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(54) **SYSTEM, ARRAY AND NON-POROUS SOLID SUPPORT COMPRISING FIXED OR IMMOBILIZED NUCLEIC ACIDS**

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See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

3,669,841 A	6/1972	Miller	195/63
3,715,278 A	2/1973	Miller	195/63
3,849,137 A	11/1974	Barzynski	
3,949,064 A	4/1976	Bornstein et al.	424/1
4,001,583 A	1/1977	Barrett	250/303
4,041,146 A	8/1977	Giaever	424/1
4,059,685 A	11/1977	Johnson	424/12
4,106,907 A	8/1978	Charlton et al.	
4,116,638 A	9/1978	Kenoff	422/99

4,166,103 A	8/1979	Wagner et al.	424/1
4,166,104 A	8/1979	Wagner et al.	424/1
4,228,237 A	10/1980	Hevey et al.	435/7
4,230,797 A	10/1980	Boguslaski et al.	435/7
4,234,563 A	11/1980	Rippe	
4,234,681 A	11/1980	DeLuca-McElroy	435/8
4,251,514 A	2/1981	Rippe	
4,254,097 A	3/1981	Rippe	
4,261,893 A	4/1981	Boguslaski et al.	260/326
4,269,933 A	5/1981	Pazos	
4,271,140 A	6/1981	Bunting	424/1
4,280,992 A	7/1981	Sugiura et al.	424/1
4,302,204 A	11/1981	Wahl et al.	23/230.3
4,312,944 A	1/1982	Mattiasson et al.	435/7
4,318,980 A	3/1982	Boguslaski et al.	435/7
4,318,981 A	3/1982	Burd et al.	435/7
4,318,982 A	3/1982	Hornby et al.	435/7
4,358,535 A	11/1982	Falkow et al.	435/5
4,374,925 A	2/1983	Litman et al.	435/7
4,376,110 A	3/1983	David et al.	436/513
4,380,580 A	4/1983	Boguslaski et al.	435/7
4,383,031 A	5/1983	Boguslaski et al.	435/7
4,391,904 A	7/1983	Litman et al.	435/7
4,446,231 A	5/1984	Self	435/7
4,483,920 A	11/1984	Gillespie et al.	435/6
4,486,539 A	12/1984	Ranki et al.	436/504
4,516,833 A	5/1985	Fusek	
4,517,338 A	5/1985	Urdea	
4,537,861 A	8/1985	Elings	

(Continued)

FOREIGN PATENT DOCUMENTS

DE P2618419 4/1976

(Continued)

OTHER PUBLICATIONS

Manuelidis et al. (1982) *Journal of Cell Biology*, vol. 95, pp. 619-625.*

(Continued)

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(57) **ABSTRACT**

Nucleic acids are fixed or immobilized to non-porous solid supports (substrates), and include systems containing such supports and arrays with fixed or immobilized nucleic acids. These compositions are useful for nucleic acid analyses and a host of applications, including, for example, detection, mutational analysis and quantification. The non-porous solid supports can be transparent or translucent, and the surfaces can be treated with agents to fix or immobilize the nucleic acids. Such agents include, for example, amine providing compounds, epoxy compounds and acid solutions. The fixed or immobilized nucleic acids can be unlabeled, or labeled with at least one non-radioactive signaling moiety, such as the case when the nucleic acids are double-stranded.

U.S. PATENT DOCUMENTS

4,542,102	A	9/1985	Dattagupta	
4,562,157	A	12/1985	Lowe	
4,563,419	A	1/1986	Ranki et al.	435/6
4,581,333	A	4/1986	Kourilsky et al.	435/6
4,358,535	A	5/1986	Falkow et al.	
4,656,127	A	4/1987	Mundy	
4,689,405	A	8/1987	Frank	
4,711,955	A	12/1987	Ward et al.	536/29
4,713,326	A	12/1987	Dattagupta	
4,724,202	A	2/1988	Dattagupta et al.	435/6
4,732,847	A *	3/1988	Stuart et al.	435/6
4,824,776	A	4/1989	Heller	435/6
4,973,493	A	11/1990	Guire	
4,994,373	A *	2/1991	Stavrianopoulos et al.	435/6
5,098,825	A	3/1992	Tchen et al.	
5,241,060	A	8/1993	Engelhardt et al.	536/27
5,260,433	A	11/1993	Engelhardt et al.	536/23.1
5,328,824	A	7/1994	Ward et al.	435/6
5,449,767	A	9/1995	Ward et al.	563/6
5,476,928	A	12/1995	Ward et al.	536/24

