The Journal of

Antimicrobial

Chemotherapy

WESTON LIBRARY

AUG 2 1 1998

J5/120 GLYRIGAT SCIENCES CENTER
600 HIGHLAND AV-MADISON, WI 53792



The British Society for Antimicrobial Chemotherapy

Volume 42

Number 2

August 1998

OXFORD UNIVERSITY PRESS

CFAD v. Anacor, IPR2015-01776 ANACOR EX. 2096 - 1/9

UK BACTERIAL ENDOCARDITIS WORKSHOPS

October/November 1998

The British Society for Antimicrobial Chemotherapy (BSAC), in collaboration with the Association of Medical Microbiologists (AMM) and the Hospital Infection Society (HIS), is holding a series of educational workshops focusing on bacterial endocarditis.

Dulwich - Monday 19 October Darlington – Wednesday 21 October Chester - Thursday 22 October

Bristol - Friday 23 October Derby - Wednesday 28 October Peterborough - Thursday 5 November Reading - Tuesday 10 November Dublin - Date to be advised Stirling - Date to be advised

Registration Fee: £25.00 (BSAC/AMM/HIS members), £100.00 (non-members)

The meetings will commence with a buffet lunch at 12 noon. The workshops will begin at 1.00pm and will close at approximately 4.30pm.

To register or for further information please contact:



Organising Secretariat: Nicole Robert/Kate Auty Gardiner-Caldwell Communications Ltd Victoria Mill, Windmill Street, Macclesfield, Cheshire SK11 7HQ, UK Tel: +44 (0)1625 664200 Fax: +44 (0)1625 664016

Association of Medical Microbiologists



The Journal of Antimicrobial Chemotherapy

Subscription information

A subscription to The Journal of Antimicrobial Chemotherapy comprises of 12 issues plus supplements, with an Annual Author and Subject Index. Subscriptions are entered on a calender year basis only. Please add sales tax to prices quoted. Price includes postage by surface mail or for subscribers in the USA and Canada by air freight or in India, Japan, Australia and New Zealand, by Air Speeded Post. Airmail rates are available on request.

Annual subscription rate (Volumes 41-42, 1998)

Insitutions: Europe and UK £375.00, Rest of World US\$670.00 Individuals*: Europe and UK £210.00, Rest of World US\$350.00 Single issue: Europe and UK £46.00, Rest of World US\$74.00 *Individual rates apply only when copies are sent to a private address and payment is made personal by cheque or credit card (American

Express, Diners, Mastercard, Visa, JCB). Back volume prices are available on request.

Orders. Orders and payments from, or on behalf of, subscribers in the various geographical areas shown below should be sent to the office indicated. The Americas: Oxford University Press, Distibution and Information Systems, 2001 Evans Road, Cary, North Carolina 27513; USA

Japan: available from the following agents: Kinokuniya Company Ltd, Journal Department, PO Box 55, Chitose, Tokyo, 156 Japan; Maruzen Company Ltd, Journal Division, PO Box 5050, Tokyo International 100-31 Japan; Usaco Corporation, 13-12, Shimbashi 1-chome, Minato-ku, Tokyo, 105 Japan.

Rest of World: Journals Subscriptions Department, Oxford University Press, Great Clarendon Street Oxford, OX2 6DP, UK

Tel: +44(0)1865 267907; telex: OXPRESS 873330; fax +44(0)1865 267485.

Advertising

To advertise in The Journal of Antimicrobial Chemotherapy, contact Peter Carpenter, Prc Associates, The Annexe, Fitznells Manor, Chessington Road, Ewell Village, Surrey KT17 1TF.

Tel:+ 44(0)181 786 7376, fax: +44(0)181 786 7262.

©The British Society of Antimicrobial Chemotherapy 1998.

