plasmid pVkl was cut with Xba I, dephosphorylated and ligated with the two fragments by standard methods. The desired reaction product has the 40 circular form: vector - Xba I - fragment 1 - Hind III - fragment 2 - Xba I - vector. Several plasmid isolates were analyzed by restriction mapping and sequencing, and one with this form chosen. This plasmid, pHuLTAC (FIG. 22), therefore contains the complete humanized light chain (FIG. 45 18) and will express high levels of the light chain when transfected into an appropriate host cell. Synthesis and affinity of humanized antibody

The plasmids pHuGTACl and pHuLTAC were transfected into mouse Sp2/0 cells, and cells that integrated the 50 plasmids were selected on the basis of resistance to mycophenolic acid and/or hygromycin B conferred by the gpt and hyg genes on the plasmids (FIGS. 21, 22) by standard methods. To verify that these cells secreted antibody that binds to the IL-2 receptor, supernatant from the cells was 55 incubated with HUT-102 cells that are known to express the IL-2 receptor. After washing, the cells were incubated with fluorescein-conjugated goat anti-human antibody, washed, and analyzed for fluorescence on a FACSCAN cytofluorometer. The results (FIG. 7A), clearly show that the humanized 60 antibody binds to these cells, but not to Jurkat T-cells that do not express the IL-2 receptor (FIG. 7D). As controls, the original mouse anti-Tac antibody was also used to stain these cells (FIG. 7B and FIG. 7C), giving similar results.

For further experiments, cells producing the humanized 65 antibody were injected into mice, and the resultant ascites collected. Humanized antibody was purified to substantial

homogeneity from the ascites by passage through an affinity column of goat anti-human immunoglobulin antibody, prepared on an Affigel-10 support (Bio-Rad Laboratories, Inc., Richmond, Calif.) according to standard techniques. To determine the affinity of the humanized antibody relative to the original ant-Tac antibody, a competitive binding experiment was performed. About 5x105 HUT-102 cells were incubated with known quantities (10-40 ng) of the anti-Tac antibody and the humanized anti-Tac antibody for 10 min at 4° C. Then 100 ng of biotinylated anti-Tac was added to the cells and incubated for 30 min at 4° C. This quantity of anti-Tac had previously been determined to be sufficient to saturate the binding sites on the cells, but not to be in large excess. Then the cells were washed twice with 2 ml of phosphate buffered saline (PBS) containing 0.1% sodium azide. The cells were then incubated for 30 min at 4° C. with 250 ng of phycocrythrin-con jugated avidin, which bound to the biotinylated anti-Tac already bound to the cells. The cells were washed again as above, fixed in PBS containing 1% paraformaldehyde, and analyzed for fluorescence on a FAC-SCAN cytofluorometer.

Use of increasing amounts (10-40 ng) of the anti-Tac antibody as competitor in the first step decreased the amount of biotinylated anti-Tac that could bind to the cells in the second step, and therefore the amount of phycoerythrin-conjugated avidin that bound in the last step, thus decreasing fluorescence (FIG. 8A). Equivalent amounts (20 ng) of anti-Tac, and humanized anti-Tac used as competitor decreased the fluorescence to approximately the same degree (FIG. 8B). This shows that these antibodies have approximately the same affinity, because if one had greater affinity, it would have more effectively competed with the biotinylated anti-Tac, thus decreasing fluorescence more. Biological properties of the humanized antibody

For optimal use in treatment of human disease, the humanized antibody should be able to destroy T-cells in the body that express the IL-2 receptor. One mechanism by which antibodies may destroy target cells is antibody-dependent cell-mediated cytotoxicity, abbreviated ADCC (Fundamental Immunology, Paul, W., Ed., Raven Press, New York (1984), at pg. 681), in which the antibody forms a biidge between the target cell and an effector cell such as a macrophage that can lyse the target. To determine whether the humanized antibody and the original mouse anti-Tac antibody can mediate ADCC, a chromium release assay was performed by standard methods. Specifically, human leukemia HUT-102 cells, which express the IL-2 receptor, were incubated with 51Cr to allow them to absorb this radionuclide. The HUT-102 cells were then incubated with an excess of either anti-Tac or humanized anti-Tac antibody. The HUT-102 cells were next incubated for 4 hrs with either a 30:1 or 100:1 ratio of effector cells, which were normal purified human peripheral blood mononuclear cells that had been activated by incubation for about 20 hrs with human recombinant IL-2. Release of 51Cr, which indicated lysis of the target HUT-102 cells, was measured and the background subtracted (Table 3). The results show that at either ratio of effector cells, anti-Tac did not lyse a significant number of the target cells (less than 5%), while the humanized antibody did (more than 20%). Hence, the humanized antibody is likely to be more efficacious than the original mouse antibody in treating T-cell leukemia or other T-cell mediated diseases.

TABLE 3

Percent 51Cr re	lease after ADCC	
		et ratio
	30:1	100:1
Antibody		
Anni-Tac	4%	<1%
Humanized anti-Tac	24%	23%

Higher level expression of the humanized anti-Tac antibody Two new plasmid vectors were prepared for expression of the humanized antibody. The plasmid pVgl (FIG. 9A) contains a human cytomegalovirus IE1 promoter and enhancer (Boshart et al., Cell 41, 521 (1985)), the human genomic Cγl segment including part of the preceding intron, and the hygromycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 9B) is similar to pVgl but contains the human genomic Cκ segment and the gpt gene.

Xba I fragments containing the humanized anti-Tac light chain and heavy chain variable regions were excised respectively from the plasmids pHuLTAC and the pHuGTAC1 and cloned into the Xba I sites of the plasmid vectors pVk and pVGI. To express the humanized anti-Tac antibody, the light chain encoding plasmid was introduced by electroporation into SP2/0 mouse myeloma cells followed by selection for gpt expression. Transfected cells expressing light chain were then transfected with the plasmid encoding the heavy chain followed by selection for hygromycin B resistance. Transfected cells producing the highest levels of humanized antibody as determined by ELISA were used for preparation of antibody. Humanized antibody was purified from culture supernatant of transfected cells by protein A sepharose chromatography.

From the foregoing, it will be appreciated that the humanlike immunoglobulins of the present invention offer numerous advantages of other human IL-2 receptor-specific antibodies. In comparison to anti-Tae mouse monoclonal antibodies, the present human-like immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

EXAMPLE 5

Design of genes for mikβ1 humanized light and heavy chains

To exert its biological effects, IL-2 interacts with a specific high-affinity membrane receptor (Greene, W., et al., 55 Progress in Hematology XIV, E. Brown, Ed., Grune and Statton, New York (1986), at pgs. 283 ff and Waldmann, Ann. Rev. Biochem 58, 875 (1989), which is incorporated herein by reference). The human IL-2 receptor is a complex multichain glycoprotein, with one chain, known as the Tac 60 peptide or alpha chain, being about 55kD in size (see, Leonard, W., et al., J. Biol. Chem. 260, 1872 (1985), which is incorporated herein by reference). The second chain is known as the p75 or beta chain (Tsudo et al., Proc. Nat. Acad. Sci. USA, 83, 9694 (1986) and Sharon et al., Science 65 234, 859 (1986), both of which are incorporated herein by reference). The p55 or Tac chain and the p75 chain each

independently bind IL-2 with low or intermediate affinity, while the IL-2 receptor complex of both chains binds IL-2 with high affinity. The p75 chain of the human IL-2 receptor will often be called herein simply the p75 protein.

Much of the elucidation of the human IL-2 receptor's structure and function is due to the development of specifically reactive monoclonal antibodies. The antibody, mik-β1, binds to the p75 chain (Tsudo et al., *Proc. Nat. Acad. Sci. USA* 86, 1982 (1989), which is incorporated herein by reference).

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindlII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC19 vector for sequencing. For mik-β1, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG. 23A and FIG. 23B. Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVgl-dhfr (FIG. 24A) contains a human cytomegalovirus IEI promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic C, I segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVgl-dhfr but contains the human genomic Cx segment and the gpt gene. Derivatives of the mik-\(\beta \) heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained Xbal sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and Xbal sites (see, C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappachain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric mik-βl antibody was shown to bind to YTJB cells, which express the p75 antigen, by flow eytometry (FlG. 25). Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDRs with the human framework be to introduce distortions into the CDRs that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence database (performed with the MicrorGenic Sequence Analysis Software (Beckman)), the

antibody Lay was chosen to provide the framework sequences for humanization of mik-\(\beta\)1.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the mik-βl variable region. The model was used to determine the amino acids in the mik-βl framework that were close enough to the CDRs to potentially interact with them (category 4 below). To design the humanized light and heavy chain mik-βl variable regions, at each position the amino acid was chosen to be the same as in the Lay antibody, unless that position fell in one or more of five categories:

- (1) The position fell within a CDR,
- (2) The Lay amino acid was unusual for human antibodies at that position, whereas the mik-βI amino acid was typical for human antibodies at that position.
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDRs).

For positions in these categories, the amino acid from the (mouse) mik- β 1 antibody was used. In addition, a position was in the fifth category if

(5) The Lay amino acid was highly unusual for human 25 antibodies at that position, and the mik-β1 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 4. 30 Some amino acids may be in more than one category. The final sequences of the humanized mik-βl light and heavy chain variable domains are shown in FIG. 26A and FIG. 26B, compared with the Lay sequences.

TABLE 4

Category	Light Chain	Heavy Chain
1	24-33, 49-55, 88-96	31-35, 50-65, 98- 108
2	13	84, 89, 90
3	30, 49	
4	70	29, 30, 72, 73
5	41	1

For the construction of genes for the humanized antibod- 45 ies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse mik-Bl chains (FIG. 23A and FIG. 23B), generally utilizing codons found in the mouse sequence. Several degenerate codons 50 were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an Xbal site at each end. Each gene was constructed from four overlapping synthetic oligonuclcotides. For each variable 55 domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 27A and FIG. 27B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 base long with about a 20 base overlap. Double stranded DNA fragments were synthesized with sequenase from each pair of oligonucleotides, digested with restriction enzymes, ligated to pBluescriptll KS (+) (Stratagene) vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the Xbal sites of the

pVgl-dhfr or pVk expression vectors. In vitro mutagenesis was used to change an Ala amino acid originally encoded by oligonucleotide wps54 to the Glu (E) at position 1 of the humanized heavy chain (FIG. 26B) by changing the nucleotides CT to AG. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by EHSA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody was eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized mik- β l antibody was characterized in comparison to the murine and chimeric antibodies. The humanized antibody bound to YTJB cells, which express p75 chain at a high level, in a fluorocytometric analysis in a manner similar to the chimeric antibody (FIG. 25), showing that it recognizes the same p75 protein.

The affinity of the humanized antibody was determined by competition with the radio-iodinated mouse mik-β1 antibody (FIG. 28). The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in *Fundamental Immunology* (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference). The binding affinity of the humanized mik-β1 antibody was within about 2-fold of the affinity of the mouse mik-β1 antibody.

The ability of humanized mik-\(\beta \) plus humanized anti-Tac antibody to inhibit IL-2 stimulated proliferation of human lymphocytes was determined. Human mononuclear cells, collected from human blood by centrifugation on Ficoll-Paque (Pharmacia), were diluted to 2×10⁶ cells/ml in RPMI mcdium+10% fetal calf serum (FCS). A 1/200 volume of phytohemagglutinin P (Difeo) was added and the cells were incubated for 4 days. The cells were incubated an additional 4 days in RPMI+10% FCS+10 u/ml IL-2. 10° of these PHA activated blasts were then incubated with or without 2 µg each of humanized mik-Bl and humanized anti-Tac in 150 μl of RPMI+ 10% FCS in wells of a 96-well plate for 1 hr, to which various dilutions of IL-2 (Amgen) were then added in 50 μl medium. The cells were incubated 48 hr, 0.5 μCi methyl-3H-thymidine (Amersham, 82 Ci/mmol) was added, and the cells were incubated 24 hr. Cells were harvested with a cell harvester and radioactivity determined. The combination of the antibodies greatly inhibited proliferation of the cells in response to IL-2 (FIG. 29), suggesting a combination of the antibodies will have strong immunosuppressive properties. Humanized mik-Bl plus humanized anti-Tac inhibited proliferation much more strongly than did either antibody alone.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other p75 specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

Design of genes for Fd79 and Fd138-80 humanized light and heavy chains

Exemplary DNA sequences coding for the polypeptide chains comprising the heavy and light chain hypervariable regions (with human framework regions) from monoclonal antibodies Fd7Q and Fd138-80, are shown in FIG. 30A through FIG. 30D.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain regions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoRI sites (scheme shown in FIG. 14). This method yields clones with authentic variable domain sequences, in contrast to other methods using mixed primers designed to anneal to the variable domain sequence (J. W. 20 Lamick et al., Bio/Technology 7, 934 (1989) and Y. L. Chiang et al., BioTech. 7, 360 (1989)). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For Fd79, two gamma-1 specific and 5 kappa specific clones were 25 sequenced. The two gamma-1 specific clones are identical in sequence. This heavy chain cDNA fragment encodes a signal peptide of 19 amino acids, a V region in mouse heavy chain subgroup IIIB, a D segment, and a JHI segment with 4 alterations compared to the genomic J_H1 sequence. The 30 deduced amino acid sequence is shown in FIG. 30A.

The five kappa specific clones belong to two groups. Two clones are identical and encode a kappa chain in which the conserved amino acid 23 cysteinc has been substituted by a tyrosine, probably representing the non-productive allele. 35 The other three clones have an identical sequence encoding a signal peptide sequence of 20 amino acids, a V region in mouse kappa chain subgroup III, and a $J_k 2$ segment with a single alteration compared to the genomic $J_k 2$ sequence (FIG. 30B). The validity of the heavy chain and the kappa 40 chain sequences was subsequently confirmed by the construction and expression of a chimeric antibody as discussed below.

The heavy chain and the kappa chain of Fdl38-80 were cloned similarly. Three clones each of the heavy chain and 45 the kappa chain were sequenced. All three heavy chain clones have an identical sequence encoding a signal peptide sequence of 19 amino acids, a V region in mouse heavy chain subgroup II, a D segment and the $J_{\rm H}3$ segment (FIG. 30C). The three kappa clones are also identical in sequence. 50 This light chain fragment encodes a signal peptide sequence of 20 amino acids, a V region gene in mouse kappa chain subgroup V and the $J_{\rm H}5$ segment (FIG. 30D). Both chains shown no irregularities in coding sequence; their validity was subsequently confirmed by construction and expression 55 of a chimeric antibody.

Construction and expression of chimeric antibodies.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVg1 (FIG. 9A) contains a human cytomegalovirus IE1 60 promoter and enhancer (M. Boshart et al., Cell 4.1, 521 (1985)), the human genomic C_v1 segment including part of the preceding intron, and the hygromycin gene (Blochlinger et al., Mol. Cell. Biol. 4, 2929 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 65 9B) is similar to pVg1 but contains the human genomic C_v segment and the gpt gene. Derivatives of the Fd79 and

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Fdl38-80 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (see., C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the XbaI sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibodies, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric Fd79 and Fd138-80 antibodies were shown to bind to HSV-1 infected vero cells by flow cytometry. Viral neutralization assays also indicated that the chimeric antibodies retain the neutralization activities of the murine antibodies (data not shown, but see below for similar results with humanized antibodies).

Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (C. Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDRs with the human framework be to introduce distortions into the CDRs that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Pom was chosen to provide the framework sequences for humanization of Fd79.

The computer program ENCAD (Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the Fd79 variable region. Inspection of the refined model of murine Fd79 revealed two amino acid residues in the framework that are close enough to have significant contacts with the CDR residues (Table 5). Lys in light chain position 49 has contacts with 3 amino acids in CDR2 of the light chain (L50 Tyr, L53 Asn, L55 Glu) and 2 amino acids in CDR3 of the heavy chain (H99 Asp, H100 Tyr). Leu in heavy chain position 93 also shows interactions with 2 amino acids in CDR2 of the heavy chain (H35 Ser, H37 Val) and an amino acid in CDR3 of the heavy chain (H100C Phe), Hence, L49 Lys and H93 Leu were retained in the construction of humanized Fd79, as their replacement with human Pom framework residues would be likely to introduce distortions into the CDRs. Also, 7 other residues in the Pom framework (5 in the light chain and 2 in the heavy chain) were substituted with common human residues (identical to the murine Fd79 sequence in 6 of the choices) because of their rare occurrence in other human antibodies. The elimination of unusual amino acids in the framework may further reduce immunogenicity. The murine Fd79 sequences and the corresponding humanized sequences are shown in FIG. 30A and FIG. 30B. Substituted residues in the Pom framework are underlined,

TABLE 5

Residues in the framework sequence showing contacts will
residues in the hypervariable regions.

Residue No.	Amino Acid	Contacting CDR residues ²
Fd79		
1.49	Lys	L50Y, L53N,L55E, H99D, H100Y
н93	Leu	H35S, H37V, H100CF
Fd138-80		
L36	His	L34V, L89Q
H27	Тут	H#2H, H34I
H30	Тут	H#2H, H53R
H48	Phc	H63F
H66	Lys	H63F
H67	Λla	H63F

The amino acid residues are numbered according to the Kabat system (E. A. Kabat et al., Sequences of Proteins of immunological interest, National Institutes of Health, Bethesda. MD (1987)): the first letter (H or L) stands for the heavy chain or light chain. The following number is the residue number. The last letter is the amino acid one letter code.

The last letter is the amino acid one letter code.

2. The hypervariable regions are defined according to Kabat: Light chain CDR1: tesidue 24-34; CDR2: 50-56; CDR3: 89-97. Heavy chain CDR1: 31-35; CDR2: 50-65; CDR3: 95-102.

Similarly, the murine heavy chain and light chain 25 sequences of Fd138-80 were subjected to sequence homology search against the NBRF protein sequence database. The sequences of the human antibody Eu were selected to provide the framework sequences for humanized Fd138-80. Inspection of a computer-generated model of Fd138-80 30 revealed 6 amino acid residues in the framework that are close enough to have important contacts with CDR residues. The residues and their contacting counterparts are listed in Table 5; these murine residues were retained in the construction of humanized Fdl 38-80. Two other residues (L87 35 Phe and H37 Met) show significant contacts with L98 Phe, which is immediately adjacent to CDR3, so these two mouse residues were also retained. Eight amino acids in the Eu framework (2 in the light chain and 6 in the heavy chain) were substituted with the murine residues (which are also consistent with the human consensus residues) because of their rare occurrence in other human antibodies. The murine Fd138-80 sequences and the corresponding humanized sequences are shown in FIG. 30C and FIG. 30D. Substituted residues in the Eu framework are underlined.

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the signal peptides, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove unde- 50 sirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an XbaI site at each end. Each gene was constructed from four overlapping synthetic oligonucleou'des. For each variable domain gene, two pairs of overlapping oligonucleotides on 55 alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal. The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with a 15 base overlap. 60 Double stranded DNA fragments were synthesized with Klenow polymerase, digested with restriction enzymes, ligated to pUC18 vector and sequenced. The two fragments with the correct sequences were then ligated into the Xbal sites of pVgl or pVk expression vectors.

The synthetic genes were then cloned into the pVgl and pVk expression vectors. For each humanized antibody con-

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structed, the heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibodies were purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibodies were eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer was exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized Fd79 and Fd138-80 antibodies were characterized in comparison to the murine and chimeric antibodies. Both humanized antibodies bind to Vero cells infected with HSV-1 in a fluorocytometric analysis in a manner similar to the chimeric antibodies (FIG. 31A and FIG. 31B), suggesting that they recognize their respective viral antigens. To more quantitatively assess the binding activity, radioiodinated murine antibodies were bound to virally infected cells and Scatchard analysis performed.

The affinitics of the humanized antibodies were determined by competition with the iodinated antibodies. Vero cells infected with HSV-1 were used as source of gB and gD antigens. Increasing amounts of competitor antibody (mouse or humanized) were added to 1.5 ng of radioiodinated tracer mouse antibody (2uCi/ug) and incubated with 4×10⁵ infected Vero cells in 0.2 ml of binding buffer (PBS+2% FCS+0.1% azide) for 1 hr. at 4° C. Cells were washed and pelleted, and their radioactivities were measured. The concentrations of bound and free tracer antibody were calculated. The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in Fundamental Immunology (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference).

