

### Notice

Medicine is an ever-changing science. As new research and clinical experience broaden our knowledge, changes in treatment and drug therapy are required. The authors and the publisher of this work have checked with sources believed to be reliable in their efforts to provide information that is complete and generally in accord with the standards accepted at the time of publication. However, in view of the possibility of human error or changes in medical sciences, neither the authors nor the publisher nor any other party who has been involved in the preparation or publication of this work warrants that the information contained herein is in every respect accurate or complete, and they disclaim all responsibility for any errors or omissions or for the results obtained from use of the information contained in this work. Readers are encouraged to confirm the information contained herein with other sources. For example and in particular, readers are advised to check the product information sheet included in the package of each drug they plan to administer to be certain that the information contained in this work is accurate and that changes have not been made in the recommended dose or in the contraindications for administration. This recommendation is of particular importance in connection with new or infrequently used drugs.

# Goodman & Gilman's The Pharmacological Basis of THERAPEUTICS

twelfth edition

### editor

Laurence L. Brunton, PhD
Professor of Pharmacology and Medicine
School of Medicine, University of California, San Diego
La Jolla, California

### associate editors

Bruce A. Chabner, MD

Professor of Medicine
Harvard Medical School
Director of Clinical Research
Massachusetts General Hospital Cancer Center
Boston, Massachusetts

Björn C. Knollmann, MD, PhD Professor of Medicine and Pharmacology

Professor of Medicine and Pharmacology
Oates Institute for Experimental Therapeutics
Division of Clinical Pharmacology
Vanderbilt University School of Medicine
Nashville, Tennessee



New York Chicago San Francisco Lisbon London Madrid Mexico City Milan New Delhi San Juan Seoul Singapore Sydney Toronto

### Goodman and Gilman's

### The Pharmacological Basis of Therapeutics, Twelfth Edition

Copyright © 2011, 2006, 2001, 1996, 1990, 1985, 1980, 1975, 1970, 1965, 1955, 1941 by The McGraw-Hill Companies, Inc. All rights reserved. Printed in China. Except as permitted under the United States Copyright Act of 1976, no part of this publication may be reproduced or distributed in any form or by any means, or stored in a database or retrieval system, without the prior written permission of the publisher.

4 5 6 7 8 9 0 CTP/CTP 17 16 15 14

ISBN 978-0-07-162442-8 MHID 0-07-162442-2

Book ISBN 978-0-07-175352-4 Book MHID 0-07-175352-4

DVD ISBN 978-0-07-175306-7 DVD MHID 0-07-175306-0

Set ISBN 978-0-07-162442-8 Set MHID 0-07-162442-2

This book was set in Times by Glyph International.

The editors were James F. Shanahan and Christie Naglieri.

The production manager was Sherri Souffrance.

Project management was provided by Rajni Pisharody, Glyph International.

The illustration manager was Armen Ovsepyan.

The designer was Janice Bielawa.

The cover art director was Anthony Landi; the cover designer was Thomas De Pierro.

The indexer was Coughlin Indexing Services.

China Translation & Printing Services, Ltd., was printer and binder.

This book is printed on acid-free paper.

### Library of Congress Cataloging-in-Publication Data

Goodman & Gilman's pharmacological basis of therapeutics.—12th ed. / editor,

Laurence L. Brunton ; associate editors, Bruce A. Chabner, Björn C. Knollmann.

p.; cm.

Other title: Goodman and Gilman's pharmacological basis of therapeutics

Other title: Pharmacological basis of therapeutics

Rev. ed. of: Goodman & Gilman's the pharmacological basis of therapeutics. 11th ed. / editor,

Laurence L. Brunton, c2006.

Includes bibliographical references and index.

ISBN-13: 978-0-07-162442-8 (hardcover : alk. paper)

ISBN-10: 0-07-162442-2

1. Pharmacology. 2. Therapeutics. I. Goodman, Louis Sanford, 1906- II. Brunton, Laurence L.

III. Chabner, Bruce. IV. Knollmann, Björn C. V. Goodman & Gilman's the pharmacological

basis of therapeutics. VI. Title: Goodman and Gilman's pharmacological basis of therapeutics.

VII. Title: Pharmacological basis of therapeutics.

[DNLM: 1. Pharmacological Phenomena. 2. Drug Therapy. QV 4 G6532 2011]

RM300.G644 2011

615'.7-dc22

2010000236

McGraw-Hill books are available at special quantity discounts to use as premiums or sales promotions, or for use in corporate training programs. To contact a representative, please email us at bulksales@mcgraw-hill.com.

In Memoriam Keith L. Parker (1954-2008)

# Contributors

### Edward P. Acosta, PharmD

Professor of Clinical Pharmacology University of Alabama, Birmingham

# Peter J. Barnes, DM, DSc, FRCP, FMedSci, FRS

Professor and Head of Respiratory Medicine National Heart & Lung Institute Imperial College, London

### Jeffrey A. Barnes, MD, PhD

Fellow in Hematology-Oncology Dana-Farber Cancer Institute Boston, Massachusetts

### Leslie Z. Benet, PhD

Professor of Bioengineering and Therapeutic Sciences Schools of Pharmacy and Medicine University of California, San Francisco

### John E. Bennett, MD

Chief of Clinical Mycology National Institute of Allergy and Infectious Diseases Bethesda, Maryland

### William Bennett, MD

Professor (Emeritus) of Medicine and Pharmacology Oregon Health & Science University, Portland

### Thomas P. Bersot, MD, PhD

Professor of Medicine; Associate Investigator Gladstone Institute of Cardiovascular Disease University of California, San Francisco

### Joseph R. Bertino, MD

Professor of Medicine and Pharmacology Robert Wood Johnson Medical School University of Medicine & Dentistry of New Jersey New Brunswick

### Donald K. Blumenthal, PhD

Associate Professor of Pharmacology & Toxicology College of Pharmacy University of Utah, Salt Lake City

### Viengngeun Bounkeua, PhD

Medical Scientist Training Program, School of Medicine University of California, San Diego

### Gregory A. Brent, MD

Professor of Medicine and Physiology Geffen School of Medicine University of California, Los Angeles

### Joan Heller Brown, PhD

Professor and Chair of Pharmacology University of California, San Diego

### Craig N. Burkhart, MD

Assistant Professor of Dermatology, School of Medicine University of North Carolina, Chapel Hill

### Iain L. O. Buxton, PharmD

Professor of Pharmacology University of Nevada School of Medicine, Reno

### Michael C. Byrns, PhD

Fellow in Pharmacology University of Pennsylvania School of Medicine, Philadelphia

### William A. Catterall, PhD

Professor and Chair of Pharmacology University of Washington School of Medicine, Seattle Cancer Center

Michael W.H. Coughtrie, PhD Professor of Biochemical Pharmacology

Division of Medical Sciences University of Dundee, Scotland David D'Alessio, MD

Jérôme Clain, PharmD, PhD

Columbia University, New York

Dana-Farber Cancer Institute

Attending Physician

Boston, Massachusetts

Professor of Endocrinology and Medicine University of Cinncinnati, Ohio

Professor of Medicine, Harvard Medical School

Director of Clinical Research, Massachusetts General Hospital

Richard T. Eastman, PhD Fellow in Microbiology Columbia University, New York

Ervin G. Erdös, MD Professor (Emeritus) of Pharmacology University of Illinois-Chicago

David A. Fidock, PhD Associate Professor of Microbiology and Medicine College of Physicians and Surgeons Columbia University, New York

Garret A. FitzGerald, MD Professor of Medicine, Pharmacology and Translational Medicine and Therapeutics; Chair of Pharmacology University of Pennsylvania School of Medicine, Philadelphia

Charles W. Flexner, MD Professor of Medicine, Pharmacology and Molecular Sciences, and International Health The Johns Hopkins University School of Medicine and Bloomberg School of Public Health Baltimore, Maryland

Peter A. Friedman, PhD

Professor of Pharmacology and Chemical Biology School of Medicine University of Pittsburgh, Pennsylvania

John W. Funder, AO, MD, BS, PhD, FRACP Professor of Medicine, Prince Henry's Institute Monash Medical Centre ClaytonVictoria, Australia

James C. Garrison, PhD Professor of Pharmacology, School of Medicine University of Virginia, Charlottesville

Kathleen M. Giacomini, PhD Professor and Chair of Biopharmaceutical Sciences School of Pharmacy University of California, San Francisco

Alfred G. Gilman, MD, PhD Professor (Emeritus) of Pharmacology University of Texas Southwestern Medical School Chief Scientific Officer, Cancer Prevention and Research Institute of Texas, Dallas

Lowell A. Goldsmith, MD, MPH Professor of Dermatology, School of Medicine University of North Carolina, Chapel Hill, North Carolina

Frank J. Gonzalez, PhD Chief, Laboratory of Metabolism Center for Cancer Research, National Cancer Institute Bethesda, Maryland

Tilo Grosser, MD Assistant Professor of Pharmacology Institute for Translational Medicine and Therapeutics University of Pennsylvania, Philadelphia

Tawanda Gumbo, MD Associate Professor of Internal Medicine University of Texas Southwestern Medical School, Dallas

Stephen R. Hammes, MD, PhD Professor of Medicine, Chief of Endocrinology and Metabolism School of Medicine and Dentistry University of Rochester, New York

R. Adron Harris, PhD

Professor of Molecular Biology; Director, Waggoner Center for Alcohol and Addiction Research University of Texas, Austin

Lisa A. Hazelwood, PhD Research Fellow, Molecular Neuropharmacology Section National Institute of Neurological Disorders and Stroke Bethesda, Maryland

Jeffrey D. Henderer, MD

Professor and Chair of Ophthalmology Temple University School of Medicine Philadelphia, Pennsylvania

Rvan E. Hibbs, PhD

Research Fellow, Vollum Institute Oregon Health & Science University, Portland

Randa Hilal-Dandan, PhD Lecturer in Pharmacology University of California, San Diego

Brian B. Hoffman, MD Professor of Medicine, Harvard Medical School Physician, VA-Boston Health Care System Boston, Massachusetts

Peter J. Hotez, MD, PhD Professor and Chair of Microbiology, Immunology, and Tropical Medicine George Washington University Washington, DC

Nina Isoherranen, PhD Assistant Professor of Pharmaceutics, School of Pharmacy University of Washington, Seattle

Edwin K. Jackson, PhD Professor of Pharmacology and Chemical Biology School of Medicine University of Pittsburgh, Pennsylvania

Allen P. Kaplan, MD Clinical Professor of Medicine Medical University of South Carolina, Charleston

Robert S. Kass, PhD Professor and Chair of Pharmacology Vice Dean for Research College of Physicians and Surgeons Columbia University, New York

Kenneth Kaushansky, MD Dean, School of Medicine and Senior Vice President of Health Sciences SUNY Stony Brook, New York