All rights reserved; no part of this publication may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, recording, or otherwise without either the prior written permission of the Publishers, or a license permitting restricted copying issued in the UK by the Copyright Licensing Agency Ltd, 90 Tottenham Court Road, London W1P 9HE, or in the USA by the Copyright Clearance Centre, 222 Rosewood Drive, Danvers, MA 01923. For those in the US/Canada not registered with CCC, articals can be obtained by fax in 48 hours by calling: WISE for Medicine™ 1-800-667-Wise. Special requests, such as copying for general distribution or for advertising or promotional purposes, should be addressed to the Production Editor, The Journal of Antimicrobial Chemotherapy, Oxford University Press, Great Clarendon Street, Oxford, OX2 6DP, UK.

The Journal of Antimicrobial Chemotherapy (ISSN 0305-7453) is published monthly by Oxford University Press, Oxford UK. Annual subscription price is US\$670.00. The Journal of Antimicrobial Chemotherapy is distributed by M.A.I.L. America, 2323 Randolph Avenue, Avenel, New Jersey, NY 07001, USA. Periodical postage paid at Rahway, New Jersey and additional mailing entry points.

US Postmaster: send address changes to The Journal of Antimicrobial Chemotherapy, c/o M.A.I.L. America, 2323 Randolph Avenue, Avenel, New Jersey, NY 07001, USA.

Journal disclaimer. All reasonable precautions have been taken by the authors, editors and publishers to verify drug names and doses, the results of experimental work and the clinical findings published in this journal. The opinions expressed are those of the authors, and not necessarily those of the editors or publishers. The ultimate responsibility for the use and dosage of drugs mentioned in the Journal and in the interpretation of published material lies with the medical practitioner and the editors and publishers can accept no liability whatsoever in respect of any claim for damages arising therefrom. Please inform the editors of any errors.

Printed by Bell and Bain Ltd, Glasgow



The Journal of Antimicrobial Chemotherapy

Volume 42 Number 2 August 1998

Contents

Antibiotic resistance R. G. Finch	125
Extended-spectrum β -lactamases in P seudomonas aeruginosa P . Nordmann and M . Guibert	128
Original articles Interactions of plaunotol with bacterial membranes T. Koga, H. Watanabe, H. Kawada, K. Takahashi, Y. Utsui, H. Domon, C. Ishii, T. Narita and H. Yasuda	133
In-vitro activity of lytic peptides, inhibitors of ion transport systems and ionophorous antibiotics against Pneumocystis carinii O. Cirioni, A. Giacometti, F. Barchiesi and G. Scalise	141
Influence of ciprofloxacin and other antimicrobial drugs on different <i>Escherichia coli</i> strains in continuous-flow cultures under aerobic and anaerobic conditions <i>H. Bernhardt, K. Schulz, K. Zimmermann and M. Knoke</i>	147
Increasing resistance of planktonic and biofilm cultures of <i>Burkholderia cepacia</i> to ciprofloxacin and ceftazidime during exponential growth M. Desai, T. Bühler, P. H. Weller and M. R. W. Brown	153
Comparison of the modified Stokes' method of susceptibility testing with results obtained using MIC methods and British Society of Antimicrobial Chemotherapy breakpoints P. E. Gosden, J. M. Andrews, K. E. Bowker, H. A. Holt, A. P. MacGowan, D. S. Reeves, J. Sunderland and R. Wise	16 1
In-vitro investigation of the antibacterial activity of agents which may be used for the oral treatment of lung infections in CF patients R. M. E. Richards, V. E. S. Hamilton and M. R. Thomas	171
The effects of increasing levels of quinolone resistance on in-vitro activity of four quinolones <i>K. S. Thomson and C. C. Sanders</i>	179
Glycopeptide tolerance in Staphylococcus aureus J. May, K. Shannon, A. King and G. French	189
Activated cell-wall synthesis is associated with vancomycin resistance in methicillin-resistant Staphylococcus aureus clinical strains Mu3 and Mu50 H. Hanaki, K. Kuwahara-Arai, S. Boyle-Vavra, R. S. Daum, H. Labischinski and K. Hiramatsu	199
The effect of a component of tea ($Camellia\ sinensis$) on methicillin resistance, PBP2' synthesis, and β -lactamase production in $Staphylococcus\ aureus$ $T.\ S.\ Yam,\ J.\ M.\ T.\ Hamilton-Miller\ and\ S.\ Shah$	211
In-vitro susceptibility of <i>Cryptococcus neoformans</i> isolates to fluconazole and itraconazole K.G. Davey, F. M. Johnson, A. D. Holmes, A. Szekely and D. W. Warnock	217

