

# A-498 (ATCC® HTB-44™)

Organism: Homo sapiens, human / Tissue: kidney / Disease: carcinoma

GENERAL INFORMATION	CHARACTERISTICS	CULTURE METHOD	SPECIFICATIONS	HISTORY	DOCUMENTATION	SHARE	EMAIL	PRINT
<b>Permits and Restrictions</b> <input type="button" value="View Permits"/>								
<b>Organism</b>	<i>Homo sapiens, human</i>							
<b>Tissue</b>	kidney							
<b>Product Format</b>	frozen							
<b>Morphology</b>	epithelial							
<b>Culture Properties</b>	adherent							
<b>Biosafety Level</b>	1							
	<i>Biosafety classification is based on <u>U.S. Public Health Service Guidelines</u>, it is the responsibility of the customer to ensure that their facilities comply with biosafety regulations for their own country.</i>							
<b>Disease</b>	carcinoma							
<b>Age</b>	52 years							
<b>Gender</b>	female							
<b>Storage Conditions</b>	liquid nitrogen vapor phase							

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<b>Derivation</b>	S. Aaronson isolated this line using techniques as described for ATCC HTB-41.							
<b>Clinical Data</b>	female							
<b>Tumorigenic</b>	Yes							
<b>Effects</b>	Yes, in nude mice; forms undifferentiated carcinoma; also forms tumors in anti thymocyte serum treated newborn mice							
<b>Comments</b>								

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<b>Complete Growth Medium</b>	The base medium for this cell line is ATCC-formulated Eagle's Minimum Essential Medium, Catalog No. 30-2003. To make the complete growth medium, add the following components to the base medium: fetal bovine serum to a final concentration of 10%.							
<b>Subculturing</b>	<ol style="list-style-type: none"><li>1. Remove and discard culture medium.</li><li>2. Briefly rinse the cell layer with 0.25% (w/v) Trypsin - 0.53 mM EDTA solution to remove all traces of serum which contains trypsin inhibitor.</li><li>3. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes). Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.</li><li>4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting.</li><li>5. Add appropriate aliquots of the cell suspension to new culture vessels.</li><li>6. Incubate cultures at 37°C.</li></ol> <p><b>Subcultivation Ratio:</b> A subcultivation ratio of 1:3 to 1:8 is recommended <b>Medium Renewal:</b> Twice per week</p>							
<b>Cryopreservation</b>	<p><b>Freeze medium:</b> culture medium, 95%; DMSO, 5% <b>Storage temperature:</b> liquid nitrogen vapor phase</p>							
<b>Culture Conditions</b>	<p><b>Atmosphere:</b> air, 95%; carbon dioxide (CO<sub>2</sub>), 5% <b>Temperature:</b> 37°C</p>							

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<b>STR Profile</b>		Amelogenin: X CSF1PO: 11,12 D13S317: 12 D16S539: 12 D5S818: 11,13 D7S820: 10,11 THO1: 6,9,3 TPOX: 8,11 vWA: 18						
<b>Isoenzymes</b>		AK-1, 1 ES-D, 2 G6PD, B GLO-I, 2 Me-2, 1 PGM1, 1-2 PGM3, 1						

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<b>Name of Depositor</b>	W Nelson-Rees							
<b>Deposited As</b>	<i>Homo sapiens</i>							
<b>References</b>	<p>Fogh J, et al. Absence of HeLa cell contamination in 169 cell lines derived from human tumors. J. Natl. Cancer Inst. 58: 209-214, 1977. PubMed: <a href="#">833871</a></p> <p>Goodfellow M, et al. One hundred and twenty-seven cultured human tumor cell lines producing tumors in nude mice. J. Natl. Cancer Inst. 59: 221-226, 1977. PubMed: <a href="#">77210034</a></p> <p>Faust JB, Meeker TC. Amplification and expression of the bcl-1 gene in human solid tumor cell lines. Cancer Res. 52: 2460-2463, 1992. PubMed: <a href="#">1568216</a></p> <p>Giard DJ, et al. In vitro cultivation of human tumors: establishment of cell lines derived from a series of solid tumors. J. Natl. Cancer Inst. 51: 1417-1423, 1973. PubMed: <a href="#">4357758</a></p> <p>Fogh J. Cultivation, characterization, and identification of human tumor cells with emphasis on kidney, testis, and bladder tumors. Natl. Cancer Inst. Monogr. 49: 5-9, 1978. PubMed: <a href="#">571047</a></p>							

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