Thomas J. Kipps, MD, PhD Professor of Medicine, Moores Cancer Center University of California, San Diego

Ronald J. Koenig, MD, PhD Professor of Metabolism, Endocrinology and Diabetes Department of Internal Medicine University of Michigan Health System, Ann Arbor

Alan M. Krensky, MD

Senior Investigator, National Cancer Institute, Bethesda, Maryland

Nora Laiken, PhD Lecturer in Pharmacology and Medicine University of California, San Diego

Andrew A. Lane, MD, PhD Fellow, Dana-Farber Cancer Institute Massachusetts General Hospital Cancer Center, Boston

Richard J. Lee, MD, PhD Professor of Medicine, Harvard Medical School Physician, Massachusetts General Hospital Boston, Massachusetts

Ellis R. Levin, MD Professor of Medicine; Chief of Endocrinology Diabetes and Metabolism University of California, Irvine, and Long Beach VA Medical Center, Long Beach

Dan L. Longo, MD Scientific Director, National Institute on Aging National Institutes of Health, Bethesda, Maryland

Alex Loukas, PhD Professor of Public Health, Tropical Medicine and Rehabilitation Sciences James Cook University, Cairns, Australia

Conan MacDougall, PharmD, MAS Associate Professor of Clinical Pharmacy School of Pharmacy University of California, San Francisco

Kenneth P. Mackie, MD Professor of Neuroscience Indiana University, Bloomington

Bradley A. Maron, MD Fellow in Cardiovascular Medicine Harvard Medical School and Brigham and Women's Hospital Boston, Massachusetts

James McCarthy, MD Associate Professor of Clinical Tropical Medicine University of Queensland Brisbane, Australia

James O. McNamara, MD Professor and Chair of Neurobiology Director of Center for Translational Neuroscience Duke University Medical Center Durham, North Carolina

CONTRIBUTORS

xiii

Boehringer Ex. 2018 Mylan v. Boehringer Ingelheim IPR2016-01564 Page 5

# xiv Jonathan M. Meyer, MD

Assistant Adjunct Professor of Psychiatry University of California, San Diego

### Thomas Michel, MD, PhD

Professor of Medicine and Biochemistry
Harvard Medical School
Senior Physician in Cardiovascular Medicine
Brigham and Women's Hospital
Boston, Massachusetts

### S. John Mihic, PhD

Professor of Neurobiology Waggoner Center for Alcohol & Addiction Research Institute for Neuroscience and Cell & Molecular Biology University of Texas, Austin

# Constantine S. Mitsiades, MD, PhD

Professor of Medical Oncology Dana-Farber Cancer Institute, Harvard Medical School Boston, Massachusetts

### Perry Molinoff, MD

Professor of Pharmacology, School of Medicine University of Pennsylvania, Philadelphia

### Dean S. Morrell, MD

Associate Professor of Dermatology University of North Carolina, Chapel Hill

### Beverly Moy, MD, MPH

Assistant Professor of Medicine Harvard Medical School Massachusetts General Hospital, Needham

### Hamza Mujagic, MD, MR. SCI, DR. SCI

Visiting Professor of Hematology and Oncology Harvard Medical School Massachusetts General Hospital, Needham

### Joel W. Neal, MD, PhD

Assistant Professor of Medicine-Oncology, Stanford University School of Medicine, Palo Alto, California

### Charles P. O'Brien, MD, PhD

Professor of Psychiatry, School of Medicine University of Pennsylvania, Philadelphia

### James O'Donnell, PhD

Professor of Behavioral Medicine and Psychiatry School of Medicine West Virginia University, Morgantown

### Erin M. Olson, MD

Fellow in Medical Oncology Dana-Farber Cancer Institute Boston, Massachusetts

### Taylor M. Ortiz, MD

Clinical Fellow in Medical Oncology Dana-Farber Cancer Institute General Hospital Cancer Center Boston, Massachusetts

# Kevin Osterhoudt, MD, MSCE, FAAP, FACMT

Associate Professor of Pediatrics
School of Medicine, University of Pennsylvania;
Medical Director, Poison Control Center, Children's Hospital
of Philadelphia, Pennsylvania

### Keith L. Parker, MD, PhD (deceased)

Professor of Internal Medicine and Pharmacology Chief of Endocrinology and Metabolism University of Texas Southwestern Medical School, Dallas

### Hemal H. Patel, PhD

Associate Professor of Anesthesiology University of California, San Diego Dean, School of Medicine and Senior Vice President of Health Sciences SUNY Stony Brook, New York

### Pivush M. Patel, MD, FRCPC

Professor of Anesthesiology University of California, San Diego

# Trevor M. Penning, PhD

Professor of Pharmacology
Director, Center of Excellence in Environmental Toxicology
School of Medicine
University of Pennsylvania, Philadelphia

### William A. Petri, Jr, MD, PhD

Professor of Medicine; Chief, Division of Infectious Diseases University of Virginia, Charlottesville

### Margaret A. Phillips, PhD

Professor of Pharmacology University of Texas Southwestern Medical School, Dallas

### Alvin C. Powers, MD

Professor of Medicine, Molecular Physiology and Biophysics Vanderbilt University Medical Center Nashville, Tennessee

### Christopher Rapuano, MD

Director, Cornea Service and Refractive Surgery Department, Wills Eye Institute Philadelphia, Pennsylvania

### Robert F. Reilly, Jr, MD

Professor of Internal Medicine University of Texas Southwestern Medical School, Dallas Chief of Nephrology VA-North Texas Health Care System, Dallas

### Mary V. Relling, PharmD

Chair of Pharmaceutical Sciences St. Jude Childrens' Research Hospital Memphis, Tennessee

### Paul G. Richardson, MD

Associate Professor of Medicine, Harvard Medical School Clinical Director, Lipper Center for Multiple Myeloma Dana-Farber Cancer Institute Boston, Massachusetts

### Suzanne M. Rivera, PhD, MSW

Assistant Professor of Clinical Sciences University of Texas Southwestern Medical Center, Dallas

### Erik Roberson, MD, PhD

Assistant Professor of Neurology and Neurobiology University of Alabama, Birmingham

### Thomas P. Rocco, MD

Associate Professor of Medicine Harvard Medical School VA-Boston Healthcare System Boston, Massachusetts

### David M. Roth, MD, PhD

Professor of Anesthesiology University of California, San Diego VA-San Diego Healthcare System

### David P. Ryan, MD

Associate Professor of Medicine Harvard Medical School Massachusetts General Hospital Cancer Center, Boston

### Kevin J. Sampson, PhD

Postdoctoral Research Scientist in Pharmacology Columbia University, New York

### Elaine Sanders-Bush, PhD

Professor (Emerita) of Pharmacology School of Medicine, Vanderbilt University Nashville, Tennessee

### Bernard P. Schimmer, PhD

Professor (Emeritus) of Medical Research and Pharmacology University of Toronto, Ontario

### Marc A. Schuckit, MD

Distinguished Professor of Psychiatry University of California, San Diego Director, Alcohol Research Center VA-San Diego Healthcare System

### Lecia Sequist, MD, MPH

Assistant Professor of Medicine Harvard Medical School, Massachusetts General Hospital Cancer Center, Boston

### Keith A. Sharkey, PhD

Professor of Physiology & Pharmacology and Medicine University of Calgary, Alberta

### Richard C. Shelton, MD

Professor of Psychiatry and Pharmacology School of Medicine, Vanderbilt University Nashville, Tennessee

### Danny Shen, PhD

Professor and Chair of Pharmacy Professor of Pharmaceutics, School of Pharmacy University of Washington, Seattle

### Randal A. Skidgel, PhD

Professor of Pharmacology and Anesthesiology College of Medicine, University of Illinois-Chicago

### Matthew R. Smith, MD, PhD

Associate Professor of Medicine, Harvard Medical School Physician, Massachusetts General Hospital, Boston

### Emer M. Smyth, PhD

Research Assistant, Professor of Pharmacology University of Pennsylvania, Philadelphia

### Peter J. Snyder, MD

Professor of Medicine University of Pennsylvania, Philadelphia

### David Standaert, MD, PhD

Professor of Neurology
Director, Center for Neurodegeneration and Experimental
Therapeutics
University of Alabama, Birmingham

### Samuel L. Stanley, Jr, MD

Professor of Medicine and President SUNY Stony Brook, New York

### Yuichi Sugiyama, PhD

Professor and Chair of Molecular Pharmacokinetics University of Tokyo, Japan

### Jeffrey G. Supko, PhD

Associate Professor of Medicine, Harvard Medical School Massachusetts General Hospital, Boston

### Palmer W. Taylor, PhD

Professor of Pharmacology, School of Medicine Dean, Skaggs School of Pharmacy and Pharmaceutical Sciences University of California, San Diego

### Kenneth E. Thummel, PhD

Professor and Chair, Department of Pharmaceutics University of Washington, Seattle

# Flavio Vincenti, MD

Professor of Clinical Medicine Medical Director, Pancreas Transplant Program University of California, San Francisco

# Joseph M. Vinetz, MD

Professor of Medicine, Division of Infectious Diseases University of California, San Diego

### Mark S. Wallace, MD

Professor of Clinical Anesthesiology University of California, San Diego

# John L. Wallace, PhD, MBA, FRSC

Professor and Director, Farncombe Family Digestive Health Research Institute

McMaster University, Hamilton, Ontario

# Jeffrey I. Weitz, MD, FRCP(C), FACP

Professor of Medicine, Biochemistry and Biomedical Sciences McMaster University

Executive Director, Thrombosis & Atherosclerosis Research Institute, Hamilton, Ontario

# David P. Westfall, PhD

Professor (Emeritus) of Pharmacology University of Nevada School of Medicine, Reno

### Thomas C. Westfall, PhD

Professor and Chair of Pharmacological and Physiological Science

St. Louis University School of Medicine, Missouri

### Wyndham Wilson, MD, PhD

Senior Investigator and Chief of Lymphoid Therapeutics Section,

Center for Cancer Research, National Cancer Institute Bethesda Maryland

# Tony L. Yaksh, PhD

Professor of Anesthesiology and Pharmacology University of California, San Diego

# Alexander C. Zambon, PhD

Assistant Professor of Pharmacology University of California, San Diego

# Preface

The publication of the twelfth edition of this book is a testament to the vision and ideals of the original authors, Alfred Gilman and Louis Goodman, who, in 1941 set forth the principles that have guided the book through eleven editions: to correlate pharmacology with related medical sciences, to reinterpret the actions and uses of drugs in light of advances in medicine and the basic biomedical sciences, to emphasize the applications of pharmacodynamics to therapeutics, and to create a book that will be useful to students of pharmacology and to physicians. These precepts continue to guide the current edition.