FOREIGN PATENT DOCUMENTS

DE	A2724486	12/1977
DE	2915082	10/1979
DE	A2915082	10/1979
EP	0046083	2/1982
EP	0 070 687	7/1982
EP	0063879 B1	11/1982
EP	0070685	1/1983
EP	0070687	1/1983
EP	0097373 B1	1/1984
EP	0103197	3/1984
GB	1548741	7/1979
GB	A2014727	7/1979
GB	1552607	9/1979
GB	2019408 *	10/1979
GB	2019408 A	10/1979
GB	A2026690	2/1980
GB	2041922 A	9/1980
GB	2045239	10/1980
GB	2125946 A	3/1984
JP	2825090	11/1998
WO	WO 83/02276	7/1983
WO	WO 83/02286	7/1983
WO	WO 8302277	7/1983
WO	WO8403564	9/1984

OTHER PUBLICATIONS

Langer et al., "Enzymatic Synthesis of Biotin-Labeled Polynucleotides: Novel Nucleic Acid Affinity Probes," *Proc. Natl. Acad. Sci. (USA)*, 78 (11):6633-6637 (Nov. 1981).

Ruth et al., "C-5 Substituted Pyrimidine Nucleosides. I. Synthesis of C-5 Allyl, Propyl, and Propenyl Uracil and Cytosine Nucleosides via Organopalladium Intermediates," *J. Org. Chem.* 43(14):2870-2876 (1978).

Bergstrom et al., "C-5 Substituted Pyrimidine Nucleosides. 2. Synthesis, via Olefin Coupling to Organopalladium Intermediates Derived from Uridine and 2'-Deoxyuridine," *J. Amer. Chem. Soc.* 100(26):8106-8112 (1978).

Bigge et al., "Palladium-Catalyzed Coupling Reactions of Uracil Nucleosides and Nucleotides," *J. Amer. Chem. Soc.* 102:2033 (1980).

Weetal et al., "Porous Glass for Affinity Chromatography Applications," *Methods in Enzymology*, vol. 34, *Affinity Techniques Enzyme Purification: Part B*, pp. 59-72 (Jakoby, W.B. and Wilchek, M., eds.) (1974).

Chard T., *An Introduction To Radioimmunoassay And Related Techniques*, Elsevier Science Publishers, B.V. (1978).

Grunstein, M., "Colony Hybridization: A Method For The Isolation of Cloned DNAs That Contain A Specific Gene," *Proc. Natl. Acad. Sci. (USA)* 72:3961-3965 (1975).

Stavrianopoulos et al., "Glycosylated DNA Probes For Hybridization/ Detection of Homologous Sequences," presented at the Third Annual Congress For Recombinant DNA Research (1983).

Singer et al., "Actin Gene Expression Visualized In Chicken Muscle Tissue Culture by Using *In Situ* Hybridization With A Biotinated Nucleotide Analog," *Proc. Nat'l Acad. Sci (USA)* 79:7331-7335 (1982).

Kennell, "Principles and Practices of Nucleic Acid Hybridization," *Progr. Nuc. Acid. Res. Mol. Biol.*, vol. 11, pp. 259-262 (1971).

Alberts et al., *Molecular Biology of the Cell*, Garland Publishers, New York and London (1983), p. 719.

Hood et al., *Immunology*, Benjamin/Cummings Publishing Co., Inc. Menlo Park, CA (1978), p. 142.

Schuurs et al., "Enzyme Immunoassay," *Clinica Chimica Acta* 81:1-40 (1977).

