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New U.S. Patent Application
Title: RAPID DISSOLUTION FORMULATION OF
A CALCIUM RECEPTOR-ACTIVE COMPOUND
Inventors: Glen LAWRENCE, Francisco J. ALVAREZ, Hung-Ren H. LIN, and Tzuchi R. JU

Sir:

We enclose the following papers for filing in the United States Patent and Trademark Office in connection with the above patent application.

Application- 55 pages, including title page, 11 independent claims with 118 claims total, and Abstract.

Applicants claim the right to priority based on Provisional Patent Application No. 60/502,219 filed September 12, 2003.

This application will be completed in accordance with 37 C.F.R. § 1.53(f) upon receiving a Notice to File Missing Parts of Application.

Please accord this application an application number and filing date.

Respectfully submitted,

FINNEGAN, HENDERSON, FARABOW,
GARRETT & DUNNER, L.L.P.

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MDS/raa
Enclosures

UNITED STATES PATENT APPLICATION

FOR

RAPID DISSOLUTION FORMULATION OF A CALCIUM RECEPTOR-ACTIVE COMPOUND

BY

GLEN LAWRENCE, FRANCISCO J. ALVAREZ, HUNG-REN H. LIN, AND TZUCHI R. JU

[001] This application claims the benefit of priority of U.S. Provisional Patent Application No. 60/502,219, filed September 12, 2003.

[002] Calcium receptor-active compounds are known in the art. One example of a calcium receptor-active compound is cinacalcet HCl, which is described, for example, in U.S. Patent No. 6,001,884. Such calcium receptor-active compounds may be insoluble or sparingly soluble in water, particularly in their non-ionized state. For example, cinacalcet has a solubility in water of less than about 1 µg/mL at neutral pH. The solubility of cinacalcet can reach about 1.6 mg/mL when the pH ranges from about 3 to about 5. However, when the pH is about 1, the solubility decreases to about 0.1 mg/mL. Such limited solubility can reduce the number of formulation and delivery options available for these calcium receptor-active compounds. Limited water solubility can also result in low bioavailability of the compounds.

[003] There is therefore a need to maximize the dissolution of the calcium receptor-active compound from a dosage form, and potentially during *in vivo* exposure. There is also a need to improve the bioavailability of the calcium receptor-active compound during *in vivo* exposure.

[004] One aspect of the present invention provides a pharmaceutical composition comprising at least one calcium receptor active compound in combination with at least one pharmaceutically acceptable carrier. Certain embodiments of the present invention are directed to a pharmaceutical composition with a defined dissolution profile.

[005] The invention also provides a method of manufacturing the pharmaceutical composition to achieve the desired dissolution profile, as well as a

method of treating a disease using the pharmaceutical composition. In addition, certain embodiments of the present invention are directed to a method for controlling dissolution rate of a formulation comprising the pharmaceutical composition.

[006] According to one aspect of the invention, the invention provides a pharmaceutical composition comprising an effective dosage amount of at least one calcium receptor-active compound and at least one pharmaceutically acceptable excipient, wherein the composition has a dissolution profile in 0.05 N HCl, measured according to a dissolution test conducted in United States Pharmacopeia (USP) - National Formulary (NF) (USP 26/NF 21), chapter 711 using a USP 2 apparatus at a temperature of 37 °C ±0.5 °C, and at a rotation speed of 75 r.p.m., which comprises from about 50% to about 125% of a target amount of the calcium receptor-active compound being released from the composition no later than about 30 minutes from the start of the test.

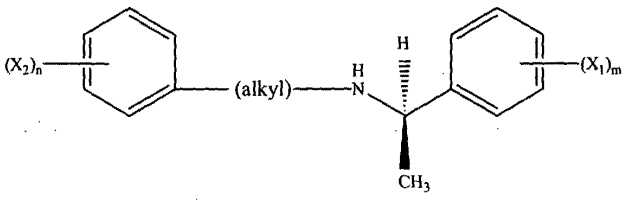
[007] According to another aspect of the invention, the invention provides a pharmaceutical composition comprising an effective dosage amount of at least one calcium receptor-active compound and at least one pharmaceutically acceptable excipient, wherein the composition has a dissolution profile in 0.05 N HCl, measured according to a dissolution test conducted in USP 26/NF 21, chapter 711 using a USP 2 apparatus at a temperature of about 37 °C, and at a rotation speed of about 75 r.p.m., which comprises from about 50% to about 125% of a target amount of the calcium receptor-active compound being released from the composition no later than about 30 minutes from the start of the test.

[008] The invention also provides a method of controlling the dissolution rate of a formulation comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient, the method comprising producing the formulation in a granulator which has a volume ranging from about 1 L to about 2000 L, and contains water in a granulation level ranging from about 10% to about 50% relative to the weight of the dry powders in the granulator.

[009] The calcium receptor-active compound useful in the claimed invention may be a calcimimetic compound or a calcilytic compound. As used herein, the term "calcimimetic compounds" refers to compounds that bind to a calcium receptor, and induce a conformational change that reduces the threshold for calcium receptor activation by the endogenous ligand Ca^{2+}, thereby reducing parathyroid hormone ("PTH") secretion. These calcimimetic compounds can also be considered allosteric modulators of the calcium receptor. As used herein, the term "calcilytic compounds" refers to compounds that act as calcium receptor antagonists, and stimulate PTH secretion.

[010] The calcimimetic compounds and calcilytic compounds useful in the present invention include those disclosed in, for example, European Patent No. 933 354; International Publication Nos. WO 01/34562, WO 93/04373, WO 94/18959, WO 95/11221, WO 96/12697, WO 97/41090; U.S. Patent Nos. 5,981,599, 6,001,884, 6,011,068, 6,031,003, 6,172,091, 6,211,244, 6,313,146, 6,342,532, 6,363,231, 6,432,656, and U.S. Patent Application Publication No. 2002/0107406. The calcimimetic compounds and/or calcilytic compounds disclosed in these patents and published applications are incorporated herein by reference.

[011] In certain embodiments, the calcium receptor-active compounds are chosen from compounds of formula (I) and pharmaceutically acceptable salts thereof



(I)

wherein:

X_1 and X_2, which may be identical or different, are each a radical chosen from CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, acetoxy, and acetyl radicals, or two of X_1 may together form an entity chosen from fused cycloaliphatic rings, fused aromatic rings, and a methylene dioxy radical, or two of X_2 may together form an entity chosen from fused cycloaliphatic rings, fused aromatic rings, and a methylene dioxy radical; provided that X_2 is not a 3-t-butyl radical;

n ranges from 0 to 5;

m ranges from 1 to 5; and

the alkyl radical is chosen from C1-C3 alkyl radicals, which are optionally substituted with at least one group chosen from saturated and unsaturated, linear, branched, and cyclic C1-C9 alkyl groups, dihydroindolyl and thiodihydroindolyl groups, and 2-, 3-, and 4-piperid(in)yl groups; and the stereoisomers thereof.

[012] Calcium receptor-active compounds useful in the present invention can be used in the form of pharmaceutically acceptable salts derived from inorganic or organic acids. The salts include, but are not limited to, the following: acetate, adipate,

alginate, citrate, aspartate, benzoate, benzenesulfonate, bisulfate, butyrate, camphorate, camphorsulfonate, digluconate, cyclopentanepropionate, dodecylsulfate, ethanesulfonate, glucoheptanoate, glycerophosphate, hemisulfate, heptanoate, hexanoate, fumarate, hydrochloride, hydrobromide, hydroiodide, 2-hydroxy-ethanesulfonate, lactate, maleate, mandelate, methansulfonate, nicotinate, 2-naphthalenesulfonate, oxalate, palmoate, pectinate, persulfate, 2-phenylpropionate, picrate, pivalate, propionate, salicylate, succinate, sulfate, tartrate, thiocyanate, tosylate, mesylate, and undecanoate. When compounds of the invention include an acidic function such as a carboxy group, then suitable pharmaceutically acceptable salts for the carboxy group are well known to those skilled in the art and include, for example, alkaline, alkaline earth, ammonium, quaternary ammonium cations and the like. For additional examples of "pharmacologically acceptable salts," see infra and Berge et al., J. Pharm. Sci. 66:1 (1977). In certain embodiments of the invention salts of hydrochloride and salts of methanesulfonic acid can be used.

[013] In some embodiments of the present invention, the calcium-receptor active compound can be chosen from cinacalcet, i.e., N-(1-(R)-(1-naphthyl)ethyl]-3-[3-(trifluoromethyl)phenyl]-1-aminopropane, cinacalcet HCl, and cinacalcet methanesulfonate. The cinacalcet HCl and cinacalcet methanesulfonate can be in various forms, such as amorphous powders, crystalline powders, and mixtures thereof. For example, the crystalline powders can be in forms including polymorphs, psuedopolymorphs, crystal habits, micromeretics, and particle morphology.

[014] The therapeutically effective amount of the calcium receptor-active compound in the compositions disclosed herein ranges from about 1 mg to about 360

mg, for example from about 5 mg to about 240 mg, or from about 20 mg to about 100 mg. As used herein, the "therapeutically effective amount" is an amount that changes in a desired manner at least one of the calcium level, the phosphorus level, the PTH level, and the calcium phosphorus product in a subject. In some embodiments, the therapeutically effective amount of cinacalcet HCl in the composition disclosed herein can be chosen from about 5 mg, about 15 mg, about 30 mg, about 50 mg, about 60 mg, about 75 mg, about 90 mg, about 120 mg, about 150 mg, about 180 mg, about 210 mg, about 240 mg, about 300 mg, or about 360 mg.

[015] While it may be possible to administer a compound of the invention alone, the compound administered will normally be present as an active ingredient in a pharmaceutical composition. Thus, a pharmaceutical composition of the invention may comprise a therapeutically effective amount of at least one calcium receptor-active compound, or an effective dosage amount of at least one calcium receptor-active compound.

[016] As used herein, an "effective dosage amount" is an amount that provides a therapeutically effective amount of the at least one calcium receptor active compound when provided as a single dose, in multiple doses, or as a partial dose. Thus, an effective dosage amount of the at least one calcium receptor active compound of the invention includes an amount less than, equal to or greater than an effective amount of the compound; for example, a pharmaceutical composition in which two or more unit dosages, such as in tablets, capsules and the like, are required to administer an effective amount of the compound, or alternatively, a multidose pharmaceutical composition, such as powders, liquids and the like, in which an effective amount of the

at least one calcium receptor-active compound is administered by administering a portion of the composition.

[017] Alternatively, a pharmaceutical composition in which two or more unit dosages, such as in tablets, capsules and the like, are required to administer an effective amount of the at least one calcium receptor active compound may be administered in less than an effective amount for one or more periods of time (i.e, a once-a-day administration, and a twice-a-day administration), for example to ascertain the effective dose for an individual subject, to desensitize an individual subject to potential side effects, to permit effective dosing readjustment or depletion of one or more other therapeutics administered to an individual subject, and/or the like.

[018] The effective dosage amount of the pharmaceutical composition disclosed herein ranges from about 1 mg to about 360 mg from a unit dosage form, for example about 5 mg, about 15 mg, about 30 mg, about 50 mg, about 60 mg, about 75 mg, about 90 mg, about 120 mg, about 150 mg, about 180 mg, about 210 mg, about 240 mg, about 300 mg, or about 360 mg from a unit dosage form.

[019] In some embodiments of the present invention, the compositions disclosed herein comprise a therapeutically effective amount of cinacalcet HCl for the treatment of hyperparathyroidism, such as primary hyperparathyroidism and secondary hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium-phosphorus product. For example, in certain embodiments, the cinacalcet HCl can be present in an amount ranging from about 1% to about 70%, such as from about 5% to about 40%, from about 10% to about 30%, or from about 15% to about 20%, by weight relative to the total weight of the composition.

[020] The compositions of the invention may contain one or more active ingredients in addition to the calcium receptor-active compound. The additional active ingredient may be another calcium receptor-active compound, or it may be an active ingredient having a different therapeutic activity. Examples of such additional active ingredients include, for example, vitamins and their analogs, such as vitamin D and analogs thereof, antibiotics, and cardiovascular agents.

[021] The cinacalcet HCl or other calcium receptor-active compound that can be used in the composition is typically present in the form of particles. These particles can have a particle D_{50} of, for example, less than or equal to about 50 μm. As used herein, the "particle D_{50}" is the particle size of the active pharmaceutical ingredient at the 50^{th} percentile of a particle size distribution. According to certain embodiments of the invention, the active pharmaceutical ingredient in the formulation has a particle D_{50} that is less than the granule D_{50} of the formulation, discussed in detail below.

[022] The particle D_{50} of the cinacalcet HCl particles can be determined by one of ordinary skill in the art using known light scattering techniques. In one embodiment of the invention, the particle D_{50} of the cinacalcet HCl particles is determined by using a particle size analyzer, such as a Malvern Mastersizer analyzer, that uses a laser to scan a suspension of particles. The particles diffract the incoming light to detectors: smaller particles diffract light at larger angles, while larger particles diffract light at smaller angles. The light intensities observed at each detector are translated into a particle size distribution based on the diameter of a sphere that has an equivalent volume to that of the measured particles.

[023] Specifically, the particle size distribution of the active pharmaceutical ingredient, for example, cinacalcet HCl, can be determined according to the following procedure. The following instrument conditions in a Malvern Mastersizer particle size analyzer are specified in its software:

Refractive Index Sample	1.630
Absorptive Index	0.1
Refractive Index Dispersant	1.375
Analysis model	General purpose spherical
Calculation sensitivity	Enhanced
Measurement snaps and time	20,000 snaps over 20 seconds
Background snaps and time	20,000 snaps over 20 seconds
Stir speed	1750 rpm

[024] While stirring, about 170 mL of a dispersion of about 0.1% sorbitan trioleate (for example Span 85®, available from Kishida Chemical) in hexane ("dispersant-B"), is added to the sampling unit, and the laser is aligned to take a background measurement of the dispersant-B.

[025] The entire suspension containing the cinacalcet HCl is added until a suitable obscuration range ranging from about 10 to about 20% is obtained. The sample is measured after the obscuration value has stabilized. After the measurement, the system is drained and rinsed once with about 170 mL of dispersant-B, the dispersant-B is drained, and the sampling unit is refilled with about 170 mL of dispersant-B. The measurement are repeated two more times with different riffled fractions. The riffling is performed on large samples to obtain small representative particle size fractions about 15 mg in size.

[026] The Obscuration, D(v,0.1), D(v,0.5), D(v,0.9) values are then calculated from these measurements. The average, standard deviation, and relative standard deviation (RSD) of the D(v,0.1), D(v,0.5), D(v,0.9) values is also calculated. The RSD (%) is calculated as follows:

$$RSD\ (\%) = \frac{100}{X}\left[\frac{\sum_{i=1}^{N}(X_i - \bar{X})^2}{N-1}\right]^{\frac{1}{2}}$$

where X, is an individual measurement in a set of N measurements and is the arithmetic mean of the set.

[027] The composition disclosed herein can be in various forms, for example, in granular form. The granules that can be used in the present invention can have a granule D_{50} ranging from about 50 μm to about 150 μm, such as from about 80 μm to about 130 μm. As defined herein, the "granule D_{50}" is the particle size of the composition at the 50th percentile of a particle size distribution. The granule D_{50} can readily be determined by one of ordinary skill in the art using sieve analysis techniques. Specifically, the granule D_{50} is determined according to the following procedure.

[028] Approximately 100 g of sample is added to sieve shaker equipped with 40 mesh, 60 mesh, 80 mesh, 100 mesh, 140 mesh, 200 mesh, 325 mesh, and the bottom pan. The sieve shaker is then turned on for about 10 minutes to separate the sample according to particle size. Each sieve is weighed to determine the amount of sample retained on each sieve and the bottom pan. The individual sieve weight is

normalized to generate sieve weight fraction. The individual sieve weight fraction is calculated by dividing each sieve weight with the sum of all sieve weights.

$$\text{Weight Fraction of each sieve} = \frac{\text{Weight of each sieve}}{\text{Sum of all sieves}}$$

[029] Before the particle size calculation, the mean size range must be determined for each sieve and the bottom pan. This mean size of each sieve screen represents the mean particle size retained on the screen. The mean size of each sieve screen is determined by the hole size of the screen (lower limit) and one sieve size larger (upper limit). In the case of the 40 mesh sieve screen, the hole size of about 1410 μm is used as an upper limit. Table 1 set forth below shows the particle size range of any retained material on each screen and the mean of the particle size range.

Table 1

Screens	Hole size of each screen (μm)	Particle size range of retained material on each screen (μm)	Median particle size of the screen (μm)
40 mesh	425	425 - 1410	918
60 mesh	250	250 - 424	337
80 mesh	180	180 - 249	215
100 mesh	150	150 - 179	165
140 mesh	106	106 – 149	128
200 mesh	75	75 – 105	90
325 mesh	45	45 – 74	60
Bottom pan	0	1 – 44	23

[030] The weight fraction of each sieve is added to generate cumulative frequency distribution starting from the bottom pan to 40 mesh screen. Once the cumulative frequency distribution is generated, the corresponding particle size at

10 percentile (D_{10}), 50-percentile (D_{50}), and 90-percentile (D_{90}) are determined. The particle size of the corresponding percentile is determined by linear interpolation between two consecutive data from the cumulative frequency distribution. For example, particle size of 50-percentile (D_{50}) is interpolated by,

$$D_{50}(\mu m) = \frac{[(50 - X_n) * d_{n+1} + (X_{n+1} - 50) * d_n]}{(X_{n+1} - X_n)}$$

where,

X_n = cumulative quantity of sample that is just below 50-percentile (in %);

d_n = mean of the particle size range from the sieve screen where X_n occurs (in mm);

X_{n+1} = next cumulative quantity of sample that is above 50-percentile (in %).

d_{n+1} = mean of the particle size range from the sieve screen where X_{n+1} occurs (in mm).

[031] According to all embodiments of the present invention, the particle size of active pharmaceutical ingredient is measured according to light scattering techniques, and the particle size of the granules of composition is measured according to sieve analysis.

[032] The compositions disclosed herein can be in a form chosen from, for example, tablets, capsules, and powders. The tablets can be made by pressing the granules into the form of tablets. The capsules can also be made using the granules.

[033] The at least one pharmaceutically acceptable excipient can be chosen from, for example, diluents such as starch, microcrystalline cellulose, dicalcium

phosphate, lactose, sorbitol, mannitol, sucrose, methyl dextrins; binders such as povidone, hydroxypropyl methylcellulose, dihydroxy propylcellulose, and sodium carboxyl methylcellulose; and disintegrants such as crospovidone, sodium starch glycolate, croscarmellose sodium, and mixtures of any of the foregoing. The at least one pharmaceutically acceptable excipient can further be chosen from lubricants such as magnesium stearate, calcium stearate, stearic acid, glyceryl behenate, hygrogenated vegetable oil, glycerine fumerate and glidants such as colloidal silicon dioxide, and mixtures thereof. In some embodiments of the present invention, the at least one pharmaceutically acceptable excipient is chosen from microcrystalline cellulose, starch, talc, povidone, crospovidone, magnesium stearate, colloidal silicon dioxide, sodium dodecyl sulfate, and mixtures of any of the foregoing. The excipients of the present invention, can be intragranular, intergranular, or mixtures thereof.

[034] In some embodiments of the present invention, the composition and/or the granules within the composition can comprise microcrystalline cellulose and starch in a weight ratio ranging from about 1:1 to about 15:1. For example, in the composition, the weight ratio of the microcrystalline cellulose and starch can range from about 1:1 to about 15:1, such as about 10:1, and in the granules within the composition, the weight ratio of the microcrystalline cellulose and starch can range from about 1:1 to about 10:1, such as about 5:1.

[035] The microcrystalline cellulose can be present in an amount ranging from about 25% to about 85%, for example from about 50% to about 80%, or from about 60% to about 75% by weight relative to the total weight of the composition. The starch can be present in an amount ranging from about 5% to about 35%, for example, from

about 5% to about 25%, or from about 5% to about 10% by weight relative to the total weight of the composition.

[036] The compositions disclosed herein can further comprise at least one ingredient chosen from coating materials that are known in the art such as, for example, hydroxypropyl methylcellulose.

[037] Certain compositions can comprise:

(a) from about 10% to about 40% by weight of a calcium receptor-active compound chosen from cinacalcet HCl and cinacalcet methanesulfonate;

(b) from about 45% to about 85% by weight of at least one diluent;

(c) from about 1% to about 5% by weight of at least one binder; and

(d) from about 1% to about 10% by weight of at least one disintegrant;

wherein the percentage by weight is relative to the total weight of the composition. The compositions can further comprise from about 0.05% to about 5% by weight, relative to the total weight of the composition, of at least one additive chosen from glidants, lubricants, and adherents. The composition can additionally comprise from about 1% to about 6% by weight of at least one coating material, relative to the total weight of the composition.

[038] In another embodiment, the composition disclosed herein comprises:

(a) from about 10% to about 40% by weight of cinacalcet HCl;

(b) from about 5% to about 10% by weight of starch;

(c) from about 40% to about 75% by weight of microcrystalline cellulose;

(d) from about 1% to about 5% by weight of povidone; and

(e) from about 1% to about 10% by weight of crospovidone;

wherein the percentage by weight is relative to the total weight of the composition.

[039] The povidone can be present in an amount ranging from about 1% to about 5%, for example, from about 1% to about 3% by weight relative to the total weight of the composition. The crospovidone can be present in an amount ranging from about 1% to about 10%, for example from about 3% to about 6%, by weight relative to the total weight of the composition.

[040] The composition can further comprise from about 0.05% to about 5% by weight, relative to the total weight of the composition, of at least one additive chosen from colloidal silicon dioxide, magnesium stearate, talc, and the like, and mixtures of any of the foregoing. In certain embodiments of the invention, the composition comprises from about 0.05% to about 1.5% of colloidal silicon dioxide, from about 0.05% to about 1.5% of magnesium stearate, from about 0.05% to about 1.5% of talc, or mixtures of any of the foregoing. The composition can even further comprise from about 1% to about 6% by weight of at least one coating material, relative to the total weight of the composition.

[041] As mentioned above, the compositions of certain embodiments of the present invention have a dissolution profile that results in about 50% to about 125% of a target amount of the calcium receptor-active compound being released from the composition no later that about 30 minutes from the start of a dissolution test that is conducted in 0.05 N HCl in a U.S.P. 2 apparatus at a temperature of 37°C ± 0.5°C at a rotation speed of 75 r.p.m. The dissolution test is conducted using a USP 2 apparatus, and according to the dissolution protocol described in USP 26/NF 21, chapter 711,

which is incorporated herein by reference. According to this embodiment using this dissolution protocol, a stated volume of the dissolution medium (±1%) is placed in the vessel of the USP 2 apparatus, the apparatus is assembled, the dissolution medium is equilibrated to 37°C ± 0.5°C, the thermometer is removed, the dosage form is placed in the vessel, and the amount of active pharmaceutical ingredient that is released as a function of time is measured.

[042] According to another embodiment of the invention, a stated volume of the dissolution medium is placed in the vessel of the USP 2 apparatus, the apparatus is assembled, the dissolution medium is equilibrated to about 37°C, the thermometer is removed, the dosage form is placed in the vessel, and the amount of active pharmaceutical ingredient that is released as a function of time is measured.

[043] The dissolution profile represents the percentage of the active pharmaceutical ingredient released based on a target amount of the active pharmaceutical ingredient in the formulation. As used herein "target amount" refers to the amount of active pharmaceutical ingredient in each formulation. In certain embodiments, the target amount refers to the label amount and/or label claim.

[044] USP 26/NF 21, chapter 905, defines a protocol used to determine the dosage-unit conformity according to the present invention, and this content uniformity protocol is incorporated herein by reference. According to this protocol, the content uniformity is determined by measuring the amount of active pharmaceutical ingredient in 10 dosage unit samples, and calculating whether the amount of active pharmaceutical ingredient in all the dosage unit samples falls within a range of 85% to 115% of the target amount. If one dosage unit sample is outside the range of 85% to 115% of the

target amount and no unit is outside a range of 75% to 125% of the target amount, or if the Relative Standard Deviation (RSD), which is the sample standard deviation expressed as a percentage of the mean, is not greater than 6%, then 20 additional dosage unit samples are tested. After treating at least 30 dosage units, the content uniformity requirement is met if not more than one dosage unit sample is outside the range of 85% to 115% of the target amount, and no unit is outside a range of 75% to 125% of the target amount, and the RSD of the at least 30 dosage units does not exceed 7.8%.

[045] In certain embodiments, the dissolution profile of the compositions disclosed herein can result in, for example, at least about 50%, at least about 70%, at least about 75%, or at least about 85%, of the target amount of the calcium receptor-active compound being released from the composition no later than about 30 minutes from the start of the test. In certain embodiments, the dissolution profile of the compositions disclosed herein can comprise at most about 125%, for example at most about 115%, at most about 110%, or at most about 100% of the target amount of the calcium receptor-active compound being released from the composition no later than about 30 minutes from the start of the test. In additional embodiments, the dissolution profile of the compositions disclosed herein can comprise from about 50% to about 125%, for example from about 70% to about 110%, of the target amount of the calcium receptor-active compound being released from the composition no later than about 30 minutes from the start of the test.

[046] Other embodiments of the present invention are directed to a method of making a pharmaceutical composition comprising:

(a) forming a granule comprising a calcium receptor-active compound and at least one pharmaceutically acceptable excipient as disclosed herein; and

(b) controlling the particle size of the granule such that from about 50% to about 125% of a target amount of calcium receptor-active compound is released from the composition no later than about 30 minutes from the start of a test in 0.05 N HCl according to a dissolution test conducted in a USP 2 apparatus at a temperature of 37 °C ±0.5 °C, and a rotation speed of 75 r.p.m.

[047] Further embodiments of the present invention are directed to a method of making a pharmaceutical composition comprising:

(b) forming a granule comprising a calcium receptor-active compound and at least one pharmaceutically acceptable excipient as disclosed herein; and

(b) controlling the particle size of the granule such that from about 50% to about 125% of a target amount of calcium receptor-active compound is released from the composition no later than about 30 minutes from the start of a test in 0.05 N HCl according to a dissolution test conducted in a USP 2 apparatus at a temperature of about 37 °C, and a rotation speed of about 75 r.p.m.

[048] The granule can be formed by any known process, such as high wet shear granulation, low wet shear granulation, fluid bed granulation, rotary granulation, extrusion-spheronization, dry granulation, roller compaction, and the like.

[049] The particle size of the granule of the composition can be controlled by various factors. In certain embodiments of the present invention, the particle size of the granule of the composition can be controlled by the amount of water added to the materials present in a granulator. For example, a desired particle size of the granule can be achieved when the granulator has a volume ranging from about 1 L to about 1200 L, such as from about 65 L to about 1200 L, or from about 300 L to about 800 L, and the amount of water added ranges from about 20% to about 40%, such as from about 30% to about 36%, relative to the amount of dry powders present in the granulator to form the granules.

[050] The granulator's impeller tip speed can also affect the particle size of the granules. In some embodiments, the impeller tip speed, measured in meters per second (m/s), can range from about 5 m/s to about 10 m/s, such as from about 7 m/s to about 9 m/s.

[051] Other embodiments of the present invention are directed to a method of making a pharmaceutical composition comprising

(a) forming a composition comprising a therapeutically effective amount of particles of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient as disclosed herein; and

(b) controlling the particle size of the calcium receptor-active compound such that from about 50% to about 125% of a target amount of the calcium receptor-active compound is released from the composition no later than about 30 minutes from the start of a test in 0.05 N HCl according to a dissolution test conducted

in a USP 2 apparatus at a temperature of 37 °C ±0.5 °C, and a rotation speed of 75 r.p.m.

[052] Additional embodiments of the present invention are directed to a method of making a pharmaceutical composition comprising

(a) forming a composition comprising a therapeutically effective amount of particles of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient as disclosed herein; and

(b) controlling the particle size of the calcium receptor-active compound such that from about 50% to about 125% of a target amount of the calcium receptor-active compound is released from the composition no later than about 30 minutes from the start of a test in 0.05 N HCl according to a dissolution test conducted in a USP 2 apparatus at a temperature of about 37 °C, and a rotation speed of about 75 r.p.m.

[053] The size of the particles is controlled during the production of the active pharmaceutical ingredient, for example, by use of a milling step, or a controlled crystallization process. For example, the active pharmaceutical ingredient can be milled using a stainless steel hammer mill with 5 mm screen and 12 hammers forward at a mill speed of 8100 ± 100 rpm, with the feed speed is set at 90 ± 10 rpm.

[054] Yet other embodiments of the present invention are directed to a method for the treatment of a disease or disorder that can be treated by altering a subject's calcium receptor activity. In some embodiments, a method for the treatment of a disease chosen from hyperparathyroidism, such as primary hyperparathyroidism and

secondary hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium-phosphorus product comprises administering to a patient, such as human, an effective dosage amount of a pharmaceutical composition comprising a calcium receptor-active compound and at least one pharmaceutically acceptable excipient as disclosed herein, wherein the composition has a dissolution profile in 0.05 N HCl, measured according to a dissolution test conducted in a USP 2 apparatus at a temperature of 37 °C ±0.5 °C, and at a rotation speed of 75 r.p.m., which comprises from about 50% to about 125% of a target amount of the calcium receptor-active compound being released from the composition in no later than about 30 minutes from the start of the test.

[055] A further embodiment of the present invention is directed to a method for the treatment of a disease chosen from hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium-phosphorus product comprises administering to a patient, such as human, an effective dosage amount of a pharmaceutical composition comprising a calcium receptor-active compound and at least one pharmaceutically acceptable excipient as disclosed herein, wherein the composition has a dissolution profile in 0.05 N HCl, measured according to a dissolution test conducted in a USP 2 apparatus at a temperature of about 37 °C, and at a rotation speed of about 75 r.p.m., which comprises from about 50% to about 125% of a target amount of the calcium receptor-active compound being released from the composition in no later than about 30 minutes from the start of the test.

[056] Reference will now be made to the following examples which are not intended to limit the invention. To the contrary, it will be appreciated that various alternatives, modifications, and equivalents may be included within the spirit and scope of the invention.

Examples

[057] Three pharmaceutical formulations with target amounts of 30mg, 60mg, and 90 mg active pharmaceutical ingredient with the following components were prepared:

	Weight % (w/w)	30 mg Tablet	60 mg Tablet	90 mg Tablet
		Amount (mg)	Amount (mg)	Amount (mg)
Cinacalcet HCl	18.367	33.06	66.12	99.18
Pregelatinized starch (Starch 1500)	33.378	60.08	120.16	180.24
Microcrystalline cellulose (Avicel PH102)	6.678	12.02	24.04	36.06
Povidone (Plasdone K29/32)	2.044	3.68	7.36	11.04
Crospovidone (Polyplasdone XL)	1.233	2.22	4.44	6.66
Purified Water[1]	----	----	----	----
Microcrystalline cellulose (Avicel PH102)	34.300	61.74	123.48	185.22
Magnesium stearate	0.500	0.90	1.80	2.70
Colloidal silicon dioxide (Colloidal anhydrous silica) (Cab-O-Sil M5P)	0.500	0.90	1.80	2.70
Crospovidone (Polyplasdone XL)	3.000	5.40	10.80	16.20
Core Tablet	**100.000**	**180.00**	**360.00**	**540.00**
Purified Water[1]	----	----	----	----
Opadry® II (colored film former)	4.000	7.20	14.40	21.60
Purified Water[1]	----	----	----	----
Opadry® Clear (clear film former)	1.500	2.70	5.40	8.10
Carnauba Wax Powder	0.010	0.018	0.036	0.054
Opacode® Ink (Black)[2]	----	----	----	----

[1] The purified Water was removed during processing.
[2] Trace quantities of ink were applied to the coated tablet.

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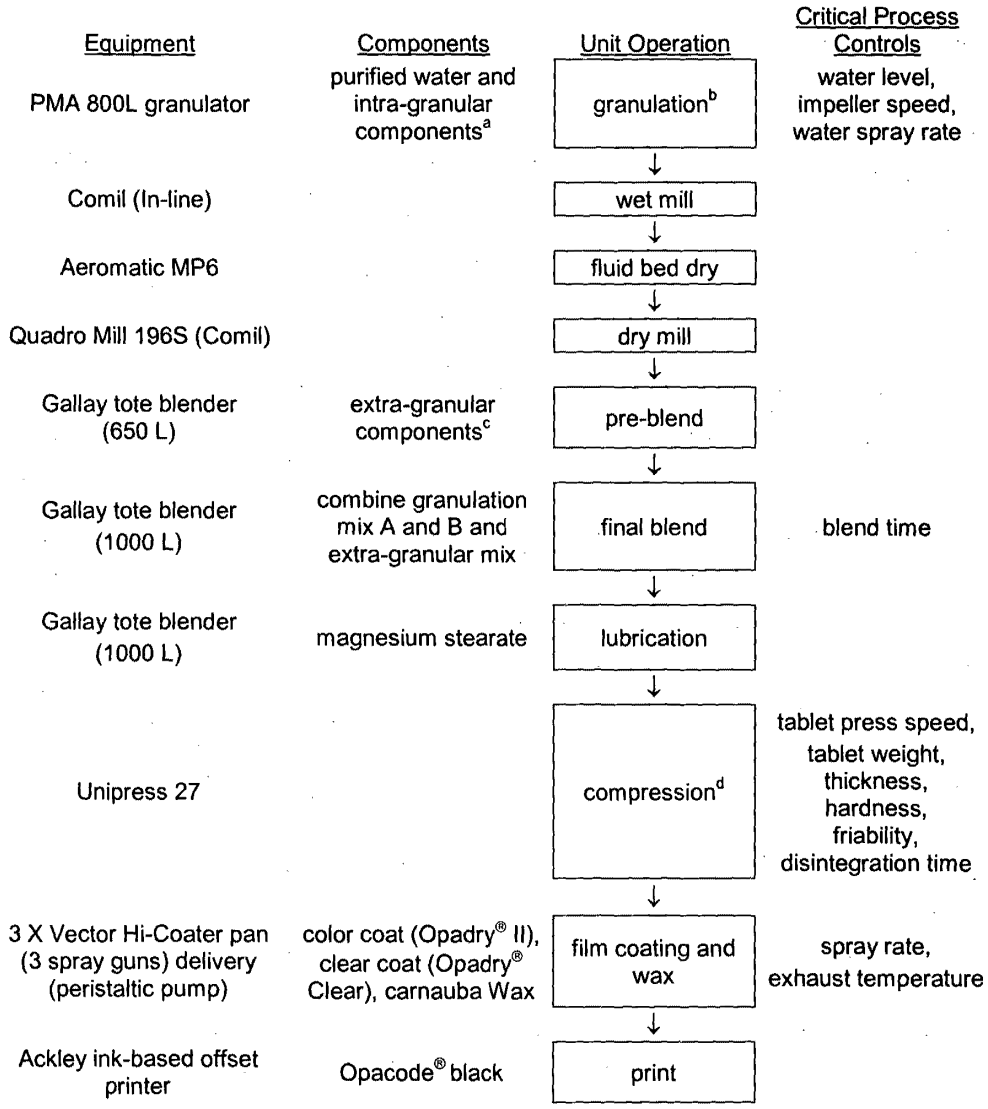
[058] The 30-, 60- and 90-mg tablets were made according to the process flow diagram depicted below.



[a] cinacalcet HCl, pregelatinized starch, microcrystalline cellulose, povidone, and crospovidone

[b] The granulation step to dry milling step is repeated to generate 2 bowls of wet granulation (Mix A and B).

[c] Extra-granular components are microcrystalline cellulose, crospovidone, and colloidal silicon dioxide

[d] Tooling dimension is dependent on tablet size and strength, (30 mg; 0.2372" x 0.3800" oval shape plain, 60 mg; 0.3000" x 0.4800" modified oval (double radius) plain, 90 mg; 0.3420" x 0.5480" modified oval (double radius) plain)

[059] The wet granulation process was conducted in a PMA 800L high-shear granulator with water serving as the granulation fluid. The cinacalcet HCl and the intra-granulation excipients (pregelatinized starch, microcrystalline cellulose, povidone, and crospovidone) were dry-mixed for 1 to 2 minutes with an impeller speed set point at 116 ± 10 rpm, followed by granulation with 30.0% to 36.0% w/w water (based on intra-granular lot size; target was 34.9% w/w) with an impeller speed set point at 116 ± 10 rpm and at a slow or fast chopper speed (target was slow speed). During the granulation process water was delivered at 9.8 ± 0.5 kg/min.

[060] Following granulation, the mixture was wet-milled using an in-line Comil equipped with a 0.375" (0.953 cm) opening screen and an impeller speed set point at 1400 ± 50 rpm. The mixture was then discharged into a fluid-bed dryer.

[061] After completion of the wet-milling process, the granulation mixture was dried in an Aeromatic MP6 fluid bed dryer with an inlet temperature set point at 70° ± 5°C. When the outlet temperature reached 37°C to 41°C, samples were taken to determine moisture levels by loss on drying (LOD). The granules were dried until the average moisture levels reached 1.0% to 2.5%.

[062] The dried granulation mixture was milled through a Quadro Mill 196S (Comil) equipped with a 0.055" (0.140 cm) opening screen at an impeller speed of 1650 ± 50 rpm into a 1000L Gallay tote.

[063] Except for magnesium stearate, the extra-granular excipients were blended in a 650 L Gallay tote blender for 7 ± 1 minutes at 12 ± 1 rpm. This mixture was further blended with the dry-milled granulation in a 1000 L Gallay tote blender for

15 ± 5 minutes at 12 ± 1 rpm, and then for 6 ± 1 minutes at 12 ± 1 rpm after magnesium stearate was added for lubrication.

[064] The final lubricated blend was compressed into tablets containing 30-, 60-, or 90 mg of the free base equivalent of active cinacalcet HCl using a Unipress 27 tablet press set to a speed of 2000 ± 300 tablets per minute and equipped with a force feeder. Throughout the compression operation, individual tablet weights (target weights of 180, 360, and 540 mg for 30-, 60-, and 90-mg tablets, respectively), the average weight of 10 tablets, tablet hardness and thickness were monitored at pre-determined intervals.

[065] The color-coating suspension and clear-coating solution were prepared by slowly adding either the Opadry® II (green) or Opadry® Clear into purified water while mixing until uniform (≥ 45 minutes). The color suspension and clear solution deaerated for ≥ 45 minutes before the spraying process began, and were used within a pre-determined time limit.

[066] Each lot was film-coated with color and clear coats in a Vector Hi-Coater 48" pan. The color-coating suspension was applied onto a moving core tablet bed (pan speed = 4 to 7 rpm) and a spray rate of 250 ± 50 grams per minute per 3 guns. The distance between the spray guns and the tablet bed was approximately 8" (20 cm) to 11" (28 cm), and the air volume was 600 ± 200 ft^3 per minute (17.1 ± 5.7 m^3 per minute) with a pan pressure differential maintained between -0.1" (-0.25 cm) to -0.3" (-0.76 cm) of water. Supply air temperature was adjusted to 80 ± 10°C to maintain an exhaust temperature of 41 ± 3°C.

[067] When the clear-coating application was completed, the heater and the air supply was turned off and the wax was spread evenly over the moving tablet bed (after it reached ≤ 37°C) with a pan speed of 4 to 7 rpm. The tablets were rotated for 5 ± 1 minutes, and after the supply air and exhaust fan were turned on, the tablets were rotated for an additional 5 ± 1 minutes with a pan speed of 4 to 7 rpm and supply air of 600 ± 200 ft^3 per minute (17.1 ± 5.7 m^3 per minute). The pan was jogged until the tablet bed temperature reached ≤ 30°C.

[068] An Ackley ink-based offset printer was used to produce 2-sided printed tablets.

[069] The dissolution profile of the three formulations were measured according the dissolution protocol described in the USP 26/NF 21, chapter 711 using a USP 2 apparatus at a temperature of about 37 °C, and at a rotation speed of about 75 r.p.m. The dissolution profile of the formulations in which at least about 75% of the cinacalcet HCl was released from the composition in no later than about 30 minutes from the start of the test is set forth in Table 2.

Table 2

Time (min)	30 mg Tablet	60 mg Tablet	90 mg Tablet
15	85.3	81.9	80.8
30	95.2	93.8	93.4
45	97.7	97.7	97.9
60	98.7	98.8	99.8

[070] The content uniformity of the three formulations were measured in accordance with USP 26/NF 21, chapter 905, described in detail above. The content uniformity and for each of the three formulations is set forth in Table 3.

Table 3

Container	30 mg Tablet		60 mg Tablet		90 mg Tablet	
	Mean (10 tablets)	% RSD	Mean (10 tablets)	% RSD	Mean (10 tablets)	% RSD
1 (beg.)	98.5	0.8	96.7	1.6	99.7	1.2
5	98.8	0.8	98.5	0.8	100.7	0.9
11	98.5	0.6	98.3	1.0	99.9	0.7
16	98.3	0.8	97.6	1.3	99.9	0.5
22	98.3	1.0	96.3	1.8	100.7	0.9
end	98.0	0.6	95.8	1.9	99.3	0.8

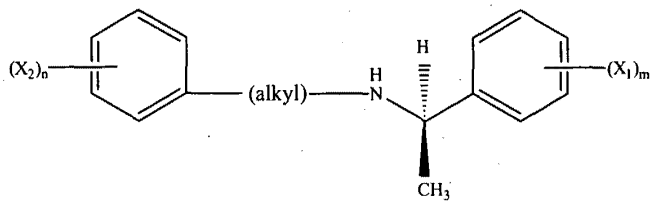
WHAT IS CLAIMED IS:

1. A pharmaceutical composition comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient,

wherein at least one dosage unit of the composition has a dissolution profile in 0.05 N HCl, measured according to a dissolution test conducted in a USP 2 apparatus at a temperature of about 37 °C, and at a rotation speed of about 75 r.p.m., which comprises from about 50% to about 125% of a target amount of the calcium receptor-active compound being released from the composition no later than about 30 minutes from the start of the test.

2. The composition according to Claim 1, wherein the calcium receptor-active compound is chosen from calcimimetic compounds and calcilytic compounds.

3. The composition according to Claim 2, wherein the calcimimetic compounds and calcilytic compounds are chosen from compounds of formula (I) and pharmaceutically acceptable salts and forms thereof



(I)

wherein:

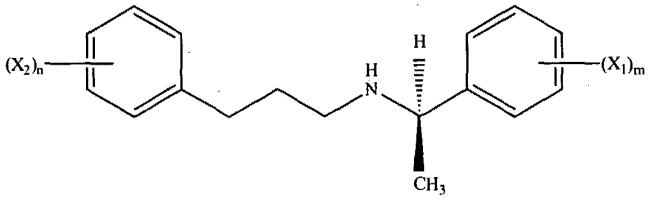
X_1 and X_2, which may be identical or different, are each a radical chosen from CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, acetoxy, and acetyl radicals, or two of X_1 may together form an entity chosen from fused cycloaliphatic rings, fused aromatic rings, and a methylene dioxy radical, or two of X_2 may together form an entity chosen from fused cycloaliphatic rings, fused aromatic rings, and a methylene dioxy radical; provided that X_2 is not a 3-t-butyl radical;

n ranges from 0 to 5;

m ranges from 1 to 5; and

the alkyl radical is chosen from C1-C3 alkyl radicals, which are optionally substituted with at least one group chosen from saturated and unsaturated, linear, branched, and cyclic C1-C9 alkyl groups, dihydroindolyl and thiodihydroindolyl groups, and 2-, 3-, and 4-piperid(in)yl groups; and the stereoisomers thereof.

4. The composition according to Claim 3, wherein the calcimimetic compounds and calcilytic compounds are chosen from compounds of formula (II) and pharmaceutically acceptable salts and forms thereof



(II)

wherein:

X_1 and X_2, which may be identical or different, are each a radical chosen from CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, acetoxy, and acetyl radicals, or two of X_1 may together form an entity chosen from fused cycloaliphatic rings, fused aromatic rings, and a methylene dioxy radical, or two of X_2 may together form an entity chosen from fused cycloaliphatic rings, fused aromatic rings, and a methylene dioxy radical; provided that X_2 is not a 3-t-butyl radical;

n ranges from 0 to 5; and

m ranges from 1 to 5.

5. The composition according to Claim 4, wherein the pharmaceutically acceptable salts and forms thereof are chosen from salts of hydrochloric acid and salts of methanesulfonic acid.

6. The composition according to Claim 4, wherein the calcimimetic compounds are chosen from cinacalcet, cinacalcet HCl, and cinacalcet methanesulfonate.

7. The composition according to Claim 1, wherein the dissolution profile comprises from about 70% to about 110% of the target amount of the calcium receptor-active compound being released from the composition no later than about 30 minutes from the start of the test.

8. The composition according to Claim 7, wherein the dissolution profile comprises at least about 75% of the target amount of the calcium receptor-active compound being released from the composition no later than about 30 minutes from the start of the test.

9. The composition according to Claim 6, wherein the dissolution profile comprises from about 70% to about 110% of the target amount of the cinacalcet HCl being released from the composition no later than about 30 minutes from the start of the test.

10. The composition according to Claim 9, wherein the dissolution profile comprises at least about 75% of the target amount of the cinacalcet HCl being released from the composition no later than about 30 minutes from the start of the test.

11. The composition according to Claim 6, wherein the cinacalcet HCl and cinacalcet methanesulfonate are in a form chosen from amorphous powders, crystalline particles and mixtures thereof.

12. The composition according to Claim 1, wherein the calcium receptor-active compound is cinacalcet HCl.

13. The composition according to Claim 12, wherein the cinacalcet HCl is in a form chosen from needle-shape particles, rod-shape particles, plate-shaped particles, and mixtures of any of the foregoing.

14. The composition according to Claim 12, wherein the particle D_{50} of the cinacalcet HCl particles is less than or equal to about 50 µm.

15. The composition according to Claim 12, wherein the cinacalcet HCl particles have a particle D_{50} effective to release from about 70% to about 110% of the target amount of the cinacalcet HCl from the composition no later than about 30 minutes from the start of the test in 0.05 N HCl.

16. The composition according to Claim 15, wherein the cinacalcet HCl particles have a particle D_{50} effective to release at least about 75% of the target amount of the cinacalcet HCl from the composition no later than about 30 minutes from the start of the test in 0.05 N HCl.

17. The composition according to Claim 1, wherein the composition is in the form of granules.

18. The composition according to Claim 1, wherein the composition is in a form chosen from tablets, capsules, and powders.

19. The composition according to Claim 17, wherein the granules have a granule D_{50} measured using a sieve analysis ranging from about 50 µm to about 150 µm.

20. The composition according to Claim 19, wherein the granules have a granule D_{50} measured using a sieve analysis ranging from about 80 µm to about 130 µm.

21. The composition according to Claim 17, wherein the granules have a granule D_{50} effective to release from about 70% to about 110% of the target amount of the calcium-receptor active compound from the composition no later than about 30 minutes from the start of the test in 0.05 N HCl.

22. The composition according to Claim 21, wherein the granules have a granule D_{50} effective to release at least about 75% of the target amount of the calcium-receptor active compound from the composition no later than about 30 minutes from the start of the test in 0.05 N HCl.

23. The composition according to Claim 12, wherein the cinacalcet HCl is present in a therapeutically effective amount for the treatment of at least one of hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium phosphorus product.

24. The composition according to Claim 12, wherein the cinacalcet HCl is present in an effective dosage amount for the treatment of at least one of hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium phosphorus product.

25. The composition according to Claim 23, wherein the hyperparathyroidism is chosen from primary hyperparathyroidism and secondary hyperparathyroidism.

26. The composition according to Claim 24, wherein the hyperparathyroidism is chosen from primary hyperparathyroidism and secondary hyperparathyroidism.

27. The composition according to Claim 12, wherein the cinacalcet HCl is present in an amount ranging from about 1% to about 70% by weight relative to the total weight of the composition.

28. The composition according to Claim 27, wherein the cinacalcet HCl is present in an amount ranging from about 5% to about 40% by weight relative to the total weight of the composition.

29. The composition according to Claim 28, wherein the cinacalcet HCl is present in an amount ranging from about 15% to about 20% by weight relative to the total weight of the composition.

30. The composition according to Claim 1, wherein the at least one pharmaceutically acceptable excipient is chosen from non-cellulose and cellulose diluents, binders, and disintegrants.

31. The composition according to Claim 1, wherein the at least one pharmaceutically acceptable excipient is chosen from microcystalline cellulose, starch,

talc, povidone, crospovidone, magnesium stearate, colloidal silicon dioxide, and sodium dodecyl sulfate and any combination thereof.

32. The composition according to claim 31, wherein crospovidone is present intergranularly, intragranularly, or a combination thereof.

33. The composition according to Claim 31, wherein crospovidone is present intergranularly.

34. The composition according to Claim 31, wherein crospovidone is present intragranularly.

35. The composition according to Claim 1, wherein the composition comprises microcystalline cellulose and starch in a weight ratio ranging from about 1:1 to about 15:1.

36. The composition according to Claim 35, wherein the composition comprises microcystalline cellulose and starch in a weight ratio of about 10:1.

37. The composition according to Claim 1, wherein the granules within the composition comprises microcystalline cellulose and starch in a weight ratio ranging from about 1:1 to about 10:1.

38. The composition according to Claim 37, wherein the weight ratio between the microcystalline cellulose and the starch in the granules with the composition is about 5:1.

39. The composition according to Claim 31, wherein the microcystalline cellulose is present in an amount ranging from about 25% to about 85% by weight relative to the total weight of the composition.

40. The composition according to Claim 31, wherein the starch is present in an amount ranging from about 5% to about 35% by weight relative to the total weight of the composition.

41. The composition according to Claim 31, wherein the povidone is present in an amount ranging from about 1% to about 5% by weight relative to the total weight of the composition.

42. The composition according to Claim 31, wherein the crospovidone is present in an amount ranging from about 1% to about 10% by weight relative to the total weight of the composition.

43. The composition according to Claim 1 comprising:

(a) from about 10% to about 40% by weight of cinacalcet HCl or cinacalcet methanesulfonate;

(b) from about 45% to about 85% by weight of at least one diluent;

(c) from about 1% to about 5% by weight of at least one binder;

(d) from about 1% to about 10% by weight of at least one disintegrant; and

(e) from about 0.05% to about 5% of at least one additive chosen from glidants, lubricants, and adherents;

wherein the percentage by weight is relative to the total weight of the composition.

44. The composition according to Claim 43 comprising from about 0.05% to about 1.5% by weight of at least one glidant relative to the total weight of the composition.

45. The composition according to Claim 43 comprising from about 0.05% to about 1.5% by weight of adherent relative to the total weight of the composition.

46. The composition according to Claim 43, further comprising at least one ingredient chosen from lubricants and clear and color coating materials.

47. The composition according to Claim 43 further comprising from about 1% to about 6% by weight of at least one coating material chosen from clear and color coating materials relative to the total weight of the composition.

48. The composition according to Claim 43 comprising

(a) from about 10% to about 40% by weight of cinacalcet HCl;

(b) from about 5% to about 10% by weight of starch;

(c) from about 40% to about 75% by weight of microcrystalline cellulose;

(d) from about 1% to about 5% by weight of povidone; and

(e) from about 1% to about 10% by weight of crospovidone;

wherein the percentage by weight is relative to the total weight of the composition.

49. The composition according to Claim 48 further comprising from about 0.05% to about 1.5% by weight of colloidal silicon dioxide relative to the total weight of the composition.

50. The composition according to Claim 48 further comprising from about 0.05% to about 1.5% by weight of magnesium stearate relative to the total weight of the composition.

51. The composition according to Claim 48 further comprising from about 1% to about 6% by weight of at least one coating material chosen from clear and color coating materials relative to the total weight of the composition.

52. The composition according to Claim 12, wherein the effective dosage amount of cinacalcet HCl ranges from about 1 mg to about 360 mg.

53. The composition according to Claim 52, wherein the effective dosage amount of cinacalcet HCl ranges from about 5 mg to about 240 mg.

54. The composition according to Claim 52,wherein the effective dosage amount of cinacalcet HCl ranges from about 20 mg to about 100 mg.

55. The composition according to claim 52, wherein the effective dosage amount of cinacalcet HCl is chosen from about 5 mg, about 15, mg, about 30 mg, about 50 mg, about 60 mg, about 75 mg, about 90 mg, about 120 mg, about 150 mg, about 180 mg, about 210 mg, about 240 mg, about 300 mg, and about 360 mg.

56. The composition according to Claim 12, wherein the therapeutically effective amount of cinacalcet HCl ranges from about 1 mg to about 360 mg.

57. The composition according to Claim 56, wherein the therapeutically effective amount of cinacalcet HCl ranges from about 5 mg to about 240 mg.

58. The composition according to Claim 56, wherein the therapeutically effective amount of cinacalcet HCl ranges from 20 mg to 100 mg.

59. The composition according to claim 56, wherein the therapeutically effective amount of cinacalcet HCl is chosen from about 5 mg, about 15, mg, about 30 mg, about 50 mg, about 60 mg, about 75 mg, about 90 mg, about 120 mg, about 150 mg, about 180 mg, about 210 mg, about 240 mg, about 300 mg, and about 360 mg.

60. A pharmaceutical composition comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient,

wherein at least one dosage unit of the composition has a dissolution profile in 0.05 N HCl, measured according to a dissolution test conducted in a USP 2 apparatus at a temperature of 37 °C ±0.5 °C, and at a rotation speed of 75 r.p.m., which comprises from 50% to 125% of a target amount of the calcium receptor-active compound being released from the composition no later than 30 minutes from the start of the test.

61. A method of making a pharmaceutical composition comprising:

(a) forming a granule comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient; and

(b) controlling the particle size of the granule such that from about 50% to about 125% of a target amount of the calcium receptor-active compound is released

from the composition no later than about 30 minutes from the start of a test in 0.05 N HCl according to a dissolution test conducted in a USP 2 apparatus at a temperature of about 37 °C, and a rotation speed of about 75 r.p.m.

62. A method of making a pharmaceutical composition comprising:

(a) forming a granule comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient; and

(b) controlling the particle size of the granule such that from about 50% to about 125% of a target amount of the calcium receptor-active compound is released from the composition no later than about 30 minutes from the start of a test in 0.05 N HCl according to a dissolution test conducted in a USP 2 apparatus at a temperature of 37 °C ±0.5 °C, and a rotation speed of 75 r.p.m.

63. A method of making a pharmaceutical composition comprising:

(a) forming a composition comprising an effective dosage amount of particles of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient; and

(b) controlling the particle size of the calcium receptor-active compound such that from about 50% to about 125% of a target amount of the calcium receptor-active compound is released from the composition no later than about 30 minutes from the start of a test in 0.05 N HCl according to a dissolution test conducted in a USP 2 apparatus at a temperature of about 37 °C, and a rotation speed of about 75 r.p.m.

64. A method of making a pharmaceutical composition comprising:

(a) forming a composition comprising an effective dosage amount of particles of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient; and

(b) controlling the particle size of the calcium receptor-active compound such that from about 50% to about 125% of a target amount of the calcium receptor-active compound is released from the composition no later than about 30 minutes from the start of a test in 0.05 N HCl according to a dissolution test conducted in a USP 2 apparatus at a temperature of 37 °C ±(0.5) °C, and a rotation speed of 75 r.p.m.

65. A method of making a pharmaceutical composition comprising forming a granule comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient in a granulator,

wherein the granulator has a volume ranging from about 1 L to about 2000 L, and

wherein the granulator contains water in a granulation level ranging from about 10% to about 50% relative to the weight of the dry powders in the granulator.

66. The method according to Claim 65, wherein the granulator has a volume ranging from about 65 L to about 1200 L.

67. The method according to Claim 65, wherein the granulator has a volume ranging from about 300 L to about 800 L.

68. The method according to Claim 65, wherein the water is in a granulation level ranging from about 20% to about 40% relative to the weight of the dry powders in the granulator.

69. The method according to Claim 65, wherein the water is in a granulation level ranging from about 30% to about 36% relative to the weight of the dry powders in the granulator.

70. The method according to Claim 65, wherein the granulator has a impeller, whose tip speed ranges from about 5 m/s to about 10 m/s.

71. The method according to Claim 70, wherein the impeller tip speed ranges from about 7 m/s to about 9 m/s.

72. A method for the treatment of at least one disease chosen from hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium phosphorus product, comprising administering to a patient in need thereof a pharmaceutical composition comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient,

wherein the composition has a dissolution profile in 0.05 N HCl, measured according to a dissolution test conducted in a USP 2 apparatus at a temperature of about 37 °C, and at a rotation speed of about 75 r.p.m., which comprises from about 50% to about 125% of a target amount of the calcium receptor-active compound being released from the composition in no later than about 30 minutes from the start of the test.

73. The method according to Claim 72, wherein the patient is human.

74. The method according to Claim 72, wherein an effective dosage amount of the pharmaceutical composition is chosen from about 5 mg, about 15 mg, about 30 mg, about 50 mg, about 60 mg, about 75 mg, about 90 mg, about 120 mg, about 150 mg, about 180 mg, about 210 mg, about 240 mg, about 300 mg, and about 360 mg.

75. A method for the treatment of at least one disease chosen from hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium phosphorus product, comprising administering to a patient in need thereof a pharmaceutical composition comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient,

wherein the composition has a dissolution profile in 0.05 N HCl, measured according to a dissolution test conducted in a USP 2 apparatus at a temperature of 37 °C ±0.5 °C, and at a rotation speed of 75 r.p.m., which comprises from about 50% to

about 125% of a target amount of the calcium receptor-active compound being released from the composition in no later than about 30 minutes from the start of the test.

76. The method according to Claim 75, wherein the patient is human.

77. The method according to Claim 75, wherein an effective dosage amount of the pharmaceutical composition is chosen from about 5 mg, about 15 mg, about 30 mg, about 50 mg, about 60 mg, about 75 mg, about 90 mg, about 120 mg, about 150 mg, about 180 mg, about 210 mg, about 240 mg, about 300 mg, and about 360 mg.

78. A pharmaceutical composition comprising

(a) from about 10% to about 40% by weight of cinacalcet HCl;

(b) from about 45% to about 85% by weight of at least one diluent; and

(c) from about 1% to about 5% by weight of at least one binder;

wherein the percentage by weight is relative to the total weight of the composition.

79. The composition according to Claim 78 further comprising from about 1% to about 10% by weight of at least one disintegrant, wherein the percentage by weight is relative to the total weight of the composition.

80. The composition according to Claim 78 further comprising from about 0.05% to about 5% of at least one additive chosen from glidants, lubricants, and adherents,

wherein the percentage by weight is relative to the total weight of the composition.

81. The composition according to Claim 80 comprising from about 0.05% to about 1.5% by weight of at least one glidant.

82. The composition according to Claim 80 comprising from about 0.05% to about 1.5% by weight of adherent.

83. The composition according to Claim 78 further comprising at least one ingredient chosen from lubricants and clear and color coating materials.

84. The composition according to Claim 78 further comprising from about 1% to about 6% by weight of at least one coating material chosen from clear and color coating materials relative to the total weight of the composition.

85. The composition according to Claim 78, wherein the cinacalcet HCl is in a form chosen from amorphous powders, crystalline particles, matrix particles, and mixtures of any of the foregoing.

86. The composition according to Claim 78, wherein the cinacalcet HCl is in a form chosen from needle-shape particles, rod-shape particles, plate-shaped particles, and mixtures of any of the foregoing.

87. The composition according to Claim 78, wherein the particle D_{50} of the cinacalcet HCl particles is less than or equal to about 50 µm.

88. The composition according to Claim 78, wherein the composition is in the form of granules.

89. The composition according to Claim 78, wherein the composition is in a form chosen from tablets, capsules, and powders.

90. The composition according to Claim 88, wherein the granules have a granule D_{50} measured using a sieve analysis ranging from about 50 µm to about 150 µm.

91. The composition according to Claim 90, wherein the granules have a granule D_{50} measured using a sieve analysis ranging from about 80 µm to about 130 µm.

92. The composition according to Claim 78, wherein the cinacalcet HCl is present in a therapeutically effective amount for the treatment of at least one of hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium phosphorus product.

93. The composition according to Claim 78, wherein the cinacalcet HCl is present in an effective dosage amount for the treatment of at least one of

hyperparathyroidism, hyperphosphonia, hypercalcemia, and elevated calcium phosphorus product.

94. The composition according to Claim 92, wherein the hyperparathyroidism is chosen from primary hyperparathyroidism and secondary hyperparathyroidism.

95. The composition according to Claim 93, wherein the hyperparathyroidism is chosen from primary hyperparathyroidism and secondary hyperparathyroidism.

96. The composition according to Claim 78, wherein the cinacalcet HCl is present in an amount ranging from about 10% to about 30% by weight relative to the total weight of the composition.

97. The composition according to Claim 96, wherein the cinacalcet HCl is present in an amount ranging from about 15% to about 20% by weight relative to the total weight of the composition.

98. The composition according to Claim 78, wherein the at least one diluent is chosen from microcystalline cellulose, starch, and mixtures thereof.

99. The composition according to claim 98, wherein the microcrystalline cellulose is present in an amount ranging from about 40% to about 75% by weight, and the starch is present in an amount ranging from about 5% to about 10% by weight, relative to the total weight of the composition.

100. The composition according to Claim 78, wherein the at least one binder is povidone.

101. The composition according to Claim 100, wherein the povidone is present in an amount ranging from about 1% to about 5% by weight, relative to the total weight of the composition.

102. The composition according to Claim 78, wherein the at least one disintegrant is crospovidone.

103. The composition according to claim 102, wherein crospovidone is present intergranularly, intragranularly, or a combination thereof.

104. The composition according to Claim 102, wherein crospovidone is present intergranularly.

105. The composition according to Claim 102, wherein crospovidone is present intragranularly.

106. The composition according to Claim 98, wherein the composition comprises microcystalline cellulose and starch in a weight ratio ranging from about 1:1 to about 15:1.

107. The composition according to Claim 106, wherein the composition comprises microcystalline cellulose and starch in a weight ratio of about 10:1.

108. The composition according to Claim 98, wherein the granules within the composition comprises microcystalline cellulose and starch in a weight ratio ranging from about 1:1 to about 10:1.

109. The composition according to Claim 108, wherein the weight ratio between the microcystalline cellulose and the starch in the granules with the composition is about 5:1.

110. The composition according to Claim 78 comprising

(a) from about 10% to about 40% by weight of cinacalcet HCl;

(b) from about 5% to about 10% by weight of starch;

(c) from about 40% to about 75% by weight of microcrystalline cellulose;

(d) from about 1% to about 5% by weight of povidone; and

(e) from about 1% to about 10% by weight of crospovidone;

wherein the percentage by weight is relative to the total weight of the composition.

111. The composition according to Claim 110 further comprising from about 0.05% to about 1.5% by weight of colloidal silicon dioxide relative to the total weight of the composition.

112. The composition according to Claim 110 further comprising from about 0.05% to about 1.5% by weight of magnesium stearate relative to the total weight of the composition.

113. A method of controlling the dissolution rate of a formulation comprising an effective dosage amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient, the method comprising producing the formulation in a granulator which has a volume ranging from about 1 L to about 2000 L, and contains water in a granulation level ranging from about 10% to 50% relative to the amount of dry powders in the granulator.

114. The method according to Claim 113, wherein the calcium receptor-active compound is cinacalcet HCl.

115. The method according to Claim 113, wherein the granulator has a volume ranging from about 65 L to about 1200 L.

116. The method according to Claim 113, wherein the granulator has a volume ranging from about 300 L to about 800 L.

117. The method according to Claim 113, wherein the water is in a granulation level ranging from about 20% to about 40% relative to the weight of the dry powders in the granulator.

118. The method according to Claim 117, wherein the water is in a granulation level ranging from about 30% to about 36% relative to the weight of the dry powders in the granulator.

ABSTRACT OF THE DISCLOSURE

The present invention relates to a pharmaceutical composition comprising a therapeutically effective amount of a calcium receptor-active compound and at least one pharmaceutically acceptable excipient, wherein the composition has a controlled dissolution profile. The present invention further relates to a method of manufacturing the pharmaceutical composition, as well as a method of treating a disease using the pharmaceutical composition.

PATENT APPLICATION FEE DETERMINATION RECORD
Effective October 1, 2004

Application or Docket Number: 10937870

CLAIMS AS FILED - PART I

	(Column 1)	(Column 2)
TOTAL CLAIMS	118	
FOR	NUMBER FILED	NUMBER EXTRA
TOTAL CHARGEABLE CLAIMS	118 minus 20=	* 98
INDEPENDENT CLAIMS	11 minus 3 =	* 8
MULTIPLE DEPENDENT CLAIM PRESENT		☐

* If the difference in column 1 is less than zero, enter "0" in column 2

SMALL ENTITY TYPE ☐		OR	OTHER THAN SMALL ENTITY	
RATE	FEE		RATE	FEE
BASIC FEE	395.00	OR	BASIC FEE	790.00
X$ 9=		OR	X$18=	116
X44=		OR	X88=	704
+150=		OR	+300=	
TOTAL		OR	TOTAL	1610

CLAIMS AS AMENDED - PART II

AMENDMENT A

	(Column 1) CLAIMS REMAINING AFTER AMENDMENT		(Column 2) HIGHEST NUMBER PREVIOUSLY PAID FOR	(Column 3) PRESENT EXTRA
Total	*	Minus	**	=
Independent	*	Minus	***	=
FIRST PRESENTATION OF MULTIPLE DEPENDENT CLAIM				☐

SMALL ENTITY		OR	OTHER THAN SMALL ENTITY	
RATE	ADDITIONAL FEE		RATE	ADDITIONAL FEE
X$ 9=		OR	X$18=	
X44=		OR	X88=	
+150=		OR	+300=	
TOTAL ADDIT. FEE		OR	TOTAL ADDIT. FEE	

AMENDMENT B

	(Column 1) CLAIMS REMAINING AFTER AMENDMENT		(Column 2) HIGHEST NUMBER PREVIOUSLY PAID FOR	(Column 3) PRESENT EXTRA
Total	*	Minus	**	=
Independent	*	Minus	***	=
FIRST PRESENTATION OF MULTIPLE DEPENDENT CLAIM				☐

RATE	ADDITIONAL FEE		RATE	ADDITIONAL FEE
X$ 9=		OR	X$18=	
X44=		OR	X88=	
+150=		OR	+300=	
TOTAL ADDIT. FEE		OR	TOTAL ADDIT. FEE	

AMENDMENT C

	(Column 1) CLAIMS REMAINING AFTER AMENDMENT		(Column 2) HIGHEST NUMBER PREVIOUSLY PAID FOR	(Column 3) PRESENT EXTRA
Total	*	Minus	**	=
Independent	*	Minus	***	=
FIRST PRESENTATION OF MULTIPLE DEPENDENT CLAIM				☐

RATE	ADDITIONAL FEE		RATE	ADDITIONAL FEE
X$ 9=		OR	X$18=	
X44=		OR	X88=	
+150=		OR	+300=	
TOTAL ADDIT. FEE		OR	TOTAL ADDIT. FEE	

* If the entry in column 1 is less than the entry in column 2, write "0" in column 3.
** If the "Highest Number Previously Paid For" IN THIS SPACE is less than 20, enter "20."
***If the "Highest Number Previously Paid For" IN THIS SPACE is less than 3, enter "3".
The "Highest Number Previously Paid For" (Total or Independent) is the highest number found in the appropriate box in column 1.

FORM PTO-875 (Rev. 10/04) — Patent and Trademark Office, U.S. DEPARTMENT OF COMMERCE

PATENT APPLICATION SERIAL NO. _____

U.S. DEPARTMENT OF COMMERCE
PATENT AND TRADEMARK OFFICE
FEE RECORD SHEET

03/29/2005 MNGUYEN 00000010 010519 10937870

01 FC:1201 1600.00 DA
02 FC:1202 4900.00 DA

UNITED STATES PATENT AND TRADEMARK OFFICE

UNITED STATES DEPARTMENT OF COMMERCE
United States Patent and Trademark Office
Address: COMMISSIONER FOR PATENTS
P.O. Box 1450
Alexandria, Virginia 22313-1450
www.uspto.gov

APPLICATION NUMBER	FILING OR 371 (c) DATE	FIRST NAMED APPLICANT	ATTORNEY DOCKET NUMBER
10/937,870	09/10/2004	Glen Lawrence	06843.0057

CONFIRMATION NO. 1696

22852
FINNEGAN, HENDERSON, FARABOW, GARRETT & DUNNER LLP
1300 I STREET, NW
WASHINGTON, DC 20005

FORMALITIES LETTER

OC000000014263302

Date Mailed: 11/02/2004

NOTICE TO FILE MISSING PARTS OF NONPROVISIONAL APPLICATION

FILED UNDER 37 CFR 1.53(b)

Filing Date Granted

Items Required To Avoid Abandonment:

An application number and filing date have been accorded to this application. The item(s) indicated below, however, are missing. Applicant is given **TWO MONTHS** from the date of this Notice within which to file all required items and pay any fees required below to avoid abandonment. Extensions of time may be obtained by filing a petition accompanied by the extension fee under the provisions of 37 CFR 1.136(a).

- The statutory basic filing fee is missing.
  Applicant must submit $ **790** *to complete the basic filing fee for a non-small entity. If appropriate, applicant may make a written assertion of entitlement to small entity status and pay the small entity filing fee (37 CFR 1.27).*
- The oath or declaration is missing.
  A properly signed oath or declaration in compliance with 37 CFR 1.63, identifying the application by the above Application Number and Filing Date, is required.
- To avoid abandonment, a late filing fee or oath or declaration surcharge as set forth in 37 CFR 1.16(e) of $130 for a non-small entity, must be submitted with the missing items identified in this letter.

The applicant needs to satisfy supplemental fees problems indicated below.

The required item(s) identified below must be timely submitted to avoid abandonment:

• Additional claim fees of **$2468** as a non-small entity, including any required multiple dependent claim fee, are required. Applicant must submit the additional claim fees or cancel the additional claims for which fees are due.

SUMMARY OF FEES DUE:

Total additional fee(s) required for this application is **$3388** for a Large Entity

- **$790** Statutory basic filing fee.
- **$130** Late oath or declaration Surcharge.

- Total additional claim fee(s) for this application is **$2468**
  - **$704** for **8** independent claims over 3.
  - **$1764** for **98** total claims over 20.

Replies should be mailed to: Mail Stop Missing Parts
Commissioner for Patents
P.O. Box 1450
Alexandria VA 22313-1450

A copy of this notice MUST be returned with the reply.

Customer Service Center
Initial Patent Examination Division (703) 308-1202
PART 3 - OFFICE COPY

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s): LAWRENCE et al.

Serial No.: 10/937,870 Group Art Unit No.: Not Yet Assigned

Filed: September 10, 2004 Examiner:

For: RAPID DISSOLUTION FORMULATION OF A CALCIUM RECEPTOR-ACTIVE COMPOUND

Docket No.: A-870

TRANSMITTAL OF DECLARATION UNDER 37 CFR 1.51 (b)(2)
RESPONSE TO NOTICE TO FILE MISSING PARTS

Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

Sir:

This is in response to a "Notice to file Missing Parts of Application" dated November 2, 2004 noting the absence of a declaration from the papers filed with the above-identified application. Applicants hereby request a three (3) month extension of time for which to respond.

Enclosed herewith are: 1) a copy of the Notice and 2) a Declaration under 37 CFR 1.51(b)(2) by the applicant(s) in this application.

Please charge Deposit Account No. 01-0519 in the amount of $130.00 for the surcharge required by § 1.16(e), as well as $1,020.00 for a three-month extension of time. An original and one copy of this paper are enclosed.

The Commissioner is hereby authorized to charge any additional fees which may be required or credit any overpayment to Deposit Account No. 01-0519.

03/14/2005 JBALINAN 00000007 010519 10937870
03 FC:1253 1020.00 DA

Respectfully submitted,

MarySusan Howard
Attorney for Applicant
Registration No.: 38,729
Phone: (805) 447-0296
Date: 8 March 05

Please send all future correspondence to:

US Patent Operations/ Olga Mekhovich
AMGEN SF, LLC

EXPRESS MAIL CERTIFICATE

"Express Mail" label number: EL 732820859 US Date of Deposit: 8 March 2005

I hereby certify that this paper or fee is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 on the date indicated above and is addressed to the Mail Stop PCT, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450.

Printed Name Signature

3-10-05

IFW

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UNITED STATES PATENT AND TRADEMARK OFFICE

UNITED STATES DEPARTMENT OF COMMERCE
United States Patent and Trademark Office
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Alexandria, Virginia 22313-1450
www.uspto.gov

APPLICATION NUMBER	FILING OR 371 (c) DATE	FIRST NAMED APPLICANT	ATTORNEY DOCKET NUMBER
10/937,870	09/10/2004	Glen Lawrence	06843.0057

CONFIRMATION NO. 1696

22852
FINNEGAN, HENDERSON, FARABOW, GARRETT & DUNNER LLP
1300 I STREET, NW
WASHINGTON, DC 20005

FORMALITIES LETTER

OC0000000014263302

Date Mailed: 11/02/2004

NOTICE TO FILE MISSING PARTS OF NONPROVISIONAL APPLICATION

03/14/2005 JBALINAN 00000007 010519 10937870

01 FC:1011 ~~300.00 DA~~
02 FC:1051 130.00 DA

FILED UNDER 37 CFR 1.53(b)

Filing Date Granted

Items Required To Avoid Abandonment:

An application number and filing date have been accorded to this application. The item(s) indicated below, however, are missing. Applicant is given **TWO MONTHS** from the date of this Notice within which to file all required items and pay any fees required below to avoid abandonment. Extensions of time may be obtained by filing a petition accompanied by the extension fee under the provisions of 37 CFR 1.136(a).

- The statutory basic filing fee is missing.
  Applicant must submit $ **790** *to complete the basic filing fee for a non-small entity. If appropriate, applicant may make a written assertion of entitlement to small entity status and pay the small entity filing fee (37 CFR 1.27).*
- The oath or declaration is missing.
  A properly signed oath or declaration in compliance with 37 CFR 1.63, identifying the application by the above Application Number and Filing Date, is required.
- To avoid abandonment, a late filing fee or oath or declaration surcharge as set forth in 37 CFR 1.16(e) of $130 for a non-small entity, must be submitted with the missing items identified in this letter.

The applicant needs to satisfy supplemental fees problems indicated below.

The required item(s) identified below must be timely submitted to avoid abandonment:

- Additional claim fees of **$2468** as a non-small entity, including any required multiple dependent claim fee, are required. Applicant must submit the additional claim fees or cancel the additional claims for which fees are due.

SUMMARY OF FEES DUE:

Total additional fee(s) required for this application is **$3388** for a Large Entity

- **$790** Statutory basic filing fee.
- **$130** Late oath or declaration Surcharge.

03/15/2005 JBALINAN 00000126 010519 10937870

01 FC:1001 790.00 DA

Total additional claim fee(s) for this application is **$2468**

- **$704** for **8** independent claims over 3.
- **$1764** for **98** total claims over 20.

Replies should be mailed to: Mail Stop Missing Parts
Commissioner for Patents
P.O. Box 1450
Alexandria VA 22313-1450

A copy of this notice MUST be returned with the reply.

Customer Service Center
Initial Patent Examination Division (703) 308-1202
PART 2 - COPY TO BE RETURNED WITH RESPONSE

Docket No.: 06843.0057-00000

DECLARATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first, and sole inventor (if only one name is listed below) or a joint inventor (if plural names are listed below) of the invention entitled

RAPID DISSOLUTION FORMULATION OF A CALCIUM RECEPTOR-ACTIVE COMPOUND

which is described and claimed in the specification which:

☐ is attached hereto.

☒ was filed on September 10, 2004
as Application Serial No. 10/937,870
and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN APPLICATION(S)			
Country	Application Number	Filing Date (day, month, year)	Priority Claimed

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below.

PROVISIONAL APPLICATION(S)	
Application Number	Filing Date (day, month, year)
60/502,219	12, September, 2003

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below and, insofar as the subject matter of this application is not disclosed in

DECLARATION (cont'd)

the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the filing date of this application:

PRIOR U.S. APPLICATIONS		
APPLICATION SERIAL NO.	FILING DATE	STATUS

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Full Name of Sole or First Inventor:	Glen Gary LAWRENCE
Inventor's Signature:	Glen Gary Lawrence Date: 11/29/04
Residence and Post Office Address:	3439 Indian Ridge Circle, Thousand Oaks, CA 91326-4949 U.S.A. (Address, City, State, Zip Code, Country)
Citizenship:	US
Full Name of Second Joint Inventor, if Any:	Francisco J. ALVAREZ
Inventor's Signature:	F. Alvarez Date: NOV 29, 2004
Residence and Post Office Address:	817 Paseo de Leon, Newbury Park, CA 91320 U.S.A. (Address, City, State, Zip Code, Country)
Citizenship:	US
Full Name of Third Joint Inventor, if Any:	Hung-Ren H. LIN
Inventor's Signature:	Hung-Ren H. Lin Date: November 29, 2004
Residence and Post Office Address:	5086 Durant Court, Oak Park, CA 91377 U.S.A. (Address, City, State, Zip Code, Country)
Citizenship:	US

DECLARATION (cont'd)

Full Name of Fourth Joint Inventor, if Any:	Tzuchi R. JU
Inventor's Signature:	_____ Date: _____
Residence and Post Office Address:	1438 N. Butler Court, Vernon Hills, IL 60061 U.S.A. (Address, City, State, Zip Code, Country)
Citizenship:	US

Docket No.: 06843.0057-00000

DECLARATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first, and sole inventor (if only one name is listed below) or a joint inventor (if plural names are listed below) of the invention entitled

RAPID DISSOLUTION FORMULATION OF A CALCIUM RECEPTOR-ACTIVE COMPOUND

which is described and claimed in the specification which:

☐ is attached hereto.

☒ was filed on September 10, 2004
as Application Serial No. 10/937,870
and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN APPLICATION(S)			
Country	Application Number	Filing Date (day, month, year)	Priority Claimed

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below.

PROVISIONAL APPLICATION(S)	
Application Number	Filing Date (day, month, year)
60/502,219	12, September, 2003

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below and, insofar as the subject matter of this application is not disclosed in

DECLARATION (cont'd)

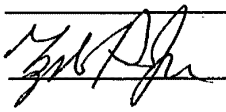
the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the filing date of this application:

PRIOR U.S. APPLICATIONS		
APPLICATION SERIAL NO.	FILING DATE	STATUS

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Full Name of Sole or First Inventor:	Glen Gary LAWRENCE
Inventor's Signature:	_____ Date: _____
Residence and Post Office Address:	3439 Indian Ridge Circle, Thousand Oaks, CA 91326-4949 U.S.A.
	(Address, City, State, Zip Code, Country)
Citizenship:	US
Full Name of Second Joint Inventor, if Any:	Francisco J. ALVAREZ
Inventor's Signature:	_____ Date: _____
Residence and Post Office Address:	817 Paseo de Leon, Newbury Park, CA 91320 U.S.A.
	(Address, City, State, Zip Code, Country)
Citizenship:	US
Full Name of Third Joint Inventor, if Any:	Hung-Ren H. LIN
Inventor's Signature:	_____ Date: _____
Residence and Post Office Address:	5086 Durant Court, Oak Park, CA 91377 U.S.A.
	(Address, City, State, Zip Code, Country)
Citizenship:	US

DECLARATION (cont'd)

Full Name of Fourth Joint Inventor, if Any:	Tzuchi R. JU
Inventor's Signature:	 Date: 12-6-04
Residence and Post Office Address:	1438 N. Butler Court, Vernon Hills, IL 60061 U.S.A. (Address, City, State, Zip Code, Country)
Citizenship:	US

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants:	LAWRENCE et al.	Docket No.:	A-870
Serial No:	10/937,870	Group Art Unit:	
Filed:	September 10, 2004	Examiner:	

For: RAPID DISSOLUTION FORMULATION OF A CALCIUM RECEPTOR-ACTIVE COMPOUND

STATEMENT UNDER 37 C.F.R. 3.73(b) AND POWER OF ATTORNEY BY ASSIGNEE

Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

Sir:

Amgen Inc. (hereinafter "Amgen"), a corporation of the State of Delaware having a place of business at One Amgen Center Drive, Thousand Oaks, California, 91320, is the assignee of the entire right, title, and interest in the above-identified U.S. patent application. Documentary evidence of a chain of title from the original owner(s) to Amgen is demonstrated by:

☒ The enclosed copy of an Assignment, the original of which is being filed for recordation by the United States Patent and Trademark Office.
OR
☐ The assignment recorded in the United States Patent and Trademark Office at Reel , Frame .

The undersigned, whose title is supplied below, is empowered to sign this statement on behalf of Amgen.

Amgen hereby appoints the registered practitioners at Customer Number **21069** to prosecute this application, to make and to transact all business in the Patent and Trademark Office connected therewith.

Please send all future correspondence and direct telephone calls to Customer Number **30174**.

Respectfully submitted,

Date March 8, 2005

By Stuart L. Watt
Stuart L. Watt
Title Vice President, Law

Enclosure: Copy of Assignment

EXPRESS MAIL CERTIFICATE

"Express Mail" label number: EL 732820859 US Date of Deposit: 8 MARCH 2005

I hereby certify that this paper or fee is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 on the date indicated above and is addressed to the Mail Stop PCT, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450.

Printed Name

Signature

Revised Form PTO-1595
(Rev. 6-93)

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PATENT & TRADEMARK OFFICE

RECORDATION FORM COVER SHEET
PATENTS ONLY

U.S. Department of Commerce
Patent and Trademark Office

Docket No.: A-870

To the Honorable Commissioner of Patents and Trademarks: Please record the attached original documents or copy thereof.

1. Name of conveying party(ies):
Glen Gary Lawrence
Francisco J. Alvarez
Hung-Ren H. Lin
Tzuchi R. Ju

Additional name(s) of conveying party(ies) attached? ☐ Yes ☒ No

2. Name and address of receiving party(ies):
Name: AMGEN INC.

Internal Address: Dept. 4300, MS 27-4-A

Street Address: One Amgen Center Drive

City: Thousand Oaks State: CA Zip: 91320-1799

Additional name(s) & address(es) attached? ☐ Yes ☒ No

3. Nature of conveyance:

☒ Assignment ☐ Merger
☐ Security Agreement ☐ Change of Name
☐ Other

Execution Date: December 6, 2004

4. Application number(s) or patent number(s):
If this document is being filed together with a new application, the execution date of the application is:

A. Patent Application No.(s)
10/937,870

B. Patent No.(s)

Additional numbers attached? ☐ Yes ☐ No

5. Name and address of party to whom correspondence concerning document should be mailed:

Name: U.S. Patent Operations/

Internal Address: Dept. 4300, M/S 27-4-A

AMGEN INC.

Street Address: One Amgen Center Drive

City: Thousand Oaks State: CA Zip: 91320-1799

6. Total number of applications and patents involved: 1

7. Total fee (37 CFR 3.41)..... $ 40.00

☐ Enclosed

☒ Authorized to be charged to deposit account.

8. Deposit account number: 01-0519

☒ The Commissioner is hereby authorized to charge any additional filing fees which may be required or credit any overpayment to Deposit Account No. 01-0519

DO NOT USE THIS SPACE

9. Statement and signature.
To the best of my knowledge and belief, the foregoing information is true and correct and any attached copy is a true copy of the original document.

MarySusan Howard
Name of Person Signing

Signature

8 March 05
Date

Total number of pages including cover sheet, attachments, and document: 5

SOLE/JOINT INVENTION
(Worldwide Rights)
Attorney Docket No. 6843.0057-00000

ASSIGNMENT

WHEREAS We, the below named inventors, [hereinafter referred to as Assignors], have made an invention entitled:

RAPID DISSOLUTION FORMULATION OF A CALCIUM RECEPTOR-ACTIVE COMPOUND

for which We executed an application for United States Letters Patent concurrently herewith or filed an application for United States Letters Patent on September 10, 2004, (Application No. 10/937,870); and

WHEREAS, **Amgen Inc.**, a corporation of Delaware, whose post office address is One Amgen Center Drive, Thousand Oaks, CA 91320-1799 (hereinafter referred to as Assignee), is desirous of securing the entire right, title, and interest in and to this invention in all countries throughout the world, and in and to the application for United States Letters Patent on this invention and the Letters Patent to be issued upon this application;

NOW THEREFORE, be it known that for good and valuable consideration the receipt of which from Assignee is hereby acknowledged, We, as Assignors, have sold, assigned, transferred, and set over, and do hereby sell, assign, transfer, and set over unto the Assignee, its lawful successors and assigns, our entire right, title, and interest in and to this invention, the U.S. Provisional Application No. 60/502,219, filed September 12, 2003, U.S. Application No. 10/937,870, and all divisions, and continuations thereof, and all Letters Patent of the United States which may be granted thereon, and all reissues thereof, and all rights to claim priority on the basis of the U.S. Provisional Application No. 60/502,219, filed September 12, 2003, and all rights to claim priority on the basis of U.S. Application No. 10/937,870, and all applications for Letters Patent which may hereafter be filed for this invention in any foreign country and all Letters Patent which may be granted on this invention in any foreign country, and all extensions, renewals, and reissues thereof; and We hereby authorize and request the Commissioner of Patents and Trademarks of the United States and any official of any foreign country whose duty it is to issue patents on applications as described above, to issue all Letters Patent for this invention to Assignee, its successors and assigns, in accordance with the terms of this Assignment;

AND, WE HEREBY covenant that We have the full right to convey the interest assigned by this Assignment, and We have not executed and will not execute any agreement in conflict with this Assignment;

AND, WE HEREBY further covenant and agree that We will, without further consideration, communicate with Assignee, its successors and assigns, any facts known to us respecting this invention, and testify in any legal proceeding, sign all lawful papers when called upon to do so, execute and deliver any and all papers that may be necessary or desirable to perfect the title to this invention in said Assignee, its successors or assigns, execute all divisional, continuation, and reissue applications, make all rightful oaths and generally do everything possible to aid Assignee, its successors and assigns, to obtain and enforce proper patent protection for this invention in the United States and any foreign country, it being understood that any expense incident to the execution of such papers shall be borne by the Assignee, its successors and assigns.

IN TESTIMONY WHEREOF, We have hereunto set our hands.

County of Ventura)
) ss.
State of California)

Name: Glen Gary LAWRENCE
Address: 3439 Indian Ridge Circle
Thousand Oaks, CA 91362-4949
By: Glen Gary Lawrence
Date 11/29/04

Subscribed and sworn to before me this 29 day of November 20 04
L. HOLTKE, Notary Public

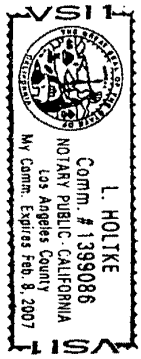
L. HOLTKE
Comm. # 1399086
NOTARY PUBLIC - CALIFORNIA
Los Angeles County
My Comm. Expires Feb. 8, 2007

SOLE/JOINT INVENTION
(Worldwide Rights)
Attorney Docket No. 6843.0057-00000

County of Ventura)
) ss.
State of California)

Name: Francisco J. ALVAREZ
Address: 817 Paseo de Leon
Newbury Park, CA 91320
By: F. Alvarez
Date: NOVEMBER 29, 2004

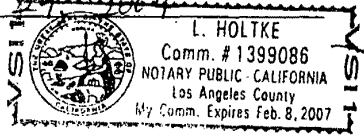
Subscribed and sworn to before me this 29 day of November, 2004
L. HOLTKE, Notary Public



County of Ventura)
) ss.
State of California)

Name: Hung-Ren H. LIN
Address: 5086 Durant Court
Oak Park, CA 91377
By: Hung-Ren H. Lin
Date: November 29, 2004

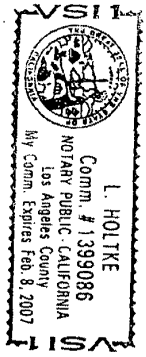
Subscribed and sworn to before me this 29 day of November, 2004
L. Holtke, Notary Public



County of _____)
) ss.
State of _____)

Name: Tzuchi R. JU
Address: 1438 N. Butler Court
Vernon Hills, IL 60061
By: _____
Date: _____

Subscribed and sworn to before me this _____ day of _____, 20____
_____, Notary Public



SOLE/JOINT INVENTION
(Worldwide Rights)
Attorney Docket No. 6843.0057-00000

ASSIGNMENT

WHEREAS We, the below named inventors, [hereinafter referred to as Assignors], have made an invention entitled:

RAPID DISSOLUTION FORMULATION OF A CALCIUM RECEPTOR-ACTIVE COMPOUND

for which We executed an application for United States Letters Patent concurrently herewith or filed an application for United States Letters Patent on September 10, 2004, (Application No. 10/937,870); and

WHEREAS, **Amgen Inc.**, a corporation of Delaware, whose post office address is One Amgen Center Drive, Thousand Oaks, CA 91320-1799 (hereinafter referred to as Assignee), is desirous of securing the entire right, title, and interest in and to this invention in all countries throughout the world, and in and to the application for United States Letters Patent on this invention and the Letters Patent to be issued upon this application;

NOW THEREFORE, be it known that for good and valuable consideration the receipt of which from Assignee is hereby acknowledged, We, as Assignors, have sold, assigned, transferred, and set over, and do hereby sell, assign, transfer, and set over unto the Assignee, its lawful successors and assigns, our entire right, title, and interest in and to this invention, the U.S. Provisional Application No. 60/502,219, filed September 12, 2003, U.S. Application No. 10/937,870, and all divisions, and continuations thereof, and all Letters Patent of the United States which may be granted thereon, and all reissues thereof, and all rights to claim priority on the basis of the U.S. Provisional Application No. 60/502,219, filed September 12, 2003, and all rights to claim priority on the basis of U.S. Application No. 10/937,870, and all applications for Letters Patent which may hereafter be filed for this invention in any foreign country and all Letters Patent which may be granted on this invention in any foreign country, and all extensions, renewals, and reissues thereof; and We hereby authorize and request the Commissioner of Patents and Trademarks of the United States and any official of any foreign country whose duty it is to issue patents on applications as described above, to issue all Letters Patent for this invention to Assignee, its successors and assigns, in accordance with the terms of this Assignment;

AND, WE HEREBY covenant that We have the full right to convey the interest assigned by this Assignment, and We have not executed and will not execute any agreement in conflict with this Assignment;

AND, WE HEREBY further covenant and agree that We will, without further consideration, communicate with Assignee, its successors and assigns, any facts known to us respecting this invention, and testify in any legal proceeding, sign all lawful papers when called upon to do so, execute and deliver any and all papers that may be necessary or desirable to perfect the title to this invention in said Assignee, its successors or assigns, execute all divisional, continuation, and reissue applications, make all rightful oaths and generally do everything possible to aid Assignee, its successors and assigns, to obtain and enforce proper patent protection for this invention in the United States and any foreign country, it being understood that any expense incident to the execution of such papers shall be borne by the Assignee, its successors and assigns.

IN TESTIMONY WHEREOF, We have hereunto set our hands.

County of _____)
) ss.
State of _____)

Name: Glen Gary LAWRENCE
Address: 3439 Indian Ridge Circle
Thousand Oaks, CA 91362-4949
By: _____
Date: _____

Subscribed and sworn to before me this _____ day of _____, 20___
_____, Notary Public

County of _____)
) ss.
)
State of _____)

Name: Francisco J. ALVAREZ
Address: 817 Paseo de Leon
Newbury Park, CA 91320
By: _____
Date: _____

Subscribed and sworn to before me this _____ day of _____, 20___
_____, Notary Public

County of _____)
) ss.
)
State of _____)

Name: Hung-Ren H. LIN
Address: 5086 Durant Court
Oak Park, CA 91377
By: _____
Date: _____

Subscribed and sworn to before me this _____ day of _____, 20___
_____, Notary Public

County of Lake)
) ss.
)
State of Illinois)

Name: Tzuchi R. JU
Address: 1438 N. Butler Court
Vernon Hills, IL 60061
By: Joseph R. Figueroa
Date: 12-6-04

Subscribed and sworn to before me this 6 day of Dec, 2004
_____, Notary Public

OFFICIAL SEAL
JOSESPH R. FIGUEROA
NOTARY PUBLIC, STATE OF ILLINOIS
My Commission Expires May 29, 2007

IFW

PATENT APPLICATION

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s):	Glen Lawrence et al.	Docket No.:	A-870 (US)
Serial No.:	10/937,870	Group Art Unit No.:	1614
Filed:	September 10, 2004	Examiner:	Unknown
For:	RAPID DISSOLUTION FORMULATION OF A CALCIUM RECEPTOR-ACTIVE COMPOUND		

INFORMATION DISCLOSURE STATEMENT

Mail Stop Amendment
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

Sir:

As a means of complying with the duty of disclosure under 37 CFR 1.97 and 1.98, applicants submit a "List of References Cited by Applicant" on a modified PTO-1449 form, along with copies of foreign patent documents and non-patent documents, for consideration by the Examiner.

The Information Disclosure Statement submitted herewith is being filed within three months of the filing date of the application or date of entry into the national stage of an international application or before the mailing date of a first Office action on the merits, or before the mailing of a first Office action after the filing of a request for continued examination under section 1.114, whichever event occurs last. 37 CFR 1.97(b).

Applicant(s) request(s) consideration of this information and passage of the application to issue.

The Commissioner is hereby authorized to charge any filing fees, which may be required or credit any overpayment to Deposit Account No. 01-0519 in the name of Amgen Inc.

Respectfully submitted,

Please send all future correspondence to:
AMGEN INC., Customer No. 30,174
1120 Veterans Boulevard
South San Francisco, CA 94112
Fax: (650) 244-2392

Olga Mekhovich
Attorney for Applicants
Registration No.: L0066
Phone: (650) 244-2000
Date: September 2, 2005

CERTIFICATE OF MAILING

I hereby certify that this correspondence is being deposited with the United States Postal Service as First Class Mail addressed to Mail Stop Amendment, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450, on the date appearing below.

September 2, 2005
Date

Julie M. Balot

Modified Form PTO-1449	Atty. Docket No. A-870 (US)	Serial No. 10/937.870
LIST OF REFERENCES CITED BY APPLICANT (Use several sheets if necessary)	Applicant GLEN LAWRENCE ET AL.	
	Filing Date Sept. 10, 2004	Group 1614

U.S. PATENT DOCUMENTS

EXAMINER'S INITIALS		DOCUMENT NUMBER	DATE	NAME	CLASS	SUB-CLASS	FILING DATE IF APPROPRIATE
	A1	5,126,145	6/30/1992	Evenstad et al.			
	A2	5,981,599	11/9/1999	Moe et al.			
	A3	6,001,884	12/14/1999	Nemeth et al.			
	A4	6,011,068	1/4/2000	Nemeth et al.			
	A5	6,031,003	2/29/2000	Nemeth et al.			
	A6	6,172,091	1/9/2001	Cohen et al.			
	A7	6,211,244	4/3/2001	Van Wagenen et al.			
	A8	6,228,807	5/8/2001	Kuchikata et al.			
	A9	6,277,788	8/21/2001	Wright			
	A10	6,313,146	11/6/2001	Van Wagenen et al.			
	A11	6,342,532	1/29/2002	Moe et al.			
	A12	6,363,231	3/26/2002	Manzer et al.			
	A13	6,387,404	5/14/2002	Oshlack et al.			
	A14	6,399,100	6/4/2002	Clancy et al.			
	A15	6,432,656	8/13/2002	Del Mar et al.			
	A16	6,495,165	12/17/2002	Thosar et al.			
	A17	2002/0015735	2/7/2002	Hedden et al.			
	A18	2002/0107406	8/8/2002	Sakai et al.			
	A19	2003/0035836	2/20/2003	Shanghvi et al.			
	A20	2003/0054041	3/20/2003	Lemmens et al.			

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EXAMINER: Initial if citation considered, whether or not citation is in conformance with MPEP 609; Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.	

Modified Form PTO-1449	Atty. Docket No. A-870 (US)	Serial No. 10/937.870
LIST OF REFERENCES CITED BY APPLICANT	Applicant GLEN LAWRENCE ET AL.	
(Use several sheets if necessary)	Filing Date Sept. 10, 2004	Group 1614

FOREIGN PATENT DOCUMENTS

EXAMINER' INITIALS		DOCUMENT NUMBER	DATE	COUNTRY	CLASS	SUB-CLASS	TRANSLATION YES	TRANSLATION NO
	B1	WO 93/04373	3/4/1993	PCT				
	B2	WO 94/18959	9/1/1994	PCT				
	B3	WO 95/11221	4/27/1995	PCT				
	B4	WO 96/12697	5/2/1996	PCT				
	B5	WO 97/41090	11/6/1997	PCT				
	B6	WO 01/34562	5/17/2001	PCT				
	B7	EP 0 933 354	8/4/1999	EUROPE				

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A-25b (Rev. 07/01/03)

Modified Form PTO-1449	Atty. Docket No. A-870 (US)	Serial No. 10/937.870
LIST OF REFERENCES CITED BY APPLICANT	Applicant GLEN LAWRENCE ET AL.	
(Use several sheets if necessary)	Filing Date Sept. 10, 2004	Group 1614

EXAMINER'S INITIALS		**OTHER DOCUMENTS (Including Publisher, Author, Title, Date, Pertinent Pages, Etc.)**
	C1	Amgen News Release, Internet Article: "Amgen Submitted New Drug Application for Cinecalcet HC1", XP002313388, URL: http://www.amgen.com/news/news03/pressRelease030908a.pdf, September 8, 2003, 2 pages.
	C2	Berge, Stephen M., et al., "Pharmaceutical Salts," *J. Pharm. Sci.*, Vol. 66, No. 1, pp. 1-19, January 1977.
	C3	Drugs in R&D, "Cinecalcet: AMG 073, Calcimimetics – Amgen/NPS Pharmaceuticals, KRN 1493, NPS 1493," Vol. 4, No. 6, pp 349-351, 2003.
	C4	Goodman, William G., et al., "The Calcimimetic Agent AMG 073 Lowers Plasma Parathyroid Hormone Levels in Hemodialysis Patients with Secondary Hyperparathyroidism," *J. Am. Soc. Nephrol*, Vol. 13, pp. 1017-1024, April 2002.
	C5	Nemeth, Edward F., et al., "Pharmacodynamics of the Type II Calcimimetic Compound Cinacalcet HCl, *J. Pharmacol. Exp Therapeutics*, Vol. 308, No. 2, pp. 627-635, February 2004.
	C6	Pattaragarn, Anirut, et al., "Effect of the Calcimimetic NPS R-467 on Furosemide-Induced Nephrocalcinosis in the Young Rat," *Kidney Internationa*, Vol. 65, pp. 1684-1689, 2004.
	C7	U.S. Food and Drug Administration, Internet Article: "FDA Approves First in a New Class of Drugs to Treat Hyperparathyroidism Associated with Renal Failure and in Patients with Parathyroid Cancer, " *FDA Talk Paper*, XP-002313389, http://www.fda.gov/bbs/topics/ANSWERS/2004/ANS01282.html), March 8, 2004, 1 page, (retrieved by EPO Searching Authority on 1/11/2005).
	C8	RxList, Inc., Internet Article: "Sensipar (Cinacalcet HCl) Tablets," XP-002313390, www.rxlist.com/cgi/generic3/sensipar.htm), August 12, 2004, 2 pages, (retrieved by EPO Searching Authority on 1/10/2005).
	C9	NPS Pharmaceuticals, Internet Article: "NPS Drug Development: Product Development Pipeline," XP-002313391, www.npsp.com/drug-development/pipline.php, September 8, 2003, 2 pages, (retrieved by EPO Searching Authority on 1/12/2005)

EXAMINER:	Date Considered:
EXAMINER: Initial if citation considered, whether or not citation is in conformance with MPEP 609; Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.	

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5 :
G01N 33/566, 33/567
C07C 211/02, 211/16, 211/27
C07H 21/00, C07K 5/00, 7/00
C12N 15/12, A61K 37/02

A1

(11) International Publication Number: WO 93/04373

(43) International Publication Date: 4 March 1993 (04.03.93)

(21) International Application Number: PCT/US92/07175

(22) International Filing Date: 21 August 1992 (21.08.92)

(30) Priority data:

749,451	23 August 1991 (23.08.91)	US
834,044	11 February 1992 (11.02.92)	US

(60) Parent Applications or Grants
(63) Related by Continuation

US	07/934,161 (CIP)
Filed on	21 August 1992 (21.08.92)
US	07/834,044 (CIP)
Filed on	11 February 1992 (11.02.92)
US	07/749,451 (CIP)
Filed on	23 August 1991 (23.08.91)

(71) Applicant *(for all designated States except US)*: NPS PHARMACEUTICALS, INC. [US/US]; 420 Chipeta Way, Salt Lake City, UT 84108 (US).

(72) Inventors; and
(75) Inventors/Applicants *(for US only)* : NEMETH, Edward, F. [US/US]; 3258 E. Fortuna Drive, Salt Lake City, UT 84124 (US). VAN WAGENEN, Bradford, C. [US/US]; 1070 East 300 South, #507, Salt Lake City, UT 84102 (US). BALANDRIN, Manuel, F. [US/US]; 5787 South 1585 East, Salt Lake City, UT 84121 (US).

(74) Agents: WARBURG, Richard, J. et al.; Lyon & Lyon, 611 West Sixth Street, 34th Floor, Los Angeles, CA 90017 (US).

(81) Designated States: AT, AU, BB, BG, BR, CA, CH, CS, DE, DK, ES, FI, GB, HU, JP, KP, KR, LK, LU, MG, MN, MW, NL, NO, PL, RO, RU, SD, SE, US , European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, SN, TD, TG).

Published
With international search report.

(54) Title: CALCIUM RECEPTOR ACTIVE MOLECULES

(57) Abstract

Method and composition useful for treating a patient having a disease characterized by an abnormal level of one or more components, the activity of which is regulated or affected by activity of one or more Ca^{2+} receptors. Novel compounds useful in these methods and compositions are also provided. The method includes administering to the patient a therapeutically effective amount of a molecule active at one or more Ca^{2+} receptors as an agonist or antagonist. Preferably, the molecule is able to act as either a selective agonist or antagonist at a Ca^{2+} receptor of one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental throphoblasts and a pharmaceutically acceptable carrier.

FOR THE PURPOSES OF INFORMATION ONLY

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DESCRIPTION

Calcium Receptor Active Molecules

Related Application

This application is a continuation-in-part of Nemeth, et al., entitled Calcium Receptor Active Molecules, filed February 11, 1992, which is a continuation-in-part of Nemeth et al., entitled Calcium Receptor Activators, U.S. Serial No. 07/749,451, filed August 23, 1991, the whole of both of which, including drawings, are hereby incorporated herein by reference.

Field of the Invention

This invention relates to the design, development, composition and use of novel calcimimetic molecules able to act in a manner analogous to extracellular calcium ions on cells, to calcilytic molecules which block the activity of extracellular calcium ions on cells, and to methods for their use and identification.

Background of the Invention

The following description provides a summary of information relevant to the present invention. It is not an admission that any of the information provided herein is prior art to the presently claimed invention, nor that any of the publications specifically or implicitly referenced are prior art to that invention.

Certain cells in the body respond not only to chemical signals, but also to ions such as extracellular calcium ions (Ca^{2+}). Changes in the concentration of extracellular Ca^{2+} (referred to herein as "$[Ca^{2+}]$") alter the functional responses of these cells. One such specialized cell is the parathyroid cell which secretes parathyroid hormone (PTH). PTH is the principal endocrine factor regulating Ca^{2+} homeostasis in the blood and extracellular fluids.

PTH, by acting on bone and kidney cells, increases the level of Ca^{2+} in the blood. This increase in $[Ca^{2+}]$ then acts as a negative feedback signal, depressing PTH secretion. The reciprocal relationship between $[Ca^{2+}]$ and PTH secretion forms the essential mechanism maintaining bodily Ca^{2+} homeostasis.

Extracellular Ca^{2+} acts directly on the parathyroid cell to regulate PTH secretion. The existence of a parathyroid cell surface protein which detects changes in $[Ca^{2+}]$ has been suggested. This protein acts as a receptor for extracellular Ca^{2+} ("the Ca^{2+} receptor"), and is suggested to detect changes in $[Ca^{2+}]$ and to initiate a functional cellular response, PTH secretion. For example, the role of Ca^{2+} receptors and extracellular Ca^{2+} in the regulation of intracellular Ca^{2+} and cell function is reviewed in Nemeth et al., 11 Cell Calcium 319, 1990; the role of Ca^{2+} receptors in parafollicular and parathyroid cells is discussed in Nemeth, 11 Cell Calcium 323, 1990; and the role of Ca^{2+} receptors on bone osteoclasts is discussed by Zaidi, 10 Bioscience Reports 493, 1990.

Other cells in the body, specifically the osteoclast in bone, the juxtaglomerular and the proximal tubule cells in the kidney, the keratinocyte in the epidermis, the parafollicular cell in the thyroid, and the trophoblast in the placenta, have the capacity to sense changes in $[Ca^{2+}]$. It has been suggested that cell surface Ca^{2+} receptors may also be present on these cells, imparting to them the ability to detect and to initiate or enable a response to changes in $[Ca^{2+}]$.

In parathyroid cells, osteoclasts, parafollicular cells (C-cells), keratinocytes, juxtaglomerular cells and trophoblasts, an increase in $[Ca^{2+}]$ evokes an increase in intracellular free Ca^{2+} concentration ("$[Ca^{2+}]_i$"). Such an increase may be caused by influx of extracellular Ca^{2+} or by mobilization of Ca^{2+} from intracellular organelles. Changes in $[Ca^{2+}]_i$ are readily monitored and quantitated

using fluorimetric indicators such as fura-2 or indo-1 (Molecular Probes, Eugene, OR). Measurement of $[Ca^{2+}]_i$ provides an assay to assess the ability of molecules to act as agonists or antagonists at the Ca^{2+} receptor.

In parathyroid cells, increases in the concentration of extracellular Ca^{2+} evoke rapid and transient increases in $[Ca^{2+}]_i$ which are followed by lower yet sustained increases in $[Ca^{2+}]_i$. The transient increases in $[Ca^{2+}]_i$ arise from the mobilization of intracellular Ca^{2+}, whereas the lower, sustained increases result from the influx of extracellular Ca^{2+}. The mobilization of intracellular Ca^{2+} is accompanied by increased formation of inositol-1,4,5-trisphosphate (IP_3) and diacylglycerol, two biochemical indicators which are associated with receptor-dependent mobilization of intracellular Ca^{2+} in various other cells.

In addition to Ca^{2+}, various other di- and trivalent cations, such as Mg^{2+}, Sr^{2+}, Ba^{2+}, La^{3+}, and Gd^{3+} also cause the mobilization of intracellular Ca^{2+} in parathyroid cells. Mg^{2+} and La^{3+} also increase the formation of IP_3; all these inorganic cations depress the secretion of PTH. The postulated Ca^{2+} receptor on the parathyroid cell is therefore promiscuous because it detects a variety of extracellular di- and trivalent cations.

The ability of various compounds to mimic extracellular Ca^{2+} in vitro is discussed by Nemeth et al., (spermine and spermidine) in "Calcium-Binding Proteins in Health and Disease", 1987, Academic Press, Inc., pp. 33-35; Brown et al., (e.g., neomycin) 128 Endocrinology 3047, 1991; Chen et al., (diltiazem and its analog, TA-3090) 5 J. Bone and Mineral Res. 581, 1990; and Zaidi et al., (verapamil) 167 Biochem Biophys Res Comm 807, 1990.

Brown et al., 6 J. Bone and Mineral Res. 11, 1991 discuss the existing theories regarding the effects of Ca^{2+} ions on parathyroid cells, and propose that the results may be explained by both a receptor-like mechanism and a receptor-independent mechanism as follows:

Polyvalent cations [e.g., divalent and trivalent cations] exert a variety of effects on parathyroid function, such as inhibition of parathyroid hormone (PTH) secretion and cAMP accumulation, stimulation of the accumulation of inositol phosphates, and elevation of the cytosolic calcium concentration. These actions are thought to be mediated through a "receptor-like" mechanism. The inhibition of agonist-stimulated cAMP accumulation by divalent and trivalent cations, for example, is blocked following preincubation with pertussis toxin. Thus, the putative polyvalent cation receptor may be coupled to inhibition of adenylate cyclase by the inhibitory guanine nucleotide regulatory (G) protein, G_i.

We recently showed that the polycationic antibiotic, neomycin, mimics the actions of di-and trivalent cations in several aspects of parathyroid function. To determine whether these actions were specific to this agent or represented a more generalized action of polycations, we tested the effects of the highly basic peptides, polyarginine and polylysine, as well as protamine on the same parameters in dispersed bovine parathyroid cells. The results demonstrate that the parathyroid cell responds to a variety of polycations as well as to polyvalent cations, potentially via similar biochemical pathways. These results are discussed in terms of the recently postulated, "receptor-independent" modulation of G proteins by polycations in other systems.

* * *

The Ca^{2+} receptor has been presumed to be analogous to other G protein-coupled receptors [e.g., a glycoprotein], but recent studies with other cell types have raised the possibility that polycations can modulate cell function by alternative or additional mechanisms. In mast cells, for example, a variety of amphipathic cations, including mastoparan, a peptide from wasp venom, 48/80, a synthetic polycation, and polylysine, enhance secretion by a pertussis toxin-sensitive mechanism, suggesting the involvement of a G protein. No classic cell surface receptor has been identified that could mediate the actions of these diverse agents. Furthermore, these same compounds have been shown to activate directly purified G proteins in solution or in artificial phospholipid vesicles. On the basis of these observations, it has been proposed that amphipathic cations activate G proteins and, in turn, mast cell secretion by a "receptor-independent" mechanism.

Polycations have also been shown to interact strongly with acidic phospholipids. Polylysines of varying chain lengths (20-1000 amino acids) bind to artificial phospholipid vesicles with dissociation constants in the range of 0.5 nM to 1.5 μM. The binding affinity is directly related to the length of the polylysine chain, with polymers of 1000 amino acids having a K_d of 0.5 nM, shorter polymers having higher Kd values, and lysine not interacting to a significant

extent. This relationship between potency and chain length is similar to that observed for the effects of polylysine $_{10,200}$, polylysine $_{3800}$, and lysine on parathyroid function.

It is possible that the binding of polycations to biomembranes produces some of their biologic actions. The permeabilization of the plasma membrane induced in some cell types by a variety of pore-forming agents, including polycations, has been postulated to be mediated by their interaction with a phosphatidylserine-like structure. In addition, the "receptor-independent" activation of purified G proteins by amphipathic cations is potentiated when these proteins are incorporated into phospholipid vesicles.

Calcium ions, in the millimolar concentration range, also produce marked changes in membrane structure. In some cases, calcium can either antagonize or potentiate the interaction of polycations with membrane lipids. These considerations raise the possibility that the actions of both polyvalent cations and polycations on parathyroid cells could involve a receptor-independent mechanism not requiring the presence of a classic, cell surface, G protein-coupled receptor. Further studies, however, are required to elucidate the molecular basis for Ca^{2+} sensing by this and other cell types. [Citations omitted.]

Shoback and Chen (6 (Supplement 1), J. Bone and Mineral Res. 1991, S135) and Racke et al. (6 (Supplement 1), J. Bone and Mineral Res. 1991, S118) describe experiments which are said to indicate that a Ca^{2+} receptor or

Ca^{2+} sensor is present in parathyroid cells. Messenger RNA isolated from such cells can be expressed in oocytes and caused to provide those oocytes with a phenotype which might be explained by the presence of a Ca^{2+} receptor protein.

Summary of the Invention

Applicant has demonstrated that Ca^{2+} receptor proteins enable certain specialized cells involved in bodily Ca^{2+} metabolism to detect and respond to changes in the concentration of extracellular Ca^{2+}. Although these receptors share certain general characteristics, they can be selectively affected by different pharmacological agents. As detailed below, certain molecules are identified with selective activity on Ca^{2+} receptors at parathyroid cells, osteoclasts, and C-cells.

Ca^{2+} receptors constitute discrete molecular targets for a new class of molecules that mimic ("calcimimetics") or antagonize ("calcilytics") the actions of extracellular Ca^{2+}. Such receptors are present on cell surfaces and have a low affinity for extracellular Ca^{2+} (apparent K_d generally greater than about 0.5 mM). Such receptors may include a free or bound effector mechanism, as defined by Cooper, Bloom and Roth, "The Biochemical Basis of Neuropharmacology", Ch. 4. Such receptors are thus distinct from intracellular Ca^{2+} receptors, e.g., calmodulin and the troponins. Calcimimetics, for example, act on Ca^{2+} receptors selectively to directly or indirectly depress the function of parathyroid cells or osteoclasts or to stimulate the function of C-cells. Calcimimetics and calcilytics of this invention allow novel therapies for hyperparathyroidism, osteoporosis and other Ca^{2+}-related diseases. This application concerns targeting Ca^{2+} receptors on each of these three cell types and other cell types that detect and respond to changes in $[Ca^{2+}]$.

Applicant is the first to demonstrate a Ca^{2+} receptor protein in parathyroid cells, and to pharmacologically

differentiate such Ca^{2+} receptors in other cells, such as C-cells and osteoclasts. Applicant is also the first to describe methods by which molecules active at these Ca^{2+} receptors can be identified and used as lead molecules in the discovery, development, design, modification and/or construction of useful calcimimetics or calcilytics which are active at Ca^{2+} receptors. Such calcimimetics or calcilytics are useful in the treatment of various disease states characterized by abnormal levels of one or more components, e.g., polypeptides such as hormones, enzymes or growth factors, the expression and/or secretion of which is regulated or affected by activity at one or more Ca^{2+} receptors. Further, the identification of different Ca^{2+} receptors in different cell types, and the specific response of such receptors to different lead molecules allows design and construction of specific molecules active in treatment of specific diseases which can be affected by action at such specific Ca^{2+} receptors. For example, abnormal levels of parathyroid hormone secretion can be affected by such specific molecules without affecting the level of secretion of other Ca^{2+} regulated hormones and the like.

Identification of such lead molecules was impeded by the prior lack of a high-throughput screening system to discover active molecules, and the absence of a structural data base upon which to design effective drug candidates. These barriers are now removed by cloning the parathyroid cell Ca^{2+} receptor and functionally related receptors, and systematically examining the structural features of certain lead molecules that activate such cloned Ca^{2+} receptors and functionally related receptors. Cloning of the Ca^{2+} receptor also enables development of transfected cell lines suitable for high-throughput screening of natural product or molecule libraries and synthetic molecules. This, together with structure-activity studies discussed below, provides the technology necessary to develop novel calcimimetics and calcilytics.

Applicant enables such procedures in this application. For example, the human parathyroid cell Ca^{2+} receptor cDNA can be cloned by screening for functional expression in Xenopus oocytes, and the structural features of organic molecules necessary for activity on the Ca^{2+} receptor can be determined through the testing of selected natural products or other molecule libraries and subsequent structure-activity studies.

Thus, in a first aspect, the invention features a pharmaceutical composition including a molecule which either mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell, or blocks an increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. The molecule has an EC_{50} of less than or equal to 5 μM, and is not protamine.

By "mimic" is meant that the molecule has one or more of the specific actions of extracellular Ca^{2+} on an extracellular Ca^{2+} responsive cell. The term does not require that all of the biological functions of extracellular Ca^{2+} are mimicked, but rather than at least one such function is mimicked. In addition it does not require that the molecule bind to the same site on the receptor as does extracellular Ca^{2+} receptor (see for example, the novel compound NPS 467 and its action in Example 20 below). By "block" is meant that one such action of Ca^{2+} is reduced or prevented by the molecule. The EC_{50} can be determined in assays as described below, where the activity mimicked is measured and the concentration of molecule which mimics at half the maximum mimicking effect is the EC_{50}. Conversely, the IC_{50} of a calcilytic is that amount which blocks half maximal activity. Preferably, such assays measure $[Ca^{2+}]_i$ increases and are confirmed to be specific to a Ca^{2+} receptor by methods described below, or their equivalent.

In preferred embodiments, bioassays described herein demonstrated that the increase in $[Ca^{2+}]_i$ in a cell is transient, having a duration of less than one minute, and the increase in $[Ca^{2+}]_i$ is rapid, occurring within thirty seconds; and the molecule also (a) evokes a sustained

increase (greater than thirty seconds) in $[Ca^{2+}]_i$, (b) evokes an increase in inositol-1,4,5-trisphosphate and/or diacylglycerol levels, e.g., within less than 60 seconds, and (c) inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation. In addition, the transient increase in $[Ca^{2+}]_i$ is abolished by pretreatment of the cell for ten minutes with 10 mM sodium fluoride, or the transient increase is diminished by brief pretreatment (not more than ten minutes) of the cell with an activator of protein kinase C, e.g., phorbol myristate acetate (PMA), mezerein or (-)indolactam V.

In a parathyroid cell, those molecules which are active in all of the assays described above are particularly useful in this invention since they are specific in their actions to a Ca^{2+} receptor of such a cell. This is particularly true for the PMA pretreatment effect described above.

In a more preferred embodiment, the cell is a parathyroid cell, and the molecule inhibits parathyroid hormone secretion from the cell; and the molecule elicits an increase in Cl^- conductance in a Xenopus oocyte injected with mRNA from a parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, keratinocyte, parafollicular thyroid cell or placental trophoblast.

In other preferred embodiments, the molecule evokes the mobilization of intracellular Ca^{2+} to cause the increase in $[Ca^{2+}]_i$; the cell is a C-cell or an osteoclast and the molecule inhibits bone resorption in vivo; the cell is an osteoclast and the molecule inhibits bone resorption in vitro; or the cell is a C-cell and the molecule stimulates calcitonin secretion in vitro or in vivo; and most preferably the molecule is either a calcimimetic or calcilytic having an EC_{50} or IC_{50} at a Ca^{2+} receptor of less than or equal to 5 μM, and even more preferably less than or equal to 1 μM, 100 nmolar, 10 nmolar, or 1 nmolar. Such lower EC_{50}'s or IC_{50}'s are

advantageous since they allow lower concentrations of molecules to be used in vivo or in vitro for therapy or diagnosis. The discovery of molecules with such low EC_{50}'s and IC_{50}'s enables the design and synthesis of similarly potent and efficacious molecules.

By "calcimimetic" molecule is meant any molecule which has one or more activities of extracellular Ca^{2+}, and preferably mimics the activity of Ca^{2+} at a Ca^{2+} receptor. For example, when used in reference to a parathyroid cell it is a molecule, which when tested on parathyroid cells, in vitro, possesses one or more, and preferably all of the following characteristics as measured by techniques well known to those in the art:

1. The molecule causes a rapid (time to peak < 5 sec) and transient increase in $[Ca^{2+}]_i$ that is refractory to inhibition by 1 μM La^{3+} or Gd^{3+}. The increase in $[Ca^{2+}]_i$ persists in the absence of extracellular Ca^{2+} but is abolished by pretreatment with ionomycin (in the absence of extracellular Ca^{2+});

2. The increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+} is not inhibited by dihydropyridines.

3. The transient increase in $[Ca^{2+}]_i$ caused by the molecule is abolished by pretreatment for 10 min. with 10 mM sodium fluoride;

4. The transient increase in $[Ca^{2+}]_i$ caused by the molecule is diminished by pretreatment with an activator of protein kinase C (PKC), such as phorbol myristate acetate (PMA), mezerein or (-)-indolactam V. The overall effect of the protein kinase C activator is to shift the concentration-response curve of the molecule to the right without affecting the maximal response;

5. The molecule causes a rapid (< 30 sec.) increase in the formation of inositol-1,4,5-trisphosphate and or diacylglycerol;

6. The molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation;

7. The molecule inhibits PTH secretion;

8. Pretreatment with pertussis toxin (100 ng/ml for > 4 hrs.) blocks the inhibitory affect of the molecule on cyclic AMP formation but does not effect increases in $[Ca^{2+}]_i$, inositol-1,4,5-trisphosphate, or diacylglycerol, nor decreases in PTH secretion;

9. The molecule elicits increases in Cl^- conductance in Xenopus oocytes injected with poly(A)$^+$- enriched mRNA from bovine or human parathyroid cells but is without effect in Xenopus oocytes injected with water or rat brain or liver mRNA; and

10. Similarly, using a cloned receptor from parathyroid cells, the molecule will elicit a response in Xenopus oocytes injected with the specific cDNA or mRNA encoding the receptor.

By "calcilytic" molecule is meant any molecule which blocks one or more of the activities of extracellular Ca^{2+} on an extracellular Ca^{2+}-sensing cell, preferably by acting as an antagonist at the Ca^{2+} receptor. For example, when used in reference to a parathyroid cell, it is a molecule which, when tested on parathyroid cells in vitro, possesses one or more, and preferably all of the following characteristics as measured by techniques well known to those in the art:

1. The molecule blocks, either partially or completely, the ability of increased concentrations of extracellular Ca^{2+} to:

a) increase $[Ca^{2+}]_i$,

b) mobilize intracellular Ca^{2+},

c) increase the formation of inositol-1,4,5-trisphosphate,

d) decrease dopamine- or isoproterenol-stimulated cyclic AMP formation, and

e) inhibit PTH secretion;

2. At low $[Ca^{2+}]$, i.e., 0.5 mM, the molecule by itself does not change $[Ca^{2+}]_i$

3. The molecule blocks increases in Cl^- conductance in Xenopus oocytes injected with poly(A)$^+$-mRNA from bovine

or human parathyroid cells elicited by extracellular Ca^{2+} or calcimimetic compounds but not in Xenopus oocytes injected with water or rat brain or liver mRNA;

4. Similarly, using a cloned receptor from parathyroid cells, the molecule will block a response in Xenopus oocytes injected with the specific cDNA or mRNA encoding the Ca^{2+} receptor, elicited by extracellular Ca^{2+} or a calcimimetic compound.

Parallel definitions of useful calcimimetics and calcilytics at Ca^{2+} receptors on other cell types are evident from the examples provided below.

The Ca^{2+} receptor is able to detect and respond to certain inorganic polycations and polycationic organic molecules. For example, the parathyroid cell is unable to distinguish increases in extracellular Ca^{2+} concentration from the addition of these organic polycations, presumably because these organic molecules act just like extracellular Ca^{2+} at the Ca^{2+} receptor. The calcimimetic molecules of this invention are particularly good agonists of the Ca^{2+} receptor and may be used as drugs that alter selected cellular functions, e.g., secretion of PTH from parathyroid cells. Unlike Ca^{2+} most of these molecules act only at one or more, but not all Ca^{2+} receptors, and thus provide an ability to specifically target one Ca^{2+} receptor.

These molecules also provide lead structures for the development of further novel therapeutics effective in the treatment of various diseases where $[Ca^{2+}]_i$ and $[Ca^{2+}]$ play a role, such as hyperparathyroidism, osteoporosis, Paget's disease, hypertension, renal disease, and cancer.

The calcimimetics and calcilytics can be formulated as pharmaceutical compositions which are useful for regulating the level of extracellular free Ca^{2+} in a patient and for mimicking the effect of extracellular Ca^{2+} on a cell selected from the group described above, by administering to the patient such a pharmaceutical composition. Prior to this invention, applicant was unaware of any such

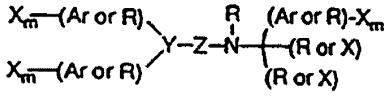
molecules acting on the Ca^{2+} receptor useful in treatment of diseases caused by irregularity in operation or regulation of a Ca^{2+} receptor or diseases in an animal having normal Ca^{2+} receptors but which can be treated by activating or deactivating such Ca^{2+} receptors.

In yet another preferred embodiment, the molecule has an EC_{50} less than or equal to 5 μM at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells (C-cells) and placental trophoblasts.

It is the specificity of action of such molecules that is particularly advantageous in this invention since it allows specific in vivo and in vitro therapy and diagnosis and discovery of additional calcimimetic or calcilytic molecules.

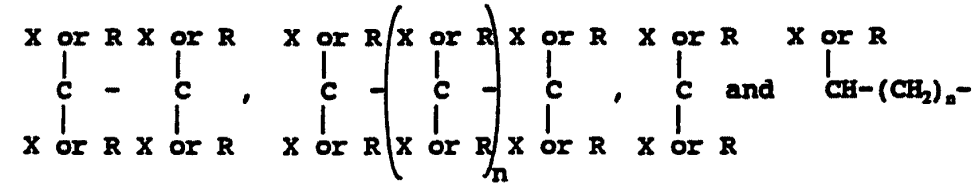
In specific preferred embodiments, the molecule is positively charged at physiological pH, and is selected from the group consisting of branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines, e.g., the branched polyamine has the formula $H_2N-(CH_2)_j-(NR_i-(CH_2)_j)_k-NH_2$ where k is an integer from 1 to 10, each j is the same or different and is an integer from 2 to 20, and each R_i is the same or different and is selected from the group consisting of hydrogen and $-(CH_2)_j-NH_2$, where j is as defined above, and at least one R_i is not hydrogen.

In an alternative embodiment, the molecule has the formula



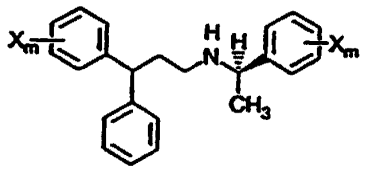
where each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, and CH_3CH_2; Ar is a hydrophobic entity; each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl,

isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, 2-, 3-, or 4-piperid(in)yl; Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon; Z is selected from the group consisting of oxygen, nitrogen, sulfur,



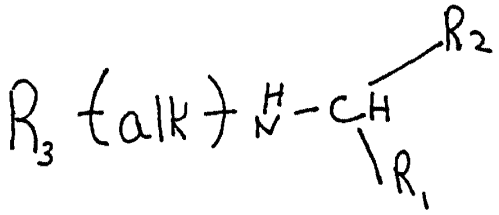
where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive. Most preferably the molecule is either a calcimimetic or calcilytic.

In preferred embodiments, the hydrophobic entity is selected from the group consisting of phenyl, 2-, 3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2-quinolinyl, 2- or 3-indolyl, benzyl, and phenoxy; the molecule is an R-diphenylpropyl-α-phenethylamine derivative, and the molecule has the formula:



with each X preferably being independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

According to a preferred aspect of the present invention, novel phenyl-α-phenethylamine analogs and derivatives are provided having the formula:



wherein alk is straight or branched chain alkylene of from 1 to 6 carbon atoms; R_1 is lower alkyl of from 1 to 3 carbon atoms or lower haloalkyl of from 1 to 3 carbon atoms substituted with from 1 to 7 halogen atoms; R_2 and R_3 are independently selected carbocyclic aryl or cycloalkyl groups, either monocyclic or bicyclic, having 5- or 6-membered rings optionally substituted with 1 to 5 substituents independently selected from lower alkyl of 1 to 3 carbon atoms, lower haloalkyl of 1 to 3 carbon atoms substituted with 1 to 7 halogen atoms, lower alkoxy of 1 to 3 carbon atoms, halogen, nitro, amino, alkylamino, amido, lower alkylamido of 1 to 3 carbon atoms, cyano, hydroxy, acyl of 2 to 4 carbon atoms lower hydroxyalkyl of 1 to 3 carbon atoms or lower thioalkyl of 1 to 3 carbon atoms. Suitable carbocyclic aryl groups are groups having one or two rings, at least one of which having aromatic character and include carbocyclic aryl groups such as phenyl and bicyclic carbocyclic aryl groups such as naphthyl. As is apparent from the above formula, the compounds encompassed therein may exist as racemic mixtures and as individual stereoisomers. Especially preferred are R-phenylpropyl

α-phenethylamine derivatives which are believed to exhibit enhanced activity in lowering serum ionized calcium.

Preferred compounds include those where alk is n-propylene. Also preferred are compounds where R_1 is methyl. Also preferred are those compounds where R_2 and R_3 are optionally substituted phenyl.

Especially preferred compounds include those where R_2 is monosubstituted phenyl, more preferably meta-substituted. Especially preferred R_3 groups include unsubstituted or monosubstituted phenyl, especially ortho-substituted. Preferred substitutents for R_2 include halogen, haloalkyl, preferably trihalomethyl, and alkoxy, preferably methoxy. Preferred substituents for R_3 include halogen.

In a second related aspect, the invention features a method for treating a patient having a disease or condition characterized by an abnormal $[Ca^{2+}]$ or $[Ca^{2+}]_i$ in one or more cells or in the blood or plasma or extracellular fluids. The method includes the step of administering to the patient a therapeutically effective amount of a molecule which either mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell or blocks an increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}.

By "abnormal" is meant that the patient, compared to the general population, has a different Ca^{2+} metabolism that is affected by one or more proteins (e.g., hormones) in the blood or extracellular body fluids, or other molecules which affect the level of extracellular and/or intracellular Ca^{2+}. Thus, the diseases include hyperparathyroidism, osteoporosis and other bone and mineral-related disorders, and the like (as described, e.g., in standard medical text books, such as "Harrison's Principles of Internal Medicine"). Such diseases are treated in this invention by molecules which mimic or block one or more of the effects of Ca^{2+} and thereby directly or indirectly affect the levels of the proteins or other molecules in the body of the patient.

By "therapeutically effective amount" is meant an amount that relieves to some extent one or more symptoms of the disease or condition in the patient. Additionally, by "therapeutically effective amount" is meant an amount that returns to normal, either partially or completely, physiological or biochemical parameters associated with or causative of the disease or condition. Generally, it is an amount between about 1 nmole and 1 μmole of the molecule, dependent on its EC_{50} and on the age, size, and disease associated with the patient.

In preferred embodiments, the molecule has an EC_{50} of less than or equal to 5 μM, and is not protamine; and most preferably interacts at a Ca^{2+} receptor as a calcimimetic or calcilytic. Most preferably the molecule is chosen from one of those described above.

In other preferred embodiments, the patient has a disease characterized by an abnormal level of one or more components the level of which is regulated or affected by activity of one or more Ca^{2+} receptors, and the molecule is active on a Ca^{2+} receptor of a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells, and placental throphoblasts.

In still other preferred embodiments, the molecule reduces the level of parathyroid hormone in the serum of the patient, e.g., to that level present in a normal individual, or to a degree sufficient to cause a decrease in plasma Ca^{2+}; and the molecule is provided in an amount sufficient to have a therapeutically relevant effect on the patient.

In a third aspect, the invention features a method for diagnosis of a disease or condition in a patient by identifying the number and/or location (and/or functional integrity) of one or more Ca^{2+} receptors within the patient and comparing that number and/or location (and/or func-

tional integrity) with that observed in normal patients as an indication of the presence of the disease or condition.

In preferred embodiments, the method is an immunoassay in which an antibody to a Ca^{2+} receptor is used to identify the number and/or location and/or functional integrity of the Ca^{2+} receptors, or the assay involves providing a labelled calcimimetic or calcilytic molecule which binds to a Ca^{2+} receptor; and the disease diagnosed is a cancer, e.g., an ectopic tumor of the parathyroid, or a condition characterized by an above normal level in the number of osteoclasts in bone or an increased level of activity of osteoclasts in bone.

In a fourth aspect, the invention features a method for identifying a molecule useful as a therapeutic molecule. The method includes screening a potentially useful molecule for either an ability to mimic the activity of extracellular Ca^{2+} in a cell, or to block an increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}, and determining whether the molecule has an EC_{50} or IC_{50} of less than or equal to 5 μM.

In other aspects, the invention features a recombinant Ca^{2+} receptor, a cell including a recombinant Ca^{2+} receptor, purified nucleic acid encoding a Ca^{2+} receptor, the biological activity and use of the molecule NPS 019, the novel compounds or compositions of matter of NPS 459, NPS 467, NPS 551, and NPS 568 (see Fig. 36) and a method for identifying a useful calcimimetic or calcilytic molecule by identifying a molecule which mimics or blocks one or more activities of Ca^{2+} at a first Ca^{2+} receptor but not at a second Ca^{2+} receptor, e.g., by use of a recombinant Ca^{2+} receptor.

By "recombinant" is meant to include any Ca^{2+} receptor produced by recombinant DNA techniques such that it is distinct from the naturally occurring Ca^{2+} receptor either in its location, purity or structure. Generally, such a receptor will be present in a cell in an amount different from those normally observed in nature.

By "purified" is meant that the antibody or nucleic acid is distinct from naturally occurring antibody or nucleic acid, being separated from antibody or nucleicacid with which it naturally occurs, e.g., in a vector system, such that it can be used to express recombinant Ca^{2+} receptor. Preferably, the antibody or nucleic acid is provided as a homogeneous preparation by standard techniques.

Such cloned receptors can be expressed in a desired cell, and isolated and crystallized to allow structure determination. Such a structure will allow design of useful molecules of this invention which can bind to the Ca^{2+} receptor. In addition, equivalent such receptors can be cloned using a first clone as a probe for clones in other cell, cDNA or genomic libraries.

Antibodies to the cloned receptor can be isolated and used as therapeutics in this invention, or as diagnostic tools for determining Ca^{2+} receptor numbers and/or locations and/or functional integrity to diagnose Ca^{2+}-related diseases or conditions. Such antibodies can also be used in vivo by intravenous administration as calcimimetics or calcilytics.

Thus, in general, the invention features calcimimetic or calcilytic molecules able to act as either selective agonists or antagonists respectively at a Ca^{2+} receptor of one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental throphoblasts. Such a composition may include any pharmaceutically acceptable carrier known to those in the art to provide a pharmaceutical composition.

The invention also features modulation of the the number of Ca^{2+} receptors in a patient by standard techniques, e.g., antisense and related technologies (e.g., ribozymes), as a therapeutic for a disease state.

This invention provides methods for identifying molecules which affect the activity of a Ca^{2+} receptor using assays, as defined below, to detect calcimimetics and/or calcilytics. Further, molecules found to be effective to reduce or enhance expression of Ca^{2+} receptor at a transcriptional or translational level by use of the assays or antibodies or other techniques described below can be defined for therapeutic uses.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof, and from the claims.

Description of the Preferred Embodiments

The drawings will first briefly be described.

Drawings

Fig. 1 depicts representative molecules useful in the invention.

Fig. 2 is a graphical representation showing increases in $[Ca^{2+}]_i$ induced by extracellular Ca^{2+} in quin-2- or fura-2-loaded bovine parathyroid cells. The initial $[Ca^{2+}]$ was 0.5 mM (using $CaCl_2$) and, at each of the arrows, was increased in 0.5 mM increments.

Fig. 3 is a graphical representation showing mobilization of $[Ca^{2+}]_i$ in bovine parathyroid cells. The initial $[Ca^{2+}]$ was 0.5 mM and was decreased to < 1 μM by the addition of EGTA as indicated. (a) Extracellular Mg^{2+} (8 mM, final) elicits an increase in $[Ca^{2+}]_i$ in the absence of extracellular Ca^{2+}. (b) Pretreatment with ionomycin (1 μM) blocks the response to Mg^{2+}. (c) Pretreatment with 5 μM molecule 1799 (a mitochondrial uncoupler) is without effect on the response to Mg^{2+}.

Fig. 4 is a graphical representation showing preferential inhibitory effects of a low concentration of Gd^{3+} on steady-state increases in $[Ca^{2+}]_i$ and that a high concentration of Gd^{3+} elicits a transient increase in $[Ca]_i$. Top panel: Control. Initial concentration of extracellular

Ca^{2+} was 0.5 mM and was increased by 0.5 mM at each of the arrowheads. Middle panel: Gd^{3+} (5 μM) blocks steady-state but not transient increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. Lower panel: Gd^{3+} (50 μM) elicits a transient increase in $[Ca^{2+}]_i$ and abolishes both transient and sustained responses to extracellular Ca^{2+}. In the middle and lower panels, just enough EGTA was added to chelate preferentially Gd^{3+}: the block of Ca^{2+} influx is removed and $[Ca^{2+}]_i$ rises promptly.

Fig. 5 is a graphical representation showing that the effects of PMA on $[Ca^{2+}]_i$, IP_3 formation, and PTH secretion are overcome by increasing concentrations of extracellular Ca^{2+}. For each variable, there is a shift to the right in the concentration-response curve for extracellular Ca^{2+}. Note also that the concentration-response curves vary sigmoidally as $[Ca^{2+}]$ increases linearly.

Fig. 6 is a graphical representation showing that increases in $[Ca^{2+}]_i$ elicited by spermine are progressively depressed by increasing $[Ca^{2+}]$. Spermine (200 μM) was added at the time shown by arrowheads. In this and all subsequent figures, the numbers accompanying the traces are $[Ca^{2+}]_i$ in nM.

Fig. 7 is a graphical representation showing that spermine mobilizes intracellular Ca^{2+} in bovine parathyroid cells. EGTA was added to reduce $[Ca^{2+}]$ to <1 μM before the addition of spermine (200 μM) as indicated (left trace). Pretreatment with ionomycin (1 μM) blocks the response to spermine (right trace).

Figs. 8A and B are graphical representations showing that spermine increases $[Ca^{2+}]_i$ and inhibits PTH secretion in bovine parathyroid cells similarly to extracellular Ca^{2+}. The data points for the spermine dose-concentration response curves are the means of two experiments.

Fig. 9 is a graphical representation showing the contrasting effects of PMA on responses to extracellular Ca^{2+} and on responses to ATPγS in bovine parathyroid cells.

Left panel: The concentration-response curve for extracellular Ca^{2+}-induced inhibition of cyclic AMP formation is shifted to the right by PMA (100 nM). Middle panel: PMA does not affect the ability of ATPγS to increases $[Ca^{2+}]_i$. Note also that the concentration-response curve to ATPγS shows classical sigmoidal behavior as a function of the log concentration, in contrast to extracellular divalent cations.

Fig. 10 is a graphical representation showing mobilization of intracellular Ca^{2+} in human parathyroid cells evoked by extracellular Mg^{2+}. Cells were obtained from an adenoma and bathed in buffer containing 0.5 mM extracellular Ca^{2+}. (a) Transient and sustained increases in $[Ca^{2+}]_i$ elicited by extracellular Mg^{2+} (10 mM, final) shows that sustained increases are not affected by nimodipine (1 μM) but are depressed by La^{3+} (1 μM) and return promptly when La^{3+} is selectively chelated by a low concentration of EGTA. (b) La^{3+} (1 μM) blocks the sustained but not the transient increase in $[Ca^{2+}]_i$ elicited by extracellular Mg^{2+}. (c) Cytosolic Ca^{2+} transients elicited by extracellular Mg^{2+} persist in the absence of extracellular Ca^{2+}.

Fig. 11 is a graphical representation showing mobilization of intracellular Ca^{2+} evoked by neomycin or protamine in bovine parathyroid cells. In all traces, the initial $[Ca^{2+}]$ and $[Mg^{2+}]$ was 0.5 and 1 mM, respectively. In trace (a) and (b), the Ca^{2+} and Mg^{2+} concentrations were increased to 2 and 8 mM, from 0.5 and 1mM respectively. In the other traces, (c) through (i), neomycin B (30 μM) or protamine (1 ug/ml) were added as indicated. La^{3+} (1 μM) EGTA (1 mM), or ionomycin (100 nM) were added as indicated. Each trace is representative of the pattern seen in 5 or more trials using at least 3 different cell preparations. Bar = 1 min.

Fig. 12 is a graphical representation showing that neomycin B blocks transient but does not block steady-state increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. Left control: $[Ca^{2+}]$ was initially 0.5 mM and was

increased in 0.5 mM increments at each of the open arrowheads before the addition of neomycin B (30 μM). Right: Neomycin B (30 μM) was added before $[Ca^{2+}]$. Bar = 1 min.

Fig. 13 is a graphical representation showing that neomycin B or protamine inhibit PTH secretion at concentrations which evoked increases in $[Ca^{2+}]_i$. Cells were incubated with the indicated concentrations of organic polycation for 30 min. in the presence of 0.5 mM extracellular Ca^{2+}. Open symbols: control responses for PTH secretion in the presence of 0.5 (circles) or 2 mM (diamonds) extracellular Ca^{2+}. Values for $[Ca^{2+}]_i$ are diamond symbols. Bovine cells were used in the experiments with protamine and human (adenoma) parathyroid cells were used in the experiments with neomycin B. Each point is the mean $\pm$ SEM of 3 experiments.

Fig. 14 is a graphical representation showing the preferential inhibitory effects of PMA on cytosolic Ca^{2+} transients elicited by spermine. Initial $[Ca^{2+}]$ was 0.5 mM; spermine (200 μM) or ATP (50 μM) were added as indicated. Bar = 1 min.

Fig. 15 is a graphical representation showing that PMA shifts to the right the concentration-response curves for extracellular Ca^{2+}- and neomycin B-induced increases in $[Ca^{2+}]_i$. Cells were pretreated with PMA for 1 min. before increasing $[Ca^{2+}]$ or before adding neomycin B as indicated. Each point is the mean $\pm$ SEM of 3 to 5 experiments.

Fig. 16 is a graphical representation showing that PMA shifts to the right the concentration-response curves for extracellular Ca^{2+}- and spermine-induced inhibition of PTH secretion. Cells were incubated with the indicated $[Ca^{2+}]$ and spermine for 30 min. in the presence (closed circles) or absence (open circles) of 100 nM PMA. Each point is the mean $\pm$ SEM of 3 experiments.

Fig. 17 is a graphical representation showing that protamine increases the formation of inositol phosphates. Parathyroid cells were incubated overnight in culture media containing 4 uCi/ml ^{3}H-myo-inositol, washed, and

incubated with the indicated concentration of protamine at 37°. After 30 sec. the reaction was terminated by the addition of $CHCl_3$:MeOH:HCl and IP_1 (circles) and IP_3 (triangles) separated by anion exchange chromatography. Each point is the mean of 2 experiments, each performed in triplicate.

Fig. 18 is a graphical representation showing that PMA depresses the formation of IP_1 evoked by extracellular Ca^{2+} or spermine. ^3H-myo-insoitol-labeled cells were exposed to the indicated [Ca^{2+}] or spermine for 30 sec. before terminating the reaction and determining IP_1 by anion exchange chromatography. Hatched columns: Cells were pretreated with PMA (100 nM) for 5 min. before increasing [Ca^{2+}] or adding spermine. Each value is the mean of 2 experiments, each performed in triplicate.

Fig. 19 is a graphical representation showing transient and sustained increases in $[Ca^{2+}]_i$ elicited by neomycin B in human (adenoma) parathyroid cells. [Ca^{2+}] was 0.5 mM. (a) The sustained increase in $[Ca^{2+}]_i$ elicited by neomycin B (10 μM) was depressed by La^{3+}. (b) The transient increase in $[Ca^{2+}]_i$ evoked by neomycin B was unaffected by La^{3+}. (c) Transient increases in $[Ca^{2+}]_i$ persisted in the absence of extracellular Ca^{2+}.

Fig. 20 is a graphical representation showing that neomycin B evokes oscillating increases the Cl^- conductance in Xenopus oocytes expressing the Ca^{2+} receptor. Upper trace from an oocyte three days after injection with human (hyperplastic) parathyroid cell poly(A)$^+$-mRNA. Lower trace from an oocyte injected with water. Neomycin B failed to elicit a response in five water-injected oocytes and carbachol elicited a response in one, which is shown. In both traces, the holding potential was -76 mV.

Fig. 21 is a graphical representation showing that neomycin B fails to affect basal or evoked increases in C-cells. Control, left trace: Fura-2-loaded rMTC 6-23 cells were initially bathed in buffer containing 1 mM Ca^{2+}

before increasing [Ca^{2+}] to 3 mM. Right trace: pretreatment with 5 mM neomycin B.

Fig. 22 is a graphical representation showing that extracellular Ca^{2+} evokes increases in $[Ca^{2+}]_i$ in rat osteoclasts. Microfluorimetric recording in a single rat osteoclast loaded with indo-1 and superfused for the indicated times (bars) with buffer containing the indicated [Ca^{2+}]. Normal buffer, superfused between the bars, contained 1 mM Ca^{2+}.

Fig. 23 is a graphical representation showing that spermine or neomycin B fail to evoke increases in $[Ca^{2+}]_i$ in rat osteoclasts. An indo-1-loaded osteoclast was superfused with the indicated concentration of spermine or neomycin B (open bars) alone or together with 20 mM Ca^{2+} (solid bars).

Fig. 24 is a graphical representation showing the differential effects of argiotoxin (shown as argiopine in the figure, structures also shown in Fig. 1). 659 and argiotoxin 636 on $[Ca^{2+}]_i$ in bovine parathyroid cells. The initial [Ca^{2+}] was 0.5 mM and was increased to 1.5 mM where indicated (right trace). Where indicated, argiotoxin 659 (300 μM) or argiotoxin 636 (400 μM) was added.

Fig. 25 is a graphical representation showing that extracellular Mg^{2+} or Gd^{3+} evoke oscillatory increases in Cl^- conductance in Xenopus oocytes injected with bovine parathyroid cell poly(A)$^+$-mRNA. In trace (a), the concentration of extracellular Ca^{2+} was < 1 μM and in trace (b), 0.7 mM. Trace (c) shows that extracellular Mg^{2+} fails to elicit a response in an oocyte injected only with the mRNA for the substance K receptor, although superfusion with substance K evokes a response. Holding potential was -70 to -80 mV.

Fig. 26 is a graphical representation showing that extracellular Ca^{2+} elicits oscillatory increases in Cl^- conductance in Xenopus oocytes injected with human (hyperplastic) parathyroid tissue poly(A)$^+$-mRNA. The oocyte was tested for responsivity to extracellular Ca^{2+} three days

after injection of 50 ng poly(A)$^+$-mRNA. Holding potential was -80 mV.

Fig. 27 is a graphical representation showing the mobilization of intracellular Ca^{2+} in bovine parathyroid cells elicited by budmunchiamine. Budmunchiamine (300 μM, structure also shown) was added where indicated.

Fig. 28 is a graphical representation showing that the ability to mobilize intracellular Ca^{2+} in parathyroid cells is stereospecific. Bovine parathyroid cells loaded with fura-2 were initially suspended in buffer containing 0.5 mM extracellular Ca^{2+} before the addition of the indicated concentration of each molecule.

Fig. 29 is a graphical representation showing effects of La^{3+} on $[Ca^{2+}]_i$ in osteoclasts. A representative trace from a single rat osteoclast loaded with indo-1 is shown. At low concentrations, La^{3+} partially blocks increases in $[Ca^{2+}]_i$, elicited by extracellular Ca^{2+}.

Figs. 30A and B are graphical representations showing the mobilization of intracellular Ca^{2+} elicited by extracellular Mn^{2+} in rat osteoclasts. Extracellular Mn^{2+} evokes concentration-dependent increases in $[Ca^{2+}]_i$ (Fig. 30A) that persist in the absence of extracellular Ca^{2+} (Fig. 30B).

Figs. 31A and 31B are graphical representations showing mobilization of $[Ca^{2+}]_i$ in rat osteoclasts elicited by a molecule termed NPS 449 (see Fig. 38). Isolated rat osteoclasts loaded with indo-1 were superfused with the indicated concentrations of NPS 449 in the presence (Fig. 31A) or absence (Fig. 31B) of 1 mM extracellular $CaCl_2$.

Fig. 32 is a graphical representation showing the mobilization of intracellular Ca^{2+} in C-cells evoked by NPS 019 (see Fig. 1). rMTC 6-23 cells were loaded with fura-2 and bathed in buffer containing 0.5 mM $[Ca^{2+}]$. Where indicated, NPS 019 was added to a final concentration of 10 μM. Representative traces show that the transient increase in $[Ca^{2+}]_i$ elicited by NPS 019 is refractory to

inhibition by La^{3+} (middle trace) and persists in the absence of extracellular Ca^{2+} (right trace).

Fig. 33 is a graphical representation showing that NPS 456 (Fig. 36) evokes oscillatory increases in Cl^- current in Xenopus oocytes which have been injected with bovine parathyroid cell poly(A)$^+$-mRNA.

Fig. 34 is a graphical representation showing that extracellular Ca^{2+} evokes oscillatory increases in Cl^- current in Xenopus oocytes which have been injected with human osteoclast mRNA. The oocyte was tested for responsivity to extracellular Ca^{2+} three days after injection of 50 ng of total poly(A)$^+$ mRNA.

Fig. 35 is a graphical representation showing that the parathyroid cell Ca^{2+} receptor is encoded by mRNA in a size range of 2.5-3.5 kb. Bovine parathyroid cell poly(A)$^+$-mRNA was size fractionated on denaturing glycerol gradients and pooled into ten fractions. Each fraction was injected (50 ng/fraction) separately into Xenopus oocytes. After three days, the oocytes were examined for their ability to respond to extracellular Ca^{2+} with oscillatory increases in the Cl^- conductance.

Fig. 36 shows the chemical structures of molecules derived from diphenylpropyl-α-phenethylamine illustrating a family of molecules which were prepared and screened to find the useful molecules of the invention.

Fig. 37 is a graphical representation showing that NPS 021 is a calcilytic compound that blocks the effects of extracellular Ca^{2+} on $[Ca^{2+}]_i$ in bovine parathyroid cells. Cells were initially bathed in buffer containing 0.5 mM $CaCl_2$ and, where indicated, the $[Ca^{2+}]$ was increased to a final of 2mM (left trace). The addition of NPS 021 (200 μM) caused no change in $[Ca^{2+}]_i$ but inhibited the increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+} (right trace).

Fig. 38 is a graph showing in vivo Ca^{2+} response to NPS 467.

Fig. 39 is a graph showing in vivo PTH response to NPS 467.

Fig. 40 is a graph showing in vivo Ca^{2+} response to 25 mg/kg NPS 467.

Figs. 41 and 42 are graphs showing in vivo Ca^{2+} responses to different enantiomers of NPS 467.

Fig. 43a depicts a reaction scheme for the preparation of fendiline or fendiline analogs or derivatives depicted in Figure 36. Fig. 43b depicts a reaction scheme for the synthesis of NPS 467.

Fig. 44 depicts a dose response curve showing that NPS 467 lowers serum ionized calcium when administered orally.

Calcimimetic and Calcilytic Molecules

Calcimimetic and calcilytic molecules useful in the invention are generally described above. These molecules can be readily identified using screening procedures to define molecules which mimic or antagonize the activity of Ca^{2+} at Ca^{2+} receptors. Examples of such procedures are provided below. These examples are not limiting in the invention but merely illustrate methods which are readily used or adapted by those skilled in the art.

Generally, calcimimetic and calcilytic molecules are identified by screening molecules which are modelled after those described below (called lead molecules). As can be seen below there are several specific calcimimetics and calcilytics useful at various Ca^{2+} receptors. Derivative molecules are readily designed by standard procedures and tested in one of many protocols known to those skilled in the art. Many molecules may be screened easily to identify the most useful in this invention.

Organic cationic molecules which mimic or antagonize the actions of Ca^{2+} in other systems contain the requisite structure for activity on a Ca^{2+} receptor. Rational design of other useful molecules involves the study of a molecule known to be calcimimetic or calcilytic and then modifying

the strucure of the known molecule. For example, polyamines are potentially calcimimetic since spermine mimics the action of Ca^{2+} in several in vitro systems. Results show that spermine does indeed cause changes in $[Ca^{2+}]_i$ and PTH secretion reminiscent of those elicited by extracellular di- and trivalent cations (see below). The experiments outlined below are therefore aimed at demonstrating that this phenomenology, obtained with spermine, involves the same mechanisms used by extracellular Ca^{2+}. To do this, the effects of spermine on a variety of physiological and biochemical parameters which characterize activation of the Ca^{2+} receptor were assessed. Those molecules having similar effects are useful in this invention and can be discovered by selecting or making molecules having a structure similar to spermine. Once another useful molecule is discovered this selection process can be readily repeated.

For clarity, below is provided a specific series of screening protocols to identify such useful molecules which are active at a parathyroid cell Ca^{2+} receptor, or which act as agonists or antagonists of the cellular response to changes in $[Ca^{2+}]$. Equivalent assays can be used for molecules active at other Ca^{2+} receptors, or which otherwise mimic or antagonize cellular functions regulated by $[Ca^{2+}]$. These assays exemplify the procedures which are useful to find calcimimetic molecules of this invention. Equivalent procedures can be used to find calcilytic molecules by screening for those molecules most antagonistic to the actions of extracellular Ca^{2+}. In vitro assays can be used to characterize the selectivity, saturability, and reversibility of these calcimimetics and calcilytics by standard techniques.

Screening Procedure

Generally, bovine parathyroid cells loaded with fura-2 are initially suspended in buffer containing 0.5 mM $CaCl_2$. The test substance is added to the cuvette in a

small volume (5-15 μl) and any change in the fluorescence signal noted. Cumulative increases in the concentration of the test substance are made in the cuvette until some predetermined concentration is achieved or changes in fluorescence noted. If no changes in fluorescence are noted, the molecule is considered inactive and no further testing is performed. In initial studies, e.g., with polyamine-type molecules, molecules were tested at concentrations as high as 5 or 10 mM. As more potent molecules are now known (see below), the ceiling concentration is lowered. For example, newer molecules are tested at concentrations up to 500 μM or less. If no changes in fluorescence are noted at this concentration, the molecule can be considered inactive.

Molecules causing increases in $[Ca^{2+}]_i$ are subjected to additional testing. The two essential characteristics of the molecule important for its consideration as a calcimimetic molecule are the mobilization of intracellular Ca^{2+} and sensitivity to PKC activators. Molecules causing the mobilization of intracellular Ca^{2+} in a PMA-sensitive manner have invariably been found to be calcimimetic molecules and to inhibit PTH secretion. Additional testing can, if needed, be performed to solidify this belief. Typically, all the various tests for calcimimetic or calcilytic activity (see above) are not performed. Rather, if a molecule causes the mobilization of intracellular Ca^{2+} in a PMA-sensitive manner, it is advanced to screening on human parathyroid cells. For example, measurements of $[Ca^{2+}]_i$ are performed to determine the EC_{50}, and to measure the ability of the molecule to inhibit PTH secretion in human parathyroid cells which have been obtained from patients undergoing surgery for primary or secondary hyperparathyroidism. The lower the EC_{50} or IC_{50} the more potent the molecule as a calcimimetic or calcilytic.

Measuring $[Ca^{2+}]_i$ with fura-2 provides a very rapid means of screening new organic molecules for activity. In

a single afternoon, 10-15 molecules can be examined and their ability to mobilize intracellular Ca^{2+} (or not) assessed. The sensitivity of any observed increase in $[Ca^{2+}]_i$ to depression by PMA can also be assessed. Moreover, a single cell preparation can provide data on $[Ca^{2+}]_i$, cyclic AMP levels, IP_3 and PTH secretion. A typical procedure is to load cells with fura-2 and then split the cell suspension in two; most of the cells are used for measurement of $[Ca^{2+}]_i$ and the remainder are incubated with molecules to assess their effects on cyclic AMP and PTH secretion. Because of the sensitivity of the radioimmunoassays for cyclic AMP and PTH, both variables can be determined in a single incubation tube containing 0.3 ml cell suspension (about 500,000 cells). Measurements of inositol phosphates are a time-consuming aspect of the screening. However, ion exchange columns eluted with chloride (rather than formate) provide a very rapid means of screening for IP_3 formation since rotary evaporation (which takes around 30 hrs) is not required. This method allows processing of nearly 100 samples in a single afternoon. Those molecules that prove interesting, as assessed by measurements of $[Ca^{2+}]_i$, cyclic AMP, IP_3, and PTH are then subjected to a more rigorous analysis by examining formation of various inositol phosphates and assessing their isomeric form by HPLC.

Interesting molecules detected in these protocols are then assessed for specificity, e.g., by examining their effects on $[Ca^{2+}]_i$ in calcitonin-secreting C-cells using, e.g., the rat MTC 6-23 cell line.

The following is illustrative of methods useful in these screening procedures. Examples of typical results for various test calcimimetic or calcilytic molecules are provided in Figs. 2-34.

Parathyroid Cell Preparation

Parathyroid glands were obtained from freshly slaughtered calves (12-15 weeks old) at a local abattoir and

transported to the laboratory in ice-cold parathyroid cell buffer (PCB) which contains (mM): NaCl, 126; KCl, 4; $MgCl_2$, 1; Na-HEPES, 20; pH 7.4; glucose, 5.6, and variable amounts of $CaCl_2$, e.g., 1.25 mM. Human parathyroid glands, obtained from patients undergoing surgical removal of parathyroid tissue for primary or uremic hyperparathyroidism (HPT), were treated similarly to bovine tissue. Glands were trimmed of excess fat and connective tissue and then minced with a fine scissors into approximate cubes of 2-3 mm. Dissociated parathyroid cells were prepared by collagenase digestion. Dissociated cells were then purified by centrifugation in Percoll buffer. The resultant parathyroid cell preparation was essentially devoid of red blood cells, adipocytes, and capillary tissue as assessed by phase contrast microscopy and Sudan black B staining. Dissociated and purified parathyroid cells were present as small clusters containing 5 to 20 cells. Cellular viability, as indexed by exclusion of trypan blue or ethidium bromide, was routinely 95%.

