# COMPETITIVE INHIBITION OF 3-HYDROXY-3-METHYLGLUTARYL COENZYME A REDUCTASE BY ML-236A AND ML-236B FUNGAL METABOLITES, HAVING HYPOCHOLESTEROLEMIC ACTIVITY

Akira ENDO, Masao KURODA and Kazuhiko TANZAWA

Fermentation Research Laboratories, Sankyo Co., Ltd., 1-2-58 Hiromachi, Shinagawa-ku, Tokyo 140, Japan

Received 15 November 1976

#### 1. Introduction

Fungal metabolites, ML-236A and ML-236B (fig.1), have been isolated from cultures of *Penicillium citrinum* as potent inhibitors of cholesterol synthesis in vitro in this laboratory [1].

These metabolites (LD<sub>50</sub> for mice > 2 g, per os) cause a marked decrease in serum cholesterol levels in rats [1], and in hens and dogs (Kitano, Tsujita and Endo, in preparation). The experiments reported in this paper demonstrate that ML-236A and ML-236B inhibit specifically 3-hydroxy-3-methylglutaryl (HMG)-CoA reductase (EC 1.1.1.34), the rate-limiting enzyme in cholesterol synthetic pathway, without affecting the rest of the enzymes involved in this pathway, and that the inhibition is competitive with respect to the substrate HMG-CoA.

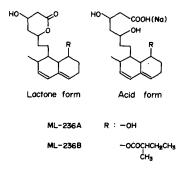


Fig.1. Structures of ML-236A and ML-236B.

North-Holland Publishing Company - Amsterdam

#### 2. Materials and methods

[1-<sup>14</sup>C] Acetate (59.5 mCi/mmol) and D,L-[2-<sup>14</sup>C] mevalonolactone (27.3 mCi/mmol) were obtained from Radiochemical Centre. [1-<sup>14</sup>C] Acetyl-CoA (49.8 mCi/mmol) and D,L-[3-<sup>14</sup>C] HMG-CoA (26.2 mCi/mmol) were purchased from New England Nuclear. Lactone forms of ML-236A and ML-236B were prepared as previously described [1]. Acid forms (sodium salts) (fig.1) of these compounds were prepared by saponification of their respective lactone forms in 0.1 N NaOH at 50°C for 2 h. Other chemicals were of the best grade commercially available.

# 2.1. Incorporation experiments

Rat liver microsomes and cytosolic enzyme fraction were isolated as described previously [2]. The reaction mixture (0.2 ml) contained: 1 mM ATP, 10 mM glucose-1-phosphate, 6 mM glutathione, 6 mM MgCl<sub>2</sub>,  $40 \mu M$  CoA, 0.25 mM NAD, 0.25 mM NADP, 100 mM potassium phosphate buffer (pH 7.4) 0.15 mg protein of microsomes, 1.5 mg protein of cytosolic enzyme fraction and 1 mM [1-14C] acetate (1.5 mCi/mmol). Where indicated, [1-14C] acetate was replaced by 0.15 mM [1-14C] acetyl-CoA (1.0 mCi/mmol), 0.13 mM D,L-[3-14C]HMG-CoA (7.3 mCi/mmol) or 0.52 mM D,L-[2-14C] mevalonate (1.26 mCi/mmol). After incubation at 37°C for 60 min, the reaction was terminated by addition of 1 ml 15% alcoholic KOH. The synthesized nonsaponifiable lipids and fatty acid were measured as described previously [2]. Under these conditions, the incorporations of radiolabeled substrates were proportional to time up to 120 min.

323



#### 2.2. HMG-CoA reductase assav

Rat liver microsomes were obtained as described previously [2], from which HMG-CoA reductase was solubilized by the method of Heller and Gould [3] and partially purified by fractionation with ammonium sulfate. The fraction precipitated by 35-50% saturation was used after dialysis for 3 h against 40 mM potassium phosphate buffer, pH 7.2, containing 100 mM sucrose, 50 mM KCl, 30 mM EDTA and 1 mM dithiothreitol. The reaction mixture (50  $\mu$ l) contained: 100 mM potassium phosphate buffer (pH 7.4) 10 mM EDTA, 10 mM dithiothreitol, 5 mM NADPH, 0.11 mM D,L-[3-14C]HMG-CoA (1.6 mCi/ mmol) and  $1-2 \mu g$  of enzyme protein. After incubation at 37°C for 20 min, the reaction was terminated by addition of 20 µl of 2 N HCl, and the mevalonolactone formed was isolated and counted as described previously [2]. The specific activity of the enzyme used was 10-17 nmol of mevalonate formed/min/mg protein under standard conditions.

Protein was determined by the method of Lowry et al. [4].

#### 3. Results

# 3.1. Incorporation of radiolabeled substrates into lipids

As reported in a previous paper [1], ML-236B lactone is far more inhibitory in the [1-14C] acetate incorporation into nonsaponifiable lipids than ML-236A lactone. The inhibitory potency was approximately doubled by the conversion of lactone forms to their respective acid forms (sodium salts). Of the four derivatives tested, ML-236B sodium salt was the most inhibitory. Concentrations required for 50% inhibition of nonsaponifiable lipid synthesis were: ML-236A 0.85  $\mu$ M, ML-236A sodium salt 0.35  $\mu$ M, ML-236B lactone 0.026  $\mu$ M, ML-236B sodium salt 0.014  $\mu$ M (0.006  $\mu$ g/ml). None of these compounds had significant effects on the fatty acid synthesis from [14C] acetate at concentrations which caused 50% or more reduction in the nonsaponifiable lipid synthesis.