Langer-Sofer et al., "Immunological method for mapping genes on Drosophila polytene chromosomes," *Proc. Natl. Acad. Sci. (USA)* 79:4381-4385 (Jul. 1982).

Guesdon et al., "The Use of Avidin-Biotin Interaction in Immunoenzymatic Techniques," *Journal of Histochemistry and Cytochemistry* 27(8):1131-1139 (1979).

Bauman et al., "A new method for fluorescence microscopical localization of specific DNA sequences by in situ hybridization of fluorochrome-labelled RNA," *Exp. Cell Res.* 128:485-490 (1980).

Avrameas et al., "Enzyme immunoassay for the measurement of antigens using peroxidase conjugates," *Biochimie* 54:837-842 (1972).

John et al., "RNA-DNA Hybrids at the Cytological Level," *Nature* 223: 582-587 (1969).

Benton, W.D. and David, R. W., "Screening Agt recombinant clones by hybridization to single plaques in situ," *Science* 196: 180 (1977).

Leary, Brigati and Ward, "Rapid and sensitive colorimetric method for visualizing biotin-labeled DNA probes hybridized to DNA or RNA immobilized on nitrocellulose: Bio-blots," *Proc. Natl. Acad. Sci (USA)* 80: 4045-4049 (Jul. 1983).

Avrameas and Guilbert, "Enzyme-immunoassay for the measurement of antigens using peroxidase conjugates," *Biochimie* 54: 837-842 (1972).

Bauman et al., "Rapid and High Resolution Detection of *in situ* Hybridization to Polytene Chromosomes Using Fluorochrome-Labeled RNA," *Chromosoma (Berl.)* 84: 1-18 (1981).

Bildwell et al., "Enzyme Immunoassays for Viral Diseases," *J. Infectious Disease: 136: S274-S278* (1977).

Broker et al., "Electron Microscopic Visualization of tRNA genes with Ferritin-Avidin: Biotin Labels," *Nucl. Acids Res.*, 5(2): 363-384 (1978).

Engvall and Perlmann, "Enzyme-linked immunosorbent assay (ELISA) Quantitative assay of immunoglobulin G," *Immunochem* 8: 871-874 (1971).

Fertel R. and Weiss, B., *Methods in Enzymol.* vol. LV II, *Biochemical Assays*, Academic Press, New York, 1978, p. 111.