Although cells can be used for experimental purposes at this point, physiological responses (suppressibility of PTH secretion and resting levels of $[Ca^{2+}]_i$) are better after culturing the cells overnight. Primary culture also has the advantage that cells can be labeled with isotopes to near isotopic equilibrium, as is necessary for studies involving measurements of inositol phosphate metabolism (see below). After purification on Percoll gradients, cells were washed several times in a 1:1 mixture of Ham's F12-Dulbecco's modified Eagles medium (GIBCO) supplemented with 50 ug/ml streptomycin, 100 U/ml penicillin, 5 ug/ml gentamicin and ITS^+. ITS^+ is a premixed solution containing insulin, transferrin, selenium, and bovine serum albumin (BSA)-linolenic acid (Collaborative Research, Bedford, MA). The cells were then transferred to plastic flasks (75 or 150 cm^2; Falcon) and incubated at 37°C in a humid atmosphere of 5% CO_2. No serum is added to these overnight cultures, since its presence allows the cells to attach to

the plastic, undergo proliferation, and dedifferentiate. Cells cultured under the above conditions were readily removed from the flasks by decanting, and show the same viability as freshly prepared cells.

Measurement of Cytosolic Ca^{2+}

Purified parathyroid cells were resuspended in 1.25 mM $CaCl_2$-2% BSA-PCB containing 1 μM fura-2-acetoxymethylester and incubated at 37°C for 20 min. The cells were then pelleted, resuspended in the same buffer lacking the ester, and incubated a further 15 min at 37°C. The cells were subsequently washed twice with PCB containing 0.5 mM $CaCl_2$ and 0.5% BSA and maintained at room temperature (about 20°C). Immediately before use, the cells were diluted five-fold with prewarmed 0.5 mM $CaCl_2$-PCB to obtain a final BSA concentration of 0.1%. The concentration of cells in the cuvette used for fluorescence recording was 1-2 x 10^6/ml.

The fluorescence of indicator-loaded cells was measured at 37°C in a spectrofluorimeter (Biomedical Instrumentation Group, University of Pennsylvania, Philadelphia, PA) equipped with a thermostated cuvette holder and magnetic stirrer using excitation and emission wavelengths of 340 and 510 nm, respectively. This fluorescence indicates the level of cytosolic Ca^{2+}. Fluorescence signals were calibrated using digitonin (50 ug/ml, final) to obtain maximum fluorescence (F_{max}), and EGTA (10 mM, pH 8.3, final) to obtain minimal fluorescence (F_{min}), and a dissociation constant of 224 nM. Leakage of dye is dependent on temperature and most occurs within the first 2 min after warming the cells in the cuvette; dye leakage increases only very slowly thereafter. To correct the calibration for dye leakage, cells were placed in the cuvette and stirred at 37°C for 2-3 min. The cell suspension was then removed, the cells pelleted, and the supernatant returned to a clean cuvette. The supernatant was then treated with digitonin and EGTA as above to obtain as estimate of dye

leakage, which is typically 10-15% of the total Ca^{2+}-dependent fluorescent signal. This estimate was subtracted from the apparent F_{min}.

Measurement of PTH Secretion

In most experiments, cells loaded with fura-2 were used in studies of PTH secretion. Loading parathyroid cells with fura-2 does not change their secretory response to extracellular Ca^{2+}. Cells were suspended in PCB containing 0.5 mM $CaCl_2$ and 0.1% BSA. Incubations were performed in plastic tubes (Falcon 2058) containing 0.3 ml of the cell suspension with or without small volumes of $CaCl_2$ and/or organic polycations. After incubation at 37°C for various times (typically 30 min), the tubes were placed on ice and the cells pelleted at 2°C. Samples of the supernatant were brought to pH 4.5 with acetic acid and stored at -70°C. This protocol was used for both bovine and human parathyroid cells.

For bovine cells, the amount of PTH in sample supernatants was determined by a homologous radioimmunoassay using GW-1 antibody or its equivalent at a final dilution of 1/45,000. ^{125}I-PTH (65-84; INCSTAR, Stillwater, MN) was used as tracer and fractions separated by dextran-activated charcoal. Counting of samples and data reduction were performed on a Packard Cobra 5005 gamma counter.

For human cells, a commercially available radioimmunoassay kit (INS-PTH; Nichols Institute, Los Angeles, CA) which recognizes intact and N-terminal human PTH was used because GW-1 antibody recognizes human PTH poorly.

Measurement of cyclic AMP

Cells were incubated as above for PTH secretion studies and at the end of incubation, a 0.15 ml sample was taken and transferred to 0.85 ml hot (70°C) water and heated at this temperature for 5-10 min. The tubes were subsequently frozen and thawed several times and the cellular debris sedimented by centrifugation. Portions of

the supernatant were acetylated and cyclic AMP concentrations determined by radioimmunoassay.

Measurement of Inositol Phosphate Formation

Membrane phospholipids were labeled by incubating parathyroid cells with 4 μCi/ml ^{3}H-*myo*-inositol for 20-24 hrs. Cells were then washed and resuspended in PCB containing 0.5 mM $CaCl_2$ and 0.1% BSA. Incubations were performed in microfuge tubes in the absence or presence of various concentrations of organic polycation for different times. Reactions were terminated by the addition of 1 ml chloroform/methanol/12 N HCl (200:100:1; v/v/v). Phytic acid hydrolysate (200 μl; 25 μg phosphate/tube) water was then added. The tubes were centrifuged and 600 μl of the aqueous phase was diluted into 10 ml water.

Inositol phosphates were separated by ion exchange chromatography using AG1-X8 in either the chloride- or formate-form. When only IP_3 levels were to be determined, the chloride-form was used, whereas the formate form was used to resolve the major inositol phosphates (IP_3, IP_2, and IP_1). For determination of just IP_3, the diluted sample was applied to the chloride-form column and the column washed with 10 ml 30 mM HCl followed by 6 ml 90 mM HCl and the IP_3 eluted with 3 ml 500 mM HCl. The last eluate was diluted and counted. For determination of all major inositol phosphates, the diluted sample was applied to the formate-form column and IP_1, IP_2, and IP_3 eluted sequentially by increasing concentrations of formate buffer. The eluted samples from the formate columns were rotary evaporated, the residues brought up in cocktail, and counted.

The isomeric forms of IP_3 were evaluated by HPLC. The reactions were terminated by the addition of 1 ml 0.45 M perchloric acid and stored on ice for 10 min. Following centrifugation, the supernatant was adjusted to pH 7-8 with $NaHCO_3$. The extract was then applied to a Partisil SAX anion-exchange column and eluted with a linear gra-

dient of ammonium formate. The various fractions were then desalted with Dowex followed by rotary evaporation prior to liquid scintillation counting in a Packard Tricarb 1500 LSC.

For all inositol phosphate separation methods, appropriate controls using authentic standards were used to determine if organic polycations interfered with the separation. If so, the samples were treated with cation-exchange resin to remove the offending molecule prior to separation of inositol phosphates.

Measurement of Cytosolic Ca^{2+} in C-cells

Neoplastic C-cells derived from a rat medullary thyroid carcinoma (rMTC 6-23) obtained from American Type Culture Collection (ATCC No. 1607) were cultured as monolayers in Dulbecco's Modified Eagle's medium (DMEM) plus 15% horse serum in the absence of antibiotics. For measurements of $[Ca^{2+}]_i$, the cells were harvested with 0.02% EDTA/0.05% trypsin, washed twice with PCB containing 1.25 mM $CaCl_2$ and 0.5% BSA, and loaded with fura-2 as described above for parathyroid cells. Measurements of $[Ca^{2+}]_i$ were performed as described above with appropriate corrections for dye leakage.

Measurement of $[Ca^{2+}]_i$ in Rat Osteoclasts

Osteoclasts were obtained from 1-2 day old Sprague-Dawley rats using aseptic conditions. The rat pups were sacrificed by decapitation, the hind legs removed, and the femora rapidly freed of soft tissue and placed in pre-warmed F-12/DMEM media (DMEM containing 10% fetal calf serum and antibiotics (penicillin-streptomycin-gentamicin; 100 U/ml-100 ug/ml-100 ug/ml)). The bones from two pups were cut lengthwise and placed in 1 ml culture medium. Bone cells were obtained by gentle trituration of the bone fragments with a plastic pipet and diluted with culture medium. The bone fragments were allowed to settle and equal portions (about 1 ml) of the medium transferred to

a 6 well culture plate containing 25 mm glass coverslips. The cells were allowed to settle for 1 hr at 37°C in a humidified 5% CO_2-air atmosphere. The coverslips were then washed 3 times with fresh media to remove nonadherent cells. Measurements of $[Ca^{2+}]_i$ in osteoclasts were performed within 6-8 hrs of removing nonadherent cells.

Cells attached to the coverslip were loaded with indo-1 by incubation with 5 μM indo-1 acetoxymethylester / 0.01% Pluronic F28 for 30 min at 37°C in F-12/DMEM lacking serum and containing instead 0.5% BSA. The coverslips were subsequently washed and incubated an additional 15 min at 37°C in F-12/DMEM lacking ester before being transferred to a superfusion chamber mounted on the stage of a Nikon Diaphot inverted microscope equipped for microfluorimetry. Osteoclasts were easily identified by their large size and presence of multiple nuclei. The cells were superfused with buffer (typically PCB containing 0.1% BSA and 1 mM Ca^{2+}) at 1 ml/min with or without test substance. The fluorescence emitted by excitation at 340 nm was directed through the video port of the microscope onto a 440 nm dichroic mirror and fluorescence intensity at 495 and 405 nm collected by photomultiplier tubes. The outputs from the photomultiplier tubes were amplified, digitized, and stored in an 80386 PC. Ratios of fluorescence intensity were used to estimate $[Ca^{2+}]_i$.

Oocyte Expression

In additional studies, Xenopus oocytes injected with mRNA from bovine or human parathyroid cells were used in screening protocols, and Cl^- conductance measured as an indirect means of monitoring increases in $[Ca^{2+}]_i$. The following is an example to test the effect of neomycin.

Oocytes were injected with poly(A)$^+$-enriched mRNA from human parathyroid tissue (hyperplastic glands from a case of secondary HPT). After 3 days, the oocytes were tested for their response to neomycin. Neomycin B evoked oscillatory increases in the Cl^- conductance which ceased upon

superfusion with drug-free saline (see Fig. 20). Responses to neomycin B were observed at concentrations between 100 μM and 10 mM. To ensure that the response evoked by neomycin B was contingent upon injection of parathyroid mRNA, the effect of neomycin B on currents in water-injected oocytes was determined. In each of five oocytes examined, neomycin B (10 mM) failed to cause any change in the current. About 40% of oocytes are known to respond to carbachol, an effect mediated by an endogenous muscarinic receptor. In five oocytes examined, one showed inward currents in response to carbachol, and this is shown in the lower trace of Fig. 20. Thus, in cells expressing a muscarinic receptor coupled to increases in $[Ca^{2+}]_i$ and Cl^- conductance, neomycin B fails to evoke a response. This shows that the response to neomycin B depends on expression of a specific protein encoded by parathyroid cell mRNA. It suggests quite strongly that in intact cells, neomycin B acts directly on the Ca^{2+} receptor to alter parathyroid cell function.

Drug Design From Lead Molecules

Certain organic molecules mimic or antagonize the action of extracellular Ca^{2+} by acting at the Ca^{2+} receptor as shown herein. The molecules tested, however, are not necessarily suitable as drug candidates, but they serve to demonstrate that the hypothesis underlying Ca^{2+} receptor-based therapies is correct. These molecules can be used to determine the structural features that enable them to act on the Ca^{2+} receptor, and thus to select molecules useful in this invention.

An example of one such analytical procedure follows: This example is detailed in the examples below, but is used here to demonstrate the rationale that can be used to design useful molecules of this invention from lead molecules discussed herein. Those in the art will recognize the analytic steps defined in the example and that analogous analysis can be conducted on other lead molecules

until the most desired calcimimetic or calcilytic is defined.

Other examples are also provided below. Together the data presented demonstrate that useful lead molecules will have aromatic groups which are preferably substituted at one or more positions, and may have branched or linear substitued or unsubstituted alkyl groups as desired. In addition, it is important to choose molecules of correct stereospecificity to ensure higher affinity for the desired Ca^{2+} receptor. These data thus point those in the art to appropriate lead molecules which can be derivatised to find optimum desired molecules of this invention, much as described below.

Although structurally diverse, molecules that are tested may have common features that can be studied. In this example, the correlation between net positive charge and potency in mobilizing intracellular Ca^{2+} was tested. Protamine (+21; EC_{50} = 40 nM) was more effective than neomycin B (+6; EC_{50} = 20 μM in human parathyroid cells and 40 μM in bovine parathyroid cells) which was more effective than spermine (+4; EC_{50} = 150 μM) in causing the mobilization of $[Ca^{2+}]_i$ in parathyroid cells. These results raise the question of whether positive charge alone determines potency, or if there are other structural features that contribute to activity on the Ca^{2+} receptor. This is important to determine at the outset because it profoundly impacts on the view that the Ca^{2+} receptor can be targeted with effective and specific therapeutic molecules. Thus, a variety of other organic polycations related to neomycin B and spermine can be studied to determine the relationship between the net positive charge of a molecule and its potency to mobilize intracellular Ca^{2+}.

The first series of molecules studied were the aminoglycosides. The molecules were examined on bovine parathyroid cells and their EC_{50}'s for the mobilization of intracellular Ca^{2+} determined. For the aminoglycosides, the rank order of potency for eliciting cytosolic Ca^{2+}

transients was neomycin B (EC_{50} = 20 or 40 μM) > gentamicin (150 μM) > bekanamycin (200 μM) > streptomycin (600 μM). Kanamycin and lincomycin were without effect when tested at a concentration of 500 μM. The net positive charge on these aminoglycosides at pH 7.3 is neomycin B (+6) > gentamicin (+5) = bekanamycin (+5) > kanamycin (average +4.5) > streptomycin (+3) > lincomycin (+1). Within the aminoglycoside series, then, there is some correlation between net positive charge but it is not absolute, and kanamycin, which would be predicted to be more potent than streptomycin, is without activity.

Testing of various polyamines revealed additional and more marked discrepancies between net positive charge and potency. Three structural classes of polyamines were examined: (1) straight chain, (2) branched chain, and (3) cyclic. The structures of the polyamines tested are provided in Fig. 1. Amongst the straight chain polyamines, spermine (+4; EC_{50} = 150 μM) was more potent than pentaethylenehexamine (+6; EC_{50} = 500 μM) and tetraethylenepentamine (+5; EC_{50} = 2.5 mM) even though the latter molecules have a greater net positive charge.

We synthesized some branched chain polyamines that have different numbers of secondary and primary amino groups and thus vary in net positive charge. Two of these molecules, NPS 381 and NPS 382, were examined for effects on $[Ca^{2+}]_i$ in bovine parathyroid cells. NPS 382 (+8; EC_{50} = 50 μM) was about twice as potent as NPS 381 (+10; EC_{50} = 100 μM) even though it contains two fewer positive charges.

A similar discrepancy between positive charge and potency was noted in experiments with cyclic polyamines. For example, hexacyclen (+6; EC_{50} = 20 μM) was more potent than NPS 383 (+8; EC_{50} = 150 μM). The results obtained with these polyamines show that positive charge is not the sole factor contributing to potency.

Additional studies provided insights into the structural features of molecules that impart activity on the

parathyroid cell Ca^{2+} receptor. One of the structurally important features is the intramolecular distance between the nitrogens (which carry the positive charge). Thus, spermine is 50-fold more potent than triethylenetetramine (EC_{50} = 8 mM) in evoking increases in $[Ca^{2+}]_i$ in bovine parathyroid cells yet both molecules carry a net positive charge of +4. The only difference in structure between these two polyamines is the number of methylenes separating the nitrogens: in spermine it is 3-4-3 whereas in triethylenetetramine it is 2-2-2. This seemingly minor change in the spacing between nitrogens has profound implications for potency and suggests that the conformational relationships of nitrogens within the molecule are critical. Supporting this are results obtained with hexacyclen and pentaethylenehexamine. The former molecule is simply the cyclic analog of the latter and contains the same number of methylenes between all nitrogens, yet the presence of the ring structure increases potency 25-fold. These results indicate that positive charge per se is not the critical factor determining the activity of an organic molecule on the Ca^{2+} receptor.

Another series of experiments reveals the importance of aromatic groups in determining activity on the Ca^{2+} receptor. The results were obtained with two arylalkylamines isolated from the venom of the spider Argiope lobata. These molecules, argiotoxin 636 and argiotoxin 659, have identical polycationic portions linked to different aromatic groups (Fig. 24). Argiotoxin 659 evoked transient increases in $[Ca^{2+}]_i$ in bovine parathyroid cells when tested at concentrations of 100 to 300 μM. In contrast, argiotoxin 636 was without effect when tested at similar concentrations (Fig. 24). The only difference in structure between these two arylalkylamines is in the aromatic portion of the molecules: argiotoxin 659 contains a 4-hydroxyindole moiety whereas argiotoxin 636 contains a 2,4-dihydroxyphenyl group. The net positive charge on these two arylalkylamines is the same (+4), so their dif-

ferent potencies must result from the different aromatic groups. This shows that net positive charge alone does not determine potency. The real importance of these findings, however, is the discovery that aromatic groups contribute significantly to the ability of molecules to activate the Ca^{2+} receptor.

Agatoxin 489 (NPS 017) and Agatoxin 505 (NPS 015) both cause the mobilization of intracellular Ca^{2+} in parathyroid cells with EC_{50}'s of 6 and 22 μM, respectively. The only difference in the structure of these molecules is a hydroxyl group on the indole moiety (Fig. 1). This shows that substitutions on the aromatic region of the molecule can influence potency. This indicates that further lead molecules to be studied will include those molecules having substituted aromatic moieties.

The structural features to be varied systematically from lead molecules described herein include (1) net positive charge, (2) number of methylenes separating nitrogens, and (3) cyclic versions of, e.g., polyamines, with and without changes in methylene spacing and net positive charge. In addition systematic variations in the structure and location of aromatic groups can be examined, e.g., in a variety of arylalkylamines isolated from the venoms of wasps and spiders; and synthetic molecules can be prepared by the coupling of commercially available aromatic moieties to the argiotoxin polyamine moiety. The argiotoxin polyamine moiety can be readily coupled to any aromatic moiety containing a carboxylic acid. Thus, it is simple to systematically screen the hydroxy and methoxy derivatives of phenylacetic acid and benzoic acid as well as the hydroxyindoleacetic acid series. Analogs containing heteroaromatic functionalities can also be prepared and assessed for activity.

Comparisons of potency and efficacy among such molecules will reveal the optimal structure and location of the aromatic group at a constant positive charge.

One of the structural variations on the polyamine motif that seems to increase potency is the presence of the cyclic version of the straight chain-parent molecule. Budmunchiamine A, isolated from the plant Albizia amara, is a cyclic derivative of spermine (Fig. 1). The addition of budmunchiamine A to bovine parathyroid cells caused a rapid and transient increase in $[Ca^{2+}]_i$ that persisted in the absence of extracellular Ca^{2+} and was blunted by pre-treatment with PMA. It therefore causes the mobilization of intracellular Ca^{2+} in parathyroid cells, probably by acting on the Ca^{2+} receptor. It is about equipotent with spermine (EC_{50} about 200 μM) yet carries one less positive charge (+3) than does spermine.

The results obtained with budmunchiamine A demonstrate the predictive power of the structure-activity studies and the novel structural information to be gained by testing natural products. Thus, screening of natural products, selected rationally on the basis of the structural information is readily performed e.g., molecules can be selected on the basis of well-established chemotaxonomic principles using appropriate data bases, such as Napralert. For example, macrocyclic polyamine alkaloids derived from papilionoid legumes related to Albizia, such as Pithecolobium and other plant-derived molecules can be screened.

Fig. 36 provides a second example of a series of molecules which were screened to determine useful molecules of this invention. These molecules were generally derived from fendiline and tested to determine their respective EC_{50}'s. Moreover, testing of related molecules, such as NPS 447 and NPS 448 reveals stereospecific effects of molecule structure. The most active compounds tested to date are the novel compounds designated NPS 467 and NPS 568 which have EC_{50} values of less than 5μM. Those in the art, by reviewing this series of molecules, can determine other suitable derivatives which can be tested in the invention.

These examples demonstrate the general design and screening process useful in this invention, and indicate that additional compound and natural product libraries can be screened as desired by those in the art to determine other useful lead molecules or novel molecules of this invention.

As discussed above, examples of molecules useful as calcimimetics include branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines. In addition, other positively charged organic molecules, including naturally occurring molecules and their analogs, are useful calcimimetics. These naturally occurring molecules and their analogs preferably have positive charge-to-mass ratios that correlate with those ratios for the molecules exemplified herein. (Examples include material isolated from marine animals, arthropod venoms, terrestrial plants and fermentation broths derived from bacteria and fungi.) It is contemplated that one group of preferred naturally occurring molecules and analogs useful as calcimimetics will have a ratio of positive charge: molecular weight (in daltons) from about 1:40 to 1:200, preferably from about 1:40 to 1:100. More specific examples of such molecules are provided below.

Polyamines

The polyamines useful as calcimimetics in this invention may be either branched or cyclic. Branched or cyclic polyamines potentially have higher calcimimetic activity than their straight-chain analogs. That is, branched or cyclic polyamines tend to have a lower EC_{50} than their corresponding linear polyamines with the same effective charge at physiological pH (see Table 1).

Table 1

Molecule	Net (+) Charge	EC_{50} (μM)
Neomycin	+6	20 or 40
Hexacyclen	+6	20
NPS 382	+8	50
NPS 381	+10	100
NPS 383	+8	150
Gentamicin	+5	150
Spermine	+4	150
Bekanamycin	+5	200
Argiotoxin-659	+4	300
Pentaethylenehexamine (PEHA)	+6	500
Streptomycin	+3	600
Spermidine	+3	2000
Tetraethylenepentamine (TEPA)	+5	2500
1,12-diaminododecane (DADD)	+2	3000
Triethylenetramine (TETA)	+4	8000

"Branched polyamines" as used herein refers to a chain molecule consisting of short alkyl bridges or alkyl groups joined together by amino linkages, and also containing points at which the chain branches. These "branch points" can be located at either a carbon atom or a nitrogen atom, preferably at a nitrogen atom. A nitrogen atom branch point is typically a tertiary amine but it may also be quaternary. A branched polyamine may have 1 to 20 branch points, preferably 1 to 10 branch points.

Generally, the alkyl bridges and alkyl branches in a branched polyamine are from 1 to 50 carbon atoms in length, preferably from 2 to 6 carbon atoms. The alkyl branches may also be interrupted by one or more heteroatoms (nitrogen, oxygen or sulfur) or substituted with functional groups such as: halo, including fluoro, chloro, bromo, or iodo; hydroxy; nitro; acyloxy (R'COO-), acylamido (R'CONH-), or alkoxy (-OR'), where R' may contain from 1 to 4 carbon atoms. The alkyl branches may

also be substituted with groups that are positively charged at physiological pH, such as amino or guanido. These functional substituents may add or change physical properties such as solubility to increase activity, delivery or bioavailability of the molecules.

The branched polyamines may have three or more chain and branch termination points. These termination points may be methyl groups or amino groups, preferably amino groups.

One preferred group of molecules is the group of branched polyamines having the formula:

$$H_2N-(CH_2)_j-(NR_i-(CH_2)_j)_k-NH_2$$

where k is an integer from 1 to 10, each j is the same or different and is an integer from 2 to 20, and each R_i is the same or different and is selected from the group consisting of hydrogen and $-(CH_2)_j-NH_2$, where j is as defined above, and at least one R_i is not hydrogen.

Particularly preferred branched polyamines of this invention are the molecules $N^1,N^1,N^5,N^{10},N^{14},N^{14}$-hexakis-(3-aminopropyl)spermine and $N^1,N^1,N^5,N^{14},N^{14}$-tetrakis-(3-aminopropyl)spermine referred to as NPS 381 and NPS 382, respectively, in Figure 1.

"Cyclic polyamines" as used herein refer to heterocycles containing two or more heteroatoms (nitrogen, oxygen or sulfur), at least two of which are nitrogen atoms. The heterocycles are generally from about 6 to about 20 atoms in circumference, preferably from about 10 to about 18 atoms in circumference. The nitrogen heteroatoms are separated by 2 to 10 carbon atoms. The heterocycles may also be substituted at the nitrogen sites with aminoalkyl or aminoaryl groups (NH_2R-), wherein R is aminoaryl or a lower alkyl of 2 to 6 carbon atoms.

Particularly preferred cyclic polyamines of this invention are shown in Figure 1 as hexacyclen (1,4,7,10,13,16-hexaaza-cyclooctadecane) and NPS 383.

Polyamino Acids

The polyamino acids useful in this invention may contain two or more positively charged amino acid residues at physiological pH. These positively charged amino acids include histidine, lysine and arginine. These polypeptides will vary in length from 2 to 800 amino acids in length, more preferably from 20 to 300 amino acids in length. These polypeptides may consist of a single repeating amino acid residue, or may have the variety of a naturally occurring protein or enzyme.

The amino acid residues comprising the polyamino acids may be any of the twenty naturally occurring amino acids, or other alternative residues. Alternative residues include, for example, the ω-amino acids of the formula $H_2N(CH_2)_nCOOH$, where n is from 2 to 6. These are neutral, nonpolar amino acids, as are sarcosine, t-butyl alanine, t-butyl glycine, N-methyl isoleucine, norleucine, phenyl glycine, citrulline, methionine sulfoxide, cyclohexyl alanine, and hydroxyproline. Ornithine is an alternative positively charged amino acid residue. The polyamino acids of this invention may also be chemically derivatized by known methods.

Particularly preferred polyamino acids of this invention include polyarginine, polylysine, and poly(argininyl-tyrosine), having 20-300 residues. Another preferred polyamino acid is protamine, or a protamine analog.

Arylalkylamines

"Arylalkylamines" as used herein refer to a class of positively-charged toxins derived from arthropod venoms. Preferred arylalkylamines of this invention include philanthotoxin-433, argiotoxin-636, and argiotoxin-659, agatoxin 505, agatoxin 489, the structures of which are shown in Figure 1, and other synthetic molecules modeled after these natural products, such as NPS 019.

Additional Components

Compositions of this invention include not only at least one calcimimetic or calcilytic, but may also include certain additional components. These additional components include targeting components, labels, and other functionalities which may be useful in the applications herein, e.g., for screening for agonists or antagonists of extracellular Ca^{2+}.

For example, an immunoglobulin or portion thereof, or a ligand specific for parathyroid cells or a Ca^{2+} receptor can be used as a target-specific component. The immunoglobulin can be a polyclonal or monoclonal antibody and may comprise whole antibodies or immunologically reactive fragments of these antibodies such as $F_{ab'}$, F_{ab}, or $(F_{ab'})_2$. Receptor-specific ligands may also be used.

The compositions of this invention may also contain components derivatized with a molecule or ion which acts as a label. A wide variety of labeling moieties can be used, including radioisotopes, chromophores, and fluorescent labels. Radioisotope labeling in particular can be readily detected in vivo. Radioisotopes may be coupled by coordination as cations in the porphyrin system. Useful cations include technetium, gallium, and indium. In the compositions, the positively charged molecule can be linked to or associated with a label.

Methods of Synthesis

Strategies for the syntheses and the modification of polyamines involve the use of a variety of amine protecting groups (phthalimido, BOC, CBZ, benzyl, and nitrile) which can be removed selectively to construct functionalized molecules. The synthetic methods involved are modelled after those used to construct argiopines 636 and 659 and other arylalkylamines derived from spider venoms.

Chain extensions of 2-4 methylenes were typically accomplished by alkylation with the corresponding N-(bromoalkyl)phthalimide. A 1:1.2 mixture of amine to the bromoalkylphthalimide was refluxed in acetonitrile in

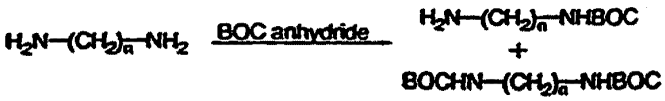
the presence of 50% KF on Celite. Chain extensions were also accomplished by alkylation of a given amine with acrylonitrile or ethylacrylate. Reaction progress was monitored by TLC and intermediates purified on silica gel using combinations of dichloromethane, methanol, and isopropylamine. Final products were purified by cation exchange (HEMA-SB) and RP-HPLC (Vydac C-18). Purity and structure verification are accomplished by ^{1}H- and ^{13}C-NMR and high-resolution mass spectrometry (EI, CI and/or FAB).

BOC protecting groups were added by the treatment of an amine (1° or 2°) with di-*tert*-butyl dicarbonate in dichloromethane in the presence of a catalytic amount of dimethylaminopyridine. Benzyl protecting groups were applied in one of two ways: (1) condensation of a 1° amine with benzaldehyde followed by sodium borohydride reduction or (2) alkylation of a 2° amine with benzylbromide in the presence of KF. Amide linkages and cyclizations were typically performed by the reaction of an amine (1° or 2°) with the N-hydroxysuccinimide ester of a given acid. This was accomplished directly (in the case of cyclizations) by treatment of the "amino acid" with dicyclohexylcarbodiimide under dilute conditions.

Deprotections of the phthalimido functionality were accomplished by reduction with hydrazine in refluxing methanol. Deprotections of the BOC functionality were accomplished in anhydrous TFA. Deprotection of benzyl, nitrile, and CBZ protecting functionalities was accomplished by reduction in glacial acetic acid under 55 psi hydrogen in the presence of a catalytic amount of palladium hydroxide on carbon. Nitrile functionalities (in the presence of benzyl and CBZ groups) were selectively reduced under hydrogen in the presence of sponge Raney nickel.

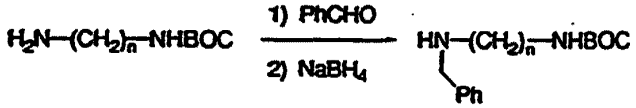
Specifically, branched polyamines are typically prepared from simple diaminoalkanes of the formula NH_2-$(CH_2)_n$-NH_2, or simple polyamines such as spermidine or spermine. One of the two primary (terminal) amines is

protected or "masked" with a protecting group such as BOC (t-butyloxycarbonyl), phthalimido, benzyl, 2-ethylnitrile (the Michael condensation production product of an amine and acrylonitrile), or amide. A typical reaction is the addition of a BOC protecting group by treatment with di-t-butyl-dicarbonate (BOC anhydride):

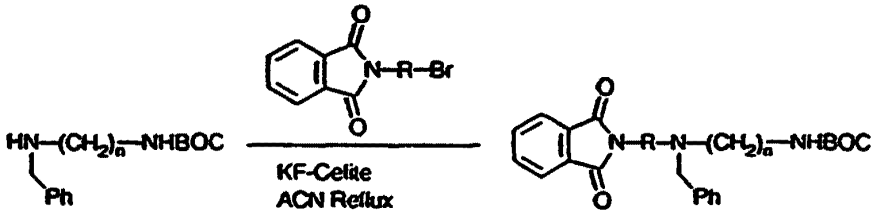


The monoprotected product is separated from the unprotected and diprotected products by simple chromatographic or distillation techniques.

The remaining free amine in the monoprotected product is then selectively alkylated (or acylated) with an alkylating (or acylating) agent. To ensure mono-alkylation, the free amine is partially protected by condensation with benzaldehyde followed by sodium borohydride reduction to form the N-benzyl derivative:

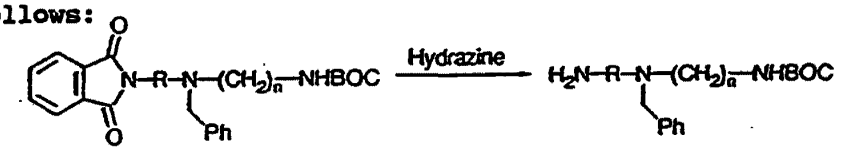


The N-benzyl derivative is then reacted with the alkylating agent. A typical alkylating agent is in an N-(bromoalkyl)phthalimide, which reacts as follows:



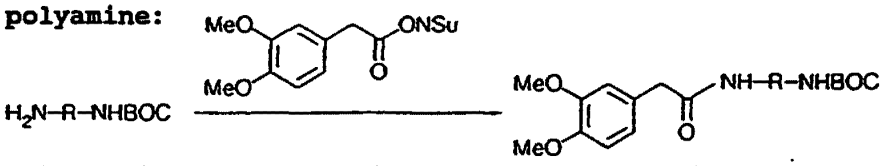
For example, N-(bromobutyl)phthalimide is used to extend or branch the chain with four methylene units. Alternatively, reaction with acrylonitrile followed by reduction of the cyano group will extend the chain by three methylenes and an amino group.

The protecting groups of the resulting chain-extended molecule can then be selectively cleaved to yield a new free amine. For example, trifluoroacetic acid is used to remove a BOC group; catalytic hydrogenation is used to reduce a nitrile functionality and remove a benzyl group; and hydrazine is used to remove phthalimido groups as follows:



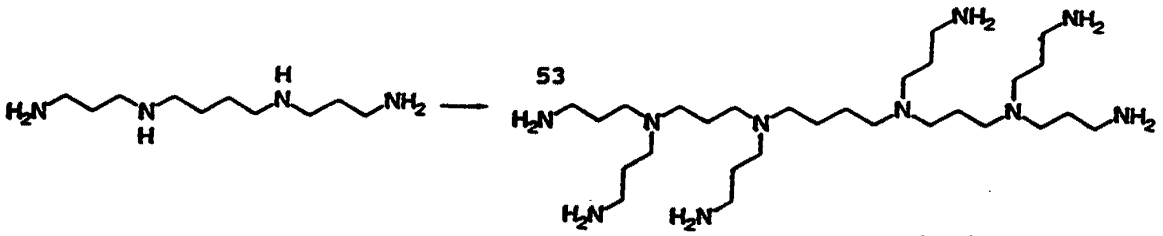
The new free amine may be alkylated (or acylated) further as above to increase the length of the polyamine. This process is repeated until the desired chain length and number of branches is obtained. In the final step, deprotection of the product results in the desired polyamine. However, further modifications may be effected at the protected end prior to deprotection in the following manner:

For example, prior to BOC-deprotection, the polyamine is acylated with the N-hydroxysuccinimide ester of 3,4-dimethoxyphenylacetic acid to yield a diprotected polyamine:



This ultimately will yield an aryl polyamine. The BOC group can then be selectively removed with trifluoroacetic acid to expose the other amino terminus which can be extended as above.

Certain branched polyamines may be formed by simultaneously alkylating or acylating the free primary and secondary amines in a polyamine formed as above. For example, treatment of spermine with excess acrylonitrile followed by catalytic reduction yields the following:



Cyclic polyamines may be prepared as above beginning with starting materials such as hexacylen (Aldrich Chem.).

The polyamino acids within the scope of the present invention can be made by recombinant techniques known in the art, or may be synthesized using standard solid-phase techniques known in the art. Solid-phase synthesis is commenced from the carboxy-terminal end of the peptide using an α-amino protected amino acid. BOC protective groups can be used for all amino groups even through other protective groups are suitable. For example, BOC-lys-OH can be esterified to chloromethylated polystyrene resin supports. The polystyrene resin support is preferably a copolymer of styrene with about 0.5 to 2% divinylbenzene as a cross-linking agent which causes the polystyrene polymer to be completely insoluble in certain organic solvents. See Stewart et al., Solid-Phase Peptide Synthesis (1969), W.H. Freeman Co., San Francisco; and Merrifield, J. Am. Chem. Soc. (1963) 85:2149-2154. These and other methods of peptide synthesis are also exemplified by U.S. Patent Nos. 3,862,925; 3,842,067; 3,972,859; and 4,105,602.

The polypeptide synthesis may use manual techniques or automatically employing, for example, an Applied Biosystems 403A Peptide Synthesizer (Foster City, California) or a Biosearch SAM II automatic peptide synthesizer (Biosearch, Inc., San Rafael, California), following the instructions provided in the instruction manual supplied by the manufacturer.

The arylalkylamines of the invention are natural products isolated by known techniques, or synthesized as described in Jasys et al., Tetrahedron Lett. (1988) 29:6223-6226, and Nason et al., Tetrahedron Lett. (1989) 30:2337-2340.

One general protocol for preparation of fendiline (or fendiline analogs shown in Fig. 36) is as follows. In a 10 ml round bottom blask equipped with a magnetic stir bar and rubber septum, 1.0 mmole 3,3'-bisphenylpropylamine (or primary alkyl amine) in 2 ml ethanol was treated with 1.1 mmole phenol and 1.0 mmole acetophenone (or substituted acetophenone). To this was added 2.0 mmoles $MgSO_4$ and 1.0 mmole $NaCNBH_3$. This was stirred under a nitrogen atmosphere at room temperature (about 20°C) for 24 hrs. The reaction was poured into 50 ml ether and washed 3 times with 1 N NaOH and once with brine. The ether layer was dried with anhydrous K_2CO_3 and reduced in vacuo. The product was then purified by column chromatography or HPLC incorporating silica stationary phase with combinations of CH_2Cl_2-Methanol-isopropylamine (typically 3% Methanol and 0.1% isopropylamine in methylene chloride).

A preferred procedure for preparing fendiline or fendiline analogs (such as those depicted in Figure 36) uses titanium(IV) isopropoxide and was modified from methods described in J. Org. Chem. 55:2552 (1990). For the synthesis of NPS 544, titanium tetrachloride (method described in Tetrahedron Letters 31:5547 (1990)) was used in place of titanium(IV) isopropoxide. The reaction scheme is depicted in Figure 43a. In Figure 43a, R,R' and R" depict hydrocarbyl groups. According to one embodiment, in a 4 ml vial, 1 mmole of amine (1) (typically a primary amine) and 1 mmole ketone or aldehyde (2) (generally acetophenone) are mixed, then treated with 1.25 mmoles titanium(IV) isopropoxide (3) and allowed to stand with occasional stirring at room temperature for about 30 minutes. Alternatively, a secondary amine may be used in place of (1). Note: some reactions will give heavy precipitates or solids which are warmed/heated (to their melting point) to allow for stirring/mixing several times over the course of the reaction. The reaction mixture is treated with 1 ml ethanol containing 1 mmole sodium cyanoborohydride (4) and the resulting mixture is then allowed

to stand at room temperature with occasional stirring for about 16 hours. After this time the reaction is quenched by the addition of about 500 µl water. The reaction mixture is then diluted to about 4 ml total volume with ethyl ether and then centrifuged. The upper organic phase is removed and reduced on a rotovapor. The resulting product, (6), is partially purified by chromatography through a short column of silica (or alternatively by using preparative TLC on silica) using combination of dichloromethane:methanol:isopropylamine (typically 95:5:0.1), prior to purification by HPLC (normal phase using silica with dichloromethane:methanol:isopropylamine or reversed phase, C-18 with 0.1% TFA with acetonitrile or methanol).

If appropriate or desired, chiral resolution may be accomplished using methods such as those described in Example 21.

Formulation and Administration

As demonstrated herein, the molecules of the invention may be used to: (a) mimic or antagonize one or more of the effect of extracellular Ca^{2+}; (b) affect the extracellular free Ca^{2+} level in an individual; and (c) treat diseases such as hyperparathyroidism, osteoporosis and hypertension. While the molecules have generally been shown to have an effect on parathyroid cells, they may also modulate the Ca^{2+} receptors on other cells, including bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells, and placental trophoblasts.

While these molecules will typically be used in therapy for human patients, they may be used to treat similar or identical diseases in other warm-blooded animal species such as other primates, farm animals such as swine, cattle, and poultry; and sports animals and pets such as horses, dogs and cats.

In therapeutic and/or diagnostic applications, the molecules of the invention can be formulated for a variety of modes of administration, including systemic and topical or localized administration. Techniques and formulations generally may be found in Remington's Pharmaceutical Sciences, Mack Publishing Co., Easton, PA.

For systemic administration, oral administration is preferred. Alternatively, injection may be used, e.g., intramuscular, intravenous, intraperitoneal, and subcutaneous. For injection, the molecules of the invention are formulated in liquid solutions, preferably in physiologically compatible buffers such as Hank's solution or Ringer's solution. In addition, the molecules may be formulated in solid form and redissolved or suspended immediately prior to use. Lyophilized forms are also included.

Systemic administration can also be by transmucosal or transdermal means, or the molecules can be administrated orally. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration bile salts and fusidic acid derivatives. In addition, detergents may be used to facilitate permeation. Transmucosal administration may be through nasal sprays, for example, or using suppositories. For oral administration, the molecules are formulated into conventional oral administration forms such as capsules, tablets, and tonics.

For topical administration, the molecules of the invention are formulated into ointments, salves, gels, or creams, as is generally known in the art.

As shown in the examples below, the amounts of various compounds of this invention which must be administered can be determined by standard procedure. Generally it is an amount between about 1 and 50 mg/kg animal to be treated.

Recombinant Ca^{2+} Receptors

Natural product screening has traditionally provided the lead structures for the development of diverse therapeutic molecules. However, high-throughput screening of natural product libraries or other molecule libraries for activity on the Ca^{2+} receptor has not previously been possible. To achieve this capability, it is best to clone the Ca^{2+} receptor cDNA and then create transfected cell lines suitable for high-throughput screening. The structure of the receptor can additionally be used to gain insight into the molecular geometry of the ligand binding site(s), and such information used to augment a rational drug design program as discussed above. Limited structure-activity studies and testing of selected natural product molecules will provide the initial structural data base necessary to guide rational natural product screening and drug design.

The bovine and human parathyroid cell Ca^{2+} receptor cDNA can be cloned by functional expression in Xenopus oocytes. It is possible to monitor an increase in intracellular Ca^{2+} in Xenopus oocytes indirectly by measuring current through the endogenous Ca^{2+}-activated Cl^- channel. The amplification of the response afforded by this signal transduction pathway enables the detection of receptor proteins encoded by mRNA at very low levels. This allows the detection of receptor-specific cDNA clones without the need for high affinity ligands, specific antisera, or protein or nucleic acid sequence information. An example of such a procedure follows.

Adult female Xenopus laevis were obtained from Xenopus I (Ann Arbor, MI) and maintained according to standard procedures. Lobes of ovary were excised from hypothermically-anesthetized toads. Clusters of oocytes were transferred into modified Barth's saline (MBS). Individual oocytes were obtained by incubation in MBS containing 2 mg/ml collagenase (Sigma, Type 1A) for 2h at 21°C and stage V-VI oocytes were selected for injection.

Glass capillary tubes (1 mm diameter) were pulled to a fine tip and manually broken to achieve a tip diameter of about 15 μM. A droplet of mRNA (1 ng/nl in diethyl-pyrocarbonate (DEPC)-treated water) was placed onto para-film and drawn into the capillary tube by suction. The capillary tube was then connected to a picospritzer (WPI Instruments) and the volume of the air-pulsed droplets adjusted to deliver 50 ng of mRNA (typically 50 nl). A 35 mm culture dish with a patch of nylon stocking fixed to the bottom was used to secure the oocytes during injection of mRNA into the vegetal pole. The injected oocytes were placed into a 35 mm culture dish containing MBS, 100 ug/ml penicillin and 100 U/ml streptomycin and incubated at 18°C for 3 days.

Following incubation, an oocyte was placed into a 100 μl plastic chamber and superfused with MBS at a flow rate of 0.5 ml/min using a peristaltic pump. Test molecules or inorganic polycations were added by rapidly moving the tubing into different buffers. Recording and current-passing electrodes were constructed from thin wall capillary tubing pulled to a resistance of 1-3 Mohms and filled with 3 M KCl. Oocytes were impaled (in the animal pole) with both electrodes under microscopic observation and connected to an Axon Instruments Axoclamp 2A voltage-clamp amplifier which was used to set the holding potential (-70 to -80 mV) and to measure the currents that were passed to maintain the holding potential. Currents were recorded directly onto a strip chart recorder.

For mRNA preparation, tissue was obtained from calves or patients with secondary HPT undergoing surgical removal of the parathyroid glands. Purified cells need not be prepared; whole pieces of gland were used to prepare mRNA that directs the expression of the Ca^{2+} receptor in Xenopus oocytes. Total cellular RNA was prepared by acid guanidinium thiocyanate/phenol extraction of homogenized glands. Oligo-dT cellulose chromatography was used to select poly(A)$^+$ mRNA by standard procedures. For size fraction-

ation of mRNA, centrifugation through glycerol gradients was used. The mRNA was denatured with 20 mM methylmercuric hydroxide and loaded (50-100 ug at a concentration of 1 mg/ml) onto a linear 15-30% glycerol gradient prepared in Beckman TLS55 tubes. Following centrifugation at 34,000 rpm for 16 hrs, 0.3 ml gradient fractions were collected and diluted in an equal volume of water containing 5mM *beta*-mercaptoethanol. mRNA was then recovered by two cycles of ethanol precipitation. If desired, the mRNA (50-100 ug of poly(A)$^+$) may be separated on a 1.2% agarose/6.0 M urea preparative gel, along with a range of RNA size markers. Following visualization of the mRNA by ethidium bromide staining, gel slices containing RNA in ~1.5-2.0 kb size steps are excised. mRNA is recovered from the agarose gel slices using RNAid binding matrix (according to the supplier's standard protocol; Stratagene, Inc.) and recovered mRNA fractions eluted into DEPC-treated water.

Amounts of recovered mRNA were quantified by UV absorbance measurement. The size range of mRNA contained within each fraction was determined by formaldehyde/agarose gel electrophoresis using a small quantity (0.5 ug) of each sample. The integrity of the mRNA was assessed by <u>in vitro</u> translation of each sample. Reticulocyte lysates (commercially available kits; BRL) were used to translate 0.05-0.5 ug of each mRNA fraction. The resulting ^{35}S-labelled proteins were analyzed by SDS-PAGE. The intact mRNA was capable of directing the synthesis of proteins of a complete size range, corresponding roughly to the sizes of the individual mRNA fractions.

A cDNA library was constructed in the vector λ ZAPII, following modifications of the technique of Gubler and Hoffman. RNA from the fraction(s) giving the best response in the oocyte assay was used as starting material. First-strand cDNA syntheses was primed with an oligo-dT/<u>Not</u>I primer-linker. Second-strand synthesis was by the RNase H/DNA Polymerase I self-priming method. Double-stranded cDNA was blunted with T4 DNA polymerase

and EcoRI adaptors blunt-end ligated to the cDNA with T4 ligase. Following NotI digestion to cleave the linker, full-length cDNA was size-selected by exclusion chromatography on Sephacryl 500 HA. First-strand cDNA was radiolabeled with α-^{32}P-dATP, and all synthesis and recovery steps monitored by following the incorporation of radioactivity. Full-length cDNA recovered from the sizing column was ligated to EcoRI/NotI digested λ ZAPII arms. The ligation mix was test packaged with commercially available high efficiency packaging extract (Stratagene, Inc.) and plated on the appropriate host strain (XL1-blue). The percentage of recombinant phage was determined by the ratio of white to blue plaques when the library is plated on IPTG and X-gal.

The average insert size was determined from ten randomly selected clones. Phage DNA "mini-preps" were digested with EcoRI and NotI to release the insert, and the size determined by agarose gel electrophoresis. The library consisted of >90% recombinant phage, and the insert size ranged from 1.5 to 4.2 kb. The recombinant ligation was packaged in large scale to generate 800,000 primary clones. The packaging mix was titered and plated at 50,000 plaques per 15 cm plate. Each pool of 50,000 clones was eluted in SM buffer and stored individually.

Plate lysate stocks of each of the clone pools were used for small scale phage DNA preparation. Phage particles are concentrated by polyethylene glycol precipitation, and phage DNA purified by proteinase K digestion followed by phenol:chloroform extraction. Twenty micrograms of DNA are digested with NotI, and used as template for *in vitro* transcription of sense-strand RNA. *In vitro* transcription is according to standard protocols, utilizing T7 RNA polymerase and 5' cap analog m^{7}GpppG in a 50μl total reaction volume. Following Dnase I/Proteinase K digestion and phenol/chloroform extraction, the RNA is concentrated by ethanol precipitation and used for oocyte injection.

Oocytes were injected with synthetic mRNA (cRNA) from each of the 16 library subpools constituting 50,000 independent clones each. After incubation for 3 to 4 days, oocytes were assayed for the ability of 10 mM neomycin to elicit a Ca^{2+} dependent Cl-current. A pool designated 6 gave a positive signal and thus contains a cDNA clone encoding a functional calcium receptor. In order to decrease the complexity of pool 6 and thus proceed towards the purification of the calcium receptor clone contained within this pool, pool 6 phage were replated at ~20,000 plaques per plate and 12 plates harvested. DNA was prepared from each of these subpools and cRNA synthesized. Again, oocytes were injected with cRNA and assayed 3-4 days later for the ability of 10 mM neomycin to elicit a Ca^{2+} dependent Cl-current. A subpool 6-3 was positive and this pool was subjected to a further round of plating reducing the complexity of pools to around 5,000 clones per pool. Pools were again assayed by preparation of cRNA and injection in oocytes. A subpool 6-3.4 was positive. In order to expedite further purification of the positive clone in pool 6-3.4, phage DNA from this pool was rescued as plasmid DNA by superinfection with the helper phage, ExAssist. Transfection of rescued plasmids into bacterial strain DH5alphaF' resulted in transformed bacterial colonies on ampicillin plates. These were harvested in pool of 900 clones each. Plasmid DNA was then prepared from each subpool and cRNA synthesized and assayed in the usual manner. Subpool 6-3.4.4 was positive. Bacteria containing the plasmid subpool 6-3.4.4 were subsequently plated in subpools of ~50 clones each. Continuation of this process is expected to result in a single clone encoding a functional calcium receptor.

Initial experiments used Xenopus oocytes injected with water or poly(A)$^+$-enriched mRNA (50 ng) from bovine parathyroid cells. After three days, the oocytes were examined for their ability to increase intracellular Ca^{2+} in response to increases in the concentration of extra-

cellular di- and trivalent cations. The oocytes were impaled with recording and current-passing electrodes and $[Ca^{2+}]_i$ was assessed indirectly by measuring currents through the endogenous Ca^{2+}-activated Cl^- channel. In oocytes injected with poly(A)$^+$-enriched mRNA from bovine (or human, Fig. 26) parathyroid cells, increasing the concentration of extracellular Ca^{2+} from 0.7 to 3, 5 or 10 mM caused a rapid and transient increase in the Cl^- conductance which then oscillated around a higher basal conductance. Increasing the concentration of extracellular Mg^{2+} from 1 to 10 mM likewise evoked oscillatory increases in Cl^- conductance. The Cl^- conductance response to extracellular Mg^{2+} persisted when the extracellular Ca^{2+} concentration was reduced to < 1 μM (Fig. 25).

The impermeant trivalent cation Gd^{3+} (600 μM) also caused oscillatory increases in the Cl^- conductance (Fig. 25). Such increases in the Cl^- conductance which oscillate and persist in the nominal absence of extracellular Ca^{2+} are noted when oocytes have been allowed to express other Ca^{2+}-mobilizing receptors and are stimulated with the appropriate ligand (e.g., substance K, Fig. 25). In these instances, the increase in Cl^- conductance reflects the mobilization of intracellular Ca^{2+}. These initial studies likewise show that extracellular polycations mobilize intracellular Ca^{2+} in parathyroid cell mRNA-injected oocytes.

Oocytes injected with water did not show any change in the Cl^- current when exposed to extracellular Ca^{2+} (10 mM) or Mg^{2+} (20 or 30 mM). In one series of experiments, oocytes were injected with the mRNA encoding the substance K receptor. In these oocytes, extracellular Mg^{2+} (20 mM) did not evoke any current but the cells responded vigorously to the addition of substance K (Fig. 25). These experiments indicate that there is no endogenous sensitivity of the oocyte to extracellular Ca^{2+} or Mg^{2+}.

Similar experiments were performed using oocytes injected with poly(A)$^+$-enriched mRNA prepared from human parathyroid glands (hyperplastic tissue from a case of secondary HPT). In these oocytes, increasing the concentration of extracellular Ca^{2+} caused a reversible increase in the Cl^- conductance which oscillated (Fig. 26). The addition of 300 μM La^{3+} likewise caused oscillatory increases in the Cl^- conductance. Increasing the concentration of extracellular Mg^{2+} from 1 to 10 mM evoked increases in the Cl^- conductance that persisted in the absence of extracellular Ca^{2+}. Additional experiments suggest that the response to extracellular Ca^{2+} is concentration dependent. Thus, in three mRNA-injected oocytes, Cl^- conductance increased to a maximum of 111 ± 22 nA at 3 mM and 233 ± 101 nA at 10 mM extracellular Ca^{2+}.

The results obtained in Xenopus oocytes demonstrate the presence of a mRNA(s) in parathyroid cells encoding a protein(s) which can impart, in normally unresponsive cells, sensitivity to extracellular Ca^{2+}. Moreover, the ability of extracellular Mg^{2+} to evoke oscillatory increases in Cl^- current in the absence of extracellular Ca^{2+} demonstrates that the Cl^- current depends on the mobilization of intracellular Ca^{2+} rather than influx of extracellular Ca^{2+}. The results obtained with La^{3+} likewise show that the expressed protein(s) is linked to the mobilization of intracellular Ca^{2+}. Together, these data show that the expressed protein(s) acts as a cell surface receptor rather than a channel. These studies provide compelling evidence for the existence of a Ca^{2+} receptor protein on the surface of parathyroid cells and demonstrate the feasibility of using the Xenopus oocyte system to achieve the molecular cloning of the Ca^{2+} receptor cDNA.

In another series of experiments, parathyroid cell mRNA, denatured with methylmercuric hydroxide, was size-fractionated by centrifugation through a glycerol gradient. Ten fractions were collected. Each group was

injected into Xenopus oocytes and after a three day incubation period the oocytes were assayed for expression of the Ca^{2+} receptor. Those oocytes injected with fractions 4-6 showed the largest and most consistent increases in Cl^- conductance in response to extracellular Ca^{2+} (Fig. 35). These results indicate that the Ca^{2+} receptor is encoded by mRNA in a size range of 2.5-3.5 kb. This indicates that a strategy using direct expression of RNA synthesized from a transcription vector cDNA library is feasible. Size-fractionation experiments of this sort were conducted and in each of three different fractionation experiments similar results were obtained.

The mRNA fractions obtained and characterized in the preceding experiments can be assayed by injection into oocytes. For each mRNA fraction, 10-20 oocytes are injected with 50 ng of RNA at a concentration of 1 ng/nl in water. Injected oocytes are maintained at 18° C for 48-72 h after which they are assessed for expression of the Ca^{2+} receptor using measurements of Cl^- current. For each group of injected oocytes the number positive for expression of the receptor, as well as the magnitude of the Ca^{2+}-dependent Cl^- current measured, is determined. As negative controls, oocytes are injected with rat liver poly(A)$^+$-enriched mRNA, yeast RNA, or water.

It is expected that an mRNA in the range of 2.5 - 3.5 kb will encode the receptor. mRNA of a larger size may necessitate a cloning approach based on hybrid depletion of parathyroid mRNA prior to oocyte injection. This strategy is not dependent upon the generation of full-length cDNA clones for success. If receptor expression is not obtained with a single size fraction of mRNA, oocytes are injected with mixed size fractions to determine a combination that does give rise to a functional receptor. If it does appear that multiple subunits are necessary for the formation of a functional receptor, the hybrid depletion expression cloning strategy is used. In this approach, clones are selected on the basis of their abil-

ity to deplete a specific mRNA species from the total mRNA population. A clone encoding a single subunit is identified by its ability to prevent the formation of the active multi-subunit complex. By exhaustive screening it is possible to identify clones encoding all of the necessary subunits.

This approach permits the isolation of clones encoding individual subunits required to form a functional receptor complex. Synthetic RNA from pools of clones are assayed for their ability to induce expression of the Ca^{2+} receptor in Xenopus oocytes by the same techniques used to analyze the original mRNA fractions. Originally, 10 pools representing 100,000 primary clones each are examined. Pools of clones showing a positive response are screened at lower (typically 4 to 10 fold) complexity, and again positive pools further subdivided and screened. This process of library sub-fractionation is followed until individual positive clones are identified. As a negative control for the oocyte expression assay, anti-sense transcripts are generated by T7 transcription of those DNA templates that induce a positive response. Anti-sense transcripts are unable to give rise to an authentic receptor, and this will control any non-specific positive signal arising from injection of synthetic RNA. Another concern is the fact that synthetic RNA can occasionally "poison" translation in injected oocytes, by an undefined mechanism. To control for this possibility, synthetic RNAs giving a negative response are co-injected at various dilutions with parathyroid cell mRNA, to determine if they are non-specifically interfering with the expression of the Ca^{2+} receptor.

When an individual clone encoding the Ca^{2+} receptor is identified, the cDNA insert will be excised from the λ vector and used for large scale production of synthetic RNA. Oocyte injection of this single RNA species allows rigorous assessment of the characteristics of the expressed receptor.

If the size of the mRNA encoding the Ca^{2+} receptor is too large for cloning by direct transcription and expression, or if multiple subunits are involved, a hybrid-depletion technique of screening pools of clones is used. cDNA insert DNA will be prepared from pools of clones from the size-selected parathyroid cell cDNA library. This DNA is hybridized to parathyroid cell mRNA under conditions that permit the formation of DNA/RNA duplexes. The unannealed, hybrid-depleted RNA is recovered and used for oocyte injections. DNA from pools of clones containing sequences representing Ca^{2+} receptor mRNA is depleted from this mRNA from the total parathyroid cell mRNA population, and expression of the receptor is reduced or absent upon oocyte injection. A process of sub-fractionation is followed on pools of clones of decreasing complexity, at each step assaying for cloned DNA that depletes Ca^{2+} receptor-encoding mRNA from the total parathyroid cell mRNA population. The use of an internal control during the hybrid depletion assays ensures that the hybrid-depleted RNA is intact and capable of being translated in the oocyte.

Human parathyroid cells express a *beta*-adrenergic receptor coupled to adenylate cyclase. This receptor can be expressed in oocytes, where it is capable of agonist-induced activation of the endogenous adenylate cyclase. During the hybrid-depletion screening for Ca^{2+} receptor clones, oocytes injected with hybrid depleted mRNA are assayed for isoproterenol-induced adenylate cyclase activation. A positive response in this assay serves to indicate that any observed inhibition of Ca^{2+} receptor response is specific, and not due to a general inhibition of the total mRNA population.

The hybrid-depletion screening strategy can result in the isolation of clones that do not contain a complete protein coding region. Positive clones isolated by this screening strategy are sequenced to determine their protein coding capacity. Northern blot analysis of human parathyroid gland RNA permits the determination of the

size of the complete mRNA corresponding to specific clones. If positive clones do not appear to be full-length, the cloned cDNA will be used as a hybridization probe to screen a parathyroid gland cDNA library for complete cDNAs.

A variety of cell lines are capable of coupling exogenously expressed receptors to endogenous functional responses. A number of these cell lines (e.g., NIH-3T3, HeLa, NG115, CHO, HEK, 293 and COS7) can be tested to confirm that they lack an endogenous Ca^{2+} receptor. Those lines lacking a response to external Ca^{2+} can be used to establish stably transfected cell lines expressing the cloned Ca^{2+} receptor.

Sequence analysis of Ca^{2+} receptor cDNA clones identified by expression cloning will delineate the open reading frame encoding the receptor protein. The coding region of the cDNA will be subcloned into multiple cloning site of the eukaryotic expression vector pMSG. This vector allows high level transcirption driven by the mouse mammary tumor virus (MMTV) promoter, and is active in a wide variety of mammalian cells. The vector also contains a gpt gene for resistance to mycophenolic acid which is under the control of the SV40 early promoter, and sequences necessary for selection and growth in bacteria. Large quantities of the expression vector/receptor cDNA plasmid construct will be grown and purified from E. coli.

The most effective method for transfection of eukaryotic cell lines with plasmid DNA varies with the given cell type. The Ca^{2+} receptor expression construct will be introduced into cultured cells by the appropriate technique, either Ca^{2+} phosphate precipitation, DEAE-dextran transfection, lipofection or electroporation. Following the transfection procedure, cells are grown in the presence of the antibiotic G418 to select for cells expressing the neomycin resistance gene. Colonies of G418 resistant transfectants will be subcloned and established as individual cell lines. Expression of the Ca^{2+} receptor protein

in G418 resistant cells will be assessed by several methods. Southern blot and slot blot analysis will confirm the presence and the copy number of the receptor cDNA sequence. Northern blot analysis will be used to demonstrate that receptor mRNA is being transcribed from the plasmid construct. Functional expression of the receptor protein will be determined by measuring the mobilization of intracellular Ca^{2+} in response to externally applied Ca^{2+} receptor agonists.

Cloning the Ca^{2+} receptor enables both structural and functional studies of this novel receptor. Recombinantly produced receptor may be crystallized for structural studies. Stably transfected cell lines expressing the receptor can be used for high-throughput screening of natural product or other compound libraries. Molecules of the requisite potency and specificity can be labeled (radioactively or fluorescently). The ability of test molecules/extracts to displace such a labeled molecule will form the basis of a high-throughput assay for screening.

Given the appropriate cells or tissues expressing other calcium receptors, these receptors may be cloned in a manner analogous to that described above for the parathyroid cell calcium receptor. For example, mRNA from human osteoclastoma tissue encodes the osteoclast calcium receptor (Figure 34). Thus, to isolate a clone for the human osteoclast receptor, one need only isolate mRNA from osteoclastoma tissue, prepared a cDNA library and assay/fractionate subpools as described above. Furthermore, the preferred receptors for drug screening are of human origin. A clone encoding a calcium receptor from one species may be used to obtain the corresponding human cDNA clone by cross-hybridization as is well known by those skilled in the art. In addition, the clone of the parathyroid cell or other cell Ca^{2+} receptor allows isolation of genes encoding similar Ca^{2+}-sensing proteins in other cells, and expression of those proteins. This is achieved by a variety of approaches. Southern blot analysis of human geno-

mic DNA, utilizing the Ca^{2+} receptor cDNA as a hybridization probe, will give an indication of the number of related sequences encoded within the genome; hybridization at varying stringencies will give an indication of the degree of divergence among the related sequences. This will provide information about the potential number of genes encoding related receptor proteins. Northern blot analysis with Ca^{2+} receptor cDNA as probe will determine if the same or related transcripts are present in various tissues. If related transcripts homologous to the parathyroid cell Ca^{2+} receptor are detected, it is a relatively simple matter to obtain clones of these mRNAs, either by screening the appropriate cDNA libraries or by polymerase chain reaction (PCR) techniques. Novel receptor clones so obtained can be assessed functionally by expression, either in oocytes or in transfected cell lines. Transfected cell lines expressing a cell-specific Ca^{2+} receptor can then provide a means of high-throughput screening for molecules that act specifically on the Ca^{2+}-sensing mechanism of, for example, osteoclasts or juxtaglomerular cells.

In an alternative method, the calcium receptor can be cloned by expression in eukaryotic cells. For example, a cDNA library can be prepared from parathyroid mRNA and cloned into the eukaryotic expression vector, pCDNA1. Subpools from this library can be transfected into eukaryotic cells such as COS7 or HEK293 cells resulting in relatively high level transient expression of encoded cDNA sequences. Cells transfected with a function calcium receptor clone will express the calcium receptor which can then be activated by calcium, neomycin or other calcimimetic compounds. If cells are first loaded with a fluorometric indicator for $[Ca^{2+}]_i$, activation of the calcium receptor results in increased fluorescence. Thus library subpools containing the calcium receptor are identified by their ability, upon transfection into eukaryotic cells, to induce a calcium or calcimimetic-specific increase in

fluorescense. This fluorescense can be detected using either a fluorimeter or a fluorescence activated cell sorter (FACS).

In an alternative method, the Ca^{2+} receptor can be cloned by use of a monoclonal antibody generated against the receptor. Monoclonal antibodies provide powerful tools for the immunoaffinity purification of specific proteins. Once purified, limited amino acid sequence data can be obtained from the protein of interest, and used to design oligonucleotide sequence probes to screen for clones of the complete cDNA sequence.

For production of hybridomas, whole bovine parathyroid gland cells are used as the immunogen. Purified, dispersed cells are obtained, and live cell preparations are injected intraperitoneally into the appropriate mouse strain, according to established procedures. Standard protocols are followed for immunization schedules and for the production of hybridomas. A two-step screening procedure is used to identify hybridomas secreting monoclonal antibodies that recognize the Ca^{2+} receptor. The initial screen will identify those monoclonals that recognize parathyroid cell surface antigens. Immunohistochemical techniques are then used to screen hybridoma supernatants for the presence of mouse antibodies that bind to the surface of parathyroid cells. This screen can be performed on fixed sections of parathyroid gland tissue, or on dispersed cells in primary culture. The techniques for this assay are well established in the literature.

This screen will identify hybridomas producing monoclonal antibodies to a variety of cell surface determinants, and monoclonals specific for the Ca^{2+} receptor would be expected to comprise only a small subset of these. To identify monoclonal antibodies that bind to the Ca^{2+} receptor, hybridoma supernatants that test positive in the initial screen are assayed for their ability to block the response of cultured parathyroid cells to Ca^{2+} receptor agonists. Some antibodies that bind to the extracellular

domain of the receptor are expected to inhibit or activate ligand binding or to otherwise interfere with or affect receptor activation.

Monoclonal antibodies positive in both screens are characterized through Western blotting, immunoprecipitation and immunohistochemistry. This permits the determination of the size of the antigen that is recognized and its tissue distribution. The appropriate monoclonal antibody is then used for purification of the Ca^{2+} receptor protein by immunoaffinity chromatography, following standard techniques. Sufficient quantities of protein are obtained to allow limited amino acid sequence determination. Degenerate oligonucleotide probes are then designed on the basis of the peptide sequence information. These probes are then used to screen parathyroid gland cDNA libraries for full length clones of the Ca^{2+} receptor. Clones obtained are characterized by DNA sequencing and by functional expression in the oocyte system and in cultured mammalian cell lines.

Alternatively, the antibodies can be used to screen expression libraries, e.g., cDNA libraries in λgt11 or its equivalent, to determine those clones expressing antigenically reactive protein. Such clones can then be sequenced to determine whether they encode a protein that might be a Ca^{2+} receptor.

It will also be appreciated by those skilled in the art that phage display libraries can be used to clone and analyze calcium receptors in place of monoclonal antibodies. In these libraries, antibody variable regions or random peptides are shotgun cloned into phage expression vectors such that the antibody regions or peptides are displayed on the surface of the phage particle. Phage which display antibody regions or peptides capable of high specific binding to calcium receptors will bind to cells which display these receptors (e.g. parathyroid cells, C-cells, osteoclasts, etc.). Millions of such phage can be panned against these cell types selecting only those

phage which can bind to these cells (which includes those phage binding to calcium receptors). In this manner, the complexity of the library can be vastly reduced. Subsequently, the screens described above for monoclonal antibodies can be used to isolate phage which display calcium receptor-binding antibody or peptide regions, and these phage can be used to isolate the calcium receptor for purposes of structural identification and cloning. Kits to prepare such phage display libraries are commercially available (e.g. Stratacyte, or Cambridge Antibody Technology Limited). Recombinant phage endowed with such calcium receptor-binding properties can also be used in lieu of monoclonal antibodies in the various analyses of calcium receptors. Such phage can also be used in high throughput binding competition screens to identify organic compounds capable of functional binding to calcium receptors which can serve as structural leads for the development of human therapeutics acting at the calcium receptor.

In another alternative, affinity cross-linking of radioligands to their receptors can be used to isolate the receptor protein as described by Pilch & Czech, 1 Receptor Biochem. Methodol. 161, 1984. Covalent attachment of a radioligand allows extensive washing to remove nonspecific binding. For example, a high affinity molecule, e.g., a random copolymer of arginine and tyrosine (MW = 22K; argtyr ratio = 4:1) which mobilizes intracellular Ca^{2+} with an EC_{50} of about 100 nM or less, is iodinated with ^{125}I, and cross-linked. Protamines, because of their much smaller size, may be preferable in cross-linking studies and can be reductively alkylated as described by Dottavio-Martin & Ravel, 87 Analyt. Biochem. 562, 1978.

Nonspecific labelling is kept to a minimum by cross-linking in the presence of unlabeled polycations and di- and trivalent cations. At high concentrations of these molecules nonspecific interactions of the label with the cell surface might be reduced.

Uses

Primary hyperparathyroidism (HPT) is characterized by hypercalcemia and elevated levels of circulating PTH. One of the major defects in HPT appears to be a diminished sensitivity of parathyroid cells to negative feedback regulation by extracellular Ca^{2+}. Thus, in tissue from patients with primary HPT, the "set-point" for extracellular Ca^{2+} is shifted to the right so that higher than normal concentrations of extracellular Ca^{2+} are required to depress PTH secretion. Moreover, in primary HPT, even high concentrations of extracellular Ca^{2+} often depress PTH secretion only partially. In secondary (uremic) HPT, a similar increase in the set-point for extracellular Ca^{2+} is observed even though the degree to which Ca^{2+} suppresses PTH secretion is normal. The changes in PTH secretion are paralleled by changes in $[Ca^{2+}]_i$: the set-point for extracellular Ca^{2+}-induced increases in $[Ca^{2+}]_i$ is shifted to the right and the magnitude of such increases is reduced. Moreover, staining of tissue with a monoclonal antibody that appears to recognize the Ca^{2+} receptor is diminished in adenomatous and hyperplastic parathyroid cells.

The Ca^{2+} receptor constitutes a discrete molecular entity for pharmacological intervention. Molecules that mimic or antagonize the action of extracellular Ca^{2+} are beneficial in the long-term management of both primary and secondary HPT. Such molecules provide the added impetus required to suppress PTH secretion which the hypercalcemic condition alone cannot achieve. Such molecules with greater efficacy than extracellular Ca^{2+} may overcome the apparent nonsuppressible component of PTH secretion which is particularly troublesome in adenomatous tissue. Alternatively or additionally, such molecules can depress synthesis of PTH, as prolonged hypercalcemia has been shown to depress the levels of preproPTH mRNA in bovine and human adenomatous parathyroid tissue. Prolonged hypercalcemia also depresses parathyroid cell proliferation in vitro, so calcimimetics can also be effective in limiting

the parathyroid cell hyperplasia characteristic of secondary HPT.

Other cells in the body can respond directly to physiological changes in the concentration of extracellular Ca^{2+}. Calcitonin secretion from parafollicular cells in the thyroid (C-cells) is regulated by changes in the concentration of extracellular Ca^{2+}. Renin secretion from juxtaglomerular cells in the kidney, like PTH secretion, is depressed by increased concentrations of extracellular Ca^{2+}. Extracellular Ca^{2+} causes the mobilization of intracellular Ca^{2+} in these cells. Isolated osteoclasts respond to increases in the concentration of extracellular Ca^{2+} with corresponding increases in $[Ca^{2+}]_i$ that arise partly from the mobilization of intracellular Ca^{2+}. Increases in $[Ca^{2+}]_i$ in osteoclasts are associated with an inhibition of functional responses (bone resorption) analogous to PTH secretion in parathyroid cells. Thus, there are sufficient indications to suggest that Ca^{2+}, in addition to its ubiquitous role as an intracellular signal, also functions as an extracellular signal to regulate the responses of certain specialized cells. Molecules of this invention can be used in the treatment of diseases associated with disrupted Ca^{2+} responses in these cells.

Cloning the Ca^{2+} receptor on parathyroid cells and other cells will allow the presence of homologous proteins in other cells to be directly assesed. A family of structurally homologous Ca^{2+} receptor proteins can thus be obtained. Such receptors will allow understanding of how these cells detect extracellular Ca^{2+} and enable evaluation of the mechanism(s) as a site of action for the therapeutics described herein effective in the treatment of HPT, osteoporosis, and hypertension, and novel therapies for other bone and mineral-related diseases.

Other uses are discussed above. For example, recombinant Ca^{2+} receptor proteins may be used in therapy, and introduced by standard methods, e.g., by transfection of nucleic acid encoding that protein. In addition, such

protein is useful in assays for calcimimetic molecules of this invention.

The following examples illustrate the invention but do not limit its scope.

Examples

In the studies described herein, a variety of organic molecules were found to mobilize intracellular Ca^{2+} and depress PTH secretion in parathyroid cells. These molecules are structurally diverse but most have a net positive charge at physiological pH. The cationic nature of the organic molecules plays an important role but is not the sole factor determining activity.

Example 1: Screening Calcimimetic Molecules on Bovine Parathyroid cells

Dissociated bovine parathyroid cells were purified on gradients of Percoll and cultured overnight in serum-free medium. The cells were subsequently loaded with fura-2 and the concentration of free intracellular Ca^{2+} measured fluorimetricly. Changes in $[Ca^{2+}]_i$ were used to screen for molecules active at the Ca^{2+} receptor. To be considered a calcimimetic, a molecule was required to show the normal effects caused by increasing extracellular Ca^{2+} and triggered by the activation of the Ca^{2+} receptor. That is,

1) The molecule must elicit an increase in $[Ca^{2+}]_i$ that persists in the absence of extracellular Ca^{2+} (demonstrating the mobilization of intracellular Ca^{2+});

2) The molecule must cause a decrease in isoproterenol-stimulated cyclic AMP formation which is blocked by pertussis toxin;

3) The molecule must inhibit PTH secretion over the same range of concentrations that cause the increase in $[Ca^{2+}]_i$; and

4) The concentration-response curves for Ca^{2+} mobilization and PTH secretion by the molecule must be

shifted to the right by a PKC activator, such as phorbol myristate acetate (PMA).

Several structurally different classes of molecules were tested: polyamines, aminoglycoside antibiotics, protamine, and polymers of lysine or arginine. The structures of these molecules are depicted in Figure 1. Included in Figure 1 are the net positive charge of the molecules and their EC_{50}'s for evoking the mobilization of intracellular Ca^{2+} in bovine parathyroid cells.

In general, the greater the net positive charge on the molecule, the greater its potency in causing the mobilization of intracellular Ca^{2+}. However, some striking exceptions to this apparent rule have been found as discussed below.

As can be seen from the figures, spermine, neomycin B, and protamine evoked rapid and transient increases in $[Ca^{2+}]_i$ in fura-2-loaded bovine parathyroid cells (Figs. 6, 7, 11). They did not, however, cause sustained, steady-state increases in $[Ca^{2+}]_i$ in bovine parathyroid cells (Fig. 6, 11), although they did in human parathyroid cells (Fig. 19). In this respect, they resembled the cytosolic Ca^{2+} response elicited by extracellular Mg^{2+}, which causes the mobilization of intracellular Ca^{2+} unaccompanied by an influx of extracellular Ca^{2+} in bovine cells (Fig. 11b). Transient increases in $[Ca^{2+}]_i$ elicited by spermine, neomycin B, or protamine were not blocked by low concentrations (1 μM) of La^{3+} or Gd^{3+} (Fig. 11f,g). Cytosolic Ca^{2+} transients elicited by the molecular polycations persisted in the absence of extracellular Ca^{2+} but were blocked when cellular stores of Ca^{2+} were depleted by pretreatment with ionomycin (Figs. 7; 11h,i). All these molecules therefore cause the mobilization of intracellular Ca^{2+} in parathyroid cells.

It was additionally shown that the molecular polycations mobilized the same pool of intracellular Ca^{2+} as that used by extracellular Ca^{2+}. Thus, increasing the concentration of extracellular Ca^{2+} progressively inhibited the

transient increases in $[Ca^{2+}]_i$ evoked by spermine (Fig. 6). Conversely, a maximally effective concentration of spermine or neomycin B (Fig. 12) blocked transient, but not steady-state increases in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+}.

Significantly, spermine, neomycin B, and protamine inhibited PTH secretion to the same extent as extracellular Ca^{2+}. These inhibitory effects on secretion were obtained at concentrations that caused the mobilization of intracellular Ca^{2+} (Figs. 8, 13). These findings are relevant to understanding the mechanisms contributing to the regulation of PTH secretion by extracellular Ca^{2+}. Because a variety of inorganic polycations all inhibit secretion, yet only extracellular Ca^{2+} causes sustained, steady-state increases in $[Ca^{2+}]_i$, such increases in $[Ca^{2+}]_i$ cannot be importantly involved in the regulation of secretion. Mobilization of intracellular Ca^{2+}, rather than the influx of extracellular Ca^{2+}, is the essential mechanism associated with the inhibition of PTH secretion. This is important because it defines the sufficient mechanism to be affected if a molecule is to affect PTH secretion; molecules stimulating selectively the influx of extracellular Ca^{2+} will be relatively ineffective in suppressing PTH secretion. In contrast, molecules causing solely the mobilization of intracellular Ca^{2+} should be just as efficacious as extracellular Ca^{2+} in suppressing PTH secretion.

Like the mobilization of intracellular Ca^{2+} elicited by extracellular Ca^{2+}, that elicited by molecular polycations was depressed by PMA. A representative experiment showing the preferential inhibitory effects of PMA on cytosolic Ca^{2+} transients elicited by spermine is shown in Fig. 14. Cytosolic Ca^{2+} transients evoked by ATP were unaffected, even when a submaximal concentration of ATP was used. The effect of PMA on cytosolic Ca^{2+} transients elicited by the molecular polycations paralleled its effect on responses to extracellular Ca^{2+}; in both cases

there was a shift to the right in the concentration-response curve (Fig. 15). The depressive effects of PMA on $[Ca^{2+}]_i$ were accompanied by potentiating effects on secretion which were overcome at higher concentrations of the organic polycations (Fig. 16).

The mobilization of intracellular Ca^{2+} elicited by molecular polycations was associated with increases in the formation of inositol phosphates. For example, protamine caused a rapid (within 30 s) increase in the formation of IP_3 which was accompanied by a rise in levels of IP_1. Both these effects were dependent on the concentration of extracellular protamine (Fig. 17). Moreover, pretreatment with PMA blunted the formation of inositol phosphates elicited by molecular polycations. Representative results obtained with spermine are presented in Fig. 18.

Spermine, neomycin B, and protamine depressed isoproterenol-induced increases in cyclic AMP. Like the inhibitory effects of extracellular Ca^{2+} on cyclic AMP formation, those caused by molecular polycations were blocked by pretreatment with pertussis toxin (Table 2).

Table 2

	cyclic AMP (% of control)	
	control	+PTx
0.5 mM Ca^{2+}	100	106 ± 8
2.0 mM Ca^{2+}	19 ± 4	94 ± 2
0.5 mM Ca^{2+}, 200 μM Spermine	23 ± 5	93 ± 6
0.5 mM Ca^{2+}, 30 μM Neomycin B	28 ± 8	87 ± 6
0.5 mM Ca^{2+}, 2 μg/ml Protamine	20 ± 4	89 ± 9

Pertussis toxin (PTx) blocks the inhibitory effects of extracellular Ca^{2+} and molecular polycations on cyclic AMP formation. Bovine parathyroid cells were cultured for 16 h with or without 100 ng/ml pertussis toxin. The cells were subsequently washed and incubated for 15 min with 10 μM isoproterenol with or without the indicated concentrations of extracellular Ca^{2+} or molecular polycations.

Total cyclic AMP (cells + supernatant) was determined by RIA and the results are expressed as a percentage of the levels obtained in 0.5 mM Ca^{2+} (112 ± 17 pmole/10^6 cells). Each value is the mean ± SEM of three experiments.

In human parathyroid cells, extracellular Mg^{2+} elicited a sustained, steady-state increase in $[Ca^{2+}]_i$ in addition to a rapid transient increase (Fig. 10). As in bovine parathyroid cells responding to extracellular Ca^{2+}, the steady-state increase in $[Ca^{2+}]_i$ evoked by Mg^{2+} in human parathyroid cells results from Ca^{2+} influx through voltage-insensitive channels (Fig. 10a). This effect of Mg^{2+} on steady-state $[Ca^{2+}]_i$ in human parathyroid cells is seen in both adenomatous and hyperplastic tissue.

Neomycin B and spermine were tested for effects on $[Ca^{2+}]_i$ in human parathyroid cells prepared from adenomatous tissue. Representative results with neomycin B are shown in Fig. 19. Neomycin B caused not only a transient but additionally a steady-state increase in $[Ca^{2+}]_i$ in human parathyroid cells (Fig. 19a). Thus, in human cells, the pattern of change in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+}, Mg^{2+} or neomycin B is very similar.

Cytosolic Ca^{2+} transients elicited by neomycin B persisted in the presence of La^{3+} (1μM) and absence of extracellular Ca^{2+}. Neomycin B therefore causes the mobilization of intracellular Ca^{2+} in human parathyroid cells. Neomycin B inhibited PTH secretion from human parathyroid cells at concentrations that caused the mobilization of intracellular Ca^{2+} (Fig. 13). There were, however, some differences in the responses of human and bovine parathyroid cells to neomycin B. The EC_{50} of neomycin B for the mobilization of intracellular Ca^{2+} was 40 μM in bovine and 20 μM in human parathyroid cells (cf. Figs. 13 and 15), whereas the potency of spermine was similar in bovine and human parathyroid cells (EC_{50} = 150

μM). Thus, although bovine cells can be used for initial studies to screen test molecules for activity, it is important to perform follow-up studies using human parathyroid cells.

To assess the effects of molecular polycations on C-cells, a neoplastic cell line, derived from a rat medullary thyroid carcinoma (rMTC 6-23 cells) was used. Both spermine (10 mM) and neomycin B (5 mM) were without effect on basal $[Ca^{2+}]_i$ in these cells. Nor did either molecule affect the response to the subsequent addition of extracellular Ca^{2+}. Representative results documenting the lack of effect of neomycin B are shown in Fig 21. Neomycin B (1 mM) or spermine (1 or 5 mM) failed to evoke any increase in $[Ca^{2+}]_i$ in osteoclasts (Fig. 23). In the trace shown, there appeared to be some potentiation of the response to a subsequent increase in the concentration of extracellular Ca^{2+}, although this was not a consistent finding. In two other cells, spermine (5 mM) was again without effect on basal $[Ca^{2+}]_i$ and caused a small inhibition (about 15%) of the extracellular Ca^{2+}-induced increase in $[Ca^{2+}]_i$. In a third cell, neomycin B (5 mM) was without effect on basal $[Ca^{2+}]_i$ and did not affect increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. The overall picture that develops from these studies is that spermine and neomycin B are without effect on basal or stimulated levels of cytosolic Ca^{2+} in osteoclasts.

The failure of the molecular polycations to affect the Ca^{2+}-sensing mechanisms of C-cells or osteoclasts demonstrates the ability to discover or design novel lead molecules that act specifically on the parathyroid cell Ca^{2+} receptor or otherwise modulate one or more functions of the parathyroid cell's normal response to $[Ca^{2+}]$.

Screening of various other molecules is described in detail below and the results summarized in Table 1.

Example 2: Polyamine Screening

Straight chain polyamines (spermine, spermidine, TETA, TEPA, and PEHA) and two derivatives thereof (NPS 381 and NPS 382) were screened as in Example 1. These molecules were all found to mobilize intracellular Ca^{2+} in bovine parathyroid cells. Their order of potency is as follows, with the net positive charge listed in parentheses:

Table 3

Molecule	EC_{50} (in μM)
NPS 382 (+8)	50
NPS 381 (+10)	100
spermine (+4)	150
PEHA (+6)	500
spermidine (+3)	2000
TEPA (+5)	2500
TETA (+4)	8000

Putrescine (+2) and cadaverine (+2) were inactive at a concentration of 2mM.

Another straight-chain polyamine, DADD, behaved somewhat differently from the other polyamines and is described in Example 7.

Example 3: Cyclic Polyamine Screening

Two cyclic polyamines, hexacyclen and NPS 383, were screened as in Example 1. Hexacyclen (+6, EC_{50} = 20 μM) is 7-fold more potent than NPS 383 (+8, EC_{50} = 150 μM). The converse would be expected based solely on net positive charge as the structural characteristic for Ca^{2+} receptor activity.

Example 4: Aminoglycoside Antibiotic Screening

Six antibiotics were screened as in Example 1. The resulting EC_{50}'s for the mobilization of intracellular Ca^{2+}, in rank order of potency, were:

Table 4

Antibiotic	EC_{50} (in μM)
neomycin (+6)	10
gentamicin (+5)	150
bekanamycin (+5)	200
streptomycin (+3)	600

Kanamycin (+4.5) and lincomycin (+1) were without effect at a concentration of 500 μM. Within the aminoglycoside series, there is a correlation between net positive charge and potency. However, neomycin is considerably more potent than various polyamines (NPS 381, NPS 382, NPS 383, PEHA) that have an equal or greater positive charge.

Example 5: Peptide and Polyamino Acid Screening

Protamine and polymers of lysine or arginine varying in peptide length were screened for their ability to mobilize intracellular Ca^{2+} as in Example 1. The resulting EC_{50}'s for the mobilization of intracellular Ca^{2+}, in rank order of potency, were:

Table 5

Peptide (MW in kD)	EC_{50} (in mM)
polyArg (100)	4
polyArg (40)	15
polyLys (27)	30
protamine (4.8)	75
polyArgTyr (22)	200
polyLys (14)	1000
polyLys (3.8)	3000

The net positive charge of these polymers increases as the MW increases. Thus, as for the aminoglycosides, there is a direct correlation between net charge and potency among this series of polyamino acids. Protamine is essentially polyArg with a net positive charge of +21.

Example 6: Arylalkylamine Screening

Molecules selected from the class of arylalkylamine toxins derived from the venoms of wasps and spiders were screened as in Example 1.

Philanthotoxin-433 (+3) was without effect at a concentration of 500 μM. It is similar in structure to the argiotoxins described below.

Argiotoxin-636 (400 μM) did not elicit increases in $[Ca^{2+}]_i$ but it did potentiate cytosolic Ca^{2+} responses to the subsequent addition of extracellular Ca^{2+}. This is a feature common to all molecules that activate the Ca^{2+} receptor and is also seen with a variety of extracellular divalent cations. This is considered in more detail in Example 7.

In contrast to argiotoxin-636, argiotoxin-659 elicited increases in $[Ca^{2+}]_i$ with an EC_{50} of 300μM. Argiotoxin-659 differs from argiotoxin-636 in having a hydroxylated indole moiety rather than a dihydroxyphenyl group. This is the only difference in the structure of these two molecules. Thus, the difference in potency lies in the nature of the aromatic group, not in the polyamine chain which carries the positive charge.

Example 7: Screening of Ca^{2+} Channel Blockers

Ca^{2+} channel blockers, i.e., those molecules which block influx of extracellular Ca^{2+} through voltage-sensitive Ca^{2+} channels, were screened as in Example 1. There are three structural classes of Ca^{2+} channel blockers: (1) dihydropyridines, (2) phenylalkylamines, and (3) benzothiazipines.

None of the dihydropyridines tested (nifedipine, nitrendipine, BAY K 8644, and (-) 202-791 and (+) 202-791) had any effect on basal $[Ca^{2+}]_i$ or increases in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+} when they were tested at 1 μM. Previous studies showed that parathyroid cells lack voltage-sensitive Ca^{2+} channels, but do have voltage-

insensitive Ca^{2+} channels that are regulated by the Ca^{2+} receptor.

The phenylalkylamines examined were verapamil, D-600 (a methoxy-derivative of verapamil), TMB-8, and an analog of TMB-8, NPS 384. The first three molecules were tested at a concentration of 100 μM. The phenylalkylamines behaved differently from other molecules examined. They evoked no change in $[Ca^{2+}]_i$ when added to cells bathed in buffer containing a low concentration of extracellular Ca^{2+} (0.5 mM). However, verapamil, D-600, and TMB-8 potentiated the mobilization of intracellular Ca^{2+} elicited by extracellular divalent cations and they additionally blocked the influx of extracellular Ca^{2+}. At intermediate levels of extracellular Ca^{2+} (1-1.5 mM), these molecules were capable of evoking a small but robust increase in $[Ca^{2+}]_i$ that arose from the mobilization of intracellular Ca^{2+}.

The phenylalkylamines act differently than organic polycations like neomycin. The data suggest that verapamil, D-600 and TMB-8 are partial agonists at the Ca^{2+} receptor, in contrast to the other molecules examined which are full agonists.

Molecule NPS 384, at a concentration of 300 μM, did not evoke an increase in $[Ca^{2+}]_i$ but it blocked influx of extracellular Ca^{2+}. Testing at higher concentrations may reveal an ability of this molecule to cause the mobilization of intracellular Ca^{2+}.

While the ability of these molecules to block influx is intriguing and not entirely unexpected, it is the ability of these molecules to evoke transient increases in $[Ca^{2+}]_i$ (arising from intracellular Ca^{2+} mobilization) that is important. Considerable experience with measurements of $[Ca^{2+}]_i$ in parathyroid cells shows that transient increases in $[Ca^{2+}]_i$ almost invariably result from the mobilization of intracellular Ca^{2+} and therefore reflects activation of the Ca^{2+} receptor.

The benzothiazipine examined, diltiazem, was similar in all respects to verapamil and D-600 and was also effective at 100 μM.

It should be mentioned that with the exception of the phenylalkylamines, all the active molecules tested above evoke increase in $[Ca^{2+}]_i$ that are of magnitude similar to that evoked by a maximally effective concentration of extracellular Ca^{2+}. This shows that these molecules are equally efficacious as extracellular divalent cations. This contrasts with the activity of phenylalkylamines, which seem to act only as partial agonists.

Amongst the phenyalkylamines, some interesting structure-activity relationships emerge. Significant is the different potencies of molecules like TMB-8 and NPS 384. TMB-8 potentiated transient increases in $[Ca^{2+}]_i$ at 100 μM whereas NPS 384 fails to do so even at 300 μM, yet these molecules carry the same net positive charge. It follows that some other structural feature, unrelated to net charge, imparts greater potency to TMB-8.

Example 8: Molecule Screening on Human Parathyroid Cells

Spermine and neomycin were tested for effects on $[Ca^{2+}]_i$ in human parathyroid cells obtained from glands removed by surgery and prepared as in Example 1. In human parathyroid cells, spermine was found to cause only a small increase in $[Ca^{2+}]_i$ when tested at a concentration of 300 μM.

Neomycin, on the other hand, evoked a large increase in $[Ca^{2+}]_i$ in human parathyroid cells when tested at a concentration of 20 μM. The magnitude of the response elicited by neomycin was equal to that evoked by a maximally effective concentration of extracellular Ca^{2+}.

Example 9: Molecule Screening on Xenopus Oocytes

Oocytes injected with mRNA from human parathyroid cells express the Ca^{2+} receptor and mobilize intracellular Ca^{2+} in response to a variety of extracellular inorganic

di- and trivalent cations. Using this screen allows one to test for an action directly on the Ca^{2+} receptor. Oocytes expressing the Ca^{2+} receptor also responded to several molecules active on intact parathyroid cells when screened as follows. Hexacyclen caused the mobilization of intracellular Ca^{2+} at a concentration of 135 μM. Neomycin (100 μM) and NPS 382 (5 mM) were also effective. This offers rather compelling evidence showing that these molecules act on the Ca^{2+} receptor or on some other protein intimately associated with its function.

For example, we have been able to detect Ca^{2+} receptor expression in oocytes by measuring $^{45}Ca^{2+}$ mobilization. In these experiments, oocytes were injected with bovine parathyroid mRNA or water and, after 72 hours exposed to serum or 10 mM neomycin. Prior to being stimulated, oocytes were loaded with $^{45}Ca^{2+}$. Stimulation with serum for 20 min resulted in intracellular $^{45}Ca^{2+}$ release representing a 45% increase compared to mock challenge with buffer. Challenge with 10 mM neomycin for 20 min. resulted in a 76% increase in $^{45}Ca^{2+}$ release. The assay is sensitive enough for use in cloning the Ca^{2+} receptor, and has the advantage of a higher throughout than the electrophysiological measurement of Ca^{2+} activated Cl^- current.

In another example, human osteoclastoma tissue was obtained from bone biopsy tissue. Oocytes injected with mRNA isolated from this tissue were challenged with 30 mM Ca^{2+}. Controls did not respond while 8 of 12 oocytes injected with osteoclastoma mRNA responded appropriately (Fig. 34). These experiments provide the first evidence that the Ca^{2+} response of osteoclasts to extracellular Ca^{2+} is in fact genetically encoded. The results also indicate that the osteoclast Ca^{2+} receptor may be cloned by expression in Xenopus oocytes.

Example 10: Molecule Screening on Rat Osteoclasts

However, the different sensitivities of parathyroid cells and osteoclasts to extracellular Ca^{2+} suggest that

their Ca^{2+} receptors are different. While parathyroid cells respond to extracellular Ca^{2+} concentrations between 0.5 and 3 mM, osteoclasts respond only when the level of extracellular Ca^{2+} increases beyond 5 mM. This rather high concentration of Ca^{2+} is nonetheless physiological for osteoclasts; as they resorb bone, the local concentration of extracellular Ca^{2+} may reach levels as high as 30 mM.

Molecule screening with osteoclasts was performed as follows. Osteoclasts were obtained from the long bones of neonatal rats. $[Ca^{2+}]_i$ was measured in single cells using the fluorimetric indicator indo-1. Spermine, spermidine, neomycin, and verapamil were tested, and none of these caused any large increase in $[Ca^{2+}]_i$ in osteoclasts (although small responses were detected).

At a concentration of 1 mM, spermidine caused a small increase in $[Ca^{2+}]_i$ (about 10% of that evoked by a maximal concentration of extracellular Ca^{2+}). Neither Neomycin (10 mM) nor Spermine (10 or 20 mM) caused increases in $[Ca^{2+}]_i$ in rat osteoclasts. Neomycin (10 mM) did not block the increase in $[Ca^{2+}]_i$ elicited by the subsequent addition of 25 mM extracellular Ca^{2+}. Pretreatment with spermine (20 mM) however, did depress the response to extracellular Ca^{2+}. Verapamil (100 μM) caused no detectable increase in $[Ca^{2+}]_i$ but it did block the response to extracellular Ca^{2+}.

Comparisons between osteoclasts and parathyroid cells show that molecules active on the latter are relatively ineffective in osteoclasts. This demonstrates that drugs that target a specific Ca^{2+} receptor without affecting those receptor types present on other Ca^{2+}-sensing cells are readily developed. Similarly, drugs active at two or more such Ca^{2+} receptors may also be developed.

Other Ca^{2+} Receptor Examples

The following examples demonstrate that, just as there are subtypes of receptors for molecular ligands, so too do there appear to be subtypes of Ca^{2+} receptors that

can be differentially affected by drugs. The parathyroid cell Ca^{2+} receptor senses levels of extracellular Ca^{2+} around 1.5 mM whereas the Ca^{2+} receptor on the osteoclast responds to levels around 10 mM (Fig. 22). Neomycin or spermine, which activate the parathyroid cell Ca^{2+} receptor, fail to affect the Ca^{2+} receptors on C-cells or osteoclasts (Figs. 21 and 23). These data constitute the first evidence for pharmacologically distinct subtypes of Ca^{2+} receptors and these data are being used to design and develop drugs that act selectively on a particular type of Ca^{2+} receptor. Indeed, testing of lead molecules demonstrate such cell-specific effects. For example, NPS 449, which elicits increases in $[Ca^{2+}]_i$ in osteoclasts is without effect on $[Ca^{2+}]_i$ in parathyroid cells. Conversely, NPS 447, which activates the parathyroid cell Ca^{2+} receptor, is effective in activating the osteoclast Ca^{2+} receptor only at concentrations 10-fold higher. Finally, agatoxin 489, although not very potent in activating the C-cell Ca^{2+} receptor (EC_{50} = 150 μM), is a quite potent activator of the parathyroid cell Ca^{2+} receptor (EC_{50} = 3 μM). The lead molecules presently under development will affect selectively the activity of a specific type of Ca^{2+}-sensing cell in vivo.

Drugs with less specificity might not necessarily be therapeutically undesirable. Thus, depressing osteoclast activity and stimulating calcitonin secretion are two different approaches to inhibiting bone resorption. Drugs that target the Ca^{2+} receptors on both of these cells might be very effective therapies for osteoporosis. Because PTH is also involved in regulating bone metabolism, drugs acting on the parathyroid cell Ca^{2+} receptor may also be useful in the treatment and/or prevention of osteoporosis.

Results of some test molecules are shown below. In Table 6, the comparative activity of calcimimetic molecules is shown. Bovine parathyroid cells and C-cells (rMT 6-23 cells) were loaded with fura-2, and rat osteoclasts with indo-1 and the potency of the indicated molecules to

mobilize intracellular Ca^{2+} determined by constructing cumulative concentration-response curves. Molecules listed as "inactive" did not alter $[Ca^{2+}]_i$ when tested at a concentration of 1 mM.

Table 6

	EC_{50} (μM)		
COMPOUND	PARATHYROID	OSTEOCLAST	C-CELL
NPS 568 (EE)	0.78	200	>300
NPS 568 (LE)	30	—	—
NPS 467 (EE)	2	>100	—
NPS 467 (LE)	>30	—	—
NPS 017	6	inactive	150
NPS 447	9	150	—
NPS 456*	15	200	>100
NPS 015	22	—	inactive
NPS 109	40	>300	5
NPS 449	inactive	150	—
NPS 468*	30	250	—
spermine	150	inactive	inactive
neomycin	40	inactive	inactive

*racemic mixture; "inactive" is defined as causing no increase in cytosolic Ca2+ at a concentration of 1-5 mM; EE is early eluting; LE is late eluting.

Example 11: Lead Molecules for Parathyroid Ca^{2+} Receptor

Structure-activity studies using polyamines and arylalkylamines led to the testing of molecules structurally akin to NPS 456. NPS 456 is a potent activator of the parathyroid cell Ca^{2+} receptor. This molecule is notable because it possess only one positive charge yet is much more potent than many polybasic molecules. Brief (2 min) pretreatment with PMA shifts the concentration-response curve for NPS 456 to the right. This indicates that NPS 456 acts through the same mechanism used by extracellular Ca^{2+}. NPS 456 evokes the mobilization of intracellular Ca^{2+} in Xenopus oocytes expressing the

parathyroid cell Ca^{2+} receptor, which demonstrates a direct action on the Ca^{2+} receptor (Fig. 33). Moreover, NPS 456 contains a chiral carbon, and therefore exists in two isomeric forms. Both isomers have been synthesized and examined for activity. The R-isomer, NPS 447, is 12-times more potent than the S-isomer, NPS 448 (Fig. 28). This is the first demonstration that a Ca^{2+} receptor can recognize an organic molecule in a stereospecific manner.

Because NPS 447 is a structurally simple molecule with selective and potent effects on the parathyroid cell Ca^{2+} receptor, structure-activity studies around this lead molecule are simple. The aim of these studies is to generate an array of related molecules with various characteristics from which the final development candidate can be selected. This effort has already revealed some of the structural domains of NPS 447 that contribute to activity and potency. For example, the novel compound NPS 459 is an analog of NPS 447 that is smaller (MW < 240) yet nearly as potent as the parent molecule, whereas several other analogs are relatively inactive. The most interesting molecules from this analog project can be put into <u>in vivo</u> testing for effects on PTH secretion and serum Ca^{2+} levels (see Examples 15, 16, 17, 18 and 23).

The novel compound NPS 467 is an even smaller molecule than NPS 447 yet the former is about 3-fold more potent than the latter in causing the mobilization of intracellular Ca^{2+} in parathyroid cells. Like NPS 456, NPS 467 is a racemic mixture. It is anticipated that Resolution of NPS 467 into its enantiomers provides an isomer of even greater potency than the racemic mixture (see Example 16). NPS 551 is another novel compound as potent as NPS 467 in causing the mobilization of intracellular Ca^{2+} in parathyroid cells. NPS 551 is a racemic mixture and it is anticipated that the resolution of NPS 551 into its enantiomers will result in an isomer that is more potent than the racemic mixture. Further structure-activity studies on molecules related to NPS 447, NPS 467, NPS 551 and NPS

568 are expected to yield pure isomers with greater potency than these molecules in their racemate forms.

Results obtained with NPS 456 (Fig. 33) show that it elicits oscillatory increases in Cl^- current at concentrations of 100 μM. NPS 456 is the most potent molecule activate on Xenopus oocytes expressing the parathyroid cell Ca^{2+} receptor. The results obtained in this expression system with neomycin and NPS 456 demonstrate that these molecules act directly on the Ca^{2+} receptor.

Example 12: Osteoclast Ca^{2+} Receptor Lead Molecules

The strategy used for elucidating the mechanism of action of extracellular Ca^{2+} on the osteoclast was similar to that proven effective in parathyroid cells. The first experiments examined the effects of La^{3+} on $[Ca^{2+}]_i$ in single rat osteoclasts loaded with the fluorimetric indicator indo-1. As described above, trivalent cations like La^{3+} are impermeant and block Ca^{2+} influx. Low micromolar concentrations of La^{3+} partially depressed extracellular Ca^{2+}-induced increases in $[Ca^{2+}]_i$ (Fig. 29). The demonstration of a La^{3+}-resistant increase in $[Ca^{2+}]_i$ provides evidence for the mobilization of intracellular Ca^{2+}. The results of these experiments parallel those obtained in parathyroid cells and suggest that similar mechanisms are used by extracellular Ca^{2+} to regulate $[Ca^{2+}]_i$ in both cell types.

Another series of experiments showed that extracellular Mn^{2+} evoked transient increases in $[Ca^{2+}]_i$ (Fig. 30(a)) that persisted in the absence of extracellular Ca^{2+} (Fig. 30B). These results are likewise indicative of the mobilization of intracellular Ca^{2+}. Although Mn^{2+} can enter some cells, it is unlikely to do so in the osteoclast because Mn^{2+} quenches the fluorescence of indo-1. Thus, if Mn^{2+} penetrated intracellularly, a decrease, not an increase in the fluorescent signal would be observed.

The results obtained with a variety of di- and trivalent cations are all consistent with the presence of a

Ca^{2+} receptor on the surface of the osteoclast that is coupled to the mobilization of intracellular Ca^{2+} and influx of extracellular Ca^{2+} through voltage-insensitive channels. Results show evidence for genetic material in human osteoclasts that encodes a Ca^{2+} receptor protein (see below). Transient increases in $[Ca^{2+}]_i$ resulting from the mobilization of intracellular Ca^{2+}, are sufficient to inhibit osteoclastic bone resorption in vitro. Thus, as with the parathyroid cell, activation of the Ca^{2+} receptor appears to be a viable means of inhibiting the activity of osteoclasts.

NPS 449 is presently the lead molecule for calcimimetic drugs on this receptor. It is a small molecule (MW < 425) and it mobilizes intracellular Ca^{2+} in rat osteoclasts with a EC_{50} of 200 μM (Figs. 31A and 31B). Although the potency of NPS 449 is relatively low, it has a simple structure with only one positive charge and is expected to have desirable pharmacodynamic and pharmacokinetic properties.

NPS 449 was examined for its ability to inhibit bone resorption in vitro. This was done by morphometric analysis of pit formation on thin slices of bovine cortical bone using scanning electron microscopy. Rat osteoclasts were incubated for 24 hours in slices of bone in the presence or absence of various concentrations of NPS 449. NPS 449 caused a concentration-dependent inhibition of bone resorption with an IC_{50} of 10 μM. The anticipated results provide the first demonstration that molecules acting at this novel site can inhibit osteoclastic bone resorption. More potent analogs of NPS 449 will be generated using synthetic chemistry and will be tested and assayed using the methods described herein.

Example 13: C-Cell Ca^{2+} Receptor Lead Molecules

Activation of the C-cell Ca^{2+} receptor stimulates the secretion of calcitonin which then acts on osteoclasts to inhibit bone resorption. Calcimimetic drugs selectively

affecting C-cells are useful in the treatment of osteoporosis.

The mobilization of intracellular Ca^{2+} is used as a functional index of Ca^{2+} receptor activity. The screening effort in C-cells is facilitated by the availability of cultured cell lines expressing the C-cell phenotype (e.g., rat medullary thyroid carcinoma cells; rMTC 6-23 cells). Selected for initial study were three arylalkylamine molecules. Two are naturally occurring (agatoxin 489 and agatoxin 505) and the other (NPS 019) is a synthetic agatoxin analog. Agatoxin 505 was found to block extracellular Ca^{2+}-induced increases in $[Ca^{2+}]_i$, with an IC_{50} of 3 μM. The inhibitory effect resulted from a block of the L-type voltage-sensitive Ca^{2+} channel present in these cells. In contrast, agatoxin 489 was found to mobilize intracellular Ca^{2+} in rMTC cells with an EC_{50} of 150 μM. This was the first organic molecule discovered that was found to activate the C-cell Ca^{2+} receptor. The synthetic analog, NPS 019, was even more potent and mobilized intracellular Ca^{2+} with an EC_{50} of 5 μM (Fig. 32). It is significant that the only structural difference between NPS 019 and agatoxin 489 is the presence or absence of an hydroxyl group. The fact that such subtle differences in structure affect profoundly the potency of molecules indicates a structurally specific binding site on the Ca^{2+} receptor. This, in turn, encourages the view that very potent and selective activators of Ca^{2+} receptors can be developed.

NPS 019, which is a small molecule (MW < 500), is a lead molecule for development of calcimimetics of the C-cell Ca^{2+} receptor and can be tested for its ability to stimulate calcitonin secretion in vitro. Subsequent in vivo testing will then determine the ability of this molecule to stimulate calcitonin secretion and inhibit bone resorption. These in vivo studies will be performed in rats. The results obtained in these studies, which are anticipated to be positive, will provide the first evi-

dence showing that a small organic molecule acting on a novel receptor can stimulate calcitonin secretion and depress bone resorption.

Example 14: Calcilytic Activity of NPS 021 on Parathyroid Cells

For a compound to be considered a calcilytic, it must block the effects of extracellular Ca^{2+} or a calcimimetic compound on an extracellular Ca^{2+}-sensing cell. An example of a calcilytic compound is NPS 021, the structure of which is provided in Fig. 1. In bovine parathyroid cells loaded with fura-2, NPS 021 blocks increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. The IC_{50} of NPS 021 for blocking this response is about 200 μM and, at concentrations around 500 μM, the increase in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+} is abolished. Significantly, NPS 021 does not by itself cause any change in $[Ca^{2+}]_i$ when tested at low $[Ca^{2+}]$ (0.5 mM; Fig. 37).

Example 15: NPS 467 Lowers Serum Ionized Calcium

Compounds shown to activate the bovine parathyroid cell Ca^{2+} receptor in vitro were tested for hypocalcemic activity in vivo. Male Sprague-Dawley rates (200 g) were maintained on a low calcium diet for one week prior to receiving test substance or vehicle as control. Blood was collected from the tail vein three hours after the intraperitoneal administration of NPS 467. Ionized Ca^{2+} in whole blood or serum was measured with a Ciba-Corning 634 Analyzer according to the instructions provided with the instrument. Serum total calcium, albumin and phosphate were measured by techniques well-known in the art.

NPS 467 caused a dose-dependent reduction in serum or whole blood Ca^{2+} (Fig. 38). The fall in blood Ca^{2+} at this time was paralleled by a proportional fall in the levels of blood total calcium. There was no change in serum albumin or phosphate levels at any of the doses examined. In preliminary studies, NPS 467, at doses effective in

lowering blood Ca^{2+}, caused a dose-dependent reduction in circulating levels of PTH (Fig. 39). The hypocalcemic effect of NPS 467 was maximal within three hours and returned toward control levels after 24 hours (Fig. 40).

NPS 467 (the EE isomer; see Example 16) was also effective in lowering serum ionized Ca^{2+} in rats maintained on a normal, calcium-replete diet. A single dose of NPS 467 (EE isomer, 10 mg/kg i.p.) caused a rapid fall in serum levels of ionized Ca^{2+} which were maximal by 1 hr (22%) decrease from the control level) and remained depressed at or near this level for up to 6 hours.

Example 16: NPS 467 Lowers Serum Ionized Calcium in a Stereospecific Manner

NPS 467 is a racemic mixture. Resolution of NPS 467 into its two enantiomers was achieved by separation on a chiral column. The EE-isomer (for "early eluting", see Example 21) was about 100-fold more potent than the LE-isomer (for "late-eluting") in activating the bovine parathyroid cell Ca^{2+} receptor in vitro as assessed by the ability of the enantiomers to evoke increases in the $[Ca^{2+}]_i$ in parathyroid cells (Fig. 41). Likewise, similar resolution of the novel compound NPS 568 into its enantiomers showed that the EE-isomer was 40-fold more potent than the LE-isomer in causing the mobilization of intracellular Ca^{2+} in bovine parathyroid cells (see Table 6, supra).

The isomers of NPS 467 were examined for effects on serum Ca^{2+} as in Example 15. Consistent with the in vitro results, the EE-isomer or NPS 467 proved to be more potent than the LE-isomer in lowering serum Ca^{2+} in vivo (Fig. 42; each compound was tested at a concentration of 5 mg/kg body weight).

Example 17: NPS 467 Lowers Serum Ionized Calcium in an in vivo Model of Secondary Hyperparathyroidism

An accepted and widely used animal model of secondary hyperparathyroidism arising from chronic renal failure is

the 5/6 nephrectomized rat. Animals receiving such surgery become initially hypocalcemic and, to maintain serum Ca^{2+} levels, there is a compensatory hyperplasia of the parathyroid glands and elevated levels of circulating PTH. Male Sprague-Dawley rats (250 g) received a 5/6 nephrectomy and were allowed to recover for 2 weeks. At this time they were normocalcemic (due to elevated levels of serum PTH). The administration of NPS 467 (EE isomer; 10 mg/kg i.p.) caused a rapid (within 2 hours) fall in serum ionized Ca^{2+} levels to 83% of controls in an animal model of secondary hyperparathyroidism. This suggests that compounds of this sort will effectively depress PTH secretion in patients with secondary hyperparathyroidism and hyperplastic parathyroid glands.

Example 18: NPS 467 Fails to Lower Serum Ionized Calcium Levels in Parathyroidectomized Animals

To determine the primary target tissue upon which NPS 467 acts to cause a hypocalcemic response, the parathyroid glands in rats were surgically removed. Animals receiving a total parathyroidectomy become hypocalcemic and are largely dependent upon dietary calcium to maintain serum Ca^{2+} homeostasis. Parathyroidectomized animals had serum ionized Ca^{2+} levels of 0.92 mM which fell gradually to 0.76 mM after 6 hours of fasting. The administration of a single dose of NPS 467 EE (10 mg/kg i.p.) did not cause any change in serum ionized Ca^{2+} levels over a period of 6 hours. These results demonstrate that intact parathyroid glands are required for the hypocalcemic effects of NPS 467 EE. The data additionally demonstrate that NPS 467 EE can target the parathyroid glands in vivo. The results are consistent with the view that NPS 467 EE acts on the parathyroid cell Ca^{2+} receptor in vivo to depress secretion of PTH and thereby cause serum levels of ionized Ca^{2+} to fall.

Example 19: NPS 467 Increases Intracellular Calcium in Human Parathyroid Glands

Dissociated parathyroid cells were prepared from a parathyroid adenoma obtained by surgery from a patient with primary hyperparathyroidism. The cells were loaded with fura-2 and $[Ca^{2+}]_i$ measured as described above. Both NPS 467 EE and NPS 568 EE caused concentration-dependent increase in $[Ca^{2+}]_i$. The EC_{50}'s for NPS 467 EE and NPS 568 EE were 20 and 3 μM, respectively. Both these compounds are thus able to increase $[Ca^{2+}]_i$ in pathological human tissue and would thus be expected to decrease serum levels of PTH and Ca^{2+} in patients with primary hyperparathyroidism.

Example 20: Mechanism of Action of NPS 467 at the Parathyroid Cell Calcium Receptor

Dissociated bovine parathyroid cells were used to further explore the mechanism of action of NPS 467 at the receptor level. In the presence of 0.5 mM extracellular Ca^{2+}, NPS 467 EE caused a rapid and transient increase in $[Ca^{2+}]_i$ which persisted in the presence of 1 μM La^{3+} and was partially depressed by pretreatment with PMA (100 nM for 2 min.). Moreover, NPS 467 (EE isomer, 30 μM) caused a rapid increase in Cl^- conductance in Xenopus oocytes injected with parathyroid cell mRNA. All these results are consistent with an action of NPS 467 on the Ca^{2+} receptor. However, the cytosolic Ca^{2+} response to NPS 467 was abolished when parathyroid cells were suspended in Ca^{2+}-free buffer. This suggests that NPS 467 cannot, by itself, cause the mobilization of intracellular Ca^{2+}. It does, however, elicit responses in parathyroid cells and in oocytes when a small amount of extracellular Ca^{2+} is present. This suggests that partial occupancy of the Ca^{2+}-binding site is required for NPS 467 to elicit a response. To test this hypothesis, parathyroid cells were suspended in Ca^{2+}-free buffer and exposed to a submaximal concentration of neomycin. Neomycin was used because it mimics, in

nearly all respects, the effects of extracellular Ca^{2+} on parathyroid cells and on Xenopus oocytes expressing the parathyroid cell Ca^{2+} receptor. The addition of 10 μM neomycin did not by itself cause an increase in $[Ca^{2+}]_i$ under these conditions. However the subsequent addition of NPS 467 EE (30 μM) now elicited a transient increase in $[Ca^{2+}]_i$ which, because there was no extracellular Ca^{2+} present, must have come from the mobilization of intracellular Ca^{2+}. When cells bathed in Ca^{2+}-free buffer were exposed to 30 μM NPS 467 there was no increase in $[Ca^{2+}]_i$. This concentration of NPS 467 is maximally effective in increasing $[Ca^{2+}]_i$ when extracellular Ca^{2+} (0.5 mM) is present. However, the subsequent addition of 10 μM neomycin now evoked a transient increase in $[Ca^{2+}]_i$. Presumably, neomycin binds to the same site as extracellular Ca^{2+} and can functionally substitute for it. Using a submaximal concentration, which by itself causes no response, achieves partial occupancy of the Ca^{2+}-binding site and allows activation of the Ca^{2+} receptor by NPS 467.

Additional studies to further define the mechanism of action of NPS 467 were performed. The cells were once again suspended in Ca^{2+}-free buffer to insure that any observed increase in $[Ca^{2+}]_i$ resulted from the mobilization of intracellular Ca^{2+}. In these experiments, however, a maximally effective concentration (100 μM) of neomycin was used. In the absence of extracellular Ca^{2+}, 100 μM neomycin evoked a rapid and transient increase in $[Ca^{2+}]_i$. The subsequent addition of 30 μM NPS 467 EE did not cause an increase in $[Ca^{2+}]_i$. In the converse experiment, 30 μM NPS 467 EE was added before 100 μM neomycin. As expected, NPS 467 EE did not cause any increase in $[Ca^{2+}]_i$. It did not, however, affect the increase in $[Ca^{2+}]_i$ evoked by the subsequent addition of 100 μM neomycin. These results, obtained with maximally effective concentrations of NPS 467 and neomycin, suggest that these two compounds do not act at the same site. Rather, the results can be suffi-

ciently explained by postulating two separate sites on the Ca^{2+} receptor, one to which extracellular Ca^{2+} and neomycin bind, and another to which NPS 467 and structurally related compounds (such as NPS 568) bind. Ligand binding to the former site can result in full activation of the Ca^{2+} receptor whereas ligand binding to the latter site can only occur and/or be functionally relevant when the extracellular Ca^{2+}-binding site is occupied to some as yet undefined degree. It is possible that ligand binding to the extracellular Ca^{2+}-binding site exposes a previously occluded binding site for NPS 467. It appears that the NPS 467-binding site is an allosteric site that augments receptor activation in response to ligand binding at the extracellular Ca^{2+}-binding site.

The data demonstrate that the parathyroid cell Ca^{2+} receptor possesses at least two distinct sites for organic ligands. One site binds the physiological ligand, extracellular Ca^{2+}, and certain organic polycations like neomycin. Binding to this site result in full activation of the Ca^{2+} receptor, an increase in $[Ca^{2+}]_i$, and the inhibition of PTH secretion. NPS 467 defines a previously unrecognized binding site on the Ca^{2+} receptor. Binding to this site can only occur and/or results in full activation of the Ca^{2+} receptor when the extracellular Ca^{2+}-binding site is partially occupied. Ligands acting at either site are effective in suppressing serum Ca^{2+} levels in vivo.

Example 21: Preparation of NPS 467

In a 250 ml round bottom flask, 10.0 g (100 mmoles) 3'-methoxy acetophenone and 13.5 g (100 mmoles) 3-phenylpropylamine were mixed and treated with 125 mmoles (35.5 g) titanium(IV) isopropoxide. The reaction mixture was stirred 30 minutes at room temperature under a nitrogen atmosphere. After this time 6.3 g (100 mmoles) sodium cyanoborohydride in 100 ml ethanol was added dropwise over the course of 2 minutes. The reaction was stirred room temperature under nitrogen for 16 hours. After this time

the reaction mixture was transferred to a 2 L separatory funnel with 1.5 L ethyl ether and 0.5 L water. The phases were equilibrated and the ether layer removed. The remaining aqueous phase was thoroughly extracted with four 1 L portions of ether. The washes were combined, dried over anhydrous potassium carbonate and reduced to a clear, light amber oil.

TLC analysis of this material on silica using chloroform-methanol-isopropylamine (100:5:1) showed product at Rf 0.65 with traces of the two starting materials at Rf 0.99 (3' methoxy acetophenone) and Rf 0.0 (3-phenylpropylamine).

The reaction mixture was chromatographed through silica (48 x 4.6 cm) using a gradient of chloroform-methanol-isopropylamine (99:1:0.1) to (90:10:0.1) which yielded 13.66 g of purified NPS 467. This material was dissolved in Hexane-isopropanol (99:1) containing 0.1% diethylamine to yield a solution with a concentration of 50 mg/ml. Chiral resolution was accomplished by chromatography of 4 ml of this solution (200 mg, maximum to achieve separation) through ChiralCel OD (25 x 2 cm) using 0.7% isopropylamine, 0.07% diethylamine in hexane at 100 ml/min, monitoring optical density at 260 nm. Under these conditions (with injections of 100 mg material) the early eluting isomer (NPS 467EE) began to emerge from the column at ~ 26 min, the late eluting isomer (NPS 467LE) began to emerge at ~34 minutes. Baseline resolution was accomplished with under these conditions. Each optical isomer (free base) was converted to the corresponding hydrochloride salt by dissolving 3 g of the free base in 100 ml ethanol and treating with it with 100 ml water containing 10 molar equivalents HCl. Lyophilization of this solution yielded a white solid.

Example 22: Preparation of NPS 568

NPS 568 was prepared using the methods described in Example 21 substituting an equivalent amount of 3-(2-

chlorophenyl)propylamine for 3-phenylpropylamine. It was found that allowing the mixture of 3'-methoxyacetophenone, 3-(2-chlorophenyl)propylamine and titanium(IV) isopropoxide to stir for 5 hours prior to treatment with $NaCNBH_3$/EtOH resulted in significantly greater yield (98%).

Example 23: NPS 467 Lowers Serum Ionized Calcium When Administered Orally

Rats (male, Sprague-Dawley, 250-300 g) were fed standard rat chow and fasted overnight prior to the experiment. NPS 467 (EE isomer) was suspended in corn oil and administered as a single oral dose through a gavage needle. Three hours later a sample of blood was taken from the tail vein and assessed for ionized Ca^{2+} levels. Fig. 44 shows that NPS 467 EE caused a dose-dependent reduction in serum levels of ionized Ca^{2+} when administered orally.

Other embodiments are within the following claims.

Claims:

1. A pharmaceutical composition comprising a molecule which mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell, or blocks an increase in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+}, said molecule having an EC_{50} of less than or equal to 5 μM, wherein said molecule is not protamine.

2. The pharmaceutical composition of claim 1 wherein said increase in $[Ca^{2+}]_i$ in a cell is transient, having a duration of less than thirty seconds.

3. The pharmaceutical composition of claim 1 or 2 wherein said increase in $[Ca^{2+}]_i$ is rapid, occurring within thirty seconds.

4. The pharmaceutical composition of claim 1 wherein said molecule evokes a sustained increase in $[Ca^{2+}]_i$, having a duration greater than thirty seconds.

5. The pharmaceutical composition of claim 1 wherein said molecule further evokes an increase in inositol-1,4,5-trisphosphate or diacylglycerol.

6. The pharmaceutical composition of claim 5 wherein said increase in said inositol-1,4,5-trisphosphate or diacylglycerol occurs within 60 seconds.

7. The pharmaceutical composition of claim 2 wherein said transient increase is diminished by pretreatment of said cell for less than 10 minutes with an activator of protein kinase C.

8. The pharmaceutical composition of claim 7 wherein said activator of protein kinase C is selected from the group consisting of phorbol myristate acetate, mezerein and (-)-indolactam V.

9. The pharmaceutical composition of claim 1 wherein said molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation.

10. The pharmaceutical composition of claim 2 wherein said transient increase in $[Ca^{2+}]_i$ is reduced by pretreatment of said cell for ten minutes with 10 mM sodium fluoride.

11. The pharmaceutical composition of claim 1 wherein said cell is a parathyroid cell and said molecule inhibits parathyroid hormone secretion from said cell.

12. The pharmaceutical composition of claim 1 wherein said molecule elicits an increase in Cl^- conductance in a Xenopus oocyte injected with mRNA from a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts.

13. The pharmaceutical composition of claim 1 wherein said molecule mobilizes intracellular Ca^{2+} to cause said increase in $[Ca^{2+}]_i$.

14. The pharmaceutical composition of claim 1 wherein said cell is a C-cell or an osteoclast and said molecule inhibits bone resorption in vivo.

15. The pharmaceutical composition of claim 1 wherein said cell is a C-cell and said molecule stimulates calcitonin secretion in vitro or in vivo.

16. The pharmaceutical composition of claim 1 wherein said molecule is either a calcimimetic or calcilytic having an EC_{50} or IC_{50} at a Ca^{2+} receptor of less than or equal to 5 μM.

17. The pharmaceutical composition of claim 1 wherein said molecule has an EC_{50} less than or equal to 5 μM at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts.

18. The pharmaceutical composition of claim 1 or 16 or 17 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 1 μM.

19. The pharmaceutical composition of claim 1 or 16, or 17 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 100 nanomolar.

20. The pharmaceutical composition of claim 1 or 16, or 17 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 10 nanomolar.

21. The pharmaceutical composition of claim 1 or 16, wherein said molecule has an EC_{50} or IC_{50} less than or equal to 1 nanomolar.

22. The pharmaceutical composition of claim 1, 16 or 17 wherein said molecule is positively charged at physiological pH.

23. The pharmaceutical composition of claim 22 wherein said molecule is selected from the group consisting of branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines.

24. The pharmaceutical composition of claim 23 wherein said branched polyamine has the formula

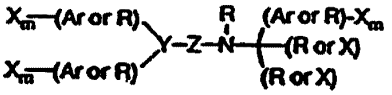
$$H_2N-(CH_2)_j-(NR_i-(CH_2)_j)_k-NH_2$$

wherein k is an integer from 1 to 10,

each j is the same or different and is an integer from 2 to 20, and

each R_i is the same or different and is selected from the group consisting of hydrogen and $-(CH_2)_j-NH_2$, wherein j is as defined above, and wherein at least one R_i is not hydrogen.

25. The pharmaceutical composition of claim 1, 16 or 17 wherein said molecule has the formula

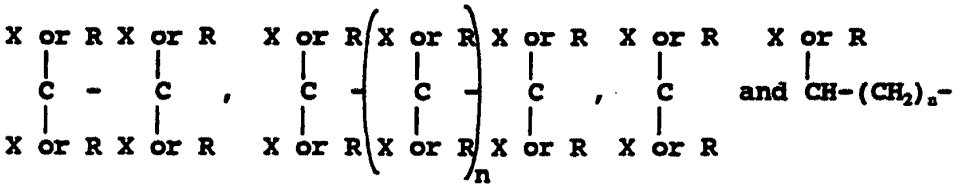


wherein each X is selected (independently) from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN-, NO_2, and CH_3CH_2; Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen, sulfur,

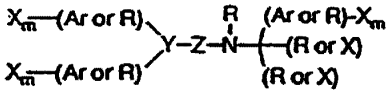


where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is either a calcimimetic or calcilytic.

26. The pharmaceutical composition of claim 25 wherein said hydrophobic entity is selected from the group consisting of phenyl, 2-, 3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2-quinolinyl, 2- or 3-indolyl, benzyl, and phenoxy.

27. A pharmaceutical composition comprising a molecule having the formula

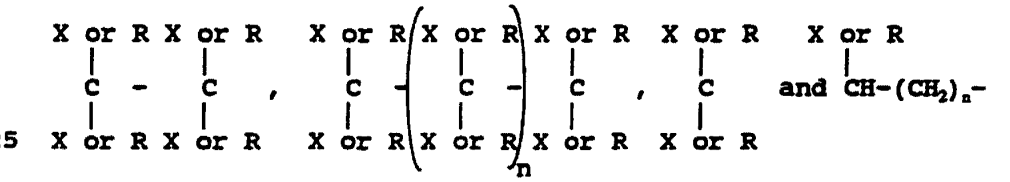


wherein each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, and CH_3CH_2; Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen sulfur,



where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is not NPS 447 or NPS 449.

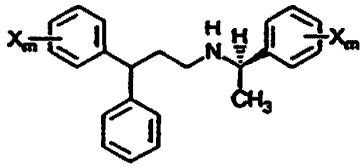
28. The pharmaceutical composition of claim 27 wherein said molecule is NPS 467.

29. The pharmaceutical composition of claim 25 wherein said molecule is NPS 459, NPS 467, NPS 551 or NPS 568.

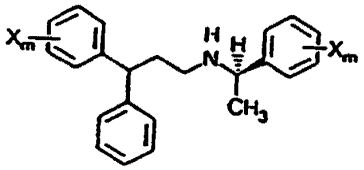
30. The pharmaceutical composition of claim 25, wherein said molecule is an R-diphenylpropyl-α-phenethylamine derivative.

31. The pharmaceutical composition of claim 27, wherein said molecule is an R-diphenylpropyl-α-phenethylamine derivative.

32. The pharmaceutical composition of claim 30 wherein said molecule has the formula:



33. The pharmaceutical composition of claim 31 wherein said molecule has the formula:



34. The pharmaceutical composition of claim 32 wherein each X is independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

35. The pharmaceutical composition of claim 33 wherein each X is independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

36. A method for treating a patient having a disease characterized by an abnormal $[Ca^{2+}]$ or $[Ca^{2+}]_i$ in one or more cells or in the blood or plasma, comprising the step of administering to said patient a therapeutically effective amount of a molecule which mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell, or blocks an increase in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+}.

37. The method of claim 36 wherein said molecule has an EC_{50} of less than or equal or 5 μM.

38. The method of claim 36 wherein said molecule is not protamine.

39. The method of claim 36 wherein said molecule interacts at a Ca^{2+} receptor as a calcimimetic or calcilytic.

40. The method of claim 36 wherein said increase in $[Ca^{2+}]_i$ in a cell is transient, having a duration of less than thirty seconds.

41. The method of claim 36 or 40 wherein said increase in $[Ca^{2+}]_i$ is rapid, occurring within thirty seconds.

42. The method of claim 36 wherein said molecule evokes a sustained increase in $[Ca^{2+}]_i$, having a duration greater than thirty seconds.

43. The method of claim 36 wherein said molecule further evokes an increase in inositol-1,4,5-trisphosphate or diacylglycerol.

44. The method of claim 43 wherein said increase in said inositol-1,4,5-trisphosphate or diacylglycerol occurs within less than 60 seconds.

45. The method of claim 40 wherein said transient increase is diminished by pretreatment of said cell with an activator of protein kinase C.

46. The method of claim 45 wherein said activator of protein kinase C is selected from the group consisting of phorbol myristate acetate, mezerein and (-)-indolactam V.

47. The method of claim 36 wherein said molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation.

48. The method of claim 40 wherein said transient increase in $[Ca^{2+}]_i$ is reduced by pretreatment of said cell for ten minutes with 10 mM sodium fluoride.

49. The method of claim 36 wherein said cell is a parathyroid cell and said molecule inhibits parathyroid hormone secretion from said cell.

50. The method of claim 36 wherein said molecule elicits an increase in Cl^- conductance in a Xenopus oocyte injected with mRNA from a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxta-glomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts.

51. The method of claim 36 wherein said molecule mobilizes Ca^{2+} within the cell to cause said increase in $[Ca^{2+}]_i$.

52. The method of claim 36 wherein said cell is a C-cell or an osteoclast and said molecule inhibits bone resorption in vivo.

53. The method of claim 36 wherein said cell is a C-cell and said molecule stimulates calcitonin secretion in vitro or in vivo.

54. The method of claim 36 wherein said molecule is either a calcimimetic or calcilytic having an EC_{50} or IC_{50} at a Ca^{2+} receptor of less than or equal to 5 μM.

55. The method of claim 36 wherein said molecule has an EC_{50} less than or equal to 5 μM at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells and placental trophoblasts.

56. The method of claim 36, 54 or 55 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 1 μM.

57. The method of claim 36, 54 or 55 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 100 nanomolar.

58. The method of claim 36, 54, or 55 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 10 nanomolar.

59. The method of claim 38, 54 or 55, wherein said molecule has an EC_{50} or IC_{50} less than or equal to 1 nanomolar.

60. The method of claim 36, 54 or 55 wherein said molecule is positively charged at physiological pH.

61. The method of claim 60 wherein said molecule is selected from the group consisting of branched or cyclic polyalkylamines, positively charged polyamino acids, and arylamines.

62. The method of claim 61 wherein said branched polyamine has the formula

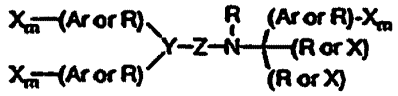
$$H_2N-(CH_2)_j-(NR_i-(CH_2)_j)_k-NH_2$$

wherein k is an integer from 1 to 10,

each j is the same or different and is an integer from 2 to 20, and

each R_i is the same or different and is selected from the group consisting of hydrogen and $-(CH_2)_j-NH_2$, wherein j is as defined above, and wherein at least one R_i is not hydrogen.

63. The method of claim 36, 54 or 55 wherein said molecule has the formula



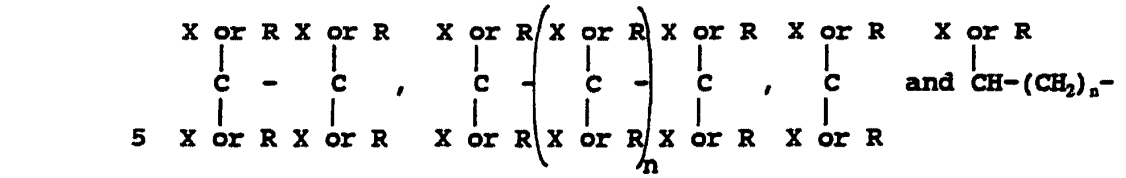
wherein each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, and CH_3CH_2;

Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen, sulfur,

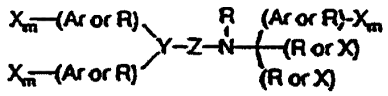


where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is either a calcimimetic or calcilytic.

64. The method of claim 63 wherein said hydrophobic entity is selected from the group consisting of phenyl, 2-3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2-quinolinyl, 2- or 3-indolyl, benzyl, and phenoxy.

65. The method of claim 36, 54 or 55 wherein said molecule has the formula



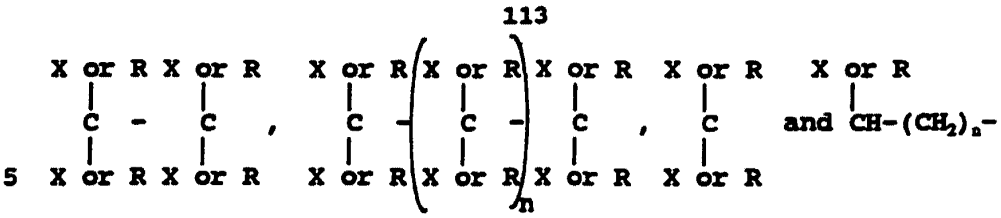
wherein each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, and CH_3CH_2;

Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen, sulfur,



where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is not NPS 447 or NPS 449.

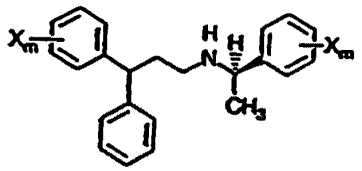
66. The method of claim 65 wherein said molecule is NPS 467 or NPS 019.

67. The method of claim 63 wherein said molecule is NPS 467, NPS 019 or NPS 456.

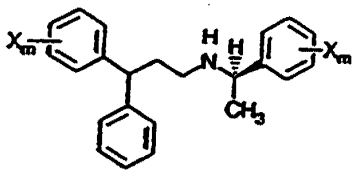
68. The method of claim 63, wherein said molecule is an R-diphenylpropyl-α-phenethylamine derivative.

69. The method of claim 65, wherein said molecule is an R-diphenylpropyl-α-phenethylamine derivative.

70. The method of claim 68 wherein said molecule has the formula:



71. The method of claim 69 wherein said molecule has the formula:



72. The method of claim 70 wherein each X is independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

73. The method of claim 71 wherein each X is independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

74. The method of claim 36 wherein said patient has a disease characterized by an abnormal level of one or more ions or substances the level of which is regulated or affected by activity of one or more Ca^{2+} receptors.

75. The method of claim 74, wherein said molecule is active on a Ca^{2+} receptor of a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, parafollicular thyroid cells, and placental trophoblasts.

76. The method of claim 36, wherein said patient has a disease selected from the group consisting of primary and secondary hyperparathyroidism, Paget's disease, hypercalcemia malignancy, osteoporosis and hypertension.

77. The method of claim 75 wherein said molecule is active only at one or more said Ca^{2+} receptors and not all said Ca^{2+} receptors.

78. The method of claim 36 wherein said molecule reduces the level of parathyroid hormone in the serum of said patient.

79. The method of claim 78, wherein said level of parathyroid hormone is reduced to that level present in a normal individual.

80. The method of claim 78 wherein said level is reduced to a degree sufficient to cause a decrease in plasma Ca^{2+}.

81. The method of claim 78 wherein said molecule is provided in an amount sufficient to have a therapeutically relevant effect on said patient.

82. The method of claim 36 wherein said molecule blocks an increase in $[Ca^{2+}]_i$ within the cell evoked by extracellular $[Ca^{2+}]$.

83. A method for diagnosis of a disease or condition in a patient, comprising the steps of identifying the number and/or location of one or more Ca^{2+} receptors within said patient and comparing said number and/or location with that observed in normal patients as an indication of the presence of a said disease or condition.

84. The method of claim 83 wherein said method is an immunoassay in which an antibody to a Ca^{2+} receptor is used to identify the number or location of said Ca^{2+} receptors.

85. The method of claim 83 wherein said assay comprises providing a labelled calcimimetic or calcilytic molecule which binds to a Ca^{2+} receptor.

86. The method of claim 83 wherein said disease is a cancer.

87. The method of claim 86 wherein said cancer is an ectopic tumor in the parathyroid.

88. The method of claim 83 wherein said condition is characterized by an increased number or activity of osteoclasts in bone.

89. A method for treatment of osteoporosis, hyperparathyroidism or hypertension, comprising the steps of administering a therapeutically effective amount of a molecule which mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell, or blocks an increase in $[Ca^{2+}]_i$ (elicited by extracellular Ca^{2+}).

90. A method for identifying a molecule useful as a therapeutic molecule, comprising the steps of screening a potentially useful molecule for its ability to mimic the activity of extracellular Ca^{2+} in a cell, or to block an increase in $[Ca^{2+}]_i$ (elicited by extracellular Ca^{2+}), and determining whether said molecule has an EC_{50} of less than or equal to 5 μM.

91. A recombinant Ca^{2+} receptor.

92. A cell comprising a recombinant Ca^{2+} receptor.

93. A method for identifying a useful calcimimetic molecule comprising the step of identifying a molecule which mimics one or more activities of Ca^{2+} at a first Ca^{2+} receptor but not at a second Ca^{2+} receptor.

94. A method for identifying a useful calcilytic molecule comprising the step of identifying a molecule which blocks one or more activities of Ca^{2+} at a first Ca^{2+} receptor but not at a second Ca^{2+} receptor.

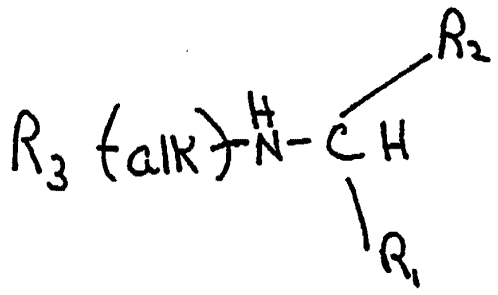
95. The method of claim 93 or 94 wherein said method includes providing a recombinant Ca^{2+} receptor useful in said identifying step.

96. Purified nucleic acid encoding a Ca^{2+} receptor.

97. The molecules NPS 459, NPS 467, NPS 544, NPS 551 and NPS 568.

98. The pharmaceutical composition of claim 25 wherein said molecule is NPS 447, NPS 448, NPS 449 or NPS 456.

99. A compound of the formula



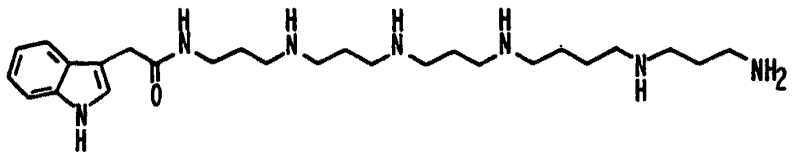
wherein alk is straight or branched-chain alkylene of from 1 to 6 carbon atoms; R_1 is lower alkyl of from 1 to 3 carbon atoms or lower haloalkyl of from 1 to 3 carbon atoms substituted with from 1 to 7 halogen atoms; and R_2 and R_3 are independently selected monocyclic or bicyclic carbocyclic aryl or cycloalkyl groups, having 5- or 6-membered rings optionally substituted with 1 to 5 substituents independently selected form lower alkyl of 1 to 3 carbon atoms, lower haloalkyl of 1 to 3 carbon atoms substituted with 1 to 7 halogen atoms, lower alkoxy of 1 to 3 carbon atoms, halogen, nitro, amino, alkylamino, amido, lower alkylamido of 1 to 3 carbon atoms, cyano, hydroxy, acyl of 2 to 4 carbon atoms or lower thioalkyl of 1 to 3 carbon atoms and pharmaceutically acceptable salts and acid addition salts thereof.

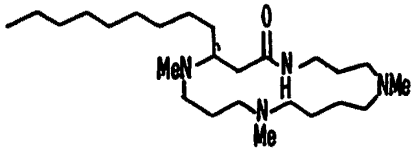
100. A compound according to claim 99 wherein alk is n-propylene.

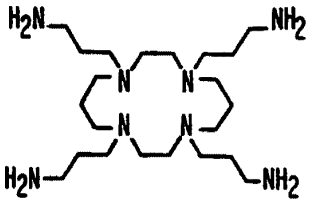
101. A compound according to claim 100 wherein R_1 is methyl.

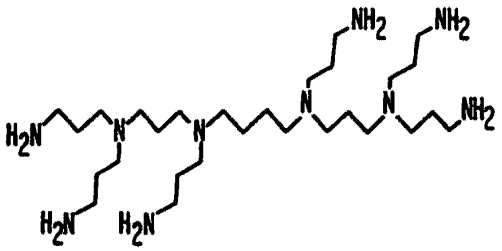
102. A compound according to claim 101 wherein R_2 and R_3 are independently selected optionally substituted phenyl groups.

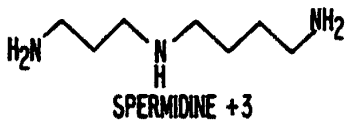
FIG. 1a.


NPS 019

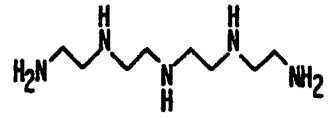

BUDMUNCHIAMINE A (+3)

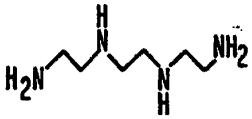

NPS 383 (+8 EC_{50}=150μM)

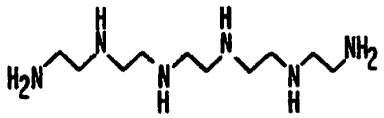

NPS 381 (+10 EC_{50}=100μM)

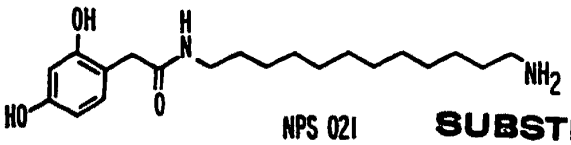

SPERMIDINE +3


1,12-DIAMINODODECANE -2
(DADD)

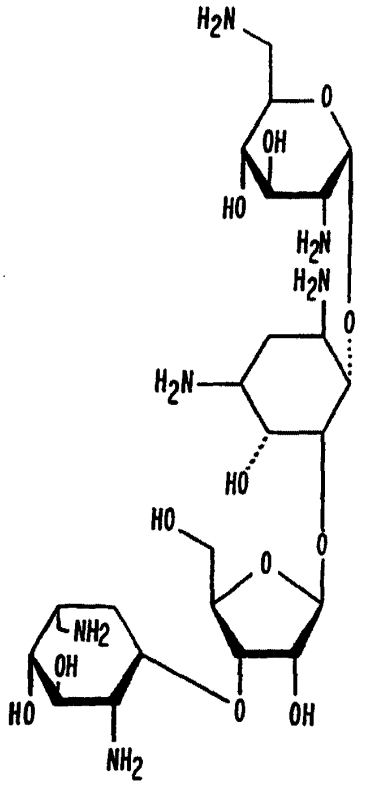

TETRAETHYLENEPENTAMINE (+5 EC_{50}=2500μM)


TRIETHYLENETETRAMINE (+4 EC_{50}=8000μM)

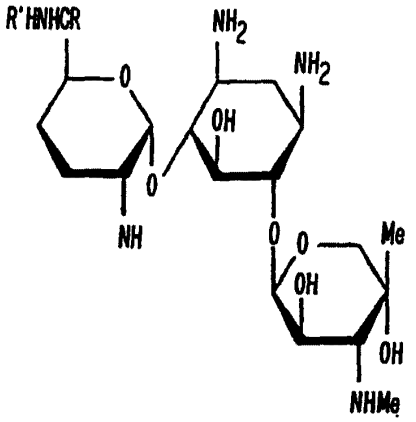

PENTAETHYLENEHEXAMINE (+6 EC_{50}=500μM)


NPS 021

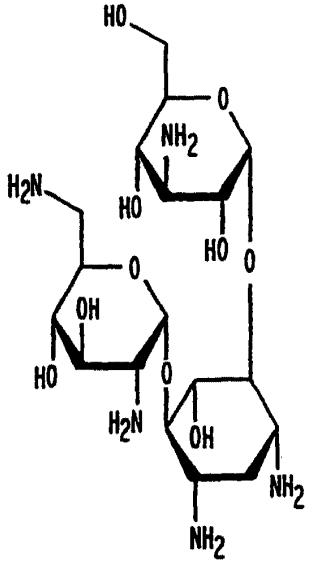
SUBSTITUTE SHEET



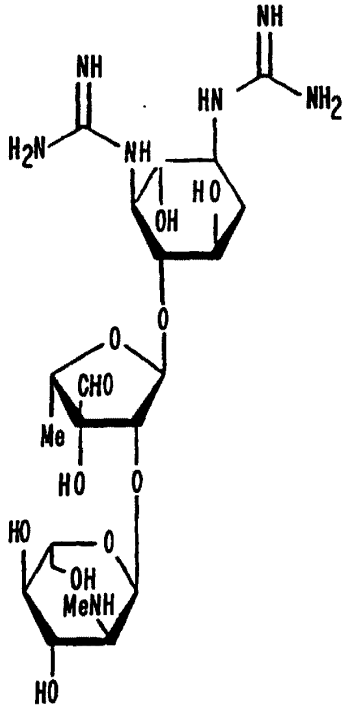
Neomycin B (+6)



Gentamicin (Complex +5)
C1 R = R' = Me
C2 R = Me; R' = H
C1aR = R' = H

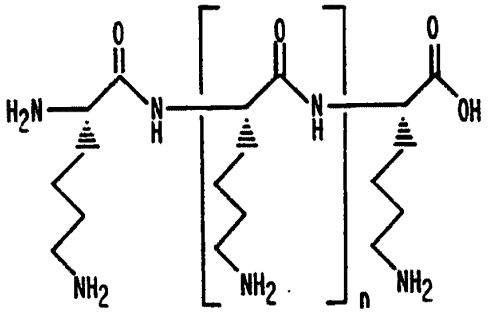


Bekanamycin (+5)

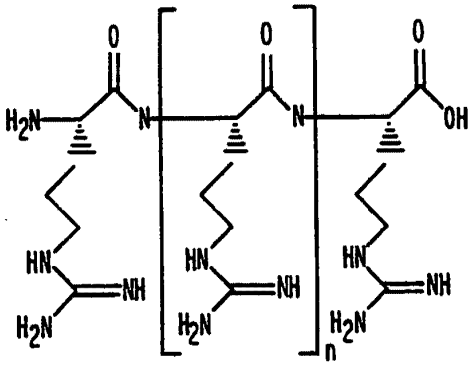


Streptomycin (+3)

FIG. 1b.



poly-Lys (27 kD) n=210 +210
poly-Lys (14 kD) n=110 +110
poly-Lys (38 kD) n=55 +55

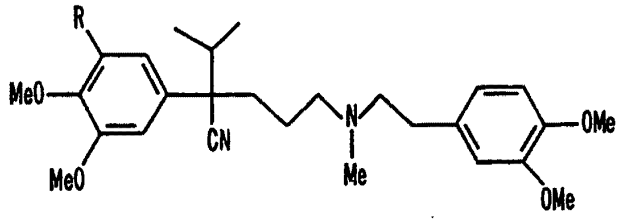


poly-Arg (100 kD) n=640 +640
poly-Arg (40 kD) n=256 +256

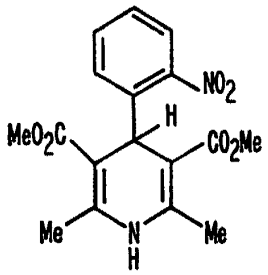
PROTAMINE +21

H N-Pro-Arg-Arg-Arg-Arg-Ser-Ser-Ser-Arg-Pro-Val-Arg-Arg-Arg-Arg-Arg-
Pro-Arg-Val-Ser-Arg-Arg-Arg-Arg-Arg-Arg-Gly-Gly-Arg-Arg-Arg-Arg-OH

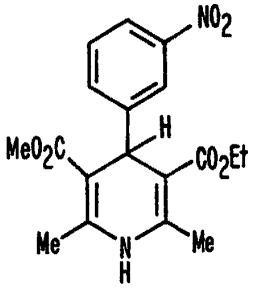
FIG. 1c.



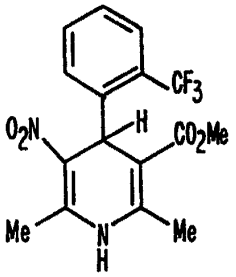
VERAPAMIL R=H +1
D-600 R=OMe +1



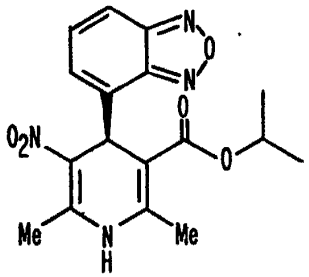
NIFEDIPINE +1



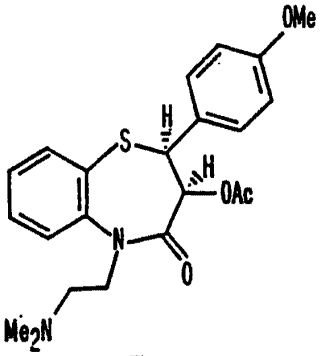
NITRENDIPINE +1



BAY K 8644 +1

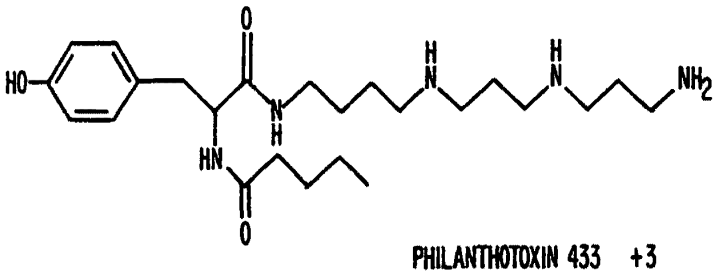


202-791 +1

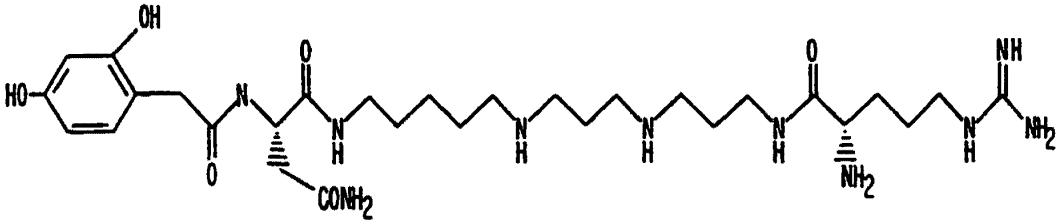


DILTIAZEM +1

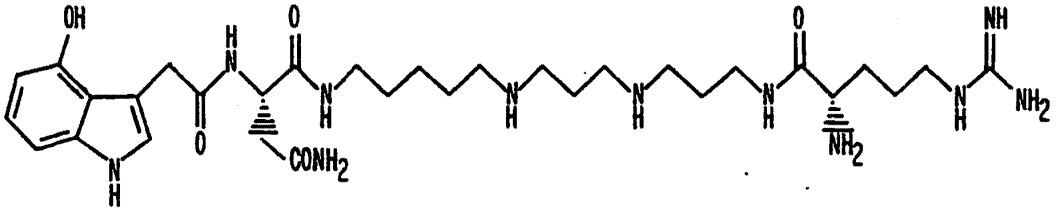
FIG. Id.



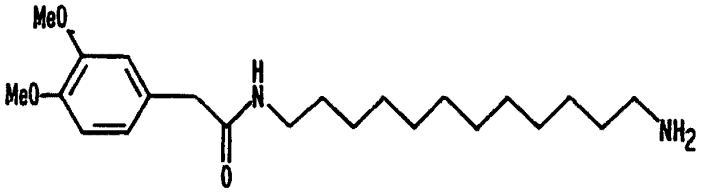
PHILANTHOTOXIN 433 +3



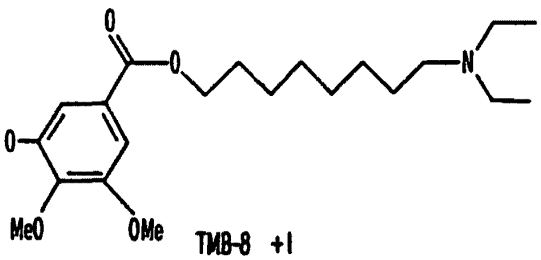
ARGIOTOXIN 636 +4



ARGIOTOXIN 659 +4



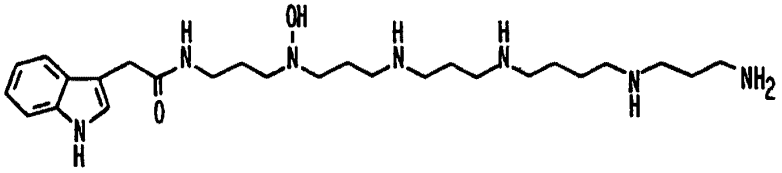
NPS 384

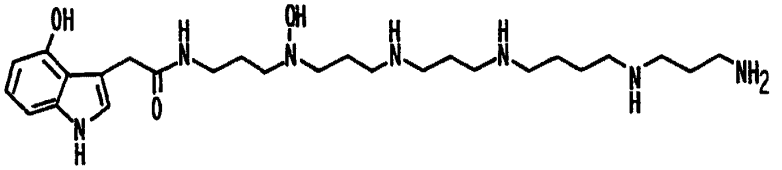


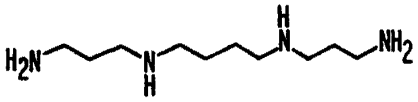
TMB-8 +1

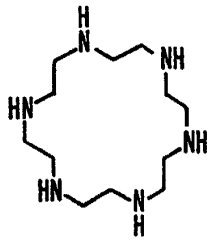
FIG. 1e.

SUBSTITUTE SHEET









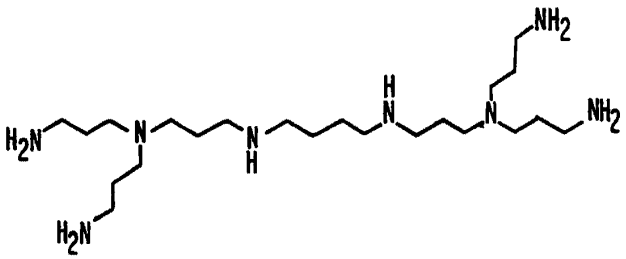


FIG. 1f.

SUBSTITUTE SHEET

FIG. 2.

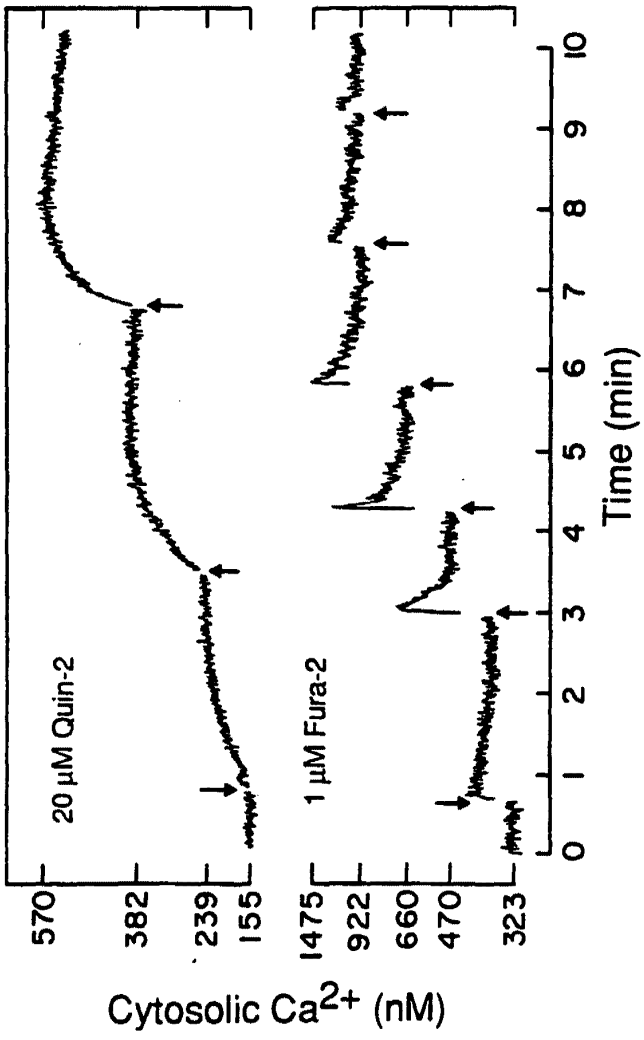

20 µM Quin-2
1 µM Fura-2
570
382
239
155
1475
922
660
470
323
0
1
2
3
4
5
6
7
8
9
10
Time (min)
Cytosolic Ca^{2+} (nM)

FIG. 3a.

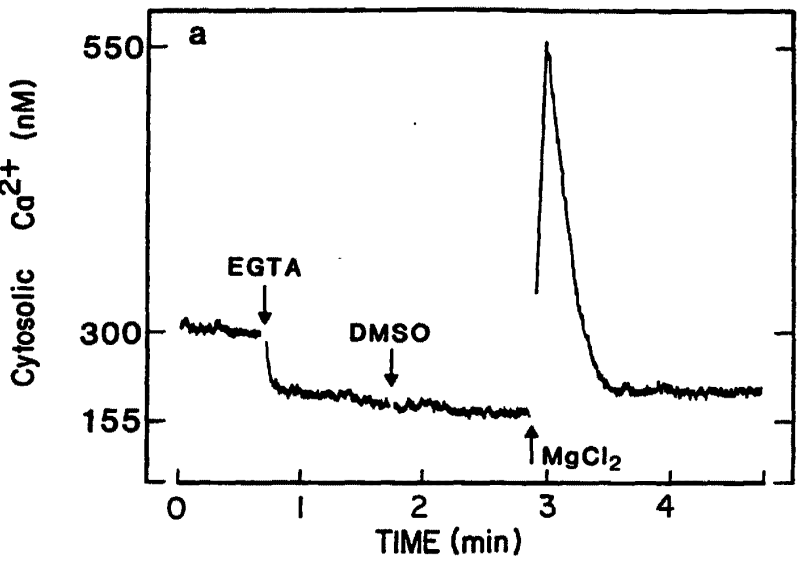


FIG. 3b.