The measurements indicate that there is no significant loss of binding affinities in the humanized antibodies (Table 6). Specifically, there is an approximately 2-fold decrease in affinity in humanized Fd79 compared to the murine Fd79 (Ka of 5.3×10⁷ M⁻¹ vs. 1.1×10⁸ M⁻¹). The affinity of humanized Fd138-80 is comparable to that of the murine antibody (Ka of 4.8×10⁷ M⁻¹ vs 5.2×10⁷ M⁻¹).

TABLE 6

Binding affinities of murine and humanized antibodies.		
Mouse K _a (M ⁻¹)	Humanized K _a (M ⁻¹)	
Fd79 (anti-gB) Fd138-80 (anti-gD)	1.1 × 10 ⁸ 5.2 × 10 ⁷	5.3×10^{7} 4.8×10^{7}

Murine Fd79 and Fd138-80 have been shown to neutralize HSV-1 in vitro without complement (J. Koga et al., Virology 151, 385 (1986)), so the neutralizing activities of the humanized antibodies were compared with the mouse antibodies. Serial dilutions of equal quantities of murine and humanized antibodies were incubated with virus for 1 hr. before inoculation onto Vero cells. After 4 days, cells were stained with neutral red to visualize plaques. Results from these plaque reduction assays indicated that both humanized Fd79 and Fd138-80 neutralize virus as efficiently as their murine counterparts (FIGS. 32A and B). Both humanized and murine Fd79 cause a 90% reduction of plaques at an antibody concentration of 10 nM (1.5 ug/ml). Similarly, humanized and murine Fd138-80 were able to cause a 90% plaque reduction at equivalent levels.

57 The antibodies were also investigated for their ability to

protect cells from viral spread in tissue culture. Vero cells

58 are known to bind to the CD33 antigen include L4B3, L1B2

and MY9 (Andrews et al., Blood 62, 124 (1983) and Griffin et al., Leukemia Research 8, 521 (1984), both of which are incorporated herein by reference).

were inoculated with virus at 0.1 pfu/cell and allowed to adsorb for 2 hrs. at 37° C. before addition of 10 ug/ml antibody. After four days, cells were stained with an anti-gB antibody for detection of viral antigens on infected cells. Results indicated that both murine and humanized Fd79 at 10 ug/ml protected culture cells from infection (FIG. 33A). However, neither murine nor humanized Fd138-80 were able to protect cells against viral spread (FIG. 33B), despite 10 their ability to neutralize virus before inoculation. Both gB and gD are thought to be associated with cell fusion and virus infectivity (W. Cai et al., J. Virol. 62, 2596 (1988) and

Another antibody that binds to CD33 is M195 (Tanimoto et al., Leukemia 3, 339 (1989) and Scheinberg et al., Leukemia 3, 440 (1989), both of which are incorporated herein by reference).

A. O. Fuller and P. G. Spear, Proc. Natl. Acad. Sci. USA 84, 5454 (1987)). However, it is possible that Fd79 blocks both 15 the infectivity and cell fusion functions of gB, while Fd138-80 blocks only the infectivity function of gD, so virus can

Cloning of heavy chain and light chain cDNA.

still spread cell-to-cell.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindllI sites, and 5' primers that hybridized to the dG tails and contained EcoR1 sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoRI and HindIII and cloned into the pUC18 vector for sequencing. For M195, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG. 34A and FIG. 34B. Construction and expression of chimeric antibody.

The binding, neutralization and protection results all indicate that the humanized Fd79 and Fd138-80 antibodies 20 have retained the binding activities and the biological properties of the murine monoclonal antibodies. The availability of humanized antibodies with specificity for HSV gB and gD, inter alia, provides an opportunity for studies of the in vivo potency and immunogenicity of humanized antibodies 25 in treating viral diseases. The recognition by Fd79 and Fd138-80 of type-common epitopes of gB and gD (J. Koga et al., Virology 151, 385 (1986)) expands the therapeutic potential to herpes simplex virus type 2 as well as type 1.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVgl-dhfr (FIG. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Ceil 41, 521 (1985)), the human genomic Cy1 segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl Acad. Sci. USA 80, 2495 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVgl-dhft but contains the human genomic Ck segment and the gpt gene. Derivatives of the M195 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained Xbal sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and XbaI sites (sec, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

The use of a combination of two or more humanized 30 antibodies in therapy is important for reducing the development of antibody resistant strains. Combination therapy of humanized antibodies with other antiviral agents such as acyclovir provides further opportunities to combat diseases when chemotherapeutic agents alone have not been effec- 35 tivc. As Fd79 and Fd138-80 reduce the frequency of viral persistence in a murine ocular model (J. F. Metcalf et al., Cur. Eye Res. 6, 173 (1987)), the humanized antibodies, typically together with other antiviral agents, are capable of reducing episodes of recurrent genital infection, an area 40 where traditional anti-viral agents have not been effective (L. Corey et al., N. Engl. J. Med. 306, 1313 (1982)). Incorporation of the human constant domains can also enhance effector functions, such as antibody-dependent cellular cytotoxicity, leading to greater therapeutic efficiency in 45 human natients.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Purified chimeric M195 antibody was shown to bind to U937 cells, which express the CD33 antigen, by flow cytometry (FIG. 35).

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other HSV specific antibodies,

Computer modeling of humanized antibodies.

In comparison to mouse monoclonal antibodies, the 50 present humanized immunoglobulin can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antibody is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysi's Software (Beckman)), the antibody Eu was chosen to provide the framework sequences for humanization of M195.

EXAMPLE 7

Design of genes for M195 humanized light and heavy chains

The p67 protein or CD33 antigen is found on the surface of progenitors of myeloid cells and of the leukemic cells of most cases of AML, but not on lymphoid cells or nonhematopoietic cells (sec, Leucocyte Typing III, ed. by A. J. 65 McMichael, Oxford University Press, pp. 622-629 (1987), which is incorporated herein by reference). Antibodies that

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the M195 variable region. The model was used to determine the amino acids in the M195 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain M195 variable regions, at each position the amino acid was chosen to be the same as in the Eu antibody, unless that position fell in one or more of four categories:

- (1) The position fell within a CDR,
- (2) The Eu amino acid was unusual for human antibodies at that position, whereas the M195 amino acid was typical for human antibodies at that position,
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human 20 sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Eu light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse M195 antibody was used: The amino acids in each category are shown in Table 7. Some amino acids may be in more than one category. The final sequences of the humanized M195 light and heavy chain variable domains are shown in FIG. 36A and FIG. 36B, compared with the Eu sequences.

TABLE 7

Category	Light Chain	Heavy Chain
1	24-38, 54-60, 93-101	31-35, 50-66, 99-105
2	10, 52, 67, 110	93, 95, 98, 106, 107, 108, 110
3	_	30, 67, 98, 106
4	40, 52, 74	27, 30, 48, 68, 98

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse M195 chains (FIG. 34A and FIG. 34B), generally utilizing codons 45 found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an Xbal site at each end. Each gene was constructed from four 50 overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 37A and FIG. 37B). The oligo- 55 the polypept de chains comprising the heavy and light chain nucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction 60 enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the XbaI sites of the pVgl-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were 65 carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells were selected for gpt expression. Clones were screened by assaying human antibody production in the culture supernatant by ELISA, and antibody was purified from the best-producing clones. Antibody was purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody was eluted with 0.2M Glycine-HCI, pH3.0 10 and neutralized with 1M Tris PH8.0. The buffer was exchanged into PBS by passing over a PD10 column (Phar-

Properties of humanized antibodies.

The humanized M195 antibody was characterized in 15 comparison to the murine and chimeric antibodies. The humanized antibody bound to U937 cells in a fluorocytometric analysis in a manner similar to the chimeric antibody (FIG. 35), showing that it recognizes the same CD33 anti-

The affinity of the humanized antibody was determined by competition with the radio-iodinated mouse M195 antibody (FIG. 38). The binding affinities were calculated according to the methods of Berzofsky (J. A. Berzofsky and I. J. Berkower, in Fundamental Immunology (ed. W. E. Paul), Raven Press (New York), 595 (1984), which is incorporated herein by reference). The mouse M195 had an affinity comparable to the published value (Tanimoto et al., op. cit.) and the humanized M195 antibody had an affinity the same as the mouse M195 to within experimental error.

Humanized M195 is useful to mediate antibody-dependent cellular cytotoxicity when human effector cells and human CD33-expressing cells are used. This is analogous to other humanized antibodies, such as reported by Junghans et al., Cancer Research 50, 1495 (1990), which is incorporated 35 herein by reference.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other CD33 specific antibodies. In comparison to mouse monoclonal antibodics, the present 40 humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

EXAMPLE 8

Design of genes for CMV5 humanized light and heavy chains

Three neutralizing antibodies to the gH glycoprotein of human cytomegalovirus (CMV) are designated CMV5, CMV109 and CMV115.

Exemplary DNA sequences, which on expression code for CDR's of monoclonal antibody CMV5 are included in FIG. 39A and FIG. 39B. Due to codon degeneracy and noncritical amino-acid substitutions, other DNA sequences can be readily substituted for those sequences, as detailed below. Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors pVk and pVgl-dhfr (FIG. 24A and 24B) using site-directed mutagenesis, such as after CHI to produce Fab fragments or after the hinge region to produce (Fab')₂ fragments.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable 5 domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoR I sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoR I and HindIII and cloned into the pUC18 vector for sequencing. For CMV5, two gamma-2a specific and two kappa specific clones were sequenced. The two gamma-2a clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced 15 amino acid sequences are shown in FIG. 39A and FIG. 39B. Similarly, by using techniques, which are well-known in the art, cDNAs for the CMV109 and CMV115 antibodies may be obtained and their sequence determined. Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVgl-dhfr (FIG. 24A) contains a human cytomegalovirus IEI promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cyl segment including part of 25 the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1983), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVgl-dhfr but contains the human genomic Ck segment and the gpt 30 gene. Derivatives of the CMV5 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained XbaI sites; the 3' primers hybridized to the last 15 nucleotides of the J 35 regions and contained splice donor signals and Xbal sites (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the cytomegalovirus 40 promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse mycloma cells by electroporation and cells selected for gp1 expression. Clones secreting a maximal amount of complete 45 antibody were detected by ELISA. Purified chimeric CMV5 antibody was shown to bind to CMV-infected cells, which express the gH antigen, by immunostaining of CMV-infected human embryonic lung fibroblasts.

Computer modeling of humanized antibodies

In order to retain high binding affinity in the humanized amibodies, the general procedures of Queen et al. were followed (See, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human anti- 55 body is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to inmoduce distortions into the CDR's that could reduce affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework 60 sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence database (performed with the MicroGenie Sequence Analysis Software (Beckman)), the antibody Wol was chosen to 65 provide the framework sequences for humanization of CMV5.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) was used to construct a model of the CMV5 variable region. The model was used to determine the amino acids in the CMV5 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain CMV5 variable regions, at each position the amino acid was chosen to be the same as in the Wol antibody, unless that position fell in one or more of five categories:

- (1) The position fell within a CDR,
- (2) The Wol amino acid was unusual for human antibodies at that position, whereas the CMV5 amino acid was typical for human antibodies at that position.
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., op. cit.) as the Wol light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse CMV5 antibody was used. In addition, a position was in the fifth category if the Wolamino acid was highly unusual for human antibodies at that position, and the CMV5 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 8. Some amino acids may be in more than one category. The final sequences of the humanized CMV5 light and heavy chain variable domains are shown in FIG. 40A and FIG. 40B, compared with the Wol sequences.

TABLE 8

Calegory	Light Chain	Heavy Chain
1	24-34, 50-56, 89-97	31-35, 50-66, 99-108
		69, 80
2		69, 80
3	49	30
4		24, 27, 28, 30, 97
5		5

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, including the same signal peptides as in the mouse CMV5 chains (FIG. 39A and FIG. 39B), generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric genes and an Xbal site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain gene, two pairs of overlapping oligonucleomides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 41A and FIG. 41B). The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction

enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences were then ligated into the Xbal sites of the pVg1-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions were carried out under conditions well-known in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids are transfected into Sp2/0 mouse myeloma cells by electroporation and cells are selected for gpt expression. Clones are screened by 10 assaying human antibody production in the culture supernatant by ELISA, and antibody purified from the best-producing clones. Antibody is purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharose CL-4B (Pharmacia). The bound antibody is 15 eluted with 0.2M Glycine-HCl, pH3.0 and neutralized with 1M Tris PH8.0. The buffer is exchanged into PBS by passing over a PD10 column (Pharmacia).

Humanized antibody was also produced by transient transfection. The heavy chain and light chain plasmids were 20 transfected into S194 cells (ATCC TIB 19) by the DEAE-dextran method (Queen et al., Mol. Cell. Biol. 4, 1043 (1984), which is incorporated herein by reference), and humanized CMV5 antibody was purified from the media supernatant as above. Antibody was quantitated by ELISA 25 assay for human lg.

Properties of humanized antibodies.

The humanized CMV5 antibody was characterized in comparison to the murine and chimeric antibodies. The humanized CMV5 antibody was shown to bind about as well 30 as the mouse and chimeric antibodies to CMV antigen, by immunostaining of CMV-infected human embryonic lung (HEL) cells (ATCC CCL 137). HEL cells monolayers in 96-well plates were infected with CMV at 0.01 pfu/cell, incubated for 4 days, dried at 37° C. and stored wrapped at 35 4° C. 100 μl blotto (5% Carnation Instant Milk in PBS at pH 7.4) was added to each well and incubated at 37° C. for 30 min. The blotto was poured off and 75 µl of a series of 2-fold dilutions of mouse, chimeric and humanized CMV5 antibody was added to the wells. The plate was incubated 1 hr 40 at 37° C. and washed twice with blotto (each wash was left on for 10 min). Then 75 µl of diluted peroxidase (HRP) conjugated goat anti-mouse or anti-human IgG (Tago) was added to each well and incubated for 1 hr at 37° C. The plate was washed 2x with PBS and 150 µl of HRP substrate 45 solution was added to each well. Color was allowed to develop at room temperature. The plates were washed with water and air dried. The wells were examined under a microscope to determine the highest dilution of the antibodies that formed a colored precipitate on the CMV-infected 50 cells. For all three antibodies, 63 ng/ml was the least amount of antibody that produced a detectable precipitate, indicating that humanized CMV5 binds about as well as the mouse and chimeric antibodies.

To compare the affinities of mouse and humanized CMV5 in another way, a competition experiment was performed. Plates of CMV-infected HEL cells as above were incubated with blotto for 30 min at 37° C. The blotto was poured off and dilutions of mouse or humanized CMV5 were added to each well in 75 µl of PBS. Then 125 µl of radio-iodinated 60 mouse CMV5 (1 µCi/µg) in PBS, containing 28,000 cpm was added to each well and incubated at 37° C. for 2.5 hr. The plate was washed 5 times with PBS, and the contents of each well were solubilized with 200 µl of 2% SDS and counted. Increasing concentrations of mouse and humanized 65 CMV5 inhibited binding of the radiolabeled CMV5 about equally well (FIG. 42), so humanized CMV5 has approxi-

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mately the same binding affinity as mouse CV5. An irrelevant antibody did not compete in this assay.

The ability of humanized CMV5 to neutralize CMV is compared to that of mouse CMV5. Mouse and humanized CMV5 are successively diluted by 2-fold in 100 µl of DME medium+2% FCS in wells of a 96-well plate. 100 µl of CMV, which has been diluted to contain 100 tissue culture infectious dose-50% (TCID50) units, are added to each well and incubated for 60 min at 37° C. Each well of antibodyvirus mixture is added to a well of subconfluent HEL cells in a 96-well plate from which the medium has been removed. The cells are incubated for 5 days and cytopathic effect (CPE) is examined in each well under a microscope. The highest dilution of antibody that inhibits CPE by 90% is a measure of the neutralizing ability of the antibody. The humanized CMV5 antibody will neutralize CMV antibody approximately as well as the mouse CMV5 antibody.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other CMV specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

EXAMPLE 9

Design of genes for AF2 human-like light and heavy chains

This example is directed to recombinant DNA segments encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding to a desired epitope of γ -IFN, such as monoclonal antibody AF2. Exemplary DNA sequences, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody AF2 are included in FIG. 43A and FIG. 43B. Due to codon degeneracy and non-critical aminoacid substitutions, other DNA sequences can be readily substituted for those sequences.

Cloning of heavy chain and light chain cDNA.

cDNAs for the heavy chain and light chain variable domain genes were cloned using anchored polymerase chain reactions (E. Y. Loh et al., Science 243, 217 (1989)), using 3' primers that hybridized to the constant regions and contained HindIII sites, and 5' primers that hybridized to the dG tails and contained EcoR I sites (scheme shown in FIG. 14). The PCR amplified fragments were digested with EcoR I and HindIII and cloned into the pUC18 vector for sequencing. For AF2, two gamma-2b specific and two kappa specific clones were sequenced. The two gamma-2b clones and two kappa clones are respectively identical in sequence. The cDNA variable domain sequences and the deduced amino acid sequences are shown in FIG. 43A and FIG. 43B. Construction and expression of chimeric antibody.

Two plasmid vectors were prepared for construction and expression of the chimeric antibody genes. The plasmid pVgl-dhfr (FIG. 24A) contains a human cytomegalovirus IE1 promoter and enhancer (M. Boshart et al., Cell 41, 521 (1985)), the human genomic Cγl segment including part of the preceding intron, and a dihydrofolate reductase (dhfr) gene (Simonsen et al., Proc. Natl. Acad. Sci. USA 80, 2495 (1984), which is incorporated herein by reference) for selection. The plasmid pVk (FIG. 24B) is similar to pVgl-dhfr but contains the human genomic Cκ segment and the gpt

gene. Derivatives of the AF2 heavy and light chain variable regions were prepared from the cDNAs by polymerase chain reaction. The 5' primers hybridized to the V regions starting at the ATG codons and contained Xbal sites; the 3' primers hybridized to the last 15 nucleotides of the J regions and contained splice donor signals and Xbal sites (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989), which is incorporated herein by reference). The modified V regions were cloned into the Xbal sites of the respective plasmid vectors between the CMV promoter and the partial introns of the constant regions.

For expression of the chimeric antibody, the heavy chain and kappa chain plasmids were transfected into Sp2/0 mouse myeloma cells by electroporation and cells scleeted for gpt expression. Clones secreting a maximal amount of complete antibody were detected by ELISA. Chimeric AF2 antibody was shown to bind to human γ -IFN by ELISA. Computer modeling of humanized antibodies.

In order to retain high binding affinity in the humanized antibodies, the general procedures of Queen et al. were followed (see, Queen et al., Proc. Natl. Acad. Sci. USA 86, 10029 (1989) and WO 90/07861, which are incorporated herein by reference). The more homologous a human antiibody is to the original murine antibody, the less likely will combining the murine CDR's with the human framework be to introduce distortions into the CDR's that could reduce 25 affinity. Normally the heavy chain and light chain from the same human antibody are chosen to provide the framework sequences, so as to reduce the possibility of incompatibility in the assembling of the two chains. Based on sequence homology search against the NBRF protein sequence data- 30 base (performed with the MicroGenic Sequence Analysis Software (Beckman)), the antibody Eu was chosen to provide the framework sequences for humanization of AF2.

The computer program ENCAD (M. Levitt, J. Mol. Biol. 168, 595 (1983), which is incorporated herein by reference) 35 was used to construct a model of the AF2 variable region. The model was used to determine the amino acids in the AF2 framework that were close enough to the CDR's to potentially interact with them (category 4 below). To design the humanized light and heavy chain AF2 variable regions, at each position the amino acid was chosen to be the same as in the Eu antibody, unless that position fell in one or more of five categories:

- (1) The position fell within a CDR,
- (2) The Eu amino acid was unusual for human antibodies at that position, whereas the AF2 amino acid was typical for human antibodics at that position,
- (3) The position was immediately adjacent to a CDR,
- (4) The model described above suggested that the amino 50 acid may be physically close to the antigen binding region (CDR's).

In category (2), "unusual" is interpreted to include amino acids that occur in less than about 20% of the human sequences in the same subgroups (as defined by Kabat et al., 55 op. cit.) as the Eu light and heavy chains, and "typical" is interpreted to include amino acids that occur in more than about 25% but generally more than 50% of the human sequences in those subgroups. For positions in these categories, the amino acid from the mouse AF2 antibody was 60 used. In addition, a position was in the fifth category if the Eu amino acid was highly unusual for human antibodies at that position, and the AF2 amino acid was different but also unusual. Then an amino acid typical for human antibodies at that position may be used.