As with editions since the second, expert scholars have contributed individual chapters. A multiauthored book of this sort grows by accretion, posing challenges to editors but also offering memorable pearls to the reader. Thus, portions of prior editions persist in the current edition, and I hasten to acknowledge the contributions of previous editors and authors, many of whom will see text that looks familiar. However, this edition differs noticeably from its immediate predecessors. Fifty new scientists, including a number from outside the U.S., have joined as contributors, and all chapters have been extensively updated. The focus on basic principles continues, with new chapters on drug invention, molecular mechanisms of drug action, drug toxicity and poisoning, principles of antimicrobial therapy, and pharmacotherapy of obstetrical and gynecological disorders. Figures are in full color. The editors have continued to standardize the organization of chapters; thus, students should easily find the basic physiology, biochemistry, and pharmacology set forth in regular type; bullet points highlight important lists within the text; the clinician and expert will find details in extract type under clear headings.

Online features now supplement the printed edition. The entire text, updates, reviews of newly approved drugs, animations of drug action, and hyperlinks to relevant text in the prior edition are available on the Goodman & Gilman section of McGraw-Hill's websites, AccessMedicine.com and AccessPharmacy.com. An Image Bank CD accompanies the book and makes all tables and figures available for use in presentations.

The process of editing brings into view many remarkable facts, theories, and realizations. Three stand out: the invention of new classes of drugs has slowed to a trickle; therapeutics has barely begun to capitalize on the information from the human genome project; and, the development of resistance to antimicrobial agents, mainly through their overuse in medicine and agriculture, threatens to return us to the pre-antibiotic era. We have the capacity and ingenuity to correct these shortcomings.

Many, in addition to the contributors, deserve thanks for their work on this edition; they are acknowledged on an accompanying page. In addition, I am grateful to Professors Bruce Chabner (Harvard Medical School/Massachusetts General Hospital) and Björn Knollmann (Vanderbilt University Medical School) for agreeing to be associate editors of this edition at a late date, necessitated by the death of my colleague and friend Keith Parker in late 2008. Keith and I worked together on the eleventh edition and on planning this edition. In anticipation of the editorial work ahead, Keith submitted his chapters before anyone else and just a few weeks before his death; thus, he is well represented in this volume, which we dedicate to his memory.

Laurence L. Brunton San Diego, California December 1, 2010 Wahl-Schott C, Biel M. HCN channels: Structure, cellular regulation and physiological function. *Cell Mol Life Sci*, **2009**, 66:470–494.

Wang X, Lupardus P, Laporte SL, Garcia KC. Structural biology of shared cytokine receptors. *Annu Rev Immunol*, **2009**, 27:79–60

Wilson NS, Dixit V, Ashkenazi A. Death receptor signal transducers: Nodes of coordination in immune signaling networks. *Nat Immunol*, **2009**, *10*:348–355.

Wong W, Scott JD. AKAP signalling complexes: Focal points in space and time. *Nat Rev Mol Cell Biol*, **2004**, 5:959–970.



# Drug Toxicity and Poisoning

Kevin C. Osterhoudt and Trevor M. Penning

Pharmacology deals with drugs and their chemical properties or characteristics, their mode of action, the physiological response to drugs, and the clinical uses of drugs. Pharmacology intersects with toxicology when the physiological response to a drug is an adverse effect. Toxicology is often regarded as the science of poisons or poisoning, but developing a strict definition for poison is problematic. A poison is any substance, including any drug, that has the capacity to harm a living organism. The Renaissance physician Paracelsus (1493-1541) is famously credited with offering the philosophical definition of poisons: "What is there that is not poison? All things are poison and nothing is without poison. Solely the dose determines that a thing is not a poison." However, poisoning inherently implies that damaging physiological effects result from exposure to pharmaceuticals, illicit drugs, or chemicals. So each drug in the pharmacopeia is a potential poison, and individual dose-, situation-, environment-, and generelated factors contribute to a drug's ability to achieve its adverse potential.

Some chemicals may inherently be poisons, such as lead, which has no known necessary physiological role in the human body, and which is known to cause neuronal injury even at very low exposure levels. Most pharmaceuticals are threshold poisons; at therapeutic dosing the drug is used to confer a health advantage, but at higher doses the drug may produce a toxic effect. For instance, iron is a nutrient essential for heme synthesis and numerous physiological enzyme functions, but overdose of ferrous sulfate can lead to life-threatening multi-organ dysfunction.

### **DOSE-RESPONSE**

Evaluation of the dose-response or the dose-effect relationship is crucially important to toxicologists. There is a graded dose-response relationship in an *individual* and a quantal dose-response relationship in the *population* (see Chapters 2 and 3). Graded doses of a drug given to

an individual usually result in a greater magnitude of response as the dose is increased. In a quantal dose-response relationship, the percentage of the population affected increases as the dose is raised; the relationship is quantal in that the effect is specified to be either present or absent in a given individual (Figure 4–1). This quantal dose-response phenomenon is extremely important in toxicology and is used to determine the *median lethal dose* (LD $_{50}$ ) of drugs and other chemicals.

The  $LD_{50}$  of a compound is determined experimentally, usually by administration of the chemical to mice or rats (orally or intraperitoneally) at several doses in the lethal range (Figure 4–1A).

To linearize such data, the response (death) can be converted to units of deviation from the mean, or probits (probability units). The probit designates the deviation from the median; a probit of 5 corresponds to a 50% response, and because each probit equals one standard deviation, a probit of 4 equals 16% and a probit of 6 equals 84%. A plot of the percentage of the population responding, in probit units, against log dose yields a straight line (Figure 4–1B). The LD $_{50}$  is determined by drawing a vertical line from the point on the line where the probit unit equals 5 (50% mortality). The slope of the dose-effect curve also is important. The LD $_{50}$  for both compounds depicted in Figure 4–1 is the same (~10 mg/kg); however, the slopes of the dose-response curves are quite different. At a dose equal to one-half the LD $_{50}$  (5 mg/kg), less than 5% of the animals exposed to compound B would die, but 30% of the animals given compound A would die.

Figure 4–2 illustrates the relationship between a quantal dose-response curve for the therapeutic effect of a drug to generate a median effective dose (ED $_{50}$ ), the concentration of drug at which 50% of the population will have the desired response, and a quantal dose-response curve for lethality by the same agent. These two curves can be used to generate a therapeutic index (TI), which quantifies the relative safety of a drug. Clearly, the higher the ratio, the safer the drug.

 $\mathrm{TI} = \mathrm{LD}_{50}/\mathrm{ED}_{50}$ 

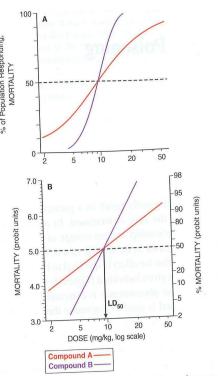


Figure 4-1. Dose-response relationships. A. The toxic response to a chemical is evaluated at several doses in the toxic or lethal range. The midpoint of the curve representing percent of population responding (response here is death) versus dose (log scale) represents the  $\mathrm{LD}_{50}$ , or the concentration of drug that is lethal in 50% of the population. **B.** A linear transformation of the data in panel A, obtained by plotting the log of the dose administered versus the percent of the population killed, in

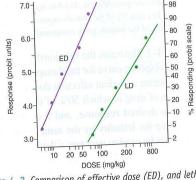


Figure 4-2. Comparison of effective dose (ED), and lethal dose (LD). See text for explanation of probit units. Note that abscissa is a logarithmic scale.

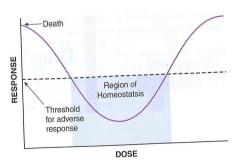


Figure 4-3. U-Shaped dose-response curve for essential metals and vitamins. Vitamins and essential metals are essential for life and their lack can cause adverse responses (plotted on the vertical axis), as can their excess, giving rise to a U-shaped concentration-dependence curve.

Drugs show a wide range of TI, from 1-2 to >100. Drugs with a low TI must be administered with caution. Agents that fall into this category include the cardiac glycoside digitalis and cancer chemotherapeutic agents. Agents with very high TI are extremely safe and include some antibiotics (e.g., penicillin), unless there is a known allergic response.

The use of median effective and median lethal doses is not without disadvantages, because median doses do not consider that the slopes of the doseresponse curves for therapeutic and lethal (toxic) effects may differ. As an alternative the ED<sub>99</sub> for the therapeutic effect can be compared to the LD1 for lethality (toxic effect), to yield a margin of safety.

# Margin of safety = $LD_1/ED_{99}$

The quantal dose-response curves described thus far represent linearization of sigmoidal dose-response curves (Figure 4-1). However, not all dose-response curves follow this shape. "U"-shaped dose-response curves can be observed for essential metals and vitamins (Figure 4-3). At low dose, adverse effects are observed since there is a deficiency of these nutrients to maintain homeostasis. As dose increases, homeostasis is achieved, and the bottom of the "U"-shaped dose-response curve is reached. As dose increases to surpass the amount required to maintain homeostasis, overdose toxicity can ensue. Thus, adverse effects are seen at both low and high dose.

# PHARMACOKINETICS VERSUS TOXICOKINETICS

The principles of pharmacokinetics (absorption, distribution, metabolism, and elimination) are described in Chapters 2, 5, and 6, and specific details are provided throughout this text. Toxicokinetics (the pharmacokinetics of a drug under circumstances that produce toxicity or excessive exposure) may differ significantly after poisoning, and these differences may profoundly alter treatment decisions and prognosis. Ingesting larger than therapeutic doses of a pharmaceutical may prolong its absorption, alter its protein binding and apparent volume of distribution, and change its metabolic fate. When confronted with a potential poisoning, two questions are foremost in the clinician's mind:

- 1. How long does an asymptomatic patient need to be monitored (drug absorption and dynamics)?
- 2. How long will it take an intoxicated patient to get better (drug elimination and dynamics)?

Drug Absorption. Aspirin poisoning is a leading cause of overdose morbidity and mortality as reported to U.S. poison control centers (Bronstein et al., 2008). In therapeutic dosing, aspirin reaches peak plasma concentrations in ~1 hour (Chapter 34). However, after overdose, several drug-related and physiology-related factors change. Aspirin overdose may cause spasm of the pyloric valve, delaying entry of the drug into the small intestine. Aspirin, especially enteric coated forms, may coalesce into bezoars, reducing the effective surface area for absorption. Peak plasma salicylate concentrations from aspirin overdose may not be reached for 4-35 hours after ingestion (Rivera et al., 2004).

Drug Elimination. After therapeutic dosing, valproic acid has an elimination half-life (tys) of ~14 hours (see Appendix II). Valproic acid poisoning may lead to coma. In predicting the duration of coma, it is important to consider that, after overdose, first-order metabolic processes appear to become saturated and the apparent elimination  $t_{1/3}$  may exceed 30-45 hours (Sztajnkrycer, 2002). Such consideration has important clinical ramifications pertaining to prognosis, resource utilization, and therapy.

