
EXHIBIT 2008

Photoplethysmography (PPG) is a noninvasive optical method accepted in the clinical use for oxygen saturation. It is widely believed that the light intensity after interaction with the bio is modulated at the heartbeat frequency mainly due to pulsatile variations of the light absorption volume pulsations. Here we report experimental observations, which are not consistent with the importance of elastic deformations of the capillary bed in the formation of the PPG waveform, a new insight on light interaction with live tissue. To explain the observations we propose a model of pulse oscillations of the arterial transmural pressure deform the connective-tissue component, causing periodical changes of both the light scattering and absorption. These local changes of the light intensity are detected as variations of the light intensity returned to a photosensitive camera. Therefore, pulse is indirectly monitored even by using the light, which slightly penetrates into the biological tissue.

Introduction

It is commonly accepted that photoplethysmography (PPG) is a non-invasive optical method for measuring blood volume changes in the microvascular bed of tissue^{1,2,3}. PPG waveforms are used for cardiac output, oxygen saturation and heart rate determination in pulse oximeters which are widely accepted for remote monitoring. Conventional pulse oximeters operate at the red and infrared light, and use the transmission mode geometry of photoplethysmography in which the light source and photodetector are situated at the opposite sides of the tissue across a fingertip or an earlobe^{2,4}. Such geometry provides efficient interaction of the light with the tissue, which includes the optical processes of multiple scattering, absorption, reflection, and transmission. The main factors of the light intensity received by the photodetector are the blood volume, blood vessel wall motion, and the presence of red blood cells^{7,8,9}. The general consensus is that the PPG waveform originates from pulse-induced changes of optical density caused by arterial pulsations^{9,10}. Capillaries are the most sensitive to the variations of their size¹¹.

Imaging photoplethysmography (iPPG) is a non-contact imaging method for mapping cardiac output. iPPG usually operate in reflection mode where both the illuminating light source and photodetector are situated alongside each other. In the reflection-mode PPG, light interaction with the tissue is more efficient than in the transmission mode. Attempts to develop imaging pulse oximeter were made, which penetrates into the tissue for several millimetres, e.g., 2.5 mm at 660 nm. In 2013, researchers from different groups reported observation of a large-amplitude modulation of the PPG waveform under green light illumination (wavelength 532 nm) in imaging photoplethysmography^{15,16,17,18,19,20,21,22}. This observation is difficult to

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Results

Mapping the areas with maximal PPG amplitude

The measurements were carried out with 58 healthy subjects. Mapping the PPG signal amplitude over the arm areas was done by using an iPPG system with lock-in pixel amplification referred to as BloodFlow, which was developed in our group^{20,25}. Before measurement the subject placed his palm on a flat surface. In a comfortable position the subject was asked to avoid hand movements during 30 seconds of video recording. A green light (wavelength of 525 nm) for illumination of the subject's arm.

Two typical examples of the PPG-amplitude distribution are shown in Fig. 1a,b where the arm is overlaid with the initial camera image to provide an anatomic reference of the respective subject. The amplitude is calculated as an AC/DC ratio of the PPG waveform where AC and DC refer to the alternating and average (slowly varying) portions of the detected photo-signals, respectively. The amplitude is represented in pseudo colours so that the red colour corresponds to the higher amplitude. It is evident from the maps that the pulsations are unevenly distributed over the subjects' arms. We found that these maps vary significantly from subject to another. Evidently, for each subject we can find an area with the maximal PPG-amplitude. We map the position of these spots in respect to the anatomical scheme of the arm separately for the left (Fig. 1c) and right (Fig. 1d) hands. In these maps, each hot spot belongs to a specific subject. It is clearly seen that the most of hot spots are situated near either radial or ulnar arteries. There are subjects in whom the hottest spots are in the thumb or in the little finger. Anyway, the hottest spots are observed in the vicinity of arteries. The amplitude of the blood pulsations in the hottest spots varies between 1.5% and 5.7%. Note that these values represent the AC/DC ratio, i.e. the amplitude of the alternating portion at the heartbeat frequency in relation to the magnitude of the slowly varying mean PPG signal. The amplitude of the hottest spot over all subjects was 2.2% (standard deviation, STD = 0.9%) for the left hand and 2.1% for the right hand.