The amino acids in each category are shown in Table 9. Some amino acids may be in more than one category. The

final sequences of the humanized AF2 light and heavy chain variable domains are shown in FIG. 44A and FIG. 44B, compared with the Eu sequences.

TABLE 9

Calegory	Light Chain	Heavy Chain
1	24-34, 50-56, 89-97	31-35, 50-66, 99-106
2	48	93, 95, 98, 107, 108, 109, 111
3		30, 98, 107
4	48, 70	27, 28, 30, 98, 107
5	63	

For the construction of genes for the humanized antibodies, nucleotide sequences were selected that encode the protein sequences of the humanized heavy and light chains, plus typical immunoglobulin signal sequences, generally utilizing codons found in the mouse sequence. Several degenerate codons were changed to create restriction sites or to remove undesirable ones. The nucleotide sequences also included the same splice donor signals used in the chimeric gencs and an Xbal site at each end. Each gene was constructed from four overlapping synthetic oligonucleotides. For each variable domain genc, two pairs of overlapping oligonucleotides on alternating strands were synthesized that encompassed the entire coding sequences as well as the signal peptide and the splice donor signal (FIG. 45A and FIG. 45B) The oligonucleotides were synthesized on an Applied Biosystems 380B DNA synthesizer. Each oligo was about 110-140 bases long with about a 15 base overlap. Double stranded DNA fragments were synthesized with Klenow polymerase from each pair of oligonucleotides, digested with restriction enzymes, ligated to the pUC18 vector and sequenced. Two fragments with the respectively correct half-sequences are then ligated into the XbaI sites of the pVg l-dhfr or pVk expression vectors in the appropriate orientations to produce the complete heavy and light chain genes. Reactions are carried out under conditions wellknown in the art (Maniatis et al., op. cit.)

The heavy chain and light chain plasmids are transfected into Sp2/0 mouse myeloma cells by electroporation and cells selected for gpt expression. Clones are screened by assaying human antibody production in the culture supernatant by ELISA, and antibody purified from the best-producing clones. Antibody is purified by passing tissue culture supernatant over a column of staphylococcal protein A-Sepharosc CL-4B (Pharmacia). The bound antibody is eluted with 0.2M Glycine-HCI, pH3.0 and neutralized with 1M Tris PH8.0. The buffer is exchanged into PBS by passing over a PD10 column (Pharmacia).

Properties of humanized antibodies.

The humanized AF2 antibody is characterized in comparison to the murine and chimeric antibodies. The humanized antibody will bind to γ -IFN in an ELISA assay in a manner similar to the mouse and chimeric antibodies, showing that it recognizes γ -IFN.

To compare the binding affinities of mouseAF2 antibody and humanized AF2 antibody, a competitive ELISA assay is performed. An ELISA plate is coated with human recombinant γ -IFN by adding 100 μ l of a 500 ng/ml solution of γ -IFN in PBS to each well and incubating overnight at 4° C. Subsequent steps are caucied out at room temperature. The γ -IFN solution is removed and 200 μ l of ELISA buffer (0.1% Tween-20, 1% Bovine serum albumin in PBS) is added to each well and incubated for 1 hr. After removing the solution, varying amounts of competitor antibody (mouse AF2 or humanizedAF2) in 100 μ l PBS is added to each well,

along with an amount of biotinylated AF2 predetermined to give a good ELISA response. The plate is incubated for 1 hr and then washed 3 times with ELISA buffer. An amount of horseradi'sh peroxidase (HRP)-con jugated strepavidin predetermined to be in excess is added in 100 µl PBS to each well and incubated for 30 min. The plate is washed 3 times in ELISA buffer, and 100 µl of substrate solution for HRP is added to each well. The plate is incubated for IO-30 min, and the optical density of each well is determined with an ELISA reader (BioRad). The decrease in optical density with increasing concentrations of competitor antibodies mouse AF2 and humanized AF2 are plotted. Mouse AF2 and humanized AF2 will compete similarly, showing that their binding affinities for γ -IFN are approximately the same. The procedures used are well known in the art (e.g., Harlow and 15 Lane, op. cit.).

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An important biological activity of γ -IFN is the induction of expression of class II HLA antigens on cells. To determine the ability of mouse and humanized AF2 to neutralize this activity, about 5×10^4 HS294T cells (Basham et al., J. Immunol. 130, 1492 (1983), which is incorporated herein by reference) are plated in 1.0 ml DMEM medium+10% FCS in each well of a 24-well plate. After overnight incubation, 0.1 nM interferon and varying amounts of mouse or humanized AF2 are added to the cells, and the plate is incubated for 25 72 hr. The cells are removed from the plate with 0.05M EDTA, stained with monoclonal antibody L243 from the

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American Type Culture Collection (ATCC) against HLA-D antigen, washed, stained with FITC conjugated goat antimouse lg and analyzed with a FACScan (Becton-Dickinson). Increasing concentrations of mouse AF2 refuce fluorescence of the cells (FIG. 46), indicating the antibody is preventing induction of HLA-D by γ-IFN. The humanized AF2 will act similarly to mouse AF2 in this assay, showing that it neutralizes the biological activity of γ-IFN.

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other γ -IFN specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins can be more economically produced and contain substantially less foreign amino acid sequences. This reduced likelihood of antigenicity after injection into a human patient represents a significant therapeutic improvement.

All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference. Although the present invention has been described in some detail by way of illustration and example for purposes of clarity and understanding, it will be apparent that certain changes and modifications may be practiced within the scope of the appended claims.

SEQUENCE LISTING

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( I ) GENERAL INFORMATION:
    ( i i i ) NUMBER OF SEQUENCES: 113
( 2 ) INFORMATION FOR SEQ ID NO:3:
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              ( A ) LENGTH: 106 amino acids
              ( B ) TYPE; amino acid
              ( C ) STRANDEDNESS: single
              ( D ) TOPOLOGY: unknown
     ( i i ) MOIECULE TYPE protein
    ( i i i ) HYPOTHETICAL: NO
     ( i x ) FEATURE:
              ( A ) NAME/KEY: Protein
              ( B ) LOCATION: 1.106
              ( D ) OTHER INFORMATION: /notc="Variable region of the mouse
     ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:1:
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1 5 10
      Glu Lys Val Thr lle Thr Cys Scr Ala Ser Ser Ser lle Ser Tyr Met 20 30
      His Trp Phe Glv Glv Lys Pro Gly Thr Ser Pro Lys Leu Trp lle Tyr 35
      Thr Thr Scr Asn Leu Ala Scr Gly Val Pro Ala Arg Phc Scr Gly Scr 50 60
      Gly Ser Gly Thr Ser Tyr Ser Lea Thr 11e Ser Arg Met Glu Ata Glu
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      Asp Ala Ala Thr Tyr Cys His Gln Arg Scr Thr Tyr Pro Leu Thr
85 90
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69 70

Pho Gly Ser Gly Thr Lys Lev Glu Leu Lys. 100

- (2) INFORMATION FOR SEQ ID NO:2:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - ('B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
 - (i i) MOLECULE TYPE: protein
 - (1 i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..107
 - (D) OTHER INFORMATION: /note="Variable region of the human Eu anti-body light chaim."
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 Met
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 Pro
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 Thr
 Leu
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 Ala
 Thr
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 Leu
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 Trp
 Trp
 Gln
 Gin
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- (2) INFORMATION FOR SEQ ID NO:3:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 am'ino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
 - (i i) MOLECULE TYPE: protein
 - (i i i) HOPOTHETICAL: NO
 - (i x) FEATURE
 - (A) NAME/KEY: Proicin
 - (B) LOCATION: 1,.116
 - (D) O'FFER INFORMATION: /noie="Variable region of the mouse anti-Tac antibody heavy chain."
 - (\times i) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Gin Val Gin Leu Gin Gin Ser Giy Ala Giu Leu Ala Lys Pro Giy Ala I Cu Val Lys Mei Ser Cys Lys Ala Ser Giy Tyr Thr Phe Thr Ser Tyr 20

Arg Mei His Trp Val Lys Gin Arg Pro Giy Gin Giy Leu Giu Trp lle 35

Giy Tyr Ile Asn Pro Ser Thr Giu Tyr Asn Gin Lys Phe 50

Lys Asp Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Ser Thr Ala Tyr 70

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Met Gln Leu Ser Ser Leu Thr Phe Glu Asp Ser Ala Val Tyr Tyr Cys
85 90 95
       Thr Val Ser Ser
                  1 1 5
(2) INFORMATION FOR SEQ ID NG:4:
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              ( B ) TYPE: amino acid
              ( C ) STRANDEDNESS: single
              ( D ) TOPOLOGY: unknown
     ( i i ) MOLECULE TYPE: protein
    ( i i i ) HYPOTHETICAL; NO
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              ( A ) NAME KEY: Protein
              ( B ) LOCATION: 1..117
              ( D ) OTHER INFORMATION: /note="Variable region of the human
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      ( x i ) SEQUENCE DESCRIPTION; $EQID NG:4:
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1 10 15
       Ser Val Lys Val Ser Cys Lys Ala Ser Gly Gly Thr Phe Ser Arg Ser 20 25 30
      Ala lie lie Trp Val Arg Gin Ala Pro Gly Gin Gly Leu Giu Trp Mei 3.5\,
      Gly Gly lie Val Pro Met Phe Gly Pro Pro Asn Tyr Ala Gla Lys Phe
50 55
      Gin Gly Arg Val Thr lic Thr Ala Asp Glu Scr Thr Asm Thr Ala Tyr
65 70 75 80
       Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Phe Tyr Phe Cys
85 90
       Ala Gly Gly Tyr Gly lle Tyr Ser Pro Glu Glu Tyr Asn Gly Gly Leu 100 105
       Val Thr Val Scr Scr
                  115
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       ( i ) SEQUENCE CHARACTERISTICS:
              ( A ) LENGTH: 116 amine acids
              ( B ) TYPE: amino acid
              ( C ) STRANDEDNESS: single
              ( D ) TOPOLOGY: unknown
      ( i i ) MOLECULE TYPE: protein
    ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
              ( A ) NAME/KEY: Protein
              ( D ) OTHER INFORMATION: /nnte="Variable region of the PDL
                     bumani wed anti-The antibody heavy chain."
      ( x i ) SEQUENCE DESCRIPTION: SEQ 1D NO:5:
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       Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser Tyr
```

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-continued

20
25
30

Arg Met His Trp Val Arg Gln Ala Pro Gly Gln Gly Leu Glu Trp Ile
35

Gly Tyr lie Asn Pro Ser Thr Gly Tyr Thr Glu Tyr Asn Gln Lys Phe
50

Lys Asp Lys Ala Thr lle Thr Ala Asp Glu Ser Thr Asn Thr Ala Tyr
65

Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val Tyr Tyr Cys
85

Ala Arg Gly Gly Gly Val Phe Asp Tyr Trp Gly Gln Gly Thr Leu Val
100

Thr Val Ser Ser

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAMERCEY: Protein
 - (B) LOCATION: 1..116
 - (D) OTHER INFORMATION: /nute="Variable region of the CDR only humanized anti-Tac antibody beavy chain."
- (x i) SEQUENCE DESCRIPTION: SEQ 1D NO.6

Gla Val Gla Leu Val Gla Ser Gly Ala Giu Val Lys Lys Pro Gly Ser los Ser Val Lys Val Lys Pro Gly Ser Tyr 20 Val Lys Val Ser Cys Lys Ala Ser Gly Gly Thr Phe Ser Ser Tyr 30 Arg Met His Trp Val Arg Gla Ala Pro Gly Glo Gly Leu Gla Trp Met 45 Gly Tyr 11e Asa Pro Ser Thr Glu Tyr Asa Gla Lys Phe 50 Asp Arg Val Thr Ile Thr Ala Asp Glu Asp Glu Ser Thr Asa Thr Ala Tyr 80 Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Phe Tyr Phe Cys 95 Ala Gly Gly Gly Gly Gly Gly Clu Val Thr Val Ser Ser

(2) IN FORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
- (i x)FEATURE:
 - (A) NAME/KEY: Protein

75

(B) LOCATION: 1..106

(D) OTHER INFORMATION: /note="Variable region of the PDL burnarized anti-Tec antibody light chain."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:7:

 Asp Ile Glu Met Thr Glu Ser Pro Ser Thr Leu Ser Alo Ser Val Gly 15

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 Thr Thr Ser Asu Leu Ala Ser Gly Val Pro Ala Arg Phe Ser Gly Ser 50

 Gly Ser Gly Thr Glu Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro Asp 65

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 Phe Gly Gln Gly Thr Lys Val Glu Val Lys 105

(2) INFORMATION FOR SEQ ID NG:8:

- (i) SEQUENCE CHARACIERISTICS:
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 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: pioter'n
- (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAMEREY: Protein
 - (B) LOCATION: 1..106
 - (D) OTHER INFORMATION: /note="Variable region of the CDR-only bumanised anti-Tac antibody light chain."
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:8:

 Asp II c
 GIn Mei Thr Scr Gln Ser Pro Ser Thr Lew Ser Aiz Ser Val Gly IS

 Asp Arg Val Thr II c
 Thr Cys Ser Ala Ser Ser Ser Ser II c
 Ser Tyr Mei 30

 His Trp Tyr 35
 Gln Gln Lys Pro Gly Lys Ala Pro Lys Lew Lew Mei Tyr 45

 Thr Thr Ser Asn Lew Ala Ser Gly Val Pro Ser Arg Phe II c Gly Ser 55

 Gly Ser Gly Thr Glu Pbc Thr Lew Thr II c Ser Ser Lew Gln Pro Asp 80

 Asp Pbc Ala Thr Tyr Tyr Cys His Gln Arg Ser Thr Tyr Pro Lew Ps

 Pbc Gly Gln Gly Thr Lys Val Glu Val Lys II c

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACIERISTICS:
 - (A) LENG'IH: 443 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: DNA

5,530,101 77 78 -continued (i i i) HYPOTHETICAL: NO (i x) FEATURE (A) NAME/KEY: misc_feature (B) LOCATION: 1.443 (D) OTHER INFORMATION: /note="Sequence encoding heavy chain variable region of CDR-unly humanized anti-Tac anybody including signal sequence." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:9: AGCTTCTAGA TGGGATGGAG CTGGATCTTT CTCTTCCTCC TGTCAGGTAC CGCGGGCGTG CACTCTCAGG TCCAGCTTGT CCAGTCTGGG GCTGAAGTCA AGAAACCTGG CTCGAGCGTG AAGGTCTCCT GCAAGGCTTC TGGCGGGACC TTTTCTAGCT ACAGGATGCA CTGGGTAAGG CAGGCCCCTG GACAGGGTCT GGAATGGATG GGATATATTA ATCCGTCGAC TGGGTATACT GAATACAATC AGAAGTTCAA GGACAGGGTC ACAATTACTG CAGACGAATC CACCAATACA GCCTACATGG AACTGAGCAG CCTGAGATCT GAGGACACCG CATTCTATTT CTGTGCAGGG GGTGGGGGAG TCTTTGACTA CGAATACAAT GGAGGGCTGG TCACAGTCTC CTCAGGTGAG TCCTTAAAAC CTCTAGACGA TAT (2) INFORMATION FOR SEQ ID NO:10: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 411 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS. single (D) TOPOLOGY: linear

- (i i) MOLECULE TYPE: DNA
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: misc_fcature
 - (B) LOCATION: 1..411
 - (D) OTHER INFORMATION: Tooc="Sequence eccoding light chain variable region of the CDR-only humanized anti-Tac antibody including signal sequence."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CAAATCTAGA TGGAGACCGA TACCCTCCTG CTATGGGTCC TCCTGCTATG GGTCCCAGGA 60
TCAACCGGAG ATATTCAGAT GACCCAGTCT CCATCTACCC TCTCTGCTAG CGTCGGGGAT 120
AGGGTCACCA TAACCTGCTC TGCCAGCTCA AGTATAAGTT ACATGCACTG GTACCAGCAG 180
AAGCCAGGCA AAGCTCCCAA GCTTCTAATG TATACCACAT CCAACCTGGC TTCTGGAGTC 240
CCTTCTCGCT TCATTGGCAG TGGATCTGGG ACCGAGTTCA CCCTCACAAT CAGCTCTCTG 300
CAGCCAGATG ATTTCGCCAC TTATTACTGC CATCAAAGGA GTACTTACCC ACTCACGTTC 360
GGTCAGGGGA CCAAGGTGGA GGTCAAACGT AAGTACACTT TTCTAGATAT A 411

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: DNA
- (i i i) HYPOTHEDCAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: misc_(cature
 - (B) LOCATION: 1..29
 - (D) OTHER INFORMATION: /standard_name="Primer me(145"

6 0

180

2 4 0

360

4 2 0

443

79 80

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( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:11:
TAATCTAGAA TTCCCCCCC CCCCCCCC
                                                                                                                 29
( 2 ) INFORMATION FOR SEQ ID NO:12:
        ( i ) SEQUENCE CHARACIERISTICS:
                ( A ) IENGTH; 46 besc peres
                ( B ) TYPE: meleic acid
( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: DNA
     ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: misc_feature
                ( B ) LOCATION: 1_46
                ( D ) OTHER INFORMATION: /standard_name="Primer roc045"
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:12:
TATAGAGCTC AAGCTTGGAT GGTGGGAAGA TGGATACAGT TGGTGC
                                                                                                                 4 6
( 2 ) INFORMATION FOR SEQ ID NO:13:
        ( i ) SEQUENCE CHARACTERETICS:
                ( A ) LENGTH: 50 base pairs
                ( B ) TYPE: nucleic acid
                ( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: DNA
     ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAMEREY: misc_leature
                ( B ) LOCATION: 1_50
                ( D ) OTHER INFORMATION: /standard_name="Primer mcG47"
      (x i) SEQUENCE DESCRIPTION: SEQ ID NO:13:
TATAGAGCTC AAGCTTCCAG TGGATAGACH GATGGGGSTG TYGTTTTGGC
( 2 ) INFORMATION FOR SEQ ID NO:14:
        ( i ) SEQUENCE CHARACIERISTICS:
                ( A ) LENGTH: 116 amino acids
                ( B ) TYPE: amino acid
                ( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: unknown
      ( i i ) MOLECULE TYPE: protein
     ( i i i ) HYPO THETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: Protein
                ( B ) LOCATION: 1..116
                ( D ) OTHER INFORMATION: /notc="Anti-fac heavy chain amino
                        acid sequence."
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:14:
       Gin Val Gin Leu Gin Gin Ser Giy Ala Giu Leu Ala Lys Pro Giy Ala
1 10 15
        Ser Val Lys Met Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser Tyr
20 25
        Arg Met His Trp Val Lys Gla Arg Pro Gly Gla Gly Leu Glu Trp Ile
35 40
        Gly Tyr lic Asn Pro Ser Thr Gly Tyr Thr Glu Tyr Asn Gln Lys Phe
```

-continued Lys Asp Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Ser Thr Ala Tyr 65 70 75 80 Ala Arg Gly Gly Val Phe Asp Tyr Trp Gly Glo Gly Thr Thr Lcu 100 110 (2) INFORMATION FOR SEQ ID NO:15: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 117 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: wiknown (i i) MOLECULE TYPE: protein (i | i) HYPUTHETICAL: NO (i x)FEATURE: (A) NAME/KEY: Protein (B) LOCATION: 1..117 (D) OTHER INFORMATION: /nou="Eu heavy chain amino acid sequence." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:15: Gln Val Gin Leu Val Gin Ser Gly Ala Glu Val Lys Lys Pro Gly Ser 1 10 15 Alm lie lie Trp Val Arg Glm Ala Pro Gly Glm Gly Leu Glu Trp Met $3.5\,$ Gly Gly Ile Val Pro Mei Phe Gly Pro Pro Asa Tyr Ala Gla Lys Phe 50 55 Gln Gly Arg Vəl Thr IIc Thr Ala Asp Glu Ser Thr Asn Thr Ala Tyr 65 70 75 80 Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Phe Tyr Phe Cys 85 90 95 Ala Gly Gly Tyr Gly 11c Tyr Ser Pro Glu Giu Tyr Asa Gly Gly Leu 100 105 Val Thr Val Scr Scr 1 1 5 (2) INFORMATION FOR SEQ ID NO:16: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 106 aroino acids (B) TYPE amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: unknown (i i) MOLDCULE TYPE- protein (I i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: Proucin (B) LOCATION: 1..106 (D) OTHER INFORMATION: /neic="Anti-Tac light chain amino acid sequence."