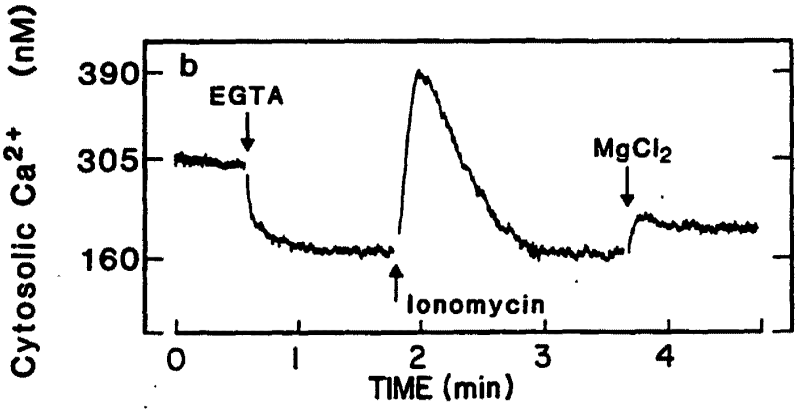
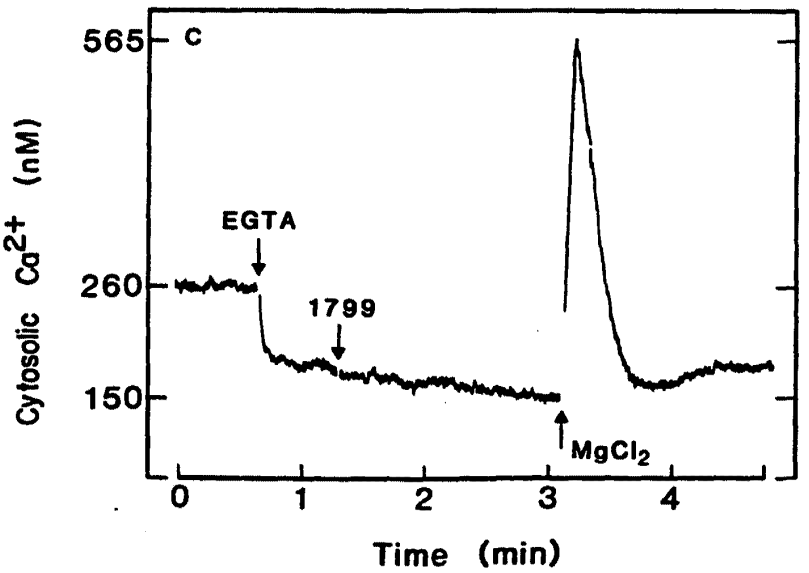


FIG. 3c.



SUBSTITUTE SHEET

FIG. 4a.

FIG. 4b.

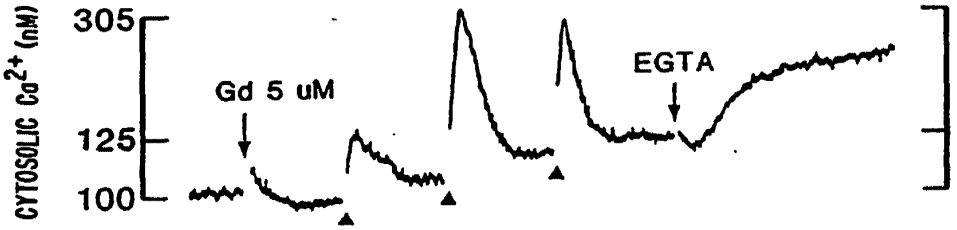


FIG. 4c.

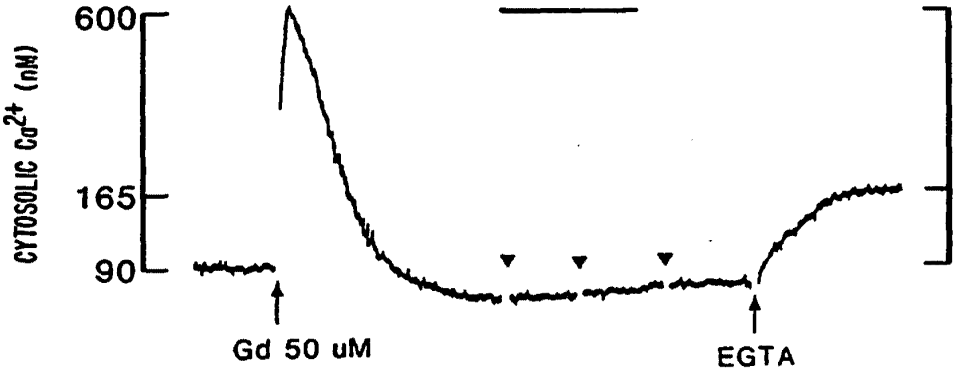


FIG. 5c.

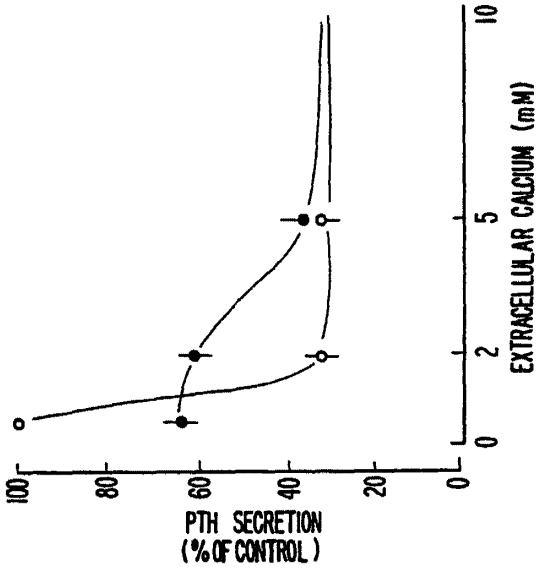


FIG. 5b.

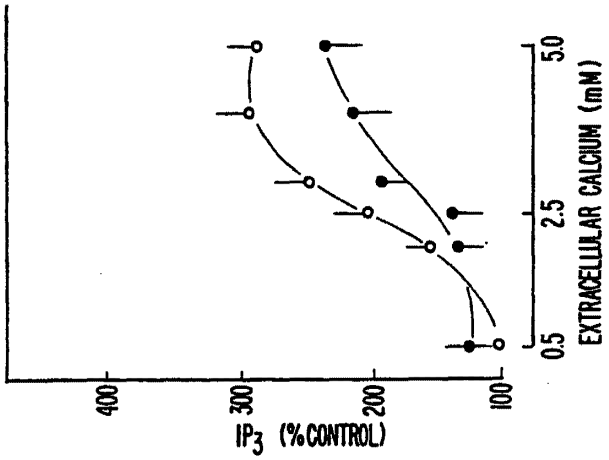
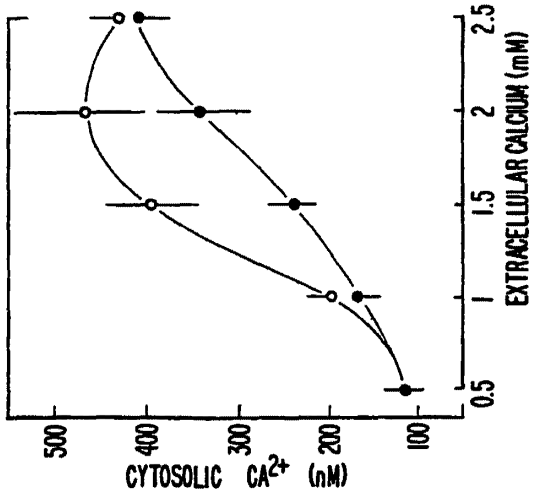


FIG. 5a.



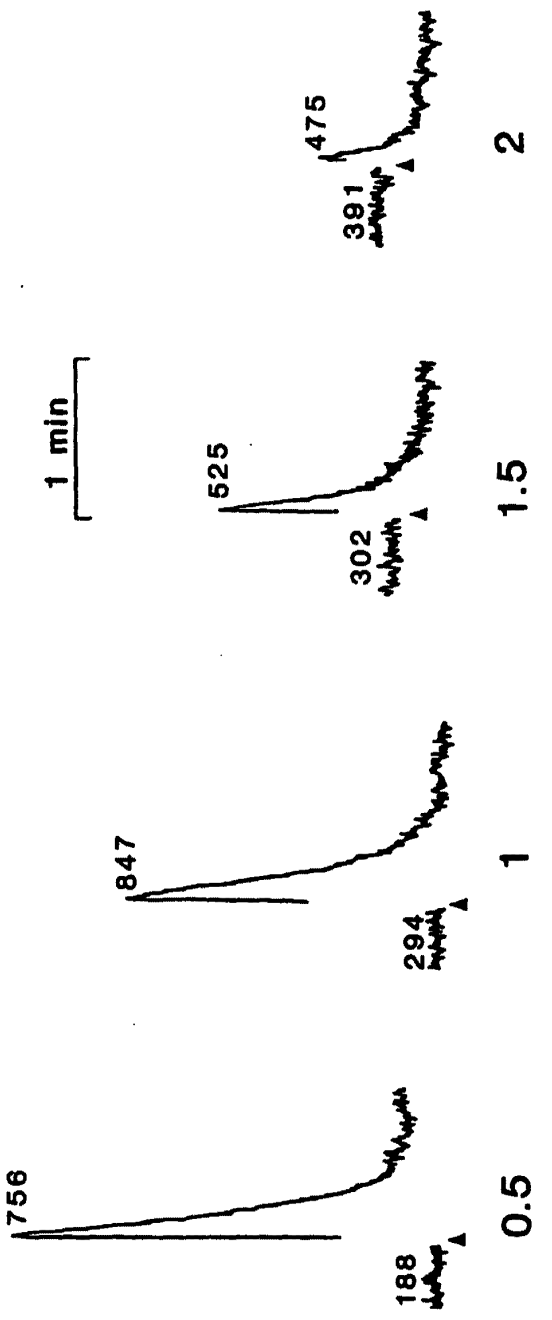

1 min
756
188
0.5
847
294
1
525
302
1.5
475
391
2
Extracellular calcium (mM)

FIG. 6.

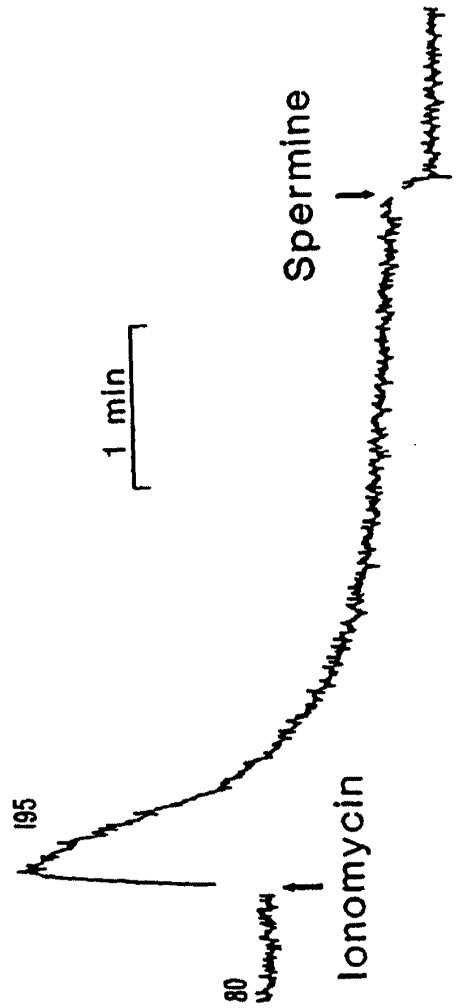
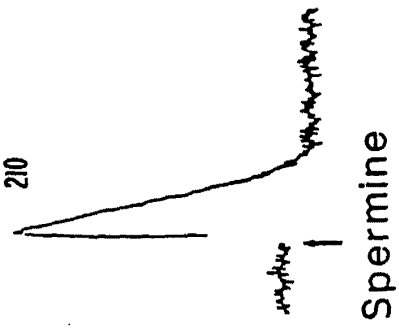


FIG. 7.



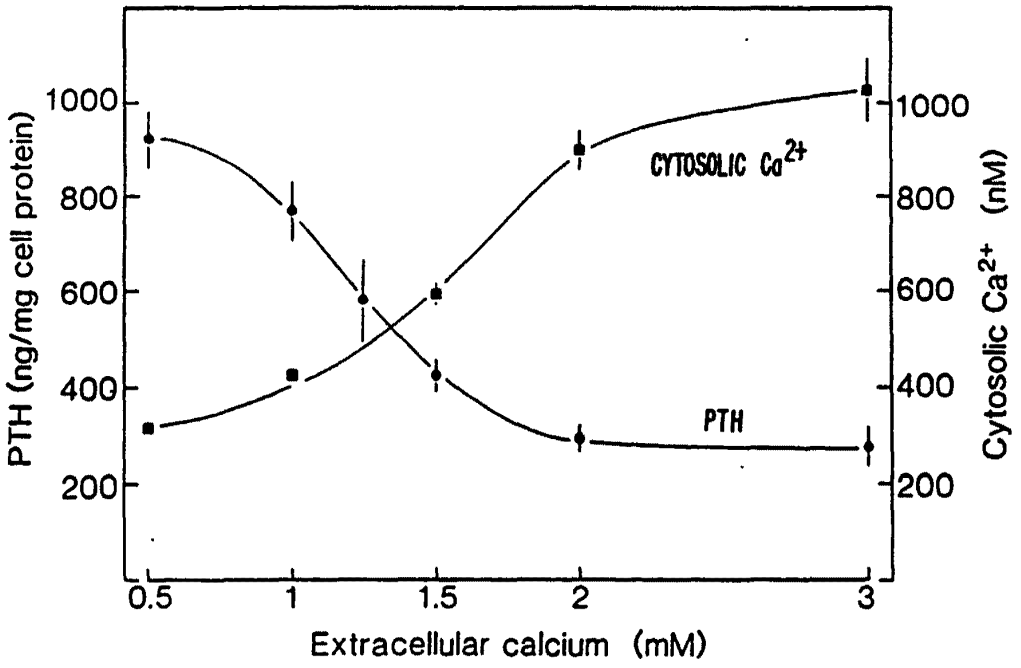

PTH (ng/mg cell protein)
1000
800
600
400
200
CYTOSOLIC Ca^{2+}
PTH
Cytosolic Ca^{2+} (nM)
1000
800
600
400
200
0.5
1
1.5
2
3
Extracellular calcium (mM)

FIG. 8a.

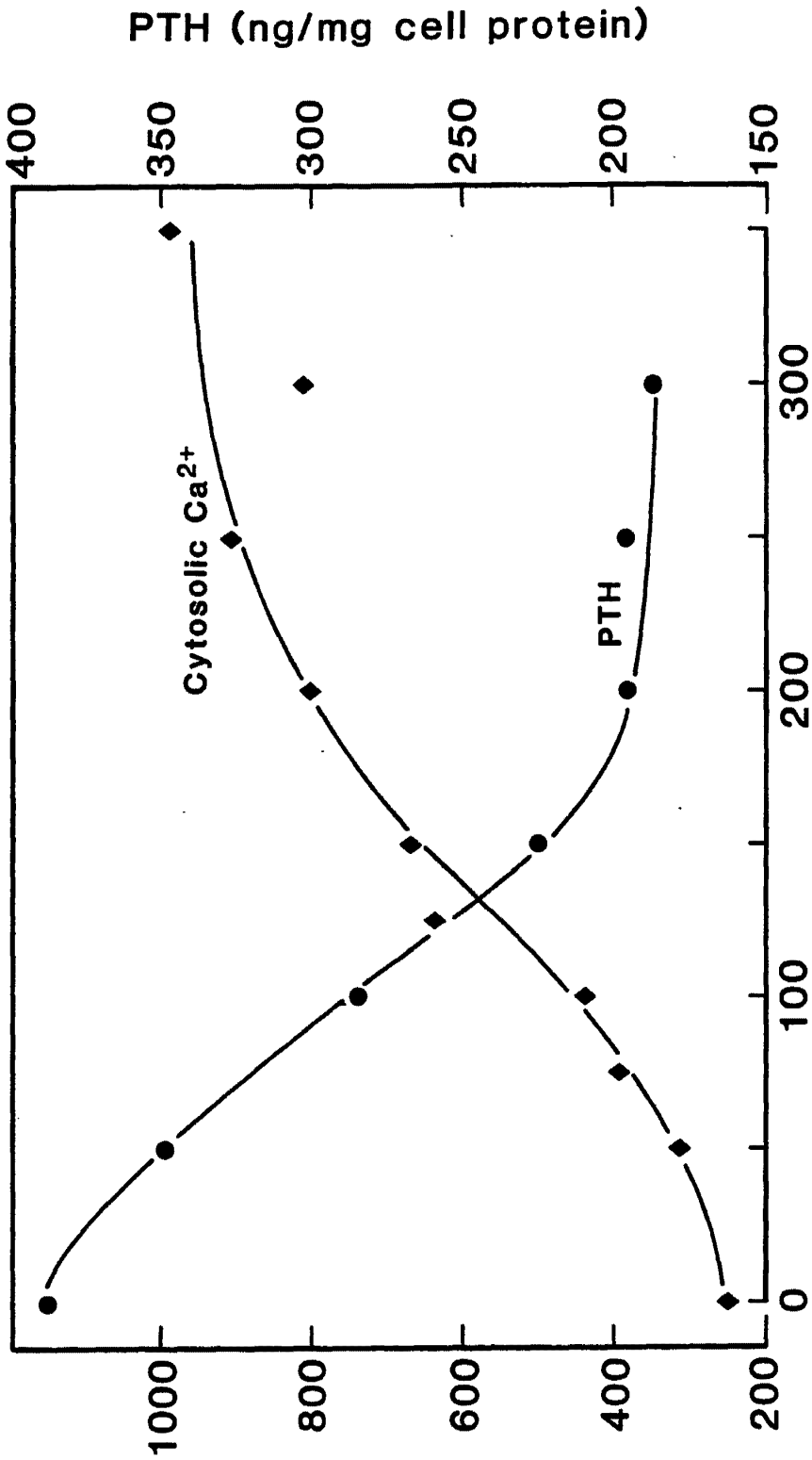

PTH (ng/mg cell protein)
400
350
300
250
200
150
Cytosolic Ca2+
PTH
300
200
100
0
Spermine (μM)
1000
800
600
400
200
Cytosolic Ca2+ (nM)

FIG. 8b.

FIG. 9a.

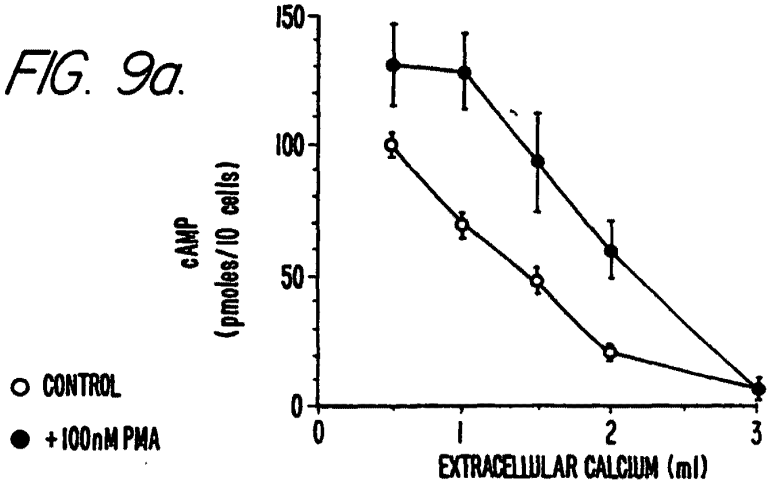


FIG. 9b.

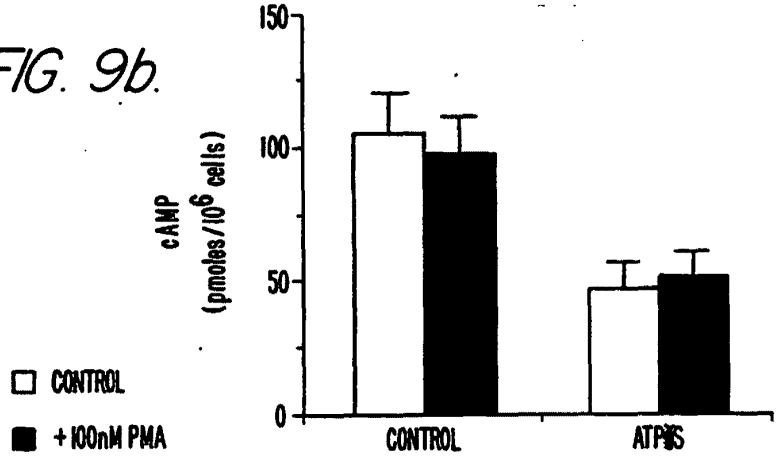
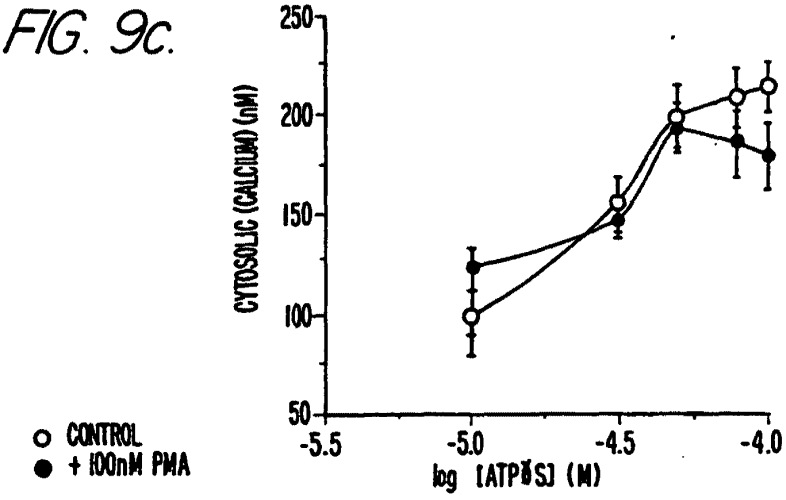


FIG. 9c.



SUBSTITUTE SHEET

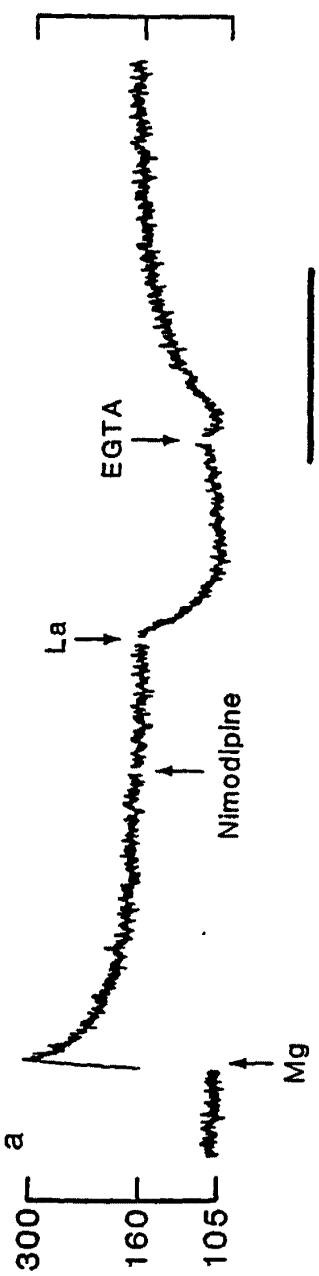


FIG. 10a.

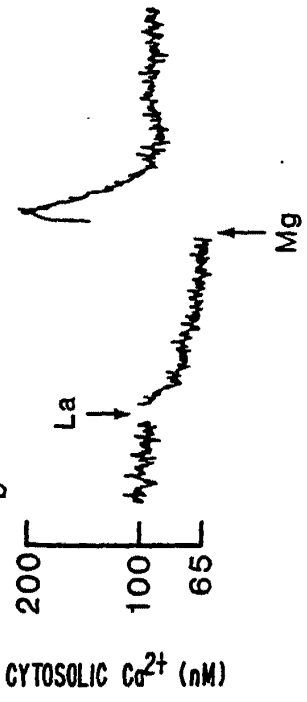


FIG. 10b.

FIG. 10c.

FIG. 11a.

FIG. 11b.

FIG. 11c.

FIG. 11d.

FIG. 11e.

FIG. 11f.

FIG. 11g.

SUBSTITUTE SHEET

FIG. 11h.

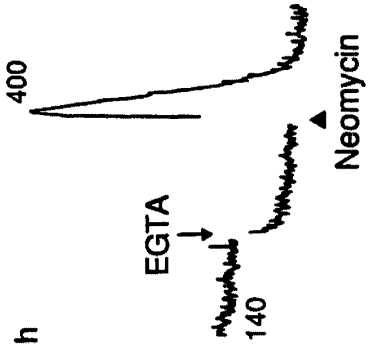


FIG. 11i.

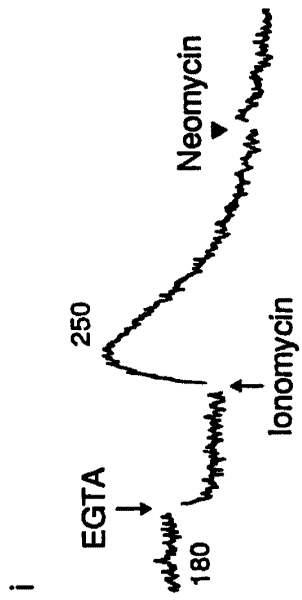
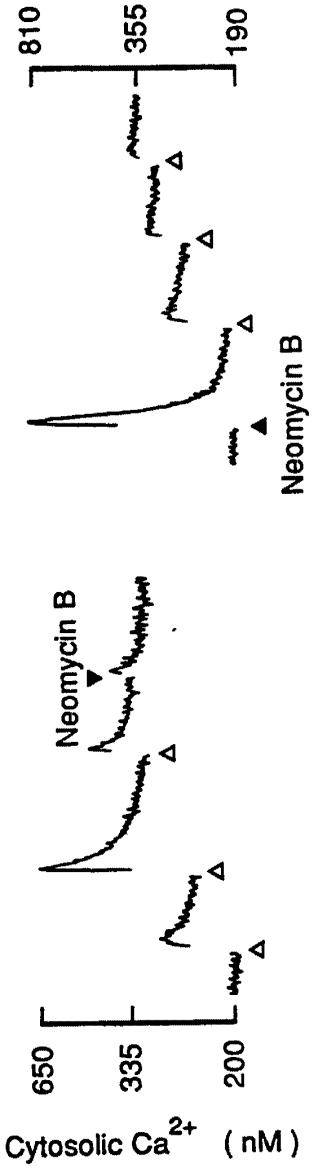


FIG. 12.



SUBSTITUTE SHEET

FIG. 13a.

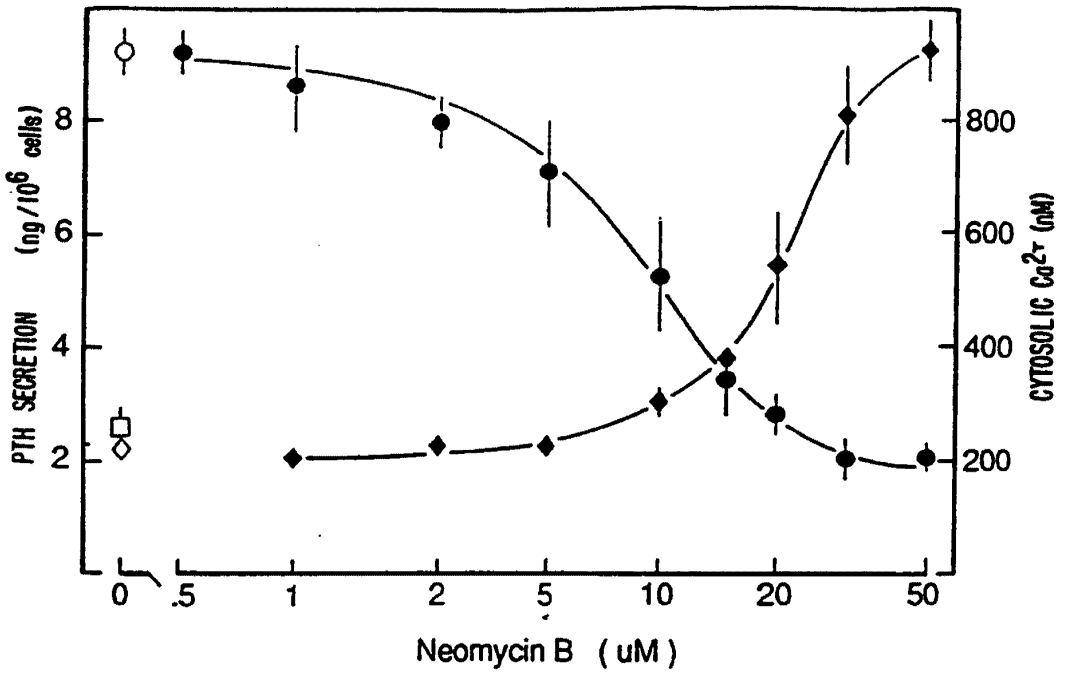
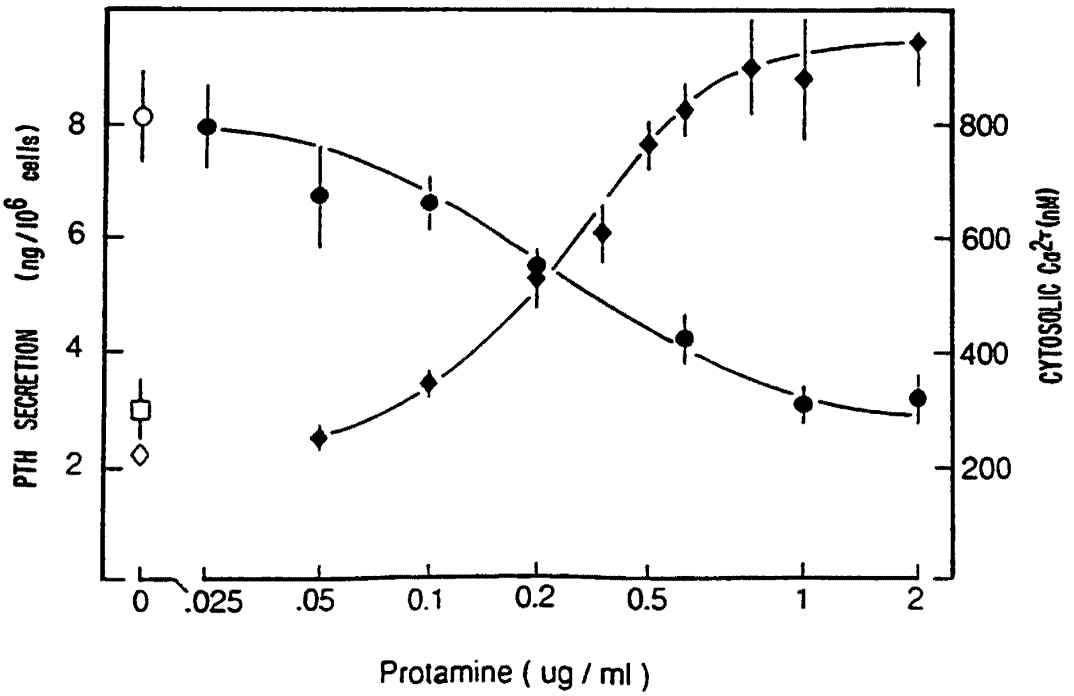
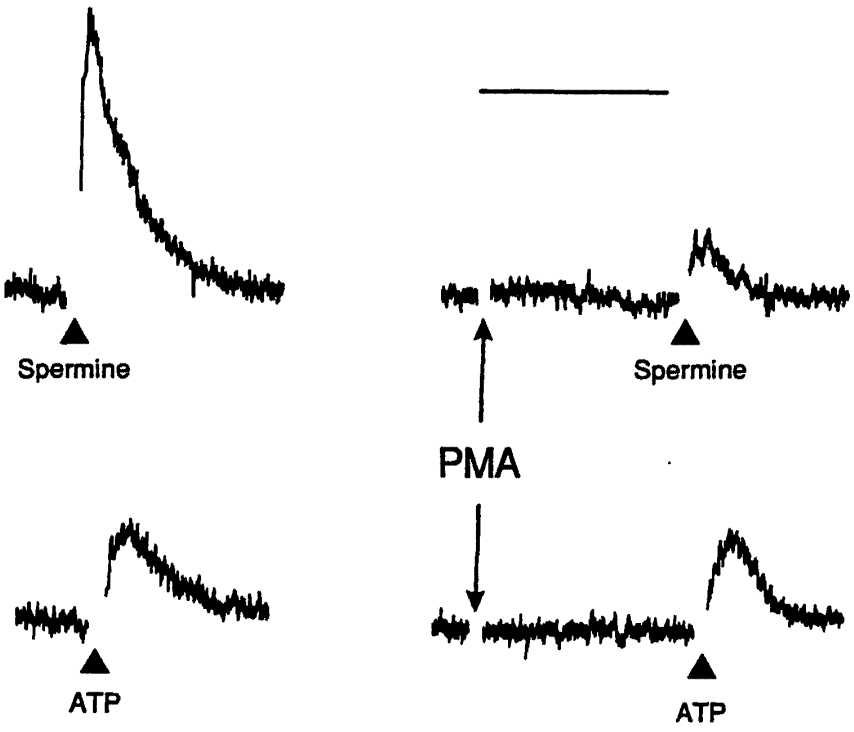


FIG. 13b.



SUBSTITUTE SHEET

FIG. 14.



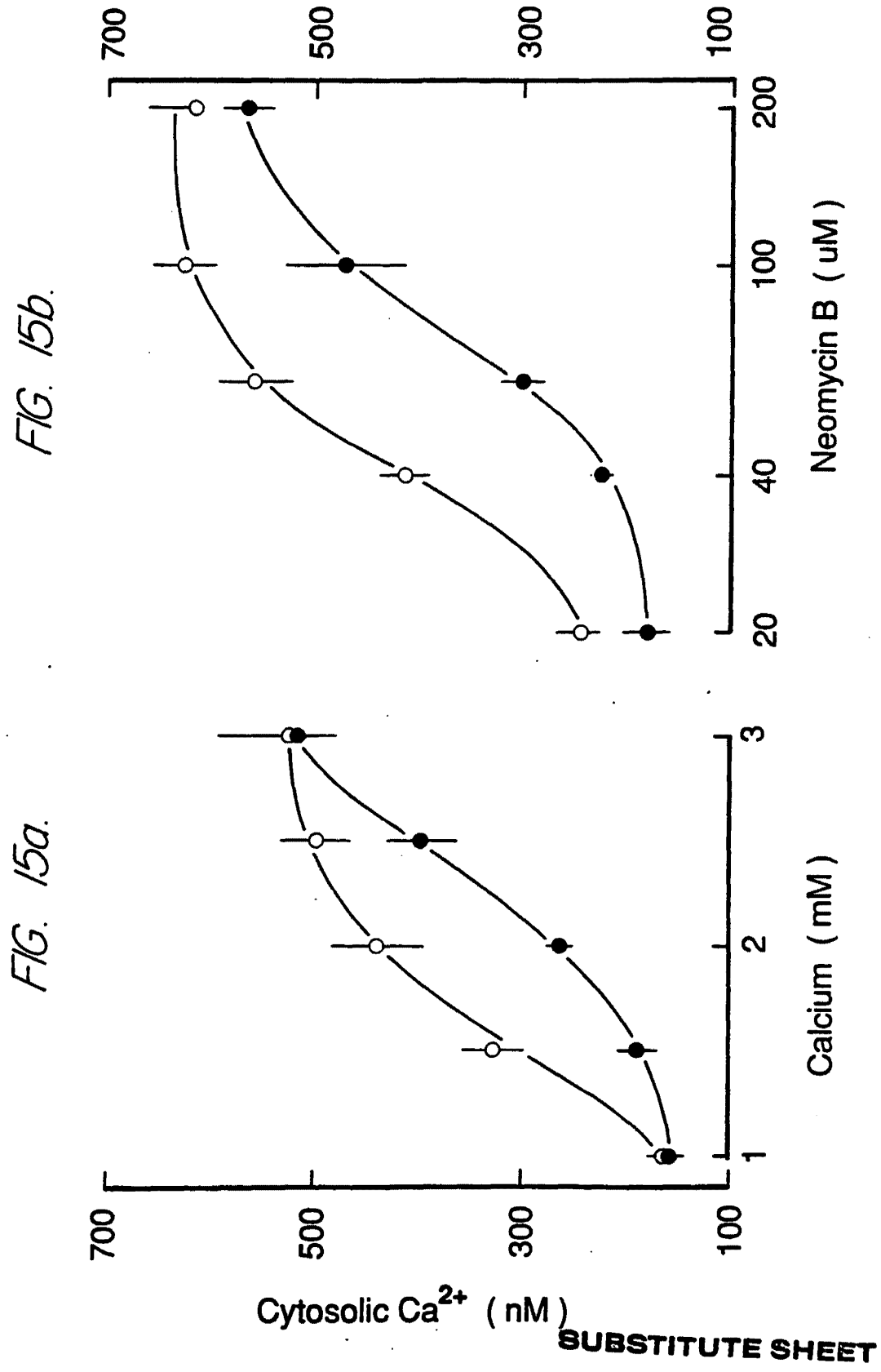


FIG. 15b.

FIG. 15a.

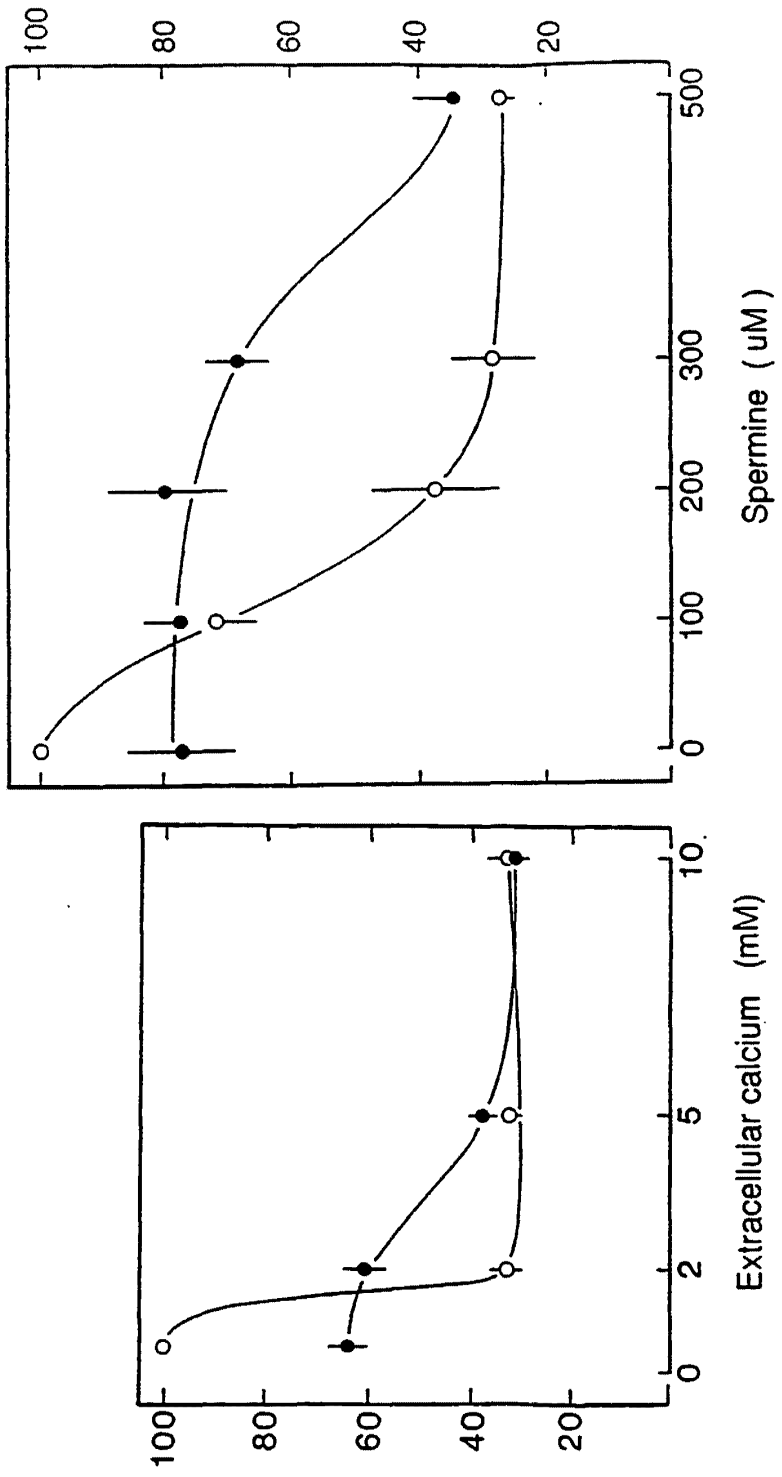


FIG. 16b.

FIG. 16a.

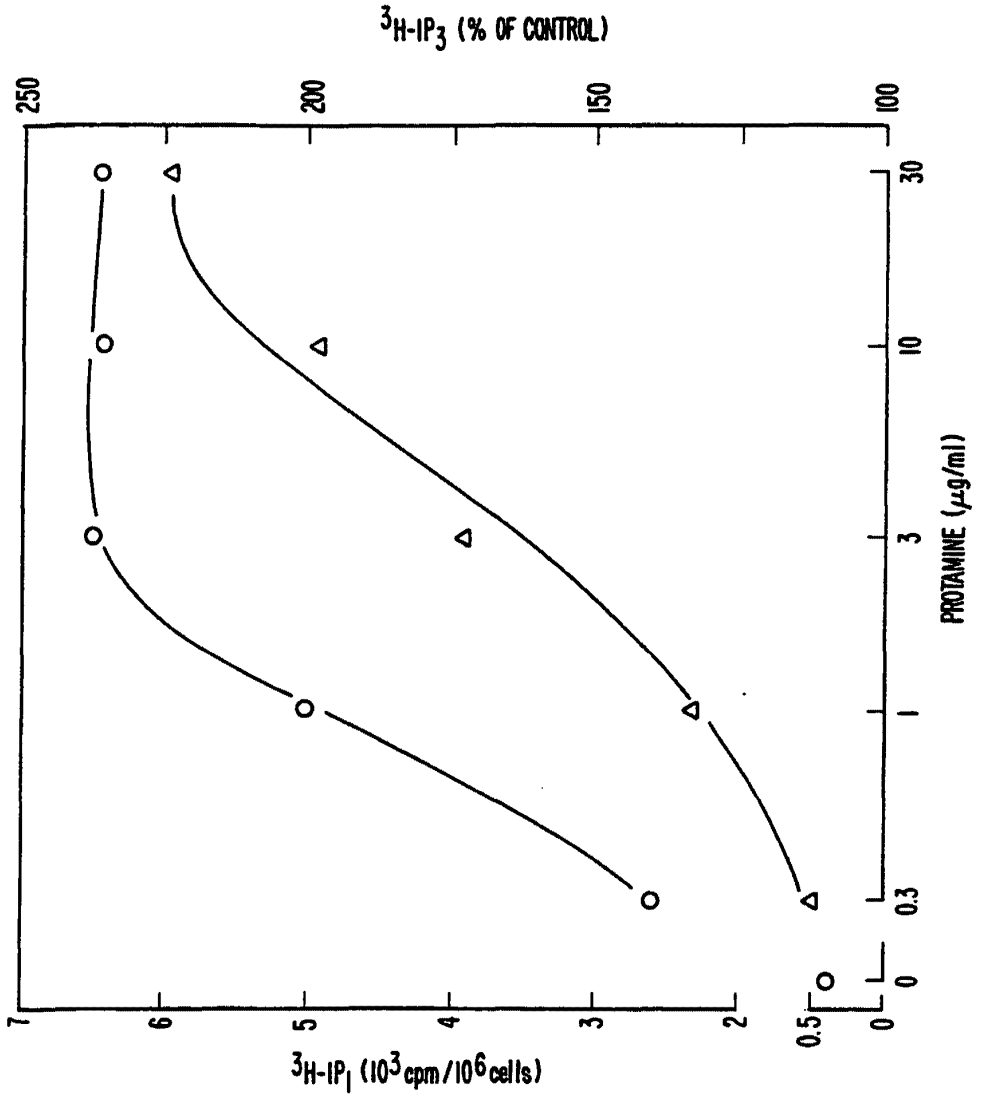


FIG. 17.

SUBSTITUTE SHEET

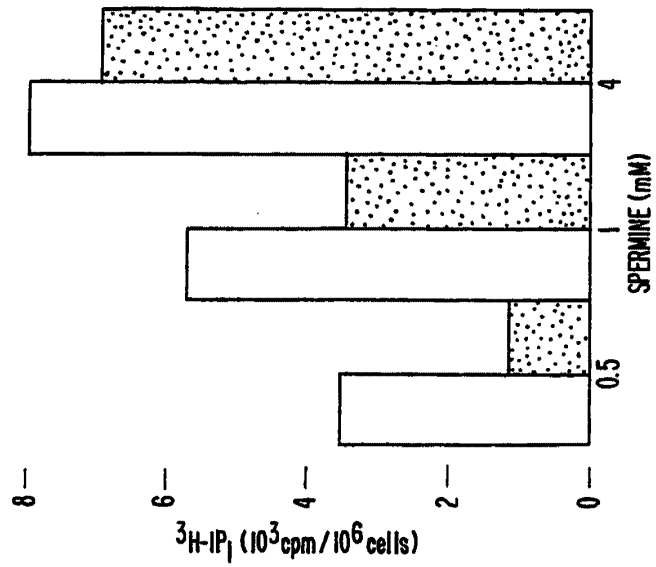


FIG. 18b.

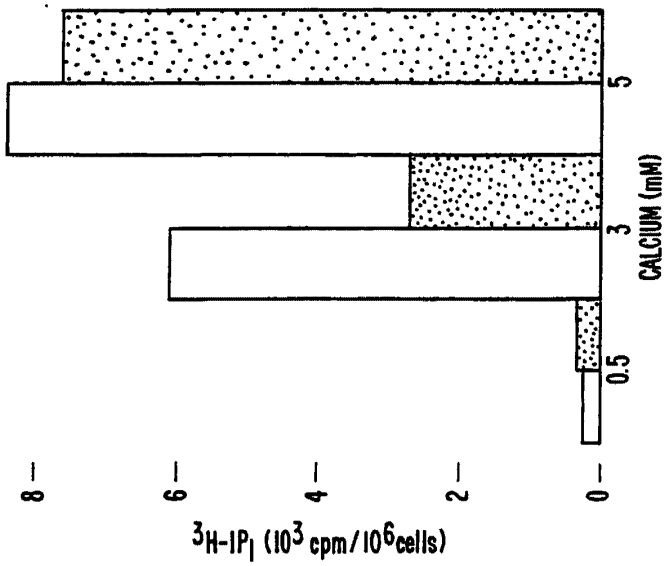


FIG. 18a.

SUBSTITUTE SHEET

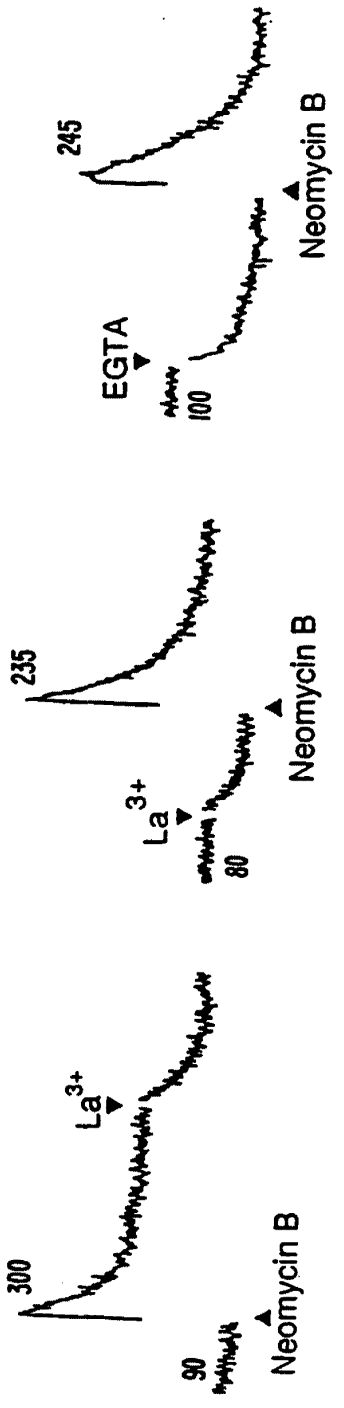

245
Neomycin B
EGTA
100
235
Neomycin B
La3+
80
300
La3+
Neomycin B
90

FIG. 19.

FIG. 20a.

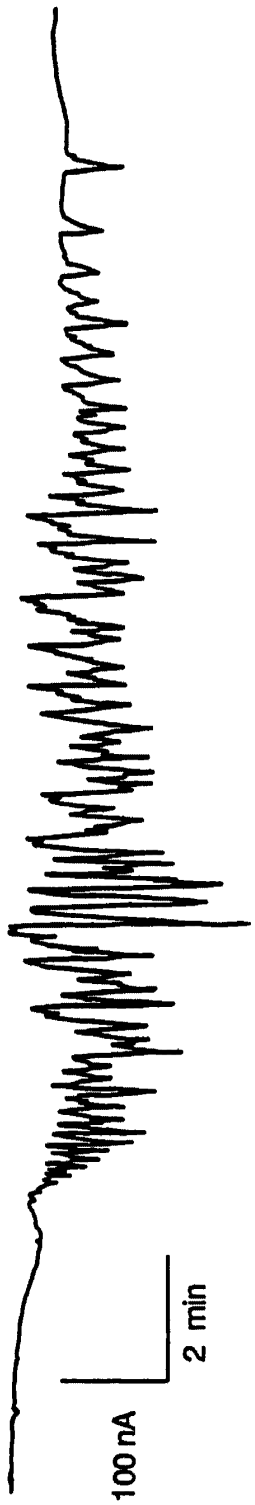
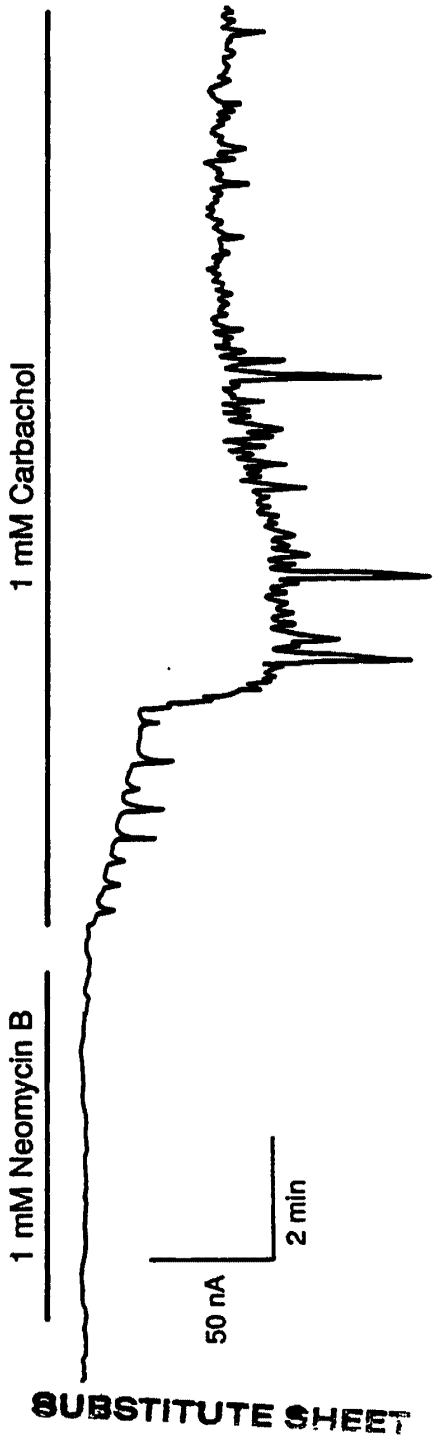


FIG. 20b.



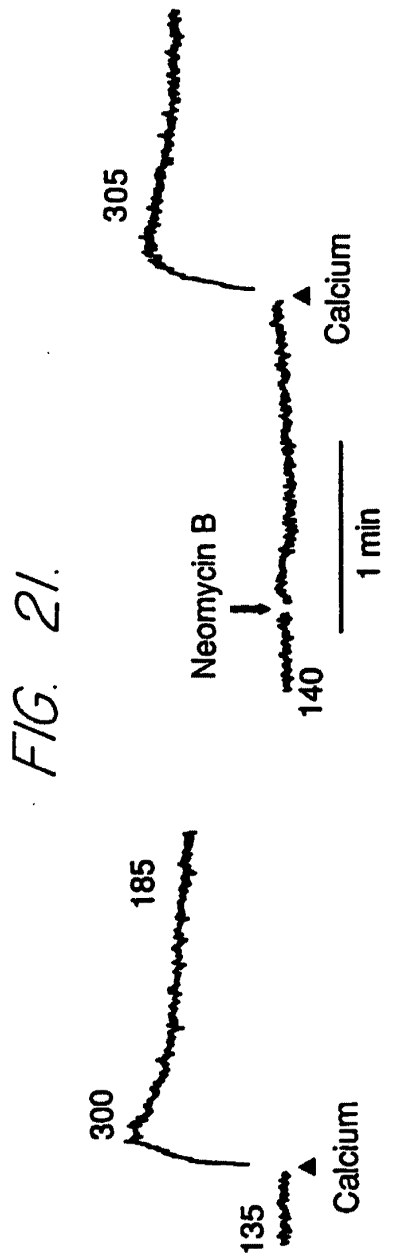


FIG. 21.

FIG. 22.

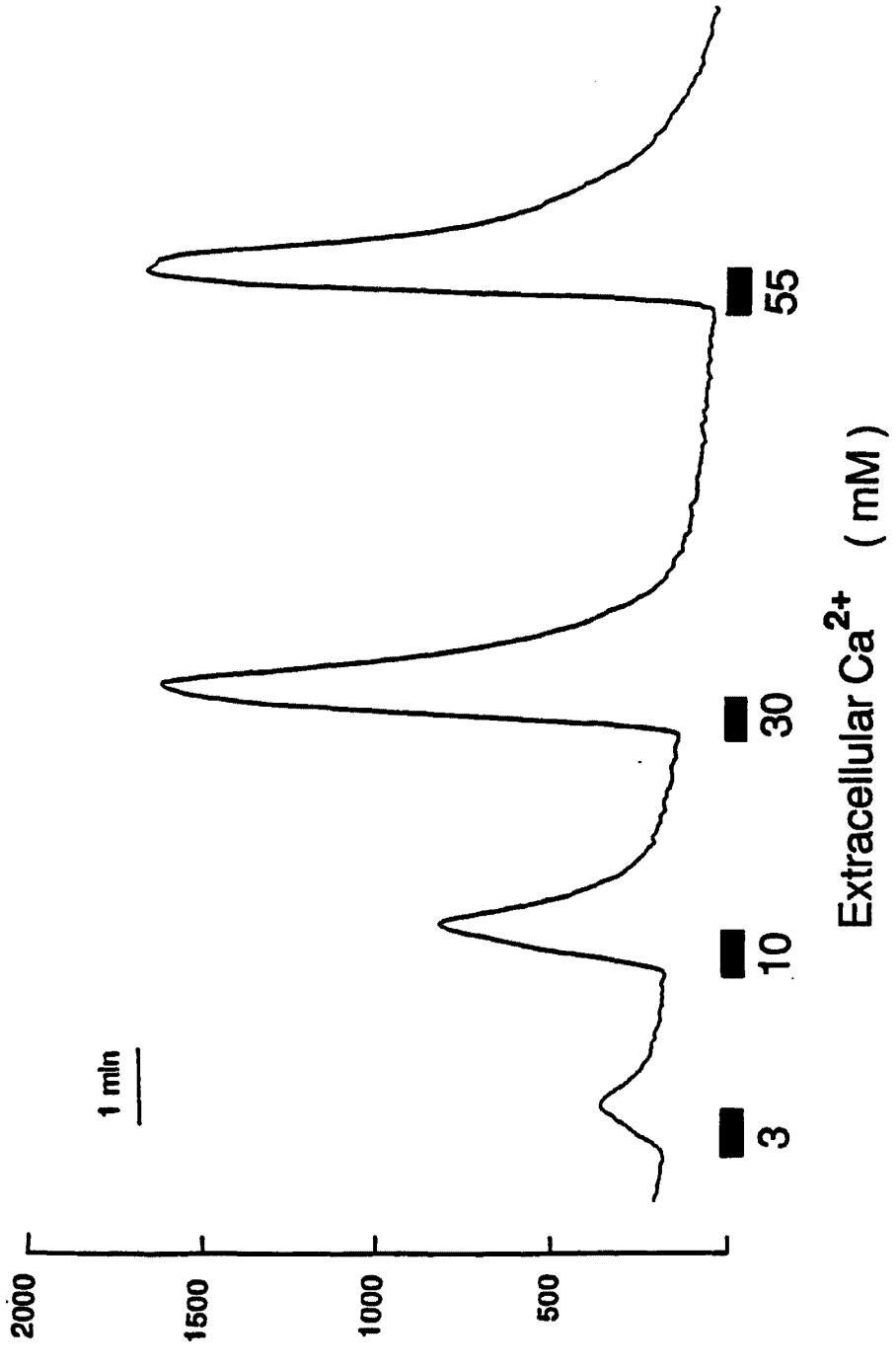

1 min
3
10
30
55
Extracellular Ca^{2+} (mM)
2000
1500
1000
500
Cytosolic Ca^{2+} (nM)

FIG. 23.

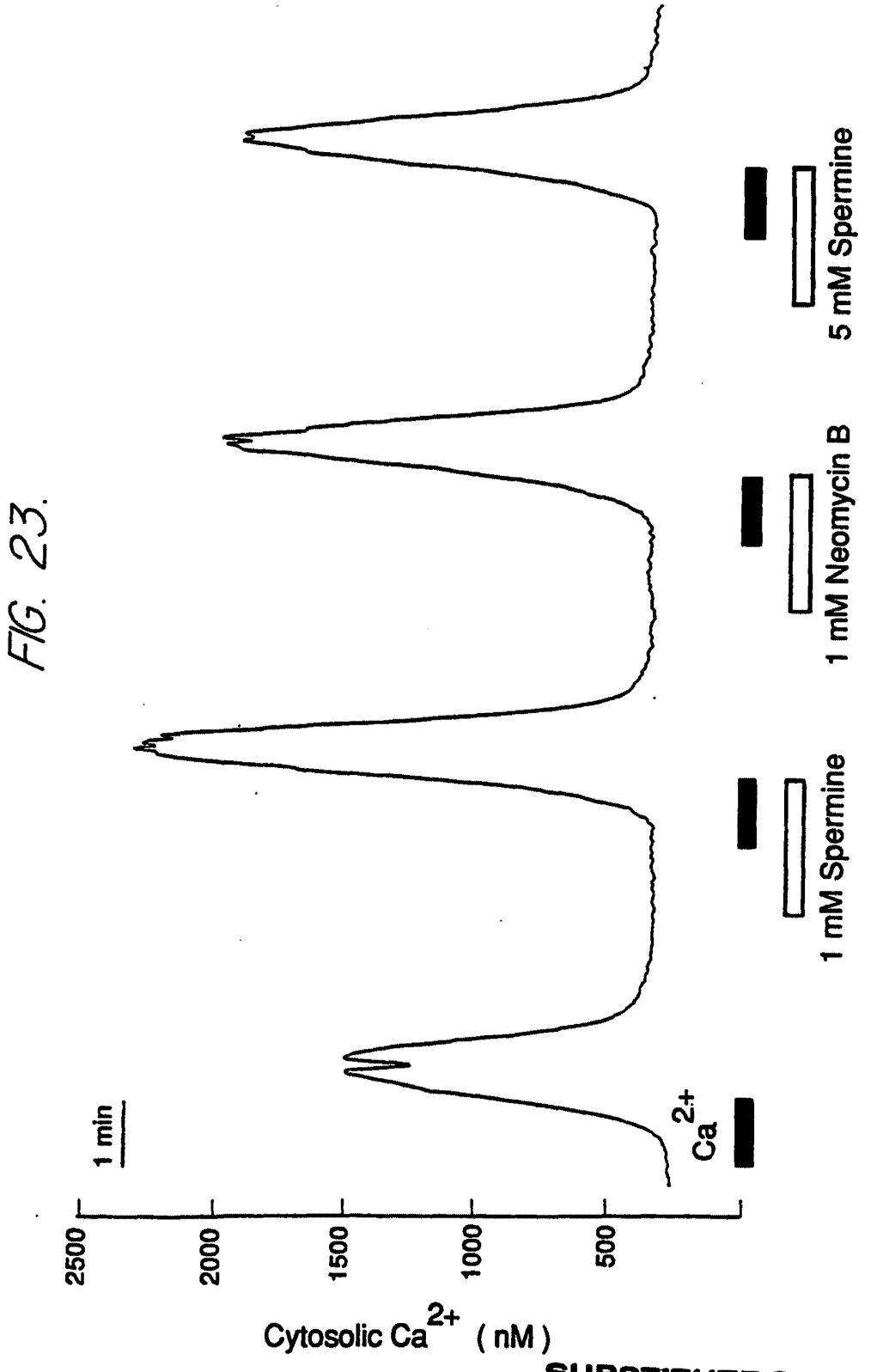
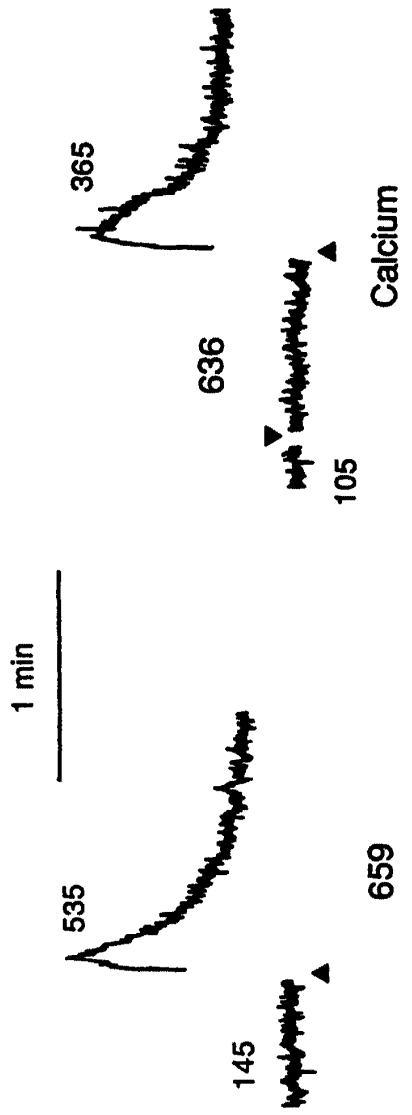

1 min
Ca2+
1 mM Spermine
1 mM Neomycin B
5 mM Spermine
2500
2000
1500
1000
500
Cytosolic Ca2+ (nM)

FIG. 24.



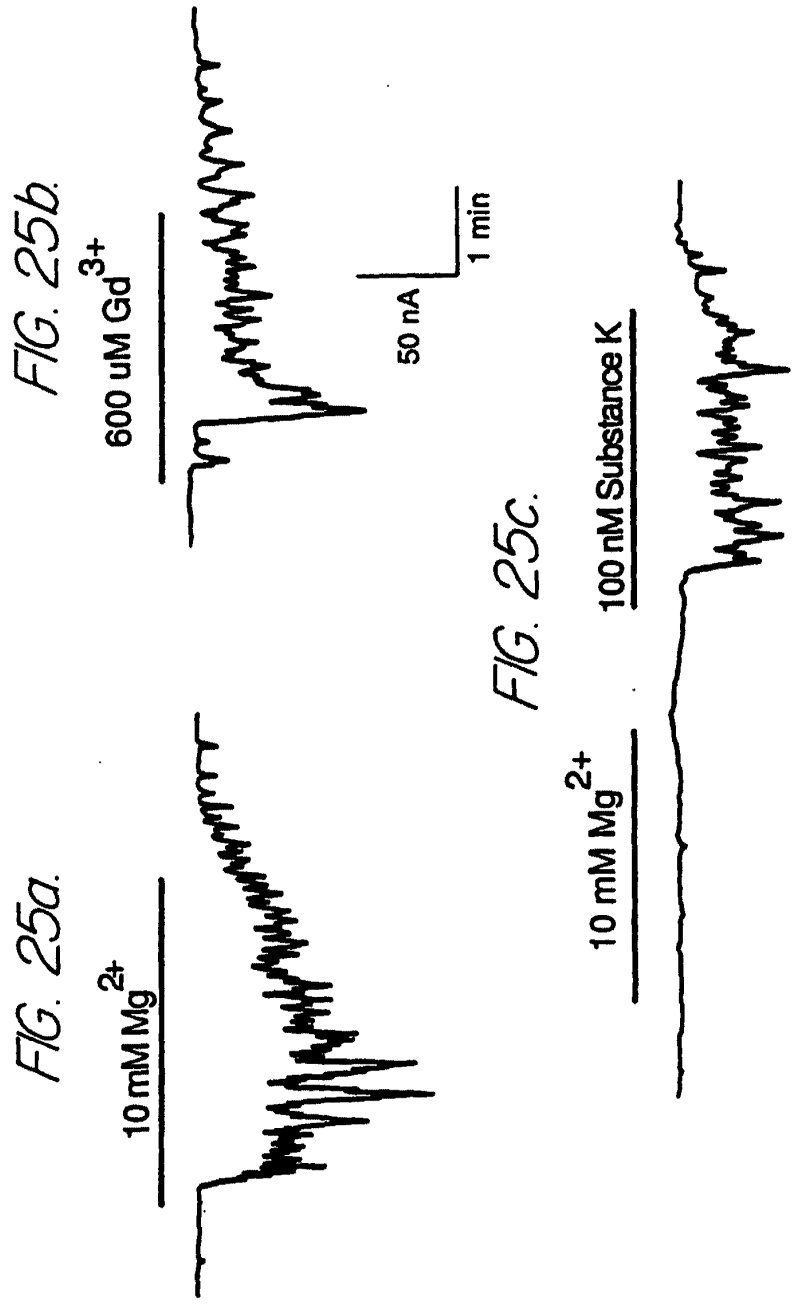


FIG. 25a.
10 mM Mg2+
FIG. 25b.
600 uM Gd3+
50 nA
1 min
FIG. 25c.
10 mM Mg2+
100 nM Substance K

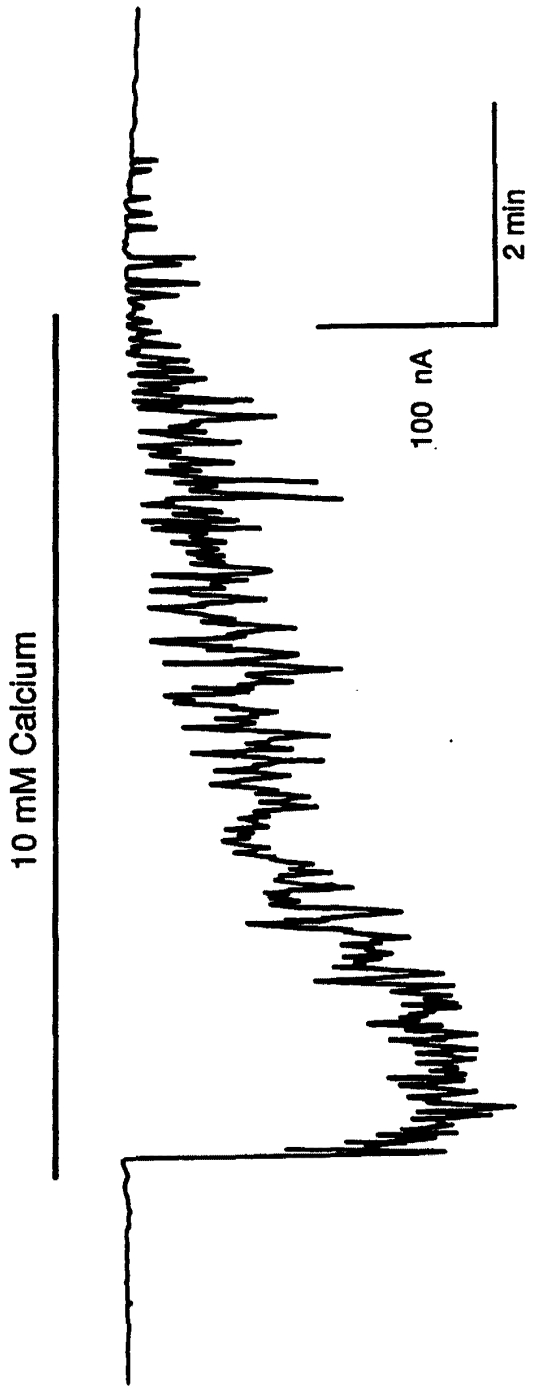


FIG. 26.

FIG. 27.

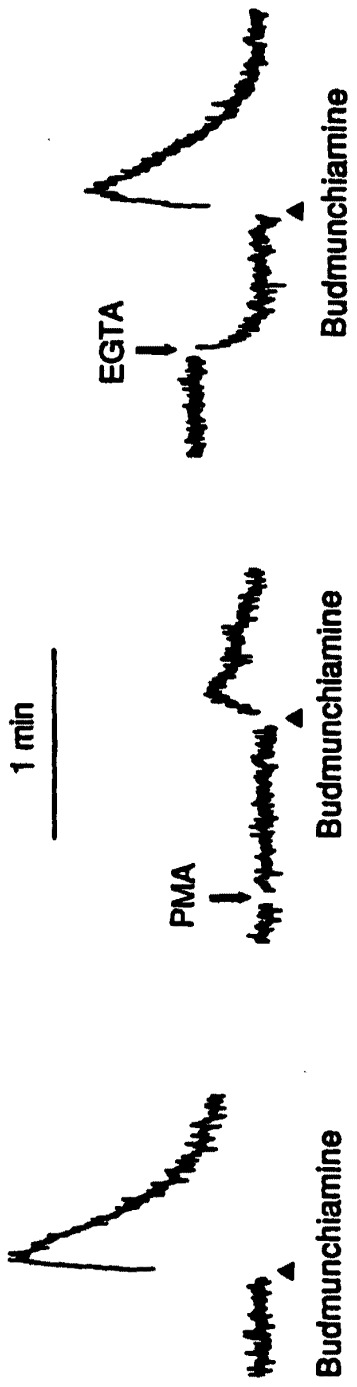
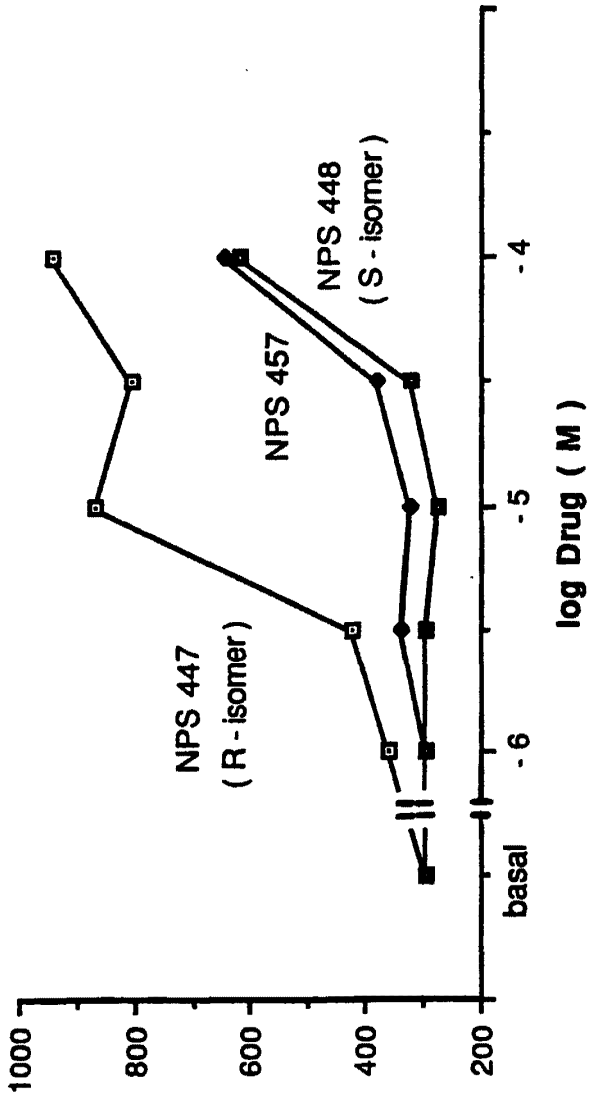

1 min
PMA
Budmunchiamine
EGTA
Budmunchiamine
Budmunchiamine

FIG. 28.


NPS 448
(S - isomer)
NPS 457
NPS 447
(R - isomer)
basal
-6
-5
-4
log Drug (M)
1000
800
600
400
200
Cytosolic Ca2+ (nM)

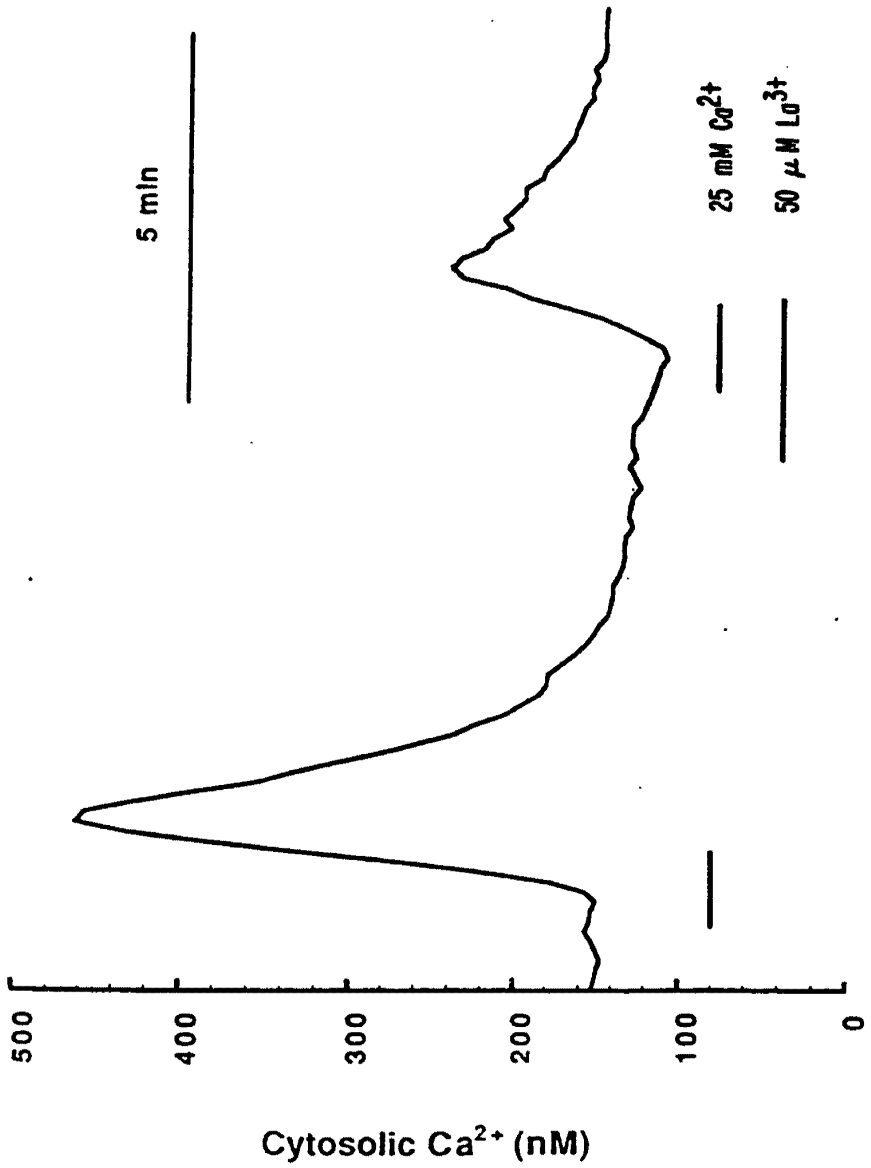
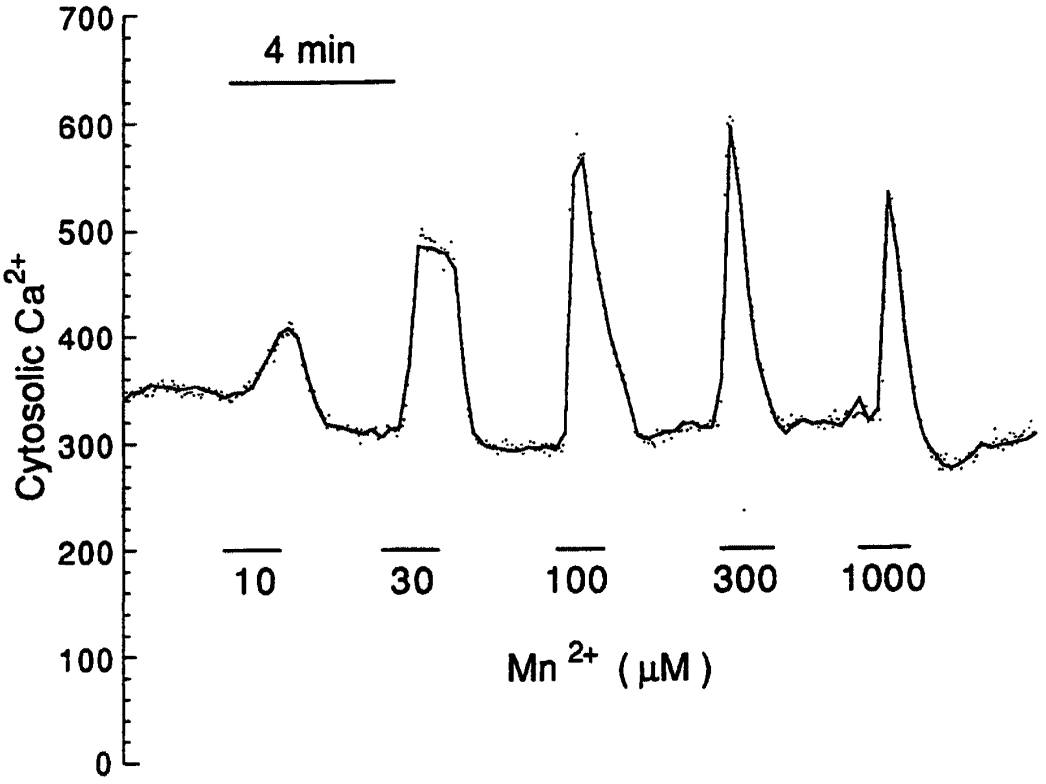


FIG. 29.

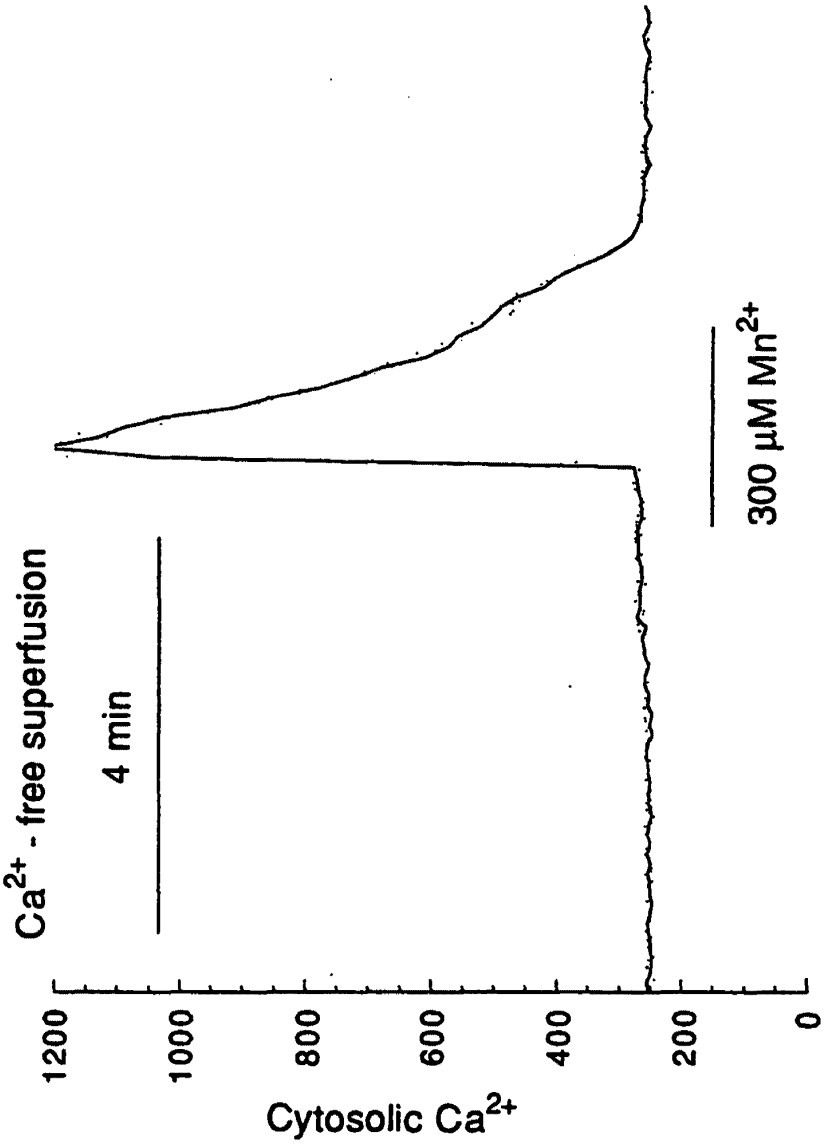
FIG. 30a.


700
600
500
400
300
200
100
0
4 min
Cytosolic Ca^{2+}
10
30
100
300
1000
Mn^{2+} (μM)

(a.)

FIG. 30b.

(b.)


Ca2+ - free superfusion
4 min
300 µM Mn2+
1200
1000
800
600
400
200
0
Cytosolic Ca2+

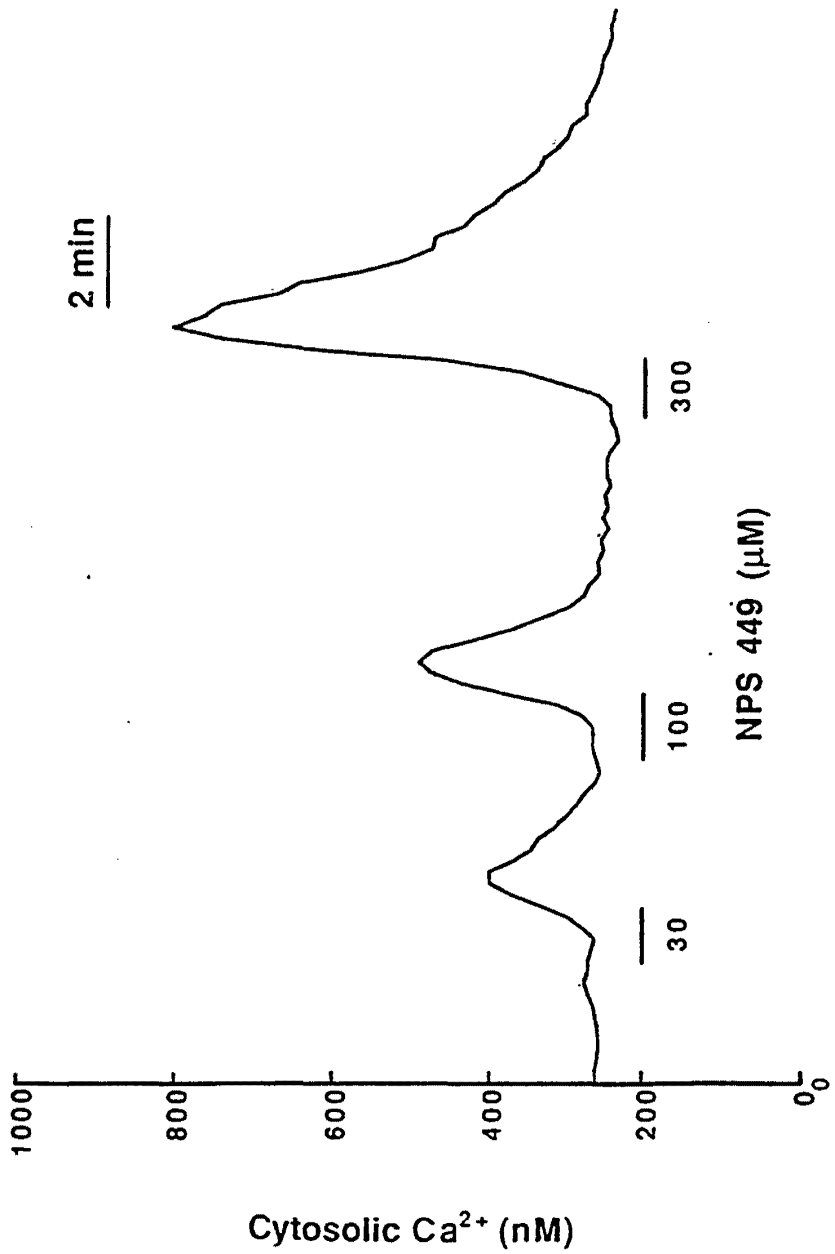

2 min
300
100
30
NPS 449 (μM)
1000
800
600
400
200
0
Cytosolic Ca^{2+} (nM)

FIG. 31a.

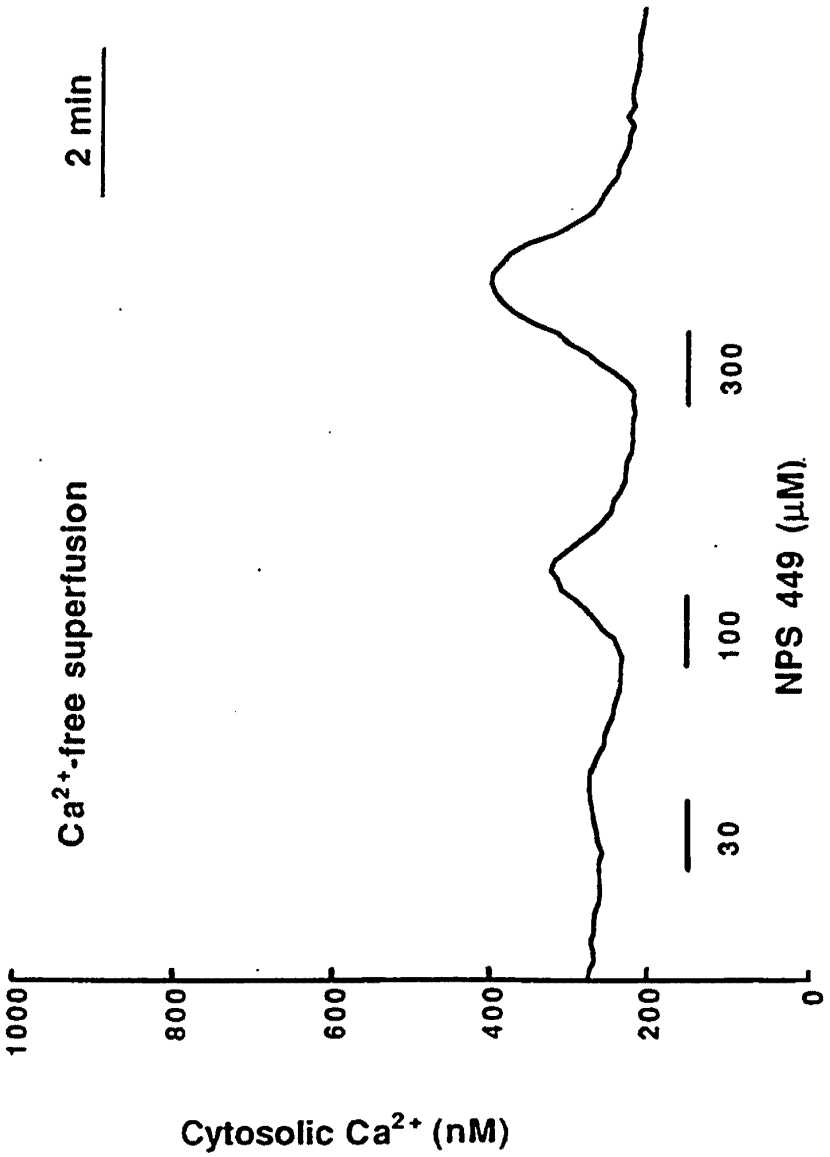

2 min
Ca2+-free superfusion
NPS 449 (μM)
30
100
300
1000
800
600
400
200
0
Cytosolic Ca2+ (nM)

FIG. 31b.

FIG. 32.

FIG. 33.

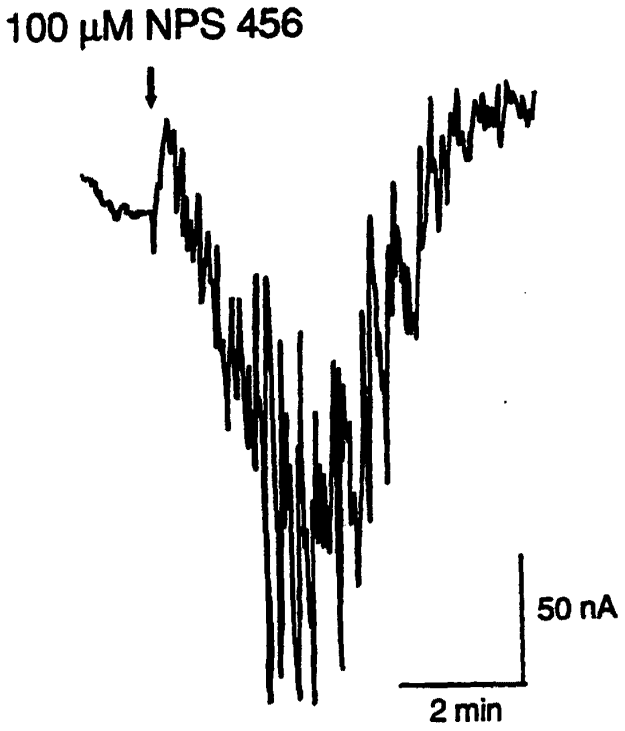
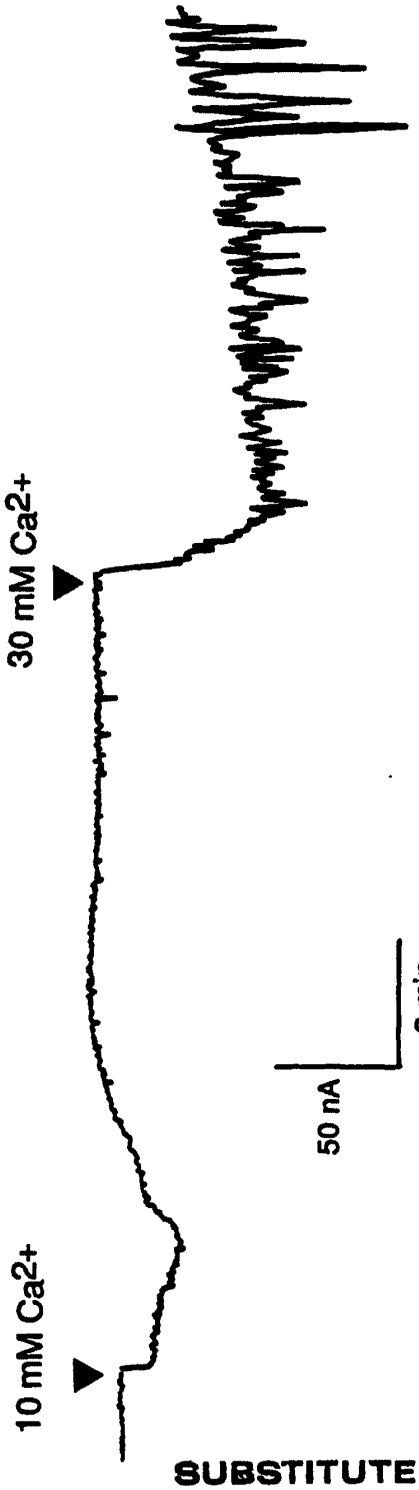


FIG. 34.



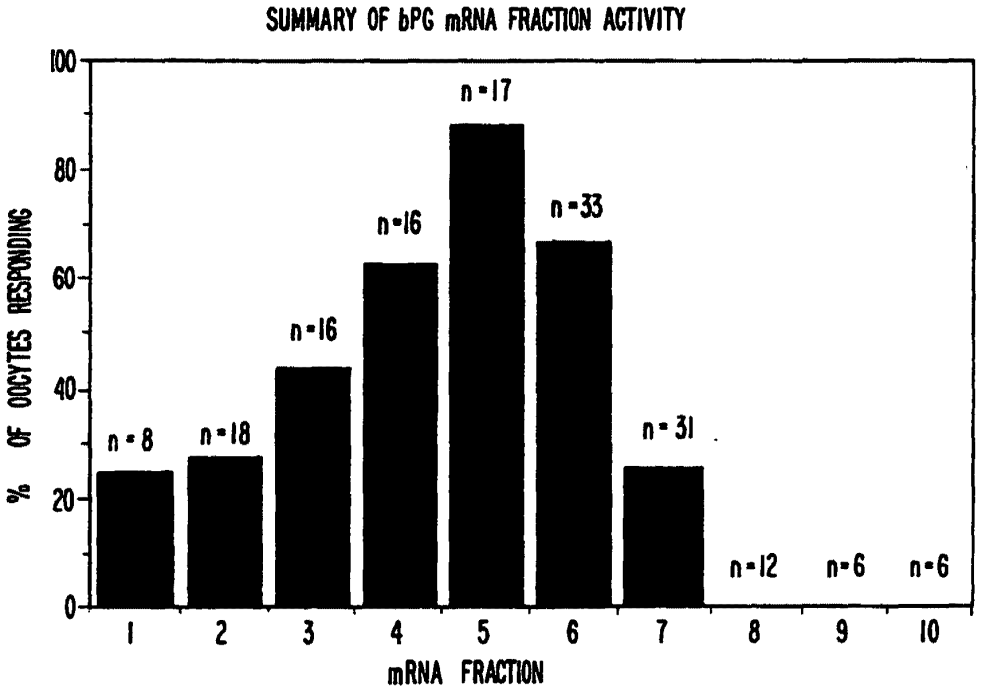
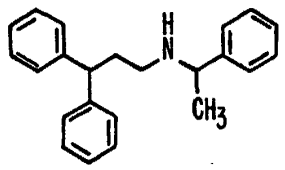
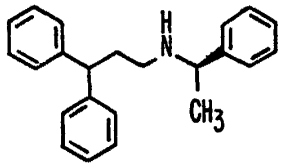

SUMMARY OF bPG mRNA FRACTION ACTIVITY
% OF OOCYTES RESPONDING
100
80
60
40
20
0
n = 8
n = 18
n = 16
n = 16
n = 17
n = 33
n = 31
n = 12
n = 6
n = 6
1
2
3
4
5
6
7
8
9
10
mRNA FRACTION

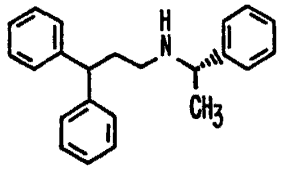
FIG. 35.



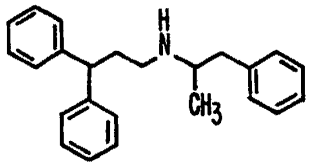
NPS #
456



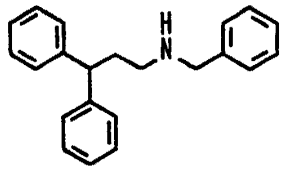
447



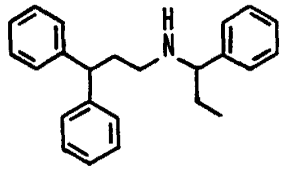
448



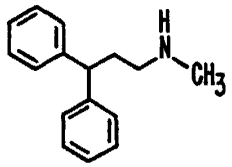
449



455



460

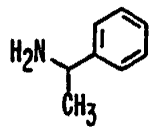


519

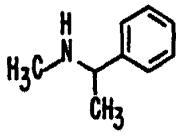
FIG. 36a.

SUBSTITUTE SHEET

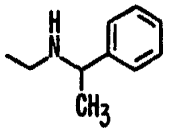
NPS #



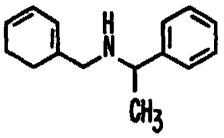
518



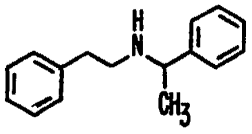
517



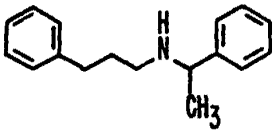
516



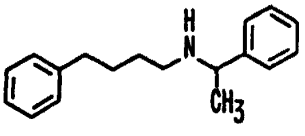
457



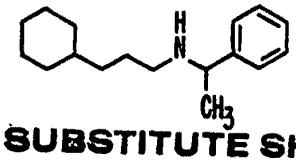
458



459



523

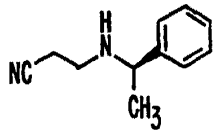


524

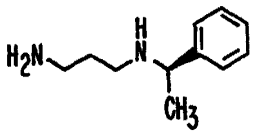
SUBSTITUTE SHEET

FIG. 36b.

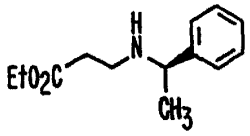
NPS #



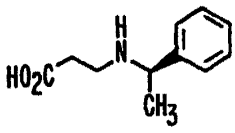
527



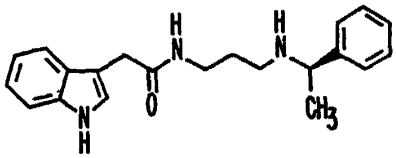
534



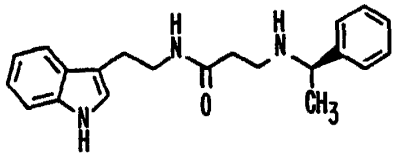
528



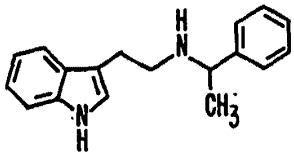
530



532



533



531

FIG. 36C.

SUBSTITUTE SHEET

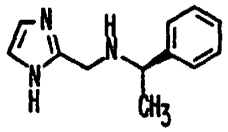
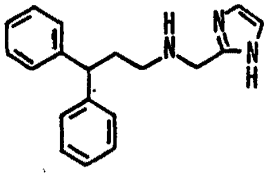
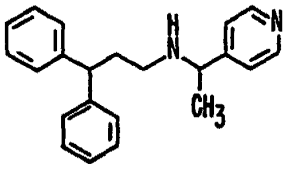
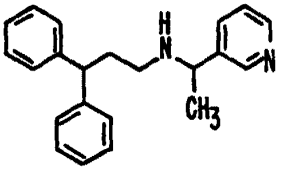
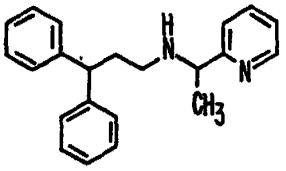
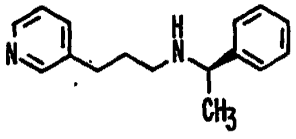
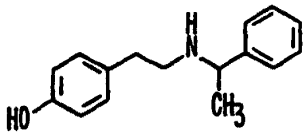
	NPS #
	526
	535
	536
	537
	538
	543
	529

FIG. 36d.

SUBSTITUTE SHEET

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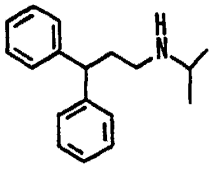
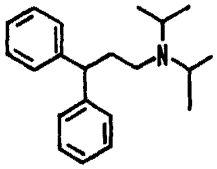
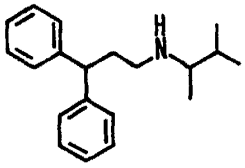
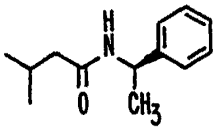
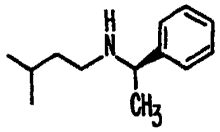
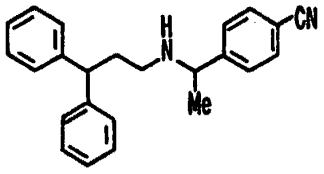
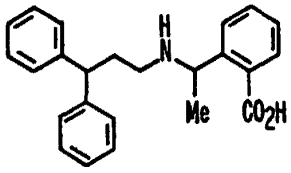
	539
	540
	542
	546
	547

FIG. 36e.

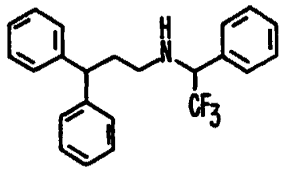
NPS #



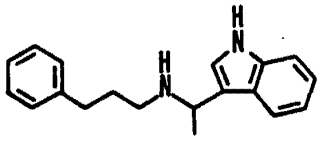
541



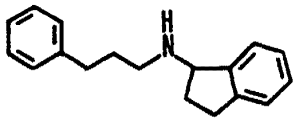
545



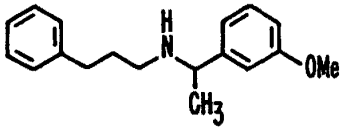
544



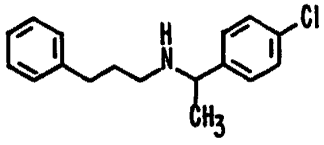
548



549



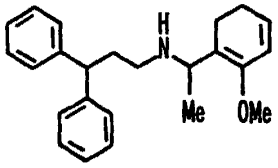
467



468

FIG. 36f.

NPS #



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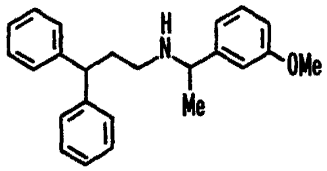

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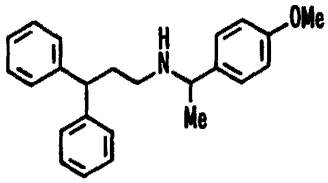
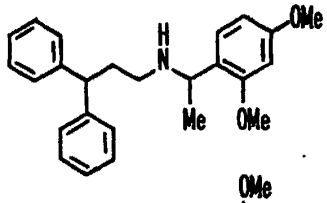


552



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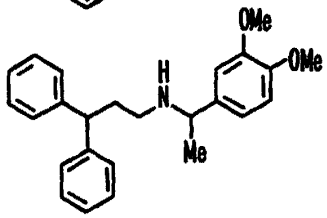

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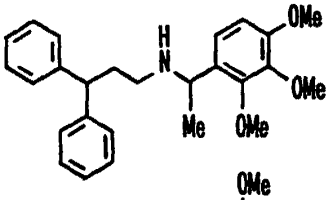

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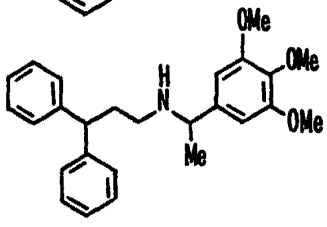

556

FIG. 36g.

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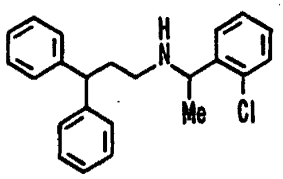
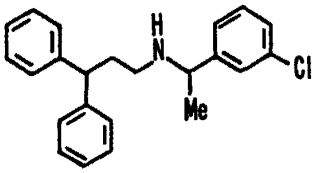
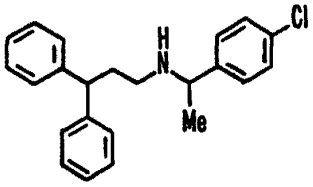
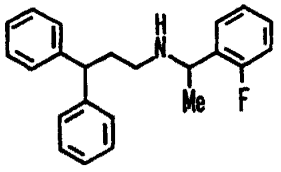
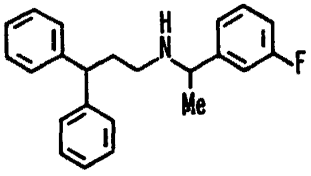
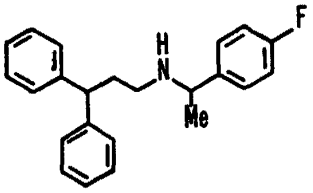
	557
	558
	559
	560
	561
	562

FIG. 36h.

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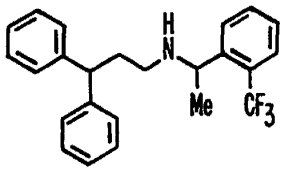
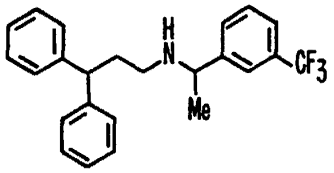
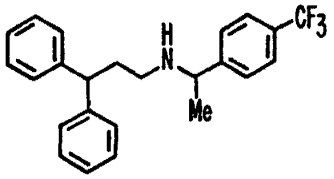
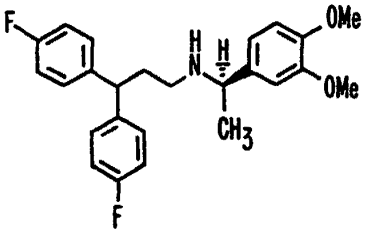
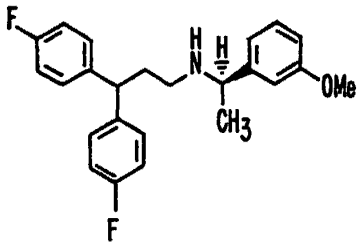
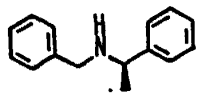
	563
	564
	565
	
	606

FIG. 36i.

NPS #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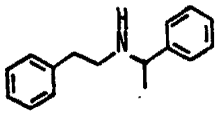
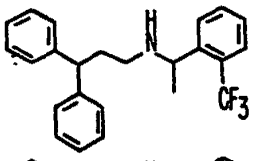

458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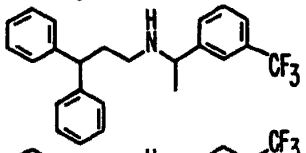
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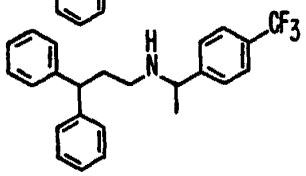
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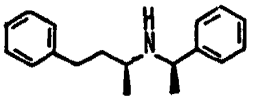
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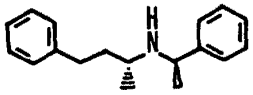
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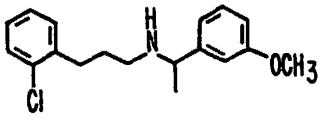
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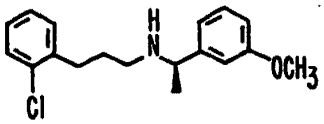

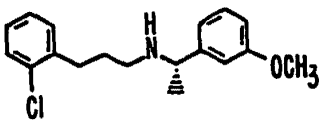

FIG. 36j.

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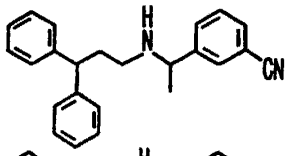
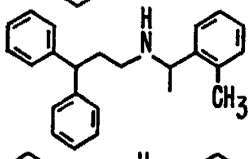
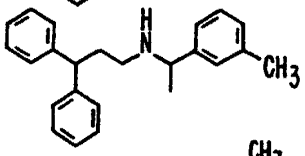
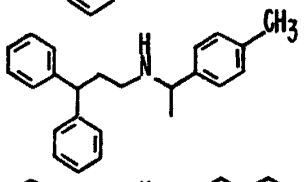
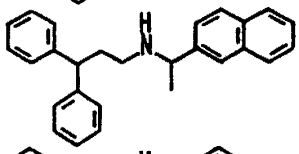
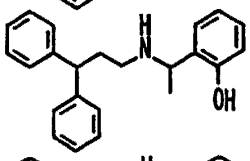
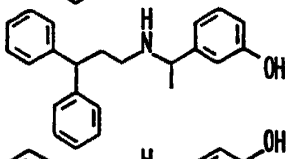
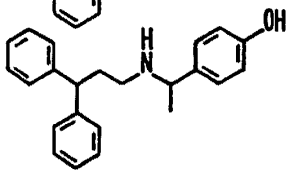
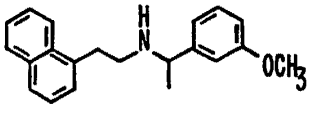
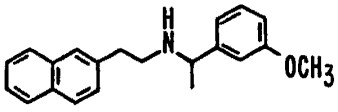
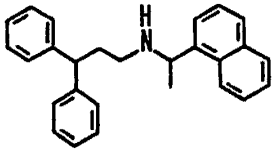
570	
571	
572	
573	
574	
575	
576	
577	
578	
579	
580	

FIG. 36k.

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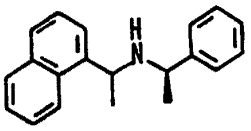
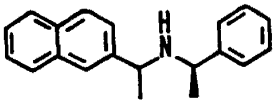
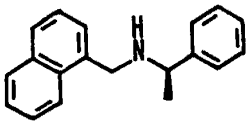
	592
	593
	594

FIG. 36L.

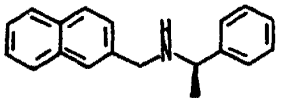
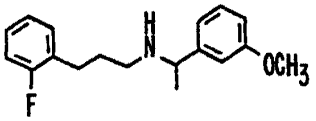
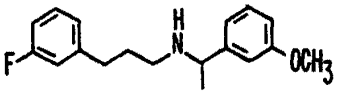
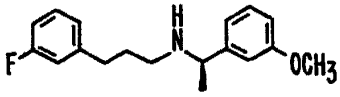
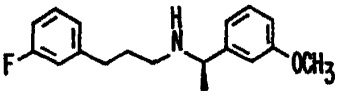
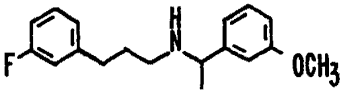
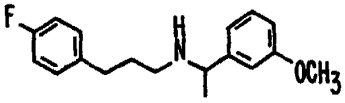
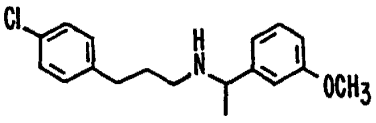
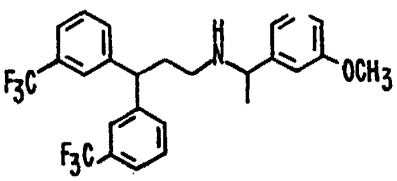
	NPS #
	595
	601
	602
	
	
	603
	604
	605
	607

FIG. 36m.

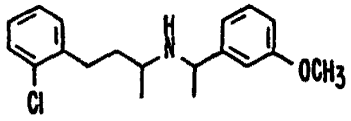
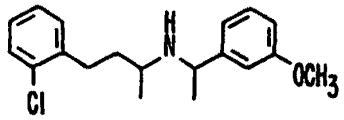
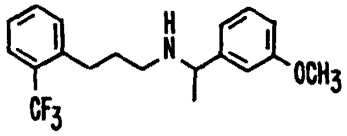
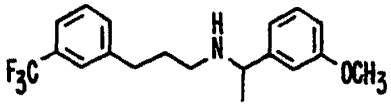
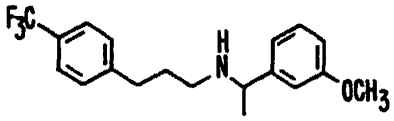
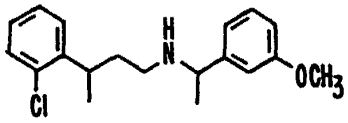
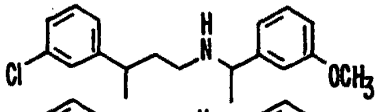
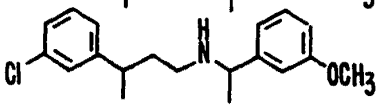
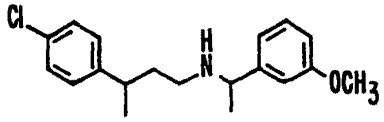
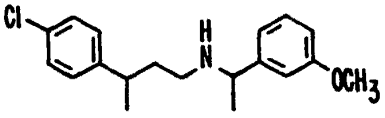
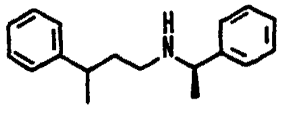
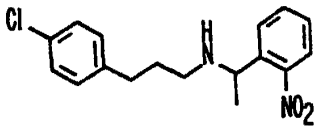
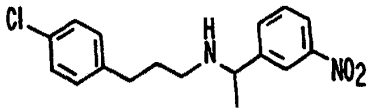
NPS #	
609	
610	
611	
612	
613	
614	
615	
616	
617	
618	
620	

FIG. 36n

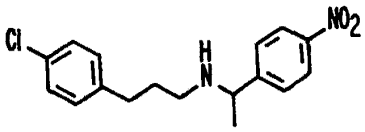
NPS #



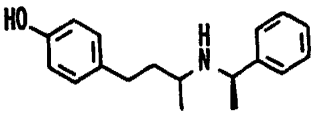
623



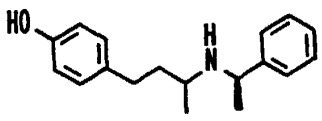
624



625



631

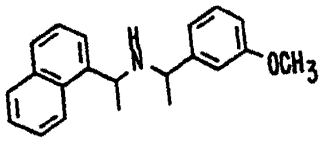


632

SUBSTITUTE SHEET FIG. 360.

NPS #

641



642

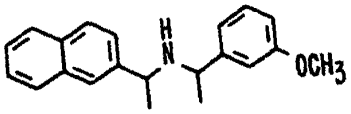
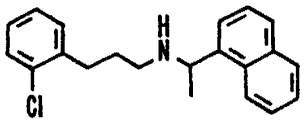
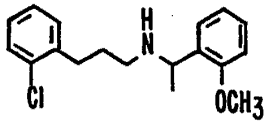


FIG. 36p.

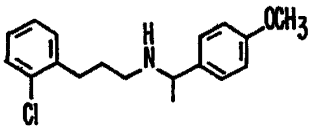


NPS #

646



650



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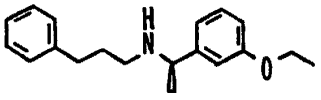
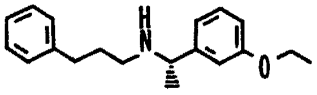
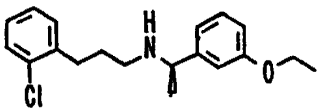
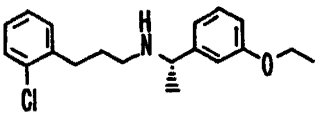




653





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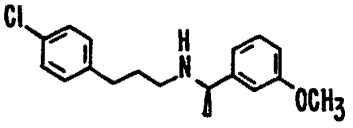
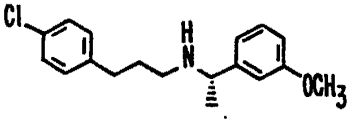


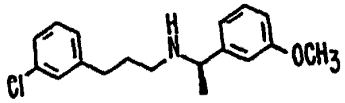


655

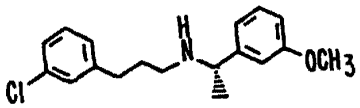
SUBSTITUTE SHEET

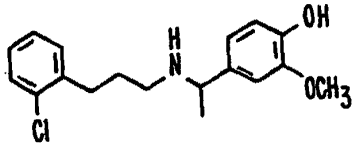
FIG. 36q.

NPS #



656



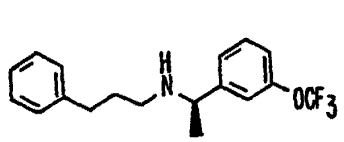


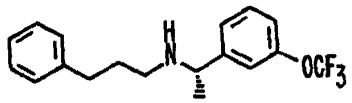
684

FIG. 36r.

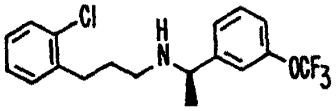
NPS #

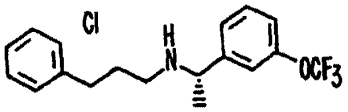
689



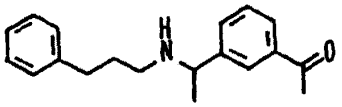


691

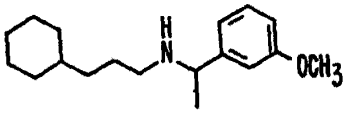




706



708



709

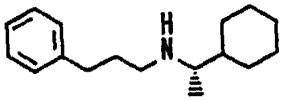


FIG. 36s.

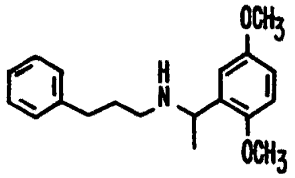
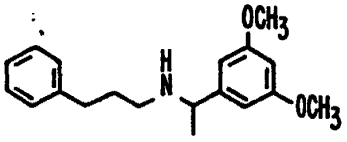
	NPS #
	710
	711

FIG. 36t.

FIG. 37b.

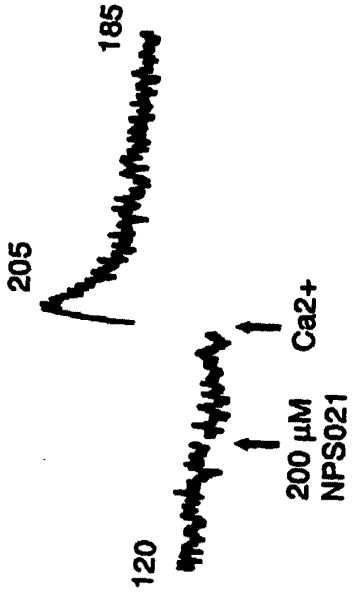


FIG. 37a.

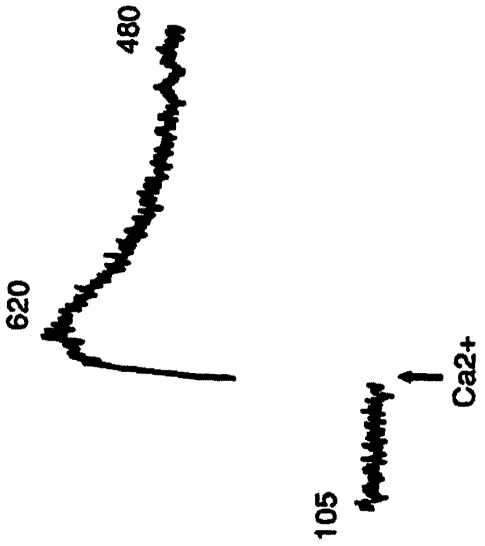
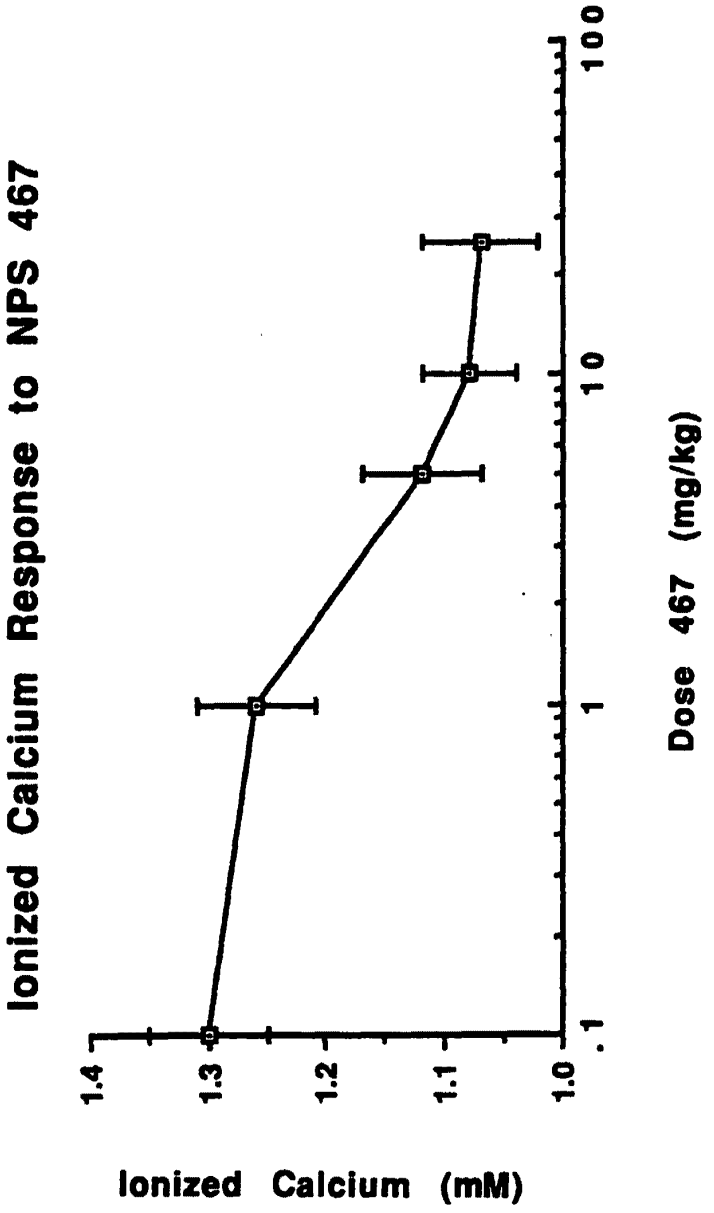


FIG. 38


Ionized Calcium Response to NPS 467
Dose 467 (mg/kg)
.1
1
10
100
1.4
1.3
1.2
1.1
1.0
Ionized Calcium (mM)

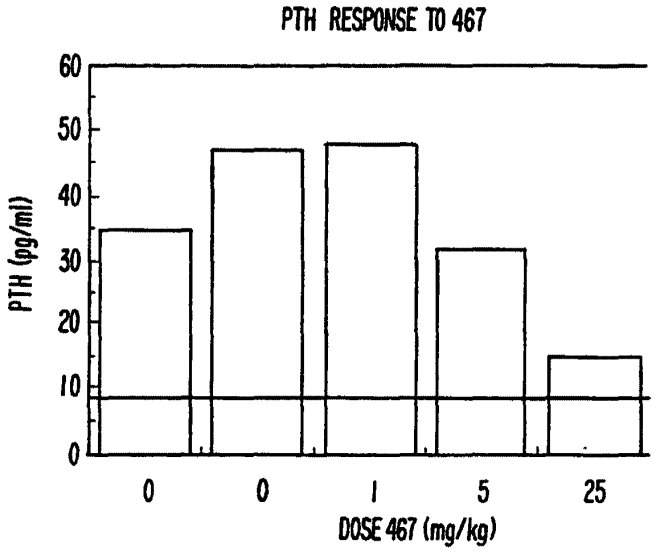

PTH RESPONSE TO 467
PTH (pg/ml)
60
50
40
30
20
10
0
0
0
1
5
25
DOSE 467 (mg/kg)

FIG. 39.

FIG. 40.

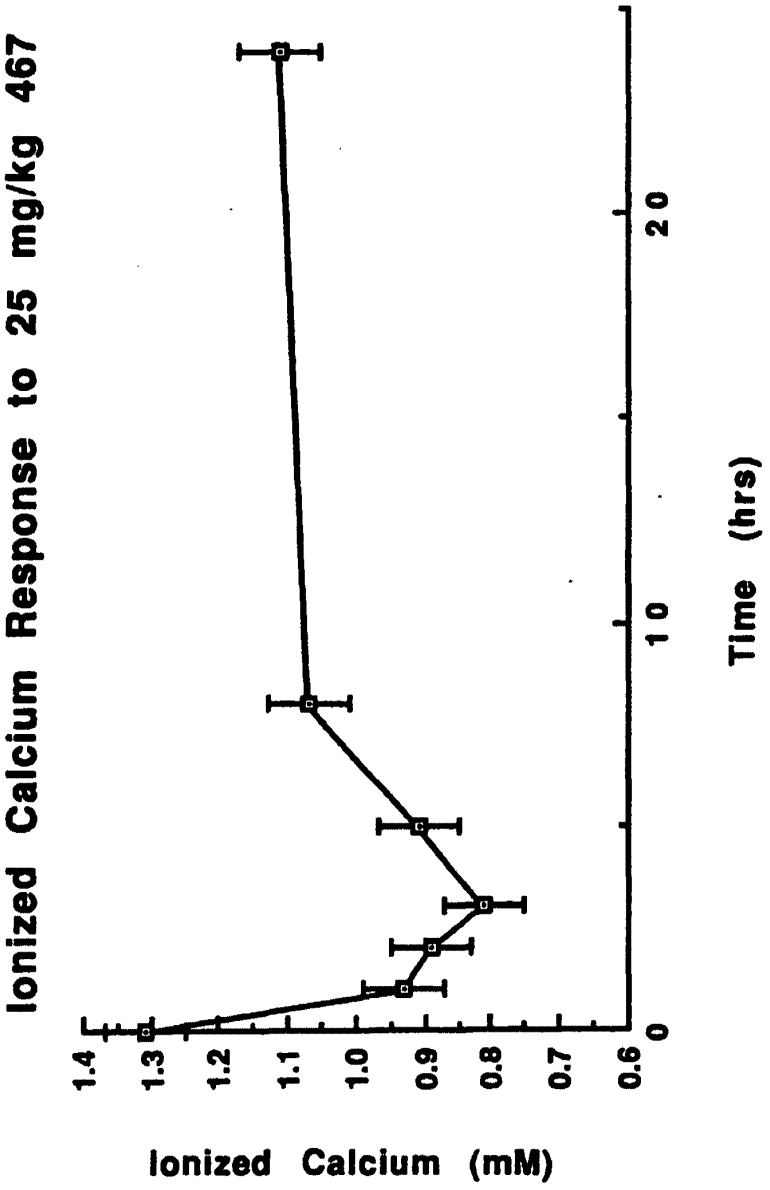
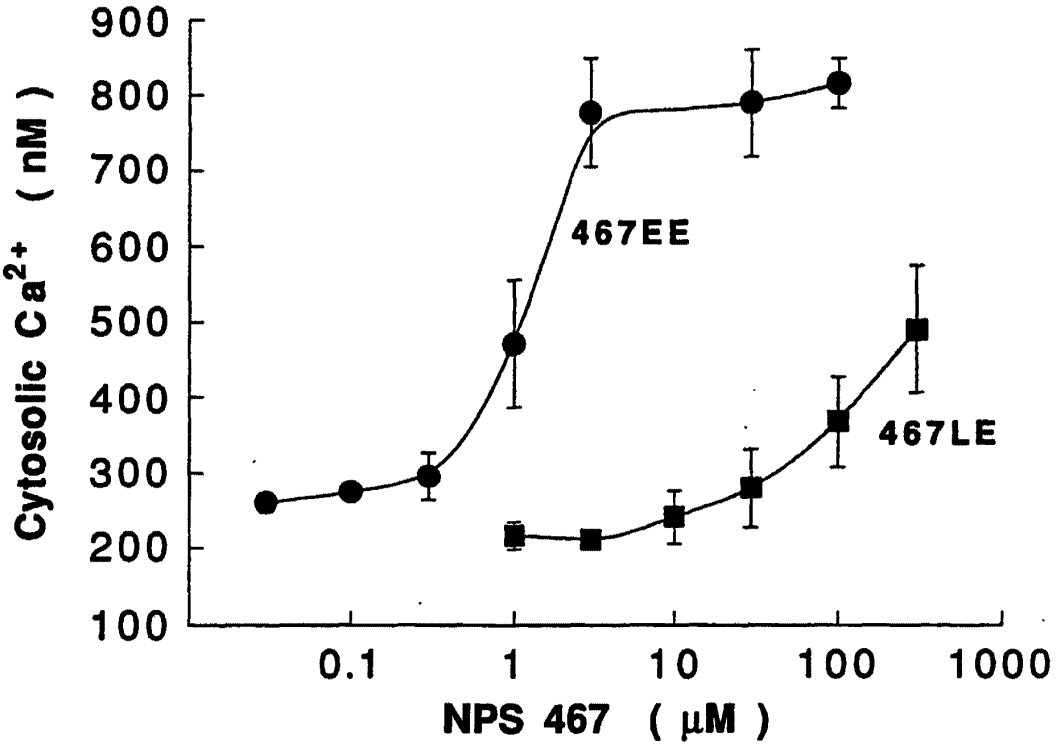

Ionized Calcium Response to 25 mg/kg 467
1.4
1.3
1.2
1.1
1.0
0.9
0.8
0.7
0.6
Ionized Calcium (mM)
0
10
20
Time (hrs)

FIG. 41.


900
800
700
600
500
400
300
200
100
Cytosolic Ca^{2+} (nM)
467EE
467LE
0.1
1
10
100
1000
NPS 467 (μM)

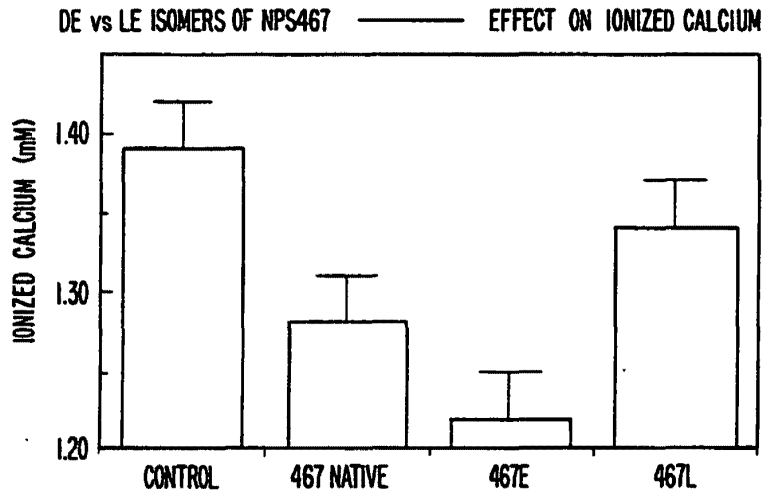

DE vs LE ISOMERS OF NPS467 ——— EFFECT ON IONIZED CALCIUM
IONIZED CALCIUM (mM)
1.40
1.30
1.20
CONTROL
467 NATIVE
467E
467L

FIG. 42.

FIG. 43a.

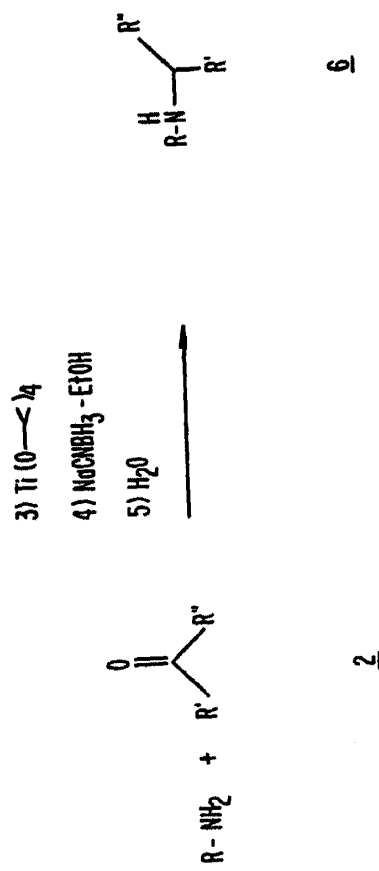
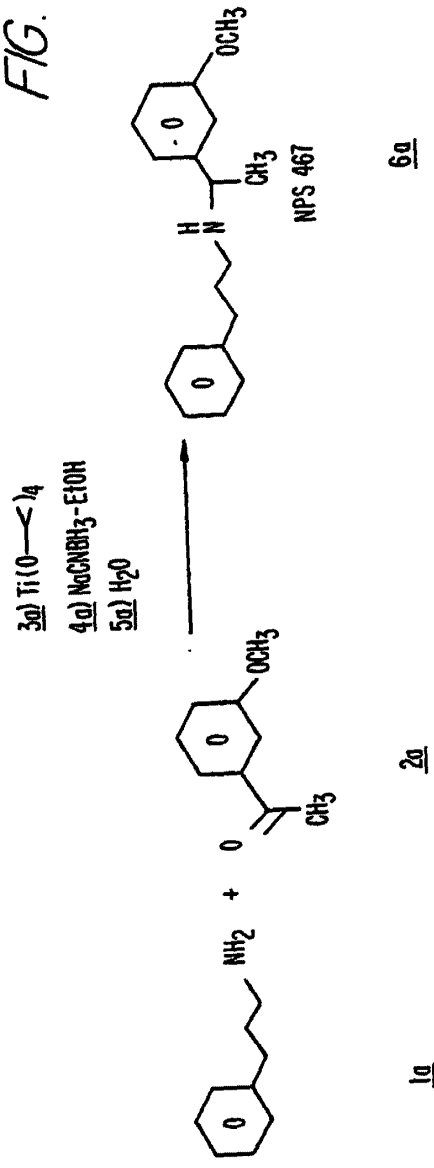


FIG. 43b.



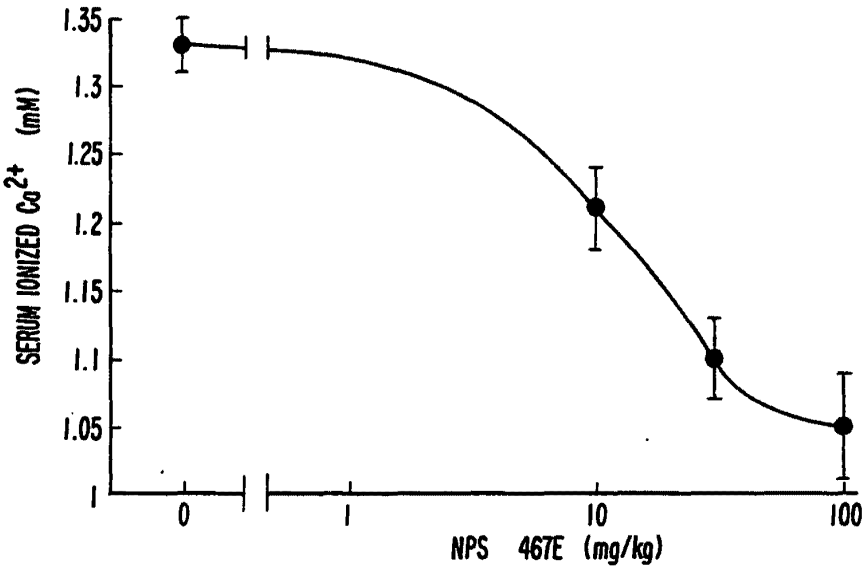

1.35
1.3
1.25
1.2
1.15
1.1
1.05
1
SERUM IONIZED Ca^{2+} (mM)
0
1
10
100
NPS 467E (mg/kg)

FIG. 44.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US92/07175

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :Please See Extra Sheet.
US CL :436/501, 503, 69.1; 536/27; 564/511, 305; 530/324, 350; 564/374
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 436/501, 503, 69.1; 536/27; 564/511, 305; 530/324, 350; 564/374

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

CAS ONLINE, APS,
search terms: structure search, calcium agonists and antagonists

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X Y	US, A, 4,988,730 (Korbonits et al) 29 January 1991, see entire document, and Table 2.	1-23, 25-27, 29-31, 36-61, 63-65, 67-69, 74-82, 89, 98, 99 32-35, 70-73
Y	CIRCULATION RESEARCH, Vol. 64, No. 3, Issued March 1989, Koenig et al, "Polyamines Mediate Androgenic Stimulation of Calcium Fluxes and Membrane transport in Rat Heart Myocytes", pages 415-426, see the abstract and page 415, column 2.	23, 24
X Y	WO, A, 89/09834 (Ellis et al) 19 October 1989, see pages 6-8, 9, 10-11, 27-28.	83, 84, 90, 92, 93, 97 85-88, 94-96
X Y	QUANTITATIVE STRUCTURE-ACTIVITY RELATIONSHIPS, Vol. 8, Issued 1989, Holtje et al, "Conformational Analysis on Calcium Channel Active Diphenylalkylamines, Diphenylbutylpiperidines, Phenylalkylamines, and Perhexiline" pages 259-265, see Figure 1.	99 1-23, 25-35

[X] Further documents are listed in the continuation of Box C. [ ] See patent family annex.

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"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

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"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
02 December 1992	09 DEC 1992
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Form PCT/ISA/210 (second sheet)(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US92/07175

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (5):

G01N 33/566, 33/567; C07C 211/02, 211/16, 211/27,; C07H 21/00; C07K 5/00, 7/00; C12N 15/12; A61K 37/02

Form PCT/ISA/210 (extra sheet)(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US92/07175

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	ANTICANCER RESEARCH, Vol. 11, Issued May-June 1991, Batra et al, "Effects of Diverse Categories of Drugs on Human Colon Tumour Cell Proliferation", pages 1221-1224, see abstract.	36-48, 50, 54, 58, 60, 61, 63-65, 68, 69, 77, 79-82
X	CARDIOVASCULAR DRUGS AND THERAPY, Vol. 2, No. 1, Issued May 1988, Opie, "Calcium Channel Antagonists Part V: Second-Generation Agents", pages 191-203, see entire article.	1-23, 25-27, 30, 31, 36-61, 63-65, 68-69, 74-77, 89
A	MEDICINAL RESEARCH REVIEWS, Vol. 9, No. 1, issued 1989, Triggle et al, "Ca2+ Channel Ligands; Structure-Function Relationships of the 1,4-Dihydropyridines", pages 123-180, see entire article.	1-99

Form PCT/ISA/210 (continuation of second sheet)(July 1992)*

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Published
With international search report.

(54) Title: CALCIUM RECEPTOR-ACTIVE MOLECULES

(57) Abstract

Method and composition useful for treating a patient having a disease characterized by an abnormal level of one or more components, the activity of which is regulated or affected by activity of one or more inorganic-ion receptor. Novel compounds useful in these methods and compositions are also provided. The method includes administering to the patient a therapeutically effective amount of a molecule active at one or more inorganic-ion receptors as an agent or antagonist. Preferably, the molecule is able to act as either a selective agonist or antagonist at a Ca^{2+} receptor of one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, distal tubule kidney cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cells), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin and glucagon secreting cells, kidney mesangial cell and mammary cell.

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CALCIUM RECEPTOR-ACTIVE MOLECULES

Field of the Invention

This invention relates to the design, development, composition and use of novel calcimimetic molecules able to act in a manner analogous to extracellular calcium ions on cells, to calcilytic molecules which block the activity of extracellular calcium ions on cells, and to methods for their use and identification.

It also relates to a novel superfamily of inorganic-ion receptors which includes, among others, calcium receptors, nucleic acids encoding such receptors, cells, tissues and animals containing such nucleic acids, antibodies to receptors, assays utilizing receptors, and methods relating to all of the foregoing.

Background of the Invention

The following description provides a summary of information relevant to the present invention. It is not an admission that any of the information provided herein is prior art to the presently claimed invention, nor that any of the publications specifically or implicitly referenced

are prior art to that invention.

Certain cells in the body respond not only to chemical signals, but also to ions such as extracellular calcium ions (Ca^{2+}). Changes in the concentration of extracellular Ca^{2+} (referred to herein as "[Ca^{2+}]") alter the functional responses of these cells. One such specialized cell is the parathyroid cell which secretes parathyroid hormone (PTH). PTH is the principal endocrine factor regulating Ca^{2+} homeostasis in the blood and extracellular fluids.

PTH, by acting on bone and kidney cells, increases the level of Ca^{2+} in the blood. This increase in [Ca^{2+}] then acts as a negative feedback signal,depressing PTH secretion. The reciprocal relationship between [Ca^{2+}] and PTH secretion forms the essential mechanism maintaining bodily Ca^{2+} homeostasis.

Extracellular Ca^{2+} acts directly on the parathy roid cell to regulate PTH secretion. The existence of a parathyroid cell surface protein which detects changes in [Ca^{2+}] has been suggested. This protein acts as a receptor for extracellular Ca^{2+} ("the Ca^{2+} receptor"), and is suggested to detect changes in [Ca^{2+}] and to initiate a functional cellular response, PTH secretion. For example, the role of Ca^{2+} receptors and extracellular Ca^{2+} in the regulation of intracellular Ca^{2+} and cell function is reviewed in Nemeth et al., 11 Cell Calcium 319, 1990; the role of Ca^{2+} receptors in parafollicular and parathyroid cells is discussed in Nemeth, 11 Cell Calcium 323, 1990; and the role of Ca^{2+} receptors on bone osteoclasts is discussed by Zaidi, 10 Bioscience Reports 493, 1990.

Other cells in the body, specifically the osteoclast in bone, the juxtaglomerular, proximal tubule cells in the kidney, the keratinocyte in the epidermis, the parafollicular cell in the thyroid, intestinal cells the trophoblast in the placenta, have the capacity to sense changes in $[Ca^{2+}]$. It has been suggested that cell surface Ca^{2+} receptors may also be present on these cells, imparting to them the ability to detect and to initiate or enable a response to changes in $[Ca^{2+}]$.

In parathyroid cells, osteoclasts, parafollicular cells (C-cells), keratinocytes, juxtaglomerular cells and trophoblasts, an increase in $[Ca^{2+}]$ evokes an increase in intracellular free Ca^{2+} concentration ("$[Ca^{2+}]_i$"). Such an increase may be caused by influx of extracellular Ca^{2+} or by mobilization of Ca^{2+} from intracellular organelles. Changes in $[Ca^{2+}]_i$ are readily monitored and quantitated using fluorimetric indicators such as fura-2 or indo-1 (Molecular Probes, Eugene, OR). Measurement of $[Ca^{2+}]_i$ provides an assay to assess the ability of molecules to act as agonists or antagonists at the Ca^{2+} receptor.

In parathyroid cells, increases in the concentration of extracellular Ca^{2+} evoke rapid and transient increases in $[Ca^{2+}]_i$ which are followed by lower yet sustained increases in $[Ca^{2+}]_i$. The transient increases in $[Ca^{2+}]_i$ arise from the mobilization of intracellular Ca^{2+}, whereas the lower, sustained increases result from the influx of extracellular Ca^{2+}. The mobilization of intracellular Ca^{2+} is accompanied by increased formation of inositol-1,4,5-trisphosphate (IP_3)

and diacylglycerol, two biochemical indicators which are associated with receptor-dependent mobilization of intracellular Ca^{2+} in various other cells.

In addition to Ca^{2+}, various other di- and trivalent cations, such as Mg^{2+}, Sr^{2+}, Ba^{2+}, La^{3+}, and Gd^{3+} also cause the mobilization of intracellular Ca^{2+} in parathyroid cells. Mg^{2+} and La^{3+} also increase the forma tion of IP_3; all these inorganic cations depress the secretion of PTH. The postulated Ca^{2+} receptor on the parathyroid cell is therefore promiscuous because it detects a variety of extracellular di- and trivalent cations.

The ability of various compounds to mimic extracellular Ca^{2+} in vitro is discussed by Nemeth et al., (spermine and spermidine) in "Calcium-Binding Proteins in Health and Disease", 1987, Academic Press, Inc., pp. 33-35; Brown et al., (e.g., neomycin) 128 Endocrinology 3047, 1991; Chen et al., (diltiazem and its analog, TA-3090) 5 J. Bone and Mineral Res. 581, 1990; and Zaidi et al., (verapamil) 167 Biochem Biophys Res Comm 807, 1990.

Brown et al., 6 J. Bone and Mineral Res. 11, 1991 discuss the existing theories regarding the effects of Ca^{2+} ions on parathyroid cells, and propose that the results may be explained by both a receptor-like mechanism and a receptor-independent mechanism as follows:

> Polyvalent cations [e.g., divalent and trivalent cations] exert a variety of effects on parathyroid function, such as inhibition of parathyroid hormone (PTH) secretion and cAMP accumulation, stimulation of the accumulation of inositol phosphates, and elevation of the cytosolic calcium

concentration. These actions are thought to be mediated through a "receptor-like" mechanism. The inhibition of agonist-stimulated cAMP accumulation by divalent and trivalent cations, for example, is blocked following preincubation with pertussis toxin. Thus, the putative polyvalent cation receptor may be coupled to inhibition of adenylate cyclase by the inhibitory guanine nucleotide regulatory (G) protein, G_i.

We recently showed that the polycationic antibiotic, neomycin, mimics the actions of di- and trivalent cations in several aspects of parathy roid function. To determine whether these actions were specific to this agent or represented a more generalized action of polycations, we tested the effects of the highly basic peptides, polyarginine and polylysine, as well as protamine on the same parameters in dispersed bovine parathyroid cells. The results demonstrate that the parathyroid cell responds to a variety of polycations as well as to polyvalent cations, potentially via similar biochemical pathways. These results are discussed in terms of the recently postulated, "receptor-independent" modulation of G proteins by polycations in other systems.

* * *

The Ca^{2+} receptor has been presumed to be analogous to other G protein-coupled receptors [e.g., a glycoprotein], but recent studies with other cell types have raised the possibility that polycations can modulate cell function by alternative or additional mechanisms. In mast cells, for example, a variety of amphipathic cations, including mastoparan, a peptide from wasp venom, 48/80, a synthetic polycation, and polylysine, enhance secretion by a pertussis

toxin-sensitive mechanism, suggesting the involvement of a G protein. No classic cell surface receptor has been identified that could mediate the actions of these diverse agents. Furthermore, these same compounds have been shown to activate directly purified G proteins in solution or in artificial phospholipid vesicles. On the basis of these observations, it has been proposed that amphipathic cations activate G proteins and, in turn, mast cell secretion by a "receptor-independent" mechanism.

Polycations have also been shown to interact strongly with acidic phospholipids. Polylysines of varying chain lengths (20-1000 amino acids) bind to artificial phospholipid vesi cles with dissociation constants in the range of 0.5 nM to 1.5 μM. The binding affinity is directly related to the length of the polylysine chain, with polymers of 1000 amino acids having a K_d of 0.5 nM, shorter polymers having higher Kd values, and lysine not interacting to a significant extent. This relationship between potency and chain length is similar to that observed for the effects of polylysine $_{10,200}$, polylysine $_{3800}$, and lysine on parathyroid function.

It is possible that the binding of polycations to biomembranes produces some of their biologic actions. The permeabilization of the plasma membrane induced in some cell types by a variety of pore-forming agents, including polycations, has been postulated to be mediated by their interaction with a phosph atidylserine-like structure. In addition, the "receptor-independent" activation of purified G proteins by amphipathic cations is potentiated when these proteins are incorporated into phospholipid vesicles.

Calcium ions, in the millimolar concentration range, also produce marked changes in membrane structure. In some cases, calcium can either antagonize or

potentiate the interaction of polycations with membrane lipids. These considerations raise the possibility that the actions of both polyvalent cations and polycations on parathyroid cells could involve a receptor-independent mechanism not requiring the presence of a classic, cell surface, G protein-coupled receptor. Further studies, however, are required to elucidate the molecular basis for Ca^{2+} sensing by this and other cell types. [Citations omitted.]

Shoback and Chen (6 (Supplement 1), J. Bone and Mineral Res. 1991, S135) and Racke et al. (6 (Supplement 1), J. Bone and Mineral Res. 1991, S118) describe experiments which are said to indicate that a Ca^{2+} receptor or Ca^{2+} sensor is present in parathyroid cells. Messenger RNA isolated from such cells can be expressed in oocytes and caused to provide those oocytes with a phenotype which might be explained by the presence of a Ca^{2+} receptor protein.

Summary of the Invention

Applicant has demonstrated that Ca^{2+} receptor proteins enable certain specialized cells involved in bodily Ca^{2+} metabolism to detect and respond to changes in the concentration of extracellular Ca^{2+}. Although these receptors share certain general characteristics, they can be selectively affected by different pharmacological agents. As detailed below, certain molecules are identified with selective activity on Ca^{2+} receptors at parathyroid cells, osteoclasts, and C-cells.

Ca^{2+} receptors constitute discrete molecular targets for a new class of molecules that mimic ("calcimimetics") or antagonize ("calcilytics") the actions of extracellular Ca^{2+}. Such receptors are present on cell

surfaces and have a low affinity for extracellular Ca^{2+} (apparent K_d generally greater than about 0.5 mM). Such receptors may include a free or bound effector mechanism, as defined by Cooper, Bloom and Roth, "The Biochemical Basis of Neuropharmacology", Ch. 4. Such receptors are thus distinct from intracellular Ca^{2+} receptors, e.g., calmodulin and the troponins. Calcimimetics, for example, act on Ca^{2+} receptors selectively to directly or indirectly depress the function of parathyroid cells or osteoclasts or to stimulate the function of C-cells. Calcimimetics and calcilytics of this invention allow novel therapies for hyperparathyroidism, osteoporosis and other Ca^{2+}-related diseases. This application concerns in one aspect targeting Ca^{2+} receptors on each of these three cell types and other cell types that detect and respond to changes in $[Ca^{2+}]$.

Applicant is the first to demonstrate a Ca^{2+} receptor protein in parathyroid cells, and to pharmacologically differentiate such Ca^{2+} receptors in other cells, such as C-cells and osteoclasts. Applicant is also the first to describe methods by which molecules active at these Ca^{2+} receptors can be identified and used as lead molecules in the discovery, development, design, modification and/or construction of useful calcimimetics or calcilytics which are active at Ca^{2+} receptors. Such calcimimetics or calcilytics are useful in the treatment of various disease states characterized by abnormal levels of one or more components, e.g., polypeptides such as hormones, enzymes or growth factors, the expression and/or secretion of which is regulated or affected by activity at one or more Ca^{2+} receptors. Further, the identification of different

Ca^{2+} receptors in different cell types, and the specific response of such receptors to different lead molecules allows design and construction of specific molecules active in treatment of specific diseases which can be affected by action at such specific Ca^{2+} receptors. For example, abnormal levels of parathyroid hormone secretion can be affected by such specific molecules without affecting the level of secretion of other Ca^{2+} regulated hormones and the like.

Identification of such lead molecules was impeded by the prior lack of a high-throughput screening system to discover active molecules, and the absence of a structural data base upon which to design effective drug candidates. These barriers are now removed by cloning the parathyroid cell Ca^{2+} receptor and functionally related receptors, and systematically examining the structural features of certain lead molecules that activate such cloned Ca^{2+} receptors and functionally related receptors. Cloning of the Ca^{2+} receptor also enables development of transfected cell lines suitable for high-throughput screening of natural product or molecule libraries and synthetic molecules. This, together with structure-activity studies discussed below, provides the technology necessary to develop novel calcimimetics and calcilytics.

Applicant enables such procedures in this application. A bovine parathyroid cell calcium receptor cDNA has been cloned and is deposited in the ATCC under accession number ATCC 75416. Using this clone, inorganic-ion receptors in other tissues and species homologs are easily obtained. For example, the human parathyroid cell Ca^{2+}

receptor cDNA can be cloned by screening nucleic acid libraries or by screening for functional expression in Xenopus oocytes, and the structural features of organic molecules necessary for activity on the Ca^{2+} receptor can be determined through the testing of selected natural products or other molecule libraries and subsequent structure-activity studies.

Thus, in a first aspect, the invention features a pharmaceutical composition including a molecule which either mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell, or blocks an increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. The molecule has an EC_{50} of less than or equal to 5 μM, and is not protamine.

By "mimic" is meant that the molecule has one or more of the specific actions of extracellular Ca^{2+} on an extracellular Ca^{2+} responsive cell. The term does not require that all of the biological functions of extracellular Ca^{2+} are mimicked, but rather that at least one such function is mimicked. In addition it does not require that the molecule bind to the same site on the Ca^{2+} receptor as does extracellular Ca^{2+} (see for example, the novel compound NPS 467 and its action in Example 20 below). By "block" is meant that one such action of Ca^{2+} is reduced or prevented by the molecule. The EC_{50} can be determined in assays as described below, where the activity mimicked is measured and the concentration of molecule which mimics at half the maximum mimicking effect is the EC_{50}. Conversely, the IC_{50} of a calcilytic is that amount which blocks half maximal activity. Preferably,

such assays measure $[Ca^{2+}]_i$ increases and are confirmed to be specific to a Ca^{2+} receptor by methods described below, or their equivalent.

In preferred embodiments, bioassays described herein demonstrate that the increase in $[Ca^{2+}]_i$ in a cell is transient, having a duration of less than one minute, and the increase in $[Ca^{2+}]_i$ is rapid, occurring within thirty seconds; and the molecule also (a) evokes a sus tained increase (greater than thirty seconds) in $[Ca^{2+}]_i$, (b) evokes an increase in inositol-1,4,5-trisphosphate and/or diacylglycerol levels, e.g., within less than 60 seconds, and (c) inhibits dopamine- or isoproterenol- stimulated cyclic AMP formation. In addition, the tran sient increase in $[Ca^{2+}]_i$ is abolished by pretreatment of the cell for ten minutes with 10 mM sodium fluoride, or the transient increase is diminished by brief pretreatment (not more than ten minutes) of the cell with an activator of protein kinase C, e.g., phorbol myristate acetate (PMA), mezerein or (-)indolactam V.

In a parathyroid cell, those molecules which are active in all of the assays described above are particularly useful in this invention since they are specific in their actions to a Ca^{2+} receptor of such a cell. This is particularly true for the PMA pretreatment effect described above.

In a more preferred embodiment, the cell is a parathyroid cell, and the molecule inhibits parathyroid hormone secretion from the cell. Other preferred embodiments include molecules that elicit an increase in

$[Ca^{2+}]_i$ as detected, for example, as an increase in Cl^- current in a Xenopus oocyte injected with mRNA from a parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cells), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin and glucagon secreting cells, kidney mesangial cell and mammary cell.

In other preferred embodiments, the molecule evokes the mobilization of intracellular Ca^{2+} to cause the increase in $[Ca^{2+}]_i$; the cell is a C-cell or an osteoclast and the molecule inhibits bone resorption in vivo; the cell is an osteoclast and the molecule inhibits bone resorption in vitro; or the cell is a C- cell and the molecule stimulates calcitonin secretion in vitro or in vivo; and most preferably the molecule is either a calcimimetic or calcilytic having an EC_{50} or IC_{50} at a Ca^{2+} receptor of less than or equal to 5 μM, and even more preferably less than or equal to 1 μM, 100 nmolar, 10 nmolar, or 1 nmolar. Such lower EC_{50}'s or IC_{50}'s are advantageous since they allow lower concentration of molecules to be used in vivo or in vitro for therapy or diagnosis. The discovery of molecules with such low EC_{50}'s and IC_{50}'s enables the design and synthesis of similarly potent and efficacious molecules.