Table 1 shows the effects of ML-236B sodium salt on the incorporation of various radiolabeled substrates into nonsaponifiable lipids. As indicated, conversions

Table 1
Inhibitory effects of ML-236B sodium salt on the incorporation of various radiolabeled substrates into nonsaponifiable lipids

| Substrate                           | ML-236B<br>sodium salt<br>(nM) | Incorporation dpm/mg <sup>a</sup> | % of control |
|-------------------------------------|--------------------------------|-----------------------------------|--------------|
| [1-14C] Acetate                     | None                           | 13 770                            |              |
|                                     | 5.0                            | 10 080                            | 73.2         |
|                                     | 50                             | 4120                              | 29.9         |
| [1- <sup>14</sup> C] Acetyl-CoA     | None                           | 8270                              |              |
|                                     | 5.0                            | 6020                              | 72.8         |
|                                     | 50                             | 2410                              | 29.2         |
| D,L-[3- <sup>14</sup> C]HMG-CoA     | None                           | 1050                              |              |
|                                     | 5.0                            | 570                               | 53.7         |
|                                     | 50                             | 270                               | 26.0         |
| D,L-[2- <sup>14</sup> C] Mevalonate | None                           | 35 870                            |              |
|                                     | 5.0                            | 34 940                            | 97.4         |
|                                     | 50                             | 34 180                            | 95.3         |

a Counts incorporated/mg protein/60 min. The values for [1-14C]acetate and [1-14C]acetyl-CoA were calculated on the assumption that 33% of the radioactivity in these substrates were converted into CO<sub>2</sub> during their incorporation into nonsaponifiable lipids.

Experimental conditions are described in Materials and methods. The results are the average of duplicate incubations.



of [14C] acetate, [14C] acetyl-CoA and D,L-[14C] HMG-CoA were inhibited to similar extent at two concentrations of the agent, 5 nM and 50 nM. On the other hand, D,L-[14C] mevalonate conversion into non-saponifiable lipids was not affected by ML-236B sodium salt at concentrations up to 50 nM, indicating that this compound inhibited specifically the enzymatic step for the conversion of HMG-CoA to mevalonate catalyzed by HMG-CoA reductase.

### 3.2. Inhibition of HMG-CoA reductase

As shown in fig.2, all the four compounds were inhibitory to HMG-CoA reductase. The acid forms (sodium salts) of both ML-236A and ML-236B were more effective in inhibiting the reductase than their respective lactone forms, and the two forms of ML-236B were more potent inhibitors than those of ML-236A. Concentrations required for 50% inhibition were: ML-236A lactone 3.4  $\mu$ M, ML-236A sodium salt 1.2  $\mu$ M, ML-236B lactone 0.10  $\mu$ M, ML 236B sodium salt 0.023  $\mu$ M. The data correlated well with the results obtained for the inhibition of nonsaponifiable lipid synthesis from [ $^{14}$ C]acetate, although higher concentrations of the compounds were required for inhibition of the reductase.

The inhibition of HMG-CoA reductase by these

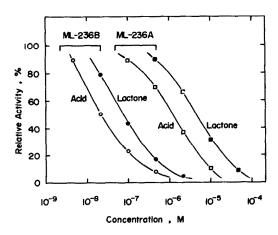


Fig. 2. Inhibition of HMG-CoA reductase by ML-236A and ML-236B. Experiments were carried out as described in Materials and methods. The results are expressed as % of control (without inhibitor). The value for control was 16.5 nmol/min/mg protein.

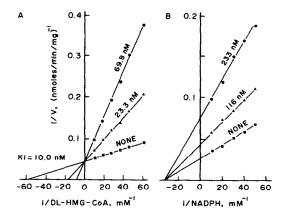


Fig. 3. Double reciprocal plots of the inhibition of HMG-CoA reductase by ML-236B sodium salt. Experiments were carried out as described in Materials and methods, except that concentrations of HMG-CoA (A) and NADPH (B) were varied as indicated.

compounds was competitive with respect to HMG-CoA and noncompetitive with respect to NADPH (fig.3). The  $K_i$  values were: ML-236A sodium salt 0.22  $\mu$ M, ML-236B sodium salt 0.010  $\mu$ M. Under these conditions,  $K_m$  values for the two substrates were: D,L-HMG-CoA 33  $\mu$ M, NADPH 40  $\mu$ M.

#### 4. Discussion

The α-methylbutyrate residue of ML-236B (fig.1) appears to play a significant role in the inhibition of HMG-CoA reductase activity, since both lactone and acid forms of ML-236A, lacking such a residue in their structure, are far less inhibitory than ML-236B analogs. The acid forms of both ML-236A and ML-236B contain a portion having a chemical structure very similar to that of 3-hydroxy-3-methylglutarate (fig.1). This is compatible with the fact that these compounds are competitive inhibitors of HMG-CoA reductase (competitive against HMG-CoA) and further that the acid forms are more potent than the corresponding lactone forms. The present results, considered together with those given in the previous paper [1], provide good evidence that a specific inhibitor of HMG-CoA reductase is effective in reducing cholesterol synthesis in vivo, and thereby in lowering cholesterol levels in blood.



Volume 72, number 2 FEBS LETTERS December 1976

## References

- [1] Endo, A., Kuroda, M. and Tsujita, Y. (1976) J. Antibiotics in the press.
- [2] Kuroda, M. and Endo, A. (1976) Biochim. Biophys. Acta 486, 70-81.
- [3] Heller, R. A. and Gould, R. G. (1973) Biochem. Biophys. Res. Commun. 50, 859-865.
- [4] Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J. (1951) J. Biol. Chem. 193, 265-275.