- Hamaguchi et al., "Enzyme-Linked Sandwich Immunoassay of Macromolecular Antigens Using the Rabbit Antibody-Coupled Glass Rod as a Solid Phase," *Eur. J. Biochem.* 7: 459-467 (1976) and *FEBS Letters*: 69(1): 11-14 (1976).
- Hofmann et al., "Iminobiotin Affinity Columns and Their Application to Retrieval of Streptavidin", *Proc. Natl. Acad. Sci. USA*, 77, No. 8, pp. 4666-4668 (1980).
- Miranda, Q.R., et al., "Solid-Phase Enzyme Immunoassay for Herpes Simplex Virus," *J. Infectious Disease* 136: S304-S310 (Oct., 1977).
- Langer and Ward, Abstract 1153: A Rapid and Sensitive Immunological Method for *In Situ* Gene Mapping in *Journal of Supramolecular Structure and Cellular Biology*, (1981).
- Langer and Ward, "A Rapid and Sensitive Immunological Method for *In Situ* Gene Mapping," in *Developmental Biology Using Purified Genes*, ed. D.D. Brown, Academic Press, pp. 647-658 (1981).
- Mosback, K., et al., "immobilized Coenzymes," *Methods in Enzymology*, vol. XLIV: 859-887 (1976).
- Nishimura et al. "Synthetic Nucleosides and Nucleotides: 5-Dimethylamino-2-oxidoisoquinolin-1 yl Diazomethane: A Novel Water Soluble Fluorescent Labelling Agent for Nucleotides," *Chem. Pharm. Bull.*, 28(6): 1695-1703 (1980).
- Rosswell, D.F. and White E.H., "The Chemiluminescence of Luminol and Related Hydrazides" in *Methods in Enzymol.* vol. LV II, *Bioluminescence and Chemiluminescence*, DeLuca, M.A. (Ed.) 409-423 (1978).
- Schott, H., et al., "A Dihydroxyboryl-Substituted Methacrylic Polymer for the Column Chromatographic Separation of Mononucleotides, Oligonucleotides, and Transfer Ribonucleic Acid," *Biochemistry* 12: 932-937 (1973).
- Rudkin and Stollar, "High Resolution Detection of DNA-RNA Hybrids *in situ* by Indirect Immunofluorescence," *Nature*, 265: 472-73 (1977).
- Sodja and Davidson, "Gene Mapping and Gene Enrichment by the Avidin-Biotin Interaction: Use of Cytochrome-C as a Polyamine Bridge," *Nucl. Acids. Res*, 5, pp. 385-400 (1978).
- Voller et al., "Enzyme immunoassays with special reference to ELISA techniques," *J. Clinical Pathology*, 31: 507-520 (1978).
- Towbin H. et al., "Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: Procedure and some applications," *Proc. Natl. Acad. Sci (USA)* 76: 4350-4354 (1979).
- Weissback, A., and Poonian, M., "Nucleic Acids Attached to Solid Matrices," *Methods in Enzymology*, vol. XXXIV, Part B: 463-475 (1974).
- Van Weemen and Schuurs, "Immunoassay Using Antigen-Enzyme Conjugates," *FEBS Letters* 15, No. 3: 232-236 (1971).
- Mathews, J.C. and Cormier, M.J., "Rapid Microassay for the Calcium-Dependent Protein Modulator of Cyclic Nucleotide Phosphodiesterase," *Methods in Enzymol.* vol. LV II, *Bioluminescence and Chemiluminescence*, DeLuca, M.A. (Ed.) 107-108 (1978).
- "Diagnostic Immunology: Current and Future Trends," *Cap Conference*, Aspen, 1978, p. 67 and 80.
- Voller et al., "A microplate method of enzyme-linked immunosorbent assay and its application to malaria," *Bull. W.H.O.*, 51: 209-211 (1974).
- Szostak et al., "Hybridization with Synthetic Oligonucleotides," *Methods in Enzymology*, 69: 416-428 (1980).
- So, et al., "Characterization of an *Escherichia coli* Plasmid Encoding for the Synthesis of Heat-Labile Toxin: Molecular Cloning of the Toxin Determinant", *Infection and Immunity*, 21: 405-11(1978).
- Reiser et al., "Transfer of Small DNA Fragments from Polyacrylamide Gels to Diazobenzyloxymethyl-paper and Detection by Hybridization with DNA Probes," *Biochem. And Biophys. Res. Comm.*, 85, No. 3, pp. 1104-1112 (1978).
- Mesulam, M. M. and Rosene, D.L., "Sensitivity in Horse-radish Peroxidase Neurohistochemistry: A Comparative Study of Nine Methods," *J. Histochem. Cytochem.* 27: No. 3, pp. 763-778 (1979).
- Land, D.B. and Jackim, E., "A New Fluorescence-Yielding Substrate for Alkaline and Acid Phosphatase," *Analytical Biochemistry*, 16: 481-486 (1966).
- Kochetkov, N.K. et al., *Organic Chemistry of Nucleic Acids, Part B*, Kochetkov, N.K., and Budovskii, E.I. (Eds.): 331-332 (1972).
- Huang and Pagano, "Nucleic Acid Hybridization Technology and Detection of Proviral Genomes," *Methods in Virology*, 6: 457-97 (1977).
- Fertel R. and Weiss, B., "Measurement of the Activity of Cyclic Nucleotide Phosphodiesterases with Firefly Luciferin-Luciferase Coupled Assay Systems," *Methods in Enzymol.* vol. LVII, *Bioluminescence and Chemiluminescence*, DeLuca, M.A. (Ed.) 94-96 (1978).
- Dallas et al. "The Characterization of an *Escherichia coli* Plasmid Determinant That Encodes for the Production of a Heat-Labile Enterotoxin," *Plasmids of Medical Environmental and Commercial Importance*, K.N. Timmis and A. Puhler, editors, Elsevier/North-Holland Biomedical Press (1979).
- Hofmann et al., "Characterization of the Functional Groups of Biotin", *J. Biol. Chem.*, 141, 207-11 (1941).
- Elisa: In The Clinical Microbiology Laboratory*, ed. T. G. Wreghitt and P. Morgan-Capner, Chapter 1, p. 9, 1990.
- Dallas and Falkow, "Molecular and Genetic Analysis of a DNA sequence Encoding for Enterotoxin Synthesis in *Escherichia coli*," *Thirteenth Joint Conference on Cholera*, The U.S.—Japan Cooperative Medical Science Program (1977).
- Stuart, W.D., et al., "Location of the 18/28S ribosomal RNA genes in two Hawaiian *Drosophila* species by monoclonal immunological identification of RNA-DNA hybrids *in situ*," *Proc. Natl. Acad. Sci. USA* 78:3751-3754 (1981).
- Haugen et al., Monoclonal antibody to aflatoxin B1-modified DNA detected by enzyme immunoassay, *PNAS*, 78, pp. 4124-4127 (1981).
- Vogelstein, B. and Gillespie, D., Preparative and Analytical Purification of DNA From Agarosa, *Proc. Natl. Acad. Sci. USA* vol. 76, No. 2, pp. 615-619, Feb. 1979.
- Munns and Liszewski, "Antibodies specific for Modified Nucleosides et al.," *Progr. Nucl. Acid Res. & Molec. Bio.*, Academic Press, Inc., vol. 24, pp. 109-165 (1980).
- Nagata et al., "Quantification of picogram levels of specific DNA immobilized in microtiter wells," *FEBS*, 183, pp. 379-382 (Apr. 1985).
- Sedlacek, H. et al., A New Method for Fluorescence Immunoassay Using Plane Surface Solid Phases (FIAPS), *J. Immun. Methods*, 26, (1979) 11-24.
- Van Der Laken et al., "Measurement of O6-ethyldeoxyguanosine and N-(deoxyguanosin-8-yl)-N-ethyl-2'-deoxyguanosine in DNA," *Methods in Enzymology*, 132: 1-12 (1986).