By "calcimimetic" molecule is meant any molecule which has one or more activities of extracellular Ca^{2+}, and preferably mimics the activity of Ca^{2+} at a Ca^{2+}

receptor. For example, when used in reference to a parathyroid cell it is a molecule which, when tested on parathyroid cells in vitro, possesses one or more, and preferably all of the following characteristics as measured by techniques well known to those in the art:

1. The molecule causes a rapid (time to peak < 5 sec) and transient increase in $[Ca^{2+}]_i$ that is refractory to inhibition by 1 μM La^{3+} or Gd^{3+}. The increase in $[Ca^{2+}]_i$ persists in the absence of extra cellular Ca^{2+} but is abolished by pretreatment with ionomycin (in the absence of extracellular Ca^{2+});

2. The molecule potentiates increases in $[Ca^{2+}]_i$ elicited by submaximal concentrations of extracellular Ca^{2+};

3. The increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+} is not inhibited by dihydropyridines;

4. The transient increase in $[Ca^{2+}]_i$ caused by the molecule is abolished by pretreatment for 10 min. with 10 mM sodium fluoride;

5. The transient increase in $[Ca^{2+}]_i$ caused by the molecule is diminished by pretreatment with an activator of protein kinase C (PKC), such as phorbol myristate acetate (PMA), mezerein or (-)-indolactam V. The overall effect of the protein kinase C activator is to shift the concentration-response curve of the molecule to the right without affecting the maximal response;

6. The molecule causes a rapid (< 30 sec.) increase in the formation of inositol-1,4,5- trisphosphate and or diacylglycerol;

7. The molecule inhibits dopamine- or isopro

terenol-stimulated cyclic AMP formation;

8. The molecule inhibits PTH secretion;

9. Pretreatment with pertussis toxin (100 ng/ml for > 4 hrs.) blocks the inhibitory effect of the molecule on cyclic AMP formation but does not effect increases in $[Ca^{2+}]_i$, inositol-1,4,5-trisphosphate, or diacylglycerol, nor decreases in PTH secretion;

10. The molecule elicits increases in $[Ca^{2+}]_i$ as detected, for example, as an increase in Cl^- current in Xenopus oocytes injected with poly(A)$^+$- enriched mRNA from bovine or human parathyroid cells but is without effect in Xenopus oocytes injected with water or rat brain or liver mRNA; and

11. Similarly, using a cloned receptor from parathyroid cells, the molecule will elicit a response in Xenopus oocytes injected with the specific cDNA, mRNA or synthetic sense RNA (cRNA) encoding the receptor.

By "calcilytic" molecule is meant any molecule which blocks one or more of the activities of extracellular Ca^{2+} on an extracellular Ca^{2+}-sensing cell, preferably by acting as an antagonist at the Ca^{2+} receptor. For example, when used in reference to a parathyroid cell, it is a molecule which, when tested on parathyroid cells in vitro, possesses one or more, and preferably all of the following characteristics as measured by techniques well known to those in the art:

1. The molecule blocks, either partially or completely, the ability of increased concentrations of extracellular Ca^{2+} to:

a) increase $[Ca^{2+}]_i$,

b) mobilize intracellular Ca^{2+},

c) increase the formation of inositol-1,4,5-trisphosphate,

d) decrease dopamine- or isoproterenol-stimulated cyclic AMP formation, and

e) inhibit PTH secretion;

2. At low [Ca^{2+}], i.e., 0.5 mM, the molecule by itself does not change $[Ca^{2+}]_i$;

3. The molecule blocks increases in Cl^- current in Xenopus oocytes injected with poly(A)$^+$- mRNA from bovine or human parathyroid cells elicited by extracellular Ca^{2+} or calcimimetic compounds but not in Xenopus oocytes injected with water or rat brain or liver mRNA;

4. Similarly, using a cloned receptor from parathyroid cells, the molecule will block a response in Xenopus oocytes injected with the specific cDNA, mRNA or cRNA encoding the Ca^{2+} receptor, elicited by extracellular Ca^{2+} or a calcimimetic compound.

Parallel definitions of useful calcimimetics and calcilytics at Ca^{2+} receptors on other cell types are evident from the examples provided below.

The Ca^{2+} receptor is able to detect and respond to certain inorganic polycations and polycationic organic molecules. For example, the parathyroid cell is unable to distinguish increases in extracellular Ca^{2+} concentration from the addition of these organic polycations, presumably because these organic molecules act just like extracellular Ca^{2+} at the Ca^{2+} receptor. The calcimimetic molecules of this invention are particularly good agonists of the

Ca^{2+} receptor and may be used as drugs that alter selected cellular functions, e.g., secretion of PTH from parathyroid cells. Unlike Ca^{2+} most of these molecules act only at one or more, but not all Ca^{2+} receptors, and thus provide an ability to specifically target one Ca^{2+} receptor.

These molecules also provide lead structures for the development of further novel therapeutics effective in the treatment of various diseases where $[Ca^{2+}]_i$ and $[Ca^{2+}]$ play a role, such as hyperparathyroidism, osteoporosis, Paget's disease, hypertension, renal disease, skin disease, cardiovascular disease, blood clotting disorders, gastorintestinal diseases, endocrine diseases, abnormalities in water metabolism and cancer.

The calcimimetics and calcilytics can be formu lated as pharmaceutical compositions which are useful for regulating the level of extracellular free Ca^{2+} in a patient and for mimicking the effect of extracellular Ca^{2+} on a cell selected from the group described above, by administering to the patient such a pharmaceutical composition. Prior to this invention, applicant was unaware of any such molecules acting on the Ca^{2+} receptor useful in treatment of diseases caused by irregularity in operation or regulation of a Ca^{2+} receptor or diseases in an animal having normal Ca^{2+} receptors but which can be treated by activating or deactivating such Ca^{2+} receptors.

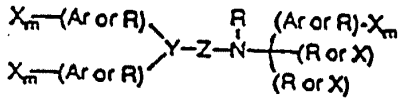
In yet another preferred embodiment, the molecule has an EC_{50} less than or equal to 5 μM at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, distal tubule kidney

cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cells), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin and glucagon secreting cells, kidney mesangial cell and mammary cell.

It is the specificity of action of such molecules that is particularly advantageous in this invention since it allows specific in vivo and in vitro therapy and diagnosis and discovery of additional calcimimetic or calcilytic molecules.

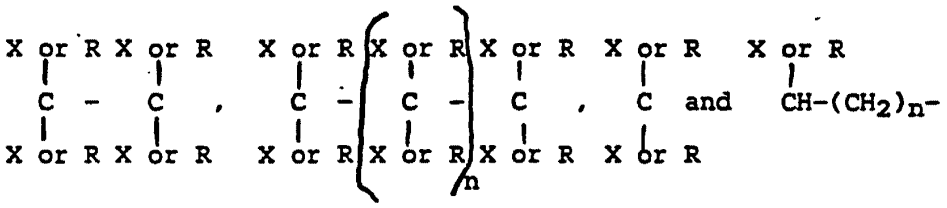
In specific preferred embodiments, the molecule is positively charged at physiological pH, and is selected from the group consisting of branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines, e.g., the branched polyamine has the formula $H_2N-(CH_2)_j-(NR_i-(CH_2)_j)_k-NH_2$ where k is an integer from 1 to 10, each j is the same or different and is an integer from 2 to 20, and each R_i is the same or different and is selected from the group consisting of hydrogen and $-(CH_2)_j-NH_2$, where j is as defined above, and at least one R_i is not hydrogen.

In an alternative embodiment, the molecule has the formula



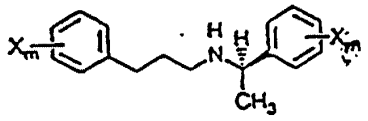
where each X independently is selected from the group

consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, and CH_3CH_2; Ar is a hydrophobic entity; each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, 2-, 3-, or 4-piperid(in)yl; Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon; Z is selected from the group consisting of oxygen, nitrogen, sulfur,



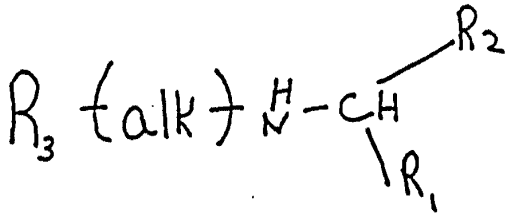
where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive. Most preferably the molecule is either a calcimimetic or calcilytic.

In preferred embodiments, the hydrophobic entity is selected from the group consisting of phenyl, 2-, 3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2- quinolinyl, 2- or 3-indolyl, benzyl, and phenoxy; the molecule is an R-phenylpropyl-α-phenethylamine derivative, and the molecule has the formula:



with each X preferably being independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

According to a preferred aspect of the present invention, novel phenylpropyl-α-phenethylamine analogs and derivatives are provided having the formula:



wherein alk is straight or branched chain alkylene of from 1 to 6 carbon atoms;
R_1 is lower alkyl of from 1 to 3 carbon atoms or lower haloalkyl of from 1 to 3 carbon atoms substituted with from 1 to 7 halogen atoms; R_2 and R_3 are independently selected carbocyclic aryl or cycloalkyl groups, either monocyclic or bicyclic, having 5- or 6-membered rings
optionally substituted with 1 to 5 substituents independently selected from lower alkyl of 1 to 3 carbon atoms, lower haloalkyl of 1 to 3 carbon atoms substituted with 1 to 7 halogen atoms, lower alkoxy of 1 to 3 carbon atoms, halogen, nitro, amino, alkylamino, amido, lower alkylamido of 1 to 3 carbon atoms, cyano, hydroxy, acyl of 2 to 4 carbon atoms lower hydroxyalkyl of 1 to 3 carbon atoms or lower thioalkyl of 1 to 3 carbon atoms. Suitable carbocyclic aryl groups are groups having one or two rings, at least one of which having aromatic character and include carbocyclic aryl groups such as phenyl and bicyclic carbocyclic aryl groups such as naphthyl. As is apparent from the above formula, the compounds encompassed therein may exist as racemic mixtures and as individual stereoisomers. Especially preferred are R-phenylpropyl α-phenethylamine derivatives which exhibit enhanced activity in lowering serum ionized calcium.

Preferred compounds include those where alk is n-propylene. Also preferred are compounds where R_1 is

methyl. Also preferred are those compounds where R_2 and R_3 are optionally substituted phenyl.

Especially preferred compounds include those where R_2 is monosubstituted phenyl, more preferably meta-substituted. Especially preferred R_3 groups include unsubstituted or monosubstituted phenyl, especially ortho-substituted. Preferred substitutents for R_2 include halogen, haloalkyl, preferably trihalomethyl, and alkoxy, preferably methoxy. Preferred substituents for R_3 include halogen, preferably chlorine.

In a second related aspect, the invention fea tures a method for treating a patient having a disease or condition characterized by an abnormal $[Ca^{2+}]$ or $[Ca^{2+}]_i$ in one or more cells or in the blood or plasma or extracellular fluids. The method includes the step of administering to the patient a therapeutically effective amount of a molecule which either mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell or blocks an increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}.

By "abnormal" is meant that the patient, compared to the general population, has a different Ca^{2+} metabolism that is affected by one or more proteins (e.g., hormones) in the blood or extracellular body fluids, or other molecules which affect the level of extracellular and/or intracellular Ca^{2+}. Thus, the diseases include hyperparathyroidism, osteoporosis and other bone and mineral-related disorders, and the like (as described, e.g., in standard medical text books, such as "Harrison's Principles of Internal Medicine"). Such diseases are treated in this invention by molecules which mimic or block one or more of the effects of Ca^{2+} and

thereby directly or indirectly affect the levels of the proteins or other molecules in the body of the patient.

By "therapeutically effective amount" is meant an amount that relieves to some extent one or more symptoms of the disease or condition in the patient. Additionally, by "therapeutically effective amount" is meant an amount that returns to normal, either partially or completely, physiological or biochemical parameters associated with or causative of the disease or condition. Generally, it is an amount between about 1 nmole and 1 μmole of the molecule, dependent on its EC_{50} and on the age, size, and disease associated with the patient.

In preferred embodiments, the molecule has an EC_{50} of less than or equal to 5 μM, and is not protamine; and most preferably interacts at a Ca^{2+} receptor as a calcimimetic or calcilytic. Most preferably the molecule is chosen from one of those described above.

In other preferred embodiments, the patient has a disease characterized by an abnormal level of one or more components the level of which is regulated or affected by activity of one or more Ca^{2+} receptors, and the molecule is active on a Ca^{2+} receptor of a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, distal tubule kidney cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cells), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin and glucagon secreting cells, kidney mesangial cell and mammary cell.

In still other preferred embodiments, the mole cule reduces the level of parathyroid hormone in the serum of the patient, e.g., to that level present in a normal individual, or to a degree sufficient to cause a decrease in plasma Ca^{2+}; and the molecule is provided in an amount sufficient to have a therapeutically relevant effect on the patient.

In a third aspect, the invention features a method for diagnosis of a disease or condition in a patient by identifying the number and/or location (and/or functional integrity) of one or more Ca^{2+} receptors within the patient and comparing that number and/or location (and/or functional integrity) with that observed in normal patients as an indication of the presence of the disease or condition.

In preferred embodiments, the method is an immunoassay in which an antibody to a Ca^{2+} receptor is used to identify the number and/or location and/or functional integrity of the Ca^{2+} receptors, or the assay involves providing a labelled calcimimetic or calcilytic molecule which binds to a Ca^{2+} receptor; and the disease diagnosed is a cancer, e.g., an ectopic tumor of the parathyroid, or a condition characterized by an above normal level in the number of osteoclasts in bone or an increased level of activity of osteoclasts in bone.

In a fourth aspect, the invention features a method for identifying a molecule useful as a therapeutic molecule. The method includes screening a potentially useful molecule for either an ability to mimic the activity of extracellular Ca^{2+} in a cell, or to block an increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}, and determining whether the molecule has an EC_{50} or IC_{50} of less than or equal to 5 μM.

In other aspects, the invention features a recombinant Ca^{2+} receptor, a cell including a recombinant Ca^{2+} receptor, purified nucleic acid encoding a Ca^{2+} receptor, the biological activity and use of the molecule NPS 019, the novel compounds or compositions of matter of NPS 459, NPS 467, and NPS 568 (see Fig. 36) and a method for identifying a useful calcimimetic or calcilytic molecule by identifying a molecule which mimics or blocks one or more activities of Ca^{2+} at a first Ca^{2+} receptor but not at a second Ca^{2+} receptor, e.g., by use of a recombinant Ca^{2+} receptor.

In another aspect, a cell including a recombinant Ca^{2+} receptor(s) from kidney is employed as a test system to screen antibiotics (e.g. aminoglycoside antibiotics) for activity at calcium receptors which produces renal toxicity in humans.

By "recombinant" is meant to include any Ca^{2+} receptor produced by recombinant DNA techniques such that it is distinct from the naturally occurring Ca^{2+} receptor either in its location, purity or structure. Generally, such a receptor will be present in a cell in an amount different from those normally observed in nature.

By "purified" is meant that the antibody or nucleic acid is distinct from naturally occurring antibody or nucleic acid, being separated from antibody or nucleic acid with which it naturally occurs, e.g., in a vector system, such that it can be used to express recombinant Ca^{2+} receptor. Preferably, the antibody or nucleic acid is provided as a homogeneous preparation by standard techniques.

Such cloned receptors can be expressed in a desired

cell, and isolated and crystallized to allow structure determination. Such a structure will allow design of useful molecules of this invention which can bind to the Ca^{2+} receptor. In addition, equivalent such receptors can be cloned using a first clone as a probe for clones in other cell, cDNA or genomic libraries.

Antibodies to the cloned receptor can be isolated and used as therapeutics in this invention, or as diagnostic tools for determining Ca^{2+} receptor numbers and/or locations and/or functional integrity to diagnose Ca^{2+}-related diseases or conditions. Such antibodies can also be used in vivo by intravenous administration as calcimimetics or calcilytics.

Thus, in general, the invention features calcimimetic or calcilytic molecules able to act as either selective agonists or antagonists respectively at a Ca^{2+} receptor of one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, distal tubule kidney cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cells), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin and glucagon secreting cells, kidney mesangial cell and mammary cell. Such a composition may include any pharmaceutically acceptable carrier known to those in the art to provide a pharmaceutical composition.

The invention also features modulation of the number of Ca^{2+} receptors in a patient by standard techniques, e.g.,

antisense and related technologies (e.g., ribozymes), as a therapeutic for a disease state.

This invention provides methods for identifying molecules which affect the activity of a Ca^{2+} receptor using assays, as defined below, to detect calcimimetics and/or calcilytics. Further, molecules found to be effective to reduce or enhance expression of Ca^{2+} receptor at a transcriptional or translational level by use of the assays or antibodies or other techniques described below can be defined for therapeutic uses.

The foregoing summary has been largely in connection with the preferred embodiment which relates generally to calcium receptors. In a further aspect, the invention relates to a novel superfamily of polypeptide receptor/sensor molecules. The cDNA clone, BoPCaR 1, disclosed herein and duly deposited, represents the first such clone described to encode a receptor/sensor which is a member of this superfamily. For purposes of the present invention, receptor/sensors belonging to this superfamily are called inorganic-ion receptors.

The novel superfamily of inorganic ion receptors includes a variety of such molecules which are related to each other by similarity of amino acid sequence, by structure and/or by function. In total, these attributes also distinguish members of this superfamily of receptors from all receptors/sensors currently known in the art. Members of this superfamily of receptors are primarily distinguished functionally by their surprising ability to detect and respond to changes in the levels of inorganic cations such as calcium, magnesium, potassium, sodium, or hydrogen ions and the like,

and upon sensing such ions, to evoke changes in cellular functions. Such changes may involve changes in second messenger levels as occur for known G-protein coupled receptors, or changes in ionic transmembrane ion flux or the like. Other members of this superfamily also exist which sense inorganic anions such as phosphate or chloride ions. All such receptors are within the scope of the current invention as described herein. Additionally, all isolated naturally occurring or synthetic ligands for receptors/sensors belonging to the superfamily of inorganic-ion receptors are within the scope of the invention. In an additional aspect, the invention features the therapeutic use of any such natural or synthetic ligands for inorganic-ion receptors.

It is a further aspect of the present invention that receptors belonging to the superfamily of inorganic-ion receptors may be activated by stimuli other than ligand binding. For example, some members of this superfamily of receptors are activated by physical forces such as stretch forces acting on membranes of cells expressing such receptors. All such receptors are within the scope of the present invention.

The inorganic-ion receptors can be used in the manner as described above in connection with the preferred receptor. Recombinant receptors expressed in a variety of tissue types including human tissue types can be used to screen (including high through-put screens) for drug discovery in methods known to those skilled in the art. Ionmimetics and ionlytics can be readily identified. The procedures can be adapted for the individual function of the receptor. For example, calcium activated chloride current may be employed

with an inorganic-ion receptor which forms a calcium channel or which nutrilizes intracellular calcium. Ligand-gated ion channels are known in the art. Inorganic-ion receptors that couple to second messenger systems permit assays as described above, including measurement of cyclic AMP and inositol phosphates. Cells that have recombinant receptors coupled to readily detectable agents also may be used such as G-protein coupling of receptors to pigment dispersion in melanophores as described in the literature. Thus, the invention fully enables methods for discovering agents that are inorganic-ion receptor mimetics or lytics, as well as such mimetics and lytics themselves, including those preferably selected from the mimetics and lytics described in detail above. Likewise, the invention embraces diagnosis and treatment of inorganic-ion receptor related diseases or conditions as described above in connection with the preferred embodiment. For example, diseases associated with elevated levels of an inorganic-ion receptor can be diagnosed based upon assays for such elevated levels. Therapeutic molecules can be identified by screening for agents that mimic the activity of the relevant native ion or that modify the effect or potency of the relevant native ion. Tissue specific expression as well as levels of expression can be evaluated and the like.

Thus, in this aspect of the invention, a novel superfamily of isolated inorganic-ion receptors is provided and unique fragments thereof also are provided. Preferably, the isolated receptors are human receptors, and most preferably the isolated receptor is a calcium receptor expressed in tissues or cells selected from the group consisting of: parathyroid, vascular, kidney, epidermis, thyroid, osteoclast, intestine, mammary, trophoblast,

platelet, gastrin secreting, glucagon secreting, cardiac, and brain, and unique fragments thereof. The invention further features polypeptide fragments of the foregoing inorganic-ion receptors which have desirable activity. For example, the fragment may include just a binding site, or a site which binds to mimics, agonists or antagonists. Other useful fragments include those that have only the external portion, membrane-spanning portions, or intracellular portion of the receptor. In addition, these fragments are useful for forming chimeric receptors with fragments of other receptors, as described in greater detail below. The invention also features muteins or analogues and other derivatives of the isolated receptors. Thus, the invention embraces not only naturally occurring proteins, but derivatives thereof.

The invention also features nucleotide sequences encoding the foregoing inorganic-ion receptors and fragments and derivatives thereof. Such nucleotide sequences may be obtained through a variety of procedures, and the disclosure of the present invention allows one of ordinary skill in the art to obtain cDNA or genomic clones encoding such receptors. For example, hybridization probes may be made based upon the nucleotide sequence of BoPCaR 1. When genomic libraries for cDNA libraries from any tissue are screened at low stringency with such probes, hybridizing clones are obtained which encode other members of the superfamily. Additionally, antibodies can readily be prepared which bind to cloned or isolated inorganic-ion receptors. Such antibodies may also be used to isolate other receptors of the invention by expression-cloning techniques known to those of ordinary skill in the art. Targeted gene walking also may be employed to identify and

clone members of the superfamily of inorganic-ion receptors. In addition, the clones may be obtained through expression cloning procedures as described above. These procedures can be adapted for the individual function of the inorganic-ion receptor of interest. For example, calcium activated chloride currents may also be employed for cloning of an inorganic-ion receptor which forms a calcium channel or which mobilizes intracellular calcium. Ligand-gated ion channels, as discussed above, are known in the art. For example, a serotonin gated ion channel has been cloned in this manner. However, the receptors of the present invention are distinct from other known ligand gated ion channels in their amino acid sequences and in that they sense inorganic ions such as calcium, magnesium, hydrogen ions, phosphate ions, and the like.

It will also be appreciated by those skilled in the art that each cloned receptor obtained in this manner provides new such DNA or antibody probes which themselves are used to identify still additional clones encoding inorganic-ion receptors. Furthermore, it will be appreciated that having obtained the sequence of more than one such receptor as outlined above, information is available pertaining to localized sequence conservation which is useful for obtaining still additional clones encoding other members of the superfamily. Such conserved sequences also may be derived from an analysis of the overall structure of BoPCaR 1, as it conventionally includes an extracellular domain, transmembrane domain and intracellular domain.

Thus, isolated nucleic acids encoding inorganic-ion receptors and unique fragments thereof are provided. The preferred receptor is a calcium receptor that is expressed in

the tissues or cells selected from the group described above. Most preferably, the nucleic acid encodes a human inorganic-ion receptor or a unique fragment thereof. The invention further provides isolation of the endogenous regulatory elements controlling the expression of the foregoing inorganic-ion receptors, and agents that are capable of agonizing or antagonizing the activity of these regulatory elements also may be identified.

The invention further provides recombinant cells expressing the nucleic acids of the invention. In such cells, the nucleic acid may be under the control of its genomic regulatory elements, or may be under the control of exogenous regulatory elements including an exogenous promoter. By "exogenous" it is meant a promoter that is not normally coupled in vivo transcriptionally to the coding sequence for the inorganic-ion receptor.

Purified antibodies to the foregoing inorganic-ion receptors are provided as well as antibodies to allosteric sites of such receptors and idiotypes of such receptors.

Inorganic-ion receptor binding agents coupled to a toxin also are provided. Such agents may be antibodies and, as such, are immunotoxins. Such immunotoxins may be used for example in killing a cell expressing an inorganic-ion receptor in vitro or in vivo.

In addition, the invention provides transgenic, nonhuman mammals containing a transgene encoding an inorganic-ion receptor or a unique fragment thereof. Such transgenes may be useful in affecting or altering the expression of an inorganic-ion receptor or in inactivating the expression of an inorganic-ion receptor through, for example,

homologous recombination. Other transgenes may be provided in such mammals, including those encoding antisense or a protein that is capable of altering the expression of a native inorganic-ion receptor gene. Such alteration may be up-regulation, down-regulation or complete inactivation.

Thus, methods are provided involving contacting a cell with an agent that binds to a cellular component for the purpose of affecting the expression of an inorganic-ion receptor. Such agents may bind to promoters, other regulatory agents acting on promoters, agents capable of binding to the receptor and nucleic acids encoding the receptor or a unique fragment thereof. Contact may be by extracellular administration, by providing a transgene encoding the agent or by any other suitable method depending upon the use to which the particular method is directed.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof, and from the claims.

Description of the Preferred Embodiments

The drawings will first briefly be described.

Drawings

Fig. 1 depicts representative molecules useful in the invention.

Fig. 2 is a graphical representation showing increases in $[Ca^{2+}]_i$ induced by extracellular Ca^{2+} in quin-2- or fura-2-loaded bovine parathyroid cells. The initial $[Ca^{2+}]$ was 0.5 mM (using $CaCl_2$) and, at each of the arrows, was increased in 0.5 mM increments.

Fig. 3 is a graphical representation showing mobilization of $[Ca^{2+}]_i$ in bovine parathyroid cells. The

initial $[Ca^{2+}]$ was 0.5 mM and was decreased to < 1 μM by the addition of EGTA as indicated. (a) Extracellular Mg^{2+} (8 mM, final) elicits an increase in $[Ca^{2+}]_i$ in the absence of extracellular Ca^{2+}. (b) Pretreatment with ionomycin (1 μM) blocks the response to Mg^{2+}. (c) Pretreatment with 5 μM molecule 1799 (a mitochondrial uncoupler) is without effect on the response to Mg^{2+}.

Fig. 4 is a graphical representation showing preferential inhibitory effects of a low concentration of Gd^{3+} on steady-state increases in $[Ca^{2+}]_i$ and that a high concentration of Gd^{3+} elicits a transient increase in $[Ca]_i$ in bovine parathyroid cells. Top panel: Control. Initial concentration of extracellular Ca^{2+} was 0.5 mM and was increased by 0.5 mM at each of the arrowheads. Middle panel: Gd^{3+} (5 μM) blocks steady-state but not transient increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. Lower panel: Gd^{3+} (50 μM) elicits a transient increase in $[Ca^{2+}]_i$ and abolishes both transient and sustained responses to extracellular Ca^{2+}. In the middle and lower panels, just enough EGTA was added to chelate preferentially Gd^{3+}: the block of Ca^{2+} influx is removed and $[Ca^{2+}]_i$ rises promptly.

Fig. 5 is a graphical representation showing that the effects of PMA on $[Ca^{2+}]_i$, IP_3 formation, and PTH secretion are overcome by increasing concentrations of extracellular Ca^{2+} in bovine parathyroid cells. For each variable, there is a shift to the right in the concentration-response curve for extracellular Ca^{2+}. Note also that the concentration- response curves vary sigmoidally as $[Ca^{2+}]$ increases linearly.

Fig. 6 is a graphical representation showing that increases in $[Ca^{2+}]_i$ elicited by spermine are progressively depressed by increasing $[Ca^{2+}]$ in bovine parathyroid cells. Spermine (200 μM) was added at the time shown by arrowheads. In this and all subsequent figures, the numbers accompanying the traces are $[Ca^{2+}]_i$ in nM.

Fig. 7 is a graphical representation showing that spermine mobilizes intracellular Ca^{2+} in bovine parathyroid cells. EGTA was added to reduce $[Ca^{2+}]$ to <1 àM before the addition of spermine (200 μM) as indicated (left trace). Pretreatment with ionomycin (1 μM) blocks the response to spermine (right trace).

Figs. 8A and B are graphical representations showing that spermine increases $[Ca^{2+}]_i$ and inhibits PTH secretion in bovine parathyroid cells similarly to extracellular Ca^{2+}. The data points for the spermine dose-concentration response curves are the means of two experiments.

Fig. 9 is a graphical representation showing the contrasting effects of PMA on responses to extracellular Ca^{2+} and on responses to ATPγS in bovine parathyroid cells. Left panel: The concentration- response curve for extracellular Ca^{2+}-induced inhibition of cyclic AMP formation is shifted to the right by PMA (100 nM). Middle panel: PMA does not affect the ability of ATPγS to increases $[Ca^{2+}]_i$. Note also that the concentration-response curve to ATPγS shows classical sigmoidal behavior as a function of the log concentration, in contrast to extracellular divalent cations.

Fig. 10 is a graphical representation showing mobilization of intracellular Ca^{2+} in human parathyroid

cells evoked by extracellular Mg^{2+}. Cells were obtained from an adenoma and bathed in buffer containing 0.5 mM extracellular Ca^{2+}. (a) Transient and sustained increases in $[Ca^{2+}]_i$ elicited by extracellular Mg^{2+} (10 mM, final) shows that sustained increases are not affected by nimodipine (1 μM) but are depressed by La^{3+} (1 μM) and return promptly when La^{3+} is selectively chelated by a low concentration of EGTA. (b) La^{3+} (1 μM) blocks the sustained but not the transient increase in $[Ca^{2+}]_i$ elicited by extracellular Mg^{2+}. (c) Cytosolic Ca^{2+} transients elicited by extracellular Mg^{2+} persist in the absence of extracellular Ca^{2+}.

Fig. 11 is a graphical representation showing mobilization of intracellular Ca^{2+} evoked by neomycin or protamine in bovine parathyroid cells. In all traces, the initial $[Ca^{2+}]$ and $[Mg^{2+}]$ was 0.5 and 1 mM, respectively. In trace (a) and (b), the Ca^{2+} and Mg^{2+} concentrations were increased to 2 and 8 mM, from 0.5 and 1 mM respectively. In the other traces, (c) through (i), neomycin B (30 μM) or protamine (1 μg/ml) were added as indicated. La^{3+} (1 μM) EGTA (1 mM), or ionomycin (100 nM) were added as indicated. Each trace is representative of the pattern seen in 5 or more trials using at least 3 different cell preparations. Bar = 1 min.

Fig. 12 is a graphical representation showing that neomycin B blocks transient but does not block steady-state increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+} in bovine parathyroid cells. Left control: $[Ca^{2+}]$ was initially 0.5 mM and was increased in 0.5 mM increments at each of the open arrowheads before the addition of neomycin B

(30 μM). Right: Neomycin B (30 μM) was added before $[Ca^{2+}]$. Bar = 1 min.

Fig. 13 is a graphical representation showing that neomycin B or protamine inhibit PTH secretion at concentrations which evoked increases in $[Ca^{2+}]_i$ in bovine parathyroid cells. Cells were incubated with the indicated concentrations of organic polycation for 30 min. in the presence of 0.5 mM extracellular Ca^{2+}. Open symbols: control responses for PTH secretion in the presence of 0.5 (circles) or 2 mM (diamonds) extracellular Ca^{2+}. Values for $[Ca^{2+}]_i$ are diamond symbols. Bovine cells were used in the experiments with protamine and human (adenoma) parathyroid cells were used in the experiments with neomycin B. Each point is the mean ± SEM of 3 experiments.

Fig. 14 is a graphical representation showing the preferential inhibitory effects of PMA on cytosolic Ca^{2+} transients elicited by spermine in bovine parathyroid cells. Initial $[Ca^{2+}]$ was 0.5 mM; spermine (200 μM) or ATP (50 μM) were added as indicated. Bar = 1 min.

Fig. 15 is a graphical representation showing that PMA shifts to the right the concentration-response curves for extracellular Ca^{2+}- and neomycin B-induced increases in $[Ca^{2+}]_i$ in bovine parathyroid cells. Cells were pretreated with PMA for 1 min. before increasing $[Ca^{2+}]$ or before adding neomycin B as indicated. Each point is the mean ± SEM of 3 to 5 experiments.

Fig. 16 is a graphical representation showing that PMA shifts to the right the concentration-response curves for extracellular Ca^{2+}- and spermine-induced inhibition of PTH secretion in bovine parathyroid cells. Cells were incubated

with the indicated [Ca^{2+}] and spermine for 30 min. in the presence (closed circles) or absence (open circles) of 100 nM PMA. Each point is the mean ± SEM of 3 experiments.

Fig. 17 is a graphical representation showing that protamine increases the formation of inositol phosphates in bovine parathyroid cells. Parathyroid cells were incubated overnight in culture media containing 4 μCi/ml ^{3}H-myo-inositol, washed, and incubated with the indicated concentration of protamine at 37°. After 30 sec. the reaction was terminated by the addition of $CHCl_3$:MeOH:HCl and IP_1 (circles) and IP_3 (triangles) separated by anion exchange chromatography. Each point is the mean of 2 experiments, each performed in triplicate.

Fig. 18 is a graphical representation showing that PMA depresses the formation of IP_1 evoked by extracellular Ca^{2+} or spermine in bovine parathyroid cells. ^{3}H-myo-inositol- labeled cells were exposed to the indicated [Ca^{2+}] or spermine for 30 sec. before terminating the reaction and determining IP_1 by anion exchange chromatography. Hatched columns: Cells were pretreated with PMA (100 nM) for 5 min. before increasing [Ca^{2+}] or adding spermine. Each value is the mean of 2 experiments, each performed in triplicate.

Fig. 19 is a graphical representation showing transient and sustained increases in $[Ca^{2+}]_i$ elicited by neomycin B in human (adenoma) parathyroid cells. [Ca^{2+}] was 0.5 mM. (a) The sustained increase in $[Ca^{2+}]_i$ elicited by neomycin B (10 μM) was depressed by La^{3+}. (b) The transient increase in $[Ca^{2+}]_i$ evoked by neomycin B was unaffected by La^{3+}. (c) Transient increases in $[Ca^{2+}]_i$

persisted in the absence of extracellular Ca^{2+}.

Fig. 20 is a graphical representation showing that neomycin B evokes oscillating increases the Cl^- current in Xenopus oocytes expressing the Ca^{2+} receptor. Upper trace from an oocyte three days after injection with human (hyperplastic) parathyroid cell poly(A)$^+$-mRNA. Lower trace from an oocyte injected with water. Neomycin B failed to elicit a response in five water-injected oocytes and carbachol elicited a response in one, which is shown. In both traces, the holding potential was -76 mV.

Fig. 21 is a graphical representation showing that neomycin B fails to affect basal or evoked increases in C-cells. Control, left trace: Fura-2- loaded rMTC 6-23 cells were initially bathed in buffer containing 1 mM Ca^{2+} before increasing [Ca^{2+}] to 3 mM. Right trace: pretreatment with 5 mM neomycin B.

Fig. 22 is a graphical representation showing that extracellular Ca^{2+} evokes increases in $[Ca^{2+}]_i$ in rat osteoclasts. Microfluorimetric recording in a single rat osteoclast loaded with indo-1 and superfused for the indicated times (bars) with buffer containing the indicated [Ca^{2+}]. Normal buffer, superfused between the bars, contained 1 mM Ca^{2+}.

Fig. 23 is a graphical representation showing that spermine or neomycin B fail to evoke increases in $[Ca^{2+}]_i$ in rat osteoclasts. An indo-1-loaded osteoclast was superfused with the indicated concentration of spermine or neomycin B (open bars) alone or together with 20 mM Ca^{2+} (solid bars).

Fig. 24 is a graphical representation showing the differential effects of argiotoxin (shown as argiopine in the figure, structures also shown in Fig. 1) 659 and argiotoxin 636 on $[Ca^{2+}]_i$ in bovine parathyroid cells. The initial $[Ca^{2+}]$ was 0.5 mM and was increased to 1.5 mM where indicated (right trace)N Where indicated, argiotoxin 659 (300 μM) or argiotoxin 636 (400 μM) was added.

Fig. 25 is a graphical representation showing that extracellular Mg^{2+} or Gd^{3+} evoke oscillatory increases in Cl^- current in Xenopus oocytes injected with bovine parathyroid cell poly(A)$^+$-mRNA. In trace (a), the concentration of extracellular Ca^{2+} was < 1 μM and in trace (b), 0.7 mM. Trace (c) shows that extracellular Mg^{2+} fails to elicit a response in an oocyte injected only with the mRNA for the substance K receptor, although superfusion with substance K evokes a response. Holding potential was -70 to -80 mV.

Fig. 26 is a graphical representation showing that extracellular Ca^{2+} elicits oscillatory increases in Cl^- current in Xenopus oocytes injected with human (hyperplastic) parathyroid tissue poly(A)$^+$-mRNA. The oocyte was tested for responsivity to extracellular Ca^{2+} three days after injection of 50 ng poly(A)$^+$-mRNA. Holding potential was -80 mV.

Fig. 27 is a graphical representation showing the mobilization of intracellular Ca^{2+} in bovine parathyroid cells elicited by budmunchiamine. Budmunchiamine (300 μM, structure also shown) was added where indicated.

Fig. 28 is a graphical representation showing that the ability to mobilize intracellular Ca^{2+} in parathyroid

cells is stereospecific. Bovine parathyroid cells loaded with fura-2 were initially suspended in buffer containing 0.5 mM extracellular Ca^{2+} before the addition of the indicated concentration of each molecule.

Fig. 29 is a graphical representation showing effects of La^{3+} on $[Ca^{2+}]_i$ in osteoclasts. A representative trace from a single rat osteoclast loaded with indo-1 is shown. At low concentrations, La^{3+} partially blocks increases in $[Ca^{2+}]_i$, elicited by extracellular Ca^{2+}.

Figs. 30A and B are graphical representations showing the mobilization of intracellular Ca^{2+} elicited by extracellular Mn^{2+} in rat osteoclasts. Extracellular Mn^{2+} evokes concentration-dependent increases in $[Ca^{2+}]_i$ (Fig. 30A) that persist in the absence of extracellular Ca^{2+} (Fig. 30B).

Figs. 31A and 31B are graphical representations showing mobilization of $[Ca^{2+}]_i$ in rat osteoclasts elicited by a molecule termed NPS 449 (see Fig. 38). Isolated rat osteoclasts loaded with indo-1 were superfused with the indicated concentrations of NPS 449 in the presence (Fig. 31A) or absence (Fig. 31B) of 1 mM extracellular $CaCl_2$.

Fig. 32 is a graphical representation showing the mobilization of intracellular Ca^{2+} in C-cells evoked by NPS 019 (see Fig. 1). rMTC 6-23 cells were loaded with fura-2 and bathed in buffer containing 0.5 mM $[Ca^{2+}]$. Where indicated, NPS 019 was added to a final concentration of 10 μM. Representative traces show that the transient increase in $[Ca^{2+}]_i$ elicited by NPS 019 is refractory to inhibition by La^{3+} (middle trace) and persists in the absence of

extracellular Ca^{2+} (right trace).

Fig. 33 is a graphical representation showing that NPS 456 (Fig. 36) evokes oscillatory increases in Cl^- current in Xenopus oocytes which have been injected with bovine parathyroid cell poly(A)$^+$-mRNA.

Fig. 34 is a graphical representation showing that extracellular Ca^{2+} evokes oscillatory increases in Cl^- current in Xenopus oocytes which have been injected with human osteoclast mRNA. The oocyte was tested for responsivity to extracellular Ca^{2+} three days after injection of 50 ng of total poly(A)$^+$ mRNA.

Fig. 35 is a graphical representation showing that the parathyroid cell Ca^{2+} receptor is encoded by mRNA in a size range of 2.5-3.5 kb. Bovine parathyroid cell poly(A)$^+$ -mRNA was size fractionated on glycerol gradients and pooled into ten fractions. Each fraction was injected (50 ng/fraction) separately into Xenopus oocytes. After three days, the oocytes were examined for their ability to respond to neomycin B (10mM) with oscillatory increases in the Cl^- current.

Fig. 36 shows the chemical structures of molecules derived from diphenylpropyl-α-phenethylamine illustrating a family of molecules which were prepared and screened to find the useful molecules of the invention.

Fig. 37 is a graphical representation showing that NPS 021 is a calcilytic compound that blocks the effects of extracellular Ca^{2+} on $[Ca^{2+}]_i$ in bovine parathyroid cells. Cells were initially bathed in buffer containing 0.5 mM $CaCl_2$ and, where indicated, the $[Ca^{2+}]$ was increased to a final of 2mM (left trace). The addition of NPS 021

(200 μM) caused no change in $[Ca^{2+}]_i$ but inhibited the increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+} (right trace).

Fig. 38 is a graph showing in vivo Ca^{2+} response to NPS R,S-467.

Fig. 39 is a graph showing in vivo PTH response to NPS R,S-467.

Fig. 40 is a graph showing in vivo serum Ca^{2+} response to 25 mg/kg NPS R,S-467.

Fig. 41 is a graph showing the in vivo response of $[Ca^{2+}]_i$ in bovine parathyroid cells to different enantiomers of NPS 467.

Fig. 42 is a graph showing the in vivo response of serum Ca^{2+} in rats to different enantiomers of NPS 467.

Fig. 43a depicts a reaction scheme for the preparation of fendiline or fendiline analogs or derivatives depicted in Figure 36. Fig. 43b depicts a reaction scheme for the synthesis of NPS 467.

Fig. 44 depicts a dose-response curve showing that NPS 467 lowers serum ionized calcium in rats when administered orally.

Fig. 45 is a restriction map of the plasmid containing BoPCaR 1, deposited with the ATCC under accession number ATCC 75416.

Fig. 46 is a restriction map of BoPCaR 1.

Fig. 47 is a nucleotide sequence corresponding (about 90% accurate) to the 2.2 Kbfragment of BoPCaR 1 sequenced.

Calcimimetic and Calcilytic Molecules

Calcimimetic and calcilytic molecules useful in the invention are generally described above. These molecules

can be readily identified using screening procedures to define molecules which mimic or antagonize the activity of Ca^{2+} at Ca^{2+} receptors. Examples of such procedures are provided below. These examples are not limiting in the invention but merely illustrate methods which are readily used or adapted by those skilled in the art.

Generally, calcimimetic and calcilytic molecules are identified by screening molecules which are modelled after those described below (called lead molecules). As can be seen below there are several specific calcimimetics and calcilytics useful at various Ca^{2+} receptors. Derivative molecules are readily designed by standard procedures and tested in one of many protocols known to those skilled in the art. Many molecules may be screened easily to identify the most useful in this invention.

Organic cationic molecules which mimic or antagonize the actions of Ca^{2+} in other systems contain the requisite structure for activity on a Ca^{2+} receptor. Rational design of other useful molecules involves the study of a molecule known to be calcimimetic or calcilytic and then modifying the structure of the known molecule. For example, polyamines are potentially calcimimetic since spermine mimics the action of Ca^{2+} in several <u>in vitro</u> systems. Results show that spermine does indeed cause changes in $[Ca^{2+}]_i$ and PTH secretion reminiscent of those elicited by extracellular di- and trivalent cations (see below). Conversely, Ga^{3+} antagonizes the effects of Gd^{3+} on the bovine parathyroid calcium receptor(s). The experiments outlined below are therefore aimed at demonstrating that this phenomenology, obtained with spermine, involves the

same mechanisms used by extracellular Ca^{2+}. To do this, the effects of spermine on a variety of physiological and biochemical parameters which characterize activation of the Ca^{2+} receptor were assessed. Those molecules having similar effects are useful in this invention and can be discovered by selecting or making molecules having a structure similar to spermine. Once another useful molecule is discovered this selection process can be readily repeated.

For clarity, below is provided a specific series of screening protocols to identify such useful molecules which are active at a parathyroid cell Ca^{2+} receptor, or which act as agonists or antagonists of the cellular response to changes in $[Ca^{2+}]$. Equivalent assays can be used for molecules active at other Ca^{2+} receptors or other inorganic-ion receptors, or which otherwise mimic or antagonize cellular functions regulated by $[Ca^{2+}]$ or other ions. These assays exemplify the procedures which are useful to find molecules, including calcimimetic molecules, of this invention. Equivalent procedures can be used to find lytic molecules, including calcilytic molecules, by screening for those molecules most antagonistic to the actions of the ion, including extracellular Ca^{2+}. In vitro assays can be used to characterize the selectivity, saturability, and reversibility of these mimetics and lytics by standard techniques.

Screening Procedure

Generally, bovine parathyroid cells loaded with fura-2 are initially suspended in buffer containing 0.5 mM $CaCl_2$. The test substance is added to the cuvette in a

small volume (5-15 µl) and any change in the fluorescence signal noted. Cumulative increases in the concentration of the test substance are made in the cuvette until some predetermined concentration is achieved or changes in fluorescence noted. If no changes in fluorescence are noted, the molecule is considered inactive and no further testing is performed. In initial studies, e.g., with polyamine-type molecules, molecules were tested at concentrations as high as 5 or 10 mM. As more potent molecules are now known (see below), the ceiling concentration is lowered. For example, newer molecules are tested at concentrations up to 500 µM or less. If no changes in fluorescence are noted at this concentration, the molecule can be considered inactive.

Molecules causing increases in $[Ca^{2+}]_i$ are subjected to additional testing. The two essential characteristics of the molecule important for its consideration as a calcimimetic molecule are the mobilization of intracellular Ca^{2+} and sensitivity to PKC activators. Molecules causing the mobilization of intracellular Ca^{2+} in a PMA-sensitive manner have invariably been found to be calcimimetic molecules and to inhibit PTH secretion. Additional testing can, if needed, be performed to solidify this belief. Typically, all the various tests for calcimimetic or calcilytic activity (see above) are not performed. Rather, if a molecule causes the mobilization of intracellular Ca^{2+} in a PMA-sensitive manner, it is advanced to screening on human parathyroid cells. For example, measurements of $[Ca^{2+}]_i$ are performed to determine the EC_{50}, and to measure the

ability of the molecule to inhibit PTH secretion in human parathyroid cells which have been obtained from patients undergoing surgery for primary or secondary hyperparathyroidism. The lower the EC_{50} or IC_{50} the more potent the molecule as a calcimimetic or calcilytic.

Measuring $[Ca^{2+}]_i$ with fura-2 provides a very rapid means of screening new organic molecules for activity. In a single afternoon, 10-15 molecules can be examined and their ability to mobilize intracellular Ca^{2+} (or not) assessed. The sensitivity of any observed increase in $[Ca^{2+}]_i$ to depression by PMA can also be assessed. Moreover, a single cell preparation can provide data on $[Ca^{2+}]_i$, cyclic AMP levels, IP_3 and PTH secretion. A typical procedure is to load cells with fura-2 and then divide the cell suspension in two; most of the cells are used for measurement of $[Ca^{2+}]_i$ and the remainder are incubated with molecules to assess their effects on cyclic AMP and PTH secretion. Because of the sensitivity of the radioimmunoassays for cyclic AMP and PTH, both variables can be determined in a single incubation tube containing 0.3 ml cell suspension (about 500,000 cells). Measurements of inositol phosphates are a time-consuming aspect of the screening. However, ion-exchange columns eluted with chloride (rather than formate) provide a very rapid means of screening for IP_3 formation since rotary evaporation (which takes around 30 hrs) is not required. This method allows processing of nearly 100 samples in a single afternoon. Those molecules that prove interesting, as assessed by measurements of $[Ca^{2+}]_i$, cyclic AMP, IP_3, and PTH are then subjected to a more rigorous analysis by examining formation of various inositol

phosphates and assessing their isomeric form by HPLC.

Interesting molecules detected in these protocols are then assessed for specificity, e.g., by examining their effects on $[Ca^{2+}]_i$ in calcitonin-secreting C-cells using, e.g., the rat MTC 6-23 cell line.

The following is illustrative of methods useful in these screening procedures. Examples of typical results for various test calcimimetic or calcilytic molecules are provided in Figs. 2-34.

Parathyroid Cell Preparation

Parathyroid glands were obtained from freshly slaughtered calves (12-15 weeks old) at a local abattoir and transported to the laboratory in ice-cold parathyroid cell buffer (PCB) which contains (mM): NaCl, 126; KCl, 4; $MgCl_2$, 1; Na-HEPES, 20; pH 7.4; glucose, 5.6, and variable amounts of $CaCl_2$, e.g., 1.25 mM. Human parathyroid glands, obtained from patients undergoing surgical removal of parathyroid tissue for primary or uremic hyperparathyroidism (HPT), were treated similarly to bovine tissue. Glands were trimmed of excess fat and connective tissue and then minced with a fine scissors into approximate cubes of 2-3 mm. Dissociated parathyroid cells were prepared by collagenase digestion. Dissociated cells were then purified by centrifugation in Percoll buffer. The resultant parathyroid cell preparation was essentially devoid of red blood cells, adipocytes, and capillary tissue as assessed by phase contrast microscopy and Sudan black B staining. Dissociated and purified parathyroid cells were present as small clusters containing 5 to 20 cells. Cellular viability, as indexed by exclusion of trypan blue

or ethidium bromide, was routinely 95%.

Although cells can be used for experimental purposes at this point, physiological responses (suppressibility of PTH secretion and resting levels of $[Ca^{2+}]_i$) are better after culturing the cells overnight. Primary culture also has the advantage that cells can be labeled with isotopes to near isotopic equilibrium, as is necessary for studies involving measurements of inositol phosphate metabolism (see below). After purification on Percoll gradients, cells were washed several times in a 1:1 mixture of Ham's F12-Dulbecco's modified Eagle's medium (GIBCO) supplemented with 50 µg/ml streptomycin, 100 U/ml penicillin, 5 µg/ml gentamicin and ITS^+. ITS^+ is a premixed solution containing insulin, transferrin, selenium, and bovine serum albumin (BSA)-linolenic acid (Collaborative Research, Bedford, MA). The cells were then transferred to plastic flasks (75 or 150 cm^2; Falcon) and incubated at 37°C in a humid atmosphere of 5% CO_2. No serum is added to these overnight cultures, since its presence allows the cells to attach to the plastic, undergo proliferation, and dedifferentiate. Cells cultured under the above conditions were readily removed from the flasks by decanting, and show the same viability as freshly prepared cells.

Measurement of Cytosolic Ca^{2+}

Purified parathyroid cells were resuspended in 1.25 mM $CaCl_2$-2% BSA-PCB containing 1 µM fura-2-acetoxymethylester and incubated at 37°C for 20 min. The cells were then pelleted, resuspended in the same buffer lacking the ester, and incubated a further 15 min at 37°C.

The cells were subsequently washed twice with PCB containing 0.5 mM $CaCl_2$ and 0.5% BSA and maintained at room temperature (about 20°C). Immediately before use, the cells were diluted five-fold with prewarmed 0.5 mM $CaCl_2$-PCB to obtain a final BSA concentration of 0.1%. The concentration of cells in the cuvette used for fluorescence recording was 1-2 x 10^6/ml.

The fluorescence of indicator-loaded cells was measured at 37°C in a spectrofluorimeter (Biomedical Instrumentation Group, University of Pennsylvania, Philadelphia, PA) equipped with a thermostated cuvette holder-and magnetic stirrer using excitation and emission wavelengths of 340 and 510 nm, respectively. This fluorescence indicates the level of cytosolic Ca^{2+}. Fluorescence signals were calibrated using digitonin (50 µg/ml, final) to obtain maximum fluorescence (F_{max}), and EGTA (10 mM, pH 8.3, final) to obtain minimal fluorescence (F_{min}), and a dissociation constant of 224 nM. Leakage of dye is dependent on temperature and most occurs within the first 2 min after warming the cells in the cuvette; dye leakage increases only very slowly thereafter. To correct the calibration for dye leakage, cells were placed in the cuvette and stirred at 37°C for 2-3 min. The cell suspension was then removed, the cells pelleted, and the supernatant returned to a clean cuvette. The supernatant was then treated with digitonin and EGTA as above to obtain as estimate of dye leakage, which is typically 10-15% of the total Ca^{2+}-dependent fluorescent signal. This estimate was subtracted from the apparent F_{min}.

Measurement of PTH Secretion

In most experiments, cells loaded with fura-2 were used in studies of PTH secretion. Loading parathyroid cells with fura-2 does not change their secretory response to extracellular Ca^{2+}. Cells were suspended in PCB containing 0.5 mM $CaCl_2$ and 0.1% BSA. Incubations were performed in plastic tubes (Falcon 2058) containing 0.3 ml of the cell suspension with or without small volumes of $CaCl_2$ and/or organic polycations. After incubation at 37°C for various times (typically 30 min), the tubes were placed on ice and the cells pelleted at 2°C. Samples of the supernatant were brought to pH 4.5 with acetic acid and stored at -70°C. This protocol was used for both bovine and human parathyroid cells.

For bovine cells, the amount of PTH in sample supernatants was determined by a homologous radioimmunoassay using GW-1 antibody or its equivalent at a final dilution of 1/45,000. ^{125}I-PTH (65-84; INCSTAR, Stillwater, MN) was used as tracer and fractions separated by dextran-activated charcoal. Counting of samples and data reduction were performed on a Packard Cobra 5005 gamma counter.

For human cells, a commercially available radioimmunoassay kit (INS-PTH; Nichols Institute, Los Angeles, CA) which recognizes intact and N-terminal human PTH was used because GW-1 antibody recognizes human PTH poorly.

Measurement of cyclic AMP

Cells were incubated as above for PTH secretion studies and at the end of incubation, a 0.15 ml sample was taken and transferred to 0.85 ml hot (70°C) water and heated

at this temperature for 5-10 min. The tubes were subsequently frozen and thawed several times and the cellular debris sedimented by centrifugation. Portions of the supernatant were acetylated and cyclic AMP concentrations determined by radioimmunoassay.

Measurement of Inositol Phosphate Formation

Membrane phospholipids were labeled by incubating parathyroid cells with 4 μCi/ml ^{3}H-myo-inositol for 20-24 hrs. Cells were then washed and resuspended in PCB containing 0.5 mM $CaCl_2$ and 0.1% BSA. Incubations were performed in microfuge tubes in the absence or presence of various concentrations of organic polycation for different times. Reactions were terminated by the addition of 1 ml chloroform/methanol/12 N HCl (200:100:1; v/v/v). Phytic acid hydrolysate (200 μl; 25 μg phosphate/tube) water was then added. The tubes were centrifuged and 600 μl of the aqueous phase was diluted into 10 ml water.

Inositol phosphates were separated by ion-exchange chromatography using AG1-X8 in either the chloride- or formate-form. When only IP_3 levels were to be determined, the chloride-form was used, whereas the formate form was used to resolve the major inositol phosphates (IP_3, IP_2, and IP_1). For determination of just IP_3, the diluted sample was applied to the chloride-form column and the column washed with 10 ml 30 mM HCl followed by 6 ml 90 mM HCl and the IP_3 eluted with 3 ml 500 mM HCl. The last eluate was diluted and counted. For determination of all major inositol phosphates, the diluted sample was applied to the formate-form column and IP_1, IP_2, and IP_3 eluted sequentially by increasing concentrations of

formate buffer. The eluted samples from the formate columns were rotary evaporated, the residues brought up in cocktail, and counted.

The isomeric forms of IP_3 were evaluated by HPLC. The reactions were terminated by the addition of 1 ml 0.45 M perchloric acid and stored on ice for 10 min. Following centrifugation, the supernatant was adjusted to pH 7-8 with $NaHCO_3$. The extract was then applied to a Partisil SAX anion-exchange column and eluted with a linear gradient of ammonium formate. The various fractions were then desalted with Dowex followed by rotary evaporation prior to liquid scintillation counting in a Packard Tri-carb 1500 LSC.

For all inositol phosphate separation methods, appropriate controls using authentic standards were used to determine if organic polycations interfered with the separation. If so, the samples were treated with cation-exchange resin to remove the offending molecule prior to separation of inositol phosphates.

Measurement of Cytosolic Ca^{2+} in C-cells

Neoplastic C-cells derived from a rat medullary thyroid carcinoma (rMTC 6-23) obtained from American Type Culture Collection (ATCC No. 1607) were cultured as monolayers in Dulbecco's Modified Eagle's medium (DMEM) plus 15% horse serum in the absence of antibiotics. For measurements of $[Ca^{2+}]_i$, the cells were harvested with 0.02% EDTA/0.05% trypsin, washed twice with PCB containing 1.25 mM $CaCl_2$ and 0.5% BSA, and loaded with fura-2 as described above for parathyroid cells. Measurements of $[Ca^{2+}]_i$ were performed as described above with

appropriate corrections for dye leakage.

Measurement of $[Ca^{2+}]_i$ in Rat Osteoclasts

Osteoclasts were obtained from 1-2 day old Sprague-Dawley rats using aseptic conditions. The rat pups were sacrificed by decapitation, the hind legs removed, and the femora rapidly freed of soft tissue and placed in prewarmed F-12/DMEM media (DMEM containing 10% fetal calf serum and antibiotics (penicillin-streptomycin-gentamicin; 100 U/ml-100 µg/ml-100 µg/ml)). The bones from two pups were cut lengthwise and placed in 1 ml culture medium. Bone cells were obtained by gentle trituration of the bone fragments with a plastic pipet and diluted with culture medium. The bone fragments were allowed to settle and equal portions (about 1 ml) of the medium transferred to a 6-well culture plate containing 25 mm glass coverslips. The cells were allowed to settle for 1 hr at 37°C in a humidified 5% CO_2-air atmosphere. The coverslips were then washed 3 times with fresh media to remove nonadherent cells. Measurements of $[Ca^{2+}]_i$ in osteoclasts were performed within 6-8 hrs of removing nonadherent cells.

Cells attached to the coverslip were loaded with indo-1 by incubation with 5 µM indo-1 acetoxymethylester / 0.01% Pluronic F28 for 30 min at 37°C in F-12/DMEM lacking serum and containing instead 0.5% BSA. The coverslips were subsequently washed and incubated an additional 15 min at 37°C in F-12/DMEM lacking ester before being transferred to a superfusion chamber mounted on the stage of a Nikon Diaphot inverted microscope equipped for microfluorimetry. Osteoclasts were easily identified by their large size and presence of multiple nuclei. The cells were superfused with

buffer (typically PCB containing 0.1% BSA and 1 mM Ca^{2+}) at 1 ml/min with or without test substance. The fluorescence emitted by excitation at 340 nm was directed through the video port of the microscope onto a 440 nm dichroic mirror and fluorescence intensity at 495 and 405 nm collected by photomultiplier tubes. The outputs from the photomultiplier tubes were amplified, digitized, and stored in an 80386 PC. Ratios of fluorescence intensity were used to estimate $[Ca^{2+}]_i$.

Oocyte Expression

In additional studies, Xenopus oocytes injected with mRNA from bovine or human parathyroid cells were used in screening protocols, and Cl^- current measured as an indirect means of monitoring increases in $[Ca^{2+}]_i$. The following is an example to test the effect of neomycin.

Oocytes were injected with poly(A)$^+$-enriched mRNA from human parathyroid tissue (hyperplastic glands from a case of secondary HPT). After 3 days, the oocytes were tested for their response to neomycin. Neomycin B evoked oscillatory increases in the Cl^- current which ceased upon superfusion with drug-free saline (see Fig. 20). Responses to neomycin B were observed at concentrations between 100 μM and 10 mM. To ensure that the response evoked by neomycin B was contingent upon injection of parathyroid mRNA, the effect of neomycin B on currents in water-injected oocytes was determined. In each of five oocytes examined, neomycin B (10 mM) failed to cause any change in the current. About 40% of oocytes are known to respond to carbachol, an effect mediated by an endogenous muscarinic receptor. In five oocytes examined, one showed inward

currents in response to carbachol, and this is shown in the lower trace of Fig. 20. Thus, in cells expressing a muscarinic receptor coupled to increases in $[Ca^{2+}]_i$ and Cl^- current, neomycin B fails to evoke a response. This shows that the response to neomycin B depends on expression of a specific protein encoded by parathyroid cell mRNA. It suggests quite strongly that in intact cells, neomycin B acts directly on the Ca^{2+} receptor to alter parathyroid cell function.

Drug Design From Lead Molecules

Certain organic molecules mimic or antagonize the action of extracellular Ca^{2+} by acting at the Ca^{2+} receptor as shown herein. The molecules tested, however, are not necessarily suitable as drug candidates, but they serve to demonstrate that the hypothesis underlying Ca^{2+} receptor-based therapies is correct. These molecules can be used to determine the structural features that enable them to act on the Ca^{2+} receptor, and thus to select molecules useful in this invention.

An example of one such analytical procedure follows: This example is detailed in the examples below, but is used here to demonstrate the rationale that can be used to design useful molecules of this invention from lead molecules discussed herein. Those in the art will recognize the analytic steps defined in the example and that analogous analysis can be conducted on other lead molecules until the most desired calcimimetic or calcilytic is defined.

Other examples are also provided below. Together the data presented demonstrate that useful lead molecules will have aromatic groups which are preferably substituted

at one or more positions, and may have branched or linear substituted or unsubstituted alkyl groups as desired. In addition, it is important to choose molecules of correct stereospecificity to ensure higher affinity for the desired Ca^{2+} receptor. These data thus point those in the art to appropriate lead molecules which can be derivatised to find optimum desired molecules of this invention, much as described below.

Although structurally diverse, molecules that are tested may have common features that can be studied. In this example, the correlation between net positive charge and potency in mobilizing intracellular Ca^{2+} was tested. Protamine (+21; EC_{50} = 40 nM) was more effective than neomycin B (+6; EC_{50} = 20 μM in human parathyroid cells and 40 μM in bovine parathyroid cells) which was more effective than spermine (+4; EC_{50} = 150 μM) in causing the mobilization of $[Ca^{2+}]_i$ in parathyroid cells. These results raise the question of whether positive charge alone determines potency, or if there are other structural features that contribute to activity on the Ca^{2+} receptor. This is important to determine at the outset because it profoundly impacts on the view that the Ca^{2+} receptor can be targeted with effective and specific therapeutic molecules. Thus, a variety of other organic polycations related to neomycin B and spermine can be studied to determine the relationship between the net positive charge of a molecule and its potency to mobilize intracellular Ca^{2+}.

The first series of molecules studied were the aminoglycosides. The molecules were examined on bovine

parathyroid cells and their EC_{50}'s for the mobilization of intracellular Ca^{2+} determined. For the aminoglycosides, the rank order of potency for eliciting cytosolic Ca^{2+} transients was neomycin B (EC_{50} = 20 or 40 μM) > gentamicin (150 μM) > bekanamycin (200 μM) > streptomycin (600 μM). Kanamycin and lincomycin were without effect when tested at a concentration of 500 μM. The net positive charge on these aminoglycosides at pH 7.3 is neomycin B (+6) > gentamicin (+5) = bekanamycin (+5) > kanamycin (average +4.5) > streptomycin (+3) > lincomycin (+1). Within the aminoglycoside series, then, there is some correlation between net positive charge but it is not absolute, and kanamycin, which would be predicted to be more potent than streptomycin, is without activity.

Testing of various polyamines revealed additional and more marked discrepancies between net positive charge and potency. Three structural classes of polyamines were examined: (1) straight chain, (2) branched chain, and (3) cyclic. The structures of the polyamines tested are provided in Fig. 1. Amongst the straight chain polyamines, spermine (+4; EC_{50} = 150 μM) was more potent than pentaethylenehexamine (+6; EC_{50} = 500 μM) and tetraethylenepentamine (+5; EC_{50} = 2.5 mM) even though the latter molecules have a greater net positive charge.

We synthesized some branched chain polyamines that have different numbers of secondary and primary amino groups and thus vary in net positive charge. Two of these molecules, NPS 381 and NPS 382, were examined for effects on $[Ca^{2+}]_i$ in bovine parathyroid cells. NPS 382 (+8; EC_{50} = 50 μM) was about twice as potent as NPS 381 (+10;

EC_{50} = 100 μM) even though it contains two fewer positive charges.

A similar discrepancy between positive charge and potency was noted in experiments with cyclic polyamines. For example, hexacyclen (+6; EC_{50} = 20 μM) was more potent than NPS 383 (+8; EC_{50} = 150 μM). The results obtained with these polyamines show that positive charge is not the sole factor contributing to potency.

Additional studies provided insights into the structural features of molecules that impart activity on the parathyroid cell Ca^{2+} receptor. One of the structurally important features is the intramolecular distance between the nitrogens (which carry the positive charge). Thus, spermine is 50-fold more potent than triethylenetetramine (EC_{50} = 8 mM) in evoking increases in $[Ca^{2+}]_i$ in bovine parathyroid cells yet both molecules carry a net positive charge of +4. The only difference in structure between these two polyamines is the number of methylenes separating the nitrogens: in spermine it is 3-4-3 whereas in triethylenetetramine it is 2-2-2. This seemingly minor change in the spacing between nitrogens has profound implications for potency and suggests that the conformational relationships of nitrogens within the molecule are critical. Supporting this are results obtained with hexacyclen and pentaethylenehexamine. The former molecule is simply the cyclic analog of the latter and contains the same number of methylenes between all nitrogens, yet the presence of the ring structure increases potency 25-fold. These results indicate that positive charge per se is not the critical factor determining the

activity of an organic molecule on the Ca^{2+} receptor.

Another series of experiments reveals the importance of aromatic groups in determining activity on the Ca^{2+} receptor. The results were obtained with two arylalkylamines isolated from the venom of the spider Argiope lobata. These molecules, argiotoxin 636 and argiotoxin 659, have identical polycationic portions linked to different aromatic groups (Fig. 24). Argiotoxin 659 evoked transient increases in $[Ca^{2+}]_i$ in bovine parathyroid cells when tested at concentrations of 100 to 300 μM. In contrast, argiotoxin 636 was without effect when tested at similar concentrations (Fig. 24). The only difference in structure between these two arylalkylamines is in the aromatic portion of the molecules: argiotoxin 659 contains a 4-hydroxyindole moiety whereas argiotoxin 636 contains a 2,4-dihydroxyphenyl group. The net positive charge on these two arylalkylamines is the same (+4), so their different potencies must result from the different aromatic groups. This shows that net positive charge alone does not determine potency. The real importance of these findings, however, is the discovery that aromatic groups contribute significantly to the ability of molecules to activate the Ca^{2+} receptor.

Agatoxin 489 (NPS 017) and Agatoxin 505 (NPS 015) both cause the mobilization of intracellular Ca^{2+} in parathyroid cells with EC_{50}'s of 6 and 22 μM, respectively. The only difference in the structure of these molecules is a hydroxyl group on the indole moiety (Fig. 1). This shows that substitutions on the aromatic region of the molecule can influence potency. This

indicates that further lead molecules to be studied will include those molecules having substituted aromatic moieties.

The structural features to be varied systematically from lead molecules described herein include (1) net positive charge, (2) number of methylenes separating nitrogens, and (3) cyclic versions of, e.g., polyamines, with and without changes in methylene spacing and net positive charge. In addition systematic variations in the structure and location of aromatic groups can be examined, e.g., in a variety of arylalkylamines isolated from the venoms of wasps and spiders; and synthetic molecules can be prepared by the coupling of commercially available aromatic moieties to the argiotoxin polyamine moiety. The argiotoxin polyamine moiety can be readily coupled to any aromatic moiety containing a carboxylic acid. Thus, it is simple to systematically screen the hydroxy and methoxy derivatives of phenylacetic acid and benzoic acid as well as the hydroxyindoleacetic acid series. Analogs containing heteroaromatic functionalities can also be prepared and assessed for activity.

Comparisons of potency and efficacy among such molecules will reveal the optimal structure and location of the aromatic group at a constant positive charge.

One of the structural variations on the polyamine motif that seems to increase potency is the presence of the cyclic version of the straight chain- parent molecule. Budmunchiamine A, isolated from the plant Albizia amara, is a cyclic derivative of spermine (Fig. 1). The addition of budmunchiamine A to bovine parathyroid cells caused a rapid and transient increase in $[Ca^{2+}]_i$ that persisted in the

absence of extracellular Ca^{2+} and was blunted by pretreatment with PMA. It therefore causes the mobilization of intracellular Ca^{2+} in parathyroid cells, probably by acting on the Ca^{2+} receptor. It is about equipotent with spermine (EC_{50} about 200 μM) yet carries one less positive charge (+3) than does spermine.

The results obtained with budmunchiamine A demonstrate the predictive power of the structure-activity studies and the novel structural information to be gained by testing natural products. Thus, screening of natural products, selected rationally on the basis of the structural information is readily performed e.g., molecules can be selected on the basis of well-established chemotaxonomic principles using appropriate data bases, such as Napralert. For example, macrocyclic polyamine alkaloids derived from papilionoid legumes related to Albizia, such as Pithecolobium and other plant-derived molecules can be screened.

Fig. 36 provides a second example of a series of molecules which were screened to determine useful molecules of this invention. These molecules were generally derived from fendiline and tested to determine their respective EC_{50}'s. Moreover, testing of related molecules, such as NPS 447 and NPS 448 reveals stereospecific effects of molecule structure. The most active compounds tested to date are the novel compounds designated NPS 467 and NPS 568 which have EC_{50} values of less than 5 μM. Those in the art, by reviewing this series of molecules, can determine other suitable derivatives which can be tested in the invention.

These examples demonstrate the general design and screening process useful in this invention, and indicate that additional compound and natural product libraries can be screened as desired by those in the art to determine other useful lead molecules or novel molecules of this invention.

As discussed above, examples of molecules useful as calcimimetics include branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines. In addition, other positively charged organic molecules, including naturally occurring molecules and their analogs, are useful calcimimetics. These naturally occurring molecules and their analogs preferably have positive charge-to-mass ratios that correlate with those ratios for the molecules exemplified herein. (Examples include material isolated from marine species, arthropod venoms, terrestrial plants and fermentation broths derived from bacteria and fungi.) It is contemplated that one group of preferred naturally occurring molecules and analogs useful as calcimimetics will have a ratio of positive charge: molecular weight (in daltons) from about 1:40 to 1:200, preferably from about 1:40 to 1:100. More specific examples of such molecules are provided below.

Polyamines

The polyamines useful as calcimimetics in this invention may be either branched or cyclic. Branched or cyclic polyamines potentially have higher calcimimetic activity than their straight-chain analogs. That is, branched or cyclic polyamines tend to have a lower EC_{50} than their corresponding linear polyamines with the same effective charge at physiological pH (see Table 1).

Table 1

Molecule	Net (+) Charge	$EC_{50}(\mu M)$
Neomycin	+6	20 or 40
Hexacyclen	+6	20
NPS 382	+8	50
NPS 381	+10	100
NPS 383	+8	150
Gentamicin	+5	150
Spermine	+4	150
Bekanamycin	+5	200
Argiotoxin-659	+4	300
Pentaethylenehexamine (PEHA)	+6	500
Streptomycin	+3	600
Spermidine	+3	2000
Tetraethylenepentamine (TEPA)	+5	2500
1,12-diaminododecane (DADD)	+2	3000
Triethylenetramine (TETA)	+4	8000

"Branched polyamines" as used herein refers to a chain molecule consisting of short alkyl bridges or alkyl groups joined together by amino linkages, and also containing points at which the chain branches. These "branch points" can be located at either a carbon atom or a nitrogen atom, preferably at a nitrogen atom. A nitrogen atom branch point is typically a tertiary amine but it may also be quaternary. A branched polyamine may have 1 to 20 branch points, preferably 1 to 10 branch points.

Generally, the alkyl bridges and alkyl branches in a branched polyamine are from 1 to 50 carbon atoms in length,

preferably from 2 to 6 carbon atoms. The alkyl branches may also be interrupted by one or more heteroatoms (nitrogen, oxygen or sulfur) or substituted with functional groups such as: halo, including fluoro, chloro, bromo, or iodo; hydroxy; nitro; acyloxy (R'COO-), acylamido (R'CONH-), or alkoxy (-OR'), where R' may contain from 1 to 4 carbon atoms. The alkyl branches may also be substituted with groups that are positively charged at physiological pH, such as amino or guanido. These functional substituents may add or change physical properties such as solubility to increase activity, delivery or bioavailability of the molecules.

The branched polyamines may have three or more chain and branch termination points. These termination points may be methyl groups or amino groups, preferably amino groups.

One preferred group of molecules is the group of branched polyamines having the formula:

$$H_2N-(CH_2)_j-(NR_i-(CH_2)_j)_k-NH_2$$

where k is an integer from 1 to 10, each j is the same or different and is an integer from 2 to 20, and each R_i is the same or different and is selected from the group consisting of hydrogen and $-(CH_2)_j-NH_2$, where j is as defined above, and at least one R_i is not hydrogen.

Particularly preferred branched polyamines of this invention are the molecules $N^1,N^1,N^5,N^{10},N^{14},N^{14}$-hexakis-(3- aminopropyl) spermine and $N^1,N^1,N^5,N^{14},N^{14}$-tetrakis-(3-aminopropyl)spermine referred to as NPS 381 and NPS 382, respectively, in Figure 1.

"Cyclic polyamines" as used herein refer to heterocycles containing two or more heteroatoms (nitrogen,

oxygen or sulfur), at least two of which are nitrogen atoms. The heterocycles are generally from about 6 to about 20 atoms in circumference, preferably from about 10 to about 18 atoms in circumference. The nitrogen heteroatoms are separated by 2 to 10 carbon atoms. The heterocycles may also be substituted at the nitrogen sites with aminoalkyl or aminoaryl groups (NH_2R-), wherein R is aminoaryl or a lower alkyl of 2 to 6 carbon atoms.

Particularly preferred cyclic polyamines of this invention are shown in Figure 1 as hexacyclen (1,4,7,10,13,16-hexaaza-cyclooctadecane) and NPS 383.

Polyamino Acids

The polyamino acids useful in this invention may contain two or more positively charged amino acid residues at physiological pH. These positively charged amino acids include histidine, lysine and arginine. These polypeptides will vary in length from 2 to 800 amino acids in length, more preferably from 20 to 300 amino acids in length. These polypeptides may consist of a single repeating amino acid residue, or may have the variety of a naturally occurring protein or enzyme.

The amino acid residues comprising the polyamino acids may be any of the twenty naturally occurring amino acids, or other alternative residues. Alternative residues include, for example, the ositions of this invention may also contain components derivatized with a molecule or ion which acts as a label. A wide variety of labeling moieties can be used, including radioisotopes, chromophores, and fluorescent labels. Radioisotope labeling in particular can be readily detected in vivo. Radioisotopes may be coupled

by coordination as cations in the porphyrin system. Useful cations include technetium, and indium. In the compositions, the positively charged molecule can be linked to or associated with a label.

Methods of Synthesis

Strategies for the syntheses and the modification of polyamines involve the use of a variety of amine protecting groups (phthalimido, BOC, CBZ, benzyl, and nitrile) which can be removed selectively to construct functionalized molecules. The synthetic methods involved are modelled after those used to construct argiopines 636 and 659 and other arylalkylamines derived from spider venoms.

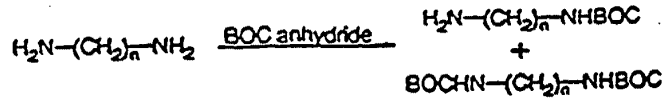
Chain extensions of 2-4 methylenes were typically accomplished by alkylation with the corresponding N-(bromoalkyl)phthalimide. A 1:1.2 mixture of amine to the bromoalkylphthalimide was refluxed in acetonitrile in the presence of 50% KF on Celite. Chain extensions were also accomplished by alkylation of a given amine with acrylonitrile or ethylacrylate. Reaction progress was monitored by TLC and intermediates purified on silica gel using combinations of dichloromethane, methanol, and isopropylamine. Final products were purified by cation exchange (HEMA-SB) and RP-HPLC (Vydac C-18). Purity and structure verification are accomplished by ^{1}H- and ^{13}C-NMR and high-resolution mass spectrometry (EI, CI and/or FAB).

BOC protecting groups were added by the treatment of an amine (1^{o} or 2^{o}) with di-tert-butyl dicarbonate in dichloromethane in the presence of a catalytic amount of dimethylaminopyridine. Benzyl protecting groups were

applied in one of two ways: (1) condensation of a 1^{o} amine with benzaldehyde followed by sodium borohydride reduction or (2) alkylation of a 2^{o} amine with benzylbromide in the presence of KF. Amide linkages and cyclizations were typically performed by the reaction of an amine (1° or 2°) with the N-hydroxysuccinimide ester of a given acid. This was accomplished directly (in the case of cyclizations) by treatment of the "amino acid" with dicyclohexylcarbodiimide under dilute conditions.

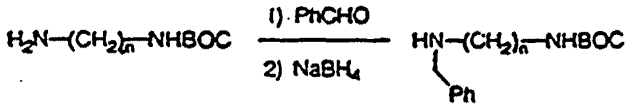
Deprotections of the phthalimido functionality were accomplished by reduction with hydrazine in refluxing methanol. Deprotections of the BOC functionality were accomplished in anhydrous TFA. Deprotection of benzyl, nitrile, and CBZ protecting functionalities was accomplished by reduction in glacial acetic acid under 55 psi hydrogen in the presence of a catalytic amount of palladium hydroxide on carbon. Nitrile functionalities (in the presence of benzyl and CBZ groups) were selectively reduced under hydrogen in the presence of sponge Raney nickel.

Specifically, branched polyamines are typically prepared from simple diaminoalkanes of the formula NH_2-$(CH_2)_n$-NH_2, or simple polyamines such as spermidine or spermine. One of the two primary (terminal) amines is protected or "masked" with a protecting group such as BOC (t-butyloxycarbonyl), phthalimido, benzyl, 2- ethylnitrile (the Michael condensation production product of an amine and acrylonitrile), or amide. A typical reaction is the addition of a BOC protecting group by treatment with di-t-butyl-dicarbonate (BOC anhydride):

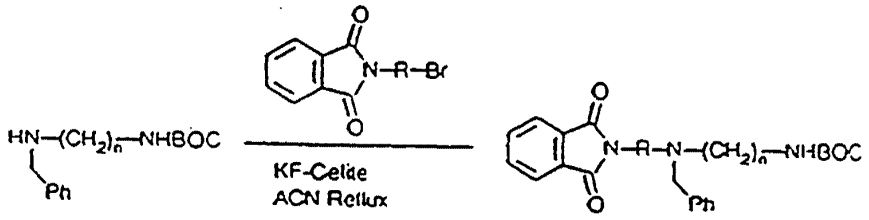


The monoprotected product is separated from the unprotected and diprotected products by simple chromatographic or distillation techniques.

The remaining free amine in the monoprotected product is then selectively alkylated (or acylated) with an alkylating (or acylating) agent. To ensure mono-alkylation, the free amine is partially protected by condensation with benzaldehyde followed by sodium borohydride reduction to form the N-benzyl derivative:



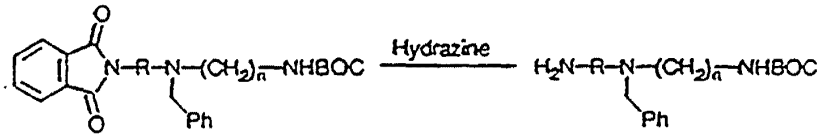
The N-benzyl derivative is then reacted with the alkylating agent. A typical alkylating agent is in an N-(bromoalkyl)phthalimide, which reacts as follows:



For example, N-(bromobutyl)phthalimide is used to extend or branch the chain with four methylene units. Alternatively,

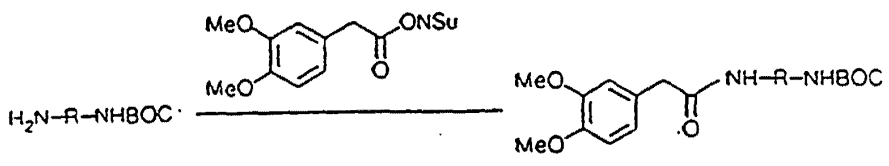
reaction with acrylonitrile followed by reduction of the cyano group will extend the chain by three methylenes and an amino group.

The protecting groups of the resulting chain-extended molecule can then be selectively cleaved to yield a new free amine. For example, trifluoroacetic acid is used to remove a BOC group; catalytic hydrogenation is used to reduce a nitrile functionality and remove a benzyl group; and hydrazine is used to remove phthalimido groups as follows:



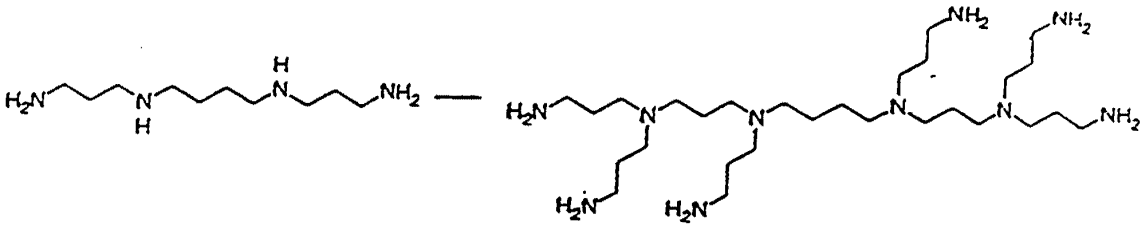
The new free amine may be alkylated (or acylated) further as above to increase the length of the polyamine. This process is repeated until the desired chain length and number of branches is obtained. In the final step, deprotection of the product results in the desired polyamine. However, further modifications may be effected at the protected end prior to deprotection in the following manner:

For example, prior to BOC-deprotection, the polyamine is acylated with the N-hydroxysuccinimide ester of 3,4-dimethoxyphenylacetic acid to yield a diprotected polyamine:



This ultimately will yield an arylalkyl polyamine. The BOC group can then be selectively removed with trifluoroacetic acid to expose the other amino terminus which can be extended as above.

Certain branched polyamines may be formed by simultaneously alkylating or acylating the free primary and secondary amines in a polyamine formed as above. For example, treatment of spermine with excess acrylonitrile followed by catalytic reduction yields the following:



Cyclic polyamines may be prepared as above beginning with starting materials such as hexacylen (Aldrich Chem.).

The polyamino acids within the scope of the present invention can be made by recombinant techniques known in the art, or may be synthesized using standard solid-phase techniques known in the art. Solid-phase synthesis is commenced from the carboxy-terminal end of the peptide using an μ-amino protected amino acid. BOC protective groups can be used for all amino groups even through other protective groups are suitable. For example, BOC-lys-OH can be esterified to chloromethylated polystyrene resin supports. The polystyrene resin support is preferably a copolymer of styrene with about 0.5 to 2% divinylbenzene as

a cross-linking agent which causes the polystyrene polymer to be completely insoluble in certain organic solvents. See Stewart et al., Solid-Phase Peptide Synthesis (1969), W.H. Freeman Co., San Francisco; and Merrifield, J. Am. Chem. Soc. (1963) 85:2149-2154. These and other methods of peptide synthesis are also exemplified by U.S. Patent Nos. 3,862,925; 3,842,067; 3,972,859; and 4,105,602.

The polypeptide synthesis may use manual techniques or automatically employing, for example, an Applied Biosystems 403A Peptide Synthesizer (Foster City, California) or a Biosearch SAM II automatic peptide synthesizer (Biosearch, Inc., San Rafael, California), following the instructions provided in the instruction manual supplied by the manufacturer.

The arylalkylamines of the invention are natural products isolated by known techniques, or synthesized as described in Jasys et al., Tetrahedron Lett. (1988) 29:6223-6226, and Nason et al., Tetrahedron Lett. (1989) 30:2337-2340.

One general protocol for preparation of fendiline (or fendiline analogs shown in Fig. 36) is as follows. In a 10 ml round bottom flask equipped with a magnetic stir bar and rubber septum, 1.0 mmole 3,3'- bisphenylpropylamine (or primary alkyl amine) in 2 ml ethanol was treated with 1.1 mmole phenol and 1.0 mmole acetophenone (or substituted acetophenone). To this was added 2.0 mmoles $MgSO_4$ and 1.0 mmole $NaCNBH_3$. This was stirred under a nitrogen atmosphere at room temperature (about 20°C) for 24 hrs. The reaction was poured into 50 ml ether and washed 3 times with 1 N NaOH and once with brine. The ether layer was dried

with anhydrous K_2CO_3 and reduced in vacuo. The product was then purified by column chromatography or HPLC incorporating osilica stationary phase with combinations of CH_2Cl_2- Methanol- isopropylamine (typically 3% Methanol and 0.1% isopropylamine in methylene chloride).

A preferred procedure for preparing fendiline or fendiline analogs (such as those depicted in Figure 36) uses titanium(IV) isopropoxide and was modified from methods described in J. Org. Chem. 55:2552 (1990). For the synthesis of NPS 544, titanium tetrachloride (method described in Tetrahedron Letters 31:5547 (1990)) was used in place of titanium(IV) isopropoxide. The reaction scheme is depicted in Figure 43a. In Figure 43a, R,R' and R" depict hydrocarbyl groups. According to one embodiment, in a 4 ml vial, 1 mmole of amine (1) (typically a primary amine) and 1 mmole ketone or aldehyde (2) (generally acetophenone) are mixed, then treated with 1.25 mmoles titanium(IV) isopropoxide (3) and allowed to stand with occasional stirring at room temperature for about 30 minutes. Alternatively, a secondary amine may be used in place of (1). Note: some reactions will give heavy precipitates or solids which are warmed/heated (to their melting point) to allow for stirring/mixing several times over the course of the reaction. The reaction mixture is treated with 1 ml ethanol containing 1 mmole sodium cyanoborohydride (4) and the resulting mixture is then allowed to stand at room temperature with occasional stirring for about 16 hours. After this time the reaction is quenched by the addition of about 500 µl water. The reaction mixture is then diluted to about 4 ml total volume with ethyl ether and then

centrifuged. The upper organic phase is removed and reduced on a rotavapor. The resulting product, (6), is partially purified by chromatography through a short column of silica (or alternatively by using preparative TLC on silica) using combination of dichloromethane:methanol:isopropylamine (typically 95:5:0.1), prior to purification by HPLC (normal phase using silica with dichloromethane:methanol: isopropylamine or reversed phase, C-18 with 0.1% TFA with acetonitrile or methanol).

If appropriate or desired, chiral resolution may be accomplished using methods such as those described in Example 21.

Formulation and Administration

As demonstrated herein, the molecules of the invention may be used to: (a) mimic or antagonize one or more of the effect of an extracellular ion, including extracellular Ca^{2+}; (b) affect the extracellular free Ca^{2+} level in an individual; and (c) treat diseases such as hyperparathyroidism, osteoporosis and hypertension. In general, diseases or conditions involving inorganic-ion receptors now can be studied, diagnosed and/or beneficially treated. While the molecules have generally been shown to have an effect on parathyroid cells, they may also modulate the Ca^{2+} receptors on other cells, including bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, distal tubule kidney cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cells), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac

atrial cell, gastrin and glucagon secreting cells, kidney mesangial cell and mammary cell.

While these molecules will typically be used in therapy for human patients, they may be used to treat similar or identical diseases in other warm-blooded animal species such as other primates, farm animals such as swine, cattle, and poultry; and sports animals and pets such as horses, dogs and cats.

In therapeutic and/or diagnostic applications, the molecules of the invention can be formulated for a variety of modes of administration, including systemic and topical or localized administration. Techniques and formulations generally may be found in Remington's Pharmaceutical Sciences, Mack Publishing Co., Easton, PA.

For systemic administration, oral administration is preferred. Alternatively, injection may be used, e.g., intramuscular, intravenous, intraperitoneal, and subcutaneous. For injection, the molecules of the invention are formulated in liquid solutions, preferably in physiologically compatible buffers such as Hank's solution or Ringer's solution. In addition, the molecules may be formulated in solid form and redissolved or suspended immediately prior to use. Lyophilized forms are also included.

Systemic administration can also be by transmucosal or transdermal means, or the molecules can be administrated orally. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal

administration bile salts and fusidic acid derivatives. In addition, detergents may be used to facilitate permeation. Transmucosal administration may be through nasal sprays, for example, or using suppositories. For oral administration, the molecules are formulated into conventional oral administration forms such as capsules, tablets, and tonics.

For topical administration, the molecules of the invention are formulated into ointments, salves, gels, or creams, as is generally known in the art.

As shown in the examples below, the amounts of various compounds of this invention which must be administered can be determined by standard procedures. Generally it is an amount between about 1 and 50 mg/kg animal to be treated.

Recombinant Receptors

Natural product screening has traditionally provided the lead structures for the development of diverse therapeutic molecules. However, high-throughput screening of natural product libraries or other molecule libraries for activity on the Ca^{2+} receptor has not previously been possible. To achieve this capability, it is best to clone the Ca^{2+} receptor cDNA and then create transfected cell lines suitable for high-throughput screening. The structure of the receptor can additionally be used to gain insight into the molecular geometry of the ligand binding site(s), and such information used to augment a rational drug design program as discussed above. Limited structure-activity studies and testing of selected natural product molecules will provide the initial structural data base necessary to guide rational natural product screening and drug design.

The discovery of a superfamily of genes permits the same advantages in connection with all inorganic-ion receptors. Although the following discussion will refer often to methodology for cloning a calcium receptor, it will be understood by those of ordinary skill in the art that the methodology generally is applicable to the cloning of all inorganic-ion receptors.

These recombinant receptors allow for the first time screening mimics and lytics, including calcimimetics and calcilytics. For example, by binding assays with the receptors. This is particularly useful in screening with the human receptors in order to identify therapeutically useful compounds.

The bovine and human parathyroid cell Ca^{2+} receptor cDNA can be cloned by functional expression in Xenopus oocytes. It is possible to monitor an increase in intracellular Ca^{2+} in Xenopus oocytes indirectly by measuring current through the endogenous Ca^{2+}-activated Cl^- channel. The amplification of the response afforded by this signal transduction pathway enables the detection of receptor proteins encoded by mRNA at very low levels. This allows the detection of receptor- specific cDNA clones without the need for high affinity ligands, specific antisera, or protein or nucleic acid sequence information. An example of such a procedure follows.

Adult female Xenopus laevis were obtained from Xenopus I (Ann Arbor, MI) and maintained according to standard procedures. Lobes of ovary were excised from hypothermically-anesthetized toads. Clusters of oocytes were transferred into modified Barth's saline (MBS).

Individual oocytes were obtained by incubation in MBS containing 2 mg/ml collagenase (Sigma, Type 1A) for 2h at 21°C and stage V-VI oocytes were selected for injection.

Glass capillary tubes (1 mm diameter) were pulled to a fine tip and manually broken to achieve a tip diameter of about 15 µM. A droplet of mRNA (1 ng/nl in diethylpyrocarbonate (DEPC)-treated water) was placed onto parafilm and drawn into the capillary tube by suction. The capillary tube was then connected to a picospritzer (WPI Instruments) and the volume of the air-pulsed droplets adjusted to deliver 50 ng of mRNA (typically 50 nl). A 35 mm culture dish with a patch of nylon stocking fixed to the bottom was used to secure the oocytes during injection of mRNA into the vegetal pole. The injected oocytes were placed into a 35 mm culture dish containing MBS, 100 µg/ml penicillin and 100 µg/ml streptomycin and incubated at 18°C for 3 days.

Following incubation, an oocyte was placed into a 100 µl plastic chamber and superfused with MBS at a flow rate of 0.5 ml/min using a peristaltic pump. Test molecules or inorganic polycations were added by rapidly moving the tubing into different buffers. Recording and current-passing electrodes were constructed from thin wall capillary tubing pulled to a resistance of 1-3 Mohms and filled with 3 M KCl. Oocytes were impaled (in the animal pole) with both electrodes under microscopic observation and connected to an Axon Instruments Axoclamp 2A voltage-clamp amplifier which was used to set the holding potential (-70 to -80 mV) and to measure the currents that were passed to maintain the holding potential. Currents were recorded

directly onto a strip chart recorder.

For mRNA preparation, tissue was obtained from calves or patients with secondary HPT undergoing surgical removal of the parathyroid glands. Purified cells need not be prepared; whole pieces of gland were used to prepare mRNA that directs the expression of the Ca^{2+} receptor in Xenopus oocytes. Total cellular RNA was prepared by acid guanidinium thiocyanate/phenol extraction of homogenized glands. Oligo-dT cellulose chromatography was used to select poly(A)$^+$ mRNA by standard procedures. For size fractionation of mRNA, centrifugation through glycerol gradients was used. The mRNA was denatured with 20 mM methylmercuric hydroxide and loaded (50-100 µg at a concentration of 1 mg/ml) onto a linear 15-30% glycerol gradient prepared in Beckman TLS55 tubes. Following centrifugation at 34,000 rpm for 16 hrs, 0.3 ml gradient fractions were collected and diluted in an equal volume of water containing 5mM beta-mercaptoethanol. mRNA was then recovered by two cycles of ethanol precipitation. If desired, the mRNA (50-100 µg of poly(A)$^+$) may be separated on a 1.2% agarose/6.0 M urea preparative gel, along with a range of RNA size markers. Following visualization of the mRNA by ethidium bromide staining, gel slices containing RNA in approximately 1 kb to 2kb size steps are excised. mRNA is recovered from the agarose gel slices using RNAid binding matrix (according to the supplier's standard protocol; Stratagene, Inc.) and recovered mRNA fractions eluted into DEPC-treated water.

Amounts of recovered mRNA were quantified by UV absorbance measurement. The size range of mRNA contained

within each fraction of the glycerol gradient was determined by formaldehyde/agarose gel electrophoresis using a small quantity (0.5 μg) of each sample. The integrity of the mRNA was assessed by in vitro translation of each sample. Reticulocyte lysates (commercially available kits; BRL) were used to translate 0.05-0.5 μg of each mRNA fraction. The resulting ^{35}S-labelled proteins were analyzed by SDS-PAGE. The intact mRNA was capable of directing the synthesis of proteins of a complete size range, corresponding roughly to the sizes of the individual mRNA fractions.

A cDNA library was constructed in the vector λ ZAPII, following modifications of the technique of Gubler and Hoffman. RNA from the fraction(s) giving the best response in the oocyte assay was used as starting material. First-strand cDNA syntheses was primed with an oligo-dT/NotI primer-linker. Second-strand synthesis was by the RNase H/DNA Polymerase I self-priming method. Double-stranded cDNA was blunted with T4 DNA polymerase and EcoRI adaptors blunt-end ligated to the cDNA with T4 ligase. Following NotI digestion to cleave the linker, full-length cDNA was size-selected by exclusion chromatography on Sephacryl 500 HA. First-strand cDNA was radiolabeled with α-^{32}P-dATP, and all synthesis and recovery steps monitored by following the incorporation of radioactivity. Full-length cDNA recovered from the sizing column was ligated to EcoRI/NotI digested λ ZAPII arms. The ligation mix was test packaged with commercially available high-efficiency packaging extract (Stratagene, Inc.) and plated on the appropriate host strain (XL1-blue). The

percentage of recombinant phage was determined by the ratio of white-to-blue plaques when the library is plated on IPTG and X-gal.

The average insert size was determined from ten randomly selected clones. Phage DNA "mini-preps" were digested with EcoRI and NotI to release the insert, and the size determined by agarose gel electrophoresis. The library consisted of >90% recombinant phage, and the insert size ranged from 1.5 to 4.2 kb. The recombinant ligation was packaged in large scale to generate 800,000 primary clones. The packaging mix was titered and plated at 50,000 plaques per 15 cm plate. Each pool of 50,000 clones was eluted in SM buffer and stored individually.

Plate lysate stocks of each of the clone pools were used for small-scale phage DNA preparation. Phage particles are concentrated by polyethylene glycol precipitation, and phage DNA purified by proteinase K digestion followed by phenol:chloroform extraction. Twenty micrograms of DNA are digested with NotI, and used as template for *in vitro* transcription of sense-strand RNA. *In vitro* transcription is according to standard protocols, utilizing T7 RNA polymerase and 5' cap analog m^7GpppG in a 50 µl total reaction volume. Following Dnase I/Proteinase K digestion and phenol/chloroform extraction, the RNA is concentrated by ethanol precipitation and used for oocyte injection.

Oocytes were injected with synthetic mRNA (cRNA) from each of the 16 library subpools constituting 50,000 independent clones each. After incubation for 3 to 4 days, oocytes were assayed for the ability of 10 mM neomycin to elicit a Ca^{2+}-dependent Cl-current. A pool designated

"pool 6" gave a positive signal and thus contains a cDNA clone encoding a functional calcium receptor. In order to decrease the complexity of pool 6 and thus proceed towards the purification of the calcium receptor clone contained within this pool, pool 6 phage were replated at μ20,000 plaques per plate and 12 plates harvested. DNA was prepared from each of these subpools and cRNA synthesized. Again, oocytes were injected with cRNA and assayed 3-4 days later for the ability of 10 mM neomycin to elicit a Ca^{2+}-dependent Cl-current. A subpool pool 6-3 was positive and this pool was subjected to a further round of plating, reducing the complexity of pools to around 5,000 clones per pool. Pools were again assayed by preparation of cRNA and injection in oocytes. A subpool pool 6-3.4 was positive. In order to expedite further purification of the positive clone in pool 6-3.4, phage DNA from this pool was rescued as plasmid DNA by superinfection with the helper phage, ExAssist (Stratagene). Transfection of rescued plasmids into bacterial strain DH5alphaF' resulted in transformed bacterial colonies on ampicillin plates. These were harvested in pool of 900 clones each. Plasmid DNA was then prepared from each subpool and cRNA synthesized and assayed in the usual manner. Subpool 6-3.4.4 was positive. Bacteria containing the plasmid subpool 6-3.4.4 were subsequently plated in subpools of μ50 clones each. Continuation of this process is expected to result in a single clone encoding a functional calcium receptor.

Alternative assays are available for detecting expression of a calcium receptor in oocytes. For example, oocytes can be loaded with $^{45}Ca^{++}$ and then treated with

a calcimimetic. Mobilization of $^{45}Ca^{++}$ from intracellular stores results in a net increase in $^{45}Ca^{++}$ efflux which is readily determined. Fluorescent Ca^{++} indicators may also be injected into oocytes. In this case, oocytes expressing a calcium receptor will exhibit increased fluorescence upon activation with a calcimimetic. These assays may be employed for cloning calcium receptors in place of the electrophysiological assay for calcium induced Cl^- current described in the above example. In addition, the calcimimetic ligand used in the cloning procedure need not be neomycin as indicated above but could instead be, for example, Gd^{+++}, Ca^{++}, Mg^{++} or other calcimimetic compound.

Initial experiments used Xenopus oocytes injected with water or poly(A)$^+$-enriched mRNA (50 ng) from bovine parathyroid cells. After three days, the oocytes were examined for their ability to increase intracellular Ca^{2+} in response to increases in the concentration of extracellular di- and trivalent cations. The oocytes were impaled with recording and current-passing electrodes and $[Ca^{2+}]_i$ was assessed indirectly by measuring currents through the endogenous Ca^{2+}-activated Cl^- channel. In oocytes injected with poly(A)$^+$-enriched mRNA from bovine (or human, Fig. 26) parathyroid cells, increasing the concentration of extracellular Ca^{2+} from 0.7 to 3, 5 or 10 mM caused a rapid and transient increase in the Cl^- current which then oscillated around a higher basal current. Increasing the concentration of extracellular Mg^{2+} from 1 to 10 mM likewise evoked oscillatory increases in Cl^- current. The Cl^- current response to

extracellular Mg^{2+} persisted when the extracellular Ca^{2+} concentration was reduced to < 1 μM (Fig. 25).

The impermeant trivalent cation Gd^{3+} (600 μM) also caused oscillatory increases in the Cl^- current (Fig. 25). Such increases in the Cl^- current which oscillate and persist in the nominal absence of extracellular Ca^{2+} are noted when oocytes have been allowed to express other Ca^{2+}-mobilizing receptors and are stimulated with the appropriate ligand (e.g., substance K, Fig. 25). In these instances, the increase in Cl^- current reflects the mobilization of intracellular Ca^{2+}. These initial studies likewise show that extracellular polycations mobilize intracellular Ca^{2+} in parathyroid cell mRNA-injected oocytes.

Oocytes injected with water did not show any change in the Cl^- current when exposed to extracellular Ca^{2+} (10 mM) or Mg^{2+} (20 or 30 mM). In one series of experiments, oocytes were injected with the mRNA encoding the substance K receptor. In these oocytes, extracellular Mg^{2+} (20 mM) did not evoke any current but the cells responded vigorously to the addition of substance K (Fig. 25). These experiments indicate that there is no endogenous sensitivity of the oocyte to extracellular Ca^{2+} or Mg^{2+}.

Similar experiments were performed using oocytes injected with poly(A)$^+$-enriched mRNA prepared from human parathyroid glands (hyperplastic tissue from a case of secondary HPT). In these oocytes, increasing the concentration of extracellular Ca^{2+} caused a reversible increase in the Cl^- current which oscillated (Fig. 26). The addition of 300 μM La^{3+} likewise caused oscillatory

increases in the Cl^- current. Increasing the concentration of extracellular Mg^{2+} from 1 to 10 mM evoked increases in the Cl^- current that persisted in the absence of extracellular Ca^{2+}. Additional experiments suggest that the response to extracellular Ca^{2+} is concentration dependent. Thus, in three mRNA-injected oocytes, Cl^- current increased to a maximum of 111 $\pm$ 22 nA at 3 mM and 233 $\pm$ 101 nA at 10 mM extracellular Ca^{2+}.

The results obtained in Xenopus oocytes demonstrate the presence of a mRNA(s) in parathyroid cells encoding a protein(s) which can impart, in normally unresponsive cells, sensitivity to extracellular Ca^{2+}. Moreover, the ability of extracellular Mg^{2+} to evoke oscillatory increases in Cl^- current in the absence of extracellular Ca^{2+} demonstrates that the Cl^- current depends on the mobilization of intracellular Ca^{2+} rather than influx of extracellular Ca^{2+}. The results obtained with La^{3+} likewise show that the expressed protein(s) is linked to the mobilization of intracellular Ca^{2+}. Together, these data show that the expressed protein(s) acts as a cell surface receptor rather than a channel. These studies provide compelling evidence for the existence of a Ca^{2+} receptor protein on the surface of parathyroid cells and demonstrate the feasibility of using the Xenopus oocyte system to achieve the molecular cloning of the Ca^{2+} receptor cDNA.

In another series of experiments, parathyroid cell mRNA was size fractionated by centrifugation through a glycerol gradient. Ten fractions were collected. Each group was injected into Xenopus oocytes and after a three day incubation period the oocytes were assayed for

expression of the Ca^{2+} receptor. Those oocytes injected with fractions 4-6 showed the largest and most consistent increases in Cl^- current in response to extracellular Ca^{2+} (Fig. 35). These results indicate that the Ca^{2+} receptor is encoded by mRNA in a size range of 2.5-3.5 kb. This indicates that a strategy using direct expression of RNA synthesized from a transcription vector cDNA library is feasible. Size-fractionation experiments of this sort were conducted and in each of three different fractionation experiments similar results were obtained.

The mRNA fractions obtained and characterized in the preceding experiments can be assayed by injection into oocytes. For each mRNA fraction, 10-20 oocytes are injected with 50 ng of RNA at a concentration of 1 ng/nl in water. Injected oocytes are maintained at 18° C for 48-72 h after which they are assessed for expression of the Ca^{2+} receptor using measurements of Cl^- current. For each group of injected oocytes the number positive for expression of the receptor, as well as the magnitude of the Ca^{2+}-dependent Cl^- current measured, is determined. As negative controls, oocytes are injected with rat liver poly(A)$^+$-enriched mRNA, yeast RNA, or water.

It is expected that an mRNA in the range of 2.5 - 3.5 kb will encode the receptor. mRNA of a larger size may necessitate a cloning approach based on hybrid depletion of parathyroid mRNA prior to oocyte injection. This strategy is not dependent upon the generation of full-length cDNA clones for success. If receptor expression is not obtained with a single size fraction of mRNA, oocytes are injected with mixed size fractions to determine a combination that

does give rise to a functional receptor. If it does appear that multiple subunits are necessary for the formation of a functional receptor, the hybrid depletion expression cloning strategy is used. In this approach, clones are selected on the basis of their ability to deplete a specific mRNA species from the total mRNA population. A clone encoding a single subunit is identified by its ability to prevent the formation of the active multi-subunit complex. By exhaustive screening it is possible to identify clones encoding all of the necessary subunits.

This approach permits the isolation of clones encoding individual subunits required to form a functional receptor complex. Synthetic RNA from pools of clones are assayed for their ability to induce expression of the Ca^{2+} receptor in Xenopus oocytes by the same techniques used to analyze the original mRNA fractions. Originally, 10 pools representing 100,000 primary clones each are examined. Pools of clones showing a positive response are screened at lower (typically 4-to-10 fold) complexity, and again positive pools further subdivided and screened. This process of library sub-fractionation is followed until individual positive clones are identified. As a negative control for the oocyte expression assay, anti-sense transcripts are generated from those DNA templates that induce a positive response. Anti-sense transcripts are unable to give rise to an authentic receptor, and this will control any non-specific positive signal arising from injection of synthetic RNA. Another concern is the fact that synthetic RNA can occasionally "poison" translation in injected oocytes, by an undefined mechanism. To control for

this possibility, synthetic RNAs giving a negative response are co-injected at various dilutions with parathyroid cell mRNA, to determine if they are non-specifically interfering with the expression of the Ca^{2+} receptor.

When an individual clone encoding the Ca^{2+} receptor is identified, the cDNA insert will be excised from the vector and used for large scale production of synthetic RNA. Oocyte injection of this single RNA species allows rigorous assessment of the characteristics of the expressed receptor.

If the size of the mRNA encoding the Ca^{2+} receptor is too large for cloning by direct transcription and expression, or if multiple subunits are involved, a hybrid-depletion technique of screening pools of clones is used. cDNA insert DNA will be prepared from pools of clones from the size-selected parathyroid cell cDNA library. This DNA is hybridized to parathyroid cell mRNA under conditions that permit the formation of DNA/RNA duplexes. The unannealed, hybrid-depleted RNA is recovered and used for oocyte injections. Other methods for hybrid-depletion or hybrid-arrest may be used alternatively and are well known by those skilled in the art. DNA from pools of clones containing sequences representing Ca^{2+}-receptor mRNA is depleted from this mRNA from the total parathyroid cell mRNA population, and expression of the receptor is reduced or absent upon oocyte injection. A process of sub-fractionation is followed on pools of clones of decreasing complexity, at each step assaying for cloned DNA that depletes Ca^{2+} receptor-encoding mRNA from the total parathyroid cell mRNA population. The use of an internal

control during the hybrid depletion assays ensures that the hybrid-depleted RNA is intact and capable of being translated in the oocyte.

Human parathyroid cells express a beta-adrenergic receptor coupled to adenylate cyclase. This receptor can be expressed in oocytes, where it is capable of agonist-induced activation of the endogenous adenylate cyclase. During the hybrid-depletion screening for Ca^{2+} receptor clones, oocytes injected with hybrid depleted mRNA are assayed for isoproterenol-induced adenylate cyclase activation. A positive response in this assay serves to indicate that any observed inhibition of Ca^{2+} receptor response is specific, and not due to a general inhibition of the total mRNA population.

The hybrid-depletion screening strategy can result in the isolation of clones that do not contain a complete protein coding region. Positive clones isolated by this screening strategy are sequenced to determine their protein coding capacity. Northern blot analysis of human parathyroid gland RNA permits the determination of the size of the complete mRNA corresponding to specific clones. If positive clones do not appear to be full length, the cloned cDNA will be used as a hybridization probe to screen a parathyroid gland cDNA library for complete cDNAs.

A variety of cell lines are capable of coupling exogenously expressed receptors to endogenous functional responses. A number of these cell lines (e.g., NIH-3T3, HeLa, NG115, CHO, HEK 293 and COS7) can be tested to confirm that they lack an endogenous Ca^{2+} receptor. Those lines lacking a response to external Ca^{2+} can be used to

establish stably transfected cell lines expressing the cloned Ca^{2+} receptor.

Production of these stable tansfectants is accomplished by transfection of an appropriate cell line with a eukaryotic expression vector, such as pMSG, in which the coding sequence for the Ca^{2+} receptor cDNA has been cloned into the multiple cloning site. These expression vectors contain a promoter region, such as the mouse mammary tumor virus promoter (MMTV), that drive high-level transcription of cDNAs in a variety of mammalian cells. In addition, these vectors contain genes for the selection of cells that stably express the cDNA of interest. The selectable marker in the pMSG vector encodes an enzyme, xanthine-guanine phosphoribosyl transferase (XGPRT), that confers resistance to a metabolic inhibitor that is added to the culture to kill the nontransfected cells. A variety of expression vectors and selection schemes are usually assessed to determine the optimal conditions for the production of Ca^{2+} receptor expressing cell lines for use in high throughput screening assays.

The most effective method for transfection of eukaryotic cell lines with plasmid DNA varies with the given cell type. The Ca^{2+} receptor expression construct will be introduced into cultured cells by the appropriate technique, either Ca^{2+} phosphate precipitation, DEAE-dextran transfection, lipofection or electroporation.

Cells that have stably incorporated the transfected DNA will be identified by their resistance to selection media, as described above, and clonal cell lines will be produced by expansion of resistant colonies. The expression

of the Ca^{2+} receptor cDNA by these cell lines will be assessed by solution hybridization and Northern blot analysis. Functional expression of the receptor protein will be determined by measuring the mobilization of intracellular Ca^{2+} in response to externally applied Ca^{2+} receptor agonists.

Cloning the Ca^{2+} receptor enables both structural and functional studies of this novel receptor. Recombinantly produced receptor may be crystallized for structural studies. Stably transfected cell lines expressing the receptor can be used for high-throughput screening of natural product or other compound libraries. Molecules of the requisite potency and specificity can be labeled (radioactively or fluorescently). The ability of test molecules/extracts to displace such a labeled molecule will form the basis of a high-throughput assay for screening.

Another strategy for cloning an inorganic-ion receptor in oocytes is as follows.

The general procedures for isolation, defolliculation and injection of frog oocytes are described below. In brief, Xenopus laevis frogs are anesthetized by immersion in 0.17% Tricaine and a lobe of the ovary removed and minced into small pieces. Individual stage V-VI oocytes are isolated using an L-shaped capillary tube after incubating the ovarian tissue in a calcium-free buffer at room temperature for 90 min. The separated oocytes [usually 200-300 per experiment] are then incubated in collagenase solution [Sigma, Type II; 2 mg/ml] for an additional 90 min following which the follicular layer can be removed with fine forceps. The defolliculated oocytes are incubated overnight

in frog ringer [ND96] at 17°C and degenerating oocytes removed [usually <2-3% of the total]. Surviving oocytes are then injected with 50 nl H_2O [control] or poly(A)$^+$RNA[15-50 ng/oocyte; in 50 nl H_2O] and then incubated for 2-4 days at 17°C. (The frog ringer, ND96, contains (mM): NaCl (96), KCl (2), $CaCl_2$ (0.5), $MgCl_2$ (0.5), HEPES (5), pyruvate (2.5)].

Total RNA is extracted from the tissue [kidney, osteoclasts, etc.] with guanidinium thiocyanate and separated on a CsCl cushion. Poly(A)$^+$RNA is then isolated by oligo(dT) cellulose chromatography [two passes through the column]. The integrity of the poly(A)$^+$RNA is assessed by agarose gel electrophoresis. The ability of 25-50 ng of poly(A)$^+$RNA to give rise to Ca^{2+}-dependent Cl^- channel activity upon exposure to extracellular Gd^{3+} is assessed [i.e., as evidence for Ca^{2+} receptor activity]. The next step is to localize the mRNA expressing Ca^{2+} receptor activity to a small size range (~1 kb). For this purpose, ~100 µg of poly(A)$^+$RNA is separated by sucrose gradient and 40 size fractions collected from the gradient. Alternatively, ~200-300 µg of poly(A)$^+$RNA can be fractionated by preparative, continuous flow agarose gel electrophoresis [Hediger, M.A.: Anal. Biochem. 159:280-86, (1986) and 70-90 fractions collected. X. laevis oocytes are injected with pools of fractions [3-5 fractions/pool] of poly(A)+RNA [0.2-0.5 ng/oocyte] fractions from the pool(s) and the ability of these pools to give rise to Ca^{2+} receptor activity assessed. Individual fractions from the pool(s) giving the highest Ca^{2+} receptor activity are injected into oocytes to define the fraction(s) within this

pool of poly(A)+RNA that give the highest expression of the Ca^{2+} receptor in oocytes.

A directional cDNA library is constructed from this poly(A)+RNA pool using the SuperScript Plasmid system [pSPORT1; BRL]. cDNA is generated using the SuperScript, MuMLV-RT, reverse transcripts [many of which will be full-length for the coding region]. cDNA is sized by gel electrophoresis and the appropriate cDNA size region [based on the poly(A)+RNA size range] is extracted using GENECLEAN II [Bio 101]. This size selected cDNA is then directionally ligated into the pSPORT1 plasmid vector using a Not I primer at the 3' end and an adapter for Sal I at the 5' end. The resulting plasmids are introduced into ELECTROMAX DH10B cells [BRL] by electroporation. The transformed bacteria are grown on nitrocellulose filters [500-800 colonies/filter] and master filters stored at 4°C [short-term] or -70°C [long-term]. Replicate filters are grown and plasmid DNA is isolated from the filters, linearized by restriction cutting, and then used as a template for sense cRNA synthesis by in vitro transcription [T7 promoter in the presence of Cap-analog]. The pools of cRNA are individually injected into oocytes which are assayed for expression of Gd^{3+}-induced [1-100μM] activation of Cl^- currents. A standard sib selection procedure is utilized to identify a clone that expresses Ca^{2+} receptor activity in oocytes.

A restriction map of the Ca^{2+} receptor cDNA is generated and suitable restriction fragments subcloned into pSPORT1. Subcloned cDNAs are bidirectionally sequenced using standard methods [Sequenase Polymerase 2® 2, USB].

Sequencing primers (18-mer) are used where necessary to sequence regions within or between subclones or to resolve compressions or ambiguous regions of sequence. The coding region for the Ca^{2+} receptor protein is determined from the longest open reading frame that has a start site preferably homologous with the Kozak consensus sequence. Hydropathy and other protein algorithms are used to generate a topology for the Ca^{2+} receptor protein. The nucleotide sequence of the cDNA, the amino acid sequence of the Ca^{2+} receptor protein, and the protein topology are compared with that for the other Ca^{2+} receptors and other known cDNAs and proteins present in the database. Homologous regions might represent domains involved in cation binding or regulation.

The presently preferred method for isolating inorganic-ion receptor nucleic acid is based upon hybridization screening.

Region specific primers or probes derived from BoPCaR 1 can be used to prime DNA synthesis and PCR amplification, as well as to identify colonies containing cloned DNA encoding a member of the inorganic ion receptor family using known methods (Innis et al., PCR Protocols, Academic Press, San Diego, CA (1990)).

When using primers that are derived from nucleic acid encoding an inorganic-ion receptor, one skilled in the art will recognize that by employing high stringency conditions, annealing at 50-60°C, sequences which are greater than about 76% homologous to the primer will be amplified. By employing lower stringency conditions, annealing at 35-37°C, sequences which are greater than about 40-50% homologous to

the primer will be amplified.

When using DNA probes derived from inorganic ion receptors for colony/plaque hybridization, one skilled in the art will recognize that by employing high stringency condition, hybridization at 50-65°C, 5X SSPC, 50% formamide, wash at 50-65°C, 0.5X SSPC, sequences having regions which are greater than about 90% homologous to the probe can be obtained, and by employing lower stringency conditions, hybridization at 35-37°C, 5X SSPC, 40-45% formamide, wash at 42°C SSPC, sequences having regions which are greater than 35-45% homologous to the probe will be obtained.

Any tissue can be used as the source for the genomic DNA encoding members of the inorganic-ion receptor family. However, with respect to RNA, the most preferred source is tissues which express elevated levels of the desired inorganic-ion receptor family member. In the present invention, oocyte injection and two-electrode whole oocyte voltage clamping was used to identify expression from such a tissue source. However, using the sequences disclosed herein, it is now possible to identify such cells using the inorganic-ion receptor sequence as a probe in northern blot or in situ hybridization procedures, thus eliminating the necessity of the procedures employed to clone the first member of this family and eliminating the need to obtain RNA from a tissue which expresses elevated levels of inorganic-ion receptor.

The present invention further provides methods of identifying cells or tissues which express a member of the inorganic-ion receptor family. A probe comprising the DNA sequence, for example, of BoPCaR 1, a fragment thereof, or a

DNA sequence encoding another member of the inorganic-ion receptor family of proteins can be used as a probe or amplification primer to detect cells which express a message homologous to the probe or primer. One skilled in the art can readily adapt currently available nucleic acid amplification or detection techniques so that it employs probes or primers based on the sequences encoding a member of the inorganic-ion receptor family.

Given the appropriate cells or tissues expressing other receptors, these receptors may be cloned in a manner analogous to that described above for the parathyroid cell calcium receptor. For example, mRNA from human osteoclastoma tissue encodes the osteoclast calcium receptor (Figure 34). Thus, to isolate a clone for the human osteoclast receptor, one need only isolate mRNA from osteoclastoma tissue, prepared a cDNA library and assay/fractionate subpools as described above. Furthermore, the preferred receptors for drug screening are of human origin. A clone encoding a receptor from one species may be used to obtain the corresponding human cDNA clone by cross-hybridization as is well known by those skilled in the art. In addition, the clone of the parathyroid cell or other cell receptor allows isolation of genes encoding similar inorganic-ion sensing proteins in other cells, and expression of those proteins. This is achieved by a variety of approaches. Southern blot analysis of human genomic DNA, utilizing the Ca^{2+} receptor cDNA as a hybridization probe, will give an indication of the number of related sequences encoded within the genome; hybridization at varying stringencies will give an indication of the degree of

divergence among the related sequences. This will provide information about the potential number of genes encoding related receptor proteins. Northern blot analysis with Ca^{2+} receptor cDNA as probe will determine if the same or related transcripts are present in various tissues. If related transcripts homologous to the parathyroid cell Ca^{2+} receptor are detected, it is a relatively simple matter to obtain clones of these mRNAs, either by screening the appropriate cDNA libraries or by polymerase chain reaction (PCR) techniques.

Targeted gene walking (TGW) is a modification of a standard polymerase chain reaction (PCR) that allows amplification of unknown DNA sequences adjacent to short segments of known sequence. Parker et al., Nucl. Acids Res., 19:3055, (1991). Unlike conventional PCR techniques that amplify DNA sequences between two known primer sites, TGW can amplify DNA adjacent to one such site. Thus, TGW can serve as a replacement for conventional cloning and library screening methods for isolating sequences upstream or downstream from known sequences. The procedure can be used to isolate genes from any starting DNA template for which a limited amount of sequence information is known.

First, several standard PCR reactions are run in parallel using one "targeted primer" and different "walking primers." The targeted primer is a sequence-specific primer exactly complementary to a known sequence on the DNA molecule of interest, and is directed towards unknown adjacent sequences. The walking primers are non-specific sequences not complementary to DNA near the target primer. The walking primers can be any oligonucleotides unrelated to

the target primer sequence. In the first series of PCRs, products are produced only when a walking primer anneals to a DNA strand contiguous with and complementary to the strand to which the targeted primer has hybridized. The PCR products of interest are preferably within the 5 kilobase size range. Amplification products are produced with as many as 60% mismatched nucleotides within the walking primer relative to DNA template. Perfect base-pairing is required only for the first two 3' nucleotides of the walking primer, but partial homology is tolerated otherwise. Annealing temperature is a key variable in determining the number of PCR products, as identified by agarose gel electrophoresis.

Second, an oligomer extension assay is performed using an "internal detection primer." This primer represents known sequences between the previous two primers, contiguous with the targeted primer. The internal detection primer is kinased with ^{32}P-gamma-ATP, then used in a single PCR cycle with DNA from the first PCR as template. This extension identifies products in the first PCR contiguous with the targeted primer. These new products are identified by agarose gel electrophoresis and autoradiography. Any products that do no hybridize to the internal detection primer represent non-contiguous amplification products produced by any subset of the primers.

Last, bands identified in the oligomer extension assay are excised from the gel, and reamplified by standard PCR using target primer and the walking primer that produced the band initially. This new PCR band is then sequenced directly to provide previously unknown sequence

information.

To extend information in the opposite direction, complements are made of the targeted and internal detection primers, and their order is reversed in the protocol.

In performing hybridization screening and cloning, the following should be considered. Since there is degeneracy in the genetic code, more than one codon exists for almost all the amino acids (except tryptophan and methionine). Moreover, the frequency of usage of any particular codon is different in non-humans as compared to humans. Taking into account codon degeneracy and human-preferred codons, oligonucleotides are synthesized. Moreover, oligonucleotides with various permutations of possible codons are also synthesized. To avoid an excessive number, not all possible sequences resulting from the degeneracy of the code need be synthesized. Rather, a subset of codons are chosen which have the highest frequency of occurrence.

Novel receptor clones so obtained can be assessed functionally by expression, either in oocytes or in transfected cell lines. Transfected cell lines expressing a cell-specific inorganic-ion receptor can then provide a means of high-throughput screening for molecules that act specifically on the ion-sensing mechanism of, for example, osteoclasts or juxtaglomerular cells.

In an alternative method, the calcium receptor can be cloned by expression in eukaryotic cells. For example, a cDNA library can be prepared from parathyroid mRNA and cloned into a eukaryotic expression vector such as, pCDNA1. Subpools from this library can be transfected into

eukaryotic cells such as COS7 or HEK293 cells resulting in relatively high-level transient expression of encoded cDNA sequences. Cells transfected with a function calcium receptor clone will express the calcium receptor which can then be activated by calcium, neomycin or other calcimimetic compounds. If cells are first loaded with a fluorometric indicator for $[Ca^{2+}]_i$, activation of the calcium receptor results in increased fluorescence. Thus library subpools containing the calcium receptor are identified by their ability, upon transfection into eukaryotic cells, to induce a calcium or calcimimetic-specific increase in fluorescence. This fluorescence can be detected using either a fluorimeter or a fluorescence-activated cell sorter (FACS).

We have also developed a "calcium trapping assay" for the detection of COS 7 cells expressing G-protein coupled receptors. In this assay COS 7 cell monolayers are transfected with cDNA clones from a bovine parathyroid cDNA library (e.g., subfractions or pools from a library prepared in pCDNA1) and are assayed for their ability to trap radioactive $^{45}Ca^{++}$ in response to treatment with an agonist for the Ca^{2+} receptor. The monolayers undergo emulsion autoradiography and cells that have trapped $^{45}Ca^{++}$ are identified by the presence of photographic grain clusters under dark-field microscopy. Library pools that produce a positive signal are then sequentially subdivided until a single cDNA that produces the signal is identified.

The calcium receptors appear to be functionally related to a class of receptors which utilize so-called "G"

proteins to couple ligand binding to intracellular signals. Such "G-coupled" receptors may elicit increases in intracellular cyclic AMP due to the stimulation of adenylyl cyclase by a receptor activated "G_s" protein, or else may elicit a decrease in cyclic AMP due to inhibition of adenylyl cyclase by a receptor activated "G_i" protein. Other receptor activated G proteins elicit changes in inositol trisphosphate levels resulting in release of Ca^{++} from intracellular stores. This latter mechanism is particularly pertinent to calcium receptors. All known G-coupled receptors are structurally related, having seven conserved transmembrane domains. A number of such receptors have been cloned based on sequence homology to previously cloned receptors. One particularly useful approach is to employ degenerate primers homologous to the conserved transmembrane domain coding regions and to amplify DNA regions encoding these sequences using polymerase chain reaction (PCR). Thus, such oligonucleotide primers are mixed with genomic DNA or cDNA to RNA isolated from the tissue of choice and PCR carried out. Some experimentation may be required to specifically amplify novel G-coupled receptor sequences from the tissue of choice since these are not necessarily identical to already known G-coupled receptors but, this is well understood by those skilled in the art [see for example, Buck, L. and Axel, R. (1991) Cell, 65:175-187].

Calcium receptors may also be cloned by such a PCR approach. Two examples of degenerate oligonucleotide primer pairs which can be used to PCR amplify sequences encoding calcium receptors are given below. The first pair is based

on the cross-homology exhibited by the majority of G-coupled receptors in the sequences encoding transmembrane domains II and VII respectively. The second primer pair is based on the cross-homology exhibited by a more divergent subgroup of G-coupled receptors which includes the calcitonin, secretin, PTH and GLP receptors. This pair corresponds to conserved sequences encoding transmembrane domains III and VII. When one or both of these two primer pairs are mixed with cDNA from parathyroid or osteoclast tissues, for example, PCR amplification results in the generation of amplified DNAs of about 500 to 800 base pairs. These DNAs can be isolated and analyzed for the presence of calcium receptor sequences. For example, each amplified sequence can be used as a probe to isolate a full-length cDNA clone which can then be assessed in one or more of the assays indicated above to determine if an inorganic-ion receptor is encoded.

PRIMER PAIR 1:

```
5'  GACTACTTCCTGGTGAACCTGGC  3'
    A   T    T  T C C   C
-------------------------------------------------
5'  TCAAIGAGCCIGTAGATGATGGGGTT  3'
     T C   AGG A  A AC CA  A
                   GC
```

PRIMER PAIR 2:

```
5' AACTACTICTGGITGCTGGTGGAGGGCCTCTA  3'
                C        A  GG G
                T
-----------------------------------------
5' GAAGCAGTAIATAATGGCAACAAAAAAGCCCTGGAA 3'
     A        G CA     G    GA
                       C     T
```

In an alternative method, the inorganic-ion receptor can be cloned by use of a monoclonal antibody

generated against the receptor. Monoclonal antibodies provide powerful tools for the immunoaffinity purification of specific proteins. Once purified, limited amino acid sequence data can be obtained from the protein of interest, and used to design oligonucleotide sequence probes to screen for clones of the complete cDNA sequence.

An example involving a calcium receptor will be described. For production of hybridomas, whole bovine parathyroid gland cells are used as the immunogen. Purified, dispersed cells are obtained, and live or fixed cell preparations are injected intraperitoneally into the appropriate mouse strain, according to established procedures. Standard protocols are followed for immunization schedules and for the production of hybridomas. A two-step screening procedure is used to identify hybridomas secreting monoclonal antibodies that recognize the Ca^{2+} receptor. The initial screen will identify those monoclonals that recognize parathyroid cell surface antigens. Immunohistochemical techniques are then used to screen hybridoma supernatants for the presence of mouse antibodies that bind to the surface of parathyroid cells. This screen can be performed on fixed sections of parathyroid gland tissue, or on dispersed cells in primary culture. The techniques for this assay are well established in the literature.

This screen will identify hybridomas producing monoclonal antibodies to a variety of cell-surface determinants, and monoclonals specific for the Ca^{2+} receptor would be expected to comprise only a small subset of these. To identify monoclonal antibodies that bind to

the Ca^{2+} receptor, hybridoma supernatants that test positive in the initial screen are assayed for their ability to block the response of cultured parathyroid cells to Ca^{2+} receptor agonists. Some antibodies that bind to the extracellular domain of the receptor are expected to inhibit or activate ligand binding or to otherwise interfere with or affect receptor activation.

Monoclonal antibodies positive in both screens are characterized through Western blotting, immunoprecipitation and immunohistochemistry. This permits the determination of the size of the antigen that is recognized and its tissue distribution. The appropriate monoclonal antibody is then used for purification of the Ca^{2+} receptor protein by immunoaffinity chromatography, following standard techniques. Sufficient quantities of protein are obtained to allow limited amino acid sequence determination. Degenerate oligonucleotide probes are then designed on the basis of the peptide sequence information. These probes are then used to screen parathyroid gland cDNA libraries for full-length clones of the Ca^{2+} receptor. Clones obtained are characterized by DNA sequencing and by functional expression in the oocyte system and in cultured mammalian cell lines.

Alternatively, the antibodies can be used to screen expression libraries, e.g., cDNA libraries in λgt11 or its equivalent, to determine those clones expressing antigenically reactive protein. Such clones can then be sequenced to determine whether they encode a protein that might be a Ca^{2+} receptor.

It will also be appreciated by those skilled in

the art that phage display libraries can be used to clone and analyze calcium receptors in place of monoclonal antibodies. In these libraries, antibody-variable regions or random peptides are shotgun cloned into phage expression vectors such that the antibody regions or peptides are displayed on the surface of the phage particle. Phage(s) which display antibody regions or peptides capable of high specific binding to calcium receptors will bind to cells which display these receptors (e.g., parathyroid cells, C-cells, osteoclasts, etc.). Hundreds of millions of such phage can be panned against these cell types preferentially selecting those phage which can bind to these cells (which includes those phage binding to calcium receptors). In this manner, the complexity of the library can be vastly reduced. Iterative repetition of this process results in a pool of phage which bind to the cell type used. Subsequently, the screens described above for monoclonal antibodies can be used to isolate phage which display calcium receptor-binding antibody or peptide regions, and these phage can be used to isolate the calcium receptor for purposes of structural identification and cloning. Kits to prepare such phage- display libraries are commercially available (e.g., Stratacyte, or Cambridge Antibody Technology Limited). Recombinant phage endowed with such calcium receptor-binding properties can also be used in lieu of monoclonal antibodies in the various analyses of calcium receptors. Such phage can also be used in high throughput binding-competition screens to identify organic compounds capable of functional binding to calcium receptors which can serve as structural leads for the development of human

therapeutics acting at the calcium receptor.

In another alternative, affinity cross-linking of radioligands to their receptors can be used to isolate the receptor protein as described by Pilch & Czech, 1 Receptor Biochem. Methodol. 161, 1984. Covalent attachment of a radioligand allows extensive washing to remove non-specific binding. For example, a high- affinity molecule, e.g., a random copolymer of arginine and tyrosine (MW = 22K; argtyr ratio = 4:1) which mobilizes intracellular Ca^{2+} with an EC_{50} of about 100 nM or less, is iodinated with ^{125}I, and cross-linked. Protamines, because of their much smaller size, may be preferable in cross-linking studies and can be reductively alkylated as described by Dottavio-Martin & Ravel, 87 Analyt. Biochem. 562, 1978.

Nonspecific labelling is kept to a minimum by cross-linking in the presence of unlabeled polycations and di- and trivalent cations. At high concentrations of these molecules nonspecific interactions of the label with the cell surface might be reduced.

The invention provides isolated nucleic acid sequences encoding inorganic-ion receptors and the isolated receptors themselves. It also provides unique fragments of the foregoing. The term "isolated" refers to a nucleic acid sequence: (i) amplified in vitro by, for example, polymerase chain reaction (PCR); (ii) synthesized by, for example, chemical synthesis; (iii) recombinantly produced by cloning; or (iv) purified, as by cleavage and gel separation. The term "isolated" when used in descriptions of polypeptide or amino acid sequences refers to polypeptides resulting from an expression system using the isolated nucleic acid

sequences of the invention, as well as polypeptides synthesized by, for example, chemical synthetic methods, and polypeptides separated from native biological materials, and then substantially purified using conventional protein analytical procedures.

Also included within the scope of the invention are unique fragments of inorganic-ion receptors. The term unique fragments refers to portions of the receptor that find no counterpart in known sequences as of the date of this invention. These polypeptide fragments can be readily identified as unique by scanning protein databases known as of the time of the invention for identical peptides. In addition to generating fragments of receptors from expression of cloned partial sequences of receptor DNA, fragments can be generated directly from the intact protein. Proteins are specifically cleaved by proteolytic enzymes, including, but not limited to, trypsin, chymotrypsin or pepsin. Each of these enzymes is specific for the type of peptide bond it attacks. Trypsin catalyzes the hydrolysis of peptide bonds whose carbonyl group is from a basic amino acid, usually arginine or lysine. Pepsin and chymotrypsin catalyze the hydrolysis of peptide bonds from aromatic amino acids, particularly tryptophan, tyrosine and phenylalanine. Alternate sets of cleaved polypeptide fragments are generated by preventing cleavage at a site which is susceptible to a proteolytic enzyme. For example, reaction of the ε-amino groups of lysine with ethyltrifluorothioacetate in mildly basic solution yields a blocked amino acid residue whose adjacent peptide bond is no longer susceptible to hydrolysis by trypsin. Goldberger et

al. Biochem., 1:401 (1962). Treatment of such a polypeptide with trypsin thus cleaves only at the arginyl residues. Polypeptides also can be modified to create peptide linkages that are susceptible to proteolytic enzyme catalyzed hydrolysis. For example, alkylation of cysteine residues with ß-haloethylamines yields peptide linkages that are hydrolyzed by trypsin. Lindley, Nature, 178: 647 (1956). In addition, chemical reagents that cleave polypeptide chains at specific residues can be used. Witcop, Adv. Protein Chem. 16: 221 (1961). For example, cyanogen bromide cleaves polypeptides at methionine residues. Gross & Witkip, J. Am Chem Soc., 83: 1510 (1961). Thus, by treating an inorganic ion receptor, such as, for example, a human calcium receptor or fragments thereof, with various combinations of modifiers, proteolytic enzymes and/or chemical reagents, numerous discrete overlapping peptides of varying sizes are generated. These peptide fragments can be isolated and purified from such digests by chromatographic methods.

Alternatively, fragments can be synthesized using an appropriate solid state synthetic procedure. Steward and Young, Solid Phase Peptide Synthesis, Freemantle, San Fansisco, CA (1968).

The fragments may be used in assays, as ion-binding agents, in the production of antibodies and the like. The fragments also may be selected to have desirable biological activities. For example, the fragment may include just the binding site, or a site which binds to agonists or antagonists (e.g. of calcium), as described herein. Such fragments are readily identified by

those of ordinary skill in the art using routine methods to detect specific binding to the fragment. For example, in the case of a calcium receptor, a fragment to be tested can be expressed, using recombinant DNA methodology, from a fragment of the gene encoding the recombinant receptor. This fragment is then contacted with calcium or another chemical under appropriate association conditions to determine whether the calcium binds to the fragment. These fragments are useful in screening assays for agonists and antagonists of calcium, and for therapeutic effect where it is useful to remove calcium from serum, or other bodily tissues.

Other useful fragments include those that have only the external portion, membrane-spanning portion, or intracellular portion of the receptor. These portions are readily identified by comparison of the amino acid sequence of the receptor with those of known receptors, or by other standard methodology. These fragments are useful for forming chimeric receptors with fragments of other receptors so that a cell lacking a receptor may be formed with an intracellular portion which performs a desired function within that cell, and an extracellular portion which causes that cell to respond to the presence of ions, or those agonists or antagonists described herein. Such chimeric receptor genes when appropriately formulated are useful genetic therapies for a variety of diseases involving dysfunction of receptors or where modulation of receptor function provides a desirable effect in the patient. In addition, chimeric receptors can be constructed such that the intracellular domain is coupled to a desired enzymatic

process which can be readily detected by colorometric, radiometric luminometric, spectrophotometric or fluorometric assays and is activated by interaction of the extracellular portion with its native ligand (e.g. calcium) or agonists and or antagonists of the invention. Cells expressing such chimeric receptors are the basis for facilitated screens for the discovery of new agonists and/or antagonists of the invention.

The invention also features muteins or analogs and other derivatives of isolated receptors, as described herein. Thus, the invention embraces not only naturally occurring proteins, but also such derivatives. Such derivatives have the desired receptor activity described herein and are generally identified by methods described herein. While examples of such proteins, and genes encoding them, are provided, these examples are not limiting in the invention but demonstrate only the variation that can be associated with such genes and proteins. Generally, the gene will have the amino acid sequence of the native receptor, but may be altered at one or more amino acid positions which do not significantly affect the receptor activity of the produced receptor protein. Thus, for example, in non-conserved regions of the calcium receptor protein (i.e., those regions not necessary for receptor activity) amino acids may be deleted, added or substituted. In more conserved regions, which are required for receptor activity, amino acids may be more conservatively substituted. For example, one or more amino acid residues within the sequence can be substituted by another amino acid of a similar polarity which acts as a functional equivalent. Substitutes for an amino acid within the

sequence may be selected from other members of the class to which the amino acid belongs. The non-polar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan and methionine. The polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine. The positively charged (basic) amino acids include arginine, lysine and histidine. The negatively charged (acidic) amino acids include asparatic acid and glutamic acid.

Those of ordinary skill in the art will recognize that standard procedures can be used to determine the conserved and non-conserved regions of the protein using in vitro mutagenesis techniques or deletion analyses, and that all such derivatives are equivalent of those described below.

Also included within the scope of the invention are receptor proteins or unique fragments or derivatives thereof which are differentially modified during or after translation, e.g., by phosphorylation, glycosylation, crosslinking, acylation, proteolytic cleavage, linkage to an antibody molecule, membrane molecule or other ligand, (Ferguson et al., 1988, Ann. Rev. Biochem. 57:285-320).

In addition, recombinant nucleic acid sequences of the invention may be engineered so as to modify processing or expression of receptor sequences. For example, and not by way of limitation, the coding sequence may be combined with a promoter sequence and/or a ribosome binding site using well characterized methods, and thereby facilitate improve expression and bioavailability.

Additionally, a given recombinant coding

sequence can be mutated in vitro or in vivo, to create variations in coding regions and/or form new restriction endonuclease sites or destroy preexisting ones, to facilitate further in vitro modification. Any technique for mutagenesis known in the art can be used including, but not limited to, in vitro site-directed mutagenesis (Hutchinson, et al., 1978, J. Biol. Chem. 253:6551), use of TAB® linkers (Pharmacia), PCR-directed mutagenesis, and the like.

In addition, codons may be modified such that while they encode an identical amino acid, that codon may be a preferred codon in the chosen expression system.

The invention also provides unique fragments of nucleic acid encoding an inorganic-ion receptor. The term "unique fragments" refers to portions of the nucleic acid that find no identical counterpart in known sequences as of the date of this invention. These fragments can be identified easily by an analysis of nucleic acid data bases existing as of the date of the invention to detect counterparts. Unique fragments are useful inter alia in cloning procedures and assays.

The invention further provides receptor binding agents including antibodies and/or fragments thereof which can be conjugated to a toxin moiety, or expressed along with a toxin moiety as a recombinant fusion protein. The toxin moiety will bind to and enter a target cell using interaction of the binding agent and the corresponding target cell surface receptor. The toxic moiety to which the agent, antibody and/or fragment is conjugated can be a protein such as, for example, pokeweed anti-viral protein, ricin, gelonin, abrin, diphtheria exotoxin, or Pseudomonas

exotoxin. The toxin moiety can also be a high energy-emitting radionuclide such as cobalt-60. The chemical structure of the toxin moiety is not intended to limit the scope of the invention in any way. Those of ordinary skill in the art will recognize that a large variety of possible moieties can be linked to the binding agents. See, for example, "Conjugate Vaccines", Contributions to Microbiology and Immunology, J.M. Cruse and R.E. Lewis, Jr (eds). Carger Press, New York, (1989). It further is art recognized that many toxin moieties, including those specifically listed above, are considered pharmaceutically acceptable.

The conjugation of the binding agent to another moiety (e.g. bacterial toxin) can be accomplished by any chemical reaction that will bind the two molecules so long as both molecules retain their respective activity. This linkage can include many chemical mechanisms, for instance covalent binding, affinity binding, intercalation, coordinate binding and complexation. The preferred binding is, however, covalent binding. The covalent binding can be achieved either by direct condensation of existing side chains or by the incorporation of external bridging molecules. Many bivalent or polyvalent linking agents are useful in coupling protein molecules, such as an antibody, to other molecules. For example, representative coupling agents can include organic compounds such as thioesters, carbodiimides, succinimide esters, diisocyanates, glutaraldehydes, diazobenzenes and hexamethylene diamines. This listing is not intended to be exhaustive of the various classes of coupling agents known in the art but, rather, is

exemplary of the more common coupling agents. (See Killen and Lindstrom 1984, "Specific killing of lymphocytes that cause experimental Autoimmune Myasthenia Gravis by toxin-acetylcholine receptor conjugates." Jour. Immun. 133:1335-2549; Jansen, F.K., H.E. Blythman, D. Carriere, P. Casella, O. Gros, P. Gros, J.C. Laurent, F. Paolucci, B. Pau, P. Poncelet, G. Richer, H. Vidal, and G.A. Voisin. 1982. "Immunotoxins: Hybrid molecules combining high specificity and potent cytotoxicity". Immunological Reviews 62:185-216; and Vitetta et al., supra).

Using recombinant DNA techniques, the present invention provides for target cells including mammalian target cells which are engineered to express inorganic-ion receptors. For example, the genes for such receptors, cloned according to the methods set forth above, may be inserted into cells which naturally express the receptors such that the recombinant gene is expressed at much higher levels.

The present invention also provides for experimental model systems for studying the physiological role of the receptors. In these model systems, the inorganic-ion receptors, fragments, or derivatives thereof, may be either supplied to the system or produced within the system. Such model systems could be used to study the effects or cell function of receptor excess or depletion. The experimental model systems may be used to study the effects in cell or tissue cultures, in whole animals, or in particular cells or tissues within whole animals or tissue culture systems, or over specified time intervals (including during embryogenesis). A preferred embodiment is assays involving the cloned receptors or parts thereof, including

particularly those useful in high through-put drug screening assays.

In additional embodiments of the invention, a recombinant gene may be used to inactivate the endogenous gene by homologous recombination, and thereby create an inorganic-ion receptor deficient cell, tissue, or animal. For example, and not by way of limitation, a recombinant gene may be engineered to contain an insertional mutation (e.g.. the neo gene) which, when inserted into the genome of a recipient cell, tissue or animal, inactivates transcription of the receptor. Such a construct may be introduced into a cell, such as an embryonic stem cell, by a technique such as transfection, transduction, injection, etc. Stem cells lacking an intact receptor sequence may generate transgenic animals deficient in the receptor.

A "transgenic animal" is an animal having cells that contain DNA which has been artificially inserted into a cell, which DNA becomes part of the genome of the animal which develops from that cell. Preferred transgenic animals are primates, mice, rats, cows, pigs, horses, goats, sheep, dogs and cats. The transgenic DNA may encode for human inorganic-ion receptors. In a further embodiment of the invention, native expression in an animal may be reduced by providing an amount of anti-sense RNA or DNA effective to reduce expression of the receptor.

A variety of methods are available for the production of transgenic animals associated with this invention. DNA can be injected into the pronucleus of a fertilized egg before fusion of the male and female pronuclei, or injected into the nucleus of an embryonic cell

(e.g., the nucleus of a two-cell embryo) following the initiation of cell division (Brinster et al., Proc. Nat. Acad. Sci. USA, 82: 4438-4442 (1985)). Embryos can be infected with viruses, especially retroviruses, modified to carry inorganic-ion receptor nucleotide sequences of the invention.

Pluripotent stem cells derived from the inner cell mass of the embryo and stabilized in culture can be manipulated in culture to incorporate nucleotide sequences of the invention. A transgenic animal can be produced from such cells through implantation into a blastocyst that is implanted into a foster mother and allowed to come to term.

Animals suitable for transgenic experiments can be obtained from standard commercial sources such as Charles River (Wilmington, MA), Taconic (Germantown, NY), Harlan Sprague Dawley (Indianapolis, IN), etc.

The procedures for manipulation of the rodent embryo and for microinjection of DNA into the pronucleus of the zygote are well known to those of ordinary skill in the art (Hogan et al., supra). Microinjection procedures for fish, amphibian eggs and birds are detailed in Houdebine and Chourrout, Experientia, 47: 897-905 (1991). Other procedures for introduction of DNA into tissues of animals are described in U.S. Patent No., 4,945,050 (Sandford et al., July 30, 1990).

By way of example only, to prepare a transgenic mouse, female mice are induced to superovulate. Females are placed with males, and the mated females are sacrificed by CO_2 asphyxiation or cervical dislocation and embryos are recovered from excised oviducts. Surrounding cumulus cells

are removed. Pronuclear embryos are then washed and stored until the time of injection. Randomly cycling adult female mice are paired with vasectomized males. Recipient females are mated at the same time as donor females. Embryos then are transferred surgically.

The procedure for generating transgenic rats is similar to that of mice. See Hammer et al., Cell, 63:1099-1112 (1990).

Methods for the culturing of embryonic stem (ES) cells and the subsequent production of transgenic animals by the introduction of DNA into ES cells using methods such as electroporation, calcium phosphate/DNA precipitation and direct injection also are well known to those of ordinary skill in the art. See, for example, Teratocarcinomas and Embryonic Stem Cells, A Practical Approach, E.J. Robertson, ed., IRL Press (1987).

In cases involving random gene integration, a clone containing the sequence(s) of the invention is co-transfected with a gene encoding resistance. Alternatively, the gene encoding neomycin resistance is physically linked to the sequence(s) of the invention. Transfection and isolation of desired clones are carried out by any one of several methods well known to those of ordinary skill in the art (E.J. Robertson, supra).

DNA molecules introduced into ES cells can also be integrated into the chromosome through the process of homologous recombination. Capecchi, Science, 244: 1288-1292 (1989). Methods for positive selection of the recombination event (i.e., neo resistance) and dual positive-negative selection (i.e., neo resistance and gancyclovir resistance)

and the subsequent identification of the desired clones by PCR have been described by Capecchi, supra and Joyner et al., Nature, 338: 153-156 (1989), the teachings of which are incorporated herein. The final phase of the procedure is to inject targeted ES cells into blastocysts and to transfer the blastocysts into pseudopregnant females. The resulting chimeric animals are bred and the offspring are analyzed by Southern blotting to identify individuals that carry the transgene.

Procedures for the production of non-rodent mammals and other animals have been discussed by others. See Houdebine and Chourrout, supra; Pursel et al., Science 244: 1281-1288 (1989); and Simms et al., Bio/Technology, 6: 179-183 (1988).

Uses

Primary hyperparathyroidism (HPT) is characterized by hypercalcemia and elevated levels of circulating PTH. One of the major defects in HPT appears to be a diminished sensitivity of parathyroid cells to negative feedback regulation by extracellular Ca^{2+}. Thus, in tissue from patients with primary HPT, the "set-point" for extracellular Ca^{2+} is shifted to the right so that higher than normal concentrations of extracellular Ca^{2+} are required to depress PTH secretion. Moreover, in primary HPT, even high concentrations of extracellular Ca^{2+} often depress PTH secretion only partially. In secondary (uremic) HPT, a similar increase in the set-point for extracellular Ca^{2+} is observed even though the degree to which Ca^{2+} suppresses PTH secretion is normal. The changes in PTH secretion are paralleled by changes in $[Ca^{2+}]_i$: the

set-point for extracellular Ca^{2+}-induced increases in $[Ca^{2+}]_i$ is shifted to the right and the magnitude of such increases is reduced. Moreover, staining of tissue with a monoclonal antibody that appears to recognize the Ca^{2+} receptor is diminished in adenomatous and hyperplastic parathyroid cells.

The Ca^{2+} receptor constitutes a discrete molecular entity for pharmacological intervention. Molecules that mimic or antagonize the action of extracellular Ca^{2+} are beneficial in the long-term management of both primary and secondary HPT. Such molecules provide the added impetus required to suppress PTH secretion which the hypercalcemic condition alone cannot achieve. Such molecules with greater efficacy than extracellular Ca^{2+} may overcome the apparent nonsuppressible component of PTH secretion which is particularly troublesome in adenomatous tissue. Alternatively or additionally, such molecules can depress synthesis of PTH, as prolonged hypercalcemia has been shown to depress the levels of preproPTH mRNA in bovine and human adenomatous parathyroid tissue. Prolonged hypercalcemia also depresses parathyroid cell proliferation in vitro, so calcimimetics can also be effective in limiting the parathyroid cell hyperplasia characteristic of secondary HPT.

Other cells in the body can respond directly to physiological changes in the concentration of extracellular Ca^{2+}. Calcitonin secretion from parafollicular cells in the thyroid (C-cells) is regulated by changes in the concentration of extracellular Ca^{2+}. Renin secretion from juxtaglomerular cells in the kidney, like PTH secretion, is

depressed by increased concentrations of extracellular Ca^{2+}. Extracellular Ca^{2+} causes the mobilization of intracellular Ca^{2+} in these cells. Other kidney cells respond to calcium as follows: elevated Ca^{2+} inhibits formation of $1,25(OH)_2$ Vitamin D by proximal tubular cells, stimulates production of calcium-binding protein in distal tubular cells, and inhibits tubular reabsorption of Ca^{2+} and Mg^{2+} and the action of vasopressin on the medullary thick ascending limb of Henle's loop (MTAL), reduces vasopressin action in the cortical collecting duct cells, and affects vascular smooth muscle cells in blood vessels of the renal glomerulus. Calcium promotes the differentiation of intestinal goblet cells, mammary cells, and skin cells. It also inhibits atrial natriuretic peptide secretion from cardiac atria, reduces cAMP accumulation in platelets, alters gastrin and glucagon secretion, and acts on vascular smooth muscle cells to modify cell secretion of vasoactive factors. Isolated osteoclasts respond to increases in the concentration of extracellular Ca^{2+} with corresponding increases in $[Ca^{2+}]_i$ that arise partly from the mobilization of intracellular Ca^{2+}. Increases in $[Ca^{2+}]_i$ in osteoclasts are associated with an inhibition of functional responses (bone resorption) analogous to PTH secretion in parathyroid cells. Release of alkaline phosphatase from bone-forming osteoblasts is directly stimulated by calcium. Thus, there are sufficient indications to suggest that Ca^{2+}, in addition to its ubiquitous role as an intracellular signal, also functions as an extracellular signal to regulate the responses of certain specialized cells. Molecules of this invention can

be used in the treatment of diseases associated with disrupted Ca^{2+} responses in these cells.

Cloning the Ca^{2+} receptor on parathyroid cells and other cells will allow the presence of homologous proteins in other cells to be directly assessed. A family of structurally homologous Ca^{2+} receptor proteins can thus be obtained. Such receptors will allow understanding of how these cells detect extracellular Ca^{2+} and enable evaluation of the mechanism(s) as a site of action for the therapeutics described herein effective in the treatment of HPT, osteoporosis, and hypertension, and novel therapies for other bone and mineral-related diseases.

Other uses are discussed above. For example, recombinant Ca^{2+} receptor proteins may be used in therapy, and introduced by standard methods, e.g., by transfection of nucleic acid encoding that protein. In addition, such protein is useful in assays for calcimimetic molecules of this invention.

The following examples illustrate the invention but do not limit its scope.

Methods for the culturing of embryonic stem (ES) cells and the subsequent production of transgenic animals by the introduction of DNA into ES cells using methods such as electroporation, calcium phosphate/DNA precipitation and direct injection also are well known to those of ordinary skill in the art. See, for example, Teratocarcinomas and Embryonic Stem Cells, A Practical Approach, E.J. Robertson, ed., IRL Press (1987).

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The Ca^{2+} receptor constitutes a discrete molecular entity for pharmacological intervention. Molecules that mimic or antagonize the action of extracellular Ca^{2+} are beneficial in the long-term management of both primary and secondary HPT. Such molecules provide the added impetus required to suppress PTH secretion which the hypercalcemic condition alone cannot achieve. Such molecules with greater efficacy than extracellular Ca^{2+} may overcome the apparent nonsuppressible component of PTH secretion which is particularly troublesome in adenomatous tissue. Alternatively or additionally, such molecules can depress

synthesis of PTH, as prolonged hypercalcemia has been shown to depress the levels of preproPTH mRNA in bovine and human adenomatous parathyroid tissue. Prolonged hypercalcemia also depresses parathyroid cell proliferation in vitro, so calcimimetics can also be effective in limiting the parathyroid cell hyperplasia characteristic of secondary HPT.

Other cells in the body can respond directly to physiological changes in the concentration of extracellular Ca^{2+}. Calcitonin secretion from parafollicular cells in the thyroid (C-cells) is regulated by changes in the concentration of extracellular Ca^{2+}. Renin secretion from juxtaglomerular cells in the kidney, like PTH secretion, is depressed by increased concentrations of extracellular Ca^{2+}. Extracellular Ca^{2+} causes the mobilization of intracellular Ca^{2+} in these cells. Other kidney cells respond to calcium as follows: elevated Ca^{2+} inhibits formation of $1,25(OH)_2$ Vitamin D by proximal tubular cells, stimulates production of calcium-binding protein in distal tubular cells, and inhibits tubular reabsorption of Ca^{2+} and Mg^{2+} and the action of vasopressin on the medullary thick ascending limb of Henle's loop (MTAL), reduces vasopressin action in the cortical collecting duct cells, and affects vascular smooth muscle cells in blood vessels of the renal glomerulus. Calcium promotes the differentiation of intestinal goblet cells, mammary cells, and skin cells. It also inhibits atrial natriuretic peptide secretion from cardiac atria, reduces cAMP accumulation in platelets, alters gastrin and glucagon secretion, and acts on vascular smooth muscle cells to modify cell secretion of vasoactive factors. Isolated osteoclasts respond to

increases in the concentration of extracellular Ca^{2+} with corresponding increases in $[Ca^{2+}]_i$ that arise partly from the mobilization of intracellular Ca^{2+}. Increases in $[Ca^{2+}]_i$ in osteoclasts are associated with an inhibition of functional responses (bone resorption) analogous to PTH secretion in parathyroid cells. Release of alkaline phosphatase from bone-forming osteoblasts is directly stimulated by calcium. Thus, there are sufficient indications to suggest that Ca^{2+}, in addition to its ubiquitous role as an intracellular signal, also functions as an extracellular signal to regulate the responses of certain specialized cells. Molecules of this invention can be used in the treatment of diseases associated with disrupted Ca^{2+} responses in these cells.

Cloning the Ca^{2+} receptor on parathyroid cells and other cells will allow the presence of homologous proteins in other cells to be directly assessed. A family of structurally homologous Ca^{2+} receptor proteins can thus be obtained. Such receptors will allow understanding of how these cells detect extracellular Ca^{2+} and enable evaluation of the mechanism(s) as a site of action for the therapeutics described herein effective in the treatment of HPT, osteoporosis, and hypertension, and novel therapies for other bone and mineral-related diseases.

Other uses are discussed above. For example, recombinant Ca^{2+} receptor proteins may be used in therapy, and introduced by standard methods, e.g., by transfection of nucleic acid encoding that protein. In addition, such protein is useful in assays for calcimimetic molecules of this invention.

The following examples illustrate the invention but do not limit its scope.

Examples

In the studies described herein, a variety of organic molecules were found to mobilize intracellular Ca^{2+} and depress PTH secretion in parathyroid cells. These molecules are structurally diverse but most have a net positive charge at physiological pH. The cationic nature of the organic molecules plays an important role but is not the sole factor determining activity.

Example 1: Screening Calcimimetic Molecules on Bovine Parathyroid cells

Dissociated bovine parathyroid cells were purified on gradients of Percoll and cultured overnight in serum- free medium. The cells were subsequently loaded with fura-2 and the concentration of free intracellular Ca^{2+} measured fluorimetricly. Changes in $[Ca^{2+}]_i$ were used to screen for molecules active at the Ca^{2+} receptor. To be considered a calcimimetic, a molecule was required to show the normal effects caused by increasing extracellular Ca^{2+} and triggered by the activation of the Ca^{2+} receptor. That is,

1) The molecule must elicit an increase in $[Ca^{2+}]_i$; the increase in $[Ca^{2+}]_i$ may persist in the absence of extracellular Ca^{2+} and/or the molecule may potentiate increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}.

2) The molecule must cause a decrease in isoproterenol-stimulated cyclic AMP formation which is blocked by pertussis toxin;

3) The molecule must inhibit PTH secretion over the same range of concentrations that cause the increase in $[Ca^{2+}]_i$; and

4) The concentration-response curves for increases in $[Ca^{2+}]_i$ and PTH secretion by the molecule must be shifted to the right by a PKC activator, such as phorbol myristate acetate (PMA).

Several structurally different classes of molecules were tested: polyamines, aminoglycoside antibiotics, protamine, and polymers of lysine or arginine. The structures of these molecules are depicted in Figure 1. Included in Figure 1 are the net positive charge of the molecules and their EC_{50}'s for evoking the mobilization of intracellular Ca^{2+} in bovine parathyroid cells.

In general, the greater the net positive charge on the molecule, the greater its potency in causing the mobilization of intracellular Ca^{2+}. However, some striking exceptions to this apparent rule have been found as discussed below.

As can be seen from the figures, spermine, neomycin B, and protamine evoked rapid and transient increases in $[Ca^{2+}]_i$ in fura-2-loaded bovine parathyroid cells (Figs. 6, 7, 11). They did not, however, cause sustained, steady-state increases in $[Ca^{2+}]_i$ in bovine parathyroid cells (Fig. 6, 11), although they did in human parathyroid cells (Fig. 19). In this respect, they resembled the cytosolic Ca^{2+} response elicited by extracellular Mg^{2+}, which causes the mobilization of intracellular Ca^{2+} unaccompanied by an influx of extracellular Ca^{2+} in bovine cells (Fig. 11b). Transient

increases in $[Ca^{2+}]_i$ elicited by spermine, neomycin B, or protamine were not blocked by low concentrations (1 μM) of La^{3+} or Gd^{3+} (Fig. 11f,g). Cytosolic Ca^{2+} transients elicited by the molecular polycations persisted in the absence of extracellular Ca^{2+} but were blocked when cellular stores of Ca^{2+} were depleted by pretreatment with ionomycin (Figs. 7; 11h,i). All these molecules therefore cause the mobilization of intracellular Ca^{2+} in parathyroid cells.

It was additionally shown that the molecular polycations mobilized the same pool of intracellular Ca^{2+} as that used by extracellular Ca^{2+}. Thus, increasing the concentration of extracellular Ca^{2+} progressively inhibited the transient increases in $[Ca^{2+}]_i$ evoked by spermine (Fig. 6). Conversely, a maximally effective concentration of spermine or neomycin B (Fig. 12) blocked transient, but not steady-state increases in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+}.