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:16:

83

Gin lle Val Leu Thr Gln Ser Pro Ala lie Met Ser Ala Ser Pro Gly 1 $$ Glu Lys Val Thr lle Thr Cys Ser Ala Ser Ser Ser lle Ser Tyr Met 20 0 25His Trp Pbc Gln Gln Lys Pro Gly Tbr Ser Pro Lys Leu Trp lle Tyr 35 40 Thr Thr Scr Asn Leu Ala Scr Gly Val Pro Ala Arg Phc Scr Gly Scr 50 Gly Ser Gly Thr Ser Tyr Ser Leu Thr lle Ser Arg Mei Glu Ala Glu 65 70 80 Asp Ala Ala Thr Tyr Tyr Cys His Gln Arg Scr Thr Tyr Pro Leu Thr 85 90 95 Pbc Gly Scr Gly Thr Lys Lcu Glu Lcu Lys 100 105

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..107
 - (D) OTHER INFORMATION: /note=Eu light chain amine acid
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Asp lle Glm Met Thr Glm Ser Pro Ser Thr Leu Ser Ala Ser Val Gly 1 $$ Asp Arg Val Thr Ilc Thr Cys Arg Ala Scr Gln Scr 11c Asn Thr Trp 20 25 Leu Ala Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Mei $3.5\,$ Tyr Lys Ala Ser Ser Leu Glu Ser Gly Val Pro Ser Arg Phe 11c Gly 50 Ser Gly Ser Gly Thr Glu Phe Thr Leu Thr lie Ser Ser Leu Gln Pro 6.5Asp Asp Phc Ala Thr Tyr Tyr Cys Gln Gln Tyr Asn Scr Asp Scr Lys 85 90 95 Mci Phe Gly Gln Gly Thr Lys Val Glu Val Lys
100

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEOURNCE CHARACTERISTICS:
 - (A) LENGTH: 433 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOTECULE TYPE: DNA
- (i i i) HYPOTHETICAL: NO
- (i a) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 6.410
 - (D) OTHER INFORMATION: /procluct="Humanized anti-Tac beavy

86

85

chain variable region. Seq 1D. 19" (x i) SEQUENCE DESCRIPTION: SEQ ID NO:18: TCTAG ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGT ACC 47 Mel Gly Trp Scr Trp 11c Pbc Lcu Pbc Lcu Leu Scr Gly Thr
1 5 GCG GGC GTG CAC TCT CAG GTC CAG CTT GTC CAG TCT GGG GCT GAA GTC Ala Gly Val His Scr Glz Val Gln Lcu Val Gln Scr Gly Ala Glu Val 15 9 5 AAG AAA CCT GGC TCG AGC GTG AAG GTC TCC TGC AAG GCT TCT GGC TAC Lys Lys Pro Gly Scr Scr Val Lys Val Scr Cys Lys Ala Scr Gly Tyr 35 40 143 ACC TTT ACT AGC TAC AGG ATG CAC TGG GTA AGG CAG GCC CCT GGA CAG
Thr Phc Thr Scr Tyr Arg Mct His Trp Val Arg Gln Ala Pro Gly Gln
50 60 GGT CTG GAA TGG ATT GGA TAT ATT AAT CCG TCG ACT GGG TAT ACT GAA Gly Lou Glu Trp Ilo Gly Tyr llo Asm Pro Scr Thr Gly Tyr Thr Glu TAC AAT CAG AAG TTC AAG GAC AAG GCA ACA ATT ACT GCA GAC GAA TCC
Tyr Asn Gin Lys Phe Lys Asp Lys Ala Thr lic Thr Ala Asp Glu Scr
80 ACC AAT ACA GCC TAC ATG GAA CTG AGC AGC CTG AGA TCT GAG GAC ACC
Thr Asn Thr Ala Tyr Mei Glu Leu Scr Scr Leu Arg Scr Glu Asp Thr
95 3 3 5 GCA GTC TAT TAC TGT GCA AGA GGG GGG GGG GTC TTT GAC TAC TGG GGC Ala Val Tyr Tyr Cys Ala Arg Gly Gly Gly Val Pbc Asp Tyr Trp Gly 115 383 CAA GGA ACC CTG GTC ACA GTC TCC TCA GGTGAGTCCT TAAAACCTCT Gin Gly Thr Lou Val Thr Val Scr Scr

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 135 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:19:

 Mcl Gly Trp Ser Trp 5
 Ile Pbc Leu Pbc Leu Leu Leu Ser Gly Tbr Ala Gly 15

 Val His Ser Glu Val Gln Leu Val Gln Ser Gly Ala Glu Vol Lys 25

 Pro Gly Ser Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Pbc 35

 Thr Ser Tyr Arg Met His Trp S5
 Val Arg Glu Ala Pro Gly Gly Gln Gly Leu 60

 Glu Trp Ile Gly Tyr Ile Asn Pro Ser Tbr Gly Tyr Thr Glu Tyr Asn 65

 Gln Lys Pbc Lys Asp Lys Ala Tbr Ile Tbr Ala Asp Glu Asp Glu Ser Thr Asn 95

 Thr Ala Tyr Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Tyr Trp Ala Val 100

 Tyr Tyr Cys Ala Arg Gly Gly Gly Gly Val Pbc Asp Tyr Trp Gly Gln Gly Gln Gly 125

 Tbr Leu Val Tbr Val Ser Ser 135

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( 2 ) INFORMATION FOR SEQ ID NO:20:
       ( i ) SEQUENCE CHARACTERISTICS:
               ( A ) LENGIH: 403 base pairs
               ( B ) TYPE: nucleic acid
               ( C ) STRANDEDNESS: single
               ( D ) TOPOLOGY: linear
     ( i i ) MOLECULE TYPE: DNA
    ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
               ( A ) NAME/KEY: CDS
               ( B ) LOCATION: 6.383
               ( D ) OTHER INFORMATION: /product="Humanized anti-Tac light
                      chain variable region: Seq ID. 21"
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:20;
TCTAG ATG GAG ACC GAT ACC CTC CTG CTA TGG GTC CTC CTG CTA TGG
Mci Glu Thr Asp Tbr Lcu Lcu Lcu Trp Val Lcu Lcu Lcu Trp
                                                                                                        47
GTC CCA GGA TCA ACC GGA GAT ATT CAG ATG ACC CAG TCT CCA TCT ACC
Val Prm Gly Ser Tbr Gly Asp IIe Gln Mel Tbr Gln Ser Pro Ser Thr

15
CTC TCT GCT AGC GTC GGG GAT AGG GTC ACC ATA ACC TGC TCT GCC AGC
                                                                                                       1 4 3
Leu Ser Ala Ser Val Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
TCA AGT ATA AGT TAC ATG CAC TGG TAC CAG CAG AAG CCA GGC AAA GCT
Ser Ser lle Ser Tyr Mel His Trp Tyr Gln Gln Lys Pro Gly Lys Ala
                                                                                                        191
CCC AAG CTT CTA ATT TAT ACC ACA TCC AAC CTG GCT TCT GGA GTC CCT Pro Lys Leu Leu IIc Tyr Thr Thr Scr Asn Leu Ala Scr Gly Val Pro
                                                                                                       239
GCT CGC TTC AGT GGC AGT GGA TCT GGG ACC GAG TTC ACC CTC ACA ATC Ala Arg Pbc Scr Gly Scr Gly Thr Glu Pbc Thr Lcu Tbr Ilc 80 90
                                                                                                       287
AGC TOT CTG CAG CCA GAT GAT TTC GCC ACT TAT TAC TGC CAT CAA AGG
                                                                                                       3 3 5
Ser Ser Leu Gln Pro Asp Asp Phe Ala Thr Tyr Tyr Cys His Gln Arg
AGT ACT TAC CCA CTC ACG TTC GGT CAG GGG ACC AAG GTG GAG GTC AAA
                                                                                                       383
Ser Thr Tyr Pro Leu Thr Phe Gly Gln Gly Thr Lys Val Glu Val Ly's
CGTAAGTACA CTTTTCTAGA
                                                                                                        403
( 2 ) INFORMATION FOR SEO ID NO:21:
       ( i ) SEQUENCE CHARACIERISTICS:
               ( A ) LENGTH: 126 amino acids
               ( B ) TYPE: amino acid
               ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: protein
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:21:
Mei Glu Thr Asp Thr Leu Leu Leu Trp Val Leu Leu Trp Val Pro
I 10
Gly Ser Thr Gly Asp IIe Gin Met Thr Gln Ser Pro Ser Thr Leu Ser 20 25 30
Ala Scr Val Gly Asp Arg Val Thr Itc Thr Cys Scr Ata Scr Scr Scr 35
lic Scr Tyr Mct His Trp Tyr Gln Gin Lys Pro Gly Lys Ala Pro Lys
50 60
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-continued
Leu Leu Ile Tyr Thr Thr Ser Asu Leu Ala Ser Gly Val Pro Ala Arg
 6 5
                                7 0
                              Ser Gly The Glu Phc The Leu The
                  Ser Gly
                               Phe Ala Thr Tyr Tyr Cys His Gln Arg
105
                              Gly Gln Gly The Lys Val Glu Val Lys
( 2 ) INFORMATION FOR SEQ ID NO:22:
        ( i ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 126 base pairs
                ( B ) TYPE: nucleic acid
                ( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: DNA
     ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: misc_[cause
                ( B ) LOCATION: 1..126
                (D) OTHER INFORMATION: /standard_name="Oligo (IES12"
                       / note='One of four oligonuclentides used to
                       synthetize the humanized anti-Tac heavy chain
                       gene."
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:22:
AGCTTCTAGA TGGGATGGAG CTGGATCTTT CTCTTCCTCC TGTCAGGTAC CGCGGGCGTG
CACTCTCAGG TCCAGCTTGT CCAGTCTGGG GCTGAAGTCA AGAAACCTGG CTCGAGCGTG
                                                                                                            120
AAGGTC
( 2 ) INFORMATION FOR SEQ ID NO:23:
        ( i ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 129 base pairs
                ( B ) TYPE: nucleic acid
                ( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: DNA
     ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: misc_fcature
                ( B ) LOCATION: 1..129
                ( D ) OTHER INFORMATION: /standard_name="Oligo HES13"
                        / note="One of four oligonucleotides used to
                        synthesize he humanized anti-Tac heavy chain
                        gene."
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:23:
CCCAGTCGAC GGATTAATAT ATCCAATCCA TTCCAGACCC TGTCCAGGGG CCTGCCTTAC
                                                                                                              60
CCAGTGCATC CTGTAGCTAG TAAAGGTGTA GCCAGAAGCC TTGCAGGAGA CCTTCACGCT
                                                                                                            120
CGAGCCAGG
                                                                                                            129
( 2 ) INFORMATION FOR SEQ ID NO:24:
        ( i ) SEQUENCE CHARACIERISTICS:
                ( A ) LENGTH: 124 base pairs
                ( B ) TYPE: nucle'ie acid
                ( C ) STRANDEDNESS: .ingle
                ( D ) TOPOLOGY: linear
```

(i i) MOLECULE TYPE: DNA

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( i i i ) HYPOTHETTCAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: misc_fcature
                ( B ) LOCATION: 1..124
                ( D ) OTHER INFORMATION: /standard_name="Origo HES14"
                         / note="One of four oligonucleotides used to
                         synthesize the humanized anti-Tae heavy chain
                         genc."
      ( x i ) SEQUENCE DESCRIPTION; SEQ ID NO:24:
TATATTAATC CGTCGACTGG GTATACTGAA TACAATCAGA AGTTCAAGGA CAAGGCAACA
                                                                                                                  60
ATTACTGCAG ACGAATCCAC CAATACAGCC TACATGGAAC TGAGCAGCCT GAGATCTGAG
                                                                                                                120
GACA
                                                                                                                124
( 2 ) INFORMATION FOR SEO ID NO:25:
        ( i ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 128 base pairs
                ( B ) TYPE: nucleic acid
                ( C ) STRANDEDNESS: stingle
                ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE; DNA
     ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
                ( A ) NAME/KEY: mise_feature
                ( B ) LOCATION: 1..128
                ( D ) OTHER INFORMATION: /standard_name="Oligo HESI5"
                         / note="One of four oligonucleorides used to
                         synthesize the burnanized anti-Tac heavy chain
                         genc."
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:25:
ATATCGTCTA GAGGTTTTAA GGACTCACCT GAGGAGACTG TGACCAGGGT TCCTTGGCCC
                                                                                                                  60
CAGTAGTCAA AGACCCCCCC CCCTCTTGCA CAGTAATAGA CTGCGGTGTC CTCAGATCTC
                                                                                                                120
                                                                                                                128
AGGCTGCT
( 2 ) INFORMATION FOR SEQ ID NO:26:
        ( i ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 120 base pairs
                 ( B ) TYPE: nucle ic acid
                 ( C ) STRANDEDNESS: single
                ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: DNA
     ( i i i ) HYPOTRETICAL. NO
      ( i x ) FEATURE:
                 ( A ) NAME/KEY: misc_feature
                 ( B ) LOCATION: 1.120
                 ( D ) OTHER INFORMATION: /standard_name="Oligo JFDI"
                         / note="One of four oligonuchotides used to
                         synthesize the humanized anti-Tue light chain
                         gene."
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:26:
CAAATCTAGA TGGAGACCGA TACCCTCCTG CTATGGGTCC TCCTGCTATG GGTCCCAGGA
                                                                                                                  60
TCAACCGGAG ATATTCAGAT GACCCAGTCT CCATCTACCC TCTCTGCTAG CGTCGGGGAT
                                                                                                                120
( 2 ) INFORMATION FOR SEQ ID NO:27:
        ( i ) SEQUENCE CHARACTERISTICS:
                ( A ) LENGTH: 114 base pairs
```

-continued (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1..114 () OTHER INFORMATION: /sundard_none"Oligo JFD2" / note="One of four oligonucleotides used to synthesize the humanized anti-Tac light chain gene." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:27: ATAAATTAGA AGCTTGGGAG CTTTGCCTGG CTTCTGCTGG TACCAGTGCA TGTAACTTAT 6 0 ACTTGAGCTG GCAGAGCAGG TTATGGTGAC CCTATCCCCG ACGCTAGCAG AGAG 114 (2) INFORMATION FOR SEQ ID NO:28: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 123 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHETICAL: NO (i x) FEATURE: (A) NAME/KEY: m'isc feature (B) LOCATION: L.123 (D) OTHER INFORMATION: /standard_namc="O'igo JFD3" / note="One of four of igonucleotides used to synthesize the humanized anti-Tac light chain genc." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:28: GCTCCCAAGC TTCTAATTTA TACCACATCC AACCTGGCTT CTGGAGTCCC TGCTCGCTTC 60 AGTGGCAGTG GATCTGGGAC CGAGTTCACC CTCACAATCA GCTCTCTGCA GCCAGATGAT 120 TTC 123 (2) INFORMATION FOR SEQ ID NO:29: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 122 base pairs (B) TYPE: pucieic acid (C) SIRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (i i i) HYPOTHED CAL: NO (i x) FEATURE: (A) NAME/KeY: misc_feature (B) LOCATION: 1..122 (D) OTHER INFORMATION: /standard_name="Oligo JFD4" / note-'One of four oligonucleatides used to synthesize the humanized anti-Tac light chain gcac." (x i) SEQUENCE DESCRIPTION: SEQ ID NO:29:

TATATCTAGA AAAGTGTACT TACGTTTGAC CTCCACCTTG GTCCCCTGAC CGAACGTGAG

TGGGTAAGTA CTCCTTTGAT GGCAGTAATA AOTGGCGAAA TCATCTGGCT GCAGAGAGCT

GA

6 0

120

122

(2) INTORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH; 384 base pairs
 - (B) TYPE: nuclei'c acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: cDNA
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:

 - (A) NAME/KEY: CDS (B) LOCATION: 1..384
 - (D) OTHER INFORMATION: /product="Light chain variable negrou of mik-betal; Seq ID No. 31"
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:30.

ATG	GAT	TTT	CAA	GTG	CAG	ATT	TTC	AGC	TTC	CTG	СТА	АТС	AGT	GCC	TCA	4 8
McI	Asp	Phe	GIn	V a l	Gln	1 1 c	Phc	Scr	Phc	Lcu	Lcu	1 lc	Scr	Ala	Scr	
1				5					1 0					I 5		
GTC	ATA	CTG	TCC	AGA	GGA	CAA	АТТ	GTT	CTC	ACC	CAG	TCT	CCA	GCA	ATC	9 6
Val	I 1 c	Lcu	Scr	Arg	Gly	G 1 n	Ilc	V a J	Lcu	The	G 1 n	Ser	Pro	Ala	1 1 c	
			2 0					2 5					3 0			
ATG	TCT	GCG	тст	CCA	GGG	GAG	AAG	GTC	ACC	ATG	ACC	TGC	AGT	GGC	AGC	1 4 4
Mcl	Scr	Ala	Scr	Pro	Gly	Glu	Lys	Val	Thr	Mc 1	Thr	Cys	Scr	Gly	Scr	
		3 5					4 0					4 5				
TCA	AGT	GTA	AGT	TTC	ATG	TAC	TGG	TAC	CAG	CAG	AGG	CCA	GGA	TCC	TCC	192
Scr	Ser	V a 1	Scr	Pbc	Mc 1	Tyr	Trp	Туг	Gln	G 1 n	Arg	Pro	G 1 y	Scr	Scr	
	5 0					5 5					6 0					
CCC	AGA	CTC	CTG	ATT	TAT	GAC	ACA	TCC	AAC	CTG	GCT	TCT	GGA	GTC	CCT	2 4 0
Pro	Arg	Lcu	Lcu	1 1 c	Tyr	Asp	Thr	Scr	Asn	Lcu	Alg	Scr	Gly	Val	Pro	
6 5					7 0					7 5					8 0	
GTT	CGC	TTC	AGT	GGC	AGT	GGG	TCT	GGG	ACC	TCT	TAC	TCT	CTC	ACA	ATC	288
V a 1	Arg	Phc	Scr	G 1 y	Scr	G 1 y	Scr	Gly	Thr	Scr	Туг	Scr	Lcu	Tbr	1 1 c	
				8 5					9 0					9 5		
AGC	CGA	ATG	GAG	GCT	GAA	GAT	GCT	GCC	ACT	TAT	TAC	TGC	CAG	CAG	TGG	3 3 6
Scr	Arg	McI	Glu	Ala	Glu	Asp	Ala	Ala	Thr	Туг	Tyr	Cys	GIn	GIn	Trp	
			i 0 0					I 0 5					110			
A G T	ACT	TAC	CCG	CTC	ACG	TTC	GOT	GCT	GGG	ACC	AAG	CTG	GAG	CTG	AAA	3 8 4
Scr	Thr	Tyr	Pro	Lcu	Thr	Phc	G 1 y	A 1 a	Gly	Thr	Lys	Lcu	G 1 u	Lcu	Lys	
		115					120					1 2 5				

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 128 amino acids (B) TYPE: amino acid

 - (D) TOPOLOGY: Timear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION; SEQ ID NO:31:

Mc I	Asp	РЬс	Gin	Val (Sin ilc	Phc So	r Phe Lei	u Leu IIe	Ser	Ala Scr 15	
V a]	I 1 c	Lcu	S c r 2 0	Arg (Gly Gln	11c V:	al Leu Th	г Сін Ѕст	Pro .	Ala ile	
Mcl	Scr	A 1 a 3 5	Scr	Рго (Gly Glu	Lys Va 40	al Thr Me	The Cys	Scr	Gly Scr	
Scr	S c r 5 0	Val	Ser	Phc A	Met Tyr	Тгр Ту	r Gin Gi	n Arg Pro 60	G 1 y	Ser Ser	
Pro 65	Arg	L c' u	L c u	Ilc 7	Tyr Asp	Thr Sc	er Asn Lei	Ala Scr	G 1 y	Val Pro	

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-continued
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 Val
 Arg
 Phc
 Scr
 Gly
 Scr
 Gly
 Scr
 Gly
 Scr
 Gly
 Thr
 Scr
 Tyr
 Scr
 Lcu
 Thr
 11c
 95