- Joseph Gall and Mary Lou Perdue, Formation and Detection of RNA-DNA Hybrid Molecules in Cytological Preparations, *Genetics*, 63: 378-383, Mar. 1969.
- Buongiorno-Nardelli, M. and Amaldi, F., Autoradiographic Detection of Molecular Hybrids Between rRNA and DNA in Tissue Sections, *Nature*, vol. 225, Mar. 7, 1970.
- Ueda R. et al., Serological Analysis of Cell Surface Antigens of Null Cell Acute Lymphocytic Leukemia by et al., *Proc. Natl. Acad. Sci. USA*, vol. 79, pp. 4386-4390, Jul. 1982.
- Method in Nucleic Acid Research, "DNA-DNA Hybridization", Kyoritsu Shuppan Co., Ltd., 1973, pp. 60-67 and 242. English Translation for Method in Nucleic Acid Research, "DNA-DNA Hybridization", Kyoritsu Shuppan Co., Ltd., 1973, pp. 60-67 and 242.
- Dictionary of Biochemistry, Tokyo Kagaku Doujin, 1984, pp. 727, 507, 450 and one other page. English Translation for Dictionary of Biochemistry, Tokyo Kagaku Doujin, 1984, pp. 727, 507, 450 and one other page.
- Mage M.G. et al, *Journal of Immunological Methods* 15:47-56 (1977).
- Wysocki L.J. and Sato, V.L., *Proc.Natl.Acad.Sci (USA)* 75(6):2844-2848 (Jun. 1978).
- Landreth K.S. et al., *roc.Natl.Acad.Sci (USA)* 79:2370-2374 (1982).
- Bunemann H. et al., *Nucleic Acids Research* 10(22):7163-7180 (1982).
- Bunemann H., *Nucleic Acids Research* 10(22):7181-7196 (1982).
- Beltz G.A. et al., *Methods in Enzymology* 100:266-285 (1983).
- Kafatos F.C et al., *Nucleic Acids Research* 7(6): 1541-1552 (1979).
- Masiakowski P. et al., *Nucleic Acids Research* 10(24):7895-7903 (1982).
- Sim G.K. et al., *Cell* 18:1303-1316 (1979).
- Frank R. et al., *Nucleic Acids Research* 11(13):4365-4377 (1983).
- Calva E. et al., *The Journal of Biological Chemistry* 255(22):11011-11016 (1980).
- Wengler G. et al., *Virology* 78:124-134 (1977).
- Coffin J.M. et al., *Cell* 13:761-773 (1978).
- Manning J.E. et al., *Chromosoma (Berl.)* 53:107-117 (1975).
- Manning J. et al., *Biochemistry* 16(7): 1364-1370 (1977).
- Maxam, A.M. and Gilbert, W., *Methods in Enzymology* 65:499-560 (1980).
- Sanger, F. et al., *Proc. Natl. Acad. Sci (USA)* 74(12):5463-5467 (1977).
- Amit B. et al., *Journal of Organic Chemistry* 39(2): 192-196 (1974).
- Amit B. et al., *Israel Journal of Chemistry* 12(1-2): 103-113 (1974).
- Barltrop J.A. et al., *Chemical Communications* 22:822-823 (1966).
- Flanders D.C. and Smith, H.I., *Applied Physics Letters* 31(7):426-428 (1977).
- Krile T.F. et al., *Applied Optics* 18(1):52-56 (1979).
- Ohtsuka E. et al., *Nucleic Acids Research* 1(10):1351-1357 (1974).
- Patchornik A. and Amit B., *Journal of the American Chemical Society* 92:6333-6335 (1970).
- Pillai V.N.R., *Synthesis* (International Journal of Methods in Synthetic Organic Chemistry Chemistry), Schill, G. et al., eds., Georg Thieme Verlag publishers, Stuttgart and New York, pp. 1-26, published 1980.
- Zehavi U. et al., *Journal of Organic Chemistry* 37(14):2281-2285 (1972).
- Seed B., *Nucleic Acids Research* 10(5):1799-1810 (1982).