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Two examples of spatial distribution of the PPG amplitude (a) and (b) overlaid with raw camera images measured in two positions of the spots in which we observed the maximal PPG amplitude for the left hand (c) and for the right hand (d) as a black circle for one subject. The colour scale on the right shows the pulsation amplitude as the AC/DC ratio of the PPG wa

Counter phase PPG waveforms

Experimental observation of the PPG waveforms pulsating in the counter phase was recently reported [1]. In this study, taking an advantage of the BPI system to visualize the spatial distribution of the relative phase of the PPG signals, in the current study, we considerably extended the cohort of subjects and confirmed that all of the PPG signals in the adjacent areas in which the PPG waveforms oscillate with the same heart rate but in the counter phase. The spatial distribution of such signals is shown in Fig. 2 where in Fig. 2a we show the map of PPG-amplitude for a subject. The area of maximal amplitude is situated nearby the radial artery. An enlarged image patch of the area containing the maximal amplitude is shown in Fig. 2b. A fragment of the PPG phase distribution of the same area is shown in Fig. 2c. The phase difference between pulsations in the adjacent areas within the black circle is equal to π . In Fig. 2d, we show three dimensional (3D) plot of the pulsation-amplitude distribution considering the relative phase of the PPG signals in the area within the black circle in Fig. 2b,c. Note that two hot spots within this circle are separated by a distance of only 3.3 mm.

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and position as the patch (b). Distribution of PPG pulsations (c) considering their relative phase in the area within the black circle. The waveforms (e) calculated in ROIs of 3×3 pixels chosen in the points with extreme amplitude within the black circle. The pulsations maps are in degrees and in per cents for phase and amplitude PPG maps, respectively.

After determination the positions of the hot spots in the PPG-amplitude map in Fig. 2a, we defined regions of interest (ROI) in the points where two extremes of the amplitude were found. In these ROIs we averaged PPG signals by the spatial averaging the pixel values within the ROIs of the size 3×3 pixels, area of $0.75 \times 0.75 \text{ mm}^2$ at the palm. These waveforms are shown in Fig. 2e. By convention used in photoplethysmography literature^{3,5}, all PPG waveforms in this paper are inverted in sign so that they correspond with varying transmural pressure. Therefore, the negative extremes of the PPG waveform correspond to the phase. As one can see, the PPG signals in these adjacent areas indeed oscillate in the counter-phase. That the red curve in Fig. 2e has a typical saw-tooth shape with the faster transition from the trough to the peak is always observed in measurements of the arterial blood pressure waveforms^{5,26}. In contrast, the PPG waveform time and its shape clearly contradicts with behaviour of the arterial pressure. Erroneously, the counter-phase PPG waveforms were attributed with asynchronous blood supply to the adjacent areas. We tried to explain their origin in the frames of the commonly accepted model of PPG in which the signal is caused by variation in the tissue optical density due to the blood-volume pulsations in arteries¹⁰. In this paper we propose the new model of the PPG signal formation suggesting indirect impact of arterial blood pulsations on PPG modulation.

Influence of the external local pressure

The experimental data presented in Figs. 1 and 2 were obtained in conditions when the subject's hand was in contact with the glass table. In the next experiment we studied the influence of the mechanical contact on the PPG signal. At the end, a subject was asked to put his hand on the glass table while the illuminator (green light) and recording camera were settled under the table. To exclude direct reflections from the glass surface, we used polarized light and a polarization filter installed before the camera with orthogonal orientation to the polarization of the incident light. There were three steps of iPPG recordings. First recording was done without contact of the skin with glass. In the second step, the contact of the palm with glass was established. In the third step, additional weight of 1.7 kg was applied to the hand in the third step. An image of the palm with this weight to the skin in the areas of the contact with the glass was es

The results of the experiment are shown in Fig. 3 where spatial distribution of PPG signals in three steps 1, 2, and 3 are presented by images a, b, and c, respectively. As or

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