The effect of dicloxacillin and fusidic acid on the extracellular and intracellular killing of <i>Staphylococcus aureus</i> S. L. Nielsen and F. T. Black	221
Bacterial concentrations in pus and infected peritoneal fluid—implications for bactericidal activity of antibiotics C. König, HP. Simmen and J. Blaser	227
Efficacy and safety of teicoplanin plus rifampicin in the treatment of bacteraemic infections caused by Staphylococcus aureus E. P. F. Yzerman, H. A. M. Boelens, M. Vogel and H. A. Verbrugh	233
Brief reports Diethylcarbamazine-related antimicrobial activity in <i>Mycobacterium tuberculosis</i> -infected blood L. W. Kitchen, C. M. Weston and S. P. Day	241
In-vitro antibiotic susceptibility and molecular analysis of anaerobic bacteria isolated in Cape Town, South Africa C. L. Koch, P. Derby and V. R. Abratt	245
Sub-MICs of sanfetrinem promote the interaction of human polymorphonuclear granulocytes with a multiply resistant strain of <i>Klebsiella pneumoniae</i> A. M. Cuffini, V. Tullio, A. I. Palarchio, A. Bonino and N. A. Carlone	249
Voriconazole against fluconazole-susceptible and resistant candida isolates: in-vitro efficacy compared with that of itraconazole and ketoconazole M. H. Nguyen and C. Y. Yu	253
Comparison of four antibiotics in a murine model of necrotizing cutaneous infections caused by toxigenic Streptococcus pyogenes and Staphylococcus aureus N. Barg	257
Comparative grepafloxacin phototoxicity in mouse skin K. Owen	261
Correspondence Current MIC breakpoints may understate the potential efficacies of carbapenems for treatment of patients with infections caused by strains of <i>Streptococcus pneumoniae</i> that are resistant or of intermediate susceptibility to penicillin J. R. Edwards, J. S. Bradley and K. P. Klugman	265
Study on the in-vitro activity of LY333328 against Gram-positive cocci	266
Activities of cefepime and five other antibiotics against nosocomial PER-1-type and/or OXA-10-type β -lactamase-producing <i>Pseudomonas aeruginosa</i> and <i>Acinetobacter</i> spp. H. Vahaboglu, S. Sarıbaş, H. Akbal, R. Ozturk and A. Yucel	269
Evaluation of the activities of two-drug combinations of rifampicin, polymyxin B and ampicillin/sulbactam against <i>Acinetobacter baumannii C. Tascini, F. Menichetti, S. Bozza, A. Del Favero and F. Bistoni</i>	270
A study of the mechanisms involved in imipenem resistance in <i>Pseudomonas aeruginosa</i> isolates from Japan <i>R. A. Stunt, C. J. Thomson, D. J. Payne and S. G. B. Amyes</i>	272
Emergence of resistance to third-generation cephalosporins amongst <i>Salmonella typhimurium</i> isolates in Greece: report of the first three cases L. S. Tzouvelekis, M. Gazouli, A. Markogiannakis, E. Paraskaki, N. J. Legakis and E. Tzelepi	273
Isolation of glycopeptide resistant <i>Streptococcus gallolyticus</i> strains with <i>vanA</i> , <i>vanB</i> , and both <i>vanA</i> and <i>vanB</i> genotypes from faecal samples of veal calves in The Netherlands <i>D. Mevius, L. Devriese, P. Butaye, P. Vandamme, M. Verschure and K. Veldman</i>	275