 Scr
 Arg
 Mci
 Glu
 Ala
 Glu
 Asp
 Ala
 Ala
 Thr
 Tyr
 Tyr
 Cys
 Glu
 Glu
 Trp

 Scr
 Thr
 Tyr
 Pro
 Lcu
 Thr
 Phc
 Gly
 Ala
 Gly
 Thr
 Lys
 Leu
 Glu
 Lcu
 Lys

 115
 120
 120
 125
 125
 125
 125

(2) INFORMATION FORSEQID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 414 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS; single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: cDNA
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1.414
 - (D) ■THER INFORMATION: /product="Heavy chain var. region of the antibody mik-betal: SeqID 33"
- (* i) SEQUENCE DESCRIPTION: SEQ ID NO:32:

ATG GCT GTC TTG GGG CTG CTC TTC TGC CTG GTG ACA TTC CCA AGC TGT 48 Met Ala Val Leu Gly Leu Leu Phe Cys Leu Val The Phe Pro Ser Cys GTC CTA TCC CAG GTG CAG CTG AAG CAG TCA GGA CCT GGC CTA GTG CAG Val Lcu Ser Gln Val Gln Lcu Lys Gln Ser Gly Pro Gly Lcu Val Gln CCC TCA CAG AGC CTG TCC ATC ACC TGC ACA GTC TCT GGT TTC TCA GTA Pro Ser Gln Ser Leu Ser Ile Thr Cys Thr Val Ser Gly Phe Ser Val ACA AGT TAT GGT GTA CAC TGG ATT CGC CAG TCT CCA GGA AAG GGT CTG The Scr Tyr Gly Val His Trp Ile Arg Gln Scr Pro Gly Lys Gly Leu GAG TGG CTG GGA GTG ATA TGG AGT GGT GGA AGC ACA GAC TAT AAT GCA 240 Glu Trp Leu Gly Val Ilc Trp Ser Gly Gly Ser Thr Asp Tyr Asn Ala GCT TTC ATA TCC AGA CTG ACC ATC AGC AAG GAC AAC TCC AAG AGC CAA 288 Ala Pho Ilo Sor Arg Lou Thr 11c Sor Lys Asp Asn Sor Lys Sor Gln GTT TTC TTT AAA GTG AAC AGT CTG CAA CCT GCT GAC ACA GCC ATA TAC 3 3 6 Val Phe Phe Lys Val Asa Ser Leu Gln Pro Ala Asp Thr Ala lle Tyr TAT TGT GCC AGA GCT GGG GAC TAT AAT TAC GAC GGT TTT GCT TAC TGG
Tyr Cys Ala Arg Ala Gly Asp Tyr Ash Tyr Asp Gly Phe Ala Tyr Trp 384 GGC CAA GGG ACT CTG GTC ACT GTC TCT GCG 4 1 4 Gly Gln Gly Tbr Lcu Val Thr Val Scr Ala 1 3 5

(2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 138 amino acids
 - (B) TYPE; amino acid
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Mes Ala Val Leu Gly Leu Leu Phe Cys Leu Val Thr Phe Pro Scr Cys 1 10 15

```
        Val
        Lcu
        Scr
        Gln
        Val
        Gln
        Lcu
        Lys
        Gln
        Scr
        Gly
        Pro
        Gly
        Lcu
        Val
        Gln
        Scr
        Lcu
        Scr
        1lc
        Tbr
        Cys
        Tbr
        Val
        Scr
        Gly
        Phc
        Scr
        Val
        Fhc
        Scr
        Fhc
        Scr
        Val
        Fhc
        Scr
        Fhc
        Scr
        Val
        Fhc
        Scr
        Fhc
        Fhc</td
```

- (2) INFORMATION FOR SEQ ID NO:34:
 - (i) SEQUENCE CHARACIERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown
 - (i i) MOLECULE TYPE: probin
 - (i i i) HYPOTHETICAL: NO
 - (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..107
 - (D) OTHER DEFORMATION: /note="Amino acid sequence of the light chain for humane Lay antibody."
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:34:

 Asp 11c
 GIn Mct 55
 Thr 55
 Gin Ser Pro 5cr Ser 10
 Leu Ser Val Ser Val Ser Val Gly
 Ser Val Gly
 Gly
 Gly
 Ser Ser 10
 Leu Ser Val Ser Val Ser Val Gly
 Gly
 Gly
 Asn Val Asn Ala Tyr

 Asp Arg Val Thr 20
 Thr Gly
 Gln Ala Ser Gly
 Gln Ala Ser Gly
 Ala Pro Lys Leu Leu Ile
 Leu Leu Ile

 Leu Asn Trp 35
 Tyr Gly
 Ala Ser Thr Arg Glu Ala Gly Val Pro Ser Arg Phe Ser Gly
 Ser Arg Phe Ser Gly
 Ser Gly
 Ser Gly

 Ser Gly
 Ser Gly
 Thr Asp Phe Thr Phe Thr Ile Ser Ser Leu Gln Pro 80
 Fro 80
 Fro 80
 Fro 80

 Thr Phe Gly
 Gln Gly Thr Lys Val Glu Val Lys 105
 Val Lys 105
 Fro 95
 Fro 95
 Fro 95

- (2) INFORMATION FOR SEQ ID NO:35:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) IENGTH: 106 amino acids (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOOY: unknown
 - (i i) MOLECULE TYPE: protei's
 - (i i i) HYPOTHETICAL: NO

(i x) FEATURE:

- (A) NAME/KEY: Procin
- (B) LOCATION: 1_106
- (D) OTHER INFORMATION: /note="Amima acid sequence of the light chain of the humanized mik-betal antibody."
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:35:

Asp IIc Gln Mct Thr Gln Scr Pro Scr Scr Leu Scr Ala Scr Val Gly
I 5 10 Asp Arg Val Thr IIe Thr Cys Scr Gly Ser Scr Scr Val Scr Phc Mci 20 25 Tyr Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Ile Tyr 35 40 45 Asp Thr Ser Asn Leu Ala Ser Gly Val Pro Ser Arg Phe Ser Gly Ser 50 55 Gly Scr Gly Thr Asp Tyr Thr Phe Thr IIe Scr Scr Leu Gln Pro Glü 65 70 75lle Ala Thr Tyr Tyr Cys Gla Gln Trp Scr Thr Tyr Pro Leu Thr 85 90 95 Pbc Gly Gln Gly Thr Lys Val Glu Val Lys 100

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACIERISTICS:
 - (A) LENGTH: 122 amino acids
 - (B) TYPE; amino acid
 - (C) STRANDEDNESS; single
 - (D) TOPOLOGY: unknown
- (i i) MOLECULE TYPE: protein
- (i i i) HYPOTHETICAL: NO
- (i x) FEATURE:
 - (A) NAME/KEY: Protein
 - (B) LOCATION: 1..122
 - (D) OTHER INFORMATION: Indic="Amino acid sequence of he heavy chain of the human Lay antibody."
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Val Gle Leu Clu Ser Gly Gly Gly Leu Val Gle Pro Gly Gly -5 Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Ala Ser 20 25 30 Ala Met Ser Trp Val Arg Glu Ala Pro Gly Lys Gly Leu Glu Trp Val 35 40 45 Ala Trp Lys Tyr Glu Asn Gly Asn Asp Lys His Tyr Ala Asp Scr Val 50 60 Asn Gly Arg Pac Thr IIc Ser Arg Asn Asp Ser Lys Asn Thr Leu Tyr 65 70 75 80 Leu Gln Met Asn Gly Leu Gln Ala Glx Val Scr Ala lie Tyr Tyr Cys 85 90 95 Ala Arg Asp Ala Gly Pro Tyr Val Ser Pro Thr Phe Phe Ala His Trp Oly Glm Gly The Leu Val The Val Ser Ser 115

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: ammo acid

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-continued
               ( C ) STRANDEDNESS: single
               ( D ) TOPOLOGY: unknown
      ( i i ) MOLECULE TYPE: protein
    ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
               ( A ) NAME/KEY: Protein
( B ) LOCATION: 1..119
               ( D ) OTHER INFORMATION: /nove="Amino acid sequence of the
                      heavy chain of the humanized mik-betal antibody."
      ( x i ) SEQUENCE DESCRIPTION: SEQ 1D NO:37:
       Glu Val Gln Leu Leu Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly l 10 15
             Leu Arg Leu Ser Cys Ala Ala Ser Giy Phe Thr Val Thr Ser Tyr 20 30
       Gly Val His Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val 3.5\, 40 45
       Gly Val lle Trp Ser Gly Gly Ser Thr Asp Tyr Asn Ala Ala Phe Ile
50 55
       See Arg Phe Thr Ile Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr Leu
65 70 75 80
       Glm Mct Asn Scr Lcu Glm Ala Glu Asp Thr Ala 11c Tyr Tyr Cys Ala 85 90 95
       Arg Ala Gly Asp Tyr Asn Tyr Asp Gly Phe Ala Tyr Trp Gly Gln Gly 100
       The Lou Val The Val Scr Scr
( 2 ) INFORMATION FOR SEQ ID NO:38:
       ( i ) SEQUENCE CHARACTERISTICS:
               ( A ) LENGTH: 107 base pairs
               ( B ) TYPE: nucleic acid
               ( C ) STRANDEDNESS: single
               ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: DNA
    ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
               ( A ) NAME/KEY: misc_feature
               ( B ) LOCATION: 1...107
               ( D ) OTHER INFORMATION: /standard_name="Oligo ve13"
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID No:38:
TTCTGCTGGT ACCAGTACAT GAAACTTACA CTTGAGCTGC CACTGCAGGT GATGGTGACG
CGGTCACCCA CTGAGGCACT GAGGCTAGAT GGAGACTGGG TCATTTG
                                                                                                         107
( 2 ) INFORMATION FOR SEQ ID NO:39:
        ( i ) SEQUENCE CHARACTERISTICS:
               ( A ) LENGTH: 136 base pairs
               ( B ) TYPE: nucleic acid
               ( C ) STRANDEDN'ESS: single
               ( D ) TOPOLOGY: linea:
      ( i i ) MOLECULE TYPE: DNA
    ( i i i ) HYPOTHETICAL: NO
      ( i x ) FEATURE:
               ( A ) NAME/KEY: misc_feature
               ( B ) LOCATION: 1..136
```

(D) OTHER INFORMATION: /standerd_name="Oligo vol4"

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(xi)SEQU	ENCE DESCRIPTION: SE	EQ ID NO:39:				
CATGTACTGG	TACCAGCAGA	AGCCAGGAAA	AGCTCCGAAA	CTTCTGATTT	ATGACACATC	6 0
CAACCTGGCT	TCTGGAGTCC	CTTCCCGCTT	CAGTGGCAGT	GGGTCTOGGA	CCGATTACAC	120
CTTTACAATC	TCTTCA					136
(2) INFORMATION E	FOR SEQ ID NO:40:					
(i)SEOU	ENCE CHARACTERISTI	CS:				
	(A) LENGTH: 137 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(ii) MOL	ECULE TYPE: DNA					
(iii) HYPC	THETICAL: NO					
	URE: (A) NAME/KEY; mist_ (B) LOCATION; 1137 (D) OTHER INFORMAT		ligo vel5"			
(xi)SEQU	IENCE DESCRIPTION: SI	EQ ID NO:40:				
TGTGTCTAGA	AAAGTGTACT	TACGTTTTAC	CTCGACCTTG	GTCCCTTGAC	COAACGTGAG	6 0
CGGOTAAGTA	CTCCACTGCT	GGCAGTAATA	AGTGGCTATA	TCTTCCGGCT	GAAGTGAAGA	120
GATTGTAAAG	GTGTAAT					1 3 7
(2) INFORMATION	FOR SEQ ID NO:41;					
	JENCE CHARACTERISTI (A) LEAGTH: 108 base (B) TYPE: nucleic acid (C) STRANDEDNESS:	pairs				
	(D) TOPOLOGY: linear	0 -				
(ii) MOLI	ECULE TYPE: DNA					
(iii) HYPO	JIHERCAL: NO					
	URE: (A) NAME/KEY: misc_ (B) LGCATION; 1108 (D) OTHER INFORMA		Oligo vc16''			
(xi)SEQU	JENCE DESCRIPTION: S	EQ 1D NO:41:				
CACATCTAGA	CCACCATGGA	TTTTCAAGTG	CAGATCTTCA	GCTTCCTGCT	AATCAGTGCC	6 0
TCAGTCATAC	TGTCCAGAGG	AGATATTCAA	ATGACCCAGT	CTCCATCT		108
(2) INFORMATION	FOR SEQ ID NO:42:					
	JENCE CHARACTERIST! (A) LENGTH: 138 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: l'incar	pairs single				
(ii) MOL	ECULE TYPE: DNA					
(iii)(HY2)	OTHERCAL: NO					
(ix)FEA						
	(A) NAME/KEY: misc_ (B) LOCATION: 1138					
	(D) OTHER INFORMA	IION: /standard_name='C	Oligo vell"			
(x i) SEQU	LIENCE DESCRIPTION: S	EQ ID NO:42:				
TAGTCTOTCG	ACCCACCACT	CCATATCACT	CCCACCCACT	CGAGTCCCTT	TCCAGGAGCC	60

	107		-continued		108	
TGGCGGACCC	AGTGTACACC	ATAACTTGTT	ACGGTGAAAC	CACTGGCGGC	ACAAGACAGT	1 2 0
CTCAGAGATC	CTCCTGGC					1 3 8
(2) INFORMATION	FOR SEQ ID NO:43:					
	UENCE CHARACTERISTI (A) LENGTH: 126 bisse (B) TYPE: oucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(i i) MOL	ECULE TYPE: DNA					
(iii)HYP	OTHETICAL: NO					
	TURE: (A) NAME/KBY: misc_; (B) LOCATION: 1,126 (D) OTHER INFORMAT		Ligo vel2"			
(x i) SEQ	UENCE DESCRIPTION: S	EQ ID NO:43:				
TGGTGGGTCG	АСЛБЛСТАТА	ATGCAGCTTT	CATATCCAGA	TTTACCATCA	GCAGAGACAA	6 0
CAGCAAGAAC	ACACTGTATC	T C C A A A 1' G A A	TAGCCTGCAA	GCCGAGGACA	CAGCCATATA	1 2 0
ттлттб						126
(2) INFORMATION	FOR SEQ ID NO:44;					
	UENCE CHARACIERISTI (A) LENGIH: 130 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(i i) MOI	ECULE TYPE: DNA					
(iii) HYP	OTHETICAL: NO		20			
	TURE: (A) NAMEASY: misc (B) LOCATION: 1130 (D) OTHER DIFORMA		ligo wps54"			
(xi)SEQ	UENCE DESCRIPTION: SI	EQ ID NO:44:				
ΛΟΛΟΤΟΤΛΟΑ	CCACCATGGC	TGTCTTGGGG	ствстсттст	GCCTGGTGAC	ATTCCCAAGC	6 0
TGTGTCCTAT	CCGCTGTCCA	GCTGCTAGAG	AGTGGTGGCG	GTCTGGTGCA	GCCAGGAGGA	120
TCTCTGAGAC						1 3 0
(2) INFORMATION	FOR SEQ ID NO:45:					
	UENCE CHARACTERISTI (A) LENGTH: 118 base (B) TYPE: nucleie acid (C) S'LRANDEDNESS: (D) TOPOLOGY: l'incar	pairs				
(ii) MOL	LECULE TYPE: DNA					
(iii)HYP	OTHETICAL: NO					
(ix)FEA	(A) NAME/KEY: misc_ (B) LOCATION: 1118		OF an incident			
(v ;) epo	(D) OTHER INFORMAT UENCE DESCRIPTION: SI		MRV Mb231			
	AGTTAGGACT		GACAGTGACC	AGAGTCCCTT	GGCCCCAGTA	6 0
	TCGTAATTAT					118
	. oo inniini		. D. OOCKCAA		0.0.0.00	

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( 2 ) INF-ORMATION FOR SEQ ID NO.-46:
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- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 111 amino acids

 - (B) TYPE: amino acid (C) SIRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: popide

(x i) SEQUENCE DESCRIPTION: SEQ 1D NO:46:

1	S	p	l	1	c	٧	2	l	L	c	u	T 5	h	г	G	1	n	S	c	r	P	r	0	-	A :	1	a		0			L	u		A	l	3	\	a	l	S		r		5	u	C	1	у
G	1	n	A	r	g	A	1	a		h		1	1	c	S	c	Γ	C	У	s	Α	r	g		2 :		a	S	С	r	-	G	п	1	S	C	г	٧	a	1	S 0	0	r	Т	ь	r	S	C	r
Т	h	r	Т	y	r		5		T	y	C	M	c	l	Н	i	2	Т	r	p	T 4	y 0	r	(G	l	n	G	l	n		L	y s		P	r	0	4	5	у	G I	l	n	P	г	D	F	r	0
L	y	5		0		L	C	u	l	1	c	L	у	S	T	У	Γ		5		S	c	r		۸ :	S	B	L	c	U		G	lu		S 6			(; 1	у	V 1	A	l	P	r	0	A	1	2
	5	g	P	h	c	S	C	r	G	1	у	S	C	T		0		P	h	c	G	1	y		TI	h	r	A	S	p		P 7	5		Т	h	r	I	. 0	u	A :	\$	п	l	l	C		i	
P	r	0	٧	a	1	G	1	u	G	; 1	Ш	G 8	1 5		٨	S	p	Т	'n	r	٧	a	1		T	h	r		0			Т	y z		C	y	S	C	1	n	Н	i	S		5		7	r	p
C	1	u	I	1	С	P	r	0			1 0	Т	b	r	P	h	С	C	1	у	G	1	У		G 1			T	h	r		L	y s	i	L	С	u	C	1	IJ	1 1			L	У	s			

(2) INFORMATION FOR SEQ ID NO:47:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 111 amino acids

 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: peptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO.47.

G 1 u 1	llc Val	Met Thr	Gin Ser Pro	Ala Thr Leu 10	Ser Val Ser	Pro Gly 15
G 1 u	Arg Ala	Thr Leu 20	Ser Cys Arg	Ala Ser Glm 25	Scr Val Scr 30	The Ser
Thr	Tyr Asn 35	Tyr Mel	His Trp Tyr	Gln Gln Lys	Pro Gly Gln 45	Scr Pro
Arg	Lcu Lcu 50	lle Lys	Tyr Ala Ser 55	Asn Lev Glu	Ser Gly 11c	Pro Ala
A r g	Phc Scr	Gly Scr	Gly Ser Gly 70	Thr Glu Phc 75	The Leu The	Ile Ser 80
Атд	Leu Glu	Scr Glu 85	Asp Phe Ala	Val Tyr Tyr 90	Cys Gla Ilis	Scr Trp 95
G 1 u	llc Pro	Tyr Thr 100	Phc Gly Gla	Gly Thr Arg	Val Glu Ilc 110	L y s

(2) INFORMATION FOR SEQ ID NO:48;

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTI-1: 122 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide

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(2) INFORMATION FOR SEQ ID NO:49:.

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 122 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: tricar
- (i i) MOLECULE TYPE: pepude
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:49:

(2) INFORMATION FOR SEQ ID NO:50:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE; amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: popude
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:SO:

Asp Arg Val Ser Ile Thr Cys Lys Ala Ser Gln Asp Val Gly Ser Ala

114

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	2 0		2 5	3 0
V a 1	Val Trp Hi	s Gln Gln Lys Ser 40	Gly Gln Ser Pro Lys 45	Leu Leu Ile
Туг	Trp Ala So	er Thr Arg His Thr 55	Gly Val Pro Asp Arg	Phc Thr Gly
S c r 6 5	Gly Scr Gl	y Thr Asp Phc Thr 70	Leu Thr Ile Thr Asn 75	Val Gin Scr 80
Glu	Asp Leu Al	a Asp Tyr Phe Cys 85	Gln Gin Tyr Scr ilc 90	Phe Pro Leu 95
Thr	Phc Gly Al	a Gly The Arg Leu	Glu Leu Lys 105	

(2) INFORMATION FOR SEQ ID NO.51:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids

 - (B) TYPE: amino acid (C) STRANDEDNESS; single
 - (D) TOPOLOGY: l'uncar

(i i) MOLECULE TYPE: popude

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:51:

Asp 1	lle Gin	Mei Thr G	ln Ser Pro Ser	Thr Leu Ser A	la Ser Val Gly 15
Asp	Arg Val	The lie T 20	br Cys Lys Ala 25	Ser Glm Asp V	al Gly Ser Ala 30
V a I	Val Trp 35	His Glm G	la Lys Pra Gly 40	Lys Ala Pro L 4:	ys Lou Lou lie 5
Туг	Trp Ala	Ser The A	rg His Thr Gly	Val Pro Scr A	rg Phe Thr Gly
S c r 6 5	Gly Ser	Gly Thr G	lu Pbc Thr Leu O	Thr lie Ser Ser 75	er Leu Gla Pro 80
Asp	Asp Pbc	Ala Thr T 85	yr Phe Cys Gln	Gla Tyr Ser 1 90	le Phe Pro Leu 95
Tbr	Рће Слу	Gln Oly T	hr Lys Val Glu 105	Val Lys	

(2) INFORMATION FOR SEQ ID NO:52:

- (i) SEQUE NCE CHARACTERISTICS:
 - (A) LENGTH: 121 amino acids (B) TYPE: amino acid

 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: pepude

(x i) SEQUENCE DESCRIPTION: SEQ ID NO 52:

Gla l	V a 1	Gla	Lcu	G 1 n C	Gln Se	r Asp	Ala	Glu 1 10	Lcu Val	I. y s	Рго	G 1 y 1 5	Ala
Scr	V a l	Lys	1 ¹ c 2 0	Scr (Cys Ly	s Val	\$ c r 2 5	Gly 1	Tyr Thr	Phe	T b r 3 0	Asp	H i s
Thr	1 1 c	H i s	Тгр	Met	Lys G1	n Arg 40	Pro	Głu (Gln Gly	L c u 4 5	GIц	Trp	Phe
G 1 y	T y r 5 0	1 1 c	Туг	Pro A	Arg As	р СІу	His	Thr	Arg Tyr	Seг	Glu	L y s	Phe
L y s	G 1 y	Lys	A 1 a	Thr	Lev Th 70	r Alz	Asp	Lys	Ser Ala 75	Ser	Thr	Ala	T y r
Mc t	His	Leu	Asn	Ser	Leu Th	г Ѕсг	G 1 u	Asp	Scr Ala	V a 1	Туг	Pho	Суѕ

Ala Arg Gly Arg Asp Ser Arg Glu Arg Asn Gly Phe Ala Tyr Trp Gly 100

Gln Gly Thr Leu Val Thr Val Ser Ala 120

- (2) INFORMATION FOR SEQ ID NO:53:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 121 amino acids
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: peptide
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:53:

- (2) INFORMATION FOR SEQ ID NO:54:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 111 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: peptide
 - (x i) SEQUENCE DESCRIPTION; SEQ ID NO:54:
 - Asp 11c
 Val
 Lcu 55
 Gln Ser Pro Ala Scr Lcu Ala Val
 Ser Lcu Gly 15

 Gln Arg Ala Thr 20
 Ilc Scr Cys Arg Ala Scr Glu Ser Val
 Asp Asn Tyr 25

 Gly Ile Ser Phe Met Asn Trp Phe 40
 Gln Gln Lys Pro Gly Gln Pro Ala 55

 Lys Leu Lcu Ile Tyr Ala Ala Scr Asn Gln Gly Scr Gly Val Pro Ala 55

 Arg Phe Scr Gly Ser Gly Scr Gly Thr Asp Phe Scr Lcu Asn Ile His 80

 Pro Met Glu Glu Asp 85
 Asp Thr Ala Met Tyr Phe Cys Gln Glu Glu Ile Lys 110

 Glu Val Pro Trp Thr Phe Gly Gly Gly Thr Lys Lcu Glu Ile Lys 110

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(2) INFORMATION FOR SEQ ID NO:55: (i) SEQUENCE CHARACTERISTICS:

- (B) TYPE: amino acid
- (A) LE VGTH: 111 amino acids (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUELNCE DESCRIPTION: SEQ ID NO:55:

lle Gln Mei Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Val Gly
5 10 Asp Arg Val Thr Ito Thr Cys Arg Ala Ser Glu Ser Val Asp Asn Tyr 20 25 30 Gly lle Ser Phe Mei Asn Trp Phe Gln Gln Lys Pro Gly Lys Ala Pro 35 Lys Leu Leu IIe Tyr Ala Ala Ser Asn Gln Gly Ser Gly Val Pro Ser 50 55 Arg Phe Scr Gly Scr Gly Scr Gly Thr Asp Phe Thr Leu Thr ilc Scr 65 75 80 Lcu Gln Pro Asp Asp Pbc Ala Thr Tyr Tyr Cys Gln Gln Scr Lys $85 \ \ \, 90 \ \ \, 95$ Glu Val Pro Trp Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys 100 105

(2) INFORMATION FOR SEQ ID NO:56:

- (i) SEOUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDN'ESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:56:
- Val Glu Leu Glu Glu Ser Gly Pro Glu Leu Val Lys Pro Gly Ala 5 $$ Ser Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asp Tyr 20 25 Asn Met His Trp Val Lys Glm Ser His Gly Lys Ser Leu Glu Trp lle 35 40 45 Gly Tyr Ile Tyr Pro Tyr Asn Gly Gly Thr Gly Tyr Asn Gln Lys Phe 50 55 Lys Ser Lys Ala Thr Leu Thr Val Asp Asn Ser Ser Ser Thr Ala Tyr 65 70 75 80 Asp Val Arg Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys 85 90 Ala Arg Gly Arg Pro Ala Met Asp Tyr Trp Gly Gla Gly The Ser Val
- (2) INFORMATION FOR SEQ ID NO:57:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: peptide

119 120