* cited by examiner

**SYSTEM, ARRAY AND NON-POROUS SOLID
SUPPORT COMPRISING FIXED OR
IMMOBILIZED NUCLEIC ACIDS**

CROSS-REFERENCE TO OTHER RELATED
APPLICATIONS

This is a continuation application of U.S. Patent Application Ser. No. 07/967,646, filed on Oct. 28, 1992, now abandoned, which application is a continuation application of U.S. Patent Application Ser. No. 07/607,347, filed on Oct. 30, 1990, also abandoned. Ser. No. 07/607,347 is a continuation of U.S. Patent Application Ser. No. 07/385,986, filed on Jul. 20, 1989, now U.S. Pat. No. 4,994,373 issued on Feb. 19, 1991. Ser. No. 07/385,986 is a continuation of U.S. Patent Application Ser. No. 06/732,374, filed on May 9, 1985, also abandoned, which application is a continuation-in-part of U.S. Patent Application Ser. No. 06/461,469, filed on Jan. 27, 1983, also abandoned.

TECHNICAL FIELD OF INVENTION

The present invention relates generally to the detection of genetic material by polynucleotide probes. More specifically, it relates to a method for quantifiably detecting a targeted polynucleotide sequence in a sample of biological and/or nonbiological material employing a probe capable of generating a soluble signal. The method and products disclosed herein in accordance with the invention are expected to be adaptable for use in many laboratory, industrial, and medical applications wherein quantifiable and efficient detection of genetic material is desired.