Significantly, spermine, neomycin B, and protamine inhibited PTH secretion to the same extent as extracellular Ca^{2+}. These inhibitory effects on secretion were obtained at concentrations that caused the mobilization of intracellular Ca^{2+} (Figs. 8, 13). These findings are relevant to understanding the mechanisms contributing to the regulation of PTH secretion by extracellular Ca^{2+}. Because a variety of inorganic polycations all inhibit secretion, yet only extracellular Ca^{2+} causes sustained, steady-state increases in $[Ca^{2+}]_i$, such increases in $[Ca^{2+}]_i$ cannot be importantly involved in the regulation of secretion. Mobilization of intracellular Ca^{2+}, rather

than the influx of extracellular Ca^{2+}, is the essential mechanism associated with the inhibition of PTH secretion. This is important because it defines the sufficient mechanism to be affected if a molecule is to affect PTH secretion; molecules stimulating selectively the influx of extracellular Ca^{2+} will be relatively ineffective in suppressing PTH secretion. In contrast, molecules causing solely the mobilization of intracellular Ca^{2+} should be just as efficacious as extracellular Ca^{2+} in suppressing PTH secretion.

Like the mobilization of intracellular Ca^{2+} elicited by extracellular Ca^{2+}, that elicited by molecular polycations was depressed by PMA. A representative experiment showing the preferential inhibitory effects of PMA on cytosolic Ca^{2+} transients elicited by spermine is shown in Fig. 14. Cytosolic Ca^{2+} transients evoked by ATP were unaffected, even when a submaximal concentration of ATP was used. The effect of PMA on cytosolic Ca^{2+} transients elicited by the molecular polycations paralleled its effect on responses to extracellular Ca^{2+}; in both cases there was a shift to the right in the concentration-response curve (Fig. 15). The depressive effects of PMA on $[Ca^{2+}]_i$ were accompanied by potentiating effects on secretion which were overcome at higher concentrations of the organic polycations (Fig. 16).

The mobilization of intracellular Ca^{2+} elicited by molecular polycations was associated with increases in the formation of inositol phosphates. For example, protamine caused a rapid (within 30 s) increase in the formation of IP_3 which was accompanied by a rise in

levels of IP_1. Both these effects were dependent on the concentration of extracellular protamine (Fig. 17). Moreover, pretreatment with PMA blunted the formation of inositol phosphates elicited by molecular polycations. Representative results obtained with spermine are presented in Fig. 18.

Spermine, neomycin B, and protamine depressed isoproterenol-induced increases in cyclic AMP. Like the inhibitory effects of extracellular Ca^{2+} on cyclic AMP formation, those caused by molecular polycations were blocked by pertussis toxin (Table 2).

Table 2

	cyclic AMP (% of control)	
	control	+PTx
0.5 mM Ca^{2+}	100	106 ± 8
2.0 mM Ca^{2+}	19 ± 4	94 ± 2
0.5 mM Ca^{2+}, 200 μM Spermine	23 ± 5	93 ± 6
0.5 mM Ca^{2+}, 30 μM Neomycin B	28 ± 8	87 ± 6
0.5 mM Ca^{2+}, 2 μg/ml Protamine	20 ± 4	89 ± 9

Pertussis toxin (PTx) blocks the inhibitory effects of extracellular Ca^{2+} and molecular polycations on cyclic AMP formation. Bovine parathyroid cells were cultured for 16 h with or without 100 ng/ml pertussis toxin. The cells were subsequently washed and incubated for 15 min with 10 μM isoproterenol with or without the indicated concentrations of extracellular Ca^{2+} or molecular polycations. Total cyclic AMP (cells + supernatant) was determined by RIA and the results are expressed as a percentage of the levels

obtained in 0.5 mM Ca^{2+} (112 ± 17 pmole/10^6 cells). Each value is the mean ± SEM of three experiments.

In human parathyroid cells, extracellular Mg^{2+} elicited a sustained, steady-state increase in $[Ca^{2+}]_i$ in addition to a rapid transient increase (Fig. 10). As in bovine parathyroid cells responding to extracellular Ca^{2+}, the steady-state increase in $[Ca^{2+}]_i$ evoked by Mg^{2+} in human parathyroid cells results from Ca^{2+} influx through voltage-insensitive channels (Fig. 10a). This effect of Mg^{2+} on steady-state $[Ca^{2+}]_i$ in human parathyroid cells is seen in both adenomatous and hyperplastic tissue.

Neomycin B and spermine were tested for effects on $[Ca^{2+}]_i$ in human parathyroid cells prepared from adenomatous tissue. Representative results with neomycin B are shown in Fig. 19. Neomycin B caused not only a transient but additionally a steady-state increase in $[Ca^{2+}]_i$ in human parathyroid cells (Fig. 19a). Thus, in human cells, the pattern of change in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+}, Mg^{2+} or neomycin B is very similar.

Cytosolic Ca^{2+} transients elicited by neomycin B persisted in the presence of La^{3+} (1μM) and absence of extracellular Ca^{2+}. Neomycin B therefore causes the mobilization of intracellular Ca^{2+} in human parathyroid cells. Neomycin B inhibited PTH secretion from human parathyroid cells at concentrations that caused the mobilization of intracellular Ca^{2+} (Fig. 13). There were, however, some differences in the responses of human and bovine parathyroid cells to neomycin B. The EC_{50} of neomycin B for the mobilization of intracellular Ca^{2+} was 40 μM in bovine and 20 μM in human parathyroid cells

(cf. Figs. 13 and 15), whereas the potency of spermine was similar in bovine and human parathyroid cells (EC_{50} = 150 μM). Thus, although bovine cells can be used for initial studies to screen test molecules for activity, it is important to perform follow-up studies using human parathyroid cells.

To assess the effects of molecular polycations on C-cells, a neoplastic cell line, derived from a rat medullary thyroid carcinoma (rMTC 6-23 cells) was used. Both spermine (10 mM) and neomycin B (5 mM) were without effect on basal $[Ca^{2+}]_i$ in these cells. Nor did either molecule affect the response to the subsequent addition of extracellular Ca^{2+}. Representative results documenting the lack of effect of neomycin B are shown in Fig 21. Neomycin B (1 mM) or spermine (1 or 5 mM) failed to evoke any increase in $[Ca^{2+}]_i$ in osteoclasts (Fig. 23). In the trace shown, there appeared to be some potentiation of the response to a subsequent increase in the concentration of extracellular Ca^{2+}, although this was not a consistent finding. In two other cells, spermine (5 mM) was again without effect on basal $[Ca^{2+}]_i$ and caused a small inhibition (about 15%) of the extracellular Ca^{2+}-induced increase in $[Ca^{2+}]_i$. In a third cell, neomycin B (5 mM) was without effect on basal $[Ca^{2+}]_i$ and did not affect increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. The overall picture that develops from these studies is that spermine and neo mycin B are without effect on basal or stimulated levels of cytosolic Ca^{2+} in osteoclasts.

The failure of the molecular polycations to affect the Ca^{2+}-sensing mechanisms of C-cells or

osteoclasts demonstrates the ability to discover or design novel lead molecules that act specifically on the parathyroid cell Ca^{2+} receptor or otherwise modulate one or more functions of the parathyroid cell's normal response to $[Ca^{2+}]$.

Screening of various other molecules is described in detail below and the results summarized in Table 1.

Example 2: Polyamine Screening

Straight-chain polyamines (spermine, spermidine, TETA, TEPA, and PEHA) and two derivatives thereof (NPS 381 and NPS 382) were screened as in Example 1. These molecules were all found to mobilize intracellular Ca^{2+} in bovine parathyroid cells. Their order of potency is as follows, with the net positive charge listed in parentheses:

Table 3

Molecule	EC_{50} (in μM)
NPS 382 (+8)	50
NPS 381 (+10)	100
spermine (+4)	150
PEHA (+6)	500
spermidine (+3)	2000
TEPA (+5)	2500
TETA (+4)	8000

Putrescine (+2) and cadaverine (+2) were inactive at a concentration of 2mM.

Another straight-chain polyamine, DADD, behaved somewhat differently from the other polyamines and is

described in Example 7.

Example 3: Cyclic Polyamine Screening

Two cyclic polyamines, hexacyclen and NPS 383, were screened as in Example 1N Hexacyclen (+6, EC_{50} = 20 μM) is 7-fold more potent than NPS 383 (+8, EC_{50} = 150 μM). The converse would be expected based solely on net positive charge as the structural characteristic for Ca^{2+} receptor activity.

Example 4: Aminoglycoside Antibiotic Screening

Six antibiotics were screened as in Example 1. The resulting EC_{50}'s for the mobilization of intracellular Ca^{2+}, in rank order of potency, were:

Table 4

Antibiotic	EC_{50} (in μM)
neomycin (+6)	10
gentamicin (+5)	150
bekanamycin (+5)	200
streptomycin (+3)	600

Kanamycin (+4.5) and lincomycin (+1) were without effect at a concentration of 500 μM. Within the aminoglycoside series, there is a correlation between net positive charge and potency. However, neomycin is considerably more potent than various polyamines (NPS 381, NPS 382, NPS 383, PEHA) that have an equal or greater positive charge. Since aminoglycoside antibiotics of this type have renal toxicity which may be related to interaction with calcium receptors in the kidney, such screening could be used to screen for toxicity in the development of new aminoglycoside antibiotics.

Example 5: Peptide and Polyamino Acid Screening

Protamine and polymers of lysine or arginine varying in peptide length were screened for their ability to mobilize intracellular Ca^{2+} as in Example 1. The resulting EC_{50}'s for the mobilization of intracellular Ca^{2+}, in rank order of potency, were:

Table 5

Peptide (MW in kD)	EC_{50} (in nM)
polyArg (100)	4
polyArg (40)	15
polyLys (27)	30
protamine (4.8)	75
polyArgTyr (22)	200
polyLys (14)	1000
polyLys (3.8)	3000

The net positive charge of these polymers increases as the MW increases. Thus, as for the aminoglycosides, there is a direct correlation between net charge and potency among this series of polyamino acids. Protamine is essentially polyArg with a net positive charge of +21.

Example 6: Arylalkylamine Screening

Molecules selected from the class of arylalkylamine toxins derived from the venoms of wasps and spiders were screened as in Example 1.

Philanthotoxin-433 (+3) was without effect at a concentration of 500 μM. It is similar in structure to the argiotoxins described below.

Argiotoxin-636 (400 μM) did not elicit increases in $[Ca^{2+}]_i$ but it did potentiate cytosolic Ca^{2+} responses to the subsequent addition of extracellular Ca^{2+}. This is a feature common to all molecules that activate the Ca^{2+} receptor and is also seen with a variety of extracellular divalent cations. This is considered in more detail in Example 7.

In contrast to argiotoxin-636, argiotoxin-659 elicited increases in $[Ca^{2+}]_i$ with an EC_{50} of 300 μM. Argiotoxin-659 differs from argiotoxin-636 in having a hydroxylated indole moiety rather than a dihydroxyphenyl group. This is the only difference in the structure of these two molecules. Thus, the difference in potency lies in the nature of the aromatic group, not in the polyamine chain which carries the positive charge.

Example 7: Screening of Ca^{2+} Channel Blockers

Ca^{2+} channel blockers, i.e., those molecules which block influx of extracellular Ca^{2+} through voltage- sensitive Ca^{2+} channels, were screened as in Example 1. There are three structural classes of Ca^{2+} channel blockers: (1) dihydropyridines, (2) phenylalkylamines, and (3) benzothiazipines.

None of the dihydropyridines tested (nifedipine, nitrendipine, BAY K 8644, and (-) 202-791 and (+) 202-791) had any effect on basal $[Ca^{2+}]_i$ or increases in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+} when they were tested at 1 μM. Previous studies showed that parathyroid cells lack voltage-sensitive Ca^{2+} channels, but do have voltage-insensitive Ca^{2+} channels that are regulated by the Ca^{2+} receptor.

The phenylalkylamines examined were verapamil, D-600 (a methoxy-derivative of verapamil), TMB-8, and an analog of TMB-8, NPS 384. The first three molecules were tested at a concentration of 100 μM. The phenylalkylamines behaved differently from other molecules examined. They evoked no change in $[Ca^{2+}]_i$ when added to cells bathed in buffer containing a low concentration of extracellular Ca^{2+} (0.5 mM). However, verapamil, D-600, and TMB-8 potentiated the mobilization of intracellular Ca^{2+} elicited by extracellular divalent cations and they additionally blocked the influx of extracellular Ca^{2+}. At intermediate levels of extracellular Ca^{2+} (1-1.5 mM), these molecules were capable of evoking a small but robust increase in $[Ca^{2+}]_i$ that arose from the mobilization of intracellular Ca^{2+}.

The phenylalkylamines act differently than organic polycations like neomycin. The data suggest that verapamil, D-600 and TMB-8 are partial agonists or allosteric activators at the Ca^{2+} receptor, in contrast to the other molecules examined which are full agonists.

Molecule NPS 384, at a concentration of 300 μM, did not evoke an increase in $[Ca^{2+}]_i$ but it blocked influx of extracellular Ca^{2+}. Testing at higher concentrations may reveal an ability of this molecule to cause the mobilization of intracellular Ca^{2+}.

While the ability of these molecules to block influx is intriguing and not entirely unexpected, it is the ability of these molecules to evoke transient

increases in $[Ca^{2+}]_i$ (arising from intracellular Ca^{2+} mobilization) that is important. Considerable experience with measurements of $[Ca^{2+}]_i$ in parathyroid cells shows that transient increases in $[Ca^{2+}]_i$ almost invariably result from the mobilization of intracellular Ca^{2+} and therefore reflects activation of the Ca^{2+} receptor.

The benzothiazipine examined, diltiazem, was similar in all respects to verapamil and D-600 and was also effective at 100 μM.

It should be mentioned that with the exception of the phenylalkylamines, all the active molecules tested above evoke increase in $[Ca^{2+}]_i$ that are of magnitude similar to that evoked by a maximally effective concentration of extracellular Ca^{2+}. This shows that these molecules are equally efficacious as extracellular divalent cations. This contrasts with the activity of phenylalkylamines, which seem to act only as partial agonists.

Amongst the phenylalkylamines, some interesting structure-activity relationships emerge. Significant is the different potencies of molecules like TMB-8 and NPS 384. TMB-8 potentiated transient increases in $[Ca^{2+}]_i$ at 100 μM whereas NPS 384 fails to do so even at 300 μM, yet these molecules carry the same net positive charge. It follows that some other structural feature, unrelated to net charge, imparts greater potency to TMB-8.

Example 8: Molecule Screening on Human Parathyroid Cells

Spermine and neomycin were tested for effects on $[Ca^{2+}]_i$ in human parathyroid cells obtained from glands removed by surgery and prepared as in Example 1. In human parathyroid cells, spermine was found to cause only a small increase in $[Ca^{2+}]_i$ when tested at a concentration of 300 μM.

Neomycin, on the other hand, evoked a large increase in $[Ca^{2+}]_i$ in human parathyroid cells when tested at a concentration of 20 μM. The magnitude of the response elicited by neomycin was equal to that evoked by a maximally effective concentration of extracellular Ca^{2+}.

Example 9: Molecule Screening on Xenopus Oocytes

Oocytes injected with mRNA from human parathyroid cells express the Ca^{2+} receptor and mobilize intracellular Ca^{2+} in response to a variety of extracellular inorganic di- and trivalent cations. Using this screen allows one to test for an action directly on the Ca^{2+} receptor. Oocytes expressing the Ca^{2+} receptor also responded to several molecules active on intact parathyroid cells when screened as follows. Hexacyclen caused the mobilization of intracellular Ca^{2+} at a concentration of 135 μM. Neomycin (100 μM) and NPS 382 (5 mM) were also effective. This offers rather compelling evidence showing that these molecules act on the Ca^{2+} receptor or on some other protein intimately associated with its function.

For example, we have been able to detect Ca^{2+}

receptor expression in oocytes by measuring $^{45}Ca^{2+}$ mobilization. In these experiments, oocytes were injected with bovine parathyroid mRNA or water and, after 72 hours exposed to serum or 10 mM neomycin. Prior to being stimulated, oocytes were loaded with $^{45}Ca^{2+}$. Stimulation with serum for 20 min resulted in intracellular $^{45}Ca^{2+}$ release representing a 45% increase compared to mock challenge with buffer. Challenge with 10 mM neomycin for 20 min. resulted in a 76% increase in $^{45}Ca^{2+}$ release. The assay is sensitive enough for use in cloning the Ca^{2+} receptor, and has the advantage of a higher throughout than the electrophysiological measurement of Ca^{2+} activated Cl^- current.

In another example, human osteoclastoma tissue was obtained from bone biopsy tissue. Oocytes injected with mRNA isolated from this tissue were challenged with 30 mM Ca^{2+}. Controls did not respond while 8 of 12 oocytes injected with osteoclastoma mRNA responded appropriately (Fig. 34). These experiments provide the first evidence that the Ca^{2+} response of osteoclasts to extracellular Ca^{2+} is in fact genetically encoded. The results also indicate that the osteoclast Ca^{2+} receptor may be cloned by expression in Xenopus oocytes.

Example 10: Molecule Screening on Rat Osteoclasts

The different sensitivities of parathyroid cells and rat osteoclasts to extracellular Ca^{2+} suggest that their Ca^{2+} receptors are different. While parathyroid cells respond to extracellular Ca^{2+} concentrations between 0.5 and 3 mM, osteoclasts respond only when the

level of extracellular Ca^{2+} increases beyond 5 mM. This rather high concentration of Ca^{2+} is nonetheless physiological for osteoclasts; as they resorb bone, the local concentration of extracellular Ca^{2+} may reach levels as high as 30 mM.

Molecule screening with rat osteoclasts was performed as follows. Osteoclasts were obtained from the long bones of neonatal rats. $[Ca^{2+}]_i$ was measured in single cells using the fluorimetric indicator indo-1. Spermine, spermidine, neomycin, and verapamil were tested, and none of these caused any large increase in $[Ca^{2+}]_i$ in osteoclasts (although small responses were detected).

At a concentration of 1 mM, spermidine caused a small increase in $[Ca^{2+}]_i$ (about 10% of that evoked by a maximal concentration of extracellular Ca^{2+}). Neither neomycin (10 mM) nor spermine (10 or 20 mM) caused increases in $[Ca^{2+}]_i$ in rat osteoclasts. Neomycin (10 mM) did not block the increase in $[Ca^{2+}]_i$ elicited by the subsequent addition of 25 mM extracellular Ca^{2+}. Pretreatment with spermine (20 mM), however, did depress the response to extracellular Ca^{2+}. Verapamil (100 μM) caused no detectable increase in $[Ca^{2+}]_i$ but it did block the response to extracellular Ca^{2+}.

Comparisons between osteoclasts and parathyroid cells show that molecules active on the latter are relatively ineffective in osteoclasts. This demonstrates that drugs that target a specific Ca^{2+} receptor without affecting those receptor types present

on other Ca^{2+}-sensing cells are readily developed. Similarly, drugs active at two or more such Ca^{2+} receptors may also be developed.

Screening for Calcimimetic and Calcilytic Activity on the Osteoclast Calcium Receptor

Compounds possessing activity on the osteoclast calcium receptor can be discovered by measuring $[Ca^{2+}]_i$ in single rat osteoclasts as described above. An improved assay enables moderate-to-high levels of compound throughout. This new method is based on the use of rabbit osteoclasts which can be obtained in high yield (10^5 per animal) and purity (95% of the cells are osteoclasts). The purity of the rabbit osteoclast preparation allows measurements of $[Ca^{2+}]_i$ to be performed on populations of cells. Because the recorded fluorescence signal is an averaged population response, intercellular variability is minimized and the precision of the assay is greatly increased. This, in turn, enables more compounds to be screened for activity.

Rabbit osteoclasts are prepared from 6-day old bunnies. The animals are sacrificed by decapitation and the long bones removed and placed into osteoclast medium (OC medium: alpha-minimum essential medium containing 5% fetal bovine serum and penicillin/streptomycin). The bones are cut into sections with a scalpel and placed in 2 ml of OC media in a 50 ml conical centrifuge tube. The bone sections are minced with scissors until a fairly homogeneous suspension of bone particles is obtained. The suspension is then diluted with 25 ml of

OC media and the preparation swirled gently ("vortexed") for 30 seconds. The bone particles are allowed to settle for 2 minutes after which the supernatant is removed and added to a 50 ml centrifuge tube. The bone particles are resuspended in OC media, swirled, sedimented and harvested as just described. The supernatants from the two harvests are combined and centrifuged and the resulting cellular pellet resuspended in Percoll. The suspension is then centrifuged and the white viscous band just below the meniscus is removed and washed with OC media. The Percoll centrifugation step results in a significant improvement in purity and allows osteoclasts to be plated at high densities, suitable for measuring $[Ca^{2+}]_i$ in populations of cells. The cells are plated onto glass cover slips appropriate for measuring $[Ca^{2+}]_i$ according to one of the methods described below. If necessary, the purity of the preparation can be improved. In this case, the cells are cultured overnight and then rinsed with Ca^{2+}- and Mg^{2+}-free buffer. The cell monolayer is then immersed in Ca^{2+}- and Mg^{2+}-free buffer containing 0.02% EDTA and 0.001% pronase for 5 min. This buffer is then removed and replaced with OC media and the cells allowed to recover for 1 to 2 hours before loading the cells with fluorimetric indicator and measuring $[Ca^{2+}]_i$ as described below.

In one embodiment, this technique allows the measurement of $[Ca^{2+}]_i$ in populations of osteoclasts using fluorescence microscopy. The purified osteoclasts

are allowed to attach to 25 mm diameter glass cover slips and then loaded with indo-1. The cover slips are secured into a superfusion chamber and placed onto the stage of a fluorescence microscope. The use of a low power objective (x4) allows a field containing 10 to 15 osteoclasts to be visualized. In one variation, the fluorescence of each cell in the field can be recorded simultaneously and stored separately for later analysis. Changes in $[Ca^{2+}]_i$ of each cell can be estimated and the average response of all cells in the field calculated. In another variation, the fluorescence from the entire field of cells can be recorded and processed immediately. In either variation, the final data is in the form of an average response from the cells present in the microscopic field. Because of this, intercellular variability is minimized and precision of the assay greatly increased. This method enables 10-20 compounds per week to be screened for activity on the osteoclast calcium receptor.

In a more preferred embodiment, this technique allows the measurement of $[Ca^{2+}]_i$ in populations of osteoclasts using a conventional fluorimeter. The purified osteoclasts are allowed to attach to rectangular glass cover slips. In one variation, a standard quartz cuvette ($1cm^2$) is used and the glass coverslips are 2 x 1.35 cm. In another variation, a microcuvette is used ($0.5\ cm^2$) and the glass coverslips are 1 x 0.75 cm. In either case the cells are loaded with fura-2 or some other suitable fluorimetric indicator for measuring $[Ca^{2+}]_i$. The

fluorescence of indicator-loaded cells is recorded as described above for bovine parathyroid cells. This method allows a higher throughput than fluorescence microscopy and enables 20-50 compounds per week to be evaluated for activity on the osteoclast calcium receptor.

In a most preferred embodiment, the technique can be used to measure $[Ca^{2+}]_i$ in osteoclasts in a 96-well plate. The purified osteoclasts are plated at high density into each well of a 96-well plate and subsequently loaded with a suitable fluorimetric indicator. The fluorescence of each well is recorded using a fluorimetric plate reader. This method has the potential of becoming fully automated using robotics a would enable high throughput screening in which 50 to 100 compounds per week could be screened.

Other Ca^{2+} Receptor Examples

The following examples demonstrate that, just as there are subtypes of receptors for molecular ligands, so too do there appear to be subtypes of Ca^{2+} receptors that can be differentially affected by drugs. The parathyroid cell Ca^{2+} receptor senses levels of extracellular Ca^{2+} around 1.5 mM whereas the Ca^{2+} receptor on the osteoclast responds to levels around 10 mM (Fig. 22). Neomycin or spermine, which activate the parathyroid cell Ca^{2+} receptor, fail to affect the Ca^{2+} receptors on C-cells or osteoclasts (Figs. 21 and 23). These data constitute the first evidence for pharmacologically distinct subtypes of Ca^{2+} receptors and these data are being used to design and develop

drugs that act selectively on a particular type of Ca^{2+} receptor. Indeed, testing of lead molecules demonstrate such cell-specific effects. For example, NPS 449, which elicits increases in $[Ca^{2+}]_i$ in osteoclasts is without effect on $[Ca^{2+}]_i$ in parathyroid cells. Conversely, NPS 447, which activates the parathyroid cell Ca^{2+} receptor, is effective in activating the osteoclast Ca^{2+} receptor only at concentrations 10-fold higher. Finally, agatoxin 489, although not very potent in activating the C-cell Ca^{2+} receptor (EC_{50} = 150 μM), is a quite potent activator of the parathyroid cell Ca^{2+} receptor (EC_{50} = 3 μM). The lead molecules presently under development will affect selectively the activity of a specific type of Ca^{2+}-sensing cell in vivo.

Drugs with less specificity might not necessarily be therapeutically undesirable. Thus, depressing osteoclast activity and stimulating calcitonin secretion are two different approaches to inhibiting bone resorption. Drugs that target the Ca^{2+} receptors on both of these cells might be very effective therapies for osteoporosis. Because PTH is also involved in regulating bone metabolism, drugs acting on the parathyroid cell Ca^{2+} receptor may also be useful in the treatment and/or prevention of osteoporosis.

Results of some test molecules are shown below. In Table 6, the comparative activity of calcimimetic molecules is shown. Bovine parathyroid cells and C-cells (rMTC 6-23 cells) were loaded with fura-2, and rat osteoclasts with indo-1 and the potency of the indicated

molecules to mobilize intracellular Ca^{2+} determined by constructing cumulative concentration-response curves. Molecules listed as "inactive" did not alter $[Ca^{2+}]_i$ when tested at a concentration of 1 mM.

Table 6

	EC_{50} (μM)		
COMPOUND	PARATHYROID	OSTEOCLAST	C-CELL
NPS R-568	0.60	200	>300
NPS S-568	30	---	---
NPS R-467	2	>100	---
NPS S-467	>30	---	---
NPS 017	6	inactive	150
NPS 447	9	150	---
NPS 456*	15	200	>100
NPS 015	22	---	inactive
NPS 109	40	>300	5
NPS 449	inactive	150	---
NPS 468*	30	250	---
spermine	150	inactive	inactive
neomycin	40	inactive	inactive

*racemic mixture; *inactive* is defined as causing no increase in cytosolic Ca^{2+} at a concentration of 1-5 mM.

*racemic mixture; "inactive" is defined as causing no increase in cytosolic Ca^{2+} at a concentration of 1-5 mM.

Example 11: Lead Molecules for Parathyroid Ca^{2+} Receptor

Structure-activity studies using polyamines and arylalkylamines led to the testing of molecules

structurally akin to NPS 456. NPS 456 is a potent activator of the parathyroid cell Ca^{2+} receptor. This molecule is notable because it possess only one positive charge yet is much more potent than many polybasic molecules. Brief (2 min) pretreatment with PMA shifts the concentration-response curve for NPS 456 to the right. This indicates that NPS 456 acts through the same mechanism used by extracellular Ca^{2+}. NPS 456 evokes the mobilization of intracellular Ca^{2+} in Xenopus oocytes expressing the parathyroid cell Ca^{2+} receptor, which demonstrates a direct action on the Ca^{2+} receptor (Fig. 33). Moreover, NPS 456 contains a chiral carbon, and therefore exists in two isomeric forms. Both isomers have been synthesized and examined for activity. The R-isomer, NPS 447, is 12 times more potent than the S-isomer, NPS 448 (Fig. 28). This is the first demonstration that a Ca^{2+} receptor can recognize an organic molecule in a stereospecific manner.

Because NPS 447 is a structurally simple molecule with selective and potent effects on the parathyroid cell Ca^{2+} receptor, structure-activity studies around this lead molecule are simple. The aim of these studies is to generate an array of related molecules with various characteristics from which the final development candidate can be selected. This effort has already revealed some of the structural domains of NPS 447 that contribute to activity and potency. For example, the novel compound NPS 459 is an analog of NPS 447 that is smaller (MW < 240) yet nearly as potent as the parent

molecule, whereas several other analogs are relatively inactive. The most interesting molecules from this analog project can be put into in vivo testing for effects on PTH secretion and serum Ca^{2+} levels (see Examples 15, 16, 17, 18 and 23).

The novel compound NPS 467 is an even smaller molecule than NPS 447 yet the former is about 3-fold more potent than the latter in causing increases in $[Ca^{2+}]_i$ in parathyroid cells. Like NPS 456, NPS 467 is a racemic mixture. Resolution of NPS 467 into its enantiomers provides an isomer of even greater potency than the racemic mixture (see Example 16). Further structure-activity studies on molecules related to NPS 447, NPS 467 and NPS 568 are expected to yield pure isomers with greater potency than these molecules in their racemic forms.

Results obtained with NPS 456 (Fig. 33) show that it elicits oscillatory increases in Cl^- current at concentrations of 100 μM. NPS 456 is the most potent molecule activate on Xenopus oocytes expressing the parathyroid cell Ca^{2+} receptor. The results obtained in this expression system with neomycin and NPS 456 demonstrate that these molecules act directly on the Ca^{2+} receptor.

Example 12: Osteoclast Ca^{2+} Receptor Lead Molecules

The strategy used for elucidating the mechanism of action of extracellular Ca^{2+} on the osteoclast was similar to that proven effective in parathyroid cells. The first experiments examined the effects of La^{3+} on $[Ca^{2+}]_i$ in single rat osteoclasts loaded with the

fluorimetric indicator indo-1. As described above, trivalent cations like La^{3+} are impermeant and block Ca^{2+} influx. Low micromolar concentrations of La^{3+} partially depressed extracellular Ca^{2+}-induced increases in $[Ca^{2+}]_i$ (Fig. 29). The demonstration of a La^{3+}-resistant increase in $[Ca^{2+}]_i$ provides evidence for the mobilization of intracellular Ca^{2+}. The results of these experiments parallel those obtained in parathyroid cells and suggest that similar mechanisms are used by extracellular Ca^{2+} to regulate $[Ca^{2+}]_i$ in both cell types.

Another series of experiments showed that extracellular Mn^{2+} evoked transient increases in $[Ca^{2+}]_i$ (Fig. 30(a)) that persisted in the absence of extracellular Ca^{2+} (Fig. 30B). These results are likewise indicative of the mobilization of intracellular Ca^{2+}. Although Mn^{2+} can enter some cells, it is unlikely to do so in the osteoclast because Mn^{2+} quenches the fluorescence of indo-1. Thus, if Mn^{2+} penetrated intracellularly, a decrease, not an increase in the fluorescent signal would be observed.

The results obtained with a variety of di- and trivalent cations are all consistent with the presence of a Ca^{2+} receptor on the surface of the osteoclast that is coupled to the mobilization of intracellular Ca^{2+} and influx of extracellular Ca^{2+} through voltage-insensitive channels. Results show evidence for genetic material in human osteoclasts that encodes a Ca^{2+} receptor protein (see below). Transient increases in $[Ca^{2+}]_i$ resulting from the mobilization

of intracellular Ca^{2+}, are sufficient to inhibit osteoclastic bone resorption in vitro. Thus, as with the parathyroid cell, activation of the Ca^{2+} receptor appears to be a viable means of inhibiting the activity of osteoclasts.

NPS 449 is presently the lead molecule for calcimimetic drugs on this receptor. It is a small molecule (MW < 425) and it mobilizes intracellular Ca^{2+} in rat osteoclasts with a EC_{50} of about 150 μM (Figs. 31A and 31B). Although the potency of NPS 449 is relatively low, it has a simple structure with only one positive charge and is expected to have desirable pharmacodynamic and pharmacokinetic properties.

NPS 449 was examined for its ability to inhibit bone resorption in vitro. This was done by morphometric analysis of pit formation on thin slices of bovine cortical bone using scanning electron microscopy. Rat osteoclasts were incubated for 24 hours in slices of bone in the presence or absence of various concentrations of NPS 449. NPS 449 caused a concentration-dependent inhibition of bone resorption with an IC_{50} of 10 μM. The anticipated results provide the first demonstration that molecules acting at this novel site can inhibit osteoclastic bone resorption. More potent analogs of NPS 449 will be generated using synthetic chemistry and will be tested and assayed using the methods described herein.

Example 13: C-Cell Ca^{2+} Receptor Lead Molecules

Activation of the C-cell Ca^{2+} receptor stimulates the secretion of calcitonin which then acts on osteoclasts to inhibit bone resorption. Calcimimetic drugs selectively affecting C-cells are useful in the treatment of osteoporosis.

The mobilization of intracellular Ca^{2+} is used as a functional index of Ca^{2+} receptor activity. The screening effort in C-cells is facilitated by the availability of cultured cell lines expressing the C-cell phenotype (e.g., rat medullary thyroid carcinoma cells; rMTC 6-23 cells). Selected for initial study were three arylalkylamine molecules. Two are naturally occurring (agatoxin 489 and agatoxin 505) and the other (NPS 019) is a synthetic agatoxin analog. Agatoxin 505 was found to block extracellular Ca^{2+}-induced increases in $[Ca^{2+}]_i$, with an IC_{50} of 3 μM. The inhibitory effect resulted from a block of the L-type voltage-sensitive Ca^{2+} channel present in these cells. In contrast, agatoxin 489 was found to mobilize intracellular Ca^{2+} in rMTC cells with an EC_{50} of 150 μM. This was the first organic molecule discovered that was found to activate the C-cell Ca^{2+} receptor. The synthetic analog, NPS 019, was even more potent and mobilized intracellular Ca^{2+} with an EC_{50} of 5 μM (Fig. 32). It is significant that the only structural difference between NPS 019 and agatoxin 489 is the presence or absence of an hydroxyl group. The fact that such subtle differences in structure affect profoundly the potency of molecules indicates a

structurally specific binding site on the Ca^{2+} receptor. This, in turn, encourages the view that very potent and selective activators of Ca^{2+} receptors can be developed.

NPS 019, which is a small molecule (MW < 500), is a lead molecule for development of calcimimetics of the C-cell Ca^{2+} receptor and can be tested for its ability to stimulate calcitonin secretion in vitro. Subsequent in vivo testing will then determine the ability of this molecule to stimulate calcitonin secretion and inhibit bone resorption. These in vivo studies will be performed in rats. The results obtained in these studies, which are anticipated to be positive, will provide the first evidence showing that a small organic molecule acting on a novel receptor can stimulate calcitonin secretion and depress bone resorption.

Example 14: Calcilytic Activity of NPS 021 on Parathyroid Cells

For a compound to be considered a calcilytic, it must block the effects of extracellular Ca^{2+} or a calcimimetic compound on an extracellular Ca^{2+}-sensing cell. An example of a calcilytic compound is NPS 021, the structure of which is provided in Fig. 1. In bovine parathyroid cells loaded with fura-2, NPS 021 blocks increases in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+}. The IC_{50} of NPS 021 for blocking this response is about 200 àM and, at concentrations around 500 àM, the increase in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+} is abolished. Significantly, NPS 021 does not by itself cause any change in $[Ca^{2+}]_i$

when tested at low [Ca^{2+}] (0.5 mM; Fig. 37). Ga^{3+} is also calcilytic to Xenopus oocytes expressing the cloned Ca^{2+} receptor: Ga^{3+} by itself has no effect on the Cl^- currents activated by Gd^{3+}, a calcimimetic, but pretreatment with Ga^{3+} blocks the action of Gd^{3+}.

Example 15: NPS 467 Lowers Serum Ionized Calcium

Compounds shown to activate the bovine parathyroid cell Ca^{2+} receptor *in vitro* were tested for hypocalcemic activity *in vivo*. Male Sprague-Dawley rates (200 g) were maintained on a low calcium diet for one week prior to receiving test substance or vehicle as control. Blood was collected from the tail vein three hours after the intra-peritoneal administration of NPS 467. Ionized Ca^{2+} in whole blood or serum was measured with a Ciba- Corning 634 Analyzer according to the instructions provided with the instrument. Serum total calcium, albumin and phosphate were measured by techniques well known in the art.

NPS 467 caused a dose-dependent reduction in serum or whole blood Ca^{2+} (Fig. 38). The fall in blood Ca^{2+} at this time was paralleled by a proportional fall in the levels of blood total calcium. There was no change in serum albumin or phosphate levels at any of the doses examined. In preliminary studies, NPS 467, at doses effective in lowering blood Ca^{2+}, caused a dose-dependent reduction in circulating levels of PTH (Fig. 39). The hypocalcemic effect of NPS 467 was maximal within three hours and returned toward control levels after 24 hours (Fig. 40).

NPS R-467 (see Example 16) was also effective in lowering serum ionized Ca^{2+} in rats maintained on a normal, calcium-replete diet. A single dose of NPS R-467 (10 mg/kg i.p.) caused a rapid fall in serum levels of ionized Ca^{2+} which were maximal by 1 hr (22% decrease from the control level) and remained depressed at or near this level for up to 6 hours.

Example 16: NPS 467 Lowers Serum Ionized Calcium in a Stereospecific Manner

NPS 467 is a racemic mixture. Resolution of NPS 467 into its two enantiomers was achieved by separation on a chiral column. The R-isomer was about 100-fold more potent than the S-isomer in activating the bovine parathyroid cell Ca^{2+} receptor *in vitro* as assessed by the ability of the enantiomers to evoke increases in $[Ca^{2+}]_i$ in parathyroid cells (Fig. 41). Likewise, similar resolution of the novel compound NPS 568 into its enantiomers showed that the R-isomer was 40-fold more potent than the S-isomer in causing the mobilization of intracellular Ca^{2+} in bovine parathyroid cells (see Table 6, *supra*).

The isomers of NPS 467 were examined for effects on serum Ca^{2+} as in Example 15. Consistent with the *in vitro* results, the R-isomer of NPS 467 proved to be more potent than the S-isomer in lowering serum Ca^{2+} *in vivo* (Fig. 42; each compound was tested at a concentration of 5 mg/kg body weight).

Example 17: NPS 467 Lowers Serum Ionized Calcium in an in vivo Model of Secondary Hyperparathyroidism

An accepted and widely used animal model of

secondary hyperparathyroidism arising from chronic renal failure is the 5/6 nephrectomized rat. Animals receiving such surgery become initially hypocalcemic and, to maintain serum Ca^{2+} levels, there is a compensatory hyperplasia of the parathyroid glands and elevated levels of circulating PTH. Male Sprague-Dawley rats (250 g) received a 5/6 nephrectomy and were allowed to recover for 2 weeks. At this time they were normocalcemic (due to elevated levels of serum PTH). The administration of NPS R-467 (10 mg/kg i.p.) caused a rapid (within 2 hours) fall in serum ionized Ca^{2+} levels to 83% of controls in an animal model of secondary hyperparathyroidism. This suggests that compounds of this sort will effectively depress PTH secretion in patients with secondary hyperparathyroidism and hyperplastic parathyroid glands.

Example 18: NPS 467 Fails to Lower Serum Ionized Calcium Levels in Parathyroidectomized Animals

To determine the primary target tissue upon which NPS 467 acts to cause a hypocalcemic response, the para thyroid glands in rats were surgically removed. Animals receiving a total parathyroidectomy become hypocalcemic and are largely dependent upon dietary calcium to maintain serum Ca^{2+} homeostasis. Parathyroidectomized animals had serum ionized Ca^{2+} levels of 0.92 mM which fell gradually to 0.76 mM after 6 hours of fasting. The administration of a single dose of NPS R-467 (10 mg/kg i.p.) did not cause any change in serum ionized Ca^{2+} levels over a period of 6 hours. These results demonstrate that intact parathyroid glands are required

for the hypocalcemic effects of NPS R-467. The data additionally demonstrate that NPS R-467 can target the parathyroid glands in vivo. The results are consistent with the view that NPS R-467 acts on the parathyroid cell Ca^{2+} receptor in vivo to depress secretion of PTH and thereby cause serum levels of ionized Ca^{2+} to fall.

Example 19: NPS 467 Increases Intracellular Calcium in Human Parathyroid Glands

Dissociated parathyroid cells were prepared from a parathyroid adenoma obtained by surgery from a patient with primary hyperparathyroidism. The cells were loaded with fura-2 and $[Ca^{2+}]_i$ measured as described above. Both NPS R-467 and NPS R-568 caused concentration- dependent increase in $[Ca^{2+}]_i$. The EC_{50}'s for NPS R-467 and NPS R-568 were 20 and 3 μM, respectively. Both these compounds are thus able to increase $[Ca^{2+}]_i$ in pathological human tissue and would thus be expected to decrease serum levels of PTH and Ca^{2+} in patients with primary hyperparathyroidism.

Example 20: Mechanism of Action of NPS 467 at the Para thyroid Cell Calcium Receptor

Dissociated bovine parathyroid cells were used to further explore the mechanism of action of NPS 467 at the receptor level. In the presence of 0.5 mM extracellular Ca^{2+}, NPS R-467 caused a rapid and transient increase in $[Ca^{2+}]_i$ which persisted in the presence of 1 μM La^{3+} and was partially depressed by pretreatment with PMA (100 nM for 2 min.). All these results are consistent with an action of NPS R-467 on the Ca^{2+} receptor. However, the cytosolic Ca^{2+}

response to NPS R-467 was abolished when parathyroid cells were suspended in Ca^{2+}-free buffer. This suggests that NPS R- 467 cannot, by itself, cause the mobilization of intracellular Ca^{2+}. It does, however, elicit responses in parathyroid cells and in oocytes when a small amount of extracellular Ca^{2+} is present. This suggests that partial occupancy of the Ca^{2+}-binding site is required for NPS R-467 to elicit a response. To test this hypothesis, parathyroid cells were suspended in Ca^{2+}-free buffer and exposed to a submaximal concentration of neomycin. Neomycin was used because it mimics, in nearly all respects, the effects of extracellular Ca^{2+} on parathyroid cells and on Xenopus oocytes expressing the parathyroid cell Ca^{2+} receptor. The addition of 10 àM neomycin did not by itself cause an increase in $[Ca^{2+}]_i$ under these conditions. However the subsequent addition of NPS R-467 (30 μM) now elicited a transient increase in $[Ca^{2+}]_i$ which, because there was no extracellular Ca^{2+} present, must have come from the mobilization of intracellular Ca^{2+}. When cells bathed in Ca^{2+}-free buffer were exposed to 30 àM NPS R-467 there was no increase in $[Ca^{2+}]_i$. This concentration of NPS R-467 is maximally effective in increasing $[Ca^{2+}]_i$ when extracellular Ca^{2+} (0.5 mM) is present. However, the subsequent addition of 10 μM neomycin now evoked a transient increase in $[Ca^{2+}]_i$. Presumably, neomycin binds to the same site as extracellular Ca^{2+} and can functionally substitute for it. Using a submaximal concentration, which by itself causes no response,

achieves partial occupancy of the Ca^{2+}-binding site and allows activation of the Ca^{2+} receptor by NPS R-467.

Additional studies to further define the mechanism of action of NPS R-467 were performed. The cells were once again suspended in Ca^{2+}-free buffer to insure that any observed increase in $[Ca^{2+}]_i$ resulted from the mobilization of intracellular Ca^{2+}. In these experiments, however, a maximally effective concentration (100 μM) of neomycin was used. In the absence of extracellular Ca^{2+}, 100 μM neomycin evoked a rapid and transient increase in $[Ca^{2+}]_i$. The subsequent addition of 30 μM NPS R-467 did not cause an increase in $[Ca^{2+}]_i$. In the converse experiment, 30 μM NPS R-467 was added before 100 μM neomycin. As expected, NPS R-467 did not cause any increase in $[Ca^{2+}]_i$. It did not, however, affect the increase in $[Ca^{2+}]_i$ evoked by the subsequent addition of 100 μM neomycin. These results, obtained with maximally effective concentrations of NPS R-467 and neomycin, suggest that these two compounds do not act at the same site. Rather, the results can be sufficiently explained by postulating two separate sites on the Ca^{2+} receptor, one to which extracellular Ca^{2+} and neomycin bind, and another to which NPS R-467 and structurally related compounds (such as NPS R-568) bind. Ligand binding to the former site can result in full activation of the Ca^{2+} receptor whereas ligand binding to the latter site can only occur and/or be functionally relevant when the extracellular

Ca^{2+}-binding site is occupied to some as yet undefined degree. It is possible that ligand binding to the extracellular Ca^{2+}- binding site exposes a previously occluded binding site for NPS R-467. It appears that the NPS R-467-binding site is an allosteric site that augments receptor activation in response to ligand binding at the extracellular Ca^{2+}-binding site.

The data demonstrate that the parathyroid cell Ca^{2+} receptor possesses at least two distinct sites for organic ligands. One site binds the physiological ligand, extracellular Ca^{2+}, and certain organic polycations like neomycin. Binding to this site result in full activation of the Ca^{2+} receptor; an increase in $[Ca^{2+}]_i$, and the inhibition of PTH secretion. NPS R-467 and NPS R-568 define a previously unrecognized binding site on the Ca^{2+} receptor. Binding to this site can only occur and/or results in full activation of the Ca^{2+} receptor when the extracellular Ca^{2+}-binding site is partially occupied. Ligands acting at either site are effective in suppressing serum Ca^{2+} levels in vivo.

Allosteric Site on Parathyroid Cell Calcium Receptor

Calcimimetic compounds that activate the bovine parathyroid cell calcium receptor, such as NPS R-467 and NPS R-568, do not cause the mobilization of intracellular Ca^{2+} in the absence of extracellular Ca^{2+}. Rather, they increase the sensitivity of the Ca^{2+} receptor to activation by extracellular Ca^{2+}, thus causing a shift to the left in the concentration-response curve for extracellular Ca^{2+}. Because of

this, it is unlikely that they act at the same site on the receptor as does extracellular Ca^{2+}. In contrast, organic and inorganic polycations do cause the mobilization of intracellular Ca^{2+} in the absence of extracellular Ca^{2+} and therefore probably act at the same site as does extracellular Ca^{2+}. Compounds like NPS R-568, presumably act in an allosteric manner and their activity is dependent on some minimal level of extracellular Ca^{2+}. This suggests that partial occupancy of the extracellular Ca^{2+}-binding site on the receptor is required for compounds like NPS R-568 to be effective. This model is consistent with the observations described in Example 20.

Other details of the mechanism of action of NPS R- 568 on the parathyroid cell Ca^{2+} receptor, however, are more accurately investigated by binding studies in which the specific binding of radiolabeled (using ^{3}H for example) NPS R-568 is assessed. There are several molecular mechanisms that could explain the activity of NPS R-568 on the parathyroid cell Ca^{2+} receptor. In one mechanism (model 1), NPS R-568 could bind to the Ca^{2+} receptor at a site that, when occupied, is not sufficient to activate the receptor functionally. Activation only occurs when some level of occupancy of the extracellular Ca^{2+}-binding site(s) is achieved. In an alterative mechanism (model 2), the occupation of the extracellular Ca^{2+}-binding site could unmask latent binding sites for compounds such as NPS R-568. Occupancy of this latent site by NPS R-568 then increases the affinity and/or efficacy of binding at the

extracellular Ca^{2+} site. Either mechanism involves a form of allosteric activation of the Ca^{2+} receptor by compounds such as NPS R-568. These are not the only possible mechanisms that could explain the effect of compounds like NPS R-568 on the parathyroid cell Ca^{2+} receptor. Other mechanisms of action may be suggested by the results of the binding studies described below.

To further investigate the mechanism of action of compounds like NPS R-568 on the parathyroid cell Ca^{2+} receptor, binding studies using ^{3}H-NPS R-568 can be performed. The specific binding of ^{3}H-NPS R-568 to intact parathyroid cells or to membranes prepared from parathyroid cells is initially investigated by techniques well-known in the art. The kinetic parameters of binding will then be measured as a function of extracellular Ca^{2+} concentrations. Specifically, Scatchard analysis of the data will reveal the number of binding sites and the apparent affinity of the receptor site for ^{3}H-NPS R-568. These parameters will then be investigated as a function of changes in the level of extracellular Ca^{2+} in the buffer used for the assay. If model 1 is correct, then a significant level of specific binding should occur in the absence of extracellular Ca^{2+}. Large changes in the kinetic parameters of binding as a function of the level of extracellular Ca^{2+} would favor model 2. It is expected that various other inorganic and organic polycations described above in other examples will cause similar changes in the binding parameters of ^{3}H-NPS R-568 as does extracellular Ca^{2+}. This would support

the view that these polycations act at the extracellular Ca^{2+}-binding site, which is distinct from that to which compound like NPS R-568 bind.

Example 21: Preparation of NPS 467

In a 250 ml round bottom flask, 10.0 g (100 mmoles) 3'-methoxy acetophenone and 13.5 g (100 mmoles) 3-phenylpropylamine were mixed and treated with 125 mmoles (35.5 g) titanium(IV) isopropoxide. The reaction mixture was stirred 30 minutes at room temperature under a nitrogen atmosphere. After this time 6.3 g (100 mmoles) sodium cyanoborohydride in 100 ml ethanol was added dropwise over the course of 2 minutes. The reaction was stirred room temperature under nitrogen for 16 hours. After this time the reaction mixture was transferred to a 2 L separator funnel with 1.5 L ethyl ether and 0.5 L water. The phases were equilibrated and the ether layer removed. The remaining aqueous phase was thoroughly extracted with four 1 L portions of ether. The washes were combined, dried over anhydrous potassium carbonate and reduced to a clear, light amber oil.

TLC analysis of this material on silica using chloroform-methanol-isopropylamine (100:5:1) showed product at R_f 0.65 with traces of the two starting materials at R_f 0.99 (3'-methoxy acetophenone) and R_f 0.0 (3-phenylpropylamine).

The reaction mixture was chromatographed through silica (48 x 4.6 cm) using a gradient of chloroform-methanol-isopropylamine (99:1:0.1) to (90:10:0.1) which yielded 13.66 g of purified NPS 467. This material was

dissolved in hexane-isopropanol (99:1) containing 0.1% diethylamine to yield a solution with a concentration of 50 mg/ml. Chiral resolution was accomplished by chroma tography of 4 ml of this solution (200 mg, maximum to achieve separation) through ChiralCel OD (25 x 2 cm) using 0.7% isopropylamine, 0.07% diethylamine in hexane at 100 ml/min, monitoring optical density at 260 nm. Under these conditions (with injections of 100 mg material) the early-eluting isomer (NPS 467R) began to emerge from the column at à26 min, the late eluting isomer (NPS 467S) began to emerge at 34 minutes. Baseline resolution was accomplished under these conditions. Each optical isomer (free base) was converted to the corresponding hydrochloride salt by dissolving 3 g of the free base in 100 ml ethanol and treating with it with 100 ml water containing 10 molar equivalents HCl. Lyophilization of this solution yielded a white solid.

NPS 568

NPS 568 is a structural analog of NPS 467 that is more potent in causing increases in $[Ca^{2+}]_i$ in bovine and human parathyroid cells. Like NPS 467, the effects of NPS 568 are stereospecific and it is the R-isomer that is the more potent enantiomer (see Table 6, supra). NPS R-568 is at present the lead calcimimetic compound with selective activity at the parathyroid cell Ca^{2+} receptor. It behaves, albeit with greater potency, similarly to NPS R-467 as described in Examples above. Thus, NPS R- 568 evokes increases in $[Ca^{2+}]_i$ in bovine parathyroid cells in

a stereospecific manner (see Table 6, supra). NPS R-568 fails to evoke increases in $[Ca^{2+}]_i$ in the absence of extracellular Ca^{2+} but it does potentiate responses to extracellular Ca^{2+}. NPS R-568 shifts the concentration-response curve for extracellular Ca^{2+} to the left.

The oral administration of NPS R-568 to rats causes a dose-dependent decrease in the levels of serum Ca^{2+} (ED_{50} = 7mg/kg). The hypocalcemic response elicited by the oral administration of NPS R-568 is rapid in onset and is paralleled by decreases in the levels of serum PTH. The hypocalcemic response evoked by the oral administration of NPS R-568 is only marginally affected by prior complete nephrectomy. However, NPS R-568 fails to elicit a hypocalcemic response in parathyroidectomized rats. NPS R-568 can thus target selectively the parathyroid cell Ca^{2+} receptor in vivo and cause an inhibition of PTH secretion. The decreases in serum levels PTH together with the resulting hypocalcemia are desirable therapeutic effects in cases of hyperparathyroidism. The synthesis of NPS R-568 is provided below.

Example 22: Preparation of NPS 568

NPS 568 was prepared using the methods described in Example 21 substituting an equivalent amount of 3-(2-chlorophenyl)propylamine for 3-phenylpropylamine. It was found that allowing the mixture of 3'-methoxyacetophenone, 3-(2-chlorophenyl)propylamine and titanium(IV) isopropoxide to stir for 5 hours prior to treatment with $NaCNBH_3$/EtOH resulted in significantly

greater yield (98%).

Example 23: NPS 467 Lowers Serum Ionized Calcium When Administered Orally

Rats (male, Sprague-Dawley, 250-300 g) were fed standard rat chow and fasted overnight prior to the experiment. NPS R-467 was suspended in corn oil and administered as a single oral dose through a gavage needle. Three hours later a sample of blood was taken from the tail vein and assessed for ionized Ca^{2+} levels. Fig. 44 shows that NPS R-467 caused a dose-dependent reduction in serum levels of ionized Ca^{2+} when administered orally.

Example 24

BOPCAR1 Cloning Method 1

An expression cloning strategy using Xenopus laevis oocytes was used to identify the cDNA encoding the bovine parathyroid calcium receptor. These techniques are described only briefly here, and a more complete description preceded this one in the sections describing techniques which may be used to clone additional forms of the Ca^{2+}-receptor from other cell types. Poly(A^+)-enriched RNA was initially prepared from bovine parathyroid glands using extraction with guanidinium thiocyanate, centrifugation through CsCl and oligo(dT) cellulose chromatography. Injection of the resultant poly(A^+)-enriched RNA into oocytes (25-50 ng/oocyte) conferred sensitivity to elevated extracellular concentrations of Ca^{2+} and the trivalent cation (1-100μM) Gd^{3+} as described herein, such that the two cations elicited calcium activated chloride

currents. No such currents were elicited in control eggs injected with water. X. laevis frogs were selected on the basis of their harvested oocytes exhibiting: (i) a high level of maturity (i.e., Stage V, VI); (ii) a high activity of Cl^- currents activated by Ca^{2+} ionophores like A23187; (iii) a high level of functional expression of Gd^{3+}-induced Cl^-current when injected with 25 ng/oocyte of total poly(A)$^+$RNA isolated from bovine parathyroid. The mRNA was then subjected to size fractionation, utilizing preparative, continuous flow agarose gel electrophoresis (Hediger, M.A., Anal. Biochem. 159:280-286 (1986)) to obtain fractions of poly(A^+)-RNA further enriched in transcripts coding for the Ca^{2+}-receptor. A peak of activity (e.g., oocytes injected with mRNA from these fractions showed enhanced expression of Gd^{3+}-activated Cl^-currents obtained with a size range of 4-5.5 kb. This RNA was used to prepare a size-selected, directional cDNA library in the plasmid pSPORT1 that was enriched in full length transcripts. Sense complementary RNA (cRNA) was then synthesized from the DNA inserts pooled from 350-500 independent clones from this library and injected into oocytes. Gd^{3+}-activated Cl^-currents were observed following injection of RNA from a single filter containing 350 colonies. Preparation and injection of cRNA from successively smaller pools of clones led to isolation of a single clone (BoPCaR 1) with a cDNA insert of 5.3 kb which expressed greatly enhanced Ca^{2+}-receptor activity following injection of its cRNA into oocytes. A plasmid containing the BoPCaR

1 cRNA (See plasmid, Figure 45; restriction map, Figure 46; and partial nucleotide sequence, Figure 47) has been deposited in the ATCC under deposit number ATCC 75416.

The BoPCaR 1 cDNA is outside the size range of the size-selected RNA found to express neomycin elicited Cl^- channel activity in Xenopus oocytes. This is consistent with the possibility that different isoforms or multiple genes may conceal other numbers of the Ca^{2+} receptor gene family.

Several pharmacological and biochemical criteria were used to identify this clone as encoding a bona fide bovine parathyroid Ca^{2+}-receptor. Oocytes expressing the cloned receptor, but not water-injected oocytes, responded to increasing concentrations of extracellular Ca^{2+} (1.5-5 mM) or Gd^{3+} (20-600μM) with large increases in Cl^- currents (up to at least 1.8 microamperes) that were several fold larger than those observed in poly(A^+)-injected oocytes. These responses increased markedly over a period of one (1) to four (4) days after injection of the eggs with cRNA prepared from the BoPCaR 1 cDNA. Furthermore, the ranges of the concentrations of the two cations eliciting this response were very similar to those shown previously to act on bovine parathyroid cells in vitroN eomycin (20-100 μM), which is known to mimic closely the effects of Ca^{2+} on parathyroid cells, produced changes in Cl^- current in oocytes essentially identical to those produced by Ca^{2+} or Gd^{3+}, and these occurred over the same range of concentrations over which this antibiotic moculates parathyroid

function in vitro. Finally, in vitro translation of RNA prepared from the clone resulted in a single major protein on polyacrylamide gels with a molecular weight of about 100 kd, whose synthesis was enhanced by inclusion of dog pancreatic microsomes, concomitant with an increase in apparent molecular weight of 10-15%. The latter suggests that the cloned receptor interacts strongly with membranes, as might be expected of an integral membrane protein receptor, and is glycosylated in its native form. Studies with the lectin concanavalin A indicate that the Ca^{2+}-receptor is likely a glycoprotein. Thus, the pharmacological properties of the cloned receptor, which is expressed at high levels in oocytes, as well as the biochemical studies carried out to date are completely consistent with its identity with the bovine parathyroid Ca^{2+} dynamics (as exemplified by changes in Ca^{2+}-activated Cl^- currents in oocytes).

Diagnostic Use

NPS R-568 or other compounds active on a calcium receptor can be used as a diagnostic tool. Specifically, a pharmaceutical preparation of such compounds is useful as a diagnostic tool. In one example, a pharmaceutical preparation containing a para thyroid cell calcimimetic compound such as NPS R-568 can be given by oral or another route of administration to hypercalcemic patients with symptoms of mental depression. If these symptoms arise from an underlying hyperparathyroid state, such as primary hyper parathyroidism, then administration of NPS R-568 or a

compound that acts similarly will alleviate those symptoms. If the symptoms do not abate, then the mental depression results from some pathological state that is not hyperparathyroidism. Thus, parathyroid cell calcimimetic compounds can be used in the differential diagnosis of mental depression.

Symptoms and signs common to hyperparathyroidism and other disorders can also be differentially diagnosed in the manner described above. Such shared signs and symptoms include, but are not limited to, hypertension, muscular weakness, and a general feeling of malaise. Alleviation of these symptoms following treatment with a parathyroid cell Ca^{2+} receptor calcimimetic compound would indicate that the problems result from the underlying hyperparathyroidism.

In another example, a compound acting as an antagonist (calcilytic) at the C-cell Ca^{2+} receptor can be administered as described above to diagnose medullary thyroid carcinoma. In this case, administration of the C-cell Ca^{2+} receptor calcilytic compound will depress serum levels of calcitonin which can be readily measured by radioimmunoassay. Certain symptoms associated with medullary thyroid carcinoma, such as diarrhea, may also be monitored to determine if they are abated or lessened following administration of the calcilytic compound.

In a third example, a compound acting as a calcimimetic at the juxtaglomerular cell Ca^{2+} receptor can be used in the differential diagnosis of hypertension. In this case, administration of the

juxtaglomerular cell Ca^{2+} receptor calcimimetic compound can be administered as described above. A decrease in blood pressure to normal levels will occur if the hypertension results mostly or exclusively from elevated levels of renin rather than from an alternative pathological state.

In another example, a compound acting as a specific calcimimetic on the osteoclast Ca^{2+} receptor can be used in the differential diagnosis of high- and low- turnover forms of osteoporosis. In this case, such a compound can be administered in a suitable pharmaceutical preparation and the levels of serum alkaline phosphatase, osteocalcin, pyridinoline and/or deoxypyridinoline crosslinks, and/or some other predictive marker of bone resorption and/or formation measured by techniques well known in the art. A large decrease in one or more of these parameters would be predictive of high-turnover osteoporosis, whereas a small or no decrease in these parameters would be predictive of low-turnover osteoporosis. Such information would dictate the appropriate treatment. Antiresorptive drugs would not be the appropriate sole therapy for low-turnover osteoporosis.

These examples are not exhaustive but serve to illustrate that specific Ca^{2+} receptors can be targeted with pharmaceutical preparations and that the observed effects of such preparations on bodily functions and/or chemical constituents can be used diagnostically. In general, calcimimetic and calcilytic compounds that act on Ca^{2+} receptors of the various

cells described above can be used in the diagnosis of the various diseases associated with the particular cell type. These diseases include but are not limited to bone and mineral-related disorders (as described in Coe et al., Disorders of Bone and Mineral Metabolism, Raven Press, 1990), kidney diseases, endocrine diseases, cancer, cardiovascular diseases, neurological diseases, gastrointestinal diseases, and diseases associated with gestation. Examples of human diseases or disorders in which such molecules may be therapeutically effective are as follows: (1) a calcimimetic is expected to ameliorate psoriasis by reducing the proliferation of the abnormal skin cells. (2) Since Ca^{2++} blocks the effect of vasopressin on MTAL and cortical collecting cells, a calcimimetic is expected to reduce water retention in states of vasoprsssin excess, such as the syndrome of inappropriate vasopressin (ADH) secretion. Conversely, calcium receptor antagonists used in states of ADH deficiency are expected to potentiate the action of any ADH present, such as in partial central diabetes insipidus. (3) Calcimimetics may be used to treat hypertension by (a) reducing renin secretion, (b) by stimulating production of vasodilators such as PTHrP (PTH-related peptide) by vascular smooth muscle. (4) Calcimimetics are expected to increase platelet aggregability, which may be useful when platelet counts are low. Conversely, calcilytics are expected to inhibit platelet function in states where there is hypercoagulability. (5) Calcium promotes differentiation of colon and mammary cells. A

calcimimetic is expected to reduce the risk of colon or breast cancer. (6) Calcium promotes urinary calcium excretion in the MTAL. A calcimimetic is expected to have a useful hypocalcemic action in the therapy of hypercalcemic disorders. The inhibitory effect of calcimimetics on osteoclasts and their stimulation of the secretion of the hypocalcemic peptide calcitonin make them expected to be useful in the therapy of hypercalcemia and its symptoms. A calcimimetic may also improve hypocalcemic symptoms by activating calcium receptors. Conversely, calcilytic is expected to reduce urinary calcium excretion and be useful in the treatment of kidney stones. In addition, calcium suppresses the formation of 1,25-dihydroxyvitamin D in the proximal renal tubule, and this vitamin D metabolite is frequently overproduced in renal stone patients and contributes to their hypercalciuria. Supression of 1,25-dihydroxyvitamin D formation by a calcimimetic is expected to be useful treating renal calcium stone disease. (7) Endogenous amines could reproduce the symptoms in uremic patients by calcimimetic or lytic actions. Calcimimetic and/or calcilytic agents are expected to improve these symptoms. (8) Some of the renal toxicity of aminoglycoside antibiotics may be mediated by interaction of these drugs with renal calcium receptors. Having the calcium receptor is expected to make it possible to carry out drug screening easily when designing new drugs of these classes to minimize renal toxicity. In addition, a renal calcium receptor antagonist would prevent or treat this renal

toxicity if it is related to this mechanism. (9) Some of the genetic component of calcium related disorders, such as osteoporosis, renal stones, and hypertension are expected to be related to inherited problems with certain forms of the receptor. These now can be studied and genetic screening/testing carried out using receptor-based reagents. The human disease, familial hypocalciuric hypercalcemia, may be due to a calcium receptor defect. Definitive diagnostic separation from cases of promary hyperparathyroidism could be carried out with receptor-based technology. (10) Calcium receptors are present in the placenta and are expected to impact on disorders of placental function and transfer of nutrients to the growing fetus.

Other embodiments are within the following claims.

We Claim:

8907V

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Nemeth, Edward F.

(ii) TITLE OF INVENTION: CALCIUM RECEPTOR-ACTIVE MOLECULES

(iii) NUMBER OF SEQUENCES: 1

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Wolf, Greenfield & Sacks, P.C.
(B) STREET: 600 Atlantic Avenue
(C) CITY: Boston
(D) STATE: Massachusetts
(E) COUNTRY: United States of America
(F) ZIP: 02210-2204

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Diskette, 5.25 inch
(B) COMPUTER: IBM-compatible
(C) OPERATING SYSTEM: MS:DOS Version 3.3
(D) SOFTWARE: WordPerfect 5.1

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: Not Available
(B) FILING DATE: Filed Herewith
(C) CLASSIFICATION: Not Available

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Gates, Edward R.
(B) REGISTRATION NUMBER: 31,616
(C) REFERENCE/DOCKET NUMBER: B0801/7012

(ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: (617) 720-3500
(B) TELEFAX: (617) 720-2441
(C) TELEX: EZEKIEL

(2) INFORMATION FOR SEQ ID NO: 1:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2148 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: No

(iv) ANTI-SENSE: No

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

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8907V

We Claim:

1. A pharmaceutical composition comprising a molecule which mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell, or blocks an increase in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+}, said molecule having an EC_{50} of less than or equal to 5 μM, wherein said molecule is not protamine.

2. The pharmaceutical composition of claim 1 wherein said increase in $[Ca^{2+}]_i$ in a cell is transient, having a duration of less than thirty seconds.

3. The pharmaceutical composition of claim 1 or 2 wherein said increase in $[Ca^{2+}]_i$ is rapid, occurring within thirty seconds.

4. The pharmaceutical composition of claim 1 wherein said molecule evokes a sustained increase in $[Ca^{2+}]_i$, having a duration greater than thirty seconds.

5. The pharmaceutical composition of claim 1 wherein said molecule further evokes an increase in inositol-1,4,5-trisphosphate or diacylglycerol.

6. The pharmaceutical composition of claim 5 wherein said increase in said inositol-1,4,5-trisphosphate or diacylglycerol occurs within 60 seconds.

7. The pharmaceutical composition of claim 2 wherein said transient increase is diminished by pretreatment of said cell for less than 10 minutes with an activator of protein kinase C.

8. The pharmaceutical composition of claim 7 wherein said activator of protein kinase C is selected

from the group consisting of phorbol myristate acetate, mezerein and (-)-indolactam V.

9. The pharmaceutical composition of claim 1 wherein said molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation.

10. The pharmaceutical composition of claim 2 wherein said transient increase in $[Ca^{2+}]_i$ is reduced by pretreatment of said cell for ten minutes with 10 mM sodium fluoride.

11. The pharmaceutical composition of claim 1 wherein said cell is a parathyroid cell and said molecule inhibits parathyroid hormone secretion from said cell.

12. The pharmaceutical composition of claim 1 wherein said molecule elicits an increase in Cl^- conductance in a Xenopus oocyte injected with mRNA from a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, perifollicular thyroid cells and placental trophoblasts.

13. The pharmaceutical composition of claim 1 wherein said molecule mobilizes intracellular Ca^{2+} to cause said increase in $[Ca^{2+}]_i$.

14. The pharmaceutical composition of claim 1 wherein said cell is a C-cell or an osteoclast and said molecule inhibits bone resorption in vivo.

15. The pharmaceutical composition of claim 1 wherein said cell is a C-cell and said molecule stimulates calcitonin secretion in vitro or in vivo.

16. The pharmaceutical composition of claim 1 wherein said molecule is either a calcimimetic or calcilytic having an EC_{50} or IC_{50} at a Ca^{2+} receptor of less than or equal to 5 μM.

17. The pharmaceutical composition of claim 1 wherein said molecule has an EC_{50} less than or equal to 5 μM at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, perifollicular thyroid cells and placental trophoblasts.

18. The pharmaceutical composition of claim 1 or 16 or 17 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 1 μM.

19. The pharmaceutical composition of claim 1 or 16, or 17 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 100 nanomolar.

20. The pharmaceutical composition of claim 1 or 16, or 17 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 10 nanomolar.

21. The pharmaceutical composition of claim 1 or 16, wherein said molecule has an EC_{50} or IC_{50} less than or equal to 1 nanomolar.

22. The pharmaceutical composition of claim 1, 16 or 17 wherein said molecule is positively charged at physiological pH.

23. The pharmaceutical composition of claim 22 wherein said molecule is selected from the group consisting of branched or cyclic polyamines, positively charged polyamino acids, and arylalkylamines.

24. The pharmaceutical composition of claim 23 wherein said branched polyamine has the formula

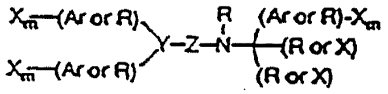
$H_2N-(CH_2)_j-(NR_i-(CH_2)_j)_k-NH_2$

wherein k is an integer from 1 to 10,

each j is the same or different and is an integer from 2 to 20, and

each R_i is the same or different and is selected from the group consisting of hydrogen and $-(CH_2)_j-NH_2$, wherein j is as defined above, and wherein at least one R_i is not hydrogen.

25. The pharmaceutical composition of claim 1, 16 or 17 wherein said molecule has the formula

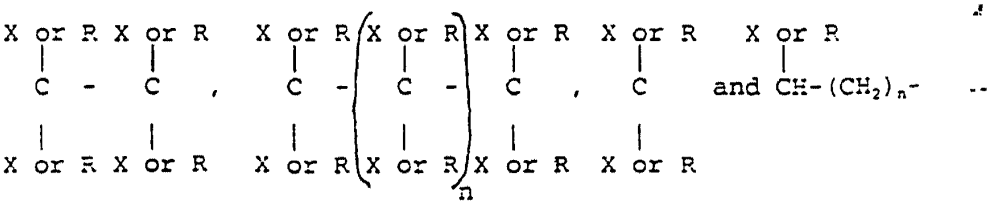


wherein each X is selected (independently) from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN-, NO_2, and CH_3CH_2; Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen, sulfur,

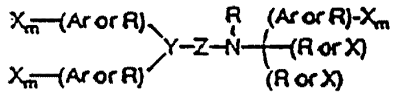


where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is either a calcimimetic or calcilytic.

26. The pharmaceutical composition of claim 25 wherein said hydrophobic entity is selected from the group consisting of phenyl, 2-, 3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2-quinolinyl, 2- or 3-indolyl, benzyl, and phenoxy.

27. A pharmaceutical composition comprising a molecule having the formula



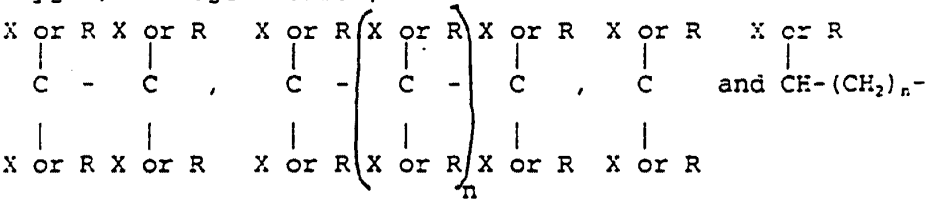
wherein each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, and CH_3CH_2;

Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen sulfur,



where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is not NPS 447 or NPS 449.

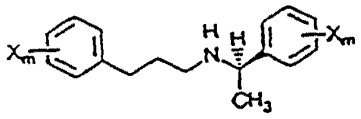
28. The pharmaceutical composition of claim 27 wherein said molecule is NPS 467.

29. The pharmaceutical composition of claim 25 wherein said molecule is NPS 459, NPS 467, or NPS 568.

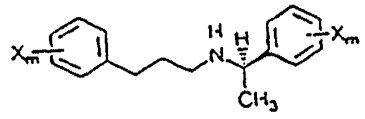
30. The pharmaceutical composition of claim 25, wherein said molecule is an R-phenylpropyl-α-phenethylamine derivative.

31. The pharmaceutical composition of claim 27, wherein said molecule is an R-phenylpropyl-α-phenethylamine derivative.

32. The pharmaceutical composition of claim 30 wherein said molecule has the formula:



33. The pharmaceutical composition of claim 31 wherein said molecule has the formula:



34. The pharmaceutical composition of claim 32 wherein each X is independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

35. The pharmaceutical composition of claim 33 wherein each X is independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

36. A method for treating a patient having a disease characterized by an abnormal $[Ca^{2+}]$ or $[Ca^{2+}]_i$ in one or more cells or in the blood or plasma, comprising the step of administering to said patient a therapeutically effective amount of a molecule which mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell, or blocks an increase in $[Ca^{2+}]_i$ evoked by extracellular Ca^{2+}.

37. The method of claim 36 wherein said molecule has an EC_{50} of less than or equal or 5 μM.

38. The method of claim 36 wherein said molecule is not protamine.

39. The method of claim 36 wherein said molecule interacts at a Ca^{2+} receptor as a calcimimetic or calcilytic.

40. The method of claim 36 wherein said increase in $[Ca^{2+}]_i$ in a cell is transient, having a duration of less than thirty seconds.

41. The method of claim 36 or 40 wherein said increase in $[Ca^{2+}]_i$ is rapid, occurring within thirty seconds.

42. The method of claim 36 wherein said molecule evokes a sustained increase in $[Ca^{2+}]_i$, having a duration greater than thirty seconds.

43. The method of claim 36 wherein said molecule further evokes an increase in inositol-1,4,5-trisphosphate or diacylglycerol.

44. The method of claim 43 wherein said increase in said inositol-1,4,5-trisphosphate or diacylglycerol occurs within less than 60 seconds.

45. The method of claim 40 wherein said transient increase is diminished by pretreatment of said cell with an activator of protein kinase C.

46. The method of claim 45 wherein said activator of protein kinase C is selected from the group consisting of phorbol myristate acetate, mezerein and (-)-indolactam V.

47. The method of claim 36 wherein said molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation.

48. The method of claim 40 wherein said transient increase in $[Ca^{2+}]_i$ is reduced by pretreatment of said cell for ten minutes with 10 mM sodium fluoride.

49. The method of claim 36 wherein said cell is a parathyroid cell and said molecule inhibits parathyroid hormone secretion from said cell.

50. The method of claim 36 wherein said molecule elicits an increase in Cl^- conductance in a Xenopus oocyte injected with mRNA from a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, perifollicular thyroid cells and placental trophoblasts.

51. The method of claim 36 wherein said molecule mobilizes Ca^{2+} within the cell to cause said increase in $[Ca^{2+}]_i$.

52. The method of claim 36 wherein said cell is a C-cell or an osteoclast and said molecule inhibits bone resorption in vivo.

53. The method of claim 36 wherein said cell is a C-cell and said molecule stimulates calcitonin secretion in vitro or in vivo.

54. The method of claim 36 wherein said molecule is either a calcimimetic or calcilytic having an EC_{50} or IC_{50} at a Ca^{2+} receptor of less than or equal to 5 μM.

55. The method of claim 36 wherein said molecule has an EC_{50} less than or equal to 5 μM at one or more but not all cells chosen from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, perifollicular thyroid cells and placental trophoblasts.

56. The method of claim 36, 54 or 55 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 1 μM.

57. The method of claim 36, 54 or 55 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 100 nanomolar.

58. The method of claim 36, 54, or 55 wherein said molecule has an EC_{50} or IC_{50} less than or equal to 10 nanomolar.

59. The method of claim 38, 54 or 55, wherein said molecule has an EC_{50} or IC_{50} less than or equal to 1 nanomolar.

60. The method of claim 36, 54 or 55 wherein said molecule is positively charged at physiological pH.

61. The method of claim 60 wherein said molecule is selected from the group consisting of branched or cyclic polyalkylamines, positively charged polyamino acids, and arylalkylamines.

62. The method of claim 61 wherein said branched polyamine has the formula

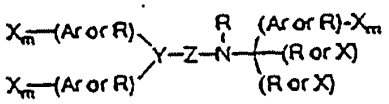
$$H_2N-(CH_2)_j-(NR_i-(CH_2)_j)_k-NH_2$$

wherein k is an integer from 1 to 10,

each j is the same or different and is an integer from 2 to 20, and

each R_i is the same or different and is selected from the group consisting of hydrogen and $-(CH_2)_j-NH_2$, wherein j is as defined above, and wherein at least one R_i is not hydrogen.

63. The method of claim 36, 54 or 55 wherein said molecule has the formula



wherein each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, and CH_3CH_2;

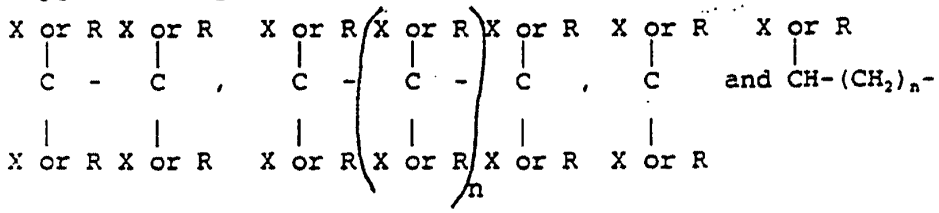
Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl,

cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen, sulfur,

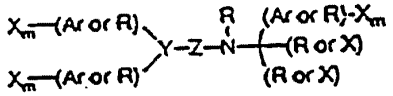


where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is either a calcimimetic or calcilytic.

64. The method of claim 63 wherein said hydrophobic entity is selected from the group consisting of phenyl, 2- 3-, or 4-pyridyl, 1- or 2-naphthyl, 1- or 2-quinolinyl, 2- or 3-indolyl, benzyl, and phenoxy.

65. The method of claim 36, 54 or 55 wherein said molecule has the formula



wherein each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, and CH_3CH_2;

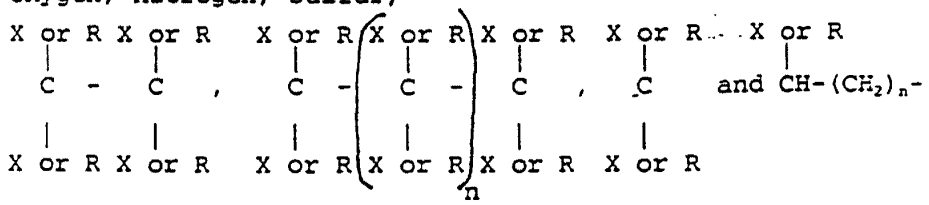
Ar is a hydrophobic entity;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, isobutyl, cyclopentyl, cyclohexyl,

cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-,3-, or 4-piperid(in)yl;

Y is selected from the group consisting of CH, nitrogen and an unsaturated carbon;

Z is selected from the group consisting of oxygen, nitrogen, sulfur,



where each n is independently between 1 and 4 inclusive, and each m is independently between 0 and 5 inclusive;

wherein said molecule is not NPS 447 or NPS 449.

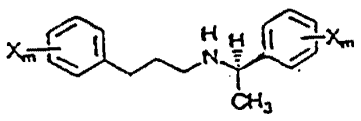
66. The method of claim 65 wherein said molecule is NPS 467 or NPS 019.

67. The method of claim 63 wherein said molecule is NPS 467 and NPS 568.

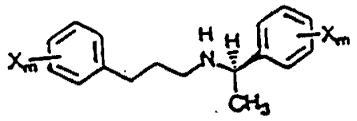
68. The method of claim 63, wherein said molecule is an R-phenylpropyl-α-phenethylamine derivative.

69. The method of claim 65, wherein said molecule is an R-phenylpropyl-α-phenethylamine derivative.

70. The method of claim 68 wherein said molecule has the formula:



71. The method of claim 69 wherein said molecule has the formula:



72. The method of claim 70 wherein each X is independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

73. The method of claim 71 wherein each X is independently selected from the group consisting of Cl, F, CF_3, CH_3, and CH_3O.

74. The method of claim 36 wherein said patient has a disease characterized by an abnormal level of one or more ions or substances, the level of which is regulated or affected by activity of one or more Ca^{2+} receptors.

75. The method of claim 74, wherein said molecule is active on a Ca^{2+} receptor of a cell selected from the group consisting of parathyroid cells, bone osteoclasts, juxtaglomerular kidney cells, proximal tubule kidney cells, keratinocytes, perifollicular thyroid cells, and placental trophoblasts.

76. The method of claim 36, wherein said patient has a disease selected from the group consisting of primary and secondary hyperparathyroidism, Paget's disease, hypercalcemia malignancy, osteoporosis and hypertension.

77. The method of claim 75 wherein said molecule is active only at one or more said Ca^{2+} receptors and not all said Ca^{2+} receptors.

78. The method of claim 36 wherein said molecule reduces the level of parathyroid hormone in the serum of said patient.

79. The method of claim 78, wherein said level of parathyroid hormone is reduced to that level present in a normal individual.

80. The method of claim 78 wherein said level is reduced to a degree sufficient to cause a decrease in plasma Ca^{2+}.

81. The method of claim 78 wherein said molecule is provided in an amount sufficient to have a therapeutically relevant effect on said patient.

82. The method of claim 36 wherein said molecule blocks an increase in $[Ca^{2+}]_i$ within the cell evoked by extracellular $[Ca^{2+}]$.

83. A method for diagnosis of a disease or condition in a patient, comprising the steps of identifying the number and/or location of one or more Ca^{2+} receptors within said patient and comparing said number and/or location with that observed in normal patients as an indication of the presence of a said disease or condition.

84. The method of claim 83 wherein said method is an immunoassay in which an antibody to a Ca^{2+} receptor is used to identify the number or location of said Ca^{2+} receptors.

85. The method of claim 83 wherein said assay comprises providing a labeled calcimimetic or calcilytic molecule which binds to a Ca^{2+} receptor.

86. The method of claim 83 wherein said disease is a cancer.

87. The method of claim 86 wherein said cancer is an ectopic tumor in the parathyroid.

88. The method of claim 83 wherein said condition is characterized by an increased number or activity of the osteoclasts in bone.

89. A method for treatment of ostioporosis, hyperparathyroidism or hypertension, comprising the steps of administering a therapeutically effective amount of a compound which mimics the activity of extracellular Ca^{2+} by evoking an increase in $[Ca^{2+}]_i$ in a cell, or blocks an increase in $[Ca^{2+}]_i$ in the presence of extracellular Ca^{2+}.

90. A method for identifying a compound useful as a therapeutic agent, comprising the steps of screening a potentially useful molecule for its ability to mimic the activity of extracellular Ca^{2+} in a cell, or to block an increase in $[Ca^{2+}]_i$ (elicited by extracellular Ca^{2+}), and determining whether said molecule has EC_{50} of less than or equal to 5 μM.

91. A recombinant Ca^{2+} receptor.

92. A cell comprising a recombinant Ca^{2+} receptor.

93. A method for identifying a useful calcimimetic compound comprising the step of identifying a compound which mimics one or more activities of Ca^{2+} at a first Ca^{2+} receptor but not at a second Ca^{2+} receptor.

94. A method for identifying a useful calcilytic compound comprising the step of identifying a compound which blocks one or more activities of Ca^{2+} at a first Ca^{2+} receptor but not at a second Ca^{2+} receptor.

95. The method of claim 93 or 94 wherein said method includes providing a recombinant Ca^{2+} receptor useful in said identifying step.

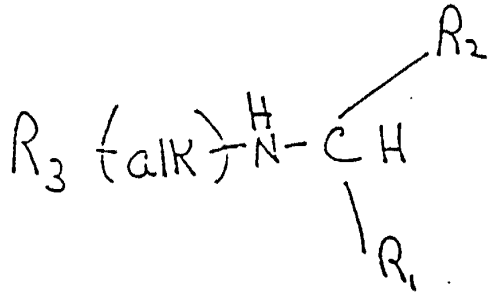
96. Purified nucleic acid encoding a Ca^{2+} receptor.

97. The compounds NPS 459, NPS 467, NPS 544 and NPS 568 and pharmaceutically acceptable acid addition salts and complexes thereof.

98. The pharmaceutical composition of claim 25 wherein said molecule is NPS 447, NPS 448, NPS 449,

NPS 456 or NPS 551.

99. A compound of the formula:



wherein alk is straight or branched-chain alkylene of from 1 to 6 carbon atoms; R_1 is lower alkyl of from 1 to 3 carbon atoms or lower haloalkyl of from 1 to 3 carbon atoms substituted with from 1 to 7 halogen atoms; and R_2 and R^3 are independently selected monocyclic or bicyclic carbocyclic aryl or cycloalkyl groups, having 5- or 6-membered rings optionally substituted with 1 to 5 substituents independently selected from lower alkyl of 1 to 3 carbon atoms, lower haloalkyl of 1 to 3 carbon atoms substituted with 1 to 7 halogen atoms, lower alkoxy of 1 to 3 carbon atoms, halogen, nitro, amino, alkylamino, amido, lower alkylamido of 1 to 3 carbon atoms, cyano, hydroxy, acyl of 2 to 4 carbon atoms or lower thioalkyl of 1 to 3 carbon atoms and pharmaceutically acceptable salts and acid addition salts thereof.

100. A compound according to claim 99 wherein alk is n-propylene.

101. A compound according to claim 100 wherein R_1 is methyl.

102. A compound according to claim 101 wherein R_2 and R_3 are independently selected optionally substituted phenyl groups.

103. A isolated nucleic acid encoding an inorganic-ion receptor or a unique fragment thereof.

104. An isolated nucleic acid as claimed in claim 103 encoding a human inorganic-ion receptor or a unique fragment thereof.

105. An isolated nucleic acid as claimed in claim 103 encoding an inorganic-ion receptor that hybridizes to Sequence 1.

106. An isolated nucleic acid as claimed in claim 103 encoding an inorganic-ion receptor that hybridizes to BoPCaR 1 or a unique fragment thereof.

107. An isolated nucleic acid claimed in claim 103 encoding a calcium receptor or a unique fragment thereof.

108. An isolated nucleic acid as claimed in claim 107 encoding a calcium receptor that is expressed in tissues or cells selected from the group consisting of:

parathyroid, vascular, kidney, epidermis, thyroid, osteoclast, intestine, mammary, trophoblast platelet, gastrin secreting, glucagon secreting, cardiac, and brain, and unique fragments thereof.

109. An isolated nucleic acid as claimed in claim 103, wherein the nucleic acid is a fragment encoding the extracellular binding domain but substantially free of transmembrane and intracellular domain portions.

110. A isolated nucleic acid as claimed in claim 103, wherein the nucleic acid is a fragment encoding the intracellular domain but substantially free of transmembrane and extracellular binding domains.

111. An isolated nucleic acid as claimed in claim 109, wherein the fragment encoding the extracellular binding domain is transcriptually coupled to nucleic acid encoding transmembrane and intracellular coding domains of a noninorganic-ion receptor.

112. An isolated nucleic acid as claimed in claim 103, comprising BoPCaR 1 or a unique fragment thereof.

113. The isolated nucleic acid of any one of claims 103, 104, 105, and 106 under the transcriptional control of an exogenous promoter.

114. A recombinant cell expressing the nucleic acid of any one of claims 103, 104, 105, 106 and 107.

115. An isolated inorganic-ion receptor, or a unique fragment thereof.

116. An isolated inorganic-ion receptor as claimed in claim 115 that is a human inorganic-ion receptor, or a unique fragment thereof.

117. An isolated inorganic-ion receptor as claimed in claim 115 or 116 wherein the receptor is a calcium receptor.

118. An isolated inorganic-ion receptor as claimed in claim 115 that is a calcium receptor expressed in tissues or cells selected from the group consisting of:

parathyroid, vascular, kidney, epidermis, thyroid, osteoclast, intestine, mammary, trophoblast platelet, gastrin secreting, glucagon secreting, cardiac, and brain, and unique fragments thereof.

119. An isolated inorganic-ion receptor as claimed in claim 115 that is the expression product of BoPCar 1, or a unique fragment thereof.

120. An isolated inorganic-ion receptor as claimed in claim 115 that is a human parathyroid calcium receptor or unique fragment thereof.

121. An isolated inorganic-ion receptor as claimed in claim 13 that is an extracellular, ion-bonding fragment substantially free of intracellular and transmembrane domains.

122. An isolated inorganic-ion receptor as claimed in claim 111 that is a calcium-binding fragment.

123. Purified antibodies to an inorganic-ion receptor.

124. Purified antibodies as claimed in claim 123 to an extracellular domain of an inorganic-ion receptor.

125. Purified antibodies as claimed in claim 123 to a calcium receptor.

126. Purified antibodies as claimed in claim 125 to a calcium receptor expressed in tissue or cells selected from the group consisting of:

parathyroid, vascular, kidney, epidermis, thyroid, osteoclast, intestine, mammary, trophoblast platelet, gastrin secreting, glucagon secreting, cardiac, and brain, and unique fragments thereof.

127. Purified antibodies as claimed in claim 124 to a calcium receptor.

128. Purified antibodies as claimed in claim 127 to an allosteric site of a calcium receptor.

129. Purified antibodies as claimed in claims 124, 125, 126 or 127 coupled to a toxin.

130. An inorganic-ion receptor binding agent coupled to a toxin.

131. An agent as claimed in claim 131 wherein the agent selectively binds a calcium receptor.

132. A transgenic, nonhuman mammal containing a transgene encoding an inorganic-ion receptor or a unique fragment thereof.

133. A transgenic mammal as claimed in claim 132 wherein the transgene encodes a calcium receptor or a unique fragment thereof.

134. A transgenic, nonhuman mammal containing a transgene that alters the expression of an inorganic-ion receptor.

135. A transgenic mammal as claimed in claim 134 wherein the transgene inactivates the expression of the inorganic-ion receptor.

136. A transgenic mammal as claimed in claim 134 wherein the transgene up-regulates or down-regulates the expression of the inorganic-ion receptor.

137. A transgenic mammal as claimed in claims 134, 135 or 136 wherein the transgene alters the expression of a calcium receptor.

138. A method for identifying nucleic acid encoding an inorganic-ion receptor comprising

contacting a library of nucleic acid with a probe selected from the group consisting of:

(a) BoPCaR 1

(b) a unique fragment of BoPCaR 1

(c) the human homolog of BoPCaR 1

(d) a set of degenerate probes of any of (a), (b) or (c) and identifying sequences within the library that hybridize to the probe.

139. A method as claimed in claim 138 further comprising

isolating the nucleic acid encoding the inorganic-ion receptor.

140. A method as claimed in claim 138 further comprising

identifying a tissue in which the inorganic-ion receptor is expressed.

141. A method for modulating the expression of an inorganic-ion receptor comprising

contacting a cell with an agent that binds to a cellular component selected from the group consisting of:

a promoter;

a regulatory agent acting on a promoter;

an agent capable of binding to the receptor; and

a nucleic acid encoding the receptor or a unique fragment thereof.

142. A method as claimed in claim 141 wherein the cell is contacted with an antisense nucleic acid.

143. A method as claimed in claim 141 wherein the cell is contacted with a DNA constructed and arranged to combine through homologous recombination with an endogenous gene encoding the receptor and to inactivate expression of the receptor in the cell.

144. A method for killing a cell expressing an inorganic-ion receptor comprising

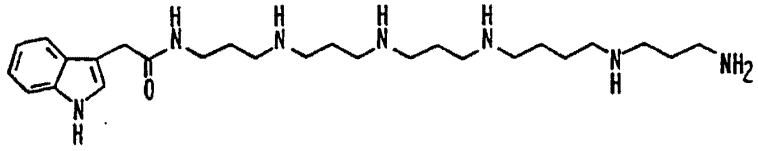
contacting the cell with an agent capable of binding to the inorganic-ion receptor, wherein the agent is conjugated to a toxin.

145. A method for identifying an agent that is an agonist or an antagonist of an inorganic-ion receptor, comprising

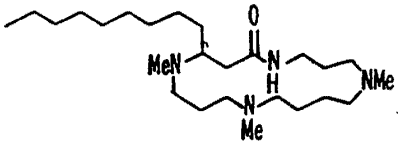
contacting a cell containing a recombinant nucleic acid encoding an inorganic-ion receptor with the agent and detecting a change in the cell.

146. A method for identifying an agent that binds to an inorganic-ion receptor comprising an assay employing an isolated inorganic-ion receptor.

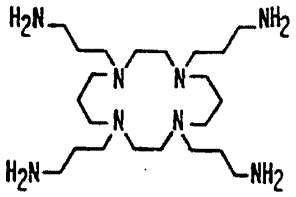
FIG. 1a.



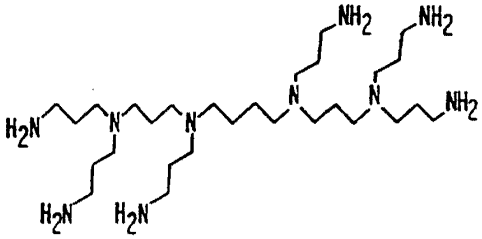
NPS 019



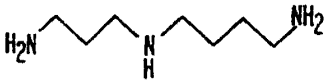
BUDMUNCHIAMINE A (+3)



NPS 383 (+8 EC_{50}=150μM)

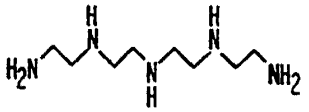


NPS 381 (+10 EC_{50}=100μM)

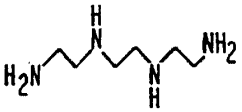


SPERMIDINE +3

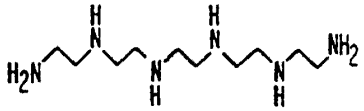
1,12-DIAMINODODECANE -2
(DADD)



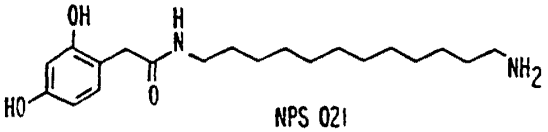
TETRAETHYLENEPENTAMINE (+5 EC_{50}=2500μM)



TRIETHYLENETETRAMINE (+4 EC_{50}=8000μM)

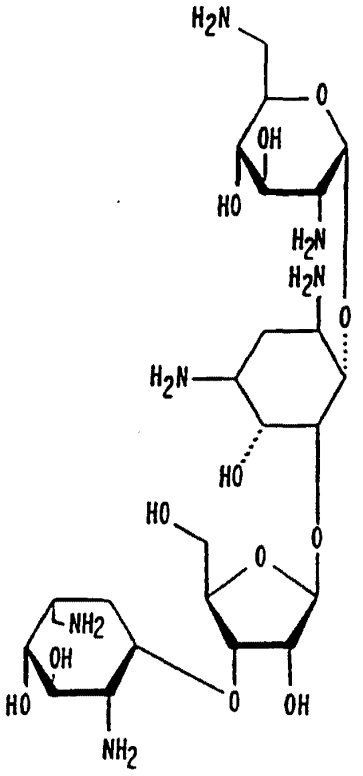


PENTAETHYLENEHEXAMINE (+6 EC_{50}=500μM)

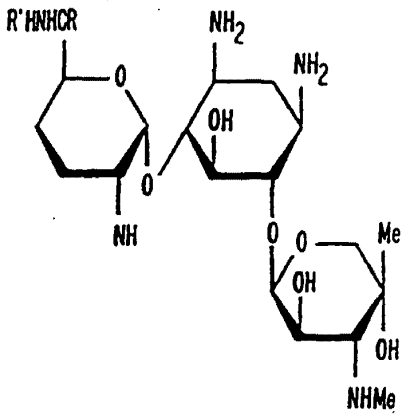


NPS 021

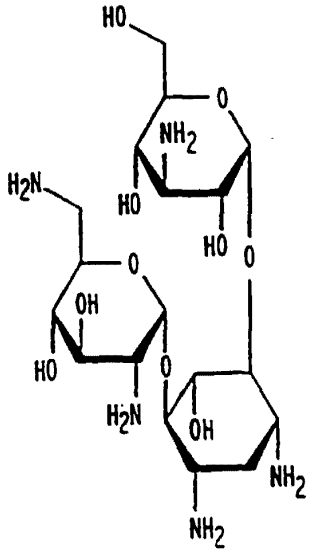
SUBSTITUTE SHEET



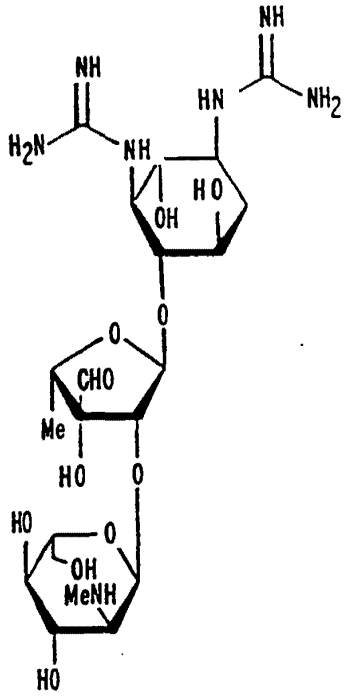
Neomycin B (+6)



Gentamicin (Complex +5)
C1 R = R' = Me
C2 R = Me; R' = H
C1aR = R' = H



Bekanamycin (+5)



Streptomycin (+3)

FIG. 1b.

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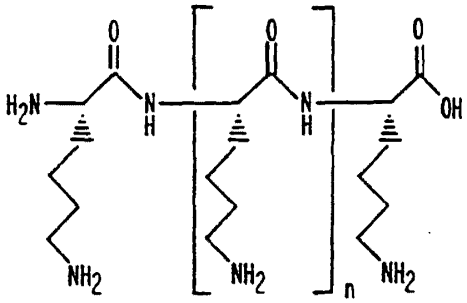
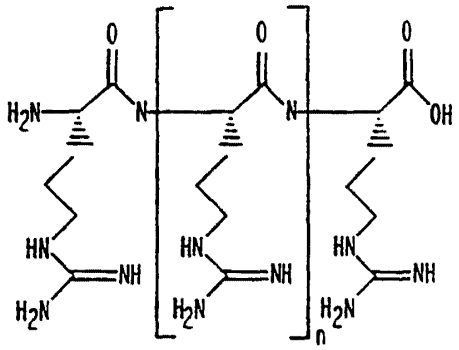


poly-Lys (27kD) n=210 +210
poly-Lys (14kD) n=110 +110
poly-Lys (38kD) n=55 +55

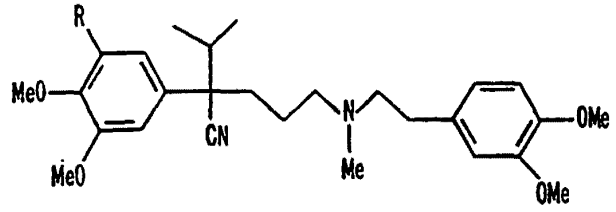


poly-Arg (100kD) n=640 +640
poly-Arg (40kD) n=256 +256

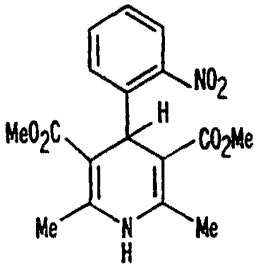
PROTAMINE +21

H N-Pro-Arg-Arg-Arg-Arg-Ser-Ser-Ser-Arg-Pro-Val-Arg-Arg-Arg-Arg-Arg-
Pro-Arg-Val-Ser-Arg-Arg-Arg-Arg-Arg-Arg-Gly-Gly-Arg-Arg-Arg-Arg-OH

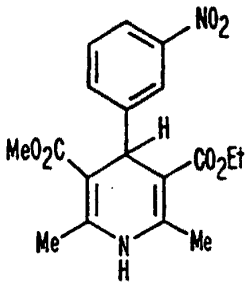
FIG. Ic.



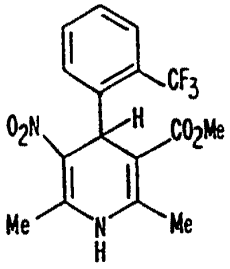
VERAPAMIL R=H +1
D-600 R=OMe +1



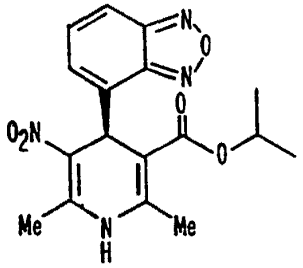
NIFEDIPINE +1



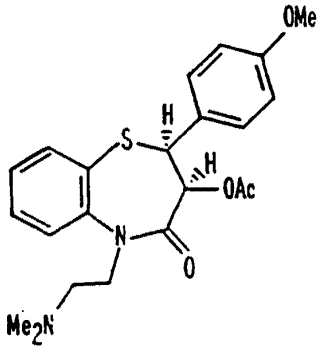
NITRENDIPINE +1



BAY K 8644 +1



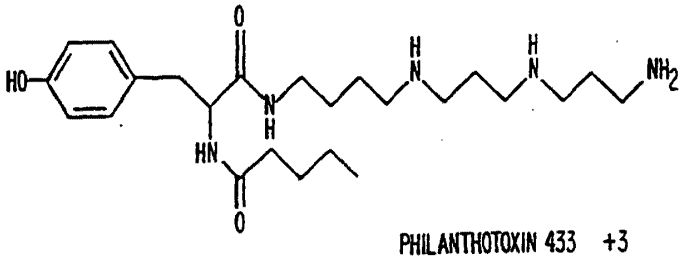
202-791 +1



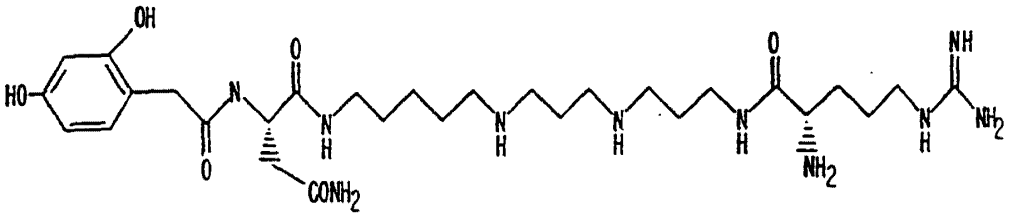
DILTIAZEM +1

FIG. 1d.

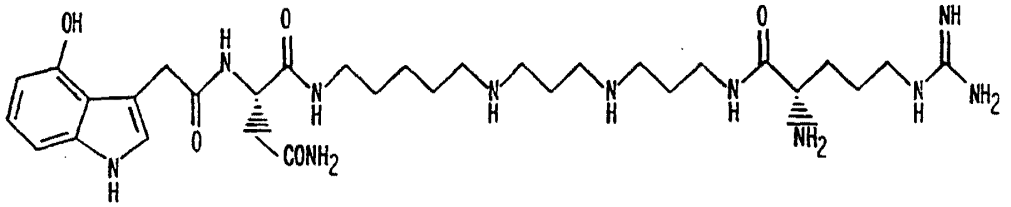
SUBSTITUTE SHEET



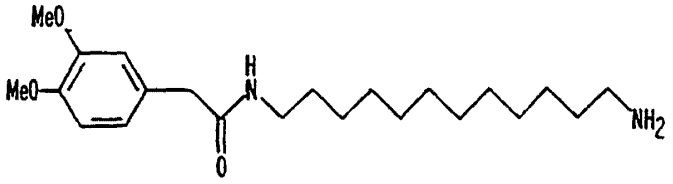
PHILANTHOTOXIN 433 +3



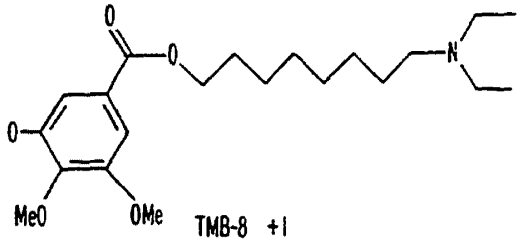
ARGIOTOXIN 636 +4



ARGIOTOXIN 659 +4



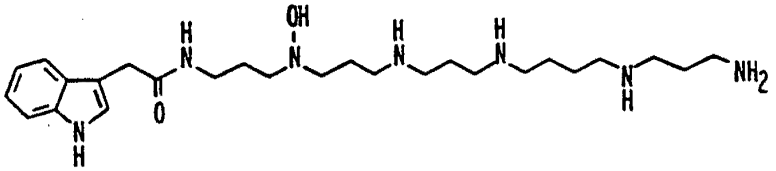
NPS 384

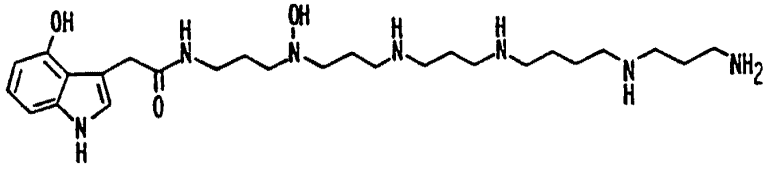


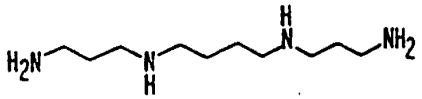
TMB-8 +1

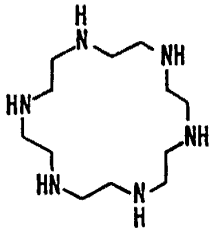
FIG. 1e.

SUBSTITUTE SHEET









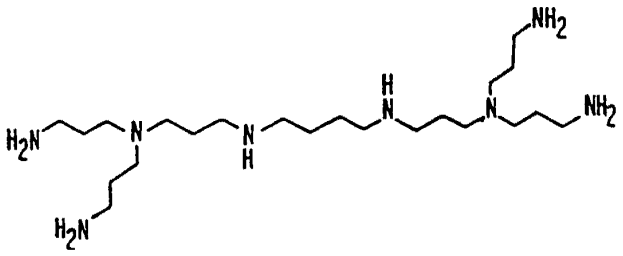


FIG. 1f.

FIG. 2.

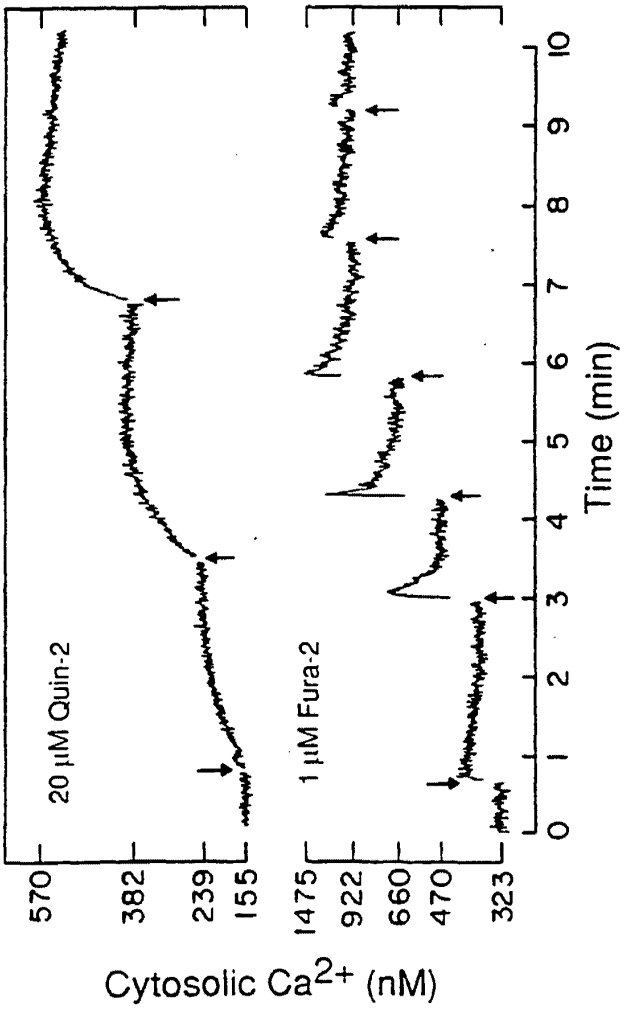

20 μM Quin-2
1 μM Fura-2
570
382
239
155
1475
922
660
470
323
0
1
2
3
4
5
6
7
8
9
10
Time (min)
Cytosolic Ca^{2+} (nM)

FIG. 3a.

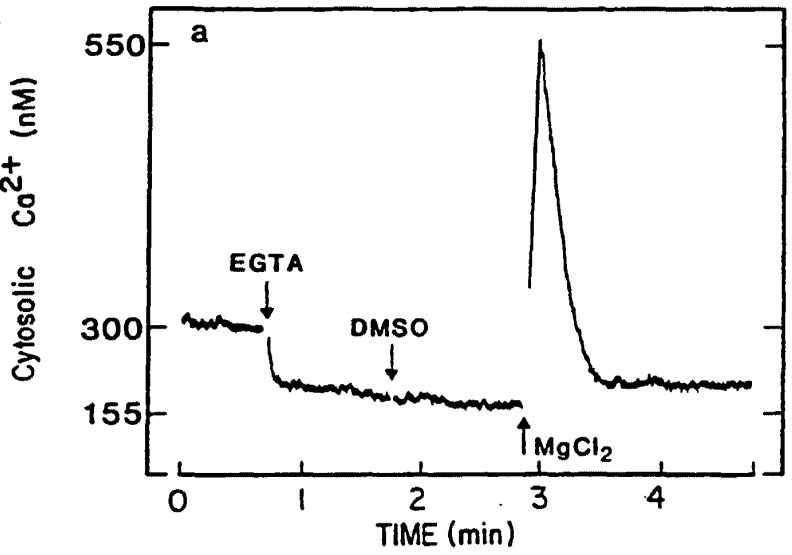


FIG. 3b.

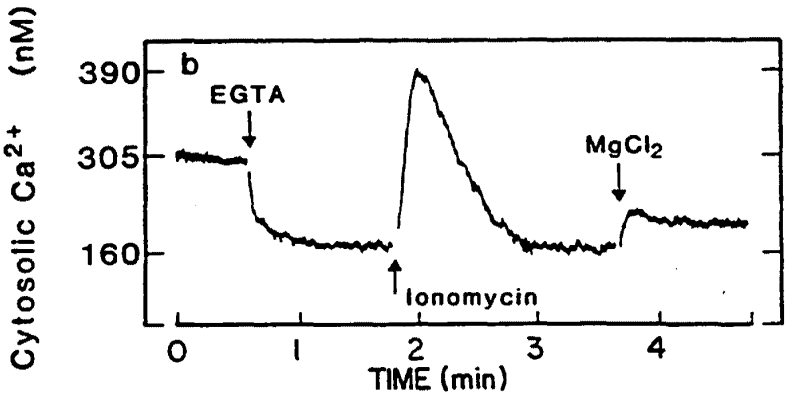
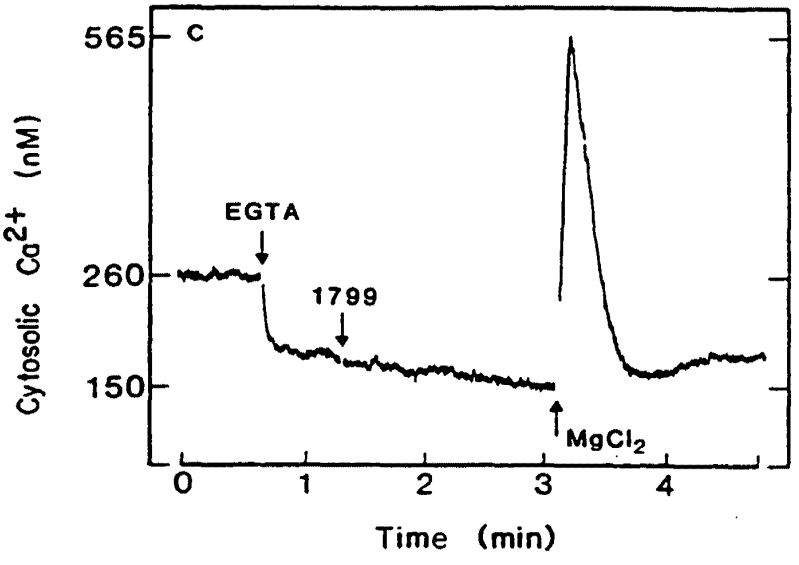


FIG. 3c.



SUBSTITUTE SHEET

FIG. 4a.

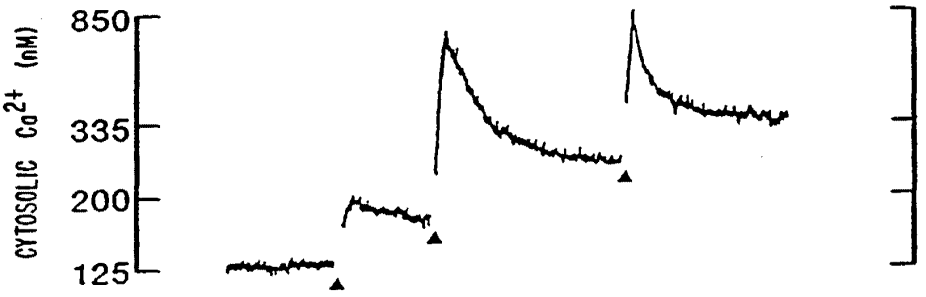


FIG. 4b.

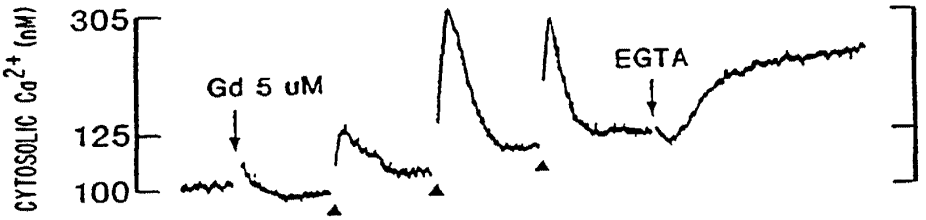


FIG. 4c.

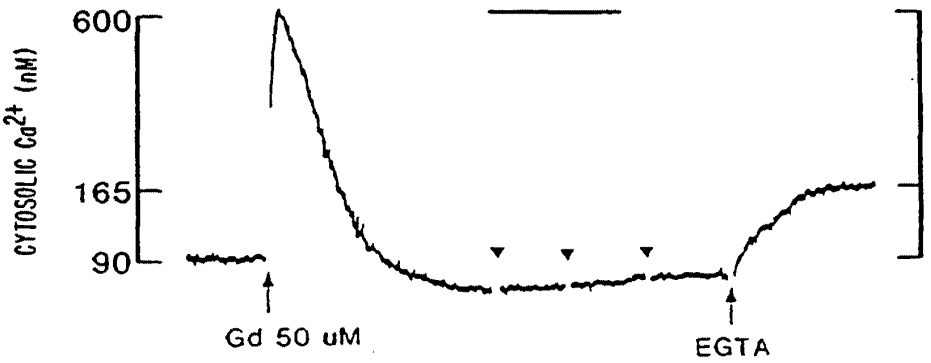


FIG. 5c.

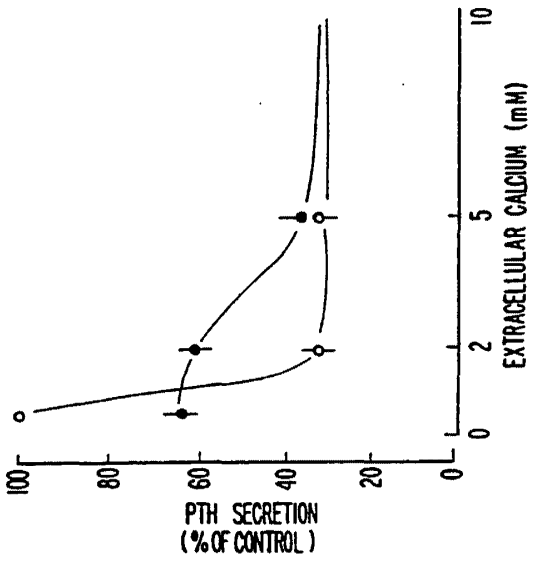


FIG. 5b.

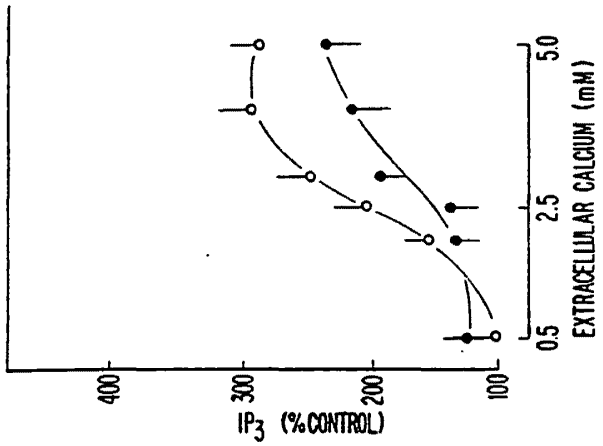
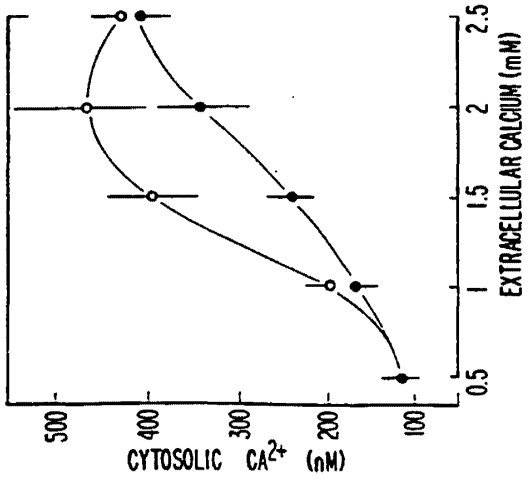


FIG. 5a.



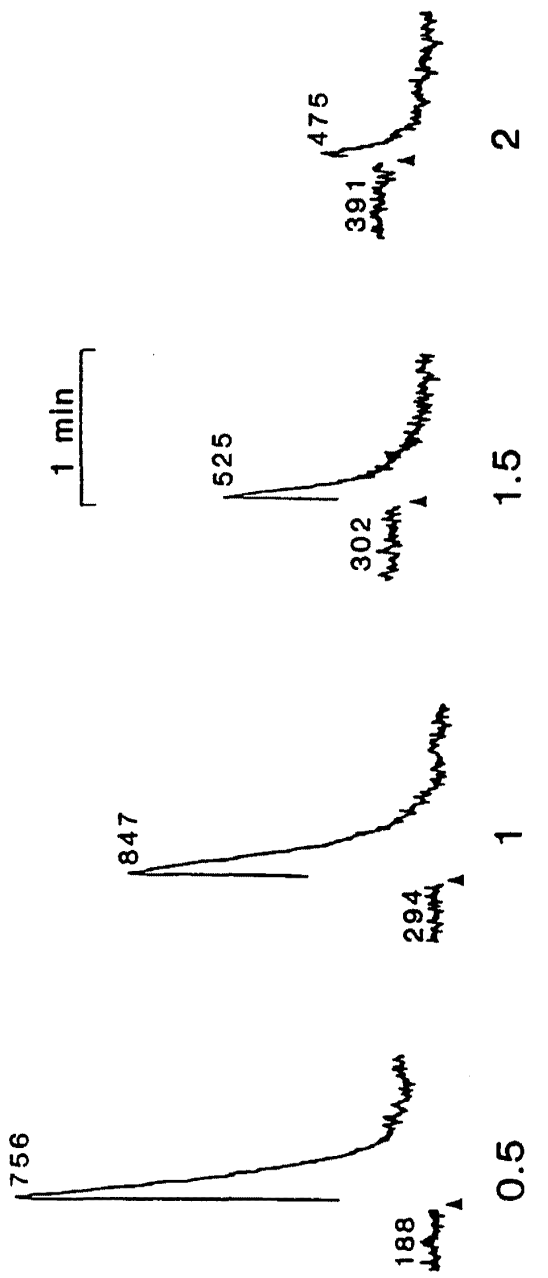

1 min
756
188
0.5
847
294
1
525
302
1.5
475
391
2
Extracellular calcium (mM)

FIG. 6.

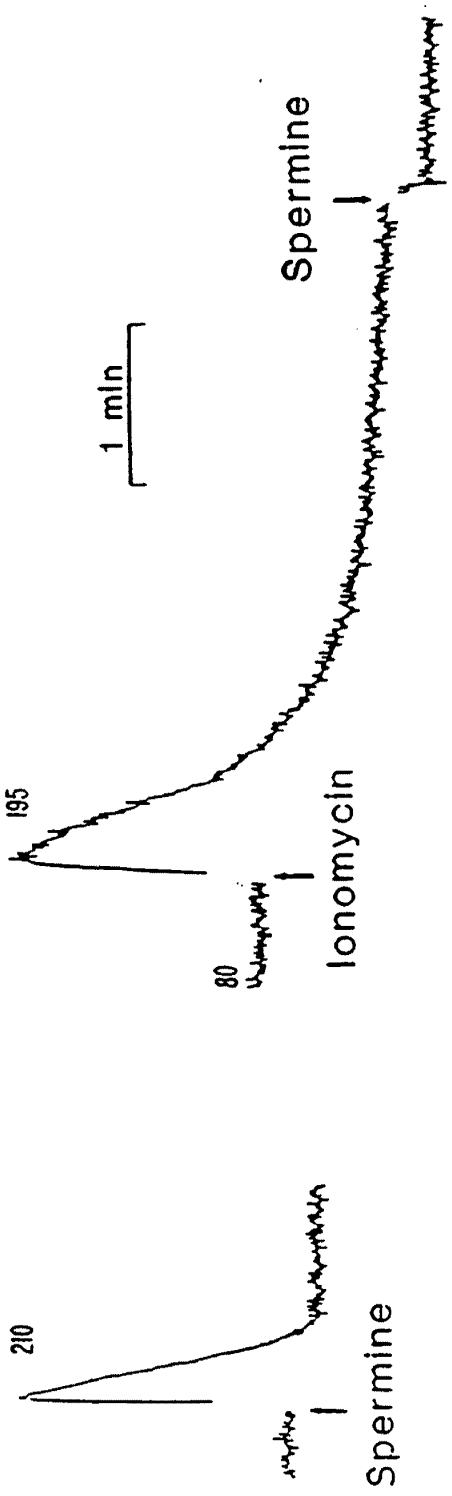


FIG. 7.

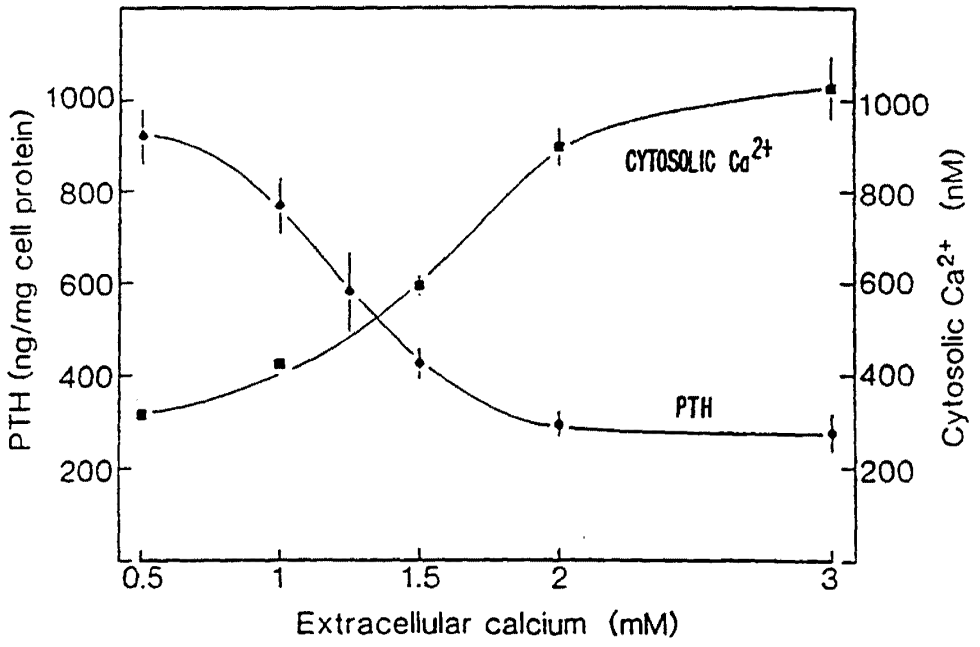

PTH (ng/mg cell protein)
1000
800
600
400
200
CYTOSOLIC Ca^{2+}
PTH
Cytosolic Ca^{2+} (nM)
1000
800
600
400
200
0.5
1
1.5
2
3
Extracellular calcium (mM)

FIG. 8a.

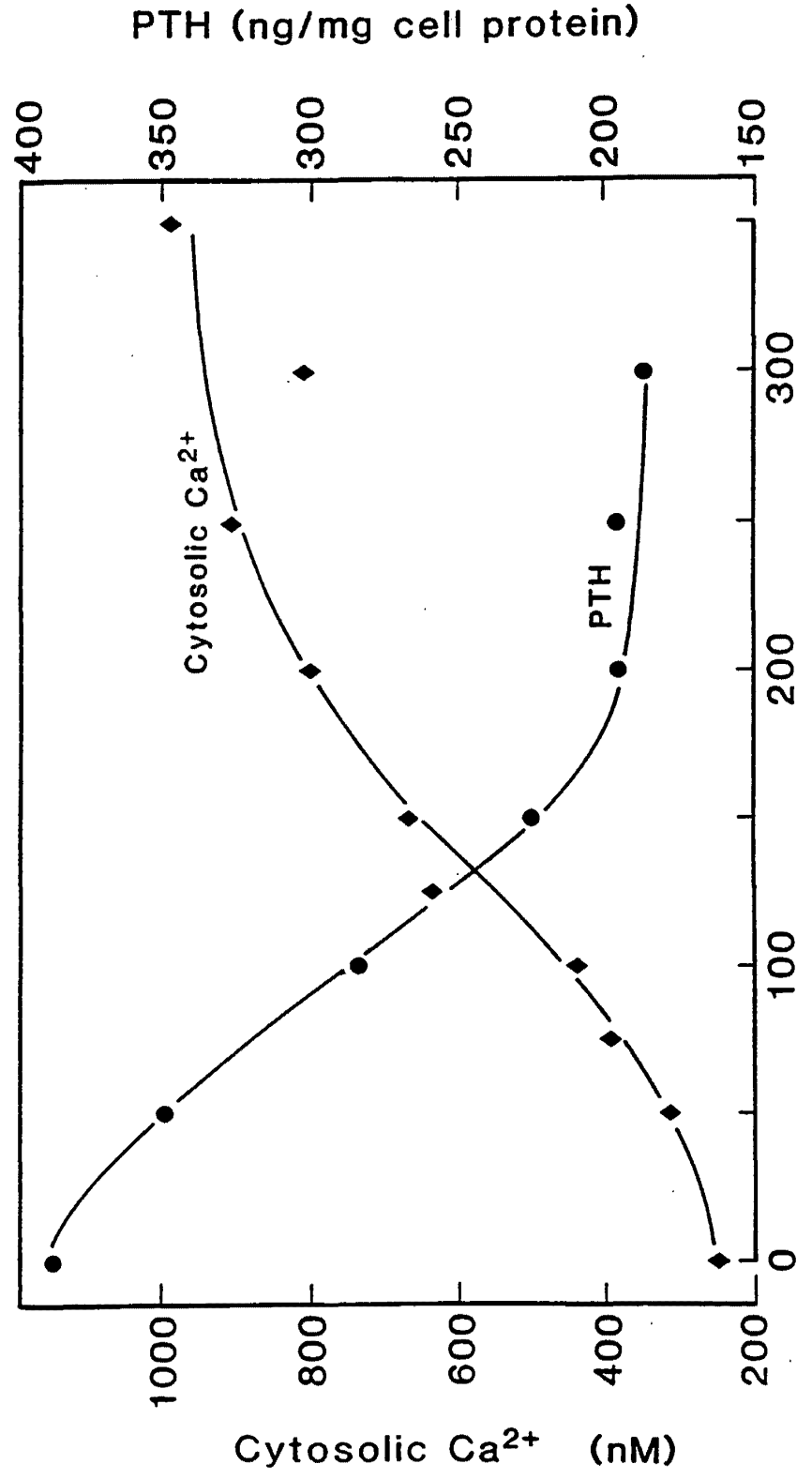

PTH (ng/mg cell protein)
400
350
300
250
200
150
Cytosolic Ca^{2+}
PTH
300
200
100
0
Spermine (μM)
1000
800
600
400
200
Cytosolic Ca^{2+} (nM)

FIG. 8b.

FIG. 9a.

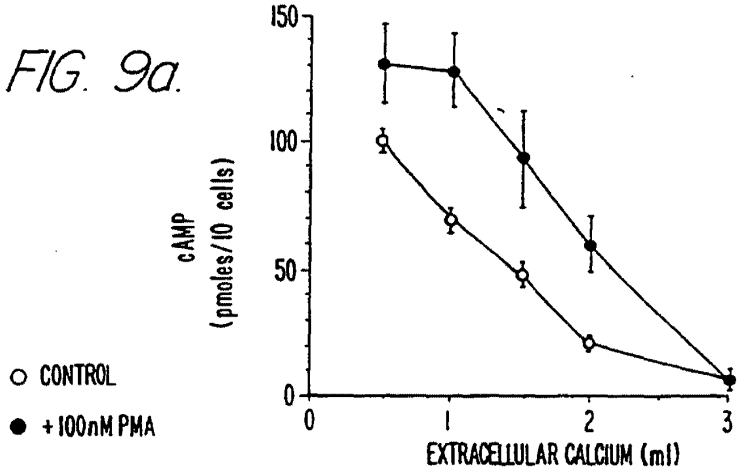


FIG. 9b.

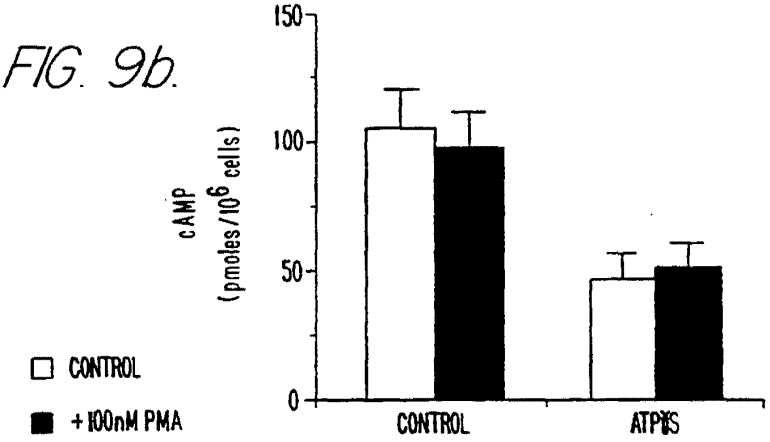
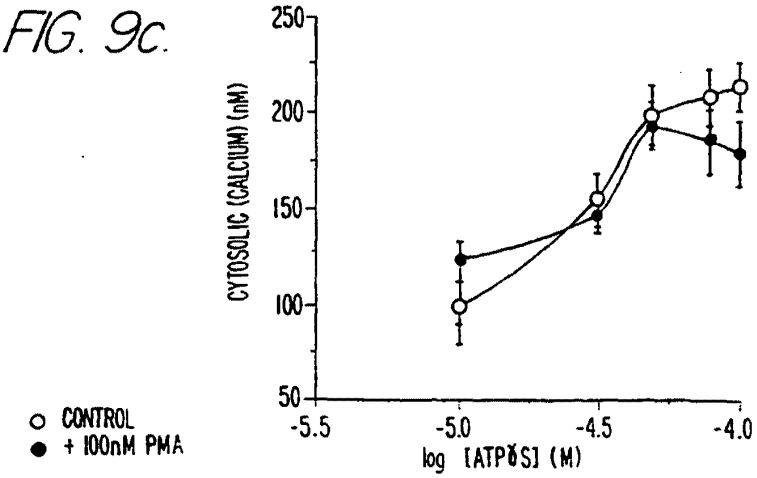


FIG. 9c.



SUBSTITUTE SHEET

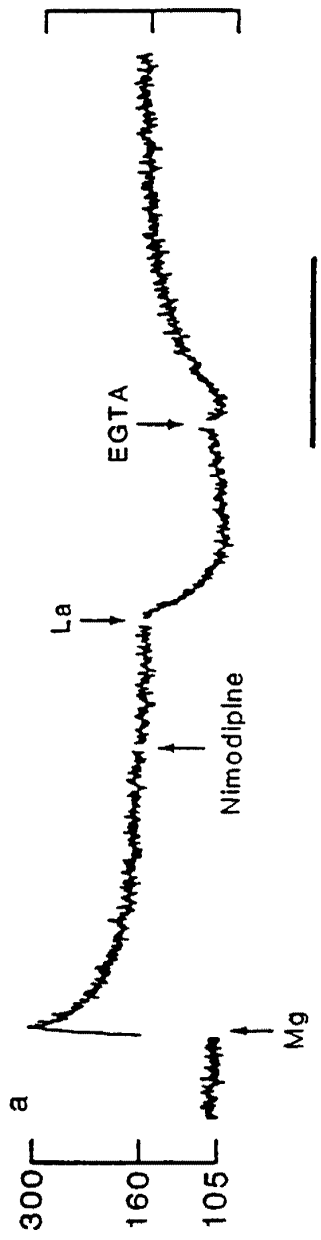


FIG. 10a.

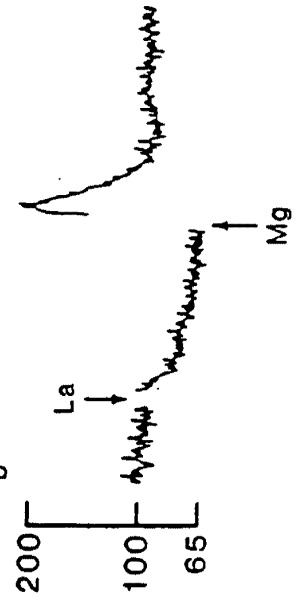


FIG. 10b.

FIG. 10c.

FIG. 11a.

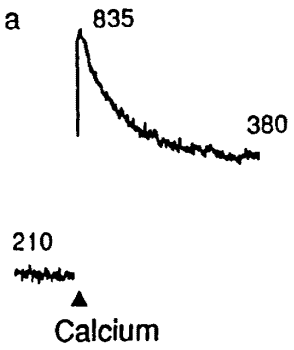


FIG. 11b.

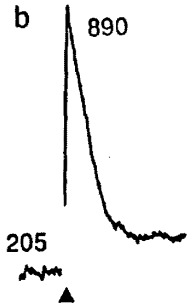


FIG. 11c.

FIG. 11d.

FIG. 11e.

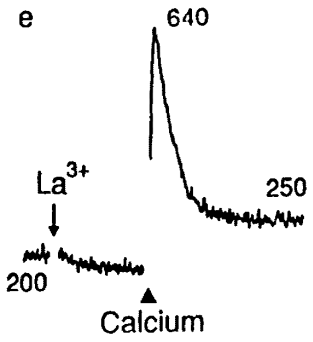


FIG. 11f.

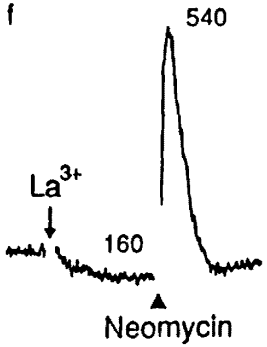


FIG. 11g.

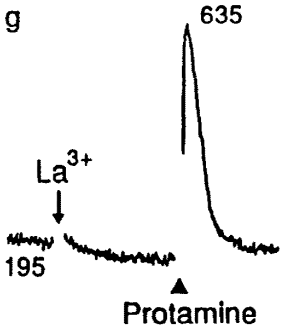


FIG. 11h.

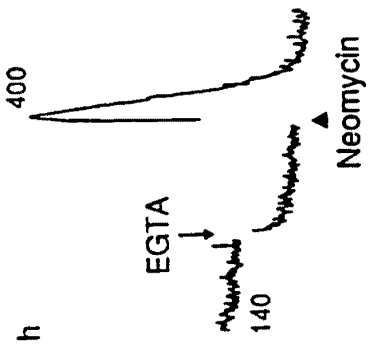


FIG. 11i.

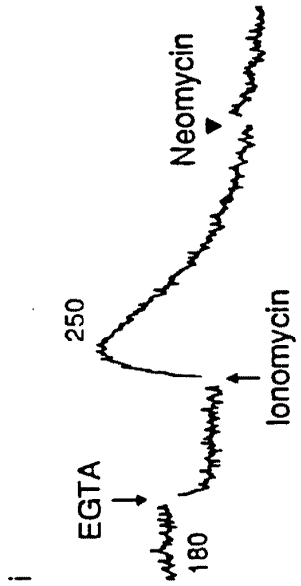


FIG. 12.

FIG. 13a.

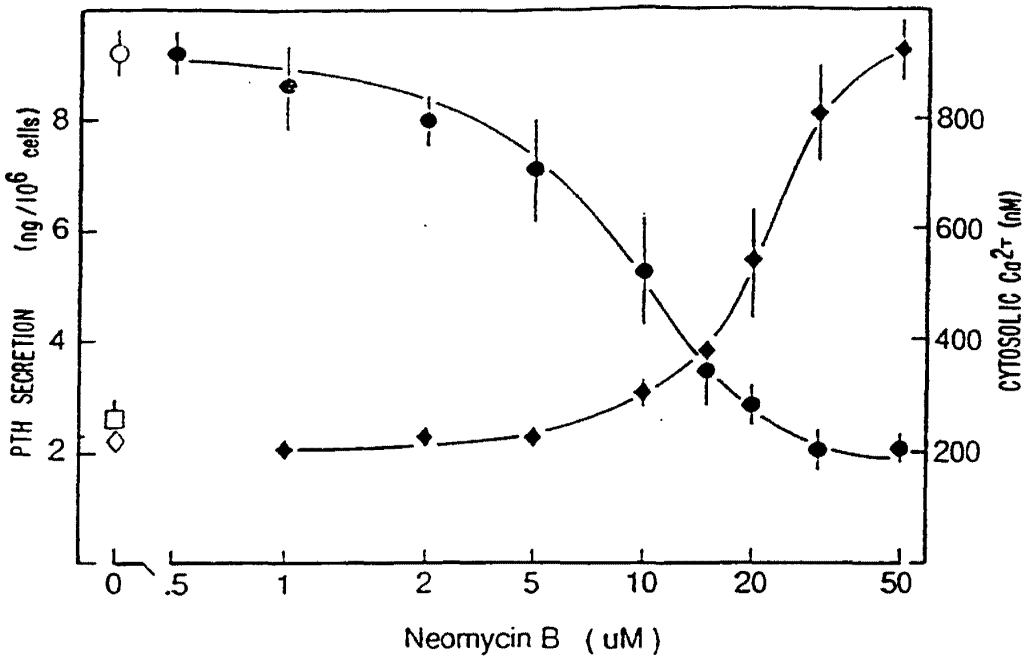
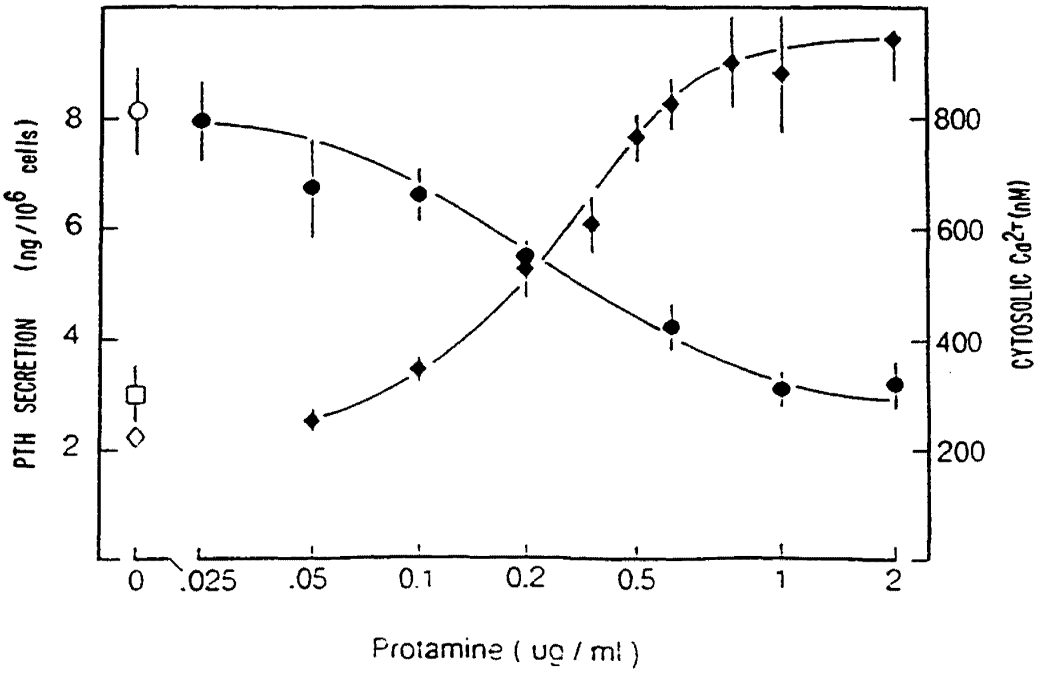
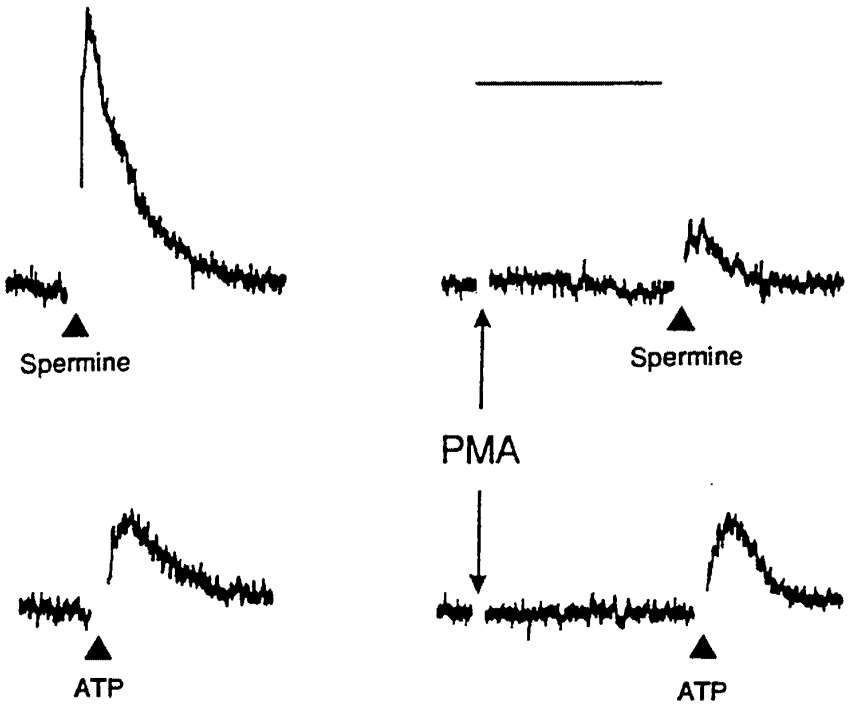


FIG. 13b.



SUBSTITUTE SHEET

FIG. 14.



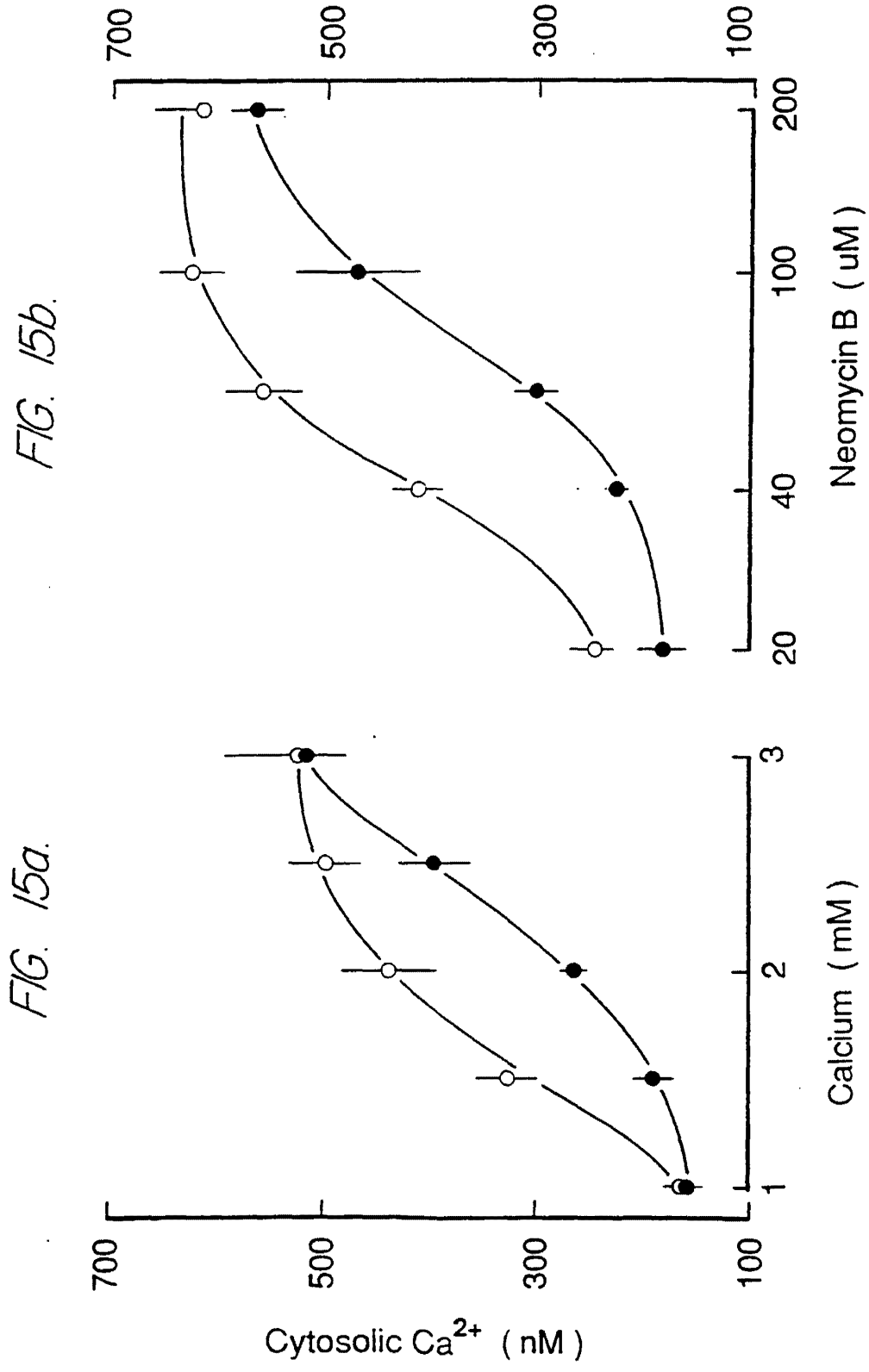


FIG. 15b.

FIG. 15a.

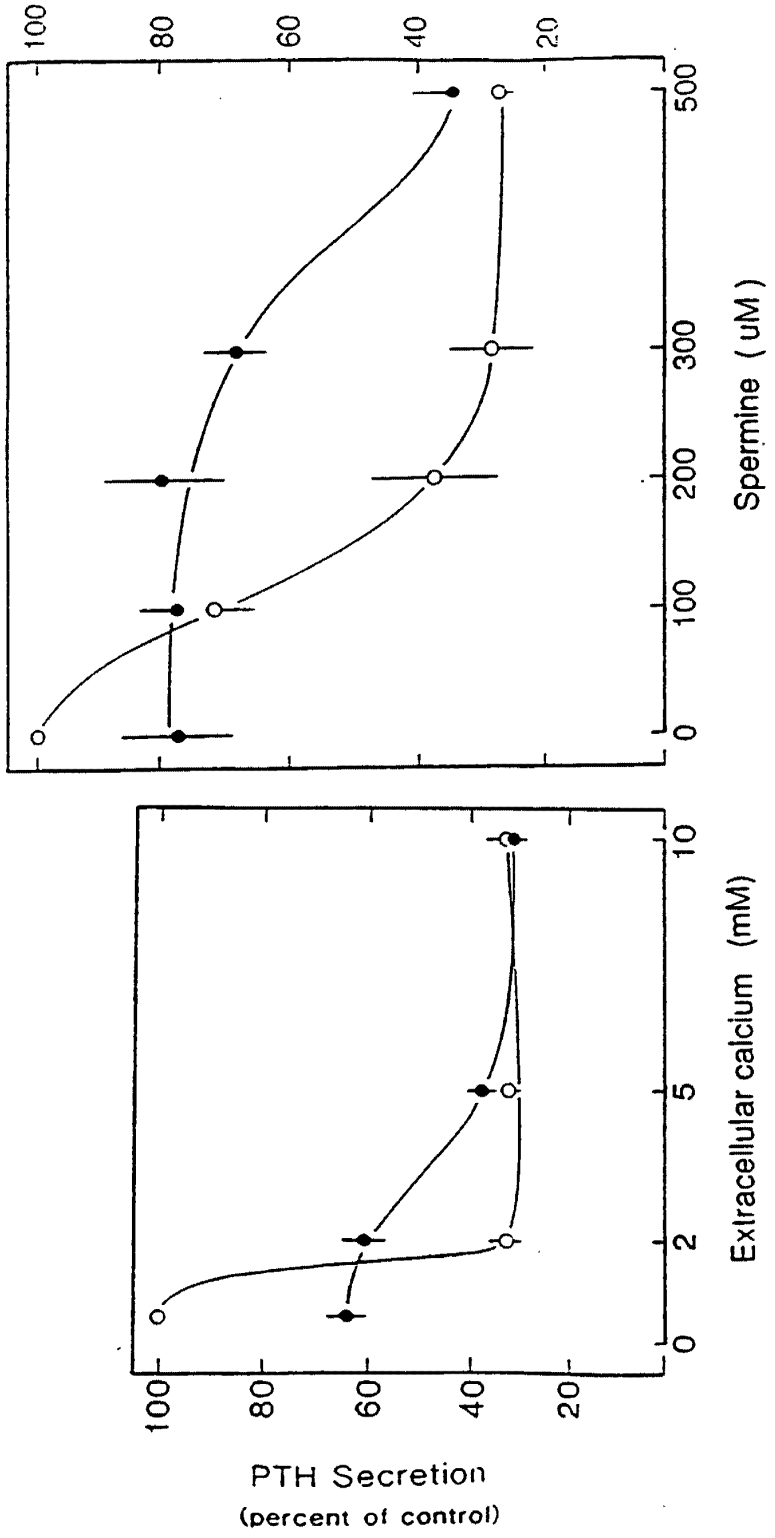


FIG. 16b.

FIG. 16a.

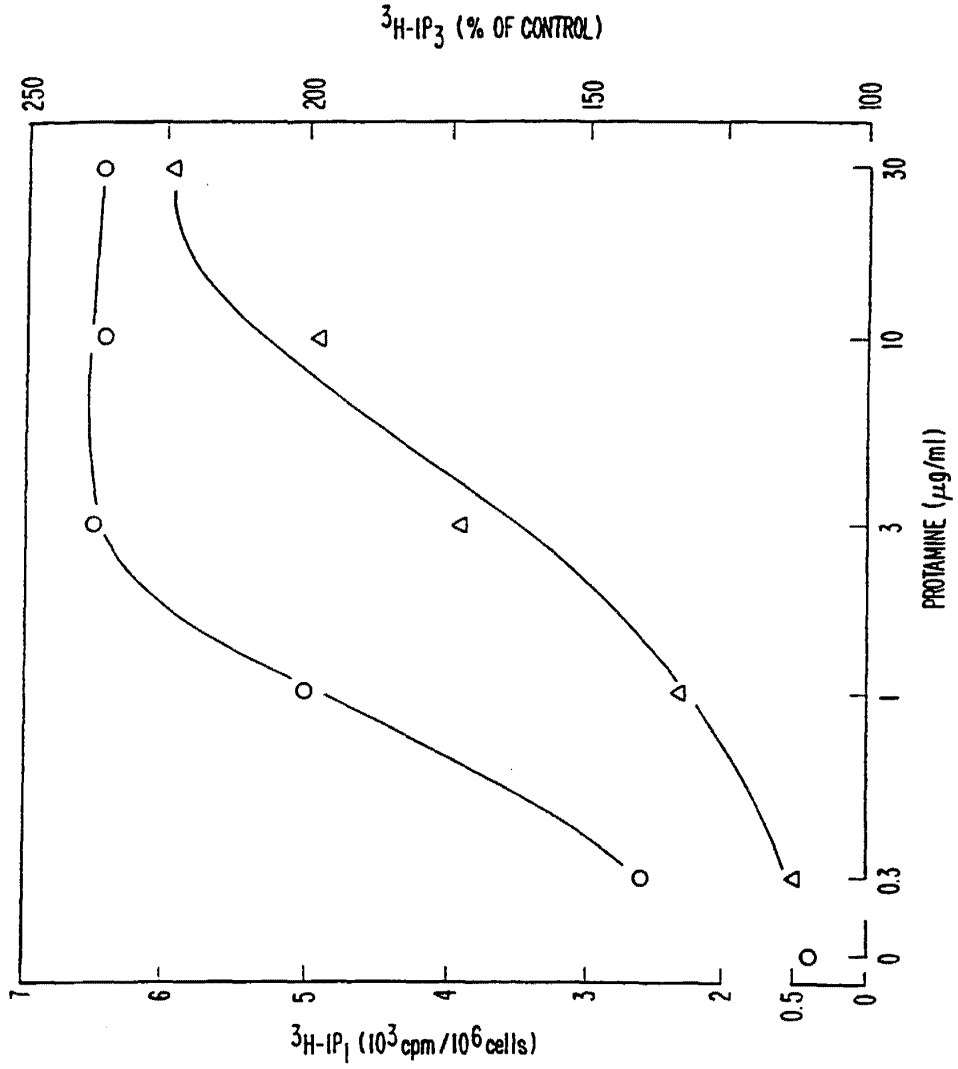

^{3}H-IP$_3$ (% OF CONTROL)
250
200
150
100
30
10
3
1
0.3
0
0.5
0
PROTAMINE (μg/ml)
7
6
5
4
3
2
^{3}H-IP$_1$ (10^3cpm/10^6cells)

FIG. 17.

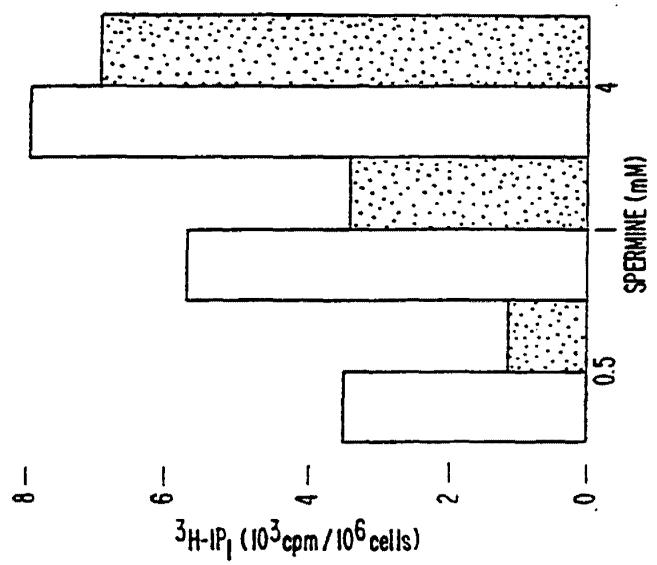


FIG. 18b.

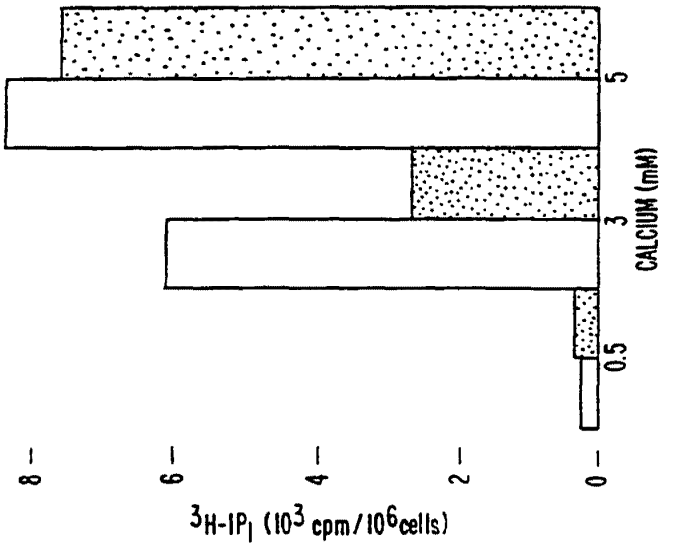


FIG. 18a.