Book reviews	281
8-methoxyquinolone C. M. Tobin, J. Sunderland, L. O. White, A. P. MacGowan and D. S. Reeves	278
An isocratic high performance liquid chromatography (HPLC) assay for moxifloxacin, a new	
Activity of nisin against <i>Streptococcus pneumoniae</i> , in vitro, and in a mouse infection model B. P. Goldstein, J. Wei, K. Greenberg and R. Novick	277



Voriconazole against fluconazole-susceptible and resistant candida isolates: in-vitro efficacy compared with that of itraconazole and ketoconazole

M. Hong Nguyen^{a,b}* and Christine Y. Yu^a

^aDepartment of Medicine, Division of Infectious Disease, University of Florida College of Medicine, PO Box 100277, JHMHC, Gainesville, FL 32610; ^bVA Medical Center, Gainesville, FL, USA

We compared the in-vitro activity of fluconazole, itraconazole, ketoconazole and voriconazole against 67 blood isolates of *Candida* spp. exhibiting a wide range of fluconazole MICs (0.125 to >64 mg/L). Voriconazole was the most potent *in vitro*, followed by itraconazole, ketoconazole and fluconazole. Itraconazole and voriconazole had in-vitro activity against fluconazole-susceptible and -resistant candida isolates. Higher itraconazole and voriconazole MICs were observed in isolates exhibiting higher fluconazole MICs, suggesting cross-resistance. Itraconazole and voriconazole MICs of ≥16 mg/L were observed only in *Candida albicans* and *Candida tropicalis*. *Candida krusei* and *Candida glabrata* exhibited itraconazole MICs of 0.5–1 mg/L and voriconazole MICs of 0.25–0.5 mg/L.

Introduction

Voriconazole is a new triazole antifungal agent which acts by inhibiting cytochrome P450 sterol 14α -demethylase, an enzyme involved in ergosterol biosynthesis. Voriconazole has potent in-vitro and in-vivo activity against *Aspergillus* spp. and other moulds. ¹⁻³ Although voriconazole has in-vitro activity against fluconazole-resistant *Candida albicans*, *Candida krusei* and *Candida glabrata*, ^{4,5} its activity against other *Candida* spp. that are fluconazole-resistant *in vitro* is unknown. Furthermore, the in-vitro activity of voriconazole has not been compared with that of itraconazole and ketoconazole. The goal of this study was to compare the in-vitro activity of fluconazole, itraconazole, ketoconazole and voriconazole against a large number of candida isolates; the isolates studied exhibited a wide range of fluconazole MICs.

Materials and methods

Sixty-seven blood isolates of *Candida* spp. collected during a prospective study of candidaemia were tested.⁶ These isolates exhibited fluconazole MICs ranging from 0.125 to >64 mg/L. These included *C. albicans* (24 isolates), *Candida tropicalis* (17), *C. glabrata* (12), *Candida parapsilosis* (8), *C. krusei* (3) and *Candida lusitaniae* (3). *C.*

parapsilosis ATCC 90018, C. albicans ATCC 90028 and 90029 and C. glabrata ATCC 90030 were incorporated into each set of experiments as quality control isolates.

The susceptibility testing was performed by a macrodilution method adhering to the National Committee for Clinical Laboratory Standards (NCCLS) protocol.⁷ Fluconazole (Pfizer Central Research, Groton, CN, USA) stock solutions of 2000 mg/L were prepared with sterile distilled water. Voriconazole (Pfizer Central Research, Groton, CN, USA) stock solutions of 4000 mg/L were prepared with dimethylsulphoxide (DMSO); subsequent dilutions were performed in water. Stock solutions of ketoconazole and itraconazole (Janssen Research Foundation, Beerse, Belgium) were prepared with 0.2 N HCl and DMSO, respectively; subsequent drug dilutions were performed according to the manufacturer's protocol. The concentrations of drugs tested were: 0.125-64 mg/L for fluconazole; 0.015-16 mg/L for itraconazole and voriconazole; and 0.03-16 mg/L for ketoconazole. Each Candida sp. was tested simultaneously against fluconazole, itraconazole, ketoconazole and voriconazole.