BACKGROUND OF THE INVENTION

In the description, the following terms are employed:

Analyte—A substance or substances, either alone or in admixtures, whose presence is to be detected and, if desired, quantitated. The analyte may be a DNA or RNA molecule of small or high molecular weight, a molecular complex including those molecules, or a biological system containing nucleic acids, such as a virus, a cell, or group of cells. Among the common analytes are nucleic acids (DNA and RNA) or segments thereof, oligonucleotides, either single- or double-stranded, viruses, bacteria, cells in culture, and the like. Bacteria, either whole or fragments thereof, including both gram positive and gram negative bacteria, fungi, algae, and other microorganisms are also analytes, as well as animal (e.g., mammalian) and plant cells and tissues.

Probe—A labelled polynucleotide or oligonucleotide sequence which is complementary to a polynucleotide or oligonucleotide sequence of a particular analyte and which hybridizes to said analyte sequence.

Label—That moiety attached to a polynucleotide or oligonucleotide sequence which comprises a signalling moiety capable of generating a signal for detection of the hybridized probe and analyte. The label may consist only of a signalling moiety, e.g., an enzyme attached directly to the sequence. Alternatively, the label may be a combination of a covalently attached bridging moiety and signalling moiety or a combination of a non-covalently bound bridging moiety and signalling moiety which gives rise to a signal which is detectable, and in some cases quantifiable.

Bridging Moiety—That portion of a label which on covalent attachment or non-covalent binding to a polynucle-

Signalling Moiety—That portion of a label which on covalent attachment or non-covalent binding to a polynucleotide or oligonucleotide sequence or to a bridging moiety attached or bound to that sequence provides a signal for detection of the label.

Signal—That characteristic of a label or signalling moiety that permits it to be detected from sequences that do not carry the label or signalling moiety.

The analysis and detection of minute quantities of substances in biological and non-biological samples has become a routine practice in clinical, diagnostic and analytical laboratories. These detection techniques can be divided into two major classes: (1) those based on ligand-receptor interactions (e.g., immunoassay-based techniques), and (2) those based on nucleic acid hybridization (polynucleotide sequence-based techniques).

Immunoassay-based techniques are characterized by a sequence of steps comprising the non-covalent binding of an antibody and antigen complementary to it. See, for example, T. Chard, *An Introduction To Radioimmunoassay And Related Techniques* (1978).

Polynucleotide sequence-based detection techniques are characterized by a sequence of steps comprising the non-covalent binding of a labelled polynucleotide sequence or probe to a complementary sequence of the analyte under hybridization conditions in accordance with the Watson-Crick base pairing of adenine (A) and thymine (T), and guanine (G) and cytosine (C), and the detection of that hybridization. [M. Grunstein and D. S. Hogness, "Cloned DNAs That Contain A Specific Gene", *Proc. Natl. Acad. Sci. USA*, 72, pp. 3961-65 (1975)]. Such polynucleotide detection techniques can involve a fixed analyte [see, e.g., U.S. Pat. No. 4,358,535 to Falkow et al], or can involve detection of an analyte in solution [see U.K. patent application 2,019, 408 A].

The primary recognition event of polynucleotide sequence-based detection techniques is the non-covalent binding of a probe to a complementary sequence of an analyte, brought about by a precise molecular alignment and interaction of complementary nucleotides of the probe and analyte. This binding event is energetically favored by the release of non-covalent bonding free energy, e.g., hydrogen bonding, stacking free energy and the like.

In addition to the primary recognition event, it is also necessary to detect when binding takes place between the labelled polynucleotide sequence and the complementary sequence of the analyte. This detection is effected through a signalling step or event. A signalling step or event allows detection in some quantitative or qualitative manner, e.g., a human or instrument detection system, of the occurrence of the primary recognition event.

The primary recognition event and the signalling event of polynucleotide sequence based detection techniques may be coupled either directly or indirectly, proportionately or inversely proportionately. Thus, in such systems as nucleic acid hybridizations with sufficient quantities of radiolabeled probes, the amount of radio-activity is usually directly proportional to the amount of analyte present. Inversely proportional techniques include, for example, competitive immuno-assays, wherein the amount of detected signal decreases with the greater amount of analyte that is present in the sample.

Amplification techniques are also employed for enhancing detection wherein the signalling event is related to the

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