Table 4-1 lists some pharmaceuticals notorious for their predilection to have initial symptoms develop after a typical 4-6 hour

### Table 4-1

### **Drugs That Commonly Manifest Initial Symptoms** More Than 4-6 Hours after Oral Overdose<sup>a</sup>

Acetaminophen Aspirin

Illicit drugs in rubber or plastic packages Monoamine oxidase inhibitors

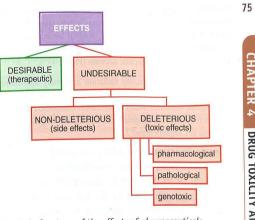
Sulfonylureas

Sustained-release formulation drugs Thyroid hormones

Valproic acid

Warfarin-like anticoagulants

<sup>a</sup>Drugs co-ingested with agents having anticholinergic activity, manifest by diminished GI motility, may also exhibit delayed onset



DRUG TOXICITY

AND

POISONING

Figure 4-4. Spectrum of the effects of pharmaceuticals.

emergency medical observation period for a suspected drug

# Types of Therapeutic Drug Toxicity

In therapeutics, a drug typically produces numerous effects, but usually only one is sought as the primary goal of treatment; most of the other effects are undesirable effects of that drug for that therapeutic indication (Figure 4-4). Side effects of drugs usually are bothersome but not deleterious; they include effects such as dry mouth occurring with tricyclic antidepressant therapy. Other undesirable effects may be characterized as toxic effects.

Dose-Dependent Reactions. Toxic effects of drugs may be classified as pharmacological, pathological, or genotoxic. Typically, the incidence and seriousness of the toxicity is proportionately related to the concentration of the drug in the body and to the duration of the exposure. Drug overdose provides a dramatic example of dose-dependent toxicities.

Pharmacological Toxicity. The CNS depression produced by barbiturates is largely predictable in a dose-dependent fashion. The progression of clinical effects goes from anxiolysis to sedation to somnolence to coma. Similarly, the degree of hypotension produced by nifedipine is related to the dose of the drug administered. Tardive dyskinesia (Chapter 16), an extrapyramidal motor disorder associated with use of antipsychotic medications, seems to be dependent upon duration of exposure. Pharmacological toxicity can also occur when the correct dose is given: there is phototoxicity associated with exposure to sunlight in patients treated with tetracyclines, sulfonamides, chlorpromazine, and nalidixic acid.

Pathological Toxicity. Acetaminophen is metabolized to nontoxic glucuronide and sulfate conjugates, and to a highly reactive metabolite N-acetyl-p-benzoquinoneimine (NAPQI) via CYP

Genotoxic Effects. Ionizing radiation and many environmental chemicals are known to injure DNA, and may lead to mutagenic or carcinogenic toxicities. Many of the cancer chemotherapeutic agents (Chapters 60-63) may be genotoxic. Discussions of genotoxicity can be found in Chapters 6 and 7.

Allergic Reactions. An allergy is an adverse reaction that results from previous sensitization to a particular chemical or to one that is structurally similar. Such reactions are mediated by the immune system. For a low-molecular-weight chemical to cause an allergic reaction, it or its metabolic product usually acts as a hapten, combining with an endogenous protein to form an antigenic complex. Such antigens induce the synthesis of antibodies, usually after a latent period of at least 1-2 weeks. Subsequent exposure to the chemical results in an antigen-antibody interaction that provokes the typical manifestations of allergy. Dose-response relationships usually are not apparent for the provocation of

> HNCOCH. HNCOCH. HNCOCH<sub>2</sub> HNCOCH

Figure 4-5. Pathways of acetaminophen metabolism and toxicity. The toxic intermediate NAPQI is *N*-acetyl-*p*-benzoquinoneimine.

allergic reactions. Allergic responses have been divided into four general categories based on the mechanism of immunological involvement.

Type I: Anaphylactic Reactions. Anaphylaxis is mediated by IgE antibodies. The Fc portion of IgE can bind to receptors on mast cells and basophils. If the Fab portion of the antibody molecule then binds antigen, various mediators (e.g., histamine, leukotrienes, and prostaglandins) are released and cause vasodilation, edema, and an inflammatory response. The main targets of this type of reaction are the gastrointestinal (GI) tract (food allergies), the skin (urticaria and atopic dermatitis), the respiratory system (rhinitis and asthma), and the vasculature (anaphylactic shock). These responses tend to occur quickly after challenge with an antigen to which the individual has been sensitized and are termed immediate hypersensitivity reactions.

Type II: Cytolytic Reactions. Type II allergies are mediated by both IgG and IgM antibodies and usually are attributed to their capacity to activate the complement system. The major target tissues for cytolytic reactions are the cells in the circulatory system. Examples of type II allergic responses include penicillin-induced hemolytic anemia, quinidine-induced thrombocytopenic purpura, and sulfonamide-induced granulocytopenia. Fortunately, these autoimmune reactions to drugs usually subside within several months after removal of the offending agent.

Type III: Arthrus Reactions. Type III allergic reactions are mediated predominantly by IgG; the mechanism involves the generation of antigen-antibody complexes that subsequently fix complement. The complexes are deposited in the vascular endothelium, where a destructive inflammatory response called serum sickness occurs. This phenomenon contrasts with the type II reaction, in which the inflammatory response is induced by antibodies directed against tissue antigens. The clinical symptoms of serum sickness include urticarial skin eruptions, arthralgia or arthritis, lymphadenopathy, and fever. Several drugs, including commonly used antibiotics, can induce serum sickness-like reactions. These reactions usually last 6-12 days and then subside after the offending agent is eliminated.

Type IV: Delayed Hypersensitivity Reactions. These reactions are

mediated by sensitized T-lymphocytes and macrophages. When sensitized cells come in contact with antigen, an inflammatory reaction is generated by the production of lymphokines and the subsequent influx of neutrophils and macrophages. An example of type IV or delayed hypersensitivity is the contact dermatitis caused by poison ivy. Idiosyncratic Reactions. Idiosyncrasy is an abnormal reactivity to a chemical that is peculiar to a given individual. The idiosyncratic response may take the form of extreme sensitivity to low doses or extreme insensitivity to high doses of drugs. Idiosyncratic reactions can result from genetic polymorphisms that cause individual differences in drug pharmacokinetics, from pharmacodynamic factors such as drug-receptor interactions (Evans and Rolling, 1999), or from variability in expression of enzyme activity. The use of genetic information to explain interindividual differences in drug responses or to individualize dosages of drugs for patients with known genetic polymorphisms is referred to as pharmacogenetics (Chapter 7).

An increased incidence of peripheral neuropathy is seen in patients with inherited deficiencies in acetylation when isoniazid is used to treat tuberculosis, e.g., slow and fast acetylators exist due to polymorphisms in N-acetyl transferase. Many black males (~10%) develop a serious hemolytic anemia when they receive primaquine as an antimalarial therapy. Such individuals have a deficiency of erythrocyte glucose-6-phosphate dehydrogenase. Genetically determined resistance to the anticoagulant action of warfarin is due to an alteration in the vitamin K epoxide reductase.

Drug-Drug Interactions. Patients are commonly treated with more than one drug, have individual dietary choices. and may also be using over-the-counter (OTC) medications, vitamins, and other "natural" supplements. This polypharmaceutical nature of healthcare requires consideration of potential drug interactions (Figure 4–6). Similar to changes in pharmacokinetics and pharmacodynamics seen after drug overdose, drug interactions may lead to altered rates of absorption, altered protein binding, or different rates of biotransformation or excretion of one or both interacting compounds. The pharmacodynamics of a drug can be altered by competition at receptors, and nonreceptor pharmacodynamic interactions can occur when two drugs have

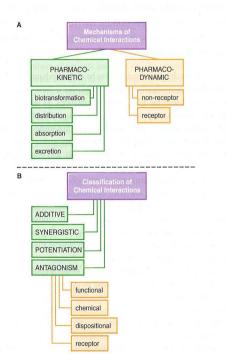


Figure 4-6. Mechanisms and classification of drug interactions.

similar actions through different cellular mechanisms. 77 Induction or inhibition of drug metabolism by CYPs (Chapter 6) are among the most clinically challenging of drug interactions.

Interaction of Absorption. A drug may cause either an increase or a decrease in the absorption of another drug from the intestinal lumen. Ranitidine, an antagonist of histamine H2 receptors, raises gastrointestinal pH and may increase the absorption of basic drugs such as triazolam (O'Connor-Semmes et al., 2001). Conversely, the bile-acid sequestrant cholestyramine leads to significantly reduced serum concentrations of propranolol, and may diminish its effect for a given dosage (Hibbard et al., 1984).

Interaction of Protein Binding. Many drugs, such as aspirin, barbiturates, phenytoin, sulfonamides, valproic acid, and warfarin, are highly protein-bound in the plasma, and it is the free (unbound) drug that produces the clinical effects. These drugs may have enhanced toxicity in overdose if protein binding sites become saturated, in physiological states that lead to hypoalbuminemia, or when displaced from plasma proteins by other drugs. The anticoagulant effects of warfarin may be enhanced by displacement from plasma proteins by simultaneous valproic acid therapy (Guthrie et al., 1995).

Interaction of Metabolism. A drug can frequently influence the metabolism of one or several other drugs (Chapter 6), and this is especially notable within hepatic CYPs. Acetaminophen is partially transformed by CYP2E1 to the toxic metabolite NAPQI (Figure 4-5). Intake of ethanol, a potent inducer of the 2E1 isoenzyme, may lead to increased susceptibility to acetaminophen poisoning after overdose (Dart et al., 2006). Similarly, a number of second-generation piperidine antihistamines (terfenadine, astemizole) were removed from the market when they were noted to lead to QT interval prolongation and tachydysrhythmias when co-administered with macrolide antibiotics.

Interaction of Receptor Binding. Buprenorphine is an opioid with partial agonist and antagonist receptor activities. It can be used as an analgesic but is more commonly used to treat opioid addiction. The drug binds opioid receptors with high affinity, and can prevent euphoria from concomitant use of narcotic drugs of abuse.

Interaction of Therapeutic Action. Aspirin is an inhibitor of platelet aggregation and heparin is an anticoagulant; given together they may increase risk for bleeding. Sulfonylureas cause hypoglycemia by stimulating pancreatic insulin release, whereas biguanide drugs (metformin) lead to decreased hepatic glucose production, and these drugs can be used together to control diabetic hyperglycemia.