Results and discussion

The fluconazole, itraconazole, ketoconazole and voriconazole MICs for the ATCC isolates were: 0.5, 0.125, 0.06

*Tel: +1-352-3794027; Fax: +1-352-3794015; E-mail: nguyemt@medicine.ufl.edu

and 0.015 mg/L, respectively, for ATCC 90018; 0.5, 0.125, 0.06 and 0.03 mg/L, respectively for ATCC 90028; 0.5, 0.125, 0.03 and 0.03 mg/L, respectively, for ATCC 90029; 16, 0.125, 0.03 and 0.03 mg/L, respectively, for ATCC 90030.

The MIC ranges, MIC₅₀s, MIC₉₀s and geometric mean MICs of ketoconazole, fluconazole, itraconazole and voriconazole for specific *Candida* spp. are presented in Table I. Using the fluconazole breakpoint values proposed by the NCCLS,⁸ 69% (46/67) of *Candida* spp. were susceptible, 9% (6/67) dose-dependently susceptible and 22% (15/67) resistant to fluconazole *in vitro*. Using the itraconazole breakpoints,⁸ 40% (27/67) were susceptible, 40% (27/67) dose-dependently susceptible and 20% (13/67) resistant to itraconazole *in vitro*.

To our knowledge, this is the first published study to compare the in-vitro efficacy of voriconazole, itraconazole and ketoconazole against a large number of *Candida* spp. with a wide range of fluconazole MICs. We included in our study not only fluconazole-resistant *C. albicans* and *C. krusei*, but also *C. tropicalis*, *C. parapsilosis* and *C. lusitaniae*. We demonstrated that voriconazole was the most potent of the azole agents against the *Candida* spp.

tested (geometric mean of 0.12 mg/L), followed by itraconazole (geometric mean of 0.30 mg/L) and ketoconazole (geometric mean of 0.75 mg/L).

Voriconazole had in-vitro activity against both fluconazole-susceptible and -resistant *Candida* spp. For fluconazole-susceptible isolates, voriconazole was significantly more potent than itraconazole and ketoconazole: the geometric mean MIC of voriconazole (0.04 mg/L) was significantly lower than that of itraconazole (0.17 mg/L; P < 0.001) and that of ketoconazole (0.43 mg/L; P < 0.001) (Table II). Moreover, 91% (42/46) of the fluconazole-susceptible *Candida* spp. exhibited voriconazole MICs of ≤ 0.125 mg/L, whereas only 52% (24/46) exhibited itraconazole MICs ≤ 0.125 mg/L, and 48% (21/46) exhibited ketoconazole MICs ≤ 0.125 mg/L.

For fluconazole-resistant or dose-dependently susceptible isolates, voriconazole also demonstrated good in-vitro activity. Sixty-two percent (13/21) of these isolates exhibited voriconazole MICs of \leq 0.5 mg/L, whereas only 43% (9/21) exhibited itraconazole MICs of \leq 0.5 mg/L, and 19% (4/21) exhibited ketoconazole MICs of \leq 0.5 mg/L. As previously noted, *C. krusei* and *C. glabrata*, species often associated with fluconazole resistance, were sus-

Table I. In-vitro activity of ketoconazole, fluconazole, itraconazole and voriconazole against *Candida* spp.