SUBSTITUTE SHEET

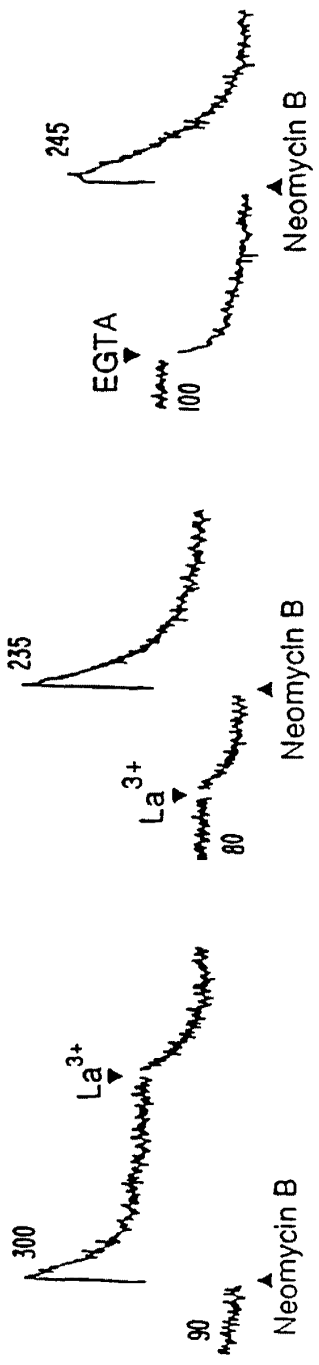
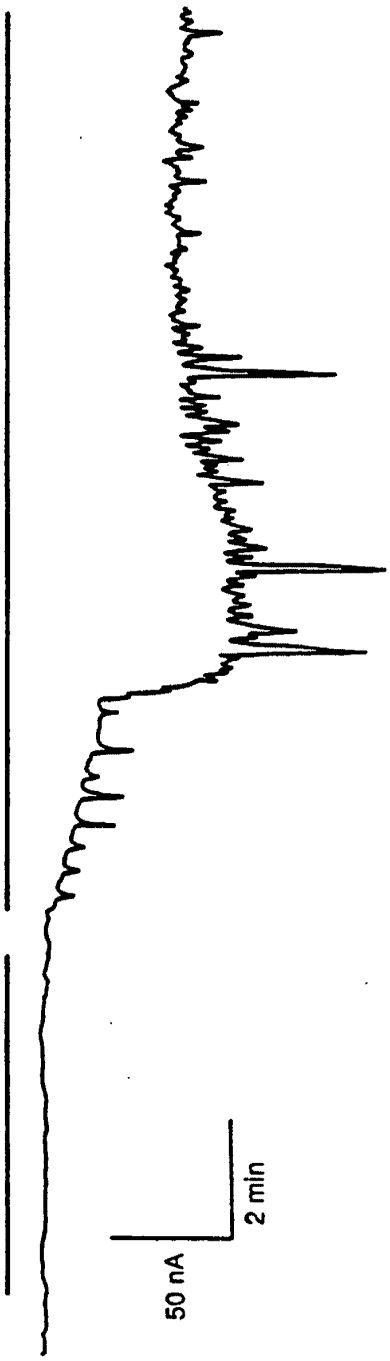

245
Neomycin B
EGTA
100
235
Neomycin B
La3+
80
300
La3+
Neomycin B
90

FIG. 19.

FIG. 20a.

FIG. 20b.



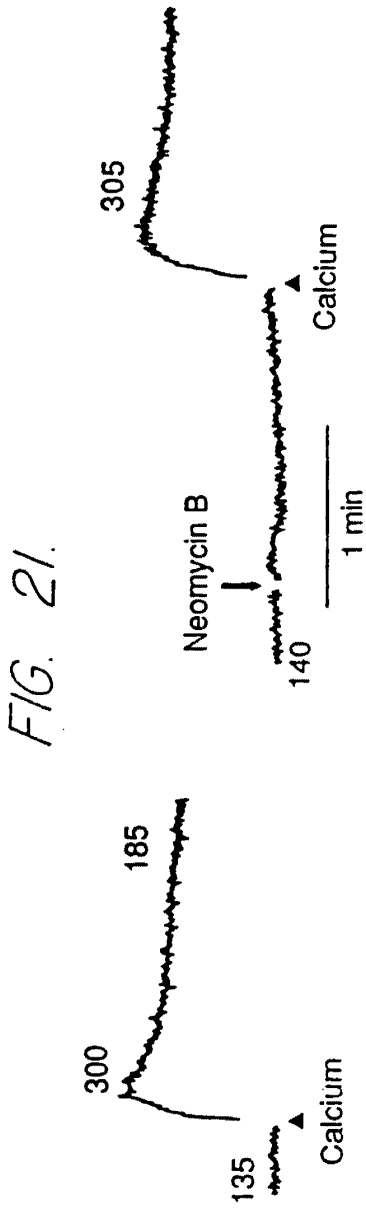


FIG. 21.

FIG. 22.

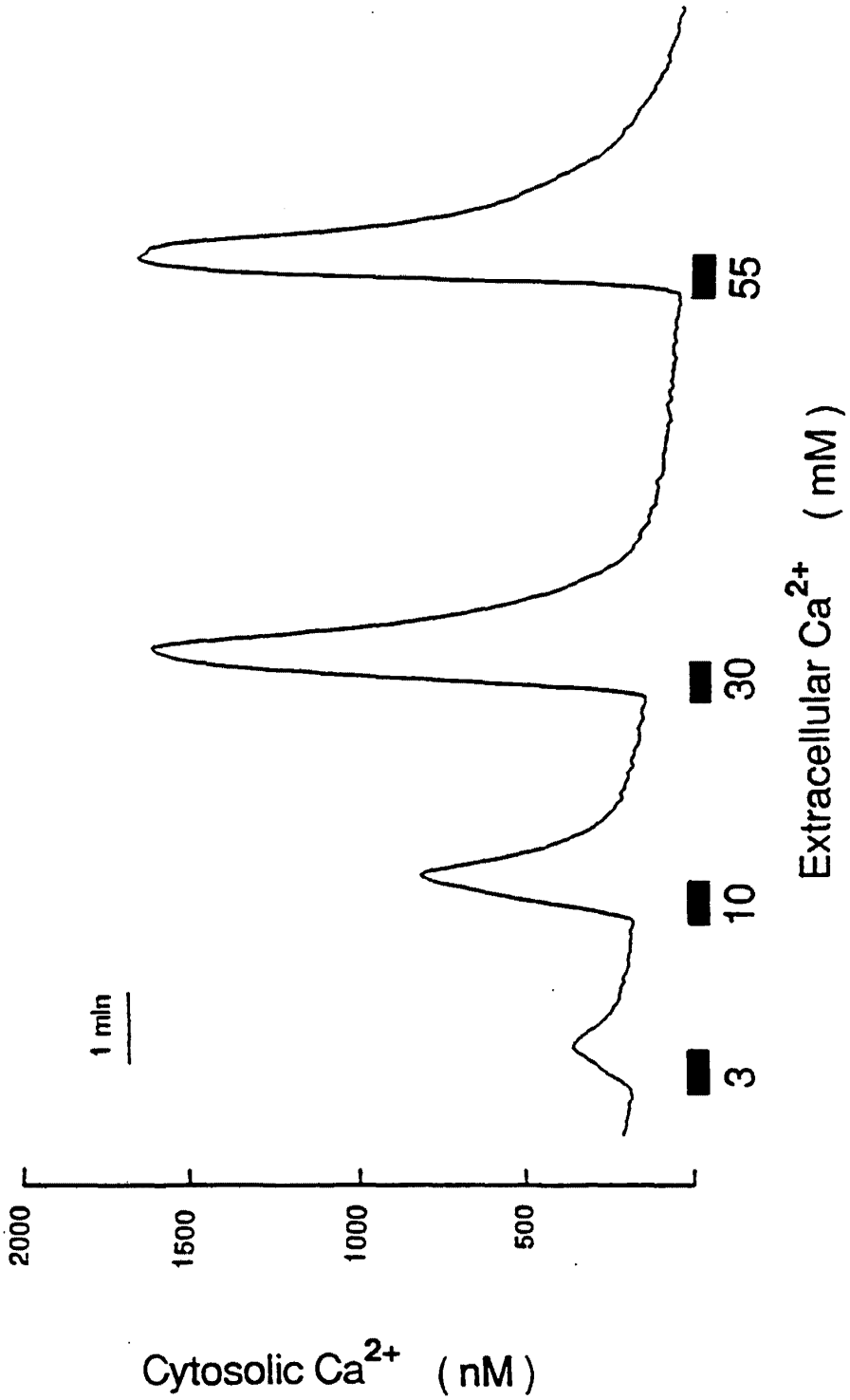
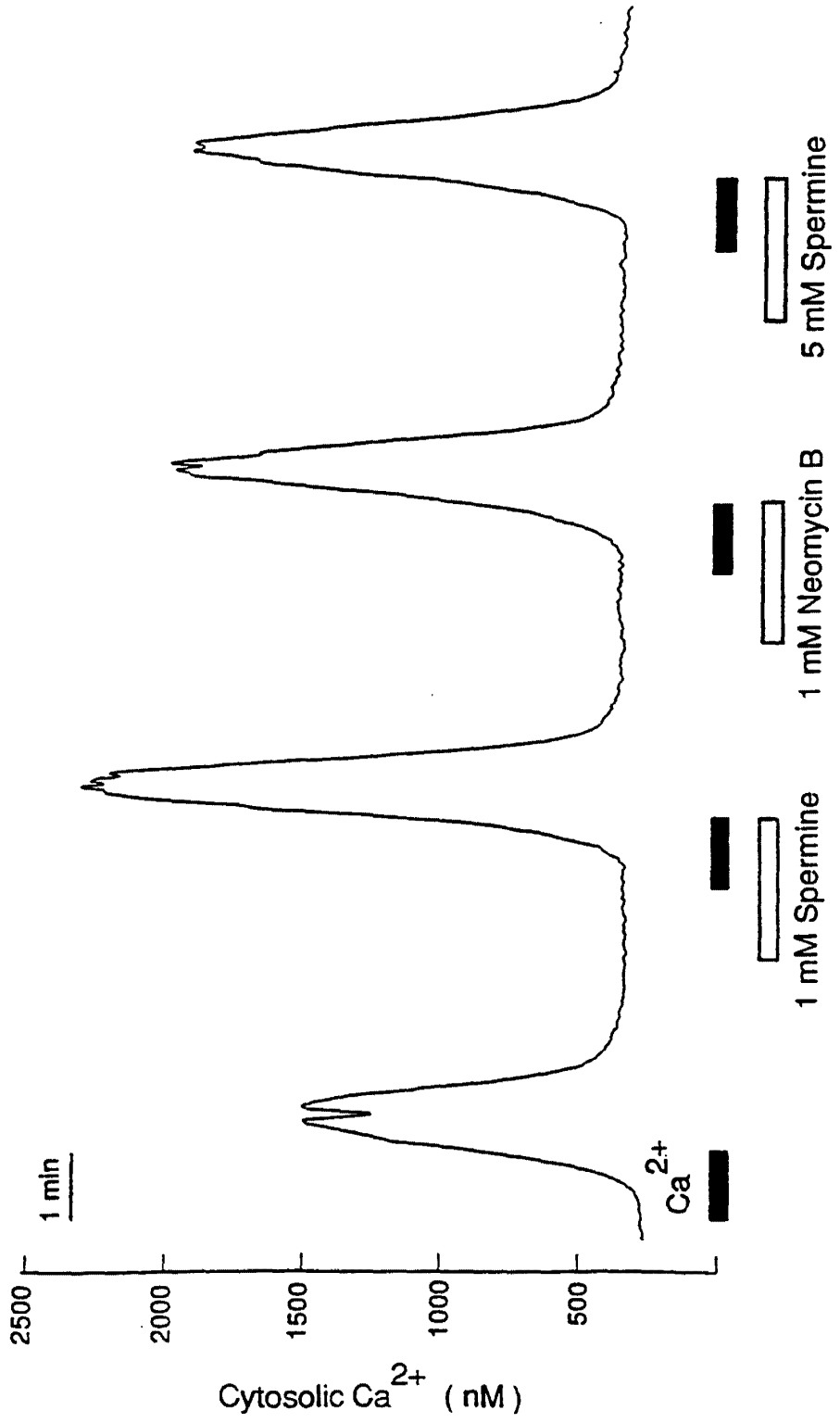
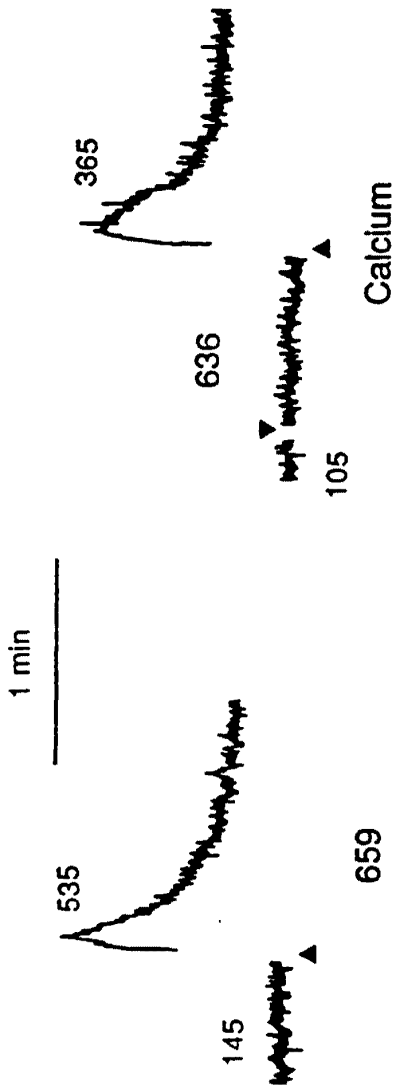

55
30
10
3
Extracellular Ca^{2+} (mM)
1 min
2000
1500
1000
500
Cytosolic Ca^{2+} (nM)

FIG. 23.



SUBSTITUTE SHEET

FIG. 24.



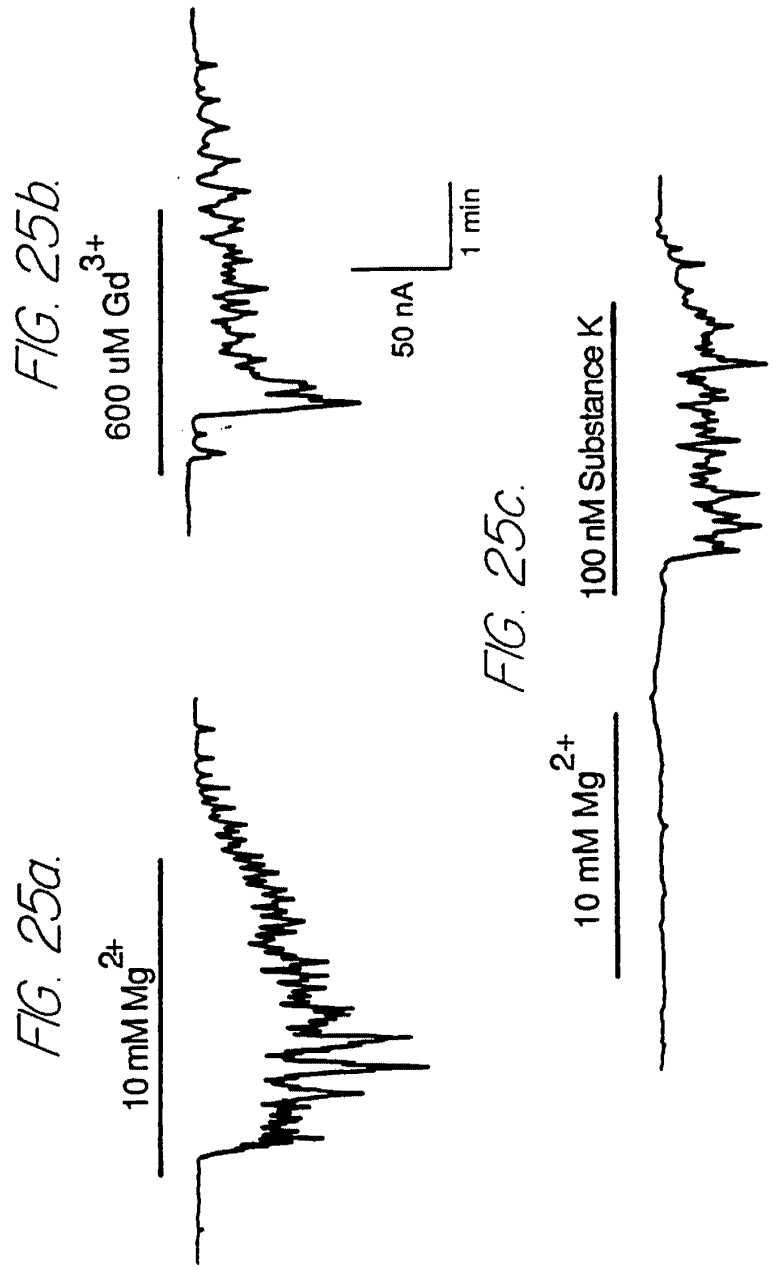


FIG. 25a.
10 mM Mg2+
FIG. 25b.
600 uM Gd3+
50 nA
1 min
FIG. 25c.
10 mM Mg2+
100 nM Substance K

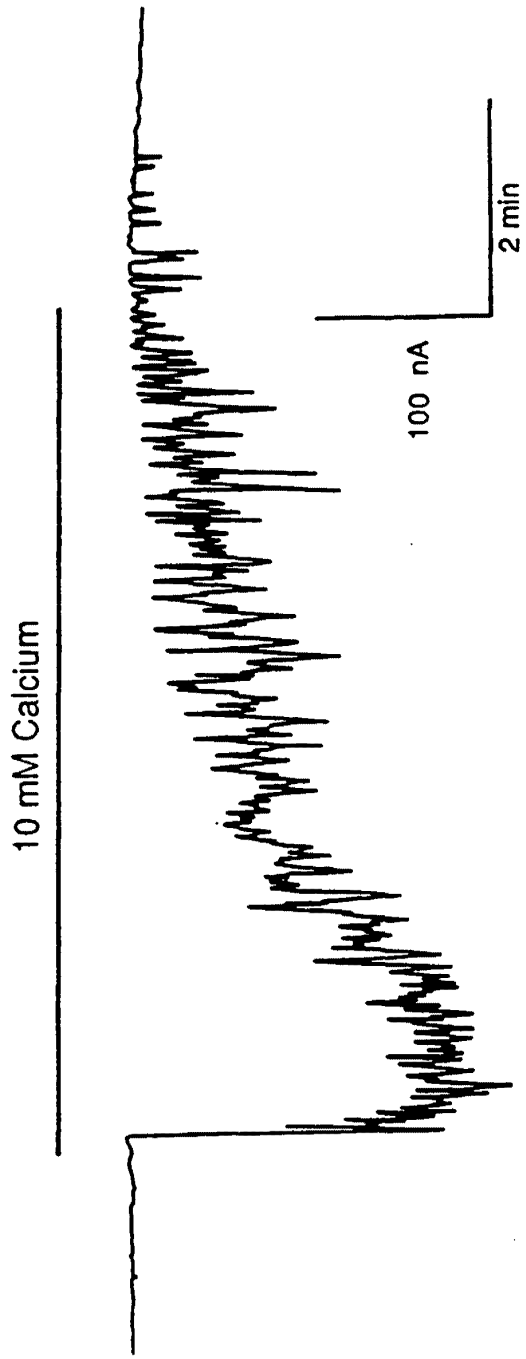
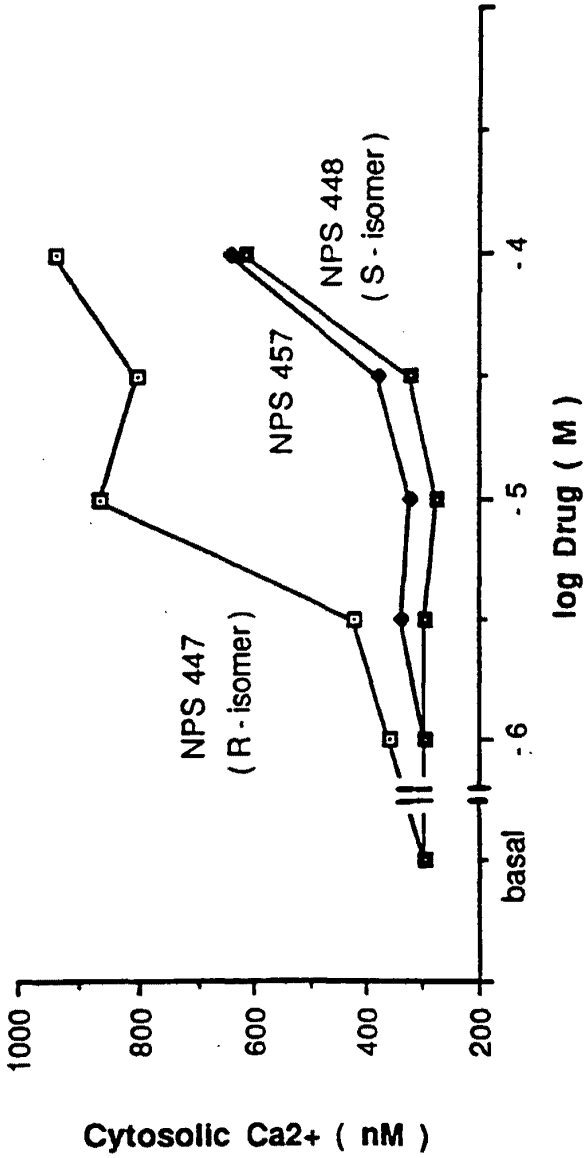


FIG. 26.


EGTA
Budmunchiamine
1 min
PMA
Budmunchiamine
Budmunchiamine

FIG. 27.

FIG. 28.


NPS 448
(S - isomer)
NPS 457
NPS 447
(R - isomer)
-4
-5
-6
basal
log Drug (M)
1000
800
600
400
200
Cytosolic Ca2+ (nM)

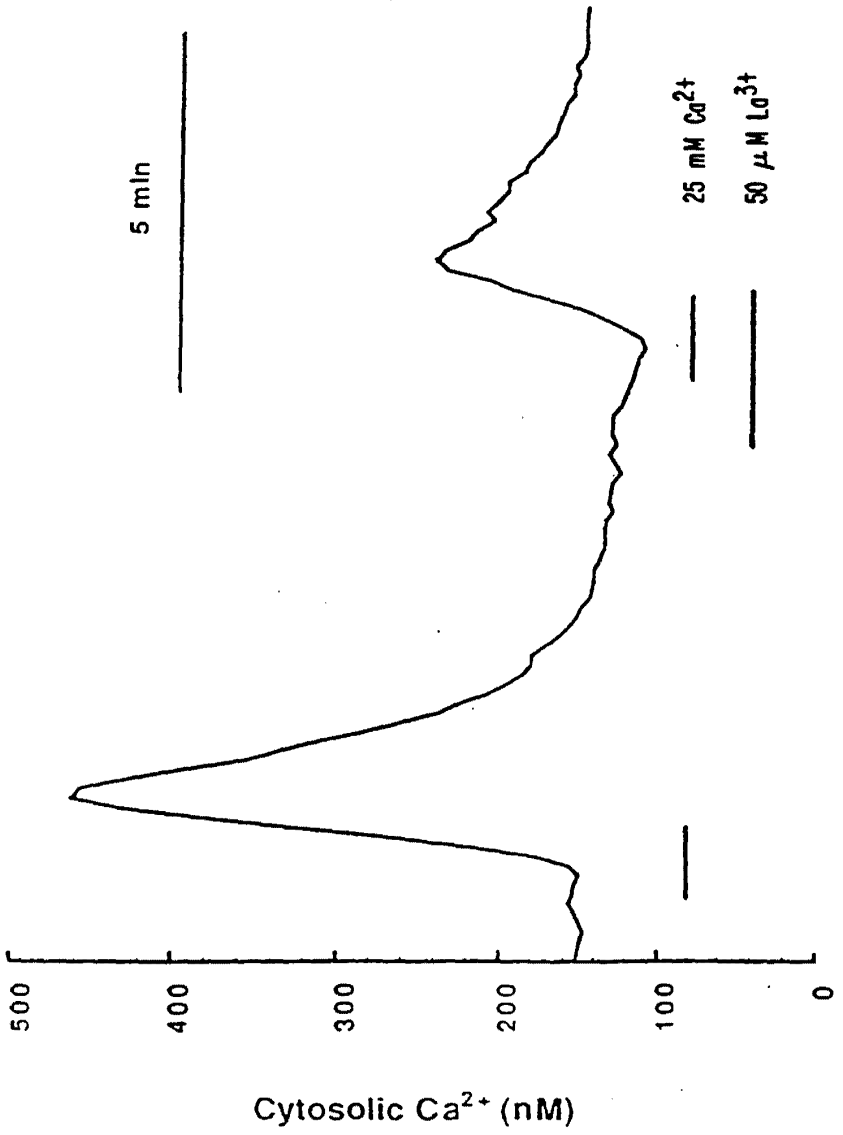
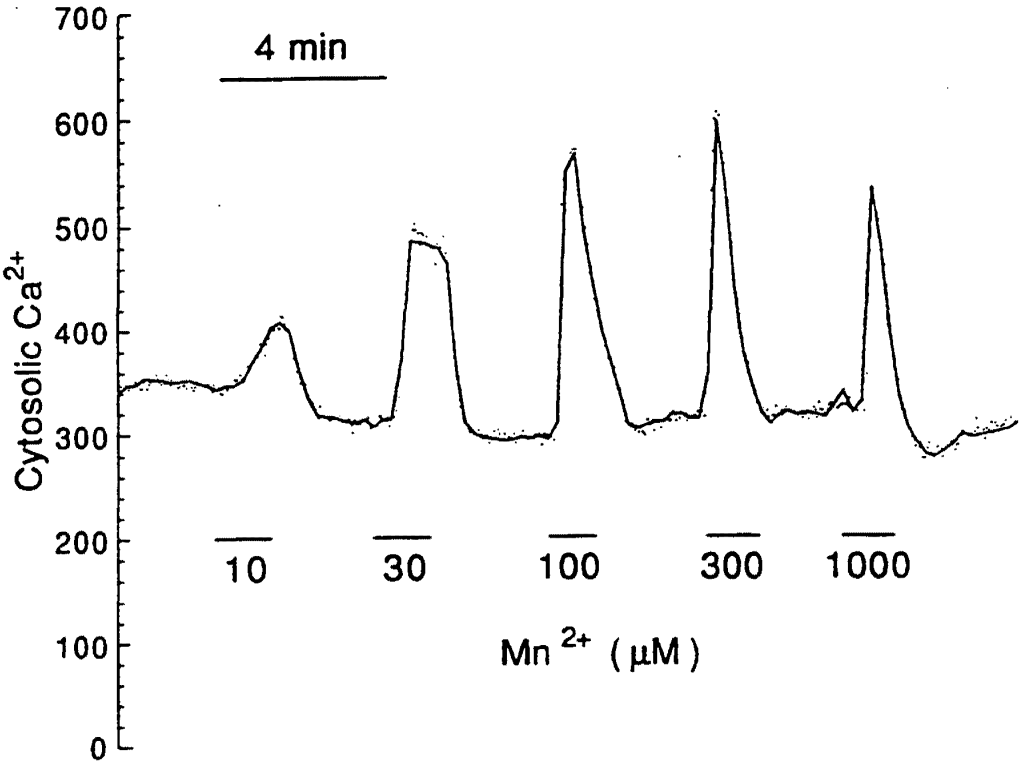


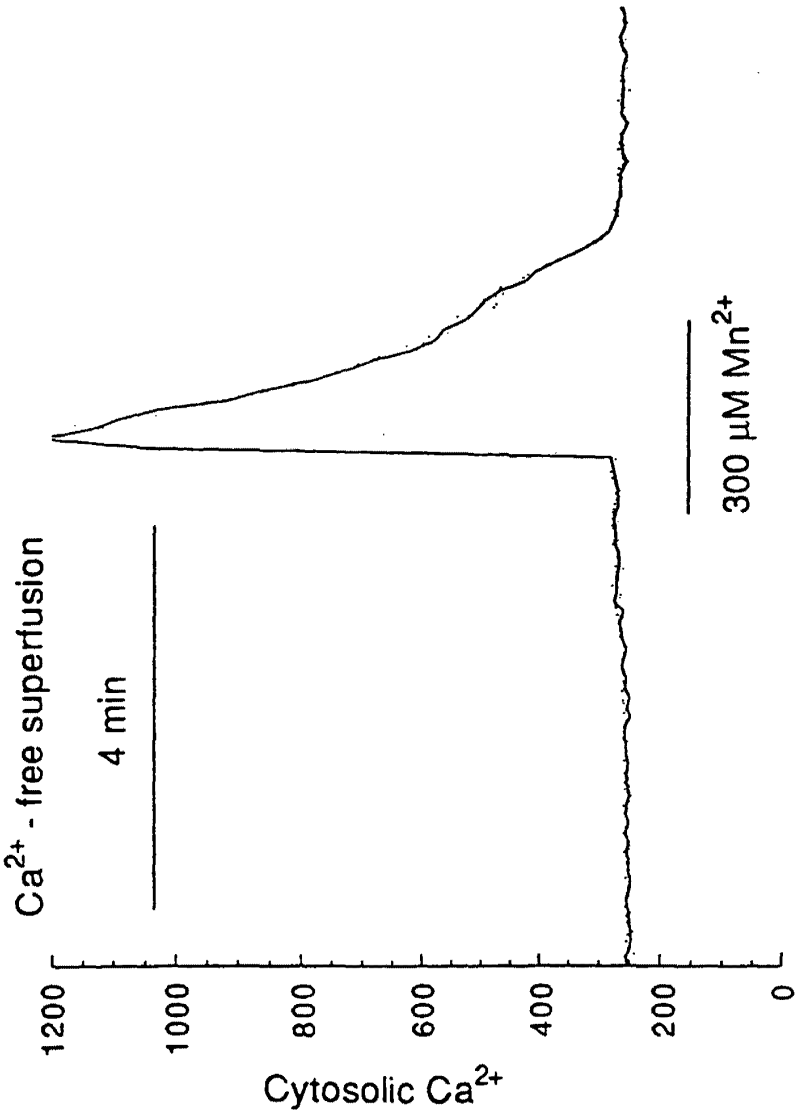
FIG. 29.

FIG. 30a.


700
600
500
400
300
200
100
0
Cytosolic Ca^{2+}
4 min
10
30
100
300
1000
Mn^{2+} (μM)

(a.)

FIG. 30b.


(b.)
Ca^{2+} - free superfusion
4 min
300 μM Mn^{2+}
1200
1000
800
600
400
200
0
Cytosolic Ca^{2+}

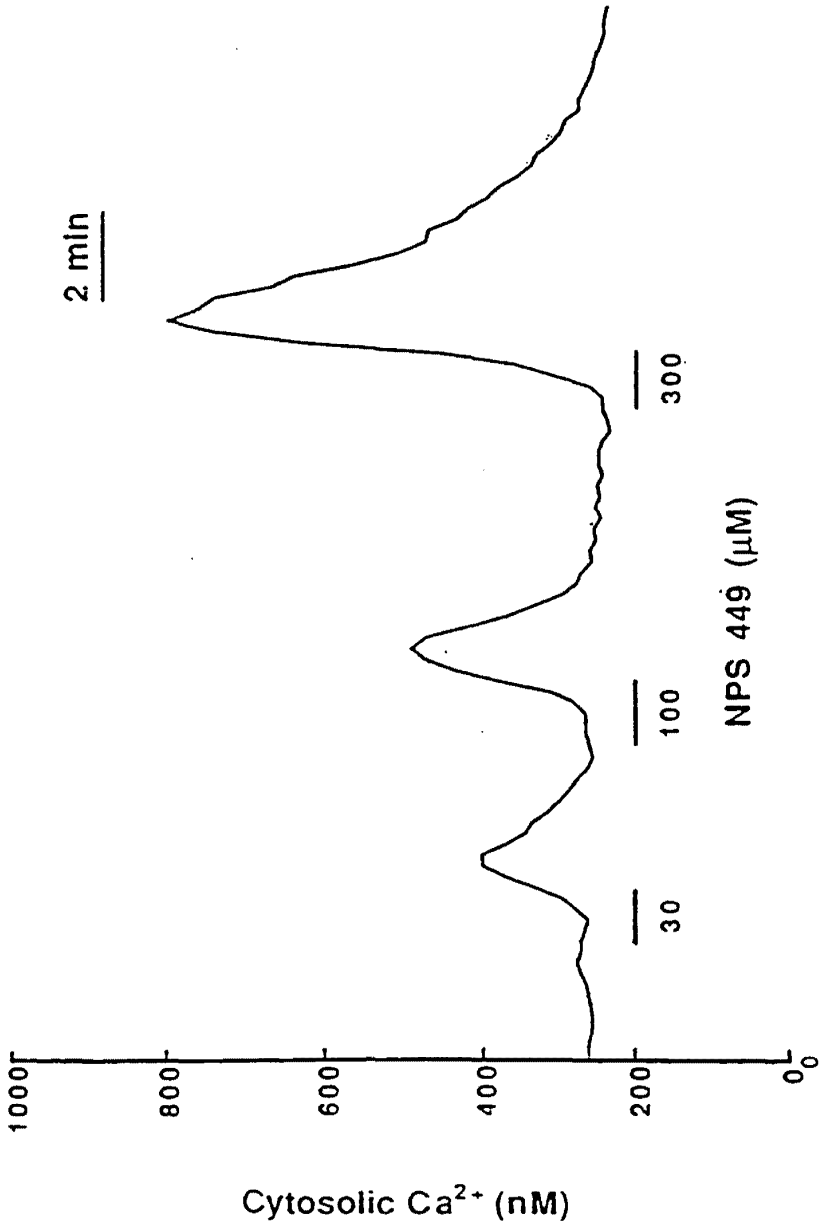

2 min
300
100
30
NPS 449 (μM)
1000
800
600
400
200
0
Cytosolic Ca^{2+} (nM)

FIG. 31a.

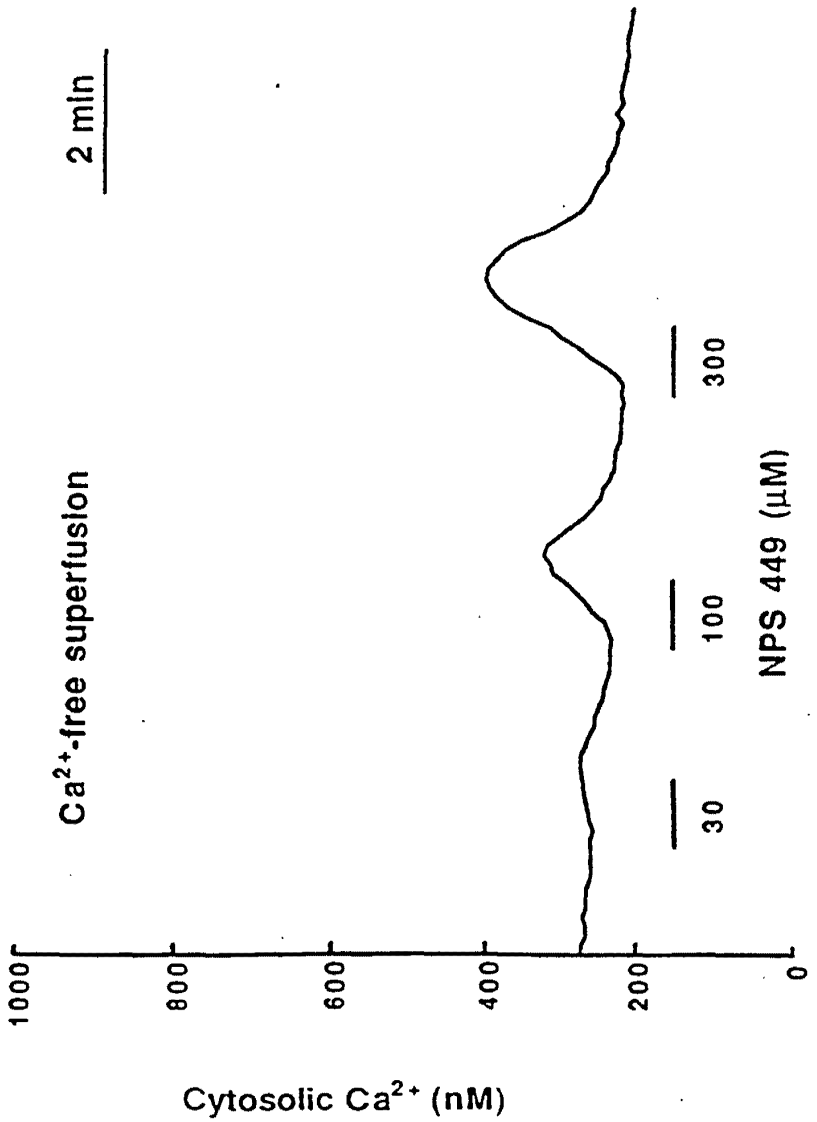

2 min
Ca2+-free superfusion
NPS 449 (μM)
30
100
300
1000
800
600
400
200
0
Cytosolic Ca2+ (nM)

FIG. 31b.

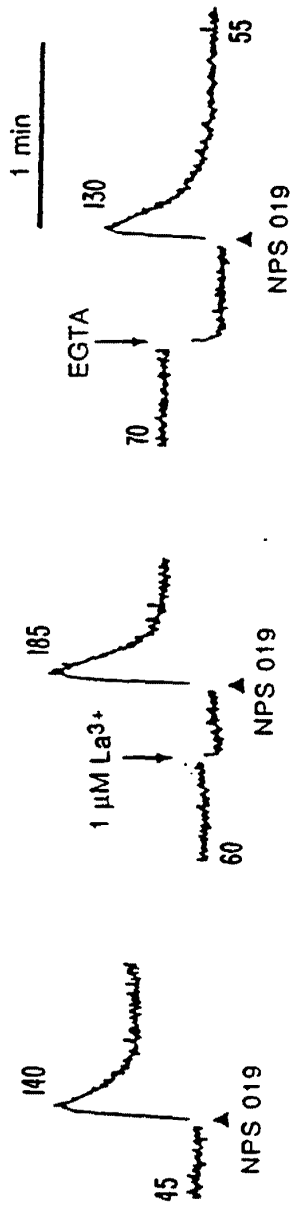


FIG. 32.

FIG. 33.

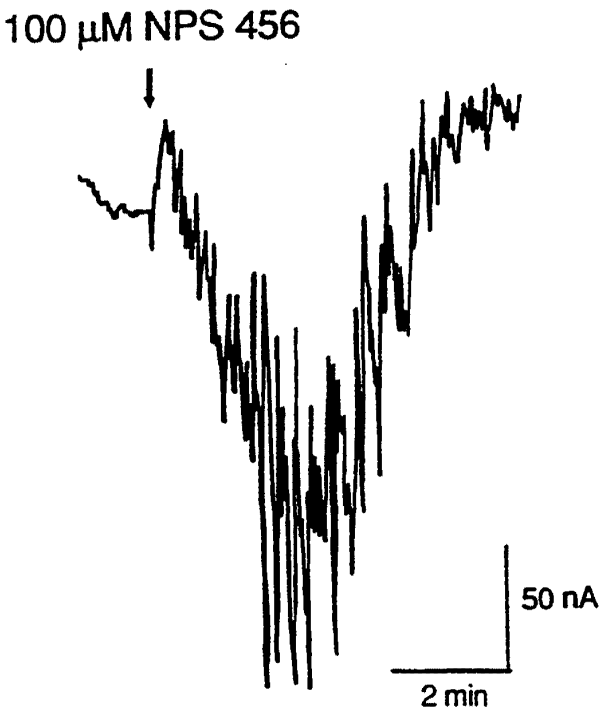
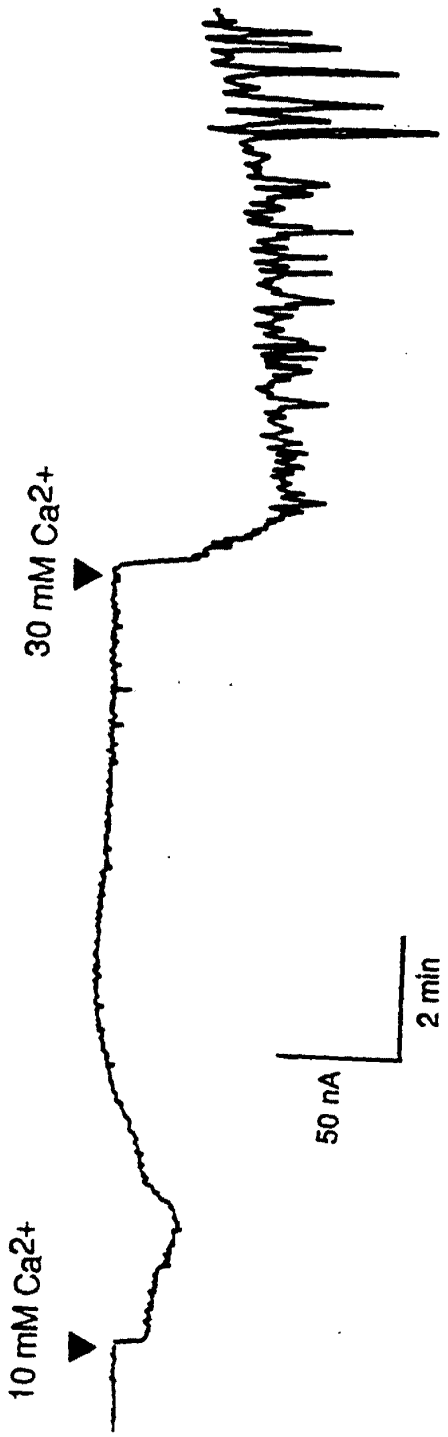


FIG. 34



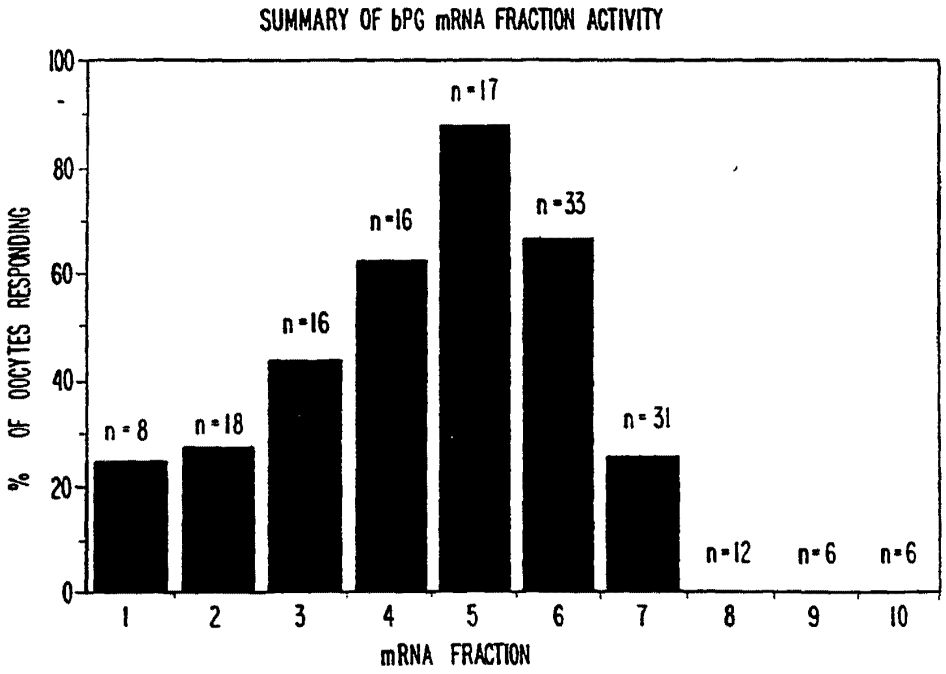
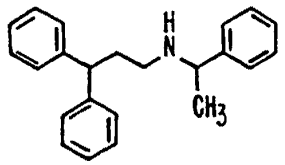
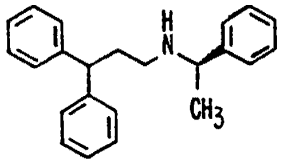

SUMMARY OF bPG mRNA FRACTION ACTIVITY
% OF OOCYTES RESPONDING
100
80
60
40
20
0
n = 8
n = 18
n = 16
n = 16
n = 17
n = 33
n = 31
n = 12
n = 6
n = 6
1
2
3
4
5
6
7
8
9
10
mRNA FRACTION

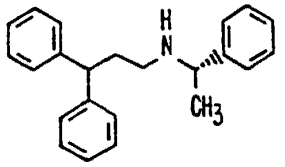
FIG. 35.



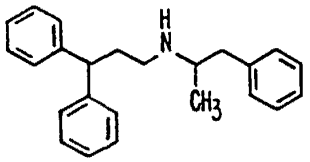
NPS # 456



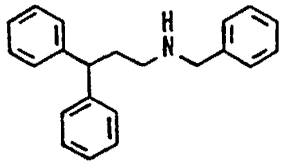
447



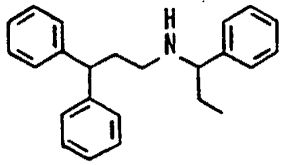
448



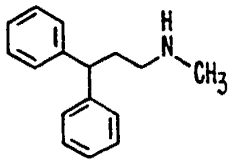
449



455



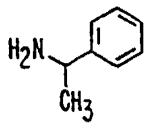
460



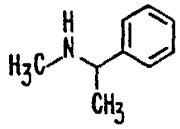
519

FIG. 36a.

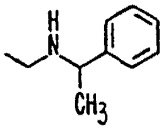
NPS #



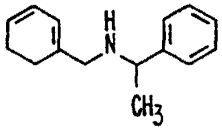
518



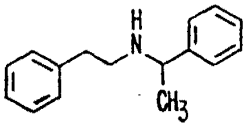
517



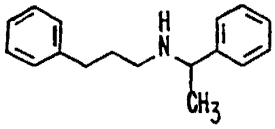
516



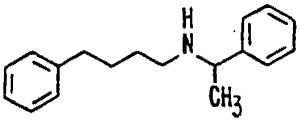
457



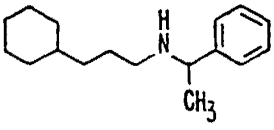
458



459



523



524

FIG. 36b.

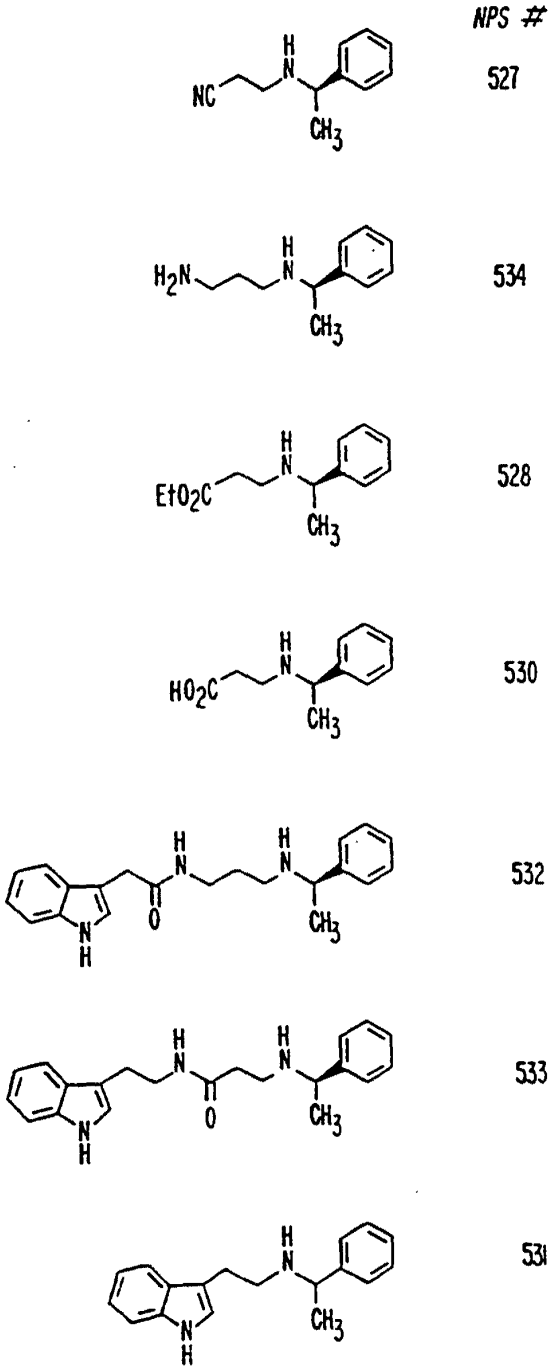


FIG. 36C.

SUBSTITUTE SHEET

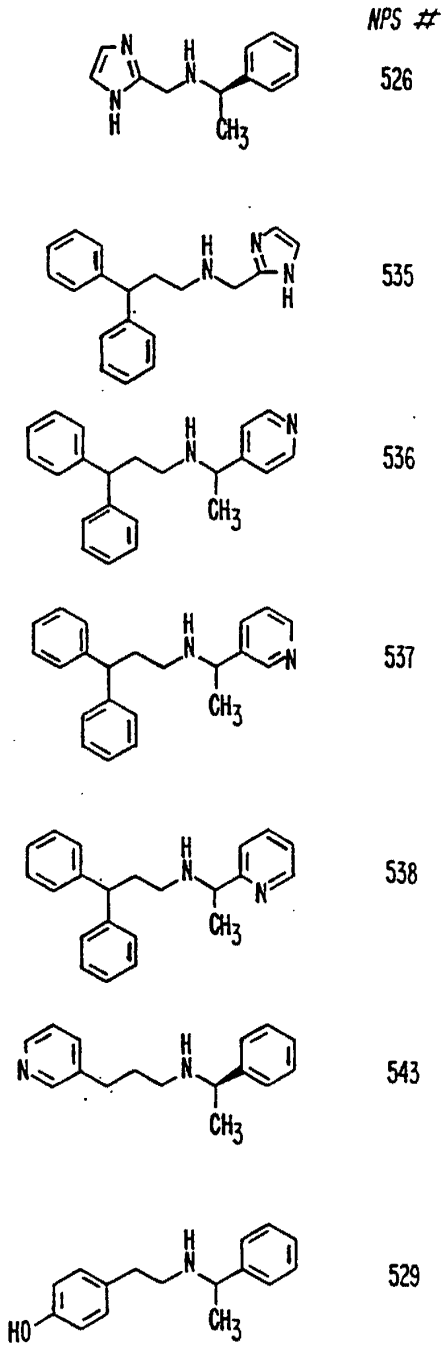


FIG. 36d.

SUBSTITUTE SHEET

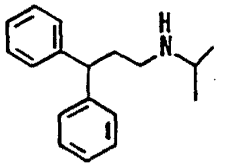
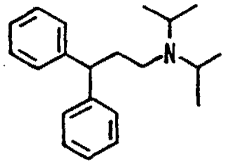
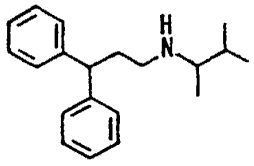
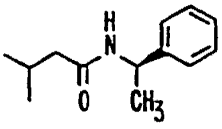
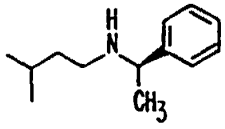
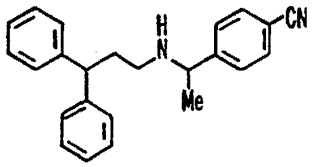
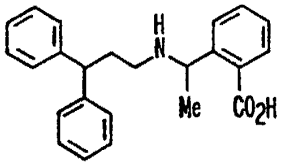
	NPS #
	539
	540
	542
	546
	547

FIG. 36e.

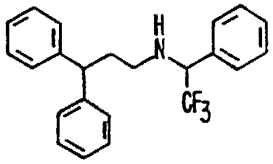
NPS #



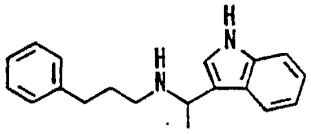
541



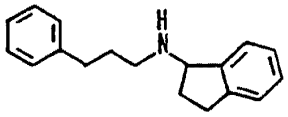
545



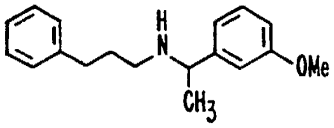
544



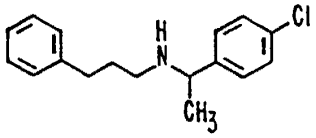
548



549



467



468

FIG. 36f.

SUBSTITUTE SHEET

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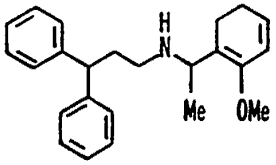
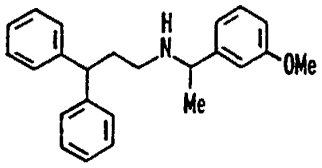
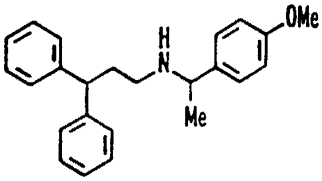
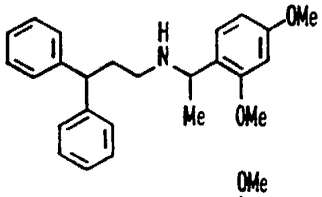
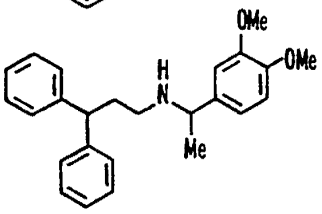
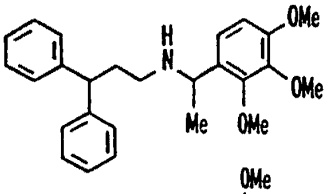
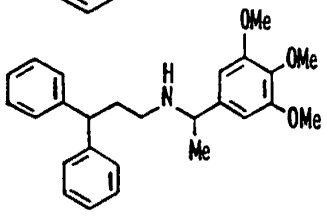
550	
551	
552	
553	
554	
555	
556	

FIG. 36g.

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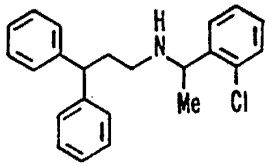
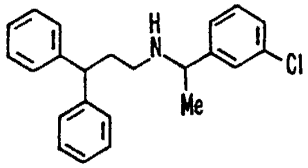
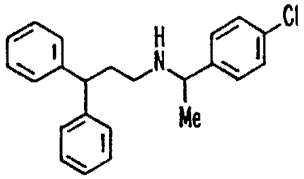
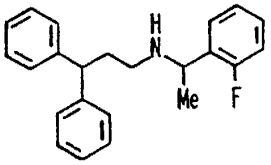
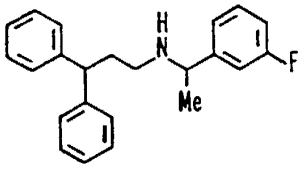
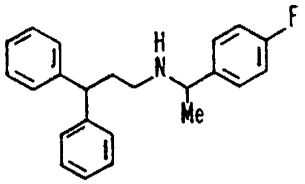
	557
	558
	559
	560
	561
	562

FIG. 36h.

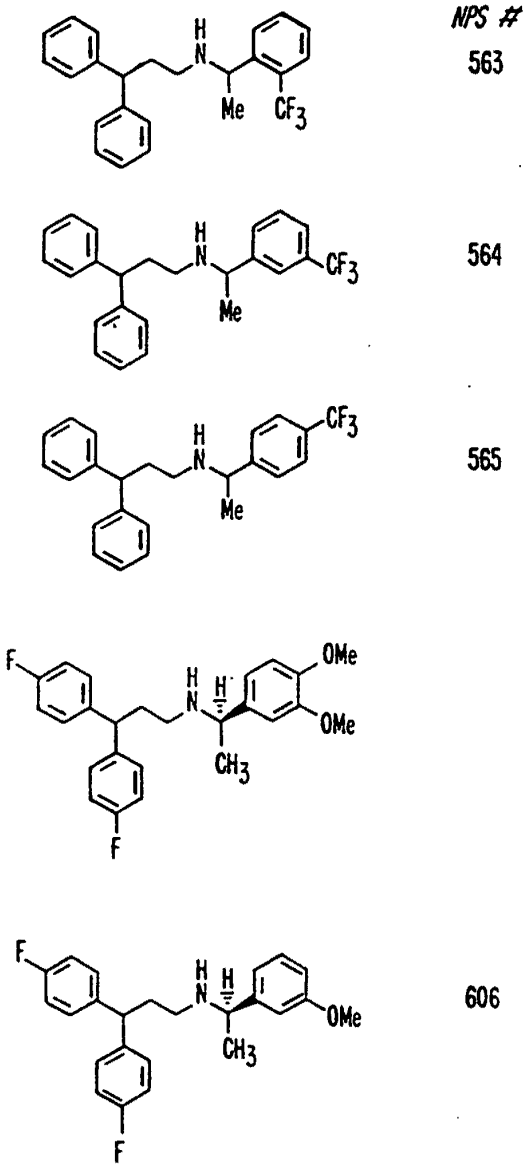
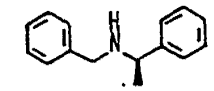


FIG. 36i.

NPS #



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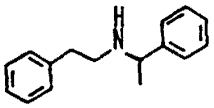

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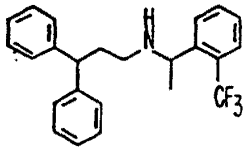

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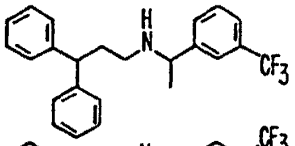

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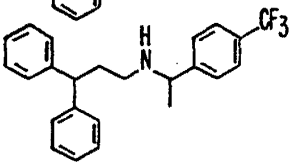

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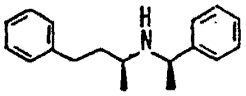

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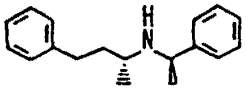

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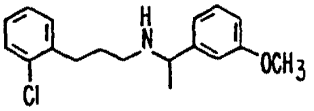

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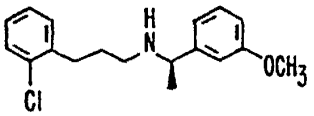

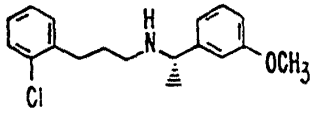

FIG. 36j.

SUBSTITUTE SHEET

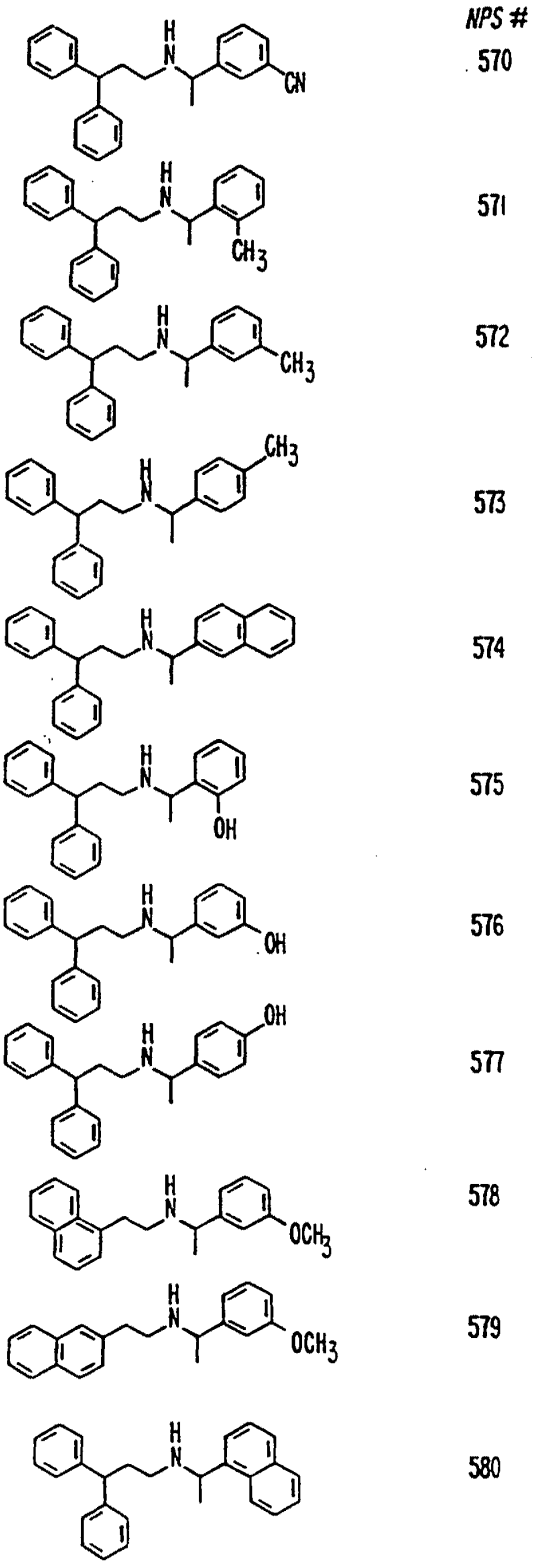

FIG. 36k.

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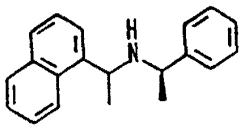
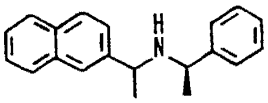
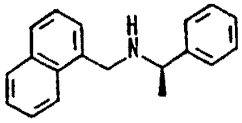
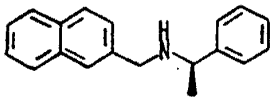
	592
	593
	594

FIG. 36L.

NPS #



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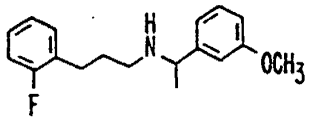

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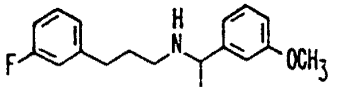

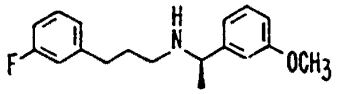

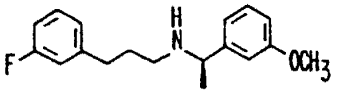

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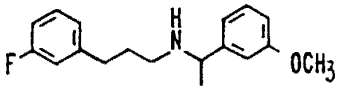

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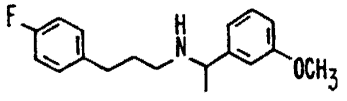

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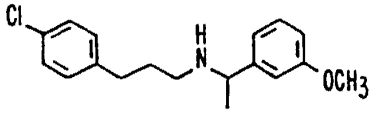
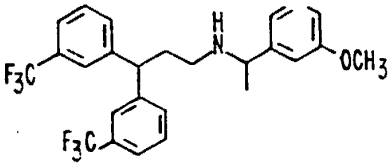


605



607

FIG. 36m.

SUBSTITUTE SHEET

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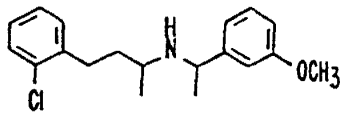

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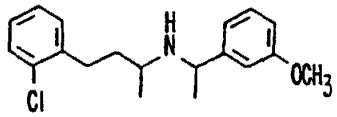

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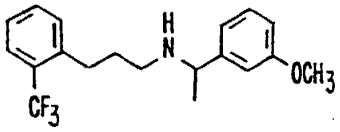

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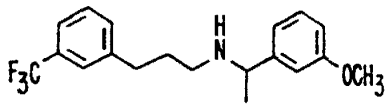

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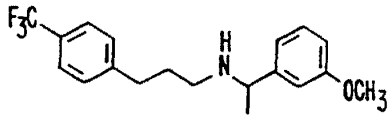

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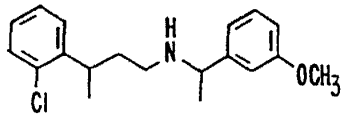

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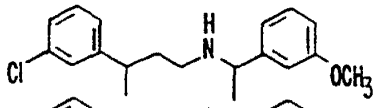
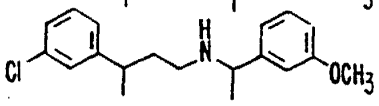
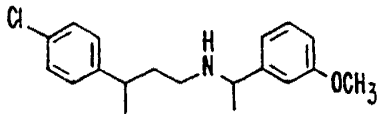


615



616



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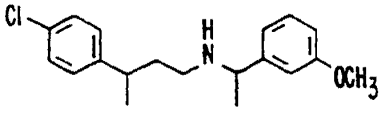

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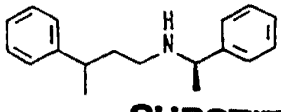
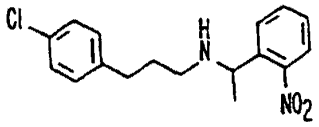


620

FIG. 36n.

SUBSTITUTE SHEET

NPS #



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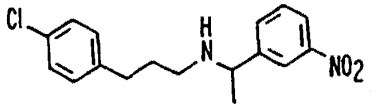

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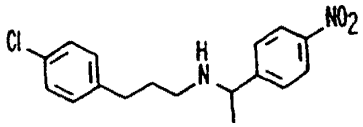

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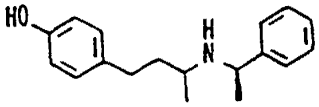

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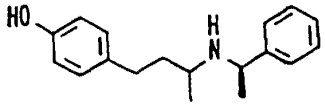

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FIG. 360.

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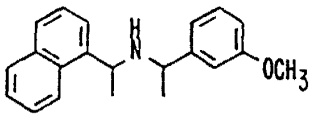

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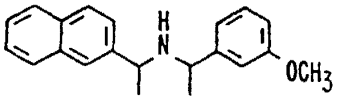

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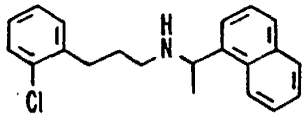
FIG. 36p.



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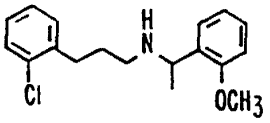

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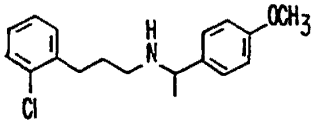

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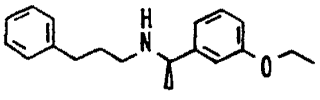

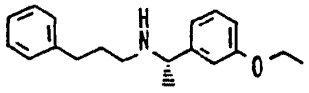

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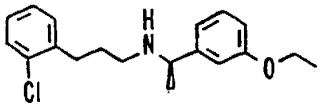

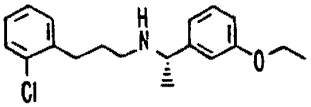

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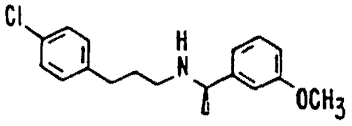

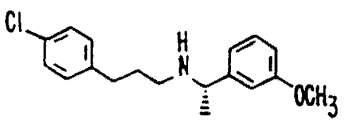

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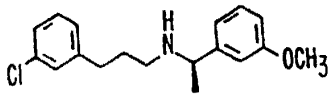
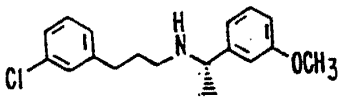
FIG. 36g

SUBSTITUTE SHEET





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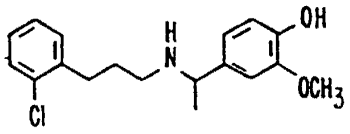
656



684

FIG. 36r.

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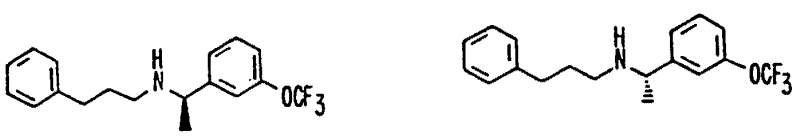
689	
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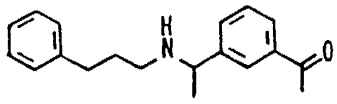
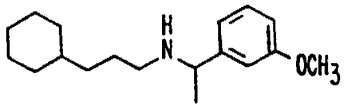
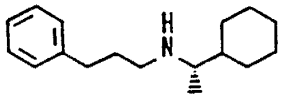
706	
708	
709	

FIG. 36s.

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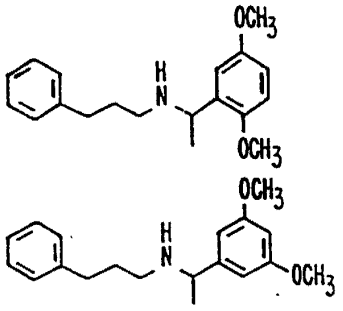


710

711

FIG. 36t.

FIG. 37b.

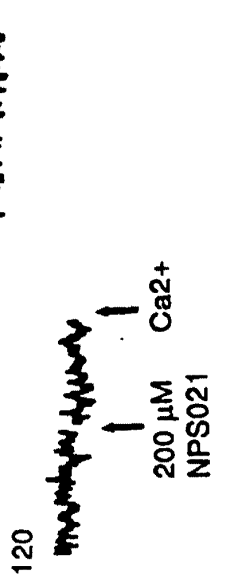


FIG. 37a.

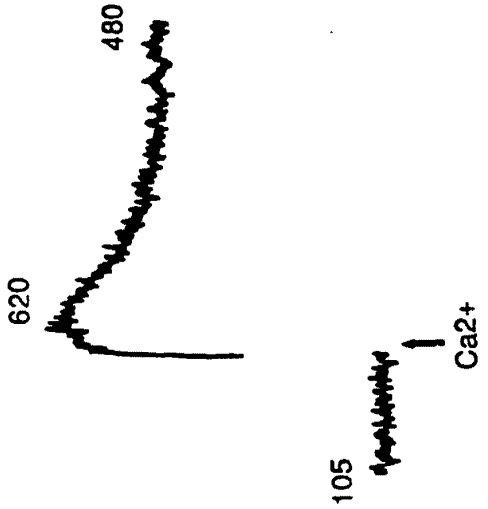
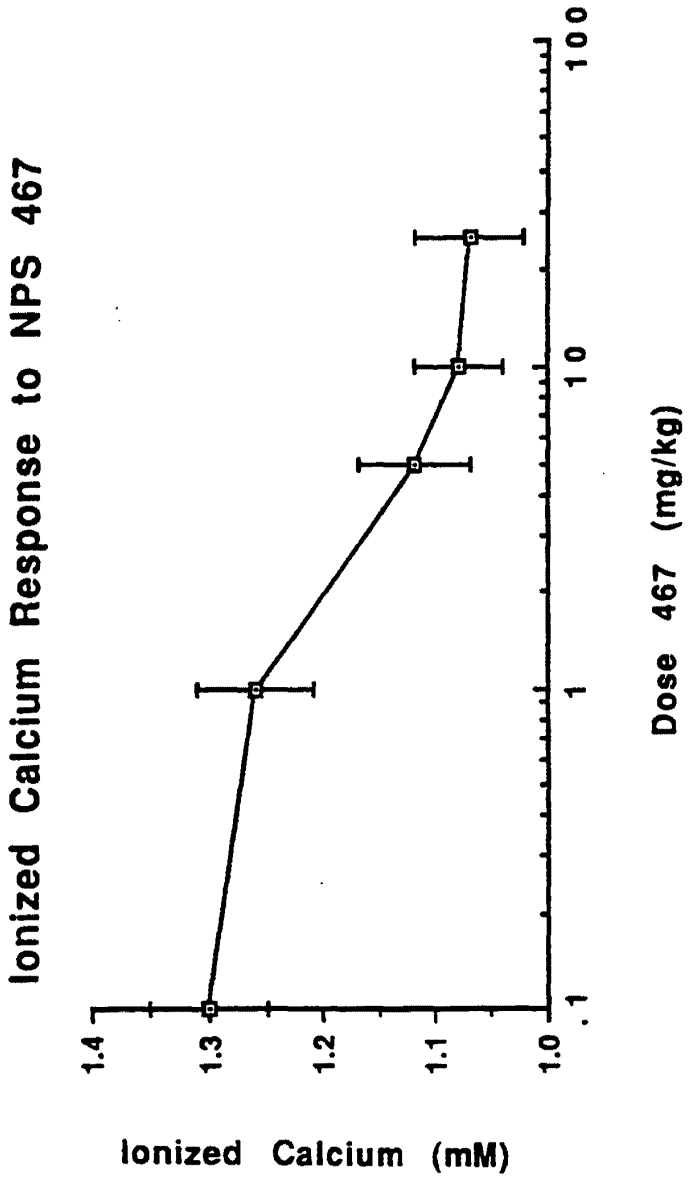


FIG. 38


Ionized Calcium Response to NPS 467
Dose 467 (mg/kg)
100
10
1
.1
1.4
1.3
1.2
1.1
1.0
Ionized Calcium (mM)

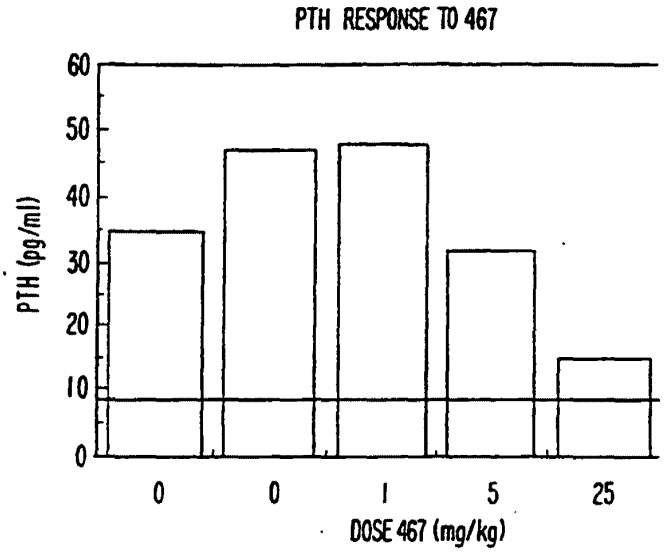

PTH RESPONSE TO 467
PTH (pg/ml)
60
50
40
30
20
10
0
0
0
1
5
25
DOSE 467 (mg/kg)

FIG. 39.

FIG. 40.

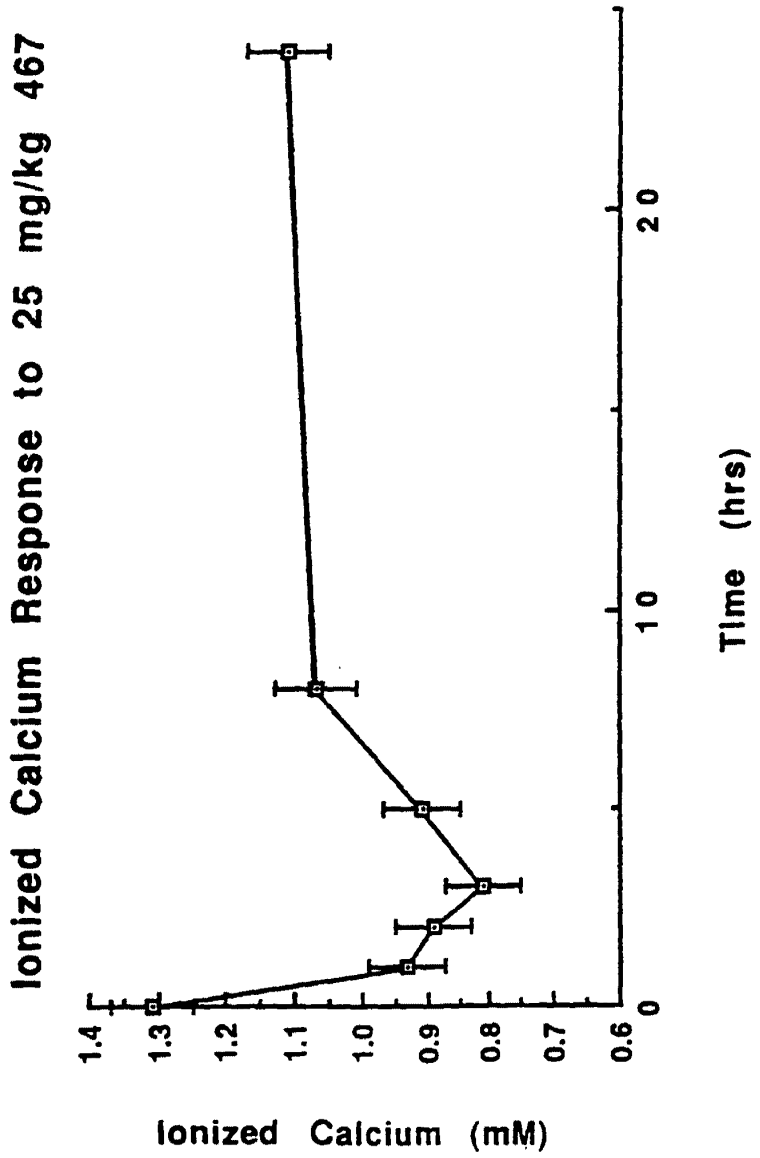
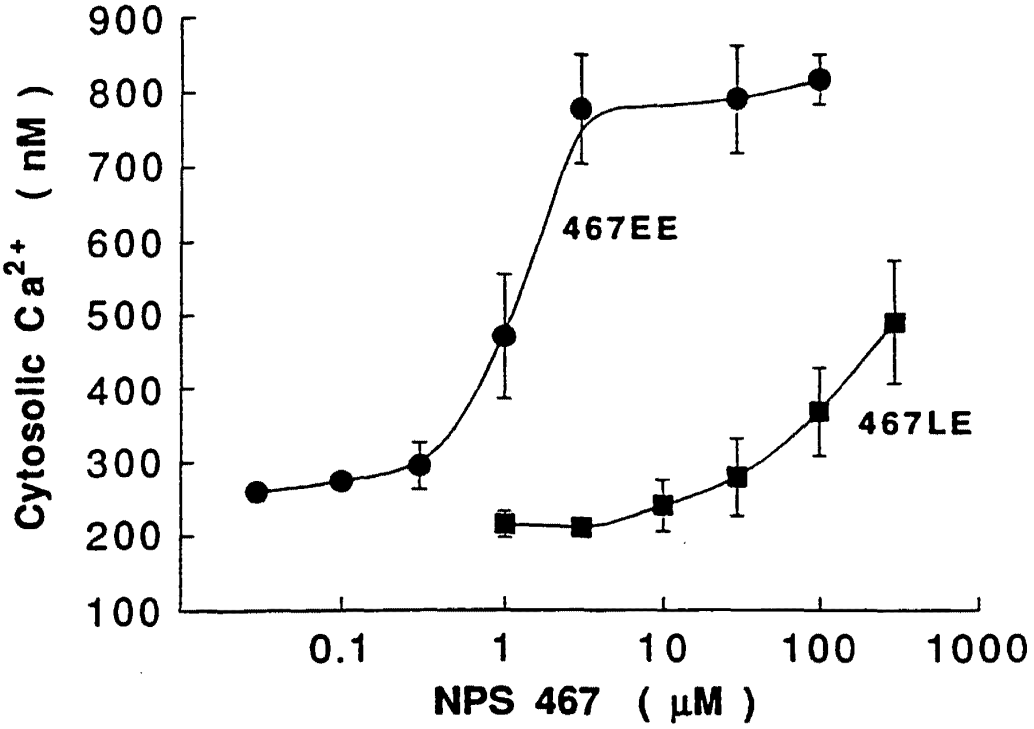

Ionized Calcium Response to 25 mg/kg 467
Ionized Calcium (mM)
1.4
1.3
1.2
1.1
1.0
0.9
0.8
0.7
0.6
0
10
20
Time (hrs)

FIG. 41.


900
800
700
600
500
400
300
200
100
Cytosolic Ca^{2+} (nM)
467EE
467LE
0.1
1
10
100
1000
NPS 467 (μM)

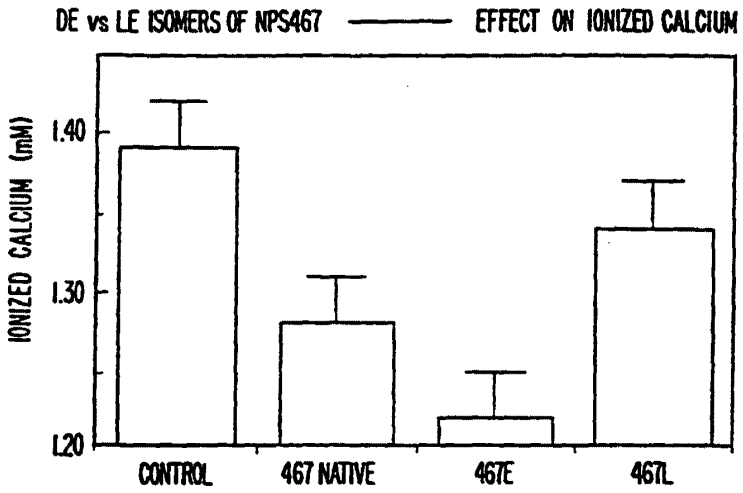

DE vs LE ISOMERS OF NPS467 ——— EFFECT ON IONIZED CALCIUM
IONIZED CALCIUM (mM)
1.40
1.30
1.20
CONTROL
467 NATIVE
467E
467L

FIG. 42.

FIG. 43a.

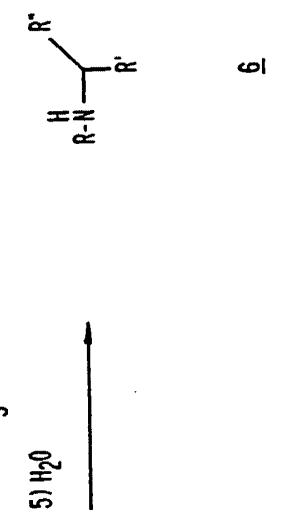
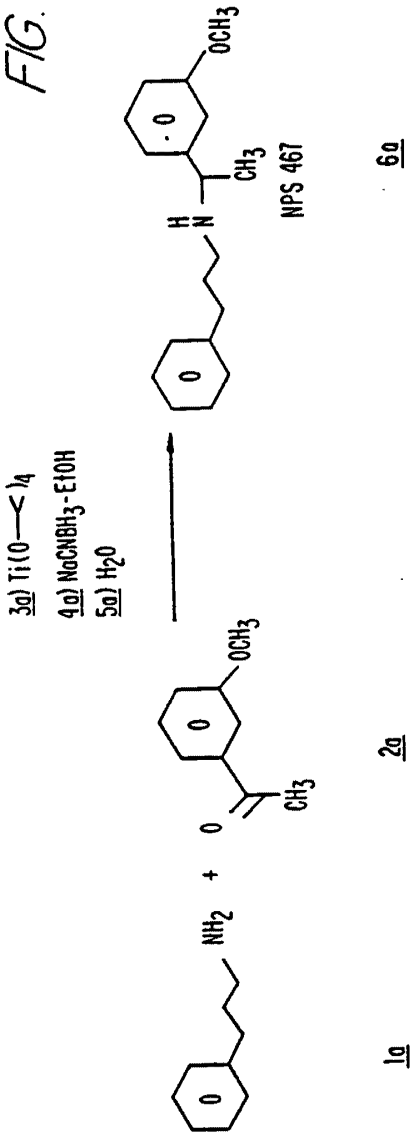


FIG. 43b.



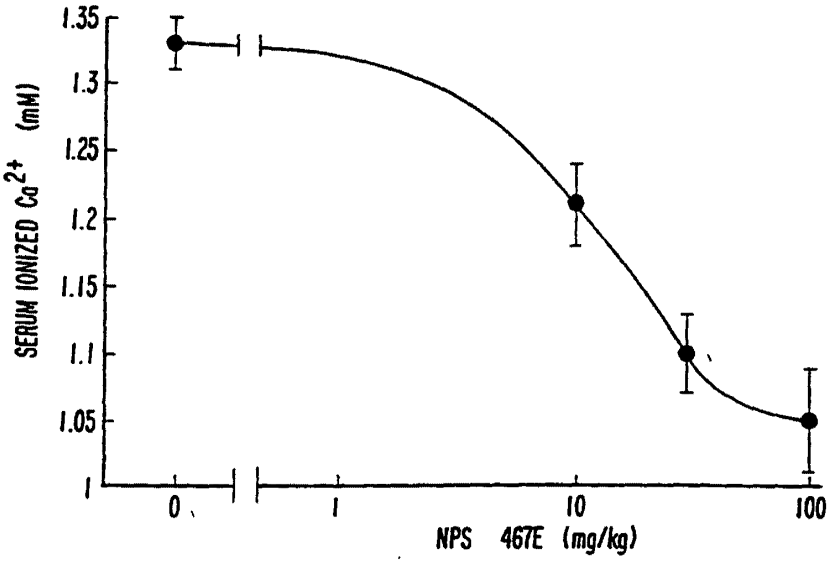

SERUM IONIZED Ca^{2+} (mM)
1.35
1.3
1.25
1.2
1.15
1.1
1.05
1
0
1
10
100
NPS 467E (mg/kg)

FIG. 44.

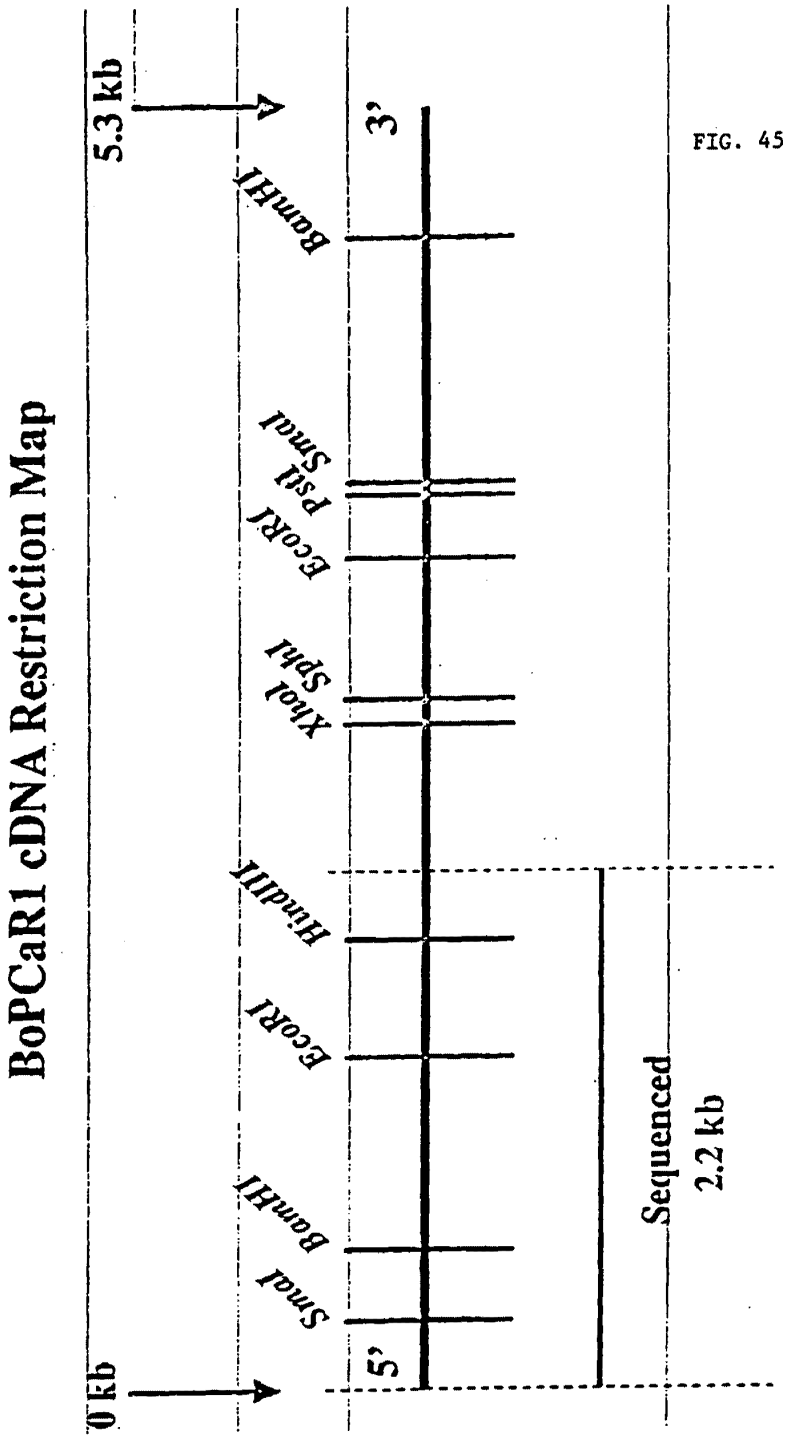

BoPCaR1 cDNA Restriction Map
5.3 kb
0 kb
5'
3'
SmaI
BamHI
EcoRI
HindIII
XhoI
SphI
PstI
SmaI
EcoRI
BamHI
Sequenced
2.2 kb

FIG. 45

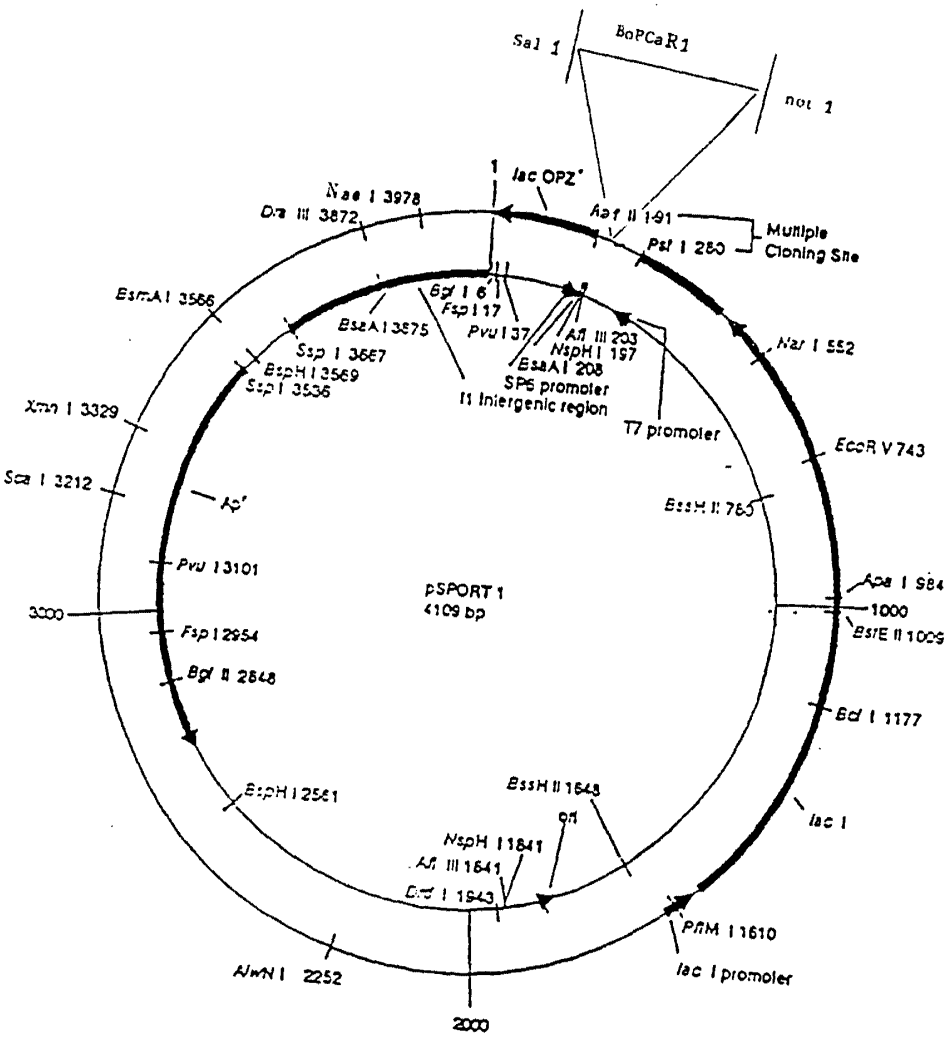

Sal 1
BoPCaR1
not 1
1
lac OPZ'
Nae I 3978
Dra III 3872
Apa II 191
Multiple Cloning Site
Pst I 250
BsmA I 3566
Bgl I 6
Fsp I 17
Pvu I 37
Afl III 203
NspH I 157
EsaA I 208
BseA I 3575
Ssp I 3567
BspH I 3569
Ssp I 3536
SPS promoter
f1 intergenic region
T7 promoter
Nar I 552
Xmn I 3329
Sca I 3212
Ap'
EcoR V 743
BssH II 765
Pvu I 3101
pSPORT 1
4109 bp
3000
Fsp I 2954
Bgl II 2648
Ape I 984
1000
BstE II 1009
Ecl I 1177
BspH I 2561
BssH II 1648
ori
lac I
NspH I 1841
Afl III 1641
Dra I 1943
PflM I 1610
lac I promoter
AlwN I 2252
2000

FIG. 46

```
   1 CCACGCGTCCGCGGA AAAAAAAAAAAAGTT CCCCACTCTAGTACA GAGAAGGTTGGCAGA

  60 GTCGTAAGCCCCCAA CCTCTTAAACTTCTC TGCATCTCCAAGGAG AAGGAGGGAAGAGGG
 120 GTTCTTTCCGACCTG AGGAGCTGGATCTGG GGTCCGAGAACCCCA AGGTAGCACCGGAAA
 180 GAACAGCACAGGAGG CGAGAGCGTGCGGTG GCCGGTKGGCGGGAG AACCAGACCCGACKY
 240 CGCGGTCCTCGGYGC TGCCGGGKYCCSGGG ACTCAGCTCAGCACG ACTGGGAAGCCGAAA
 300 GTACTACACACGGTC TCTGCATGATGTGAC TTCTGAAGACTCAAG AGCCACCCACTTCAC
 360 TAGTCTGCAATGGAG AAGGCAGAAATGGAA AGTCAAACCCCACGG TTCCATTCTATTAAT
 420 TCTGTAGACATGTGC CCCCACTGCAGGGAG TGAGTCGCACCAAGG GGGAAAGTCCTCAGG
 480 GGCCCCCAGACCACC AGCGCTTGAGTCCCT CTTCCTGGAGAGAAA GCAGAACTATGGCAC
 540 TTTATAGCTGCTGTT GGATCCTCTTGGCTT TTTTCTACCTGGTGC ACTTCCGCCTATGGG
 600 CCTGACCAGCGAGCC CAAAAGAAAGGGGAC ATTATCCTCGGGGGG CTCTTTCCTATTCAT
 660 TTTGGGGTTGMMGTG AAAGATCAGGATCTA AAGTCGAGGCCGGAG TCCGTGGAGTGTATC
 720 AGGTATAATTTCCGA GGATTTCGCTGGTTA CAAGCTATGATATTT GCCATAGAGGAAATA
 780 AACAGCAGTCCAGCC CTTCTTCCCAACATG ACCCTGGGATACAGG ATATTCGACACTTGT
 840 AACACCGTCTCTAAA GCCTTGGAGGCCACC CTGAGTTTTGTGGCC CAGAACAAAATTGAC
 900 TCTTTGAACCTTGAT GAGTTCTGCAACTGC TCAGAGCACATCCCC TCTACCATCGCAGTG
 960 GTGGGAGCTACTGGC TCGGGCATCTCCACA GCAGTGGCCAACCTG CTGGGGCTCTTCTAC
1020 ATCCCCCAGGTCAGC TATGCCTCCTCCAGC AGACTCCTCAGCAAC AAGAATCAATTCAAG
1080 TCCTTCCTCCGCACC ATACCCAATGATGAA CACCAGGCCACGGCC ATGGCTGACATCATC
1140 GAGTACTTCCGCTGG AACTGGGTGGGCACA ATTGCAGCTGACGAT GACTATGGCCGGCCA
1200 GGGATCGAGAAGTTT CGAGAGGAAGCTGAG GAGAGGGACATCTGC ATCGACTTCAGCGAG
1260 CTCATCTCCCAATAC TCTGATGAGGAAAAG ATCCAGCAGGTGGTG GAGGTGATCCAGAAT
1320 TCCACCGCCAAAGTC ATTGTCCGTCTTCTC CAGCGGCCCAGACCT GGAACCCCTCATCAA
1380 AGAGATCGTCCGGCG CAATATCACAGGCAG GATCTGGCTGGCCAG CGAGSCTGGSCAGCT
1440 CTTCCCTGATTGCTA TGCCCGAGTATTTCC ATGTGGTCGGAGGCA CCATTGGGTTTGGTT
1500 TGAAAGCTGGGCAGA TCCCAGGCTTCCGGG AATTCCTGCAGAAAG TCCACCCCAGGAAGT
1560 CTGTCCACAATGGTT TTGCCAAGGAGTTTT GGGAAGAAACATTTA ACTGCCACCTGCAAG
1620 AGGGTGCTAAAGGCC CATTACCGGTGGACA CCTTCCTGAGAGGTC ACGAAGAAGGAGGTG
1680 CCAGGTTAAGCAACA GTCCCACTGCCTTCC GACCTCTGTGCACTG GGGAGGAGAACATCA
1740 GCAGTGTCGAGACTC CTTACATGGATTATA CACATTTACGGATAT CCTACAACGTCTACT
1800 TAGCCGTCTACTCCA TTGCTCATGCCCTAC AAGATATATACACCT GCATACCTGGGAGAG
1860 GGCTCTTCACCAACG GTTCCTGCGCAGATA TCAAGAAGGTTGAAG CTTGGCAGGTCCTGA
1920 AACACCTGCGGCACC TAAATTTTACCAGCA ATATGGGGGAGCAAG TAACTTTCGATGAAT
1980 GTGGAGACCTGGCAG GGAACTATTCCATCA TCAACTGGCACCTCT CCCCAGAGGACGGCT
2040 CCATAGTGTTTAAGG AAGTTGGATATTACA ATGTCTATGCCAAGA AAGGAGAGAGACTCT
2100 TCATCAATGATGAAA AAATTCTGTGGAGTG GATTCTCAAGGGAGG TGC 2148
```

FIG. 47

SUBSTITUTE SHEET

ATTORNEY'S DOCKET NO.: B0801/7012WO

ERG/lfr
3/9/93
8985V/

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants: Brigham and Women's Hospital
75 Francis Street
Boston, Massachusetts USA 02115

NPS Pharmaceuticals, Inc.
420 Chipeta Way
Salt Lake City, Utah USA 84108

Inventors: NEMETH, Edward F. et al.

International Serial No: PCT/US93/01642
Filing Date: 23 February 1993
For: Calcium Receptor-Active Molecules

BOX PCT
Hon. Commissioner of Patents and Trademarks
Washington, D. C. 20231

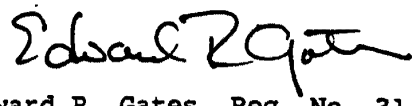
Sir:

REMARKS

Enclosed are substitute pages 9, 41, and 166 in connection with the above-identified patent application. Also enclosed is the American Type Culture Collection (ATCC), Rockville, Maryland receipt indicating that the deposit made in connection with the instant application was received on 19 February 1993 and was assigned ATCC number 75416.

The undersigned attorney had not yet been informed by the ATCC as to the particular deposit number at the time of filing the application. Accordingly, substitute pages including the correct number are enclosed.

Respectfully submitted,



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INTERNATIONAL SEARCH REPORT

Int. ional Application No
PCT/US 93/01642

A. CLASSIFICATION OF SUBJECT MATTER
IPC 5 A61K31/00 A61K31/13 A61K31/135 A61K31/405 C07K15/00
C12N15/12 C12N15/11 C12N5/10 C07K13/00 C07K15/28
A01K67/027

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 5 A61K C07K C12N A01K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
E	WO,A,93 04373 (NPS PHARMACEUTICALS INC.) 4 March 1993 see the whole document ---	1-82,89, 98-103
X	EP,A,0 253 327 (CHINOIN) 20 January 1988 see the whole document ---	1-23, 25-27
X	WO,A,91 00853 (NEW YORK UNIVERSITY) 24 January 1991 see claims ---	1-23
X	WO,A,92 14709 (CAMBRIDGE NEUROSCIENCE INC.) 3 September 1992 see claims --- -/--	1-23

[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

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"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
15 November 1993	23. 12. 93

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax (+31-70) 340-3016	KLAVER, T

Form PCT/ISA/210 (second sheet) (July 1992)

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP,A,0 395 357 (PFIZER INC.) 31 October 1990 see claims	1-23,36, 38,39
X	GB,A,2 113 089 (UNIVERSITY OF NEW YORK) 3 August 1983 * whole document, esp. page 2 *	1-24
A	BIOSCIENCE REP. vol. 10, no. 6 , 1990 pages 493 - 507 M. ZAIDI '"Calcium receptors" on eukaryotic cells with special reference to the osteoclast.' cited in the application	
A	CELL CALCIUM vol. 11, no. 5 , 1990 pages 319 - 321 E.F. NEMETH ET AL. 'The role of extracellular calcium in the regulation of intracellular calcium and cell function.' cited in the application	
A	CELL CALCIUM vol. 11, no. 5 , 1990 pages 323 - 327 E.F. NEMETH 'Regulation of cytosolic calcium by extracellular divalent cations in C-cells and parathyroid cells.' cited in the application	
A	NORMAN, VANAMAN, MEANS , EDS. 'calcium-binding proteins in health and disease' 1987 , ACADEMIC PRESS , SAN DIEGO Evidence for the presence of a novel calcium-binding protein calcium receptor on the surface of parathyroid cells. Auth. E.F. Nemeth see page 36 - page 38	

2

INTERNATIONAL SEARCH REPORT

Int. ational application No.
PCT/US93/01642

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. [X] Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
REMARK: Although claims 36-89 are directed to a method of treatment of or a diagnostic method practised on the human/animal body the search has been carried out based on the alleged effects of the compound/composition.

2. [X] Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
Claims searched incompletely: 1-22,25,27,36-60,63,65,83.
For further information see annex.

3. [ ] Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. [ ] As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. [ ] As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. [ ] As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. [ ] No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

[ ] The additional search fees were accompanied by the applicant's protest.

[ ] No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet (1)) (July 1992)

FURTHER INFORMATION CONTINUED FROM PCT/ISA/206

a. A compound cannot be sufficiently described by its psycho-chemical or pharmachological characteristics. The search has therefor been restricted to compounds mentioned in the claims and the description (cl.1-22,36-60).

b. In view of the large number of compounds defined by the general formula of claims 25,27,63 and 65 the search has been limited to the inventive part of the molecule and/or the compounds mentioned in the description and/or in the claims (PCT Art. 6).

c. The subject matter of claim 83 is not clear, because the actual matter is not revealed.

INTERNATIONAL SEARCH REPORT

Information on patent family members

Int. onal Application No
PCT/US 93/01642

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A-9304373	04-03-93	AU-A-	2588992	16-03-93
EP-A-0253327	20-01-88	AU-B-	599117	12-07-90
		AU-A-	7552687	14-01-88
		DE-A-	3778770	11-06-92
		JP-A-	63072657	02-04-88
		SU-A-	1597096	30-09-90
		SU-A-	1588740	30-08-90
		US-A-	4988730	29-01-91
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		JP-A-	58131914	06-08-83

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A1

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08/292,827	19 August 1994 (19.08.94)	US

(60) Parent Applications or Grants
(63) Related by Continuation

US	07/749,451 (CIP)
Filed on	23 August 1991 (23.08.91)
US	07/834,044 (CIP)
Filed on	11 February 1992 (11.02.92)
US	07/934,161 (CIP)
Filed on	21 August 1992 (21.08.92)
US	08/017,127 (CIP)
Filed on	12 February 1993 (12.02.93)
US	08/009,389 (CIP)
Filed on	23 February 1993 (23.02.93)
US	08/141,248 (CIP)
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US	08/292,827 (CIP)
Filed on	19 August 1994 (19.08.94)

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(72) Inventors; and
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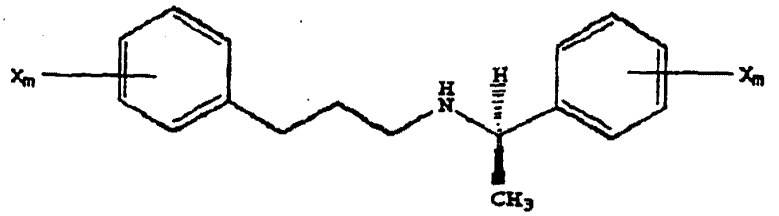
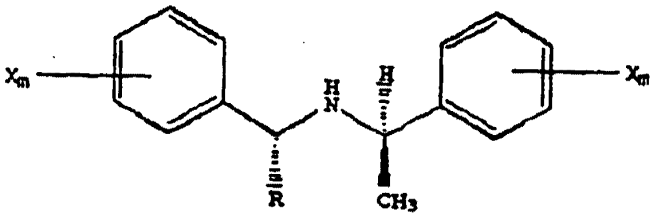
(81) Designated States: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MN, MW, NL, NO, NZ, PL, PT, RO, RU, SD, SE, SI, SK, TJ, TT, UA, US, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ).

Published
With international search report.

(54) Title: CALCIUM RECEPTOR-ACTIVE ARYLALKYL AMINES



(I)



(II)

(57) Abstract

The present invention features molecules with general structure (I) or (II) which can modulate on or activities of an inorganic ion receptor. Preferably, the molecule can mimic or block the effect of extracellular Ca^{2+} on a calcium receptor. The preferred use of such molecules is to treat diseases or disorders by altering inorganic ion receptor activity, preferably calcium receptor activity.

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DESCRIPTION

Calcium receptor-active arylalkyl amines

FIELD OF THE INVENTION

This invention relates to the design, development, composition and use of novel molecules able to modulate the activity of inorganic ion receptor.

BACKGROUND OF THE INVENTION

Certain cells in the body respond not only to chemical signals, but also to ions such as extracellular calcium ions (Ca^{2+}). Changes in the concentration of extracellular Ca^{2+} (referred to herein as "$[Ca^{2+}]$") alter the functional responses of these cells. One such specialized cell is the parathyroid cell which secretes parathyroid hormone (PTH). PTH is the principal endocrine factor regulating Ca^{2+} homeostasis in the blood and extracellular fluids.

PTH, by acting on bone and kidney cells, increases the level of Ca^{2+} in the blood. This increase in $[Ca^{2+}]$ then acts as a negative feedback signal, depressing PTH secretion. The reciprocal relationship between $[Ca^{2+}]$ and PTH secretion forms the essential mechanism maintaining bodily Ca^{2+} homeostasis.

Extracellular Ca^{2+} acts directly on parathyroid cells to regulate PTH secretion. The existence of a parathyroid cell surface protein which detects changes in $[Ca^{2+}]$ has been confirmed. Brown *et al.*, 366 Nature 574, 1993. In parathyroid cells, this protein acts as a receptor for extracellular Ca^{2+} ("the calcium receptor"), and detects changes in $[Ca^{2+}]$ and to initiate a functional cellular response, PTH secretion.

Extracellular Ca^{2+} can exert effects on different cell functions, reviewed in Nemeth *et al.*, 11 Cell Calcium 319, 1990. The role of extracellular Ca^{2+} in parafollicular (C-cells) and parathyroid cells is discussed in Nemeth, 11 Cell Calcium 323, 1990. These cells have been shown to express similar Ca^{2+} receptor. Brown *et al.*, 366 Nature 574, 1993; Mithal et al., 9 Suppl. 1 J. Bone and Mineral

Res. s282, 1994; Rogers *et al.*, 9 Suppl. 1 J. Bone and Mineral Res. s409, 1994; Garrett *et al.*, 9 Suppl. 1 J. Bone and Mineral Res. s409, 1994. The role of extracellular Ca^{2+} on bone osteoclasts is discussed by Zaidi, 10 Bioscience Reports 493, 1990. In addition keratinocytes, juxtaglomerular cells, trophoblasts, pancreatic beta cells and fat/adipose cells all respond to increases in extracellular calcium which likely reflects activation of calcium receptors of these cells.

The ability of various compounds to mimic extracellular Ca^{2+} *in vitro* is discussed by Nemeth *et al.*, (spermine and spermidine) in "Calcium-Binding Proteins in Health and Disease," 1987, Academic Press, Inc., pp. 33-35; Brown *et al.*, (*e.g.*, neomycin) 128 Endocrinology 3047, 1991; Chen *et al.*, (diltiazem and its analog, TA-3090) 5 J. Bone and Mineral Res. 581, 1990; and Zaidi *et al.*, (verapamil) 167 Biochem. Biophys. Res. Commun. 807, 1990. Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959, and Nemeth *et al.*, PCT/US92/07175, International Publication Number WO 93/04373, describe various compounds which can modulate the effect of an inorganic ion on a cell having an inorganic ion receptor, preferably modulate the effects of calcium on a calcium receptor.

The references provided in the background are not admitted to be prior art.

SUMMARY OF THE INVENTION

The present invention features molecules which can modulate one or activities of an inorganic ion receptor. Preferably, the molecule can mimic or block the effect of extracellular Ca^{2+} on a calcium receptor. The preferred use of such molecules is to treat diseases or disorders by altering inorganic ion receptor activity, preferably calcium receptor activity.

Extracellular Ca^{2+} is under tight homeostatic control and controls various processes such as blood clotting, nerve and muscle excitability, and proper bone formation.

Calcium receptor proteins enable certain specialized cells to respond to changes in extracellular Ca^{2+} concentration. For example, extracellular Ca^{2+} inhibits the secretion of parathyroid hormone from parathyroid cells, inhibits bone resorption by osteoclasts, and stimulates secretion of calcitonin from C-cells.

Compounds modulating inorganic ion receptor activity can be used to treat diseases or disorders by affecting one or more activities of an inorganic ion receptor resulting in a beneficial effect to the patient. For example, osteoporosis is an age related disorder characterized by loss of bone mass and increased risk of bone fracture. Compounds blocking osteoclastic bone resorption either directly (*e.g.*, a osteoclast ionmimetic compound) or indirectly by increasing endogenous calcitonin levels (*e.g.*, a C-cell ionmimetic), and/or by decreasing parathyroid hormone levels (*e.g.*, a parathyroid cell ionmimetic) can retard bone loss and, thus, result in beneficial effects to patients suffering from osteoporosis.

In addition, it is known that intermittent low dosing with PTH results in an anabolic effect on bone mass and appropriate bone remodeling. Thus, compounds and dosing regiments evoking transient increases in parathyroid hormone (*e.g.*, intermittent dosing with a parathyroid cell ionlytic) can increase bone mass in patients suffering from osteoporosis.

Additionally, diseases or disorders characterized by a defect in one or more inorganic ion receptor activities may be treated by the present invention. For example, certain forms of primary hyperparathyroidism are characterized by abnormally high levels of parathyroid hormone and decreased parathyroid gland responsiveness to circulating calcium. Calcium receptor modulating agents can be used to modulate parathyroid cell responsiveness to calcium.

Preferably, the compound modulates calcium receptor activity and is used in the treatment of diseases or disorders which can be affected by modulating one or more activities of a calcium receptor. Preferably, the disease or disorder is characterized by abnormal bone and mineral homeostasis, more preferably calcium homeostasis.

Abnormal calcium homeostasis is characterized by one or more of the following activities: (1) an abnormal increase or decrease in serum calcium; (2) an abnormal increase or decrease in urinary excretion of calcium; (3) an abnormal increase or decrease in bone calcium levels, for example, as assessed by bone mineral density measurements; (4) an abnormal absorption of dietary calcium; and (5) an abnormal increase or decrease in the production and/or release of circulating messengers or hormones which affect calcium homeostasis such as parathyroid hormone and calcitonin. The abnormal increase or decrease in these different aspects of calcium homeostasis is relative to that occurring in the general population and is generally associated with a disease or disorder.

More generally, a molecule which modulates the activity of an inorganic ion receptor is useful in the treatment of diseases characterized by abnormal inorganic ion homeostasis. Preferably, the molecule modulates one or more effects of an inorganic ion receptor. Inorganic ion receptor modulating agents include ionmimetics, ionlytics, calcimimetics, and calcilytics.

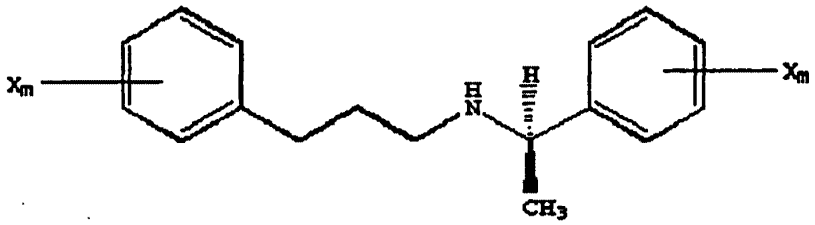
Ionmimetics are molecules which mimic the effects of increasing ion concentration at an inorganic ion receptor. Preferably, the molecule affects one or more calcium receptor activities. Calcimimetics are ionmimetics which affect one or more calcium receptor activities and preferably binds to a calcium receptor.

Ionlytics are molecules which reduce or block one or more activities caused by an inorganic ion on an inorganic ion receptor. Preferably, the molecule inhibits one or

more calcium receptor activities. Calcilytics are ionlytics which inhibit one or more calcium receptor activities evoked by extracellular calcium and preferably bind to a calcium receptor.

Inorganic ion receptor modulating agents can be formulated as pharmacological agents or compositions to facilitate administration in a patient. Pharmacological agents or compositions are agents or compositions in a form suitable for administration into a mammal, preferably a human. Considerations concerning forms suitable for administration are known in the art and include toxic effects, solubility, route of administration, and maintaining activity.

Thus, a first aspect the invention features an inorganic ion receptor modulating agent comprising a molecule which either evokes one or more inorganic ion receptor activities, or blocks one or more inorganic ion receptor activity caused by an extracellular inorganic ion. The molecule has the formula:



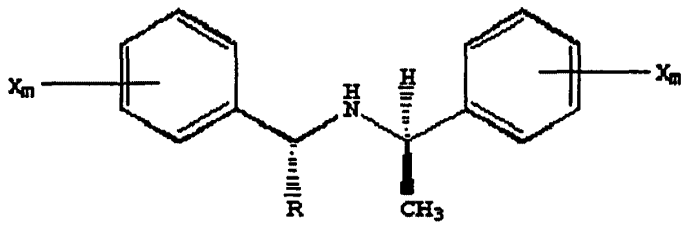
where each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, an aliphatic ring and an attached or fused aromatic ring; and

each m is independently between 0 and 5 inclusive.

Preferably, the aromatic and aliphatic rings have 5 to 7 members. More preferably, the aromatic and aliphatic rings contain only carbon atoms (i.e., the ring is not a heterocyclic ring).

Preferably, the molecule either evokes one or more calcium receptor activities, or blocks one or more calcium receptor activities caused by extracellular calcium.

Another aspect of the present invention features an inorganic ion receptor modulating agent having the formula:



where each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, acetoxy, aliphatic ring and an attached or fused aromatic ring;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, allyl, isobutyl, t-butyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-, 3-, or 4- piperid(in)yl; and

each m is independently between 0 and 5 inclusive.

The molecule either evokes one or more inorganic ion receptor activities, or blocks one or more inorganic ion receptor activities caused by an extracellular inorganic ion. Preferably, the molecule either evokes one or more calcium receptor activities, or blocks one or more calcium receptor activities caused by extracellular calcium.

In preferred embodiments R is either H, CH_3, ethyl, or isopropyl, and each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, aliphatic ring and an attached or fused aromatic ring. Preferably, the aliphatic ring and attached or fused aromatic ring have 5 to 7 members. More preferably, the aromatic and aliphatic rings contain only carbon atoms.

Another aspect of the present invention features an inorganic ion receptor modulating agent comprising a molecule selected from the group consisting of compound 4L, compound 8J, compound 8U, compound 9R, compound 11X, compound 12U, compound 12V, compound 12Z, compound 14U, compound 16M and compound 16P.

Other aspects of the present invention feature methods for using the agents described herein for treating diseases or disorders by modulating inorganic ion receptor activity. Patients in need of such treatments can be identified by standard medical techniques, such as routine blood analysis. For example, by detecting a deficiency of protein whose production or secretion is affected by changes in inorganic ion concentrations, or by detecting abnormal levels of inorganic ions or hormones which effect inorganic ion homeostasis.

Therapeutic methods involve administering to the patient a therapeutically effective amount of an inorganic ion receptor modulating agent. In preferred embodiments these methods are used to treat a disease or disorder characterized by abnormal inorganic ion homeostasis, more preferably a disease or disorder characterized by abnormal calcium homeostasis. Diseases and disorders characterized by abnormal calcium homeostasis include hyperparathyroidism, osteoporosis and other bone and mineral-related disorders, and the like (as described, *e.g.*, in standard medical text books, such as "Harrison's Principles of Internal Medicine"). Such diseases and disorders are treated using calcium receptor modulating agents which mimic or block one or more of the effects of Ca^{2+} and, thereby, directly or indirectly affect the levels of proteins or other molecules in the body of the patient.

By "therapeutically effective amount" is meant an amount of an agent which relieves to some extent one or more symptoms of the disease or disorder in the patient; or returns to normal either partially or completely one or

more physiological or biochemical parameters associated with or causative of the disease or disorder.

In a preferred embodiment, the patient has a disease or disorder characterized by an abnormal level of one or more calcium receptor regulated components and the molecule is active on a calcium receptor of a cell selected from the group consisting of parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, central nervous system cell, peripheral nervous system cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cell), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin-secreting cell, glucagon-secreting cell, kidney mesangial cell, mammary cell, beta cell, fat/adipose cell, immune cell and GI tract cell.

More preferably, the cell is a parathyroid cell and the molecule reduces the level of parathyroid hormone in the serum of the patient, even more preferably the level is reduced to a degree sufficient to cause a decrease in plasma Ca^{2+}, most preferably the parathyroid hormone level is reduced to that present in a normal individual.

Thus, the present invention features agents and methods useful in the treatment of diseases and disorders by modulating inorganic ion receptor activity. For example, the molecules of the present invention can be used to target calcium receptors on different cell types that detect and respond to changes to external calcium. For example, molecules mimicking external calcium may be used to selectively depress secretion of parathyroid hormone from parathyroid cells, or depress bone resorption by osteoclasts, or stimulate secretion of calcitonin from C-cells. Such molecules can be used to treat diseases or disorders characterized by abnormal calcium homeostasis such as hyperparathyroidism and osteoporosis.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof and from the claims.

BRIEF DESCRIPTION OF THE DRAWING

Figs. 1A-D, 2A-D, 3A-E, 4A-E, 5A-D, 6A-E, 7A-E, 8A-E, 9A-F, 10A-E, 11A-E, 12A-D, 13A-D, 14A-D, 15A-D, 16A-D, 17A-D, 18A-E, 19A-D, and 20A-D show the chemical structures of molecules derived from diphenylpropyl-α-phenethylamine illustrating a family of molecules which were prepared and screened to find the useful molecules of the invention.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention describes inorganic ion receptor modulating agents able to mimic or block an effect of an inorganic ion at an inorganic ion receptor. The preferred use of inorganic ion receptor modulating agents is to treat a disease or disorder by modulating inorganic ion receptor activity. Preferably, the molecules are used to treat diseases or disorders characterized by abnormal ion homeostasis, more preferably abnormal calcium homeostasis. Other uses of inorganic ion receptor modulating agents, such as diagnostics uses, are known in the art. Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.

I. CALCIUM RECEPTORS

Calcium receptors and nucleic acid encoding calcium receptors are described by Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959. Calcium receptors are present on different cell types such as parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, central nervous system cell, peripheral nervous system cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cell), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin-

secreting cell, glucagon-secreting cell, kidney mesangial cell, mammary cell, beta cell, fat/adipose cell, immune cell, and GI tract cell. The calcium receptor on these

cell types may be different. It is also possible that a cell can have more than one type of calcium receptor.

Comparison of calcium receptor activities and amino acid sequences from different cells indicate that distinct calcium receptor types exist. For example, calcium receptors can respond to a variety of di- and trivalent cations. The parathyroid calcium receptor responds to calcium and Gd^{3+}, while osteoclasts respond to divalent cations such as calcium but does not respond to Gd^{3+}. Thus, the parathyroid calcium receptor is pharmacologically distinct from calcium receptor on the osteoclast.

On the other hand, the nucleic acid sequences encoding calcium receptors present in parathyroid cells and C-cells indicate that these receptors have a very similar amino acid structure. Nevertheless, calcimimetic compounds exhibit differential pharmacology and regulate different activities at parathyroid cells and C-cells. Thus, pharmacological properties of calcium receptors may vary significantly depending upon the cell type or organ in which they are expressed even though the calcium receptors may have similar structures.

Calcium receptors, in general, have a low affinity for extracellular Ca^{2+} (apparent K_d generally greater than about 0.5 mM). Calcium receptors may include a free or bound effector mechanism as defined by Cooper, Bloom and Roth, "The Biochemical Basis of Neuropharmacology", Ch. 4, and are thus distinct from intracellular calcium receptors, *e.g.*, calmodulin and the troponins.

Calcium receptors respond to changes in extracellular calcium levels. The exact changes depend on the particular receptor and cell line containing the receptor. For example, the *in vitro* effect of calcium on the calcium receptor in a parathyroid cell include the following:

1. An increase in internal calcium. The increase is due to the influx of external calcium and/or

mobilization of internal calcium. Characteristics of the increase in internal calcium include the following:

(a) A rapid (time to peak < 5 seconds) and transient increase in $[Ca^{2+}]_i$ that is refractory to inhibition by 1 μM La^{3+} or 1 μM Gd^{3+} and is abolished by pretreatment with ionomycin (in the absence of extracellular Ca^{2+});

(b) The increase is not inhibited by dihydropyridines;

(c) The transient increase is abolished by pretreatment for 10 minutes with 10 mM sodium fluoride;

(d) The transient increase is diminished by pretreatment with an activator of protein kinase C (PKC), such as phorbol myristate acetate (PMA), mezerein or (-)-indolactam V. The overall effect of the protein kinase C activator is to shift the concentration-response curve to calcium to the right without affecting the maximal response; and

(e) Treatment with pertussis toxin (100 ng/ml for > 4 hours) does not affect the increase.

2. A rapid (< 30 seconds) increase in the formation of inositol-1,4,5-triphosphate or diacylglycerol. Treatment with pertussis toxin (100 ng/ml for > 4 hours) does not affect this increase;

3. The inhibition of dopamine- and isoproterenol-stimulated cyclic AMP formation. This effect is blocked by pretreatment with pertussis toxin (100 ng/ml for > 4 hours); and

4. The inhibition of PTH secretion. Treatment with pertussis toxin (100 ng/ml for > 4 hours) does not affect the inhibition in PTH secretion.

Using techniques known in the art, the effect of calcium on other calcium receptors in different cells can be readily determined. Such effects may be similar in regard to the increase in internal calcium observed in parathyroid cells. However, the effect is expected to

differ in other aspects, such as causing or inhibiting the release of a hormone other than parathyroid hormone.

II. INORGANIC ION RECEPTOR MODULATING AGENTS

Inorganic ion receptor modulating agents either evokes one or more inorganic ion receptor activities, or blocks one or more inorganic ion receptor activities caused by an extracellular inorganic ion. Calcium receptor modulating agents can mimic or block an effect of extracellular Ca^{2+} on a calcium receptor. Preferred calcium receptor modulating agents are calcimimetics and calcilytics. Generic and specific structures of inorganic ion receptor modulating agents are provided in the Summary *supra*, and in Figure 1.

Inorganic ion receptor modulating agents can be identified by screening molecules which are modelled after a molecule shown to have a particular activity (i.e., a lead molecule). Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.

Preferred inorganic ion receptor modulation agents described by the present invention are compounds are 8J, 8U, 9R, 11X, 12U, 12V, 12Z, 14U, 16M, and 16P. These compounds all have EC_{50} values of less than 5 μM.

The EC_{50} is the concentration of the molecule which evokes a half-maximal effect. The IC_{50} is the concentration of molecule which causes a half-maximal blocking effect. The EC_{50} or IC_{50} can be determined by assaying one or more of the activities of an inorganic ion at an inorganic ion receptor. Preferably, such assays are specific to a particular calcium receptor. For example, assays which measure hormones whose production or secretion is modulated by a particular inorganic ion receptor are preferred.

Increases in $[Ca^{2+}]_i$ can be detected using standard techniques such as by using fluorimetric indicators or by measuring an increase in Cl^- current in a *Xenopus* oocyte injected with nucleic acid coding for a calcium receptor. Nemeth *et al.*, PCT/US93/01642, International Publication

Number WO 94/18959. For example, poly(A)$^+$ mRNA can be obtained from cells expressing a calcium receptor, such as a parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cell), intestinal cell, central nervous cell, peripheral nervous system cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin-secreting cell, glucagon-secreting cell, kidney mesangial cell, mammary cell, beta cell, fat/adipose cell, immune cell, and GI tract cell. Preferably, the nucleic acid is from a parathyroid cell, C-cell, or osteoclast. More preferably, the nucleic acid encodes a calcium receptor and is present on a plasmid or vector.

Preferably, the molecule is either a calcimimetic or calcilytic having an EC_{50} or IC_{50} at a calcium receptor of less than or equal to 5 μM, and even more preferably less than or equal to 1 μM, 100 nmolar, 10 nmolar, or 1 nmolar. Such lower EC_{50}'s or IC_{50}'s are advantageous since they allow lower concentrations of molecules to be used *in vivo* or *in vitro* for therapy or diagnosis. The discovery of molecules with such low EC_{50}'s and IC_{50}'s enables the design and synthesis of additional molecules having similar potency and effectiveness.

In preferred embodiments the calcium receptor modulating agent is a calcimimetic which inhibits parathyroid hormone secretion from a parathyroid cell *in vitro* and decreases PTH secretion *in vivo*; stimulates calcitonin secretion from a C-cell *in vitro* and elevates calcitonin levels *in vivo*; or blocks osteoclastic bone resorption *in vitro* and inhibits bone resorption *in vivo*.

In another preferred embodiment the calcium receptor modulating agent is a calcilytic which evokes the secretion of parathyroid hormone from parathyroid cells *in*

vitro and elevates the level of parathyroid hormone *in vivo*.

Preferably, the agent selectively targets inorganic ion receptor activity, more preferably calcium receptor activity, in a particular cell. By "selectively" is meant that the molecule exerts a greater effect on inorganic ion receptor activity in one cell type than at another cell type for a given concentration of agent. Preferably, the differential effect is 10-fold or greater. Preferably, the concentration refers to blood plasma concentration and the measured effect is the production of extracellular messengers such as plasma calcitonin, parathyroid hormone or plasma calcium. For example, in a preferred embodiment, the agent selectively targets PTH secretion over calcitonin secretion.

In another preferred embodiment, the molecule has an EC_{50} or IC_{50} less than or equal to 5 μM at one or more, but not all cells chosen from the group consisting of parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, central nervous system cell, peripheral nervous system cell, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cell), intestinal cell, trophoblast in the placenta, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin-secreting cell, glucagon-secreting cell, kidney mesangial cell, mammary cell, beta cell, fat/adipose cell, immune cell and GI tract cell.

Preferably, inorganic ion receptor modulating agents mimic or block all of the effects of extracellular ion in a cell having an inorganic ion receptor. For example, calcium receptor modulating agents preferably mimic or block all of the effects of extracellular ion in a cell having a calcium receptor. Calcimimetics need not possess all the biological activities of extracellular Ca^{2+}, but, rather, at least one such activity is mimicked.

Similarly, calcilytics need not reduce or prevent all of the activities caused by extracellular calcium. Additionally, different calcimimetics and different calcilytics do not need to bind to the same site on the calcium receptor as does extracellular Ca^{2+} to exert their effects.

A. Calcimimetics

The ability of molecules to mimic or block the activity of Ca^{2+} at calcium receptors can be determined using procedures known in the art and described by Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959. For example, calcimimetics possess one or more and preferably all of the following activities when tested on parathyroid cells *in vitro*:

1. The molecule causes a rapid (time to peak < 5 seconds) and transient increase in $[Ca^{2+}]_i$ that is refractory to inhibition by 1 μM La^{3+} or 1 μM Gd^{3+}. The increase in $[Ca^{2+}]_i$ persists in the absence of extracellular Ca^{2+} but is abolished by pretreatment with ionomycin (in the absence of extracellular Ca^{2+});

2. The molecule potentiates increases in $[Ca^{2+}]_i$ elicited by submaximal concentrations of extracellular Ca^{2+};

3. The increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+} is not inhibited by dihydropyridines;

4. The transient increase in $[Ca^{2+}]_i$ caused by the molecule is abolished by pretreatment for 10 minutes with 10 mM sodium fluoride;

5. The transient increase in $[Ca^{2+}]_i$ caused by the molecule is diminished by pretreatment with an activator of protein kinase C (PKC), such as phorbol myristate acetate (PMA), mezerein or (-)-indolactam V. The overall effect of the protein kinase C activator is to shift the concentration-response curve of the molecule to the right without affecting the maximal response;

6. The molecule causes a rapid (< 30 seconds) increase in the formation of inositol-1,4,5-triphosphate and/or diacylglycerol;

7. The molecule inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation;

8. The molecule inhibits PTH secretion;

9. Pretreatment with pertussis toxin (100 ng/ml for > 4 hours) blocks the inhibitory effect of the molecule on cyclic AMP formation but does not effect increases in $[Ca^{2+}]_i$, inositol-1,4,5-triphosphate, or diacylglycerol, nor decreases in PTH secretion;

10. The molecule elicits increases in Cl^- current in *Xenopus* oocytes injected with poly(A)$^+$- enriched mRNA from bovine or human parathyroid cells, but is without effect in *Xenopus* oocytes injected with water, or rat brain or liver mRNA; and

11. Similarly, using a cloned calcium receptor from a parathyroid cell, the molecule will elicit a response in *Xenopus* oocytes injected with the specific cDNA or mRNA encoding the receptor.

Different calcium activities can be measured using available techniques. Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959. Parallel definitions of molecules mimicking Ca^{2+} activity on other calcium responsive cell, preferably at a calcium receptor, are evident from the examples provided herein and Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.

Preferably, the agent as measured by the bioassays described herein, or by Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959, has one or more, more preferably all of the following activities: evokes a transient increase in internal calcium, having a duration of less that 30 seconds (preferably by mobilizing internal calcium); evokes a rapid increase in $[Ca^{2+}]_i$, occurring within thirty seconds; evokes a sustained increase (greater than thirty seconds) in $[Ca^{2+}]_i$

(preferably by causing an influx of external calcium); evokes an increase in inositol-1,4,5-triphosphate or diacylglycerol levels, preferably within less than 60 seconds; and inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation.

The transient increase in $[Ca^{2+}]_i$ is preferably abolished by pretreatment of the cell for ten minutes with 10 mM sodium fluoride, or the transient increase is diminished by brief pretreatment (not more than ten minutes) of the cell with an activator of protein kinase C, preferably, phorbol myristate acetate (PMA), mezerein or (-) indolactam V.

B. Calcilytics

The ability of a molecule to block the activity of external calcium can be determined using standard techniques. Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959. For example, molecules which block the effect of external calcium, when used in reference to a parathyroid cell, possess one or more, and preferably all of the following characteristics when tested on parathyroid cells *in vitro*:

1. The molecule blocks, either partially or completely, the ability of increased concentrations of extracellular Ca^{2+} to:

(a) increase $[Ca^{2+}]_i$

(b) mobilize intracellular Ca^{2+},

(c) increase the formation of inositol-1,4,5-triphosphate,

(d) decrease dopamine- or isoproterenol-stimulated cyclic AMP formation, and

(e) inhibit PTH secretion;

2. The molecule blocks increases in Cl^- current in *Xenopus* oocytes injected with poly(A)$^+$ mRNA from bovine or human parathyroid cells elicited by extracellular Ca^{2+} or calcimimetic compounds, but not in *Xenopus* oocytes injected with water or liver mRNA;

3. Similarly, using a cloned calcium receptor from a parathyroid cell, the molecule will block a response in *Xenopus* oocytes injected with the specific cDNA, mRNA or cRNA encoding the calcium receptor, elicited by extracellular Ca^{2+} or a calcimimetic compound.

Parallel definitions of molecules blocking Ca^{2+} activity on a calcium responsive cell, preferably at a calcium receptor, are evident from the examples provided herein and Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959..

III. TREATMENT OF DISEASES OR DISORDERS

A preferred use of the compounds described by the present invention is in the treatment or prevention of different diseases or disorders by modulating inorganic ion receptor activity. The inorganic ion receptor modulating agents of the present invention can exert an affect on a inorganic ion receptor causing one or more cellular effects ultimately producing a therapeutic effect.

Different diseases and disorders can be treated by the present invention by targeting cells having an inorganic ion receptor, such as a calcium receptor. For example, primary hyperparathyroidism (HPT) is characterized by hypercalcemia and elevated levels of circulating PTH. A defect associated with the major type of HPT is a diminished sensitivity of parathyroid cells to negative feedback regulation by extracellular Ca^{2+}. Thus, in tissue from patients with primary HPT, the "set-point" for extracellular Ca^{2+} is shifted to the right so that higher than normal concentrations of extracellular Ca^{2+} are required to depress PTH secretion. Moreover, in primary HPT, even high concentrations of extracellular Ca^{2+} often depress PTH secretion only partially. In secondary (uremic) HPT, a similar increase in the set-point for extracellular Ca^{2+} is observed even though the degree to which Ca^{2+} suppresses PTH secretion is normal. The changes in PTH secretion are paralleled by changes in $[Ca^{2+}]_i$: the

set-point for extracellular Ca^{2+}-induced increases in $[Ca^{2+}]_i$ is shifted to the right and the magnitude of such increases is reduced.

Molecules that mimic the action of extracellular Ca^{2+} are beneficial in the long-term management of both primary and secondary HPT. Such molecules provide the added impetus required to suppress PTH secretion which the hypercalcemic condition alone cannot achieve and, thereby, help to relieve the hypercalcemic condition. Molecules with greater efficacy than extracellular Ca^{2+} may overcome the apparent nonsuppressible component of PTH secretion which is particularly troublesome in adenomatous tissue. Alternatively or additionally, such molecules can depress synthesis of PTH, as prolonged hypercalcemia has been shown to depress the levels of preproPTH mRNA in bovine and human adenomatous parathyroid tissue. Prolonged hypercalcemia also depresses parathyroid cell proliferation *in vitro*, so calcimimetics can also be effective in limiting the parathyroid cell hyperplasia characteristic of secondary HPT.

Cells other than parathyroid cells can respond directly to physiological changes in the concentration of extracellular Ca^{2+}. For example, calcitonin secretion from parafollicular cells in the thyroid (C-cells) is regulated by changes in the concentration of extracellular Ca^{2+}.

Isolated osteoclasts respond to increases in the concentration of extracellular Ca^{2+} with corresponding increases in $[Ca^{2+}]_i$ that arise partly from the mobilization of intracellular Ca^{2+}. Increases in $[Ca^{2+}]_i$ in osteoclasts are associated with the inhibition of bone resorption. Release of alkaline phosphatase from bone-forming osteoblasts is directly stimulated by calcium.

Renin secretion from juxtaglomerular cells in the kidney, like PTH secretion, is depressed by increased concentrations of extracellular Ca^{2+}. Extracellular Ca^{2+} causes the mobilization of intracellular Ca^{2+} in these cells. Other kidney cells respond to calcium as follows:

elevated Ca^{2+} inhibits formation of $1,25(OH)_2$-vitamin D by proximal tubular cells, stimulates production of calcium-binding protein in distal tubular cells, and inhibits tubular reabsorption of Ca^{2+} and Mg^{2+} and the action of vasopressin on the medullary thick ascending limb of Henle's loop (MTAL), reduces vasopressin action in the cortical collecting duct cells, and affects vascular smooth muscle cells in blood vessels of the renal glomerulus.

Calcium also promotes the differentiation of intestinal goblet cells, mammary cells, and skin cells; inhibits atrial natriuretic peptide secretion from cardiac atria; reduces cAMP accumulation in platelets; alters gastrin and glucagon secretion; acts on vascular smooth muscle cells to modify cell secretion of vasoactive factors; and affects cells of the central nervous system and peripheral nervous system.

Thus, there are sufficient indications to suggest that Ca^{2+}, in addition to its ubiquitous role as an intracellular signal, also functions as an extracellular signal to regulate the responses of certain specialized cells. Molecules of this invention can be used in the treatment of diseases or disorders associated with disrupted Ca^{2+} responses in these cells.

Specific diseases and disorders which might be treated or prevented, based upon the affected cells, also include those of the central nervous system such as seizures, stroke, head trauma, spinal cord injury, hypoxia-induced nerve cell damage such as in cardiac arrest or neonatal distress, epilepsy, neurodegenerative diseases such as Alzheimer's disease, Huntington's disease and Parkinson's disease, dementia, muscle tension, depression, anxiety, panic disorder, obsessive-compulsive disorder, post-traumatic stress disorder, schizophrenia, neuroleptic malignant syndrome, and Tourette's syndrome; diseases involving excess water reabsorption by the kidney such as syndrome of inappropriate ADH secretion (SIAH),

cirrhosis, heart failure, and nephrosis; hypertension; preventing and/or decreasing renal toxicity from cationic antibiotics (*e.g.*, aminoglycoside antibiotics); gut motility disorders such as diarrhea, and spastic colon; GI ulcer diseases; GI absorption diseases such as sarcoidosis; and autoimmune diseases and organ transplant rejection.

While inorganic ion receptor modulating agents of the present invention will typically be used in therapy for human patients, they may be used to treat similar or identical diseases or disorders in other warm-blooded animal species such as other primates, farm animals such as swine, cattle, and poultry; and sports animals and pets such as horses, dogs and cats.

IV. ADMINISTRATION

The molecules of the invention can be formulated for a variety of modes of administration to treat patients by modulating inorganic ion receptor activity. Techniques and formulations for administration of compounds generally may be found in Remington's Pharmaceutical Sciences, Mack Publishing Co., Easton, PA. Administration of ionmimetics and ionlytics is discussed by Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.

Suitable forms, in part, depend upon the use or the route of entry, for example oral, transdermal, or by injection. Such forms should allow the agent to reach a target cell whether the target cell is present in a multicellular host or in culture. For example, pharmacological agents or compositions injected into the blood stream should be soluble in the concentrations used. Other factors are known in the art, and include considerations such as toxicity and forms which prevent the agent or composition from exerting its effect.

Agents can also be formulated as pharmaceutically acceptable salts (*e.g.*, acid addition salts) and complexes thereof. The preparation of such salts can facilitate the

pharmacological use by altering the physical characteristics of the agent without preventing it from exerting its physiological effect. Examples of useful alterations in physical properties include lowering the melting point to facilitate transmucosal administration and increasing the solubility to facilitate administering higher concentrations of the drug.

For systemic administration, oral administration is preferred. Alternatively, injection may be used, *e.g.*, intramuscular, intravenous, intraperitoneal, and subcutaneous. For injection, the molecules of the invention are formulated in liquid solutions, preferably in physiologically compatible buffers such as Hank's solution or Ringer's solution. In addition, the molecules may be formulated in solid form and redissolved or suspended immediately prior to use. Lyophilized forms can also be produced.

Systemic administration can also be by transmucosal or transdermal means, or the molecules can be administered orally. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, bile salts and fusidic acid derivatives. In addition, detergents may be used to facilitate permeation. Transmucosal administration may be through nasal sprays, for example, or using suppositories. For oral administration, the molecules are formulated into conventional oral administration dosage forms such as capsules, tablets, and tonics.

For topical administration, the molecules of the invention are formulated into ointments, salves, gels, or creams, as is generally known in the art.

Generally, a therapeutically effective amount is between about 1 nmole and 3 μmole of the molecule, preferably 0.1 nmole and 1 μmole depending on its EC_{50} or IC_{50} and on the age and size of the patient, and the

disease or disorder associated with the patient. Generally it is an amount between about 0.1 and 50 mg/kg, preferably 0.01 and 20 mg/kg, animal to be treated.

V. EXAMPLES

The compounds described herein can be synthesized using standard techniques such as those described by Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959. Examples describing the syntheses of compounds 4L, 8J, 8U, 9R, 11X, 12U, 12V, 12Z, 14U, 16M and 16P are provided below. Compounds 4L, 8J, 8U, 11X and 16M were prepared from the condensation of a primary amine with an aldehyde or ketone in the presence of titanium(IV) isopropoxide. The resulting intermediate imines were then reduced *in situ* by the action of sodium cyanoborohydride, sodium borohydride, or sodium triacetoxyborohydride. The intermediate enamine for the synthesis of compound 8U was catalytically reduced using palladium hydroxide.

Synthesis of compounds 9R, 14U, and 16P were prepared by reductive amination of a commercially available aldehyde or ketone with a primary amine in the presence of sodium cyanoborohydride or sodium triacetoxyborohydride. It was found for the syntheses of these three compounds (9R, 14U, and 16P) that sodium triacetoxyborohydride afforded the desired diastereomers with greater diastereoselectivity than using sodium cyanoborohydride. The enriched mixtures were further purified to a single diastereomer by normal-phase HPLC or by recystallization from organic solvents.

Compounds 12U, 12V and 12Z were prepared by a diisobutylaluminum hydride (DIBAL-H) mediated condensation of an amine with a nitrile. The resulting intermediate imine is reduced *in situ* by the action of sodium cyanoborohydride or sodium borohydride. The intermediate alkenes (compounds 12U and 12V) were reduced by catalytic hydrogenation in EtOH using palladium on carbon. Compounds which were converted to their corresponding

hydrochloride were done so by treatment of the free base with ethereal HCl to afford white solids.

The amines in these syntheses were (1) purchased from Aldrich Chemical Co., Milwaukee, WI, (2) purchased from Celgene Corp., Warren, NJ, or (3) prepared synthetically using standard techniques. All other reagent chemicals were purchased from Aldrich Chemical Co.

Example 1: Synthesis of Compound 4L

N-3-Phenyl-1-propyl-1-(1-naphthyl)ethylamine

A mixture of 3-phenyl-1-propylamine (135 mg, 1 mmol), 1'-acetonaphthone (170 mg, 1 mmol), and titanium (IV) isopropoxide (355 mg, 1.3 mmol) was stirred at room temperature for 1 hour. The reaction was treated with 1 M ethanolic sodium cyanoborohydride (1 mL) and stirred at room temperature for 16 hours. The reaction was diluted with ether and treated with water (0.1 mL). The reaction was centrifuged and the ether layer removed and concentrated to a milky oil. A small portion of this material (10 mg) was purified by HPLC (Phenomenex, 1.0 x 25 cm, 5 μM silica) using a gradient of dichloromethane to 10% methanol in dichloromethane containing 0.1% isopropylamine. This afforded the product (free base) as a single component by GC/El-MS (R_t= 10.48 min) m/z (rel. int.) 289 (M^+,11), 274 (63), 184 (5), 162 (5), 155 (100), 141 (18), 115 (8), 91 (45), 77(5).

Example 2: Synthesis of Compound 8J

N-(3-phenylpropyl)-1-(3-thiomethylphenyl)ethylamine hydrochloride

3'-Aminoacetophenone (2.7 g, 20 mmol) was dissolved in 4 mL of concentrated HCl, 4 g of ice and 8 mL of water. The solution was cooled to 0°C, and sodium nitrite (1.45 g, 21 mmol) dissolved in 3-5 mL of water was added over 5 minutes while maintaining the temperature below 6°C. Sodium thiomethoxide (1.75 g, 25 mmol) was dissolved in 5 mL of water and cooled to 0°C. To this solution was added

the diazonium salt over 10 minutes while maintaining the temperature below 10°C. The reaction was stirred for an additional hour while allowing the temperature to rise to ambient. The reaction mixture was partitioned between ether and water. The ether layer was separated and washed with sodium bicarbonate and sodium chloride, and dried over sodium sulfate. The ether was evaporated to give a 74% yield of 3′-thiomethylacetophenone. The crude material was purified by distillation at reduced pressure.

3-Phenylpropylamine (0.13 g, 1 mmol), 3′-thiomethylacetophenone (0.17 g, 1 mmol), and titanium (IV) isopropoxide (0.36 g, 1.25 mmol) were mixed together and allowed to stand for 4 hours. Ethanol (1 mL) and sodium cyanoborohydride (0.063 g, 1 mmol) were added and the reaction was stirred overnight. The reaction was worked up by the addition of 4 mL of ether and 200 μL of water. The mixture was vortexed and then spun in a centrifuge to separate the solids. The ether layer was separated from the precipitate, and the solvent removed *in vacuo*. The oil was redissolved in dichloromethane and the compound purified by preparative TLC on silica gel eluted with 3% methanol/dichloromethane to yield the title compound as a pure oil: GC/EI-MS(R_t=7.64 min) m/z (rel. int.)285 (M^+,18), 270(90), 180(17), 151(100), 136(32), 104(17), 91(54), 77(13).

Example 3: Synthesis of Compound 8U

N-3-(2-methoxyphenyl)-1-propyl-(R)-3-methoxy-α-methylbenzylamine hydrochloride

A mixture of (*R*)-(+)-3-methoxy-α-methylbenzylamine (3.02 g, 20 mmol), 2-methoxycinnamaldehyde (3.24 g, 20 mmol), and titanium (IV) isopropoxide (8.53 g, 30 mmol, 1.5 Eq.) was stirred 2 hours at room temperature and treated with 1 M (20 mL) ethanolic sodium cyanoborohydride. The reaction was stirred overnight (16 hours), diluted with diethylether, and treated with water (1.44 mL, 80 mmol, 4 Eq.). After mixing for 1 hour the

reaction mixture was centrifuged and the ether layer removed and concentrated to an oil. This material was dissolved in glacial acetic acid, shaken with palladium hydroxide and hydrogenated under 60 p.s.i. hydrogen for 2 hours at room temperature. The catalyst was removed by filtration and the resulting solution concentrated to a thick oil. This material was dissolved in dichloromethane and neutralized with 1 N NaOH. The dichloromethane solution was separated from the aqueous phase, dried over anhydrous potassium carbonate and concentrated to an oil. This material was dissolved in ether and treated with 1 M HCl in diethylether. The resulting precipitate (white solid) was collected, washed with diethylether, and air dried. GC/El-MS (R_t = 9.69 min) of this material (free base) showed a single component: m/z (rel. int.) 299 (M+, 21), 284 (100), 164 (17), 150 (8), 135 (81), 121 (40), 102 (17), 91 (43), 77 (18).

Example 4: Synthesis of Compound 9R

(R)-N-(1-(2-naphthyl)ethyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride

A mixture of (*R*)-(+)-1-(1-naphthyl)ethylamine (10.0 g, 58 mmol), 2'-acetonaphthone (9.4 g, 56 mmol), titanium (IV) isopropoxide (20.7 g, 73.0 mmol), and EtOH (abs.) (100 mL) was heated to 60°C for 3 hours. Sodium cyanoborohydride ($NaCNBH_3$) (3.67 g, 58.4 mmol) was then added. The reaction mixture was stirred at room temperature for 18 hours. Ether (1 L) and H_20 (10 mL) were added to the reaction mixture and the resulting precipitate was then removed by centrifugation. The supernatant was evaporated under vacuum and the crude product was recrystallized four times from hot hexane, to provide 1.5 g of pure (98+%) diastereomer. The free base was dissolved in hexane, filtered, and then ethereal HCl was added to precipitate the product as a white solid (1.1 g, 6 % yield), m.p.: softens 200-240 °C (dec.).

Example 5: Synthesis of Compound 11X

N-(4-Isopropylbenzyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride

A mixture of (*R*)-(+)-1-(1-naphthyl)ethylamine (1.06 g, 6.2 mmol), 4-isopropylbenzaldehyde (0.92 g, 6.2 mmol), and titanium (IV) isopropoxide (2.2 g, 7.7 mmol) was heated to 100°C for 5 min then allowed to stir at room temperature for 4 hours. Sodium cyanoborohydride ($NaCNBH_3$) (0.39 g, 6.2 mmol) was then added followed by EtOH (1 mL). The reaction mixture was stirred at room temperature for 18 hours. Ether (100 mL) and H_2O (1 mL) were added to the reaction mixture and the resulting precipitate was then removed by centrifugation. The supernatant was evaporated under vacuum and the crude product was chromatographed on silica gel (50 mm X 30 cm column) (elution with 1% $MeOH/CHCl_3$). The chromatographed material was then dissolved in hexane and ethereal HCl was added to precipitate the product as a white solid (0.67 g, 35 % yield), m.p.; 257-259°C.

Example 6: Synthesis of Compound 12U

N-3-(2-methylphenyl)-1-propyl-(R)-3-methoxy-α-methylbenzylamine hydrochloride

A solution of 2-methylcinnamonitrile (1.43 g, 10 mmol) in dichloromethane (10 mL) was cooled to 0°C and treated dropwise (15 minutes) with 1 M diisobutylaluminum hydride (10 mL, dichloromethane). The reaction was stirred at 0°C for 15 minutes and treated dropwise (15 minutes) with a 1 M solution of (*R*)-(+)-3-methoxy-α-methylbenzylamine (1.51 g, 10 mmol) in dichloromethane (10 mL). The reaction was stirred 1 hours at 0°C and poured into a solution of ethanol (100 mL) containing sodium cyanoborohydride (1 g, 16 mmol). The reaction mixture was stirred 48 hour at room temperature. The reaction was diluted with ether and neutralized with 1 N NaOH. The ether layer was removed, dried over anhydrous potassium carbonate and concentrated to an oil. This material was

chromatographed through silica using a gradient of dichloromethane to 5% methanol in dichloromethane to afford the unsaturated intermediate, a single component by GC/El-MS (R_t=10.06 min) m/z (rel. int.) 281 (M+, 17), 266 (59), 176 (19), 146 (65), 135 (73), 131 (100), 91 (21), 77 (13).

The unsaturated intermediate in ethanol was hydrogenated (1 atm H_2) in the presence of palladium on carbon for 16 hours at room temperature. The product from this reaction was converted to the hydrochloride salt by treatment with 1 M HCl in diethylether. GC/El-MS (R_t = 9.31 min) of this material (free base) showed a single component: m/z (rel. int.) 283 (M+, 21), 268 (100), 164 (12), 148 (8), 135 (85), 121 (12), 105 (49), 91 (23), 77 (21).

Example 7: Synthesis of Compound 12V

N-3-(3-methylphenyl)-1-propyl-(R)-3-methoxy-α-methylbenzylamine hydrochloride

The compound was prepared following the procedure described in Example 6, but using 2-methylcinnamonitrile. The unsaturated intermediate was a single component by GC/EI-MS (R_t = 10.21 min) m/z (rel. int.) 281 (M+, 57), 266 (86), 146 (98), 135 (88), 131 (100), 115 (43), 102 (26), 91 (43), 77 (18). Reduction of this material and hydrochloride formation using the procedure described Example 6 afforded the product. GC/EI-MS (R_t = 9.18 min) of this material (free base) showed a single component; m/z (rel. int.) 283 (M+, 19), 268 (100), 164 (11), 148 (8), 135 (76), 121 (16), 105 (45), 91 (23), 77 (21).

Example 8: Synthesis of Compound 12Z

N-3-(2-chlorophenyl)-1-propyl-(R)-1-(1-naphthyl)ethylamine hydrochloride

The compound was prepared following the procedures described in Example 6, but using 2-chlorohydrocinnamonitrile and (*R*)-(+)-1-(1-

naphthyl)ethylamine on a 10 mmol scale. Chromatography through silica using a gradient of dichloromethane to 5% methanol in dichloromethane afforded the product as a single component by TLC analysis (5% methanol in dichloromethane). The hydrochloride was prepared by treatment with 1 M HCl in diethylether.

Example 9: Synthesis of Compound 14U

(R)-N-(1-(4-methoxyphenyl)ethyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride

A mixture of (*R*)-(+)-1-(1-naphthyl)ethylamine (1.1 g, 6.2 mmol), 4'-methoxyacetophenone (0.93 g, 6.2 mmol), titanium (IV) isopropoxide (2.2 g, 7.7 mmol), and EtOH (abs.) (1 mL) was heated to 60°C for 3 hours. Sodium cyanoborohydride ($NaCNBH_3$) (0.39 g, 6.2 mmol) was then added, and the reaction mixture was stirred at room temperature for 18 hours. Ether (200 mL) and H_2O (2 mL) were added to the reaction mixture and the resulting precipitate was then removed by centrifugation. The supernatant was evaporated under vacuum and the crude product was chromatographed on silica gel (25 mm X 25 cm column) (elution with 1% MeOH/$CHCl_3$). A portion of this material was HPLC chromatographed [Selectosil, 5 μM silica gel; 25 cm x 10.0 mm (Phenomenex, Torrance, CA), 4 mL per minute; UV det. 275 nM; 12% ethyl acetate-88% hexane (elution time 12.0 min)]. The HPLC purified diastereomer was then dissolved in hexanes and ethereal HCl was added to precipitate the product as a white solid (20 mg), m.p.: 209-210°C(dec.).

Example 10: Synthesis of Compound 16M

N-(3-chloro-4-methoxybenzyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride

A mixture of (*R*)-(+)-1-(1-naphthyl)ethylamine (6.6 g, 39 mmol), 3'-chloro-4'-methoxybenzaldehyde (6.6 g, 39 mmol), and titanium (IV) isopropoxide (13.8 g, 48.8 mmol), and EtOH (abs.) (30 mL) was heated to 80°C for 30 minutes

then allowed to stir at room temperature for 3 hours. Sodium cyanoborohydride ($NaCNBH_3$) (2.45 g, 39 mmol) was then added. The reaction mixture was stirred at room temperature for 18 hours. Ether (100 mL) and H_2O (2 mL) were added to the reaction mixture and the resulting precipitate was then removed by centrifugation. The supernatant was evaporated under vacuum and the crude product was chromatographed on silica gel (50 mm X 30 cm column) (elution with CH_2Cl_2). The chromatographed material was then dissolved in hexane (500 mL), decolorized with Norit® filtered (0.2 μM), and then ethereal HCl was added to precipitate the product as a while solid (10.2 g, 56 % yield), m.p.: 241-242°C (dec.).

Example 11: Synthesis of Compound 16P

4-Methoxy-3-methylacetophenone [16P Precursor]

A mixture of 4'-hydroxy-3'-methylacetophenone (5.0 g, 33.3 mmol), iodomethane (5.7 g, 40.0 mmol), K_2CO_3 (granular, anhydrous) (23.0 g, 167 mmol), and acetone (250 mL) was refluxed for 3 hours. The reaction mixture was then cooled to room temperature, filtered to remove the inorganic salts, and evaporated under vacuum. The crude product was dissolved in ether (100 mL) and washed with H_2O (2 x 20 mL). The organic layer was dried (Na_2SO_4) and evaporated to yield 4.5 g, 82.4% yield. The ketone was used in the following reaction without further purification.

(R)-N-(1-(4-Methoxy-3-methylphenyl)ethyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride [Compound 16P]

A mixture of (*R*)-(+)-1-(1-naphthyl)ethylamine (4.24 g, 24.8 mmol), 4'-methoxy-3'-methylacetophenone (4.06 g, 24.8 mmol), and titanium (IV) isopropoxide(8.8 g, 30.9 mmol), and EtOH (abs.) (1 mL) was heated to 100°C for 2 hours. Isopropanol (45 mL) was added and the reaction was then cooled to 10°C in an ice bath. Sodium triacetoxyborohydride ($NaHB(O_2CCH_3)_3$) (10.5 g, 49.5 mmol)

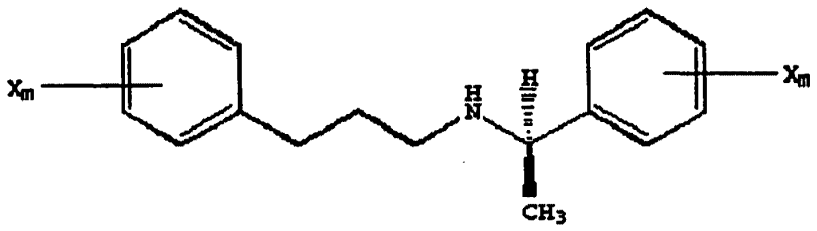
was then added in portions over 15 minutes. The reaction mixture was then heated to 70°C for 18 hours. The mixture was cooled to room temperature and poured into ether (400 mL). The suspension was centrifuged, the supernatant was collected and the pellet was washed with ether (400 mL). The combined organic washings were evaporated under vacuum. The residue was dissolved in ether (400 mL) and washed with 1 N NaOH (4 x 50 mL) and H_2O (2 x 50 mL). The organic layer was dried (Na_2SO_4), filtered and evaporated under vacuum. EtOH (abs.) was added to the wet residue which was then dried thoroughly on a rotary evaporator to provide an oil. The mixture was then chromatographed on silica gel (50 mm x 30 cm) [elution with (1% MeOH:1% IPA:$CHCl_3$) to give 4.8 g of an oil].