A drug interaction is said to be *additive* when the combined effect of two drugs equals the sum of the effect of each agent given alone. A synergistic effect is one in which the combined effect exceeds the sum of the effects of each drug given alone. Potentiation describes the creation of a toxic effect from one drug due to the presence of another drug. Antagonism is the interference of one drug with the action of another. Drug antagonism may confer a therapeutic advantage when one drug is to be used as an antidote for the toxicity of another drug. Functional or physiological

78 antagonism occurs when two chemicals produce opposite effects on the same physiological function; this is the basis for most supportive care provided to patients treated for drug overdose poisoning. Chemical antagonism, or inactivation, is a reaction between two chemicals to neutralize their effects, such as is seen with chelation therapy. Dispositional antagonism is the alteration of the disposition of a substance (its absorption, biotransformation, distribution, or excretion) so that less of the agent reaches the target organ or its persistence in the target organ is reduced. Receptor antagonism entails the blockade of the effect of a drug with another drug that competes at the receptor site.

# DESCRIPTIVE TOXICITY TESTING IN ANIMALS

Two main principles underlie all descriptive toxicity tests performed in animals. First, those effects of chemicals produced in laboratory animals, when properly qualified, apply to human toxicity. When calculated on the basis of dose per unit of body surface, toxic effects in human beings usually are encountered in the same range of concentrations as those in experimental animals. On the basis of body weight, human beings generally are more vulnerable than experimental animals. Such information is used to select dosages for clinical trials of candidate therapeutic agents and to attempt to set limits on permissible exposure to environmental toxicants.

Second, exposure of experimental animals to toxic agents in high doses is a necessary and valid method to discover possible hazards to human beings who are exposed to much lower doses. This principle is based on the quantal dose-response concept. As a matter of practicality, the number of animals used in experiments on toxic materials usually will be small compared with the size of human populations potentially at risk. For example, 0.01% incidence of a serious toxic effect (such as cancer) represents 25,000 people in a population of 250 million. Such an incidence is unacceptably high. Yet, detecting an incidence of 0.01% experimentally probably would require a minimum of 30,000 animals. To estimate risk at low dosage, large doses must be given to relatively small groups. The validity of the necessary extrapolation is clearly a crucial question.

Chemicals are first tested for toxicity by estimation of the LD<sub>50</sub> in two animal species by two routes of administration; one of these is the expected route of exposure of human beings to the chemical being tested. The number of animals that die in a 14-day period after a single dose is recorded. The animals also are examined

for signs of intoxication, lethargy, behavioral modification, and morbidity. The chemical is next tested for toxicity by repeat exposure, usually for 90 days. This study is performed most often in two species by the route of intended use or exposure with at least three doses. A number of parameters are monitored during this period, and at the end of the study, organs and tissues are examined by a pathologist.

Long-term or chronic studies are carried out in animals at the same time that clinical trials are undertaken. For drugs, the length of exposure depends somewhat on the intended clinical use. If the drug normally would be used for short periods under medical supervision, as would an antimicrobial agent, a chronic exposure of animals for 6 months might suffice. If the drug would be used in human beings for longer periods, a study of chronic use for 2 years may be required.

Studies of chronic exposure often are used to determine the carcinogenic potential of chemicals. These studies usually are performed in rats and mice for the average lifetime of the species. Other tests are designed to evaluate teratogenicity (congenital malformations), perinatal and postnatal toxicity, and effects on fertility. Teratogenicity studies usually are performed by administering drugs to pregnant rats and rabbits during the period of organogenesis. As noted in Chapter 6, in silico computational methods of chemical biology systems may soon contribute to such studies.

# TOXICOLOGY, SAFETY TESTING, AND CLINICAL TRIALS

Fewer than one-third of the drugs tested in clinical trials reach the marketplace. Federal law in the U.S. and ethical considerations require that the study of new drugs in humans be conducted in accordance with stringent guidelines. Such studies apply the principles of toxicology mentioned earlier.

Once a drug is judged ready to be studied in humans, a Notice of Claimed Investigational Exemption for a New Drug (IND) must be filed with the FDA. The IND includes: 1) information on the composition and source of the drug; 2) chemical and manufacturing information; 3) all data from animal studies; 4) proposed clinical plans and protocols; 5) the names and credentials of physicians who will conduct the clinical trials; and 6) a compilation of the key data relevant to study the drug in man made available to investigators and their institutional review boards (IRBs).

It often requires 4-6 years of clinical testing to accumulate and analyze all required data. Testing in humans is begun after sufficient acute and subacute animal toxicity studies have been completed. Chronic safety testing in animals, including carcinogenicity studies, is usually done concurrently with clinical trials. In each of the three formal phases of clinical trials, volunteers or patients must be informed of the investigational status of the drug as well as the possible risks and must be allowed to decline or to consent to participate and receive the drug. These regulations are based on the ethical principles set forth in the Declaration of Helsinki. In addition to the approval of the sponsoring organization and the FDA, an interdisciplinary IRB at the facility where the clinical drug trial will be conducted must review and approve the scientific and ethical plans for testing in humans.

In phase 1, the effects of the drug as a function of dosage are established in a small number (20-100) of healthy volunteers. Although a goal is to find the maximum tolerated dose, the study is designed to prevent severe toxicity. If the drug is expected to have significant toxicity, as may be the case in cancer and AIDS therapy, volunteer patients with the disease rather than disease-free volunteers are used in phase 1. Phase 1 trials are designed to determine the probable limits of the safe clinical dosage range. These trials may be non-blind or "open"; i.e., both the investigators and the subjects know what is being given. Alternatively, they may be "blinded" and/or

placebo-controlled. The choice of design depends on the drug, disease, goals of investigators, and ethical considerations. Many predictable toxicities are detected in this phase. Pharmacokinetic measurements of absorption,  $t_{\mbox{\tiny LS}},$  and metabolism are often conducted. Phase 1 studies are usually performed in research centers by clinical pharmacologists.

In phase 2, the drug is studied in patients with the target disease to determine its efficacy ("proof of concept"), and the doses to be used in any follow-on trials. A modest number of patients (100-200) are studied in detail. A single-blind design may be used, with an inert placebo medication and an established active drug (positive control) in addition to the investigational agent. Phase 2 trials are usually done in special clinical centers (e.g., university hospitals). A broader range of toxicities may be detected in this phase. Phase 2 trials have the highest rate of drug failures, and only ~25% of drug candidates move on to phase 3.

In phase 3, the drug is evaluated in much larger numbers of patients with the target disease—usually thousands—to further establish and confirm safety and efficacy. Using information gathered in phases 1 and 2, phase 3 trials are designed to minimize errors caused by placebo effects, variable course of the disease, etc. Therefore, double-blind and crossover techniques are frequently used. Phase 3 trials are usually performed in settings similar to those anticipated for the ultimate use of the drug. Phase 3 studies can be difficult to design and execute and are usually expensive because of the large numbers of patients involved and the amount of data that must be collected and analyzed. The drug is formulated as intended for the market. The investigators are usually specialists in the disease being treated. Certain toxic effects, especially those caused by immunologic processes, may first become apparent in phase 3 trials.

If phase 3 results meet expectations, application is made to the FDA for permission to market the new agent. Marketing approval requires submission of a New Drug Application (NDA) (or for biologicals, a Biological License Application [BLA]) to the FDA. Note that even at this point in the process, experience with the new drug is limited to information gathered from a few thousand patients. Thus, post-marketing surveillance is crucial in determining the actual toxicity of the drug as it is administered to a much larger population in which low-frequency adverse reactions will be noticeable.

# EPIDEMIOLOGY OF ADVERSE DRUG RESPONSES AND PHARMACEUTICAL POISONING

Poisoning can occur in many ways following both therapeutic and nontherapeutic drug or chemical exposures (Table 4-2). The incidence of serious and Table 4-2

# Potential Scenarios for the Occurrence of Poisoning

Therapeutic drug toxicity Exploratory exposure by young children Environmental exposure Occupational exposure Recreational abuse Medication error Prescribing error Dispensing error Administration error Purposeful administration for self-harm

Purposeful administration to harm another

fatal adverse drug reactions in U.S. hospitals is extremely high (Institute of Medicine, 1999; Lazarou et al., 1998). It is estimated that ~2 million hospitalized patients have serious adverse drug reactions each year, and ~100,000 have fatal adverse drug reactions. Use of good principles of prescribing, as described in Appendix I and Table 4-6, can aid in avoiding such adverse outcomes.

Some toxicities of pharmaceuticals can be predicted based upon their known pharmacological mechanism; however, it is often not until the post-marketing period that the therapeutic toxicity profile of a drug becomes fully appreciated. In the U.S., the approval system for new drugs typically uses only 500 to 3000 exposed subjects. Such a system is likely to identify toxicities occurring in 1% or more of patients receiving a drug (Strom, 2004). The Adverse Event Reporting System of the FDA relies upon two signals to detect rarer adverse drug events. First, the FDA requires (Code of Federal Regulations, Title 21, Volume 5, Section 314.80) drug manufacturers to perform post-marketing surveillance of prescription drugs, and similar regulations exist for nonprescription products. Second, the FDA operates a voluntary reporting system (MedWatch, online at http://www.fda.gov/Safety/MedWatch/ default.htm) available to both health professionals and consumers. Hospitals may also support adverse drug event committees to investigate potential adverse drug events, and these investigations may be made available to industry and the government. Unfortunately, any national dataset will significantly underestimate the morbidity and mortality attributable to adverse drug events because of under-reporting and because it is difficult to estimate the denominator of total patient exposures for each event reported once a drug is available on the open market.

As an example, post-marketing surveillance identified the toxicity associated with the serotonin receptor-modulating drug cisapride. This drug was known to enhance GI motility and was marketed in the U.S. as a treatment for gastroesophageal reflux. Postmarketing surveillance revealed that cisapride was associated with prolongation of the QT interval and predisposition to ventricular

DRUG

arrhythmia. It was withdrawn from the market, and subsequent casecontrol studies demonstrated increased risk of arrhythmia (Hennessy et al., 2008). Cisapride is now limited in its distribution through an investigational access program managed by the manufacturer. In another instance, selective serotonin reuptake inhibitors (SSRIs, Chapter 15) are believed to be safer than monoamine oxidase inhibitors and tricyclic antidepressants, and are popularly prescribed for treatment of depression; however, current post-marketing studies are looking to see if the use of SSRIs may predispose adolescent and young adult patients to suicidality (Barbui et al., 2009). Thus, the determination of drug toxicity carries beyond the stages of drug development.

Therapeutic drug toxicity is only a subset of poisoning, as noted in Table 4-2. Misuse and abuse of both prescription and illicit drugs is a major public health problem. The incidence of unintentional, noniatrogenic poisoning is bimodal, primarily affecting exploratory young children, ages 1-5 years, and the elderly. Intentional overdose with pharmaceuticals is most common in adolescence and through adulthood. Fifty-one percent of poison exposures reported to the American Association of Poison Control Centers (AAPCC) involve children ≤5 years, but this group accounts for <3% of reported fatalities, demonstrating that intentional poisonings are inherently more dangerous than exploratory or inadvertent exposures (Bronstein et al., 2008).