Species	n	Antimicrobial agent	48 h MIC (mg/L)				
			range	50%	90%	geometric mean	
C. albicans	24	ketoconazole	0.03->16	8	>16	1.10	
		fluconazole	0.125 - > 64	0.5	>64	1.30	
		itraconazole	0.06 - > 16	0.125	0.5	0.22	
		voriconazole	≤0.015->16	≤0.015	0.25	0.06	
C. tropicalis	17	ketoconazole	0.03 - > 16	4	>16	1.75	
		fluconazole	0.5 - > 64	8	>64	9.02	
		itraconazole	0.015 - > 16	0.25	>16	0.54	
		voriconazole	≤0.015->16	0.125	>16	0.33	
C. glabrata	12	ketoconazole	0.03-1	1	1	0.47	
		fluconazole	2–32	8	32	8.00	
		itraconazole	0.25-1	0.5	1	0.56	
		voriconazole	0.06-0.5	0.125	0.25	0.16	
C. parapsilosis	8	ketoconazole	0.03-1	0.125	1	0.19	
		fluconazole	0.5 - > 64	2	32	3.35	
		itraconazole	0.125 - 0.5	0.125	0.25	0.19	
		voriconazole	0.015-1	0.03	0.25	0.06	
C. krusei	3	ketoconazole	0.5-1	0.5	0.5	0.63	
		fluconazole	>64	>64	>64	64.07	
		itraconazole	0.25 - 0.5	0.5	0.5	0.40	
		voriconazole	0.5	0.5	0.5	0.50	
C. lusitaniae	3	ketoconazole	0.03-0.5	0.03	0.5	0.08	
		fluconazole	0.125 - 32	2	32	1.99	
		itraconazole	0.125 - 0.5	0.125	0.5	0.20	
		voriconazole	0.015 - 0.5	0.015	0.5	0.06	

Candida susceptibility to voriconazole in vitro

Table II. Geometric means, MIC₅₀ and MIC₉₀ for fluconazole-susceptible and -resistant *Candida* species against itraconazole, ketoconazole and voriconazole

Azole agent	Geometric mean MIC (1	ole:	MIC_{50}/MIC_{90} (mg/L) for fluconazole		
	susceptible isolates	resistant isolates	P value	susceptible isolates	resistant isolates
Itraconazole		į.			
all Candida spp	. 0.17	1.39	< 0.001	0.125/0.5	1/16
C. albicans	0.13	3.97	< 0.001	0.125/0.25	16/16
C. tropicalis	0.13	2.51	0.001	0.125/0.25	1/16
C. glabrata	0.46	1.00	0.01	0.5/0.5	1/1
C. parapsilosis	0.18	0.25	NS	0.125/0.25	0.125/0.5
C. krusei	_	0.40	_	– ,	0.5/0.5
Ketoconazole					
all Candida spp	. 0.43	2.51	0.003	0.25/16	4/16
C. albicans	0.70	11.36	0.06	0.5/16	8/16
C. tropicalis	1.06	3.06	NS	0.5/16	8/16
C. glabrata	0.37	1.00	NS	0.5/1	1/1
C. parapsilosis	0.11	1.00	0.01	0.125/0.25	1/1
C. krusei	-	0.63	_	_	0.5/0.5
Voriconazole					
all Candida spp	. 0.04	1.14	< 0.001	0.03/0.125	0.5/16
C. albicans	0.02	5.64	< 0.001	0.015/0.06	16/16
C. tropicalis	0.08	1.68	0.003	0.06/0.125	0.25/16
C. glabrata	0.13	0.25	NS	0.125/0.25	0.25/0.5
C. parapsilosis	0.03	0.50	0.002	0.03/0.06	0.25/1
C. krusei	_	0.50	_	_	0.5/0.5

NS, not significant.

ceptible *in vitro* to itraconazole, ^{9–10} to voriconazole⁴ with MICs of 0.25–0.5 mg/L, and to ketoconazole with MICs of 0.5–1 mg/L (Table II).

Despite these promising results, there was cross-resistance between fluconazole and voriconazole for some *Candida* spp. For example, isolates with higher fluconazole MICs were associated with higher voriconazole MICs (P < 0.001, linear regression). There was also cross-resistance between fluconazole, itraconazole and keto-conazole: the higher fluconazole MICs were associated with higher itraconazole and keto-conazole MICs (P < 0.001, and 0.003, respectively). This pattern of cross-resistance has been previously described, and may result from the similar mechanisms of actions of these agents. 9,10

Six (38%) of the 16 Candida spp. with fluconazole MICs of \geq 64 mg/L displayed itraconazole MICs of \geq 16 mg/L and ketoconazole MICs of \geq 8 mg/L. All of these isolates had voriconazole MICs of \geq 16 mg/L. In our study, these high levels of resistance to multiple azole agents (MICs \geq 8 mg/L) were seen only for C. albicans and C. tropicalis isolates. Fluconazole-resistant C. krusei, C. glabrata, C. parapsilosis and C. lusitaniae isolates, on the other hand, did not display high-level resistance to itraconazole, ketoconazole and voriconazole.