```
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

Glo Vai Gln Leu Vai Gln Ser Giy Ala Glu Val Lys Lys Pro Gly Scr lo lo Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asp Tyr 20

Asn Mel His Trp Val Arg Gln Ala Pro Gly Gln Gly Leu Glu Trp lle 35

Gly Tyr lle Tyr Pro Tyr Asn Gly Gly Thr Gly Tyr Asn Gln Lys Phe 50

Lys Ser Lys Ala Thr lle Thr Ala Asp Glu Ser Thr Asn Thr Ala Tyr 65

Met Glu Leu Ser Ser Leu Arg Ser Glu Asp Thr Ala Val Tyr Tyr Cys 85

Ala Arg Gly Arg Pro Ala Met Asp Tyr Trp Gly Gln Gly Thr Leu Val 105

Thr Val Ser Ser
```

(2) INFORMATION FOR SEQ ID NO:58:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: popuido
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:58:

Gin lie Val Leu Thr Gin Ser Pro Ala lie Mei Ser Ala Ser Pro Gly

Glu Lys Val Thr Mei Thr Cys Ser Gly Ser Ser Ser Val Ser Pho Mei

Tyr Trp Tyr Gln Gln Arg Pro Gly Ser Ser Pro Arg Leu Leu lle Tyr

Asp Thr Ser Asn Leu Ala Ser Gly Val Pro Val Arg Pho Ser Gly Ser

Gly Ser Gly Thr Ser Tyr Ser Leu Thr lle Ser Arg Mei Glu Ala Glu

65

Asp Ala Ala Thr Tyr Tyr Cys Gln Gln Trp Ser Thr Tyr Pro Leu Thr

105

Phe Gly Ala Gly Thr Lys Leu Glu Leu Lys

(2) INFORMATION FOR SEQ ID NO.59.

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 106 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: pe pide
- (x i) SEQUENCE DESCRIPTION: SE♠ 1D NO:59:

Asp Arg Val Thr IIc Thr Cys Scr Gly Scr Scr Scr Val Scr Phc Mci

Tyr Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu lle Tyr 45

Asp Thr Ser Asn Leu Ala Ser Gly Val Pro Ser Arg Phe Ser Gly Ser 50

Gly Ser Gly Thr Asp Tyr Thr Phe Thr lle Ser Ser Leu Gln Pro Glu 75

Asp lle Ala Thr Tyr Tyr Cys Gln Gln Trp Ser Thr Tyr Pro Leu Thr 85

Phe Gly Gln Gly Thr Lys Val Glu Val Lys 105

(2) INFORMATION FOR SEQ ID NO:60.

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: am'mo acid
 - (C) STRANDEDN'ESS: single
 - (D) TOPOLOGY: funcar

(i i) MOLECULE TYPE: populdo

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:60:

Gin Val Gin Leu Lys Gin Ser Giy Pro Giy Leu Val Gin Pro Ser Gin Ser Gin 10

Ser Leu Ser lic The Cys The Val Ser Giy Pho Giy Leu Val Gin Pro Ser Tyr 25

Gly Val His Trp lic Arg Gin Ser Pro Giy Lys Giy Leu Giu Trp Leu 45

Gly Val His Trp Ser Giy Giy Ser The Asp Tyr Asn Ala Ala Phe lic 50

Ser Arg Leu The lic Ser Lys Asp Asn Ser Lys Ser Gin Val Phe 80

Lys Val Asn Ser Leu Gin Pro Ala Asp The Ala Ile Tyr Tyr Cys Ala 90

Arg Ala Giy Asp Tyr Asn Tyr Asp Giy Phe Ala Tyr Trp Giy Gin Giy Giy Leu Val The Val Ser Ala 115

The Leu Val The Val Ser Ala

(2) INFORMATION FOR SEQ ID NO:61:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGIH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDA'ESS: single (D) TOPOLOGY: linear
- (1 i) MOLECULE TYPE: pepude
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:61:

Giu Val Gin Leu Leu Giu Ser Giy Giy Giy Leu Val Gin Pro Giy Giy Giy Leu Val Gin Pro Giy Giy Giy Ser Leu Arg Leu Ser Cys Aia Ala Ser Giy Phe Thr Val Thr Ser Tyr 25 Giy Val His Trp Val Arg Gin Ala Pro Giy Lys Giy Leu Giu Trp Vai 35 Giy Vai IIc Trp Ser Giy Giy Ser Thr Asp Tyr Asn Ala Ala Phe IIc 50 Arg Phe Thr 11e Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr Leu 80

123

Gln Mei Asn Ser Leu Gln Ala Glu Asp Thr Aln lle Tyr Tyr Cys Ala 85

Arg Ala Gly Asp Tyr Asn Tyr Asp Gly Phe Ala Tyr Trp Gly Gln Gly 100

The Leu Val The Val Ser Ser

(2) INFORMATION FOR SEQ ID NO:62:

- (i) SEQUENCE CHARACIERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS; single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: populat

(x i) SEQUENCE DESCRIPTION: SEQ ID NO.62:

 Asp
 11c
 Val
 Lcu
 Thr
 Gln
 Scr
 Pro
 Ala
 Thr
 Lcu
 Scr
 Val
 Thr
 Pro
 Ala
 Thr
 Lcu
 Scr
 Val
 Thr
 Pro
 Ala
 Thr
 Lcu
 Scr
 Pro
 Ala
 Scr
 Val
 Thr
 Pro
 Asn
 A

(2) INFORMATION FOR SEQ ID NO:63:

- (i) SEQUENCE CHARACIERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: poptide

(x i) SEQUENCE DESCRIPTION: SEQ ID NO:63:

(2) INFORMATION FOR SEQ ID NO:64:

- (i) SEQUENCE CHARACIERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: am ino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i l) MOLECULE TYPE: pepilde
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:64:
- Ser Mei Lys lle Ser Cys Lys Ala Ser Val Tyr Ser Phe Thr Gly Tyr $20\,$
- Thr McL Asn Trp Val Lys Glo Ser His Gly Gln Asn Leu Glu Trp IIc 35 40 45
- Gly Leu lle Asn Pro Tyr Asn Gly Gly Thr Scr Tyr Asn Gla Lys Phe 50 60
- Lys Gly Lys Ala Thr Leu Thr Val Asp Lys Scr Scr Asp Thr Ala Tyr 65 70 75 80
- Met Glu Leu Leu Ser Leu Thr Ser Ala Asp Ser Ala Val Tyr Tyr Cys 85 90
- The Arg Arg Gly Phc Arg Asp Tyr Scr Mct Asp Tyr Trp Gly Gln Gly 100
- Thr Scr Val Thr Val Scr Scr 115
- (2) INFORMATION FOR SEQ ID NO:65:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 areino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: pepulde
 - (\mathbf{x} i) SEQUENCE DESCRIPTION: SEQ ID NO:65:
 - Gin Val Gin Leu Val Gin Ser Gly Ala Glu Val Lys Lys Pro Gly Ser 1 10 15
 - Ser Val Arg Val Ser Cys Lys Ala Ser Gly Tyr Ser Phe Thr Gly Tyr
 20 25 30
 - Thr Mcl Asp Trp Val Arg Gin Ala Pro Gly Lys Gly Leu Glu Trp Val 35 40 45
 - Gly Lew IIc Asn Pro Tyr Asn Gly Gly Thr Scr Tyr Asn Gln Lys Phe 50
 - Lys Gly Arg Val Thr Val Ser Leu Lys Pro Ser Phe Asn Gln Ala Tyr 65 70 75 80
 - Met Glu Leu Ser Ser Leu Phe Ser Glu Asp Thr Ala Val Tyr Tyr Cys 85 90
 - Thr Arg Arg Gly Phe Arg Asp Tyr Ser Met Asp Tyr Trp Gly Gln Gly
 - Thr Leu Val Thr Val Ser Ser
- (2) INFORMATION FOR SEQ ID NO:66:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 393 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear

-continued (i i) MGLECULE TYPE: cDNA (i x) FEATURE: (A) NA VIEIKEY: CDS (B) LOCATION: 1,.393 (x i) SEQUENCE DESCRIPTION: SEQ 1D NO:66: ATG GAG AAA GAC ACA CTC CTG CTA TGG GTC CTG CTT CTC TGG GTT CCA Met Glu Lys Asp Thr Leu Leu Leu Trp Val Leu Leu Trp Val Pro GGT TCC ACA GGT GAC ATT GTG CTG ACC CAA TCT CCA GCT TCT TTG GCT Gly Ser Thr Gly Asp lie Val Leu Thr Gln Ser Pro Ala Ser Leu Ala GTG TCT CTA GGG CAG AGG GCC ACC ATC TCC TGC AGA GCC AGC GAA AGT 144 Val Ser Leu Gly Gln Arg Ala Thr lie Ser Cys Arg Ala Ser Glu Ser GTT GAT AAT TAT GGC AFT AGT TTT ATG AAC TGG TTC CAA CAG AAA CCA Val Asp Asn Tyr Gly lic Scr Phc Mcl Asn Trp Phc Gln Gln Lys Pro 50 55 192 GGA CAG CCA CCC AAA CTC CTC ATC TAT GCT GCA TCC AAC CAA GGA TCC Gly Gln Pro Pro Lys Leu Leu lle Tyr Ala Ala Scr Asn Gln Gly Scr 2 4 0 GGG GTC CCT GCC AGG TTT AGT GGC AGT GGG TCT GGG ACA GAC TTC AGC Gly Val Pro Ala Arg Phc Scr Gly Scr Gly Scr Gly Thr Asp Phc Scr 85 288 CTC AAC ATC CAT CCT ATG GAG GAG GAT GAT ACT GCA ATG TAT TTC TGT Leu Asn llc His Pro Mei Glu Glu Asp Asp Thr Ala Mei Tyr Phe Cys 3 3 6 CAG CAA AGT AAG GAG GTT CCG TGG ACG TTC GGT GGA GGC ACC AAG CTG Gln Gln Ser Lys Glu Val Pro Trp Thr Phe Gly Gly Gly Thr Lys Leu GAA ATC AAA 393 Clu Ile Lys 130

- (2) INFORMATION FOR SEQ ID NO:67:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 131 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: procin
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:67:
- Met
 Glu
 Lys
 Asp
 Thr
 Leu
 Leu
 Leu
 Trp
 Val
 Leu
 Leu
 Leu
 Trp
 Val
 Leu
 Leu</th

-continued Glu Ilc Lys 130 (2) INFORMATION FOR SEO ID NO.68: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 405 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPGLOGY: linear (i i) MOLECULE TYPE: cDNA (i x) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1..405 (x i) SEQUENCE DESCRIPTION: SEQ ID NO:68: ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT GCA GGC Mct Gly Trp Scr Trp lie Phe Leu Phe Leu Leu Scr Gly Thr Ala Gly GTC CAC TCT GAG GTC CAG CTT CAG CAG TCA GGA CCT GAG CTG GTG AAA Val His Ser Glu Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Lys 209 6 CCT GGG GCC TCA GTG AAG ATA TCC TGC AAG GCT TCT GGA TAC ACA TTC
Pro Gly Ala Scr Val Lys lie Scr Cys Lys Ala Scr Gly Tyr Tbr Pbc ACT GAC TAC AAC ATG CAC TGG GTG AAG CAG AGC CAT GGA AAG AGC CTT
Thr Asp Tyr Asm Mct His Trp Val Lys Glm Ser His Gly Lys Ser Leu
50
60 GAG TGG ATT GGA TAT ATT TAT CCT TAC AAT GGT GGT ACT GGC TAC AAC Gla Trp llc Gly Tyr Ilc Tyr Pro Tyr Asn Gly Gly Thr Gly Tyr Asn CAG AAG TTC AAG AGC AAG GCC ACA TTG ACT GTA GAC AAT TCC TCC AGC Gln Lys Phc Lys Scr Lys Ala Thr Leu Thr Val Asp Asn Scr Scr Scr ACA GCC TAC ATG GAC GTC CGC AGC CTG ACA TCT GAG GAC TCT GCA GTC Tbr Ala Tyr Mcl Asp Val Arg Scr Lcu Tbr Ser Glu Asp Scr Ala Val 3 3 6 TAT TAC TGT GCA AGA GGG CGC CCC GCT ATG GAC TAC TGG GGT CAA GGA Tyr Tyr Cys Ala Arg Gly Arg Pro Ala Met Asp Tyr Trp Gly Gin Gly 115 384 ACC TCA GTC ACC GTC TCC TCA
Thr Scr Val Thr Val Scr Scr 405 (2) INFORMATION FOR SEQ ID NO.69: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 135 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (i i) MOLECULE TYP E: protein (x i) SEQUENCE DESCRIPTION: SEQ ID NO. 69: McL Gly Trp Scr Trp llc Phe Leu Phe Leu Leu Scr Gly Thr Ala Gly
1 10 Val His Scr Glu Val Gln Lcu Gln Gln Scr Gly Pro Glu Lcu Val Lys 20 25Pro Gly Ala Scr Val Lys lic Scr Cys Lys Ala Scr Gly Tyr Thr Pbc 35

The Asp Tyr Asm Met His Trp Val Lys Gin Ser His Gly Lys Ser Leu 50 55

5,530,101

131 132

Glu Trp llc Gly Tyr IIc Tyr Pro Tyr Asn Gly Gly Thr Gly Tyr Asn 65

Gln Lys Phc Lys Scr Lys Ala Thr Lcu Thr Val Asp Asn Scr Scr Scr 95

Thr Ala Tyr Mcl Asp Val Arg Scr Lcu Thr Scr Glu Asp Scr Ala Val 100

Tyr Tyr Cys Ala Arg Gly Arg Pro Ala Mcl Asp Tyr Trp Gly Glu Gly 125

Thr Scr Val Thr Val Scr Scr 135

(2) INTORMATION FOR SEQ 1D NO:70:

- (1) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE; amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:70:

 Asp
 lie
 Gin
 Mcl
 Thr
 Gln
 Ser
 Pro
 Ser
 Thr
 Leu
 Ser
 Ala
 Ser
 Val
 Gly

 Asp
 Arg
 Val
 Thr
 lle
 Thr
 Cys
 Arg
 Ala
 Ser
 Gln
 Ser
 Ile
 Asn
 Thr
 Trp

 Leu
 Ala
 Trp
 Tyr
 Gln
 Gln
 Lys
 Pro
 Gly
 Lys
 Ala
 Pro
 Leu
 Leu
 Met

 Tyr
 Lys
 Ala
 Ser
 Ser
 Leu
 Gly
 Ser
 Gly
 Val
 Pro
 Ser
 Arg
 Phe
 lle
 Gly

 Ser
 Gly
 Ser
 Leu
 Gly
 Ser
 Gly
 Val
 Pro
 Ser
 Arg
 Phe
 lle
 Gly

 Ser
 Gly
 Ser
 Leu
 Thr
 Leu
 Thr
 Leu
 Thr
 Leu
 Thr
 Leu
 Thr
 Leu
 Thr

(2) INFORMATION FOR SEQ ID NO:71:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 111 amino acids
 - (B) TYPE: amimo acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECU1E TYPE: pepuide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:71:

 Asp 11e
 Glm
 Met
 Thr 5
 Glm
 Ser Pro
 Ser Ser Leu Ser Leu Ser Ala Ser Val Gly 15

 Asp Arg Val
 Thr Ile Thr Cys Arg Ala Ser Glu Ser Val Asp Asn Tyr 20

 Gly Ile Ser Phe Met Asn Trp Phe Gln Gln Lys Pro Gly Lys Ala Pro 35

 Lys Leu Leu Ile Tyr Ala Ala Ser Asn Gln Gly Ser Gly Val Pro Ser 50

 Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser 65

 Ser Leu Glu Pro Asp Asp Phe Ala Thr Tyr Tyr Cys Gln Gln Ser Lys 95

133

Glu Val Pro Trp Thr Phc Gly Gln Gly Thr Lys Val Glu Ilc Lys
100 110

- (2) INFORMATION FOR SEQ ID NO:72:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 117 amino acids (B) TYPE: amino acid
 - (B) TYPE: amino scid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: peptide
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:72:

Val Thr Val Scr Scr 115

- (2) INFORMATION FOR SEQ ID No:73:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 116 amino acids
 - (B) TYPE: smino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: popuide
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:73:

Gin Val Gin Leu Val Gin Ser Giy Ala Giu Val Lys Lys Pro Gly Ser IS

Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asp Tyr Asn Met His Trp Val Arg Gln Ala Pro Gly Gly Gln Gly Leu Glu Trp lle

Gly Tyr lle Tyr Pro Tyr Asn Gly Gly Thr Gly Tyr Asn Gln Lys Phe

Sor Lys Ala Thr IIc Thr Ala Asp Glu Ser Thr Asn Thr Ala Tyr 75

Ala Arg Gly Arg Pro Ala Met Asp Tyr Trp Gly Gln Gly Thr Leu Val

The Val Ser Ser

(2) INFORMATION FOR SEQ ID NO:74:

	155		-continued		150	
	JENCE CHARACTERISTI (A) LENGTH: 132 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	p airs single				
(ii) MOL	ECULE TYPE: DNA (oligo	onucleolide)				
(xi)SEQU	JENCE DESCRIPTION: SI	EQ ID NO:74:				
TATATCTAGA	CCACCATGGG	ATGGAGCTGG	ATCTTTCTCT	тсстсстстс	AGGAACTGCT	6 0
GGCGTCCACT	CTCAGGTTCA	GCTGGTGCAG	TCTGGAGCTG	AGGTGAAOAA	GCCTGGGAGC	1 2 0
TCAGTGAAGG	тт					I 3 2
(2) INFORMATION	FOR SEQ ID NO:75;					
	JENCE CHARACIERISTI (A) LENOTH: 133 base (B) TYPE: nucleic acid (C) S'INANDEDNESS: (D) TOPOLOGY: lincor	pairs single				
(i i) MOL	ECULE TYPE: DNA (oligo	onucteo ide)				
(x i)SEQU	JENCE DESCRIPTION: SI	EQ ID NO:75:				
AGCCGGTACC	ACCATTGTAA	GGATAAATAT	ATCCAATCCA	TTCCAGGCCT	TGGCCAGGAG	6 0
CCTGCCTCAC	CCAGTGCATG	TTGTAGTCAG	TGAAGGTGTA	GCCAGAAGCT	TTGCAGGAAA	120
CCTTCACTGA	GCT					1 3 3
(2) INFORMATION	FOR SEQ ID NO:76:					
	DENCE CHARACTERISTI (A) LENGTH: 112 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs single				
(ii) MOL	ECULE TYPE: DNA (oligo	onuclco ide)				
(xi)SEQU	JENCE DESCRIPTION: SI	EQ ID NO:76:				
TGGTGGTACC	GGCTACAACC	AGAAGTTCAA	GAGCAAGGCC	ACAATTACAG	CAGACGAGAG	6 U
TACTAACACA	GCCTACATGG	AACTCTCCAG	CCTGAGGTCT	GAGGACACTG	CA	1 2
(2) INFORMATION	FOR SEQ ID NO:77:					
	JENCE CHARACTERISTI (A) LENGTH: 111 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (O) TOPOLOGY: linear	Pairs single				
(i i) MOL	ECULE TYPE: DNA (oligo	onucleolide)				
(x i) SEQU	CENCE DESCRIPTION: S	EQ ID NO:77:				
TATATCTAGA	GGCCATTCTT	ACCTGAAGAG	ACAGTGACCA	GAGTCCCTTG	GCCCCAGTAG	6 0
TCCATAGCGG	GGCGCCCTCT	TGCGCAGTAA	TAGACTGCAG	TGTCCTCAGA	C	1 1 1
(2) INFORMATION	FOR SEQ ID NO:78:					
	DENCE CHARACTERUSTI (A) LENGTH: 122 base (B) TYPE: nucleic acid (C) STRANDEDN'ESS: (D) TOPOLOGY: l'uncar	pairs singile				

(i i) MOLECULE TYPE: DNA (oligenucleatide)

	137		-continued	1	.38	
(7 ;) \$501	JENCE DESCRIPTION: SE	EO ID NO.78:	-condition			
	CCACCATGGA		CTCCTGCTAT	GGGTCCTGCT	TOTOTOGOTT	6 0
	CAGGTGACAT					
	CAGGIGACAI	TCAGATGACC	CAGTCTCCGA	GCTCTCTGTC	CGCATCAGIA	1 2 0
GG						1 2 2
(2) INFORMATION	FOR SEQ ID NO:79:					
	FENCE CHARACTERISTI (A) LENGTH: 122 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pair; single				
(i i) MOL	ECULE TYPE: DNA (oligo	onuclcoude)				
(x i) SEQI	JENCE DESCRIPTION: SE	EQ ID NO:79:				
TCAGAAGCTT	AGGAGCCTTC	CCGGGTTTCT	GTTGGAACCA	GTTCATAAAG	CTAATGCCAT	6 0
AATTGTCGAC	ACTTTCGCTG	GCTCTGCATG	TGATGGTGAC	CCTGTCTCCT	ACTGATGCGG	1 2 0
A C						1 2 2
/ 2) INTORNACION	EOD DEO ED NO.00.					
(2) INFORMATION						
	UENCE CHARACTERISTI (A) LENGTH: 119 hase (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY; hinear	pair: single				
(i i) MOL	ECULE TYPE: DNA (olig	onucleolide)				
(x i) SEQ	UENCE DESCRIPTION: S	EQID NO:80.				
TCCTAAGCTT	CTGATTTACG	CTGCATCCAA	CCAAGGCTCC	GGGGTACCCT	CTCGCTTCTC	6 0
AGGCAGTGGA	TCTGGGACAG	ACTTCACTCT	CACCATTTCA	TCTCTGCAGC	CTGATGACT	1 1 9
(2) INFORMATION	FOR SECULD MO-81-					
	UENCE CHARACTERISTI (A) LENGTH: 118 base (B) TYPE: nucleic acid (C) STRANDEDN'ESS: (D) TOPOLOGY: Fincar	single single				
(i i) MOC	ECULE TYPE: DNA (olig	onucleotide)				
(xi)SEQ	UENCE DESCRIPTION: S	EQ TO NO:81:				
ТАТАТСТАСА	CTTTGGATTC	TACTTACGTT	TGATCTCCAC	CTTGGTCCCT	TGACCGAACG	6 0
TCCACGGAAC	CTCCTTACTT	TGCTGACAGT	AATAGGTTGC	GAAGTCATCA	GGCTGCAG	1 1 8
(2) INFORMATION	FOR SEQ ID NO:82:					
(i)SEQ	UENCE CHARACTERIST (A) LENGTH: 381 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: incar	: pairi single				
(i i) MOI	LECULE TYPE: cDNA					
(ix)FEA	TURE: (A) NAME/KEY: CDS (B) LOCATION: 1.381					
(-:) 000	UENCE DESCRIPTION: S					
			GGA CTT ATC	CTT TTT	C ATT TCA	
	C ACA CCT C					4 8

139

								-60	IIU IIUC	ш						
1				5					1 0					1 5		
GCC	тсс	AGA	GGT	GAT	АТТ	GTG	CTA	ACT	CAG	TCT	CCA	GCC	ACC	CTG	TCT	96
A] 2	Scr	Arg	G I y 2 0	Asp	llc	Val	Lcu	Thr 25	G 1 n	Scr	Pro	Ala	T b r	Lси	Scr	
GTG	ACT	CCG	GGA	GAT	AGC	GTC	AGT	CTT	TCC	TGC	AGG	GCC	AGC	CAA	AGT	I 4 4
V a l	Thr	Pro 35	Gly	Asp	Ser	Val	Scr 40	Leu	Scr	Cys	Arg	A I a 4 5	Scr	G 1 n	Scr	
ATT	AGC	AAC	AAC	CTA	CAC	TGG	TAT	CAA	CAA	AAA	TCA	CAT	GAG	TCT	CCA	192
1 1 c		Asn	Asn	Lcu	His		Tyr	G 1 n	Gln	Lys		His	Glu	Scr	Pro	
	5 0					5 5					6 0					
A G G	CTT	CTC	ATC	AAG	TAT	GCT	TCC	CAG	TCC	ATC	TCT	GGG	ATC	CCC	TCC	2 4 0
A r g	Lcu	Leu	llc	Lys	T y r 7 0	Ala	Scr	G 1 n	Scr	1 l c 7 5	Scr	Gly	1 1 c	Pro	Scr 80	
A G G	TTC	AGT	GGC	AGT	GGA	TCA	GGG	ΛСΛ	GAT	TTC	ACT	CTC	AGT	GTC	AAC	288
Arg	Phc	Scr	Gly		Gly	Scr	Gly	Thr		Phc	Thr	Lcu	Scr		Asn	
				\$ 5				-	9 0					9 5		
GGT	GTG	GΛG	ΛCΤ	GΛΛ	GAT	TTT	GGA	ATG	TAT	TTC	TGT	CAA	CAG	AGT	AAC	3 3 6
Gly	V a l	Glu	Thr	Glu	Asp	Phc	Gly		Tyr	Phc	Cys	G 1 n		Scr	Asn	
			1 0 0					1 0 5					1 1 0			
AGT	TGG	CCT	CAT	ΛCG	TTC	$GG\Lambda$	GGG	GGG	A C C	$\Lambda \Lambda G$	CTG	GAA	ATA	AAA		3 8 1
Scr	Trp		H i s	Thr	Phc	Gly		Gly	Thr	Lys	Lсш		1 1 c	Lys		
		115					120					125				

(2) INFORMATION FOR SEQ ID NO:83:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 127 amino soids
 - (B) TYPE: am'mo acid
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:83:

 Mcl
 Val
 Phc
 Thr
 Pro
 Gin
 11c
 Lcu
 Gly
 Lcu
 Met
 Leu
 Phc
 Trp
 Ilc
 Scr

 A1z
 Scr
 Arg
 Gly
 Asp
 Ilc
 Val
 Lcu
 Thr
 Gln
 Scr
 Pro
 Ala
 Thr
 Lcu
 Scr

 Val
 Thr
 Pro
 Asp
 Scr
 Val
 Scr
 Lcu
 Scr
 Lcu
 Scr
 Gln
 Scr
 Lcu
 Scr
 Gln
 Scr
 His
 Gln
 Scr
 Pro

 Arg
 Lcu
 Lcu
 Lic
 Lys
 Tyr
 Ala
 Scr
 Gln
 Scr
 His
 Glu
 Scr
 Pro

 Arg
 Phc
 Scr
 Gly
 Scr
 Gly
 Scr
 Gly
 Thr
 Asp
 Phc
 Thr
 Asp
 Phc
 Thr
 Asp
 Phc
 Cys
 Gln
 Gln
 Scr
 Asp
 Asp
 Asp
 Flow
 Gly

(2) INFORMATION FOR SEQ ID NO:84:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 414 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULETYPE: cDNA
- (i x) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: I..414

141 142

(x i) SEQUENCE DESCRIPTION: SEQ 1D NO:84: ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT GCA GGT Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly 1 5 10 48 GTC CAC TCT GAG GTC CAG CTG CAA CAG TCT GGA CCT GAG CTG GTG AAG Val His Scr Glu Val Gln Lcu Gln Gln Scr Gly Pro Glu Lcu Val Lys 96 CCT GGA GCT TCA ATG AAG ATA TCC TGC AAG GCT TCT GTT TAC TCA TTC Pro Gly Ala Scr McL Lys IIc Scr Cys Lys Ala Scr Val Tyr Scr Phc 35 144 ACT GGC TAC ACC ATG AAC TGG GTG AAG CAG AGC CAT GGA CAG AAC CTT Tbr Gly Tyr Thr Mci Asn Trp Val Lys Gln Scr His Gly Gln Asn Leu 50 60 192 GAG TGG ATT GGA CTT ATT A.AT CCT TAC AAT GGT GGT ACT AGC TAC AAC
Glu Trp llc Gly Lcu llc Asn Pro Tyr Asn Gly Gly Thr Scr Tyr Asn
65 CAG AAG TTC AAG GGG AAG GCC ACA TTA ACT GTA GAC AAG TCA TCC AAC GIn Lys Pho Lys Gly Lys Ala Thr Lou Thr Val Asp Lys Scr Scr Asn 90 ACA GCC TAC ATG GAG CTC CTC AGT CTG ACA TCT GCG GAC TCT GCA GTC
Thr Ala Tyr Mci Glu Lcu Lcu Scr Lcu Thr Scr Ala Asp Scr Ala Val 3 3 6 TAT TAC TGT ACA AGA CGG GGG TTT CGA GAC TAT TCT ATG GAC TAC TGG
Tyr Tyr Cys Tbr Arg Arg Gly Phc Arg Asp Tyr Scr Mci Asp Tyr Trp
115 3 8 4 GGT CAA GGA ACC TCA GTC ACC GTC TCC TCA 4 1 4 Gly Gin Gly Thr Ser Val Thr Val Ser Ser

(2) INFORMATION FOR SEQ ID NO:85:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 138 amino acids
 - (B) TYPE: amino acid (B) TOPOLOGY: linear
- (i i) MOLECULE TYPE: protein
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:SS:

 Mei
 Gly
 Trp
 Ser
 Trp
 He
 Phe
 Leu
 Phe
 Leu
 Ser
 Gly
 Thr
 Ala
 Gly

 Val
 His
 Ser
 Glu
 Val
 Gln
 Leu
 Gln
 Ser
 Gly
 Pro
 Glu
 Leu
 Val
 Lys

 Pro
 Gly
 Ala
 Ser
 Met
 Lys
 Ile
 Ser
 Cys
 Lys
 Ala
 Ser
 Val
 Tyr
 Ser
 Phe

 Thr
 Gly
 Tyr
 Thr
 Met
 Asn
 Trp
 Val
 Lys
 Gln
 Ser
 His
 Gly
 Asn
 Leu

 Glu
 Trp
 Ile
 Gly
 Leu
 Ile
 Asn
 Pro
 Tyr
 Asn
 Gly
 Gln
 Ser
 Tyr
 Asn

 Glu
 Lys
 Gly
 Lys
 Ala
 Thr
 Leu
 Tyr
 Val
 Asp
 Lys
 Ser
 Ser
 Asn

 Thr
 <td

(2) INFORMATION FOR SEQ ID NO:86:

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( i ) SEQUENCE CHARACTERISTICS:
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- (A) LENGTH: 108 amino acids (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: l'incar
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:86:

Giu ile Val Leu Thr Gin Ser Pro Gly Thr Leu Ser Leu Ser Pro Gly
1 10 15

Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gin Ser Val Ser Ser Gly

Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gln Ser Val Ser Ser Gly
20 25 30

Tyr Leu Gly Trp Tyr Glя Glя Lys Pra Gly Glя Ala Pra Arg Leu Leu 35 40 45

Ile Tyr Gly Ala Ser Ser Arg Ala Thr Gly Ile Pro Asp Arg Phe Ser 50

Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr lie Ser Arg Leu Glu 65 70 75

Pro Glu Asp Phc Ala Vel Tyr Tyr Cys Gin Gin Tyr Gly Scr Lcu Gly 85 90

Arg Thr Phe Gly Gin Gly Thr Lys Val Glu Ile Lys 100 $\,$

- (2) INFORMATION FOR SEQ ID NO:87:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGITH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (| i) MOLECULE TYPE: popude
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO.87:

Glu Ile Val Leu Thr Gin Ser Pro Gly Thr Leu Ser Leu Ser Pro Gly 1 $$

Glu Arg Ala Thr Leu Scr Cys Arg Ala Scr Gin Scr lle Scr Asn Asn $^{\circ}$ 25

Leu His Tro Tyr Gln Gln Lys Pro Gly Gln Ala Pro Arg Leu Leu lle 35 40 45

Lys Tyr Als Scr Gln Ser IIe Scr Gly lle Pro Asp Arg Phe Scr Gly 50

Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Arg Leu Glu Pro 65 70 75

Glu Asp Phc Ala Val Tyr Tyr Cys Gln Gln Scr Asn Scr Trp Pro His 85

Thr Phe Gly Gin Gly Thr Lys Val Glu lie Lys

- (2) information for SEQ ID NO:88:
 - (;) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 122 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: pepside
 - (× i) SEQUENCE DESCRIPTION: SEQ ID NO:88:
 - Gin Val Gin Leu Met Gin Ser Gly Ala Giu Val Lys Lys Pro Gly Ser

-continued

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Ser Val Arg Val Ser Cys Lys Thr Ser Gly Gly Thr Phe Val Asp Tyr 25

Lys Gly Leu Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val 40

Gly Gln Ile Pro Leu Arg Phe Asn Gly Glu Val Lys Asn Pro Gly Ser 60

Val Val Arg Val Ser Val Ser Leu Lys Pro Ser Phe Asn Gln Ala His 65

Met Glu Leu Ser Ser Leu Phe Ser Glu Asp Thr Ala Val Tyr Tyr Cys 95

Ala Arg Glu Tyr Gly Phe Asp Thr Ser Asp Tyr Tyr Tyr Tyr Tyr Trp 100

Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ico
```

(2) INFORMATION FOR SEQ ID NO:89:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: am ino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(i i) MOCECULE TYPE: pcpude

(xi) SEQUENCE DESCRIPTION: SEQ ID NO.89:

G 1 n	Val Gln	Leu Val	Gln Ser	Gly Ala	Glu Val 10	Lys Lys	Pro Gly Scr 15
Ѕст	Val Arg	Val Ser 20	Cys Lys	Ala Scr 25	Gly Tyr	Scr Phc	The Gly Tye
Thr	Met Asn 35	Trp Val	Arg Gln	Ala Pra 40	Oly Lys	Gly Leu 45	Glu Trp Val
Gly	Leu Ile 50	Asn Pro	Tyr Asn S5	Gly Gly	Thr Ser	Tyr Asn 60	Gln Lys Phe
L y s 6 5	Gly Arg	Val Thr	Val Scr 70	Lcu Lys	Pro Ser 75	Phc Asn	Gln Ala Tyr 80
McL	Glu Lcu	Ser Ser 85	Leu Phe	Ser Glu	Asp Thr 90	Ala Val	Tyr Tyr Cys
Тһт	Arg Arg	Gly Phe 100	Arg Asp	Tyr Ser 105	Мец Азр	Tyr Trp	Gly Gln Gly
Thr	Leu Val	The Val	Ser Ser				

(2) INTORMATION FOR SEQ ID NO:90:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 129 base pairs
 - (B) TYPE: nuclcic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE: DNA (oligonucleotide)
- (x i 1 SEQUENCE DESCRIPTION: SEQ ID NO:90:

TAGATCTAGA CCACCATGGT TTTCACACCT CAGATACTCA GACTCATGCT CTTCTGGATT 60

TCAGCCTCCA GAGGTGAAAT TGTGCTAACT CAGTCTCCAG GCACCCTAAG CTTATCACCG 120

GGAGAAAGG

			-continued			
(2) IN FORMATION	FORSEQ ID NO:91:					
	UENCE CHARACTERISTI (A) LENGTH: 128 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				
(i i) MOL	ECULE TYPE: DNA (oligo	onucleotide)			·	
(x i) SEQ	UENCE DESCRIPTION: SE	EQ ID NO:91:			35	
TAGACAGAAT	TCACGCGTAC	T T G A T A A G T A	GACGTGGAGC	TTGTCCAGGT	TTTTGTTGGT	6 0
ACCAGTGTAG	GTTGTTGCTA	A T A C T T T G G C	TGGCCCTGCA	GGAAAGTGTA	GCCCTTTCTC	1 2 0
CCGGTGAT						1 2 8
(2) INFORMATION	FOR SEQ ID NO:92:					
	UENCE CHARACIERISTI (A) LENG'III: 113 base (B) TYPE: nucleic neid (C) STRANDEDN'ESS: (D) TOPOLOGY: linear	pairs				
(ii)MOL	ECULE TYPE: DNA (of ag	onucleotide)				
(xi)SEQ	UENCE DESCRIPITON: SE	EQ ID NO:92:				
AAGAGAATTC	ACGCGTCCCA	GTCCATCTCT	GGAATACCCG	ATAGGTTCAG	TGGCAGTGGA	6 0
TCAGGGACAG	ATTTCACTCT	CACAATAAGT	AGGCTCGAGC	CGGAAGATTT	TGC	113
(2) INFORMATION	FOR SEQ ID NO:93:					
(i)SEQ	UENCE CHARACIERISTI (A) LENGTH: 116 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pa`us				
(ii) MOI	ECULE TYPE: DNA (oligo	vnucleutide)				
(xi)SEQ	UENCE DESCRIPTION: SI	EQ ID NO:93:				
TAGATCTAGA	GTTGAGAAGA	CTACTTACGT	TTTATTTCTA	CCTTGGTCCC	TTGTCCGAAC	6 0
GTATGAGGCC	AACTGTTACT	CTGTTGACAA	TAATACACAG	CAAAATCTTC	CGGCTC	116
(2) INFORMATION	FOR SEQ ID NO:94					
	UENCE CHARACIERISTI (A) LENGTH: 134 base (B) TYPE: nucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs				4
(i i) MO1	ECULE TYPE: DNA (oligo	onucleatide)				
(xi)SEQ	UENCE DESCRIPTION: SI	EQ ID NO:94:				
TATATCTAGA	CCACCATGGG	ATGGAGCTGG	ATCTTTCTCT	TCCTCCTGTC	AGGAACTGCA	6 0
GGTGTCCACT	CTCAAGTCCA	ACTGGTACAG	TCTGGAGCTG	AGGTTAAAAA	GCCTGGAAGT	1 2 0
TCAGTAAGAG	тттс					I 3 4
(2) INFORMATION	FOR SEQ ID NO:95:					
(i) SEQ	UENCE CHARACIERISTI (A) LENGTH: 134 base (B) TYPE: mucleic acid (C) STRANDEDNESS: (D) TOPOLOGY: linear	pairs single				

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	-continued
(i i) MGLECULE TYPE: DNA (oligonucleoside)	
(x i) SEQUENCE DESCREPTION: SEQ ID NO:95:	
TATATAGGTA CCACCATGGG ATGGAGCTGG	ATCTTTCTCT TCCTCCTGTC AGGAACTGCA 60
TGCCTGTCTC ACCCAGTTCA TGGTATACCC	AGTGAATGAG TATCCGGAAG CTTTGCAGGA 120
AACTCTTACT GAAC	134
(2) INFORMATION FOR SEQ ID NO.96:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 116 base pain; (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(i i) MOLECULE TYPE: DNA (oligonucle olide)	
(\times i) SEQUENCE DESCRIPTION: SEQ ID NO:96:	
TATATAGGTA CCAGCTACAA CCAGAAGTTC	AAGGGCAGAG TTACAGTTTC TTTGAAGCCT 60
TCATTTAACC AGGCCTACAT GGAGCTCAGT	AGTCTGTTTT CTGAAGACAC TGCAGT 116
(2) INFORMATION FOR SEQ ID NO:97:	
(i) SEQUENCE CHARACTERISTICS: (A) IENGTH: 116 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(i i) MOLECULE TYPE; DNA (oligonucleoide)	
(x i) SEQUENCE DESCRIPTION: SEQ ID NO.97:	
TATATCTAGA GGCCATTCTT ACCTGAGGAG	ACGGTGACTA AGGTTCCTTG ACCCCAGTAG 60
TCCATAGAAT AGTCTCGAAA CCCCCGTCTT	GTACAGTAAT AGACTGCAGT GTCTTC 116
(2) INFORMATION FOR SEQ ID NO:98:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 408 base pairs (B) TYPE: necleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(i i) MOLECULE TYPE:cDNA	
(i x) FEATURE: (A) NAME/KPY: CDS	
(B)LOCATION: 1,408	
(x i) SEQUENCE DESCRIPTION: SEQ ID NO:98:	
ATG CAT CAG ACC AGC ATG GGC ATC A Mct His GI m Thr Scr Mct Gly IIc L 1	
TTC ATA TCC ATA CTG CTC TGG TTA T	AT GGT GCT GAT GGG AAC ATT GTT 96
Phe lle Ser lle Leu Leu Trp Leu T 20	
ATG ACC CAA TCT CCC AAA TCC ATG T Met Thr Gin Ser Pro Lys Ser Met T 35	
ACC TTG AGC TGC AAG GCC AGT GAA A Thr Leu Ser Cys Lys Ala Ser Glu A 50 55	
TAT CAA CAG AAA CCA GAG CAG TCT C Tyr Gln Gln Lys Pro Glu Gln Ser P 65 70	

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151
                                                                                152
                                                -continued
TCC AAC CGG TAC ACT GGG GTC CCC GAT CGC TTC ACG GGC AGT GGA TCT
                                                                                                   288
Scr Asm Arg Tyr Thr Gly Val Pro Asp Arg Phe Thr Gly Scr Gly Scr
GCA ACA GAT TTC ACT CTG ACC ATC AGC AGT GTG CAG OCT GAA GAC CTT Ala Thr Asp Phe Thr Leu Thr Ile Ser Ser Val Gin Ala Giu Asp Leu 100
                                                                                                   3 3 6
GCA GAT TAT CAC TGT GGA CAG AGT TAC AAC TAT CCA TTC ACG TTC GGC Ala Asp Tyr His Cys Gly Glu Ser Tyr Asn Tyr Pro Phc Tbr Pbc Gly
TCG GGG ACA AAG TTG GAA ATA AAG
                                                                                                   408
Ser Gly Thr Lys Len Glo Ile Lys
( 2 ) INFORMATION FOR SEQ ID NO:99:
       ( i ) SEQUENCE CHARACTERISTICS:
               ( A ) LENGTH: 136 amino acids
               ( B ) TYPE: amino acid
              ( D ) TOPOLOGY: linear
      ( i i ) MOLECULE TYPE: protes n
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:99:
Mcc His Gla Thr Ser Mec Gly 11c Lys Mei Gla Ser Gla Thr Lea Val
Phe II c Ser II e Leu Leu Trp Leu Tyr Gly Ala Asp Gly Asn II c Val 20
Met Thr Gln Ser Pro Lys Ser Met Tyr Val Ser 11c Gly Glu Arg Val
Thr Leu Ser Cys Lys Ala Ser Glu Asn Val Asp Thr Tyr Val Ser Trp
Tyr Gln Gln Lys Pro Glu Gln Scr Pro Lys Lcu Lcu 11e Tyr Gly Ala
65 70 80
Ser Asn Arg Tyr Thr Gly Val Pro Asp Arg Phe Thr Gly Scr Gly Ser 95
Ala Thr Asp Phe Thr Leu Thr lie Ser Ser Val Gln Ala Glu Asp Leu
100 105
                                             1 0 5
Ala Asp Tyr His Cys Gly Gla Ser Tyr Asa Tyr Pro Phe Thr Phe Gly
115 120
Scr Gly Thr Lys Leu Glu lic Lys
( 2 ) INFORMATION FOR SEQ ID NO:100:
       ( i ) SEQUENCE CHARACTERISTICS:
              ( A ) LENGTH: 456 base Pairs
              ( B ) TYPE: nucleic acid
              ( C ) STRANDEDNESS: single
              ( D ) TOPOLOGY: l'incar
      ( i i ) MOLECULE TYPE: cDNA
      ( i x ) FEATURE:
               ( A ) NAMEKEY: CDS
               ( B ) LOCATION: 1.456
      ( x i ) SEQUENCE DESCRIPTION: SEQ ID NO:100:
ATG ACA TCA CTG TTC TCT CTA CAG TTA CCG AGC ACA CAG GAC CTC GCC Mei Thr Ser Leu Phe Ser Leu Gln Leu Pro Ser Thr Gln Asp Leu Ala
                                                                                                     4 8
ATG GGA TGG AGC TGT ATC ATC CTC TTC TTG GTA GCA ACA GCT ACA GGT Mci Gly Trp Scr Cys Iic lle Leu Phe Leu Val Ala Thr Ala Thr Gly
```

(2) INFGRMATION FOR SEQ ID NO:101:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 152 am ino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(i i) MOLECULE TYPE: protein

(\times i) SEQUENCE DESCRIPTION: SEQ ID NG:101:

 McI
 Thr
 Scr
 Lcu
 Phc
 Scr
 Lcu
 Gln
 Lcu
 Pro
 Scr
 Thr
 Gln
 Asp
 Lcu
 Ala

 McI
 Gly
 Trp
 Scr
 Cys
 llc
 llc
 Lcu
 Phc
 Lcu
 Val
 Ala
 Thr
 Aln
 Thr
 Aln
 Thr
 Gly
 Aln
 Thr
 Gly
 Aln
 Thr
 Gly
 Aln
 Thr
 Gly
 Aln
 Thr
 Aln
 Thr
 Gly
 Aln
 Aln
 Thr
 Gly
 Aln
 Aln

(2) INFORMATION FOR SEQ ID NO:102:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 107 amino acids

155 156

(B) TYPE: amino acid

- (C) STRANDEDNESS: single
- (O) TOPOLOGY: linear
- (i i) MOLECULE TYPE: peptide
- (x i) SEQUENCE DESCRIPTION: SEQ ID NO:102:

 Asp | 11 c | Gln | Mc | Thr | Gln | Scr | Pro | Scr | Thr | Leu | Scr | Ala | Scr | Val | Gly | 15

 Asp | Arg | Val | Thr | Ilc | Thr | Cys | Arg | Ala | Scr | Gln | Scr | Ilc | Asn | Thr | Trp | 20

 Leu | Ala | Trp | Tyr | Gln | Gln | Lys | Pro | Gly | Lys | Ala | Pro | Lys | Leu | Leu | Mc | 45

 Tyr | Lys | Ala | Scr | Scr | Leu | Glu | Scr | Gly | Val | Pro | Scr | Arg | Phe | Ilc | Gly | Scr | Gly | Scr | Gly | Thr | Glu | Phe | Thr | Leu | Thr | Ilc | Scr | Scr | Leu | Gln | Pro | Scr | Asp | Scr | Asp | Scr | Lys | Scr | Asp | Scr | Lys | Scr | Cly | Cly

- (2) INFORMATION FOR SEQ ID NO:103:
 - (i) SEQUENCE CHARACIERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: anino scid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: l'incar
 - (i i) MOLECULE TYPE: pe pide
 - (x i) SEQUENCE DESCRIPTION: SE€ ID NO:103:

 Asp II c
 Gln Met
 Thr S
 Gln Scr Pro Scr Thr IO
 Leu Scr Ala Scr Ala Scr Val Gly

 Asp Arg Val Thr II c
 Thr Cys Lys Ala Scr Glu Asn Val Asp Thr Tyr

 Val Scr Trp 35
 Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu II c

 Tyr Gly Ala Scr Asn Arg Tyr Thr Gly Val Pro Scr Arg Pbc Ser Gly 55

 Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Scr Scr Leu Gln Pro 65

 Asp Pbc Ala Thr Tyr Tyr Cys Gly Gln Scr Tyr Asn Tyr Pro 95

 Thr Pbc Gly Gln Gln Gly Thr Lys Val Glu Val Lys 105

- (2) INFORMATION FOR SEQ ID NO:104:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 117 amino acids
 - (B) TYPE: animo acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
 - (i i) MOLECULE TYPE: peptide
 - (x i) SEQUENCE DESCRIPTION: SEQ ID NO:104:

-continued Ile lle Trp Val Arg Gln Ala Pro Gly Gln Gly Leu Glu Trp Met 35 40 45 Gly Oly Tyr Gly ile Tyr Ser Pro Glu Glu Tyr Asm Gly Gly Leu 100 105 110 (2) INFORMATION FOR SEQ ID NO:105: (i) SEQUENCE CHARACIERISTICS: (A) LENGTH: 117 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: hinear (i i) MOLECULE TYPE: popude (* i) SEQUENCE DESCRIPTION: SEQ ID NO:105: Gin Val Gin Leu Val Gin Ser Gly Ala Glu Val Lys Lys Pro Gly Ser l 10 15 Val Lys Val Ser Cys Lys Ala Ser Gly Tyr lle Phe Thr Ser Ser 20 25 30 Trp Ile Asn Trp Vei Arg Gln Ala Pro Gly Gln Gly Leu Glu Trp Mei 35 40 45 Gly Arg llc Asp Pro Ser Asp Gly Glu Val His Tyr As π Gln Asp Pbc 50 60 Val Thr Val Ser Ser 1 1 5 (2) INFORMATION FOR SEQ ID NO:106. (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 115 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligenucleolide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:106:

TTTTTTCTAG ACCACCATGG AGACCGATAC CCTCCTGCTA TGGGTCCTCC TGCTATGGGT 6 0 CCCAGGATCA ACCGGAGATA TTCAGATGAC CCAGTCTCCG TCGACCCTCT CTGCT 115

(2) INFORMATION FOR SEO ID NO:107:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGIH: 120 hase pairs
 - (B) TYPE: nucleic acid

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159 160

-continued (C) ST'RANDEDNESS: single (D) TOPOLOOY: linear (i i) MOLECULE TYPE: DNA (oligonucleou'de) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:107; TTTTAAGCTT GGGAGCTTTG CCTGGCTTCT GCTGATACCA GGATACATAA GTATCCACAT 6 0 TTTCACTGGC CTTGCAGGTT ATGGTGACCC TATCCCCGAC GCTAGCAGAG AGGGTCGACG 120 (2) INTORMATION FOR SEQ ID NO:108: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 118 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS; single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:108: TTTTAAGCTT CTAATTTATG GGGCATCCAA CCGGTACACT GGGGTACCTT CACGCTTCAG 6 0 TGGCAGTGGA TCTGGGACCG ATTTCACCCT CACAATCAGC TCTCTGCAGC CAGATGAT 1 1 8 (2) INFORMATION FOR SEQ ID NO:109: (i) SEQUENCE CHARACIERISTICS: (A) LENGTH: 120 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS; single (D) TOPOLOGY. linear (i i) MOLECULE TYPE: DNA (oligonucleoride) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:109: TTTTTTCTAG AGCAAAAGTC TACTTACGTT TGACCTCCAC CTTGGTCCCC TGACCGAACG 6 0 TGAATGGATA GTTGTAACTC TGTCCGCAGT AATAAGTGGC GAAATCATCT GGCTGCAGAG 120 (2) INFORMATION FOR SEQ ID NO:110: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 114 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleotide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:110: 6 0 TTTTTCTAGA CCACCATGGG ATGGAGCTGG ATCTTTCTCT TCCTCCTGTC AGGTACCGCG GGCGTGCACT CTCAGGTCCA GCTTGTCCAG TCTGGGGCTG AAGTCAAGAA ACCT 114 (2) INFORMATION FOR SEQ ID NO:111: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 121 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (i i) MOLECULE TYPE: DNA (oligonucleoide) (x i) SEQUENCE DESCRIPTION: SEQ ID NO:111: TTTTGAATTC TCGAGACCCT GTCCAGGGGC CTGCCTTACC CAGTTTATCC AGGAGCTAGT 6 0 AAAGATGTAG CCAGAAGCTT TGCAGGAOAC CTTCACGGAG CTCCCAGGTT TCTTGACTTC 120 A 121

CAGGCTGCTC	AGTT					1 3 4		
GTCAGCAAAC	CAGGGCAGAA	ATCCTCTTGC	АСАСТААТАС	ACTGCAGTGT	CCTCTGATCT	120		
TTTTCTAGA	GGTTTTAAGG	ACTCACCTGA	GGAGACTGTG	ACCAGGGTTC	CTTGGCCCCA	6 0		
(x i) SEQU	IENCE DESCRIPTION: SI	EQ ID NO:113:						
(ii) MOL	ECUTE TYPE: DNA (oligo	procleotide)						
(D) TOPOLOGY: linear								
(C) STRANDEDNESS: single								
(A) LENGH: 134 base pairs (B) TYPE: nucleic acid								
(i) SEQUENCE CHARACIERISTICS:								
(2) INFORMATION	FOR SEQ ID NO:113:							
GAACTGAGCA	GCCTGAG					1 3 7		
CAAGATTTCA	AGGACCGTGT	TACAATTACA	GCAGACGAAT	CCACCAATAC	AGCCTACATG	120		
TTTTGAATTC	TCGAGTGGAT	GGGAAGGATT	GATCCTTCCG	ATGGTGAAGT	TCACTACAAT	6 0		
(x i) SEQU	ENCE DESCRIPTION: SI	EQ ID NO:112:						
(ii) MOL	ECULE TYPE: DNA (eligo	enuciceúde)						
1	(D) TOPOLOGY: linear							
(C) STRANDEDNESS: single								
	(B) TYPE: model's acid							
(i) SEQUENCE CHARACIERISTICS: (A) LENGTH: 137 base pairs								
4 1 1 1 1 1 1 1 1 1	T. C.							
(2) INFORMATION I	FOR SEQ ID NO:112:							

What is claimed is:

- 1. A humanized immunoglobulin having complementarity determining regions (CDRs) from a donor immunoglobulin and heavy and light chain variable region frameworks from human acceptor immunoglobulin heavy and light chain frameworks, which humanized immunoglobulin specifically binds to a human interleukin-2 receptor with an affinity constant of at least 10⁷ M⁻¹ and no greater than about two-fold that of the donor immunoglobulin, wherein the sequence of the humanized immunoglobulin heavy chain variable region framework is 65% to 95% identical to the sequence of the donor immunoglobulin heavy chain variable region framework.
- 2. A humanized immunoglobulin according to claim 1 which is an antibody comprising two light chain/heavy chain dimers 50
- A humanized immunoglobulin according to claim 1, which specifically binds to a human interleukin-2 receptor with an affinity of at least 10⁸ M⁻¹.
- A humanized immunoglobulin according to claim 1, wherein said human interleukin-2 receptor is the p55 Tac interleukin-2 receptor.
- 5. A humanized immunoglobulin according to claim 1, wherein said human interleukin-2 receptor is the p75 interleukin-2 receptor.
 - 6. A humanized immunoglobulin according to claim 1,

- 95 wherein said donor immunoglobulin is the anti-Tac antibody.
 - 7. A humanized immunoglobulin according to claim 1, wherein said acceptor immunoglobulin heavy and light chain frameworks are from the same human antibody.
 - A humanized immunoglobulin according to claim 7, wherein said human antibody is the Eu human antibody.
 - 9. A humanized immunoglobulin according to claim 1 which is substantially pure.
 - 10. A humanized immunoglobulin according to claim 1 that competitively inhibits the binding of interleukin-2 to said human interleukin-2 receptor.
- 11. A recombinant immunoglobulin which specifically binds to the p55 chain of the human IL-2 receptor, wherein the amino acid sequence of the mature light chain variable region is:
- DI QMTQSPSTLSASVGDRVTITCSASSSIS YMHWYQQKPGKAPKLLIYTTSNLASGVPAR FS GS GSGTEFTLTISSLQPDDFATYYCHQR STYP LTFGQGTKVEVK.
- 12. A recombinant immunoglobulin which specifically binds to the p55 chain of the human IL-2 receptor, wherein the amino acid sequence of the mature heavy chain variable region is:

Q V Q L V Q S G A E V K K P G S S V K V S C K A S G Y T F T S Y R M H W V R Q A P G Q G L E W I G Y I N P S T G Y T E Y N Q K F K D K A T I T A D E S T N T A Y M E L S S L R S E D T A V Y Y C A R G G G V F D Y W G Q G T L V T V S S.

13. An immunoglobulin which specifically binds to the p55 chain of the human IL-2 receptor, said immunoglobulin having two pairs of heavy and light chains wherein:

each light chain comprises a complete human kappa chain constant region sequence and a variable region sequence:

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and

each heavy chain comprises a complete human γ chain constant region sequence and a variable region sequence:

Q V Q L V Q S G A E V K K P G S S V K V S C K A S G Y T F T S Y R M H W V R Q A P G Q G L E W I G Y I N P S T G Y T E Y N Q K F K D K A T 1 T A D E S T N T A Y M E L S S L R S E D T A V Y Y C A R G G G V F D Y W G Q G T L V T V S S.

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