Exact quantification of nontherapeutic poisoning occurrences remains elusive. The National Institute of Drug Abuse (NIDA) commissioned the University of Michigan to perform the "Monitoring the Future" survey of U.S. high school children regarding drug use patterns (NIDA, 2009). In 2008, 47% of surveyed high school seniors reported that they had previously used an illicit drug. Among pharmaceutical abuse, 13% reported having abused a non-heroin narcotic, 10% an amphetamine drug, 8.9% a benzodiazepine, antidepressant, or antipsychotic medicine, and 8.5% a barbiturate. NIDA also monitors emergency department (ED) visits through its Drug Abuse Warning Network (DAWN). In 2006, ~250 ED visits per 100,000 population were related to drug toxicity and 49% of these visits were related to the nonmedical use and abuse of pharmaceutical agents (U.S. DHHS). The top five drugs involved in drug-related deaths reported to DAWN-participating medical examiners' offices in 2005 are presented in Table 4-3.

The AAPCC (www.aapcc.org) offers a toll-free poisoning information phone hotline throughout the U.S., and has been collecting voluntary reports of potential poison exposures for over 25 years. It currently has over 46 million human exposure case records in its database. In 2007, nearly 2.5 million cases, including 1239 fatal poisonings, were voluntarily reported to the AAPCC's National Poison Data System (NPDS). Eighty-three percent of human poison exposures reported to the NPDS were unintentional, 13% were intentional, and 2.5% were adverse drug reactions (Bronstein et al., 2008). The substances most frequently involved in human exposures and fatalities are presented in Tables 4-4 and 4-5.

# Top Five Agents Involved in Drug-Related Deaths

Cocaine **Opioids** Benzodiazepines Alcohol Antidepressants Source: U.S. DHHS.

# Substances Most Frequently Involved in Human Poisoning Exposures has all various and summer than some

Polsoning Exposures	%
Substance	
Analgesics	9.1
Personal care products	
	6.2
Sedatives/hypnotics/antipsychotics	5.1
Foreign bodies Tonical preparations	1.5
Topical preparations  Cold and cough medications	15
Antidepressants	4.0
Source: Data From Bronstein et al., 2008.	

### Table 4-5

# Poisons Associated with the Largest Number of Human Fatalities

Sedatives/hypnotics/antipsychotics Antidepressants Cardiovascular drugs Stimulants and street drugs Source: Bronstein et al., 2008.

# PREVENTION OF POISONING

Reduction of Medication Errors. Over the past decade considerable attention has been given to the reduction of medication errors and adverse drug events (ADEs). Medication errors can occur in any part of the medication prescribing or use process, while ADEs are injuries related to the use or nonuse of medications. It is believed that medication errors are 50-100 times

more common than ADEs (Bates et al., 1995). Some ADEs, such as previously unknown allergies, are unpreventable, but most ADEs can be prevented. Traditionally, the "5 Rights" of safe medication administration have been taught on hospital wards:

Right drug, right patient, right dose, right route, right time.

However, accomplishing a reduction in medication errors involves scrutiny of the systems involved in prescribing, documenting, transcribing, dispensing, administering, and monitoring a therapy, as presented in Appendix I. Good medication use practices have mandatory and redundant checkpoints, such as having a pharmacist, a doctor, and a nurse all review and confirm that an ordered dose of a medication is appropriate for a patient prior to the drug's administration. In such a system, medication errors occur only when several "holes" in the medication administration safeguards exist and are simultaneously aligned (Figure 4–7). Several practical strategies have been suggested to reduce medication errors within hospitals and other healthcare settings (Table 4-6), and these strategies are being constantly revised.

Poisoning Prevention in the Home. Table 4-2 demonstrates that there are several contexts into which poisoning prevention can be directed. Depression and suicidal ideation need to be identified and treated. Exposure to hazards in the home, outdoor, and work environments need to be reduced to reasonably achievable levels.

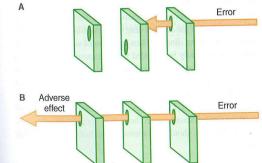


Figure 4-7. The "Swiss cheese" model of medication error. Several checkpoints typically exist to identify and prevent an adverse drug event, and that adverse event can only occur if holes in several systems align. A. One systematic error does not lead to an adverse event, because it is prevented by another check in the system. B. Several systematic errors align to allow an adverse event to occur. (Adapted from Reason, 2000.)

### Table 4-6

### **Best Practice Recommendations to Reduce** Medication Administration Errors

- · Maintain unit-dose distribution systems for nonemergency medications
- · Have pharmacies prepare intravenous solutions

81

DRUG

TOXICITY

AND

POISONING

- · Remove inherently dangerous medications (e.g., concentrated KCl) from patient care areas
- Develop special procedures for high-risk drugs
- Improve drug-related clinical information
- resources
- · Improve medication administration education for clinicians
- · Educate patients about the safe and accurate use of medications
- · Improve access of bedside clinicians to pharmacists

### Long Term

Implement technology-based safeguards:

- Computerized order entry
- · Computerized dose and allergy checking
- Computerized medication tracking
- · Use of bar codes or electronic readers for medication preparation and administration

<sup>a</sup>See Massachusetts Hospital Association.

Poisoning prevention strategies may be categorized as being passive, requiring no behavior change on the part of the individual, or active, requiring sustained adaptation to be successful. Passive prevention strategies are the most effective, and several types of passive poisoning prevention are described in Table 4–7.

### **Passive Poisoning Prevention Strategies** and Examples

Reduce manufacture/sale of poisons Withdrawal of phenformin from U.S. pharmaceutical

Decrease amount of poison in a consumer product Limiting number of pills in a single bottle of baby

Prevent access to poison

The use of child-resistant packaging Change product formulation

Removing ethanol from mouthwash

The incidence of poisoning in children has decreased dramatically over the past four decades. This favorable trend is largely due to improved safety packaging of drugs, drain cleaners, turpentine, and other household chemicals; improved medical training and care; and increased public awareness of potential poisons.

From 1958-1973, aspirin ingestion was a common cause of childhood poisoning death. In 1973, regulations were instituted requiring child-resistant packaging for aspirin consumer products, and this change in packaging was associated with a 34% reduction in the aspirin-related child mortality rate (Rodgers, 2002). In 2007, no aspirin deaths among children aged less than 6 years were reported to the AAPCC (Bronstein et al., 2008). From 1983-1990, iron was the single most frequent cause of unintentional pharmaceutical ingestion fatality in children younger than 6 years, and accounted for almost one-third of such deaths. In 1997, drug products with 30 mg or more of elemental iron per dosage unit were required to be placed into unit-dose packaging. This packaging change was associated with a decrease in mortality (odds ratio of 0.07; 95% confidence interval, 0.52-0.01) (Tenenbein, 2005).

# PRINCIPLES OF TREATMENT OF POISONING

The majority of poisoning exposures reported to U.S. poison control centers are judged to be nontoxic or only minimally toxic (Bronstein et al., 2008). When toxicity is expected, or does occur, the priority of poisoning treatment is to support vital functions until the drug or chemical is eliminated from the body. Because of the acute onset of action and finite duration of action of most drugs, the treatment of poisoning must be prompt and goal-directed. The first goal is to maintain vital physiological functions from impairment. The second goal is to keep the concentration of poison in tissues as low as possible by preventing absorption and enhancing elimination. The third goal is to combat the toxicological effects of the poison at the effector sites.

Initial Stabilization of the Poisoned Patient. The "ABC" mnemonic of emergency care is popularly taught and applies to the treatment of acute poisoning (Table 4-8). In severe cases, endotracheal intubation, mechanical ventilation, pharmacological blood pressure support, and/or extracorporeal circulatory support may be necessary and appropriate.

Identification of Clinical Patterns of Toxicity. A carefully obtained medical history may allow for the creation of a list of available medications or chemicals that might be implicated in a poisoning event. Often, an observation of physical symptoms and signs may be the only additional clues to a poisoning diagnosis.

Table 4-8

## **ABCDE: Initial Treatment Approach for Acute Poisoning**

Airway Breathing

Maintain patency Maintain adequate oxygenation and

ventilation

Circulation Maintain perfusion of vital organs Assess for central nervous system

> dysfunction If neurological disability is noted,

- Oxygen administration (check pulse oximetry)
- · Dextrose administration (check blood glucose concentration)
- Naloxone administration (consider
- Thiamine (for adult patients receiving dextrose)

Exposure Assess "toxidrome" (see Table 4–9)

Groups of physical signs and symptoms associated with specific poisoning syndromes are known as toxidromes (Erickson, 2007; Osterhoudt, 2004). Table 4-9 describes commonly encountered toxidromes.

The most typically available urine drug toxicology test is an immunoassay designed to detect common drugs of abuse such as amphetamines, barbiturates, benzodiazepines, cannabis, cocaine, and opiates. Acute poisoning with these substances can usually be determined on clinical grounds, and the results of these assays are infrequently available fast enough to guide stabilization. Additionally, detection of drugs or their metabolites on a urine immunoassay does not mean that the detected drug is responsible for the currently observed poisoning illness. When ingestion of acetaminophen or aspirin cannot clearly be excluded via the exposure history, serum quantification of these drugs is recommended. An electrocardiogram (ECG) may be useful at detecting heart blocks, Na+ channel blockade, or K+ channel blockade associated with specific medication classes (Table 4-10). Further laboratory analysis, such as use of blood gas determinations, serum chemistries, complete blood counts, and other testing, should be tailored to the individual poisoning

Decontamination of the Poisoned Patient. Poisoning exposures may be by inhalation, by dermal or mucosal absorption, by injection, or by ingestion. The first step in preventing absorption of poison is to stop any ongoing exposure. If necessary, eyes and skin should

Common Toxidrom	es management	South West board	ling	usols.	MARI	mylis	and the con-	ELEGEBRICA REPORTED FOR THE STATE OF THE STA
DRUG CLASS	EXAMPLE(S)	MENTAL STATUS	HR	BP	RR	T	PUPIL SIZE	OTHER STATE STATES
Sympathomimetic	Cocaine Amphetamine	Agitation	1	1			onditions (Ten	Tremor, diaphoresis
Anticholinergic	Diphenhydramine Belladonna atropa	Delirium	1	1		1	Construction of the constr	Ileus, flushing
Cholinergic	Organophosphates	Somnolence/coma			1		1	SLUDGE <sup>a</sup> , fasciculation
Opioid	Heroin Oxycodone	Somnolence/coma	1		1		Joseph Parking	
Sedative-hypnotic	Benzodiazepines Barbiturates	Somnolence/coma		1	<b>\</b>			
Salicylate	Aspirin Month	Confusion	1		1	1		Diaphoresis, vomiting
Ca <sup>2+</sup> channel blocker	Verapamil		1	1				

HR, heart rate; BP, blood pressure; RR, respiratory rate; T, temperature. aSLUDGE, muscarinic effects of Salivation, Lacrimation, Urination, Defecation, Gastric cramping, and Emesis.

be washed copiously. Gastrointestinal decontamination is the process of preventing or reducing absorption of a substance after it has been ingested. The primary strategies for GI decontamination are gastric emptying, adsorption of poison, and catharsis. Minimal indications for considering GI decontamination include: 1) the poison must be potentially dangerous; 2) the poison must still be unabsorbed in the stomach or intestine, so it must be soon after ingestion; and 3) the procedure must be able to be performed safely and with proper technique. Gastric emptying is rarely recommended anymore, but the administration of activated charcoal and the performance of whole bowel irrigation remain therapeutic options.