In conclusion, voriconazole has potent in-vitro activity against *Candida* spp., including those that were dose-dependently fluconazole-susceptible or fluconazole-resistant. This finding suggests that voriconazole might be effective in the treatment of refractory candidosis caused by fluconazole-resistant strains. However, cross-resistance with fluconazole exists in a small subset of *Candida* spp. Given the high oral bioavailability and the well tolerated nature of voriconazole, this drug may become an important addition to the armamentarium of systemic antifungal agents. This promise, however, requires to be confirmed in the clinical setting.

References

- **1.** McGinnis, M. R., Pasarell, L., Sutton, D. A., Fothergill, A. W., Cooper, C. R. & Rinaldi, M. G. (1997). In-vitro evaluation of voriconazole against some clinically important fungi. *Antimicrobial Agents and Chemotherapy* **41**, 1832–4.
- **2.** George, D., Miniter, P. & Andriole, V. T. (1996). Efficacy of UK-109496, a new azole antifungal agent, in an experimental model of invasive aspergillosis. *Antimicrobial Agents and Chemotherapy* **40**, 86–91.

- **3.** Radford, S. A., Johnson, E. M. & Warnock, D. W. (1997). In-vitro studies of activity of voriconazole (UK-109,496), a new triazole antifungal agent, against emerging and less common mold pathogens. *Antimicrobial Agents and Chemotherapy* **41**, 841–3.
- **4.** Barry, A. L. & Brown, S. D. (1996). In-vitro studies of two triazole antifungal agents (voriconazole [UK-109,496] and fluconazole) against *Candida* species. *Antimicrobial Agents and Chemotherapy* **40**, 1948–9
- **5.** Ruhnke, M., Schmidt-Westhausen, A. & Trautmann, M. (1997). In-vitro activities of voriconazole (UK-109,496) against fluconazole-susceptible and -resistant *Candida albicans* isolates from oral cavities of patients with human immunodeficiency virus infection. *Antimicrobial Agents and Chemotherapy* **41**, 575–7.
- **6.** Nguyen, M. H., Peacock, J. E., Morris, A. J., Tanner, D. C., Nguyen, M. L., Snydman, D. C. *et al.* (1996). The changing face of candidemia: emergence of non-*Candida albicans* species and antifungal resistance. *American Journal of Medicine* **100**. 617–23.
- 7. National Committee for Clinical Laboratory Standards. (1995). Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts. Tentative Standard M27-T. NCCLS, Wayne, PA.

- **8.** Rex, J. H., Pfaller, M. A., Galgiani, J. N., Bartlett, M. S., Espinel-Ingroff, A., Ghannoum, M. A. *et al.* (1997). Development of interpretive breakpoints for antifungal susceptibility testing: conceptual framework and analysis of *in vitro–in vivo* correlation data for fluconazole, itraconazole, and candida infections. *Clinical Infectious Diseases* **24**, 235–47.
- **9.** Barchiesi, F., Colombo, A. L., McGough, D. A., Fothergill, A. W. & Rinaldi, M. G. (1994). In-vitro activity of itraconazole against fluconazole-susceptible and -resistant *Candida albicans* isolates from oral cavities of patients infected with human immunodeficiency virus. *Antimicrobial Agents and Chemotherapy* **38**, 1530–3.
- **10.** St-Germain, G., Dion, C., Espinel-Ingroff, A., Ratelle, J. & de Repentigny, L. (1995). Ketoconazole and itraconazole susceptibility of *Candida albicans* isolated from patients infected with HIV. *Journal of Antimicrobial Chemotherapy* **36**, 109–18.

Received 17 October 1997; returned 21 January 1998; revised 12 February 1998; accepted 4 March 1998