The desired diastereomer was further purified by HPLC chromatography [SUPELCOSIL™ PLC-Si, 18 μM silica gel; 25 cm x 21.2 mm (Supelco, Inc., Bellefonte, PA), 7 mL per minute; UV det. 275 nM: 20% EtOAc-80% hexane (elution time 9.5 - 11.0 min)]. Injections (800 μL aliquots) of the mixture (100 mg/mL solution in eluent) provided 65 mg of the desired isomer. Multiple HPLC injections provided 1.0 g of purified material. The HPLC chromatographed material was dissolved in hexane (50 mL) and the hydrochloride salt was precipitated with ethereal HCl. The salt was collected on fritted glass and washed with hexane to provide 1.0 g of a white solid, mp 204-205°C.

Other embodiments are within the following claims.

Claims

1. An inorganic ion receptor modulating agent comprising a molecule having the formula:

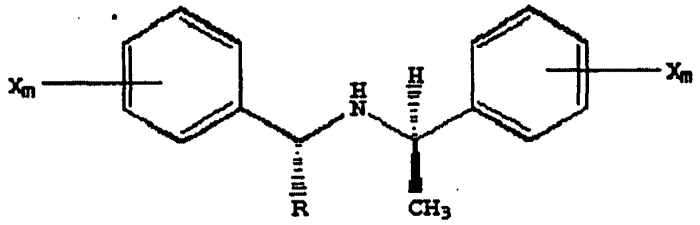


wherein each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O an aliphatic ring and an attached or fused aromatic ring; and

each m is independently between 0 and 5 inclusive; wherein said molecule either evokes one or more inorganic ion receptor activities, or blocks one or more inorganic ion receptor activities caused by an extracellular inorganic ion.

2. The agent of claim 1 wherein said molecule is a calcimimetic and said inorganic ion receptor activity is a calcium receptor activity.

3. An inorganic ion receptor modulating agent comprising a molecule having the formula:



wherein each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN,

NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, acetoxy, aliphatic ring and an attached or fused aromatic ring;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, allyl, isobutyl, t-butyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-, 3-, or 4- piperid(in)yl; and

each m is independently between 0 and 5 inclusive; wherein said molecule either evokes one or more inorganic ion receptor activities, or blocks one or more inorganic ion receptor activities caused by an extracellular inorganic ion.

4. The agent of claim 3 wherein said molecule is a calcimimetic and said inorganic ion receptor activity is a calcium receptor activity.

5. The agent of claim 4 wherein each R is independently selected from the group consisting of H, CH_3, ethyl, and isopropyl.

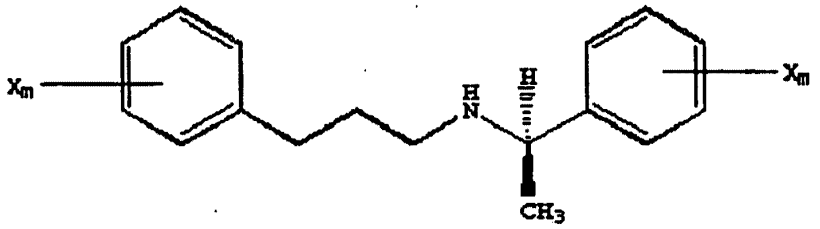
6. The agent of claim 4 wherein each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, aliphatic ring and an attached or fused aromatic ring.

7. The agent of claim 5 wherein each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, aliphatic ring and an attached or fused aromatic ring.

8. An inorganic ion receptor modulation agent comprising a molecule selected from the group consisting of compound 4L, compound 8J, compound 8U, compound 9R, compound 11X, compound 12U, compound 12V, compound 12Z, compound 14U, compound 16M, and compound 16P.

9. An agent of any claim 1-8 further comprising a physiologically acceptable carrier.

10. A method for treating a patient comprising the step of administering a therapeutically effective amount of an inorganic ion receptor modulating agent to a patient in need of such treatment, said agent comprising a molecule having the formula:



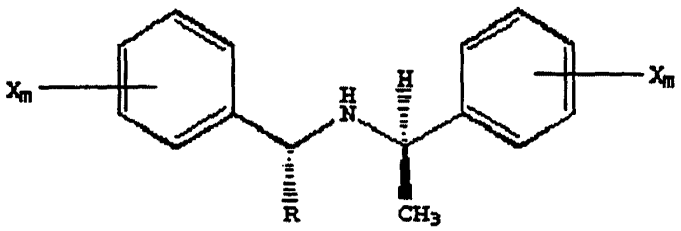
wherein each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O an aliphatic ring and an attached or fused aromatic ring; and

each m is independently between 0 and 5 inclusive; wherein said molecule either evokes in one or more inorganic ion receptor activities, or blocks one or more inorganic ion receptor activities caused by an extracellular inorganic ion.

11. The agent of claim 10 wherein said molecule is a calcimimetic and said inorganic ion receptor activity is a calcium receptor activity.

12. A method for treating a patient comprising the step of administering a therapeutically effective amount of an inorganic ion receptor modulating agent to a patient in need of such treatment, said agent comprising a molecule having the formula:

35



wherein each X independently is selected from the group consisting of H, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, CF_3, CHF_2, CH_2F, CF_3O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, acetoxy, aliphatic ring and an attached or fused aromatic ring;

each R independently is selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, butyl, allyl, isobutyl, *t*-butyl, cyclopentyl, cyclohexyl, cycloheptyl, cyclooctyl, indenyl, indanyl, dihydroindolyl, thiodihydroindolyl, and 2-, 3-, or 4- piperid(in)yl; and

each m is independently between 0 and 5 inclusive;

wherein said molecule either evokes one or more inorganic ion receptor activities, or blocks one or more inorganic ion receptor activities caused by an extracellular inorganic ion.

13. The method of claim 12 wherein said patient has a disease or disorder characterized by abnormal calcium homeostasis, said molecule is a calcimimetic and said inorganic ion receptor activity is a calcium receptor activity.

14. The method of claim 13 wherein each R is independently selected from the group consisting of H, CH_3, ethyl, and isopropyl.

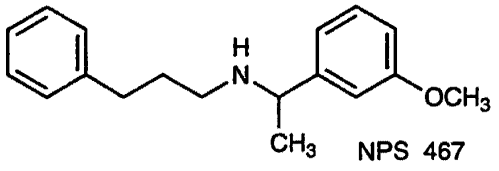
15. The method of claim 13 wherein each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, aliphatic ring and an attached or fused aromatic ring.

16. The method of claim 14 wherein each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, aliphatic ring and an attached or fused aromatic ring.

17. A method for treating a patient by modulating inorganic ion receptor activity comprising the step of administering to said patient a therapeutically effective amount of a molecule selected from the group consisting of compound 4L, compound 8J, compound 8U, compound 9R, compound 11X, compound 12U, compound 12V, compound 12Z, compound 14U, compound 16M, and compound 16P.

18. The method of claim 17 wherein said patient has a disease or disorder characterized by abnormal calcium homeostasis.

19. The method of any claim 12-17 wherein said patient has a disease selected from the group consisting of primary and secondary hyperparathyroidism, Paget's disease, hypercalcemia malignancy, osteoporosis and hypertension.



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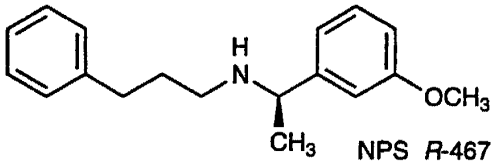
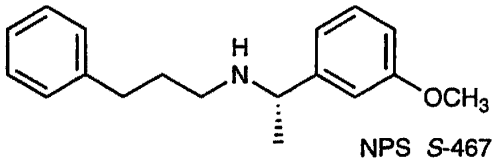
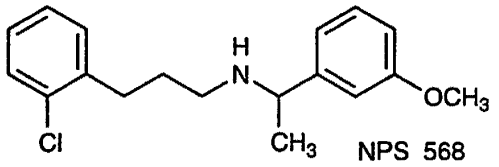


NPS *R*-467



NPS *S*-467



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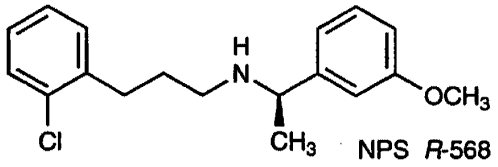
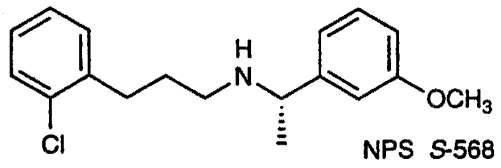


NPS *R*-568



NPS *S*-568

FIG. 1A.

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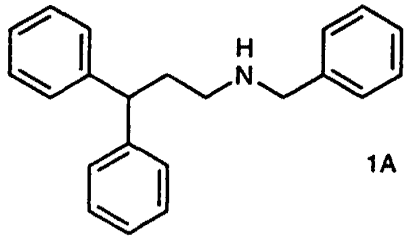

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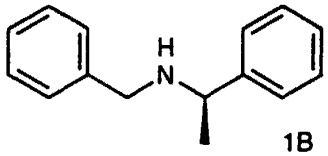
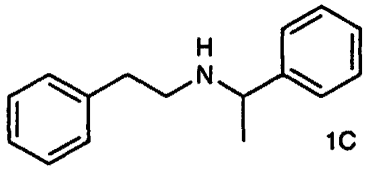


1B



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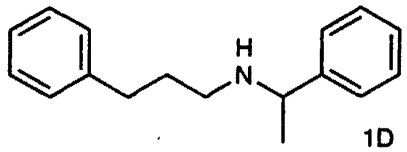

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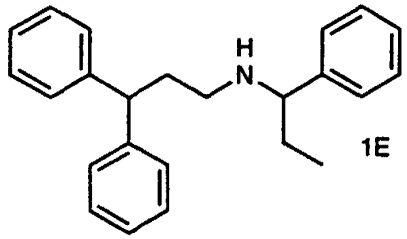
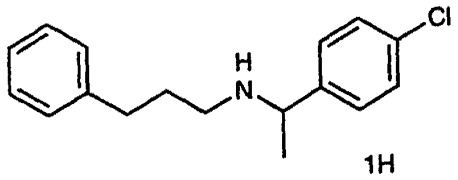
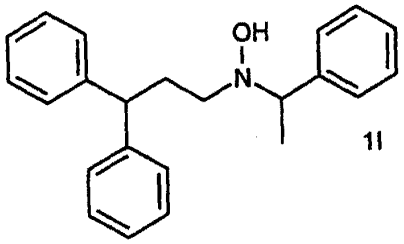


1E

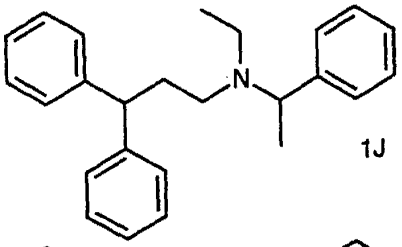
FIG. 1B.



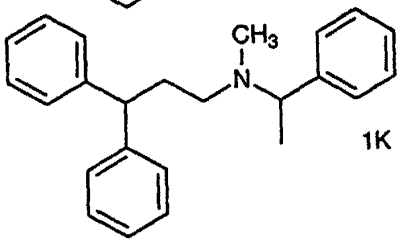
1H



1I



1J



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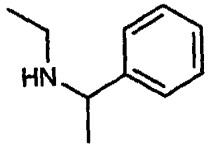
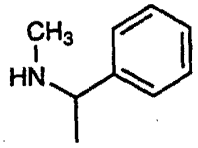
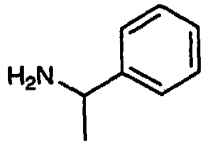


1L



1M



1N

FIG. 1C.

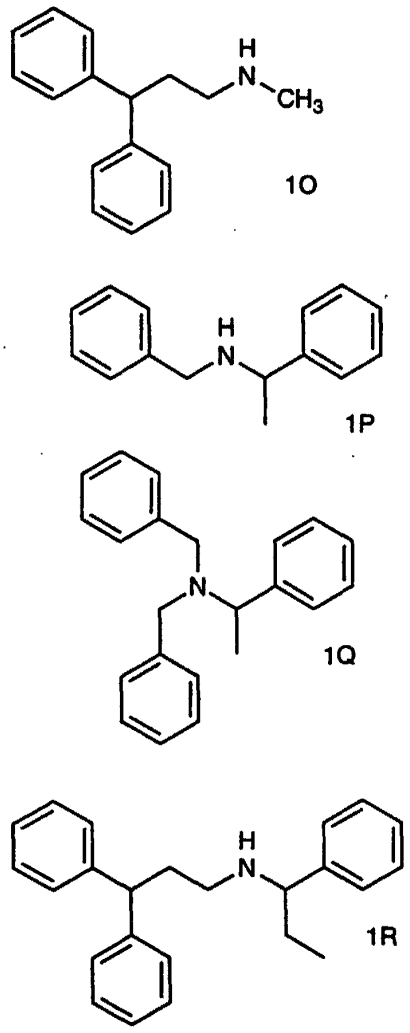


FIG. 1D.

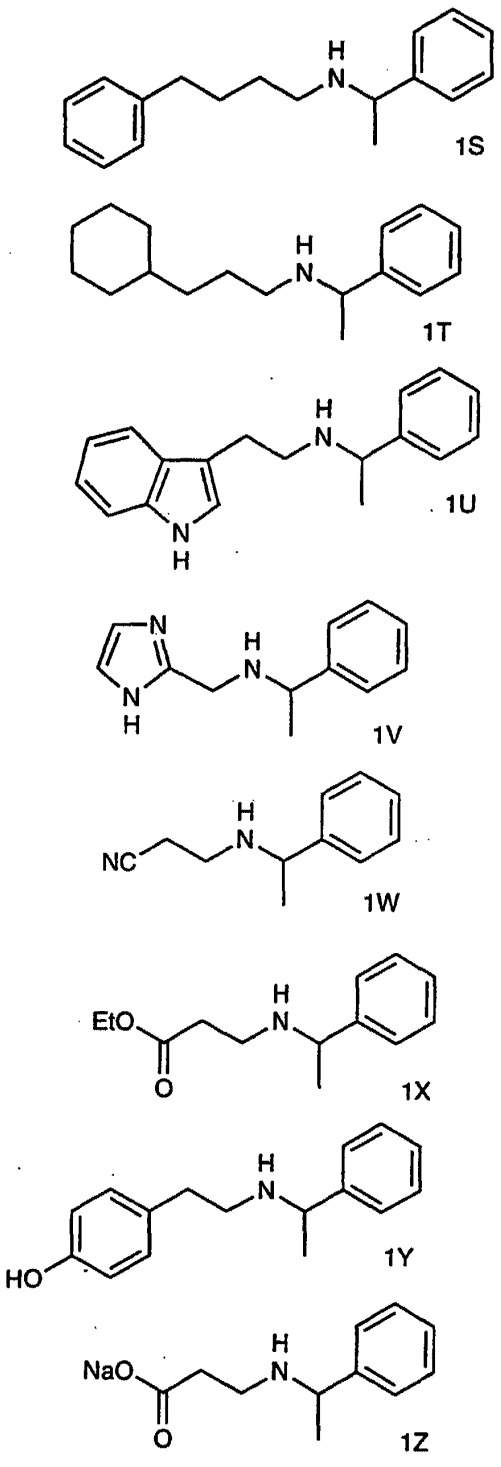


FIG. 1E.

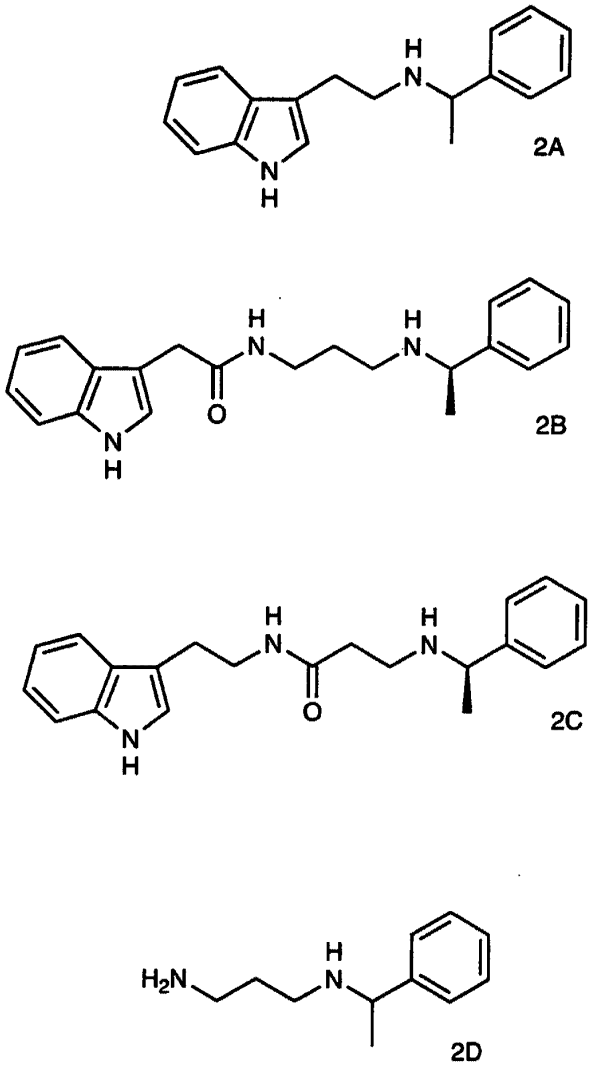


FIG. 2A.

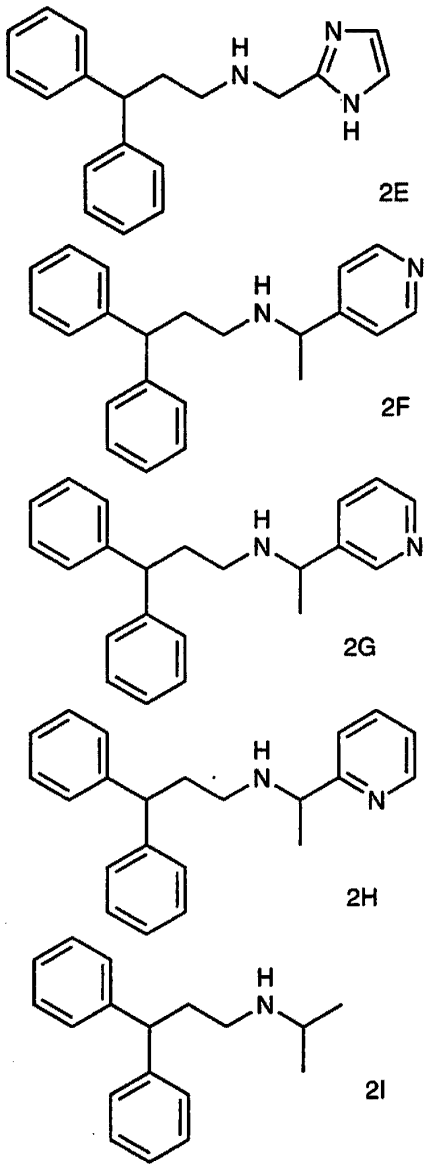


FIG.2B.

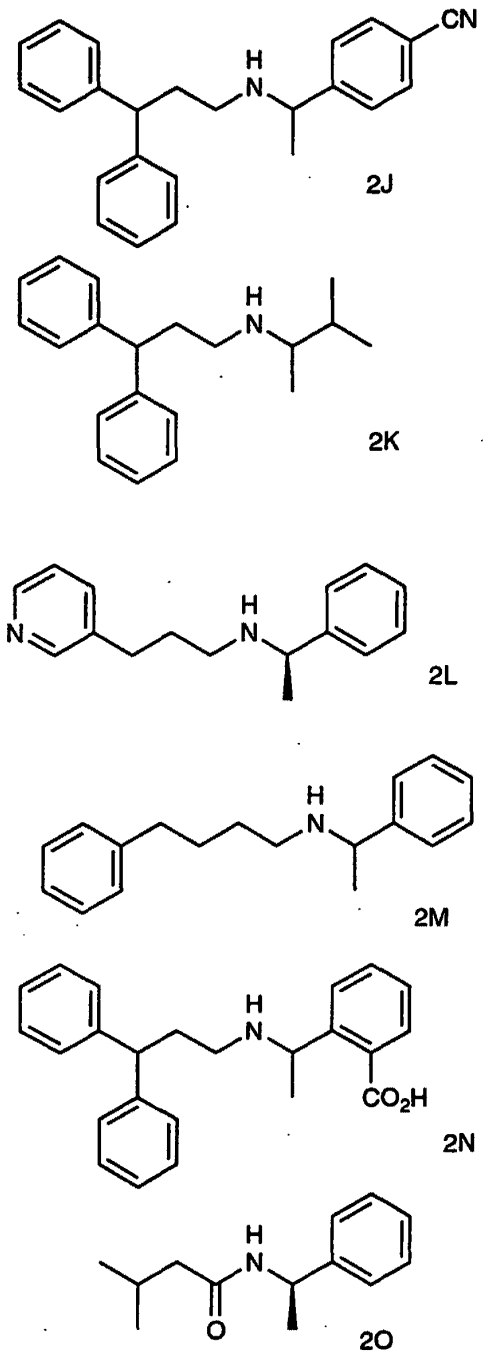


FIG. 2C.

RECTIFIED SHEET (RULE 91)
ISA / EP

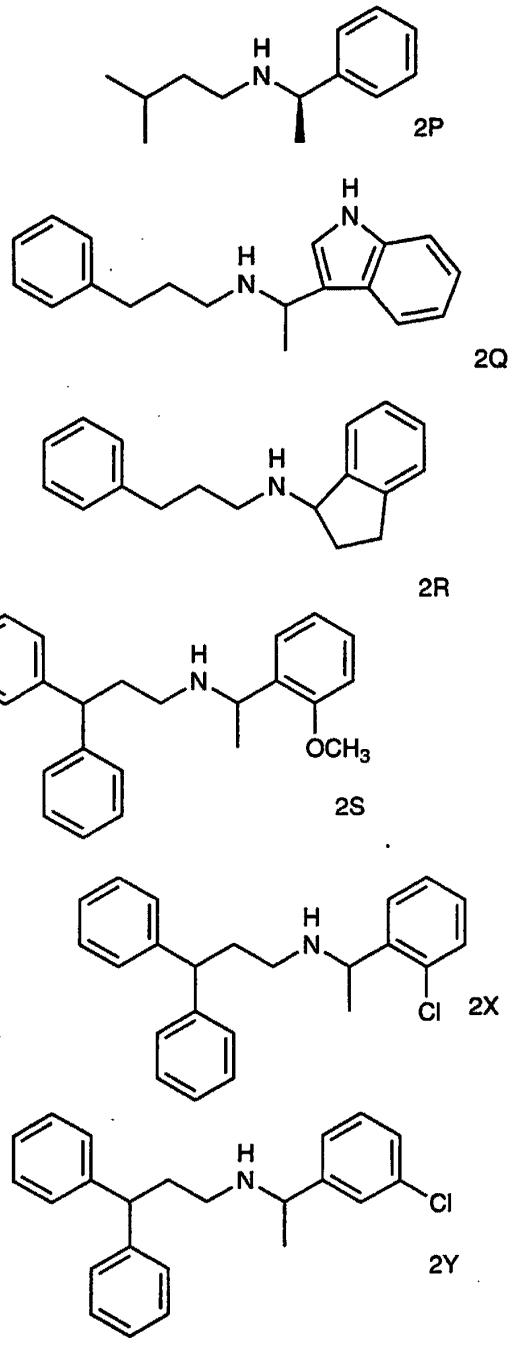


FIG. 2D.

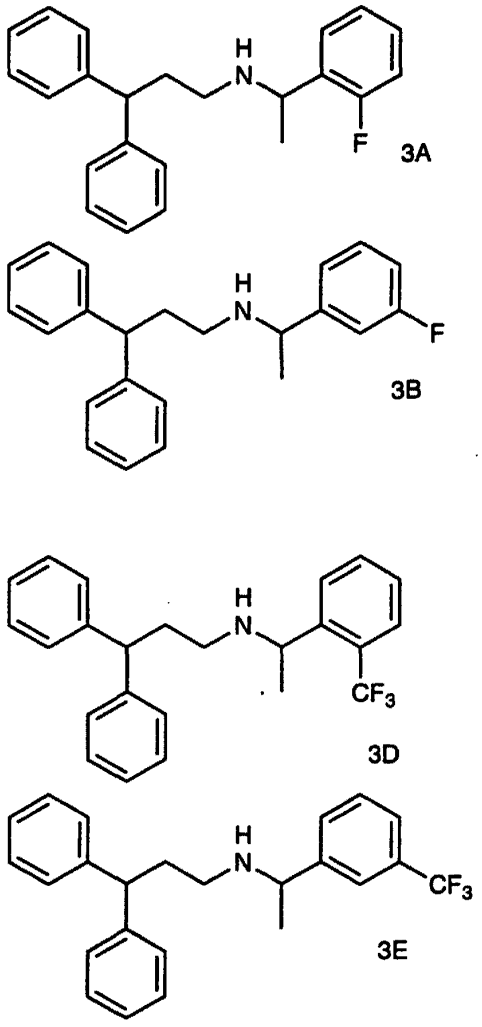
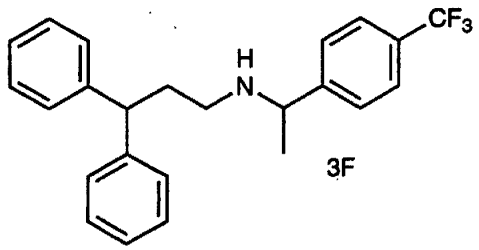
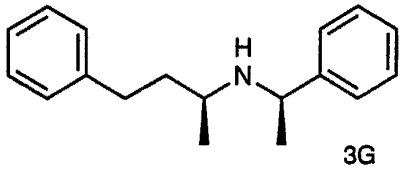


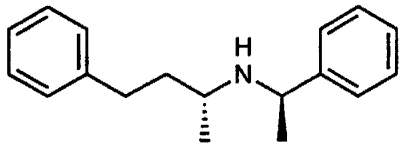
FIG. 3A.



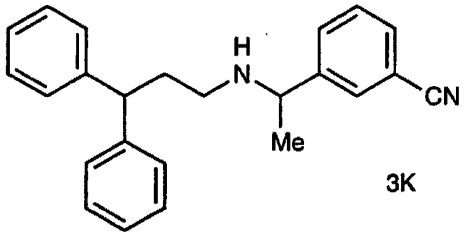
3F



3G

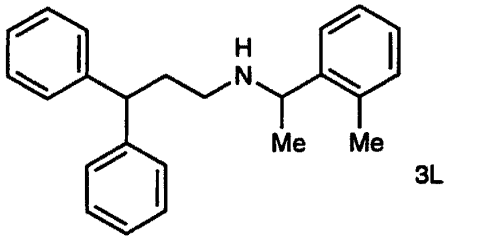


3H

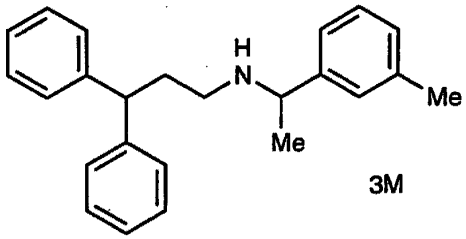


3K

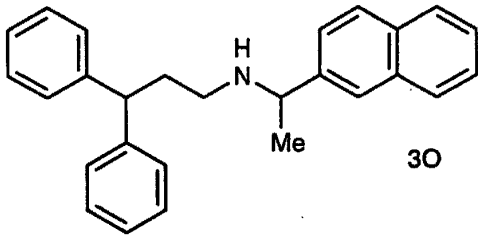
FIG. 3B.



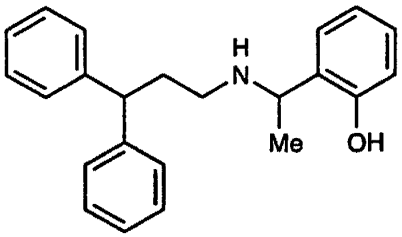
3L



3M



3O



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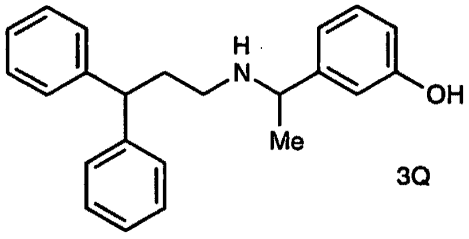


3Q

FIG. 3C.

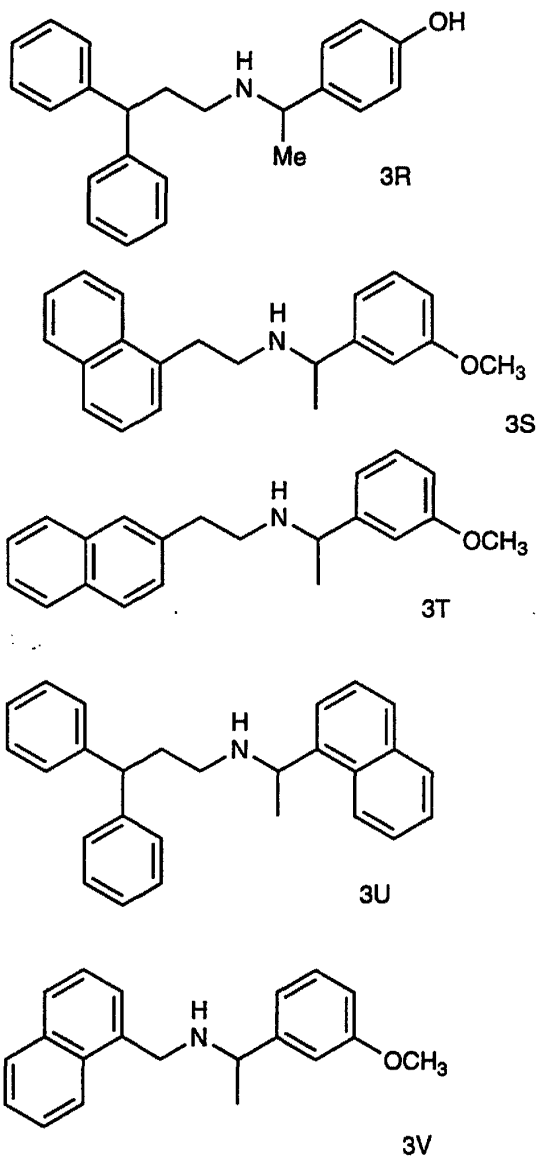


FIG. 3D.

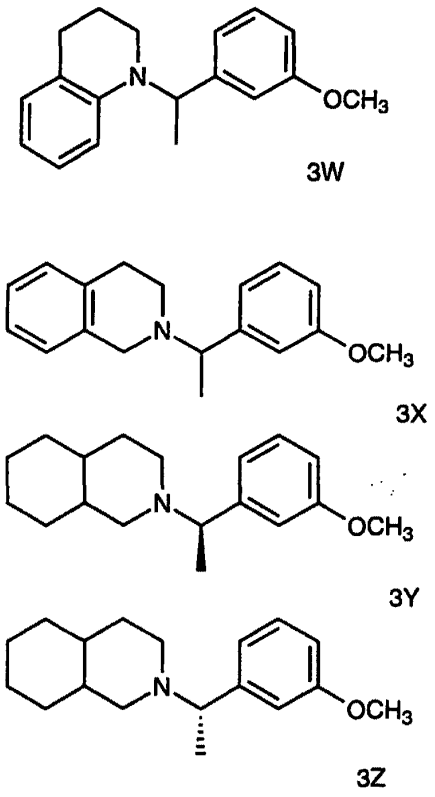
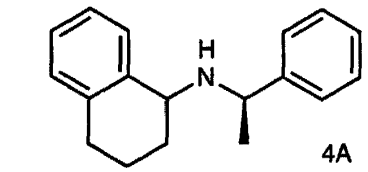
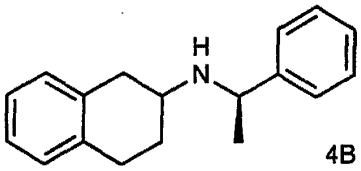


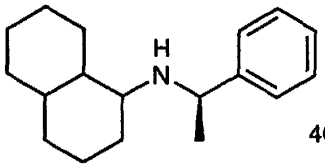
FIG. 3E.



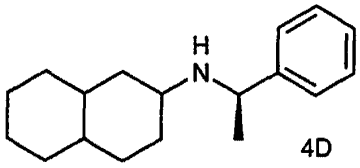
4A



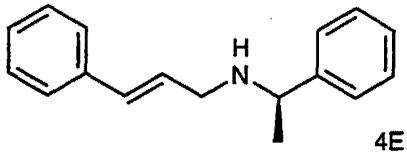
4B



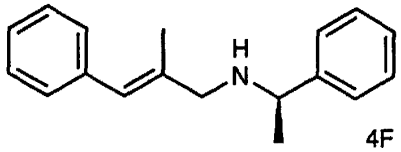
4C



4D



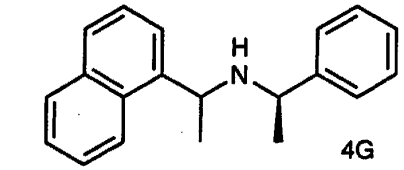
4E



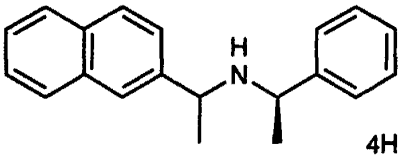
4F

FIG. 4A.

RECTIFIED SHEET (RULE 91)
ISA / EP



4G



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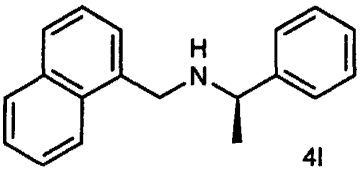

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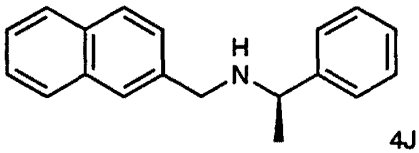
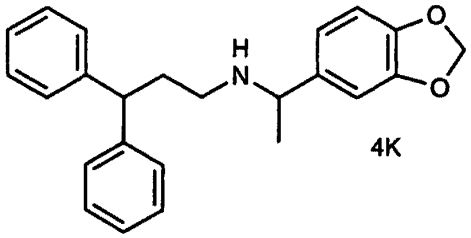
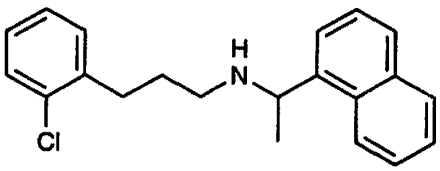


4J

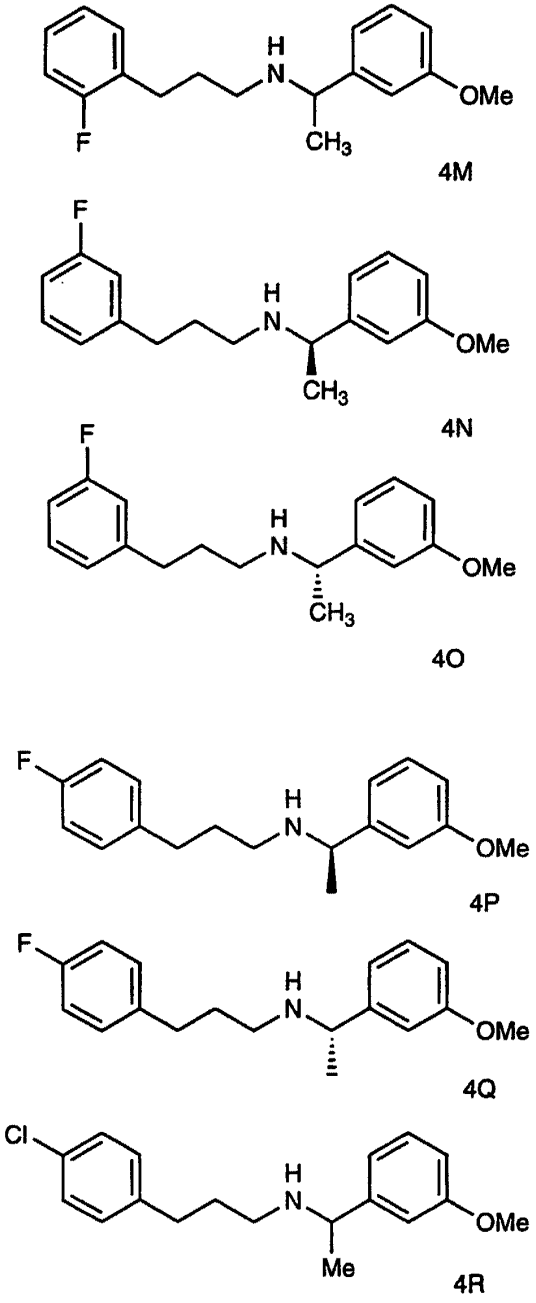


4K



4L

FIG. 4B.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4M
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CH3
4N
F
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OMe
CH3
4O
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4P
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OMe
4Q
Cl
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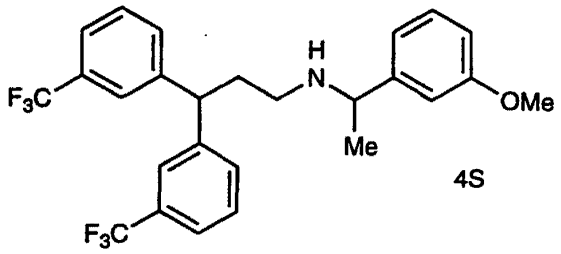
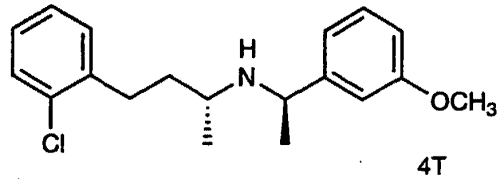
OMe
Me
4R

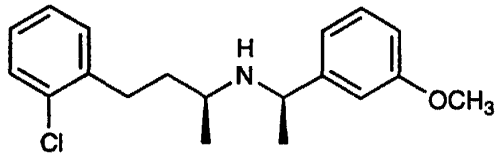
FIG. 4C.



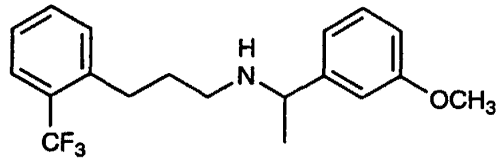
4S



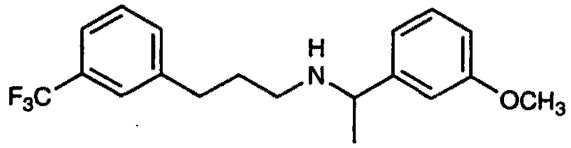
4T



4U



4V



4W

FIG. 4D.

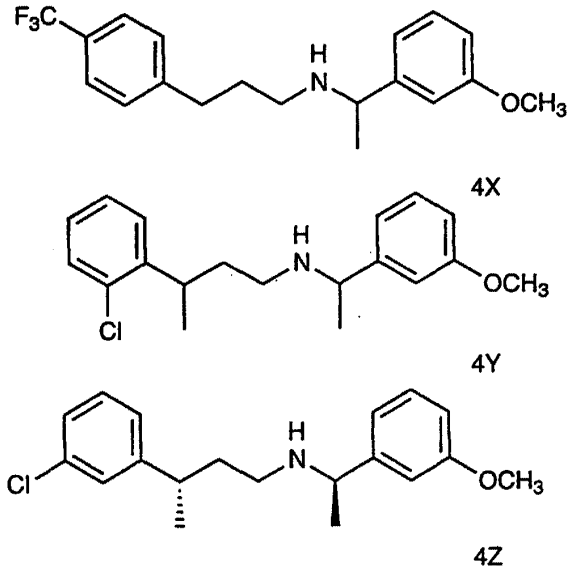


FIG. 4E.

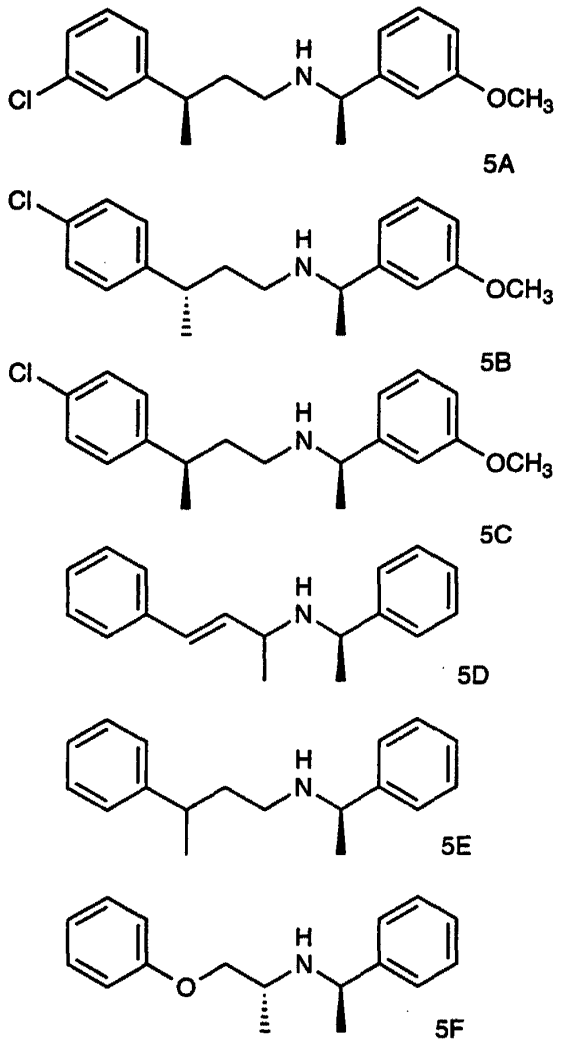


FIG. 5A.

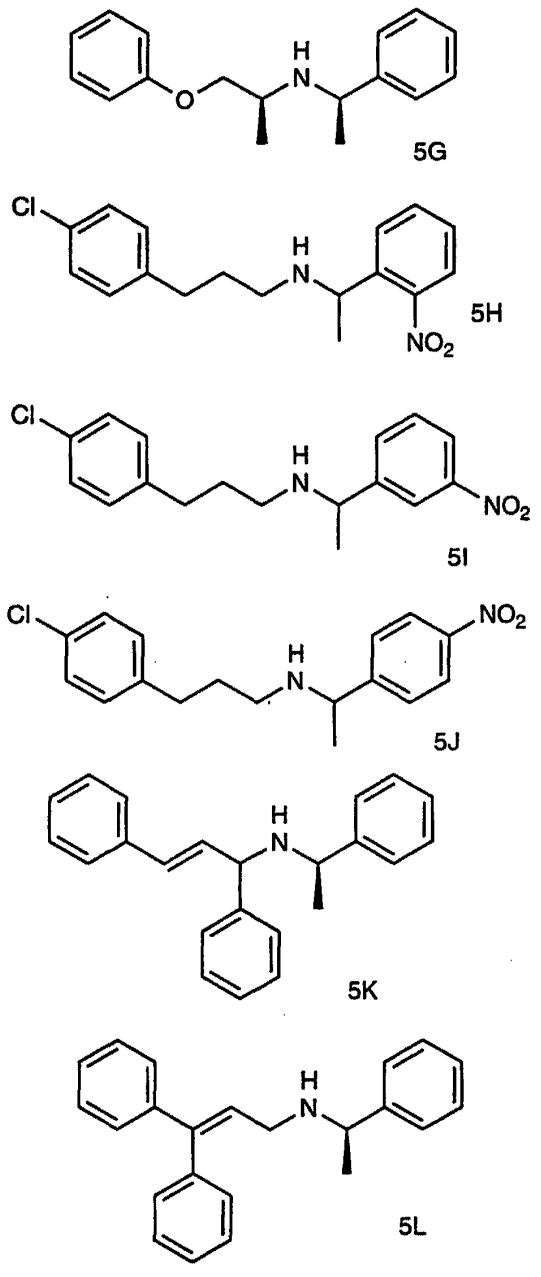


FIG. 5B.

RECTIFIED SHEET (RULE 91)
ISA / EP

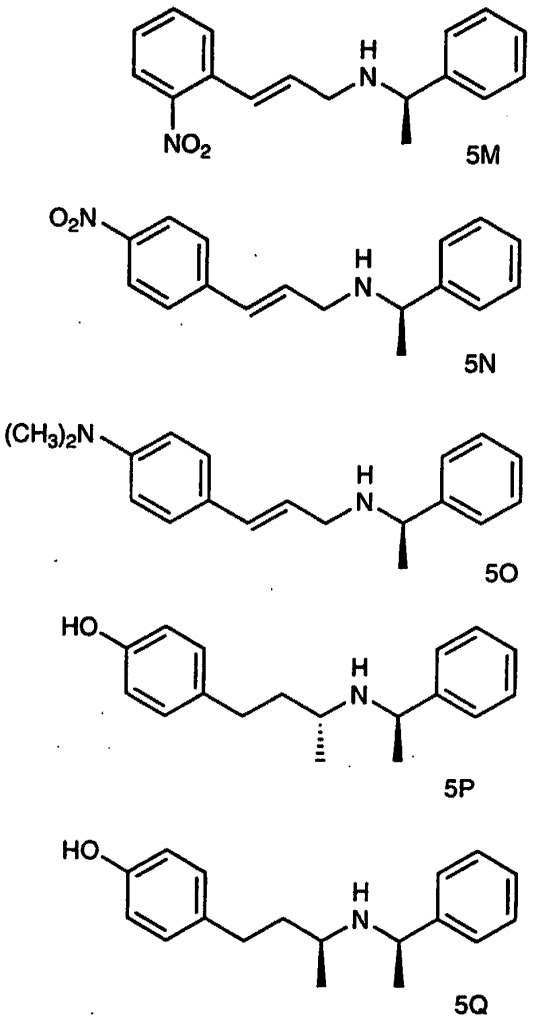
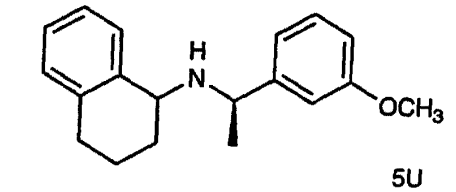
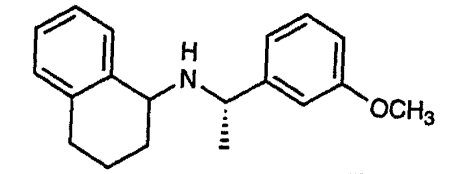


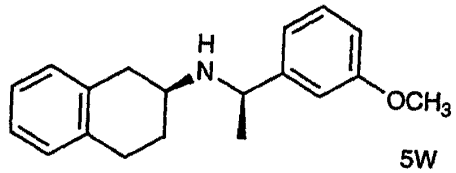
FIG. 5C.



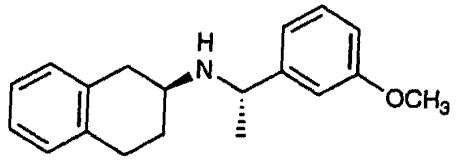
5U



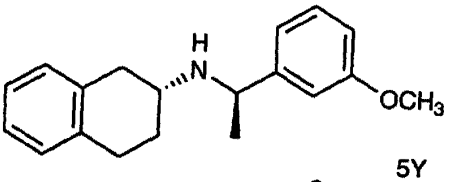
5V



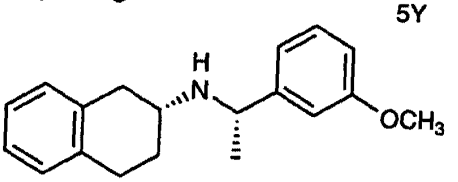
5W



5X



5Y



5Z

FIG. 5D.

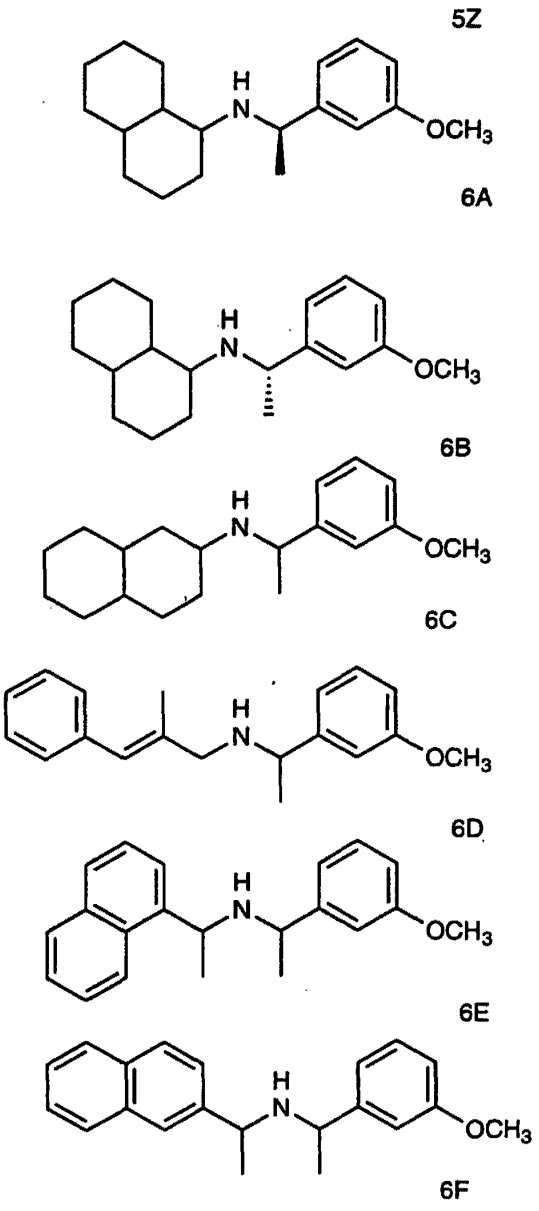
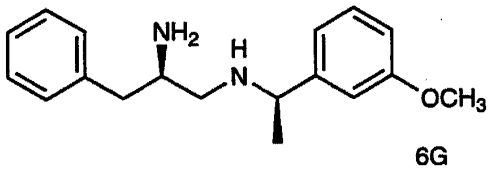
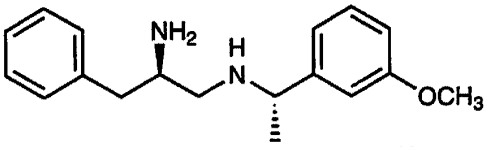


FIG. 6A.

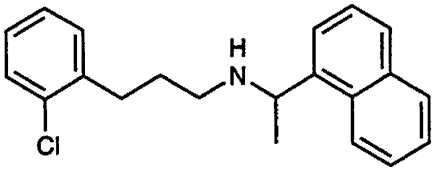
RECTIFIED SHEET (RULE 91)
ISA / EP



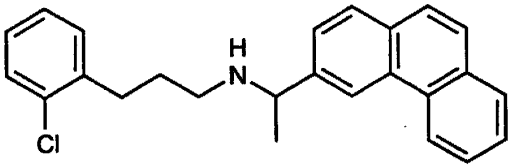
6G



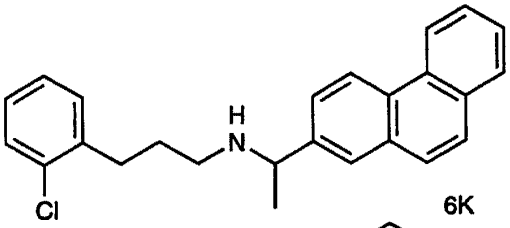
6H



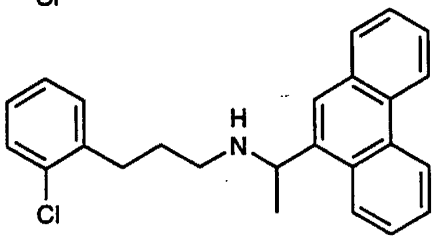
6I



6J



6K



6L

FIG. 6B.

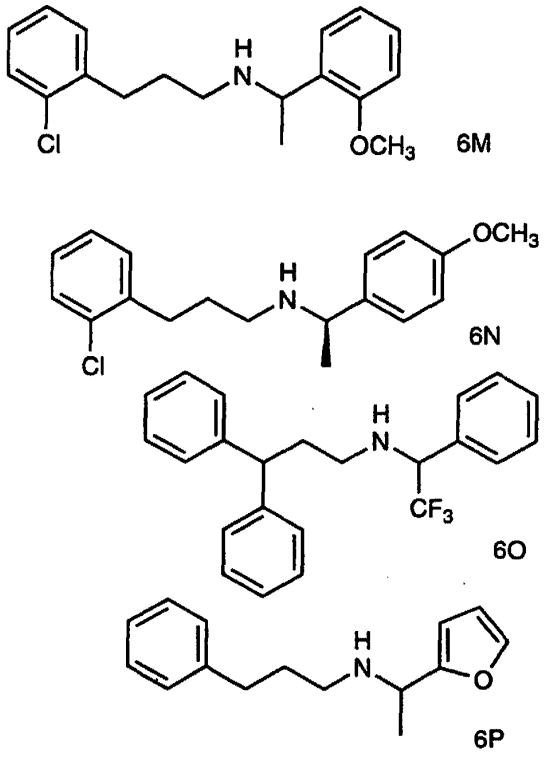


FIG. 6C.

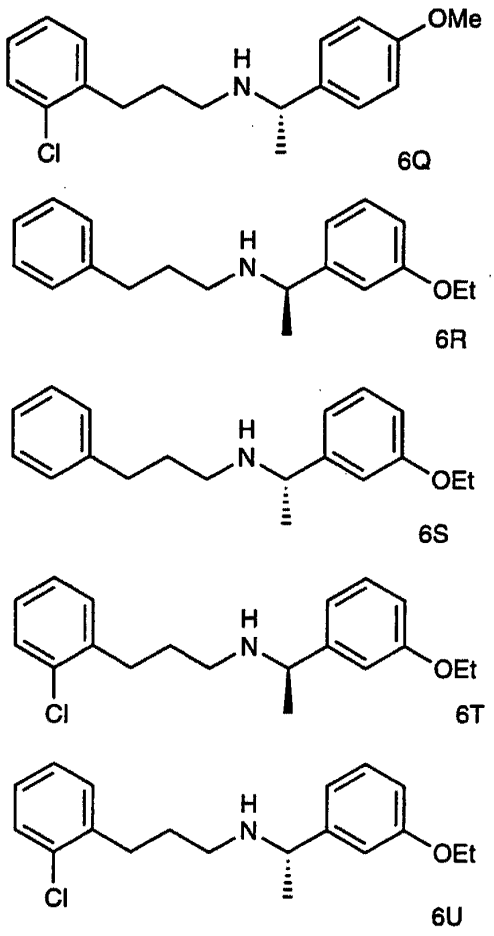


FIG. 6D.

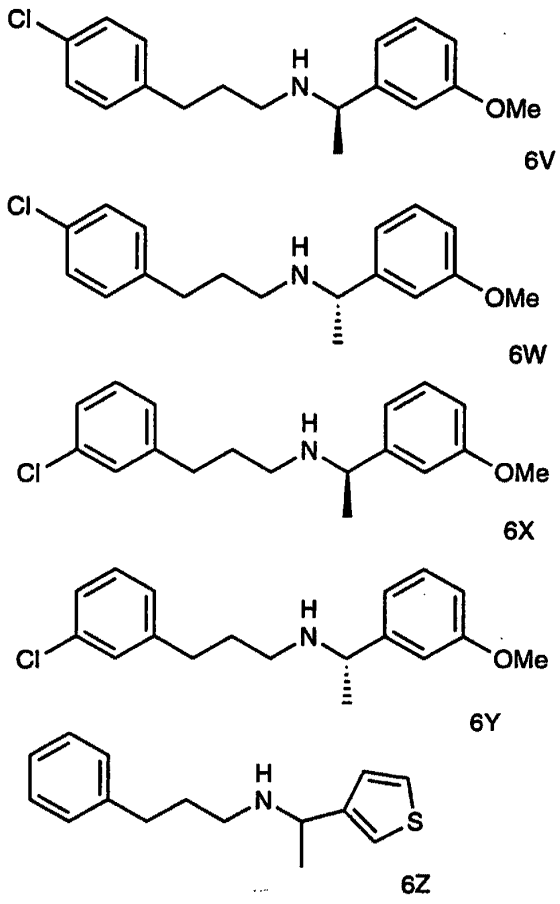


FIG. 6E.

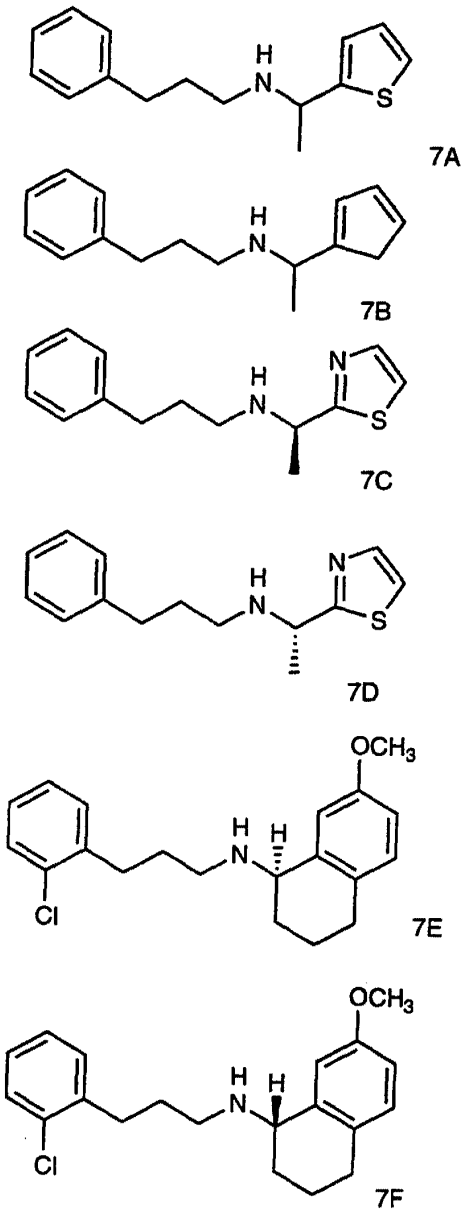


FIG. 7A.

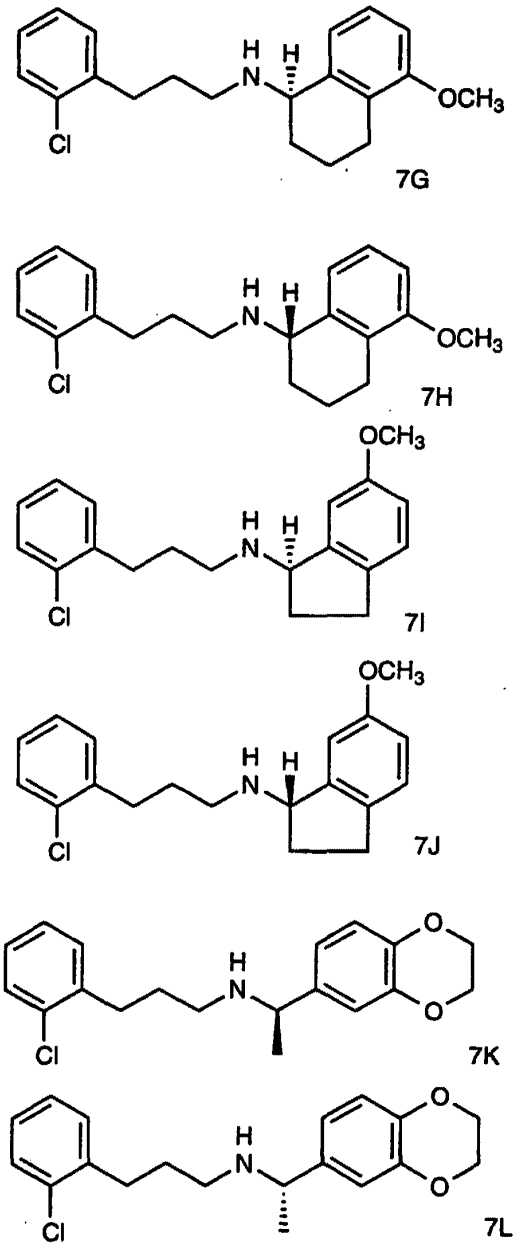


FIG. 7B.

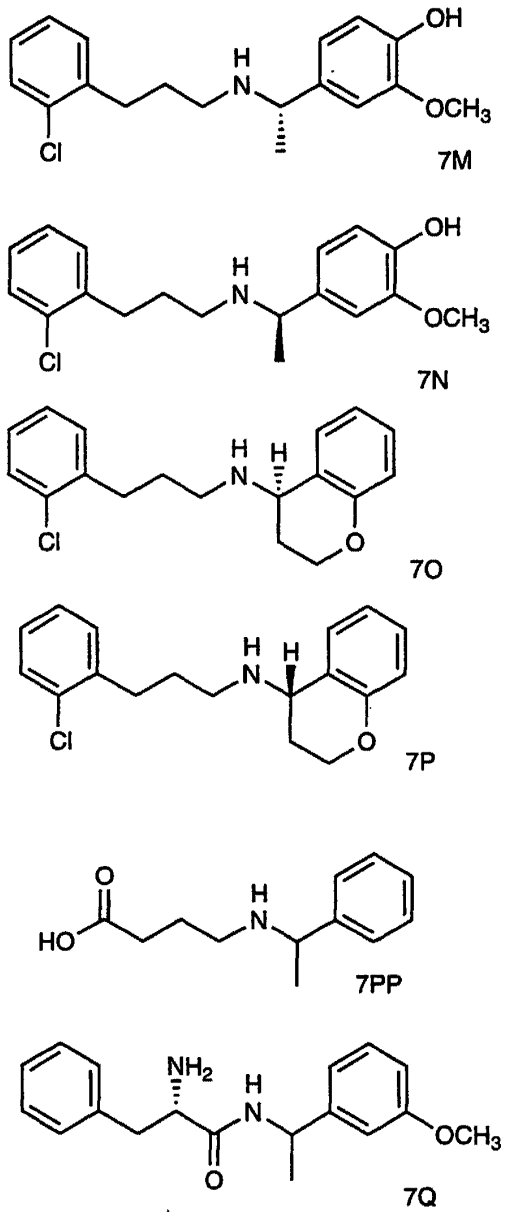

OH
H
N
OCH3
Cl
7M
OH
H
N
OCH3
Cl
7N
H
H
N
Cl
O
7O
H
H
N
Cl
O
7P
O
HO
H
N
7PP
NH2
H
N
OCH3
O
7Q

FIG. 7C.

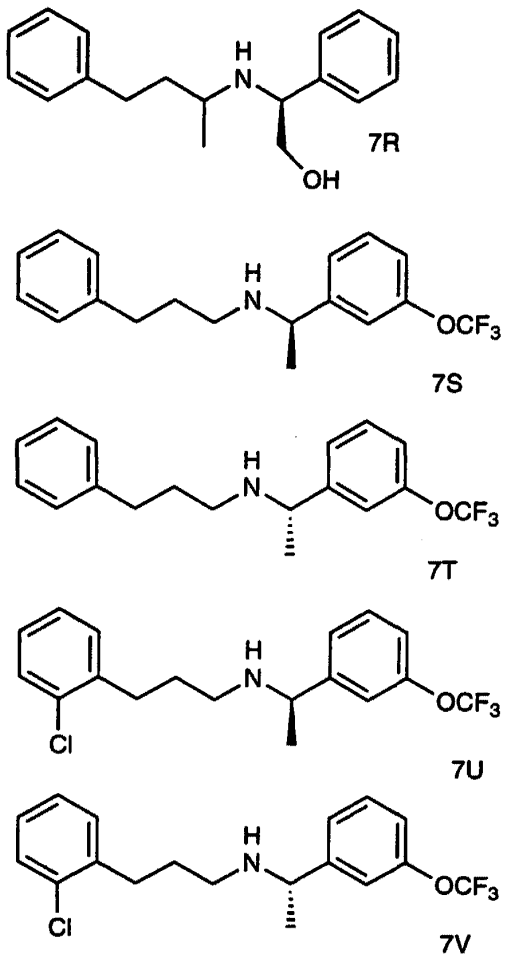


FIG. 7D.

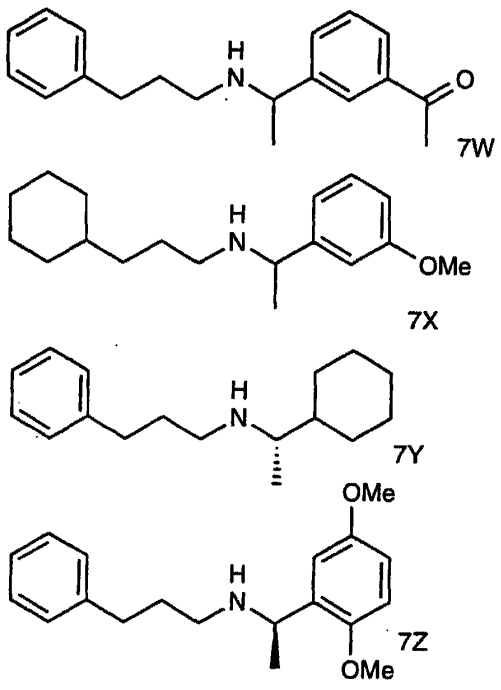


FIG. 7E.

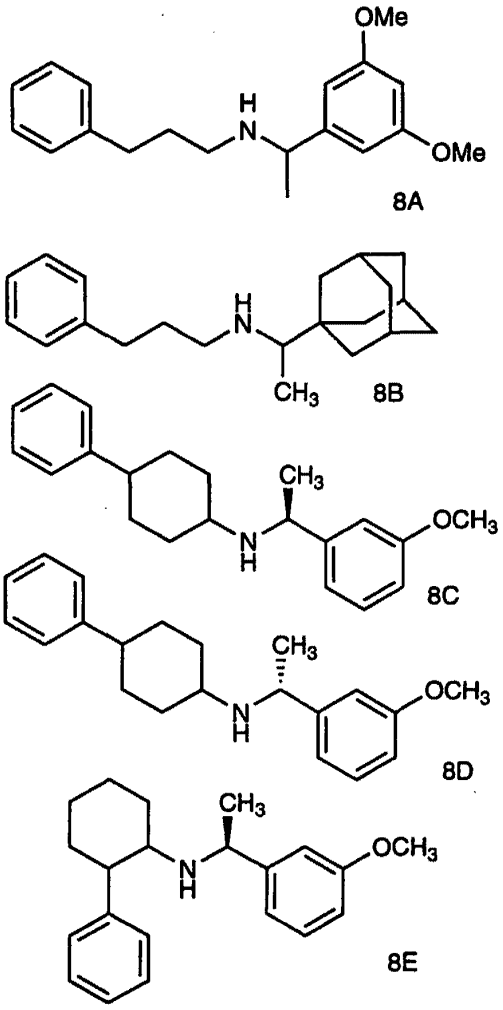

OMe
H
N
OMe
8A
H
N
CH3
8B
CH3
N
H
OCH3
8C
CH3
N
H
OCH3
8D
CH3
N
H
OCH3
8E

FIG. 8A.

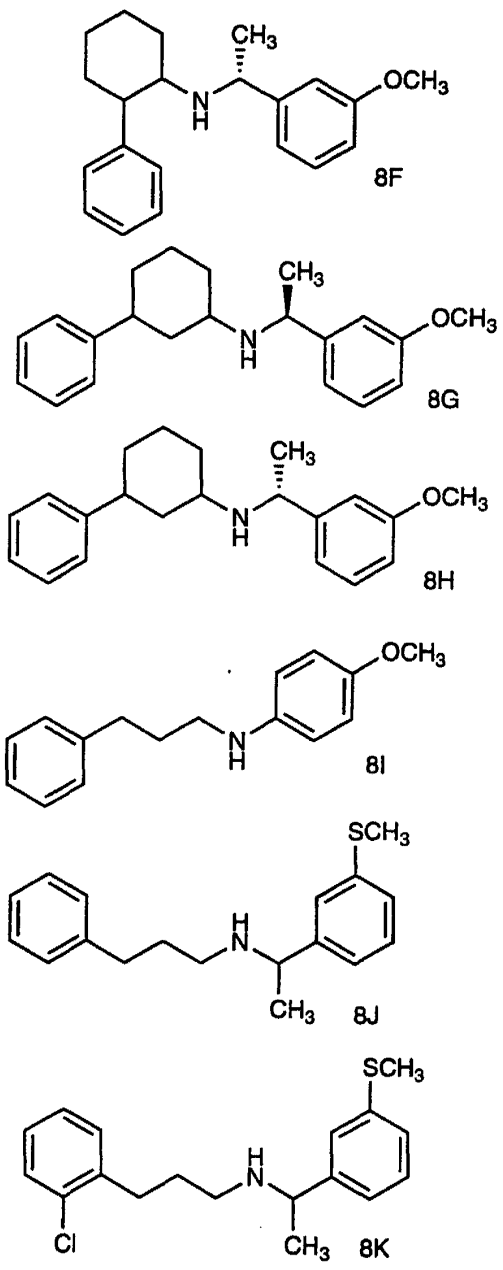


FIG. 8B.

RECTIFIED SHEET (RULE 91)
ISA / EP

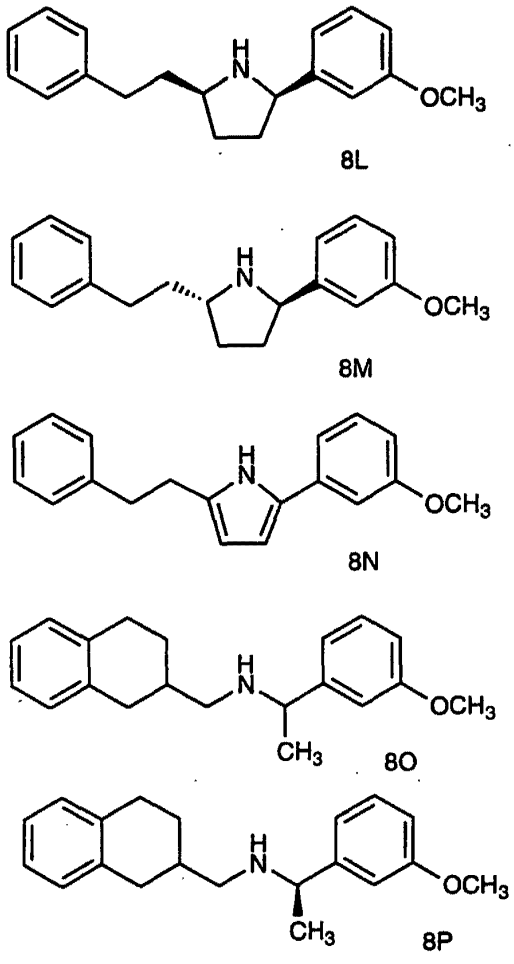


FIG. 8C.

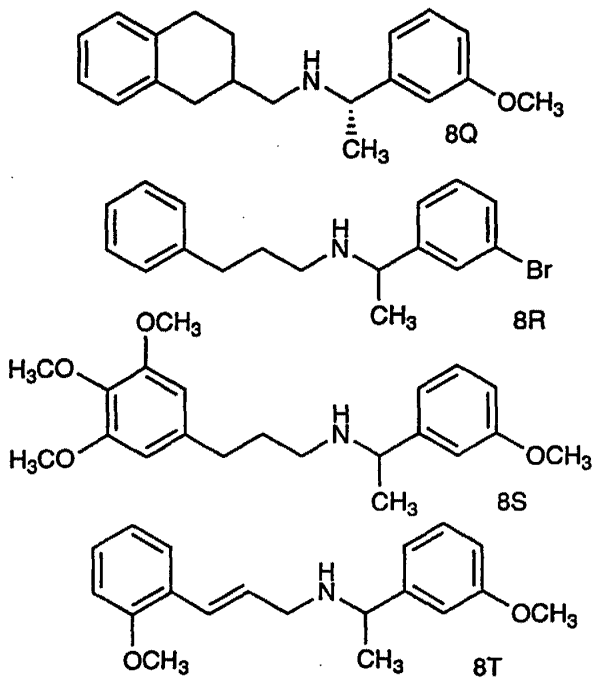

OCH3
CH3
8Q
Br
CH3
8R
OCH3
H3CO
H3CO
OCH3
CH3
8S
OCH3
OCH3
CH3
8T

FIG. 8D.

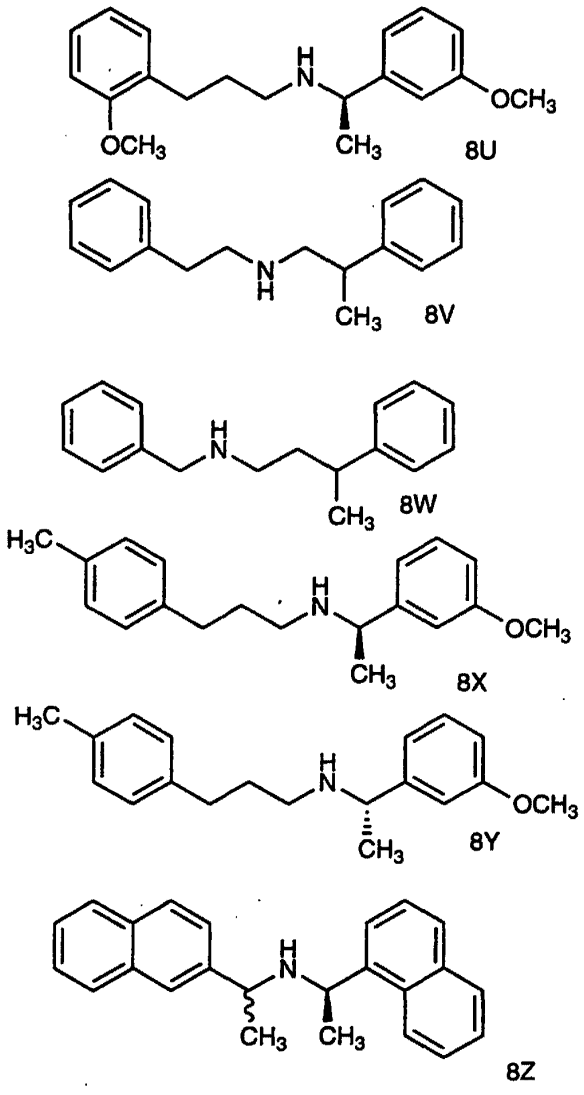

OCH3
OCH3
CH3
8U
CH3
8V
CH3
8W
H3C
OCH3
CH3
8X
H3C
OCH3
CH3
8Y
CH3
CH3
8Z

FIG. 8E.

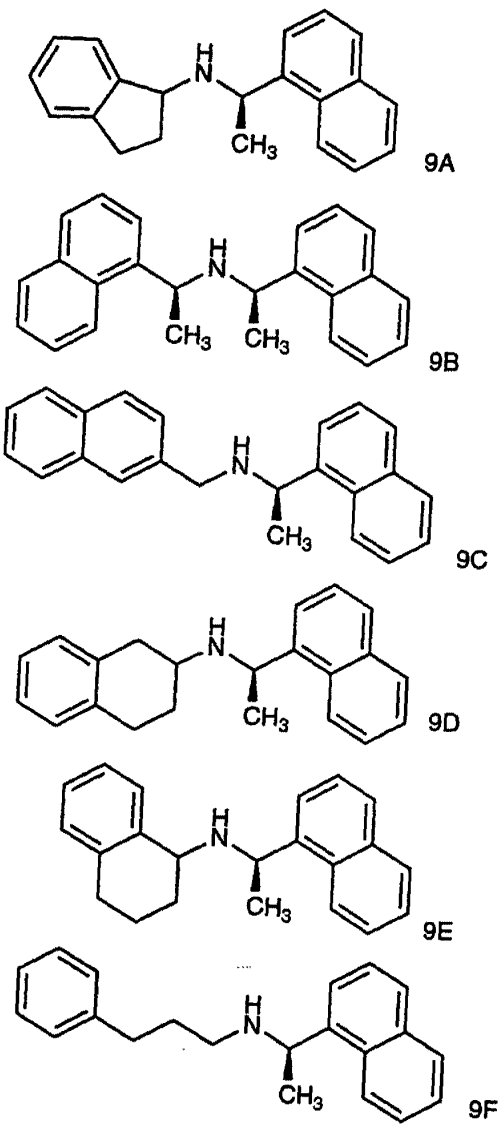


FIG. 9A.

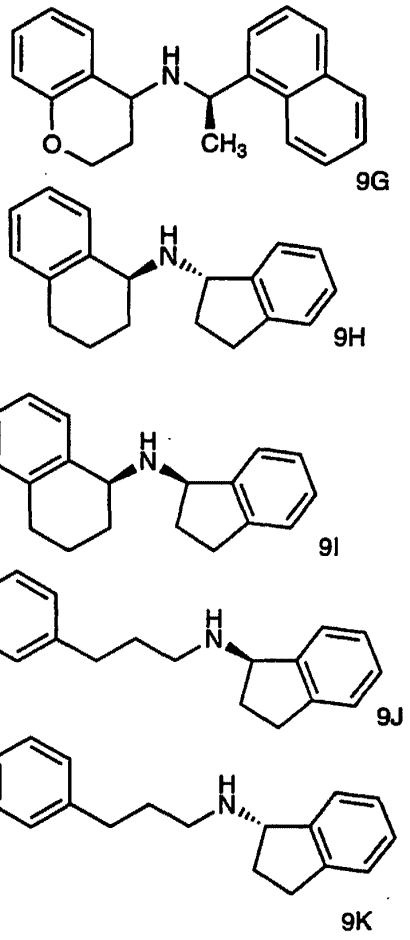


FIG. 9B.

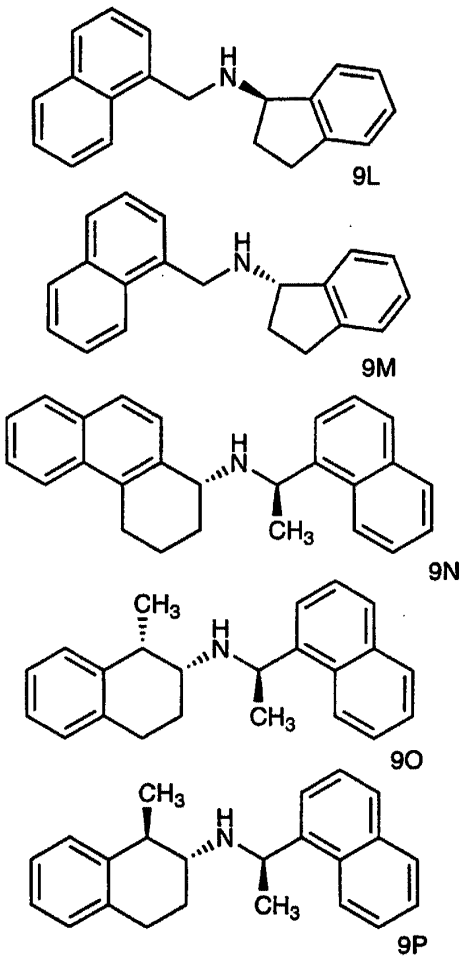


FIG. 9C.

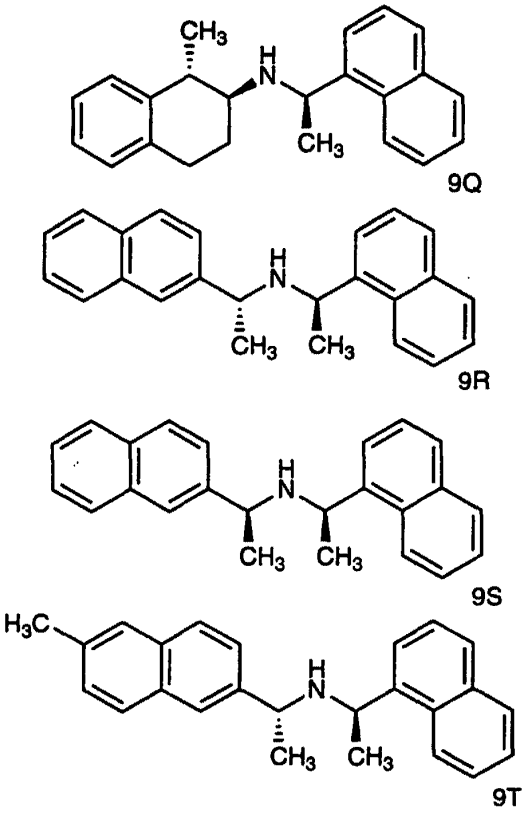


FIG. 9D.

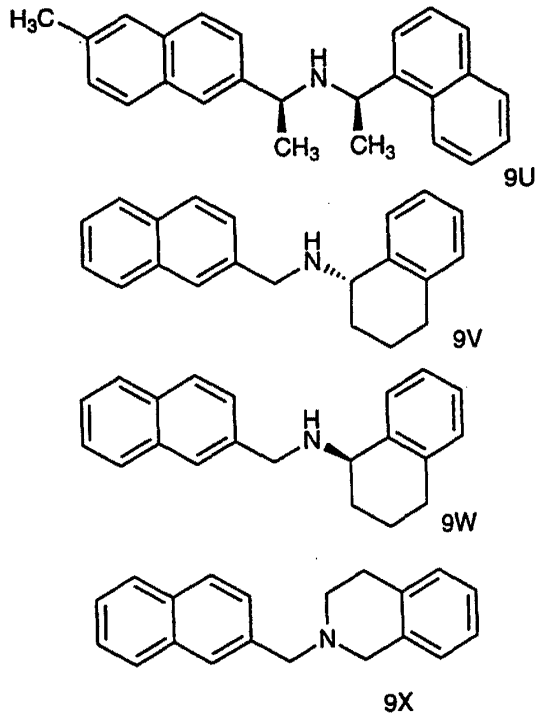
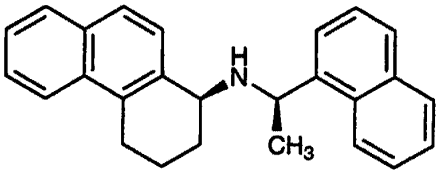
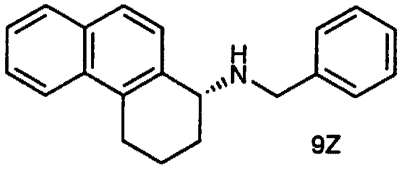


FIG. 9E.

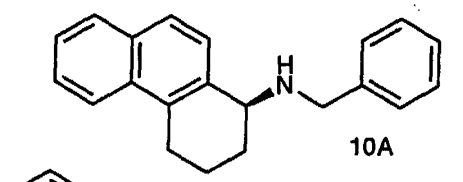


9Y

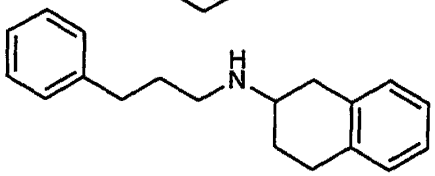


9Z

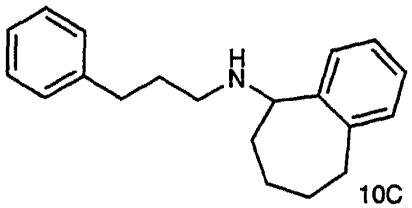
FIG. 9F.



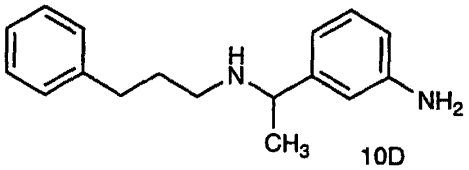
10A



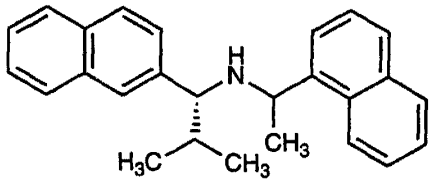
10B



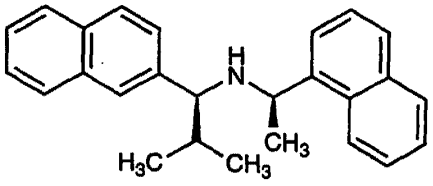
10C



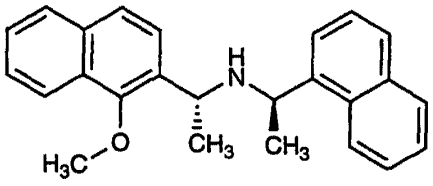
10D



10E

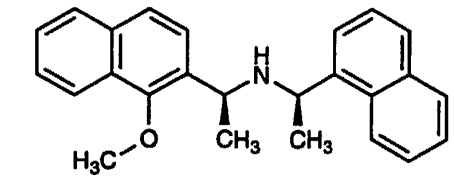


10F

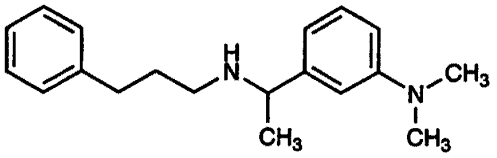


10G

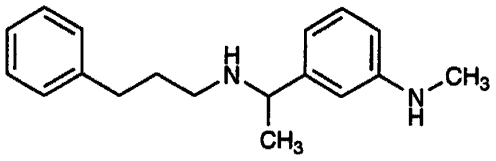
FIG. 10A.



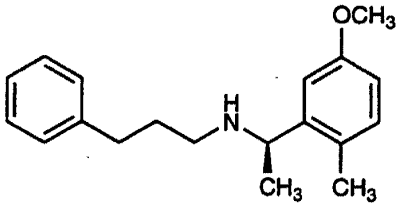
10H



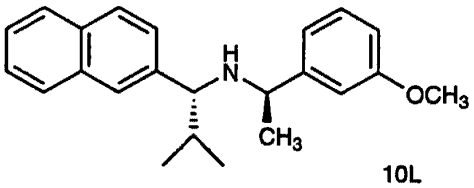
10I



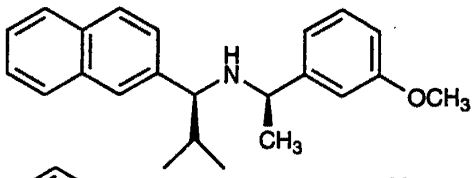
10J



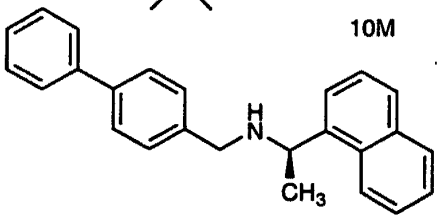
10K



10L

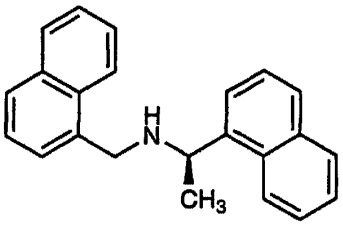


10M

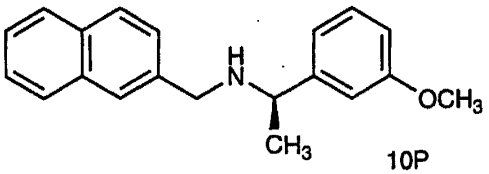


10N

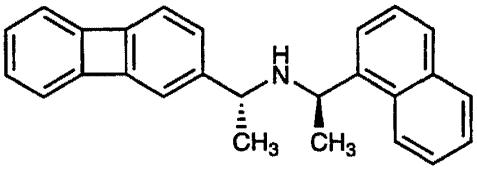
FIG. 10B.



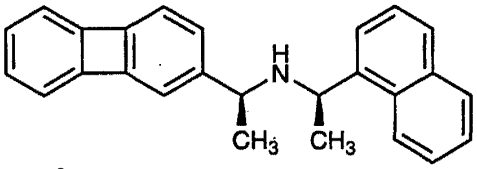
10O



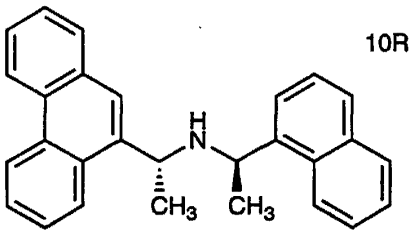
10P



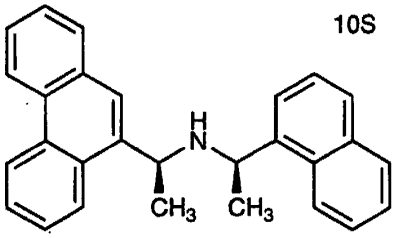
10Q



10R

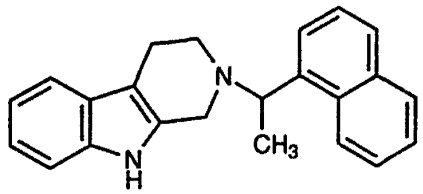


10S

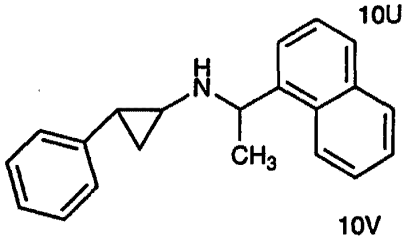


10T

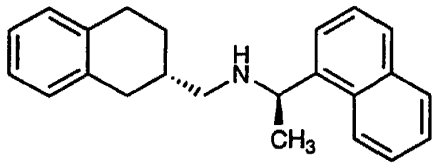
FIG.10C.



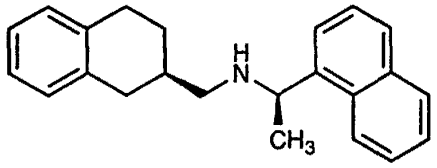
10U



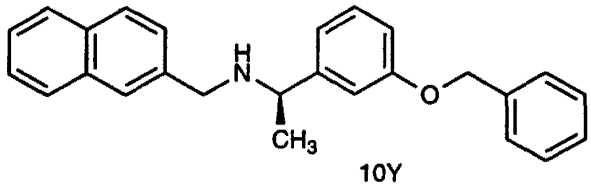
10V



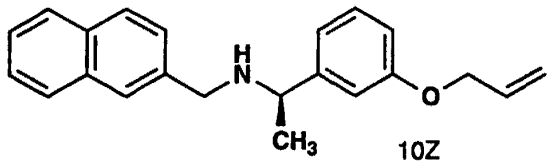
10W



10X



10Y



10Z

FIG. 10D.

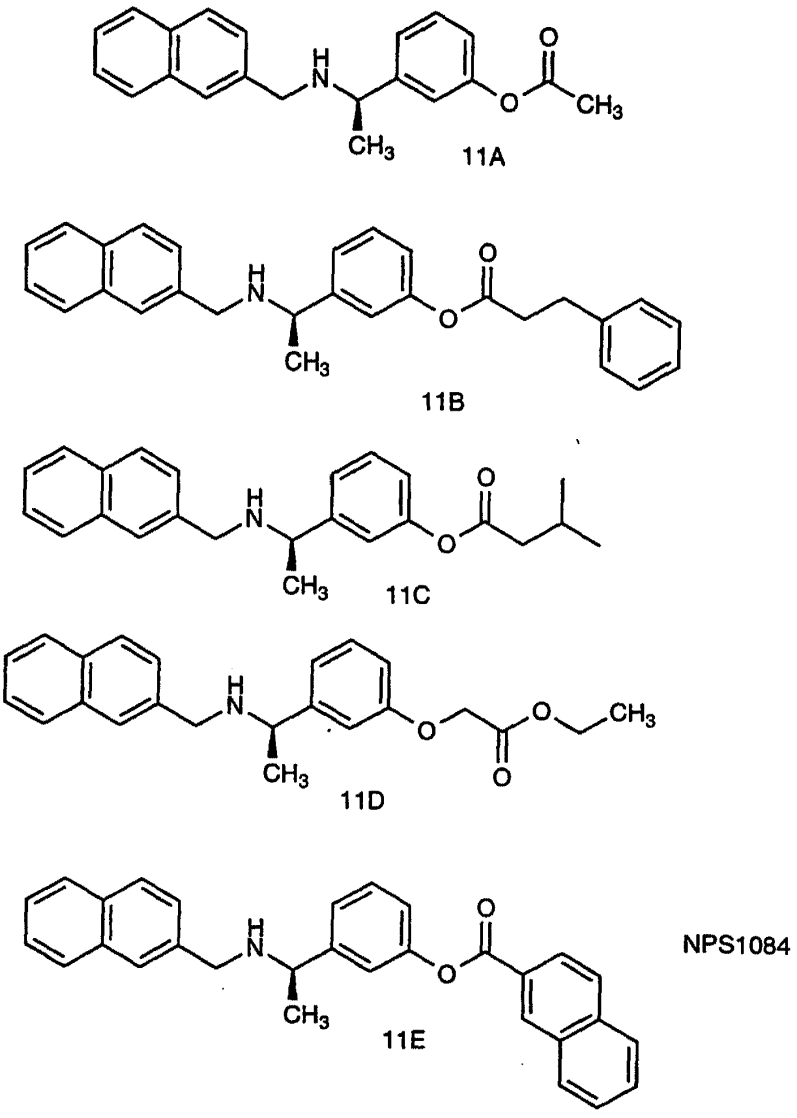


FIG. 11A.

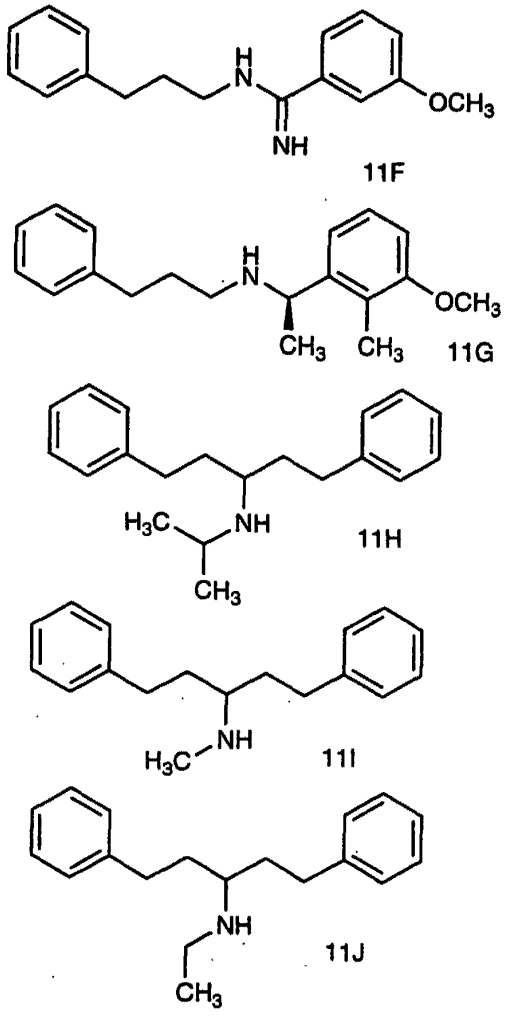
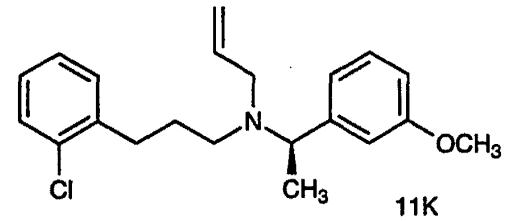
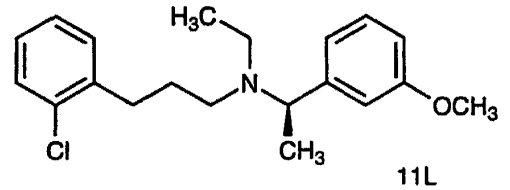

H
N
NH
OCH3
11F
H
N
OCH3
CH3
CH3
11G
H3C
NH
CH3
11H
NH
H3C
11I
NH
CH3
11J

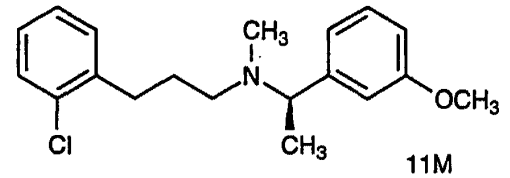
FIG. 11B.



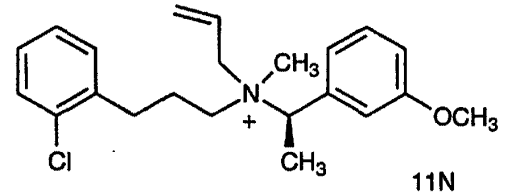
11K



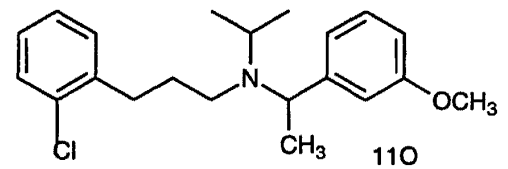
11L



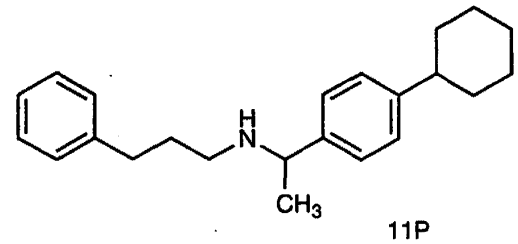
11M



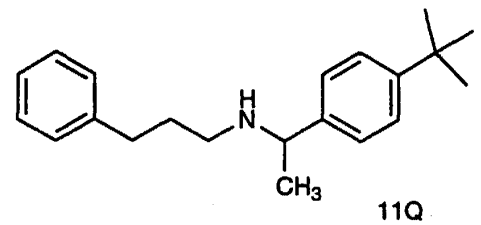
11N



11O



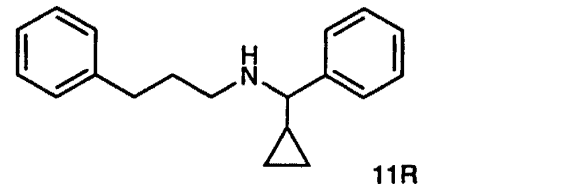
11P



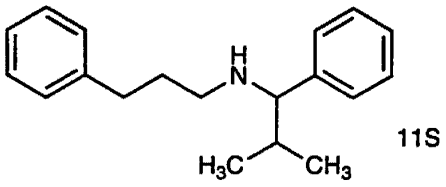
11Q

FIG. 11C.

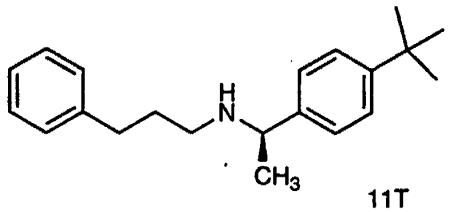
RECTIFIED SHEET (RULE 91)
ISA / EP



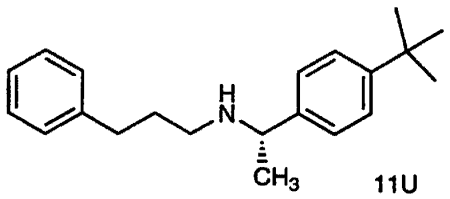
11R



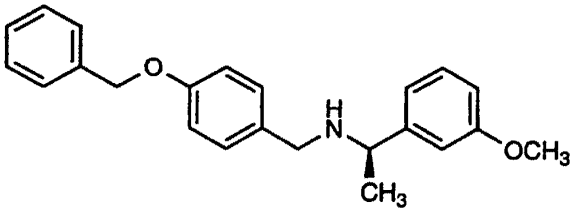
11S



11T

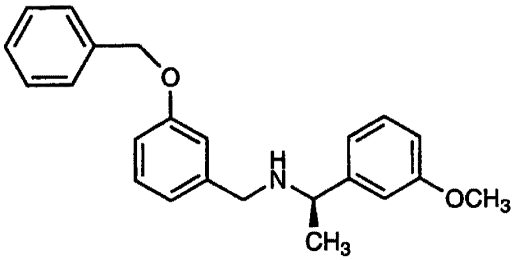


11U

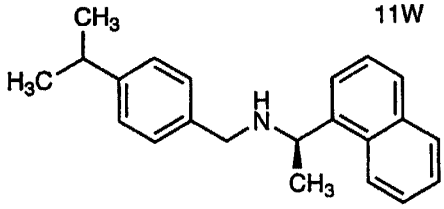


11V

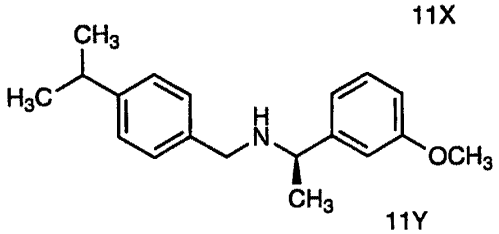
FIG. 11D.



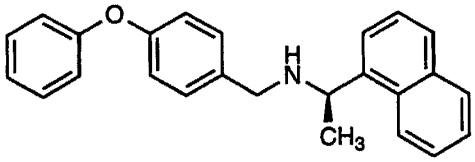
11W



11X

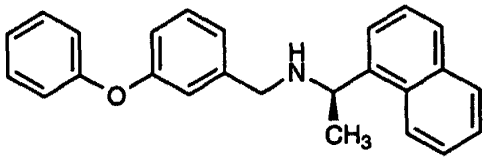


11Y

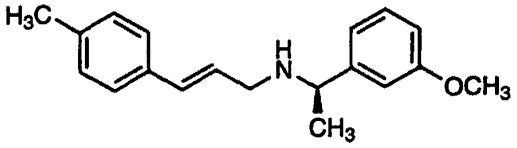


11Z

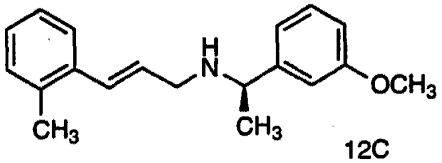
FIG. 11E.



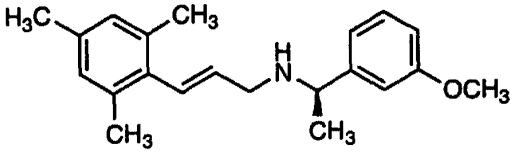
12A



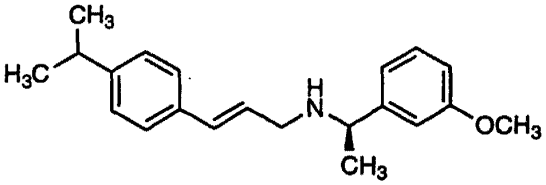
12B



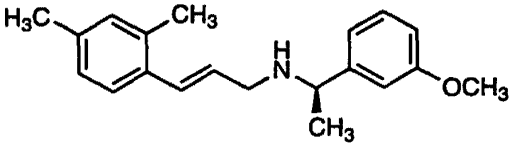
12C



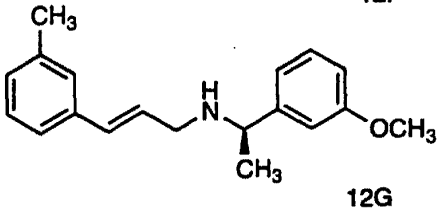
12D



12E

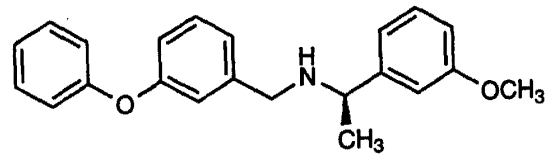


12F

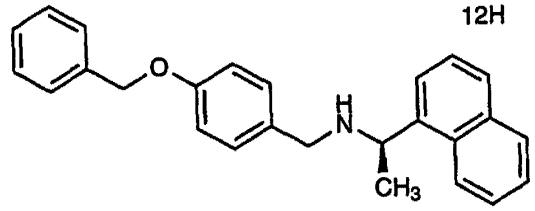


12G

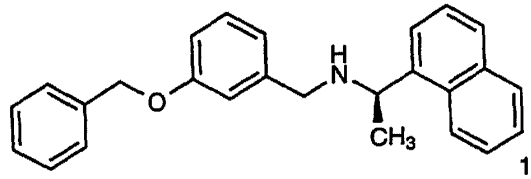
FIG. 12A.



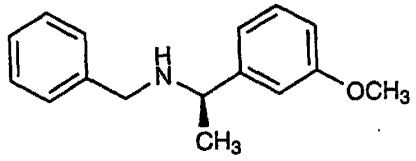
12H



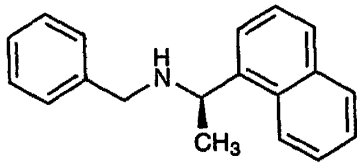
12I



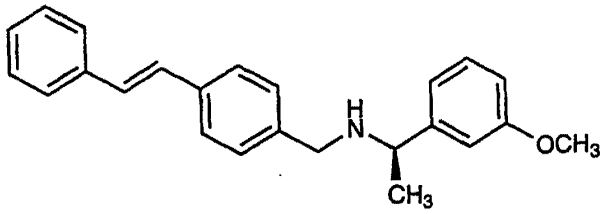
12J



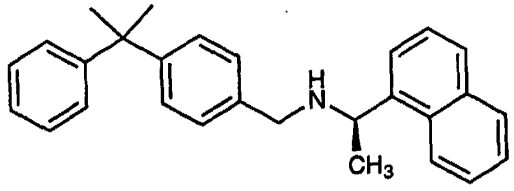
12K



12L

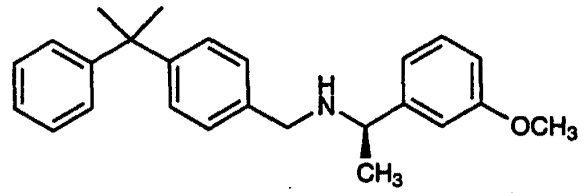


12M

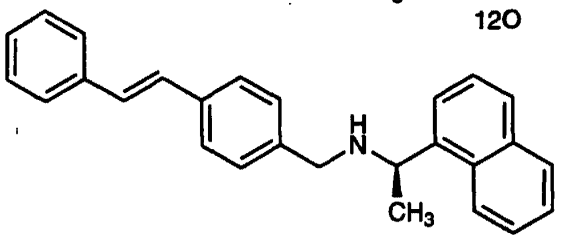


12N

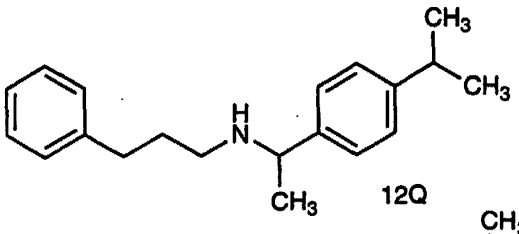
FIG. 12B.



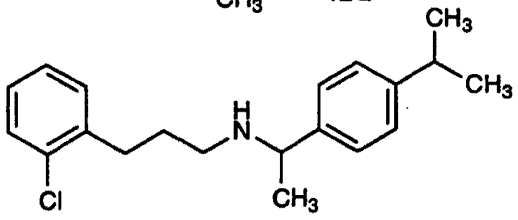
12O



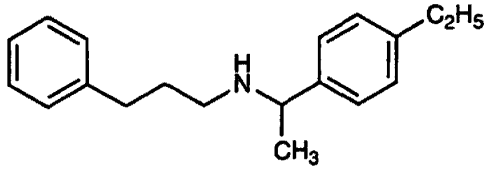
12P



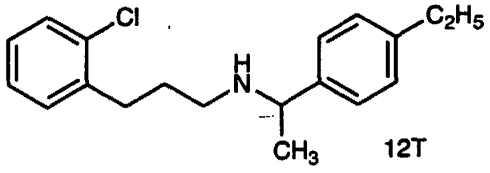
12Q



12R

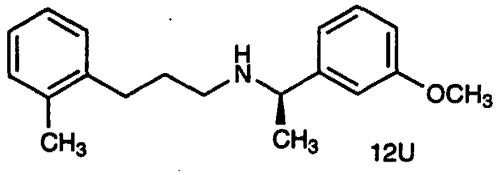


12S

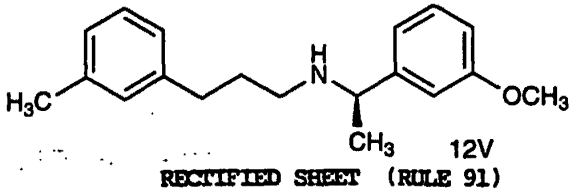


12T

FIG. 12C.

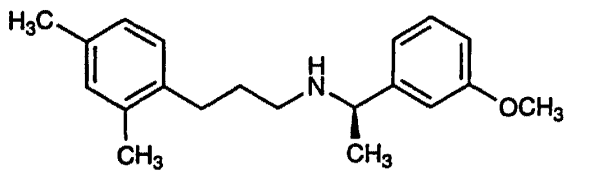


12U

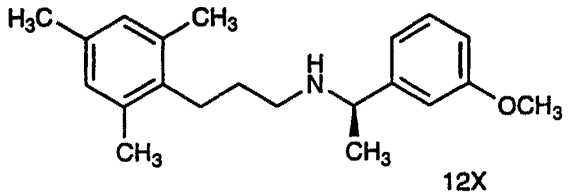


12V

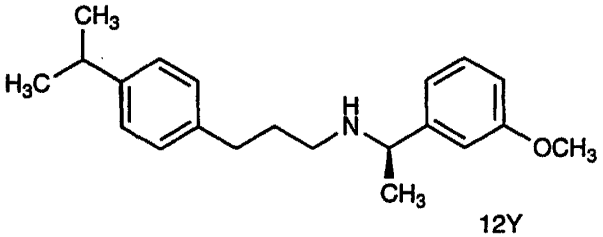
RECTIFIED SHEET (RULE 91)
ISA / EP



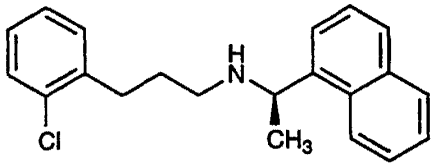
12W



12X



12Y



12Z

FIG. 12D.

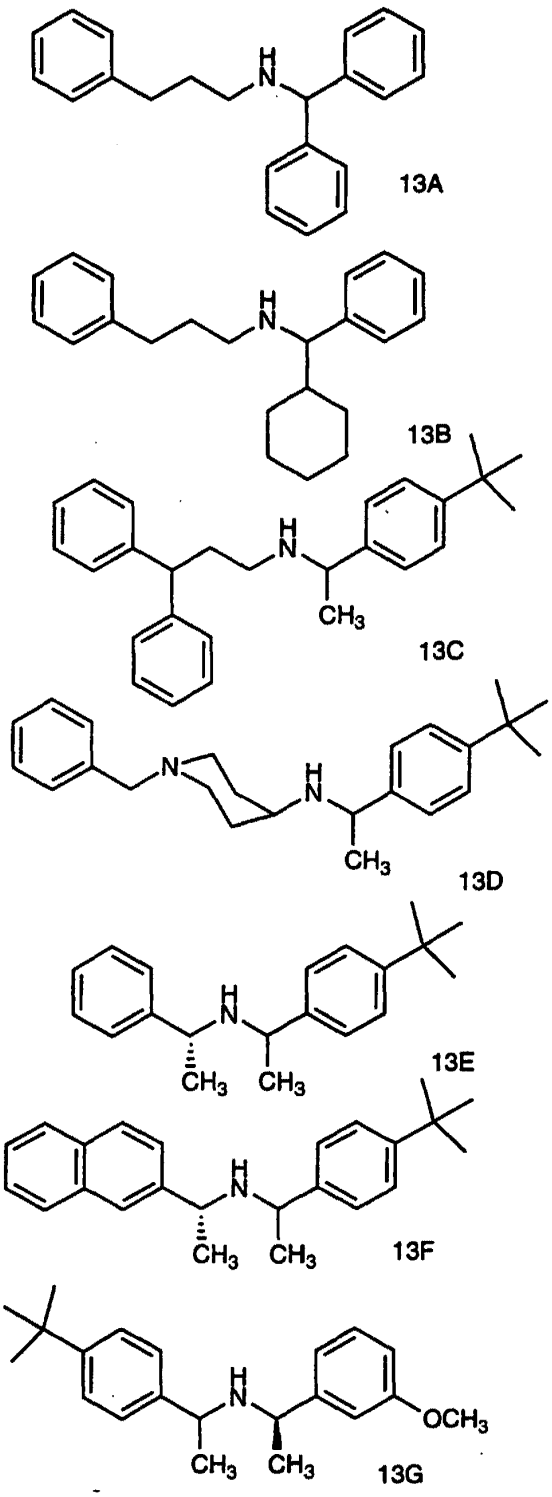
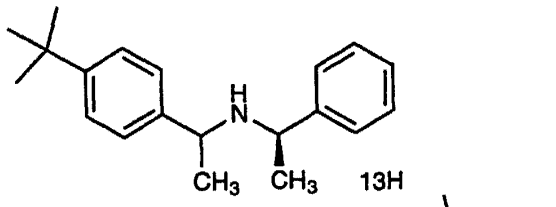
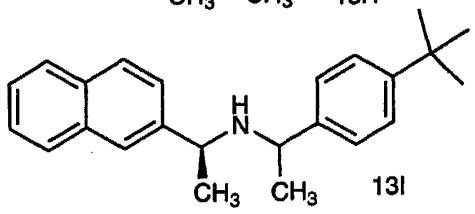


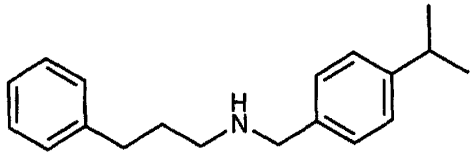
FIG. 13A.



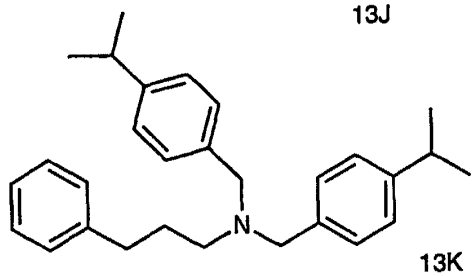
13H



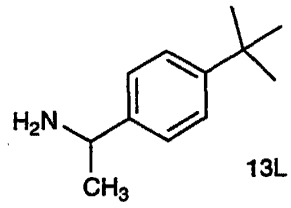
13I



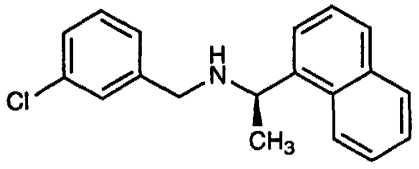
13J



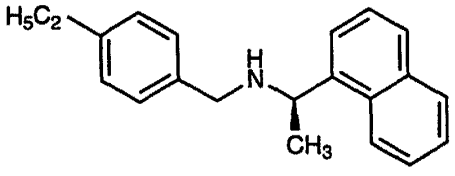
13K



13L



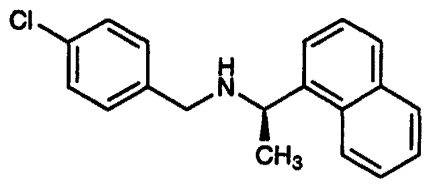
13M



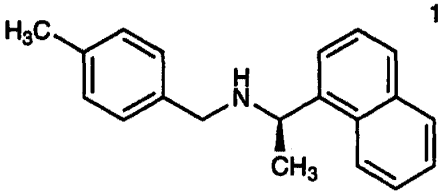
13N

FIG. 13B.

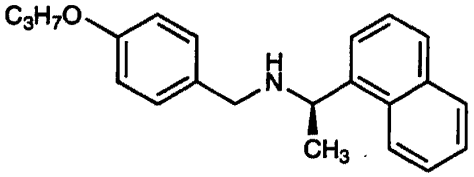
RECTIFIED SHEET (RULE 91)
ISA / EP



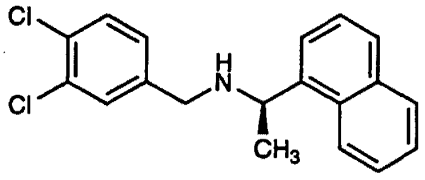
13O



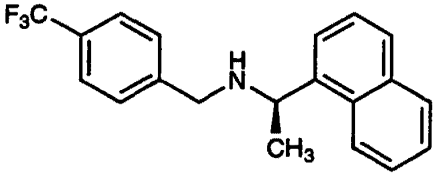
13P



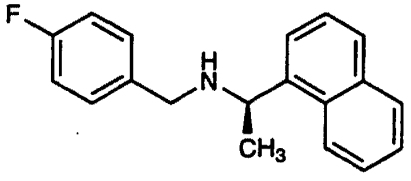
13Q



13R



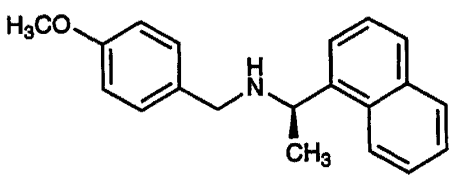
13S



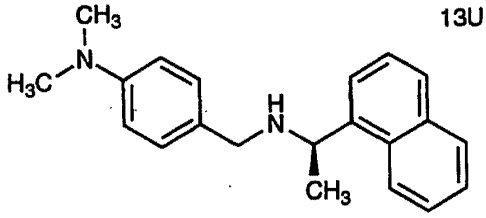
13T

FIG. 13C.

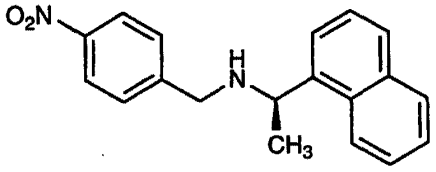
RECTIFIED SHEET (RULE 91)
ISA / EP



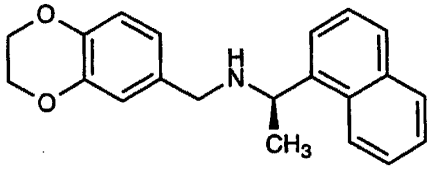
13U



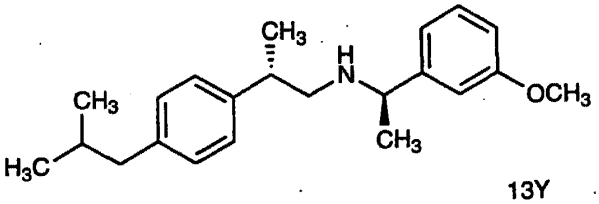
13V



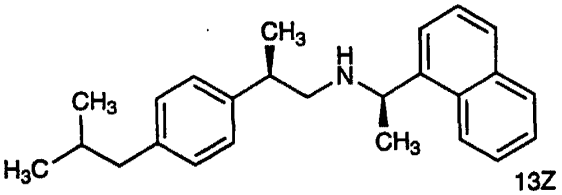
13W



13X

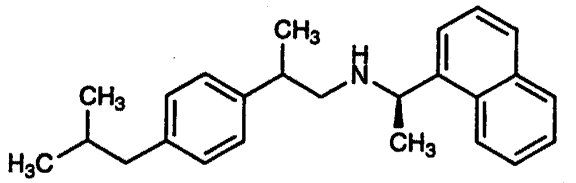


13Y

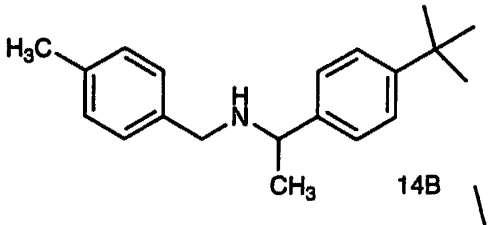


13Z

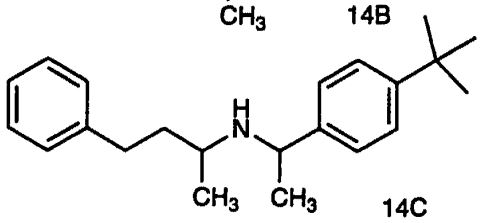
FIG. 13D.



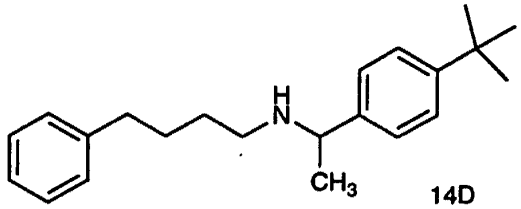
14A



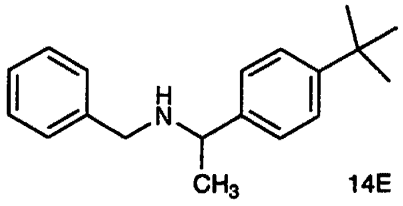
14B



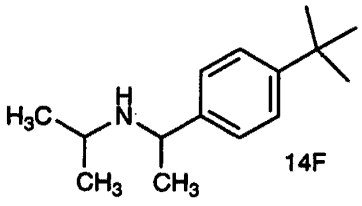
14C



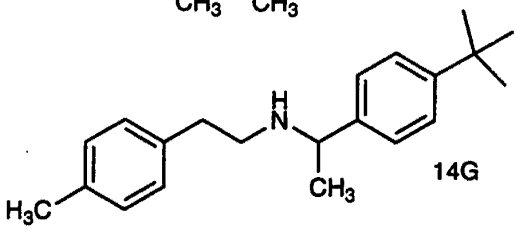
14D



14E



14F



14G

FIG. 14A.

RECTIFIED SHEET (RULE 91)
ISA / EP

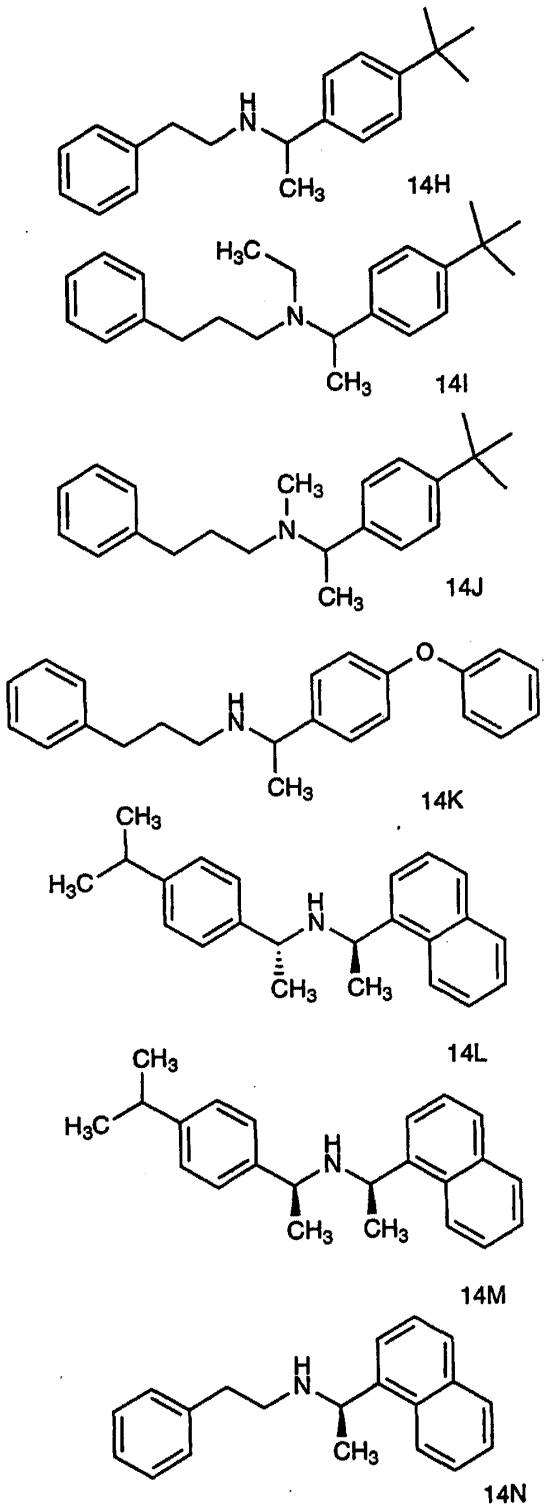
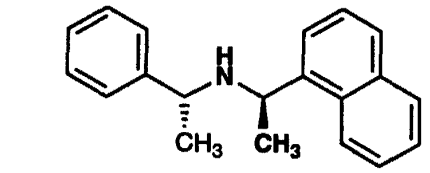
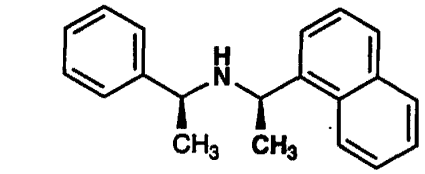


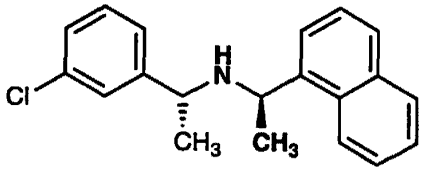
FIG. 14B.



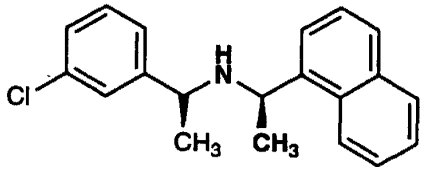
14O



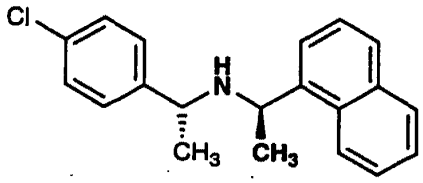
14P



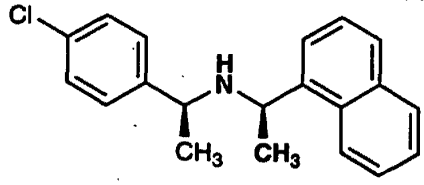
14Q



14R

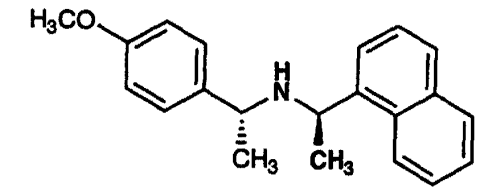


14S

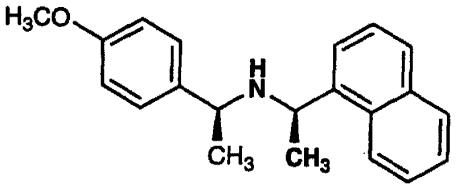


14T

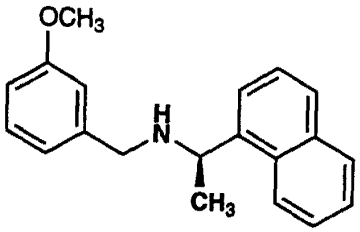
FIG. 14C.



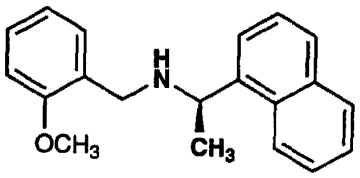
14U



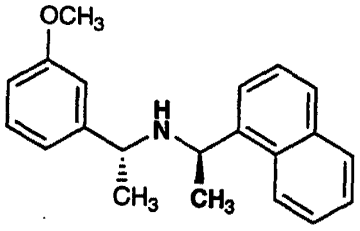
14V



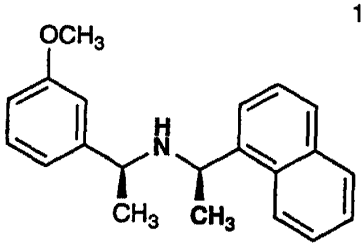
14W



14X



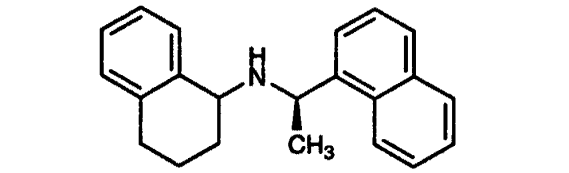
14Y



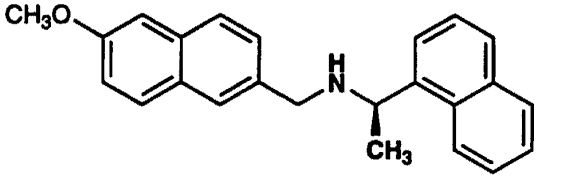
14Z

FIG. 14D.

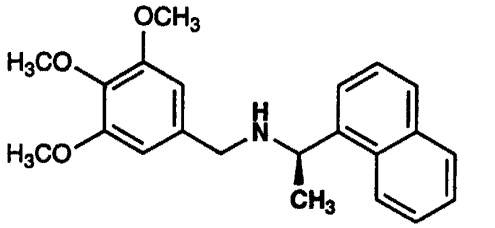
RECTIFIED SHEET (RULE 91)
ISA / EP



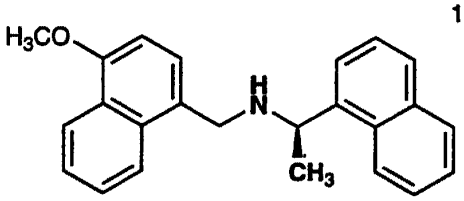
15A



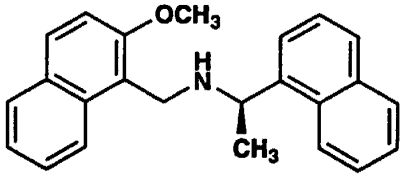
15B



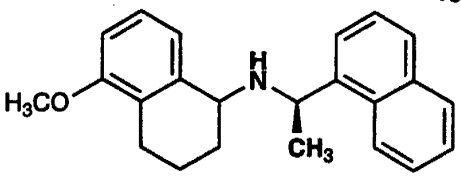
15C



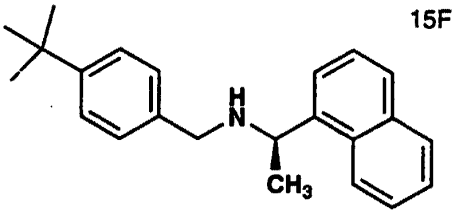
15D



15E



15F



15G

FIG. 15A.

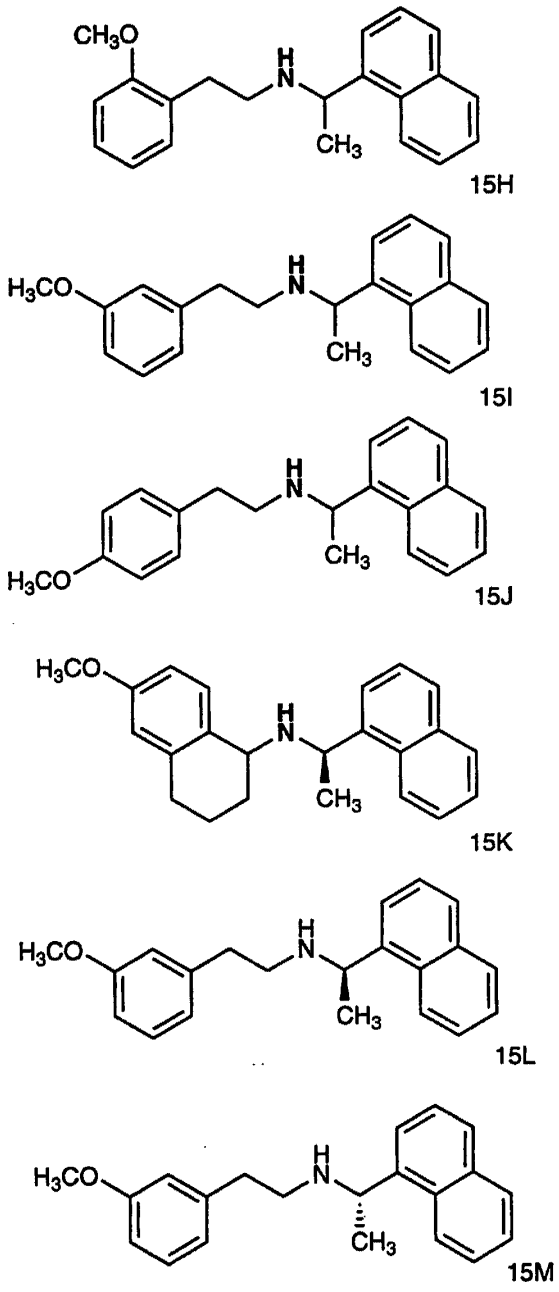
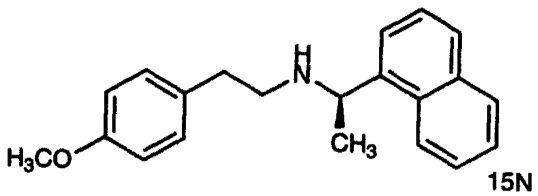
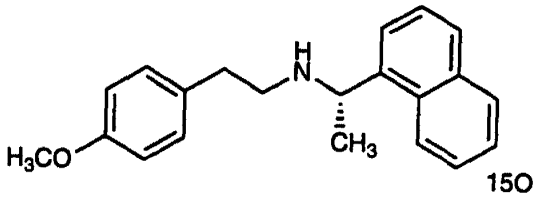

CH3O
H
N
CH3
15H
H3CO
H
N
CH3
15I
H3CO
H
N
CH3
15J
H3CO
H
N
CH3
15K
H3CO
H
N
CH3
15L
H3CO
H
N
CH3
15M

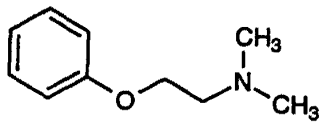
FIG. 15B.



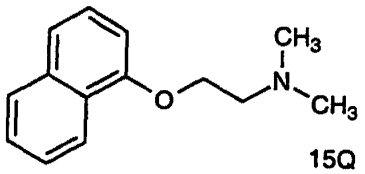
15N



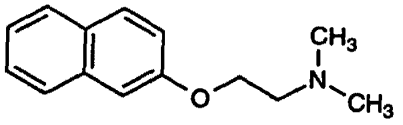
15O



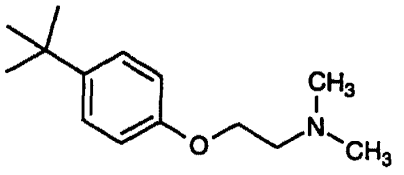
15P



15Q

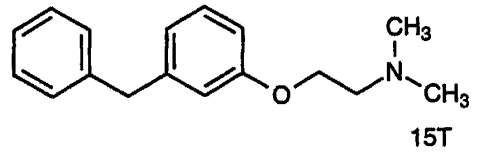


15R

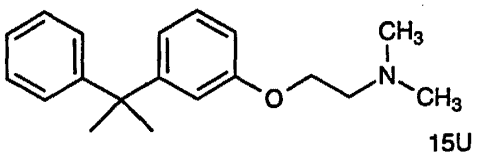


15S

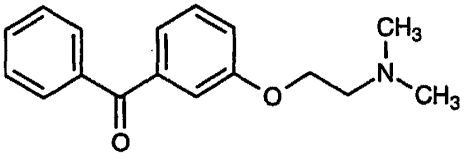
FIG. 15C.



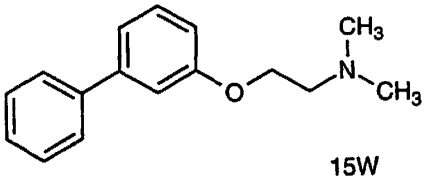
15T



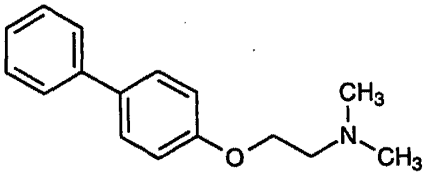
15U



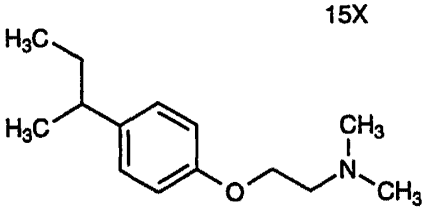
15V



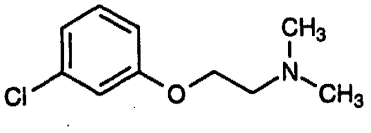
15W



15X

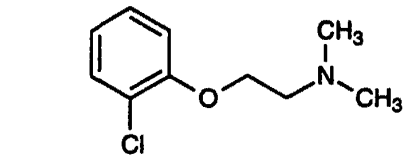


15Y

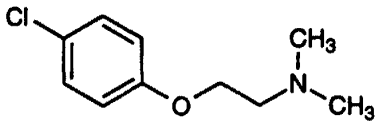


15Z

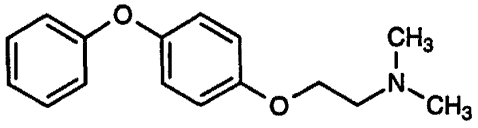
FIG. 15D.



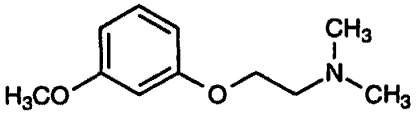
16A



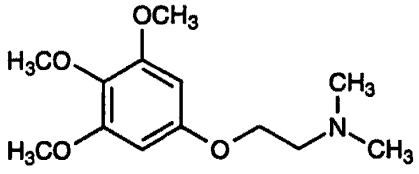
16B



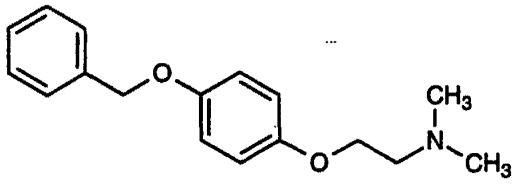
16C



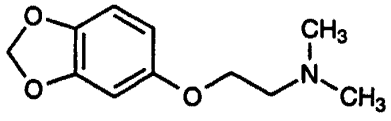
16D



16E



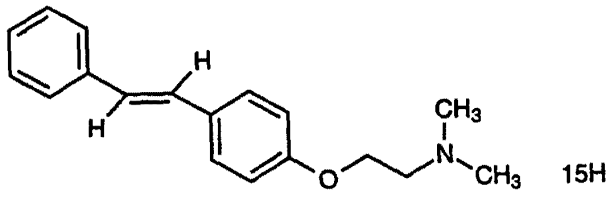
16F



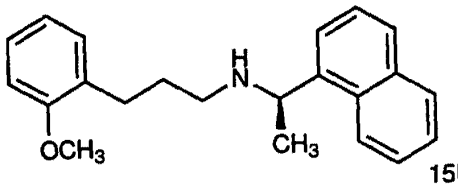
16G

FIG. 16A.

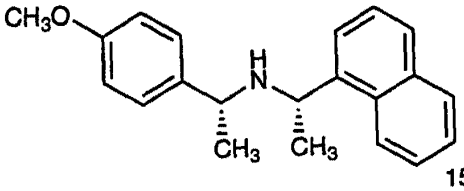
RECTIFIED SHEET (RULE 91)
ISA / EP



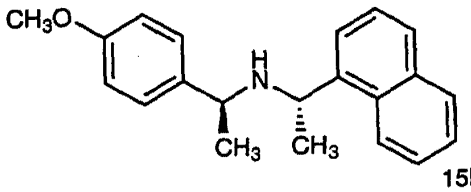
15H



15I

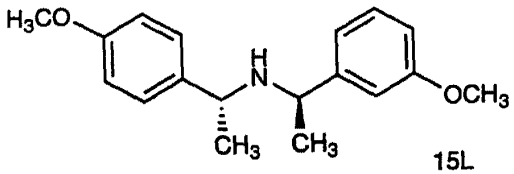


15J

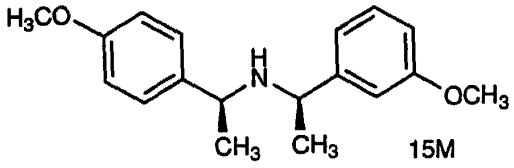


15K

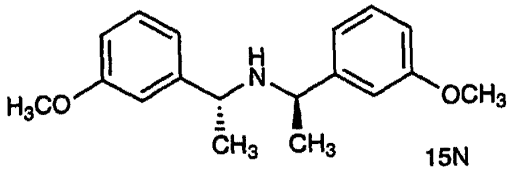
FIG. 16B.



15L

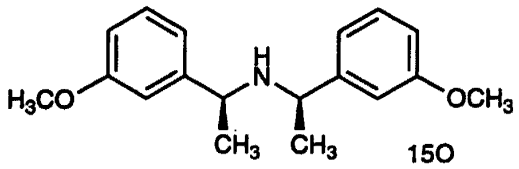


15M

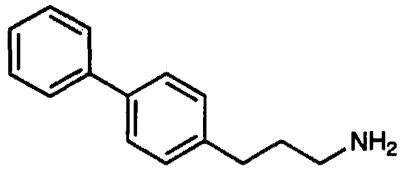


15N

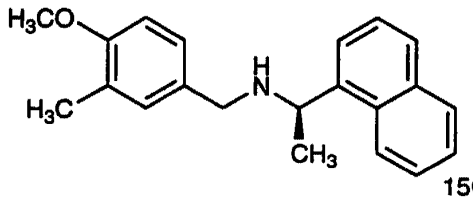
RECTIFIED SHEET (RULE 91)
ISA / EP



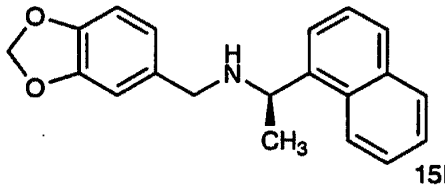
15O



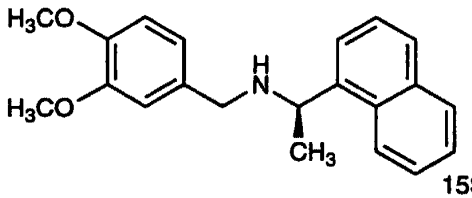
15P



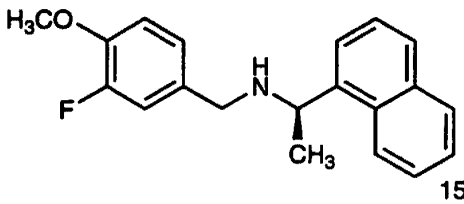
15Q



15R



15S



15T

FIG. 16C.

RECTIFIED SHEET (RULE 91)
ISA / EP

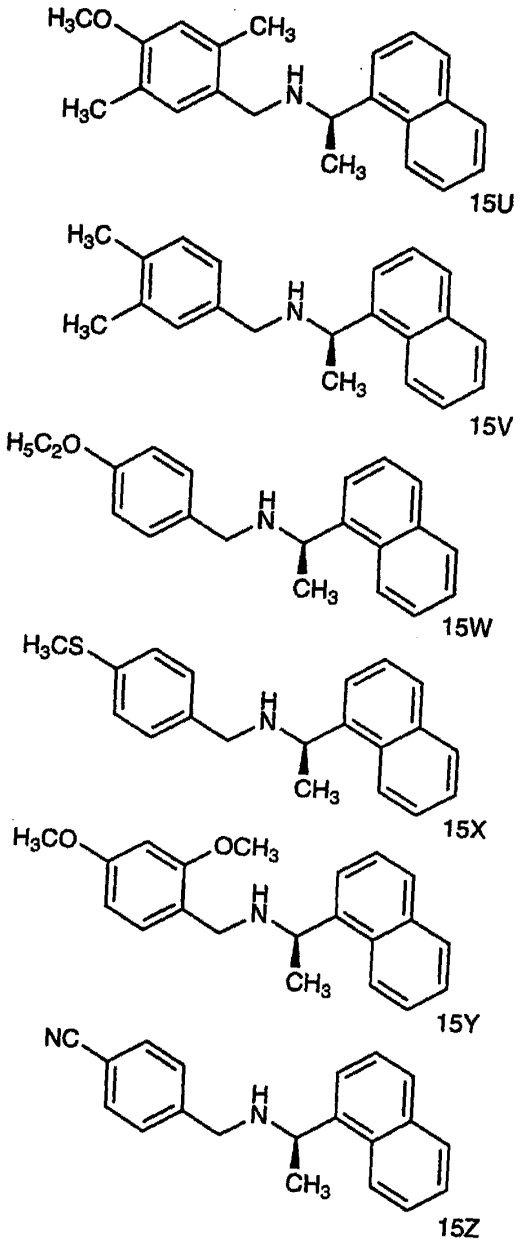

H3CO
CH3
H3C
H
N
CH3
15U
H3C
H3C
H
N
CH3
15V
H5C2O
H
N
CH3
15W
H3CS
H
N
CH3
15X
H3CO
OCH3
H
N
CH3
15Y
NC
H
N
CH3
15Z

FIG. 16D.

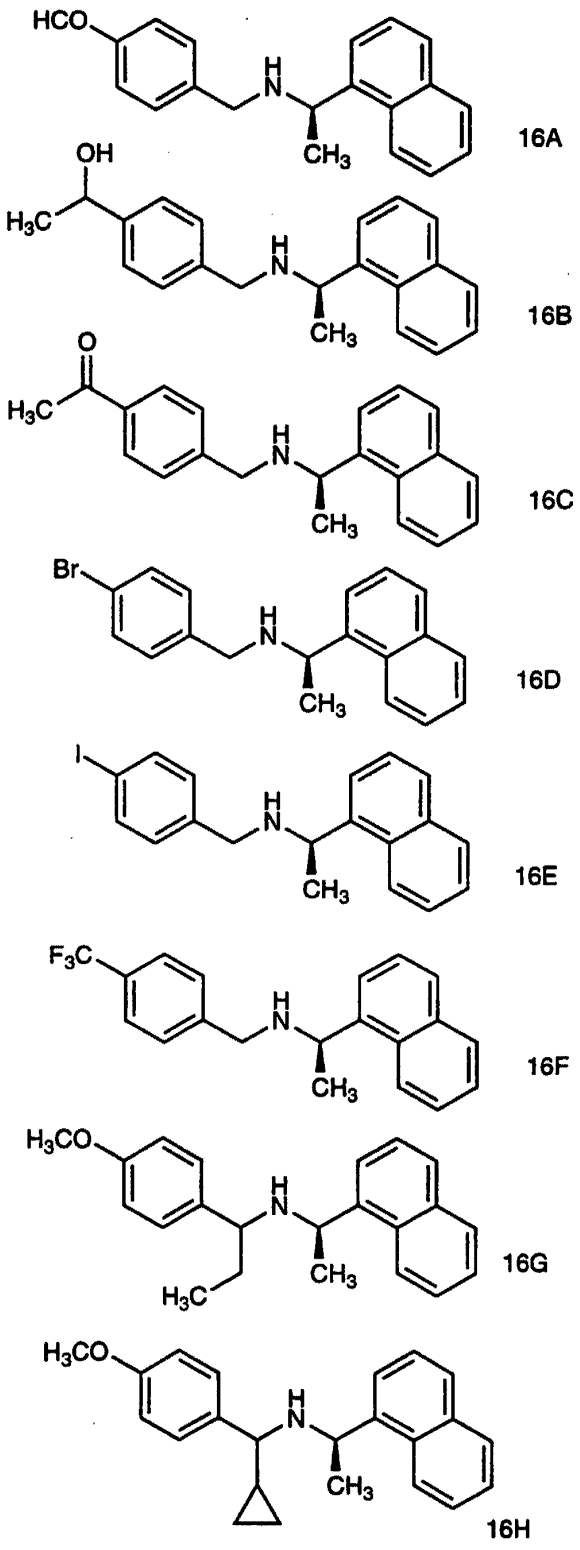
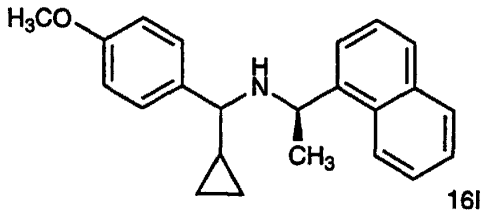
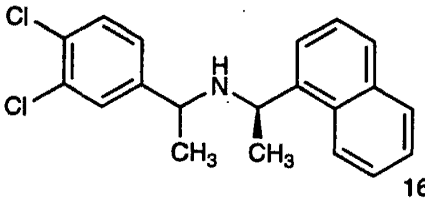

HCO
H
N
CH3
16A
OH
H3C
16B
O
H3C
16C
Br
16D
I
16E
F3C
16F
H3CO
H3C
16G
H3CO
16H

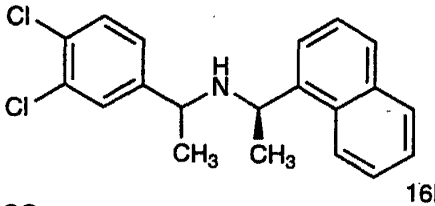
FIG. 17A.



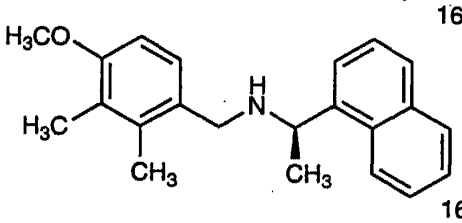
16I



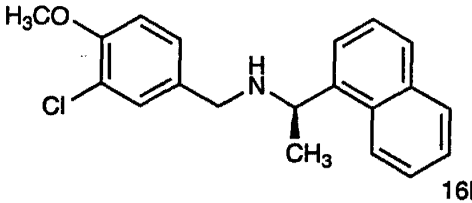
16J



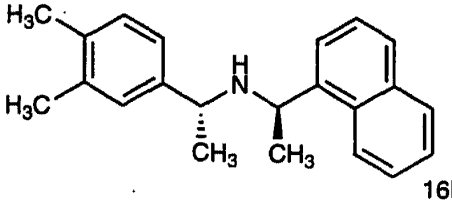
16K



16L



16M



16N

FIG. 17B.

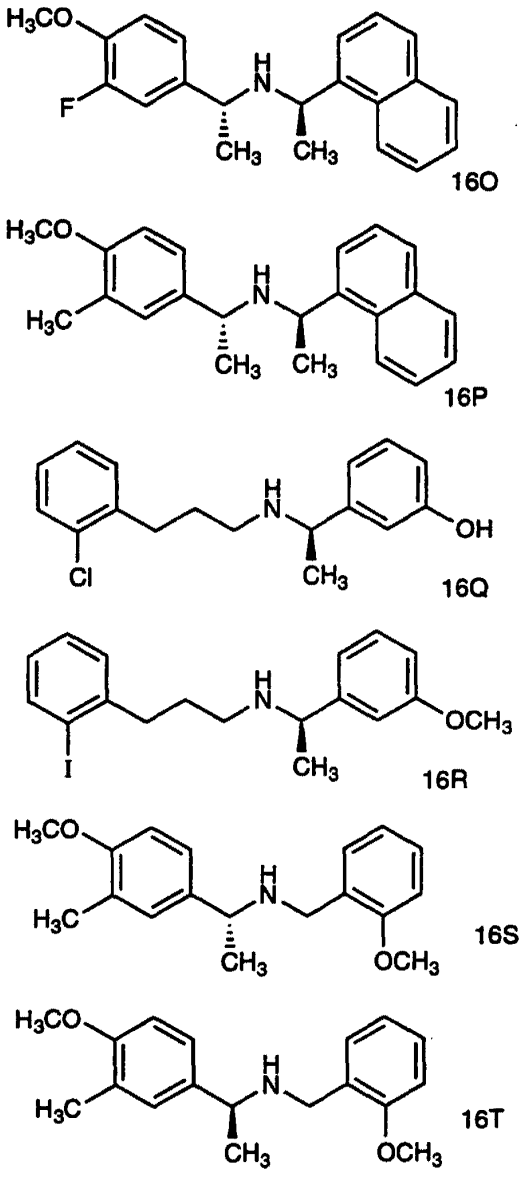

H3CO
F
H
N
CH3
CH3
16O
H3CO
H3C
H
N
CH3
CH3
16P
Cl
H
N
OH
CH3
16Q
I
H
N
OCH3
CH3
16R
H3CO
H3C
H
N
CH3
OCH3
16S
H3CO
H3C
H
N
CH3
OCH3
16T

FIG. 17C.

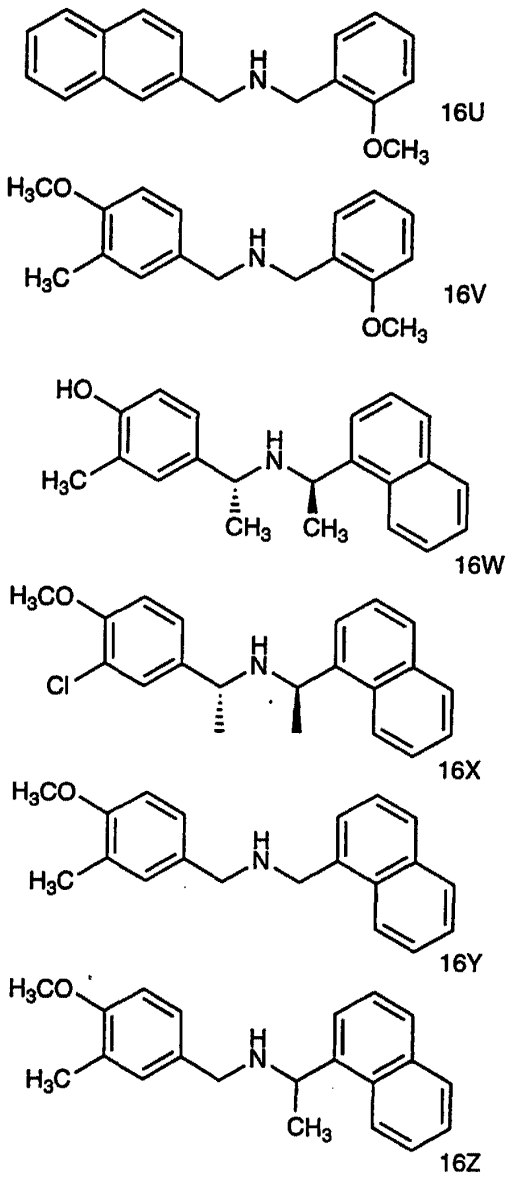
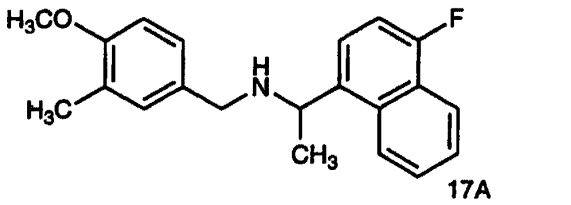
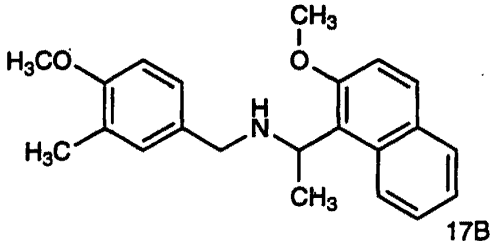


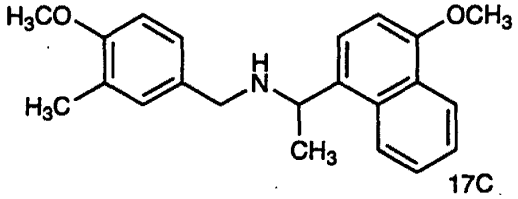
FIG. 17D.



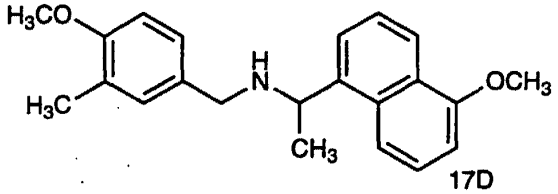
17A



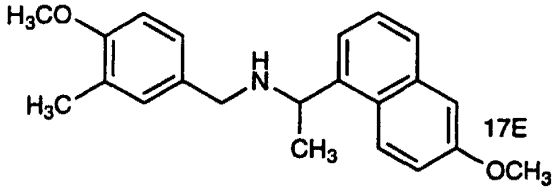
17B

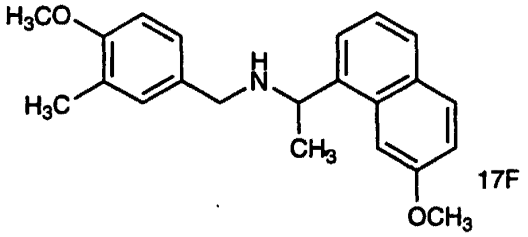


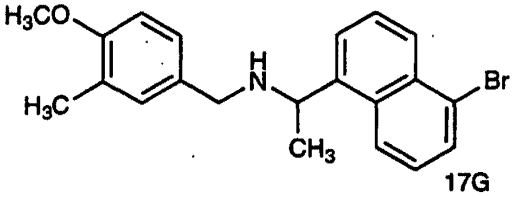
17C



17D







17G

FIG. 18A.