Differential Poisoning Diagnosi BRADYCARDIA/HEART BLOCK	QRS INTERVAL PROLON	IGATION	QTC INTERVAL PROLONGATION
Cholinergic agents Physostigmine Neostigmine Organophosphates, carbamates	Antiarrhythmia drugs Bupropion Chloroquine Diphenhydramine Lamotrigine		(See Arizona Center for Education and Research on Therapeutics website, http://www.azcert.org/medical-pros/ drug-lists/drug-lists.cfm)
Sympatholytic agents	Phenothiazines		
β receptor antagonists	Propranolol		
Clonidine Clonidine	Tricyclic antidepressa	nts about 1 A sou	
Opioids			
Other			
Digoxin  Ca <sup>2+</sup> channel blockers			
Lithium			

AND

(Bronstein et al., 2008).

Syrup of Ipecac. The U.S. FDA approved syrup of ipecac for sale without a prescription in 1965. The alkaloids cephaeline and emetine within syrup of ipecac act as emetics because of both a local irritant effect on the enteric tract and a central effect on the chemoreceptor trigger zone in the area postrema of the medulla. Syrup of ipecae is available in 0.5- and 1-fluid ounce containers. Ipecae is given orally at a dose of 15 mL for children up to 12 years, and 30 mL for older children and adults. Administration of ipecac is typically followed by a drink of water, and reliably produces emesis in 15-30 minutes. Contraindications for syrup of ipecac administration include existing or impending CNS depression, ingestion of a corrosive or hydrocarbon drug (due to the emergence of chemical pneumonia), or presence of a medical condition that might be exacerbated by vomiting. Ipecac has been misused by bulimic patients; in cases of Munchausen syndrome by proxy and with chronic abuse, ipecac may cause serum electrolyte abnormalities, cardiomyopathy, ventricular arrhythmia, and death.

Gastric Lavage. The procedure for gastric lavage involves passing an orogastric tube (24-French for small children, up to 40-French for adults) into the stomach with the patient in the left-lateral decubitus position with head lower than feet. Preferably, the tube will have been designed for lavage purposes, and will have sizable side holes in the tubing. After withdrawing stomach contents, 10 to 15 mL/kg (up to 250 mL) of saline lavage fluid is administered and withdrawn. This process continues until the lavage fluid returns clear. Complications of the procedure include mechanical trauma to the stomach or esophagus, pulmonary aspiration of stomach contents, and yagus nerve stimulation.

Adsorption of a poison refers to the binding of a poison to the surface of another substance. An adsorbed poison may be less available for absorption into the body. It is well known that the fullness of the stomach from a meal affects a drug's absorption kinetics. Fuller's earth has been suggested as an adsorbent for paraquat, Prussian blue binds thallium and cesium, and sodium polystyrene can adsorb lithium. The most

common adsorbent used in the treatment of acute drug overdose is activated charcoal.

Volunteer studies suggest that activated charcoal is more effective at reducing drug absorption than either induced emesis or gastric lavage (Tenebein, 1987). In a position paper of the American Academy of Clinical Toxicology on the use of single-dose activated charcoal, the opinion is given that single-dose charcoal should not be administered routinely in the management of poisoned patients, and that it should only be considered if a patient has ingested a potentially toxic amount of poison up to 1 hour before charcoal administration (AACT, 2005). In 2007, charcoal was used in 4.3% of cases reported to American poison control centers (Bronstein et al., 2008). Clinical evidence of improved patient parameters from treatment with activated charcoal are slowly emerging (Buckley et al., 1999; Isbister et al., 2007; Page et al., 2009), but good outcome data from clinical trials are still lacking.

Activated Charcoal. Charcoal is created through controlled pyrolysis of organic matter, and is *activated* through steam or chemical treatment that increases its internal pore structure and adsorptive surface capacity. The surface of activated charcoal contains carbon moieties, such as carbonyl and hydroxyl groups, that are capable of binding poisons. The recommended dose is typically 0.5-2 g/kg of body weight, up to a maximum tolerated dose of ~75-100 g. As a rough estimate, 10 g of activated charcoal is expected to bind ~1 g of drug. The efficacy of activated charcoal at adsorbing ingested drug diminishes over time. Alcohols, corrosives, hydrocarbons, and metals are not believed to be well adsorbed by charcoal.

Complications of activated charcoal therapy include vomiting, constipation, pulmonary aspiration, and death. Charcoal slurries are black and gritty; in a series of children offered charcoal in a pediatric emergency department, only 44% of the children <6 years accepted the agent orally (Osterhoudt et al., 2004a). Nasogastric administration of charcoal increases the incidence of vomiting (Osterhoudt et al., 2004b), and may increase the risk for pulmonary aspiration. Charcoal should not be given to patients with suspected GI perforation, or to patients who may be candidates for endoscopy.

Whole Bowel Irrigation. Whole bowel irrigation (WBI) involves the enteral administration of large amounts of a high molecular weight, iso-osmotic polyethylene glycol electrolyte solution with the goal of passing poison by the rectum before it can be absorbed. Potential candidates for WBI include: 1) "body-packers" with intestinal packets of illicit drugs; 2) patients with iron overdose; 3) patients who have ingested patch pharmaceuticals; and 4) patients with overdoses of sustained-release or bezoar-forming drugs.

Polyethylene glycol electrolyte solution is typically administered at a rate of 25 to 40 mL/kg/h until the rectal effluent is clear and no more drug is being passed. To achieve these high

administration rates a nasogastric tube may be used. Large doses have been administered without adversely affecting serum electrolyte concentrations. WBI is contraindicated in the presence of bowel obstruction or perforation, and may be complicated by abdominal distention or pulmonary aspiration.

Cathartics. The two most common categories of simple cathartics are the magnesium salts, such as magnesium citrate and magnesium sulfate, and the nondigestible carbohydrates, such as sorbitol. The use of simple cathartics has been abandoned as a GI decontamination strategy, although sorbitol is sometimes administered with single-dose activated charcoal in an effort to add sweetness and reduce its predilection toward constipation.

Enhancing the Elimination of Poisons. Once absorbed, the deleterious toxicodynamic effects of some drugs may be reduced by methods that hasten their elimination from the body. Urinary excretion of some drugs may be enhanced by the process of ion-trapping in alkaline urine. Gastrointestinal excretion of some drugs may be enhanced through use of multiple doses of activated charcoal. Some drugs may be removed from the body by extracorporeal techniques such as peritoneal dialysis, hemodialysis, or hemoperfusion.

Manipulating Urinary pH: Urinary Alkalinization. Drugs subject to renal clearance are excreted into the urine by glomerular filtration and active tubular secretion (Chapter 2); non-ionized compounds may be reabsorbed far more rapidly than ionized polar molecules. Weakly acidic drugs are susceptible to "ion-trapping" in the urine. Aspirin is a weak acid with a  $pK_a = 3.0$ . As the pH of the urine increases, more salicylate is in its ionized form at equilibrium, and more salicylic acid is diffused into the tubular lumen of the kidney. Urinary alkalinization is also believed to speed clearance of phenobarbital, chlorpropamide, methotrexate, and chlorphenoxy herbicides. The American Academy of Clinical Toxicologists recommends urine alkalinization as first-line treatment only for moderately severe salicylate poisoning that does not meet criteria for hemodialysis (Proudfoot et al., 2004). To achieve alkalinization of the urine, 100-150 mEq of sodium bicarbonate in 1L of D5W is infused intravenously at twice the maintenance fluid requirements and then titrated to effect. Hypokalemia should be treated since it will hamper efforts to alkalinize the urine due to H+-K+ exchange in the kidney. Urine alkalinization is contraindicated in the presence of renal failure, or when the fluid administration may worsen pulmonary edema or congestive heart failure. Acetazolamide is not used to alkalinize urine as it promotes acidemia.

Multiple-Dose Activated Charcoal. Activated charcoal adsorbs drug to its surface and promotes enteral elimination. Multiple doses of activated charcoal can speed elimination of absorbed drug by two mechanisms. Charcoal may interrupt enterohepatic circulation of hepatically metabolized drug excreted in the bile, and charcoal may create a diffusion gradient across the GI mucosa and promote movement of drug from the bloodstream onto the charcoal in the intestinal lumen. Activated charcoal may be administered in multiple doses, 12.5 g/h every 1, 2, or 4 hours (smaller doses may be used for children). Complications of therapy are similar to those listed for

single-dose activated charcoal. Charcoal enhances the clearance of many drugs of low molecular weight, small volume of distribution, and long elimination t<sub>1/2</sub>. In the absence of good clinical outcomes data, multiple-dose activated charcoal is believed to have the most potential utility in overdoses of carbamazepine, dapsone, phenobarbital, quinine, theophylline, and yellow oleander (AACT, 1999; de Silva et al., 2003).

Extracorporeal Drug Removal. The ideal drug amenable to removal by hemodialysis has a low molecular weight, a low volume of distribution, high solubility in water, and minimal protein binding. Hemoperfusion involves passing blood through a cartridge containing adsorbent particles. The most common poisonings for which hemodialysis is sometimes used include salicylate, methanol, ethylene glycol, lithium, carbamazepine, and valproic acid. For a more exhaustive list of drugs amenable to hemodialyis or hemoperfusion, see Winchester (2002).

Antidotal Therapies. Antidotal therapy involves antagonism or chemical inactivation of an absorbed poison. The pharmacodyamics of a poison can be altered by competition at a receptor, as in the antagonism provided by naloxone therapy in the setting of heroin overdose. A physiological antidote may use a different cellular mechanism to overcome the effects of a poison, as in the use of glucagon to stimulate an alternative to the blocked  $\beta$  adrenergic receptor and increase cellular cyclic AMP in the setting of propranolol overdose. Antivenoms and chelating agents bind and directly inactivate poisons. The biotransformation of a drug can also be altered by an antidote; for instance, fomepizole will inhibit alcohol dehydrogenase and stop the formation of toxic acid metabolites from ethylene glycol and methanol. Many drugs used in the supportive care of a poisoned patient (anticonvulsants, vasoconstricting agents, etc.) may be considered nonspecific functional antidotes.

The mainstay of therapy for poisoning is good support of the airway, breathing, circulation, and vital metabolic processes of the poisoned patient until the poison is eliminated from the body; specific antidotes are uncommonly needed. Among the most common specific antidotes used are N-acetyl-L-cysteine for acetaminophen poisoning, opioid antagonists for opioid overdose, and chelating agents for poisoning from certain metal ions. A listing of other commonly used antidotes is presented in Table 4–11.

# IMPORTANT RESOURCES FOR INFORMATION RELATED TO DRUG TOXICITY AND POISONING

Pharmacology textbooks offer important information pertaining to the toxic nature of drugs, but they may lack

Benztropine Bicarbonate, sodium Bromocriptine Calcium gluconate or chloride Carnitine Crotalidae polyvalent

immune Fab Dantrolene Deferoxamine Digoxin immune Fab Diphenhydramine Dimercaprol (BAL) EDTA, CaNa, Ethanol Fomepizole

Flumazenil Glucagon hydrochloride Hydroxocobalamin hydrochloride Insulin (high dose) Leucovorin calcium Methylene blue

Octreotide acetate

Oxygen, hyperbaric Penicillamine Physostigmine salicylate Pralidoxime chloride (2-PAM) Pyridoxine hydrochloride Succimer (DMSA) Thiosulfate, sodium Vitamin K.

(phytonadione)

Some Common Antidotes and Their Indications

POISONING INDICATION(S) Acetaminophen Organophosporus and carbamate pesticides Drug-induced dystonia

Na+ channel blocking drugs Neuroleptic malignant syndrome Ca2+ channel blocking drugs, Fluoride Valproate hyperammonemia North American crotaline snake envenomation

Malignant hyperthermia Cardiac glycosides Drug-induced dystonia Lead, mercury, arsenic

Methanol, ethylene glycol Methanol, ethylene glycol Benzodiazepines β adrenergic antagonists Cyanide

Ca2+ channel blockers Methotrexate Methemoglobinemia Opioids Naloxone hydrochloride Sulfonylurea-induced hypoglycemia Carbon monoxide Lead, mercury, copper Anticholinergic syndrome

Isoniazid seizures

Lead, mercury, arsenic Cyanide Coumarin, indanedione

Organophosphorus pesticides

discussion of household, industrial, or environmental chemicals, and they may lack detailed discourse of prevention, identification, and treatment of overdose. Additional information on poisoning from drugs and chemicals can be found in many dedicated books of toxicology (Flomenbaum, 2006; Klaassen, 2007; Olson, 2007; Shannon et al., 2007). A popular computer database

for information on toxic substances is POISINDEX (Micromedex, Inc., Denver, CO).

The National Library of Medicine offers information on toxicology and environmental health (http://sis. nlm.nih.gov/enviro.html), including a link to ToxNet (http://toxnet.nlm.nih.gov/), a cluster of full-text and bibliographic databases on toxicology, hazardous chemicals, and related areas.

Regional poison control centers are a resource for valuable poisoning information, and can be reached from anywhere within the U.S. through a national PoisonHelp hotline: 1-800-222-1222. Poison centers also collect epidemiological data regarding poisoning, perform all hazards surveillance, provide education, and work collaboratively with other agencies to effect poisoning prevention.

### BIBLIOGRAPHY

American Academy of Clinical Toxicology, and the European Association of Poisons Centres and Clinical Toxicologists. Position statement and practice guidelines on the use of multidose activated charcoal in the treatment of acute poisoning. Clin Toxicol, 1999, 37:731-751.

American Academy of Clinical Toxicology, and the European Association of Poisons Centres and Clinical Toxicologists. Position paper: Gastric lavage. J Toxicol Clin Toxicol, 2004, 42:933-943.

American Academy of Clinical Toxicology, and the European Association of Poisons Centres and Clinical Toxicologists. Position paper: Single-dose activated charcoal. Clin Toxicol, **2005**, 43:61-87.

American Academy of Pediatrics Committee on Injury, Violence, and Poison Prevention. Poison treatment in the home. Pediatrics, 2003, 112:1182-1185.

Arizona Center for Education and Research on Therapeutics. Drugs that prolong the QT interal and/or induce torsades de pointes ventricular arrhythmia. http://www.azcert.org/medicalpros/drug-lists/drug-lists.cfm. Accessed January 23, 2010.

Barbui C, Esposito E, Cipriai A. Selective serotonin reuptake inhibitors and risk of suicide: A systematic review of observational studies. Can Med Assoc J, 2009, 180:291–297.

Bates DW, Boyle DL, Vander Bliet MB, et al. Relationship between medication errors and adverse drug events. J Gen Intern Med, 1995, 10:199-205.

Bronstein AC, Spyker DA, Cantilena LR, et al. 2007 Annual Report of the American Association of Poison Control Centers' National Poison Data System (NPDS): 25th annual report. Clin Toxicol, 2008, 46:927-1057.

Buckley NA, Whyte IM, O'Connell DL, Dawson AH. Activated charcoal reduces the need for N-acetylcysteine treatment after acetaminophen overdose. Clin Toxicol, 1999,

Dart RC, Erdman AR, Olson KR, et al. Acetaminophen poisoning: an evidence-based consensus guideline for out-ofhospital management. Clin Toxicol, 2006, 44:1-18.

Erickson TE, Thompson TM, L'u JJ. The approach to the patient with an unknown overdose. Emerg Med Clin North Am, 2007, 25:249-281

de Silva HA, Foneska MM, Pathmeswaran A, et al. Multipledose activated charcoal for treatment of yellow oleander poisoning: A single-blind, randomized, placebo-controlled trial. Lancet, 2003, 361:1935-1938.

Evans WE, Rolling MV. Pharmacogenomics: translating functional genomics into rational therapeutics. Science, 1999, 286:487-491.

Flomenbaum NE, Goldfrank LR, Hoffman RS, et al., eds. Goldfrank's Toxicologic Emergencies, 8th ed. McGraw-Hill, New York, 2006.

Guthrie SK, Stoysich AM, Bader G, Hilleman DE. Hypothesized interaction between valproic acid and warfarin. J Clin Psychopharmacol, 1995, 15:138-139.

Hennessy S, Leonard CE, Newcomb C, et al. Cisapride and ventricular arrhythmia. Br J Clin Pharmacol, 2008, 66:375-385. Hibbard DM, Peters JR, Hunninghake DB. Effects of cholestyramine and colestipol on the plasma concentrations of propralolol. Br J Clin Pharmacol, 1984, 18:337-342.

Institute of Medicine. To Err Is Human: Building a Safer Health System. National Academy Press, Washington, DC, 1999.

Isbister GK, Friberg LE, Stokes B, et al. Activated charcoal decreases the risk of QT prolongation after citalopram overdose. Ann Emerg Med, 2007, 50:593-600.

Klaassen CD, ed. Casarett and Doull's Toxicology: The Basic Science of Poisons, 7th ed. McGraw-Hill, New York, 2007.

Lazarou J, Pomeranz BH, Corey PN. Incidence of adverse drug reactions in hospitalized patients: A meta-analysis of prospective studies, JAMA, 1998, 279:1200-1205.

Manoguerra AS, Cobaugh DJ, Guidelines for the Management of Poisonings Consensus Panel. Clin Toxicol, 2005, 43:1-10.

Massachusetts Hospital Association. MHA best practice recommendations to reduce medication errors. Available at: http://macoalition.org/documents/Best\_Practice\_Medication\_ Errors.pdf. Also see: http://www.macoalition.org/ initiatives. shtml. Accessed April 9, 2010.

National Institute on Drug Abuse. Johnston LD, O'Malley PM, Bachman JG, Schulenberg JE. Monitoring the future: National results on adolescent drug use—overview of key findings 2008. Available at: http://www.monitoringthefuture.org/ pubs/ monographs/overview2008.pdf. See also: http://www.drugabuse. gov/DrugPages/MTF.html. January 23, 2010.

O'Connor-Semmes RL, Kersey K, Williams DH, et al. Effect of ranitidine on the pharmacokinetics of triazolam and alphahydroxytriazolam in both young and older people. Clin Pharmacol Ther, 2001, 70:126-131.

Olson KR, ed. Poisoning & Drug Overdose, 5th ed. McGraw-Hill, 87 New York, 2007.

Osterhoudt KC. No sympathy for a boy with obtundation. Pediatr Emerg Care, 2004, 20:403-406.

Osterhoudt KC. The lexiconography of toxicology. J Med Toxicol, 2006, 2:1-3.

Osterhoudt KC, Alpern ER, Durbin D, et al. Activated charcoal administration in a pediatric emergency department. Pediatr Emerg Care, 2004a, 20:493-498.

Osterhoudt KC, Durbin D, Alpern ER, Henretig FM. Risk factors for emesis after therapeutic use of activated charcoal in acutely poisoned children. Pediatrics, 2004b, 113:806-810.

Page CB, Duffull SB, Whyte IM, Isbister GK. Promethazine overdose: Clinical effects, predicting delirium and the effect of charcoal. QJ Med, 2009, 102:123-131.

Pond SM, Lewis-Driver DJ, Williams GM, et al. Gatric emptying in acute overdose: A prospective randomized trial. Med J Australia, 1995, 163:345-349.

Proudfoot AT, Krenzelok EP, Vale JA. Position paper on urine alkalinization. J Toxicol Clin Toxicol, 2004, 42:1-26.

Reason J. Human error: Models and management. Br Med J, 2000, 320:768-770

Rivera W, Kleinschmidt KC, Velez LI, et al. Delayed salicylate toxicity at 35 hours without early manifestations following a single salicylate ingestion. Ann Pharmacother, 2004, 38:1186-1188.

Rodgers GB. The effectiveness of child-resistant packaging for aspirin. Arch Pediatr Adolesc Med, 2002, 156:929-933.

Shannon MW, Borron SW, Burns MJ, eds. Haddad and Winchester's Clinical Management of Poisoning and Drug Overdose, 4th ed. Saunders/Elsevier, Philadelphia, 2007.

Strom BL. Potential for conflict of interest in the evaluation of suspected adverse drug reactions. JAMA, 2004, 292:2643-

Sztajnkrycer MD. Valproic acid toxicity: Overview and management. Clin Toxicol, 2002, 40:789-801.

Tenenbein M. Unit-dose packaging of iron supplements and reduction of iron poisoning in young children. Arch Pediatr Adolesc Med, 2005, 159:557-560.

Tenenbein M, Cohen S, Sitar DS. Efficacy of ipecac-induced emesis, orogastric lavage, and activated charcoal for acute drug overdose. Ann Emerg Med, 1987, 16:838-841.

U.S. Department of Health and Human Services. Drug abuse warning network. Available at: https://dawninfo.samhsa.gov/ default.asp. Accessed April 9, 2010.

Winchester JF. Dialysis and hemoperfusion in poisoning. Adv Renal Replace Ther, 2002, 9:26-30.