RECTIFIED SHEET (RULE 91)
ISA / EP

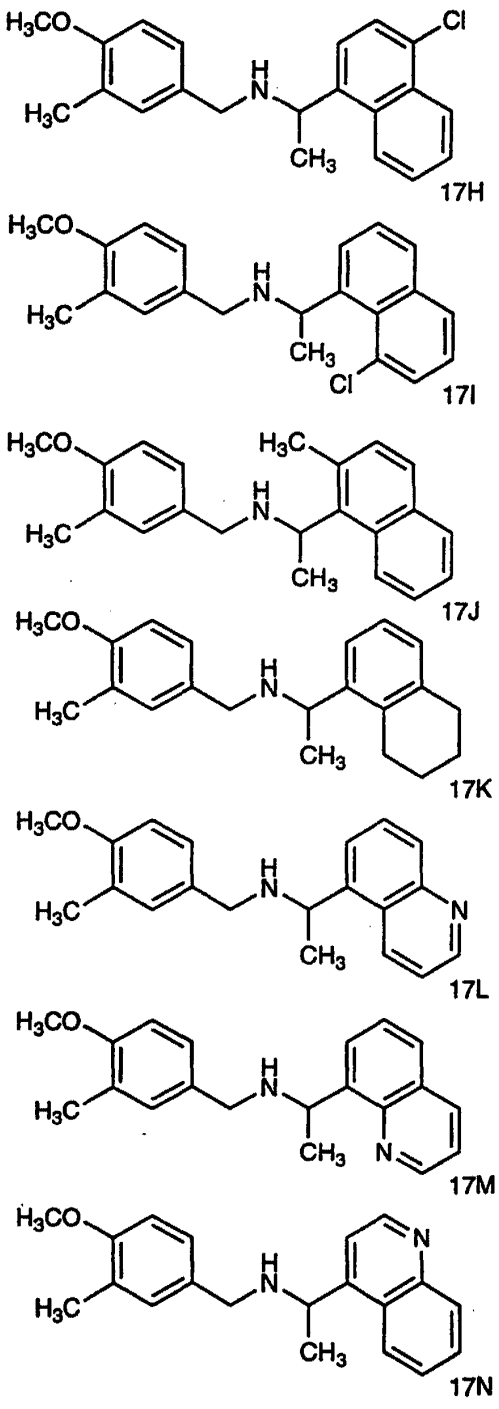
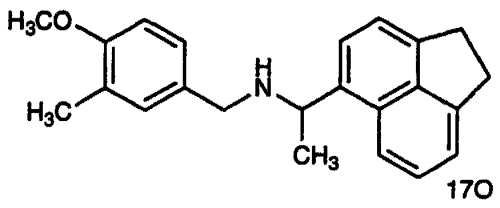
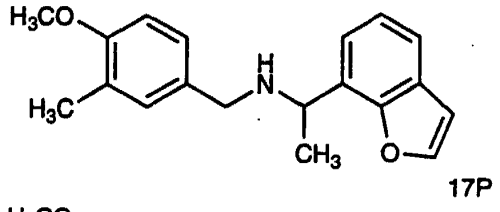


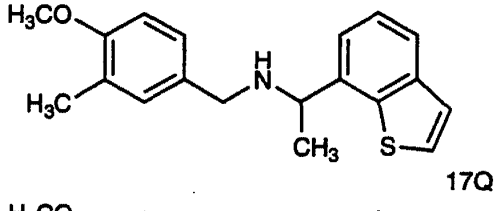
FIG. 18B.



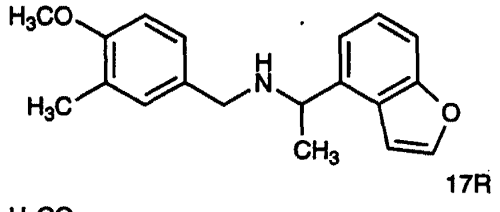
17O



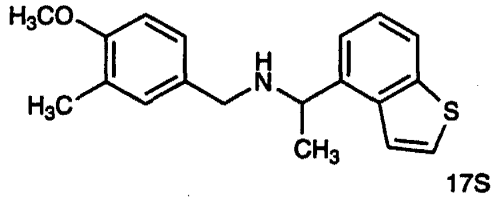
17P



17Q



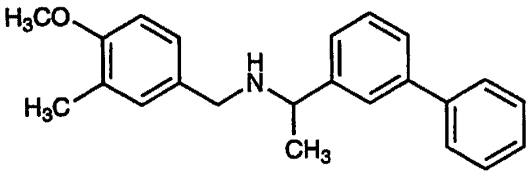
17R



17S

FIG. 18C.

RECTIFIED SHEET (RULE 91)
ISA / EP



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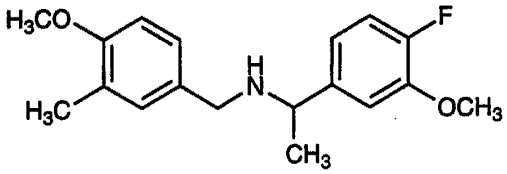

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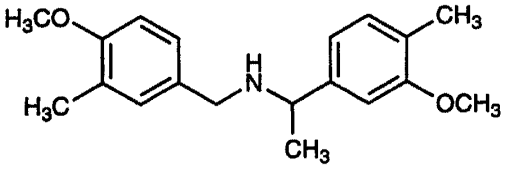

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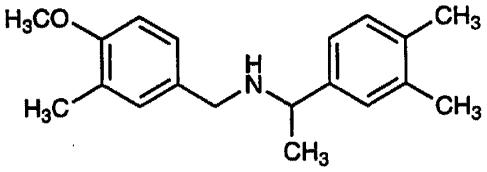

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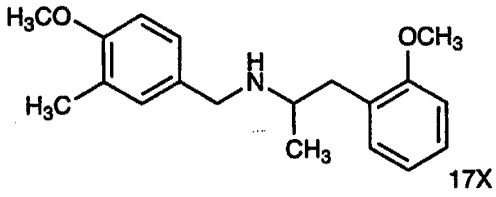

17X

FIG. 18D.

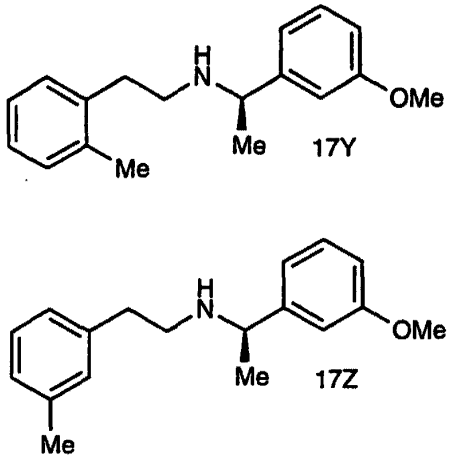

FIG. 18E.

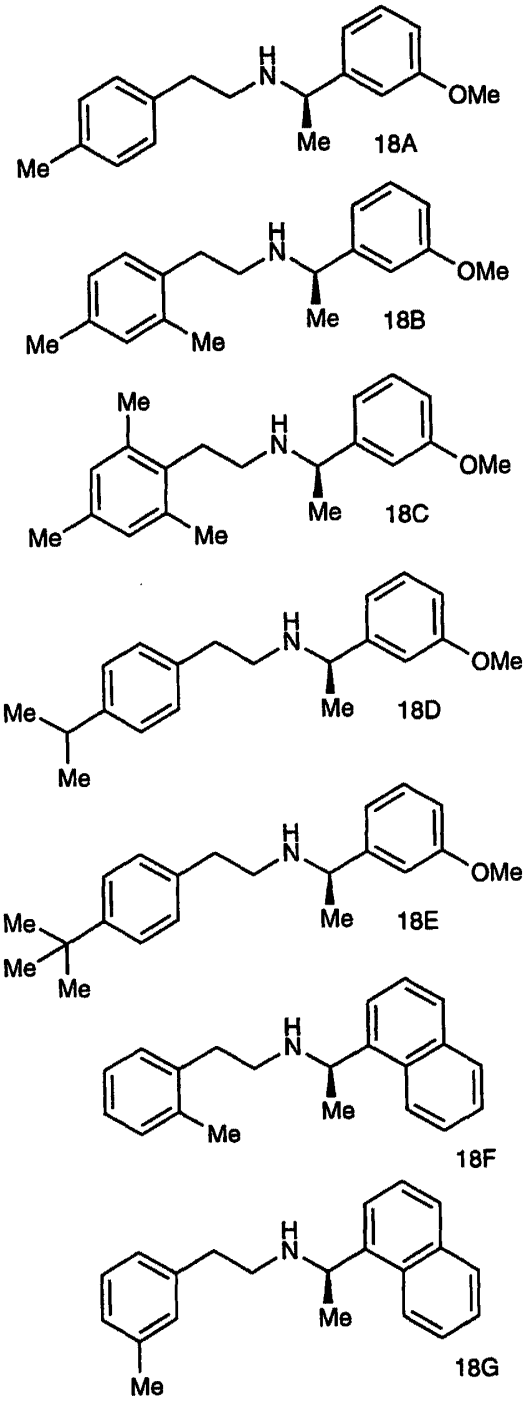
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FIG. 19A.

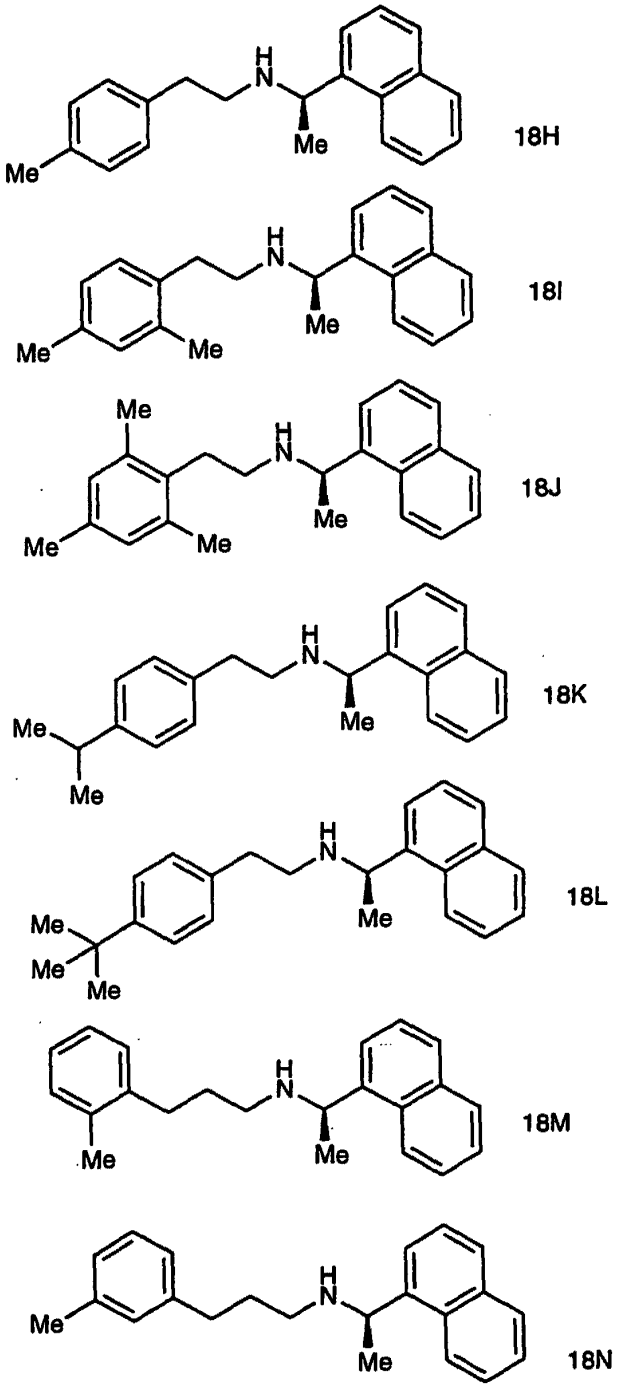
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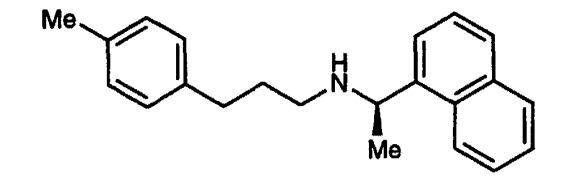
FIG. 19B.



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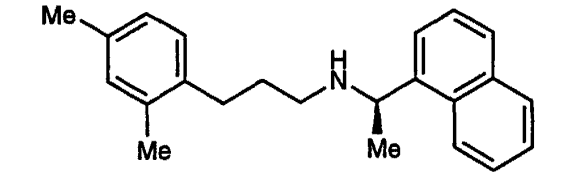

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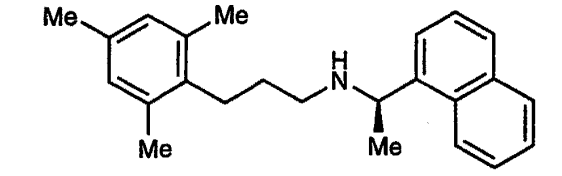

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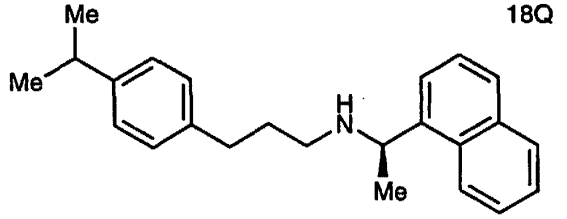

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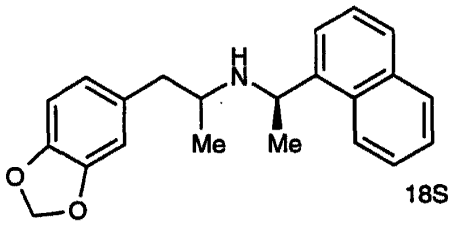

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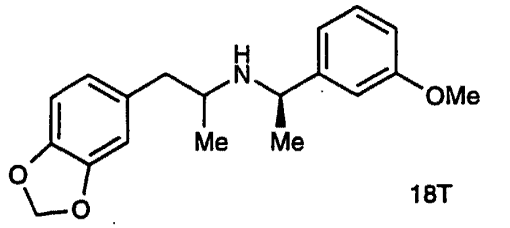
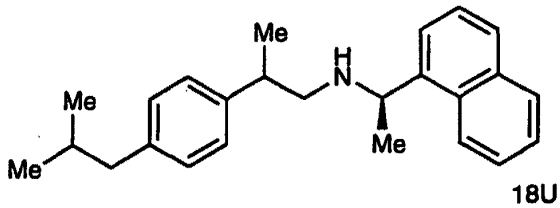


18T

FIG. 19C.

RECTIFIED SHEET (RULE 91)
ISA / EP



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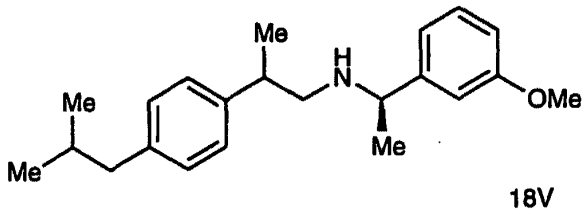

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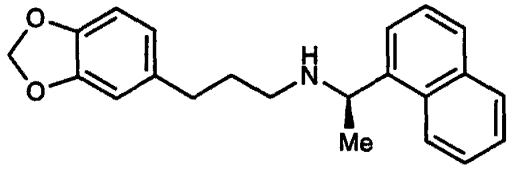

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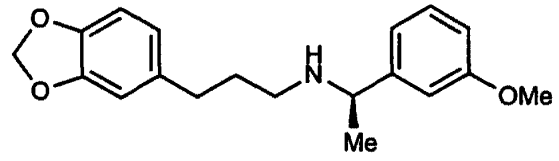

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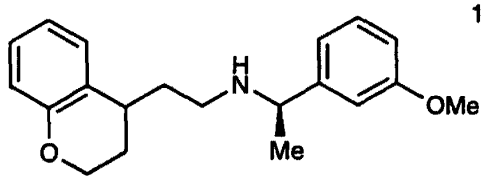
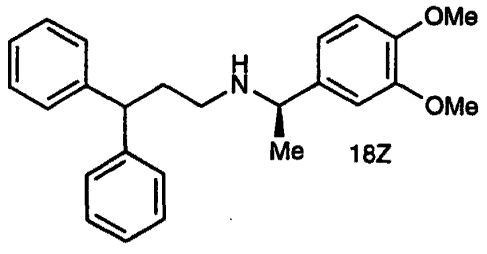


18Y

FIG. 19D.



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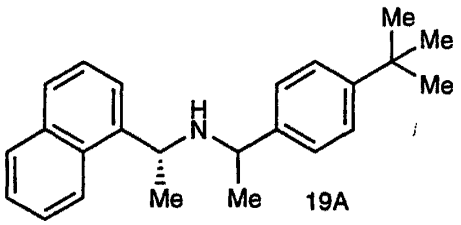

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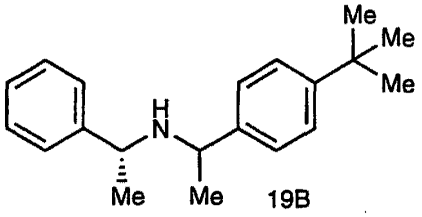

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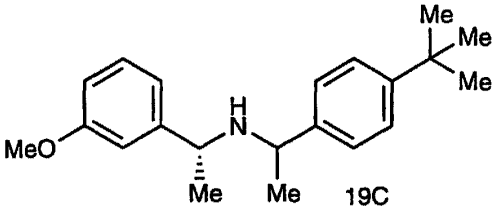

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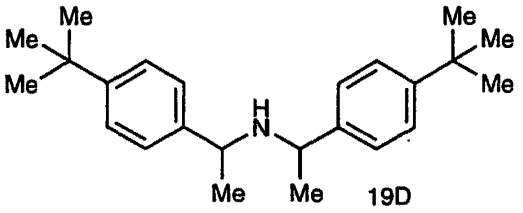

19D

FIG. 20A.

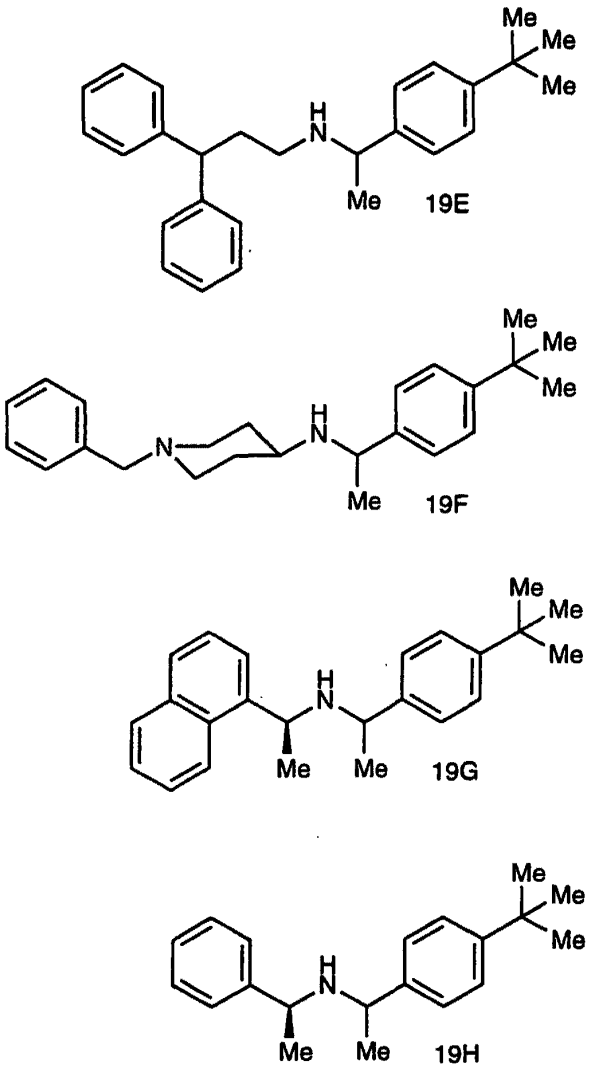
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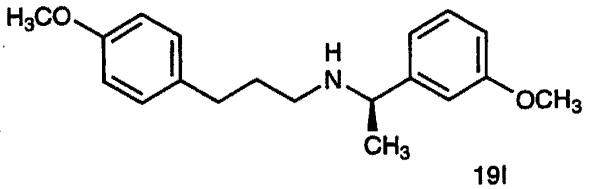
FIG. 20B.



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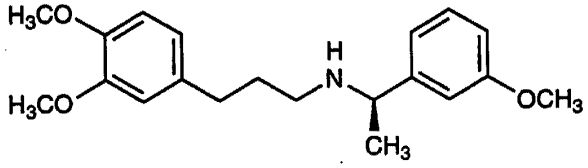

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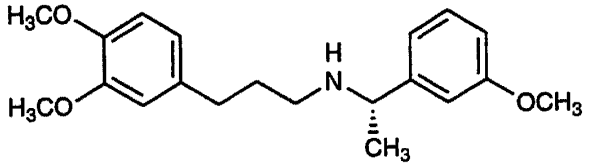

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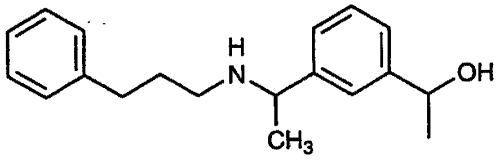

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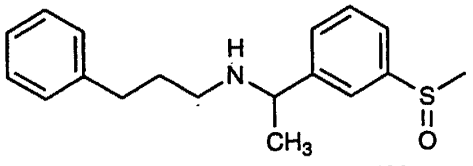

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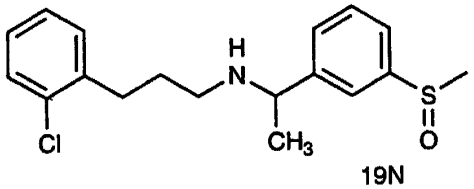

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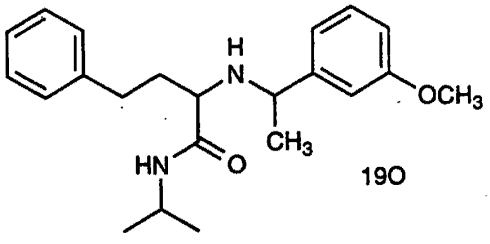

19O

FIG. 20C.

RECTIFIED SHEET (RULE 91)
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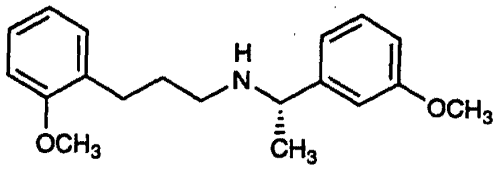

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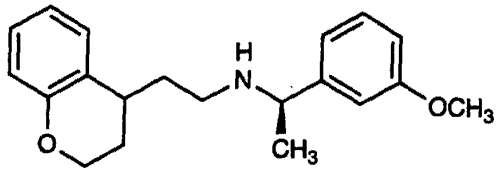

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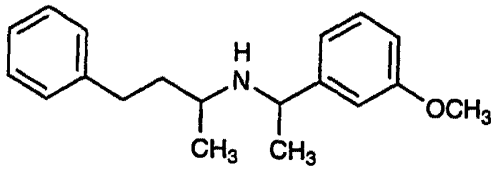

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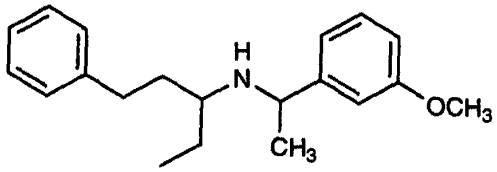

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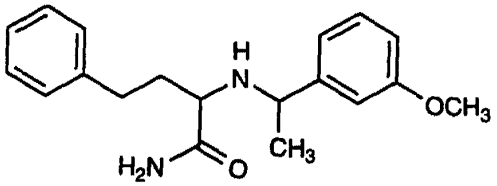

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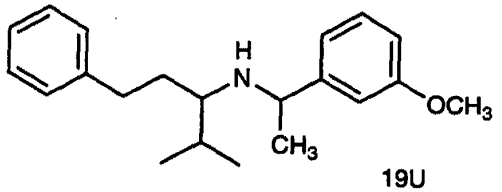
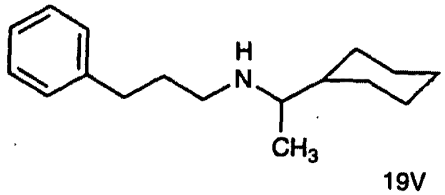


19U

FIG. 20D.



19V

INTERNATIONAL SEARCH REPORT

Interna al Application No
PCT/US 94/12117

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C07C211/30 C07C217/62 C07C217/58 C07C323/32 A61K31/13

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C07C

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO,A,93 04373 (NPS PHARMACEUTICALS) 4 March 1993 cited in the application see page 14, line 27 - page 17, line 14; claims 1,25,36,63,99 ---	1-19
X	TETRAHEDRON: ASYMMETRY, vol.2, no.3, 1991, OXFORD GB pages 183 - 186 S. G. DAVIES, O. ICHIHARA 'Asymmetric Synthesis of R-.beta.-Amino Butanoic Acid and S-.beta.-Tyrosine: Homochiral Lithium Amide Equivalents for Michael Addition to .alpha.,.beta.-Unsaturated Esters' see page 184, compounds 3,4,5 --- -/--	3

[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

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"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
31 January 1995	-9. 02. 95

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax (+31-70) 340-3016	Seufert, G

Form PCT/ISA/210 (second sheet) (July 1992)

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	TETRAHEDRON, vol.41, no.24, 1985, OXFORD GB pages 6005 - 6011 J. C. G. VAN NIEL, U. K. PANDIT 'NADH Models XXI. Stereoselective Reduction of Chiral Imines with Hantzsch Ester' see page 6006, compounds 5a-e	3
X	JOURNAL OF ORGANIC CHEMISTRY, vol.51, no.25, 1986, EASTON US pages 4953 - 4959 S. H. BERTZ ET AL. 'Asymmetric Induction with Amidocuprates' see page 4955, table II, entry 11	3
X	GB,A,2 213 818 (E. R. SQUIBB & SONS INC.) 23 August 1989 see examples 1,3-15	3
X	LIEBIGS ANNALEN DER CHEMIE, no.8, 1990, WEINHEIM DE pages 795 - 805 G. BRINGMANN ET AL. 'The Enantioselective Synthesis of Optically Active, Benzene Nucleus-Substituted 1-Phenylethylamines from the corresponding Acetophenones' see page 798, table 2, compound ent-9k	3
X	SYNLETT, no.5, 1990, STUTTGART DE pages 253 - 255 G. BRINGMANN ET AL. 'Enantiomerically Pure N-Boc Protected .beta.-Keto-.gamma.-Amino Acid Esters from Simple Keto Precursors: Novel, Stereocontrolled Approach to Statine Derivatives with Any Desired Configuration' see page 254, compound ent-7a	3
X	TETRAHEDRON: ASYMMETRY, vol.3, no.2, 1992, OXFORD GB pages 235 - 238 CORNELIS LENSINK, J. G. DE VRIES 'Improving Enantioselectivity by Using a Mono-Sulphonated Diphosphine as Ligand for Homogeneous Imine Hydrogenation' see page 235, compound 3	3

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2

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CHEMICAL ABSTRACTS, vol. 116, no. 2, 13 January 1992, Columbus, Ohio, US; abstract no. 14742u, A. G. BECALSKI ET AL. page 558 ; see abstract and RN 137769-86-9 & INORGANIC CHEMISTRY, vol.30, no.26, EASTON US pages 5002 - 5008 ---	3
X	EP,A,0 508 307 (SUMITOMO CHEMICAL COMPANY) 14 October 1992 see claim 1; examples ---	3
P,X	WO,A,94 18959 (NPS PHARMACEUTICALS) 4 March 1993 cited in the application see page 17, line 23 - page 19, line 23 see claims 1, 25-27, 30-36, 63-65, 68-73, 97, 99 see also RN 159149-96-9, 125275-99-2 ---	1-19
P,X	SYNTHESIS, no.12, December 1993, STUTTGART DE pages 1243 - 1246 E. JUARISTI ET AL. 'Use of N,N'-Dimethylpropenylurea (DMPU) as Solvent in the Efficient Preparation of Enatiomerically Pure Secondary Amines' see page 1245, line 17 - line 25 ---	3
P,X	CHEMICAL ABSTRACTS, vol. 121, no. 19, 7 November 1994, Columbus, Ohio, US; abstract no. 230462, Y. KOMEYOSHI, J. KUDO page 1060 ; see abstract and RN 158075-03-7, 158075-02-6,158075-01-5, 158074-98-7, 158074-97-6, 158074-96-5, 158074-95-4 & JP,A,6 116 214 (SUMITOMO CHEMICAL COMPANY) 26 April 1994 ------	3

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/12117

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Although claims 12-19 are directed to a method of treatment of the human body, the search has been carried out and based on the alleged effects of the compounds/compositions.

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. ☐ As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest.

☐ No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet (1)) (July 1992)

INTERNATIONAL SEARCH REPORT

information on patent family members

Inter nal Application No
PCT/US 94/12117

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO-A-9304373	04-03-93	AU-A-	2588992	16-03-93
		CA-A-	2115828	04-03-93
		JP-T-	6510531	24-11-94
		NO-A-	940581	25-04-94
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		GB-A,B	2245270	02-01-92
		US-A-	5097043	17-03-92
EP-A-0508307	14-10-92	JP-A-	5201938	10-08-93
		US-A-	5298660	29-03-94
WO-A-9418959	01-09-94	AU-B-	3777093	14-09-94
JP-A-6116214	26-04-94	NONE		

Form PCT/ISA/210 (patent family annex) (July 1992)

CORRECTED VERSION*

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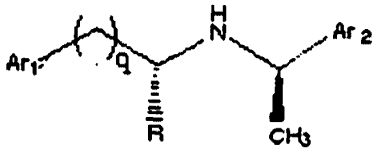
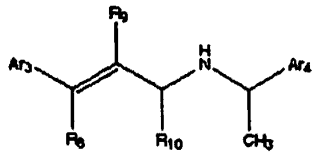


(a)



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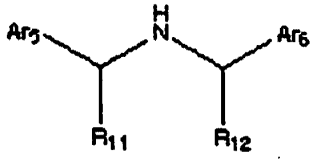

(c)

(57) Abstract

The present invention features compounds of general formulae a), b), c), able to modulate one or more activities of an inorganic ion receptor and methods for treating diseases or disorders by modulating inorganic ion receptor activity. Preferably, the compound can mimic or block the effect of extracellular Ca^{2+} on a calcium receptor.

*(Referred to in PCT Gazette No. 34/2000, Section II)

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DESCRIPTION

Calcium Receptor-Active Compounds

Field of the Invention

This invention relates to the design, development, composition and use of compounds able to modulate one or more inorganic ion receptor activities.

Background of the Invention

Certain cells in the body respond not only to chemical signals, but also to ions such as extracellular calcium ions (Ca^{2+}). Changes in the concentration of extracellular Ca^{2+} (referred to herein as "$[Ca^{2+}]$") alter the functional responses of these cells. One such specialized cell is the parathyroid cell which secretes parathyroid hormone (PTH). PTH is the principal endocrine factor regulating Ca^{2+} homeostasis in the blood and extracellular fluids.

PTH, by acting on bone and kidney cells, increases the level of Ca^{2+} in the blood. This increase in $[Ca^{2+}]$ then acts as a negative feedback signal, depressing PTH secretion. The reciprocal relationship between $[Ca^{2+}]$ and PTH secretion forms the essential mechanism maintaining bodily Ca^{2+} homeostasis.

Extracellular Ca^{2+} acts directly on parathyroid cells to regulate PTH secretion. The existence of a parathyroid cell surface protein which detects changes in $[Ca^{2+}]$ has been confirmed. Brown *et al.*, 366 Nature 574, 1993. In parathyroid cells, this protein acts as a receptor for extracellular Ca^{2+} ("the calcium receptor"), and detects changes in $[Ca^{2+}]$ and to initiate a functional cellular response, PTH secretion.

Extracellular Ca^{2+} can exert effects on different cell functions, reviewed in Nemeth *et al.*, 11 Cell Calcium 319, 1990. The role of extracellular Ca^{2+} in parafollicular (C-cells) and parathyroid cells is discussed in Nemeth, 11

Cell Calcium 323, 1990. These cells have been shown to express similar Ca^{2+} receptor. Brown *et al.*, 366 Nature 574, 1993; Mithal et al., 9 Suppl. 1 J. Bone and Mineral Res. s282, 1994; Rogers *et al.*, 9 Suppl. 1 J. Bone and Mineral Res. s409, 1994; Garrett *et al.*, 9 Suppl. 1 J. Bone and Mineral Res. s409, 1994. The role of extracellular Ca^{2+} on bone osteoclasts is discussed by Zaidi, 10 Bioscience Reports 493, 1990. In addition keratinocytes, juxtaglomerular cells, trophoblasts, pancreatic beta cells and fat/adipose cells all respond to increases in extracellular calcium which likely reflects activation of calcium receptors of these cells.

The ability of various compounds to mimic extracellular Ca^{2+} *in vitro* is discussed by Nemeth *et al.*, (spermine and spermidine) in "Calcium-Binding Proteins in Health and Disease," 1987, Academic Press, Inc., pp. 33-35; Brown *et al.*, (*e.g.*, neomycin) 128 Endocrinology 3047, 1991; Chen *et al.*, (diltiazem and its analog, TA-3090) 5 J. Bone and Mineral Res. 581, 1990; and Zaidi *et al.*, (verapamil) 167 Biochem. Biophys. Res. Commun. 807, 1990. Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959, and Nemeth *et al.*, PCT/US92/07175, International Publication Number WO 93/04373, describe various compounds which can modulate the effect of an inorganic ion on a cell having an inorganic ion receptor.

The references provided in the background are not admitted to be prior art.

Summary of the Invention

The present invention features compounds able to modulate one or more activities of an inorganic ion receptor and methods for treating diseases or disorders by modulating inorganic ion receptor activity. Preferred compounds can mimic or block the effect of extracellular calcium on a cell surface calcium receptor.

Diseases or disorders which can be treated by modulating inorganic ion receptor activity include one or more of the following types: (1) those characterized by abnormal inorganic ion homeostasis, preferably calcium homeostasis; (2) those characterized by an abnormal amount of an extracellular or intracellular messenger whose production can be affected by inorganic ion receptor activity, preferably calcium receptor activity; (3) those characterized by an abnormal effect (*e.g.*, a different effect in kind or magnitude) of an intracellular or extracellular messenger which can itself be ameliorated by inorganic ion receptor activity, preferably calcium receptor activity; and (4) other diseases or disorders in which modulation of inorganic ion receptor activity, preferably calcium receptor activity will exert a beneficial effect, for example, in diseases or disorders where the production of an intracellular or extracellular messenger stimulated by receptor activity compensates for an abnormal amount of a different messenger. Examples of extracellular messengers whose secretion and/or effect can be affected by modulating inorganic ion receptor activity include inorganic ions, hormones, neurotransmitters, growth factors, and chemokines. Examples of intracellular messengers include cAMP, cGMP, IP_3, and diacylglycerol.

Thus, a compound of this invention preferably modulates calcium receptor activity and is used in the treatment of diseases or disorders which can be affected by modulating one or more activities of a calcium receptor. Calcium receptor proteins enable certain specialized cells to respond to changes in extracellular Ca^{2+} concentration. For example, extracellular Ca^{2+} inhibits the secretion of parathyroid hormone from parathyroid cells, inhibits bone resorption by osteoclasts, and stimulates secretion of calcitonin from C-cells.

In a preferred embodiment, the compound is used to treat a disease or disorder characterized by abnormal bone and mineral homeostasis, more preferably calcium homeo-

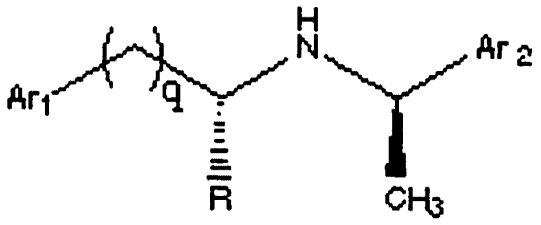
stasis. Extracellular Ca^{2+} is under tight homeostatic control and controls various processes such as blood clotting, nerve and muscle excitability, and proper bone formation. Abnormal calcium homeostasis is characterized by one or more of the following activities: (1) an abnormal increase or decrease in serum calcium; (2) an abnormal increase or decrease in urinary excretion of calcium; (3) an abnormal increase or decrease in bone calcium levels, for example, as assessed by bone mineral density measurements; (4) an abnormal absorption of dietary calcium; (5) an abnormal increase or decrease in the production and/or release of messengers which affect serum calcium levels such as parathyroid hormone and calcitonin; and (6) an abnormal change in the response elicited by messengers which affect serum calcium levels. The abnormal increase or decrease in these different aspects of calcium homeostasis is relative to that occurring in the general population and is generally associated with a disease or disorder.

Diseases and disorders characterized by abnormal calcium homeostasis can be due to different cellular defects such as a defective calcium receptor activity, a defective number of calcium receptors, or a defective intracellular protein acted on by a calcium receptor. For example, in parathyroid cells, the calcium receptor is coupled to the G_i protein which in turn inhibits cyclic AMP production. Defects in G_i protein can affect its ability to inhibit cyclic AMP production.

Thus, a first aspect the invention features an inorganic ion receptor modulating compound having the formula:

5

STRUCTURE I



where Ar_1 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, $N(CH_3)_2$, phenyl, phenoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, acetyl, ethylene dioxy;

Ar_2 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, and acetoxy;

q is 0, 1, 2, or 3; and

R is either H, or lower alkyl;

and pharmaceutically salts and complexes thereof.

Compounds of this invention have preferred stereochemistry. The CH_3 shown in Structure I is at a chiral center and provides an α-(*R*)-methyl structure. When R is CH_3, the R shown in Structure I is also at chiral center which provides an (*R*)-methyl structure. Thus, when R is CH_3, the Structure I compound has (*R,R*) stereochemistry.

Inorganic ion receptor activities are those processes brought about as a result of inorganic ion receptor activation. Such processes include the production of molecules which can act as intracellular or extracellular messengers.

Inorganic ion receptor-modulating compound include ionomimetics, ionolytics, calcimimetics, and calcilytics. Ionomimetics are compounds which bind to an inorganic ion receptor and mimic (*i.e.*, evoke or potentiate) the effects of an inorganic ion at an inorganic ion receptor. Preferably, the compound affects one or more calcium receptor activities. Calcimimetics are ionomimetics which effects one or more calcium receptor activities and bind to a calcium receptor.

Ionolytics are compounds which bind to an inorganic ion receptor and block (*i.e.*, inhibit or diminish) one or more activities caused by an inorganic ion at an inorganic ion receptor. Preferably, the compound affects one or more calcium receptor activities. Calcilytics are ionolytics which block one or more calcium receptor activities evoked by extracellular calcium and bind to a calcium receptor.

Ionomimetics and ionolytics may bind at the same receptor site as the native inorganic ion ligand binds or can bind at a different site (*e.g.*, allosteric site). For example, NPS *R*-467 binding to a calcium receptor results in calcium receptor activity and, thus, NPS *R*-467 is classified as a calcimimetic. However, NPS *R*-467 binds to the calcium receptor at a different site (*i.e.*, an allosteric site) than extracellular calcium.

A measure of a compounds effectiveness can be determined by calculating the EC_{50} or IC_{50} for that compound. The EC_{50} is the concentration of a compound which causes a half maximal mimicking effect. The IC_{50} is the concentration of compound which causes a half-maximal blocking effect. EC_{50} and IC_{50} for compounds at a calcium receptor can be determined by assaying one or more of the activities of extracellular calcium at a calcium receptor. Examples of assays for measuring EC_{50}, and IC_{50} are described Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959, and Nemeth *et al.*, PCT/US92/07175, International Publication Number WO

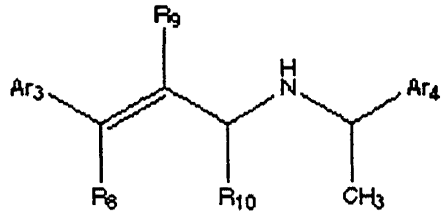
93/04373, (both of these publications are hereby incorporated by reference here) and below. Such assays include oocyte expression assays and measuring increases in intracellular calcium ion concentration ($[Ca^{2+}]_i$) due to calcium receptor activity. Preferably, such assays measure the release or inhibition of a particular hormone associated with activity of a calcium receptor.

An inorganic ion receptor-modulating compound preferably selectively targets inorganic ion receptor activity in a particular cell. For example, selective targeting of a calcium receptor activity is achieved by a compound exerting a greater effect on a calcium receptor activity in one cell type than at another cell type for a given concentration of compound. Preferably, the differential effect is 10-fold or greater as measured *in vivo* or *in vitro*. More preferably, the differential effect is measured *in vivo* and the compound concentration is measured as the plasma concentration or extracellular fluid concentration and the measured effect is the production of extracellular messengers such as plasma calcitonin, parathyroid hormone, or plasma calcium. For example, in a preferred embodiment, the compound selectively targets PTH secretion over calcitonin secretion.

Preferably, the compound is either a calcimimetic or calcilytic having an EC_{50} or IC_{50} at a calcium receptor of less than or equal to 5 μM, and even more preferably less than or equal to 1 μM, 100 nmolar, 10 nmolar, or 1 nmolar using one of the assays described below. More preferably, the assay measures intracellular Ca^{2+} in HEK 293 cells transformed with nucleic acid expressing the human parathyroid calcium receptor and loaded with fura-2. Lower EC_{50}'s or IC_{50}'s are advantageous since they allow lower concentrations of compounds to be used *in vivo* or *in vitro*. The discovery of compounds with low EC_{50}'s and IC_{50}'s enables the design and synthesis of additional compounds having similar or improved potency, effectiveness, and/or selectivity.

Another aspect of the present invention features an inorganic ion receptor modulating compound having the formula:

STRUCTURE II



where Ar_3 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, $N(CH_3)_2$, acetyl, ethylene dioxy.

Ar_4 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, and acetoxy;

R_8 is either hydrogen or phenyl;

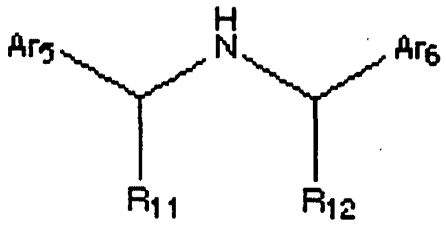
R_9 is either hydrogen or methyl; and

R_{10} is either hydrogen, methyl, or phenyl;

or pharmaceutically acceptable salts and complexes thereof.

Another aspect of the present invention features an inorganic ion receptor modulating compound having the formula:

STRUCTURE III



where Ar_5 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, acetyl, ethylene dioxy, -CH=CH-phenyl;

Ar_6 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, acetyl, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, carbomethoxy, $OCH_2C(O)C_2H_5$ and acetoxy;

R_{11} is hydrogen or methyl; and

R_{12} is hydrogen or methyl.

Another aspect of the present invention features a pharmaceutical composition made up of an inorganic ion receptor-modulating compound described herein and a physiologically acceptable carrier. A "pharmacological composition" refers to a composition in a form suitable for administration into a mammal, preferably a human. Preferably, the pharmaceutical composition contains a sufficient amount of a calcium receptor modulating compound in a proper pharmaceutical form to exert a therapeutic effect on a human.

Considerations concerning forms suitable for administration are known in the art and include toxic effects, solubility, route of administration, and maintaining

activity. For example, pharmacological compositions injected into the blood stream should be soluble.

Pharmaceutical compositions can also be formulated as pharmaceutically acceptable salts (*e.g.*, acid addition salts) and complexes thereof. The preparation of such salts can facilitate the pharmacological use of a compound by altering its physical characteristics without preventing it from exerting a physiological effect.

Another aspect the present invention features a method for treating a patient by modulating inorganic ion receptor activity using inorganic ion receptor modulating compounds described herein. The method involves administering to the patient a pharmaceutical composition containing a therapeutically effective amount of an inorganic ion receptor-modulating compound. In a preferred embodiment, the disease or disorder is treated by modulating calcium receptor activity by administering to the patient a therapeutically effective amount of a calcium receptor-modulating compound.

Inorganic ion receptor-modulating compounds, and compositions containing the compounds, can be used to treat patients. A "patient" refers to a mammal in which modulation of an inorganic ion receptor will have a beneficial effect. Patients in need of treatment involving modulation of inorganic ion receptors can be identified using standard techniques known to those in the medical profession.

Preferably, a patient is a human having a disease or disorder characterized by one more of the following: (1) abnormal inorganic ion homeostasis, more preferably abnormal calcium homeostasis; (2) an abnormal level of a messenger whose production or secretion is affected by inorganic ion receptor activity, more preferably affected by calcium receptor activity; and (3) an abnormal level or activity of a messenger whose function is affected by inorganic ion receptor activity, more preferably affected by calcium receptor activity.

Diseases characterized by abnormal calcium homeostasis include hyperparathyroidism, osteoporosis and other bone and mineral-related disorders, and the like (as described, *e.g.*, in standard medical text books, such as "Harrison's Principles of Internal Medicine"). Such diseases are treated using calcium receptor-modulating compounds which mimic or block one or more of the effects of extracellular Ca^{2+} on a calcium receptor and, thereby, directly or indirectly affect the levels of proteins or other compounds in the body of the patient.

By "therapeutically effective amount" is meant an amount of a compound which relieves to some extent one or more symptoms of the disease or disorder in the patient; or returns to normal either partially or completely one or more physiological or biochemical parameters associated with or causative of the disease or disorder.

In a preferred embodiment, the patient has a disease or disorder characterized by an abnormal level of one or more calcium receptor-regulated components and the compound is active on a calcium receptor of a cell selected from the group consisting of: parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, central nervous system cell, peripheral nervous system cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cell), intestinal cell, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin-secreting cell, glucagon-secreting cell, kidney mesangial cell, mammary cell, beta cell, fat/adipose cell, immune cell, GI tract cell, skin cell, adrenal cell, pituitary cell, hypothalamic cell and cell of the subfornical organ.

More preferably, the cells are chosen from the group consisting of: parathyroid cell, central nervous system cell, peripheral nervous system cell, cell of the thick ascending limb of Henle's loop and/or collecting duct in the kidney, parafollicular cell in the thyroid (C-cell),

intestinal cell, GI tract cell, pituitary cell, hypothalamic cell and cell of the subfornical organ.

In a preferred embodiment, the compound is a calcimimetic acting on a parathyroid cell calcium receptor and reduces the level of parathyroid hormone in the serum of the patient. More preferably, the level is reduced to a degree sufficient to cause a decrease in plasma Ca^{2+}. Most preferably, the parathyroid hormone level is reduced to that present in a normal individual.

In another preferred embodiment, the compound is a calcilytic acting on a parathyroid cell calcium receptor and increases the level of parathyroid hormone in the serum of the patient. More preferably, the level is increased to a degree sufficient to cause an increase in bone mineral density of a patient.

Patients in need of such treatments can be identified by standard medical techniques, such as blood or urine analysis. For example, by detecting a deficiency of protein whose production or secretion is affected by changes in inorganic ion concentrations, or by detecting abnormal levels of inorganic ions or hormones which effect inorganic ion homeostasis.

Various examples are used throughout the application. These examples are not intended in any way to limit the invention.

Other features and advantages of the invention will be apparent from the following figures, detailed description of the invention, examples, and the claims.

Brief Description of the Drawings

Figs. 1a-1r, show the chemical structures of different compounds.

Figs. 2-131 provided physical data for representative compounds herein described.

Description of the Preferred Embodiments

The present invention features compounds able to modulate one or more inorganic ion receptor activities, preferably the compound can mimic or block an effect of an extracellular ion on a cell having an inorganic ion receptor, more preferably the extracellular ion is Ca^{2+} and the effect is on a cell having a calcium receptor. Publications concerned with the calcium activity, calcium receptor and/or calcium receptor modulating compounds include the following: Brown *et al.*, Nature 366: 574, 1993; Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959; Nemeth *et al.*, PCT/US92/07175, International Publication Number WO 93/04373; Shoback and Chen, J. Bone Mineral Res. 9: 293 (1994); and Racke *et al.*, FEBS Lett. 333: 132, (1993). These publications are not admitted to be prior art to the claimed invention.

I. Calcium Receptors

Calcium receptors are present on different cell types and can have different activities in different cell types. The pharmacological effects of the following cells, in response to calcium, is consistent with the presence of a calcium receptor: parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, central nervous system cell, peripheral nervous system cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cell), intestinal cell, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin-secreting cell, glucagon-secreting cell, kidney mesangial cell, mammary cell, beta cell, fat/adipose cell, immune cell, GI tract cell, skin cell, adrenal cell, pituitary cell, hypothalamic cell and cell of the subfornical organ. In addition, the presence of calcium receptors on parathyroid cell, central nervous system cell, peripheral nervous system

cell, cell of the thick ascending limb of Henle's loop and/or collecting duct in the kidney, parafollicular cell in the thyroid (C-cell), intestinal cell, GI tract cell, pituitary cell, hypothalamic cell and cell of the subfornical organ, has been confirmed by physical data.

The calcium receptor on these different cell types may be different. It is also possible that a cell can have more than one type of calcium receptor. Comparison of calcium receptor activities and amino acid sequences from different cells indicate that distinct calcium receptor types exist. For example, calcium receptors can respond to a variety of di- and trivalent cations. The parathyroid calcium receptor responds to calcium and Gd^{3+}, while osteoclasts respond to divalent cations such as calcium, but do not respond to Gd^{3+}. Thus, the parathyroid calcium receptor is pharmacologically distinct from the calcium receptor on the osteoclast.

On the other hand, the nucleic acid sequences encoding calcium receptors present in parathyroid cells and C-cells indicate that these receptors have a very similar amino acid structure. Nevertheless, calcimimetic compounds exhibit differential pharmacology and regulate different activities at parathyroid cells and C-cells. Thus, pharmacological properties of calcium receptors may vary significantly depending upon the cell type or organ in which they are expressed even though the calcium receptors may have similar or even identical structures.

Calcium receptors, in general, have a low affinity for extracellular Ca^{2+} (apparent K_d generally greater than about 0.5 mM). Calcium receptors may include a free or bound effector mechanism as defined by Cooper, Bloom and Roth, "The Biochemical Basis of Neuropharmacology", Ch. 4, and are thus distinct from intracellular calcium receptors, *e.g.*, calmodulin and the troponins.

Calcium receptors respond to changes in extracellular calcium levels. The exact changes depend on the particular receptor and cell line containing the receptor. For

example, the *in vitro* effect of calcium on the calcium receptor in a parathyroid cell includes the following:

1. An increase in internal calcium. The increase is due to the influx of external calcium and/or to mobilization of internal calcium. Characteristics of the increase in internal calcium include the following:

(a) A rapid (time to peak < 5 seconds) and transient increase in $[Ca^{2+}]_i$ that is refractory to inhibition by 1 μM La^{3+} or 1 μM Gd^{3+} and is abolished by pretreatment with ionomycin (in the absence of extracellular Ca^{2+});

(b) The increase is not inhibited by dihydropyridines;

(c) The transient increase is abolished by pretreatment for 10 minutes with 10 mM sodium fluoride;

(d) The transient increase is diminished by pretreatment with an activator of protein kinase C (PKC), such as phorbol myristate acetate (PMA), mezerein or (-)-indolactam V. The overall effect of the protein kinase C activator is to shift the concentration-response curve of calcium to the right without affecting the maximal response; and

(e) Pretreatment with pertussis toxin (100 ng/ml for > 4 hours) does not affect the increase.

2. A rapid (< 30 seconds) increase in the formation of inositol-1,4,5-triphosphate or diacylglycerol. Pretreatment with pertussis toxin (100 ng/ml for > 4 hours) does not affect this increase;

3. The inhibition of dopamine- and isoproterenol-stimulated cyclic AMP formation. This effect is blocked by pretreatment with pertussis toxin (100 ng/ml for > 4 hours); and

4. The inhibition of PTH secretion. Pretreatment with pertussis toxin (100 ng/ml for > 4 hours) does not affect the inhibition in PTH secretion.

Using techniques known in the art, the effect of calcium on other calcium receptors in different cells can

be readily determined. Such effects may be similar in regard to the increase in internal calcium observed in parathyroid cells. However, the effect is expected to differ in other aspects, such as causing or inhibiting the release of a hormone other than parathyroid hormone.

II. Inorganic Ion Receptor Modulating Compounds

Inorganic ion receptor modulating compounds modulate one or more inorganic ion receptor activities. Preferred calcium receptor modulating compounds are calcimimetics and calcilytics. Inorganic ion receptor modulating compounds can be identified by screening compounds which are modelled after a compound shown to have a particular activity (*i.e.*, a lead compound).

A preferred method of measuring calcium receptor activity is to measure changes in $[Ca^{2+}]_i$. Changes in $[Ca^{2+}]_i$ can be measured using different techniques such by using HEK 293 cells transduced with nucleic acid expressing the human parathyroid calcium receptor and loaded with fura-2; and by measuring an increase in Cl^- current in a *Xenopus* oocyte injected with nucleic acid coding for a calcium receptor. (*See* Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.) For example, poly(A)$^+$ mRNA can be obtained from cells expressing a calcium receptor, such as a parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epidermis, parafollicular cell in the thyroid (C-cell), intestinal cell, central nervous cell, peripheral nervous system cell, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin-secreting cell, glucagon-secreting cell, kidney mesangial cell, mammary cell, beta cell, fat/adipose cell, immune cell, and GI tract cell. Preferably, the nucleic acid is from a parathyroid cell, C-cell, or osteoclast. More preferably,

the nucleic acid encodes a calcium receptor and is present on a plasmid or vector.

In preferred embodiments the calcium receptor modulating compound is a calcimimetic which inhibits bone resorption *in vivo* by an osteoclast; inhibits bone resorption *in vitro* by an osteoclast; stimulates calcitonin secretion *in vitro* or *in vivo* from a c-cell; inhibits parathyroid hormone secretion from a parathyroid cell *in vitro* and decreases PTH secretion *in vivo*; elevates calcitonin levels *in vivo*; or blocks osteoclastic bone resorption *in vitro* and inhibits bone resorption *in vivo*.

In another preferred embodiment the calcium receptor modulating compound is a calcilytic which evokes the secretion of parathyroid hormone from parathyroid cells *in vitro* and elevates the level of parathyroid hormone *in vivo*.

Preferably, the compound selectively targets inorganic ion receptor activity, more preferably calcium receptor activity, in a particular cell. By "selectively" is meant that the compound exerts a greater effect on inorganic ion receptor activity in one cell type than at another cell type for a given concentration of compound. Preferably, the differential effect is 10-fold or greater. Preferably, the concentration refers to blood plasma concentration and the measured effect is the production of extracellular messengers such as plasma calcitonin, parathyroid hormone or plasma calcium. For example, in a preferred embodiment, the compound selectively targets PTH secretion over calcitonin secretion.

In another preferred embodiment, the compound has an EC_{50} or IC_{50} less than or equal to 5 μM at one or more, but not all cells chosen from the group consisting of: parathyroid cell, bone osteoclast, juxtaglomerular kidney cell, proximal tubule kidney cell, distal tubule kidney cell, central nervous system cell, peripheral nervous system cell, cell of the thick ascending limb of Henle's loop and/or collecting duct, keratinocyte in the epi-

dermis, parafollicular cell in the thyroid (C-cell), intestinal cell, platelet, vascular smooth muscle cell, cardiac atrial cell, gastrin-secreting cell, glucagon-secreting cell, kidney mesangial cell, mammary cell, beta cell, fat/adipose cell, immune cell, GI tract cell, skin cell, adrenal cell, pituitary cell, hypothalamic cell and cell of the subfornical organ. More preferably, the cells are chosen from the group consisting of parathyroid cell, central nervous system cell, peripheral nervous system cell, cell of the thick ascending limb of Henle's loop and/or collecting duct in the kidney, parafollicular cell in the thyroid (C-cell), intestinal cell, GI tract cell, pituitary cell, hypothalamic cell and cell of the subfornical organ. The presence of a calcium receptor in this group of cells has been confirmed by physical data such as *in situ* hybridization and antibody staining.

Preferably, inorganic ion receptor modulating compounds mimic or block the effects of an extracellular ion on a cell having an inorganic ion receptor, such that the compounds achieve a therapeutic effect. Inorganic ion receptor modulating compounds may have the same, or different, effects on cells having different types of inorganic ion receptor morphology (*e.g.*, such as cells having normal inorganic ion receptors, a normal number of inorganic ion receptor, an abnormal inorganic ion receptor, and an abnormal number of inorganic ion receptors).

Calcium receptor modulating compounds preferably mimic or block all of the effects of extracellular ion in a cell having a calcium receptor. However, calcimimetics need not possess all the biological activities of extracellular Ca^{2+}. Similarly, calcilytics need not block all of the activities caused by extracellular calcium. Additionally, different calcimimetics and different calcilytics do not need to bind to the same site on the calcium receptor as does extracellular Ca^{2+} to exert their effects.

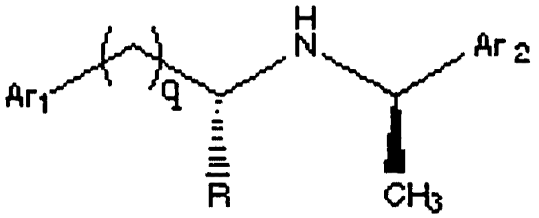
Inorganic modulating compounds need not effect inorganic receptor activity to the same extent or in exactly

the same manner as the natural ligand. For example, a calcimimetic may effect calcium receptor activity to a different extent, to a different duration, by binding to a different binding site, or by having a different affinity, compared to calcium acting at a calcium receptor.

A. Calcimimetics

1. Structure I Compounds

Structure I compounds able to modulate calcium receptor activity have the following formula:



where, Ar_1 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, $N(CH_3)_2$, phenyl, phenoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, acetyl, ethylene dioxy, preferably each substituent is independently selected from the group consisting of, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, I, CF_3, CHF_2, CH_2F, CF_3O, CF_3CH_2O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, and acetoxy. More preferably, Ar_1 is either a naphthyl or a phenyl having 1-5 substituents each independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, I, Cl, F, CF_3, and CH_3, more preferably CF_3O, I, Cl, F, and CF_3;

Ar_2 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl,

halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, and acetoxy, preferably each substituent is independently selected from the group consisting of, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, I, CF_3, CHF_2, CH_2F, CF_3O, CF_3CH_2O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, and acetoxy. More preferably, Ar_2 is either a naphthyl or a phenyl having 1-5 substituents each independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, I, Cl, F, CF_3, and CH_3, more preferably CF_3O, I, Cl, F, CH_3O, and CF_3.

q is 0, 1, 2, or 3; and

R is either H, or CH_3;

and pharmaceutically salts and complexes thereof.

"Lower alkyl" refers to a saturated hydrocarbon having 1-4 carbons, preferably 1-3 carbon atoms, which may be straight chain or branched.

"Lower alkoxy" refers to "O-lower alkyl". Where "O" is an oxygen joined to a lower alkyl.

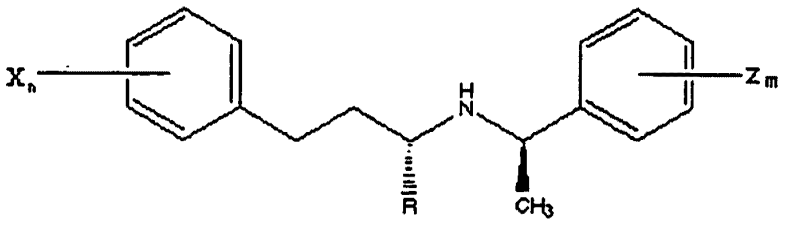
"Lower thioalkyl" refers to "S-lower alkyl". Where "S" is a sulfur joined to a lower alkyl.

"Lower haloalkyl" refers to a lower alkyl substituted with at least one halogen. Preferably, only the terminal carbon of the lower haloalkyl is substituted with a halogen and 1 to 3 halogens are present. More preferably, the lower haloalkyl contains 1 carbon. Preferably, the halogen substitutions are either Cl or F.

"Lower haloalkoxy" refers to "O-lower haloalkyl". Where "O" is an oxygen joined to a lower haloalkyl.

a. <u>Ar_1 and Ar_2 are Both Optionally Substituted Phenyls</u>

In a preferred embodiment both Ar_1 and Ar_2 are optionally substituted phenyls and the compound has following formula:



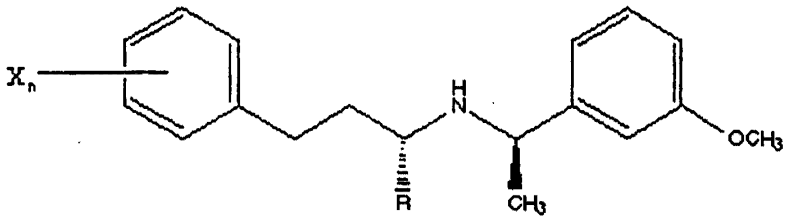
where R is hydrogen or methyl

m and n are each independently 0, 1, 2, 3, 4, or 5;

each X is independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, $N(CH_3)_2$, phenyl, phenoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, acetyl, ethylene dioxy. Preferably each X is independently selected from the group consisting of, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, I, CF_3, CHF_2, CH_2F, CF_3O, CF_3CH_2O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, *t*-butyl, and acetoxy. More preferably, each X is independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3O, I, Cl, F, CF_3, and CH_3, more preferably CF_3O, I, Cl, F, and CF_3;

each Z is independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, and acetoxy. Preferably each Z is independently selected from the group consisting of, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, I, CF_3, CHF_2, CH_2F, CF_3O, CF_3CH_2O, CH_3S, OH, CH_2OH, $CONH_2$, CN, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, *t*-butyl, and acetoxy. More preferably, each Z is independently selected from the group consisting of, isopropyl, CH_3O, CH_3S, CF_3O, CF_3, I, Cl, F, and CH_3.

In a more preferred embodiment, at least one of the Z substituents is in the *meta* position. More preferably, the compound has the following formula:

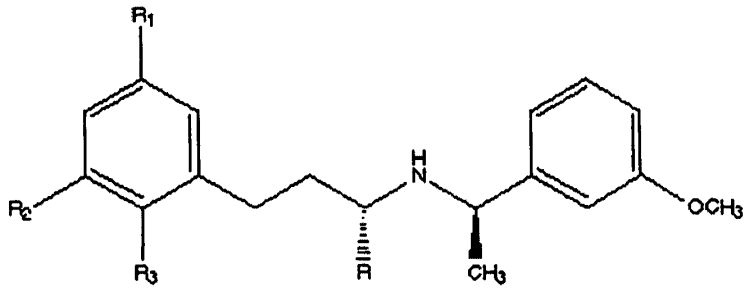


where R is either hydrogen or methyl;

m is 0, 1, 2, 3, 4, or 5, preferably 1 or 2;

and each X is independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, $N(CH_3)_2$, phenyl, phenoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, acetyl, ethylene dioxy, preferably each substituent is independently selected from the group consisting of, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, I, CF_3, CHF_2, CH_2F, CF_3O, CF_3CH_2O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, and acetoxy, more preferably, isopropyl, CH_3O, CH_3S, CF_3O, CF_3, I, Cl, F, and CH_3.

More preferably, the compound has the formula:



where R is either hydrogen or methyl;

R_1 is either halogen or hydrogen, preferably R_1 is either F, or hydrogen;

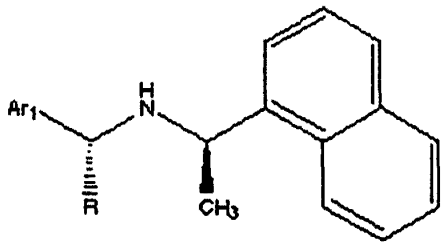
R_2 is either hydrogen, halogen, lower alkyl, lower haloalkyl, or lower haloalkoxy, preferably, R_2 is either hydrogen, CF_3, CH_3, OCF_3, or F, and

R_3 is either hydrogen, halogen, or alkoxy, preferably, R_3 is either Cl, F, hydrogen, or methoxy, more preferably methoxy.

In alternative more preferred combinations; at least two of R_1, R_2, and R_3 is halogen, preferably F and R is hydrogen or CH_3; R is hydrogen or CH_3, R_2 is either lower haloalkyl, or lower haloalkoxy, preferably OCF_3 or CF_3, and R_1 and R_3 is hydrogen; and R is CH_3, R_3 is halogen, preferably Cl, R_1 is either halogen or hydrogen, preferably F or hydrogen, and R_2 is either hydrogen, lower alkyl, lower haloalkyl, or lower haloalkoxy, preferably, hydrogen, CF_3, CH_3, OCF_3, or F.

b. Ar_2 is Naphthyl and q is 0

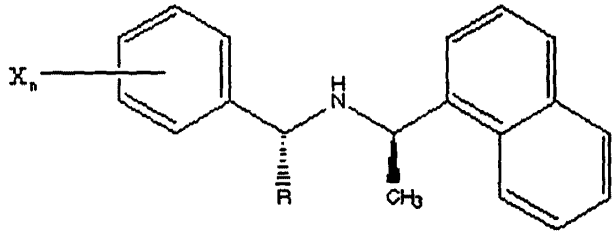
In another preferred embodiment, Ar_2 is naphthyl, q is 0, and the compound has the formula:



where Ar_1 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, $N(CH_3)_2$, phenyl, phenoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, acetyl, ethylene dioxy, preferably each substituent is independently selected from the group consisting of, CH_3, CH_3O, CH_3CH_2O,

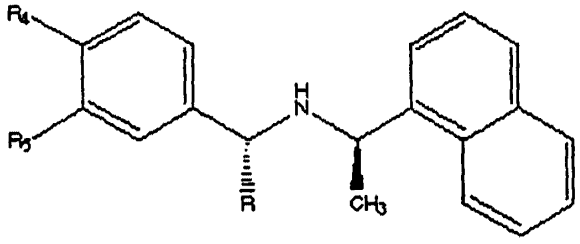
methylene dioxy, Br, Cl, F, I, CF_3, CHF_2, CH_2F, CF_3O, CF_3CH_2O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, and acetoxy. More preferably, Ar_1 is either a naphthyl or a phenyl having 1-5 substituents each independently selected from the group consisting of isopropyl, CH_3O, CH_3S, CF_3, CF_3O I, Cl, F, and CH_3

More preferably, Ar_1 is an optional substituted phenyl where the compound has the formula:



where X_n represents the optional substituents for the optionally substituted phenyl as described above (with the preferred substituents and number of substituents as described above).

Even more preferably the compound has the formula:



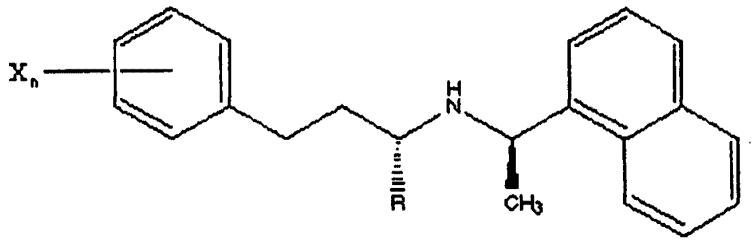
where R is either CH_3 or hydrogen;

R_4 is either lower alkyl, halogen, or alkoxy, preferably isopropyl, chlorine, or methoxy; and

R_5 is either hydrogen, lower alkyl, or halogen, preferably methyl, CH_3, Br, or Cl.

c. Ar_2 is Naphthyl and q is 2

In another preferred embodiment, Ar_1 is a substituted phenyl, Ar_2 is naphthyl, q is 2 and the compound has the formula:

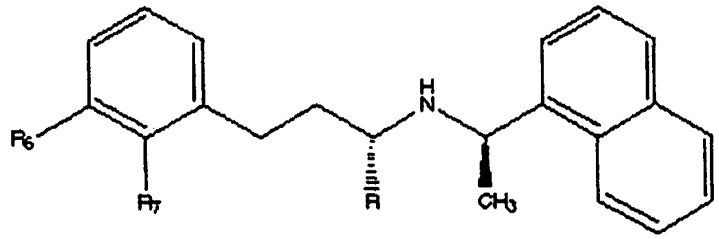


where R is either hydrogen or CH_3;

n is 0, 1, 2, 3, 4, or 5, preferably 1 or 2; and

each X is independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, $N(CH_3)_2$, phenyl, phenoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, acetyl, ethylene dioxy, preferably each substituent is independently selected from the group consisting of, CH_3, CH_3O, CH_3CH_2O, methylene dioxy, Br, Cl, F, I, CF_3, CHF_2, CH_2F, CF_3O, CF_3CH_2O, CH_3S, OH, CH_2OH, $CONH_2$, CN, NO_2, CH_3CH_2, propyl, isopropyl, butyl, isobutyl, t-butyl, and acetoxy, more preferably, isopropyl, CH_3O, CH_3S, CF_3O, CF_3, I, Cl, F, and CH_3.

More preferably, the compound has the formula:



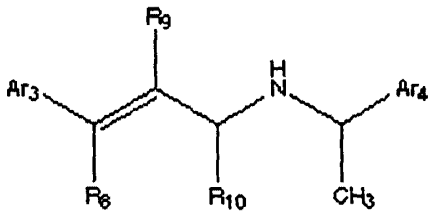
where R_6 is either is either hydrogen, lower haloalkyl, or lower haloalkoxy, preferably hydrogen, OCF_3 or CF_3; and

R_7 is either halogen or hydrogen, preferably chlorine or hydrogen.

In other embodiments R, R_6 and R_7 are as described above (with the preferred substituents as described above), provided that when both R and R_6 are hydrogen, R_7 is not Cl; and R is CH_3, and R_6 and R_7 is as described above (with the preferred substituents as described above).

2. Structure II Compounds

Structure II compounds have the formula:



where Ar_3 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, $N(CH_3)_2$, acetyl, ethylene dioxy, preferably $N(CH_3)_2$, lower alkoxy, or lower alkyl;

Ar_4 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, and acetoxy, preferably lower alkoxy, more preferably methoxy;

R_8 is either hydrogen or phenyl, preferably hydrogen;

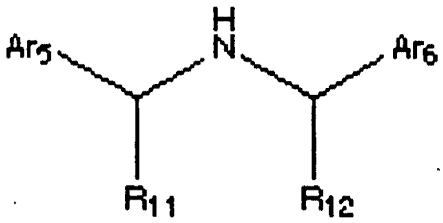
R_9 is either hydrogen or methyl; and

R_{10} is either hydrogen, methyl, or phenyl, more preferably when R_{10} is methyl the chiral carbon it is attached to is the (*R*) stereoisomer.

Preferably, the α-methyl in Structure II is an (*R*)-α-methyl.

3. Structure III Compounds

Structure III compounds have the formula:



where Ar_5 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, acetoxy, benzyl, benzyloxy, α,α-dimethylbenzyl, NO_2, CHO, $CH_3CH(OH)$, acetyl, ethylene dioxy, -CH=CH-phenyl, preferably, lower alkyl, phenoxy, -CH=CH-phenyl, dimethylbenzyl, methoxy, methylene, or ethylene;

Ar_6 is either naphthyl or phenyl optionally substituted with 0 to 5 substituents each independently selected from the group consisting of, acetyl, lower alkyl, halogen, lower alkoxy, lower thioalkyl, methylene dioxy, lower haloalkyl, lower haloalkoxy, OH, CH_2OH, $CONH_2$, CN, carbomethoxy, $OCH_2C(O)C_2H_5$ and acetoxy, preferably methoxy, lower alkyl, phenyl, halogen, CF_3, CN, carbomethoxy or, $OCH_2C(O)C_2H_5$;

R_{11} is hydrogen or methyl, preferably when R_{11} is methyl the carbon to which it is attached is an (*R*) stereoisomer; and

R_{12} is hydrogen or methyl, preferably when R_{12} is methyl the carbon to which it is attached is an (*R*) stereoisomer.

4. Calcimimetic Activity

The ability of compounds to mimic the activity of Ca^{2+} at calcium receptors can be determined using procedures known in the art and described by Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959. For example, calcimimetics possess one or more and preferably all of the following activities when tested on parathyroid cells *in vitro*:

1. The compound causes a rapid (time to peak < 5 seconds) and transient increase in intracellular calcium concentration that is refractory to inhibition by 1 μM La^{3+} or 1 μM Gd^{3+}. The increase in $[Ca^{2+}]_i$ persists in the absence of extracellular Ca^{2+}, but is abolished by pre-treatment with ionomycin (in the absence of extracellular Ca^{2+});

2. The compound potentiates increases in $[Ca^{2+}]_i$ elicited by submaximal concentrations of extracellular Ca^{2+};

3. The increase in $[Ca^{2+}]_i$ elicited by extracellular Ca^{2+} is not inhibited by dihydropyridines;

4. The transient increase in $[Ca^{2+}]_i$ caused by the compound is abolished by pretreatment for 10 minutes with 10 mM sodium fluoride;

5. The transient increase in $[Ca^{2+}]_i$ caused by the compound is diminished by pretreatment with an activator of protein kinase C (PKC), such as phorbol myristate acetate (PMA), mezerein or (-)-indolactam V. The overall effect of the protein kinase C activator is to shift the concentration-response curve of the compound to the right without affecting the maximal response;

6. The compound causes a rapid (< 30 seconds) increase in the formation of inositol-1,4,5-triphosphate and/or diacylglycerol;

7. The compound inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation;

8. The compound inhibits PTH secretion;

9. Pretreatment with pertussis toxin (100 ng/ml for > 4 hours) blocks the inhibitory effect of the compound on cyclic AMP formation, but does not effect increases in $[Ca^{2+}]_i$, inositol-1,4,5-triphosphate, or diacylglycerol, nor decreases in PTH secretion;

10. The compound elicits increases in Cl^- current in *Xenopus* oocytes injected with poly(A)$^+$-enriched mRNA from bovine or human parathyroid cells, but is without effect in *Xenopus* oocytes injected with water, or liver mRNA; and

11. Similarly, using a cloned calcium receptor from a parathyroid cell, the compound will elicit a response in *Xenopus* oocytes injected with the specific cDNA or mRNA encoding the receptor.

Different calcium activities can be measured using available techniques. (*See*, Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.) Parallel definitions of compounds mimicking Ca^{2+} activity on other calcium responsive cell, preferably at a calcium receptor, are evident from the examples provided herein and Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.

Preferably, the compound as measured by the bioassays described herein, or by Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959, has one or more, more preferably all of the following activities: evokes a transient increase in internal calcium, having a duration of less that 30 seconds (preferably by mobilizing internal calcium); evokes a rapid increase in $[Ca^{2+}]_i$, occurring within thirty seconds; evokes a sustained increase (greater than thirty seconds) in $[Ca^{2+}]_i$ (preferably by causing an influx of external calcium); evokes an increase in inositol-1,4,5-triphosphate or diacylglycerol levels, preferably within less than 60 seconds; and

inhibits dopamine- or isoproterenol-stimulated cyclic AMP formation.

The transient increase in $[Ca^{2+}]_i$ is preferably abolished by pretreatment of the cell for ten minutes with 10 mM sodium fluoride, or the transient increase is diminished by brief pretreatment (not more than ten minutes) of the cell with an activator of protein kinase C, preferably, phorbol myristate acetate (PMA), mezerein or (-) indolactam V.

C. Calcilytics

The ability of a compound to block the activity of extracellular calcium at a calcium receptor can be determined using standard techniques based on the present disclosure. (*See*, also Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.) For example, compounds which block the effect of extracellular calcium, when used in reference to a parathyroid cell, possess one or more, and preferably all of the following characteristics when tested on parathyroid cells *in vitro*:

1. The compound blocks, either partially or completely, the ability of increased concentrations of extracellular Ca^{2+} to:

(a) increase $[Ca^{2+}]_i$,

(b) mobilize intracellular Ca^{2+},

(c) increase the formation of inositol-1,4,5-triphosphate,

(d) decrease dopamine- or isoproterenol-stimulated cyclic AMP formation, and

(e) inhibit PTH secretion;

2. The compound blocks increases in Cl^- current in *Xenopus* oocytes injected with poly(A)$^+$-mRNA from bovine or human parathyroid cells elicited by extracellular Ca^{2+} or calcimimetic compounds, but not in *Xenopus* oocytes injected with water or liver mRNA;

3. Similarly, using a cloned calcium receptor from a parathyroid cell, the compound will block a response in

Xenopus oocytes injected with the specific cDNA, mRNA or cRNA encoding the calcium receptor, elicited by extracellular Ca^{2+} or a calcimimetic compound.

Parallel definitions of compounds blocking Ca^{2+} activity on a calcium responsive cell, preferably at a calcium receptor, are evident from the examples provided herein and Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959.

III. TREATMENT OF DISEASES OR DISORDERS

Diseases or disorders which can be treated by modulating calcium receptor activity are known in the art. For example, diseases or disorders which can be treated by modulating calcium receptor activity can be identified based on the functional responses of cells regulated by calcium receptor activity. Functional responses of cells regulated by calcium receptor are know in the art, including PTH secretion by parathyroid cells, calcitonin secretion by C-cells, and bone resorption by osteoclasts.

Such functional responses are associated with different diseases or disorders. For example, hyperparathyroidism results in elevated levels of PTH in the plasma. Decreasing the plasma levels of PTH offers an effective means of treating hyperparathyroidism. Likewise, increasing plasma levels of calcitonin is associated with an inhibition of bone resorption. Inhibiting bone resorption is an effective treatment for osteoporosis. Thus, modulation of calcium receptor activity can be used to treat diseases such as hyperparathyroidism, and osteoporosis.

Those compounds modulating inorganic ion receptor activity, preferably calcium receptor activity, can be used to confer beneficial effects to patients suffering from a variety of diseases or disorders. For example, osteoporosis is an age-related disorder characterized by loss of bone mass and increased risk of bone fracture. Compounds can be used to block osteoclastic bone resorption either directly (*e.g.*, an osteoclast ionomimetic

compound) or indirectly by increasing endogenous calcitonin levels (*e.g.*, a C-cell calcimimetic). Alternatively, a calcilytic active on the parathyroid cell calcium receptor will increase circulating levels of parathyroid hormone, stimulating bone formation. All three of these approaches will result in beneficial effects to patients suffering from osteoporosis.

In addition, it is known that intermittent low dosing with PTH results in an anabolic effect on bone mass and appropriate bone remodeling. Thus, compounds and dosing regimens evoking transient increases in parathyroid hormone (*e.g.*, intermittent dosing with a parathyroid cell ionolytic) can increase bone mass in patients suffering from osteoporosis.

Additional diseases or disorders can be identified by identifying additional cellular functional responses, associated with a disease or disorder, which are regulated by calcium receptor activity. Diseases or disorder which can be treated by modulating other inorganic ion receptors can be identified in an analogous manner.

The inorganic ion receptor-modulating compounds of the present invention can exert an affect at an inorganic ion receptor causing one or more cellular effects ultimately producing a therapeutic effect. Calcium receptor-modulating compounds of the present invention can exert an effect on calcium receptor causing one or more cellular effects ultimately producing a therapeutic effect. Different diseases can be treated by the present invention by targeting cells having a calcium receptor.

For example, primary hyperparathyroidism (HPT) is characterized by hypercalcemia and abnormal elevated levels of circulating PTH. A defect associated with the major type of HPT is a diminished sensitivity of parathyroid cells to negative feedback regulation by extracellular Ca^{2+}. Thus, in tissue from patients with primary HPT, the "set-point" for extracellular Ca^{2+} is shifted to the right so that higher than normal concentrations of

extracellular Ca^{2+} are required to depress PTH secretion. Moreover, in primary HPT, even high concentrations of extracellular Ca^{2+} often depress PTH secretion only partially. In secondary (uremic) HPT, a similar increase in the set-point for extracellular Ca^{2+} is observed even though the degree to which Ca^{2+} suppresses PTH secretion is normal. The changes in PTH secretion are paralleled by changes in $[Ca^{2+}]_i$: the set-point for extracellular Ca^{2+}-induced increases in $[Ca^{2+}]_i$ is shifted to the right and the magnitude of such increases is reduced.

Patients suffering from secondary HPT may also have renal osteodystrophy. Calcimimetics appear to be useful for treating both abnormal PTH secretion and osteodystrophy in such patients.

Compounds that mimic the action of extracellular Ca^{2+} are beneficial in the long-term management of both primary and secondary HPT. Such compounds provide the added impetus required to suppress PTH secretion which the hypercalcemic condition alone cannot achieve and, thereby, help to relieve the hypercalcemic condition. Compounds with greater efficacy than extracellular Ca^{2+} may overcome the apparent nonsuppressible component of PTH secretion which is particularly troublesome in the major form of primary HPT caused by adenoma of the parathyroid gland. Alternatively or additionally, such compounds can depress synthesis of PTH, as prolonged hypercalcemia has been shown to depress the levels of preproPTH mRNA in bovine and human adenomatous parathyroid tissue. Prolonged hypercalcemia also depresses parathyroid cell proliferation *in vitro*, so calcimimetics can also be effective in limiting the parathyroid cell hyperplasia characteristic of secondary HPT.

Cells other than parathyroid cells can respond directly to physiological changes in the concentration of extracellular Ca^{2+}. For example, calcitonin secretion from parafollicular cells in the thyroid (C-cells) is regulated by changes in the concentration of extracellular Ca^{2+}.

Isolated osteoclasts respond to increases in the concentration of extracellular Ca^{2+} with corresponding increases in $[Ca^{2+}]_i$ that arise partly from the mobilization of intracellular Ca^{2+}. Increases in $[Ca^{2+}]_i$ in osteoclasts are associated with the inhibition of bone resorption. Release of alkaline phosphatase from bone-forming osteoblasts is directly stimulated by calcium.

Renin secretion from juxtaglomerular cells in the kidney, like PTH secretion, is depressed by increased concentrations of extracellular Ca^{2+}. Extracellular Ca^{2+} causes the mobilization of intracellular Ca^{2+} in these cells. Other kidney cells respond to calcium as follows: elevated Ca^{2+} inhibits formation of $1,25(OH)_2$-vitamin D by proximal tubule cells, stimulates production of calcium-binding protein in distal tubule cells, and inhibits tubular reabsorption of Ca^{2+} and Mg^{2+} and the action of vasopressin on the thick ascending limb of Henle's loop (MTAL), reduces vasopressin action in the cortical collecting duct cells, and affects vascular smooth muscle cells in blood vessels of the renal glomerulus.

Calcium also promotes the differentiation of intestinal goblet cells, mammary cells, and skin cells; inhibits atrial natriuretic peptide secretion from cardiac atria; reduces cAMP accumulation in platelets; alters gastrin and glucagon secretion; acts on vascular smooth muscle cells to modify cell secretion of vasoactive factors; and affects cells of the central nervous system and peripheral nervous system.

Thus, there are sufficient indications to suggest that Ca^{2+}, in addition to its ubiquitous role as an intracellular signal, also functions as an extracellular signal to regulate the responses of certain specialized cells. Compounds of this invention can be used in the treatment of diseases or disorders associated with disrupted Ca^{2+} responses in these cells.

Specific diseases and disorders which might be treated or prevented, based upon the affected cells, also

include those of the central nervous system such as seizures, stroke, head trauma, spinal cord injury, hypoxia-induced nerve cell damage such as in cardiac arrest or neonatal distress, epilepsy, neurodegenerative diseases such as Alzheimer's disease, Huntington's disease and Parkinson's disease, dementia, muscle tension, depression, anxiety, panic disorder, obsessive-compulsive disorder, post-traumatic stress disorder, schizophrenia, neuroleptic malignant syndrome, and Tourette's syndrome; diseases involving excess water reabsorption by the kidney such as syndrome of inappropriate ADH secretion (SIADH), cirrhosis, congestive heart failure, and nephrosis; hypertension; preventing and/or decreasing renal toxicity from cationic antibiotics (*e.g.*, aminoglycoside antibiotics); gut motility disorders such as diarrhea, and spastic colon; GI ulcer diseases; GI diseases with excessive calcium absorption such as sarcoidosis; and autoimmune diseases and organ transplant rejection.

While calcium receptor-modulating compounds of the present invention will typically be used in therapy for human patients, they may also be used to treat similar or identical diseases in other warm-blooded animal species such as other primates, farm animals such as swine, cattle, and poultry; and sports animals and pets such as horses, dogs and cats.

IV. Administration

The different compounds described by the present invention can be used to treat different diseases or disorders by modulating inorganic ion receptor activity, preferably calcium receptor activity. The compounds of the invention can be formulated for a variety of modes of administration, including systemic and topical or localized administration. Techniques and formulations generally may be found in Remington's Pharmaceutical Sciences, Mack Publishing Co., Easton, PA. Administration of ionomimetics and ionolytics is discussed by Nemeth *et al.*,

PCT/US93/01642, International Publication Number WO 94/18959.

Suitable dosage forms, in part, depend upon the use or the route of entry, for example oral, transdermal, or by injection. Such dosage forms should allow the compound to reach a target cell whether the target cell is present in a multicellular host or in culture. For example, pharmacological compounds or compositions injected into the blood stream should be soluble. Other factors are known in the art, and include considerations such as toxicity and dosage form which retard the compound or composition from exerting its effect.

Compounds can also be formulated as pharmaceutically acceptable salts (*e.g.*, acid addition salts) and complexes thereof. Pharmaceutically acceptable salts are non-toxic salts at the concentration at which they are administered. The preparation of such salts can facilitate the pharmacological use by altering the physical characteristic of the compound without preventing it from exerting its physiological effect. Useful alterations in physical properties include lowering the melting point to facilitate transmucosal administration and increasing the solubility to facilitate administering higher concentrations of the drug.

Pharmaceutically acceptable salts include acid addition salts such as those containing sulfate, hydrochloride, maleate, phosphate, sulfamate, acetate, citrate, lactate, tartrate, methanesulfonate, ethanesulfonate, benzenesulfonate, *p*-toluenesulfonate, cyclohexylsulfamate and quinate. (*See e.g.*, PCT/US92/03736, hereby incorporated by reference herein.) Pharmaceutically acceptable salts can be obtained from acids such as hydrochloric acid, maleic acid, sulfuric acid, phosphoric acid, sulfamic acid, acetic acid, citric acid, lactic acid, tartaric acid, malonic acid, methanesulfonic acid, ethanesulfonic acid, benzenesulfonic acid, *p*-toluenesulfonic acid, cyclohexylsulfamic acid, and quinic acid.

Pharmaceutically acceptable salts can be prepared by standard techniques. For example, the free base form of a compound is dissolved in a suitable solvent, such as an aqueous or aqueous-alcohol solution, containing the appropriate acid and then isolated by evaporating the solution. In another example, a salt is prepared by reacting the free base and acid in an organic solvent.

Carriers or excipients can also be used to facilitate administration of the compound. Examples of carriers and excipients include calcium carbonate, calcium phosphate, various sugars such as lactose, glucose, or sucrose, or types of starch, cellulose derivatives, gelatin, vegetable oils, polyethylene glycols and physiologically compatible solvents. The compositions or pharmaceutical composition can be administered by different routes including intravenously, intraperitoneal, subcutaneous, and intramuscular, orally, topically, or transmucosally.

For systemic administration, oral administration is preferred. Alternatively, injection may be used, *e.g.*, intramuscular, intravenous, intraperitoneal, and subcutaneous. For injection, the compounds of the invention are formulated in liquid solutions, preferably in physiologically compatible buffers such as Hank's solution or Ringer's solution. In addition, the compounds may be formulated in solid form and redissolved or suspended immediately prior to use. Lyophilized forms can also be produced.

Systemic administration can also be by transmucosal or transdermal means, or the compounds can be administered orally. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, bile salts and fusidic acid derivatives. In addition, detergents may be used to facilitate permeation. Transmucosal administration may be through nasal sprays, for example, or using suppositories. For

oral administration, the compounds can be formulated into conventional oral administration dosage forms such as capsules, tablets, and liquid preparations.

For topical administration, the compounds of the invention can be formulated into ointments, salves, gels, or creams, as is generally known in the art.

The amounts of various compounds of this invention to be administered can be determined by standard procedures. Generally, a therapeutically effective amount is between about 1 nmole and 3 μmole of the compound, preferably 0.1 nmole and 1 μmole depending on its EC_{50} or IC_{50} and on the age and size of the patient, and the disease or disorder associated with the patient. Generally, it is an amount between about 0.1 and 50 mg/kg, preferably 0.01 and 20 mg/kg of the animal to be treated.

V. Examples

Examples are provided below illustrating different aspects and embodiments of the present invention. These examples are not intended to limit the claimed invention.

Example 1: Cloning of Human Parathyroid Calcium Receptor From a Human Parathyroid Gland Adenoma Tumor

This example describes the cloning of a human parathyroid calcium receptor from a human parathyroid gland adenoma tumor using pBoPCaR1 as a hybridization probe (*See*, Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959). The probe was used to identify nucleic acid encoding human parathyroid gland calcium receptor by cross-hybridization at reduced stringency.

Messenger RNA was prepared from a human parathyroid gland adenoma tumor removed from a 39-year-old Caucasian male diagnosed with primary hyperparathyroidism. Northern blot analysis of this mRNA using pBoPCaR1 as a hybridization probe identified calcium receptor transcripts of about 5 Kb and about 4 Kb. A cDNA library was constructed

from the mRNA. Double-stranded cDNA larger than 3 Kbp were size-selected on an agarose gel and ligated into the cloning vector lambda ZapII. Five hundred thousand primary recombinant phage were screened with the 5.2 Kbp cDNA insert of pBoPCaR1 as a hybridization probe. The pBoPCaR1 insert was labeled by random-primed synthesis using [^{32}P]-dCTP to a specific activity of 1 x 10^9 cpm/μg.

Library screening was performed at a hybridization stringency of 400 mM Na^+, 50% formamide at a temperature of 38°C. Plaque lift filters were hybridized at a probe concentration of 500,000 cpm/ml for 20 hours. Following hybridization, filters were washed in 1 x SSC at 40°C for 1 hr.

The primary screen identified about 250 positive clones identified by hybridization to pBoPCaR1. Seven of these clones were taken through secondary and tertiary screens to isolate single clones that hybridized to the pBoPCaR1 probe. These seven clones were analyzed by restriction enzyme mapping and Southern blot analysis. Three of the clones contained cDNA inserts of about 5 Kbp and appear to be full-length clones corresponding to the 5 Kb mRNA. Two of the clones contain cDNA inserts of about 4 Kbp and appear to be full-length clones corresponding to the 4 Kb mRNA.

Restriction enzyme mapping of the two different sized inserts indicate that they share regions of sequence similarity in their 5' ends, but diverge in their 3' end sequences. DNA sequence analyses indicate that the smaller insert may result from alternative polyadenylation upstream of the polyadenylation site used in the larger insert.

Representative cDNA inserts for both size classes were subcloned into the plasmid vector pBluescript SK. Linearization followed by *in vitro* transcription using T7 RNA polymerase produced cRNA transcripts. The cRNA transcripts were injected into *Xenopus* oocytes (150 ng/μl RNA; 50 nl/oocyte) for functional analysis. Following

incubation periods of 2-4 days, the oocytes were assayed for the presence of functional calcium receptors. Both clone types gave rise to functional calcium receptors as assessed by the stimulation of calcium-activated chloride currents upon addition of appropriate calcium receptor agonists. Known calcium receptor agonists, including NPS *R*-467 and NPS *R*-568 (*see*, Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959), activated the oocyte-expressed receptor at about the same concentrations known to be effective for the native parathyroid cell receptor. Thus, both clones encode a functional, human parathyroid cell calcium receptor.

Plasmids were prepared by subcloning each size class of insert into pBluescript thereby producing pHuPCaR 5.2 and pHuCaR 4.0. The nucleic acid sequence, and amino acid sequence, of the inserts are shown in SEQ. ID. Nos. 1 and 2.

Several differences were observed between the nucleic acid sequences of the two cDNA inserts. Sequence analyses of the two cDNA inserts indicate the existence of at least two sequence variants differing in the 3' untranslated region and which may result from alternative polyadenylation. In addition, sequence variation exists at the 5' end of the inserts. These distinct sequences correspond to untranslated regions and may have arisen due to alternative transcriptional initiation and/or splicing.

Three additional sites of sequence variation are observed within the coding regions of cDNA clones pHuPCaR5.2 and pHuPCaR4.0 (see SEQ. ID. NOs. 1 and 2) demonstrating that these cDNA clones encode distinct proteins. Sequence analysis of the human CaR gene indicates that the additional 30 base pairs of DNA in cDNA clone pHuPCaR5.2, as compared to the pHuPCaR 4.0 cDNA clone, results from alternative mRNA splicing. The alternative mRNA splicing is predicted to insert 10 additional amino acids into the CaR polypeptide encoded by the pHuPCaR5.2 cDNA at a site between aa#536 and aa#537 in

polypeptide encoded by pHuPCaR4.0 cDNA. In addition, pHuPCaR4.0 encodes glutamine (Gln) at aa#925 and glycine (Gly) at position 990 whereas pHuPCaR5.2 encodes arg (Arg) at both equivalent positions. The human CaR gene encodes for Gln and Arg, respectively, at these positions. The difference between the pHuPCaR4.0 cDNA compared to human DNA appears to represent a true sequence polymorphism within the human population while the single base change in pHuPCaR5.2 probably reflects a mutation which occurred during its cloning. Both cDNAs encode functional calcium receptors as demonstrated by the ability of *Xenopus* oocytes injected with cRNA prepared from these cDNA clones to respond to 10 mM extracellular calcium as ascertained by Cl- conductance. However, it is possible that these two receptor isoforms are functionally and/or pharmacologically distinct.

Example 2: Selection of Stable Recombinant Cells Expressing the Calcium Receptor

Clonal cell lines that stably express the two human and the bovine calcium receptors have been isolated. Calcium receptor cDNAs were subcloned in two different, commercially available expression vectors; pMSG (obtained from Pharmacia) and Cep4B (obtained from Invitrogen). The first vector contains the selectable marker gene for xanthine-guanine phosphoribosyltransferase (gpt) allowing stably transfected cells to overcome the blockade of the purine biosynthetic pathway imposed by addition of 2 μg/ml aminopterin and 25 μg/ml mycophenolic acid. The second vector encodes a gene conferring resistance to the antibiotic hygromycin (used at 200 μg/ml). HuPCaR 5.2 and HuPCaR 4.0 cDNAs (SEQ. ID. NOs. 1 and 2, respectively) were removed from the parent bluescript plasmid with Not I and Hind III restriction enzymes and then either ligated directly into Not I + Hind III digested Cep4B or treated with the klenow fragment of DNA polymerase prior to blunt-end ligation into Sma I digested pMSG.

The pMSG subclone containing the HuPCaR 5.2 insert was transfected into CHO cells as discussed above. Selection has resulted in 20 resistant clones which are being characterized. The Cep4B subclone containing the HuPCaR 5.2 insert was transfected into HEK 293 cells as described above. Selection with hygromycin resulted in a pool of stable clones. Clones expressing the HuPCaR 4.0 receptor isoform were prepared similarly.

Cells obtained from the pool of hygromycin selected HEK 293 cells transfected with Cep4B containing the HuPCaR 5.2 insert were plated on collagen coated Aklar squares which had been placed into individual wells of 12-well tissue culture plates. Two to six days later, medium was removed and the cells washed with balanced salt solution and 1 ml of buffer containing 1 μM fura2-AM, 1 mM $CaCl_2$ and 0.1% BSA and 1 mM $CaCl_2$. Measurements of fluorescence in response to calcium receptor agonists were performed at 37°C in a spectrofluorimeter using excitation and emission wavelengths of 340 and 510 nm, respectively. For signal calibration, Fmax was determined after addition of ionomycin (40 μM) and the apparent Fmin was determined by addition of 0.3 M EGTA, 2.5 M Tris-HCl; pH 10. Robust increases in $[Ca^{2+}]_i$ were observed in response to the addition of the following calcium receptor agonists: Ca^{2+} (10 mM), Mg^{2+} (20 mM) and NPS *R*-467. Control cells expressing functional substance K receptors did not respond to these calcimimetic compounds.

Additional clonal isolates of HEK 293 cells transfected with pHuPCaR4.0 sequence were obtained. These were tested for responsiveness to calcimimetics as described above except that the cells were tested while in suspension.

Example 3: Using Fura-2 Loaded Parathyroid cells To Measure to Calcium Receptor Activity

This section describes procedures used to obtain parathyroid cells from calves and humans, and to use the parathyroid cells to measure calcium receptor activity.

Parathyroid glands were obtained from freshly slaughtered calves (12-15 weeks old) at a local abattoir and transported to the laboratory in ice-cold parathyroid cell buffer (PCB) which contains (mM): NaCl, 126; KCl, 4; $MgCl_2$, 1; Na-HEPES, 20; pH 7.4; glucose, 5.6, and variable amounts of $CaCl_2$, *e.g.*, 1.25 mM. Human parathyroid glands, were obtained from patients undergoing surgical removal of parathyroid tissue for primary or uremic hyperparathyroidism (uremic HPT), and were treated similarly to bovine tissue.

Glands were trimmed of excess fat and connective tissue and then minced with fine scissors into cubes approximately 2-3 mm on a side. Dissociated parathyroid cells were prepared by collagenase digestion and then purified by centrifugation in Percoll buffer. The resultant parathyroid cell preparation was essentially devoid of red blood cells, adipocytes, and capillary tissue as assessed by phase contrast microscopy and Sudan black B staining. Dissociated and purified parathyroid cells were present as small clusters containing 5 to 20 cells. Cellular viability, as indexed by exclusion of trypan blue or ethidium bromide, was routinely 95%.

Although cells can be used for experimental purposes at this point, physiological responses (*e.g.*, suppressibility of PTH secretion and resting levels of $[Ca^{2+}]_i$) should be determined after culturing the cells overnight. Primary culture also has the advantage that cells can be labeled with isotopes to near isotopic equilibrium, as is necessary for studies involving measurements of inositol phosphate metabolism.

After purification on Percoll gradients, cells were washed several times in a 1:1 mixture of Ham's F12-

Dulbecco's modified Eagle's medium (GIBCO) supplemented with 50 μg/ml streptomycin, 100 U/ml penicillin, 5 μg/ml gentamicin and ITS⁺. ITS⁺ is a premixed solution containing insulin, transferrin, selenium, and bovine serum albumin (BSA)-linolenic acid (Collaborative Research, Bedford, MA). The cells were then transferred to plastic flasks (75 or 150 cm^2; Falcon) and incubated overnight at 37°C in a humid atmosphere of 5% CO_2. No serum is added to these overnight cultures, since its presence allows the cells to attach to the plastic, undergo proliferation, and dedifferentiate. Cells cultured under the above conditions were readily removed from the flasks by decanting, and show the same viability as freshly prepared cells.

Purified parathyroid cells were resuspended in 1.25 mM $CaCl_2$-2% BSA-PCB containing 1 μM fura-2-acetoxymethyl-ester and incubated at 37°C for 20 minutes. The cells were then pelleted, resuspended in the same buffer, but lacking the ester, and incubated a further 15 minutes at 37°C. The cells were subsequently washed twice with PCB containing 0.5 mM $CaCl_2$ and 0.5% BSA and maintained at room temperature (about 20°C). Immediately before use, the cells were diluted five-fold with prewarmed 0.5 mM $CaCl_2$-PCB to obtain a final BSA concentration of 0.1%. The concentration of cells in the cuvette used for fluorescence recording was 1-2 x 10^6/ml.

The fluorescence of indicator-loaded cells was measured at 37°C in a spectrofluorimeter (Biomedical Instrumentation Group, University of Pennsylvania, Philadelphia, PA) equipped with a thermostated cuvette holder and magnetic stirrer using excitation and emission wavelengths of 340 and 510 nm, respectively. This fluorescence indicates the level of cytosolic Ca^{2+}. Fluorescence signals were calibrated using digitonin (50 μg/ml, final) to obtain maximum fluorescence (F_{max}), and EGTA (10 mM, pH 8.3, final) to obtain minimal fluorescence (F_{min}), and a dissociation constant of 224 nM. Leakage of dye is dependent on temperature and most occurs within the

first 2 minutes after warming the cells in the cuvette. Dye leakage increases only very slowly thereafter. To correct the calibration for dye leakage, cells were placed in the cuvette and stirred at 37°C for 2-3 minutes. The cell suspension was then removed, the cells pelleted, and the supernatant returned to a clean cuvette. The supernatant was then treated with digitonin and EGTA to estimate dye leakage, which is typically 10-15% of the total Ca^{2+}-dependent fluorescent signal. This estimate was subtracted from the apparent F_{min}.

Example 4: Using Fura-2 Loaded HEK 293/pHuPCaR4.0 Cells To Measure to Calcium Receptor Activity

This section describes procedures used to assay calcium receptor activity using fura-2 loaded HEK 293/pHuPCaR4.0 cells. HEK 293 cells transfected with pHuPCaR4.0 were loaded with fura-2 by incubating the cells in Dulbecco's modified Eagle's media buffered with 20 mM HEPES containing about 5 μM fluo-3/AM for one hour at room temperature. Cell were then rinsed with Hank's balanced salt solution buffered with 20 mM HEPES containing 1 mM $CaCl_2$ and 1 mM $MgCl_2$. Compounds to be tested were then added to the cells and fluorescence was measured (excitation and emission wavelengths of 340 and 510 nm, respectively).

Example 5: Measuring the Ability of Compounds to Modulate Calcium Receptor Activity

The ability of different compounds to modulate calcium receptor activity was assayed by measuring increases in $[Ca^{2+}]_i$ in HEK 293 cells transfected with nucleic acid encoding pHuPCaR4.0 using fura-2 loaded cells or using parathyroid cells loaded with using fura-2 loaded cells. Results of different experiments are summarized in Tables 1.a, 1.b.1, 1.b.2, 1.c., and 2. Tables 1.a, 1.b.1, 1.b.2, and 1.c summarizes the effects of compounds, at different concentrations, on calcium receptor activity assayed as

described in Example 4 (*i.e.*, using HEK 293 cells transfected with nucleic acid encoding pHuPCaR4.0, which were loaded with fura-2).

Table 2, summarizes the results of different experiments where the EC_{50} was calculated either parathyroid cells, or HEK 293/pHuPCaR4.0, loaded with fura-2. Cells were loaded with fura-2 and assayed as described in Example 2 (for parathyroid cells) or Example 3 (for HEK 293/pHuPCaR4.0 cells).

Table 1.a. Calcimimetic compounds which produce greater than 40% response at 3.3 ng/mL in HEK-293 cells expressing the human calcium receptor.

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
Reference compounds				
R-568		95	69	24
17P		101	86	54
17X		105	93	51
24X	126	109	124	109
24Y	119	120	127	102
17J	116	118	122	102
25A	122	120	114	92
17E	116	110	110	92
24Z	138	138	135	90
14S	116	106	105	88
25E	132	129	122	85
17G	125	128	119	77
14T	126	125	117	77
17H	126	124	111	74
14O	119	119	102	74
25I	119	113	114	74
12J	131	130	113	68

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
12I	115	111	93	68
25G	130	115	99	66
9R		108	101	64
12F	118	110	101	63
12O	110	117	94	62
23Z	129	126	100	61
17M		115	99	59
16V		114	102	58
25O	126	115	96	57
25J	119	123	105	56
16L	146	138	98	56
12N	115	106	102	55
16T		97	88	55
25U	107	107	95	55
17P		101	86	54
16Q		110	88	53
23E	137	113	102	53
17C	113	120	99	52
25L	97	97	85	52
8Z		101	97	52
17X		105	93	51
13R		132	98	51
17O		112	96	51
23Q	122	114	98	51
16X		111	96	51
24V	127	98	71	50
13O		115	94	50
17N		108	86	49
21V	122	116	99	48
24M	132	134	99	48
13U		108	79	47

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
24P	140	138	110	46
17Y	109	94	79	46
11X		100	76	45
25H	115	107	89	45
22J		99	71	45
9C		104	82	45
13S		102	87	45
10Q	103	100	84	44
13P		110	83	44
8K		98	81	44
13N		114	88	43
10N	106	97	77	43
12H	114	115	94	43
25P	90	81	75	41
18A		111	88	40
14L		109	78	40

Table 1.b.1. Calcimimetic compounds which produce greater than 40% response at 33 ng/mL in HEK-293 cells expressing the human calcium receptor

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
Reference compounds				
R-568		95	69	24
17P		101	86	54
17X		105	93	51
12C	134	125	98	39
16I	121	117	96	36

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
17D		108	91	38
17F		111	90	28
24C	116	113	87	32
25K	124	107	86	35
13F	125	122	85	38
21F		109	85	36
21S	132	131	85	34
10F		96	84	27
14R	106	107	84	37
13G	111	128	82	29
14Z	118	103	82	20
16N	122	159	82	8
8U	123	129	82	11
23W	117	97	81	25
12G	139	139	81	35
15G		113	80	32
25M	118	100	79	25
13V		110	79	33
14P	112	103	78	30
6T	123	129	78	15
14Q		101	78	35
17L	111	104	78	31
24K		106	78	30
24U	106	106	78	25
25Q	116	95	77	20
8J		104	77	39
23H	121	114	77	28
21C=4U	134	114	76	17
25F	97	85	76	28
16R		100	76	25
17I	118	97	76	18

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Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
24J		103	75	31
21O		109	75	37
24G	109	94	75	22
15I	111	93	75	24
21D		104	75	17
20Y	117	95	74	24
10P		102	74	8
23M	113	97	74	26
14Y		109	73	17
17K	98	97	73	37
12E	117	121	73	23
17Z		99	73	37
16W		102	73	4
23K	106	107	72	24
25X	96	94	72	22
13W		109	71	12
23P	125	99	70	22
18B	111	96	69	26
21Y		100	68	36
17W		92	67	13
23A		103	67	24
23G	127	93	67	13
13M		92	66	15
21U	104	104	66	18
21R		100	66	15
10S/10T		86	65	13
17R		98	65	13
13X		102	65	13
4N		100	65	13
21E		94	64	4
15J	80	75	64	13

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
22Y		114	64	28
21G		88	63	18
24L		105	62	10
10V		99	62	8
10W/10X		98	61	9
17B		92	61	19
23Y	106	87	61	16
11Y		103	61	20

Table 1.b.2 Calcimimetic compounds which produce greater than 40% response at 33 ng/mL in HEK-293 cells expressing the human calcium receptor

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
reference compounds				
R568		95	69	24
17P		101	86	54
17X		105	93	51
18C	99	87	60	18
23T	102	74	60	31
4V		93	59	
8G		84	59	6
23I		102	58	3
21M		102	58	17
24O	137	114	58	8
3U		89	57	
9A		82	56	6
12M	98	86	56	11
12B	130	110	56	4

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
21P		92	56	13
8T		85	55	13
10L/10M		99	55	4
24I	109	84	55	11
14N		89	55	15
23R	104	86	54	13
23S		97	53	3
21T	133	112	53	3
10W/10X		81	53	4
13T		90	53	6
6R		94	52	7
20I		87	52	12
24A	122	85	52	9
12D	128	109	52	5
6X		84	52	10
18T	99	74	52	14
21X	119	101	51	2
23J	102	61	51	29
10Z		96	51	5
16Z		88	51	9
23N		96	50	2
16U		85	50	4
11D		96	50	4
23X		94	49	1
17A		88	49	7
20J		80	48	8
22X		86	48	10
23U		87	48	3
9Z		74	48	4
16J	92	76	47	31
25N	94	73	46	8
4P		81	46	8

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
23O	111	79	46	13
13Q		95	46	5
4G		83	46	
12Y		80	46	10
12L		88	45	10
23F		82	45	5
11W		81	44	2
8H		88	44	7
25V	89	59	43	26
25W	95	69	42	8
10R		82	42	7
21N	124	98	42	4
8S		73	42	7
8X		75	40	19
13E	123	94	40	2

Table 1.c. Calcimimetic compounds which produce greater than 40% response at 330 ng/mL in HEK-293 cells expressing the human calcium receptor

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
reference compounds				
R568		95	69	24
17P		101	86	54
17X		105	93	51
7X		85		
3H		84		
3L		81	28	
16O	129	81	21	2
8O/8Q	124	80	14	0

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
14A	98	78	10	7
23L	107	77	37	9
1T		76		
7W		76		
4H		77	37	
8D		75		
5M		73	21	
4U		72		
24E	94	71	35	6
16M	130	68	11	4
4M		68	34	
2S		67	29	
17V	91	66	27	-1
2X		66	15	
23D	91	66	35	13
4P		65	32	
5B/5C		65	20	
3M		64	19	
16K	78	62	36	8
5D		62	18	
4D		61	13	
24B	76	61	34	11
24H	81	60	32	13
5L		60	16	
2Y		59	10	
5G		58	16	
3V		56	14	
2Q		56	4	
14B	75	55	11	4
13Z	93	54	22	5
8A		54		
24D	87	53	34	39

Compound Code	% activity at four concentrations (ng/mL)			
	3300	330	33	3.3
1D		53		
13I	85	52	3	1
3B		52	15	
8C		51		
14H	112	49	5	5
7U		49		
5E		48	7	
13H	88	48	36	12
13Y	106	47	2	4
4J		47	8	
14I	80	45	11	7
4B		45	8	
3D		45	4	
3R		45	2	
3A		41	7	
14J	55	41	6	5
4I		40	9	

TABLE 2

Arylalkylamine Calcimimetics from Figure 1 Active at the Parathyroid Cell Calcium Receptor *In Vitro* ($EC_{50} \leq 5$ μM)

Compound Code (from Fig. 1)	EC_{50} (μM)	Compound Code (from Fig. 1)	EC_{50} (μM)
NPS *R*-467	2.0	11X	0.83
NPS *R*-568	0.60	11Y	2.8
3U	0.64	12L	1.7
3V	1.8	12U	1.2
4A	1.4	12V	0.42
4B	2.0	12W	3.2

4C	2.0	12Y	2.0
4D	4.4	12Z	0.11
4G	1.8	13Q	*ca.* 0.8
4H	≥3.0	13R	0.25
4J	2.2	13S	<0.13
4M	2.1	13U	0.19
4N	0.8	13X	<0.75
4P	1.6	14L	0.26
4R/6V	4.2	14Q	0:47
4S	3.3	14U	0.13
4T/4U	1.6	14V	1.7
4V	2.5	14Y	0.38
4W	2.3	15G	*ca.* 0.5
4Y	1.3	16Q	0.04
4Z/5A	4.4	16R	0.36
5B/5C	2.8	16T	0.04
5W/5Y	3.6	16V	<0.13
6E	2.7	16W	0.59
6F(*R,R-*)	0.83	16X	0.10
6R	3.4	17M	0.15
6T	2.9	17O	0.04
6X	2.5	17P	0.04
7W	3.2	17R	0.39
7X	1.1	17W	0.43
8D	2.5	17X	0.02
8J	0.78	20F	<1.0
8K	1.3	20I	>1.0
8R	2.6	20J	>3.0
8S	1.7	20R	2.4
8T	1.8	20S	4.2
8U	0.44	21D	3.0
8X	0.76	21F	0.38
8Z	0.40	21G	1.1

9C	0.60	21O	0.26
9D	1.4	21P	0.43
9R	0.25	21Q	1.4
9S	4.8	21R	0.37
10F	0.89	25C	> 2
11D	1.8	25D	0.019

Examples 6-17: Synthesis of Compounds

The compounds described herein can be synthesized using standard techniques such as those described by Nemeth *et al.*, PCT/US93/01642, International Publication Number WO 94/18959. Examples describing representative syntheses of compounds described in the text are provided below.

Synthesis of compounds 9R, 14U, and 17P were prepared by reductive amination of a commercially available aldehyde or ketone with a primary amine in the presence of sodium cyanoborohydride or sodium triacetoxyborohydride. Compounds 11Y, 12H, 12K, 12M, 14S, 14T, 16L-O, 17E, 17G, 17J, 24X, 24Y, 25A, 25E-25K, and 25O were prepared in a similar manner.

It was found for the syntheses of these three compounds (9R, 14U, and 16P) that sodium triacetoxyborohydride afforded the desired diastereoisomers with greater diastereoselectivity than using sodium cyanoborohydride. The enriched mixtures were further purified to a single diastereomer by normal-phase HPLC or by recystallization from organic solvents.

Compounds 8J, 8U, 11X, 17M, and 25Y were prepared from the condensation of a primary amine with an aldehyde or ketone in the presence of titanium(IV) isopropoxide. The resulting intermediate imines were then reduced *in situ* by the action of sodium cyanoborohydride, sodium borohydride, or sodium triacetoxyborohydride. The intermediate enamine for the synthesis of compound 8U was

catalytically reduced using or palladium dihydroxide on carbon.

Compounds 12U, 12V and 12Z were prepared by a diisobutylaluminum hydride (DIBAL-H) mediated condensation of an amine with a nitrile. The resulting intermediate imine is reduced *in situ* by the action of sodium cyanoborohydride or sodium borohydride. The intermediate alkenes (compounds 12U and 12V) were reduced by catalytic hydrogenation in EtOH using palladium on carbon. Compounds which were converted to their corresponding hydrochloride were done so by treatment of the free base with ethereal HCl to afford white solids.

The amines in these syntheses were purchased from Aldrich Chemical Co., Milwaukee, WI, or from Celgene Corp., Warren, NJ, or were prepared synthetically using standard techniques. All other reagent chemicals were purchased from Aldrich Chemical Co.

Example 6: Synthesis of Compound 25Y

N-(3-(2-Phenyl)propyl)-1-(1-naphthyl)ethylamine

A mixture of 3-phenyl-1-propylamine (135 mg, 1 mmol), 1'-acetonaphthone (170 mg, 1 mmol), and titanium (IV) isopropoxide (355 mg, 1.3 mmol) was stirred at room temperature for 1 hour. The reaction was treated with 1 M ethanolic sodium cyanoborohydride (1 mL) and stirred at room temperature for 16 hours. The reaction was diluted with ether and treated with water (0.1 mL). The reaction was centrifuged and the ether layer removed and concentrated to a milky oil. A small portion of this material (10 mg) was purified by HPLC (Phenomenex, 1.0 x 25 cm, 5 μM silica) using a gradient of dichloromethane to 10% methanol in dichloromethane containing 0.1% isopropylamine. This afforded the product (free base) as a single component by GC/El-MS (R_t= 10.48 min) m/z (rel. int.) 289 (M^+,11), 274 (63), 184 (5), 162 (5), 155 (100), 141 (18), 115 (8), 91 (45), 77(5).

Example 7: Synthesis of Compound 8J

N-(3-phenylpropyl)-1-(3-thiomethylphenyl)ethylamine hydrochloride

3'-Aminoacetophenone (2.7 g, 20 mmol) was dissolved in 4 mL of concentrated HCl, 4 g of ice and 8 mL of water. The solution was cooled to 0°C, and sodium nitrite (1.45 g, 21 mmol) dissolved in 3-5 mL of water was added over 5 minutes while maintaining the temperature below 6°C. Sodium thiomethoxide (1.75 g, 25 mmol) was dissolved in 5 mL of water and cooled to 0°C. To this solution was added the diazonium salt over 10 minutes while maintaining the temperature below 10°C. The reaction was stirred for an additional hour while allowing the temperature to rise to ambient. The reaction mixture was partitioned between ether and water. The ether layer was separated and washed with sodium bicarbonate and sodium chloride, and dried over sodium sulfate. The ether was evaporated to give a 74% yield of 3'-thiomethylacetophenone. The crude material was purified by distillation at reduced pressure.

3-Phenylpropylamine (0.13 g, 1 mmol), 3'-thiomethylacetophenone (0.17 g, 1 mmol), and titanium (IV) isopropoxide (0.36 g, 1.25 mmol) were mixed together and allowed to stand for 4 hours. Ethanol (1 mL) and sodium cyanoborohydride (0.063 g, 1 mmol) were added and the reaction was stirred overnight. The reaction was worked up by the addition of 4 mL of ether and 200 μL of water. The mixture was vortexed and then spun in a centrifuge to separate the solids. The ether layer was separated from the precipitate, and the solvent removed *in vacuo*. The oil was redissolved in dichloromethane and the compound purified by preparative TLC on silica gel eluted with 3% methanol/dichloromethane to yield the title compound as a pure oil: GC/EI-MS(R_t=7.64 min) m/z (rel. int.)285 (M^+,18), 270(90), 180(17), 151(100), 136(32), 104(17), 91(54), 77(13).

Example 8: Synthesis of Compound 8U

N-3-(2-methoxyphenyl)-1-propyl-(R)-3-methoxy-α-methylbenzylamine hydrochloride

A mixture of (R)-(+)-3-methoxy-α-methylbenzylamine (3.02 g, 20 mmol), 2-methoxycinnamaldehyde (3.24 g, 20 mmol), and titanium (IV) isopropoxide (8.53 g, 30 mmol, 1.5 Eq.) was stirred 2 hours at room temperature and treated with 1 M (20 mL) ethanolic sodium cyanoborohydride. The reaction was stirred overnight (16 hours), diluted with diethylether, and treated with water (1.44 mL, 80 mmol, 4 Eq.). After mixing for 1 hour the reaction mixture was centrifuged and the ether layer removed and concentrated to an oil. This material was dissolved in glacial acetic acid, shaken with palladium hydroxide and hydrogenated under 60 p.s.i. hydrogen for 2 hours at room temperature. The catalyst was removed by filtration and the resulting solution concentrated to a thick oil. This material was dissolved in dichloromethane and neutralized with 1 N NaOH. The dichloromethane solution was separated from the aqueous phase, dried over anhydrous potassium carbonate and concentrated to an oil. This material was dissolved in ether and treated with 1 M HCl in diethylether. The resulting precipitate (white solid) was collected, washed with diethylether, and air dried. GC/El-MS (R_t = 9.69 min) of this material (free base) showed a single component: m/z (rel. int.) 299 (M+, 21), 284 (100), 164 (17), 150 (8), 135 (81), 121 (40), 102 (17), 91 (43), 77 (18).

Example 9: Synthesis of Compound 9R

(R)-N-(1-(2-naphthyl)ethyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride

A mixture of (R)-(+)-1-(1-naphthyl)ethylamine (10.0 g, 58 mmol), 2'-acetonaphthone (9.4 g, 56 mmol), titanium (IV) isopropoxide (20.7 g, 73.0 mmol), and EtOH (abs.) (100 mL) was heated to 60°C for 3 hours. Sodium cyanoborohydride ($NaCNBH_3$) (3.67 g, 58.4 mmol) was then added.

The reaction mixture was stirred at room temperature for 18 hours. Ether (1 L) and H_2O (10 mL) were added to the reaction mixture and the resulting precipitate was then removed by centrifugation. The supernatant was evaporated under vacuum and the crude product was recrystallized four times from hot hexane, to provide 1.5 g of pure (98+%) diastereomer. The free base was dissolved in hexane, filtered, and then ethereal HCl was added to precipitate the product as a white solid (1.1 g, 6 % yield), m.p.: softens 200-240°C (dec.).

Example 10: Synthesis of Compound 11X

N-(4-Isopropylbenzyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride

A mixture of (*R*)-(+)-1-(1-naphthyl)ethylamine (1.06 g, 6.2 mmol), 4-isopropylbenzaldehyde (0.92 g, 6.2 mmol), and titanium (IV) isopropoxide (2.2 g, 7.7 mmol) was heated to 100°C for 5 min then allowed to stir at room temperature for 4 hours. Sodium cyanoborohydride ($NaCNBH_3$) (0.39 g, 6.2 mmol) was then added followed by EtOH (1 mL). The reaction mixture was stirred at room temperature for 18 hours. Ether (100 mL) and H_2O (1 mL) were added to the reaction mixture and the resulting precipitate was then removed by centrifugation. The supernatant was evaporated under vacuum and the crude product was chromatographed on silica gel (50 mm X 30 cm column) (elution with 1% MeOH/ $CHCl_3$). The chromatographed material was then dissolved in hexane and ethereal HCl was added to precipitate the product as a white solid (0.67 g, 35 % yield), m.p.; 257-259°C.

Example 11: Synthesis of Compound 12U

N-3-(2-methylphenyl)-1-propyl-(R)-3-methoxy-α-methylbenzylamine hydrochloride

A solution of 2-methylcinnamonitrile (1.43 g, 10 mmol) in dichloromethane (10 mL) was cooled to 0°C and treated dropwise (15 minutes) with 1 M diisobutylaluminum

hydride (10 mL, dichloromethane). The reaction was stirred at 0°C for 15 minutes and treated dropwise (15 minutes) with a 1 M solution of (*R*)-(+)-3-methoxy-α-methylbenzylamine (1.51 g, 10 mmol) in dichloromethane (10 mL). The reaction was stirred 1 hours at 0°C and poured into a solution of ethanol (100 mL) containing sodium cyanoborohydride (1 g, 16 mmol). The reaction mixture was stirred 48 hour at room temperature. The reaction was diluted with ether and neutralized with 1 N NaOH. The ether layer was removed, dried over anhydrous potassium carbonate and concentrated to an oil. This material was chromatographed through silica using a gradient of dichloromethane to 5% methanol in dichloromethane to afford the unsaturated intermediate, a single component by GC/El-MS (R_t=10.06 min) m/z (rel. int.) 281 (M+, 17), 266 (59), 176 (19), 146 (65), 135 (73), 131 (100), 91 (21), 77 (13).

The unsaturated intermediate in ethanol was hydrogenated (1 atm H_2) in the presence of palladium on carbon for 16 hours at room temperature. The product from this reaction was converted to the hydrochloride salt by treatment with 1 M HCl in diethylether. GC/El-MS (R_t = 9.31 min) of this material (free base) showed a single component: m/z (rel. int.) 283 (M+, 21), 268 (100), 164 (12), 148 (8), 135 (85), 121 (12), 105 (49), 91 (23), 77 (21).

Example 12: Synthesis of Compound 12V

N-3-(3-methylphenyl)-1-propyl-(R)-3-methoxy-α-methylbenzylamine hydrochloride

The compound was prepared following the procedure described in Example 11, but using 2-methylcinnamonitrile. The unsaturated intermediate was a single component by GC/EI-MS (R_t = 10.21 min) m/z (rel. int.) 281 (M+, 57), 266 (86), 146 (98), 135 (88), 131 (100), 115 (43), 102 (26), 91 (43), 77 (18). Reduction of this material and hydrochloride formation using the procedure described Example

11 afforded the product. GC/EI-MS (R_t = 9.18 min) of this material (free base) showed a single component; m/z (rel. int.) 283 (M+, 19), 268 (100), 164 (11), 148 (8), 135 (76), 121 (16), 105 (45), 91 (23), 77 (21).

Example 13: Synthesis of Compound 12Z

N-3-(2-chlorophenyl)-1-propyl-(R)-1-(1-naphthyl)ethylamine hydrochloride

The compound was prepared following the procedures described in Example 11, but using 2-chlorohydrocinnamonitrile and (*R*)-(+)-1-(1-naphthyl)ethylamine on a 10 mmol scale. Chromatography through silica using a gradient of dichloromethane to 5% methanol in dichloromethane afforded the product as a single component by TLC analysis (5% methanol in dichloromethane). The hydrochloride was prepared by treatment with 1 M HCl in diethylether.

Example 14: Synthesis of Compound 14U

(R)-N-(1-(4-methoxyphenyl)ethyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride

A mixture of (*R*)-(+)-1-(1-naphthyl)ethylamine (1.1 g, 6.2 mmol), 4'-methoxyacetophenone (0.93 g, 6.2 mmol), titanium (IV) isopropoxide (2.2 g, 7.7 mmol), and EtOH (abs.) (1 mL) was heated to 60°C for 3 hours. Sodium cyanoborohydride ($NaCNBH_3$) (0.39 g, 6.2 mmol) was then added, and the reaction mixture was stirred at room temperature for 18 hours. Ether (200 mL) and H_2O (2 mL) were added to the reaction mixture and the resulting precipitate was then removed by centrifugation. The supernatant was evaporated under vacuum and the crude product was chromatographed on silica gel (25 mm X 25 cm column) (elution with 1% MeOH/$CHCl_3$). A portion of this material was HPLC chromatographed [Selectosil, 5 μM silica gel; 25 cm x 10.0 mm (Phenomenex, Torrance, CA), 4 mL per minute; UV det. 275 nM; 12% ethyl acetate-88% hexane (elution time 12.0 min)]. The HPLC purified diastereomer was then dissolved in hexanes and ethereal HCl was added

to precipitate the product as a white solid (20 mg), m.p.: 209-210°C(dec.).

Example 15: Synthesis of Compound 17M

N-(3-chloro-4-methoxybenzyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride

A mixture of *(R)*-(+)-1-(1-naphthyl)ethylamine (6.6 g, 39 mmol), 3'-chloro-4'-methoxybenzaldehyde (6.6 g, 39 mmol), and titanium (IV) isopropoxide (13.8 g, 48.8 mmol), and EtOH (abs.) (30 mL) was heated to 80°C for 30 minutes then allowed to stir at room temperature for 3 hours. Sodium cyanoborohydride ($NaCNBH_3$) (2.45 g, 39 mmol) was then added. The reaction mixture was stirred at room temperature for 18 hours. Ether (100 mL) and H_2O (2 mL) were added to the reaction mixture and the resulting precipitate was then removed by centrifugation. The supernatant was evaporated under vacuum and the crude product was chromatographed on silica gel (50 mm X 30 cm column) (elution with CH_2Cl_2). The chromatographed material was then dissolved in hexane (500 mL), decolorized with Norit® filtered (0.2 μM), and then ethereal HCl was added to precipitate the product as a while solid (10.2 g, 56 % yield), m.p.: 241-242°C (dec.).

Example 16: Synthesis of Compound 17P

4-Methoxy-3-methylacetophenone [17P Precursor]

A mixture of 4'-hydroxy-3'-methylacetophenone (5.0 g, 33.3 mmol), iodomethane (5.7 g, 40.0 mmol), K_2CO_3 (granular, anhydrous) (23.0 g, 167 mmol), and acetone (250 mL) was refluxed for 3 hours. The reaction mixture was then cooled to room temperature, filtered to remove the inorganic salts, and evaporated under vacuum. The crude product was dissolved in ether (100 mL) and washed with H_2O (2 x 20 mL). The organic layer was dried (Na_2SO_4) and evaporated to yield 4.5 g, 82.4% yield. The ketone was used in the following reaction without further purification.

(R)-N-(1-(4-Methoxy-3-methylphenyl)ethyl)-(R)-1-(1-naphthyl)ethylamine hydrochloride [Compound 17P]

A mixture of (*R*)-(+)-1-(1-naphthyl)ethylamine (4.24 g, 24.8 mmol), 4'-methoxy-3'-methylacetophenone (4.06 g, 24.8 mmol), and titanium (IV) isopropoxide(8.8 g, 30.9 mmol), and EtOH (abs.) (1 mL) was heated to 100°C for 2 hours. Isopropanol (45 mL) was added and the reaction was then cooled to 10°C in an ice bath. Sodium triacetoxyborohydride ($NaHB(O_2CCH_3)_3$) (10.5 g, 49.5 mmol) was then added in portions over 15 minutes. The reaction mixture was then heated to 70°C for 18 hours. The mixture was cooled to room temperature and poured into ether (400 mL). The suspension was centrifuged, the supernatant was collected and the pellet was washed with ether (400 mL). The combined organic washings were evaporated under vacuum. The residue was dissolved in ether (400 mL) and washed with 1 N NaOH (4 x 50 mL) and H_2O (2 x 50 mL). The organic layer was dried (Na_2SO_4), filtered and evaporated under vacuum. EtOH (abs.) was added to the wet residue which was then dried thoroughly on a rotary evaporator to provide an oil. The mixture was then chromatographed on silica gel (50 mm x 30 cm) [elution with (1% MeOH:1% IPA:$CHCl_3$) to give 4.8 g of an oil].

The desired diastereomer was further purified by HPLC chromatography [SUPELCOSIL™ PLC-Si, 18 μM silica gel; 25 cm x 21.2 mm (Supelco, Inc., Bellefonte, PA), 7 mL per minute; UV det. 275 nM: 20% EtOAc-80% hexane (elution time 9.5 - 11.0 min)]. Injections (800 μL aliquots) of the mixture (100 mg/mL solution in eluent) provided 65 mg of the desired isomer. Multiple HPLC injections provided 1.0 g of purified material. The HPLC chromatographed material was dissolved in hexane (50 mL) and the hydrochloride salt was precipitated with ethereal HCl. The salt was collected on fritted glass and washed with hexane to provide 1.0 g of a white solid, mp 204-205°C.

Example 17: Synthesis of Compound 17X

3-Chloro-4-methoxybenzaldehyde

A mixture of 3-chloro-4-hydroxybenzaldehyde (25 g, 160 mmol), iodomethane (27.25 g, 192 mmol), K_2CO_3 (granular, anhydrous) (110.6 g, 800 mmol), and acetone (300 mL) was refluxed for 3 hours. The reaction mixture was then cooled to room temperature. Diethyl ether (500 mL) was added and the mixture was filtered through paper to remove the inorganic solids. The filtrate was evaporated under reduced pressure, dissolved in diethyl ether (800 mL), and washed with 0.1 *N* NaOH (3 x 100 mL). The organic layer was dried (Na_2SO_4) and evaporated under vacuum to yield 24 g, 92% yield of crude product. This material was further purified by chromatography on silica gel (50 mm x 30 cm) (elution with hexane-EtOAc, 5:1) to give 15.02 g, 56% yield of a white solid: TLC (hexane-EtOAc, 5:1) R_f=0.24; GC R_t=4.75 min; MS (EI) *m/z* 170(M^+), 172(M+2).

1-Methyl-(3'-chloro-4'-methoxybenzyl) alcohol

A mixture of 3-chloro-4-methoxybenzaldehyde (13 g, 76.5 mmol), methylmagnesium chloride (52 g, 153 mmol), and THF (300 mL) was refluxed for 3 hours. The reaction mixture was cooled to room temperature. NH_4Cl (satd. soln., 6 mL) was added dropwise followed by diethyl ether (500 mL) and the mixture was filtered through paper to remove the inorganic solids. The filtrate was evaporated under reduced pressure and the resulting solid was dissolved in diethyl ether (300 mL) and washed with water (4 x 25 mL). The organic layer was dried (Na_2SO_4) and evaporated under vacuum to yield 11.3 g, 80% yield of crude product. This material was further purified by chromatography on silica gel (50 mm x 30 cm) (elution with CH_2Cl_2) to yield 11.3 g, 63% yield of an oil; TLC (CH_2Cl_2) R_f=0.25; GC R_t=5.30 min; MS (EI) *m/z* 186(M^+), 188(M+2).

3'-Chloro-4'-methoxyacetophenone

A mixture of 1-methyl-(3'-Chloro-4'-methoxybenzyl) alcohol (7.6 g, 41 mmol), pyridinium chlorochromate (PCC) (13.16 g, 61.5 mmol), and CH_2Cl_2 (300 mL) was allowed to stir at room temperature for 2 hours. Diethyl ether (1000 mL) was added and the resulting mixture was placed on a chromatography column of silica gel (50 mm x 30 cm) (elution with diethyl ether) to yield 7.3 g, 97% yield of crude solid product. GC analysis of this material showed it to be 99% pure and it was used in the following reaction without further purification. TLC (diethyl ether) R_f=1.0; GC R_t=5.3 min; MS (EI) *m/z* 184(M^+), 184(M+2).

(R,R)-N-(1-Ethyl-4'-methoxy-3'-chlorophenyl)-1-(1-naphthylethyl)amine

A mixture of 3'-chloro-4'-methoxyacetophenone (5.3 g, 29 mmol), (*R*)-(+)-1-(1-naphthyl)ethylamine (4.98 g, 29 mmol), titanium (IV) isopropoxide (10.2 g, 36 mmol), and isopropanol (20 mL) was heated to 100°C for 3 hours. Sodium triacetoxyborohydride ($NaB(O_2CCH_3)_3$; 12.29 g, 58 mmol) was added in portions over 10 minutes. The reaction mixture was heated to reflux for 30 minutes and was then allowed to stir at room temperature for 18 hours. The mixture was then poured into diethyl ether (500 mL); H_2O (2 mL) was added and the suspension was centrifuged to remove the fine precipitate of titanium salts. The supernatant was collected and the pellet was washed with ether (500 mL). The combined organic layers were dried (Na_2SO_4) and evaporated under vacuum to yield 6.81 g, 70% of crude product.

This material was further purified by chromatography on silica gel (50 mm x 30 cm) (elution with 3% MeOH-97% CH_2Cl_2) to give 2.01 g of an oil. The diastereomer was further purified by recrystallization. The free base (1.98 g) was converted to its HCl salt with ethereal HCl. This salt was dissolved in hot isopropanol (65 mL) and the solution was filtered through paper. The filtrate was

evaporated under vacuum and the resulting solid dissolved in isopropanol (30 mL). After standing at room temperature for 18 hours, the crystalline solid was collected, washed with cold isopropanol (20 mL), and dried to yield 0.87 g, 40% (from free base) of the diastereomerically pure hydrochloride salt: mp 236-237½C (dec); TLC (MeOH-CH_2Cl_2 [99:1]) R_f=0.25; GC R_t=11.06 min; FTIR (KBr pellet, cm^{-1}) 3433, 2950, 2931, 2853, 2803, 2659, 2608, 2497, 1604, 1595, 1504, 1461, 1444, 1268, 1260, 1067, 1021, 802, 781, 733; MS (EI) *m/z* 339(M^+), 341(M+2).

Example 18: Additional Synthesis Protocol

Preparation of 22Z and 23A

A stirred solution of sodium hydride (2.173 g, 60% in oil, 54.325 mmol) in dimethylformamide (100ml) was treated dropwise with triethyl phosphonoacetate (12.47 g, 55.65 mmol) and stirred 30 min at rt. After this time, a solution of *m*-trifluoromethoxy benzaldehyde (10.0 g, 52.6 mmol) in dimethylformamide (50 ml) was added dropwise and the solution stirred 30 min at rt and 30 min at 100°C. The reaction was quenched by the addition of water and transferred to a separatory funnel using diethyl ether (500 ml). The ether solution was washed with saturated ammonium chloride (4 x 500 ml), dried over anhydrous magnesium sulfate, filtered and concentrated to afford ethyl *m*-trifluoromethoxycinnamate as an oil; m/z (rel. int.) 260 (M+, 19), 232 (16), 215 (100), 187 (21), 101 (28).

The ethyl ester in ethanol (100 ml) was reduced under 60 p.s.i. hydrogen using a catalytic amount (10% by weight) palladium hydroxide. After reduction (2 hr, rt) the reaction was filtered and concentrated to afford ethyl *m*-trifluoromethoxyhydrocinnamate as an oil; m/z (rel. int.) 262 (M+, 16), 217 (7), 188 (100), 175 (28), 103 (31), 91 (18), 77 (23).

The saturated ethyl ester was hydrolyzed in a solution of ethanol-10 M sodium hydroxide (1:1) for 16 hr at rt. After this time the solution was acidified and the

product extracted into diethyl ether. The ether solution was dried over anhydrous magnesium sulfate and concentrated to afford *m*-trifluoromethoxyhydrocinnamic acid as a solid; m/z (rel. int.) 234 (M+, 46), 188 (100), 174 (65), 103 (27), 91 (12), 77 (17).

The acid was stirred in excess thionyl chloride for 4 hr at rt. The excess thionyl chloride was evaporated at reduced pressure (100°C) to afford *m*-trifluoromethoxyhydrocinnamyl chloride as an oil. The product was used without further purification.

A solution of *m*-trifluoromethoxyhydrocinnamyl chloride (9.8 g, 39 mmol) in tetrahydrofuran was cooled to -78°C and treated dropwise with a solution (13 ml of 3 M in tetrahydrofuran) of methylmagnesium bromide (39 mmol). The reaction was stirred 4 hr at -78°C, 8 hr at rt, and quenched with dilute HCl. The reaction mixture was extracted with diethyl ether. The ether was dried over anhydrous magnesium sulfate, filtered and concentrated to an oil. Chromatography of this material through silica using a gradient of hexane to acetone afforded 4-(3-trifluoromethoxyphenyl)-2-butanone as an oil; m/z (rel. int.) 232 (M+, 68), 217 (7), 189 (59), 175 (31), 103 (28), 43 (100).

A solution of 4-(3-trifluoromethoxyphenyl)-2-butanone (2.32 g, 10 mmol), (*R*)-1-(3-methoxyphenyl)ethylamine (1.51 g, 10 mmol), and titanium (IV) isopropoxide (3.55 g, 12.5 mmol) were stirred 4 hr at rt. The reaction mixture was then treated with a solution (10 ml of 1 M) of ethanolic sodium cyanoborohydride (10 mmol) and stirred 16 hr at rt. The reaction was diluted with diethyl ether (50 ml) and treated with water (0.72 ml, 40 mmol). After mixing thoroughly the solution was centrifuged and the ether layer decanted and concentrated to an oily solid. The solid was suspended in diethyl ether, filtered through 0.45 μM CR PTFE Acrodisc and concentrated to give a clear oil. Repetitive preparative thin-layer chromatography using 5% methanol in chloroform afforded the two

diasteriomers, (*S,R*)-N-[4-(3-trifluoromethoxyphenyl)-2-butyl]-1-(3-methoxyphenyl)ethylamine, 22Z [m/z (rel. int.) 367 (M+,3), 352 (20), 232 (4), 178 (47), 135 (100), 105 (14), 91 (10), 77 (11)] and (*R,R*)-*N*-[4-(3-trifluoromethoxyphenyl)-2-butyl]-1-(3-methoxyphenyl)ethylamine, 23A; m/z (rel. int.) 367 (M+, 3), 352 (19), 232 (7), 178 (43), 135 (100), 105 (19), 91 (10), 77 (11).

Preparation of 22X and 22Y

In a similar fashion an equal molar amount of 4-(3-trifluoromethoxyphenyl)-2-butanone, (*R*)-1-(1-naphthyl) ethylamine and 1.25 equivalents titanium (IV) isopropoxide were mixed and the intermediate imine reduced with ethanolic sodium cyanoborohydride. Work-up and repetitive preparative thin-layer chromatography using 5% methanol in chloroform afforded (*S,R*)-*N*-[4-(3-trifluoromethoxyphenyl)-2-butyl]-1-(1-naphthyl)ethylamine, 22X; m/z (rel. int.) 387 (M+,3), 372 (15), 198 (15), 176 (12), 155 (100), 128 (8), 115 (6), 109 (4), 103 (5), 77 (8) and (*R,R*)-*N*-[4-(3-trifluoromethoxyphenyl)-2-butyl]-1-(1-naphthyl)ethylamine, 22Y; m/z (rel. int.) 387 (M+,2), 372 (12), 198 (16), 176 (11), 155 (100), 128 (8), 115 (6), 109 (4), 103 (5), 77 (8).

Preparation of 4T

In a similar fashion an equal molar amount of 4-(2-chlorophenyl)-2-butanone, prepared from *o*-chlorobezaldehyde, (*R*)-1(3-methoxyphenyl)ethylamine and 1.25 equivalents titanium (IV) isopropoxide were mixed and the intermediate imine reduced with ethanolic sodium cyanoborohydride. Work-up and repetitive preparative thin-layer chromatography using 5% methanol in chloroform afforded (*R,R*)-*N*-[4-(2-chlorophenyl)-2-butyl]-1-(3-methoxyphenyl) ethylamine, 4T; m/z (rel. int.) 317 (M+,3), 302 (16), 178 (62), 178 (62), 135 (100), 125 (15), 105 (10), 91 (6), 77 (8).

Preparation of 21Y

In a similar fashion an equal molar amount of 4-(3-trifluoromethylphenyl)-2-butanone, prepared from *m*-trifluoromethylbezaldehyde, (*R*)-1-(3-methoxyphenyl) ethylamine and 1.25 equivalents titanium (IV) isopropoxide were mixed and the intermediate imine reduced with ethanolic sodium cyanoborohydride. Work-up and repetitive preparative thin-layer chromatography using 5% methanol in chloroform afforded (*R,R*)-*N*-[4-(3-trifluoromethylphenyl)-2-butyl]-1-(3-methoxyphenyl)ethylamine, 21Y [m/z (rel. int.) 351 (M+,2), 336 (18), 216 (4), 202 (3), 178 (45), 135 (100), 105 (13), 91 (9), 77 (8)] and (S,R)-N-[4-(3-trifluoromethylphenyl)-2-butyl]-1-(3-methoxyphenyl) ethylamine, 21X.

Preparation of 25C and 25D

In a similar fashion an equal molar amount of 4-(3-trifluoromethylphenyl)-2-butanone, (R)-1-(1-naphthyl) ethylamine and 1.25 equivalents titanium (IV) isopropoxide were mixed and the intermediate imine reduced with ethanolic sodium cyanoborohydride. Work-up and repetitive preparative thin-layer chromatography using 5% methanol in chloroform afforded (S,R)-N-[4-(3-trifluoromethylphenyl)-2-butyl]-1-(1-naphthyl)ethylamine, 25C [m/z (rel. int.) 371 (M^+, 3), 356 (16), 198 (15), 155 (100), 129 (8), 115 (5), 109 (3), 77 (2)] and (R,R)-N-[4-(3-trifluoromethylphenyl)-2-butyl]-1-(1-naphthyl)ethylamine, 25D; m/z (rel. int.) 371 (M^+, 3), 356 (16), 198 (15), 155 (100), 129 (8), 115 (5), 109 (3), 77 (2).

Preparation of 21D

In a similar fashion an equal molar amount of 4-phenyl-2-butanone (Aldrich Chemical Co.), (R)-1-(3-methoxyphenyl)ethylamine and 1.25 equivalents titanium (IV) isopropoxide were mixed and the intermediate imine reduced with ethanolic sodium cyanoborohydride. Work-up and repetitive preparative thin-layer chromatography using 5%

methanol in chloroform afforded (R,R)-N-(4-phenyl-2-butyl)-1-(3-methoxyphenyl)ethylamine, 21D [m/z (rel. int.) 283 (M^+, 4), 268 (13), 178 (45), 135 (100), 105 (15), 91 (43), 77 (11)] and (S,R)-N-(4-phenyl-2-butyl)-1-(3-methoxyphenyl)ethylamine, 21E.

Preparation of 21F

In a similar fashion an equal molar amount of 4-phenyl-2-butanone (Aldrich Chemical Co.), (R)-1-(1-naphthyl)ethylamine and 1.25 equivalents titanium (IV) isopropoxide were mixed and the intermediate imine reduced with ethanolic sodium cyanoborohydride. Work-up and repetitive preparative thin-layer chromatography using 5% methanol in chloroform afforded (R,R)-N-(4-phenyl-2-butyl)-1-(1-naphthyl)ethylamine, 21F; m/z (rel. int.) 303 (M^+, 6), 288 (14), 198 (22), 155 (100), 129 (8), 115 (5), 91 (19), 77 (4).

Preparation of 12Z

A stirred solution of 2-chlorohydrocinnamonitrile (Aldrich Chemical Co., 1.66 g, 10 mmol) in dichloromethane (100 ml) was cooled to -78°C and treated dropwise with diisobutylaluminum hydride (1.42 g, 10 mmol). The reaction was stirred 1 hr at rt, cooled to -78 °C and treated with a solution of 1-(1-naphthyl)ethylamine (1.71 g, 10 mmol) in dichloromethane (25 ml). The reaction was transferred to an ice bath and stirred 2 hr. After this time the reaction was poured directly into a stirred solution of ethanolic sodium borohydride (50 ml of 0.2 M, 10 mmol). The mixture was stirred 30 min at rt and the excess sodium borohydride quenched by the addition of 10% HCl. The solution was then made basic by the addition of 10 *N* NaOH and transferred to a separatory funnel washing with diethyl ether (300 ml). The aqueous phase was removed and the remaining organic layer washed with 1 *N* NaOH (3 x 100 ml). The organic layer was dried over anhydrous magnesium sulfate, and concentrated to an oil. Chromatography of

this material through silica gel using a gradient of chloroform to 10% methanol-chloroform afforded 2.34g (72% yield) of (*R*)-N-[3-(2-chlorophenyl)propyl]-1-(1-naphthyl)ethylamine, 12Z, as a clear oil; m/z (rel. int.) 323 (M+, 2), 308 (63), 288 (7), 196 (5), 184 (5), 155 (100), 125 (24), 115 (8), 103 (4), 91 (3), 77 (7).

Preparation of 12B

In a similar fashion, 4-methylcinnamonitrile was treated with diisobutyl aluminum hydride and the intermediate aluminum-imine complex treated with (*R*)-1-(3-methoxyphenyl)ethylamine. The intermediate imine was treated with ethanolic sodium borohydride. Work-up and chromatography yielded (*R*)-*N*-[3-(4-methylphenyl)prop-2-enyl]-1-(3-methoxyphenyl)ethylamine, 12B, as a clear, colorless oil; m/z (rel. int.) 281 (M+, 6), 266 (5), 176 (27), 146 (75), 135 (63), 131 (100), 115 (25), 105 (21), 91 (21), 77 (21).

Preparation of 12C

In a similar fashion, 2-methylcinnamonitrile was treated with diisobutyl aluminum hydride and the intermediate aluminum-imine complex treated with (*R*)-1-(3-methoxyphenyl)ethylamine. The intermediate imine was treated with ethanolic sodium borohydride. Work-up and chromatography yielded (*R*)-*N*-[3-(2-methylphenyl)prop-2-enyl]-1-(3-methoxyphenyl)ethylamine, 12C, as a clear, colorless oil; m/z (rel. int.) 281 (M+, 4), 266 (15), 176 (18), 146 (62), 135 (58), 131 (100), 115 (23), 105 (19), 91 (38), 77 (17).

Preparation of 12D

In a similar fashion, 2,4,6-trimethylcinnamonitrile was treated with diisobutyl aluminum hydride and the intermediate aluminum-imine complex treated with (*R*)-1-(3-methoxyphenyl)ethylamine. The intermediate imine was treated with ethanolic sodium borohydride. Work-up and

chromatography yielded (*R*)-*N*-[3-(2,4,6-trimethylphenyl) prop-2-enyl]-1-(3-methoxyphenyl)ethylamine, 12D, as a clear, colorless oil; m/z (rel. int.) 309 (M+,8), 294 (25), 174 (82), 159 (100), 135 (52), 129 (29), 105 (21), 91 (17), 77 (14).

Preparation of 12E

In a similar fashion, 4-isopropylcinnamonitrile was treated with diisobutyl aluminum hydride and the intermediate aluminum-imine complex treated with (R)-1-(3-methoxyphenyl)ethylamine. The intermediate imine was treated with ethanolic sodium borohydride. Work-up and chromatography yielded (R)-N-[3-(4-isopropylphenyl)prop-2-enyl]-1-(3-methoxyphenyl)ethylamine, 12E, as a clear, colorless oil; m/z (rel. int.) 309 (M^+, 9), 294 (7), 174 (98), 159 (22), 135 (80), 117 (100), 105 (35), 91 (37), 77 (19).

Preparation of 12F

In a similar fashion, 2,4-dimethylcinnamonitrile was treated with diisobutyl aluminum hydride and the intermediate aluminum-imine complex treated with (R)-1-(3-methoxyphenyl)ethylamine. The intermediate imine was treated with ethanolic sodium borohydride. Work-up and chromatography yielded (R)-N-[3-(2,4-dimethylphenyl)prop-2-enyl]-1-(3-methoxyphenyl)ethylamine, 12F, as a clear, colorless oil; m/z (rel. int.) 295 (M^+, 8), 294 (15), 174 (29), 160 (75), 145 (100), 135 (68), 117 (21), 105 (30), 91 (26), 77 (19).

Preparation of 12G

In a similar fashion, 3-methylcinnamonitrile was treated with diisobutyl aluminum hydride and the intermediate aluminum-imine complex treated with (R)-1-(3-methoxyphenyl)ethylamine. The intermediate imine was treated with ethanolic sodium borohydride. Work-up and chromatography yielded (R)-N-[3-(3-methylphenyl)prop-2-

enyl]-1-(3-methoxyphenyl)ethylamine, 12G, as a clear, colorless oil; m/z (rel. int.) 281 (M+, 5), 266 (9), 176 (24), 146 (71), 135 (62), 131 (100), 115 (23), 105 (19), 91 (41), 77 (18).

Preparation of 25E

In a similar fashion, cinnamonitrile was treated with diisobutyl aluminum hydride and the intermediate aluminum-imine complex treated with (R)-1-(3-methoxyphenyl)ethylamine. The intermediate imine was treated with ethanolic sodium borohydride. Work-up and chromatography yielded (R)-N-(3-phenylprop-2-enyl)-1-(3-methoxyphenyl)ethylamine, 25E, as a clear colorless oil; m/z (rel. int.) 267 (M+, 3), 252 (14), 176 (17), 135 (62), 117 (100), 105 (28), 91 (56), 77 (33).

Preparation of 25G

In a similar fashion, α-methylcinnamonitrile was treated with diisobutyl aluminum hydride and the intermediate aluminum-imine complex treated with (R)-1-(3-methoxyphenyl)ethylamine. The intermediate imine was treated with ethanolic sodium borohydride. Work-up and chromatography yielded *(R)-N*-(2-methyl-3-phenylprop-2-enyl)-1-(3-methoxyphenyl)ethylamine, 25G, as a clear, colorless oil; m/z (rel. int.) 281 (M+,5), 266 (18), 190 (12), 146 (78), 135 (82), 131 (100), 115 (21), 105 (21), 91 (62), 77 (19).

Preparation of 6X

A stirred solution of sodium hydride (1.8 g, 75 mmol) in dimethylformamide (150 ml) was treated with a solution of diethylcyanomethyl phosphonate (13.3 g, 75 mmol) in dimethylformamide (50 ml). The reaction was stirred 30 min at rt. After this time the reaction was treated with 3-chlorobenzaldehyde (10.54 g, 75 mmol) and stirred 1 hr at rt and 30 min at 60°C. The reaction was then quenched by the addition of water (200 ml). The reaction mixture

was transferred to a separatory funnel using diethyl ether (300 ml) and the resulting organic phase washed with water (5 x 300 ml) and brine. The organic layer was dried over anhydrous potassium carbonate and concentrated to yield 3-chlorocinnamonitrile (11.06 g) as a solid. The solid was dissolved in tetrahydrofuran (50 ml) and treated with excess diborane and stirred 30 min at rt. The reaction was poured over ice/10% HCl. The acidic aqueous phase was washed with diethyl ether (2 x 200 ml). The aqueous phase was made basic by the addition of 10 *N* NaOH and extracted with diethyl ether (200 ml). The ether extract was dried over anhydrous potassium carbonate and concentrated to afford 3-(3-chlorophenyl)propylamine as an oil (0.6 g, 3.54 mmol). The 3-(3-chlorophenyl)propylamine (0.60 g, 3.54 mmol), 3'-methoxyacetophenone (0.53 g, 3.54 mmol) and 1.25 molar equivalents titanium (IV) isopropoxide (1.26 g, 4.43 mmol) were mixed 4 hr at rt and the intermediate imine treated with an ethanolic sodium cyanoborohydride (5 ml of 1 M, 5 mmol). The reaction was stirred 16 hr at rt, diluted with diethyl ether (50 ml) and treated with water (0.32 ml, 17.7 mmol). After mixing thoroughly the solution was centrifuged and the ether layer concentrated to a milky solid. This material was suspended in diethyl ether and filtered through a 0.45 μM CR PTFE Acrodisc. The ether wash was concentrated to an oil. Chromatography of this material (silica, preparative thin-layer chromatography) using 3% methanol-dichloromethane (containing 0.1% isopropylamine) afforded *N*-[3-(3-chlorophenyl)propyl]-1-(3-methoxyphenyl)ethylamine, 6X; m/z (rel. int.) 303 (M+, 3), 288 (40), 196 (3), 164 (8), 135 (100), 125 (46), 103 (26), 91 (29), 77 (29).

Preparation of 6V

An equal molar amount of 3-(4-chlorophenyl) propylamine (prepared in a similar fashion from 4-chlorobenzaldehyde as above) 3'-methoxyacetophenone and 1.25 molar equivalents titanium (IV) isopropoxide were

mixed 4 hr at rt and the intermediate imine treated with an ethanolic sodium cyanoborohydride (5 ml of 1M, 5 mmol). Work-up and chromatography afforded *N*-[3-(4-chlorophenyl) propyl]-1-(3-methoxyphenyl)ethylamine, 6V, as an oil; m/z (rel. int.) 303 (M+,8), 288 (91), 196 (4), 164 (10), 135 (100), 125 (61), 103 (21), 91 (21), 77 (18).

Preparation of 20A

In a similar fashion, an equal molar amount of 1-(1-methoxyphenyl)ethylamine, 4-*t*-butylacetophenone and 1.25 molar equivalents titanium (IV) isopropoxide were mixed 4 hr at rt and the intermediate imine treated with an ethanolic sodium cyanoborohydride (5 ml of 1M, 5 mmol). Work-up and chromatography afforded (*R*)-*N*-[1-(4-*t*-butylphenyl) ethyl]-1-(1-naphthyl)ethylamine, 20A, as an oil; m/z (rel. int.) 331 (M+, 12), 316 (29), 161 (70), 155 (100), 131 (14), 127 (13), 115 (10), 105 (6), 91 (10), 77 (7).

Preparation of 25H and 25I

In a similar fashion, an equal molar amount of (*R*)-1-(3-methoxyphenyl)ethylamine, *trans*-4-phenyl-3-butene-2-one and 1.25 molar equivalents titanium (IV) isopropoxide were mixed 4 hr at rt and the intermediate imine treated with an ethanolic sodium cyanoborohydride (5 ml of 1 M, 5 mmol). Work-up and chromatography afforded (*R,R*)-*N*-(2-methyl-4-phenybut-3-enyl)-1-(3-methoxyphenyl)ethylamine, 25H, as an oil; m/z (rel. int.) 283 (M+, 4), 268 (13), 178 (40), 135 (100), 105 (15), 91 (47), 77 (13) and (*S,R*)-*N*-(2-methyl-4-phenybut-3-enyl)-1-(3-methoxyphenyl) ethylamine, 25I, as an oil; m/z (rel. int.) 283 (M+,4), 268 (13), 178 (40), 135 (100), 105 (15), 91 (47), 77 (13).

Preparation of 16L and 16M

In a similar fashion, an equal molar amount of (*R*)-1-(3-methoxyphenyl)ethylamine, 3-methoxyacetophenone and 1.25 molar equivalents titanium (IV) isopropoxide were mixed 4 hr at rt and the intermediate imine treated with

an ethanolic sodium cyanoborohydride (5 ml of 1 M, 5 mmol). Work-up and chromatography afforded (*R,R*)-*N*-[1-(4-methoxyphenyl)ethyl]-1-(3-methoxyphenyl)ethylamine, 16L, as an oil; m/z (rel. int.) 284 (M-1, 1), 270 (85), 150 (83), 135 (100), 120 (12), 105 (28), 91 (25), 77 (23) and (*S,R*)-*N*-[1-(4-methoxyphenyl)ethyl]-1-(3-methoxyphenyl) ethylamine, 16M, as an oil; m/z (rel. int.) 284 (M-1, 1), 270 (53), 150 (98), 135 (100), 120 (11), 105 (33), 91 (25), 77 (23).

Preparation of 5B/5C

In a similar fashion, 4-chloroacetophenone was used to prepare 3-methyl-3-(4-chlorophenyl)cinnamonitrile. The nitrile was catalytically reduced (palladium hydroxide, acetic acid, 60 p.s.i. hydrogen 2 hr) to generate 3-methyl-3-(4-chlorophenyl)propylamine. An equal molar amount of the amine, 3'-methoxyacetophenone and 1.25 molar equivalents titanium (IV) isopropoxide were mixed 4 hr at rt and the intermediate imine treated with an ethanolic sodium cyanoborohydride (5 ml of 1 M, 5 mmol). Work-up and chromatography afforded *N*-(3-methyl-3-(4-chlorophenyl)propyl]-1-(3-methoxyphenyl)ethylamine, 5B/5C as an oil; m/z (rel. int.) 317 (M+, 12), 302 (74), 210 (2), 182 (4), 164 (12), 135 (100), 121 (25), 103 (40), 91 (19), 77 (28).

Preparation of 4Z/5A

In a similar fashion, 3-chloroacetophenone was used to prepare 3-methyl-3-(3-chlorophenyl)cinnamonitrile. The nitrile was catalytically reduced (palladium hydroxide, acetic acid, 60 p.s.i. hydrogen 2 hr) to generate 3-methyl-3-(3-chlorophenyl)propylamine. An equal molar amount of the amine, 3'-methoxyacetophenone and 1.25 molar equivalents titanium (IV) isopropoxide were mixed 4 hr at rt and the intermediate imine treated with an ethanolic sodium cyanoborohydride (5 ml of 1 M, 5 mmol). Work-up and chromatography afforded *N*-[3-methyl-3-(3-chlorophenyl)